

1 **The crucial role of genome-wide genetic variation in conservation**

2 Marty Kardos^{1*}, Ellie E. Armstrong², Sarah Fitzpatrick³, Samantha Hauser⁴, Philip W. Hedrick⁵,
3 Joshua M. Miller^{6,7,8}, David A. Tallmon⁹, W. Chris Funk¹⁰

4
5 1. Northwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and
6 Atmospheric Administration, Seattle, Washington 98112, USA

7
8 2. Department of Biology, Stanford University, 327 Campus Drive, Stanford, CA 94305, USA

9
10 3. W.K. Kellogg Biological Station; Department of Integrative Biology; Ecology, Evolution, and
11 Behavior Program, Michigan State University, 3700 Gull Lake Drive, Hickory Corners, MI 49060,
12 USA

13
14 4. Department of Biological Sciences, University of Wisconsin – Milwaukee, P.O.
15 Box 413, Milwaukee, WI 53211, USA

16
17 5. School of Life Sciences, Arizona State University, Tempe, AZ 85287, USA

18
19 6. San Diego Zoo Wildlife Alliance, 15600 San Pasqual Valley Road, Escondido, CA 92027, USA

20
21 7. Polar Bears International, PO Box 3008, Bozeman, MT, USA

22
23 8. Department of Biological Sciences, MacEwan University, Edmonton, Alberta, CA T5J 4S2

24
25 9. Biology and Marine Biology Program, University of Alaska Southeast, 11066 Auke Lake Way
26 Juneau, AK 99801, USA

27
28 10. Department of Biology, Graduate Degree Program in Ecology, Colorado State University, Fort
29 Collins, CO 80523, USA

30
31 * Address correspondence to martin.kardos@noaa.gov

32
33 **Keywords:** genomics, population dynamics, inbreeding depression, adaptative potential, extinction
34

43 **Abstract**

44 The unprecedented rate of extinction calls for efficient use of genetics to help conserve
45 biodiversity. Several recent genomic and simulation-based studies have argued that the field of
46 conservation biology has placed too much focus on conserving genome-wide genetic variation,
47 and that the field should instead focus on managing the subset of functional genetic variation that
48 is thought to affect fitness. Here, we critically evaluate the feasibility and likely benefits of this
49 approach in conservation. We find that population genetics theory and empirical results show that
50 conserving genome-wide genetic variation is generally the best approach to prevent inbreeding
51 depression and loss of adaptive potential from driving populations towards extinction. Focusing
52 conservation efforts on presumably functional genetic variation will only be feasible occasionally,
53 often misleading, and counterproductive when prioritized over genome-wide genetic variation.
54 Given the increasing rate of habitat loss and other environmental changes, failure to recognize the
55 detrimental effects of lost genome-wide genetic variation on long-term population viability will
56 only worsen the biodiversity crisis.

57

58 **Introduction**

59 Decades of theoretical (1) and empirical (2, 3) research suggest that conserving genome-wide
60 genetic variation improves population viability. Maintaining genetic variation, *adaptive potential*
61 (see Glossary), and avoiding *inbreeding depression* are central motivations for maintaining large,
62 connected natural populations. Principles of genetics and evolution have therefore played a large
63 role in conservation biology since its inception (4, 5). The genomics revolution has inspired
64 biologists to leverage genome analysis to advance conservation beyond what was possible with
65 traditional genetics. Numerous studies have sequenced genomes of non-model organisms of
66 conservation concern to understand population history, *inbreeding* depression, and the genetic
67 basis of adaptation. A particularly exciting area of research has been to determine when and how
68 functional genetic information can advance conservation.

69 Several recent studies suggest that too much emphasis has been placed on genome-wide
70 genetic variation in conservation biology. For example, persistence of small populations for long
71 periods of time despite low genetic variation, and the collapse of the Isle Royale wolf population
72 after the infusion of genetic variation via immigration, have been interpreted as a challenge to the
73 idea that genetic variation generally increases population viability (6-12). Additionally, a weak
74 relationship between conservation status and genetic variation has been used to argue that genome-

75 wide (presumably neutral) genetic variation is of little importance to conservation (11). Several
76 authors have thus advocated for an approach that focuses on functional genetic variation that is
77 thought to directly affect fitness (including minimizing deleterious genetic variation) in place of
78 the traditional emphasis on conserving genome-wide genetic variation (6-8, 11).

79 Here, we evaluate the theoretical and empirical basis of this challenge to the importance of
80 genome-wide genetic variation and show that its premise is inconsistent with population genetic
81 theory and empirical findings. While it is clear that functional genetic information can advance
82 conservation, deemphasizing the maintenance of genome-wide genetic variation would increase
83 the extinction risk of threatened populations.

84

85 **1. Is genetic variation predictive of inbreeding and inbreeding depression?**

86 Inbreeding depression is thought to be driven mainly by homozygous and *identical-by-descent*
87 deleterious, partially recessive alleles (13), with lethal and small effect deleterious alleles
88 contributing substantially (14). The constant input of new deleterious mutations (15-19) makes
89 inbreeding depression a ubiquitous phenomenon that can push populations toward extinction (2,
90 20-23). One of the foundational predictions of theoretical population genetics is that the rate of
91 loss of heterozygosity (H) per generation ($\Delta\bar{H}=1/2N_e$) is identical to the rate of increase in mean
92 individual inbreeding (F), which is $\Delta\bar{F}=1/2N_e$ (24). \bar{H} is therefore expected to be entirely
93 predictive of \bar{F} (24-29).

94 A more difficult, but crucial question is whether genome-wide genetic variation (π) is
95 predictive of inbreeding depression. Deleterious alleles are lost in small populations due to
96 selection and genetic drift (30, 31), but they are also more often expressed in homozygotes in
97 smaller populations due to inbreeding. Selective *purging* of large effect deleterious alleles
98 following inbreeding combined with genetic drift may therefore result in low *inbreeding load* and
99 little inbreeding depression in the most highly inbred populations with the lowest π . However, the
100 presence of purging does not imply that high fitness is maintained in small populations with low π .

101 Population genetics theory predicts that larger populations will have higher neutral (24)
102 and deleterious genetic variation (32, 33). This is illustrated in Fig. 1, where simulated large
103 populations have higher π (24) and higher inbreeding load (32-34) arising from segregating
104 partially recessive deleterious alleles. These simulations assume empirically supported models of
105 fitness and dominance (h) effects (*SI Appendix*). Smaller populations have lower π due to genetic

106 drift, and fewer *lethal equivalents* due to genetic drift and purging. However, despite having fewer
107 lethal equivalents, chronically smaller populations have lower mean fitness due to partially
108 recessive deleterious alleles being expressed following inbreeding, and some reaching high
109 frequency or fixation (i.e., high *drift load*). Therefore, a negative relationship is expected between
110 π and drift load for populations at mutation-drift-selection equilibrium.

111
112
113 Equilibrium levels of π and drift load are not expected in populations with fluctuating
114 population size or immigration rate. A common scenario with high conservation relevance is
115 isolated populations that have experienced recent bottlenecks. The simulated data in Fig. 2 shows
116 that genome-wide π declines over time following a bottleneck, as expected from classical theory
117 (24) (Fig. 2A). This pattern is paralleled by lethal equivalents (Fig. 2B) owing to the loss of
118 deleterious alleles via genetic drift and purging of deleterious alleles expressed in homozygotes
119 due to inbreeding (30, 31). However, the deleterious alleles remaining after a bottleneck often go
120 to high frequency or fixation. This results in individuals being homozygous for increasingly more
121 deleterious alleles (higher drift load, Fig. 2C) as π declines inexorably during a sustained
122 bottleneck, the same pattern expected for small populations at equilibrium (Fig. 1). It is notable,
123 though, that π , inbreeding load, and drift load can change at substantially different rates following
124 a bottleneck. For example, drift load can become quite high before π declines substantially
125 following a bottleneck (Fig. 2A, 2C). However, small populations that already have low π are also
126 expected to have low mean fitness due to ever-increasing drift load, which demonstrates that π is a
127 good indicator of drift load and mean fitness. Occasional immigration can be sufficient to maintain
128 high π and low drift load in small populations (Fig. 2). This is one reason why maintaining
129 connectivity is a priority in conservation biology, and why *genetic rescue* is an effective tool for
130 managing small, isolated populations (30, 35, 36).

131 Empirical data show that purging does not eliminate the extinction threat posed by
132 inbreeding. Pedigree-based studies have yielded mixed results with regard to purging, with
133 typically only a small portion of inbreeding depression being removed after sustained inbreeding
134 in small populations (37-39). Analyses of 60 genomes from seven ibex species found that species
135 which went through the most severe bottlenecks had more deleterious alleles (40). Alpine ibex,
136 which were once reduced to 100 individuals, had fewer highly deleterious alleles but more mildly

137 deleterious alleles compared to Iberian ibex (bottleneck size 1,000 individuals). Empirical genetic
138 data suggest small populations have higher drift load (40-42) which has resulted in lower
139 population growth in populations with lower genetic variation (2, 3). In agreement with
140 theoretical expectations outlined above, these data suggest that purging is insufficient to maintain
141 high fitness in the face of strong genetic drift and inbreeding. Thus, the presence of genomic
142 signatures of purging should not be taken as evidence for the absence of inbreeding depression, or
143 for demographic stability of small populations.

144 The relationship between π and fitness is obviously complicated, particularly immediately
145 after a bottleneck (Fig. 2). Populations with the lowest π and highest inbreeding will also have the
146 lowest inbreeding load on average due to reduced deleterious genetic variation via genetic drift
147 and purging. However, these same genetically depauperate populations will typically have lower
148 fitness than larger, genetically diverse populations on average due to ever-increasing drift load
149 (Fig. 1 & 2). The bottom line is that reduced fitness is generally expected in small, isolated,
150 genetically depauperate populations due to inbreeding depression and the accumulation of drift
151 load, and that maintaining genetic variation and population connectivity will increase long term
152 viability.

153

154 **2. Is genome-wide genetic variation predictive of adaptive potential?**

155 The ability of populations to adapt to changing environmental conditions (*adaptive potential*) is
156 fundamental for persisting through environmental change (43, 44). A core insight from theoretical
157 genetics is that adaptation requires additive genetic variance (V_a) for the selected trait(s) (45). A
158 lack of V_a can limit a population's response to selection and eventually lead to extinction (43, 44,
159 46). As with other types of genetic variation, V_a is affected by mutation at loci affecting the trait,
160 selection, migration, and genetic drift (47). We therefore expect from first principles that larger
161 populations will have higher π and higher V_a than small populations *on average* (Fig. 1), and thus
162 that π should be correlated with V_a . Despite strong theoretical support, determining the strength
163 and importance of this relationship in real populations, especially those of conservation concern,
164 has generated longstanding controversy (48).

165 Basic population genetic theory shows that population size and connectivity play major
166 roles in determining V_a , and thus adaptive potential. Isolated populations below a certain size
167 should lose V_a due to genetic drift more rapidly than it is replenished via mutation (47).

168 Additionally, recently bottlenecked populations that have lost π will eventually also lose V_a and
169 evolutionary potential in the absence of immigration (Fig. 2). However, while the eventual
170 reduction in V_a in small populations is inevitable, the initial effects of a bottleneck on V_a can be
171 complex. Recently bottlenecked populations may show decreases, stability, or even short-term
172 increases in V_a due to the conversion of dominant or epistatic variance into V_a as allele frequencies
173 change due to genetic drift (49-51). This potential conversion of nonadditive to additive variation
174 in bottlenecked populations is highly stochastic across traits and populations, and is one of the
175 processes that can cloud the relationship between molecular and quantitative trait variation (52).
176 Nonetheless, the two important takeaways are: 1) although bottlenecks can complicate the
177 prediction of declining V_a for any given trait in small populations, V_a will be reduced on average,
178 especially for traits with primarily additive inheritance; and 2) eventually, the inexorable decline in
179 π in very small populations means that all small populations will eventually lose V_a and their
180 ability to adapt to environmental change. Adaptive potential in such populations will be severely
181 limited unless V_a is replenished by new mutations or migration from differentiated populations
182 (35) (Fig. 2).

183 The hypothesis that small populations harbor less V_a has been tested empirically in both
184 laboratory and field settings. Most experimental studies show declines in V_a and weaker responses
185 to selection in small populations or following bottlenecks (53-55). On the other hand, field studies
186 often find a weak association between V_a and genome-wide genetic variation when comparing
187 across populations (48, 56); this weak relationship is likely due to a combination of factors, none
188 of which refute the two takeaways described above.

189 As discussed above, empirical results suggest that V_a may initially increase after a
190 bottleneck due to the conversion of epistatic and dominance variance to V_a (50, 57), and then
191 decline after substantial inbreeding accumulates. Further, V_a is expected to vary among traits and
192 populations depending on genetic architecture, mutation rate, and the mode and history of
193 selection. In practice, most studies are unable to account for these factors and are generally only
194 able to assess a few traits per species/population. Estimates of V_a for each trait are also typically
195 based on a modest number of families. Although the number of traits, populations, and species
196 studied has increased, determining the total V_a for fitness in a given population of conservation
197 concern is not an attainable goal. Additionally, the vast majority of the best-characterized species
198 with respect to V_a in the wild (i.e., most of the species included in (48, 56) meta-analyses) are

199 common. The species and populations in which the relationship between V_a and genetic variation
200 is expected to be strongest, namely, declining species of conservation concern, tend to be most
201 difficult to characterize.

202 Arguably the most important point is that the loss of genetic variation in small and/or
203 bottlenecked populations is inevitable and will eventually lead to reduced V_a and reduce adaptive
204 potential, regardless of short-term and stochastic outcomes. Isolated populations that remain small
205 are unlikely to recover substantial V_a due to the slow rate of mutation and the counteracting loss of
206 variation to genetic drift, and the lack of adaptive potential is problematic for long term viability
207 (43, 44, 47).

208

209 **3. What is the relationship between genome-wide genetic variation and population viability?**

210 The central question regarding the role of genetic variation in conservation is whether populations
211 with lower π are less likely to persist. Genetic effects on the persistence of a particular population
212 are difficult to predict with certainty because there are many factors involved that are difficult to
213 evaluate, including mating system and demographic history (32, 33), current and future
214 environmental conditions (58), and the extent to which *soft selection* versus *hard selection*
215 predominate (59, 60). Additionally, the highly stochastic demography of small populations, which
216 is exacerbated by inbreeding depression (61), means that widely divergent outcomes can be
217 expected across populations with the same environmental, demographic, and genetic starting
218 conditions. However, theoretical empirical studies have yielded broadly applicable insights into
219 the effects of genetic variation and inbreeding on population viability.

220 Population genetics theory predicts that small, isolated populations with low genetic
221 variation are more likely to go extinct due to genetic effects than larger, more genetically diverse
222 populations under empirically supported mutational assumptions (19, 22, 23, 62). *De novo*
223 mutations following a bottleneck are expected to cause eventual extinction of very small,
224 genetically depauperate populations via *mutational meltdown* (*SI Appendix* Fig. S1) (19). The
225 average time to extinction is shorter under the more realistic scenario where bottlenecked
226 populations carry deleterious mutations at the outset (Fig. 3). However, the extinction rate depends
227 strongly on bottleneck duration, with longer restrictions conferring increased extinction due to
228 both demographic stochasticity and the constant increase in drift load. Short-lived bottlenecks are
229 one scenario where viability may sometimes be higher for historically smaller, less genetically
230 diverse populations that have fewer deleterious alleles at the outset of the bottleneck due to

231 historical genetic drift and purging (Fig. 1, 3A, 3B). However, this assumes inbreeding depression
232 is the only genetic challenge operating, and simultaneous selection caused by environmental
233 change may reverse this relationship. Longer bottlenecks in isolated populations are expected to
234 result in very high extinction rates due to mutational meltdown regardless of the abundance of
235 deleterious alleles at the outset (19) (Fig. 3C).

236 Empirical studies of population dynamics arguably provide the strongest evidence for the
237 broad benefits of increased genetic variation for population viability. Numerous studies have
238 almost universally found that populations with higher genetic variation have increased population
239 growth and viability (63). For example, lower genetic variation was associated with reduced
240 population growth in alpine ibex (3) and increased local extinction in Glanville fritillary butterflies
241 (2). Inbred laboratory lines of animals, which quickly lose genetic variation, often become extinct
242 substantially more rapidly than control lines (64, 65). Additionally, the infusion of genetic
243 variation via natural (66) and facilitated immigration ('genetic rescue') nearly always increases
244 population growth (35, 36, 67, 68) either by masking of deleterious recessive alleles, or by
245 infusing adaptive genetic variation.

246 The collapse of the Isle Royale wolf population after a mainland male immigrated to the
247 small population has been interpreted as a counter-example to the efficacy of genetic rescue (8).
248 However, detailed documentation indicates that results from this unusual system are unsuitable as
249 a general example of the likely demographic outcome of genetic rescue attempts (67, 69, 70). The
250 immigration of only a single male into Isle Royale makes is unusual in the context of managed
251 genetic rescue attempts which typically involve translocation of multiple individuals into a small
252 population, e.g., (71-73). The single migrant male wolf dominated and increased reproduction,
253 resulting in genetic rescue (an increase in population size following outbreeding). However, his
254 extremely high reproduction resulted in very high inbreeding within two generations and the
255 subsequent dramatic population decline (67, 69, 70). This male was likely just an opportunistic,
256 successful migrant from the nearest population. It is unclear whether he carried an exceptional
257 number of deleterious alleles that drove the subsequent decline, or if inbreeding following
258 exceptionally high reproduction of any individual would have led to a similar demographic
259 outcome.

260 Recovery of some populations from severe bottlenecks, and persistence of some
261 populations despite small N_e and low genetic variation is often cited as a challenge to the idea that
262 low genetic variation and inbreeding reduce population viability (6, 8, 9, 11, 74-77). Soulé (5) [p.

263 178] pointed out the fundamental flaw of this argument, which he referred to as the “fallacy of the
264 accident” nearly 35 years ago: the only observable populations that have experienced bottlenecks
265 are those that survived. The potentially numerous populations that went extinct under similar
266 conditions are unobservable. Counting extant, genetically depauperate populations is therefore an
267 unreliable metric of the extinction risk posed by lost genetic variation and inbreeding. Theoretical
268 population genetics and population ecology both predict that some populations will survive
269 bottlenecks, and some lucky ones will persist for long periods at small population size. However,
270 such cases are likely the rare exception, the lottery winners so-to-speak (5, 67).

271 The most immediate threats to small, genetically depauperate populations are demographic
272 stochasticity and inbreeding depression. However, long term population persistence will in most
273 cases require populations to adapt to environmental change (e.g., climate change, novel diseases,
274 invasive species, etc.) (44, 78). Rapid adaptation to new conditions is possible, but requires
275 sufficient genetic variation and relatively large population size (53, 79). All of the material above
276 highlights the fundamental importance of maintaining large, connected, genetically diverse
277 populations. Long term population viability requires having both manageable *genetic load* and
278 adaptive potential associated with genome-wide genetic variation.

279

280 **4. Simulation-based inferences of the effects of genetic variation and inbreeding on** 281 **population viability**

282 Simulation-based studies showed long ago that inbreeding depression can substantially increase
283 extinction risk (23, 80). However, our increasing understanding of deleterious mutation parameters
284 (e.g., deleterious mutation rates, and the distribution of fitness effects [DFE]) combined with the
285 availability of sophisticated, user-friendly simulation software (81) will likely advance our
286 understanding of inbreeding depression and purging within the field of conservation.

287 While there is much to learn about deleterious mutation parameters, a lot is known about
288 the most important elements. First, deleterious mutations arise frequently (15, 16, 82-84), and
289 large effect deleterious alleles appear to be a major driver of inbreeding depression (14, 85-87).
290 For example, lethal alleles arose via mutation at a rate of ~3% per diploid genome in *Drosophila*
291 (14). Inbreeding depression appeared to be largely due to highly deleterious alleles originating in a
292 subset of pedigree founders in sheep and mice (86, 87). Lethal and other large effect deleterious
293 alleles are frequently observed in small natural populations, humans, and model organisms (14, 83,
294 85, 88-90). The majority of humans and *Drosophila* likely carry one or more recessive lethal

295 alleles (85, 89, 90). Deleterious mutations appeared at a rate of $U=1.2$ /diploid genome/generation
296 in *Drosophila* (15) and $U = 1.6$ in hominids (16). Mutation accumulation studies show that the
297 DFE for deleterious mutations is strongly bimodal, with most mutations having small to moderate
298 effects (e.g. $|s|<0.25$) and a minority being lethal or semi-lethal (82).

299 Second, the degree of dominance (h) is strongly related to mutation effect size. Direct
300 observation of dominance effects in yeast and *Drosophila* suggest that nearly neutral deleterious
301 mutations are slightly recessive on average (h slightly less than 0.5), and highly deleterious
302 mutations (e.g., $|s|>0.25$) are nearly fully recessive (h very near zero), with h declining
303 exponentially as s increases in size (14, 91, 92). There is still much uncertainty regarding
304 deleterious mutation parameters (see discussion below). However, the best available information
305 suggests that reasonable values of U are >1 , the DFE is strongly bimodal, and dominance declines
306 substantially with increasing size of s . These findings guide the simulations presented above
307 (details in *SI Appendix*).

308 Recently, results from genetically explicit simulations were used to argue that genome-
309 wide genetic variation is of little importance to population viability, and that purging is likely to
310 prevent extinction (8, 11, 74). However, these studies excluded large effect deleterious mutations
311 (*SI Appendix* Fig. S2) and assumed values of U that were between 2.6 and 92.3 times lower than
312 the best estimate of U in *Drosophila* (Table 1). As a result, these models (8, 11, 74) produce
313 substantially weaker inbreeding depression (<0.05 to approximately 1 lethal equivalent) than
314 observed in real populations, where the median number of lethal equivalents for juvenile survival
315 in captive mammals was 3.1 (93), and 12 for total fitness in wild mammals (23) (*SI Appendix* Fig.
316 S3). There is substantial uncertainty in deleterious mutation rates, and the DFE, particularly for
317 non-model organisms. However, the discrepancy between the assumed mutation parameters and
318 the resulting inbreeding depression in the aforementioned studies (8, 11, 74) and the best available
319 empirical estimates (Table 1, *SI Appendix* Fig. S3), yield results that underestimate the importance
320 of genetic variation in conservation, and the efficacy of genetic rescue as a tool in conservation.

321
322 **5. Is the relationship between genetic variation and conservation status informative of the**
323 **importance of genetic variation to population viability?**

324 It has been suggested that a weak relationship between genetic variation and conservation status
325 (e.g., IUCN Red List) means that genome-wide genetic variation is uninformative of extinction

326 risk (11). However, this relationship is not universally expected, even though extinction risk is
327 strongly affected by genome-wide genetic variation.

328 First, a lag is expected between reduced population size and the loss of genetic variation.
329 Most threatened populations initially decline due to non-genetic factors (e.g., habitat loss, disease,
330 climate change). Thus, multiple generations are required for a substantial reduction in genetic
331 variation, even after severe bottlenecks (Fig. 2A). Threatened populations that became small due
332 to non-genetic factors may still have high genetic variation due to this lag. Second, failing to
333 control for other factors that influence genetic variation (e.g., N_e , dispersal, generation time, and
334 mutation rate (11)) can obscure the relationship between genetic variation and conservation status.
335 In contrast, a study controlling for phylogeny (a proxy for the aforementioned confounding
336 factors) showed a significant relationship between genetic variation and conservation status (94).

337 Differences among studies in the measures of genetic variation can further obscure true
338 relationships between genetic variation and conservation status. Estimates of genetic variation for
339 different species used in comparative studies vary widely in the number of sampled individuals
340 and populations, and in the regions of the genome analyzed. Some studies estimate species-wide
341 genetic diversity from a single individual (11, 95, 96) and compare different genetic data types
342 across species (6, 96). Using single genomes to estimate species-wide genetic diversity is
343 problematic because the individuals chosen may not be representative of the species as a whole
344 (e.g., captive individuals (95)). Rather, multiple individuals and populations are necessary to
345 accurately reflect a species' distribution of genetic variation (97, 98). Additionally, estimates of
346 genetic diversity are affected by reference genome quality (99), mapping bias (100, 101), the
347 methods used to measure genetic variation (e.g., whole genome sequencing, RNAseq, RADseq),
348 and bioinformatics approaches (98, 99). Thus, sampling, genetic markers, and analyses should be
349 standardized when measuring the relationship between genetic variation and conservation status.

350 Lastly, IUCN Red List status is an imperfect index of extinction risk because it is a
351 subjective measure of population viability. The IUCN Red List is important for monitoring
352 biodiversity, but the guidelines used to categorize threat levels within the Red List are subject to
353 user interpretation, which can lead to inconsistent assessments (102-106). The imperfect
354 relationship between IUCN Red List status and extinction risk means that Red List status is an
355 inappropriate surrogate for extinction risk in assessing the relationship between genome-wide
356 diversity and extinction risk. Together these issues suggest that the weak relationship between

357 genetic variation and conservation status has little bearing on the importance of genome-wide
358 genetic variation for extinction risk.

359

360 **6. What is the role of functional genetic variation in conservation?**

361 The widespread availability of genomic data for non-model organisms has rapidly advanced our
362 understanding of the genetic basis and evolution of fitness-related traits in natural populations,
363 e.g., (107-111). This revolution has raised the question of how to effectively integrate functional
364 genetic information into conservation practice (112-115). It has repeatedly been suggested that
365 genetic assessment and management of threatened populations should be focused on variation at
366 particular loci that affect particular fitness traits (11, 116-118). However, such gene-targeted
367 conservation approaches are always difficult, and prone to failure for several reasons.

368 First, understanding the genetic basis of fitness remains extremely complicated and
369 challenging (112, 114). While some important traits in natural populations are affected by loci
370 with very large effects, most traits are determined by many small-effect loci (119-121). A
371 comprehensive understanding of the genetic basis of such traits is out of reach for non-model
372 organisms (122). To accurately understand the locus-specific effects on a trait and fitness requires
373 information on dominance and pleiotropy, epistasis, genotype-by-environment interactions, and
374 the amount of linkage disequilibrium with other loci influencing the trait or other fitness
375 components (112). These factors are expected to vary among traits and to differ for the same trait
376 among species and potentially among populations within a species, e.g., (107). Therefore,
377 substantial effort is necessary to understand the conservation relevance of a particular genetic
378 variant and predict whether the benefits of gene-targeted conservation actions outweigh potential
379 detrimental effects (112, 114).

380 A classic example of the potential for undesirable outcomes of gene-targeted conservation
381 management is the suggestion that genetic management of captive and wild populations should be
382 designed around maintaining genetic variation at the major histocompatibility complex (MHC)
383 (11, 116, 117, 123). The MHC has been studied in great detail in humans because of its
384 importance in immunity, organ transplantation, and autoimmune disease, but its organization is
385 poorly understood in most other vertebrates. Although there is strong evidence for its adaptive
386 importance, some variants have detrimental effects, and the adaptive effects of other variants
387 appear to be environmentally dependent (124). Detailed examination of the fitness effects of MHC

388 alleles and haplotypes is necessary to determine how much maintaining MHC variation enhances
389 fitness.

390 Additionally, as highlighted multiple times over the last 35 years (112, 125-129), basing
391 conservation management on a small subset of loci risks increasing the loss of genetic variation
392 elsewhere in the genome. Such efforts would be counterproductive unless the gain in mean fitness
393 associated with gene-targeted management is greater than the loss in fitness associated with lost
394 genome-wide genetic variation (112). This highlights the challenges and pitfalls of gene-targeted
395 conservation. When recommendations for maintaining genome-wide genetic variation versus
396 particular adaptive variants are in conflict, a cost-benefit analysis of the two approaches should be
397 performed and a composite solution identified (112). Recent cases where genomic analyses have
398 revealed that large effect loci play a key role in traits of conservation importance, e.g., (107, 108,
399 110, 130) will be the first to empirically test the efficacy of gene-targeted conservation
400 approaches.

401

402 **Discussion**

403 Genomic data should be used to challenge findings from population genetics theory and previous
404 empirical data that form the basis for genetic management of small populations. Recent genomic
405 studies provide useful fodder to determine how to effectively use genomic data to improve
406 conservation in ways that were previously impossible. Examples are emerging of how
407 understanding functional genetic variation could improve recommendations to conserve imperiled
408 populations (107, 108, 110, 130), making genomic data more useful for conservation than ever
409 before. However, genomic data have not discredited the decades worth of evidence that inbreeding
410 depression, mutational meltdown, and loss of adaptive potential are major threats to conservation.

411 Identifying genetic variants that affect fitness traits undoubtedly advances understanding of
412 the genetic basis of adaptation, and that is important in itself (131). However, placing conservation
413 priority on a small, apparently adaptive portion of the genome ignores what may be the vast
414 majority of variation elsewhere in the genome that will fuel adaptation to unpredictable future
415 conditions (112, 114, 125, 126). This approach is reminiscent of the “adaptationist programme”
416 that Gould & Lewontin (132) criticized >40 years ago for being overly enamored with adaptive
417 explanations for interesting traits (‘spandrels’) without considering that they might have arisen by
418 accident, and that they are but one part of the whole, complex organism (114). Now, as then, we
419 should avoid the temptation to place undue priority on putatively adaptive loci (‘molecular

420 spandrels' (133)) without first considering the rest of the genome. Our inability to predict future
421 changes in genotype-by-environment interactions should lead us to recognize the importance of
422 genome-wide genetic variation (including presently neutral variation), and more importantly, the
423 factors that make it possible – large livable habitats and natural patterns of connectivity among
424 them. Conserving genetic variation across the whole genome is almost certainly the most reliable
425 approach to conserve the genetic variation that matters.

426 We know of no convincing evidence that supports abandoning the focus on genome-wide
427 genetic variation in exchange for a focus on functional variation. The recent simulation studies that
428 have been used to discount the importance of genome-wide genetic variation in conservation (8,
429 11, 74) are based on assumptions that are inconsistent with the preponderance of empirical data on
430 the genetics of inbreeding depression and its effect on population viability (see above). Some
431 small populations may not suffer strong inbreeding depression, and some may not rebound
432 following the introduction of genetic variation. However, as pointed out in the formative years of
433 conservation biology, we must resist the temptation to dismiss the extinction risks associated with
434 lost genetic variation in small populations (5).

435 Although population genetics theory has done a remarkably good job of predicting patterns
436 now observable in genomic data, many questions remain unanswered that will improve the utility
437 of genomic data in conservation. For example, how prevalent is soft selection? The presence of
438 soft selection could help explain some of the instances where populations persist for long periods
439 despite inbreeding (59, 60). How much do U and the distribution of fitness effects for deleterious
440 mutations vary among taxa? U may be rather consistent within some taxonomic groups (e.g.,
441 mammals) where the number of genes is strongly conserved (134). Nevertheless, variation among
442 taxa in gene number, mutation rate, and the amount of intergenic DNA that is subject to
443 deleterious mutation is an important consideration for assessing the fitness effects of inbreeding.
444 Lastly, while it is clear that the distribution of mutation fitness effects is bimodal (82),
445 understanding the specific shape of this distribution, and how much this varies among taxa, is
446 important for our understanding of the extinction risks associated with small population size and
447 inbreeding.

448 Genomic data will undoubtedly continue to be used to revisit and refine insights gained
449 since genetics was first applied to conservation and to understand the extinction process (4, 5, 46,
450 135). So far, genomic data have reinforced earlier findings showing that genome-wide genetic
451 variation is key to population viability. Given the increasing rate of habitat loss and fragmentation,

452 failing to recognize and mitigate the effects of lost genome-wide genetic variation would only
453 exacerbate the biodiversity crisis.

454

455 **Acknowledgements**

456 We thank P. Hohenlohe, R. Waples, A. García-Dorado, and three anonymous reviewers for
457 comments on previous drafts. SWF was supported by National Science Foundation grant DEB
458 2016569. DAT was supported by National Institute of General Medical Sciences of the National
459 Institutes of Health under award number RL5GM118990. WCF was supported by National
460 Science Foundation grants DEB 1413925, DEB 1754821, and DEB 1838282. SSH was supported
461 by University of Wisconsin-Milwaukee College of Letters and Science.

462

463 **Data availability**

464 Materials to replicate the simulations are available at <https://doi.org/10.5281/zenodo.5513957>.

465

466 **References**

- 467 1. R. Lande, S. Shannon, The role of genetic variation in adaptation and population
468 persistence in a changing environment. *Evolution*, 434-437 (1996).
- 469 2. I. Saccheri *et al.*, Inbreeding and extinction in a butterfly metapopulation. *Nature* **392**, 491-
470 494 (1998).
- 471 3. C. Bozzuto, I. Biebach, S. Muff, A. R. Ives, L. F. Keller, Inbreeding reduces long-term
472 growth of Alpine ibex populations. *Nature Ecology & Evolution* **3**, 1359-1364 (2019).
- 473 4. O. Frankel, M. E. Soulé, *Conservation and Evolution* (CUP Archive, 1981).
- 474 5. M. E. Soulé, *Viable Populations for Conservation* (Cambridge University Press, 1987).
- 475 6. J. A. Robinson *et al.*, Genomic flatlining in the endangered island fox. *Current Biology* **26**,
476 1183-1189 (2016).
- 477 7. J. A. Robinson *et al.*, Genomic signatures of extensive inbreeding in Isle Royale wolves, a
478 population on the threshold of extinction. *Science Advances* **5**, eaau0757 (2019).
- 479 8. C. C. Kyriazis, R. K. Wayne, K. E. Lohmueller, Strongly deleterious mutations are a
480 primary determinant of extinction risk due to inbreeding depression. *Evolution Letters* **5**,
481 33-47 (2021).
- 482 9. P. A. Morin *et al.*, Reference genome and demographic history of the most endangered
483 marine mammal, the vaquita. *Molecular Ecology Resources* 10.1111/1755-0998.13284
484 (2020).
- 485 10. M. V. Westbury *et al.*, Hyena paleogenomes reveal a complex evolutionary history of
486 cross-continental gene flow between spotted and cave hyena. *Science Advances* **6**,
487 eaay0456 (2020).
- 488 11. J. C. Teixeira, C. D. Huber, The inflated significance of neutral genetic diversity in
489 conservation genetics. *Proceedings of the National Academy of Sciences* **118**,
490 e2015096118 (2021).
- 491 12. Y. Xue *et al.*, Mountain gorilla genomes reveal the impact of long-term population decline
492 and inbreeding. *Science* **348**, 242-245 (2015).

- 493 13. D. Charlesworth, J. H. Willis, The genetics of inbreeding depression. *Nature Reviews*
494 *Genetics* **10**, 783-796 (2009).
- 495 14. M. J. Simmons, J. F. Crow, Mutations affecting fitness in *Drosophila* populations. *Annual*
496 *review of genetics* **11**, 49-78 (1977).
- 497 15. C. Haag-Liautard *et al.*, Direct estimation of per nucleotide and genomic deleterious
498 mutation rates in *Drosophila*. *Nature* **445**, 82-85 (2007).
- 499 16. A. Eyre-Walker, P. D. Keightley, High genomic deleterious mutation rates in hominids.
500 *Nature* **397**, 344-347 (1999).
- 501 17. M. Lynch, Rate, molecular spectrum, and consequences of human mutation. *Proceedings*
502 *of the National Academy of Sciences* **107**, 961-968 (2010).
- 503 18. J. Haldane, The effect of variation of fitness. *The American Naturalist* **71**, 337-349 (1937).
- 504 19. M. Lynch, J. Conery, R. Burger, Mutation accumulation and the extinction of small
505 populations. *The American Naturalist* **146**, 489-518 (1995).
- 506 20. L. F. Keller, D. M. Waller, Inbreeding effects in wild populations. *Trends in Ecology &*
507 *Evolution* **17**, 230-241 (2002).
- 508 21. R. Frankham, Genetics and extinction. *Biological conservation* **126**, 131-140 (2005).
- 509 22. L. S. Mills, P. E. Smouse, Demographic consequences of inbreeding in remnant
510 populations. *The American Naturalist* **144**, 412-431 (1994).
- 511 23. J. J. O'Grady *et al.*, Realistic levels of inbreeding depression strongly affect extinction risk
512 in wild populations. *Biological Conservation* **133**, 42-51 (2006).
- 513 24. S. Wright, Evolution in Mendelian populations *Genetics* **16**, 97-159 (1931).
- 514 25. J. E. Powell, P. M. Visscher, M. E. Goddard, Reconciling the analysis of IBD and IBS in
515 complex trait studies. *Nature Reviews Genetics* **11**, 800-805 (2010).
- 516 26. D. Speed, D. J. Balding, Relatedness in the post-genomic era: is it still useful? *Nature*
517 *Reviews Genetics* **16**, 33-44 (2015).
- 518 27. A. Jacquard, Inbreeding: One word, several meanings. *Theoretical Population Biology* **7**,
519 338-363 (1975).
- 520 28. G. Malécot, *The Mathematics of Heredity* (W. H. Freeman, 1970).
- 521 29. M. Kardos *et al.*, Genomic consequences of intensive inbreeding in an isolated wolf
522 population *Nature Ecology & Evolution* **2**, 124-131 (2018).
- 523 30. P. W. Hedrick, A. Garcia-Dorado, Understanding inbreeding depression, purging, and
524 genetic rescue. *Trends in Ecology & Evolution* **31**, 940-952 (2016).
- 525 31. A. García-Dorado, Understanding and predicting the fitness decline of shrunk populations:
526 inbreeding, purging, mutation, and standard selection. *Genetics* **190**, 1461-1476 (2012).
- 527 32. M. C. Whitlock, Selection, load and inbreeding depression in a large metapopulation.
528 *Genetics* **160**, 1191-1202 (2002).
- 529 33. D. Charlesworth, B. Charlesworth, Inbreeding depression and its evolutionary
530 consequences. *Annual Review of Ecology and Systematics* **18**, 237-268 (1987).
- 531 34. N. E. Morton, J. F. Crow, H. J. Muller, An estimate of the mutational damage in man from
532 data on consanguineous marriages. *Proceedings of the National Academy of Sciences* **42**,
533 855-863 (1956).
- 534 35. A. R. Whiteley, S. W. Fitzpatrick, W. C. Funk, D. A. Tallmon, Genetic rescue to the
535 rescue. *Trends in Ecology & Evolution* **30**, 42-49 (2015).
- 536 36. R. Frankham, Genetic rescue of small inbred populations: Meta-analysis reveals large and
537 consistent benefits of gene flow. *Molecular Ecology* **24**, 2610-2618 (2015).
- 538 37. R. Frankham, D. M. Gilligan, D. Morris, D. A. Briscoe, Inbreeding and extinction: effects
539 of purging. *Conservation Genetics* **2**, 279-284 (2001).

- 540 38. R. C. Lacy, J. D. Ballou, Effectiveness of selection in reducing the genetic load in
541 populations of *Peromyscus polionotus* during generations of inbreeding. *Evolution* **52**, 900-
542 909 (1998).
- 543 39. P. Crnokrak, S. C. Barrett, Perspective: purging the genetic load: a review of the
544 experimental evidence. *Evolution* **56**, 2347-2358 (2002).
- 545 40. C. Grossen, F. Guillaume, L. F. Keller, D. Croll, Purging of highly deleterious mutations
546 through severe bottlenecks in Alpine ibex. *Nature Communications* **11**, 1-12 (2020).
- 547 41. S. Mathur, J. A. DeWoody, Genetic load has potential in large populations but is realized
548 in small inbred populations. *Evolutionary Applications* (2021).
- 549 42. A. Khan *et al.*, Genomic evidence for inbreeding depression and purging of deleterious
550 genetic variation in Indian tigers. *Proceedings of the National Academy of Sciences* (In
551 review).
- 552 43. M. Lynch, R. Lande, "Evolution and extinction in response to environmental change" in
553 Biotic interactions and global change, P. M. Kareiva, J. G. Kingsolver, R. B. Huey, Eds.
554 (Sinauer, Sunderland, MA., 1993), pp. 234-250.
- 555 44. M. Kardos, G. Luikart, The genomic architecture of fitness is a major driver of population
556 viability during rapid environmental change. *The American Naturalist* **197**, 511-525
557 (2021).
- 558 45. D. S. Falconer, T. F. C. Mackay, *Introduction to quantitative genetics* (Pearson, ed. 4,
559 1996).
- 560 46. R. Bürger, M. Lynch, Evolution and extinction in a changing environment: a quantitative-
561 genetic analysis. *Evolution* **49**, 151-163 (1995).
- 562 47. R. Lande, G. Barrowclough, "Effective population size, genetic variation, and their use in
563 population management" in Viable populations for conservation, M. Soulé, Ed.
564 (Cambridge University Press, Cambridge, 1987), chap. 6, pp. 87-123.
- 565 48. D. H. Reed, R. Frankham, How closely correlated are molecular and quantitative measures
566 of genetic variation? A meta-analysis. *Evolution* **55**, 1095-1103 (2001).
- 567 49. C. J. Goodnight, Epistasis and the effect of founder events on the additive genetic variance.
568 *Evolution* **42**, 441-454 (1988).
- 569 50. E. H. Bryant, S. A. McCommas, L. M. Combs, The effect of an experimental bottleneck
570 upon quantitative genetic variation in the housefly. *Genetics* **114**, 1191-1211 (1986).
- 571 51. C. J. Goodnight, On the effect of founder events on epistatic genetic variance. *Evolution*
572 **41**, 80-91 (1987).
- 573 52. N. Barton, M. Turelli, Effects of genetic drift on variance components under a general
574 model of epistasis. *Evolution* **58**, 2111-2132 (2004).
- 575 53. R. Frankham *et al.*, Do population size bottlenecks reduce evolutionary potential? *Animal*
576 *Conservation* **2**, 255-260 (1999).
- 577 54. Y. Willi, J. Van Buskirk, A. A. Hoffmann, Limits to the adaptive potential of small
578 populations. *Annual Review of Ecology, Evolution, and Systematics* **37**, 433-458 (2006).
- 579 55. P. de Villemereuil *et al.*, Little adaptive potential in a threatened passerine bird. *Current*
580 *Biology* **29**, 889-894. e883 (2019).
- 581 56. E. A. Mittell, S. Nakagawa, J. D. Hadfield, Are molecular markers useful predictors of
582 adaptive potential? *Ecology Letters* **18**, 772-778 (2015).
- 583 57. R. C. Lacy, A. F. Malo, G. Alaks, Maintenance of genetic variation in quantitative traits of
584 a woodland rodent during generations of captive breeding. *Conservation Genetics* **19**, 789-
585 802 (2018).

- 586 58. S. Meagher, D. J. Penn, W. K. Potts, Male–male competition magnifies inbreeding
587 depression in wild house mice. *Proceedings of the National Academy of Sciences* **97**, 3324-
588 3329 (2000).
- 589 59. A. F. Agrawal, Ecological determinants of mutation load and inbreeding depression in
590 subdivided populations. *The American Naturalist* **176**, 111-122 (2010).
- 591 60. D. A. Bell, R. P. Kovach, Z. L. Robinson, A. R. Whiteley, T. E. Reed, The ecological
592 causes and consequences of hard and soft selection. *Ecology Letters*
593 <https://doi.org/10.1111/ele.13754> (2021).
- 594 61. D. Goodman, "The demography of chance extinction" in *Viable populations for*
595 *conservation*, M. E. Soulé, Ed. (Cambridge University Press, Cambridge, 1987), pp. 11-
596 34.
- 597 62. R. Lande, Risk of population extinction from fixation of new deleterious mutations.
598 *Evolution* **48**, 1460-1469 (1994).
- 599 63. R. C. Lacy, Importance of genetic variation to the viability of mammalian populations.
600 *Journal of Mammalogy* **78**, 320-335 (1997).
- 601 64. J. Bowman, D. Falconer, Inbreeding depression and heterosis of litter size in mice.
602 *Genetics Research* **1**, 262-274 (1960).
- 603 65. L. I. Wright, T. Tregenza, D. J. Hosken, Inbreeding, inbreeding depression and extinction.
604 *Conservation Genetics* **9**, 833 (2008).
- 605 66. M. Åkesson *et al.*, Genetic rescue in a severely inbred wolf population. *Molecular Ecology*
606 **25**, 4745-4756 (2016).
- 607 67. K. Ralls, P. Sunnucks, R. C. Lacy, R. Frankham, Genetic rescue: A critique of the evidence
608 supports maximizing genetic diversity rather than minimizing the introduction of
609 putatively harmful genetic variation. *Biological Conservation* **251**, 108784 (2020).
- 610 68. S. W. Fitzpatrick *et al.*, Genomic and fitness consequences of genetic rescue in wild
611 populations. *Current Biology* **30**, 517-522. e515 (2020).
- 612 69. P. Hedrick, J. Robinson, R. O. Peterson, J. A. Vucetich, Genetics and extinction and the
613 example of Isle Royale wolves. *Animal Conservation* **22**, 302-309 (2019).
- 614 70. P. W. Hedrick, R. O. Peterson, L. M. Vucetich, J. R. Adams, J. A. Vucetich, Genetic
615 rescue in Isle Royale wolves: genetic analysis and the collapse of the population.
616 *Conservation genetics* **15**, 1111-1121 (2014).
- 617 71. J. Hogg, S. Forbes, B. Steele, G. Luikart, Genetic rescue of an insular population of large
618 mammals. *Proceedings of the Royal Society B: Biological Sciences* **273**, 1491 - 1499
619 (2006).
- 620 72. W. E. Johnson *et al.*, Genetic Restoration of the Florida Panther. *Science* **329**, 1641-1645
621 (2010).
- 622 73. R. L. Westemeier *et al.*, Tracking the long-term decline and recovery of an isolated
623 population. *Science* **282**, 1695-1698 (1998).
- 624 74. J. A. Robinson, C. Brown, B. Y. Kim, K. E. Lohmueller, R. K. Wayne, Purging of strongly
625 deleterious mutations explains long-term persistence and absence of inbreeding depression
626 in island foxes. *Current Biology* **28**, 3487-3494. e3484 (2018).
- 627 75. T. Caro, M. K. Laurenson, Ecological and genetic factors in conservation: a cautionary
628 tale. *Science* **263**, 485-486 (1994).
- 629 76. D. Simberloff, The contribution of population and community biology to conservation
630 science. *Annual Review of Ecology and Systematics* **19**, 473-511 (1988).
- 631 77. A. Harcourt, Population viability estimates: theory and practice for a wild gorilla
632 population. *Conservation Biology* **9**, 134-142 (1995).

- 633 78. O. H. Frankel, Genetic conservation: our evolutionary responsibility. *Genetics* **78**, 53-65
634 (1974).
- 635 79. M. Ørsted, A. A. Hoffmann, E. Sverrisdóttir, K. L. Nielsen, T. N. Kristensen, Genomic
636 variation predicts adaptive evolutionary responses better than population bottleneck
637 history. *PLoS Genetics* **15**, e1008205 (2019).
- 638 80. R. C. Lacy, VORTEX: a computer simulation model for population viability analysis.
639 *Wildlife Research* **20**, 45-65 (1993).
- 640 81. B. C. Haller, P. W. Messer, SLiM 2: Flexible, interactive forward genetic simulations.
641 *Molecular Biology and Evolution* **34**, 230-240 (2017).
- 642 82. A. Eyre-Walker, P. D. Keightley, The distribution of fitness effects of new mutations.
643 *Nature Reviews Genetics* **8**, 610-618 (2007).
- 644 83. A. E. Trask *et al.*, Evidence of the phenotypic expression of a lethal recessive allele under
645 inbreeding in a wild population of conservation concern. *Journal of Animal Ecology* **85**,
646 879-891 (2016).
- 647 84. K. Ralls, J. D. Ballou, B. A. Rideout, R. Frankham, Genetic management of
648 chondrodystrophy in California condors. *Animal Conservation* **3**, 145-153 (2000).
- 649 85. M. A. Ballinger, M. A. Noor, Are lethal alleles too abundant in humans? *Trends in*
650 *Genetics* **34**, 87-89 (2018).
- 651 86. R. C. Lacy, G. Alaks, A. Walsh, Hierarchical analysis of inbreeding depression in
652 *Peromyscus polionotus*. *Evolution* **50**, 2187-2200 (1996).
- 653 87. J. Casellas, J. Piedrafita, G. Caja, L. Varona, Analysis of founder-specific inbreeding
654 depression on birth weight in Ripollesa lambs. *Journal of Animal Science* **87**, 72-79
655 (2009).
- 656 88. K. Ralls, J. D. Ballou, Genetic status and management of California condors. *The Condor*
657 **106**, 215-228 (2004).
- 658 89. Z. Gao, D. Waggoner, M. Stephens, C. Ober, M. Przeworski, An estimate of the average
659 number of recessive lethal mutations carried by humans. *Genetics* **199**, 1243-1254 (2015).
- 660 90. V. M. Narasimhan *et al.*, Health and population effects of rare gene knockouts in adult
661 humans with related parents. *Science* **352**, 474-477 (2016).
- 662 91. A. F. Agrawal, M. C. Whitlock, Inferences about the distribution of dominance drawn from
663 yeast gene knockout data. *Genetics* **187**, 553-566 (2011).
- 664 92. H.-W. Deng, M. Lynch, Estimation of deleterious-mutation parameters in natural
665 populations. *Genetics* **144**, 349-360 (1996).
- 666 93. K. Ralls, J. D. Ballou, A. Templeton, Estimates of lethal equivalents and the cost of
667 inbreeding in mammals. *Conservation Biology* **2**, 185-193 (1988).
- 668 94. D. Spielman, B. W. Brook, R. Frankham, Most species are not driven to extinction before
669 genetic factors impact them. *Proceedings of the National Academy of Sciences* **101**, 15261-
670 15264 (2004).
- 671 95. Z. Consortium, A comparative genomics multitool for scientific discovery and
672 conservation. *Nature* **587**, 240-245 (2020).
- 673 96. R. B. Corbett-Detig, D. L. Hartl, T. B. Sackton, Natural selection constrains neutral
674 diversity across a wide range of species. *PloS Biology* **13**, e1002112 (2015).
- 675 97. P. Pečnerová *et al.*, High genetic diversity and low differentiation reflect the ecological
676 versatility of the African leopard. *Current Biology* **31**, 1862-1871 (2021).
- 677 98. E. E. Armstrong *et al.*, Long live the king: chromosome-level assembly of the lion
678 (*Panthera leo*) using linked-read, Hi-C, and long-read data. *BMC Biology* **18**, 1-14 (2020).

- 679 99. A. Prasad, E. D. Lorenzen, M. V. Westbury, Evaluating the role of reference-genome
680 phylogenetic distance on evolutionary inference. *bioRxiv* 10.1101/2021.03.03.433733
681 (2021).
- 682 100. D. Y. Brandt *et al.*, Mapping bias overestimates reference allele frequencies at the HLA
683 genes in the 1000 genomes project phase I data. *G3: Genes, Genomes, Genetics* **5**, 931-941
684 (2015).
- 685 101. S. Gopalakrishnan *et al.*, The wolf reference genome sequence (*Canis lupus lupus*) and its
686 implications for *Canis* spp. population genomics. *BMC genomics* **18**, 1-11 (2017).
- 687 102. J. A. Clark, R. M. May, Taxonomic bias in conservation research. *Science* **297**, 191-193
688 (2002).
- 689 103. P. Cardoso, P. A. Borges, K. A. Triantis, M. A. Ferrández, J. L. Martín, Adapting the
690 IUCN Red List criteria for invertebrates. *Biological Conservation* **144**, 2432-2440 (2011).
- 691 104. H. R. Akçakaya, S. H. Butchart, G. M. Mace, S. N. Stuart, C. Hilton-Taylor, Use and
692 misuse of the IUCN Red List Criteria in projecting climate change impacts on biodiversity.
693 *Global Change Biology* **12**, 2037-2043 (2006).
- 694 105. N. Trull, M. Böhm, J. Carr, Patterns and biases of climate change threats in the IUCN Red
695 List. *Conservation Biology* **32**, 135-147 (2018).
- 696 106. M. W. Hayward *et al.*, Ambiguity in guideline definitions introduces assessor bias and
697 influences consistency in IUCN Red List status assessments. *Frontiers in Ecology and*
698 *Evolution* **3** (2015).
- 699 107. N. F. Thompson *et al.*, A complex phenotype in salmon controlled by a simple change in
700 migratory timing. *Science* **370**, 609-613 (2020).
- 701 108. N. J. Barson *et al.*, Sex-dependent dominance at a single locus maintains variation in age at
702 maturity in salmon. *Nature* **528**, 405-408 (2015).
- 703 109. D. E. Pearse *et al.*, Sex-dependent dominance maintains migration supergene in rainbow
704 trout. *Nature Ecology & Evolution* **3**, 1731-1742 (2019).
- 705 110. B. Epstein *et al.*, Rapid evolutionary response to a transmissible cancer in Tasmanian
706 devils. *Nature Communications* **7**, 12684 (2016).
- 707 111. C. Küpper *et al.*, A supergene determines highly divergent male reproductive morphs in the
708 ruff. *Nature Genetics* **48**, 79-83 (2015).
- 709 112. M. Kardos, A. Shafer, The peril of gene-targeted conservation. *Trends in Ecology &*
710 *Evolution* **33**, 827-839 (2018).
- 711 113. A. B. Shafer *et al.*, Genomics and the challenging translation into conservation practice.
712 *Trends in Ecology & Evolution* **30**, 78-87 (2015).
- 713 114. D. Pearse, Saving the spandrels? Adaptive genomic variation in conservation and fisheries
714 management. *Journal of Fish Biology* **89**, 2697-2716 (2016).
- 715 115. W. Funk, B. R. Forester, S. J. Converse, C. Darst, S. Morey, Improving conservation
716 policy with genomics: a guide to integrating adaptive potential into US Endangered
717 Species Act decisions for conservation practitioners and geneticists. *Conservation Genetics*
718 **20**, 115-134 (2018).
- 719 116. A. L. Hughes, MHC polymorphism and the design of captive breeding programs.
720 *Conservation Biology* **5**, 249-251 (1991).
- 721 117. O. Manlik *et al.*, Is MHC diversity a better marker for conservation than neutral genetic
722 diversity? A case study of two contrasting dolphin populations. *Ecology and Evolution* **9**,
723 6986-6998 (2019).
- 724 118. L. Laikre, Hereditary defects and conservation genetic management of captive populations.
725 *Zoo Biology* **18**, 81-99 (1999).

- 726 119. E. A. Boyle, Y. I. Li, J. K. Pritchard, An expanded view of complex traits: from polygenic
727 to omnigenic. *Cell* **169**, 1177-1186 (2017).
- 728 120. T. A. Manolio *et al.*, Finding the missing heritability of complex diseases. *Nature* **461**,
729 747-753 (2009).
- 730 121. J. Yang *et al.*, Common SNPs explain a large proportion of the heritability for human
731 height. *Nature Genetics* **42**, 565-569 (2010).
- 732 122. M. Kardos, A. Husby, S. E. McFarlane, A. Qvarnström, H. Ellegren, Whole-genome
733 resequencing of extreme phenotypes in collared flycatchers highlights the difficulty of
734 detecting quantitative trait loci in natural populations. *Molecular Ecology Resources* **16**,
735 726-741 (2016).
- 736 123. M. K. Oliver, S. B. Piertney, Selection maintains MHC diversity through a natural
737 population bottleneck. *Molecular Biology and Evolution* **29**, 1713-1720 (2012).
- 738 124. D. Garrigan, P. W. Hedrick, Perspective: detecting adaptive molecular polymorphism:
739 lessons from the MHC. *Evolution* **57**, 1707-1722 (2003).
- 740 125. P. S. Miller, P. W. Hedrick, MHC polymorphism and the design of captive breeding
741 programs: simple solutions are not the answer. *Conservation Biology* **5**, 556-558 (1991).
- 742 126. R. C. Vrijenhoek, P. L. Leberg, Let's not throw the baby out with the bathwater: a
743 comment on management for MHC diversity in captive populations. *Conservation Biology*
744 **5**, 252-254 (1991).
- 745 127. S. M. Haig, J. D. Ballou, S. R. Derrickson, Management options for preserving genetic
746 diversity: reintroduction of Guam rails to the wild. *Conservation Biology* **4**, 290-300
747 (1990).
- 748 128. P. W. Hedrick, P. F. Brussard, F. W. Allendorf, J. A. Beardmore, S. Orzack, Protein
749 variation, fitness, and captive propagation. *Zoo Biology* **5**, 91-99 (1986).
- 750 129. R. C. Lacy, Should we select genetic alleles in our conservation breeding programs? *Zoo*
751 *Biology* **19**, 279-282 (2000).
- 752 130. M. R. Jones *et al.*, Adaptive introgression underlies polymorphic seasonal camouflage in
753 snowshoe hares. *Science* **360**, 1355-1358 (2018).
- 754 131. H. Ellegren, B. C. Sheldon, Genetic basis of fitness differences in natural populations.
755 *Nature* **452**, 169-175 (2008).
- 756 132. S. J. Gould, R. C. Lewontin, The spandrels of San Marco and the Panglossian paradigm: a
757 critique of the adaptationist programme. *Proceedings of the Royal Society B* **205**, 581-598
758 (1979).
- 759 133. R. D. Barrett, H. E. Hoekstra, Molecular spandrels: tests of adaptation at the genetic level.
760 *Nature Reviews Genetics* **12**, 767-780 (2011).
- 761 134. J. P. Demuth, T. De Bie, J. E. Stajich, N. Cristianini, M. W. Hahn, The evolution of
762 mammalian gene families. *PloS one* **1**, e85 (2006).
- 763 135. F. W. Allendorf, Genetics and the conservation of natural populations: allozymes to
764 genomes. *Molecular Ecology* **26**, 420-430 (2017).

765
766
767
768 **Glossary:**

769 ***Adaptive potential:*** The ability of a population to evolve adaptively in response to selection.
770 Usually measured as narrow sense heritability (the proportion of phenotypic variance attributed to
771 additive genetic effects).

773 **Drift load:** The reduction in mean fitness of a population due to homozygosity for deleterious
774 alleles.

775 **F:** The individual inbreeding coefficient: the identical-by-descent fraction of an individual's
776 genome.
777

778 **Genetic load:** The reduction in fitness due to all genetic effects arising from both segregating and
779 fixed deleterious alleles.
780

781 **Genetic rescue:** Increase in population growth or reduction in genetic load arising from the
782 immigration of individuals with new alleles.
783

784 **h:** The dominance coefficient. A derived allele is recessive when $h=0$ (heterozygous genotypes
785 have the same mean fitness as homozygous wildtypes), and dominant when $h=1$ (heterozygous
786 genotypes have the same mean fitness as homozygous derived allele genotype), and additive when
787 $h=0.5$ (heterozygous genotypes have fitness midway between the alternative homozygous
788 genotypes).

789

790 **H:** Heterozygous fraction of an individual's genome.
791

792 **Hard selection:** Where an individual's absolute fitness depends only on its phenotype or genotype
793 and is independent of the phenotypes or genotypes of other individuals in the population.
794

795 **Identical-by-descent:** Two segments of DNA are identical-by-descent when they both descend
796 from a single haploid genome in a recent ancestor.
797

798 **Inbreeding:** Mating between relatives.
799

800 **Inbreeding depression:** Reduced fitness of individuals whose parents are related.
801

802 **Inbreeding load:** A measure of the potential for inbreeding to reduce fitness, measured by the
803 number of **Lethal equivalents**, which is a set of alleles that would on average cause death when
804 homozygous.
805

806 **Mutational meltdown:** Extinction of a population due to the synergistic interactions of population
807 decline, genetic drift, and the accumulation of deleterious alleles.
808

809 **π :** Nucleotide diversity: expected proportion of nucleotide differences between randomly chosen
810 pairs of haploid genomes in a population.
811

812 **Purging:** Increased selective elimination of deleterious, partially recessive alleles that are exposed
813 to purifying selection via inbreeding.
814

815 **Soft selection:** Selection where an individual's fitness depends on its phenotype or genotype
816 relative to others in the same population.
817
818

819 **Figure Legends**

820
821 **Figure 1.** Relationship of nucleotide diversity (π) with the inbreeding load (lethal equivalents)
822 (A), drift load (B), and additive genetic variance in a quantitative trait (V_a) (C). The data are from
823 the 1,000th generation of 10 simulated populations with 9 different constant effective population
824 sizes (N_e).

825
826
827 **Figure 2.** Genetic effects of bottlenecks with and without immigration. Nucleotide diversity (π)
828 (A), number of lethal equivalents (B), drift load (C), and the additive genetic variance in a
829 quantitative trait (V_a) (D) are shown for 100 generations after a simulated bottleneck in isolated
830 populations (orange) and with 5 immigrants every 2 generations up to generation 50 (blue).
831 Population size was held constant at $N_e=1,000$ for 1,000 generations before the bottleneck and then
832 at $N_e=25$ starting at generation 0. The thin lines show the results from 25 replicates. The thick lines
833 represent the mean across 25 replicates. Immigrants during the first 50 generations are from a
834 population with $N_e=500$ that split from the receiving population the generation of the bottleneck.
835 Details of the simulation model and parameters are provided in the *SI Appendix*.

836
837 **Figure 3.** Population viability during bottlenecks from carrying capacity $K=1,000$ (left column)
838 and $K=500$ (right column) to $K=100$. The bottlenecks were 2 (A), 10 (B), and 50 (C) generations
839 in length. The black line shows the proportion of extant populations. Gray lines show population
840 size for each of 50 replicate simulations in each scenario.

841
842
843
844
845
846
847
848
849
850
851

852 **Table legends**

853

854 **Table 1.** Deleterious mutation rates used in previous simulation-based analyses of inbreeding
855 depression and genetic rescue.

856