

**Towards more robust hydroacoustic estimates of fish abundance in the presence of  
pelagic macroinvertebrates**

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## Abstract

The inclusion of unwanted targets in hydroacoustic surveys biases estimates of fish abundance. Thus, knowledge of frequency-dependent responses of unwanted targets (e.g., pelagic macroinvertebrates) can help ensure that transducer frequencies are used that minimize this bias. We determined how fish density estimates varied across multiple frequencies when the larval stage of a midge, *Chaoborus*, was present in the water column. We hypothesized that fish density estimates would increase with increasing transducer frequency, owing to greater backscattering by *Chaoborus* at higher frequencies than lower ones, which allows it to be included with the backscattering caused by fish. We found that fish density estimates were always greater at higher frequencies (e.g., 120 and 200 kHz) compared to a lower one (70 kHz) in several productive north-temperate reservoirs. Furthermore, pairwise comparisons of total (i.e., fish plus *Chaoborus*) backscattering showed that significantly more backscattering occurred at higher rather than lower frequencies. We also found that fish density estimates varied between spring and summer, partially owing to inter-seasonal size variation in *Chaoborus* that influenced its backscattering. Beyond demonstrating why the presence of pelagic macroinvertebrates needs to be considered when estimating fish abundance with hydroacoustics, we provide methods to identify and reduce this bias.

**Key words:** phantom midge, *Mysis*, fish acoustics, *Dorosoma cepedianum*, diel vertical migration, geostatistics

## 1. Introduction

As with all sampling methods used to estimate fish abundance, hydroacoustic surveys are not without bias. One potential problem is that hydroacoustic surveys do not exclusively detect the targets of interest (i.e., fish). For example, many macroinvertebrates, such as *Mysis* spp. (Rudstam et al., 2008; Axenrot et al., 2009), corixids (Kubecka et al., 2000), chironomids (Kubecka et al., 2000), *Chaoborus* spp. (Knudsen et al., 2006), jellyfish (Colombo et al., 2009), and krill (Simard and Lavoie, 1999) have measurable backscattering at frequencies commonly used during fish hydroacoustic assessment surveys. Therefore, fish abundance estimates could include contributions from non-target organisms, resulting in overestimates of fish abundance (Jurvelius et al., 2008; Rudstam et al., 2008). Owing to the commonality of pelagic macroinvertebrates in small natural lakes (Schindler et al., 1993), the Great Lakes (Beeton, 1960), man-made reservoirs (Chimney et al., 1981), estuaries (Moriarty et al., 2012), and oceans (Atkinson et al., 2004), the need to quantify and understand this bias seems paramount, regardless of ecosystem type.

The degree of bias from pelagic macroinvertebrates can be influenced by the design of the hydroacoustic survey, as well as the behaviors and biological attributes of the macroinvertebrates. Hydroacoustic surveys of fish abundance are routinely conducted at night because fish typically disperse from schools in reduced light, allowing for more precise estimates of fish abundance (Drastik et al., 2009). However, many other (non-fish) organisms, such as benthic-pelagic macroinvertebrates, also often reside in the water column at night. For example, *Mysis* spp. (Rudstam et al., 1989), *Chaoborus* spp. (La Row and Marzolf, 1970), and krill (Cotte and Simard, 2005) engage in diel vertical

71 migration behavior, where individuals reduce predation risk by occupying deeper, darker  
72 water (or sediments) during the day and moving into the water column at night to feed  
73 and/or find more suitable habitat. Other macroinvertebrates, such as chironomids, also  
74 emerge from the benthos during the day and are present in the water column and at the  
75 surface at night (Sjöberg and Danell, 1982). Bias from macroinvertebrates is further  
76 enhanced by organisms with an air bladder (i.e., *Chaoborus*), which may resonate at  
77 certain transducer frequencies and therefore contribute more backscatter than expected  
78 based on their size alone (Jones and Xie, 1994; Eckmann, 1998).

79 Not accounting for macroinvertebrates in the water column during a hydroacoustic  
80 survey of fish abundance can result in a biased fish estimate. For example, smelt  
81 (*Osmerus eperlanus*) density in Lake Hiidenvesi, Finland was overestimated by up to  
82 55% when the presence of *Chaoborus* was not considered (Malinen et al., 2005). This  
83 bias can be reduced, however, by conducting hydroacoustic surveys using frequencies  
84 at which the backscattering from non-target organisms is lower. Knudsen et al. (2006)  
85 evaluated the acoustic backscattering response of *Chaoborus* at six different  
86 frequencies, ranging 38–710 kHz. These authors detected the greatest response at 200  
87 kHz and attributed the higher backscattering at that frequency to resonance by the air  
88 sacs of *Chaoborus*. Further, these authors found that the backscattering strength varied  
89 with *Chaoborus* size. For this reason, Knudsen et al. (2006) recommended using lower  
90 frequencies (e.g., <200 kHz) during fish assessments in the presence of *Chaoborus*.  
91 Studies in other ecosystems (e.g., Malinen et al., 2005; Jurvelius et al., 2008; Knudsen  
92 and Larsson, 2009) have also concluded that fish abundance estimates measured at



200 kHz are biased high when macroinvertebrates (e.g., *Chaoborus*) are present, and that lower frequencies should be used.

Herein, we sought to evaluate the potential for a vertically migrating macroinvertebrate, *Chaoborus*, to bias hydroacoustic surveys of fish density in shallow, eutrophic reservoirs common to the eastern United States, as well as provide methods to reduce any observed bias. Specifically, we sampled with 70-, 120-, 200-, and 430-kHz frequencies during two seasons (spring and summer) to identify how estimates of fish density change with frequency in the presence of *Chaoborus*, an abundant organism in these ecosystems. To determine whether backscatter from *Chaoborus* contributed to total acoustic backscatter in our study reservoirs, we first examined the ability of our hydroacoustics system to estimate *Chaoborus* abundance. To further understand the influence of *Chaoborus* on our acoustic surveys, we also: 1) compared total (i.e., both fish and *Chaoborus*) acoustic backscattering (NASC,  $\text{m}^2\cdot\text{nmi}^{-2}$ ) from every pairwise combination of frequencies; and 2) determined the frequency-response of reservoir organisms (i.e., both fish and *Chaoborus*) during spring and summer using four different frequencies (70, 120, 200, and 430 kHz). We then evaluated how estimates of fish density varied with transducer frequency and the season sampled. Overall, our findings show frequency-dependent and season-specific biases that should be considered when developing hydroacoustic assessment protocols to estimate fish abundance in the presence of pelagic macroinvertebrates such as *Chaoborus*.

## 2. Methods

### 2.1 Study ecosystems and species

We conducted mobile hydroacoustic surveys and trawled for fish at night to estimate fish density in three Ohio (USA) reservoirs (i.e., Acton Lake, Alum Creek Lake, and Hoover Reservoir; Figure 1) during spring (May or June) and summer (August) of 2017. As with other Ohio reservoirs, our three study ecosystems are small, shallow, warm, and biologically productive (Table 1), and have seasonal hypolimnetic hypoxia during the summer through early fall. The high productivity of these reservoirs stems from their agricultural- and urban-based watersheds (Vanni et al., 2005).

These reservoirs support a fish community consisting of piscivores and planktivores. Top predators in these reservoirs are typified by largemouth bass (*Micropterus salmoides*) and saugeye (*Sander vitreus canadensis*), with their forage base primarily composed of planktivorous fishes, including gizzard shad (*Dorosoma cepedianum*), crappies (*Pomoxis* spp.), sunfishes (*Lepomis* spp.), and brook silverside (*Labidesthes sicculus*). The main species of interest for this study was gizzard shad (*Dorosoma cepedianum*), which has historically been the most abundant prey species in Ohio reservoirs (Sieber Denlinger et al., 2006; Hale et al., 2008; Dillon et al., 2019).

All of our study reservoirs have populations of the vertically migrating, macroinvertebrate *Chaoborus punctipennis*. *Chaoborus* tend to reside in bottom sediments during daylight hours and move into the water column at night (La Row and Marzolf, 1970). When in the water column at night, resonance from their air sacs holds the potential to contribute to total backscattering estimates, which has been shown to bias estimates of fish density in other ecosystems (e.g., Malinen et al., 2005; Knudsen

et al. 2006). Because this potential bias from *Chaoborus* has not been measured or considered in annual hydroacoustic surveys of fish abundance by the Ohio Department of Natural Resources – Division of Wildlife (ODNR-DOW), which oversees fisheries management in Ohio reservoirs, the need to estimate this bias is paramount (JDC, co-author, ODNR-DOW, pers. comm.)

## **2.2 Abiotic data collection**

Contemporaneously to hydroacoustic surveys and trawls (see sections 2.3 and 2.4), we collected abiotic data at three sites (Figure 1) to inform reservoir condition at the time of fish sampling. Vertical profiles of temperature (nearest 0.1 °C) and dissolved oxygen concentration (nearest 0.1 mg·L<sup>-1</sup>) were determined with a multi-parameter sonde (Model 6600, YSI, Inc., Yellow Springs, Ohio, USA) at 1-m depth increments from the surface to the bottom at each site (n = 3) in each reservoir.

## **2.3 Hydroacoustic survey design**

Hydroacoustic data were collected with a BioSonics echosounder (DT-X; Seattle, Washington, USA) operating four split-beam transducers with different frequencies: 70-kHz transducer (6.5 °, 3 dB beam angle; 3.7 m, 2x near-field); 120-kHz transducer (7.2 °, 3 dB beam angle; 1.7 m, 2x near-field); 200-kHz transducer (6.9 °, 3 dB beam angle; 1.1 m, 2x near-field); and 430-kHz transducer (6.9 °, 3 dB beam angle; 0.6 m, 2x near-field). Even though each frequency transducer had a different near-field distance, the same exact portion of the water column was used within each analysis to enable fair comparisons across frequencies (see sections 2.5-2.7). For every survey, all transducers were oriented downward on a fixed plate secured to the side of the boat, and positioned 0.5 m below the water surface.

Hydroacoustic data collection followed procedures outlined in Dillon et al. (2019). Settings included a  $-130$  dB threshold,  $0.2$ -ms pulse duration, pulse rate of  $10$  pings $\cdot$ s $^{-1}$ , start range of  $1$  m, and a stop range that varied with reservoir depth. Surveys were conducted (depth permitting) in a zig-zag pattern (Figure 1). All mobile surveys began  $0.5$  h after sunset, were conducted at a speed of  $7$  km $\cdot$ h $^{-1}$  or slower, and were completed at least one hour before sunrise. We collected passive data at night for the length of one transect while moving at survey speed to quantify background noise in each reservoir (Parker-Stetter et al., 2009; Dillon et al., 2019). In our reservoirs, mean volume backscattering strength ( $S_v$  in dB) noise ranged  $-119.2$  to  $-91.5$  dB, whereas target strength (TS in dB) noise ranged  $-148.5$  to  $-99.8$  dB.

We calibrated each transducer with a frequency-specific standard tungsten-carbide reference sphere of known TS ( $38.1$  mm diameter for  $70$  kHz;  $38.1$  mm diameter for  $120$  kHz;  $36$  mm diameter for  $200$  kHz;  $17.5$ -mm diameter for  $430$  kHz; Foote et al., 1987) during spring and summer. Calibration offsets were then applied to the survey data based on the results from the closest date of field calibration (typically within one month). Using the equations of Del Grosso and Mader (1972) and Francois and Garrison (1982), we calculated speed of sound and absorption coefficients using the average temperature from the entire water column at the deepest site sampled in each reservoir.

## **2.4 Biological collections**

We collected fish for species identification and total length (TL) measurements by towing (at  $12$ – $14$  km $\cdot$ h $^{-1}$ ) a  $1$ -m high x  $2$ -m wide neuston net ( $6,350$ - $\mu$ m mesh) at the surface, and a  $1$ -m high x  $1$ -m wide framed trawl ( $3,175$ - $\mu$ m mesh) at the surface and at

specific depths. Depths of sub-surface tows were chosen based on the greatest apparent abundance of fish (i.e., observed backscatter) observed in the concurrent hydroacoustic survey. Individual fish collected with surface net tows may not directly match those sampled by hydroacoustics at deeper depths. However, for the reasons outlined in Dillon et al. (2019), we are justified in the use of our surface net tows, as their main purpose was to collect as many individuals as possible. We conducted three tows with each net in each reservoir, with our net sampling spanning a distance of approximately 500 m (Figure 1). All captured individuals were euthanized, preserved in 95% ethanol, and returned to the laboratory where they were identified to species and measured. We measured TL (nearest 1 mm) and wet mass (nearest 1 g) for a subset of up to 50 randomly selected individuals per species per trawl.

We collected *Chaoborus* with a pump (TD5-300, Tsurumi Pump, Glendale Heights, Illinois, USA) at the same three sites sampled for abiotic data in each reservoir (Figure 1). *Chaoborus* were sampled at three discrete depths, with the depth of each sample chosen based on site-specific temperature and dissolved oxygen profiles such that individuals were collected in the epilimnion, metalimnion, and hypolimnion (Dillon et al., in review). Each pump sample filtered approximately 1 m<sup>3</sup> of water. Up to 25 randomly selected individuals from each discrete-depth sample at each site were measured (nearest 0.1 mm), and lengths were converted to dry mass (nearest 0.001 mg) using an established regression (Chimney et al., 2007). Total *Chaoborus* biomass was calculated by multiplying the density of the sample by the average dry mass of the measured individuals. We tested for differences in mean length between *Chaoborus* captured during spring (May or June) and summer (August) with a Mann-Whitney U

Test, as Bartlett's Test and visual examination of the data (histograms and Q-Q plots) revealed that the data were non-normally distributed.

## **2.5 Hydroacoustic estimates of *Chaoborus***

To determine whether backscatter from *Chaoborus* contributed to total acoustic backscatter in our study reservoirs, we first examined the ability of our hydroacoustics system to estimate *Chaoborus* abundance. To do this, we developed a relationship between a commonly used hydroacoustic index of biomass,  $S_v$ , and the pumped biomass estimate of *Chaoborus* by matching the same layer of the hydroacoustic survey to the same layer of the discrete-depth pump sample. Data from additional, but similar, sampling used to complete this analysis (section 2.5) is described in Appendix C.

### **2.5.1 Hydroacoustic processing**

All hydroacoustic analyses were conducted in Echoview (Versions 7.1–8, Myriax Pty Ltd, Hobart, Tasmania, Australia) following the methods outlined in Dillon et al. (2019), which we briefly describe here. Estimates of background noise at 1-m depth were calculated using equations from Parker-Stetter et al. (2009) and then subtracted from the survey data in the linear domain. We also removed other instances of noise (i.e., from boat wakes, bubbles, methane gas, etc.) by manually scanning the echograms and delineating these areas as “bad data” in Echoview.

We used the hydroacoustic data from the 70-, 200-, and 430- kHz frequencies to estimate the abundance of *Chaoborus* through a masking technique similar to one used by Rudstam et al. (2008), who separated echoes of a diel migrating invertebrate (*Mysis* spp.) from that of fish in Lake Ontario. First, a fish-exclusion threshold was applied to

the 70-kHz data (Rudstam et al. 2008). Only data from the 70-kHz frequency was thresholded for this analysis, as this frequency: 1) included the smallest acoustic contribution from *Chaoborus* (Knudsen et al., 2006; this study); 2) was used as the frequency that produced the most reliable estimate of fish density (see Results); and 3) was used as the frequency for masking fish echoes from the other higher frequencies (see below).

The minimum TS threshold (Table A1) was chosen by examining the *in situ* single-target distributions from -75 to -30 dB and selecting a valley in the distribution. If no clear valley was observed in the *in situ* single target distribution, then we chose the minimum TS value by converting TL of individual fish captured in net/rawl tows to TS with a general dorsal-aspect equation (Love, 1977). Single targets were identified using a maximum beam compensation of 6 dB and the settings listed in Dillon et al. (2019). A range-dependent  $S_v$  threshold was set 6 dB less than the selected minimum TS value and input as the “Minimum TS threshold” setting in each  $S_v$  variable for every reservoir sampled (see Parker-Stetter et al., 2009).

Pings between the 70- and either 200- or 430-kHz data were then matched for complete data overlap. In the  $S_v$  variable, the data were resampled as 10 data pixels·m<sup>-1</sup> and dilated to allow masking areas around each fish target. A data range bitmap was then applied to the dilated data, which removed fish targets (and the area immediately around each fish target); we replaced the values in these areas as no data. Results from the bitmap were then applied as a mask over the 200- or 430-kHz  $S_v$  data so that only echoes from *Chaoborus* remained.

## 2.5.2 Analysis

We examined the relationship between the average pump biomass estimate and the hydroacoustic estimate of *Chaoborus* biomass ( $S_v$ ) using least-squares linear regression. We developed four predictive models corresponding to the two frequencies (200 and 430 kHz) and seasons (spring: May or June; summer: August) used. We generated separate models for each period, owing to observed *Chaoborus* size differences between sampling seasons. We then used the intercept from these models to determine the TS of 1 g dry wt·m<sup>-3</sup> of *Chaoborus*. Because of the linearity principle in fisheries acoustics, the slope of the relationship between the log<sub>10</sub> of biomass or density and  $S_v$  in dB should have a slope of 0.1, if the target size is not density-dependent (Simmonds and MacLennan, 2005). We tested if the slope of each model was significantly different from 0.1 by determining if the 95% CI of the estimated slope encompassed 0.1. When the slope of the model was not significantly different from 0.1, we used the intercept estimates from these models to determine the TS (dB) of 1 g dry wt·m<sup>-3</sup> of *Chaoborus*. If the slope was significantly different from 0.1, we forced the slope of the line to equal 0.1 to get an intercept estimate, and then determined the TS (dB) of 1 g dry wt·m<sup>-3</sup> of *Chaoborus*.

## **2.6 Transducer frequency comparisons**

We explored how total backscattering (i.e., from fish and *Chaoborus*) differed among frequencies to help explain any observed differences in estimates of fish density among them. We compared cell-specific total backscattering responses (nautical area scattering coefficient, NASC, m<sup>2</sup>·nautical mile<sup>-2</sup> [nmi<sup>-2</sup>]) from every pairwise combination of frequencies (i.e., 70, 120, 200, and 430 kHz).

### **2.6.1 Hydroacoustic processing**



The hydroacoustic data processing differed in some ways from that described in section 2.5. While we subtracted noise estimates from the hydroacoustic data, as above, for the analyses conducted here only, data were not thresholded and we used a horizontal cell size of 250 m and vertical cell size of 2.5 m. This cell size was chosen for these analyses because it is beyond the distance where notable spatial correlation was present in our data (Figures A1–A3), and we were able to smooth over any areas where large fluctuations in the acoustic response might exist. We included all data from the depth of the 70-kHz transducer near-field (as this was the deepest near-field from any frequency transducer sampled) to the bottom exclusion zone (0.2 m off the detected bottom) for these analyses only, as *Chaoborus* were found at all depths in our reservoirs.

#### 2.6.2 Analysis

Frequency-specific and cell-specific NASC values ( $\text{m}^2 \cdot \text{nmi}^{-2}$ ) were compared to one another with major-axis regressions (Warton et al., 2006) using the R-package “lmodel2” (Legendre, 2018). In so doing, we could test for common slopes and their fit to a 1:1 line (slope = 1, intercept = 0). A significant ( $\alpha = 0.05$ ) relationship between each pairwise combination of frequencies was tested using data from 99 permutations.

Using the same data (i.e., 250- x 2.5- m cell size, noise removed, no thresholding), we identified which frequencies (70, 120, 200, 430 kHz) resulted in the highest backscattering of reservoir organisms (i.e., both fish and *Chaoborus* together) by examining the frequency-response of our data. We determined the frequency-response during both spring and summer to account for potential differences in the frequency-dependent backscattering response associated with changes in organism

(i.e., fish and/or *Chaoborus*) size. The frequency-response ( $\pm 1$  standard error, SE) was calculated as the average cell-specific (from a random selection of one third of the number of cells sampled in each reservoir) acoustic scattering (NASC,  $\text{m}^2 \cdot \text{nmi}^{-2}$ ) from each frequency sampled.

## **2.7 Fish density estimation**

Only the three lower frequencies (70, 120, and 200 kHz) were used to calculate fish density because they are commonly used in fish abundance assessment surveys used by agencies, including in Ohio (Hale et al., 2008; Godlewska et al., 2009).

### **2.7.1 Hydroacoustic processing**

Hydroacoustic data processing followed similar methods to those described above (sections 2.5 and 2.6), with noise being removed. Additionally, frequency-specific TS thresholds were applied to the data, using the methods described above (section 2.5). We consider thresholding to be the most appropriate method for generating fish density estimates at each frequency, given the high density of *Chaoborus* observed in our ecosystems (Baran et al., 2019), as well as the lack of overlap in TS between known fish and *Chaoborus* at the lower frequencies sampled (see Discussion).

Data processing was standardized to 50-m horizontal x 1-m vertical cells within each transect, regardless of transducer frequency or reservoir, and only focused on the 1-m vertical interval from 4.2 to 5.2 m. We chose this 1-m depth-layer for analysis as its upper limit (4.2 m) corresponds to the shallowest depth surveyed by the 70-kHz transducer, owing to its 3.7 m, 2x near-field, and position below the water surface, and the lower limit (5.2 m) corresponds to the starting depth of the hypoxic hypolimnion (dissolved oxygen  $< 2.0 \text{ mg} \cdot \text{L}^{-1}$ ), which was present during August surveys (Dillon et al.,

in review). We know from previous research that fish avoid hypolimnetic hypoxia in our study reservoirs (Burbacher 2011; Dillon et al., in review). We used Sawada et al.'s (1993)  $N_v$  index to identify cells with potentially biased *in situ* TS estimates, owing to overlapping single-target detections, and replaced those cells with the average TS from the same depth-layer (Dillon et al., 2019).

## 2.7.2 Analysis

We estimated cell-specific fish density via echo-integration, scaling the area backscattering coefficient (ABC,  $m^2 \cdot m^{-2}$ ) by the average backscattering cross section ( $\sigma_{bs}$ ,  $m^2$ ). Average fish density estimates (number of individuals  $\cdot m^{-2}$ ) were then calculated using a geostatistical model that accounted for the spatial characteristics of our sampling design. Because we used a non-random sampling design (i.e., zig-zag transects; see Figure 1), classical statistical methods are inappropriate, as assumptions about randomized sampling (i.e., independence) were violated (Simard et al., 1992; Petitgas, 2001). By contrast, a geostatistical model-based approach does not require a randomized survey design as it explicitly models inherent spatial trends and correlation in the data (Rivoirard et al., 2000). For this reason, studies have evaluated the efficacy of using geostatistical methods and found that they provide more robust (e.g., more precise variance estimates) estimates of fish abundance than traditional statistical methods (e.g., Petitgas, 2001; Taylor et al., 2005).

We describe our geostatistical models (hereafter referred to as spatial models), in brief, with details available in Appendix B. To satisfy assumptions of a Gaussian spatial model, density estimates from every frequency, season (spring or summer), and reservoir sampled were shifted (add 0.5 to each density measurement) and  $\log_{10}$

transformed. The spatial model included a spatial trend term, as well as an error covariance that assumed an exponential covariance between different locations (measured in km). Our spatial models also included a nugget term in the error covariance, to account for possible measurement error or fine-scale spatial dependencies. Spatial trends were represented by terms for Northing (km), Easting (km), and their interaction, as exploratory data analyses indicated these trends were apparent in our data (PFC and RAD, unpub. data). The spatial model parameters were estimated using maximum likelihood. A spatial model was appropriate for our data analyses, as Akaike information criterion (AIC) analysis indicated preference for spatial models over non-spatial models for most surveys (Supplementary Material; Table S1). Summaries of the spatial models showed that estimates of model terms (i.e., intercept, Northing, Easting, and their interaction), and the significance of these terms varied with the reservoir and season sampled (Supplementary Material: Tables S2–S4). Further, while mean density estimates were similar between the spatial and non-spatial models, the error (SE) estimates from the spatial model were generally smaller than those calculated from the non-spatial model (Supplementary Material: Table S5, Figure S1). Thus, based on the non-random sampling design, the AIC values, and lower observed SE of the estimates, we used the spatial model to calculate average fish density for all of our surveys.

To quantify how much the spatially-varying fish density estimates ( $\log_{10}$  transformed) differed among frequencies and between seasons, we developed a functional analysis of variance (ANOVA) model (Kaufman and Sain, 2010) for each reservoir. We used this approach given that assumptions of independence and equal

variances were not met, which precluded the use of a simpler ANOVA. Each functional ANOVA model included a spatial trend (with terms for Northing, Easting, and their interaction), a spatially-varying term for frequency, a spatially-varying term for season, and a spatially-varying term for the errors. We assumed an exponential covariance for each spatially varying term. We only included a nugget term for the error term. The functional ANOVA model was fit using maximum likelihood (see Appendix B for further details).

### **3. Results**

#### **3.1 Fish collections**

The majority (by abundance) of the trawl catch was gizzard shad in Acton Lake (100%), whereas brook silverside was caught in greater abundance in Alum Creek Lake (78%) and Hoover Reservoir (74%). The mean TL ( $\pm 1$  SD) of gizzard shad captured in Acton Lake was  $54 \pm 19$  mm, in Alum Creek Lake was  $89 \pm 8$  mm, and in Hoover Reservoir was  $66 \pm 18$  mm. The mean TL of brook silverside was  $52 \pm 28$  mm in Alum Creek Lake, and  $59 \pm 15$  mm in Hoover Reservoir. The range of fish TLs recorded (regardless of species) was 20–95 mm in Acton Lake, 24–95 mm in Alum Creek Lake, and 30–90 mm in Hoover Reservoir.

#### **3.2 *Chaoborus* collections**

Depth-specific pump sampling confirmed the presence of *Chaoborus* in our study ecosystems (Table 2; Figure 2). We captured a wide size-range of *Chaoborus* in conjunction with our hydroacoustic surveys (Table 2; Figure 2). Similar patterns in the mean TL ( $\pm 1$  SD) of *Chaoborus* were observed across the three reservoirs sampled,

with a higher abundance of smaller *Chaoborus* being captured during summer ( $3.3 \pm 2.1$  mm) than during spring ( $5.9 \pm 2.9$  mm) (Mann-Whitney test;  $W = 483380$ ;  $P < 0.01$ ; Table 2; Figure 2). Further, more individuals were captured during summer (mean density ranged 256 to 688 individuals·m<sup>-2</sup>) than during spring (mean density ranged 43 to 229 individuals·m<sup>-2</sup>) in all three reservoirs (Table 2).

### 3.3 Hydroacoustic estimates of *Chaoborus*

The relationship between the hydroacoustic estimate of *Chaoborus* abundance and the estimate of *Chaoborus* biomass from pump samples varied with the frequency of transducer (200 vs. 430 kHz) and sampling season (Table 3; Figure 3). However, simple least-squares regression results indicated a significant relationship between the hydroacoustic and biomass estimates of *Chaoborus* for every frequency and time period sampled (Table 3). The models with the highest coefficients of determination ( $R^2$  ranged 0.94–0.97) included data from spring (May or June), whereas models from summer (August) had  $R^2$  values that ranged 0.36–0.52 (Table 3). The resultant hydroacoustic portrayal of *Chaoborus* is shown in Figure 4 (right panels).

Estimated slopes of these regression lines ranged 0.05–0.13 (Table 3) and significantly differed from 0.1 for all models. During spring, the TS of 1 g dry wt·m<sup>-3</sup> of *Chaoborus* was larger at 200 kHz (TS = -21.1 dB) than at 430 kHz (TS = -45.8 dB). The opposite pattern was found during summer, where a larger TS value was observed at 430 (TS = -13.1 dB) than at 200 kHz (TS = -37.2 dB). When all of the data were pooled (between seasons), the overall TS value for 1 g dry wt·m<sup>-3</sup> of *Chaoborus* was -35.4 dB at 200 kHz and -26.8 dB at 430 kHz.

### 3.4 Transducer frequency comparisons

Pairwise comparisons of total (i.e., from both fish and *Chaoborus*) NASC ( $\text{m}^2 \cdot \text{nm}^{-2}$ ) in the same cell between frequencies showed a consistent pattern of more backscattering at higher frequencies (Figure 5). Our analyses revealed significant differences ( $P = 0.01$ ) between total backscattering for every comparison, with the calculated slopes indicating that backscattering was greater at higher frequencies than at lower ones (Table 4). A visual representation of this finding can be seen in our snapshots of the frequency-specific thresholded echograms (Figure 4), where higher  $S_v$  values were observed at higher frequencies.

The frequency-dependent response of total (i.e., from both fish and *Chaoborus*) NASC was similar among reservoirs, but varied between seasons (Figure 6). During spring (May or June), we observed a hump-shaped curve in the frequency-response, with a peak at 200 kHz. However, during summer (August), we found an increasing trend in the frequency-response, with the echo energy return at 430 kHz almost twice that at 200 kHz (Figure 6). NASC values were highest in Hoover Reservoir during spring and in Acton Lake during summer, with Alum Creek Lake always being the lowest (Figure 6).

### **3.5 Fish density estimates**

Based on the observed increase in area backscattering with frequency (see Figures 4–6), we were not surprised that estimates of average fish density increased with the transducer frequency used during both seasons (Table 5). The average fish density calculated for Acton Lake during August at 200 kHz was 5.1 times greater than at 70 kHz. For Alum Creek Lake, it was 2.6 times greater, and for Hoover Reservoir, it was 3.6 times greater (Table 5). The magnitude of the difference in fish density

estimates between 120-kHz and 200-kHz transducers was usually smaller than when comparing density estimates calculated at 70 kHz to either of the two higher frequencies. No clear pattern among frequencies or among reservoirs was evident in observed differences in fish density estimates between spring and summer (Table 5). Generally, the standard error of the fish density estimate increased with increasing frequency during both seasons (Table 5).

Results from the functional ANOVA showed that differences in fish density estimates were attributable to multiple factors, with the amount of variance explained by each spatially varying parameter differing among reservoirs (Table 6). Across all three reservoirs, sampling season explained more variance in the fish density estimates than the transducer frequency (comparing partial sill parameters; Table 6). These changing patterns of acoustic backscattering between spring and summer likely represent changing population demographics (e.g., abundance, size distributions) of fish, *Chaoborus*, or both. Interestingly, spatial patterns in the fish density estimates differed among frequencies, as the largest range parameters and correlation at 100 m were found for the frequency term (except the range parameter for Acton Lake; see Table 5). The calculated nugget was similar among reservoirs (see Table 6), which was likely due to measurement error or other small-scale spatial features.

#### **4. Discussion**

The primary goal of our study was to improve the ability of fishery assessment biologists and researchers to use hydroacoustics to estimate fish density in the presence of pelagic macroinvertebrates that are residing in the water column. Towards



this end, we explored how estimates of fish density varied with hydroacoustic transducer frequency (70, 120, and 200 kHz) and sampling season (spring vs. summer) in three Ohio reservoirs with an abundance of *Chaoborus*, a vertically migrating macroinvertebrate that has air sacs, which resonate much like the air bladder of fish. Collectively, our findings demonstrate that pelagic macroinvertebrates can bias estimates of fish density, especially when a high-frequency transducer (e.g., 200 or 430 kHz) is used, and that the degree of bias is season-dependent, owing to changes in the demographics of *Chaoborus*. Below, we discuss these findings in detail and offer recommendations that can benefit the design and analysis of hydroacoustics assessment surveys of fish populations in the presence of non-target organisms such as *Chaoborus*.

#### **4.1 Bias associated with high-frequency transducers**

Our calculated estimates of fish density varied with transducer frequency. At 200 kHz, fish density estimates ranged 1.8- to 5.1-fold greater than those made with a 70-kHz transducer. Estimated fish densities at 120 kHz were intermediate to the 70- and 200-kHz transducers, though most were similar to those at 200 kHz. We attribute the differences in fish density among frequencies to the presence of *Chaoborus* in our study ecosystems. Because our assessment of total (i.e., both fish and *Chaoborus*) backscattering increased with increasing transducer frequency, we recommend using a lower frequency transducer (e.g., 70 kHz) when estimating fish density in the presence of *Chaoborus*. Our findings indicate that doing so would reduce bias, and hence, improve estimates of fish density.

Such wide differences in density estimates among frequencies were initially surprising, as recent work has shown that different frequencies can produce similar fish abundance estimates (Godlewska et al., 2009), especially when fish density is low (Guillard et al., 2014; Mouget et al., 2019). However, consideration of the presence of *Chaoborus* in our study ecosystems helps explain the observed differences in estimates of fish density. Previous studies have found that strong backscattering by *Chaoborus* can result in fish estimates that are biased high (see Eckmann, 1998; Malinen et al., 2005; Jurvelius et al., 2008; Knudsen and Larsson, 2009). The ability to calculate bias-free and robust estimates of fish density is important, as it can help understand fish population and food web dynamics, which also explains why it is a top priority for management agencies such as the ODNR-DOW (*sensu* Drastik et al., 2017; Dillon et al., 2019; JDC, ODNR-DOW, co-author, pers. comm.).

We are confident in our conclusion that *Chaoborus* is responsible for the discrepancy in fish density estimates across frequencies because no other known macroinvertebrate “scatterers” are known to reside in our surveyed ecosystems. While zooplankton were captured in our discrete-depth pump samples (Dillon et al., in review), Ohio reservoirs are generally characterized by small zooplankton taxa (e.g., Rotifera, *Eubosmina*; Bremigan and Stein, 1994; Vanni et al., 2005) that are unlikely to contribute acoustic backscattering. Furthermore, the larger cladoceran and copepod zooplankton that are present in our reservoirs have been found to contribute negligible amounts of backscatter at the frequencies investigated herein (Northcote, 1964; Frouzova et al., 2004). Instead, we argue that the observed increase in backscattering with increasing frequency resulted from *Chaoborus*, a macroinvertebrate known to migrate into the

water column at night, and which also has two pairs of air sacs (Teraguchi, 1975) that likely resonate at the higher frequencies used herein.

#### **4.2 Removal of bias using thresholding**

The method that we presented in this study to generate a less-biased estimate of fish density in the presence of macroinvertebrates involved simple thresholding with a low-frequency (70 kHz) transducer. As with many hydroacoustic analysis decisions, our ability to derive a bias-free estimate of fish density in the presence of *Chaoborus* can be viewed as a compromise between excluding backscattering from *Chaoborus*, and including that from fish (Malinen et al., 2001; *sensu* Simmonds and MacLennan, 2005). While other methods to achieve this goal are available in the literature (Eckmann, 1998; Wagner-Dobler and Jacobs, 1988; Malinen et al., 2005; Jurvelius et al., 2008), most of these methods require fish and macroinvertebrates to be separated in space or to have different target strength (TS) distributions (but, see Baran et al., 2019). These conditions, however, were never met for any hydroacoustic survey in our ecosystems. Even though fewer *Chaoborus* were present in the water column during the day compared to at night, we still observed *Chaoborus* at all depths at all times (Dillon et al., in review). Additionally, the schooling behavior of our main fish species of interest (gizzard shad), precluded us from sampling during the day, as it is established practice to estimate abundance of gizzard shad at night (Vondracek and Degan, 1995).

Our method of thresholding to generate a fish density estimate appears robust, as our selection of frequency-specific minimum TS values did not bias the differences in fish density that were observed among frequencies. According to hydroacoustic theory, fish density should increase as TS values become smaller. However, we selected a

smaller minimum TS value at 70 kHz than at either 120 or 200 kHz (Table A1). Thus, the higher estimates of fish density observed at either 120 or 200 kHz did not stem from our selection of the minimum TS value, and is more likely attributable to interference from *Chaoborus*. Further, across our reservoirs and both sampling seasons, the largest selected minimum TS value (-61 dB; Table A1) corresponded to an estimated fish total length (TL) of 17 mm (Love, 1977), which is a smaller-sized fish than was captured by our nets. The size-class of fish observed in Ohio reservoirs during the time of sampling (20–95 mm) is not sufficiently different from the known TS of *Chaoborus*; a 20 mm fish (TS = -59.9 dB at 200 kHz and -59.7 dB at 120 kHz; Love, 1977) would be within 6 dB of the TS of *Chaoborus* at both 200 kHz (TS = -60 to -64 dB; Jones and Xie, 1994; Knudsen et al., 2006) and 120 kHz (TS = -64 to -66 dB; Baran et al., 2019). Thus, 200 and 120 kHz appear to be *inappropriate* frequencies to use for estimation of fish abundance in the presence of *Chaoborus* (Knudsen et al., 2006; Jurvelius et al., 2008; Knudsen and Larsson, 2009; this study). Further, our transducer comparison identified 70 kHz as the frequency with the lowest total backscattering response and thus, most appropriate frequency to use for fish density estimation (of the frequencies tested).

Although we only estimated fish density within a 1-m layer of the water column (from 4.2 to 5.2 m depth), we would expect to find the same pattern (i.e., increasing fish density with increasing transducer frequency) had a larger portion of the water column been included in the analysis. We expect this pattern to hold as both gizzard shad and *Chaoborus* abundance are known to be greater at shallower rather than deeper depths at the time of sampling (i.e., at night; Dillon et al., 2019; Dillon et al., in review; JDC,

ODNR-DOW, coauthor, unpublished data). Thus, potentially even more bias from *Chaoborus* in fish density estimates would have been observed at shallower depths.

Unfortunately, we were constricted to conducting our analyses in a 1-m depth layer, as the acoustic near-field increases with decreasing transducer frequency, reducing the proportion of the water column sampled. In our case, the 2x near-field distance of the 70-kHz transducer was calculated to be 3.7 m, restricting our measurements to waters deeper than 4.2 m (depth location of the transducer plus the 2x near-field). Thus, to avoid bias from *Chaoborus*, at least in our ecosystems, our hydroacoustic estimates of fish may be biased low, due to the availability bias of our 70-kHz transducer (i.e., fish occupying depths of the water column that are within the near-field of the 70-kHz transducer). This tradeoff in biases (availability of fish versus *Chaoborus* backscatter) is important to be aware of, even though we have clearly demonstrated the importance of considering bias from *Chaoborus*. We suggest that an unbiased, relative index of fish abundance (i.e., reducing bias from *Chaoborus* by using a 70-kHz transducer) is better than a biased, absolute fish abundance estimate (i.e., using a 200-kHz transducer that samples a larger portion of the water column), as it would provide a more accurate estimate of fish abundance in the ecosystem. Once the extent of bias from *Chaoborus* in fish density estimates at higher compared to lower frequencies is quantified, as shown in this study, additional steps can then be taken to address the fish availability bias. Two potential solutions that hold the potential to overcome biases associated with the deep near-field associated with low-frequency (e.g., 70 kHz) transducers include: 1) using a transducer with a wide beam angle, which will shrink the near-field of the transducer; or 2) using an upward-facing rather than

downward-looking transducer (e.g., Baran et al., 2019). Collectively, these results highlight the need to be mindful of potential tradeoffs (e.g., other biases in fish density estimation) that might emerge when seeking to eliminate biases associated with pelagic macroinvertebrates such as *Chaoborus*. Recognition of these tradeoffs in choosing acoustics gear (i.e., transducers) is applicable to any ecosystem.

#### **4.3 Influence of season on frequency-dependent biases**

Interestingly, along with the observed transducer frequency responses, fish density estimates varied with the time period of sampling in our three study reservoirs. This result is not surprising when we consider that *Chaoborus* population demographics (e.g., length distributions, abundances) differed between spring (May or June) and summer (August); average lengths decreased from spring to summer. Our expectation is that these changes would influence estimates of fish density, with scattering at a lower frequency being greater during spring (when *Chaoborus* individuals are large) than during summer (when *Chaoborus* individuals are small). A size-dependent scattering-response of *Chaoborus* is unsurprising, as Knudsen et al. (2006) found that longer individuals had greater mean TS values relative to shorter individuals. Additionally, we found more *Chaoborus* during summer when backscattering was lower, than during spring, suggesting that size is more important than density when accounting for potential biases associated with *Chaoborus* backscattering.

Some research (e.g., Knudsen et al., 2006; Wagner-Dobler and Jacobs, 1988) has been skeptical regarding the use of acoustics to estimate *Chaoborus* densities. Contrary to this skepticism, however, we observed a significant relationship between *Chaoborus* biomass calculated from discrete-depth pump samples and the

corresponding hydroacoustic estimates of biomass ( $S_v$ ) in those depth-layers. While this relationship varied with transducer frequency (200 or 430 kHz) and sampling season, the observed predictive relationships were all strong and positive, with  $R^2$  values ranging 0.36–0.97. Collectively, the observed patterns in the frequency-dependent response, combined with our predictive *Chaoborus* hydroacoustic abundance models, provide added confidence that *Chaoborus* are driving variation in estimates of fish density among transducer frequencies. Variation in the frequency-dependent response between spring and summer also indicates that seasonal changes in estimated fish density can arise, not only because of demographic changes in the fish population, but also because of demographic variation in the resident *Chaoborus* population. These findings demonstrate the need to consider the dynamics of *Chaoborus*, or other pelagic macroinvertebrates, when choosing what season to assess the target fish population with hydroacoustics.

## 5. Conclusions

Hydroacoustic estimates of fish density varied considerably across our study ecosystems, owing to both characteristics of the sampling gear used (i.e., transducer frequency) and the resident biota (i.e., *Chaoborus*). The large observed differences in estimates of fish density at different frequencies appear primarily due to the frequency-dependent backscattering response of *Chaoborus*. Specifically, we found estimates of fish density to be 1.8- to 5.1-fold higher with a 200-kHz transducer than with a 70-kHz transducer across our three study reservoirs. We also found that total (i.e., combined fish and *Chaoborus*) backscattering increased with increasing transducer frequency,

with the greatest total backscattering always observed at the higher frequencies (200 kHz and 430 kHz, depending on the season of sampling). We are confident that *Chaoborus* in the water column is the source of our overestimation of fish density, as we found high *Chaoborus* densities in our study ecosystems, as well as strong correlations between hydroacoustic estimates of *Chaoborus* and observed biomass in pump samples. Overall, we stress the need to consider and reduce bias associated with the presence of pelagic macroinvertebrates such as *Chaoborus* during hydroacoustic surveys, which may change seasonally due to changes in their demographics (e.g., length, abundance). Because both empirical and theoretical data suggest that small *Chaoborus* individuals resonate less than larger individuals, even when at a high density, we recommend conducting fish abundance surveys during times when *Chaoborus* are small in size, if practical. Additionally, we recommend choosing a low-frequency (e.g., 70 kHz) transducer for sampling fish wherever possible—perhaps mounted in an upward-facing direction in shallow ecosystems—to also help reduce potential bias from vertically migrating *Chaoborus* from the outset. Following these recommendations would offer researchers and fishery management agencies alike a means to generate more robust estimates of fish abundance in ecosystems that also support large populations of pelagic macroinvertebrates.

## **Acknowledgments**

This project was funded by the Federal Aid in Sport Fish Restoration Program (F-69-P, Fish Management in Ohio) administered jointly by the United States Fish and Wildlife Service and the Division of Wildlife-Ohio Department of Natural Resources



641 (ODNR-DOW; project FADR79 to SAL, JDC, and RAD). Additional support came from  
642 the US National Science Foundation (NSF) under grants NSF-DMS-1407604 and NSF-  
643 SES-1424481 (to PFG). Thank you to the many people involved in completing the fish  
644 assessments, including staff from the Aquatic Ecology Laboratory and ODNR-DOW.  
645 Thanks to the University of Toledo for the use of their 430-kHz transducer. We  
646 appreciate the constructive criticism received from one anonymous reviewer, which  
647 greatly improved a previous version of this manuscript.

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## TABLES

**Table 1.** Characteristics of the three Ohio reservoirs sampled for this study during 2017.

Surface area, average depth ( $z_{Avg}$ ), and maximum ( $z_{Max}$ ) depth are reported for the entire reservoir. Total phosphorus concentration (TP) and chlorophyll *a* concentration (Chl*a*) were measured as part of standard Ohio Department of Natural Resources-Division of Wildlife water quality surveys.

Reservoir	Surface area (km <sup>2</sup> )	$z_{Avg}$ (m)	$z_{Max}$ (m)	TP (µg·L)	Chl <i>a</i> (µg·L)	Survey date
Acton Lake	2.4	3.4	9.5	85.7	57.7	May 30
						August 14
Alum Creek Lake	13.5	6.6	19.3	19.8	8.8	May 25
						August 28
Hoover Reservoir	11.7	5.7	20.9	40.3	24.4	June 8
						August 31

**Table 2.** Mean density (# individuals·m<sup>-2</sup> ± 1 SD) and mean total length (mm ± 1 SD) of *Chaoborus* captured at night in discrete-depth pump samples in Acton Lake, Alum Creek Lake, and Hoover Reservoir (Ohio) during spring (May or June) and summer (August), 2017.

Reservoir	Attribute	Spring	Summer
Acton Lake	density	229 ± 14	688 ± 273
	length	7.3 ± 1.8	3.7 ± 2.5
Alum Creek Lake	density	43 ± 52	309 ± 35
	length	4.0 ± 2.9	2.8 ± 1.5
Hoover Reservoir	density	82 ± 28	256 ± 91
	length	6.2 ± 3.0	3.5 ± 2.0

805 **Table 3.** Results from least-squares regressions used to quantify the relationship between hydroacoustic estimates of  
806 *Chaoborus* acoustic energy ( $S_v$ , dB; a proxy for biomass) and *Chaoborus* biomass ( $\log_{10}$  (g dry wt·m<sup>-3</sup>)) from discrete-  
807 depth pump samples collected in three Ohio reservoirs (Acton Lake, Alum Creek Lake, and Hoover Reservoir) during  
808 2017. Four regressions (see “Data” column) were conducted corresponding to the two frequencies (200 and 430 kHz) and  
809 two seasons (Spring: May or June; Summer: August) of hydroacoustic data collection. We also calculated the target  
810 strength (TS, dB) of 1 g dry wt·m<sup>-3</sup> of *Chaoborus* for each dataset when the slope of the line did not significantly differ from  
811 0.1. TS values with an asterisk (\*) were calculated after forcing the slope of the regression line for that model (August, 200  
812 kHz) to equal 0.1, as the original slope differed from 0.1. Significant terms ( $\alpha = 0.05$ ) are presented in bold.

Data	Variable	Slope $\pm$ 1 SE	P	Overall model	R <sup>2</sup>	TS
Spring, 200 kHz	Intercept	<b>2.11 <math>\pm</math> 0.64</b>	<b>0.03</b>	$F_{1,4} = 59.08$ , $P < 0.01$	0.94	-21.1
	$S_v$	<b>0.08 <math>\pm</math> 0.01</b>	<b>&lt; 0.01</b>			
Spring, 430 kHz	Intercept	<b>4.58 <math>\pm</math> 0.68</b>	<b>&lt; 0.01</b>	$F_{1,4} = 115.74$ , $P < 0.01$	0.97	-45.8
	$S_v$	<b>0.13 <math>\pm</math> 0.01</b>	<b>&lt; 0.01</b>			
Summer, 200 kHz	Intercept	0.50 $\pm$ 1.11	0.66	$F_{1,11} = 6.11$ , $P = 0.03$	0.36	-37.2*
	$S_v$	<b>0.05 <math>\pm</math> 0.02</b>	<b>0.03</b>			
Summer, 430 kHz	Intercept	1.31 $\pm$ 1.33	0.36	$F_{1,7} = 7.57$ , $P = 0.03$	0.52	-13.1
	$S_v$	<b>0.07 <math>\pm</math> 0.03</b>	<b>0.03</b>			

813

814

**Table 4.** Statistics describing pairwise comparisons (x-axis:y-axis) of frequency-specific (70, 120, 200, and 430 kHz) estimates of total nautical area scattering coefficient (NASC, m<sup>2</sup>·nmi<sup>-2</sup>), calculated with major-axis regression. All slopes were significantly greater than one ( $\alpha = 0.05$ ). NASC includes all scattering after noise was subtracted from the data. Data were not thresholded. CI = confidence interval. All data were collected in three Ohio reservoirs during spring (May or June) and summer (August), 2017.

Comparison	R <sup>2</sup>	P	Slope	Slope: 95% CI	Intercept	Intercept: 95% CI
70:120 kHz	0.74	0.01	2.93	2.83, 3.04	3.45	-2.87, 9.39
70:200 kHz	0.79	0.01	3.67	3.56, 3.78	30.49	23.54, 37.04
70:430 kHz	0.38	0.01	8.86	8.23, 9.59	-92.90	-136.32, -55.58
120:200 kHz	0.78	0.01	1.30	1.26, 1.35	16.34	8.84, 23.54
120:430 kHz	0.47	0.01	2.79	2.62, 2.97	-61.85	-95.30, -31.97
200:430 kHz	0.66	0.01	1.87	1.80, 1.96	-32.11	-52.80, -12.77

**Table 5.** Statistics from the spatial models used to describe frequency-specific (70, 120, and 200 kHz) estimates of mean fish density (# individuals·m<sup>-2</sup>) in Acton Lake, Alum Creek Lake, and Hoover Reservoir (Ohio) during spring (May or June) and summer (August), 2017. Thresholds applied are in Table A1. SE = standard error; CI = confidence interval.

Reservoir	Frequency	Spring			Summer		
		Density	SE	95% CI	Density	SE	95% CI
Acton Lake	70 kHz	0.38	0.01	0.37, 0.39	0.47	0.01	0.44, 0.49
	120 kHz	0.77	0.03	0.72, 0.82	0.68	0.03	0.63, 0.74
	200 kHz	0.69	0.03	0.64, 0.74	2.40	0.20	2.05, 2.83
Alum Creek Lake	70 kHz	1.06	0.08	0.91, 1.23	0.32	0.00	0.31, 0.33
	120 kHz	1.37	0.07	1.25, 1.51	0.58	0.02	0.54, 0.62
	200 kHz	3.02	0.22	2.62, 3.47	0.84	0.04	0.77, 0.91
Hoover Reservoir	70 kHz	0.69	0.03	0.63, 0.76	0.61	0.02	0.57, 0.64
	120 kHz	1.41	0.04	1.34, 1.48	2.18	0.13	1.93, 2.44
	200 kHz	1.74	0.08	1.60, 1.90	2.19	0.12	1.95, 2.44

**Table 6.** Results from the functional ANOVA used to determine the difference between fish density estimates (# individuals·m<sup>-2</sup>; see Table 4) calculated at the three different transducer frequencies (70, 120, and 200 kHz) during two seasons (Spring: May or June; Summer: August) in Acton Lake, Alum Creek Lake, and Hoover Reservoir, 2017. The partial sill indicates the variance accounted for by each spatial term in the functional ANOVA model, and the range parameter and correlation at 100 m show the relationship between the terms in the model and distance. A larger range parameter indicates strong spatial covariance of that term in the model. The estimated nugget term for the error terms are: Acton Lake = 0.23; Alum Creek Lake = 0.12; and Hoover Reservoir = 0.16.

Reservoir	Parameter	Partial Sill	Range Parameter	Correlation at 100 m
Acton Lake	Intercept	<0.01	0.21	0.98
	Season	0.16	0.21	0.62
	Frequency	0.05	0.19	0.58
	Error	0.18	2.38	0.41
Alum Creek Lake	Intercept	0.04	0.15	0.52
	Season	0.21	<0.01	<0.01
	Frequency	0.09	1.59 x 10 <sup>5</sup>	1.00
	Error	0.20	0.12	0.27
Hoover Reservoir	Intercept	0.08	0.03	0.05
	Season	0.20	0.04	0.09
	Frequency	<0.01	11.16	0.99
	Error	0.02	5390.01	0.09

## FIGURE CAPTIONS

**Figure 1.** Location of Ohio in North America (A), and the survey design in Acton Lake (B), Alum Creek Lake (C), and Hoover Reservoir (D). The hydroacoustic (solid lines), trawling (dashed lines with circle end caps), and abiotic/discrete-depth pump sampling (squares) sites are displayed on each map.

**Figure 2.** Length distribution of *Chaoborus* from discrete-depth pump samples collected in Acton Lake, Alum Creek Lake, and Hoover Reservoir (Ohio) during 2017. These data were used to build the predictive models between observed *Chaoborus* biomass and hydroacoustic estimates of *Chaoborus* abundance. Each column represents a different season (Spring: May or June; Summer: August) sampled and each row is a specific reservoir. Sampling in the additional reservoirs is described in Appendix C.

**Figure 3.** Relationships between hydroacoustic estimates of *Chaoborus* mean volume backscattering strength ( $S_v$ ), which is a proxy for biomass, and estimated *Chaoborus* biomass (dry wt) in discrete-depth pump samples collected in Acton Lake, Alum Creek Lake, and Hoover Reservoir (Ohio) during 2017. Data are presented from multiple reservoirs, years, times of day, frequencies (200 kHz = left panels; 430 kHz = right panels), and seasons (spring: May or June = top panels; summer: August = bottom panels).

**Figure 4.** Hydroacoustic echograms showing the mean volume backscattering strength ( $S_v$ ) of fish and *Chaoborus* at multiple transducer frequencies (70, 120, and 200 kHz for fish; 430 kHz for *Chaoborus*) in Acton Lake (top panels), Alum Creek Lake (middle panels), and Hoover Reservoir (bottom panels) during August 2017. Fish



echograms were thresholded with frequency-specific minimum TS values (Table A1). The *Chaoborus* echogram was created by masking fish from the 70-kHz echogram over  $S_v$  from the 430-kHz echogram. Columns correspond to transducer frequency, which increase from left to right. The horizontal axis is 250 m in length. Note: the near-field of each transducer is frequency-specific and not indicated on the echograms.

**Figure 5.** Relationships between total (i.e., both fish and *Chaoborus*) cell-specific NASC for pairwise combinations of transducer frequencies (70, 120, 200 and 430 kHz) during spring (May or June; black circle) and summer (August; open square) in Acton Lake, Alum Creek Lake, and Hoover Reservoir (Ohio) during 2017. The solid line is the best-fit major-axis regression line for data from both spring and summer. Data were not thresholded, although background noise was removed.

**Figure 6.** Frequency-specific (70, 120, and 200 kHz ) response of total backscattering (i.e., both fish and *Chaoborus*) in Acton Lake, Alum Creek Lake, and Hoover Reservoir (Ohio) during 2017. Values are mean  $\pm$  1 SE NASC from one third of the number of cells sampled in each reservoir at night during spring (May or June) and summer (August). Data were not thresholded, although background noise was removed.

881 **Appendix A.**

882 **Table A.1.** The frequency-specific (70, 120, and 200 kHz) fish target strength threshold (TS min; dB) applied in Acton

883 Lake, Alum Creek Lake, and Hoover Reservoir (Ohio) during Spring (May or June) and Summer (August), 2017.

Reservoir	Survey date	TS min: 70 kHz	TS min: 120 kHz	TS min: 200 kHz
Acton Lake	May 30	-61	-60	-63
	August 14	-65	-60	-63
Alum Creek Lake	May 25	-63	-62	-63
	August 28	-63	-62	-63
Hoover Reservoir	June 8	-62	-60	-63
	August 31	-63	-62	-64

## A. FIGURE CAPTIONS

**Figure A.1.** Estimated covariance structure of the geostatistical model for all transducer frequencies (70, 120, and 200 kHz) used to sample fish during May and August 2017 in Acton Lake, Ohio. Except for the 200-kHz transducer in August, the spatial correlation was close to zero at 0.2 km.

**Figure A.2.** Estimated covariance structure of the geostatistical model for all transducer frequencies (70, 120, and 200 kHz) used to sample fish during May and August 2017 in Alum Creek Lake, Ohio. Except for the 70-kHz and 200-kHz transducer in May, the spatial correlation is close to zero at 0.2 km.

**Figure A.3.** Estimated covariance structure of the geostatistical model for all transducer frequencies (70, 120, and 200 kHz) used to sample fish in during June and August 2017 in Hoover Reservoir, Ohio. For all frequencies and both months, the spatial correlation was close to zero at 0.2 km.

## Appendix B.

### B.1. Spatial modeling of fish abundance

In a given reservoir, we recorded locations  $s = (n, e)^T$  in Northing ( $n$ ) and Easting ( $e$ ) coordinates in units of km, measured relative to the centroid of the reservoir. Our spatial domain of interest was  $D \subset \mathbf{R}^2$ , a contiguous and convex set of locations with a water depth of at least 4 m.

Then for a given reservoir, frequency, and season, we had  $m$  fish density estimates  $(Y(s_i) : i = 1, \dots, m)$ , at locations  $s_i = (n_i, e_i)^T$ . Then,  $Z(s_i) = \log(Y(s_i) + 0.5)$  denoted our shifted and transformed estimates with  $Z = (Z(s_1), \dots, Z(s_m))^T$ .

We assumed that the transformed ( $\log_{10}$ ) density estimates over the region  $D, \{Z(s) : s \in D\}$ , was a geostatistical process. More specifically, we had  $\{Z(s)\}$  as a Gaussian process (GP) with mean  $\mu(s)$  and covariance  $C(s, s')$  for locations  $s$  and  $s'$  in  $D$ . The spatial trend model we used at location  $s$  was:

$$\mu(s_i) = \beta_0 + \beta_1 n_i + \beta_2 e_i + \beta_3 n_i e_i.$$

The covariance between locations  $s$  and  $s'$  had the form

$$C(s, s') = \theta_1 \exp(-d(s, s')/\theta_2) + \theta_3 I(s = s'), \quad (\text{B.1})$$

where  $\theta_1$  was the partial sill parameter,  $\theta_2$  the range parameter,  $\theta_3$  the nugget parameter,  $I(\cdot)$  the indicator function, and  $d(s, s')$  was the Euclidean distance between  $s$  and  $s'$ .

We estimated the trend parameters  $\beta = (\beta_0, \beta_1, \beta_2, \beta_3)^T$  and spatial covariance parameters  $\theta = (\theta_1, \theta_2, \theta_3)^T$  from the data  $Z$  using maximum likelihood methods (e.g., Cressie, 1991). We used R code (R Core Team, 2018), available from

924 <https://github.com/petercraigmile/GSP>, to fit the geostatistical models to each reservoir,  
 925 frequency, and time period sampled.

926

## 927 **B.2. Predicting average fish density**

928 Letting  $|D| = \int_{s \in D} ds$  denote the area of  $D$ , the average fish density over  $D$ , calculated  
 929 from the geostatistical density process  $\{Y(s)\}$  was

$$930 \quad \frac{1}{|D|} \int_{s \in D} Y(s) ds.$$

931 Using a set of  $K$  prediction locations  $p_1, \dots, p_K$  spaced 50 m apart, but restricted to cover  
 932  $D$ , we estimated this integral using

$$933 \quad \frac{1}{K} \sum_{k=1}^K Y(p_k).$$

934 However, we do not know the actual values of the process  $Y(\cdot)$  at all locations in  $D$ .

935 Instead, using our model, we repeatedly obtained predictions of the shifted and  $\log_{10}$ -  
 936 transformed process  $Z(\cdot)$  at the prediction locations  $p_1, \dots, p_K$ , transformed the  
 937 predictions back to the original scale, and then summarized these simulations.

938 Then, let  $\mu = (\mu(s_1), \dots, \mu(s_m))^T$  denote the spatial trend at the data locations, and

939  $\eta = (\mu(s_1), \dots, \mu(s_K))^T$  denote the spatial trend at the predicted locations. Let  $V =$

940  $[C(s_i, s_j) : i = 1, \dots, m, j = 1, \dots, m]$  denote the covariance matrix for the data  $Z, P =$

941  $[C(p_i, p_j) : i = 1, \dots, K, j = 1, \dots, K]$  denote the covariance matrix at the predicted

942 locations, and  $C = [C(p_i, s_j) : i = 1, \dots, K, j = 1, \dots, m]$  denote the covariance between the

943 predicted and data locations.

Then, a prediction of the shifted and log-transformed process at all the prediction locations simultaneously,  $\tilde{Z}$ , was drawn from a  $K$ -variate normal distribution with mean

$$\eta + CV^{-1}[Z - \mu]$$

and covariance

$$P - CV^{-1}C^T.$$

To provide an unbiased estimate of the abundance with measures of uncertainty (SEs and CIs), we repeatedly obtained 1000 sets of predicted values of this process,

$\tilde{Z}^{(l)}, l = 1, \dots, 1000$ . Then,  $\tilde{Z}^{(l)}(p_i)$  denoted the  $l$ th prediction at location  $p_i$  and

$$\tilde{Y}^{(l)}(p_i) = \exp\left(\tilde{Z}^{(l)}(p_i)\right) - 0.5$$

was the back-transformed prediction on the original density scale. Our estimate of the average fish density from the  $l$ th prediction was:

$$A_l = \frac{1}{K} \sum_{k=1}^K \tilde{Y}^{(l)}(p_k).$$

We used the average of the 1000  $A_l$  values as our estimate of the average fish density over  $D$  and the standard deviation of the values as our SE for the average fish density over  $D$ . We obtained a 95% confidence interval (CI) for the average fish density by using 0.025 and 0.975 quantiles of the  $A_l$  values.

### B.3. Functional ANOVA model

We used a functional analysis of variance (ANOVA) (Kaufman and Sain, 2010) to model the spatially-varying relationship between frequency and season. While Kaufman

964 and Sain (2010) used a Bayesian framework, we used a frequentist approach, fitting the  
 965 model in R using maximum likelihood.

966 For each reservoir, we let  $D$  denote the spatial domain of interest in the reservoir.  
 967 We let  $i = 1, 2$  denote spring (May or June) or summer (August) and let  $j = 1, 2, 3$  denote  
 968 70, 120, and 200 kHz. We then let  $Z_{ij}(s)$  be our shifted and  $\log_{10}$  transformed fish  
 969 density estimate at location  $s \in D$ . We modeled the following contrasts:

$$970 \quad W_{11}(s) = Z_{12}(s) - Z_{11}(s) \quad (120 \text{ kHz minus } 70 \text{ kHz, spring});$$

$$971 \quad W_{12}(s) = Z_{13}(s) - Z_{11}(s) \quad (200 \text{ kHz minus } 70 \text{ kHz, spring});$$

$$972 \quad W_{21}(s) = Z_{22}(s) - Z_{21}(s) \quad (120 \text{ kHz minus } 70 \text{ kHz, summer});$$

$$973 \quad W_{22}(s) = Z_{23}(s) - Z_{21}(s) \quad (200 \text{ kHz minus } 70 \text{ kHz, summer});$$

974 with the functional ANOVA model

$$975 \quad W_{ij}(s) = \mu(s) + \alpha_i(s) + \delta_j(s) + \epsilon_{ij}(s), \quad i = 1, 2, j = 1, 2, s \in D.$$

976 We then defined the different spatially-varying terms in the model. We assumed  
 977 that the spatially-varying intercept term,  $\{\mu(s) : s \in D\}$ , was a Gaussian process (GP)  
 978 with mean  $k_\mu(s)$  and an exponential covariance with partial sill  $\theta_{\mu,1}$  and range parameter  
 979  $\theta_{\mu,2}$ :

$$980 \quad C_{\theta_\mu}(s, s') = \theta_{\mu,1} \exp(-d(s, s') / \theta_{\mu,2}) \quad (\text{B.2})$$

981 The mean term  $k_\mu(s)$  was used to capture covariate effects over space; again, we  
 982 included the Northing, Easting, and their interaction.

983 Following Kaufman and Sain (2010), we supposed that the spatially-varying  
 984 season effect  $\{\alpha_i(s) : s \in D, i = 1, 2\}$  was a GP, with zero mean and covariance

$$\text{cov}(\alpha_i(s), \alpha_{i'}(s')) = \begin{cases} \frac{C_{\theta_\alpha}(s, s')}{2}, & i = i'; \\ \frac{-C_{\theta_\alpha}(s, s')}{2}, & i \neq i', \end{cases}$$

where  $C_{\theta_\alpha}$  was an exponential covariance with partial sill  $\theta_{\alpha,1}$  and range parameter  $\theta_{\alpha,2}$  (defined similarly to (B.2)). Similarly, we assumed the spatially-varying frequency contrast effects  $\{\delta_j(s) : s \in D, j = 1, 2\}$  was a GP with zero mean and covariance

$$\text{cov}(\delta_j(s), \delta_{j'}(s')) = \begin{cases} \frac{C_{\theta_\delta}(s, s')}{2}, & j = j'; \\ \frac{-C_{\theta_\delta}(s, s')}{2}, & j \neq j', \end{cases}$$

where  $C_{\theta_\delta}$  was an exponential covariance with partial sill  $\theta_{\delta,1}$  and range parameter  $\theta_{\delta,2}$  (defined similarly to equation B.2).

To complete the model, we assumed that the spatially-varying error term  $\{\epsilon_{ij}(s) : s \in D\}$  was an independent GP (over  $i$  and  $j$ ) with mean zero and exponential covariance with partial sill  $\theta_{\epsilon,1}$ , range parameter  $\theta_{\epsilon,2}$ , and nugget parameter  $\theta_{\epsilon,3}$ , (defined similarly to equation B. 1)). Note that only the error term  $\{\epsilon_{ij}(s)\}$  contained a nugget term to account for possible measurement error, or short-range spatial effects.

## References

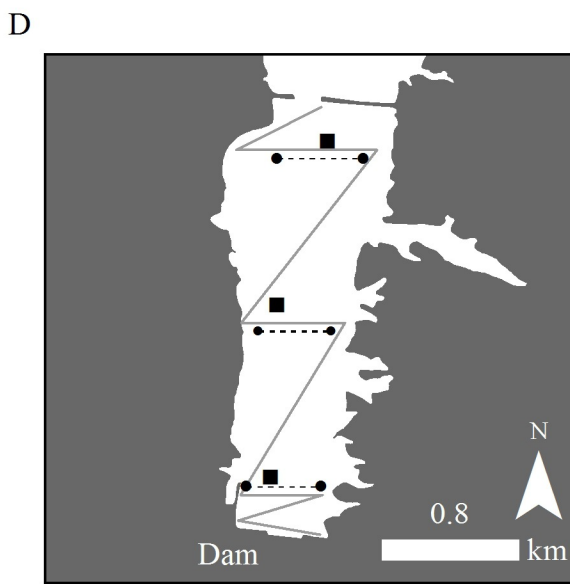
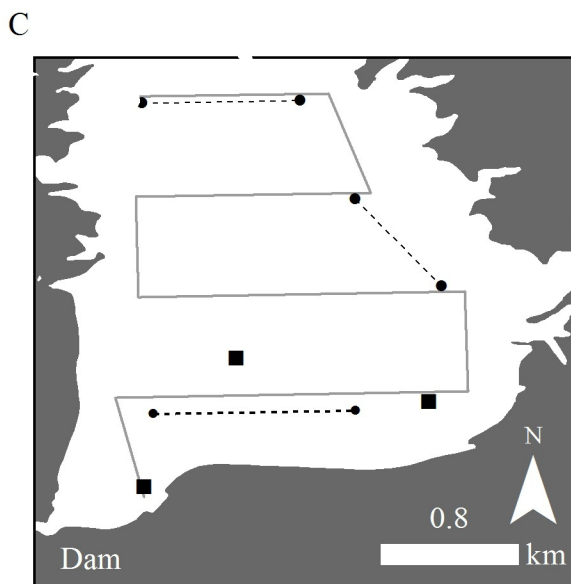
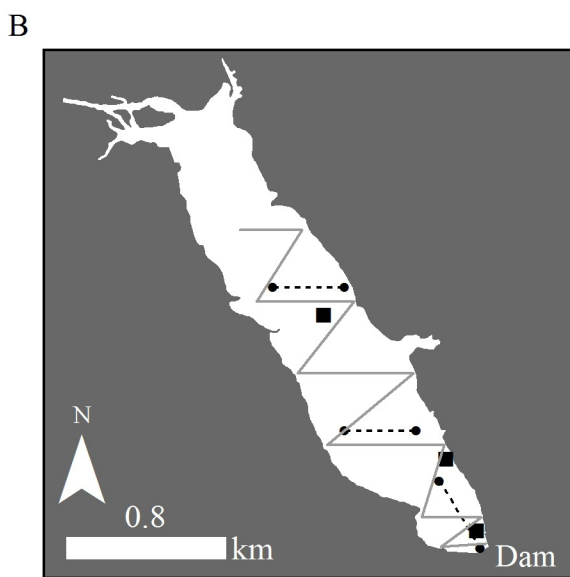
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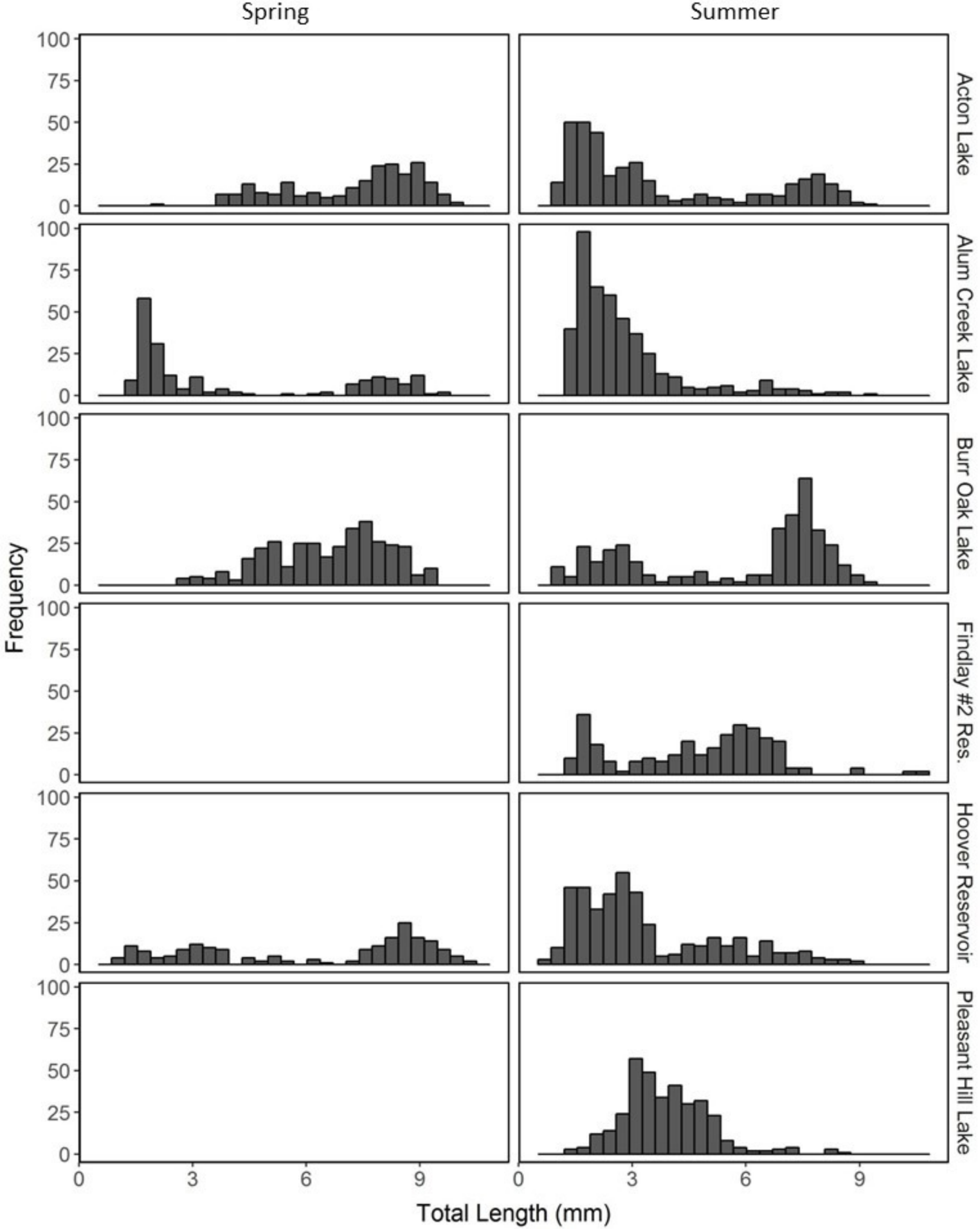


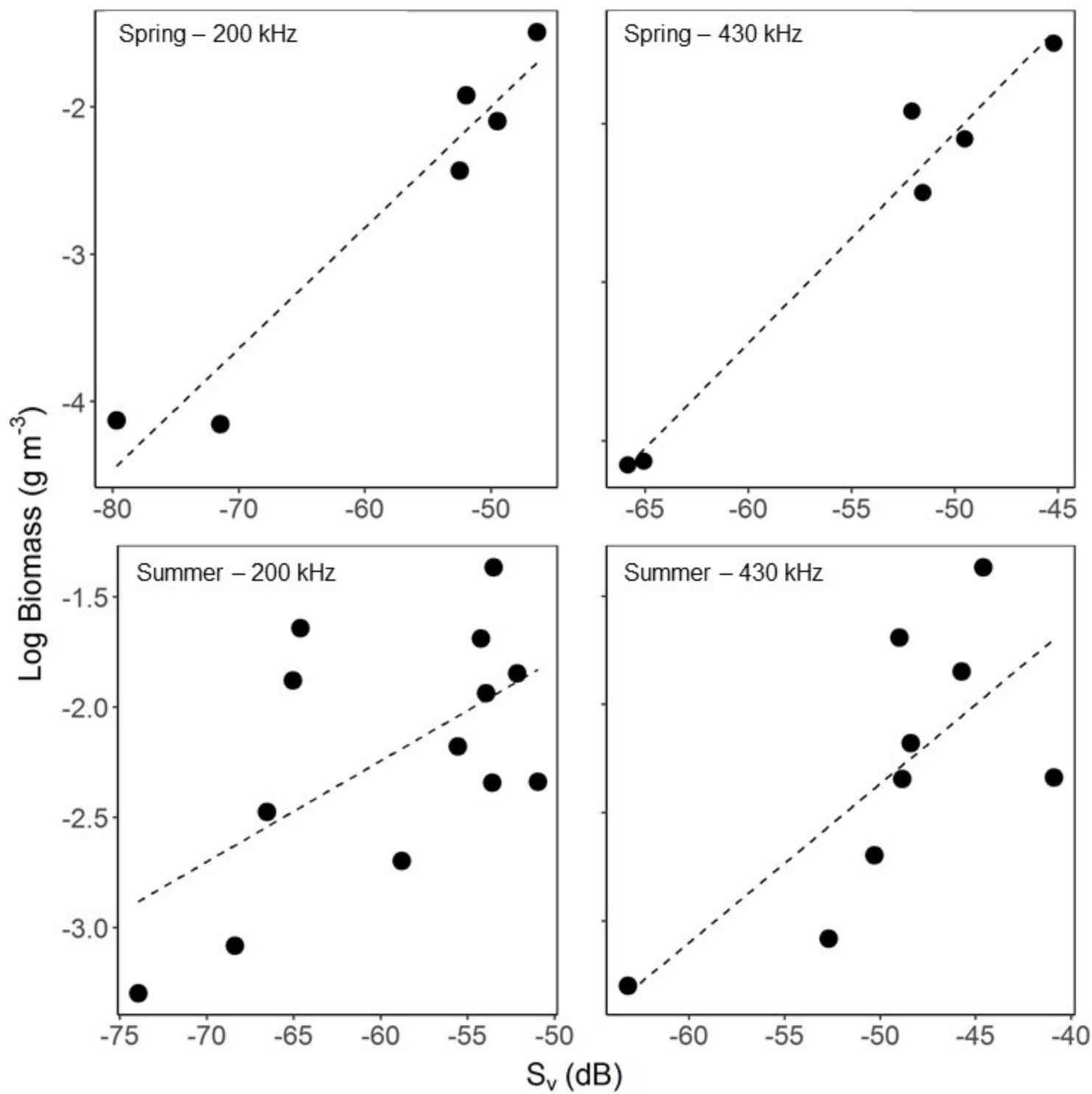
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1005 **Foundation for Statistical Computing, Vienna, Austria. URL [https://www.R-](https://www.R-project.org/)**  
1006 **[project.org/](https://www.R-project.org/).**  
1007

## Appendix C.

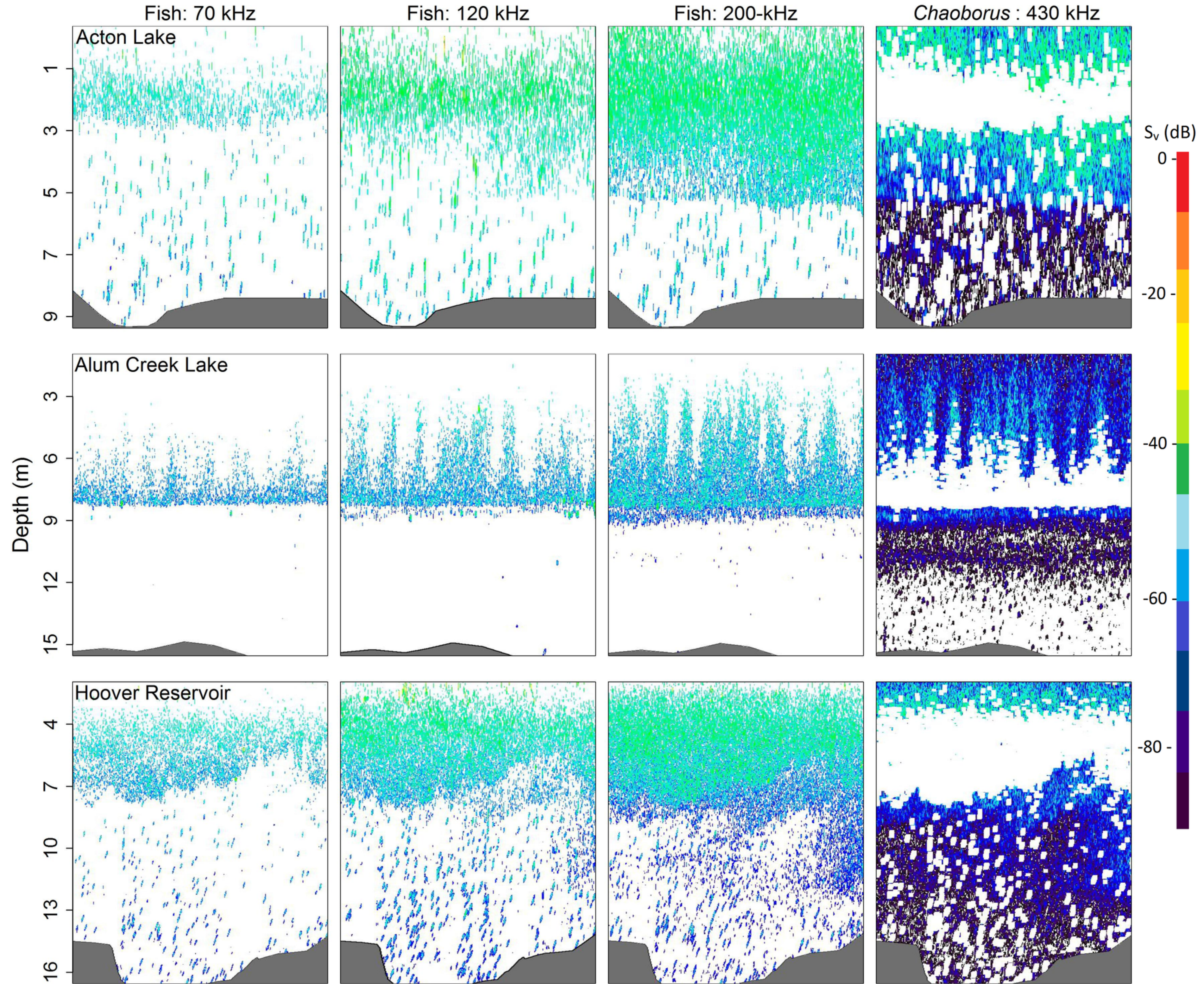
To develop our hydroacoustic method to estimate *Chaoborus* abundance, we supplemented the data collected during 2017, as described in this study, with additional collections. Specifically, we also conducted hydroacoustic surveys and trawling during the day (as well as at night) for the reservoirs and dates that were described in the main text (section 2.1). Alum Creek Lake and Hoover Reservoir were also sampled in their entirety at night during August 2016, not just in the lower dam as with our 2017 sampling. An additional four reservoirs were sampled at night during August 2016 and 2017, including Burr Oak Lake, Pleasant Hill Lake, and Findlay #2 Reservoir (Findlay #2 Reservoir was only sampled during 2017). Similar hydroacoustic survey designs (i.e., zig-zag pattern; section 2.3) to the main manuscript were used in all additional reservoirs.



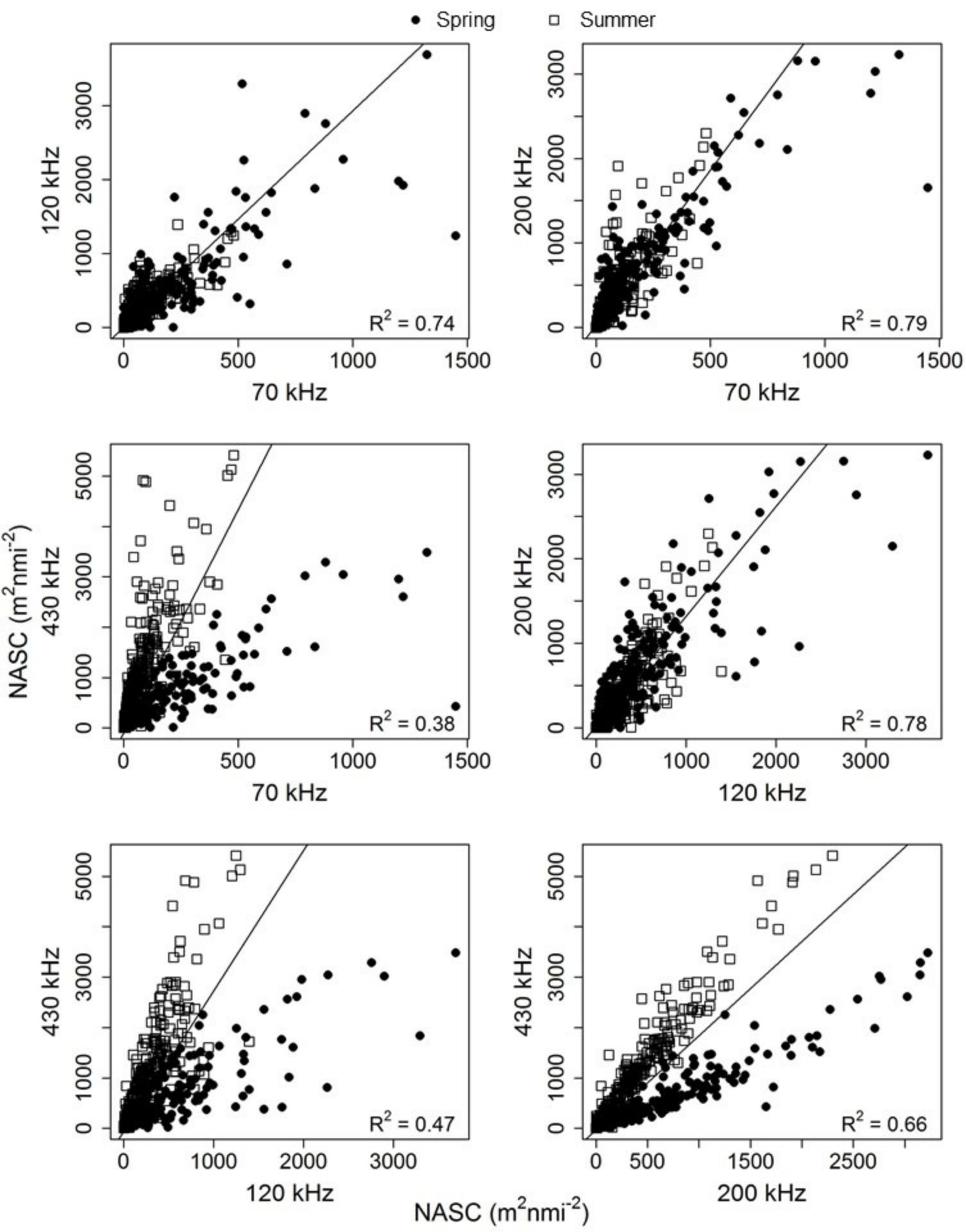












Spring

Summer

