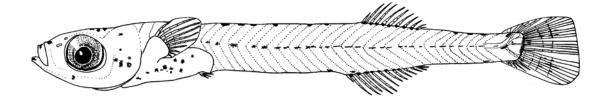
NOAA Technical Memorandum NMFS-SEFSC-481



PRELIMINARY GUIDE TO THE IDENTIFICATION OF THE EARLY LIFE HISTORY STAGES OF ATHERINID FISHES OF THE WESTERN CENTRAL NORTH ATLANTIC

 $\mathbf{B}\mathbf{Y}$

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ATHERINIDAE: True Silversides

The fishes of the family Atherinidae, commonly known as Old World or True silversides, were recently separated from the atherinopsids by Saeed et al. (1994). Dyer & Chernoff (1986b) confirmed this decision based on twenty diagnostic characters. In the western central Atlantic (WCA) area, there are only 3 species of this family (*Alepidomus evermanni, Hypoatherina harringtonensis* and *Atherinomorus stipes*) all pertaining to the subfamily Atherinomorinae (Chernoff, 2001).

Atherinids are small fishes <100 mm SL, slender, elongate, and moderately compressed posteriorly, with large eyes. They have an incomplete lateral line, broad silvery lateral band, and the pelvic fins are usually abdominal. The characters that separate this family from the atherinopsids are: premaxilla not protactile; distal end of the premaxilla not expanded; premaxilla with a postmaxillary process; anterior infraorbital sensory canal connected to preopercular canal, and two well-separated dorsal fins, the first with II to V spines (Chernoff, 2001).

Atherinids are small schooling fishes, associated with surface waters during all stages of life, and are extremely abundant in inshore regions, such as lagoons and estuaries. Most atherinids are marine, a few species are found around coral reefs, and other species are confined to freshwater. For example, 19 of the 24 species of the

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Craterocephalus are restricted to freshwater in Australia and New Guinea (Nelson, 1994).

The atherinids produce demersal eggs that are usually covered with adhesive filaments which attach to vegetation and debris. Newly hatched atherinid larvae (ca. 3 mm) are slender, with an extremely short gut, and pigmented eyes. A line of melanophores is present along the dorsal and lateral midline from head to the caudal region and dorsally on the gut. The only information on atherinid larvae is for one species of the Indo-Pacific coral reef, *Hypoatherina tropicalis* (Schmitt, 1983). This species has larval characters very similar to atherinopsid larvae.

In this guide, we include information on the atherinid larvae of *Atherinomorus stipes*. We identified and illustrated a larval series of *A. stipes* based on meristic and pigmentation characters. It is the most commonly caught atherinid in embayments and lagoons located in waters of Mexican Caribbean waters. A separate paper in preparation will provide a complete description of the early life history stages of this species (Vásquez-Yeomans & Valdéz-Moreno, in prep.). Tables Atherinidae 1 & 2 provide meristic and distribution data. This will aid in identifying the undescribed ELH stages of the two other species. However, care must be taken not to confuse these with the atherinopsids that are very similar.

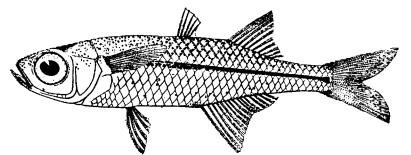
 Table Atherinidae 1. Meristic characters for the atherinid species found in the western cental North

 Atlantic. Counts were obtained from authors listed in Literature Cited.

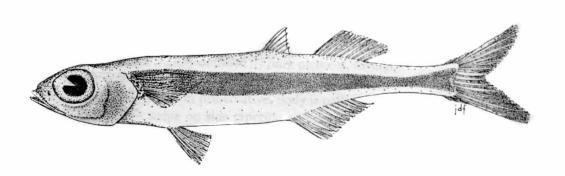
Species		Fin Rays	5			
	First	Second	Anal	Pectoral	Gill Rakers	Vertebrae
	Dorsal	Dorsal				
Alepidomus evermanni	V	I,9-11	I,12-15			
Atherinomorus stipes	IV-VI	I,8-10	I,11-13	14-16		37-40
Hypoatherina harringtonensis	V-VII	I,9-11	I,11-12	14-15	5-6+20-22	43-45

Table Atherinidae 2. Distribution & habitat of atherinids. Data from authors listed in Literature Cited.

Species	Geographical Distribution & Habitat	
Alepidomus evermanni	Pinar del Rio & San Cristobal, Cuba primarily in freshwater,	
	occasionally in estuaries or on coast near river mouths &	
	flooded mangroves.	
Atherinomorus stipes	From southern Florida to Brazil, including eastern & southern	
	Gulf of Mexico, Yucatan, Bahamas, and Caribbean Sea in coastal habitats over turtle grass beds and upper portions of coral reefs.	
Hypoatherina harringtonensis	From southern Florida, Bahamas, Bermuda, & the West Indies	
	including the southern Gulf of Mexico & Yucatan in coastal and offshore habitats, entering turtle grass beds in evening.	



Alepidomus evermanni (Eigenmann 1903). Original illustration (probably 45 mm adult).



Hypoatherina harringtonensis (Goode 1877). Illustration from Lavenberg & Chernoff (1995). Drawing done in Fishbase by Robbie Cada (size unknown).

MERISTICS

Vertebrae:	
Total	37-40
Number of Fin Spines and Rays:	
First Dorsal	IV-VI
Second Dorsal I,8-10	
Total 13-17	
Anal	I,11-13
Pectoral	14-16
Pelvic	
Caudal	
Principal	9+8
Gill Rakers 5-6+19-21	

LIFE HISTORY

Range: From southern Florida to Brazil, including eatern & southern Gulf of Mexico, Yucatan, Bahamas, & Caribbean.

Habitat: Shallow waters, freshwater, brackish, marine (reef associated).

ELH Pattern: oviparous, demersal eggs, planktonic larvae.

Spawning:

Season: Possibly year-around.

LITERATURE

Aguilera 1998; Avilés Torres A. 2002, Castro-Aguirre et al. 1999, Cervignon 1991, Cervignon et al. 1992, Chernoff 2001, Claro 1994, Eschmeyer 1996, Froese & Pauly 2001, McEAchran & Fechhelm 1998, Robins & Ray 1986, Schomburgk 1848, Smith 1997, Schmitter et al. 2000.

ILLUSTRATIONS

Original: Early preflexion larva, 3.8 mm, lateral and dorsal views; early flexion larva, 5.7 mm, lateral, dorsal and ventral views; postflexion larva, 11.7 mm, lateral view; late postflexion larva, 15.4 mm, lateral view, developing scales not shown. Larvae collected at Rio Huache, Quintana Roo, Mexico.

EARLY LIFE HISTORY DESCRIPTION

EGGS: Unknown

LARVAE:

Length at hatching: <3.8 mm

Length at flexion:5.5-6.7 through 8.3-9.0 mm

Length at Transformation: 15-20 mm

Sequence of fin development:C1, D2 & A, P1, C2, P2, D1:

- Pigmentation: Head: Initially 3 large melanophores over head (2 over midbrain & 1 over hindbrain) during preflexion stage; melanophores increase over head, 3 small on opercle, usually 1 small melanophore on posterior margin of dentary during flexion; 2 over froebrain,2 each at nostril, 2 on premaxillary, & 2 on anterior margin of dentary by 10.1 mm; increase in number over forebrain and midbrain, on snout and at dentary by 15.4 mm. Trunk: Dorsum with little pigment, 2-3 melanophores on dorsal margin of last myomere & over tipof notochord in larvae <4.5 mm; increase to 9-22 melanophores ondorsal midline, under notochord & on hypural margin during flexion stage; 2-3 melanophores along dorsal finbase inlate flexion stage increasing to 8-9 through postflexion stage; small melanophores anteriorly over dorsum at 12.6 mm, extending to posterior part of the tail >14.5 mm. Lateral midline unpirmented in larvae <4.3 mm; 10-19 melanophores during late preflexion stage; internal series above & below vertebral column on last vertebraeby 11.7 mm; series of melanophores above lateral midline posterior of tail by 12 mm increasing to anterior part of body through postflexion. Ventral pigment absent inlarvae <4.0 mm; 4-13 melanophores along ventral margin on tail during late preflexion stage; extending as double row from pelvic-fin bases to anal-fin base by 12 mm; increase number on anal-fin base in postflexion stage. Melanophores dorsally on gut & 3-5 in mid-gut through preflexion & flexion stages, incresing ventrally in postflexion stage.
- Diagnostic Characters: Myomeres 36-40 (usually 37-38; 3 melanophores on head (2 on midbrain, 1 hindbrain); 3 small melanophores onopercle inlate flexion stage; 1 small pigment spot on posterior part of dentary; 1-4 small melanophores over tip of notochord in larvae <4.0 mm, & below notochord inlarvae ~4.5-15.0 mm; dorsally on gut & 3-5 melanophores at mid-gut; scales appear at ~14 mm.

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EARLY LIFE HISTORY DESCRIPTION

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LARVAE:

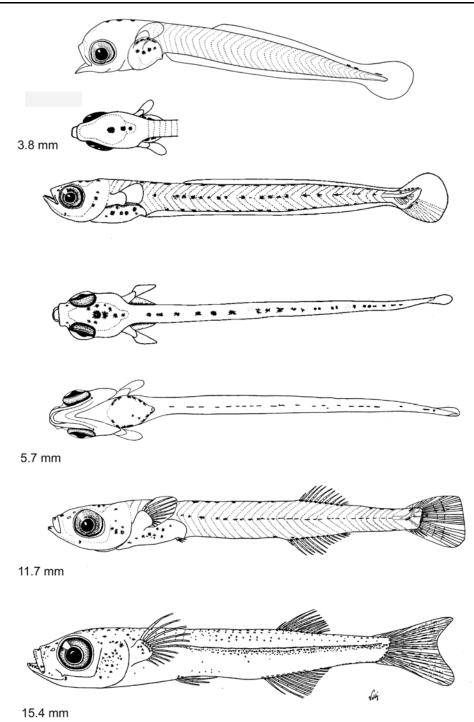
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