

Closing the genome of *Teredinibacter turnerae* T7902 by long-read nanopore sequencing

Mark T. Gasser,¹ Annie Liu,¹ Ron Flatau,² Marvin A. Altamia,² Claire Marie Filone,¹ Daniel L. Distel²

AUTHOR AFFILIATIONS See affiliation list on p. 3.

ABSTRACT We present the complete closed circular genome sequence derived from the Oxford Nanopore sequencing of the shipworm endosymbiont, *Teredinibacter turnerae* T7902 (DSM 15152, ATCC 39867), originally isolated from the shipworm, *Lyrodus pedicellatus* (1). This sequence will aid in the comparative genomics of shipworm endosymbionts and the understanding of the host–symbiont evolution.

KEYWORDS shipworm, symbiosis, endosymbionts, intracellular bacteria

Teredinibacter turnerae is a cellulolytic gammaproteobacterium (Cellvibrionaceae) that occurs as an intracellular endosymbiont of wood-boring bivalves (Teredinidae) (1–4) commonly known as shipworms. Strain T7902, representing *T. turnerae* clade II (5, 6), was isolated from the gills of the shipworm, *Lyrodus pedicellatus*, collected in Long Beach, CA in 1979 (1). Briefly, the gills were homogenized in shipworm basal medium (SBM) (7) and streaked on 0.9% agar SBM plates supplemented with 0.2% w/v powdered cellulose (SigmaCell Type 100; Sigma-Aldrich) and 5 mM NH₄Cl. Colonies were then serially re-streaked to obtain a clonal isolate. The original genome sequence of *T. turnerae* T7902 was published to GenBank (GCA_000379165.1) but was not described in peer-reviewed literature. This sequence was completed on 2012–05–22 at the DOE Joint Genome Institute under award 10.46936/10.25585/60001419 using 454 GS FLX Titanium and Illumina HiSeq 2000 sequencing platforms. It was assembled using Velvet v. 1.0.13 (8) and ALLPATHS v. R40295 (9), resulting in an improved high-quality draft assembly comprising 72 scaffolds with 76 contigs. As of 2024–05–10, eight fragmented genomes and one closed circular genome (T7901, Clade I, GenBank: GCA_000023025.1) were available for strains of *T. turnerae*. Here, we present the re-sequencing and completed genome of strain T7902 from nanopore-only sequencing (Table 1).

A colony of *T. turnerae* strain T7902 grown at 30°C on SBM (7) plates supplemented with 0.025% NH₄Cl and 0.2% cellulose (SigmaCell Type 101; Sigma-Aldrich) was used to inoculate a 6 mL liquid culture of SBM supplemented with 0.025% NH₄Cl and 0.2% carboxymethyl cellulose medium and grown at 30°C, 100 rpm for 4 days. Bacterial cells were harvested by centrifugation (10 min, 4°C, 4,000×g), and high-molecular-weight DNA was isolated from the cell pellet using the Wizard HMW DNA Extraction Kit (Promega, US) according to the manufacturer's protocol. The DNA quality and length were assessed on a TapeStation DNA Analyzer (Agilent Technologies, US). Nanopore sequencing (Oxford Nanopore Technologies, UK) was performed by ligation using the Nanopore Q20+ Chemistry Kit v14 according to the manufacturer's protocol and sequenced on a MinION instrument using an R10.4 flow cell (FLO-MIN112). Bases

TABLE 1 Assemblies of *Teredinibacter turnerae* T7902

| GenBank Assembly | Scaffolds (contigs) | Size (bp) | GC% | CDS |
|-----------------------------|---------------------|-----------|------|------|
| GCA_000379165.1 | 72 (76) | 5,387,817 | 50.8 | 4268 |
| GCA_037935975.1 (this work) | 1 (1) | 5,348,823 | 50.9 | 4212 |

Editor Irene L. G. Newton, Indiana University, Bloomington, Indiana, USA

Address correspondence to Daniel L. Distel, d.distel@northeastern.edu.

The authors declare no conflict of interest.

See the funding table on p. 3.

Received 10 May 2024

Accepted 6 November 2024

Published 10 December 2024

Copyright © 2024 Gasser et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

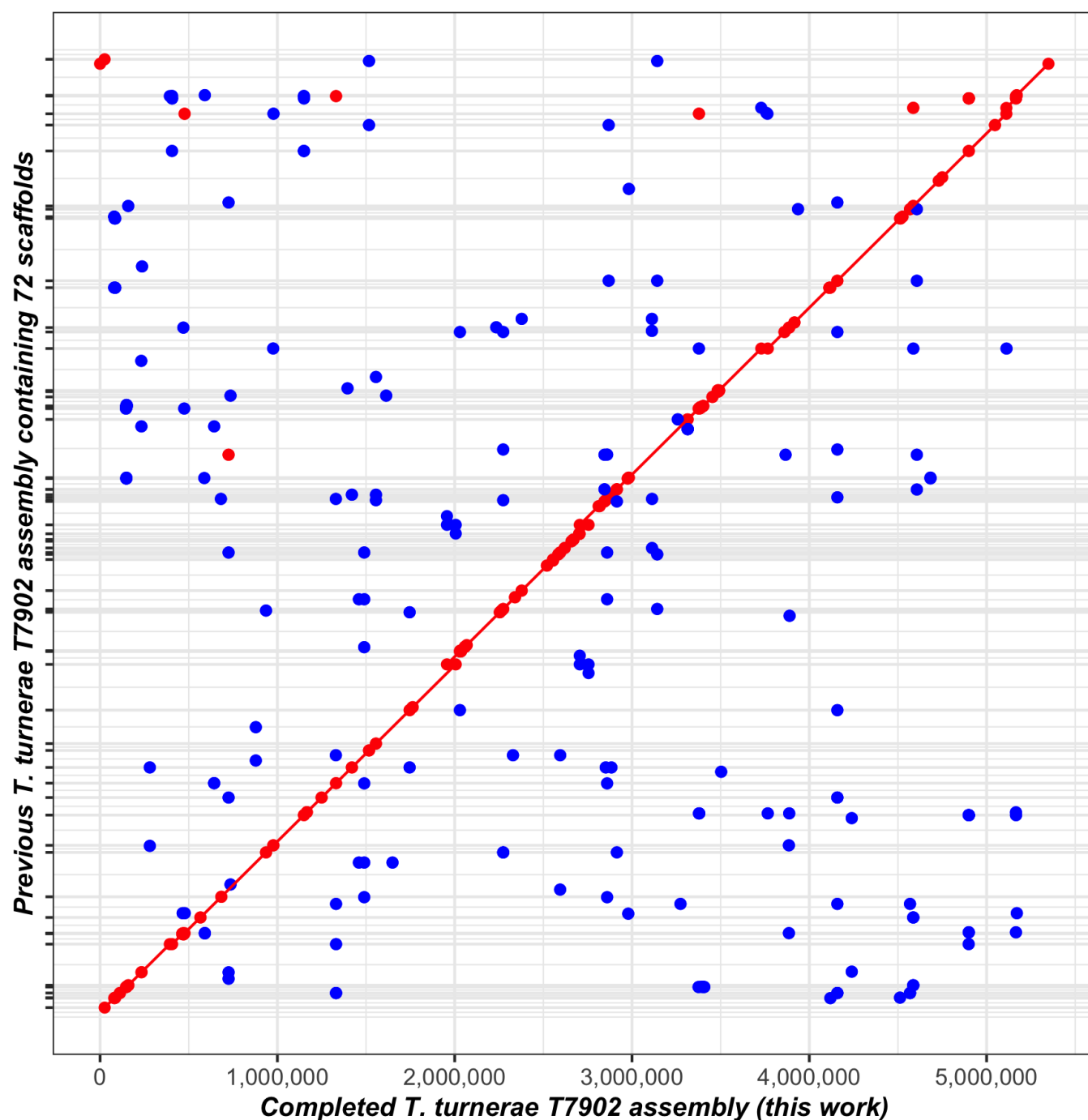


FIG 1 Synteny plot comparing the previously published genome of *Teredinibacter turnerae* T7902 (GCA_000379165.1) and the new genome sequence and assembly presented here (GCA_037935975.1). A MUMmer3 plot was generated with NUCmer v3.1 (15) using default settings to assess synteny and completion. Minimum exact matches of 20 bp are represented as a dot, with lines representing exact match lengths > 20 bp. Forward matches are displayed in red, while reverse matches are shown in blue.

were called using Guppy v6.5.7 with the super-accurate (SUP) algorithm and default-quality read filtering generating 300,309 reads (N_{50} = 8,763 bp). *De novo* assembly was performed with Flye v2.9.2 (<https://github.com/fenderglass/Flye>) (10), followed by contig correction and consensus generation with Medaka v1.8.0 (<https://github.com/nanoporetech/medaka>). To circularize, overlaps were identified and removed before the assembly was rotated to the gene predicted by Prodigal v2.6.3 (11) nearest the middle

of the contig with Circlator v1.5.5 (<https://github.com/sanger-pathogens/circlator>) (12). The resulting chromosomal assembly (278.0× coverage) was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (13). The new assembly shares 99.99% average nucleotide identity (14) and is highly syntenic (15) with the original (Fig. 1) but reduces the genome size by 38,994 to 5,348,823 bp, contains 56 fewer predicted CDS, and resolves several assembly errors. For all software, default parameters were used, except where otherwise noted.

ACKNOWLEDGMENTS

The research reported in this publication was supported by the following awards to DLD: National Oceanic and Atmospheric Administration (NA19OAR0110303), Gordon and Betty Moore Foundation (GBMF 9339), National Institutes of Health (1R01AI162943-01A1, subaward: 10062083-NE), and Johns Hopkins University Applied Physics Laboratory internal research and development funds to MTG. The National Science Foundation (DBI 1722553) also funded some equipment used in this research. The funders had no role in the study design, data collection and interpretation, or the decision to submit the work for publication.

AUTHOR AFFILIATIONS

¹Johns Hopkins University Applied Physics Laboratory, Laurel, Maryland, USA

²Ocean Genome Legacy Center, Northeastern University, Nahant, Massachusetts, USA

AUTHOR ORCIDs

Mark T. Gasser  <http://orcid.org/0000-0002-9396-4954>

Annie Liu  <http://orcid.org/0009-0002-3224-6008>

Ron Flatau  <http://orcid.org/0000-0002-4357-5870>

Marvin A. Altamia  <http://orcid.org/0000-0002-8625-767X>

Claire Marie Filone  <http://orcid.org/0000-0002-4041-3948>

Daniel L. Distel  <http://orcid.org/0000-0002-3860-194X>

FUNDING

| Funder | Grant(s) | Author(s) |
|--|-------------------------------|------------------|
| DOC National Oceanic and Atmospheric Administration (NOAA) | NA19OAR0110303 | Daniel L. Distel |
| Gordon and Betty Moore Foundation (GBMF) | GBMF 9339 | Daniel L. Distel |
| HHS National Institutes of Health (NIH) | 1R01AI162943-01A1,10062083-NE | Daniel L. Distel |
| National Science Foundation (NSF) | DBI 1722553 | Daniel L. Distel |

AUTHOR CONTRIBUTIONS

Mark T. Gasser, Conceptualization, Formal analysis, Writing – original draft, Writing – review and editing | Annie Liu, Methodology, Writing – original draft | Ron Flatau, Methodology, Writing – original draft | Marvin A. Altamia, Methodology, Writing – original draft | Claire Marie Filone, Funding acquisition, Supervision, Writing – review and editing | Daniel L. Distel, Funding acquisition, Supervision, Writing – original draft, Writing – review and editing

DATA AVAILABILITY

The complete genome sequence of T7902 has been deposited in GenBank under the accession number [CP149817](#). The Oxford Nanopore sequencing reads are available from the NCBI Sequence Read Archive (SRA) under the accession number [SRR28421272](#).

REFERENCES

1. Distel DL, Morrill W, MacLaren-Toussaint N, Franks D, Waterbury J. 2002. *Teredinibacter turnerae* gen. nov., sp. nov., a dinitrogen-fixing, cellulolytic, endosymbiotic gamma-proteobacterium isolated from the gills of wood-boring molluscs (Bivalvia: Teredinidae). *Int J Syst Evol Microbiol* 52:2261–2269. <https://doi.org/10.1099/00207713-52-6-2261>
2. Altamia MA, Shipway JR, Stein D, Betcher MA, Fung JM, Jospin G, Eisen J, Haygood MG, Distel DL. 2020. *Teredinibacter waterburyi* sp. nov., a marine, cellulolytic endosymbiotic bacterium isolated from the gills of the wood-boring mollusc *Bankia setacea* (Bivalvia: Teredinidae) and emended description of the genus *Teredinibacter*. *Int J Syst Evol Microbiol* 70:2388–2394. <https://doi.org/10.1099/ijsem.0.004049>
3. Altamia MA, Shipway JR, Stein D, Betcher MA, Fung JM, Jospin G, Eisen J, Haygood MG, Distel DL. 2021. *Teredinibacter haidensis* sp. nov., *Teredinibacter purpureus* sp. nov. and *Teredinibacter franksiae* sp. nov., marine, cellulolytic endosymbiotic bacteria isolated from the gills of the wood-boring mollusc *Bankia setacea* (Bivalvia: Teredinidae) and emended description of the genus *Teredinibacter*. *Int J Syst Evol Microbiol* 71:004627. <https://doi.org/10.1099/ijsem.0.004627>
4. Yang JC, Madupu R, Durkin AS, Ekborg NA, Pedamallu CS, Hostetler JB, Radune D, Toms BS, Henrissat B, Coutinho PM, et al. 2009. The complete genome of *Teredinibacter turnerae* T7901: an intracellular endosymbiont of marine wood-boring bivalves (shipworms). *PLoS ONE* 4:e6085. <https://doi.org/10.1371/journal.pone.0006085>
5. Altamia MA, Lin Z, Trindade-Silva AE, Uy ID, Shipway JR, Wilke DV, Concepcion GP, Distel DL, Schmidt EW, Haygood MG. 2020. Secondary metabolism in the gill microbiota of shipworms (Teredinidae) as revealed by comparison of metagenomes and nearly complete symbiont genomes. *mSystems* 5:e00261-20. <https://doi.org/10.1128/mSystems.00261-20>
6. Altamia MA, Wood N, Fung JM, Dedrick S, Linton EW, Concepcion GP, Haygood MG, Distel DL. 2014. Genetic differentiation among isolates of *Teredinibacter turnerae*, a widely occurring intracellular endosymbiont of shipworms. *Mol Ecol* 23:1418–1432. <https://doi.org/10.1111/mec.12667>
7. Waterbury JB, Calloway CB, Turner RD. 1983. A cellulolytic nitrogen-fixing bacterium cultured from the gland of deshayes in shipworms (Bivalvia: Teredinidae). *Science* 221:1401–1403. <https://doi.org/10.1126/science.221.4618.1401>
8. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <https://doi.org/10.1101/gr.074492.107>
9. Butler J, MacCallum I, Kleber M, Shlyakhter IA, Belmonte MK, Lander ES, Nusbaum C, Jaffe DB. 2008. ALLPATHS: de novo assembly of whole-genome shotgun microreads. *Genome Res* 18:810–820. <https://doi.org/10.1101/gr.7337908>
10. Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>
11. Hyatt D, Chen G-L, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <https://doi.org/10.1186/1471-2105-11-119>
12. Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. *Genome Biol* 16:294. <https://doi.org/10.1186/s13059-015-0849-0>
13. Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the prokaryotic genome annotation pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>
14. Rodriguez-R LM, Konstantinidis KT. 2016. The enveomics collection: a toolbox for specialized analyses of microbial genomes and metagenomes. *PeerJ*. <https://doi.org/10.7287/peerj.preprints.1900v1>
15. Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. *Genome Biol* 5:R12. <https://doi.org/10.1186/gb-2004-5-2-r12>