

Genomic inference of contemporary effective population size in a large island population of collared flycatchers (*Ficedula albicollis*)

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Abstract

Due to its central importance to many aspects of evolutionary biology and population genetics, the long-term effective population size (N_e) has been estimated for numerous species and populations. However, estimating contemporary N_e is difficult and in practice this parameter is often unknown. In principle, contemporary N_e can be estimated using either analyses of temporal changes in allele frequencies, or the extent of linkage disequilibrium (LD) between unlinked markers. We applied these approaches to estimate contemporary N_e of a relatively recently founded island population of collared flycatchers (*Ficedula albicollis*). We sequenced the genomes of 85 birds sampled in 1993 and 2015, and applied several temporal methods to estimate N_e at a few thousand (4000–7000). The approach based on LD provided higher estimates of N_e (20,000–32,000) and was associated with high variance, often resulting in infinite N_e . We conclude that whole-genome sequencing data offers new possibilities to estimate high (>1000) contemporary N_e , but also note that such estimates remain challenging, in particular for LD-based methods for contemporary N_e estimation.

KEY WORDS

contemporary N_e , genome sequencing, linkage disequilibrium, temporal method

1 | INTRODUCTION

Effective population size (N_e) is one of the key parameters in evolutionary biology. It is defined as the size of an idealized population (i.e., Wright-Fisher population; Fisher, 1930; Wright, 1931) that would have the same rate of genetic drift and inbreeding as the population in focus. N_e determines the amount and distribution of genetic variation in a population in interaction with several evolutionary forces like mutation, recombination, selection and migration

(Crow & Kimura, 1970). As a consequence, it is a good indicator of evolutionary potential and fitness (Lynch et al., 1995). It is also necessary to predict fixation probabilities of deleterious and beneficial alleles (Robertson, 1961), and is essential to infer demographic history (Hsieh et al., 2016; Juric et al., 2016; Ostrander et al., 2017). Several ways to estimate N_e based on genetic data have been developed (reviewed in Luikart et al., 2010; Wang, 2005; Wang et al., 2016). These approaches differ in the mathematical framework used and range from approaches based on classic population genetic theory (Crow &

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Kimura, 1970; Evans, 1979) to coalescent theory (Wakeley, 2008) to estimate either contemporary (very recent) or historical (long-term) N_e (Wang, 2005).

Methods that infer historical N_e have been applied to understand demographic history in a wide range of species (Charlesworth, 2009). The advent of next-generation sequencing (NGS) techniques and whole-genome resequencing protocols has allowed attempts to infer complex demographic scenarios (Beichman et al., 2018). The advances of demographic history inference include, for example, estimation of N_e of multiple nonequilibrium populations with different levels of connectivity (Steinrücken et al., 2019) and inference of historical N_e fluctuations over relatively short or long periods of time (Barbato et al., 2015; Browning & Browning, 2015; Li & Durbin, 2011; Santiago et al., 2020; Terhorst et al., 2017). In contrast, contemporary N_e remains difficult to estimate in nature, especially for large populations (Gilbert & Whitlock, 2015; Marandel et al., 2019; Serbezov et al., 2012).

In principle, by having information on demographic and life history parameters (such as census size, sex ratio, variance in reproductive success, mating system and/or pedigree) it is possible to estimate current N_e (Caballero, 1994; Wang, 2005; Wang & Caballero, 1999; Waples & England, 2011). However, detailed data on these parameters are rarely available and generally difficult to collect. In practice, genetic methods therefore have to be widely used (Palstra & Fraser, 2012). Contemporary N_e can be estimated by examining changes in allele frequencies over time (temporal methods; Hui & Burt, 2015; Jorde & Ryman, 1995, 2007; Krimbas & Tsakas, 1974; Nei & Tajima, 1981; Pollak, 1983; Waples, 1989). N_e can also be estimated from the extent of linkage disequilibrium (LD) between unlinked loci, as genetic drift generates correlated alleles at different loci (Hill, 1981; Waples & Do, 2010).

Temporal methods should give the most direct estimate of drift but come with the challenge of sampling a population of interest several generations apart, which can be especially problematic for organisms with long generation time. All genetic methods to estimate contemporary N_e are more powerful when N_e is small and when the signal of drift can be seen, even with small sample sizes and limited number of loci/alleles. In contrast, estimating current N_e in large populations ($N_e > 1000$) has proven to be challenging, at least when the number of markers is low, and often results in N_e estimates indistinguishable from infinity (Marandel et al., 2019; Waples & Do, 2010). However, in theory, data from a large number of loci should harbor sufficient information to provide information on N_e of large populations (Luikart et al., 2010; Wang, 2016; Waples & Do, 2010). How much data is needed to precisely estimate N_e in such cases remains unclear and conclusions are often based on simulation studies.

Here, we use large-scale genomic data to estimate contemporary N_e of an island population of collared flycatchers (*Ficedula albicollis*). We sampled 85 individuals at two time points 22 years (about nine generations) apart on Gotland, a Baltic Sea island that is thought to relatively recently have been colonized by collared flycatchers and with approximately 4500 current breeding pairs (L. Gustafsson, personal observation). Based on high coverage, whole-genome

resequencing data, we used both temporal and LD methods to estimate contemporary N_e with data at a scale that rarely has been applied to natural populations before.

2 | MATERIALS AND METHODS

2.1 | Study population

We sampled 85 collared flycatcher (*Ficedula albicollis*) individuals from the Baltic island of Gotland. Forty-five adult birds were sampled in 1993 and another 40 in 2015 (22 years or approximately nine generations apart). The collared flycatcher is a small passerine bird that breeds mainly in southeast Europe and southwest Asia but isolated populations are also found at two Swedish islands in the Baltic Sea (Gotland and Öland). Gotland has been inhabited for at least 150 years but the detailed colonization history remains unknown (Lundberg & Alatalo, 1992).

2.2 | Sequencing and data filtering

DNA was extracted from blood following established protocols (described in e.g., Burri et al., 2015). All individuals were sequenced with a paired-end approach on an Illumina HiSeqX instrument for a read length of 150 bp and an insert size of 350 bp. Reads were mapped to a repeat-masked collared flycatcher reference genome assembly, version FicAlb1.5 (GenBank Accession GCA_000247815.2), using BWA mem 0.7.13 (Li & Durbin, 2009) and further processed with Samtools 1.3 (Li et al., 2009). Reads were deduplicated with PICARD 2.0.1 (<http://broadinstitute.github.io/picard/>), and realigned and re-calibrated with GATK3.6 (DePristo et al., 2011). Variants were called independently for each time sample with GATK's HaplotypeCaller and GenotypeGVCFs 3. The mean genome-wide coverage varied from 30 to 50 among all sequenced individuals. After variant calling the data consisted of 19.8 million single nucleotide polymorphisms (SNPs) in the 1993 sample and 19.3 million in the 2015 sample. Aiming for a very high quality data set, we applied a strict filtering using VCFtools (Danecek et al., 2011). Specifically, we conservatively removed all SNPs where any of the individuals had a coverage lower than 10 or higher than 100, or a mapping quality below 20. Additionally, we removed all sites within 1 kb from scaffold ends. We considered variants that were segregating in both time data sets as well as variants that were only segregating in one of the two cohorts.

To obtain a large number of independent markers in a computationally efficient way, we proceeded in two steps. We first sampled 1,000,000 SNPs (from 29 chromosomes and the two largest unassigned linkage groups) before further removing all sites with neighbouring SNPs within a distance of 2 kb. This distance is known to be the approximate distance at which linkage gets back to background levels in the collared flycatcher (Ellegren et al., 2012, Figure S1; total map length has been estimated at 3132 cM [mean recombination rate for the whole genome equals

3.1 but ranges from 2.0–11.1 cM/Mb]; large chromosomes have smaller recombination rate; Kawakami et al., 2014). We used VCFtools to investigate genetic structure within and between time cohorts using a principal component analysis (PCA). F_{ST} (Weir & Cockerham, 1984) was also obtained between time cohorts, confidence intervals were obtained by resampling the data set 500 times before computing F_{ST} .

2.3 | Contemporary N_e estimation

We used the temporal and LD-based methods to estimate contemporary N_e . Temporal methods use changes in allele frequencies over several generations to estimate recent N_e . This approach relies on the idea that the variance in allele frequency change between generations is a function of N_e (Krimbas & Tsakas, 1974; Waples, 1989). The larger changes in allele frequencies over time, the smaller the inferred N_e . Linkage disequilibrium methods are based on the fact that random genetic drift in a finite population creates associations between linked and unlinked alleles and is therefore informative about N_e (Waples, 1989; Waples & Do, 2010). For both types of analyses, we filtered the data by excluding annotated genes, conserved elements (Craig et al., 2018), and regions with estimated recombination rate of zero (Kawakami et al., 2014).

We used three different temporal N_e estimators: a likelihood based \hat{N}_B by Hui and Burt (2015), and two different F -statistics: F_s by Jorde and Ryman (2007) and F_c by Nei and Tajima (1981). The likelihood-based estimator uses a computationally efficient hidden Markov algorithm and continuous approximation of allele frequencies. This approach makes the method well suited for estimation of larger N_e . The method is implemented in r package (NB) and we used a slightly modified version where we allow for a noninteger number of generations. We used a generation time of 2.5 years, N_e prior ranging from 50 to 100,000. The two additional F -statistics are moment-based estimators and can be calculated by obtaining standardized variance of allele frequencies changes. Both F -statistics were calculated in NEESTIMATOR v2.1 (Do et al., 2014). We used the plan I sampling procedure (sampling adults after the reproduction or before reproduction but returning them to population; Waples, 1989). To estimate LD-based N_e we used an approach developed by Waples and Do (2008), LDNe, implemented in NEESTIMATOR v2. LDNe is based on the mean of squared interlocus correlations of allele frequencies obtained from the Burrows method (Waples, 2006; Weir, 1996). We ran LDNe using two variations of the method, first including comparisons between all the SNPs in the data set and then omitting comparisons of loci on the same chromosomes while still comparing each SNP to all the SNPs on different chromosomes. The latter removed any physical linkage but greatly reduced the number of comparisons. We performed analysis for each time cohort independently. We performed jackknife to estimate 95% confidence intervals for F -statistics and LD-based estimate. In all methods we ignored SNPs

with a frequency lower than 5%. The number of SNPs after filtering was 78,636 and the median distance between neighbouring SNPs was >3 kb.

Additionally, in order to investigate the power of all methods, we created several smaller data sets for both time cohorts by varying the number of SNPs from 1000 to 78,636 in increments of 500 sites. To further investigate the influence of physical linkage, we varied the minimum distance between SNPs in the analysed data sets from 1 to 40 kb with increments of 200 bp, thus creating 196 additional data sets per time cohort. Importantly, those data sets dramatically vary in number of SNPs as number of SNPs and average distance between SNPs are inescapably correlated.

2.4 | Recent N_e changes over time

We used the GONE method to infer recent changes of N_e over time from linkage disequilibrium and SNP data (Santiago et al., 2020). We performed the analysis for each time cohort separately and applied no frequency filters to the SNP data sets. We used recombination rates from a collared flycatcher linkage map (Kawakami et al., 2014) and set maximum recombination rate between pairs of analysed SNPs to 0.01 (hc = 0.01).

3 | RESULTS

3.1 | Summary of data

We performed whole-genome resequencing of 85 adults of collared flycatcher from the Baltic Sea island Gotland at a mean coverage of 38.6 \times (range: 29.5 \times –50.2 \times). Forty-five birds were sampled in 1993 and 40 in 2015. We stringently filtered genotypes based on the coverage and mapping quality before removing any SNPs with missing data. Additional filtering of annotated genes, conserved elements, regions with estimated recombination rate of zero and non-independent SNPs based on physical proximity reduced our data to 131,902 SNPs.

This data set was used in all downstream analysis and filtered to 78,636 SNPs according to software settings. The average LD between the pairs of SNPs was $r^2 = .0228$ and $r^2 = .0257$ for 1993 and 2015 time cohorts, respectively. Additional data sets were created to test LD-based method performance as described above.

3.2 | Genetic structure

A PCA of genetic variation (Figure 1) suggested that there was no clear structure either between or within the time samples. This was corroborated by the observation of very low genetic differentiation between the cohorts ($F_{ST} = 2.6 \times 10^{-4}$; 95% CI: 1.73×10^{-4} – 3.34×10^{-4}). The genomic F_{ST} landscape was flat and no F_{ST} peaks were visible on any of the chromosomes (Figure S1).

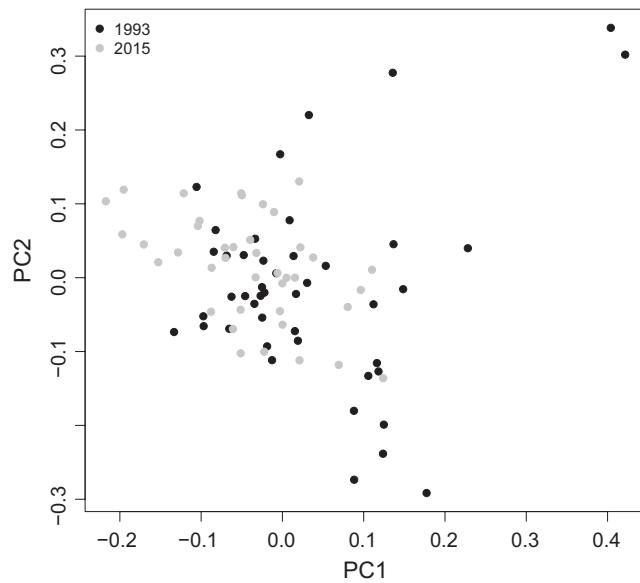


FIGURE 1 Principal component analysis (PCA) demonstrating the lack of structure between the two time samples of collared flycatchers from the Baltic Sea island Gotland. Individuals sampled in 1993 are represented in black and individuals sampled in 2015 are in grey

3.3 | Contemporary N_e estimation

Contemporary N_e estimates are summarized in Table 1. In general, all temporal methods gave relatively similar results of contemporary N_e ranging from approximately 4000 to 7000 individuals. The likelihood-based estimator provided the highest estimate of $\hat{N}_B = 6921$ (95% CI: 5015–11,079; Figure 2). Both F -statistics provided lower point estimates, $F_s = 5804$ (95% CI: 4837–10,341) and $F_c = 3921$ (95% CI: 3148–5198). In concordance with the results from temporal methods, GONE analysis showed that N_e for the most recent past equalled approximately few thousands (Figure 3). Additionally, the analysis indicated that the collared flycatcher population used to be higher (approximately 10,000) and slightly declined over the last 100 generations.

The LD-based method (with no linkage information included) provided higher estimates of contemporary N_e and varied from approximately 20,000 for the 2015 cohort ($N_e = 20,094$; 95%

CI: 8430–infinity) to 33,000 individuals for the 1993 cohort ($N_e = 32,534$; 95% CI: 10,670–infinity). We applied two corrections presented by Waples et al. (2016, Equations 1a and 1b) to account for linkage between SNPs in the data set. The corrected N_e estimates equaled approximately 38,000 and 23,000 for 1993 and 2015 time cohorts, respectively, for both corrections. The LD-based method that is restricted to interchromosomal comparisons, effectively taking into account physical linkage information by comparing each SNP to all SNPs on the other chromosomes, returned negative N_e with infinite credible estimates suggesting large N_e .

We varied the number of SNPs using both types of methods (temporal and LD) to explore the effect of sampling on N_e estimation (Figure 4 and Figure S2). Data sets with less than 30,000–40,000 SNPs provided N_e estimates with high variance, ranging from approximately 1000 to infinity. Data sets with a higher number of SNPs provided similar results to that obtained in the full analyses.

We varied the minimal distance between SNPs (1–40 kb; LD method) to explore the influence of physical linkage on N_e estimation (Figures S3 and S4). The median distance was always higher than the minimal distance and ranged from 1.8 to 63 kb. We observed a large variance in the obtained N_e estimates ranging from 9921 (2015 time cohort) and 12,642 (1993 time cohort) to infinity. Approximately half of the data sets with a distance of >10 kb between SNPs gave infinite N_e (44% and 53% for 1993 and 2015 time cohorts). The variance among data sets with SNPs with larger distance was also higher.

4 | DISCUSSION

Effective population size is a key concept in population genetics and evolutionary biology. Paradoxically, it is at the same time one of the most difficult parameters to estimate, especially for contemporary N_e and when N_e is large (1000 or larger). In this study, we estimated contemporary N_e in a wild bird population using methods based on linkage disequilibrium and temporal comparisons of allele frequencies. Our study is one of very few applying whole-genome resequencing data to estimate current N_e and using temporal and LD methods to estimate effective population size in a large natural population.

TABLE 1 Summary of N_e estimates using temporal and LD methods

Method	Data set	N_e	Low 95%	High 95%
Temporal - F_s^a	1993 and 2015	3921	3148	5198
Temporal - F_c^b	1993 and 2015	5804	4837	10,341
Temporal - \hat{N}_B^c	1993 and 2015	6921	5015	11,079
Linkage disequilibrium ^d	1993 cohort	32,534	10,670	Infinity
Linkage disequilibrium ^d	2015 cohort	20,094	8430	Infinity

^aJorde and Ryman (2007).

^bNei and Tajima (1981).

^cHui and Burt (2015).

^dWaples and Do (2008).

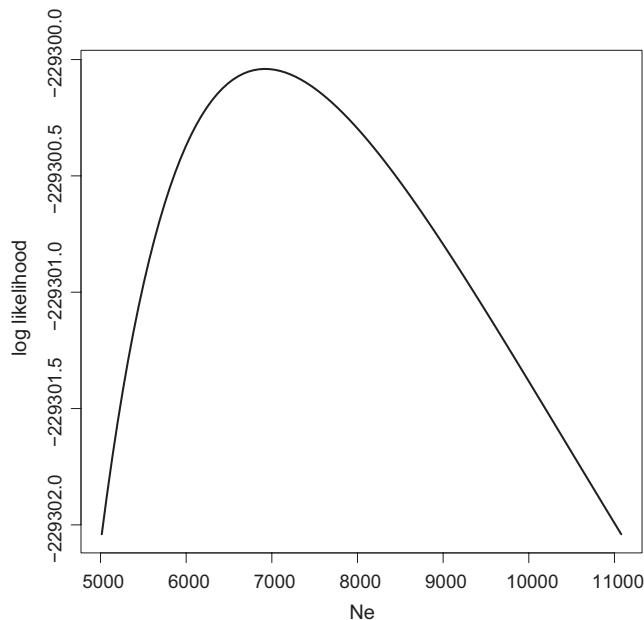


FIGURE 2 Likelihood-based N_e estimation (N_B) using temporal data

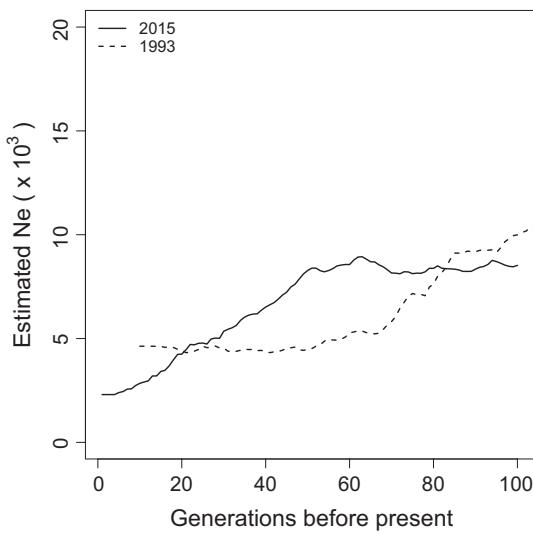


FIGURE 3 Recent changes in N_e inferred in GONE analysis

4.1 | Effective population size of Gotland collared flycatchers

We have previously estimated the historical, long-term N_e of collared flycatchers using Approximate Bayesian Computation modeling (Nadachowska-Brzyska et al., 2013; Nater et al., 2015) and the pairwise sequentially Markovian coalescent (PSMC; Nadachowska-Brzyska et al., 2016). About 200,000 years ago, N_e was large, $\sim 500,000$ – $600,000$. Populations of collared flycatchers decreased towards the middle of the Last Glacial Period (50,000 years ago), and then showed signs of steady increase towards 10,000 years ago. In contrast to mainland collared flycatcher populations, the Baltic

population declined to a level below 200,000 individuals around Last Glacial Maximum. This difference may reflect different ancestry of current collared flycatcher populations. It is important to note that PSMC analysis did not provide information on recent N_e , reflects historical N_e of an ancestral population and is sensitive to ancestral population structure and admixture (Li & Durbin, 2011).

The colonization history of collared flycatchers on Gotland is unknown, including lack of knowledge about when (after the Last Glacial Period) colonization took place and if it was associated with a severe bottleneck. The distance to the central European mainland is about 400 km, and the distance to breeding areas further south where collared flycatchers are abundant is larger than that. Collared flycatchers were registered on Gotland some 150 years ago (Lundberg & Alatalo, 1992), while Carl von Linné (Carl Linnaeus) did not make notes of the species when visiting Gotland in the summer of 1741 (Linnaeus, 1745). However, Linnaeus arrived on Gotland on one of the first days of July, when flycatchers no longer sing and are less conspicuous (unless seen feeding nestlings).

Whole-genome resequencing data indicated that the degree of genetic diversity in this island population (4.5×10^{-3}) is comparable to mainland populations (Burri et al., 2015). Both temporal and LD-based methods as well as GONE indicated a contemporary N_e of at least few thousands. This is in line with the results from a detailed analysis of identity by descent (IBD) segments of collared flycatchers from the nearby island Öland (Kardos et al., 2017), an island that was probably colonized by flycatchers from the Gotland population about 50 years ago. That analysis indicated that ancestral N_e , which probably reflects the N_e of the source (Gotland) population, was at least 5000. All these results suggest that the Baltic Sea population has been large for a relatively long time, and there was no strong bottleneck associated with the colonization event on Gotland island. Similarly, although GONE analysis indicated some decline of Gotland population over the last 100 generations, no drastic bottleneck was detected.

4.2 | Large N_e - performance of genetic methods to estimate contemporary N_e

Genetic methods to estimate N_e rely on the genetic drift signal present in the data. It follows that the larger the population the more difficult it is to estimate its effective population size. In practice, the estimates for large populations may be indistinguishable from infinity, especially when the number of individuals and loci analyzed are small (Waples & Do, 2010). Simulation studies have been used to evaluate the performance of methods based on temporal approaches and linkage disequilibrium, and usually considered populations of small to moderate size (Wang, 2002; Waples, 2006; Waples & Do, 2010; Waples & England, 2011). The amount of information used by temporal methods increases linearly with the number of loci. For LD-based methods, it increases with the square of the number of loci as it uses information on LD across all loci. Wang (2016) explored the possibility of estimating contemporary N_e of

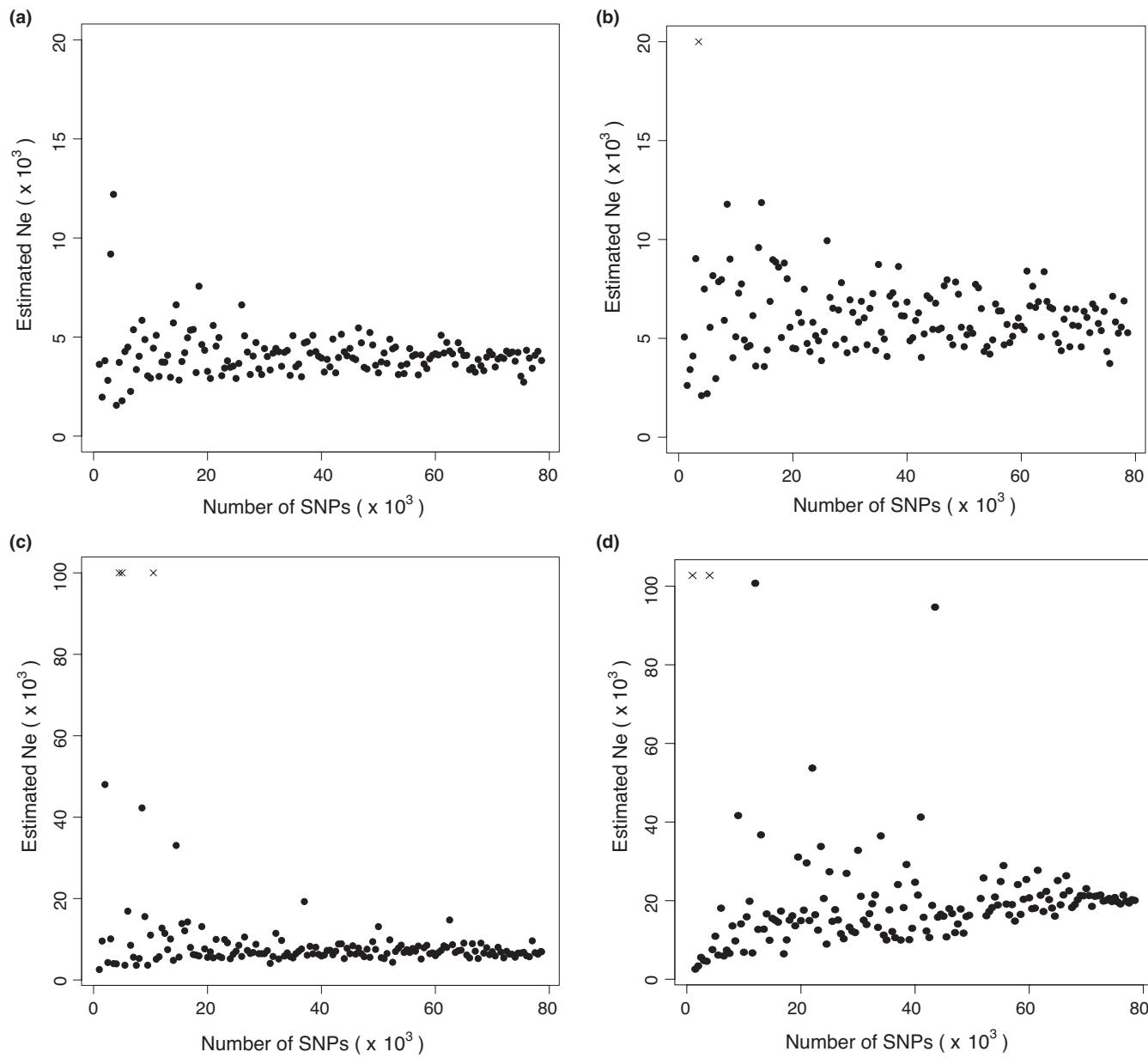


FIGURE 4 The relationship between the number of SNPs used in the analysis and N_e estimation using temporal methods and LD-based methods. (a) F_s (Jorde & Ryman, 2007); (b) F_c (Nei & Tajima, 1981); (c) \hat{N}_B (Hui & Burt, 2015); (d) LDNe (Waples & Do, 2008) for 2015 cohort. X indicates estimates that equal infinity

large populations and showed that LD-based methods can provide reasonably good estimates of N_e even for populations of a size as high as 30,000. These results were obtained from simulations with a sample size of 100 individuals and only 20 microsatellite loci. Another simulation-based study (Waples & Do, 2010) indicated that LD methods work well for small populations (100–200 or less), and temporal methods are more precise in contemporary N_e estimation. The study indicated that it is challenging to obtain estimates for large ($N_e > 1000$) populations due to very weak signal of drift. Similarly, results from a recent study by Waples et al. (2020) suggested that precision of LD based methods does not increase much with data sets larger than a few thousand loci. All studies mentioned above were based on simulations.

We investigated the possibility of estimating relatively large N_e in a natural population and tested a similar number of individuals but many more loci (78,636 SNPs after filtering) than in the simulations of Wang (2016). Using temporal methods, we were able to estimate contemporary effective population size with quite narrow credible intervals. All three temporal methods gave similar results suggesting that N_e of an order of a few thousand can be estimated when temporal genomic data are available. On the other hand, LD-based results were higher and were associated with much higher uncertainty. Several estimates obtained using LD-based methods on different number of SNPs and variable physical distance between neighbouring SNPs resulted in infinitive credible intervals. This result is in line with simulation studies indicating low power of LD-based methods

for contemporary N_e estimation when applied to large populations and with limited number of loci. Nonindependence of loci used in LD-based approaches may not only bias results but significantly also limit the power of LD-based methods due to pseudoreplication (Waples et al., 2020).

4.3 | Confounding factors

Genetic drift is not the only evolutionary force that changes allele frequencies over time and thereby affects patterns of LD along the genome. In the case of contemporary N_e estimation, migration is a force that can drastically influence the distribution of alleles in the population. Importantly, migration may bias N_e estimation in different ways. We expect overestimation of N_e when there is immigration from populations with limited differentiation to the focal population; we expect to see less drift due to the influx of alleles that are already present in the population. In this case the estimated N_e reflects the N_e of a metapopulation. Alternatively, we expect underestimation of LD-based N_e when the population exchanges migrants with substantially differentiated populations. In this case many foreign alleles enter the population, giving the impression of stronger drift.

The magnitude of the bias depends on the amount of migration between populations. Several simulation studies have evaluated the influence of migration on contemporary N_e estimation (Gilbert & Whitlock, 2015; Ryman et al., 2014) and concluded that migration rates below 1% ($m = 0.01$) does not have a strong effect on estimates (in some cases even migration of the order of 5%–10% did not lead to substantial bias; Waples & England, 2011). In the case of collared flycatchers on Gotland, a 1% migration rate would correspond to a large number of new birds coming to the island every year. The closest flycatcher population that could potentially serve as a migration source is located at Öland island. A substantial part of the Öland population is ringed and ringed individuals are extremely rare on Gotland island. While this observation excludes extensive migration between islands one cannot exclude immigration from other mainland populations.

Other factors that can potentially bias N_e estimation include selection and overlapping generations in the studied population. Selection creates linkage disequilibrium and changes in allele frequencies at genomic region under selection. We sought to minimize this effect by conservatively filtering functional regions potentially under strong selection pressure (genes, conserved elements) and regions of very low recombination rate where linked selection might be prevalent. When applying temporal methods, a bias might arise from using age-structured populations (overlapping generations). This can be minimized by taking temporal samples several generations apart, at least 3–5 generations apart (Waples & Yokota, 2007). With about nine generations between the two time samples analysed in this study, results comparable to that obtained with nonoverlapping generations can be expected.

It was suggested that whole-genome sequencing and/or genotyping of several thousands of loci can potentially overcome problems

associated with estimation of high contemporary effective population size (Luikart et al., 2010; Wang, 2016; Waples & Do, 2010). Our results indicated that several classic (F -statistics) as well as new (likelihood-based) temporal methods can provide reliable estimates of high contemporary effective population size. Nevertheless, the contemporary N_e estimation remains challenging. In particular, temporal methods need population samples taken several generations apart and such data may be unavailable for many populations of interest. Single sample estimators that rely on LD information do not have this limitation, but the results may provide very wide credible intervals and be difficult to interpret. The LD-based results for contemporary N_e estimation should always be interpreted with caution and preferably be augmented with another method for estimating contemporary N_e or recent population dynamics.

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CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

AUTHOR CONTRIBUTION

Krystyna Nadachowska-Brzyska, Ludovic Dutoit and Hans Ellegren conceived of the study and wrote the manuscript. Ludovic Dutoit performed all main analyses. Linnéa Smeds provided scripts and analysed raw data. Martin Kardos assisted with interpretation of the results and writing the manuscript. Lars Gustafsson provided samples.

DATA AVAILABILITY STATEMENT

The genome resequencing data have been made freely available in EMBL-EBI European Nucleotide Archive (<http://www.ebi.ac.uk/ena>) under accession number PRJEB22864.

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REFERENCES

Barbato, M., Orozco-terWengel, P., Tapió, M., & Bruford, M. W. (2015). SNeP: A tool to estimate trends in recent effective population size trajectories using genome-wide SNP data. *Frontiers in Genetics*, 6, 1–6. <https://doi.org/10.3389/fgene.2015.00109>

Beichman, A. C., Huerta-Sánchez, E., & Lohmueller, K. E. (2018). Using genomic data to infer historic population dynamics of nonmodel organisms. *Annual Review of Ecology, Evolution, and Systematics*, 49, 433–456. <https://doi.org/10.1146/annurev-ecolsys-110617-062431>

Browning, S. R., & Browning, B. L. (2015). Accurate non-parametric estimation of recent effective population size from segments of identity by descent. *American Journal of Human Genetics*, 97(3), 404–418. <https://doi.org/10.1016/j.ajhg.2015.07.012>

Burri, R., Nater, A., Kawakami, T., Mugal, C. F., Olason, P. I., Smeds, L., & Ellegren, H. (2015). Linked selection and recombination rate variation drive the evolution of the genomic landscape of differentiation across the speciation continuum of *Ficedula* flycatchers. *Genome Research*, 25, 1656–1665. <https://doi.org/10.1101/gr.196485.115>

Caballero, A. (1994). Developments in the prediction of effective population size. *Heredity*, 73, 657–679. <https://doi.org/10.1038/hdy.1994.174>

Charlesworth, B. (2009). Effective population size and patterns of molecular evolution and variation. *Nature Review Genetics*, 10, 195–205. <https://doi.org/10.1038/nrg2526>

Craig, R. J., Suh, A., Wang, M., & Ellegren, H. (2018). Natural selection beyond genes: Identification and analyses of evolutionarily conserved elements in the genome of the collared flycatcher (*Ficedula albicollis*). *Molecular Ecology*, 27, 476–492. <https://doi.org/10.1111/mec.14462>

Crow, J., & Kimura, M. (1970). *An introduction to population genetics theory*. Harper and Row.

Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., Handsaker, R. E., Lunter, G., Marth, G. T., Sherry, S. T., McVean, G., & Durbin, R. (2011). The variant call format and VCFtools. *Bioinformatics*, 27, 2156–2158. <https://doi.org/10.1093/bioinformatics/btr330>

DePristo, M. A., Banks, E., Poplin, R., Garimella, K. V., Maguire, J. R., Hartl, C., Philippakis, A. A., del Angel, G., Rivas, M. A., Hanna, M., McKenna, A., Fennell, T. J., Kernytsky, A. M., Sivachenko, A. Y., Cibulskis, K., Gabriel, S. B., Altshuler, D., & Daly, M. J. (2011). A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nature Genetics*, 43, 491–498. <https://doi.org/10.1038/ng.806>

Do, C., Waples, R. S., Peel, D., Macbeth, G. M., Tillett, B. J., & Ovenden, J. R. (2014). NeEstimator v2: Re-implementation of software for the estimation of contemporary effective population size (Ne) from genetic data. *Molecular Ecology Resources*, 14, 209–214. <https://doi.org/10.1111/1755-0998.12157>

Ellegren, H., Smeds, L., Burri, R., Olason, P. I., Backström, N., Kawakami, T., Künstner, A., Mäkinen, H., Nadachowska-Brzyska, K., Qvarnström, A., Uebbing, S., & Wolf, J. B. W. (2012). The genomic landscape of species divergence in *Ficedula* flycatchers. *Nature*, 491, 756–760. <https://doi.org/10.1038/nature11584>

Evans, W. (1979). *Mathematical population genetics*. Springer.

Fisher, R. A. (1930). *The genetical theory of natural selection*. Oxford University Press.

Gilbert, K. J., & Whitlock, M. C. (2015). Evaluating methods for estimating local effective population size with and without migration. *Evolution*, 69, 2154–2166. <https://doi.org/10.1111/evo.12713>

Hill, W. (1981). Estimation of effective population size from data on linkage disequilibrium. *Genetics Research*, 38, 209–216. <https://doi.org/10.1017/S0016672300020553>

Hsieh, P., Veeramah, K. R., Lachance, J., Tishkoff, S. A., Wall, J. D., Hammer, M. F., & Gutenkunst, R. N. (2016). Whole-genome sequence analyses of Western Central African Pygmy hunter-gatherers reveal a complex demographic history and identify candidate genes under positive natural selection. *Genome Research*, 26, 279–290. <https://doi.org/10.1101/gr.192971.115>

Hui, T., & Burt, A. (2015). Estimating effective population size from temporally spaced samples with a novel, efficient maximum-likelihood algorithm. *Genetics*, 200, 285–293. <https://doi.org/10.1534/genetics.115.174904>

Jorde, P. E., & Ryman, N. (1995). Temporal allele frequency change and estimation of effective size in populations with overlapping generations. *Genetics*, 139, 1077–1090. <https://doi.org/10.1093/genetics/139.2.1077>

Jorde, P. E., & Ryman, N. (2007). Unbiased estimator for genetic drift and effective population size. *Genetics*, 935, 927–935. <https://doi.org/10.1534/genetics.107.075481>

Juric, I., Aeschbacher, S., & Coop, G. (2016). The strength of selection against Neanderthal introgression. *PLoS Genetics*, 12, 1–25. <https://doi.org/10.1371/journal.pgen.1006340>

Kardos, M., Qvarnström, A., & Ellegren, H. (2017). Inferring individual inbreeding and demographic history from segments of identity by descent in *Ficedula* flycatcher genome sequences. *Genetics*, 205, 1319–1334. <https://doi.org/10.1534/genetics.116.198861>

Kawakami, T., Smeds, L., Backström, N., Husby, A., Qvarnström, A., Mugal, C. F., Olason, P., & Ellegren, H. (2014). A high-density linkage map enables a second-generation collared flycatcher genome assembly and reveals the patterns of avian recombination rate variation and chromosomal evolution. *Molecular Ecology*, 23, 4035–4058. <https://doi.org/10.1111/mec.12810>

Krimbas, C., & Tsakas, S. (1974). The genetics of *Dacus oleae* V. Changes of esterase polymorphism in a natural population following insecticide control - Selection or drift? *Evolution*, 25, 454–460. <https://doi.org/10.1002/cbic.200300625>

Li, H., & Durbin, R. (2009). Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*, 25, 1754–1760. <https://doi.org/10.1093/bioinformatics/btp324>

Li, H., & Durbin, R. (2011). Inference of human population history from individual whole-genome sequences. *Nature*, 475, 493–496. <https://doi.org/10.1038/nature10231>

Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., & Durbin, R. (2009). The sequence alignment/map format and SAMtools. *Bioinformatics*, 25, 2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>

Linnaeus, C. (1745). *Öländska och Gotländska Resa*. Kiesewetter Gottfried.

Luikart, G., Ryman, N., Tallmon, D. A., Schwartz, M. K., & Allendorf, F. W. (2010). Estimation of census and effective population sizes: The increasing usefulness of DNA-based approaches. *Conservation Genetics*, 11, 355–373. <https://doi.org/10.1007/s10592-010-0050-7>

Lundberg, A., & Alatalo, R. (1992). *The pied flycatcher*. T&AD Poyser.

Lynch, M., Conery, J., & Burger, R. (1995). Mutation accumulation and the extinction of small populations. *The American Naturalist*, 146, 489. <https://doi.org/10.1086/285812>

Marandel, F., Lorance, P., Berthelé, O., Trenkel, V. M., Waples, R. S., & Lamy, J. (2019). Estimating effective population size of large marine populations, is it feasible? *Fish and Fisheries*, 20, 189–198. <https://doi.org/10.1111/faf.12338>

Nadachowska-Brzyska, K., Burri, R., Olason, P. I., Kawakami, T., Smeds, L., & Ellegren, H. (2013). Demographic divergence history of pied flycatcher and collared flycatcher inferred from whole-genome re-sequencing data. *PLoS Genetics*, 9, e1003942. <https://doi.org/10.1371/journal.pgen.1003942>

Nadachowska-Brzyska, K., Burri, R., Smeds, L., & Ellegren, H. (2016). PSMC analysis of effective population sizes in molecular ecology and its application to black-and-white *Ficedula* flycatchers. *Molecular Ecology*, 25, 1058–1072. <https://doi.org/10.1111/mec.13540>

Nater, A., Burri, R., Kawakami, T., Smeds, L., & Ellegren, H. (2015). Resolving evolutionary relationships in closely related species with whole-genome sequencing data. *Systematic Biology*, 64, 1000–1017. <https://doi.org/10.1093/sysbio/syv045>

Nei, M., & Tajima, F. (1981). Genetic drift and estimation of effective population size. *Genetics*, 98, 625–640. <https://doi.org/10.1093/genetics/98.3.625>

Ostrander, E. A., Wayne, R. K., Freedman, A. H., & Davis, B. W. (2017). Demographic history, selection and functional diversity of the canine genome. *Nature Review Genetics*, 18, 705–720. <https://doi.org/10.1038/nrg.2017.67>

Palstra, F. P., & Fraser, D. J. (2012). Effective / census population size ratio estimation: A compendium and appraisal. *Ecology and Evolution*, 2, 2357–2365. <https://doi.org/10.1002/ece3.329>

Pollak, E. (1983). A new method for estimating the effective population size from allele frequency changes. *Genetics*, 104, 531–548. <https://doi.org/10.1093/genetics/104.3.531>

Robertson, A. (1961). Inbreeding in artificial selection programs. *Genetical Research*, 2, 189–194.

Ryman, N., Allendorf, F. W., Jorde, P. E., Laikre, L., & Hössjer, O. (2014). Samples from subdivided populations yield biased estimates of effective size that overestimate the rate of loss of genetic variation. *Molecular Ecology Resources*, 14, 87–99. <https://doi.org/10.1111/1755-0998.12154>

Santiago, E., Novo, I., Pardiñas, A. F., Saura, M., Wang, J., & Caballero, A. (2020). Recent demographic history inferred by high-resolution analysis of linkage disequilibrium. *Molecular Biology and Evolution*, 37(12), 3642–3653. <https://doi.org/10.1093/molbev/msaa169>

Serbezov, D., Jorde, P. E., Bernatchez, L., Olsen, E., & Vøllestad, A. (2012). Short-term genetic changes: Evaluating effective population size estimates in a comprehensively described brown trout (*Salmo trutta*) population. *Genetics*, 191, 579–592. <https://doi.org/10.1534/genetics.111.136580>

Steinrücken, M., Kamm, J., Spence, J. P., & Song, Y. S. (2019). Inference of complex population histories using whole-genome sequences from multiple populations. *Proceedings of the National Academy of Sciences of the United States of America*, 116, 17115–17120. <https://doi.org/10.1073/pnas.1905060116>

Terhorst, J., Kamm, J. A., & Song, Y. S. (2017). Robust and scalable inference of population history from hundreds of unphased whole genomes. *Nature Genetics*, 49, 303–309. <https://doi.org/10.1038/ng.3748>

Wakeley, J. (2008). *Coalescent theory*. Roberts and Company Publishers.

Wang, J. (2002). A pseudo-likelihood method for estimating effective population size from temporally spaced samples. *Genetics Research*, 78, 243–257. <https://doi.org/10.1017/S0016672301005286>

Wang, J. (2005). Estimation of effective population sizes from data on genetic markers. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 360, 1395–1409. <https://doi.org/10.1098/rstb.2005.1682>

Wang, J. (2016). A comparison of single-sample estimators of effective population sizes from genetic marker data. *Molecular Ecology*, 25, 4692–4711. <https://doi.org/10.1111/mec.13725>

Wang, J., & Caballero, A. (1999). Developments in predicting the effective size of subdivided populations. *Heredity*, 82, 212–226. <https://doi.org/10.1038/sj.hdy.6884670>

Wang, J., Santiago, E., & Caballero, A. (2016). Prediction and estimation of effective population size. *Heredity*, 117, 193–206. <https://doi.org/10.1038/hdy.2016.43>

Waples, R. S. (1989). A generalized approach for estimating effective population size from temporal changes in allele frequency. *Genetics*, 121, 379–391. <https://doi.org/10.1093/genetics/121.2.379>

Waples, R. S. (2006). A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. *Conservation Genetics*, 7, 167–184. <https://doi.org/10.1007/s10592-005-9100-y>

Waples, R. S., & Do, C. (2008). LDNE: A program for estimating effective population size from data on linkage disequilibrium. *Molecular Ecology Resources*, 8, 753–756. <https://doi.org/10.1111/j.1755-0998.2007.02061.x>

Waples, R. S., & Do, C. (2010). Linkage disequilibrium estimates of contemporary N_e using highly variable genetic markers: A largely untapped resource for applied conservation and evolution. *Evolutionary Applications*, 3, 244–262. <https://doi.org/10.1111/j.1752-4571.2009.00104.x>

Waples, R. S., & England, P. R. (2011). Estimating contemporary effective population size on the basis of linkage disequilibrium in the face of migration. *Genetics*, 189, 633–644. <https://doi.org/10.1534/genetics.111.132233>

Waples, R. K., Larson, W. A., & Waples, R. S. (2016). Estimating contemporary effective population size in non-model species using linkage disequilibrium across thousands of loci. *Heredity*, 117, 233–240. <https://doi.org/10.1038/hdy.2016.60>

Waples, R. S., Waples, R. K., & Ward, E. J. (2020). Pseudoreplication in genomics-scale datasets. *Biorxiv*. <https://doi.org/10.1101/2020.11.12.380410>

Waples, R. S., & Yokota, M. (2007). Temporal estimates of effective population size in species with overlapping generations. *Genetics*, 233, 219–233. <https://doi.org/10.1534/genetics.106.065300>

Weir, B. (1996). *Genetic data analysis II* (2nd ed.). Sinauer Associates.

Weir, B. S., & Cockerham, C. C. (1984). Estimating F-statistics for the analysis of population. *Evolution*, 38(6), 1358–1370. <https://doi.org/10.2307/2408641>

Wright, S. (1931). Evolution in Mendelian populations. *Genetics*, 16, 97–159. <https://doi.org/10.1093/genetics/16.2.97>

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