

# Author Manuscript

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1111/GCB.15362](https://doi.org/10.1111/GCB.15362)

This article is protected by copyright. All rights reserved

## Key rules of life and the fading cryosphere:

### impacts in alpine lakes and streams

Running title: *Rules of life and the cryosphere*

James J Elser, Flathead Lake Biological Station, University of Montana, Polson, Montana 59860, USA

Chenxi Wu, State Key Laboratory of Freshwater Ecology and Biotechnology, Institute of Hydrobiology, Chinese Academy of Sciences, Wuhan 430072, China

Angélica L González, Department of Biology & Center for Computational and Integrative Biology, Rutgers University, Camden, NJ 08102, USA

Daniel H Shain, Department of Biology & Center for Computational and Integrative Biology, Rutgers University, Camden, NJ 08102, USA

Heidi J Smith, Center for Biofilm Engineering, Montana State University, Bozeman, MT 59717, USA

Ruben Sommaruga, Department of Ecology, Lake and Glacier Research Group, University of Innsbruck, Innsbruck, Austria

Craig E Williamson, Department of Biology, Miami University, Oxford, Ohio 45056, USA

Janice Brahney, Department of Watershed Sciences, Utah State University, Logan UT 84322

Scott Hotaling, School of Biological Sciences, Washington State University, Pullman, WA 99164, USA

Joseph Vanderwall, Flathead Lake Biological Station, University of Montana, Polson, MT 59860, USA

Jinlei Yu, State Key Laboratory of Lake Science and Environment, Nanjing Institute of Geography and Limnology, Chinese Academy of Science, Nanjing 210008, China

Vladimir Aizen, Department of Geography, University of Idaho, Moscow, ID 83844, USA

Elena Aizen, Department of Geography, University of Idaho, Moscow, ID 83844, USA

28 Tom J Battin, Stream Biofilm and Ecosystem Research Laboratory, School of Architecture, Civil  
and Environmental Engineering, Ecole Polytechnique Fédérale Lausanne, Lausanne,  
30 Switzerland

Roberto Camassa, Department of Mathematics, Carolina Center for Interdisciplinary Applied  
32 Mathematics, University of North Carolina, USA

Xiu Feng, Key Laboratory of Aquatic Biodiversity and Conservation, Institute of Hydrobiology,  
34 Chinese Academy of Sciences, Wuhan 430072, China

Hongchen Jiang, State Key Lab of Biogeology and Environmental Geology, China University of  
36 Geosciences, Wuhan 430074, China

Lixin Lu, Cooperative Institute for Research in the Atmosphere, Colorado State University, Fort  
38 Collins, CO 80526, USA

John J Qu, Global Environment and Natural Resources Institute (GENRI) and Department of  
40 Geography and GeoInformation Science (GGS), George Mason University, Fairfax, VA  
22030, USA

42 Ze Ren, Flathead Lake Biological Station, University of Montana, Polson, Montana 59860, USA

Jun Wen, Sichuan Key Laboratory of Plateau Atmosphere and Environment, College of  
44 Atmospheric Sciences, Chengdu University of Information Technology, Chendu 610225,  
China

46 Lijuan Wen, Key Laboratory of Land Surface Process and Climate Change in Cold and Arid  
Region, Northwest Institute of Eco-Environment and Resources, Chinese Academy of  
48 Sciences, Lanzhou 730000, China

H. Arthur Woods, Division of Biological Sciences, University of Montana, Missoula, Montana  
50 59812, USA

*Xiong Xiong, State Key Laboratory of Freshwater Ecology and Biotechnology, Institute of*

52 *Hydrobiology, Chinese Academy of Sciences, Wuhan 430072, China*

*Jun Xu, State Key Laboratory of Freshwater Ecology and Biotechnology, Institute of*

54 *Hydrobiology, Chinese Academy of Sciences, Wuhan 430072, China*

*Gongliang Yu, Key Laboratory of Algal Biology, Institute of Hydrobiology, Chinese Academy of*

56 *Sciences, Wuhan 430072, China*

*Joel T Harper, Department of Geosciences, University of Montana, Missoula, Montana 59812,*

58 *USA*

*Jasmine E Saros, School of Biology and Ecology, Climate Change Institute, University of Maine,*

60 *Orono, Maine 04469, USA*

62

## Abstract

Alpine regions are changing rapidly due to loss of snow and ice in response to ongoing climate change. While studies have documented ecological responses in alpine lakes and streams to these changes, our ability to predict such outcomes is limited. We propose that the application of fundamental rules of life can help develop necessary predictive frameworks. We focus on four key rules of life and their interactions: the temperature dependence of biotic processes from enzymes to evolution; the wavelength-dependence of the effects of solar radiation on biological and ecological processes; the ramifications of the non-arbitrary elemental stoichiometry of life; and maximization of limiting resource use efficiency across scales. As the cryosphere melts and thaws, alpine lakes and streams will experience major changes in temperature regimes, absolute and relative inputs of solar radiation in ultraviolet and photosynthetically active radiation, and relative supplies of resources (e.g., carbon, nitrogen, and phosphorus), leading to nonlinear and interactive effects on particular biota, as well as on community and ecosystem properties. We propose that applying these key rules of life to cryosphere-influenced ecosystems will reduce uncertainties about the impacts of global change and help develop an integrated global view of rapidly changing alpine environments. However, doing so will require intensive interdisciplinary collaboration and international cooperation. More broadly, the alpine cryosphere is an example of a system where improving our understanding of mechanistic underpinnings of living systems might transform our ability to predict and mitigate the impacts of ongoing global change across the daunting scope of diversity in Earth's biota and environments.

Keywords: rules of life; cryosphere; lakes; streams, glaciers;

## 1. Introduction

General rules govern life on Earth. At its core, life replicates and evolves, even in challenging, ice- and snow-covered mountain environments that harbor communities uniquely shaped by their extreme nature. However, rapid environmental change is challenging the persistence of these living systems. Among Earth system components, the cryosphere is undergoing some of the most rapid changes, driven largely by anthropogenic impacts (WGMS, 2017). These changes are predicted to be particularly acute in mountain regions (Wang et al., 2016), where permafrost, permanent ice, and snow are rapidly receding, spurring widespread environmental changes (Milner et al., 2017; Shugar et al., 2017; Huss & Hock, 2018). Indeed, this retreat is accelerating commensurate with climate warming during recent decades (WGMS, 2017; Maurer et al., 2019). Crucial changes in snowpack are also occurring, for example, in mountainous areas of the western USA where snowmelt is responsible for 70% of total runoff (Li et al., 2017). Thus, cryosphere-influenced ecosystems and biota are now under threat. Understanding and predicting the impacts of these changes on alpine aquatic ecosystems and their ecological communities poses an urgent challenge for ecologists and environmental scientists (Hotelling et al., 2017a; Moser et al., 2019) because alpine aquatic ecosystems provide key services upon which human well-being depends, especially for the inhabitants of mountain regions (Viviroli et al., 2007).

To move beyond merely documenting ecological changes into making predictions requires a grasp of the fundamental mechanisms that drive biological and ecological systems. These mechanisms involve essential “rules of life” that pertain to all or most living things. Here we highlight four potential rules of life that we propose are especially important in understanding how alpine aquatic ecosystems respond to a rapidly changing cryosphere. These key rules are:

temperature dependence of metabolism (Brown et al., 2004); the spectral dependence of biotic responses to solar radiation (Williamson & Rose, 2010); biological stoichiometry (Elser et al., 2000); and resource use efficiency (Hodapp et al., 2019). We describe these key rules and propose them as a framework for understanding and predicting future changes in ecological structure and function in cryosphere-influenced lakes and streams (see Figure 1). Our emphasis will be on the influence of glaciers and snowpack; alpine permafrost is an important component but beyond the scope of this article. Achieving a better grasp on fundamental properties of living systems will greatly enhance our ability not only to understand the impacts of these changes in the cryosphere but also to forecast their consequences.

## **2. Key rules of life and ongoing shifts in alpine lakes and streams**

*2.1 What is a “rule of life”?* We propose that a rule of life describes a major mechanism, phenomenon, or process that structures most or all of life on Earth. Such rules generally hinge on inescapable aspects of our physicochemical world and can produce emergent structural features of interacting networks of biological units at all levels of biological organization. Here we delineate four such rules that, while relevant in all environments, are especially germane for alpine lakes and streams. We recognize that a variety of other relevant candidate rules could be proposed, including the ubiquity of Darwinian evolution as mentioned above, and may also be of relevance here. However, we contend that the four rules we discuss are especially useful in understanding and predicting the dynamics of alpine lakes and streams in a changing climate given the well-known effects of ice and snow on temperature, light environments, nutrient regimes, and habitat succession.

### *2.2 Four key rules*

We propose four rules of life necessary for obtaining a fundamental and thus predictive

understanding of how aquatic biota and ecosystems in alpine environments will respond to a changing cryosphere under ongoing climate change.

*Key rule 1: Temperature.* Temperature has a fundamental effect on nearly all biological activities due to the underlying physics of biochemical processes (red arrows in Figure 1, center).

*Key rule 2: Wavelength dependence.* Biological systems are differentially affected by photosynthetically active radiation (PAR) and ultraviolet radiation (UVR) from molecules to ecosystems (Figure 1, top).

*Key rule 3: Biological stoichiometry.* Earth's species comprise a non-random assemblage of chemical elements that reflects their evolved life histories and shapes their distribution and dynamics (Figure 1, gray bars).

*Key rule 4: Efficiency.* At different levels of organization, biological systems maximize use efficiency of limiting resources.

#### 2.2.1 *Key rule 1: Temperature.*

Metabolic theory, based on over a century of theoretical and empirical study, describes rate process kinematics via the Boltzmann relationship:

$$R = B_0 M^\alpha e^{(-E/kT)}$$

This function applies to nearly all critical biological functions and represents a rule of life to which all organisms adhere. In this formulation,  $R$  is a biological rate process,  $B_0$  is a normalization constant that is independent of body size and temperature,  $M$  is body mass,  $\alpha$  is an allometric scaling exponent, and  $e^{-E/kT}$  is the exponential Arrhenius function, where  $E$  is the activation energy,  $k$  is the Boltzmann constant, and  $T$  is temperature in Kelvin (Brown et al., 2004; West & Brown, 2005). This relationship implies that, across a moderate range below the



point at which temperature becomes stressful, increasing temperature increases enzyme activity, metabolic rate, and all biological rate processes that depend on metabolism, including growth. Via these mechanisms, higher temperatures can drive generation times and even rates of evolution (Allen et al. 2006). However, as discussed below, such general large-scale trends may not apply across narrower ranges of temperature and especially at low temperatures close to the freezing point of water.

At low temperatures, organisms face unique challenges related to molecular flexibility, metabolism, and increased viscosity. To circumvent these bottlenecks, evolutionary trends are observed that increase molecular flexibility (D'Amico et al., 2006) and loosen molecular bonds (Siddiqui & Cavicchioli, 2006), while increasing readily available energy (i.e., ATP supply) (Napolitano & Shain, 2005) to compensate for reduced molecular motion and rates of diffusion. Although these responses are well-suited to low temperatures, as the cryosphere in mountain regions disappears, temperatures will rise and aquatic thermal regimes will increasingly be coupled to air temperature, with reduced buffering from glacier and perennial snowfield melt (Brown et al., 2006). While key habitats for ice-dependent life (Anesio & Laybourn-Parry, 2012) will be lost, downstream aquatic habitats may also undergo dramatic reorganizations leading to changes in temperature as well as the creation or loss of lakes and reorganization of stream channels (Milner & Bailey, 1989; Clague & Evans, 2000; Brahney et al., 2010; Shugar et al., 2017).

Under future temperature regimes, costly adaptations to cold will become useless and could lead to widespread extinction unless evolutionary responses are rapid. Specifically, cold-adapted enzymes, which typically evolve higher flexibility via weaker intramolecular interactions, may denature more readily in novel, warmer environments (Siddiqui & Cavicchioli, 2006).

Likewise, likewise, cold-adapted membranes may fail at modestly high temperatures, with  
186 severe disruption to ion and energy homeostasis. The extreme sensitivity of such biota is  
exemplified by several species of snow algae. These key players in biogeochemical cycling and  
188 food webs in snow habitats display narrow optimal growth temperatures ranging between 0°C  
and 4°C (Hoham et al., 2008). Similarly, freshwater alpine stream invertebrates, including  
190 species of Chironomidae, Ephemeroptera, and Trichoptera, have optimal temperatures between  
1.2°C and 10.6°C (Niedrist & Füreder, 2020). Taken together, these features of organismal  
192 adaptation to low temperature suggest that even subtle increases in temperature above the  
freezing point of water in cryosphere-influenced aquatic ecosystems will likely lead to an overall  
194 decline in diversity of cold-adapted biota as they are thrust into thermal environments for which  
they are ill-suited. However, recent evidence from studies of glacier-associated insects provides  
196 a caveat. Some cold-adapted aquatic taxa can persist despite deglaciation, at least on  
contemporary time scales (Muhlfeld et al., 2020), with access to persistent low-temperature  
198 habitats associated with snowpack or cold seeps and springs. Furthermore, recent studies  
indicate that some cold-adapted insects have relatively broad temperature performance optima  
200 (Pallarés et al., 2020; Treanor et al., 2013). This work highlights the need to examine even basic  
assumptions about the impacts of cryosphere loss on associated biodiversity and that other key  
202 rules beyond temperature dependence may play an important role.

204 Despite the predicted losses of cold-adapted taxa, a number of biological rates are likely to  
increase with increasing temperatures as the cryosphere declines, including productivity and  
206 evolutionary rate given the fundamental temperature-dependence of rate processes (Rule 1).  
Furthermore, newly created habitats (e.g., meltwater lakes) as well as formerly ice- and snow-  
208 influenced streams and lakes will be invaded by more temperate-adapted species (Shah et al.,  
2020), dramatically changing community composition (Tiberti et al., 2019) and function.

Ultimately, this will homogenize biodiversity at the level of the alpine landscape (i.e., decrease beta diversity), as has been shown for both insects and microbes in glacier-fed streams (Jacobsen et al., 2012; Wilhelm et al., 2013; Hotaling et al., 2019). Finally, organisms living in the cryosphere itself may themselves contribute to feedbacks that will accelerate the decline of cryosphere environments. One example is decreased albedo triggered by warming-induced increases in biomass of organisms such as snow algae, which populate ice and snow at high altitudes and latitudes (Takeuchi et al., 2006; Lutz et al., 2016).

One challenge for future research is to increase our understanding of thermal performance curves, including lower and upper temperature thresholds, for a variety of cold-adapted taxa. Similarly, data on thermal acclimation responses is lacking for the vast majority of cold-adapted species. However, this information is critical for predicting organismal responses to changes in their climatic niches, via adaptation or through changes in their distributions, in the face of a melting cryosphere. Such efforts would also be greatly advanced by improving our assessments of whole-genomes of cold-adapted biota as well as metagenomic and metatranscriptomic properties of microbial communities in alpine environments across a broad range of temperature regimes.

#### *2.2.2 Key rule 2: Wavelength dependence.*

Rule 2 states simply that wavelength matters. At a molecular level, photosynthetically active radiation (PAR; wavelengths from 400 - 700 nm) drives photosynthesis while ultraviolet radiation (UVR; solar wavelengths from 290-400 nm) can damage DNA and other cell constituents. At the ecosystem level, both UVR and PAR underlie basal resource availability. For example, PAR drives primary production whereas UVR can transform refractory organic matter (autochthonous

or terrestrially-derived) into energy resources through photodegradation, fueling ecosystem

236 respiration. PAR and UVR also enable many organisms to orient visually to their environments,  
enabling them to forage, avoid predators, and engage in reproductive behaviors (Williamson &  
238 Rose, 2010; Williamson et al., 2014, 2019).

240 As terrestrial landscapes in mountains transition from predominantly snow and ice cover to  
vegetation, wavelength-dependent effects of sunlight in lakes and streams also are changing  
242 (Cannone et al., 2008; Figure 2). Reductions in the duration of snow and ice cover on lakes and  
streams (Benson et al., 2012) cause strong increases in exposure to sunlight. The shortest and  
244 most damaging wavelengths of UVR increase with elevation much more rapidly than does PAR,  
increasing by 20% or more per 1,000 m (Blumthaler et al., 1997). In glacierized regions,  
246 scouring leads to lakes and streams with high turbidity that greatly reduces PAR as well as UVR  
(Sommaruga, 2015). Thus, in places where glacial turbidity has subsided, the resulting clear-  
248 water lakes and streams at high elevations are among the most UVR-exposed environments in  
the world (Figure 2) (Sommaruga, 2001; Rose et al., 2009). Extremely high incident UVR:PAR  
250 ratios in alpine and subalpine ecosystems lead to photoinhibition of photosynthesis and  
photodamage of DNA (Cooke et al., 2006; Williamson et al., 2010). Over many decades, UVR  
252 penetration in lakes and streams declines due to higher concentrations of DOM from increases  
in terrestrial vegetation (Figure 2) and lakes shift to low UVR:PAR conditions that favor primary  
254 production (Engstrom et al., 2000; Williamson et al., 2001b).

256 As remote as many high-elevation lakes and streams are, they can be highly susceptible to  
contamination from the development of more human-dominated lower-elevation landscapes due  
258 to a combination of high UVR exposure levels and a phenomenon known as the alpine cold  
distillery. The mechanism involved here is that precipitation-borne contaminants become

increasingly concentrated in high elevation ice and snow due to sublimation and low temperatures that slow contaminant degradation (Daly & Wania, 2005). This concentrating effect can lead to increased levels of contaminants such as polycyclic aromatic hydrocarbons, the toxicity of which is amplified through phototoxicity by exposure to high UVR (Larson & Berenbaum, 1988). As terrestrial vegetation increases over the longer term with climate warming, UVR-absorbing DOM inputs reduce UVR:PAR ratios, photodegradation of organic matter, photoinhibition of photosynthesis, phototoxicity, and photodamage of DNA and other cellular components (Figure 2) (Williamson et al., 2001a, b).

Ultimately, the loss of high UVR:PAR ecosystems as DOM increases with increasing terrestrial vegetation in the catchment (Figure 2b) will likely lead to the loss of unique species that are adapted to these extreme conditions. For example, aquatic invertebrate predators with low UVR tolerance can expand into regions where higher DOM provides a refuge from UVR (Lindholm et al., 2016). In many alpine lakes with high UVR exposure, only one species of highly pigmented crustacean zooplankton species exists (Marinone et al., 2006). Lakes just above treeline are most sensitive to these increases in DOM and reduced UVR (Vinebrooke & Leavitt, 2005).

Several critical knowledge gaps provide a compelling need for further research to understand how changes in the spectral composition of sunlight, and the UVR:PAR ratio in particular will play an important role in a fading cryosphere. Most urgent is the need for more data on the UVR and PAR exposure levels and transparency of high elevation lakes and streams because current UVR transparency data in particular are very sparse. Second is the need for more data on the UVR tolerance levels of both indigenous high elevation as well as potentially invasive lowland species. Third, high elevation ecosystems are nutrient poor and cold at the same time that they are exposed to high levels of UVR, highlighting the need to understand the shifting

roles of temperature vs. nutrient vs. light limitation vs. photodamage of aquatic organisms

across trophic levels as alpine regions warm. Fundamental shifts in the structure and function of these unique aquatic ecosystems are likely due to differential responses to these drivers across trophic levels in the warming alpine cryosphere.

### 2.2.3 Key rule 3: Biological stoichiometry.

Due to a shared evolutionary history, organisms have a common biochemistry constructed from a limited suite of chemical elements (Da Silva & Williams, 2001). However, they can differ considerably in the proportion of these elements within and among species (Fagan et al., 2002; Sterner & Elser, 2002; Woods et al., 2003; Gonzalez et al., 2011, 2018). These stoichiometric differences ultimately reflect the evolution of diverse life history strategies and body plans (Elser et al., 2000; Sterner & Elser, 2002; Kay et al., 2005). For example, fast-growing biota tend to have low C:P and N:P ratios due to increased allocation to P-rich ribosomal RNA (Elser et al., 2003). Further, nutrient supplies vary both spatially and temporally, particularly in the mountain cryosphere (Ren et al., 2019), and are often not present in a balanced supply in relation to biotic demands, inducing nutrient limitation that drives local diversity, community assembly, and ecosystem dynamics. Stoichiometric plasticity and stoichiometric adaptations are key mechanisms by which organisms can respond to such challenges in the biogeochemical environment (Quigg et al., 2003, 2011; Litchman et al., 2012). However, evolutionary constraints acting upon the basic biochemical machinery of living organisms may also impose limits on those stoichiometric responses (Zimmerman et al., 2014).

Taking a stoichiometric perspective on the living things that inhabit alpine lakes and streams may be of particular importance in considering the impacts of cryosphere change because glacial meltwaters can be geochemically distinct from other source waters. For example,

depending on catchment geology, concentrations of available forms of P can increase, while concentrations of dissolved inorganic and organic nitrogen decrease with increasing catchment glacial coverage (Tockner et al., 2002; Hood et al., 2009). This pattern emerges even when bedrock sources of P are low, potentially owing to the atmospheric deposition of dust (Vandeberg & Vanlooy, 2016). Impacts of glaciers on nitrogen supplies are less clear. In the central Rockies of North America, glacial meltwaters are enriched in nitrate (Wynn et al., 2007; Saros et al., 2010), while studies in other regions have reported variable relationships of N to glacier cover (Tranter et al., 1994; Hood & Scott, 2008). The mechanisms underpinning regional differences are uncertain, but may relate to patterns of atmospheric deposition or microbial activity in glacial sub-habitats (Hamilton et al., 2013; Fegelman et al., 2016; Hotelling et al., 2017b).

Less is known about how shifts in snowpack regimes affect nutrient supplies to alpine lakes and streams (Parker et al., 2008; Green, 2012). This is unfortunate as shifts from permanent to seasonal snowpack and from seasonal snow to rain-dominated precipitation are likely to have major effects on catchment biogeochemistry. However, we might expect the loss of permanent snowpack as the snowline increases in altitude (Marty et al., 2017) and an increased mobilization of accumulated nutrients in that snow. In addition, weathering of newly exposed bedrock, which releases nutrients, should increase through both biological and physical mechanisms. The relative balance of N and P in that meltwater will likely depend on regional differences in the balance of dust and atmospheric N deposition that has accumulated (Brahney et al., 2015). This requires further research.

Changes in nutrient regimes due to cryosphere loss are expected to affect ecological dynamics in alpine aquatic ecosystems. For example, increases in N:P ratios due to disproportionate runoff of inorganic N from melting glaciers (Saros et al., 2010; Slemmons & Saros, 2012;

Colombo et al., 2019; Figure 3, left), will likely induce P limitation and negatively affect fast-growing, P-rich organisms (Figure 3, right). The effects of nutrient limitation may be particularly severe given the short growing season of alpine environments, which impose strong selection pressures for rapid growth and development. Nevertheless, glacial melting is expected to lengthen the growing season (Fell et al., 2017), which could ultimately benefit slow-growing species and disadvantage fast-growing organisms with high P requirements, especially in low P supply environments (Elser et al., 2000; Kay et al., 2005).

A variety of research opportunities exist to gain a better understanding of stoichiometric dimensions of cryosphere change in mountain regions. For example, as discussed above, some emerging data indicate strong differences in N:P ratios in runoff from glaciers vs snowpack, but regional differences exist. Thus, to develop a clearer picture we need more data about N and P concentrations and ratios from a wider range of glacial environments. We also need more biogeochemical data from other components of the cryosphere, such as rock glaciers and alpine permafrost. Another opportunity would be to investigate nutrients other than N and P, such as changes in the availability of Fe and Ni due to glacial melt. Finally, a key challenge for future research is to understand whether cold-adapted organisms have different stoichiometric niches compared to related species from warmer habitats.

#### 2.2.4 Key rule 4: Resource use efficiency.

Rule 4 states that, at different levels of organization, biological systems maximize the use efficiency of limiting resources. *Resource use efficiency* (RUE) relates to the fraction of supplied or available resources converted into new biomass and thus links potential and real productivity (Hodapp et al., 2019) with major implications for biogeochemical fluxes. Maximizing RUE of a



limiting resource is relevant from cells to ecosystems, although it will act at different temporal scales (Hodapp et al., 2019). In resource competition at the community scale (Tilman 1982, Grover 1997), cellular mechanisms contribute to increased RUE via the minimal cell quota of the best competitors, which is the cellular content of a limiting resource required to achieve zero net growth rate. Other examples of mechanisms that maximize use efficiency of limiting nutrients such as phosphorus are known for different levels of organization (Sterner and Elser 2002). For example, at the molecular level, organisms can replace phospholipids with sulfolipids under P limitation (Elser et al., 2010). At the community level, low-P taxa (e.g., *Bosmina*) can replace high-P species (e.g., *Daphnia*) when ecosystem P limitation results in seston with high C:P ratio (Demott and Gulati, 1999).

Resource use efficiency is also strongly linked to that of biodiversity. For example, according to the biodiversity-ecosystem functioning hypothesis (Loreau et al., 2001), higher taxonomic diversity is expected to increase RUE (Hooper et al., 2005). However, considering species functional diversity (i.e., traits) is also important in this relationship because RUE can also decrease with increasing taxonomic diversity when functional diversity does not increase concomitantly (Hodapp et al., 2019). Functional redundancy, which describes the ability of several taxa to perform the same function, may provide insurance against alteration in ecosystem functioning upon changes in community composition or loss of species (Miki et al., 2013). Indeed, large changes in biodiversity have been observed in alpine streams and lakes when they shift from their turbid to clear state (Jacobsen et al., 2012; Peter and Sommaruga, 2016; Cauvy-Fraunié & Dangles, 2019) but we do not know how those changes affect ecosystem functioning. In any case, it is expected that the factors discussed above (i.e., temperature, light, and stoichiometry) will predictably influence succession in ecosystem diversity, functionality and productivity within alpine ecosystems as the influence of the

cryosphere declines. These changes will reflect broad shifts in limiting resources and thus in RUE across space and time as alpine aquatic ecosystems and especially lakes experience large changes in light/UVR and nutrient conditions (Figure 4). For example, the magnitude and frequency of environmental changes or disturbance caused by the discharge of glacial meltwaters will likely drive RUE as community composition and diversity change during the ecosystem's ontogeny. This contention is based on previous studies showing that relief of P limitation increases the abundance of small-bodied consumers in communities (Mulder & Elser, 2009; Ott et al., 2014; Jochum et al., 2017) and enhances nutrient transfer efficiency from small to larger organisms. Thus, a converse situation in which cryosphere loss enhances P limitation (see discussion of Rule 3) should favor large-bodied organisms and decrease carbon, but increase P transfer efficiencies through food webs.

Considering the four rules of life discussed, Rule 4 is perhaps where we most lack empirical evidence. Thus, future research should include studies on how RUE changes during ecosystem ontogeny as the cryosphere vanishes. Further, we urgently need to understand how changes in biodiversity of alpine aquatic ecosystems affect ecosystem function as this link is intrinsically related to RUE. One of the few assessments of this link in lakes affected by a vanishing glacier showed no evidence for multifunctional redundancy in bacterial communities (Peter & Sommaruga, 2016). However, no information is available for other communities.

### *2.3 Interactions among the key rules*

The rules just described do not operate independently and indeed in alpine ecosystems such interactions may be critical as lakes and streams experience cryosphere change. For example, temperature acclimatization (Rule 1) can affect biomass stoichiometry (Rule 3) (Woods et al., 2003; Cross et al., 2015; Yvon-Durocher et al., 2015; Phillips et al., 2017; Yvon-Durocher et al.,

2017). Organismal N and P contents can increase and N:P ratios decrease at lower temperatures due to higher allocation to ribosomes under lower temperatures to compensate for decreased reaction rates (Farewell & Neidhardt, 1998; Woods et al., 2003; Toseland et al., 2013; Daines et al., 2014; Yvon-Durocher et al., 2015, 2017). Stoichiometry (Rule 3) and UVR (Rule 2) also interact. Increased UVR exposure following deglaciation and the loss of ice and snow cover leads to photoinhibition of photosynthesis that may decrease food quantity for grazers, but higher UV may simultaneously lower C:P ratios of primary producers and thus improve food quality (Xenopoulos et al., 2002; Sommaruga, 2015; Stenzel et al., 2017).

Interactions among these key rules extend beyond stoichiometric dimensions. For example, there are also important interactions between temperature (Rule 1) and UVR (Rule 2). Springtime increases in water temperature generally lag increases in UVR exposure (Figure 5). Exposure to high UVR at cold temperatures decreases the potential for molecular repair of DNA damage due to the temperature dependence of repair enzymes (Macfadyen et al., 2004). The combination of higher UVR exposure and earlier ice-out, often timed near summer solstice, the annual peak in sunlight, can lead to very high net UVR damage potential (Figure 3). Note that the patterns we discuss encompass alpine regions in temperate latitudes. Seasonality in alpine temperature and UVR exposure will be dampened in the tropics, but amplified towards the poles. The inhibitory effects of UVR on photosynthetic biota (Rule 2) are moderated at higher temperatures (Rule 1) because DNA repair processes are performed by temperature-dependent enzymes (Beardall et al., 2014). Meanwhile, temperature - PAR interactions may arise in well-illuminated situations because of the well-known temperature dependence of Rubisco activity. Thus, higher temperatures may allow increases in maximum photosynthetic capacity (Davison, 1991; Galmes et al., 2015). An important open question, however, is how these interacting

processes ultimately influence emergent resource use efficiency (Rule 4) in communities and ecosystems under cryosphere change.

### **3. Conclusions: Meeting the scientific challenges of the changing alpine cryosphere**

Existing studies have documented the impacts of cryosphere change on the ecology of alpine lakes and streams (see various citations throughout). While important, such efforts do not necessarily provide the ability to forecast future impacts of cryosphere change, especially across diverse environments in the world's mountain regions. Better forecasts across broad spatial scales will require us to understand the fundamental mechanisms that underpin ecological change. These mechanisms likely arise from core biological properties of living things - rules of life. In this paper, we have applied four key rules of life to better understand ongoing ecological changes in cryosphere-influenced aquatic ecosystems in alpine environments (Figure 1) and to predict future transitions.

Achieving better insights is important because ongoing changes in the cryosphere have many impacts on ecosystem services that are valuable in mountain regions (Viviroli et al., 2007; Klein et al., 2019). Mitigating these impacts, or adapting to them if mitigation is impossible, is complicated by considerable regional variability and temporal unpredictability of these impacts. For example, the mechanisms behind variation in the provisioning of nutrients (N, P) from glacial systems are not well understood, but likely involve differences in regional geology and land-use, climate, land-atmosphere interactions, microbial activity, and glacial drainage patterns. Importantly, we need to document not only ongoing ecological responses to such shifts, but also to predict those outcomes based on mechanistic understanding. For example, as delineated earlier, while exposure to DNA-damaging UVR increases with increasing elevation, air and water temperatures decrease, reducing the effectiveness of photoenzymatic repair of DNA damage (as demonstrated in zooplankton; Cooke et al., 2006). The inability to repair UVR-

induced DNA damage will in turn require other adaptive responses such as high levels of photoprotective compounds or behavioral avoidance of UVR exposure through diel vertical migration, responses that may vary among taxa of zooplankton (Ekvall et al., 2015; Tartarotti et al., 2017) with important implications for other ecological interactions such as visual predation (Hylander et al., 2009). Likewise, understanding fundamental temperature responses of metabolism and performance, especially at low temperatures as water warms above its freezing point, will permit prediction of biotic responses to forecasted changes in lake and stream temperatures (Jones et al. 2017) as glaciers and snowpack recede in the alpine landscape.

To develop such a predictive, mechanistic understanding and to obtain broadly applicable insights into unfolding global patterns of cryosphere change and impact on alpine lakes and streams, multilateral international cooperation is needed so that studies can be integrated across regions. For example, the vast mountain cryosphere domain of central Asia remains relatively understudied from an ecological perspective and should be a focus of increasing international attention. Furthermore, interdisciplinary integration across atmospheric, earth, and life sciences is essential. Only with such integration can the most ecologically relevant geophysical changes be identified and tracked and their impacts predicted from a foundation of fundamental biological knowledge.

**Acknowledgements:** None of the authors have any real or perceived conflicts of interest relevant to this article. This paper is a product of a US-China workshop “Rules of Life: A Fading Cryosphere Shifting Temperature and Stoichiometry in Mountain Lakes and Streams” held at the Flathead Lake Biological Station (University of Montana) supported by the US National Science Foundation (DMS-1834494) and the Natural National Science Foundation of China (41981220290). CEW acknowledges support from NSF DEB-1754276 and RS from the Austrian

Science Fund (FWF P24442-B25). SH was supported by NSF OPP-1906015. We thank M.

Skidmore (Montana State University), R. McLaughlin (North Carolina State University), and C. Florentine (USGS) for stimulating presentations and discussions during the initial days of the workshop, and Z. Dong (Northwest Institute of Eco-Environment and Resources, Chinese Academy of Sciences), J. Li (Northwest Institute of Eco-Environment and Resources, Chinese Academy of Sciences), W. Tan (Central China Normal University), and Z. Song (The National Natural Science Foundation of China) for sharing ideas and findings during the workshop. We are also grateful to the staff of the Flathead Lake Biological Station for their hospitality. C. Muhlfeld and an anonymous reviewer provided useful comments on an earlier draft.

**Author Contributions.** Contributions are as follows. Workshop conceptualization: JJE, CW, JTH, JES. Funding Acquisition: JJE, CW. Project administration: JJE, CW. Supervision: JJE, CW. Visualization: JJE, CEW, RS, HJS, JB, SH, ALG, JY. Writing – Original Draft: JJE, CW, ALG, DHS, HJS, RS, CEW. Writing – review and editing: JJE, CW, ALG, DHS, HJS, RS, CEW, JB, SH, JV, JY, VA, EA, TJB, RC, XF, HJ, LL, JJQ, ZR, JW, LW, HAW, XX, JX, GY, JTH, JES.

**Data Sharing and Data Accessibility.** No original data are included in this paper.

## References

- Allen, A. P., Gillooly, J. F., Savage, V. M. & Brown, J. H. (2006). Kinetic effects of temperature on rates of genetic divergence and speciation. *Proceedings of the National Academy of Sciences of the United States of America*, 103, 9130-9135.
- Anesio, A. M. & Laybourn-Parry, J. (2012). Glaciers and ice sheets as a biome. *Trends in Ecology & Evolution*, 27, 219-225.

- 510 Beardall, J., Stojkovic, S. & Gao, K. (2014). Interactive effects of nutrient supply and other  
environmental factors on the sensitivity of marine primary producers to ultraviolet radiation:  
512 implications for the impacts of global change. *Aquatic Biology*, 22, 5-23.
- Benson, B. J., Magnuson, J. J., Jensen, O. P., Card, V. M., Hodgkins, G. & Korhonen, J., et al.  
514 (2012). Extreme events, trends, and variability in Northern Hemisphere lake-ice phenology  
(1855-2005). *Climatic Change*, 112, 299-323.
- 516 Blumthaler, M., Ambach, W. & Ellinger, R. (1997). Increase in solar UV radiation with altitude.  
*Journal of Photochemistry and Photobiology B-Biology*, 39, 130-134.
- 518 Brahney, J., Clague, J. J., Edwards, T. W. D. & Menounos, B. (2010). Late Holocene  
paleohydrology of Kluane Lake, Yukon Territory, Canada. *Journal of Paleolimnology*, 44,  
520 873-885.
- Brahney, J., Mahowald, N., Ward, D. S., Ballantyne, A. P. & Neff, J. C. (2015). Is atmospheric  
522 phosphorus pollution altering global alpine lake stoichiometry? *Global Biogeochemical  
Cycles*, 29, 1369-1383.
- 524 Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M. & West, G. B. (2004). Toward a  
metabolic theory of ecology. *Ecology*, 85, 1771-1789.
- 526 Brown, L. E., Hannah, D. M. & Milner, A. M. (2006). Hydroclimatological influences on water  
column and streambed thermal dynamics in an alpine river system. *Journal of Hydrology*,  
528 325, 1-20.
- Cannone, N., Diolaiuti, G., Guglielmin, M. & Smiraglia, C. (2008). Accelerating climate change  
530 impacts on alpine glacier forefield ecosystems in the European Alps. *Ecological  
Applications*, 18, 637-648.
- 532 Cauvy-Fraunié, S. & Dangles, O. (2019). A global synthesis of biodiversity responses to glacier  
retreat. *Nature Ecology & Evolution* 3, 1675–1685.

- 534 Clague, J. J. & Evans, S. G. (2000). A review of catastrophic drainage of moraine-dammed  
lakes in British Columbia. *Quaternary Science Reviews*, 19, 1763-1783.
- 536 Colombo, N., Bocchiola, D., Martin, M., Confortola, G., Salerno, F., Godone, D., D'Amico, M.E.,  
& Freppaz, M. (2019). High export of nitrogen and dissolved organic carbon from an alpine  
538 glacier (Indren Glacier, NW Italian Alps). *Aquatic Sciences* 81:74
- Cooke, S. L., Williamson, C. E. & Saros, J. E. (2006). How do temperature, dissolved organic  
540 matter and nutrients influence the response of *Leptodiatomus ashlandi* to UV radiation in a  
subalpine lake? *Freshwater Biology*, 51, 1827-1837.
- 542 Cross, W. F., Hood, J. M., Benstead, J. P., Huryn, A. D. & Nelson, D. (2015). Interactions  
between temperature and nutrients across levels of ecological organization. *Global Change*  
544 *Biology*, 21, 1025-1240.
- D'Amico, S., Collins, T., Marx, J. C., Feller, G., and Gerday, C. (2006). Psychrophilic  
546 microorganisms: Challenges for life. *EMBO Rep.*, 7, 385–389.
- Da Silva, J. F. & Williams, R. J. P. (2001). *The biological chemistry of the elements: the*  
548 *inorganic chemistry of life*. Oxford University Press, Oxford.
- Daines, S. J., Clark, J. R. & Lenton, T.M. (2014). Multiple environmental controls on  
550 phytoplankton growth strategies determine adaptive responses of the N:P ratio. *Ecology*  
*Letters*, 17, 414-25.
- 552 Daly, G. L. & Wania, F. (2005). Organic contaminants in mountains. *Environmental Science &*  
*Technology*, 39, 385-398.
- 554 Davison, I. R. (1991). Environmental effects on algal photosynthesis: temperature. *Journal of*  
*Phycology*, 27, 2-8.
- 556 DeMott, W. R., & Gulati, R. D. (1999). Phosphorus limitation in *Daphnia*: evidence from a long  
term study of three hypereutrophic Dutch lakes. *Limnology and Oceanography*, 44, 1557-  
558 1564.



Ekvall, M. T., Hylander, S., Walles, T., Yang, X. & Hansson, L. (2015). Diel vertical migration,  
size distribution and photoprotection in zooplankton as response to UV-A radiation.

*Limnology and Oceanography*, 60, 2048-2058.

Elser, J. J., Acharya, K., Kyle, M., Cotner, J., Makino, W. & Markow, T., et al. (2003). Growth  
rate-stoichiometry couplings in diverse biota. *Ecology Letters*, 6, 936-943.

Elser, J. J., Sterner, R. W., Gorokhova, E., Fagan, W. F., Markow, T. A. & Cotner, J. B., et al.  
(2000). Biological stoichiometry from genes to ecosystems. *Ecology Letters*, 3, 540-550.

Elser, J. J., Fagan, W. F., Kerkhoff, A. J., Swenson, N. G., & Enquist, B. J. (2010). Biological  
stoichiometry of plant production: metabolism, scaling and ecological response to global  
change. *New Phytologist*, 186, 593-608.

Engstrom, D. R., Fritz, S. C., Almendinger, J. E. & Juggins, S. (2000). Chemical and biological  
trends during lake evolution in recently deglaciated terrain. *Nature*, 408, 161-166.

Fagan, W. F., Siemann, E., Mitter, C., Denno, R. F., Huberty, A. F. & Woods, H. A., et al.  
(2002). Nitrogen in insects: implications for trophic complexity and species diversification.  
*American Naturalist*, 160, 784-802.

Farewell, A. & Neidhardt, F. C. (1998). Effect of temperature on *in vivo* protein synthetic  
capacity in *Escherichia coli*. *Journal of Bacteriology*, 180, 4704-4710.

Fegel, T. S., Baron, J. S., Fountain, A. G., Johnson, G. F. & Hall, E. K. (2016). The differing  
biogeochemical and microbial signatures of glaciers and rock glaciers. *Journal of  
Geophysical Research-Biogeosciences*, 121, 919-932.

Fell, S. C., Carrivick, J. L. & Brown, L. E. (2017). The multitrophic effects of climate change and  
glacier retreat in mountain rivers. *Bioscience*, 67, 897-911.

Galmes, J., Kapralov, M. V., Copolovici, L. O., Hermida-Carrera, C. & Niinemets, U. (2015).

Temperature responses of the Rubisco maximum carboxylase activity across domains of

life: phylogenetic signals, trade-offs, and importance for carbon gain. *Photosynthesis*

*Research*, 123, 183-201.

Gonzalez, A. L., Cereghino, R., Dezerald, O., Farjalla, V. F., Leroy, C. & Richardson, B. A., et al. (2018). Ecological mechanisms and phylogeny shape invertebrate stoichiometry: A test using detritus-based communities across Central and South America. *Functional Ecology*, 32, 2448-2463.

Gonzalez, A. L., Miguel Farina, J., Kay, A. D., Pinto, R. & Marquet, P. A. (2011). Exploring patterns and mechanisms of interspecific and intraspecific variation in body elemental composition of desert consumers. *Oikos*, 120, 1247-1255.

Green, K. (2012). Intra- and inter-annual changes in chemistry of Australian glacial lakes. *Marine and Freshwater Research*, 63, 513-527.

Grover, J. P. 1997. *Resource Competition*, Chapman & Hall.

Hamilton, T. L., Peters, J. W., Skidmore, M. L. & Boyd, E. S. (2013). Molecular evidence for an active endogenous microbiome beneath glacial ice. *The ISME Journal*, 7, 1402-1412.

Hodapp, D., Hillebrand, H. & Striebel, M. (2019). "Unifying" the concept of resource use efficiency in ecology. *Frontiers in Ecology and Evolution*, 6, doi: 10.3389/fevo.2018.00233.

Hoham, R. W., Frey, F. M., Mohn, W. W., Felio, J. H., Todd, S., Duncan, J. E., & Banghart, J. B. (2008). Optimum growth temperatures of three species of green *Chloromonas* snow algae from upstate New York and the White Mountains, Arizona. *Arctic, Antarctic, and Alpine Research*, 40, 355–363.

Hooper, D. U., Chapin, F. S., Ewel, J. J., Hector, A., Inchausti, P. & Lavorel, S., et al.

(2005). Effects of biodiversity on ecosystem functioning: A consensus of current knowledge. *Ecological Monographs*, 75, 3-35.

Hood, E., & Scott, D. (2008). Riverine organic matter and nutrients in southeast Alaska affected by glacial coverage. *Nature Geoscience*, 1, 583–587.

608 Hood, E., Fellman, J., Spencer, R. G., Hernes, P. J., Edwards, R., D'Amore, D., & Scott, D.  
 (2009). Glaciers as a source of ancient and labile organic matter to the marine environment.  
 610 *Nature*, 462, 1044-1047.

Hotaling, S., Finn, D. S., Giersch, J. J., Weisrock, D. W. & Jacobsen, D. (2017a). Climate  
 612 change and alpine stream biology: progress, challenges, and opportunities for the future.  
*Biological Reviews*, 92, 2024-2045.

614 Hotaling, S., Foley, M. E., Zeglin, L. H., Finn, D. S., Tronstad, L. M., Giersch, J. J., C.C.  
 Muhlfeld, & Weisrock, D. W. (2019). Microbial assemblages reflect environmental  
 616 heterogeneity in alpine streams. *Global Change Biology*, 25, 2576-2590.  
<https://doi.org/10.1111/gcb.14683>

618 Hotaling, S., Hood, E. & Hamilton, T. L. (2017b). Microbial ecology of mountain glacier  
 ecosystems: biodiversity, ecological connections and implications of a warming climate.  
 620 *Environmental Microbiology*, 19, 2935-2948.

Huss, M. & Hock, R. (2018). Global-scale hydrological response to future glacier mass loss.  
 622 *Nature Climate Change*, 8, 135-140.

Hylander, S., Larsson, N. & Hansson, L. (2009). Zooplankton vertical migration and plasticity of  
 624 pigmentation arising from simultaneous UV and predation threats. *Limnology and  
 Oceanography*, 54, 483-491.

626 Jacobsen, D., Milner, A. M., Brown, L. E., & Dangles, O. (2012). Biodiversity under threat in  
 glacier-fed river systems. *Nature Climate Change*, 2, 361-364.

628 Jochum, M., Barnes, A. D., Weigelt, P., Ott, D., Rembold, K. & Farajallah, A., et al. (2017).  
 Resource stoichiometry and availability modulate species richness and biomass of tropical  
 630 litter macro-invertebrates. *Journal of Animal Ecology*, 86, 1114-1123.

Jones, L. A., C. C. Muhlfeld, and L. A. Marshall. 2017. Projected warming portends seasonal shifts of stream temperatures in the Crown of the Continent Ecosystem, USA and Canada. *Climatic Change*, 144, 641–655.

Kay, A. D., Ashton, I. W., Gorokhova, E., Kerkhoff, A. J., Liess, A. & Litchman, E. (2005). Toward a stoichiometric framework for evolutionary biology. *Oikos*, 109, 6-17.

Klein, J. A., C. M. Tucker, A. W. Nolin, K. A. Hopping, R. S. Reid, C. Steger, A. Grêt-Regamey, S. Lavorel, B. Müller, E. T. Yeh, R. B. Boone, P. Bourgeron, V. Butsic, E. Castellanos, X. Chen, S. K. Dong, G. Greenwood, M. Keiler, R. Marchant, R. Seidl, T. Spies, J. Thorn, and K. Yager. 2019. Catalyzing transformations to sustainability in the world's mountains. *Earth's Future*, 7, 2018EF001024.

Larson, R. A. & Berenbaum, M. R. (1988). Environmental phototoxicity. *Environmental Science & Technology*, 22, 354-360.

Li, D., Wrzesien, M. L., Durand, M., Adam, J. & Lettenmaier, D. P. (2017). How much runoff originates as snow in the western United States, and how will that change in the future? *Geophysical Research Letters*, 44, 6163-6172.

Lindholm, M., Wolf, R., Finstad, A. & Hessen, D. O. (2016). Water browning mediates predatory decimation of the Arctic fairy shrimp *Branchinecta paludosa*. *Freshwater Biology*, 61, 340-347.

Litchman, E., Edwards, K. F., Klausmeier, C. A. & Thomas, M. K. (2012). Phytoplankton niches, traits and eco-evolutionary responses to global environmental change. *Marine Ecology Progress Series*, 470, 235-248.

Loreau, M., Naeem, S., Inchausti, P., Bengtsson, J., Grime, J. P. & Hector, A., et al. (2001). Biodiversity and ecosystem functioning: current knowledge and future challenges. *Science*, 294, 804-808.

Lutz, S., Anesio, A. M., Raiswell, R., Edwards, A., Newton, R. J. & Gill, F., et al. (2016). The  
656 biogeography of red snow microbiomes and their role in melting arctic glaciers. *Nature  
Communications*, 7, e11968.

Macfadyen, E. J., Williamson, C. E., Grad, G., Lowery, M., Jeffrey, W. H. & Mitchell, D. L.  
(2004). Molecular response to climate change: temperature dependence of UV-induced  
660 DNA damage and repair in the freshwater crustacean *Daphnia pulex*. *Global Change  
Biology*, 10, 408-416.

Marinone, M. C., Marque, S. M., Suarez, D. A., Dieguez, M. D. C., Perez, P. & Rios, P. D. L., et  
al. (2006). UV radiation as a potential driving force for zooplankton community structure in  
664 Patagonian lakes. *Photochemistry and Photobiology*, 82, 962-971.

Marty, C., Tilg, A. M. & Jonas, T. (2017). Recent evidence of large-scale receding snow water  
666 equivalents in the European Alps. *Journal of Hydrometeorology*, 18, 1021-1031.

Maurer, J.M., Schaefer, J.M., Rupper, S. and Corley, A., (2019). Acceleration of ice loss across  
668 the Himalayas over the past 40 years. *Science advances*, 5, 6.

Miki T, Yokokawa T, & Matsui K. (2013). Biodiversity and multifunctionality in a microbial  
670 community: a novel theoretical approach to quantify functional redundancy. *Proceedings  
Royal Society B Biology Sciences*, 281, 20132498.

Milner, A. M. & Bailey, R. G. (1989). Salmonid colonization of new streams in Glacier Bay  
672 National Park, Alaska. *Aquaculture Research*, 20, 179-192.

Milner, A. M., Khamis, K., Battin, T. J., Brittain, J. E., Barrand, N. E. & Fuehrer, L., et al.  
(2017). Glacier shrinkage driving global changes in downstream systems. *Proceedings of  
676 the National Academy of Sciences of the United States of America*, 114, 9770-9778.

Moser, K. A., Baron, J. S., Brahney, J., Oleksy, I. A., Saros, J. E. & Hundey, E. J., et al. (2019).  
678 Mountain lakes: Eyes on global environmental change. *Global and Planetary Change*, 178,  
77-95.

680 Muhlfeld CC, Cline TJ, Giersch JJ, Peitzsch E, Florentine C, Jacobsen D, Hotelling S. (2020)  
 Specialized meltwater biodiversity persists despite widespread deglaciation. *PNAS*, 117,  
 682 2208-12214.

Mulder, C. & Elser, J. J. (2009). Soil acidity, ecological stoichiometry and allometric scaling in  
 684 grassland food webs. *Global Change Biology*, 15, 2730-2738.

Napolitano, M. J. & Shain, D. H. (2005). Distinctions in adenylate metabolism among organisms  
 686 inhabiting temperature extremes. *Extremophiles*, 9, 93-98.

Niedrist, G. H., & Füreder, L. (2020). Real-time warming of alpine streams: (re)defining  
 688 invertebrates' temperature preferences. *River Research and Applications*,  
<https://doi.org/10.1002/rra.3638>

690 Ott, D., Digel, C., Klärner, B., Maraun, M., Pollierer, M. & Rall, B. C., et al. (2014). Litter  
 elemental stoichiometry and biomass densities of forest soil invertebrates. *Oikos*, 123, 1212-  
 692 1223.

Pallarés, S., Millán, A., Mirón, J. M., Velasco, J., Sánchez-Fernández, D., Botella-Cruz, M., &  
 694 Abellán, P. (2020). Assessing the capacity of endemic alpine water beetles to face climate  
 change. *Insect Conservation and Diversity / Royal Entomological Society of London*, 13,  
 696 271–282.

Parker, B. R., Vinebrooke, R. D. & Schindler, D. W. (2008). Recent climate extremes alter  
 698 alpine lake ecosystems. *Proceedings of the National Academy of Sciences of the United  
 States of America*, 105, 12927-31.

700 Peter, H. & Sommaruga, R. (2016). Shifts in diversity and function of lake bacterial communities  
 upon glacier retreat. *The ISME Journal*, 10, 1545–1554.

702 Phillips, K. N., Godwin, C. M. & Cotner, J. B. (2017). The effects of nutrient imbalances and  
 temperature on the biomass stoichiometry of freshwater bacteria. *Frontiers in Microbiology*,  
 704 8, e1692.

Quigg, A., Finkel, Z. V., Irwin, A. J., Rosenthal, Y., Ho, T. Y. & Reinfelder, J. R., et al. (2003).

706 The evolutionary inheritance of elemental stoichiometry in marine phytoplankton. *Nature*,  
425, 291-294.

708 Quigg, A., Irwin, A. J. & Finkel, Z. V. (2011). Evolutionary inheritance of elemental stoichiometry  
in phytoplankton. *Proceedings of the Royal Society B: Biological Sciences*, 278, 526-534.

710 Ren, Z., Martyniuk, N., Oleksy, I. A., Swain, A., & Hotaling, S. (2019). Ecological stoichiometry  
of the mountain cryosphere. *Frontiers in Ecology and Evolution*, 7, 360.

712 Rose, K. C., Williamson, C. E., Saros, J. E., Sommaruga, R. & Fischer, J. M. (2009).  
Differences in UV transparency and thermal structure between alpine and subalpine lakes:  
714 implications for organisms. *Photochemical & Photobiological Sciences*, 8, 1244-1256.

Saros, J. E., Rose, K. C., Clow, D. W., Stephens, V. C., Nurse, A. B. & Arnett, H. A., et al.  
716 (2010). Melting alpine glaciers enrich high-elevation lakes with reactive nitrogen.  
*Environmental Science & Technology*, 44, 4891-4896.

718 Shah, A.A., Dillon, M.E., Hotaling, S., & Woods, H.A. (2020). High elevation insect communities  
face shifting ecological and evolutionary landscapes. *Current Opinion in Insect Science*, 41,  
720 10.1016/j.cois.2020.04.002.

Shugar, D. H., Clague, J. J., Best, J. L., Schoof, C., Willis, M. J. & Copland, L., et al. (2017).  
722 River piracy and drainage basin reorganization led by climate-driven glacier retreat. *Nature*  
*Geoscience*, 10, 370-375.

724 Siddiqui, K. S. & Cavicchioli, R. (2006). Cold-adapted enzymes. *Annual Review of*  
*Biochemistry*, 75, 403-433.

726 Slemmons, K. E. H. & Saros, J. E. (2012). Implications of nitrogen-rich glacial meltwater for  
phytoplankton diversity and productivity in alpine lakes. *Limnology and Oceanography*, 57,  
728 1651-1663.

Sommaruga, R. (2001). The role of solar UV radiation in the ecology of alpine lakes. *Journal of Photochemistry and Photobiology B-Biology*, 62, 35-42.

Sommaruga, R. (2015). When glaciers and ice sheets melt: consequences for planktonic organisms. *Journal of Plankton Research*, 37, 509-518.

Stenzel, B., Rofner, C., Perez, M. T. & Sommaruga, R. (2017). Stoichiometry of natural bacterial assemblages from lakes located across an elevational gradient. *Scientific Reports*, 7, e5875.

Sterner, R. W. & Elser, J. J. (2002). *Ecological Stoichiometry*. Princeton University Press, Princeton.

Takeuchi, N., Uetake, J., Fujita, K., Aizen, V. B. & Nikitin, S. D. (2006). A snow algal community on Akkem glacier in the Russian Altai mountains. *Annals of Glaciology*, 43, 378-384.

Tartarotti, B., Trattner, F., Remias, D., Saul, N., Steinberg, C. E. W. & Sommaruga, R. (2017). Distribution and UV protection strategies of zooplankton in clear and glacier-fed alpine lakes. *Scientific Reports*, 7, 4487.

Thrane, J., Hessen, D. O. & Andersen, T. (2016). The impact of irradiance on optimal and cellular nitrogen to phosphorus ratios in phytoplankton. *Ecology Letters*, 19, 880-888.

Tiberti, R., Buscaglia, F., Callieri, C., Rogora, M., Tartari, G., & Sommaruga, R. (2019). Food web complexity of high mountain lakes is largely affected by glacial retreat. *Ecosystems*, <https://doi.org/10.1007/s10021-019-00457-8>

Tilman, D. 1982. *Resource Competition and Community Structure*, Princeton University Press.

Tockner, K., Malard, F., Uehlinger, U. & Ward, J. V. (2002). Nutrients and organic matter in a glacial river-floodplain system (Val Roseg, Switzerland). *Limnology and Oceanography*, 47, 266-277.



- 752 Toseland, A., Daines, S. J., Clark, J. R., Kirkham, A., Strauss, J. & Uhlig, C., et al. (2013). The  
impact of temperature on marine phytoplankton resource allocation and metabolism. *Nature*  
754 *Climate Change*, 3, 979-984.
- Tranter, M., Brown, G. H., Hodson, A., Gurnell, A. M. & Sharp, M. J. (1994). Variations in the  
756 nitrate concentration of glacial runoff in Alpine and sub-Polar environments. *IAHS*  
*Publications-Series of Proceedings and Reports-Intern Assoc Hydrological Sciences*, 223,  
758 299-312.
- Treanor, H. B., Giersch, J. J., Kappenman, K. M., Muhlfeld, C. C., & Webb, M. A. H. (2013).  
760 Thermal tolerance of meltwater stonefly *Lednia tumana* nymphs from an alpine stream in  
Waterton–Glacier International Peace Park, Montana, USA. *Freshwater Science*, 32, 597–  
762 605.
- Vandeberg, G. S. & Vanlooy, J. A. (2016). Continental glacier meltwater contributions to late  
764 summer stream flow and water quality in the northern Wind River Range, Wyoming, USA.  
*Environmental Earth Sciences*, 75, 389. doi.org/10.1007/s1266.
- 766 Vinebrooke, R. D. & Leavitt, P. R. (2005). Mountain lakes as indicators of the cumulative  
impacts of ultraviolet radiation and other global stressors. In: *Global Change and Mountain*  
768 *Regions* (pp. 437-447). Springer, Dordrecht.
- Viviroli, D., Durr, H. H., Messerli, B., Meybeck, M. & Weingartner, R. (2007). Mountains of the  
770 world, water towers for humanity: Typology, mapping, and global significance. *Water*  
*Resources Research*, 43, W074477.
- 772 Wang, Q., Fan, X. & Wang, M. (2016). Evidence of high-elevation amplification versus Arctic  
amplification. *Scientific Reports*, 6, e19219.
- 774 West, G. B. & Brown, J. H. (2005). The origin of allometric scaling laws in biology from  
genomes to ecosystems: towards a quantitative unifying theory of biological structure and  
776 organization. *Journal of Experimental Biology*, 208, 1575-1592.

WGMS. (2017). Global Glacier Change Bulletin No.2 (2014-2015). Zemp, M., Nussbaumer, S.

778 U., Gärtner, Roer, I., Huber, J., Machguth, H., Paul, F., and Hoelzle, M. (eds.),  
ICSU(WDS)/IUGG(IACS)/UNEP/UNESCO/WMO, World Glacier Monitoring Service, Zurich,  
780 Switzerland, 244 pp., publication based on database version: doi:10.5904/wgms-fog-2017-  
10.

782 Wilhelm, L., Singer, G. A., Fasching, C., Battin, T. J., & Besemer, K. (2013). Microbial  
biodiversity in glacier-fed streams. *The ISME journal*, 7, 1651.

784 Williamson, C. E. & Rose, K.C. (2010). When UV meets freshwater. *Science*, 329, 637-639.

Williamson, C. E., Grad, G., De Lange, H. J., Gilroy, S. & Karapelou, D. M. (2002).

786 Temperature-dependent ultraviolet responses in zooplankton: Implications of climate  
change. *Limnology and Oceanography*, 47, 1844-1848.

788 Williamson, C. E., Neale, P. J., Grad, G., De Lange, H. J. & Hargreaves, B. R. (2001a).

Beneficial and detrimental effects of UV on aquatic organisms: Implications of spectral  
790 variation. *Ecological Applications*, 11, 1843-1857.

Williamson, C. E., Neale, P. J., Hylander, S., Rose, K. C., Figueroa, F. L. & Robinson, S. A., et

792 al. (2019). The interactive effects of stratospheric ozone depletion, UV radiation, and climate  
change on aquatic ecosystems. *Photochemical & Photobiological Sciences*, 18, 717-746.

794 Williamson, C. E., Olson, O. G., Lott, S. E., Walker, N. D., Engstrom, D. R. & Hargreaves, B. R.

(2001b). Ultraviolet radiation and zooplankton community structure following deglaciation in  
796 Glacier Bay, Alaska. *Ecology*, 82, 1748-1760.

Williamson, C. E., C. Salm, S. L. Cooke, and J. E. Saros. (2010). How do UV radiation,

798 temperature, and zooplankton influence the dynamics of alpine phytoplankton communities?  
*Hydrobiologia*, 648, 73-81.

800 Williamson, C. E., Zepp, R. G., Lucas, R. M., Madronich, S., Austin, A. T. & Ballare, C. L., et al.

(2014). Solar ultraviolet radiation in a changing climate. *Nature Climate Change*, 4, 434-441.

- Woods, H. A., Makino, W., Cotner, J. B., Hobbie, S. E., Harrison, J. F. & Acharya, K., et al. (2003). Temperature and the chemical composition of poikilothermic organisms. *Functional Ecology*, 17, 237-245.
- Wynn, P. M., Hodson, A. J., Heaton, T. H. E. & Chenery, S. R. (2007). Nitrate production beneath a high Arctic glacier, Svalbard. *Chemical Geology*, 244, 88-102.
- Xenopoulos, M. A., Frost, P. C. & Elser, J. J. (2002). Joint effects of ultraviolet radiation and phosphorus supply on phytoplankton growth rate and elemental composition. *Ecology*, 83, 423-435.
- Yvon-Durocher, G., Dossena, M., Trimmer, M., Woodward, G. & Allen, A. P. (2015). Temperature and the biogeography of algal stoichiometry. *Global Ecology and Biogeography*, 24, 562-570.
- Yvon-Durocher, G., Schaum, C. E. & Trimmer, M. (2017). The temperature dependence of phytoplankton stoichiometry: investigating the roles of species sorting and local adaptation. *Frontiers in Microbiology*, 8, e2003.
- Zimmerman, A. E., Allison, S. D. & Martiny, A. C. (2014). Phylogenetic constraints on elemental stoichiometry and resource allocation in heterotrophic marine bacteria. *Environmental Microbiology*, 16, 1398-1410.

## Figure Legends

**Figure 1.** A cross section of a lake or a stream channel illustrating major pathways by which four key rules of life mediate the responses of ecological systems to a changing cryosphere in aquatic alpine environments. Temperature modulates molecular processes connected to metabolism and biosynthesis (Rule 1). Solar radiation modulates biotic responses in a wavelength-dependent manner (Rule 2) via effects of photosynthetically active radiation (PAR)

on photosynthesis and consumer behavior while UVR drives photodamage and avoidance

828 responses. Element supplies, temperature, and light affect the variable C:N:P stoichiometry of  
photoautotroph biomass, which influences the dynamics of food quality for, and nutrient  
830 recycling by, stoichiometrically constrained consumers (Rule 3). As a result of interactions  
among these key rules, resource use efficiency (RUE) emerges at different levels of  
832 organization and modulates the efficiency of transformations of energy and materials through  
the ecological system (Rule 4).

834 **Figure 2.** (a) A schematic showing the sequence of expected changes in the light environment

836 and associated light-dependent responses as the alpine cryosphere diminishes in a glaciated  
catchment. Light attenuation in turbid, young, glacier-fed lakes is dominated by mineral-rich  
838 glacial flour particles that attenuate light largely through scattering that is not highly wavelength-  
selective. After glacial influence subsides, very clear lakes remain in a largely bedrock and till  
840 landscape with very little organic matter. In high alpine regions these are the highest ultraviolet  
radiation (UVR) exposure environments in the world. Incident UVR is high in these

842 environments, and both UVR and photosynthetically active radiation (PAR) show minimal  
attenuation with depth due to the very high water clarity. As terrestrial vegetation develops in the  
844 landscape over time, terrestrially-derived dissolved organic matter (DOM) is leached into the  
lakes, selectively attenuating UVR relative to PAR. (b) Irradiance (percent of subsurface

846 irradiance) vs. depth profiles for 320 nm UVR (violet line) and (PAR, green line) in three lakes in  
a glacial chronosequence in Glacier Bay, Alaska. Data from Silty Bruce (a glacier-fed lake still in  
848 contact with glacier), Little Esker (~10 years old), and Klotz Hills (~90 years old) lakes  
(Williamson et al., 2001b).

**Figure 3.** Predicted shifts in (a) nutrient supply and (b) biological demand of nutrients (i.e., stoichiometric ratios) as glaciers and permanent snowpack in the catchment fade. It is anticipated that SRP will decline as DIN and DOC fluxes increase (Hood et al., 2009; Saros et al., 2010). Cold, turbid systems with increased P availability and short growing seasons are expected to favor fast-growing organisms with greater P requirements (Elser et al., 2000, 2003). These fast-growing biota will be disadvantaged as glacier coverage declines and ecosystem P-limitation emerges.

**Figure 4.** Hypothetical changes in resource use efficiency (RUE) and the factors impacting RUE during lake ontogeny. Initially, with a high degree of glacial connectivity lakes are highly turbid and light limited (Sommaruga, 2015). System turbidity will decrease over time following loss of hydrological connectivity with the glacier. As glacial connectivity recedes, light limitation decreases and in turn RUE increases. We hypothesize a temporary decrease in RUE as the system shifts from a turbid to a clear state largely due to community restructuring. PAR, UVR, and external supplies of nitrogen and organic carbon are all expected to increase over time while phosphorus will decrease (as described in Rule 3). Community assembly will have low functional diversity initially and be dominated by mixotrophic and organisms adapted to low light. Functional diversity will increase over time and be greatest at intermediate levels of disturbance (i.e., the transition between turbid and clear states). Following the loss of turbidity, a major disturbance, functional diversity will decrease and be dominated by fewer and specialized keystone taxa during the highly clear state. A similar trajectory may be followed in glacier-dominated streams as turbidity conditions change.

**Figure 5:** Examples of differences in the phenology of exposure to (a) potentially damaging UVR (320 nm, derived from a radiative transfer model), (b) surface water temperature (T), and

(c) the potential for UVR damage (UVR:T ratio) in a clear-water lowland temperate zone lake

(dashed lines, data from Williamson et al., 2002), and a high elevation temperate zone lake

(solid lines, 4,000 m, representing the Tibetan Plateau, the largest high elevation region in the

world). UVR exposure in the alpine lake is estimated by using the data from the lowland lake

and an 11% increase per 1000 m in 320 nm solar UVR (Blumthaler et al., 1997). Alpine surface

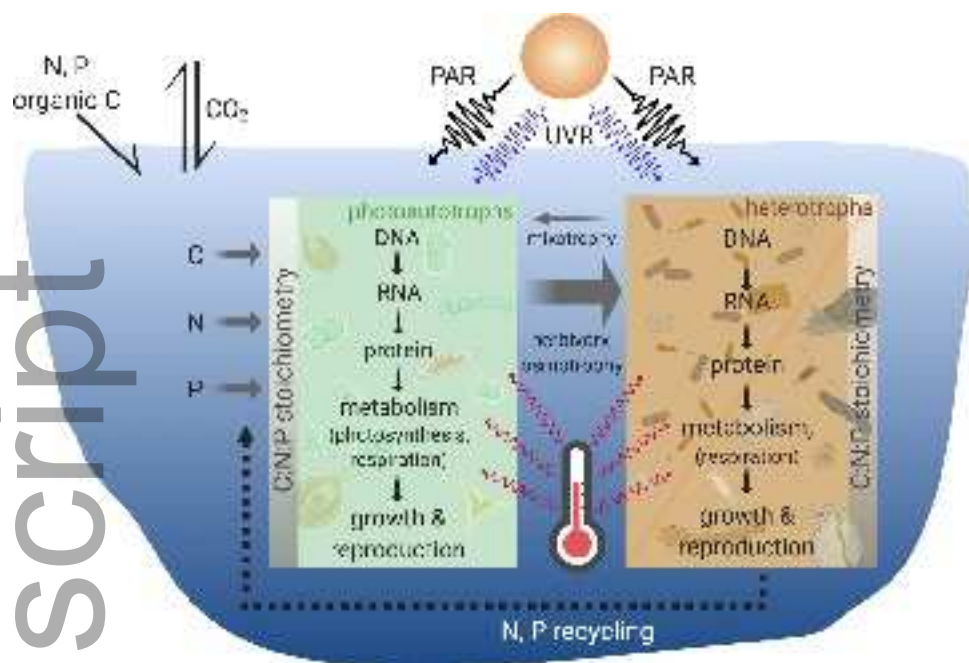
water temperatures are hypothetical, based on ice-out soon after the summer solstice.

Temperature data are from multiple temperate-zone alpine lakes around the world (C.E.W.,

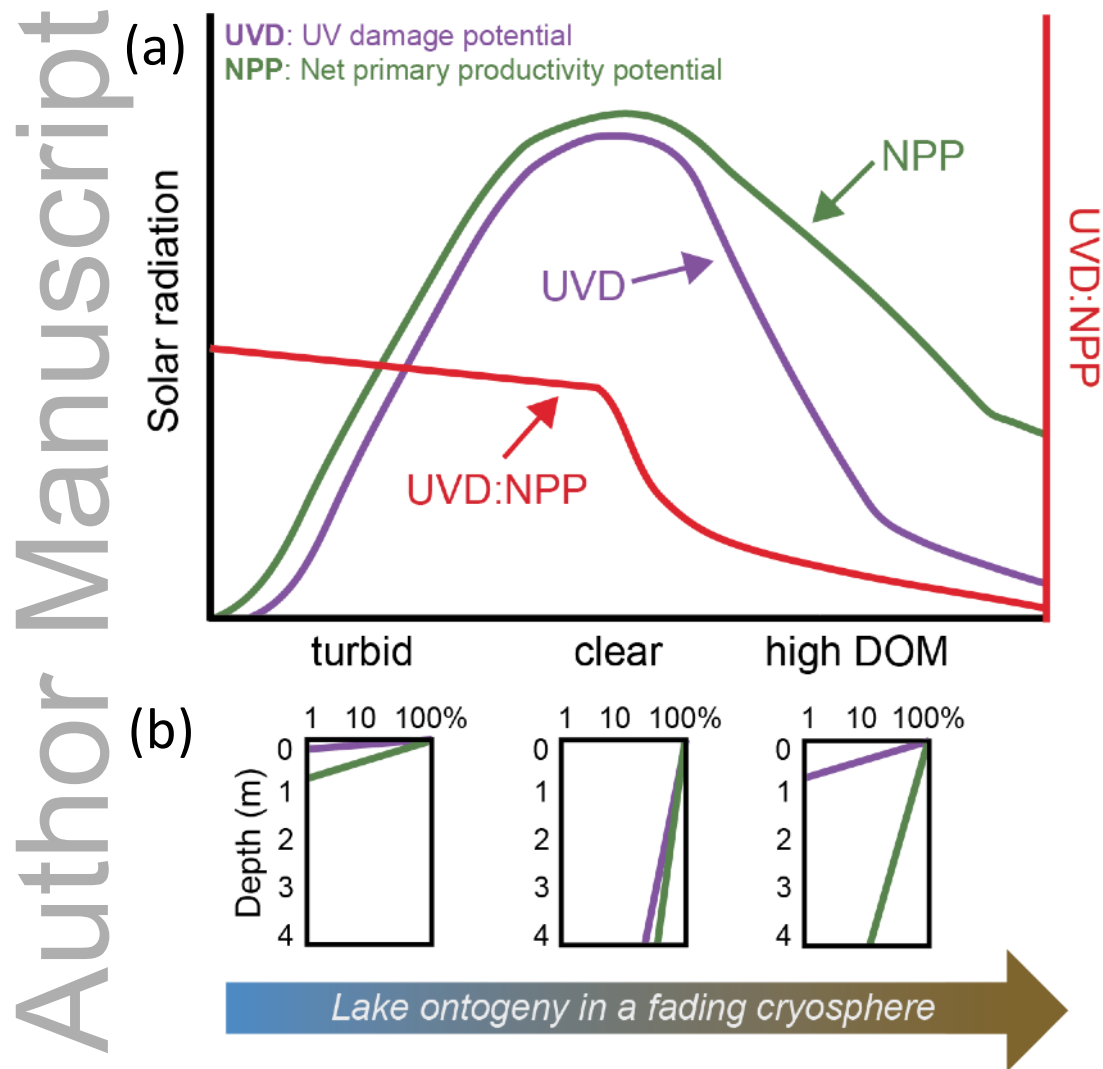
unpublished data). Incident UVR will increase, and seasonal variations in UVR and T decrease

with decreasing latitude into more tropical regions, while the reverse patterns will be observed in

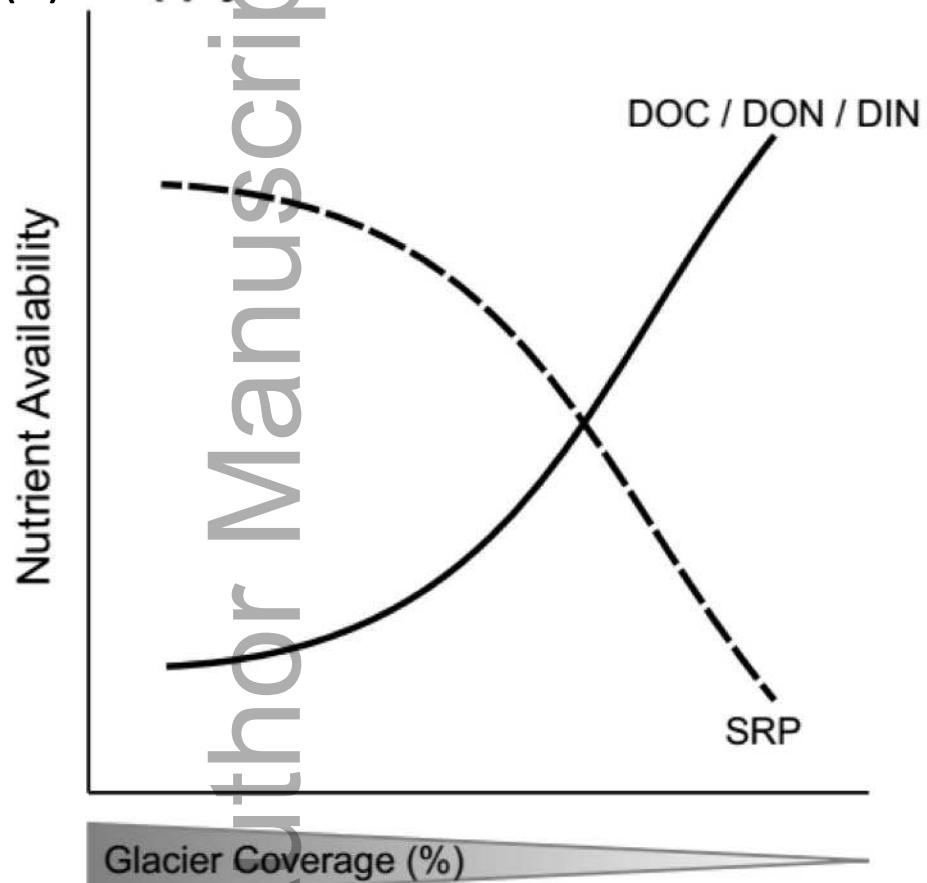
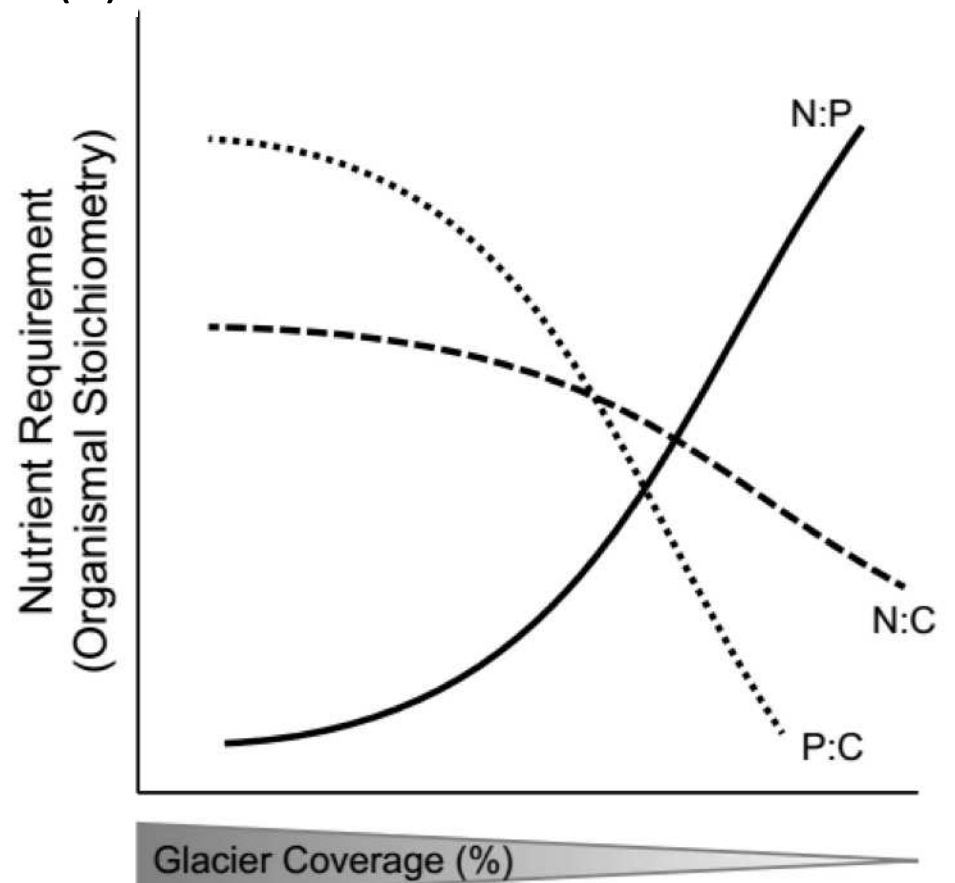
higher latitude boreal to polar regions.



gcb\_15362\_f1.png





(a) **Supply**(b) **Demand**

TIME/ONTOGENY

PAR

UV

P

DIN & DOC

Community  
Assembly

RUE

Turbid

Clear

TIME/ONTOGENY

