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Under the Alaska Salmon Research Task Force Act, NOAA Fisheries, on behalf of the Secretary of Commerce, in collaboration with the State of Alaska, was required to convene a task force to review existing Pacific salmon research in Alaska and identify applied research needed to better understand the increased variability and declining salmon returns in some regions in order to support sustainable salmon runs in Alaska. NOAA Fisheries and the Governor of Alaska were required to each appoint a representative to serve on the task force. The Task Force was made up of a diverse group of Alaska salmon knowledge holders, including members from federal, state, tribal, university, industry, and non-governmental organizations. This report was written by the Task Force and its Arctic-Yukon-Kuskokwim Working Group. The views, opinions, and recommendations expressed are only those of the Task Force and the Arctic-Yukon-Kuskokwim Working Group.

Executive Summary

Pacific salmon are an essential part of Alaska's cultural, commercial, recreational, and subsistence fisheries, providing economic opportunities to communities in Alaska as well as supplying important food and traditional and cultural practices for tens of thousands of Indigenous and rural people and communities. Due to its sheer size, Alaska maintains some of the best freshwater and marine habitats for salmon health and resilience. Despite this, some Alaska salmon populations are facing sustained and dramatic declines, with devastating impacts to food security and traditional ways of life for the people that depend on them. In addition, some salmon stocks are experiencing wide fluctuations in returns that lead to high inter-annual variation creating uncertain economic outcomes. Because of these sustained declines and increased variability in salmon returns, Congress enacted the Alaska Salmon Research Task Force Act (Appendix 1: Task Force Act) to identify the gaps in knowledge

that are needed to understand the variability and declining trends. The purposes of the act are to: 1) ensure that Pacific salmon productivity and abundance trends in Alaska are characterized and that research needs are identified; 2) prioritize scientific research needs for Pacific salmon in Alaska; 3) address the increased variability or decline in Pacific salmon returns in Alaska by creating a coordinated salmon research strategy; and 4) support collaboration and coordination for Pacific salmon conservation efforts in Alaska.



Critical Need to Understand Shifts in Alaska Salmon Productivity

Alaska is warming at a rate two times faster than the Lower 48 contiguous states, which is having a profound effect on Alaska salmon populations. Improved understanding of the mechanisms that regulate the distribution, migration, diversity and abundance of Alaska salmon will promote their conservation, allow for better projections, characterize uncertainty for production trends under climate warming, and enhance the sustainable fisheries management, food security, and economic security for Alaskans.

These dramatic shifts in Alaska salmon productivity are occurring even though freshwater habitats where Alaska salmon reside during their life history are relatively pristine, especially when compared to habitats that salmon stocks encounter at lower latitudes. Existing knowledge regarding Alaska salmon ecology indicates that warming in both freshwater and marine habitats is creating divergent impacts on salmon species and stocks where some are responding positively to warming (i.e., abundance is increasing) and others are declining in response. For example, residents of the Yukon River drainage have experienced declines in returning Chinook salmon since 2000 with minimal improvement and periodic crashes in chum salmon, most recently in 2020-2023. These declines have led to the cessation of in-river commercial fisheries for Chinook salmon and severe restrictions to subsistence fishing for all salmon species, including a complete closure of subsistence fishing for Chum and Chinook salmon in districts 5 and 6 in the upper Yukon and Tanana rivers in 2020 to 2023.

It is critical that we take action now to understand mechanisms driving Alaska salmon production and to provide a path for mitigating negative impacts. Acting now allows for a response before there is further decline, making immediate action more successful and cost effective than waiting until stocks become critically low or reach depleted status.

As such, the Task Force members were acutely aware of the impact the declining Chinook and chum salmon returns in western Alaska are having on culture and food security in that region. Through the Task Force Act, the Task Force formed the Arctic Yukon Kuskokwim Working Group (AYK WG) to identify priority research needs for salmon in that region. Public testimony often included recommendations for action that can be done now, such as management actions to close fisheries where western Alaska salmon are harvested as bycatch or interception. Although the Task Force

understood it would be impossible to meet all expectations, our commitment was to develop priority research needs that would enable decision makers at local and regional levels to act quickly in response to research needs.

It is through this lens that the Task Force set out to recommend a coordinated research strategy based on the review of existing Pacific salmon research and the identification of knowledge gaps, and applied research needed to better understand why Alaska salmon are experiencing increased variability and declines.

Gravel to Gravel (G2G) life history research strategy

Salmon life begins and ends in the gravel and throughout their life history they depend on freshwater, nearshore estuarine, and marine habitats to grow and thrive. Through G2G, Tribes, State and Federal agencies and institutions, and others work together to build a strong foundation for co-stewardship, where Traditional and Indigenous Knowledge along with western science play important parts in maintaining resilient habitats and communities within Alaska.

Potential drivers influencing Pacific salmon production in Alaska and recommended applied research needs

Based on a review of existing knowledge and public comment, the Task Force identified the following potential drivers influencing production within the Pacific salmon life cycle in Alaska and associated priority research needs. The list of potential drivers are provided in order of the number of Pacific salmon life history stages impacted from all to fewer. While a larger number of research needs were discussed, the list here illustrates the priority research needs decided through discussions within the Task Force and from input through Public comment.

Warming climate and extreme events

• Research to understand and quantify the effects of natural environmental variability and warming climate on Alaska salmon distribution and abundance.

Salmon health and condition

- Research to understand the connections between freshwater, estuarine, and the marine environment that lead to pathogens or declines in some vitamin levels for salmon as these changes could affect the ability of returning adults to successfully reach the spawning grounds, successfully spawn, the numbers of eggs produced and fertilized, and their ability to produce viable offspring.
- Research to understand prey quality and quantity on health and condition of salmon in marine estuarine and freshwater habitats.
- Research to understand the mechanism(s) behind declining size at age as these declines impact the amount of food available per fish, the size and number of eggs per female for future generations, and can contribute to declining run sizes.
- Research to understand the effects of ocean acidification on health, behavior, prey availability/quality for salmon in marine environments.

Predators

- Research to understand potential conflicts between predator or endangered predators and prey (salmon) within freshwater, estuarine, and marine habitats.
- Research to address the role hatchery salmon release sites have on drawing in assemblages of predator species that otherwise would not be present in coastal nurseries, thereby potentially increasing predation pressures on wild stocks that may also inhabit these nurseries.



Marine food limitations

- Research to understand the implications of habitat use by Alaska salmon populations at various levels of abundance, the productive capacity of habitats for each life stage, and the potential implications of density-dependent effects.
- Research to better understand the role of salmon in pelagic communities, the food availability for salmon and the nutritional quality of prey organisms, including harmful algal blooms, to better understand inter-and intra-specific competition among salmon at sea.

Marine harvest and bycatch

- Research to reduce bycatch, interception, and Illegal, Unreported, and Unregulated (IUU) fishing through improved understanding of distribution and migration patterns of Alaska salmon stocks to better predict and avoid incidental harvest in the migratory corridors for Alaska salmon including Bering Sea, Aleutian Island, and Gulf of Alaska areas and regions in the North Pacific where there is increased potential for IUU fishing.
- Research to improve our ability to determine the stock origin (wild and hatchery) of chum and Chinook salmon taken in marine harvest, bycatch, and interception.
- Research to understand the frequency of occurrence and mortality rate (direct and discard) attributed to unobserved fishing mortality (e.g., IUU, unreported catch, incidental catch/mixed stocks); and once this information is known, determine the impact to the populations.
- Research to better define how all sources of marine harvest, including mixed stock fisheries, influence salmon abundance and how this varies by species, stock, and region.
- Research to understand genetic diversity and fitness effects from fishing or hatchery influence that may reduce a population's resilience and ability to recover from climate induced depression of population abundance or low productivity.

Freshwater habitat changes

- Research to develop meaningful measures of ecosystem performance (space and time scales) that supports biological diversity of Alaska salmon to maintain and conserve the processes that confer resilience (habitat and/or genetic diversity) in the face of ongoing environmental change.
- Research that improves our understanding of the impact of hatchery strays on wild salmon where intermingling with those stocks in freshwater has the potential to reduce genetic diversity, reproductive success, suffocation from carcass loading, and resilience to climate variability and warming.

• Research to explore range expansion and colonization of new habitats.

Freshwater harvest

- Research that addresses mortality rate attributable to unobserved freshwater fishing mortality due to release, incomplete capture, unreported catch, and illegal fishing and how this source of mortality impacts the populations.
- Research that documents production changes, spawning success, and affects on other species in the high Arctic as salmon begin to return in larger numbers to the region.

Recommended Applied Strategies to Address Priority Research Needs

Improved understanding of the social impacts

Develop/improve understanding of the impact of declining salmon returns to food security, nutrition and physical/ mental health, social cohesiveness, culture and language/ knowledge transfers, and traditional ways of life.

Improved stock identification methods

Develop/improve novel stock and fish identification methods at a finer scale than is currently available from genetic mixed stock analysis. In some areas, like western Alaska, current stock groupings based on genetic distinctions cover wide geographical areas that do not allow a full understanding of the impacts of marine harvest at the finer resolution used for management and impact assessment.

Better characterization of ocean distributions and marine migration routes

Develop/improve/expand research to understand the migratory routes and ocean distributions for western Alaska salmon to reduce bycatch potential (*see recommendations by the Alaska Bycatch Review Task Force report dated November* 2022; Appendix 6) and interception and to understand potential for impacts on their health from shifting food webs and competitive pressures.

Expanded ocean ecosystem surveys

Develop/improve/continue ocean surveys to understand how shifts in climate and ocean conditions (climate warming/extreme events) impact the food web and health and condition of juvenile salmon populations, with the primary research goal to identify additional marine management actions, with the secondary goal of improving forecasts of short- and long-term prospects for decision makers.

Strategies to minimize human impacts on freshwater and coastal habitats

Develop/improve/expand strategies to prioritize actions that reduce human impacts on freshwater and coastal ecosystems with the goal of maximizing the number, diversity, and health of wild smolts and spawners.

Making use of new technologies

Develop/improve/expand use of new technologies and advanced analytical methods for Alaska salmon research, including molecular identification, genomics, environmental DNA, mass marking, intelligent tags, salmon observation systems, biochronologies and life history reconstruction, and remote sensing/ autonomous vehicles.

More effective monitoring of salmon indicator stocks

Develop/improve/continue to identify indicator stocks for Chinook and chum salmon that can be monitored and tracked throughout their life cycle to better understand mechanisms impacting survival in marine and freshwater habitats and provide an early warning system.

Improved stock assessments for in-season management

Develop/improve/continue stock assessment programs that allow for timely in season management decisions to mitigate uncertainties in adult salmon return strength.

Life-cycle modeling and management strategy evaluations for climate resilience

Develop/improve/expand approaches to modeling biological impacts of climate warming and extreme events on the full life cycle of Pacific salmon that include management strategy evaluations to test how different management actions may impact production and to address adaptive research/ management strategies in relation to climate scenarios that consider management approaches that facilitate adaptation, ensemble models to characterize uncertainty in climate impact projections, and ocean intelligence systems for targeted information on impacts of climate warming and extreme events on ocean ecosystems and salmon growth and health.

Better data management and sharing

Develop a salmon data information/knowledge portal with FAIR (Findable, Accessible, Interoperable, and Reusable) and CARE (Collective Benefit, Authority to Control, Responsibility, and Ethics) principles such as envisioned by the *Research Networking Activity for Sustained Coordinated Observations of Arctic Change (RNA CoObs) – Supporting* the Roadmap for Arctic Observing & Data Systems (Arctic ROADS) for data sharing among communities, researchers, and managers. An open access salmon information/ knowledge portal such as this would enable the proposed G2G assessment approach of coordinated research where individual projects, regardless of whether they are led by state, federal, university, tribal, or non-profit entities, will share information with all other projects, with intentional integration of data and information across research projects.

Recommended Framework

Research project development should include a framework that involves "salmon people" across tribal, federal, state, non-profit, international, and other entities. One example would be to initiate "Two-Eyed Seeing" framework that embraces "learning to see from one eye with the strengths of Indigenous knowledge and ways of knowing, and from the other eye with the strengths of mainstream knowledge and ways of knowing

"KRITFC emphasizes that efforts to narrow these data gaps and pursue research under these (or other) themes should include Tribal entities as partners and collaborators from the outset of the project development and funding application process. Local and Tribal people should be employed in these projects as much as possible. A coordinated salmon research strategy should consider gravel-to-gravel impacts to salmon health and viability. It should account for salmon throughout their life cycle and across management and research jurisdictions. This strategy would be inherently ecosystem-based in nature and account for interactions and relationships between salmon and all other species and ecosystem elements they encounter, including weather and climate, predators, prey, similar pelagic foragers, seabirds, vegetation, and humans - those that harvest salmon intentionally (i.e., subsistence fishers) and unintentionally (i.e., bycatch). Western scientific, social scientific, and Traditional research would be supported and included in this strategy.

Tribes and Tribal organizations, NGOs, state agencies, federal agencies, and international entities should band together to understand salmon, and what is happening to salmon. These research efforts should be led by Salmon People – primarily Alaska Native people and organizations – and should be centered on Indigenous data, knowledge, and wisdom as well as Western science. Stable, non-competitive funding should support



these freshwater-to-marine research efforts, and substantial amounts of this funding (i.e., more than 50%) should be dedicated to Tribal governments and (inter)Tribal organizations to employ Tribal citizens in monitoring work.

Further, this gravel-to-gravel salmon research strategy should connect with policy to protect and restore salmon to abundance. Research efforts should not just gather information (i.e., about climate impacts, habitat changes, or fine-scale genetics) for the sake of understanding, but to inform Tribal, state, and federal decision-makers of the best pathways forward to recovering salmon stocks. Informing and effecting meaningful policy and management actions should be the primary focus for the research strategy. KRITFC appreciates that this Task Force has endeavored to be expansive and inclusive. Accepting a wide variety of experts to the Task Force, an unlimited number of members to the AYK Working Group, and regular public comment is encouraging to our Tribes.



We emphasize that partnerships amongst all research and management entities are key for expanding our understanding of salmon. Tribal entities are engaged and ready to collaborate with federal, state, and NGO entities to protect our salmon and ways of life – neither of which can be separated from the other." — Jonathan Samuelson (Chair of the Kuskokwim River Inter-Tribal Fish Commission, KRITFC; Quote, March 15, 2024)



Coordinated Research Strategy Gravel to Gravel / Life History Figure



Salmon life begins and ends in the gravel (Gravel to Gravel – G2G) and throughout their life history they depend on freshwater, estuarine, and marine habitats to grow and thrive. It is through the utilization of diverse, pristine habitats found in Alaska that salmon can gain resiliency to the effects of climate warming. However, Alaska salmon are facing greater challenges than those at lower latitudes due to the pace of warming (two times faster than lower latitudes) and extreme events (marine heat waves, loss of seasonal sea ice, drought, spikes in freshwater temperatures, exposure to metals in freshwater rivers due to melting permafrost, loss of aquatic vegetation and forage fishes, etc.) that can have dramatic physical, chemical, and food web impacts that affect their survival. Through G2G federal agencies, Tribes, state agencies and institutions, and others can work together to build a strong foundation for co-stewardship, where traditional and Indigenous Knowledge along with western science play important parts in support of resilient habitats and communities within Alaska.



We propose a G2G assessment approach of coordinated research where individual projects, regardless of whether they are led by state, federal, university, tribal, or NGO entities, will share information with all other projects. This strategy includes an intentional integration of data and information across research projects. Prior salmon research efforts have undoubtedly enabled important advancements in our knowledge and understanding of salmon abundance patterns across Alaska. However, when each research project is advanced and understood in isolation, which is the norm, we often fail to develop a synthesized and holistic perspective across the entire salmon life cycle. Consequently, it becomes difficult to develop an integrated and unified picture of the nature of where salmon bottlenecks are, and to clearly identify the most important drivers of these bottlenecks. Perceived progress takes a long time to develop, and it becomes challenging for experts with different perspectives to coalesce under a common understanding.

To do this, for each stock selected for study, an intensive suite of studies will need to be implemented concurrently over a 5 to 6-year period. Together, the suite of projects should address all relevant potential drivers for salmon abundance identified in the Task Force report and all life stages, ideally with overlap across multiple projects. In addition to data collection studies, retrospective analyses and modeling efforts will be useful to "bring it all together" by consolidating data across suites of projects and integrating data from separate studies. This synthesis will highlight critical factors or life stages limiting salmon adult run abundance and inform potential policy and management actions. Research approaches that are intensive and holistic, employing coordinated and focused examinations of all potential drivers at once, have been particularly successful for identifying factors most important to survival and productivity of salmon in other areas (e.g., Salish Sea Marine Survival Project), and it is expected that the G2G assessment approach would be similarly successful.

Potential Drivers for Alaska Salmon Production

"I think there's a lot of things that we should be looking at...drivers of the system... bigger ecosystem kind of approach." — Charles Lean, Public Testimony Day 1, 3:06:15

Based on a review of existing knowledge, the Task Force identified the following potential drivers influencing production throughout the Pacific salmon life cycle in Alaska (listed in order of the number of salmon life history stages impacted (all to less); no priority assigned): warming climate/extreme events, salmon health and condition, predators, marine food limitations, marine harvest and bycatch, freshwater habitat changes, and freshwater harvest. Research gaps were then identified within these potential drivers of Alaska salmon productivity which set the stage for the Task Force to develop priority research needs and applied strategies (listed in the Executive Summary) to address each potential driver.

Warming Climate/Extreme Events (impacts all life stages)

"Temperature was in the 90s three or four days...Our river was too warm for those salmon to survive... salmons were being overheated and they floated down the river." — James Nicori, Public Testimony Day 1, 2:52:25

Alaska is warming at a rate two times faster than the lower latitudes, and this warming is affecting all aspects of the salmon life cycle. Indirect impacts of climate-related phenomena include changes to timing of salmon smolt outmigration and upriver migration, reduced fitness in response to shifts to lower quality prey as well as shifts in their ocean migration and distribution patterns. Warming is also increasing the frequency of extreme events that have profound negative impacts on some species and stocks of salmon depending on what freshwater systems are utilized for spawning and the migration routes taken during their marine life history. Extreme events can lead to short term (days to weeks) changes in water flow, oxygen levels in freshwater, spikes in freshwater temperatures, and wildfires as well as seasonal changes (months) such as loss of seasonal sea and lake/river ice, and longer-term phenomena (months to years) such as marine heat waves. These extreme events have direct physiological impacts on salmon stocks, such as heat stress that can compromise the success of returning adult spawners or juveniles in the ocean and reduce health and condition of salmon in the marine environment in response to shifts to lower quality prey as well as shifts in their ocean migration and distribution patterns.

Salmon Health and Condition (impacts all life stages)

"...we're learning very quickly...that several of these salmon stocks are just becoming far less productive..." — Vanessa von Biela, Public Testimony Day 1, 4:08:20

Changes to the health and condition of Pacific salmon in Alaska have been noted by fishers and biologists. Across Alaska, Chinook salmon and other species of salmon have been experiencing declining size at age as well as shifts in age at maturity (to younger age). Other changes include increased presence of ichthyophonus (a fungal-like infection) and a reduction in thiamine levels in Chinook salmon, both of which are believed to come from their marine prey. Other changes that are found for Alaska salmon include deficiencies in fat stores, particularly during early marine life history stages and potentially during adult spawning migration. All of these changes could increase mortality rates during marine life history stages, affect the ability of returning adults to reach spawning grounds and spawn successfully, and decrease their ability to produce viable offspring.

Predators (impacts all life stages)

"On my hunting operation where I guide out of Huslia...grayling ate (them - salmon eggs)." — Virgil Umphenour, Public Testimony Day 1, 3:32:18

Predation occurs throughout the Pacific salmon life cycle, but can be difficult to assess, especially in estuarine and marine environments. Salmon sharks are estimated to consume 73 to 146 million Pacific salmon each year. Many marine mammal predators of salmon, particularly seals and sea lions, have increased in abundance in recent decades. Changes in subsistence practices along some Alaska rivers, for example, the decreased need to harvest fish such as northern pike to feed dog teams, has resulted in increasing abundance of freshwater predators in some places, as has the expansion of pike beyond its native range.

Socio-economic changes since the 1970s have also affected the ecological system. As snow machines replaced dog teams for transportation, community residents kept fewer dogs and needed fewer salmon and non-salmon fish species, such as sheefish and Northern pike, to feed them. Some researchers have theorized that the decreased harvests of these piscivorous fish have led to increased predation on juvenile salmon in the freshwater environment.

Marine Food Limitations (impacts marine life history stages)

"The ladies cutting the fish... say, "Come look at this fish." The intestines were welded to their meat and to their bones. They can't figure out why that is happening. And sometimes... growth on the body... and they smell differently." — James Nicori, Public Testimony Day 1, 2:50:25

Many Alaskan salmon stocks are experiencing particularly poor marine survival, which could be due to marine food limitations. Pacific salmon move from rivers to the ocean to take advantage of better feeding and growing opportunities there. However, there are several indications that critical changes have occurred for Alaskan salmon marine feeding and growth opportunities that adversely impact their marine survival and/or their health and condition when they return to their natal spawning rivers. For instance, local and Indigenous knowledge holders are raising concerns about deteriorating fat content and health of returning salmon. Additionally, a growing body of scientific literature associates many of these abundance or size declines with competition among salmon species, including those of hatchery origin. Therefore, it is critical to understand the mechanisms and degree to which marine food limitations may be causing poor returns of Alaskan salmon, and to understand what actions could possibly mediate these conditions.

Marine Harvest and Bycatch (immature and maturing life stages)

"My big concerns are... bycatch in the pollock industry...and Area M fishery." — Brooke Woods, Public Testimony Day 2, 2:19:45

Marine harvest of Pacific salmon occurs at multiple scales (i.e., international, national, state, regional, etc.) and there are many complexities to reporting and attributing catch (limits to genetic stock identification) at these various levels. In addition, most ocean fisheries have some amount of bycatch or interception of Pacific salmon as part of the harvest process. There are challenges in tracing bycaught salmon to their stock of origin given that the various stocks intermingle in the marine environment. This is evident within the eastern Bering Sea where a majority of the prohibited species catch for chum salmon is Asian origin, much of which are produced in hatcheries, and the difficulty in separating western Alaska summer chum salmon stocks among rivers due to the limits imposed by current methods of genetic stock separation.

Freshwater Habitat Changes (spawning adult to smolt life stages)

"... the beavers, they dam the river where the spawners can't even go through the dam...seeing... lots of beaver dams. There used to be no beavers in our area (Kuskokwim River)...migrating into the lower streams...down to the coast now." — James Nicori, Public Testimony Day 2, 2:26:30

Much of Alaska's freshwater habitat is considered relatively pristine compared to those in lower latitudes. While intact landscapes are most likely to support biological diversity and the reliable delivery of salmon to ecosystems and people, they remain subject to large-scale drivers such as warming climate. For example, intact freshwater landscapes help to buffer environmental variability and contribute to long-term stability of salmon populations through differing responses to varying conditions (much like the stabilizing effect of asset diversity on financial portfolios), yet the buffering can be overwhelmed in times of drought or changing water tables. In some regions, Pacific salmon freshwater habitat has been impacted by human actions such as mine-tailings, oil spills, and construction over salmon habitat. In addition, some rivers in Alaska are turning bright orange and can be as acidic as vinegar due to metals that escape from melting permafrost.

In some regions of Alaska, dramatic changes to freshwater habitat have been brought on by glacial recession, isostatic rebound, and tectonic forces. These changes to the landscape impact freshwater habitats in countless ways, both positive and negative, across the state. Intact habitat allows for salmon populations to respond positively in some cases, such as in Glacier Bay in Southeast Alaska, where receding glaciers have resulted in new freshwater habitats and colonization by salmon. In other cases, invasive species such as Elodea present a significant risk to salmon streams in some regions of Alaska as the plant affects the quality of habitat for juvenile salmon. Hatchery adults also stray into streams where wild stocks are spawning and have been known to intermingle with those stocks potentially reducing genetic diversity, reproductive success, and resilience to climate variability and change. The presence of hatchery strays can also make it difficult to monitor escapements of wild salmon by inflating aerial and foot survey counts, and has resulted in reductions in geographic coverage of wild stock escapement indices in some areas where high hatchery stray proportions have been documented.

Traditional Knowledge holders often communicate that sport fishing can interfere with salmon survival, both through the physical disturbances caused by sport fishers walking in streams and on riverbanks, as well as through overall disrespectful behavior towards salmon often described as "playing with one's food." In addition, more roads along freshwater river systems can create new challenges: "A chemical sprayed on tires actually kills salmon...driving on roads...can affect the salmon."

Freshwater Harvest (adult life stage)

"If the fishing got real low, people would pull their nets on their own." — Victor Lord, Public Testimony Day 1, 3:58:00

The effect and magnitude of historical freshwater harvest (commercial, sport, and subsistence) on salmon populations is not well understood. Even though freshwater harvest has been reduced or eliminated in some areas because of recent declines, the productivity of stocks continues to decline, which suggests that freshwater harvest is not the primary driver on the abundance relative to other influences. The biggest information gap for subsistence harvest is in the Arctic, but overall this is not seen as a major influence on abundance relative to other influences. Beyond harvest levels, traditional knowledge about freshwater harvest provide important clues about the environment as a whole and ecological connections sometimes undetected in the biological sciences. For example, local observations of natural indicators, or those environmental signals that fishers have connected to salmon run timing and abundance can be fertile areas for scientific inquiry.

"When the fish come in ... it depends on the wind. If it's from the east, the more fish will be on the west side because the west side is high water. When it's from the west, the fish are across there along the east riverbank of Kuskokwim because more water ... And when there's north wind, the tide not come in that much, it'll be low water." — Adolph Lupie, Tuntutuliak

For subsistence harvest, traditional and Indigenous Knowledge illustrates that salmon provides not only an essential food source for families, but also supports important cultural, linguistic, and family traditions that encourage health and wellbeing within communities. Changing patterns in the harvest and use of salmon continue to drive disconcerting social changes in the region, such as reducing the time that families spend together at fish camps and the resulting challenges to passing on cultural knowledge between older and younger generations.

"We have to respect them and keep what you do catch very clean and handle them carefully...The animal, fish, or something we take into our home the women they take care of it right away...with no complaining...because the person who hunts will get more not less...if we leave it... will not catch anymore while others are catching more. We have to respect them and keep what you do catch very clean and handle them carefully." — Adolph Lupie, Tuntutuliak

"...our people are into sharing. Elders first. Families first. We never kept the first king. We shared it." — Evon Waska, Task Force Interview, 7:26

"Our cultural and our self-identity was giving to the Elders and sharing our catch."

- Evon Waska, Task Force Interview, 7:26



Existing Knowledge

Because of salmon's importance to food security as well as its cultural and economic value, there is a lot known about what salmon need to thrive, particularly in the freshwater phase of their life cycle. Adult salmon require the ability to move from the ocean to freshwater habitats, which must provide the conditions that support the healthy development of eggs, fry, and juveniles. Collectively, freshwater habitat quality can be characterized by the 4Cs: "clean, cool, complex, and connected". Spawning beds must consist of clean gravel that is free of silt, or there must be sufficient movement of water between the stream and its gravel bed ("hyporheic flow", or upwelling/downwelling) to prevent eggs and embryos from suffocating. Streams and lakes must be free of toxic levels of heavy metals, pesticides, and other pollutants. Complex habitats, such as rivers with healthy floodplains, are important for fueling food webs and giving juvenile salmon the ability to move into side channels that might have better feeding conditions or fewer predators. Habitat features such as undercut banks and logiams provide small salmon protection from high flows and from predators, and lakes and beaver ponds can provide good overwintering habitat for species such as coho salmon that spend at least a full year in freshwater before going to sea. Rivers with good forest cover, intact floodplains, or that contain a mixture of local and regional groundwater and stream water can also provide salmon the opportunity to seek out waters that are neither too cold in the winter nor too warm in the summer. Finally, young salmon must be able to migrate downstream to the ocean without being blocked by dams or culverts or entrained in diversions, and returning adult salmon must also be able to travel unimpeded to their spawning grounds.

Each female salmon can produce several thousands of eggs during her one chance at spawning. Because of this, salmon can support high levels of non-industrial harvest (i.e., nearshore and in-river; commercial, sport, and subsistence) when freshwater and ocean conditions are good. They can also rebound from population declines when the climate is favorable and freshwater habitats have not been impacted by damming, logging, or floodplain development. Healthy salmon populations seem to do best when they are subject to an intermediate level of harvest. Too little harvest can allow too many adults on the spawning grounds, which can reduce egg survival, and when too many juvenile salmon are produced they can compete with each other for food. When harvest is too high, freshwater habitats suffer from the lack of gravel cleaning by spawning salmon and a loss of nutrients that are brought in from the ocean to spawning grounds by adult salmon. Very small populations can also lose genetic diversity, which can jeopardize their ability to evolve in response to external stressors such as climate warming. These ecological concepts align well with an ethic that is shared among many Indigenous peoples in Alaska, which is to harvest salmon when they make themselves available, take only what is needed, and to leave salmon alone when they are not doing well.

The ocean is where salmon spend most of their lives, and where they put on 99% of their body weight. However, this is a challenging place to study, and we only have a broad understanding of this phase of their life cycle. Upon entering the ocean, salmon generally move in a counterclockwise direction around the North Pacific, cycling between productive summer habitat in the Bering Sea and ice-free waters of the Gulf of Alaska during the winter. Salmon feed on a variety of prey, including zooplankton (e.g., copepods, krill, crab larvae) when small and moving up to squid and forage fishes, such as herring, as they grow. Salmon growth and survival is the result of complex and poorly understood interactions among climate, food, predators, and competitors. Large-scale climate variation, such as shifts in the Aleutian Low Pressure system, which controls the oceanic currents, ocean temperatures, and weather patterns that set up food production in the ocean - phytoplankton and zooplankton - and oscillations between conditions favorable and unfavorable for salmon can be seen throughout Alaska's history (e.g., the 1976/77 "regime shift" that was a boon for Alaska salmon fisheries). The number of hatchery salmon released into the ocean is also at an all-time high, likely leading to reduced food for wild salmon in certain places and times.

In recent years, we have seen the emergence of extreme events such as drought, short term spikes in river temperatures, and marine heatwaves, which manifest as dramatic and persistent increases in temperatures across broad regions of the North Pacific. Marine heat waves profoundly alter marine ecosystems, including affecting what and how much food is available for salmon and the kinds of predators they face, and they are accompanied by hot weather over land as well, leading to river temperatures high enough to weaken or kill migrating salmon even as far north as the Yukon River. Extreme events such as heat waves complicate our ability to maintain optimal levels of salmon harvest, because the methods for setting harvest are usually developed against some average background level of natural mortality and are not well-equipped to deal with big shifts in natural mortality.

Following these scientific observations, we acknowledge that Western science is not the only knowledge source useful for understanding the complex relationships between salmon, their environments, their life-cycle needs, climate, and other factors. Indigenous and traditional knowledge often goes beyond that which is directly related to ecological aspects of the natural world and includes values associated with the entire world view, such as relationship, responsibility, reciprocity, and redistribution (4 Rs). Empirical observation is critical but in ways that focus on and teach appropriate human action as an integral part of the natural world. For example, beliefs about reciprocal relationships of care between humans and fish teach culturally appropriate behavior around concepts of salmon return and conservation.

With regards to existing knowledge about salmon, Indigenous and traditional knowledge have existed, been passed down, and been built upon for thousands of years.



"Growing up in the village, we lived from the land, river and sea. While engaging in subsistence activities, passed and taught from generation to generation, we continue to do so with great respect to the environment we live in, which can be unforgiving if not taught to survive in it. They taught us how to travel on different land, river, weather and sea ice conditions which can change in a heartbeat.

All that they taught us is woven into the fabric of our culture, to be able to survive and perpetuate the life of our culture. We were taught how to relate to the environment we were born to, as well as relating to other persons within our culture. For example, the great respect we have for our parents, aunts and uncles. To share our hunting catches with widows and those that need help. Translating the nuances of our culture and life is a challenge at times, from one language to another. I think that Indigenous knowledge better conveys all those things passed on that are deep at the core of the subsistence lifestyle we live. The permanency and perpetuity of our culture. Closely too is traditional knowledge, that conveys the timelessness of cultural traditions and subsistence practices. To continue, unbroken, with our subsistence way of life." — Oscar Evon, Tribal member of the Native Village of Kwigillingok



This knowledge can appear in many forms and from multiple cultural traditions and is commonly derived from keen, longterm observation of and interaction with local landscapes. Ultimately, we need an intertwined and holistic approach to understanding Alaska salmon, including relationships between Alaska salmon and all living and nonliving things, that includes observations from multiple knowledge types and from a variety of experiences from scientific research to generational observation and harvest.

"...'Traditional Knowledge,' it includes both Western and Indigenous knowledge...[there] are non-natives and have a lot of Traditional Knowledge regarding fish and game. So, I personally feel that using the term "Traditional Knowledge" is an inclusive term which is what this Task Force was required to use. Incorporating Indigenous Traditional Knowledge would benefit knowledge from the locals throughout the state also using Traditional Knowledge from local fisherman regardless of ethnicity." — Jacob Ivanoff, Tribal member of the Village of Unalakleet As such, Indigenous and Traditional Knowledge is included in this report to the best of our ability. Information sources include a few ethnographic interviews, conducted as part of the Task Force's work, public testimony, and literature sources focused on the social science of traditional knowledge (Table 1; Appendix 3). In addition, information and recommendations provided by the Arctic-Yukon-Kuskokwim Working Group, which included local members from each of the Arctic, Yukon, and Kuskokwim regions of Alaska, provided another opportunity to include traditional knowledge which will be presented later in this report.

"I believe in traditional knowledge. I wish to learn more on salmon in today in the ocean and in our Kuskokwim River ... If you stay here long enough, you'll learn ... in another village, they'll tell you more. [We need to] integrate Traditional Knowledge and talk to more [people] that are not fluent in English. Talk to ones that have knowledge with their language ... They use words to describe fish and their situation going on" — Adolph Lupie, Tuntutuliak

Table 1. Examples of Indigenous and Traditional Knowledge captured through the Task Force process. Context to these and all Indigenous and Traditional Knowledge captured is provided in Appendix 3. Examples provided below are a select few for illustrative purposes.

Source	Knowledge
Interview	15:12 Adolph Lupie – "There will be a lot of mosquitoes before the salmon come in."
Interview	15:26 Adolph Lupie – "Mountainsthere'll be lots of snow and they'll be floodingmore water means more fish coming."
Interview	17:55 Adolph Lupie – "salmon will come in really fast when we have a good stormy weather from the southit's bringing them in from the ocean."
Interview	25:44 Adolph Lupie – Explaining Traditional Knowledge to Westerners with an example – "My dad used towhen he's carpenteringhe used a stick to measure and eyeball. I told himwe could use a measuring tapebut he said that the stick measure is more accuratewhen you're doing carpentry. Experiencing with it for long time you get used to it and you can eyeball how good to cut it."
Public Testimony Day 1	2:45:40 James Nicori – "Predicting the salmonstarting winterThey check the thickness of the ice and how much snow we have on the hillsIn the springtime they really pay attention to birds that are coming and when the birds are plentiful they are happy to have salmon this summer."
Public Testimony Day 1	2:47:00 James Nicori – "Observe the storm coming inwhen we have strong west wind it pushes some of the Yukon fish into the Kuskokwim and if we have really south winds it pushes somethat are supposed to be going into the Kuskokwim into the Yukon."

Productivity Trends

North Pacific

The North Pacific Anadromous Fish Commission (NPAFC) collates Pacific salmon commercial catch data and the number of hatchery salmon released into the North Pacific Ocean each year. These data come from Canada, Japan, Korea, Russia and the United States, where Alaska salmon harvest is separated from Washington, Oregon and California. [Note that catch numbers are an imperfect metric of salmon abundance, because they also depend on fishing effort.] What is noticeable within this near 100-year time series is the number of Pacific salmon harvested each year by all five nations is at historically high levels since the 1990s

(Figure 1). Peak commercial catches of 600 million Pacific salmon by all five nations occurred several times during the mid-2000s to present. Hatchery salmon releases began during the 1950s, but the numbers of salmon released into the North Pacific Ocean increased during the 1970s and has peaked at around 5 billion salmon each year from 1987 to present (Figure 2). Overall, Japan, the United States, and Russia release the highest number of hatchery salmon into the ocean each year when compared with Canada and Korea.



Commerical Catch of Pacific Salmon (numbers)

Figure 1. Commercial catch (millions of fish; 1925 to 2022) of Pacific salmon within nearshore and rivers for Canada (light yellow), Japan (orange), Korea (light blue), Russia (grey), Alaska (green), and the Washington, Oregon, California (WA/OR/CA, dark blue).





Hatchery Release of Pacific Salmon (numbers)

Figure 2. Hatchery releases of Pacific salmon (billions of fish) by Washington, Oregon, California (WOCI; dark blue), Canada (yellow), Alaska (green), Russia (light blue), Japan (orange), and Korea (grey).

State of Alaska

The Alaska Department of Fish and Game manages commercial salmon harvest within four regions of Alaska including Southeast, Central, Westward, and Arctic-Yukon-Kuskokwim Regions (Figure 3). The time series of salmon harvest by species and region are shown in Figures (4-8). The time period shown (1959–2022) illustrates the variability in harvest since statehood and includes the higher production period starting in 1976/77, the onset of increased releases of salmon from hatcheries, and recent declines in productivity for some species and stocks. Chinook salmon commercial harvest averaged around 600 thousand from the late 1950s to mid-1970s, then increased to roughly 800 thousand during the early 1980s and has since gradually declined to around 250 thousand. In general, the downward trend in Chinook salmon commercial harvest since the mid-1980s includes dramatic declines within the AYK, Central and Southeast regions and high variability in harvest within the Westward region starting around 2007. Chum salmon commercial harvest was around 5 million from the late 1950s to 1980 but then increased to around 18 million through 2018. The



Figure 3. Alaska Department of Fish and Game commercial salmon regions including the Southeast (green), Central (red), Westward (yellow) and Arctic Yukon Kuskokwim (orange) regions.

commercial harvest of chum salmon has recently declined to levels seen during the mid-1980s, and subsistence harvest to lowest levels on record, with the AYK region having the largest decline. Sockeye salmon commercial harvest in Alaska has varied between 10 to 60 million. Commercial harvest of sockeye salmon has been strong during recent years with proportionally higher harvest coming from Bristol Bay in the Central Region. There is also large regional variability in sockeye salmon harvest with recent declines in some regions and record harvests in others.

Pink salmon are characterized by considerable variability and commercial harvest has varied between 50 to 210 million annually since the mid-1980s. The lowest catches occurred during 1959 and 1978 and the highest catches occurred during 2013 and 2015. Much of the pink salmon harvest occurs within the Gulf of Alaska regions including Southeast Alaska, Prince William Sound and Kodiak. Harvest within the Bering Sea is considerably lower in comparison. Pink salmon are caught in sport and subsistence fisheries, but those numbers are small compared to commercial harvest. Coho salmon commercial harvest has ranged between 2.5 to nearly 10 million, following the 1976/77 shift with the peak commercial harvest occurring during the early 1990s and the lowest commercial harvest occurring during 2020. Much of the recent decline in coho salmon harvest is within the Southeast and AYK regions.



Figure 4. Number (thousands) of Chinook salmon harvested in Alaska (1959–2022). Series include total commercial harvest (main panel), commercial harvest for ADF&G Commercial Fisheries Regions (lower panels), and sport and subsistence harvest through 2021 (side panels). Note change in scale of y-axis. Data source: ADF&G, adapted from NPAFC (2023).



Figure 5. Number (millions) of chum salmon harvested in Alaska (1959–2022). Series include total commercial harvest (main panel), commercial harvest for ADF&G Commercial Fisheries Regions (lower panels), and sport and subsistence harvest through 2021 (side panels). Note change in scale of y-axis. Data source: ADF&G, adapted from NPAFC (2023).



Figure 6. Number (millions) of sockeye salmon harvested in Alaska (1959–2022). Series include total commercial harvest (main panel), commercial harvest for ADF&G Commercial Fisheries Regions (lower panels), and sport and subsistence harvest through 2021 (side panels). Note change in scale of y-axis. Data source: ADF&G, adapted from NPAFC (2023).



Figure 7. Number (millions) of pink salmon harvested in Alaska (1959-2022) as reported annually to NPAFC. Series include total commercial harvest (main panel), commercial harvest for ADF&G Commercial Fisheries Regions (lower panels), and sport and subsistence harvest through 2021 (side panels). Note change in scale of y-axis. Data source: ADF&G, adapted from NPAFC (2023).



Figure 8. Number (millions) of coho salmon harvested in Alaska (1959-2022). Series include total commercial harvest (main panel), commercial harvest for ADF&G Commercial Fisheries Regions (lower panels), and sport and subsistence harvest through 2021 (side panels). Note change in scale of y-axis. Data source: ADF&G, adapted from NPAFC (2023).

Arctic Yukon Kuskokwim Working Group Report

Executive Summary

- **1.** The most pronounced declines of chum salmon and Chinook salmon in Alaska have occurred in the Arctic-Yukon-Kuskokwim (AYK) region, a vast and remote area dominated by the Yukon and Kuskokwim rivers, and including habitat throughout Norton Sound and into the western Arctic. Communities throughout this region have been intimately dependent on salmon for subsistence and culture for millennia and are currently suffering immense hardship due to restrictions on fishing intended to protect dwindling AYK salmon populations. The AYK Working Group (WG) of the Alaska Salmon Research Task Force (Task Force) included 42 volunteer members (no volunteers were excluded) representing a wide variety of knowledge holders, from salmon harvesters and processors to agency and academic scientists, with extensive experience with salmon in this region. The goal of the WG activities was to develop a prioritized list of research needs for understanding the causes of recent declines in AYK chum salmon and Chinook salmon populations.
- **2.** The AYK-WG held virtual meetings twice-monthly in the autumn of 2023 to develop a process for assembling a list of potential concerns contributing to recent declines in AYK salmon and for translating these into a set of prioritized research themes. The WG adopted the framework developed by the Task Force that organized potential research themes around the life cycles of salmon. A variety of criteria were used by WG members to establish the prioritization, including whether the research could provide new insights in the short-term and whether the knowledge derived from this research had the potential to be actionable in fisheries management. There was not a consensus across diverse WG members regarding the most likely causes of salmon declines, and priorities described here represent research that a majority of WG members felt would help support resilient salmon populations.
- **3.** The two top priority research themes identified by the WG were to **better understand impacts of marine harvest on AYK salmon** and **changes in the quantity and quality of marine food for AYK salmon**. For marine harvest, emphasis was placed on improving: 1) the sampling of marine fisheries (state, federal, and foreign) for incidental harvest of salmon, 2) methods for identifying the stock of origin of chum and Chinook salmon caught in these fisheries, and 3) the escapement monitoring needed to quantify the consequences for salmon populations

throughout the AYK region. Research to improve understanding of changes in the quantity and quality of food for AYK salmon in marine environments included understanding climate-related changes to salmon food resources, as well as the impacts of hatchery-origin pink salmon and chum salmon, and high abundance sockeye salmon from Bristol Bay, on feeding, growth, and survival of wild AYK chum salmon and Chinook salmon.

- **4.** Additional top priority research themes included understanding changes in the health of migrating and spawning adult salmon and how climate change is affecting freshwater and marine ecosystems. The health of migrating and spawning adults theme included particular emphasis on understanding the interacting effects of reduced body size and physiological condition, changes in disease prevalence and parasite loads, and changes in the hydrology and water temperatures experienced by adult fish in freshwater habitats. These interacting factors are expected to affect both the survival of fish during their spawning migrations and their fitness once they have reached spawning grounds, yet these effects on population dynamics of AYK salmon are currently not clear. The climate change theme emphasized interactions between changing physical features of freshwater and marine ecosystems (e.g., hydrology, water temperature) and nearly all other themes described in this report. Additional research topics that were also highlighted included understanding how climate change affects the incidence of harmful algae blooms, changes in sea ice, and melting permafrost - all of which could have important impacts on AYK salmon ecosystems.
- 5. The three lowest priority research themes were: 1) changing freshwater conditions (beyond effects on spawner health), 2) historical freshwater harvest, and 3) marine and freshwater predators. While considered important by many WG participants, these were not considered as high a priority as the themes mentioned above. Within these research themes were topics such as estimating the effects of increases in freshwater predators because of reduced harvest on these species, changes in flow regimes affecting egg incubation and juvenile rearing, and increased predation by expanding marine mammal populations.
- **6.** The WG also identified critical activities to improve coordination of research across AYK and to develop more equitable opportunities for people of this region to have more meaningful engagement in the fishery science,

management, and regulatory processes. In particular, the WG was united in its support to develop formal approaches to integrate Indigenous and western ways-of-knowing in the management process. There was also widespread emphasis on research that explored the efficacy of alternative management approaches for achieving sustainability, given the inevitable uncertainties in data and understanding. Other emphases for improving research coordination across the AYK included developing robust and publicly accessible databases that could incorporate data collected by communities, agencies, and academic institutions. Last, the WG believes that more emphasis should be placed on synthetic research approaches, such as life-cycle modeling, that would provide the platform for better synthesis of research that is widely spread across geography, time, and life stages of AYK salmon.

Overview of the charge to the Working Group as Part of Act

Congress passed the Alaska Salmon Research Task Force Act during December 2022 to form an Alaska Salmon Research Task Force (Task Force) to characterize trends in the productivity and abundance of Pacific salmon in Alaska, identify and prioritize research needs with respect to understanding increased variability or decline in Pacific salmon returns to Alaska, and to establish a coordinated research strategy to address salmon returns that are in decline or experiencing increased variability. One requirement within the Act was for the Task Force to establish a work group (by July 2023) focused specifically on the research needs associated with salmon returns in the Arctic-Yukon-Kuskokwim (AYK) regions of western Alaska.

Overview of the AYK region and its salmon

The Arctic-Yukon-Kuskokwim (AYK) region of western Alaska is located north of Bristol Bay and is dominated by the watersheds of the Kuskokwim and Yukon rivers which drain into the Eastern Bering Sea. The region also includes smaller rivers draining to Norton Sound and Kotzebue Sound, and extends into the Alaska Arctic where rivers drain to the Chukchi and Beaufort seas. The headwaters of the Yukon River extend into Canada where approximately 40% of the watershed is located (Figure 1).

Five species of anadromous Pacific salmon spawn in AYK rivers, though the most important species for fisheries and the cultures of the people living in the region are chum salmon and Chinook salmon. In the Alaska Arctic, north of Kotzebue Sound, salmon are not historically the preferred subsistence fish (although attitudes may be in transition with expanding salmon distributions); hence, research on salmon in this vast area remains low.

The AYK region includes the traditional homelands of several Alaska Native groups, with Inupiat and Yup'ik people typically living in coastal regions, and Athabascan people living in upriver regions for millennia (Langdon 2002, Wolfe and Spaeder 2009). The cultures, economies and nutritional foundation of tribes throughout the AYK region are intimately woven with salmon, particularly chum salmon and Chinook salmon. Sockeye salmon, pink salmon, and coho salmon are also harvested, along with a variety of freshwater resident species such as northern pike, burbot, and whitefish.

Fisheries in the AYK region are primarily subsistence fisheries. The region has supported in-river commercial fisheries for Chinook salmon and chum salmon over the last century, and some sport fisheries exist as well. Due to severe salmon population declines over the last two decades, there have been increasing restrictions on commercial, sport, and subsistence fisheries to protect spawning populations of these species in all rivers. Chum salmon populations in the AYK region showed severely depressed populations in 1999-2001, and again in 2020 - 2023. Chinook salmon populations in the region have shown steady declines since the early 2000s. Closed and restricted salmon fishing infringes on the opportunity for rural residents to maintain a subsistence way of life (Alaska National Interest Lands Conservation Act [ANILCA] of 1980, 16 U.S.C. § 3101-3233) and opportunity to pass down subsistence culture to younger generations.

The vast watersheds of rivers that drain the AYK region remain largely undeveloped and remote, which presents serious impediments to western science approaches to understanding the ecology of the watersheds and their salmon. Management of fishery resources is also hampered by these geographical challenges because of the inevitably high levels of uncertainty in stock assessments of fish populations, and the widely dispersed nature of fishing which makes it difficult to monitor.

The cultures, economies, and food security of people who live throughout the AYK region are being seriously impacted by declining salmon returns and the associated restrictions on fisheries. Climate change is also producing new challenges for people living in the AYK region. These changes are affecting the cultures and sustainability of AYK communities that primarily lead salmon-centered subsistence lifestyles. The AYK Working Group emphasizes the urgency of the need to take action to maintain vibrant and sustainable communities that are robust to the inevitable changes in the salmon resources that form the foundation of these communities.



Figure 1. Map of Alaska with the major watersheds and communities (black dots). The Arctic-Yukon- Kuskokwim region is located north of Bristol Bay north through Norton and Kotzebue Sounds into the Alaska Arctic. Watershed data available from the United States Geological Survey (https://www.sciencebase.gov/catalog/item/5a1632bae4b09fc93dd1721f). Community locations provided by the U.S. Census Bureau.

Background

The Alaska Salmon Research Task Force Act identified the need to convene an AYK Working Group (hereafter AYK WG), to address the research needs related to salmon declines in the Arctic-Yukon-Kuskokwim region. Fifteen members of the Task Force who had knowledge and expertise for this region volunteered to serve on this AYK WG. However, it was acknowledged that the information needs and expertise for the AYK WG should be far broader than those represented by Task Force members alone. Nominations for public members of the working group (non-task force members) were solicited in July 2023. All nominated public members who agreed to serve on the AYK WG were accepted as working group members. In total, 42 individuals agreed to serve on the AYK WG, including 15 Task Force members and 27 public members (Appendix 1). The Task Force appointed Katie Howard (Alaska Department of Fish & Game) to lead the AYK WG on behalf of the task force, and Daniel Schindler (University of Washington) was elected by public members of the AYK WG to co-chair the working group.

A strength of the convened AYK WG was the diversity of perspectives represented by the 42 members (Appendix #). These members are knowledge holders from Kuskokwim Bay, Kuskokwim River, lower, middle and upper parts of the Yukon River, Norton Sound, North Slope, academia, federal and state management agencies, environmental and fisheries non-profits, tribal organizations, inter-tribal fish commissions, and the commercial fishing industry.

Many AYK WG members repeatedly expressed the value and need for voices at the table to equitably include western science and local and Indigenous knowledge holder insights and expertise. Due to the short timelines stipulated for this endeavor, the large geographic scope of the AYK region, and limited resources to support in-person communications, the work and collaboration among working group members was heavily reliant on technology and digital forms of communications (e.g., video conferences, Excel and Word documents through cloud sharing, and email). It should be acknowledged that technology-heavy communications create their own inequities, particularly for a region where internet access can be limited and computer-based information sharing may be somewhat foreign to some members, particularly elders. This was a significant challenge for the AYK WG and it is recommended that, if another group is similarly convened in the future, resources and planning be used to allow for in-person engagement as a more equitable means of communication and collaboration among knowledge holders.

Working Group Process

The AYK WG met by virtual teleconference about every 2 weeks from September through December 2023 to discuss potential explanations for AYK salmon declines, other research needs in the region, and AYK-specific research priorities. Initial meetings were an open discussion for WG members to express ideas and concerns, which guided the co-chairs in developing structures and agendas for subsequent meetings.

The AYK WG adopted the conceptual framework developed by the Task Force for the possible explanations of AYK salmon decline for the following themes: 1) spawner health; 2) freshwater harvest; 3) freshwater predators; 4) marine predators; 5) freshwater conditions for eggs and juvenile rearing and migration; 6) marine food limitation; 7) climate change; and 8) marine harvest (focused primarily on bycatch in federal fisheries, harvest and interceptions in state-managed salmon fisheries, and illegal, unreported and unregulated foreign fisheries). It was clear from initial scoping meetings that AYK WG members wished to express concerns and suggest research priorities that were not strictly evaluating reasons for AYK salmon declines, so additional categories and discussion topics were created, and research priorities were not confined to evidence for salmon declines.

Shared spreadsheets allowed AYK WG members to formalize questions and hypotheses they had within each of these different research themes. AYK WG members who found access to the shared spreadsheets challenging were encouraged to reach out to the co-chairs to ensure their input was captured on the spreadsheets by working with other AYK WG members. AYK WG members with better technology access and comfort were encouraged to work with those who found these forms of communication challenging, so that as many perspectives as possible were represented.

After discussing research questions and hypotheses that fell under each of the research themes specified by the Task Force, the AYK WG discussed how best to prioritize these research themes. The AYK WG agreed that each member should have the independence to assess each research priority based on their own knowledge base and criteria. Examples of criteria that AYK WG members cited in their assessments were: a) whether reducing a specific scientific uncertainty would lead to actionable management changes, b) whether reducing a scientific uncertainty was a short-term or long-term goal, c) whether research on a specific topic would improve synthesis of AYK salmon ecology, d) whether research on a specific topic would improve community engagement and knowledge sharing, e) would research lead to more holistic science such as what Tribes currently use rather than the typically narrow focus of western science, f) would research on a topic produce knowledge that was immediately applicable, g) whether research would provide immediate information with high potential benefit, h) whether the research would benefit management of salmon ecosystems in the Arctic region of the AYK, and i) whether the research would produce knowledge that would be applicable beyond the AYK region.

The primary goal of the AYK WG was to prioritize research needs where research could fill knowledge gaps in understanding recent AYK salmon declines. To establish these priorities, members of the AYK WG were presented with the list of research questions and hypotheses developed by the group (Appendix 2). As a way for individuals to express their own perspectives about how to prioritize potential research, each AYK WG member was asked to assign a total of 20 points across all potential hypotheses or questions, with the constraint that the maximum score they could assign to an individual question or hypothesis was 10. We received scores from the majority of AYK WG members (29 of 42) and these scores were summed across individual AYK WG members and across hypotheses and questions to provide weights to each of the eight Task Force research themes. We also summarized scores by tallying the number of AYK WG members who assigned a score of at least 1 to any of the questions or hypotheses within each of the eight Task Force research themes. The intention of this exercise was not to treat individual hypotheses or questions as competing alternative explanations for the recent AYK salmon declines. Rather, it was a process for summarizing the variety of perspectives on what were the highest priority research themes across the entire spectrum of beliefs held by members of the AYK WG. Last, we ranked individual questions and hypotheses based on the total number of points assigned to them by all the AYK WG members who participated in the survey. All hypotheses and questions and their associated scores are provided in Appendix 2. Here we highlight the top nine as these attracted the majority of attention from across the AYK WG.

Subsequent to AYK WG discussions, co-chairs drafted a report on behalf of the WG, with a first draft provided to the AYK WG for review by March 11, 2024. Co-chairs revised the report based on AYK WG member feedback and delivered a final report to the Task Force on March 21, 2024.

Overview of AYK WG general priorities for future research needs on AYK salmon

Results from the survey of AYK WG members about priorities for future research provided one mechanism for summarizing across the range of perspectives within the AYK WG. AYK WG members developed a set of questions that could be explored by future research to support resilient AYK

chum salmon and Chinook salmon populations. The two specific questions that received the highest scores across AYK WG members both deal with the availability and quality of marine food for AYK salmon (Figure 2, purple bars). The top specific hypothesis focused on competition between AYK salmon and hatchery pink and chum salmon, and with high abundances of sockeye salmon from Bristol Bay. The second most popular question was concerned with whether changes in climate and marine food webs may be limiting growth and survival of AYK salmon.

Four of the top nine questions dealt with bycatch and interceptions of AYK fish in federal, state, and foreign fisheries (Figure 2, blue bars). Three of these hypotheses focused on understanding the biological impacts of these marine harvests on AYK salmon populations. One of these questions was concerned with increasing the analytical resolution to distinguish the stock of origin of AYK fish captured in marine fisheries.

Climate change, particularly as an interacting stressor on all other drivers of change in AYK salmon populations, was one of the nine top questions discussed by the AYK WG (Figure 2, yellow bar). Two of the top nine specific hypotheses and questions focused on the implications of changes in spawner health and quality for AYK salmon populations (Figure 2, red bars). All other specific questions received less than 20 points from the scoring exercise performed by the AYK WG (Appendix 2).



Figure 2. Total assigned scores to the top nine individual research questions considered by the AYK WG, colored by the associated research theme. All other questions or hypotheses received less than 20 total points. Hypotheses and questions are colored according to: 1) Marine Food Limitation theme (purple), 2) Marine Harvest theme (blue), 3) Climate Change theme (yellow), and 4) Spawner Health theme (red).

When summing AYK WG scores across all questions within each research theme, marine harvest, marine food limitation, climate change and spawner health were the top research priorities (Figure 3, black bars). The highest priority research theme was to better quantify the impacts of marine harvest on AYK chum salmon and Chinook salmon stocks (Figure 3, black bars). The second highest research priority identified by this approach was to understand the consequences of changes in marine food limitation for AYK salmon growth and survival. The third highest priority research theme was to understand the effects of a variety of stressors affecting the health and quality of migrating and spawning adult salmon and how these translate into population dynamics. Understanding climate change, particularly through interactions with other stressors of AYK salmon, was the fourth highest priority research theme identified by the AYK WG scoring exercise. We note, however, that there were elements

of climate change in research priorities that were included under other themes. The remaining four research themes all received some level of support for prioritization in the following descending order: freshwater conditions that affect egg incubation and juvenile growth and survival, freshwater harvest, marine predators, and freshwater predators. These last four categories received only about 22% of the total weight assigned by the AYK WG to the eight research themes. The top four research themes collectively received 78% of the total weight of scores from the AYK WG (Figure 3).

As a complementary way to summarize research priorities across the diverse members of the AYK WG, we also tallied the number of AYK WG members who assigned any degree of weight to each of the Task Force research themes (Figure 3, gray bars). This method of summarizing perspectives from across the AYK WG generally reinforced the priority

list established by the weighted scoring method, except for two notable differences. First, marine harvest and marine food limitation were ranked equally as the two top research priorities. Spawner health and climate change were the next two priorities and their order in the rankings did not change. Of the remaining four research themes, marine predators received more than twice as much weight via this second method of ranking themes compared to the first method. However, the combined weight of the lowest priority four research themes was still only about 32% compared to 68% for the four top research themes (Figure 3). In the sections below we discuss each of these general research themes and the distribution of support for individual questions or hypotheses that were aggregated under each theme.



Figure 3. Summary of the AYK Working Group research priorities organized by themes established by the Task Force. Black bars show the distribution of relative weights assigned by WG members to hypotheses or questions that were aligned within each theme. Values are relative weights that are proportional to the sums across scores assigned to individual hypotheses and questions under each research theme. Gray bars are weights that are proportional to the number of WG members who expressed any level of weight to each of the research themes.

In the section below, in decreasing order of priority by research theme, we describe the details of individual questions or hypotheses that could guide research within each of the general themes.

A) Marine Harvest

Support for research quantifying and mitigating marine harvest impacts on AYK salmon was a dominant topic throughout AYK WG discussions. Within the theme of marine harvest, there were relatively high scores prioritizing research that further quantified the number of AYK salmon harvested in state, federal, and foreign marine fisheries, and improved data and methods to compare these numbers to the abundances of AYK salmon stocks as a way to estimate the biological and social consequences of marine fishery catches. Emphasis was also placed on improving the resolution of current techniques (i.e., genetic stock identification, GSI) for estimating stock-specific impact rates of marine harvest on individual AYK salmon stocks. Overall, there was roughly equal emphasis placed on understanding interceptions in state, federal and foreign fisheries. There was recognition of the amount of effort currently being applied towards rigorously quantifying Chinook and chum salmon bycatch in federal marine fisheries, though weak stock resolution in current GSI and limited coverage of Chinook and chum salmon escapements throughout the AYK region place limits on the desired resolution of these efforts to understand their biological impacts. The scoring was similarly high for more research to understand the biological implications of both chum salmon and Chinook salmon catches in state and foreign fisheries.

B) Marine Food Limitation

Research on marine food limitation for AYK chum salmon and Chinook salmon was identified as another top priority research theme by the WG. The highest score for all individual research questions and hypotheses was in this theme. The AYK WG expressed particular support for research understanding competition between AYK salmon and increasing pink salmon, chum salmon, and sockeye salmon abundance from other regions, and especially in consideration of hatchery-produced competitors. This research theme also received relatively high scores for research to determine whether there were climate-induced changes in the quantity and quality of marine food for AYK salmon in the Bering Sea, and whether changes in nearshore habitat conditions were reducing the survival of AYK smolts upon their migration to the nearshore ocean.

C) Spawner Health

Research to understand causes and consequences of adult salmon health status while migrating and spawning was a relatively complex theme as WG members identified many dimensions of this problem that affect the survival of fish as they migrate from the ocean to spawning grounds, and their subsequent success on the spawning grounds. In particular, emphasis was placed on understanding the effects of changing climate and ocean conditions on en route and pre-spawn mortality of adult fish. Whether changes in the incidence of parasites and diseases (such as with Ichthyophonous), and nutritional health exacerbated these effects on reproductive health and stock productivity. Warming river temperatures with climate change was also identified as a poorly understood interactor with other factors that affect the health of adult fish in freshwater habitats. Other concerns within this broad theme emphasized quantification of thresholds or conditions where genetic population sizes were so low that there was increased risk of extirpation of individual sub-populations in AYK watersheds, and quantifying how widespread these thresholds were surpassed at present. Also identified as an important research activity was to systematically review existing in-river stock assessments to determine whether there was adequate precision and accuracy to quantify en route mortality in the large AYK rivers, particularly in the Yukon River where there is an international commitment to meet an escapement goal for Chinook and chum salmon that spawn in Canadian components of the watershed. Last, there was concern that little was known or acknowledged about the effects of handling fish for research purposes on their stress levels and eventual success on the spawning grounds.

D) Climate Change

The AYK WG widely emphasized that the consequence of climate change in freshwater and marine habitats was an overriding priority because climate change is likely modifying or amplifying nearly all other stressors identified in other research themes. Beyond climate change as an amplifier of other stressors, the AYK WG identified other research questions to be pursued. These include understanding linkages between climate stressors in the ocean and in freshwater habitats, whether climate-driven changes in sea ice have affected plankton phenology and potential mismatches with AYK salmon, and whether climate change has increased the incidence and intensity of harmful algal blooms that affect juvenile salmon growth and survival in the ocean. There was also interest in understanding whether the expansion of anadromous salmon into the Arctic was affecting the ecology of resident fishes in rivers that historically did not have salmon in them, though this question did not receive any support in the scoring exercise.

E) Freshwater Conditions

Changing freshwater habitat conditions that affect egg incubation success, and juvenile salmon rearing and migration conditions, rated to be of intermediate priority, focused on issues related to whether changes in watershed habitat productivity and capacity were reducing the fitness and abundance of salmon smolts leaving AYK rivers. Related to this question was whether changes in hydrology may be affecting the complexity, connectivity and geomorphology of freshwater habitats in ways that affect freshwater food webs that support juvenile salmon growth, and hydrologic effects on egg incubation conditions. Additional questions were focused on whether climate driven effects on floods, spring ice breakup, thermal regimes, and permafrost loss were contributing to a degradation of freshwater habitat for juvenile salmon. Other considerations included asking whether the expansion of beavers was altering salmon habitat in substantial ways, and whether the declines of marine-derived nutrients from declines in abundant species (e.g., pink salmon and chum salmon) were reducing the productivity of freshwater habitats.

F) Freshwater Harvest

One of the lower priority research themes was associated with current and legacy effects of harvest in freshwater fisheries. The research question that received the highest score within this theme was focused on understanding whether the current escapement goals were still valid given the observed changes in the environment and the different constraints on population productivity that are expressed in freshwater versus marine ecosystems. Other important concerns were focused on understanding whether there were legacy effects of historical freshwater fisheries, particularly commercial fisheries, on the current demographic structure of AYK salmon populations, which have shown a pronounced decline in average body size and age-at-maturity for Chinook salmon in particular. Other questions were focused on whether large fish that become entangled in but drop out from small mesh gill nets actually survive to reproduce, as is typically assumed. While it is often assumed that the consequences of managing to the higher end of an escapement goal range has the same biological consequences as managing to achieve the bottom end of an escapement goal range, several AYK WG members believed this assumption needed to be more thoroughly explored. Further, there was interest to explore the consequences for harvest on weak stocks when they mingle with dominant or highly abundant stocks within the same river, and whether there were ways to develop more stock specificity in freshwater fisheries. Last, because of the recent proliferation of early-maturing males ("jacks") in Chinook salmon, some AYK WG members believed that research should explore whether there were potential negative consequences of harvesting these small individuals from a population, though this question also received no support in the scoring exercise.

G) Marine and Freshwater Predators

The two research themes determined to be the lowest research priority by the AYK WG were associated with changes in the predation rates on salmon in marine and freshwater habitats. In marine habitats there was concern that increasing apex predators such as resident killer whales, seals, and salmon sharks may be reducing marine survival of AYK salmon and generating new evolutionary pressures on large fish that may be related to declining body size in AYK salmon, particularly Chinook salmon. In freshwater habitats, there was concern that piscivores may be increasing because of declining harvest of resident species (such as northern pike) that could be translating into increased predation rates on juvenile salmon. Other questions were focused on whether the proliferation of beavers was enhancing predation on juvenile salmon through changes in habitat that facilitate predators such as pike and juvenile coho salmon, and whether climate-induced changes in freshwater habitats were increasing vulnerability of juvenile salmon to freshwater predators.



Other research needs and priorities for improving science and management of AYK salmon

The AYK WG also identified critical activities to improve coordination of research across the AYK and to develop more equitable opportunities for people of this region to have more meaningful engagement in the fishery science, management, and regulatory processes. These concerns complement similar existing efforts to prioritize research on Chinook salmon and chum salmon in the AYK region by the Arctic-Yukon-Kuskokwim Sustainable Salmon Initiative (Schindler et al. 2013). In addition to discussing the eight research themes of the Task Force as they relate to understanding the recent declines in AYK chum salmon and Chinook salmon, the AYK WG discussed several other research needs for improving the science and management of AYK salmon. These additional themes are listed and described briefly below.

A) Knowledge integration and alternative approaches towards fishery management

The AYK WG was united in its support to develop and explore the efficacy of formal approaches to integrate traditional, Indigenous, and western ways-of-knowing in the science and management process. At present, there is little integration of alternative sources of knowledge and perspectives about how the AYK salmon ecosystems function despite a considerable amount of knowledge held by the individuals who live throughout the region. The challenge is to find ways to weave different types of knowledge and different perspectives into a coherent framework for informing management and conservation efforts. The AYK WG believes that exploring alternative ways of accomplishing this integration is itself a research priority that has the distinct potential to both improve understanding of AYK salmon and their ecosystems, and to improve integration of a variety of knowledge types to better inform the management process.

B) Need for synthetic life-cycle approaches to improve integration

As AYK salmon complete their life cycles, their biology integrates across a wide variety of habitats, from freshwater streams, lakes and wetlands, to estuaries and the coastal ocean, to the Bering Sea and Gulf of Alaska. Understanding how changes in management and the environment affect salmon populations is seriously challenged by this complexity. Most research on AYK salmon has focused on individual life stages in specific habitats which, as the AYK WG expressed, has hampered scientific progress towards developing a holistic understanding of the drivers of population dynamics. The AYK WG believes that more emphasis should be placed on synthetic research approaches, such

as life-cycle modeling, that would better integrate across geography, time, and life stages of AYK salmon. Additional emphasis would be placed on integrating life cycle modeling, field research, traditional knowledge, and management activities into a coherent framework to enhance knowledge generation and improve management outcomes.

C) Management under uncertainty

Fisheries management is an imperfect process that makes decisions despite incomplete understanding of ecosystems and fish populations, including their responses to management actions. These challenges are especially acute in the AYK region of Alaska because ecosystems are so vast, remote, and heterogeneous which hinders comprehensive monitoring needed to reduce key uncertainties in data and models used in management. People who rely on salmon fisheries also hold a wide range of values and goals for assessing the success or failure of management decisions. Thus, there was widespread support for research that evaluates trade-offs associated with various management strategies for achieving a variety of goals for stakeholders, given the inevitable uncertainties in data and understanding. Such research would use combinations of simulation modeling and community engagement to co-develop projects that explicitly quantify trade-offs associated with the potential risks and rewards of alternative management strategies given uncertainties in how ecosystems function and how they may change in the future.

D) Database coordination

A final emphasis for improving research coordination across the AYK included developing robust and publicly accessible databases that could incorporate data collected by communities, agencies, and academic institutions. This is a non-trivial task given the heterogeneity in types of data relevant to salmon populations and aquatic ecosystems throughout AYK. Research is needed to develop ways to both capture the vast amounts of historical data on AYK salmon and ecosystems, and to seamlessly add new data streams as further research is pursued. Both serious design considerations and substantial funding will be needed to accomplish this ambitious goal, but the invaluable payoff would be increased transparency in research and management, and greater leveraging of data to improve ecological understanding. Such an effort would need to proactively plan for the sustainability of such a database, and to make specific policies to acknowledge and respect data sovereignty.



References

Langdon, S.J. 2002. The Native People of Alaska. Greatland Graphics, Anchorage, Alaska.

Schindler, D., C. Krueger, P. Bisson, M. Bradford, B. Clark, J. Conitz, K. Howard, M. Jones, J. Murphy, K. Myers, M. Scheuerell, E. Volk, and J. Winton. 2013. Arctic-Yukon-Kuskokwim Chinook Salmon Research Action Plan: Evidence of Decline of Chinook Salmon Populations and Recommendations for Future Research. Prepared for the AYK Sustainable Salmon Initiative (Anchorage, AK). v + 70pp.

Wolfe and Spaeder. 2009. People and salmon of the Yukon and Kuskokwim drainages and Norton Sound in Alaska: fishery harvests, culture changes, and local knowledge systems. American Fisheries Society Symposium 70: 349-379.

See Appendices 9 – 10

Appendix 1: Task Force Act Engrossed in Senate (12/14/2022) 117th CONGRESS 2d Session

S. 3429

An Act to Establish an Alaska Salmon Research Task Force

Be it enacted by the Senate and House of Representatives of the United States of America in Congress assembled,

SECTION 1. SHORT TITLE.

This Act may be cited as the "Alaska Salmon Research Task Force Act".

SEC. 2. PURPOSES.

The purposes of this Act are—

- to ensure that Pacific salmon trends in Alaska regarding productivity and abundance are characterized and that research needs are identified;
- (2) to prioritize scientific research needs for Pacific salmon in Alaska;
- (3) to address the increased variability or decline in Pacific salmon returns in Alaska by creating a coordinated salmon research strategy; and
- (4) to support collaboration and coordination for Pacific salmon conservation efforts in Alaska.

SEC. 3. SENSE OF CONGRESS.

It is the sense of Congress that-

- (1) salmon are an essential part of Alaska's fisheries, including subsistence, commercial, and recreational uses, and there is an urgent need to better understand the freshwater and marine biology and ecology of salmon, a migratory species that crosses many borders, and for a coordinated salmon research strategy to address salmon returns that are in decline or experiencing increased variability;
- (2) salmon are an essential element for the well-being and health of Alaskans; and

(3) there is a unique relationship between people of Indigenous heritage and the salmon they rely on for subsistence and traditional and cultural practices.

SEC. 4. ALASKA SALMON RESEARCH TASK FORCE.

- (a) In General.—Not later than 90 days after the date of enactment of this Act, the Secretary of Commerce, in consultation with the Governor of Alaska, shall convene an Alaska Salmon Research Task Force (referred to in this section as the "Research Task Force") to—
- (1) review existing Pacific salmon research in Alaska;
- (2) identify applied research needed to better understand the increased variability and declining salmon returns in some regions of Alaska; and
- (3) support sustainable salmon runs in Alaska.
 - (b) Composition And Appointment.—
- IN GENERAL.—The Research Task Force shall be composed of not fewer than 13 and not more than 19 members, who shall be appointed under paragraphs (2) and (3).
- (2) APPOINTMENT BY SECRETARY.—The Secretary of Commerce shall appoint members to the Research Task Force as follows:
- (A) One representative from each of the following:

(i) The National Oceanic and Atmospheric Administration who is knowledgeable about salmon and salmon research efforts in Alaska.

(ii) The North Pacific Fishery Management Council.

(iii) The United States section of the Pacific Salmon Commission.

(B) Not less than 2 and not more than 5 representatives from each of the following categories, at least 2 of whom shall represent Alaska Natives who possess personal knowledge of, and direct experience with, subsistence uses in rural Alaska, to be appointed with due regard to differences in regional perspectives and experience:

(i) Residents of Alaska who possess personal knowledge of, and direct experience with, subsistence uses in rural Alaska.

(ii) Alaska fishing industry representatives throughout the salmon supply chain, including from—

- (I) directed commercial fishing;
- (II) recreational fishing;
- (III) charter fishing;
- (IV) seafood processors;

(V) salmon prohibited species catch (bycatch) users; or

- (VI) hatcheries.
- (C) 5 representatives who are academic experts in salmon biology, salmon ecology (marine and freshwater), salmon habitat restoration and conservation, or comprehensive marine research planning in the North Pacific.

(3) APPOINTMENT BY THE GOVERNOR OF ALASKA.—The Governor of Alaska shall appoint to the Research Task Force one representative from the State of Alaska who is knowledgeable about the State of Alaska's salmon research efforts.

(c) Duties.—

- (1) REVIEW.—The Research Task Force shall—
- (A) conduct a review of Pacific salmon science relevant to understanding salmon returns in Alaska, including an examination of—

 (i) traditional ecological knowledge of salmon populations and their ecosystems;

(ii) marine carrying capacity and density dependent constraints, including an examination of interactions with other salmon species, and with forage base in marine ecosystems;

(iii) life-cycle and stage-specific mortality;

(iv) genetic sampling and categorization of population structure within salmon species in Alaska;

 (v) methods for predicting run-timing and stock sizes;
 (v) methods for predicting run-timing and stock sizes;

((vi) oceanographic models that provide insight into stock distribution, growth, and survival;

(vii) freshwater, estuarine, and marine processes that affect survival of smolts;

(viii) climate effects on freshwater and marine habitats;

 (ix) predator/prey interactions between salmon and marine mammals or other predators; and

(x) salmon productivity trends in other regions, both domestic and international, that put Alaska salmon populations in a broader geographic context; and

(B) identify scientific research gaps in understanding the Pacific salmon life cycle in Alaska.

- (2) REPORT.—Not later than 1 year after the date the Research Task Force is convened, the Research Task Force shall submit to the Secretary of Commerce, the Committee on Commerce, Science, and Transportation of the Senate, the Committee on Environment and Public Works of the Senate, the Subcommittee on Commerce, Justice, Science, and Related Agencies of the Committee on Appropriations of the Senate, the Committee on Natural Resources of the House of Representatives, the Subcommittee on Commerce, Justice, Science, and Related Agencies of the Committee on Appropriations of the House of Representatives, and the Alaska State Legislature, and make publicly available, a report-
- (A) describing the review conducted under paragraph (1); and

(B) that includes—

(i) recommendations on filling knowledge gaps that warrant further scientific inquiry; and

(ii) findings from the reports of work groups submitted under subsection (d) (2)(C).

(d) Administrative Matters.-

(1) CHAIRPERSON AND VICE CHAIRPER-SON.—The Research Task Force shall select a Chair and Vice Chair by vote from among the members of the Research Task Force.

(2) WORK GROUPS.-

(A) IN GENERAL.—The Research Task Force—

(i) not later than 30 days after the date of the establishment of the Research Task Force, shall establish a work group focused specifically on the research needs associated with salmon returns in the AYK (Arctic-Yukon-Kuskokwim) regions of Western Alaska; and

(ii) may establish additional regionally or stock focused work groups within the Research Task Force, as members determine appropriate.

((B) COMPOSITION.—Each work group established under this subsection shall—

(i) consist of not less than 5 individuals who—

- (I) are knowledgeable about the stock or region under consideration; and
- (II) need not be members of the Research Task Force; and
- (ii) be balanced in terms of stakeholder representation, including commercial, recreational, and subsistence fisheries, as well as experts in statistical, biological, economic, social, or other scientific information as relevant to the work group's focus.
- (C) REPORTS.—Not later than 9 months after the date the Research Task Force is convened, each work group established under this subsection shall submit a report with the work group's findings to the Research Task Force.
- (3) COMPENSATION.—Each member of the Research Task Force shall serve without compensation.
- (4) ADMINISTRATIVE SUPPORT.—The Secretary of Commerce shall provide such administrative support as is necessary for the Research Task Force and its work groups to carry out their duties, which may include support for virtual or in-person participation and travel expenses.
- (e) Federal Advisory Committee Act.—The Federal Advisory Committee Act (5 U.S.C. App.) shall not apply to the Research Task Force.

SEC. 5. DEFINITION OF PACIFIC SALMON.

In this Act, the term "Pacific salmon" means salmon that originates in Alaskan waters.

Passed the Senate December 14, 2022.

Appendix 2: Task Force Timeline and Milestones

Alaska Salmon Research Task Force

During the first meeting in June 2023, the Task Force members agreed on an approach/milestone timeline for the objectives provided in the ACT (Table 1).

Table 1. Date, approach and milestones the Alaska Salmon Research Task Force used to address objectives within the Alaska Salmon Research Task Force Act.

Date	Approach/Milestones
June 28, 2023	Task Force Establishes Regional Teams (TF members; address ALL of ALASKA) to build EXISTING KNOWLEDGE and begin to discuss RESEARCH GAPS/NEEDS
July 27, 2023	Task Force Meeting; Establish the ARCTIC YUKON KUSKOKWIM WORKING GROUP
August 18, 2023	Begin to close our REVIEW OF EXISTING KNOWLEDGE; Final comments on DRAFT REPORT OUTLINE
October 2023	DRAFT document on EXISTING KNOWLEDGE – and initial list of RESEARCH GAPS/NEEDS (place on website for Public review)
November 2023	FINAL DRAFT EXISTING KNOWLEDGE; Continue to list RESEARCH GAPS and NEEDS (Public input)
April 2024	DRAFT FINAL REPORT; Begin one month Public Review
May 2024	FINAL DRAFT of REPORT
June 2024	FINAL REPORT

Figure 1 illustrates the process the AKSTRF utilized to complete the objectives and prepare a Final Report for the June 2024 deadline. First, the Task Force formed Regional Teams (Southeast, Central, Westward, and Arctic-Yukon-Kuskokwim) with the task of: 1) providing reference materials to understand salmon returns in Alaska; 2) identifying gaps in understanding the Pacific salmon life cycle in Alaska; and 3) recommendations on filling knowledge gaps that warrant further scientific inquiry. In addition, the Task Force formed an Arctic-Yukon-Kuskokwim (AYK) Working Group (WG) to focus on the research needs associated with salmon returns in the AYK regions of Western Alaska. The AYK WG merged the AYK Regional Team members (15) from the Task Force with 30 other members from the Arctic, Yukon, and Kuskokwim region (see AYK WG Report section for more details).



Figure 1. The process the Alaska Salmon Research Task Force used to complete Act objectives and the Final Report.

In consideration of the ACT objectives, the Task Force scheduled bimonthly (every other month; see Table 2) public meetings to discuss current progress and solicit feedback from the public. Public comment on these tasks was solicited throughout the process during bimonthly meetings, online through our Task Force web page and during the in person/hybrid meeting held in Anchorage,

AK on November 14 and 15, 2023. The DRAFT report was also provided on the web page for Public comment and input during mid-October to mid-November (prior to our November 14 and 15, 2023 in person hybrid meeting in Anchorage, AK) and during the month of April 2024.

Task Force Meetings

 Table 2
 Dates and focus of the Alaska Salmon Research Task Force meetings open to the public.

Date	Format and Primary Focus
7/27/2023	Virtual. Establish the ARCTIC YUKON KUSKOKWIM WORKING GROUP
9/19/23	Virtual. Discuss report outline and progress toward existing knowledge and research gaps
11/14-15/23	Hybrid in Anchorage, AK. Existing knowledge and gaps, Research needs and Public comment/testimony
1/25/24	Virtual. Report on progress toward DRAFT Report
3/27/24	Virtual. Report on DRAFT FINAL REPORT
5/22/24	Virtual. Comment on DRAFT FINAL REPORT and incorporation of public comments

Appendix 3: References for Existing Knowledge

Indigenous Knowledge/Traditional Knowledge

This knowledge was captured through part of the Task Force process, which included interviews, public testimony, and literature review focused on the social science of traditional knowledge. Specifically, efforts were made to conduct interviews from each region of the AYK, with initial outreach through inter-Tribal organizations within each region (Kawerak, KRITFC, and Tanana Chiefs Conference). Those that were contacted either declined, accepted, referred to another individual, or preferred to provide public testimony in lieu of an interview. Referred individuals were reached out to as well and, if willing and able, were captured through the Task Force process.

What is specifically documented in this appendix is knowledge as it relates to each potential impact (Climate Warming and extreme events, Predation, Freshwater habitat change, Freshwater harvest) of Alaska salmon production identified. In addition, powerful quotes from recent events are included to provide more impact and speak to the human element of those that rely on and are in relationship with salmon. Lastly, research priorities listed are meant to supplement what is already provided by the AYK WG.

Potential Impact	Indigenous & Traditional Knowledge	Potential Impact	Indigenous & Traditional Knowledge
Climate Warming	29:30 Adolph Lupie – "Climate changewe are experiencing it and handling itintegrating the science and Traditional Knowledge."	Climate Warming	Public Testimony Day 1 – 4:08:20 Vanessa von Biela – "…we're learning very quickly…that several of these salmon stocks are just becoming far less productive…with changes to climate."
Climate Warming Godduhn et al. 2020 – Another fisher shared similar sentiments regarding the effects of climate change on the fishery: "You have climate change, the biology of the water, the salmon ecosystem is changing, the acidity is increasing, the tempera- ture is increasing which changes their food that they eat and have available."		Predation	Mikow et al. 2019 – Some respondents described how the sloughs and tributaries of the Kuskokwim River are constantly changing. A beaver dam or sandbar could block a free-flowing slough, creating a dead end slough: "And those are full of [salmon] carcasses. And then the predators are there: the
Climate Godduhn et al. 2020 – One of the more themes among responses pertained to t	Godduhn et al. 2020 – One of the more common themes among responses pertained to the effects		crazy in those areas"
of climate change. Many respondents cited late freeze-ups, early breakups, and an overall lack of snow and ice for the past several years.		Predation	Public Testimony Day 1 – 3:32:18 Virgil Umphenour – "On my hunting operation where I guide out of Husliagraylingwould be digging up the salmon
Climate	Public Testimony Day 2 – 2:28:30 James Nicori		eggsand grayling ate (them)."
warming	 "One summer we hadtemperatures in the low 90s, upper 80s a couple weeks. They were starting to have fish floating down the river haven't reached spawning grounds." 	Freshwater Habitat Change	Mikow et al. 2019 – Key respondents also de- scribed their perceptions that sport fishers inter- fere with salmon spawning and contribute to the physical disturbance of Chinook salmon spawning
Climate Warming Public Testimony Day 1 – 2:52:25 James Nicori – "Temperature was in the 90s three or four daysOur river was too warm for those salmon to survive salmons were being overheated and they floated down the river"			habitat by walking in streams and on riverbanks.

Potential Impact	Indigenous & Traditional Knowledge	Potential Impact	Indigenous & Traditional Knowledge
Freshwater Habitat ChangePublic Testimony Day 2 – 2:26:30 James Nicori – " the beavers, they dam the river where the spawners can't even go through the damseeing lots of beaver dams. There used to be no beavers in our area (Kuskokwim River)migrating into the lower streamsdown to the coast now."		Freshwater Harvest	McDevitt & Koster 2021 – The most common gear types for harvesting salmon include drift gillnets, set gillnets, fish wheels, and rod and reel. Although both set and drift gillnets are used drainage-wide, disparate physical characteristics between the three regions of the river typically demand different gear types in each region
Freshwater Habitat Change	Public Testimony Day 2 – 2:35:50 Jacob Ivanoff – "A chemical sprayed on tires actually kills…driving on roads…can affect the salmon."	Freshwater Harvest	Godduhn et al. 2020 – These (Kuskokwim River) communities are all highly subsistence-dependent with strong traditions surrounding the use of
Freshwater Harvest	13:25 Adolph Lupie – "We have to respect them and keep what you do catch very clean and handle them carefullyThe animal, fish, or something we	Harvest	salmon. Godduhn et al. 2020 –harvest of wild foods is
	take into our home the women they take of it right awaywith no complainingbecause the person who hunts will get more not lessif we leave it will not catch anymore while others are catching more.We have to respect them and keep what you do catch very clean and handle them carefully."	Freshwater Harvest	still a primary way of life for most residents. Godduhn et al. 2020 – Residents affirm that salmon provides not only an essential food source for families, but also supports important cultural, linguistic, and family traditions that encourage health and wellbeing within communities. Chang-
Freshwater Harvest	18:44 Adolph Lupie – "When it's too hot we don't try to fish for subsistence."	ing patterns in the harvest and use on Lupie – "When it's too hot we don't or subsistence." ing patterns in the harvest and use continue to drive disconcerting social the region, such as reducing the time	
Freshwater Harvest	water est6:59 Evon Waska – "Wood boatersAs soon as ice went out the setnet went in. King setnetthose weapons were 8.5 to 8 inches."		lies spend together at fish camps and the lack of transmission of cultural knowledge between older and younger generations.
Freshwater Harvest	7:26 Evon Waska – "…our people are into sharing. Elders first. Families first. We never kept the first king. We shared it."	Freshwater Harvest	in Godduhn et al. 2020 – Limited archeological documentation in the region suggests that Kus- kokwim River residents historically exploited
Freshwater Harvest	r 7:46 Evon Waska – "Our cultural and our self-iden- tity was giving to the Elders and sharing our catch."		consistent salmon runs that provided the most reliable element of the resource base, certainly for the last 3,000 years (Shaw 1998).
Freshwater Harvest	10:36 Evon Waska – "The moms would teach the daughters how to cut fish."	Freshwater Harvest	in Godduhn et al. 2020 – Prior to Alaska statehood in 1959, commercial fishing in the Kuskokwim Area
Freshwater Harvest	12:37 Evon Waska – "Mom and dad were boss. They would say enough of those." – meaning time to stop fishing.		was regulated by quota, and subsistence fishing was unregulated and mostly undocumented fishers either kept fish they caught for subsistence use or if a commercial buyer were available, they
Freshwater Harvest	Inwater estMcDevitt & Koster 2021 – Many respondents attribute the decline in the use of fish camps to increased restrictions on fishing opportunity and an associated increase in fishing costsFreshwater HarvestInwaterMcDevitt & Koster 2021 – Although thousands		could sell the fish (Ikuta et al. 2013). Mikow et al. 2019 – Fishers in all study communi-
Freshwater			ties explained that they maintain close commu- nication with family and friends downstream to obtain news about the arrival timing of Chinook
Harvest	of residents throughout the drainage harvest salmon each season, several factors differen- tiate one region of the river to the next. These include differences in the physical nature of the river through its course, species distribution and abundance, types of gear used by fishers, and population sizes of communities.	Freshwater Harvest	salmon. Mikow et al. 2019 – Almost all key respondents observed a decline in Chinook salmon abundance in fishing areas near their communities, and some have decided to no longer target the species as a conservation measure.

Potential Impact	Indigenous & Traditional Knowledge
Freshwater Harvest	Ikuta et al. 2013 – Salmon have been a vital source of protein and a cultural and economic resource since time immemorial.
Freshwater Harvest	Ikuta et al. 2013 – Many fishing traditions related to avoiding waste were described by research respondents. People do not catch more fish than can be processed in a timely manner, and avoid cutting in the hottest time of day, when the quality of the meat degrades.
Freshwater Harvest	Ikuta et al. 2013 – Historical methods of harvesting salmon near Kwethluk include gillnets, fish spears, fish traps, and dipnets.
Freshwater Harvest	Brown et al. 2012 – The most widely used resource category was fishwhich was also the resource most commonly harvested and the one making up the bulk of the total subsistence harvest.
Freshwater Harvest	Public Testimony Day 2 – 2:30:00 James Nicori – "In the older daysthere weren't any biologists going aroundthe fish cutters, the wiveswere fish biologiststold us no more fishing."
Freshwater Harvest	Public Testimony Day 1 – 2:40:25 James Nicori – " just tell us (take) what you can and let the rest go so we can have more to come in later years."
Freshwater Harvest	Public Testimony Day 1 – 2:40:40 James Nicori – "When the salmon comes in, fish on the first run that are comingthe first ones are mostly male and they go up to the headwaters for the females. Sowhen you have enoughthe females pass by."
Freshwater Harvest	Public Testimony Day 1 – 3:58:00 Victor Lord – "If the fishing got real low, people would pull their nets on their own."

Source	Powerful Quotes
Senate Committee on Indian Affairs	18:38 Lisa Murkowski – "We don't fish. we don't eat." & "Multiple other factors, including management structures we have to look at"
Senate Committee on Indian Affairs	31:20 Brian Ridley – "130,000 salmon, 1-3% prohibited species catch, but we know every salmon counts"
Senate Committee on Indian Affairs	37:20 Jonathan Samuelson – "serve on all coun- cils to represent leverage holistic approach, etc."
Senate Committee on Indian Affairs	44:10 Karma Ulvi – "claim that Aleutian fish- eries taking fish from western Alaska nothing new" since 1905 officially
Senate Committee on Indian Affairs	49:50 Dan Winkelman – "Not just negatively affecting our culture and wellbeing, but our good health."
Senate Committee on Indian Affairs	1:05:00 Charles Menadelook – "In my opinion, if we don't do anything we'll run out of salmon and/or marine mammals in 5-6 years."
Senate Committee on Indian Affairs	1:07:00 Charles Menadelook – "I think that the main thing we need to do is change the way the salmon is managedmanaged towards maximum commercial yield instead of other yieldThat's what's killing salmon."
Senate Committee on Indian Affairs	1 1:17:30 Brian Ridley – "I had an Elder tell me that they lived through the Great Depression but didn't even know it because they had everything they needed."
Senate Committee on Indian Affairs	1:20:00 Jonathan Samuelson – "My brother is the provider in my family. He had Covid and went out anyways to get a moose. He had no choice. The provider role in our communities is so important."
Senate Committee on Indian Affairs	1:24:30 DanWinkelman – "This area (Y-K Delta) has one of the highest rates of traditional food consumption in the entire state"
Task Force Interview	4:44 Adolph Lupie – "I'm turning 70My par- ents are gone. They told me that I'll be reaching this era when we will get no more fish on the river swimming, but we have to deal with it withscience."
Task Force Interview	10:51 Adolph Lupie – "today we are hurting and suffering, sacrificing fish."
Source	Powerful Quotes
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Task Force Interview	2:36 Evon Waska – "We came, our mom and dad, (to look) forward to the return of the salmon, and that's the way I grew up."
Task Force Interview	3:27 Evon Waska – "Most of the year it's winter in Alaska. To sustain us through the long winter months…our parents taught us how to live the way of life of how important our salmon was."
Task Force Interview	12:22 Evon Waska – "Sometimes that king salmon would be the only dinner we had."
AKSRFT Interview	14:59 Evon Waska – "I'm into commercial fishing that was a great helpwe entered the cash economy and didn't get richUnem- ployment, lack of jobs getting heating oil, billscommercial fishing ended in 2014that was their incomeGot to stop hardship."
Task Force Interview	21:20 Evon Waska – "no more commercial fishing milk is now 10, maybe 10-12 bucks, maybe 15 bucks a gallonyou have to barge 'em and freight air freightmy people are going to turn to subsistence."
Task Force	2:18:36 Brooke Woods – "the crisis we're in.
Public Testimony Nov. 15	This is a cultural crisis."
Task Force Public Testimony Nov. 14	2:31:25 Stanislaus Sheppard – "North Pacific Fishery(ies) (Management) Council member said, "Sounds like we're in a humanitar[ian] crisis."
Task Force Public Testimony Nov. 14	2:56:25 James Nicori – "We work together. We have to work together."
Task Force Public Testimony Nov. 14	3:27:40 Virgil Umphenour – "Whenever there's no salmon (I think - hard to hear - might say famine) it affects the entire ecosystem."
Source	Research Priorities based on Indigenous & Traditional Knowledge
Task Force Interview	5:28 Adolph Lupie – "I wish to learn more salmon in the ocean and our Kuskokwim River."
Task Force Interview	30:22 Adolph Lupie – In regards to salmon ecol- ogy – "How come we don't include things like photosynthesis, plants, food, light…" referring to the fact that we don't look at everything, just the fish.
Task Force Interview	40:33 Adolph Lupie – "Drop the scientific pro- tocols and integrate Traditional Knowledge."

Source	Powerful Quotes
Task Force Interview	40:53 Adolph Lupie – "Talk to more that are not fluent in English. Talk to ones that have more knowledge with their language. They have better words to describe the fish and the situation going on."
Task Force Interview	41:17 Adolph Lupie – "What I mean by (sci- entific) protocolssomeone once had to do research on not catching more fish while the other one was catching a lotcan we share that too?" – Look into how we can share fish that are collected for research purposes."
Task Force Interview	35:38 Evon Waska – "My suggestion is, why don't you tag 'em and see where they go some electronic device."
Task Force Interview	36:40 Evon Waska – * "The interceptions are reaching Western Alaska." – research into interceptions of Western Alaska salmon
Task Force Public Testimony Nov. 15	2:11:46 Dan Gillikin – "Personally would love to see nothing more than a large scale program for collecting stream temperature data."
Task Force Public Testimony Nov. 15	2:17:45 Brooke Woods – "I just don't think we're looking at the physical and mental im- pacts of no salmon to our dietIt's essential."
Task Force Public Testimony Nov. 15	2:20:40 Brooke Woods – "We need to look at the marine environment and see what we can do as far as management decisions."
Task Force Public Testimony Nov. 15	2:20:55 Brooke Woods – "'looking at the health and wellbeing of our people and what salmon means."
Task Force Public Testimony Nov. 15	2:22:50 Brooke Woods – "Pink salmon hatchery productionneeds to be addressed."
Task Force Public Testimony Nov. 15	2:32:00 Gabe Canfield – "The impacts that are happening to Alaska Pacific salmon and Alaska Pacific salmon habitat. Beyond the more well known ones some more localized human impactssuperfund sites and areas of impact- ed water qualityincreased road creations human impact(s) to critical salmon habitat should be a priorityheard from community visits across community (Yukon River) this summer."

Source	Powerful Quotes
Task Force	2:37:00 Stanislaus Sheppard – "We're all fight-
Public Testimony Nov. 14	ing for the same thing. That's salmonWe're talking about protecting the migratory routes just like birds."
Task Force	2:42:00 James Nicori – "If it is possible in any
Public Testimony Nov. 14	way, create a bufferwest coast of Alaska where the salmon travels12 mile buffer zoneso trawlers and bycatch cannot be op- eratingCreate a buffer zone."
Task Force	2:53:55 James Nicori – "six inches (net) can
Public Testimony Nov. 14	catch the bigger salmons but they hangbefore you pull in your net they come loose and they fall down into the bottom of the riverThat's salmon reproduce being wastedWe have to look at too."
Task Force	3:01:30 Charles Lean – "I think that should be
Public Testimony Nov. 14	a research priority. At what point do we throw in the towel and say that stocks done? It's never coming back." - referring to addressing sustainable escapement threshold
Task Force	3:04:30 Charles Lean – "I'd like to see these
Public Testimony Nov. 14	research efforts focus on existing science I'd like to see synthesis papersreach som conclusionssomething you can act on."
Task Force Public Testimony Nov. 14	3:05:40 Charles Lean – "Most importantly taking action and evaluating that action to- wards efficacy."
Task Force Public Testimony Nov. 14	3:07:58 Tiffany Agayar – "I was wondering if anybody has ever compiled all the data to- gether with how they run, returning back to their streams."
Task Force Public Testimony Nov. 14	3:11:15 Tiffany Agayar – "I was just wondering if they were thinking of making a reconciliation of tagged fishwith different areas where fish are." - asking about synthesis and compilation of tagged fish data (e.g., migration)
Task Force Public Testimony Nov. 14	3:12:20 Tiffany Agayar – "I believe that tracking would help with being able to point how to and when to regulate just like they do on the Yukon."
Task Force Public Testimony Nov. 14	3:15:20 Kathleen Demientieff – "I would rather have the Native (community - not sure - indiscernible) approvewe have Traditional Knowledge."

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Source	Powerful Quotes
Task Force	3:59:45 Daniel Schindler (Dan Gillikin) – "
Public Testimony Nov. 14	freshwater stream temperature regime"
Task Force Public Testimony Nov. 14	4:02:35 Vanessa von Biela – "We do have a lot of water temperature data from across the state one of the major problemswe're thinking about them in terms of averages but is that an accurate reflection of what ani- mals are experiencing?often missing events because we're doing things like taking aver- ages." - saying we need more detailed water temperature data
Task Force Public Testimony Nov. 14	4:04:45 Dan Gillikin – "Second, is the loss of MDN subsidies to our streamsWhat has been the total loss of biomass contributed by returning salmon in these systems over the last 10, 20, 30, 40 (years). This loss has had to of reduced the carrying capacity of these systemsCan this be correlated to juvenile growth inferred from scale analysis?"
Task Force Public Testimony Nov. 14	4:08:40 Vanessa von Biela – "What does that mean for the way we manage salmon if the productivity of the ecosystem is gonna be reduced?"
Task Force Public Testimony Nov. 14	4:09:55 Vanessa von Biela – "The one research gap that I can identify is we really don't have the funding to do all the things that we need to do."

References

References were gathered by the Alaska Salmon Research Task Force Members and through Public emails and comments.

Reference Title

2020 KRITFC Community-Based Harvest Monitoring Program Summary & Report

2020 Takotna River Salmon Run Timing and Abundance

2021 KRITFC Community-Based Harvest Monitoring Program Summary & Report

2021 Kuskokwim River Salmon Situation Report

2021 Takotna River Salmon Run Timing and Abundance

2022 Kuskokwim River Salmon Situation Report

2022 Takotna River Salmon Run Timing and Abundance

A

Ackerman, M. W., C. Habicht, and L. W. Seeb. 2011. SNPs under diversifying selection provide increased accuracy and precision in mixed stock analyses of sockeye salmon from Copper River, Alaska. (PDF) Transactions of the American Fisheries Society 140: 865-881.

Ackerman, M.W., W.D. Templin, J.E. Seeb, and L.W. Seeb. 2013. Landscape heterogeneity and local adaptation define the spatial genetic structure of Pacific salmon in a pristine environment. Conservation Genetics 14: 483-498.

ADF&G. 2022. A study of the interactions between hatchery and natural pink and chum salmon in Southeast Alaska and Prince William Sound streams Progress Synopsis May 2022 ADF&G Chinook Salmon Research Team. 2013. Chinook salmon stock assessment and research plan, 2013. Alaska Department of Fish and Game, Special Publication No. 13-01, Anchorage. <u>http://www.adfg.alaska.</u> gov/FedAidPDFs/SP13-01.pdf

Alaska Department of Fish and Game Staff. 2022. Preliminary Harvest Rates of Western Alaska and Alaska Peninsula Chum Salmon Stocks in South Alaska Peninsula Fisheries, 2022. Regional Information Report No. 5J23-02.

Alaska Department of Fish and Game Staff. 2023. Chignik River Sockeye Salmon Action Plan, 2023. RC 4 Alaska Board of Fisheries. Alaska Department of Fish and Game. February 2023

Alaska Department of Fish and Game Staff. 2023. Chignik River King Salmon Action Plan, 2023. RC 5 Alaska Board of Fisheries. Alaska Department of Fish and Game. February 2023. Adkison, M.D. 2002. Preseason forecasts of pink salmon harvests in southeast Alaska using Bayesian model averaging. Alaska Fishery Research Bulletin 9, 1–8.Agler, B.A., G.T. Ruggerone, L.I. Wilson, and F.J. Mueter. 2013. Historical growth of Bristol Bay and Yukon River, Alaska chum salmon in relation to climate and inter-and intraspecific competition. Deep-Sea Research II 94:165-177.

Agler, B.A., G.T. Ruggerone, L.I. Wilson, and F.J. Mueter. 2013. Historical growth of Bristol Bay and Yukon River, Alaska chum salmon (*Oncorhynchus keta*) in relation to climate and inter- and intraspecific competition. Deep Sea Research Part II: Topical Studies in Oceanography 94, 165–177. <u>https://doi.org/10.1016/j</u>. dsr2.2013.03.028.

Alexander, R.F., and K. K. English. 2022. Preliminary assessment of the Canadian and Alaskan Sockeye stocks harvested in the northern boundary fisheries using run reconstruction techniques, 2009-2021. Prepared by LGL Limited, Sidney, BC, for the Pacific Salmon Commission, Vancouver, BC, and Fisheries and Oceans, Canada, Prince Rupert, BC. 127p.

Alexandersdottir, M. 1987. Life history of pink salmon (*Onco-rhynchus gorbuscha*) in Southeast Alaska and implications for management. Ph.D. Thesis. University of Washington, Seattle.

Andersen, D. B., B. Retherford, and C. Brown. 2013. Climate Change and Impacts on Subsistence Fisheries in the Yukon River Drainage, Alaska. Fisheries Resource Monitoring Program 10-250 final report.

Anderson, J.H., P.L. Faulds, W.I. Atlas, and T.P. Quinn. 2013. Reproductive success of captively bred and naturally spawned Chinook salmon colonizing newly accessible habitat. Evolutionary Applications, 6(2), pp.165-179.

Anderson, J.H., E.J. Ward, and S.M. Carlson. 2011. A model for estimating the minimum number of offspring to sample in studies of reproductive success. Journal of Heredity, 102(5), pp.567-576.

Anderton, I., and P. Frost. 2002. "Traditional/Local Knowledge Salmon Survey." Yukon River Panel Project CRE-16-02 Final Report (2002).

Andrews, A.G, E.V. Farley, Jr., J.H. Moss, J.M. Murphy, and E.F. Husoe. 2009. Energy density and length of juvenile Pink salmon in the eastern Bering Sea from 2004 to 2007: a period of relatively warm and cool sea surface temperatures. North Pacific Anadromous Fish Commission Bulletin 5:183-189.

Araki, H., and M.S. Blouin. 2005. Unbiased estimation of relative reproductive success of different groups: evaluation and correction of bias caused by parentage assignment errors. Molecular Ecology, 14(13), pp.4097-4109.

Azuma, T., 1992. Diel feeding habits of sockeye and chum salmon in the Bering Sea during the summer. Nippon Suisan Gakkaishi 58, 2019–2025.

Azumaya, T., and Y. Ishida. 2000. Density interactions between pink salmon (*Oncorhynchus gorbuscha*) and chum salmon (O. keta) and their possible effects on distribution and growth in the North Pacific Ocean and Bering Sea. NPAFC Bulletin 2, 165–174.

В

Baker, M.R., and A.B. Hollowed. 2014. Delineating ecological regions in marine systems: Integrating physical structure and community composition to inform spatial management in the eastern Bering Sea. Deep Sea Research Part II: Topical Studies in Oceanography 109, 215–240. https://doi.org/10.1016/j.dsr2.2014.03.001.

Baker, T.T., A. Wertheimer, R.D. Burkett, R. Dunlap, D.M. Eggers, E.I. Fritts, A.J. Gharrett, R.A. Holmes, and R.L Wilmot. 1996. Status of Pacific salmon and steelhead escapements in Southeastern Alaska. Fisheries 21, 6–18.

Barclay, A.W., P.A. Crane, D.B. Young, H.A. Hoyt, and C. Habicht. 2017. Current status of genetic studies of coho salmon from Southcentral Alaska and evaluations for mixed-stock analysis in Cook Inlet. Alaska Department of Fish and Game, Fishery Manuscript Series No. 17-01, Anchorage.

Barclay, A.W., and C. Habicht. 2019. Genetic baseline for Cook Inlet coho salmon and evaluations for mixed stock analysis. Alaska Department of Fish and Game, Fishery Manuscript Series No. 19-19, Anchorage.

Barclay, A.W., D.F. Evenson, and C. Habicht. 2019. New genetic baseline for Upper Cook Inlet Chinook salmon allows for the identification of more stocks in mixed stock fisheries: 413 loci and 67 populations. Alaska Department of Fish and Game, Fishery Manuscript Series No. 19-06, Anchorage.

Barclay, A.W., M. Schuster, C.M. Kerkvliet, M.D. Booz, B.J. Failor, and C. Habicht. 2019. Coded wire tag augmented genetic mixed stock analysis of Chinook salmon harvested in Cook Inlet marine sport fishery, 2014-2017. Alaska Department of Fish and Game, Fishery Manuscript No. 19-04, Anchorage.

Barclay, A.W., and E. L. Chenoweth. 2021. Genetic stock identification of Upper Cook Inlet sockeye salmon harvest, 2020. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report No. 5J21-04, Anchorage.

Barclay, A.W., H.A. Hoyt, and C. Habicht. 2021. Genetic population structure of Chinook salmon from Middle and Upper Susitna River: A report to Alaska Energy Authority, Susitna-Watana hydroelectric project (submitted July 25, 2017). Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report No. 5J21-02, Anchorage.

Barclay, A.W., S. Gilk-Baumer, K. Shedd, J. Botz, and C. Habicht. 2022. Genetic stock composition of the commercial harvest of Chinook salmon in Copper River District, 2018-2021. Alaska Department of Fish and Game, Fishery Data Series No. 22-35, Anchorage.

Bataille, C.P., Brennan, S.R., Hartmann, J., Moosdorf, N., Wooller, M.J., Bowen, G.J., 2014. A geostatistical framework for predicting variability in strontium concentrations and isotope ratios in Alaskan rivers. Chemical Geology 389, 1–15. <u>https://doi.org/10.1016/j</u>. chemgeo.2014.08.030

Batten, S.D., G.T. Ruggerone, and I. Ortiz. 2018. Pink salmon induce a trophic cascade in plankton populations in the southern Bering Sea and around the Aleutian Islands. Fisheries Oceanography doi:10.111/fog.12276.

Baumer, J., J. Wadle, and R. Dublin. 2022 Chignik King Salmon Stock of Concern Draft Action Plan. Oral Presentation to Alaska Board of Fisheries. Alaska Department of Fish and Game.

Beacham, T.D., C.B. Murray, and R.E. Withler. 1989. Age, morphology, and biochemical genetic variation of Yukon River Chinook salmon. Transactions of the American Fisheries Society 118: 46-63. https://doi.org/10.1577/1548-8659(1989)118<0046 :AMABGV>2.3.CO;2.

Beacham, T.D., J.R. Candy, C. Wallace, S. Urawa, S. Sato, N.V. Varnavskaya, K.D. Le, M. Wetklo. 2009. Microsatellite stock identification of Chum Salmon on a Pacific Rim basis. North American Journal of Fisheries Management 29:1757–1776. https://doi. org/10.1577/M08-188.1.

Beacham, T. D., M. Wetklo, L. Deng, and C. MacConnachie. 2011. Coho Salmon Population Structure in North America Determined from Microsatellites, Transactions of the American Fisheries Society, 140(2):253-270.

Beamish, R.J., B.E. Riddell, K.L. Lange, E. Farley Jr., S. Kang, T. Nagasawa, V. Radchenko, O. Temnykh, and S. Urawa. 2009. A long-term research and monitoring plan (LRMP) for Pacific salmon in the North Pacific Ocean. North Pacific Anadromous Fish Commission Special Publication No. 1, 44 p.

Beamish, R. J., editor. 2018 The ocean ecology of Pacific salmon and trout. American Fisheries Society, Bethesda, Maryland.

Beaudreau, A.H., M.N. Chan, and P.A. Loring. 2018. Harvest portfolio diversification and emergent conservation challenges in an Alaskan recreational fishery. Biological Conservation 222, 268–277. https://doi.org/10.1016/j.biocon.2018.04.010.

Beechie, T.J., C. Fogel, C. Nicol, J. Jorgensen, B. Timpane-Padgham, and P. Kiffney. 2022. How does habitat restoration influence resilience of salmon populations to climate change? Ecosphere 14(2):e4402. doi:10.1002/ecs2.4402.

Bellmore, J.R., J.B. Fellman, E. Hood, M.R. Dunkle, and R.T. Edwards. 2022. A melting cryosphere constrains fish growth by synchronizing the seasonal phenology of river food webs. Global Change Biology 28(16):4807–4818. https://doi.org/10.1111/gcb.16273.

Berkman, S.A., T.M. Sutton, F.J. Mueter, and B.W. Elliott. 2021. Effects of early-life stage and environmental factors on the freshwater and marine survival of Chinook salmon (*Oncorhynchus tshawytscha*) in rivers of southeast Alaska. Fishery Bulletin 119:201–215. https://doi.org/10.7755/FB.119.4.1.

Bernard, D.R. 1983. Variance and bias of catch allocations that use the age composition of escapements. Alaska Department of Fish and Game, Division of Commercial Fisheries, Informational Leaflet No. 227, Juneau.

Bond, N., J.E. Overland, M. Spillane, and P. Stabeno. 2003. Recent shifts in the state of the North Pacific. Geophysical Research Letters 30:2–5. https://doi.org/10.1029/2003GL018597.

Botz, J., and M.A. Sommerville. 2021. Management of salmon stocks in the Copper River, 2018-2020 -- a report to the Alaska Board of Fisheries. Alaska Department of Fish and Game, Special Publication No. 21-08, Anchorage.

Bowen L, V.R. von Biela, S.D. McCormick, A.M. Regish, S.C. Waters, B. Durbin-Johnson, M. Britton, M.L. Settles, D.S. Donnelly, S.M. Laske, et al. 2020. Transcriptomic response to elevated water temperatures in adult migrating Yukon River Chinook salmon (*Oncorhynchus tshawytscha*). Conservation Physiology 8:1–22.

Brabets, T.P., and M.A. Walvoord. 2009. Trends in streamflow in the Yukon River Basin from 1944 to 2005 and the influence of the Pacific Decadal Oscillation. Journal of Hydrology 371:108–119.

Bradford, M.J., J. Duncan, and J.W. Jang. 2009. Downstream migrations of juvenile salmon and other fishes in the upper yukon river. Arctic 61(3):255-264.

Bradford, M.J., J.A. Grout, and S. Moodie. 2001. Ecology of juvenile Chinook salmon in a small non-natal stream of the Yukon River drainage and the role of ice conditions on their distribution and survival. Canadian Journal of Zoology. 79(11):2043-2054. https:// doi.org/10.1139/z01-165.

Bradley, P. T., M.D. Evans, and A.C. Seitz. 2015. Characterizing the Juvenile Fish Community in Turbid Alaskan Rivers to Assess Potential Interactions with Hydrokinetic Devices. Transactions of the American Fisheries Society 144(5):1058–1069.

Braem, N.M., E. Mikow, A. Goddhun, A.R. Brenner, A. Trainor, S.J. Wllson and M.L. Kostick. 2017. Key Subsistence Fisheries in Northwest Alaska, 2012-2014. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 433, Fairbanks.

Briscoe, R.J., M.D. Adkison, A. Wertheimer, and S.G. Taylor. 2005. Biophysical factors associated with the marine survival of Auke Creek, Alaska, coho salmon. Transactions of the American Fisheries Society 134:817–828.

Brock, M., and P. Coiley-Kenner. 2009. A compilation of traditional knowledge about the fisheries of Southeast Alaska. Alaska Department of Fish and Game, Division of Subsistence, Technical Paper No. 332. Bromaghin, J.F., 2005. A versatile net selectivity model, with application to Pacific salmon and freshwater species of the Yukon River, Alaska. Fisheries Research 74:157–168. https://doi.org/10.1016/j. fishres.2005.03.004.

Bromaghin, J.F., 2008. Bels: Backward elimination locus selection for studies of mixture composition or individual assignment. Molecular Ecology Resources 8:568–571. https://doi.org/10.1111/j.1471-8286.2007.02010.x.

Bromaghin, J.F., R.M. Nielson, and J.J. Hard. 2008. An investigation of the potential effects of selective exploitation on the demography and productivity of Yukon River Chinook salmon. Alaska Fisheries Technical Report Number 100. USFWS.

Bromaghin, J.F, D.F. Evenson, T.H. McLain, and B.G. Flannery. 2011. Using a Genetic Mixture Model to Study Phenotypic Traits: Differential Fecundity among Yukon River Chinook Salmon. Transactions of the American Fisheries Society 140:235–249. https:// doi.org/ 10.1080/00028487.2011.558776.

Bromaghin, J.F., R.M. Nielson, and J.J. Hard. 2011. A model of Chinook salmon population dynamics incorporating size-selective exploitation and inheritance of polygenic correlated traits. Natural Resource Modeling 24:1–47. https://doi. org/10.1111/j.1939-7445.2010.00077.x.

Brown, C.L., J.S. Maganz, D.S. Koster, and N.M. Braem. 2009. Subsistence Harvests in 8 Communities in the Central Kuskokwim Drainage, 2009. Alaska Department of Fish and Game, Division of Subsistence, Technical Paper 365, Juneau.

Brown, C.L., and M.L. Kostick, editors. 2017. Harvest and Use of Subsistence Resources in 4 Communities in the Nenana Basin, 2015. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 429, Fairbanks.

Brown, C.L., H. Cold, L. Hutchinson-Scarbrough, B. Jones, J.M. Keating, B.M. McDavid, M. Urquia, J. Park, L.A. Sill, and T. Barnett. 2022. Alaska Subsistence and Personal Use Salmon Fisheries 2019 Annual Report. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 490, Anchorage.

Brown, R.J., A. von Finster, R.J. Henszey, and J.H. Eiler. 2017. Catalog of Chinook salmon spawning areas in Yukon River Basin in Canada and United States. Journal of Fish and Wildlife Management 8(2):558 - 586; e1944-687X. doi:10.3996/052017-JFWM-045.

Brown, R.J., Bradley, C., Melegari, J.L., 2020. Population trends for Chinook and summer chum salmon in two Yukon River tributaries in Alaska. Journal of Fish and Wildlife Management 11:377–400. https://doi.org/10.3996/072019-JFWM-064.

Brown, Z.W., G.L. van Dijken, and K.R. Arrigo. 2011. A reassessment of primary production and environmental change in the Bering Sea. Journal of Geophysical Research 116, C08014. https://doi. org/10.1029/2010JC006766. Brown, Z.W., and K.R. Arrigo. 2013. Sea ice impacts on spring bloom dynamics and net primary production in the Eastern Bering Sea. Journal of Geophysical Research: Oceans 118:43–62. https://doi. org/10.1029/2012JC008034.

Bryant, M.D., and F.H. Everest. 1998. Management and condition of watersheds in southeast Alaska: the persistence of anadromous salmon. Northwest Science 72:249–267.

Bryant, M.D., 2000. Estimating Fish Populations by Removal Methods with Minnow Traps in Southeast Alaska Streams. North American Journal of Fisheries Management 20:923–930. https:// doi.org/10.1577/1548-8675(2000)020<0923:EFPBRM>2.0.CO;2

Bryant, M.D., N.D. Zymonas, and B.E. Wright. 2004. Salmonids on the fringe: abundance, species, composition, and habitat use of salmonids in high-gradient headwater streams, southeast Alaska. Transactions of the American Fisheries Society 133:1529–1538.

Bryant, M.D., 2009. Global climate change and potential effects on Pacific salmonids in freshwater ecosystems of southeast Alaska. Climatic Change 95:169–193.

Bryant, M.D., and R.D. Woodsmith. 2009. The response of salmon populations to geomorphic measurements at three scales. North American Journal of Fisheries Management 29:549–559.

Bue, B.G., D.B. Molyneaux, and K.L. Schaberg. 2008. Kuskokwim River chum salmon run reconstruction. Alaska Department of Fish and Game, Anchorage, Fishery Data Series No. 08-64, Anchorage.

Bue, B.G., K.L. Schaberg, Z.W. Liller, and D.B. Molyneaux. 2012. Estimates of the historic run and escapement for the Chinook salmon stock returning to the Kuskokwim River, 1976-2011. Fishery Data Series 12–49, 1–47.

Buffington, J.M., T.E. Lisle, R.D. Woodsmith, and S. Hilton. 2002. Controls on the size and occurrence of pools in coarse-grained forest rivers. River Research and Applications 18:507–531. https:// doi.org/10.1002/rra.693.

Bugaev, A.V., E.A. Zavolokina, L.O. Zavarina, A.O. Shubin, S.F. Zolotukhin, N.F. Kaplanova, N. Volobuev. 2006. Identification of local stocks of chum salmon *Oncorhynchus keta* in the western Bering Sea in trawl catches of R/V TINRO in September-October, 2002 and 2003. Izvestia TINRO 146:3-34 (In Russian).

Bugaev, A.V., and K.W. Myers. 2009. Stock-specific distribution and abundance of immature Chinook salmon in the western Bering Sea in summer and fall 2002-2004. North Pacific Anadromous Fisheries Commission Bulletin 5:87–97.

Bugaev, A.B., E.A. Zavolokina, L.O. Zavarina, E.H. Kiryeev, A.O. Shubin, Yo.E. Ignatev, S.F. Zolotukhi, H.F. Kaplanova, and M.B. Volobuev. 2009. Origin and distribution of local stocks of chum salmon in the western Bering Sea based on trawl surveys of the R/V TINRO in 2004 and 2006. North Pacific Anadromous Fish Commission Doc. 1271. 30p.

Buklis, L.S. 1999. A description of economic changes in commercial salmon fisheries in a region of mixed subsistence and market economies. Arctic 52:40–48.

Burkett, B., J. Koenings, M. Haddix, and D. Barto. 1989. Cooperative ADF&G, FRED Division/U.S. Forest Service lake enrichment program for Southeast Alaska. Alaska Department of Fish and Game, FRED Report No. 98.

Burnside, C., and B. A. Fuerst. 2023. Chignik Management Area salmon annual management report, 2022. Alaska Department of Fish and Game, Fishery Management Report No. 23-02, Anchorage.

Burril, S.E., V.R. von Biela, N. Hillgruber, and C.E. Zimmerman. 2018. Energy allocation and feeding ecology of juvenile chum salmon (*Oncorhynchus keta*) during transition from freshwater to saltwater. Polar Biology 41(7):1–15.

Buschman, V.Q, and E. Sudlovenick. 2023. Indigenous-led conservation in the Arctic supports global conservation practices. Arctic Science 9:714-719 dx.doi.org/10.1139/AS-2022-0025.

С

Carey M.P., V.R. von Biela, A. Dunker, K.D. Keith, M. Schelske, C. Lean, and C.E. Zimmerman. 2021. Egg retention of high-latitude sockeye salmon (*Oncorhynchus nerka*) in the Pilgrim River, Alaska, during the Pacific marine heatwave of 2014–2016. Polar Biology 44:1643–1654.

Carothers, C., S. Cotton, and K. Moerlein. 2013. Subsistence Use and Knowledge of Salmon in Barrow and Nuiqsut, Alaska. Final Report for OCS Study BOEM 2013-0015. University of Alaska Coastal Marine Institute, Fairbanks.

Carothers, C., T.L. Sformo, S. Cotton, J.C. George, and P.A.H. Westley. 2019. Pacific salmon in the rapidly changing Arctic: Exploring local knowledge and emerging fisheries in Utqiagvik and Nuiqsut, Alaska. Arctic 72(3):273-288.

Carothers, C., J. Black, S.J. Langdon and others. 2021. Indigenous peoples and salmon stewardship: a critical relationship. Ecology and Society 26(1):16.

Carvalho, K.S., T.E. Smith, and S. Wang. 2021. Bering Sea marine heatwaves: Patterns, trends and connections with the Arctic. Journal of Hydrology 600:126462. https://doi.org/10.1016/j. jhydrol.2021.126462.

Celewycz, A.G., A.C. Wertheimer, J.A. Orsi, and J.L. Lum. 1994. Nearshore distribution and residency of pink salmon (*Oncorhynchus gorbuscha*) and chum salmon (*O. keta*) fry and their predators in Auke Bay and Gastineau Channel, southeast Alaska. AFSC Processed Report 94-05.

Chaloner, D.T., K.M. Martin, M.S. Wipfli, P.H. Ostrom, and G.A. Lamberti. 1996. Marine carbon and nitrogen in southeastern Alaska stream food webs: evidence from artificial and natural streams. Canadian Journal of Fisheries and Aquatic Sciences 59:1257–1265.

Chasco, B., R. Hilborn, and G.T. Ruggerone. 2004. Chignik salmon studies investigations of salmon populations, hydrology, and limnology of the Chignik lakes, Alaska, during 2003-2004. Technical Report, School of Aquatic and Fishery Science, Fisheries Research Institute, Washington University Issue 0304.

Chasco, B.E., and others. 2017. Competing tradeoffs between increasing marine mammal predation and fisheries harvest of Chinook salmon. Nature/Scientific Reports. doi:10.1038/s41598-017-14984-8

Cheng, W., C. Habicht, W.D. Templin, Z.D. Grauvogel, S.D. Moffitt, R.E. Brenner, R.P. Josephson, and A.J. Gharrett. 2013. Population genetic structure of odd-year pink salmon from Prince William Sound based on a single year. Alaska Hatchery Research Group Technical Document No. 14.

Cheng, W., E. Curchitser, C. Ladd, P. Stabeno, and M. Wang. 2014. Influences of sea ice on the Eastern Bering Sea: NCAR CESM simulations and comparison with observations. Deep Sea Research Part II: Topical Studies in Oceanography 109:27–38. https://doi. org/10.1016/j.dsr2.2014.03.002.

Chenoweth E.M., and K.R. Criddle. 2019. The Economic Impacts of Humpback Whale Depredation on Hatchery-Released Juvenile Pacific Salmon in Southeast Alaska. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 11:62–75.

Chenoweth E.M., J.M. Straley, M.V. McPhee, S. Atkinson, and S. Reifenstuhl. 2017. Humpback whales feed on hatchery-released juvenile salmon. Royal Society Open Science 4:170180. <u>http://dx.doi.org/10.1098/</u> rsos.170180.

Cieciel, K., E.V. Farley, L.B. Eisner. 2009. Jellyfish and Juvenile Salmon Associations with Oceanographic Characteristics during Warm and Cool Years in the Eastern Bering Sea. North Pacific Anadromous Fish Commission Bulletin 5:209–224.

Clark, R., M. Willette, S. Fleischman, and D. Eggers. 2007. Biological and fishery-related aspects of over escapement in Alaska sockeye salmon *Onchorynchus nerka*. Alaska Department of Fish and Game, Special Publication No. 07-17, Anchorage.

Cline, T.J., J. Ohlberger, and D.E. Schindler. 2019. Effects of warming climate and competition in the ocean for life-histories of Pacific salmon. Nature Ecology and Evolution 3:935–942.

Connors, B., M.J. Malick, G.T. Ruggerone, P. Rand, M. Adkison, J.R. Irvine, R. Campbell, and K. Gorman. 2020. Climate and competition influence sockeye salmon population dynamics across the Northeast Pacific Ocean. Canadian Journal of Fisheries and Aquatic Sciences 77:943-949.

Connors, B.M., M.R. Siegle, J. Harding, S. Rossi, B.A. Staton, M.L. Jones, M.J. Bradford, R. Brown, B. Bechtol, B. Boherty, S. Cox, and B.J.G. Sutherland. 2022. Chinook salmon diversity contributes to fishery stability and trade-offs with mixed-stock harvest. Ecological Applications 32(8):e2709. https://doi.org/10.1002/eap.2709.

Conrad, S., and W. Davidson. 2013. Overview of the 2012 Southeast Alaska and Yakutat commercial, personal use, and subsistence salmon fisheries. Alaska Department of Fish and Game, Fishery Management Report No. 13-03, Anchorage.

Cook, M.E.A., and M.V. Sturdevant. 2013. Diet Composition and Feeding Behavior of Juvenile Salmonids Collected in the Northern Bering Sea from August to October, 2009 – 2011. North Pacific Anadromous Fish Commission, Technical Report No. 9:118–126.

Copeman, L.A., C.D. Salant, M.A. Stowell, M.L. Spencer, D.G. Kimmel, A.I. Pinchuk, and B.J. Laurel. 2022. Annual and spatial variation in the condition and lipid storage of juvenile Chukchi Sea gadids during a recent period of environmental warming (2012 to 2019). Deep Sea Research Part II: Topical Studies in Oceanography 20:105180. https://doi.org/10.1016/j.dsr2.2022.105180.

Courtney, M.B., M. Evans, K.R. Shedd, et al. 2021. Understanding the behavior and ecology of Chinook salmon (*Oncorhynchus tshawytscha*) on an important feeding ground in the Gulf of Alaska. Environmental Biology of Fishes 104:357–373. https://doi. org/10.1007/ s10641-021-01083-x

Cox, C.J., R.S. Stone, D.C. Douglas, D.M. Stanitski, and M.R. Gallagher. 2019. The Aleutian Low-Beaufort Sea Anticyclone: A Climate Index Correlated With the Timing of Springtime Melt in the Pacific Arctic Cryosphere. Geophysical Research Letters 46:7464–7473. https://doi.org/10.1029/2019GL083306.

Coyle, K.O., A.I. Pinchuk, L.B. Eisner, and J.M. Napp. 2008. Zooplankton species composition, abundance and biomass on the eastern Bering Sea shelf during summer: The potential role of water-column stability and nutrients in structuring the zooplankton community. Deep Sea Research Part II: Topical Studies in Oceanography 55:1775–1791. <u>https://doi.org/10.1016/j.dsr2.2008.04.029</u>.

Coyle, K.O., L.B. Eisner, F.J. Mueter, A.I. Pinchuk, M.A. Janout, K.D. Cieciel, E.V. Farley, and A.G. Andrews. 2011. Climate change in the southeastern Bering Sea: impacts on pollock stocks and implications for the oscillating control hypothesis. Fisheries Oceanography 20:139–156. https://doi.org/10.1111/j.1365-2419.2011.00574.x.

Creelman, E., L. Hauser, R. Simmons, W. Templin, and L. Seeb. 2011. Temporal and geographic genetic divergence: Characterizing sockeye salmon populations in the Chignik watershed, Alaska, using single nucleotide polymorphisms. Transactions of the American Fisheries Society 140: 749-762.

Crone, R.A., and C.E. Bond. 1976. Life history of coho salmon, *Oncorhynchus kisutch*, in Sashin Creek, Southeastern Alaska. Fisheries Bulletin 74:897–923.

Crowell, A.L., and M. Arimitsu. 2023. Climate change and pulse migration: intermittent Chugach Inuit occupation of glacial fjords on the Kenai Coast, Alaska. Frontiers in Environmental Archaeology 1:1145220. doi: 10.3389/fearc.2023.1145220.

Crozier, L.G., and J.E. Siegel. 2023. A comprehensive review of the impacts of climate change on salmon: strengths and weaknesses of the literature by life stage. Fishes 8(6): 319. https//doi. org/10.3390/fishes8060319.

Cunningham, C.J., T.A. Branch, T.H. Dann, M. Smith, J.E. Seeb, L.W. Seeb, and R. Hilborn. 2017. A General Model for Salmon Run Reconstruction That Accounts for Interception and Differences in Availability to Harvest. Canadian Journal of Fisheries and Aquatic Sciences 75(3):439-451. Online. doi: 10.1139/cjfas-2016-0360.

Cunningham, C.J., G.T. Ruggerone, and T.P. Quinn. 2013. Size selectivity of predation by brown bears depends on the density of their sockeye salmon prey. The American Naturalist 181(5):663-673. doi:10.1086/670026.

Cunningham, C.J., P.A.H. Westley, and M.D. Adkison. 2018. Signals of large scale climatic drivers, hatchery enhancement, and marine factors in Yukon River Chinook salmon survival revealed with a Bayesian life history model. Global Change Biology 24(9):4399-4416. doi:10.111/gcb.14315

D

Dann, T.H., C. Habicht, J.R. Jasper, H.A. Hoyt, A.W. Barclay, W.D. Templin, T.T. Baker, F.W. West, and L.F. Fair. 2009. Genetic stock composition of the commercial harvest of sockeye salmon in Bristol Bay, Alaska, 2006-2008. Alaska Department of Fish and Game, Fishery Manuscript Series No. 09-06, Anchorage.

Dann, T.H., W.W. Smoker, J.J. Hard, and A.J. Gharrett. 2010. Outbreeding depression after two generations of hybridizing Southeast Alaska coho salmon populations. Transactions of the American Fisheries Society 139:1292–1305. https://doi.org/10.1577/T09-203.1

Dann, T.H., A. Barclay and C. Habicht. 2012. Western Alaska Salmon Stock Identification Program Technical Document 5: Status of the SNP baseline for sockeye salmon. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J12-10, Anchorage.

Dann, T.H., C. Habicht, J.R. Jasper, E.K.C. Fox, H.A. Hoyt, H.L. Liller, E.S. Lardizabal, P.A. Kuriscak, Z.D. Grauvogel, and W.D. Templin. 2012. Sockeye salmon baseline for the Western Alaska Salmon Stock Identification Program. Alaska Department of Fish and Game, Special Publication No. 12-12, Anchorage.

Dann, T.H., C. Habicht, T.T. Baker, and J.E. Seeb. 2013. Exploiting genetic diversity to balance conservation and harvest of migratory salmon. Canadian Journal of Fisheries and Aquatic Sciences. 70:785-793.

Dann, T.H., C. Habicht, W.D. Templin, L.W. Seeb, G. McKinney, and J.E. Seeb. 2018. Identification of genetic markers useful for mixed stock analysis of Chinook salmon in Cook Inlet, Alaska. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J18-04, Anchorage. Dann, TH. 2021. Stock composition of subsistence harvests and total return of sockeye salmon from the Kvichak River, Dryad, Dataset, https://doi.org/10.5061/dryad.j0zpc86fm

Dann, T.H., H. Hoyt, E. Lee, E. Fox, and M.B. Foster. 2023. Genetic Stock Composition of Chum Salmon Harvested in Commercial Salmon Fisheries of the South Alaska Peninsula, 2022. Alaska Department of Fish and Game, Special Publication No. 23-07, Anchorage.

Daum, D. and B. Flannery. 2011. Canadian-origin Chinook salmon rearing in nonnatal U.S. tributary streams of the Yukon River, Alaska. Transactions of the American Fisheries Society 140:207-220. 10.1080/00028487.2011.545004.

Davidson, B., R. Bachman, T. Thynes, D. Gordon, A. Piston, K. Jensen, K. Monagle, and S. Walker. 2017. Annual Management Report of the 2011 Southeast Alaska Commercial Purse Seine and Drift Gillnet Fisheries. Alaska Department of Fish and Game, Fishery Management Report No. 11-27, Anchorage.

Davis, N.C.D., 2003. Feeding Ecology of Pacific Salmon (*Oncorhynchus* spp.) in the Central North Pacific Ocean and Central Bering Sea, 1991-2000. Hokkaido University. PhD Dissertation, 205p.

Davis, N.D., J.L. Armstrong, and K.W. Myers. 2003. Bering Sea salmon food habits: Diet overlap in fall and potential for interactions among salmon. Final Report to the Yukon River Drainage Fisheries Association. SAFS-UW-0311. Fisheries Research Institute, School of Aquatic and Fisheries Sciences, University of Washington, Seattle. 34 p.

Davis, N.D., M. Fukuwaka, J.L. Armstrong, and K.W. Myers. 2004. Salmon Food Habits Studies in the Bering Sea, 1960 to Present. North Pacific Anadromous Fish Commission Technical Report No. 6:24–28.

Davis, N.D., A.V. Volkov, A.Y. Efimkin, N.A. Kuznetsova, J.L. Armstrong, and O. Sakai, , 2009. Review of BASIS Salmon Food Habits Studies. North Pacific Anadromous Fish Commission, Bulletin No. 5:197–208.

De Robertis, A., and E.D. Cokelet. 2012. Distribution of fish and macrozooplankton in ice-covered and open-water areas of the eastern Bering Sea. Deep Sea Research Part II: Topical Studies in Oceanography 65–70:217–229. <u>https://doi.org/10.1016/j.</u> dsr2.2012.02.005

Decovich, N.A, and K.G. Howard. 2011. Genetic Stock Identification of Chinook Salmon Harvest on the Yukon River 2010. Alaska Department of Fish and Game, Fishery Data Series No. 11-65, Anchorage.

Deeg, C.M., et al. 2022. Pathogens and stressors of overwintering salmon in the Gulf of Alaska. North Pacific Anadromous Fish Commission Technical Report No. 18:47-52.

Denman, R.A., and G.T. Ruggerone. 1994. Effects of beaver colonization on the hydrology and spawning habitat of sockeye salmon in the Chignik Lakes, Alaska. Natural Resources Consultants, Inc report, Seattle, 65p.

Denton, C., 1988. Marine survival of Chinook salmon, *Oncorhynchus tshawytscha*, reared at three densities. Alaska Department of Fish and Game FRED Report 88. Juneau, AK. 10 p.

Der Hovanisian, J., J.S. McPherson, E. Jones, P. Richards, R. Chapell, B. Elliott, T. Johnson, and S. Fleischman. 2011. Chinook salmon status and escapement goals for stocks in Southeast Alaska. Alaska Department of Fish and Game, Special Publication No. 11-19, Anchorage.

Dickerson, B., K. Brinck, M. Willson, P. Bentzen, and T.P. Quinn. 2005. Relative importance of salmon body size and arrival time at breeding grounds to reproductive success. Ecology 86:347–352.

Dickerson, B.R., T.P. Quinn, and M.F. Willson. 2002. Body size, arrival date, and reproductive success of pink salmon, *Oncorhynchus gorbuscha*. Ethology Ecology & Evolution 14:29–44. https://doi. org/10.1080/08927014.2002.9522759.

Division of Commercial Fisheries Staff 2023. Preliminary Harvest Rates of Western Alaska and Alaska Peninsula Chum Salmon Stocks in South Alaska Peninsula Fisheries, 2022. A Report to the Alaska Board of Fisheries February 2023. Alaska Department of Fish and Game

Doctor, K.K., R. Hilborn, M. Rowse, and T. Quinn, 2009. Spatial and temporal patterns of upriver migration by sockeye salmon populations in the Wood River system, Bristol, Bay, Alaska. Transactions of the American Fisheries Society 139:80-91. DOI: 10.1577/T08-227.1

Donkersloot, R., J.C. Black, C. Carothers, D. Pinger, W. Justin, P.M. Clay, M.R. Poe, E.R. Gavenus, W. Voinot-Baron, C. Stevens, and M. Williams. 2020. Assessing the sustainability and equity of Alaska salmon fisheries through a well-being framework. Ecology and Society 25(2):18.

Donnellan, S.J., and A.R. Munro, editors. 2023. Run forecasts and harvest projections for 2023 Alaska salmon fisheries and review of the 2022 season. Alaska Department of Fish and Game, Special Publication No. 23-10, Anchorage.

Dube, M., B. Muldoon, J. Wilson, and K.B. Maracle. 2012. Accumulated state of the Yukon River watershed: Part 1 Critical Review of Literature. Integrated Environmental Assessment and Management 9(3):426-438

Dube, M., J.E. Wilson, and J. Waterhouse. 2012. Accumulated state assessment of the Yukon River watershed: Part II Quantitative effects-based analysis integrating western science and traditional ecological knowledge. Integrated Environmental Assessment and Management 9(3):439-455.

DuBois, L., Z.W. Liller. 2010. Yukon River chinook salmon aging consistency. Alaska Department of Fish and Game, Fishery Data Series No. 10-45, Anchorage.

Duffy-Anderson, J.T., P.J. Stabeno, E.C. Siddon, A.G. Andrews, D.W. Cooper, L.B. Eisner, E.V. Farley, C.E. Harpold, R.A. Heintz, D.G. Kimmel, F.F. Sewall, A.H. Spear, and E.C. Yasumiishi, 2017. Return of warm conditions in the southeastern Bering Sea: Physics to fluorescence. PLoS ONE 12:1–21. https://doi.org/10.1371/ journal. pone.0185464.

Duncan, D.H., and A.H. Beaudreau. 2019. Spatiotemporal Variation and Size-Selective Predation on Hatchery- and Wild-Born Juvenile Chum Salmon at Marine Entry by Nearshore Fishes in Southeast Alaska. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 11:372–390.

Dunmall, K.M., J.D. Reist, E.C. Carmack, J.A. Babaluk, M.P. Heide-Jørgensen, and M.F. Docker. 2013. Pacific Salmon in the Arctic: Harbingers of Change. In: F.J. Mueter, D.M.S. Dickson, H.P. Huntington, J.R. Irvine, E.A.Logerwell, S.A. MacLean, L.T. Quakenbush, and C. Rosa (*eds.*), Responses of Arctic Marine Ecosystems to Climate Change. Alaska Sea Grant, University of Alaska Fairbanks. doi:10.4027/ ramecc.2013.07

Dunmall, K.M., N.J. Mochnacz, C.E. Zimmerman, C. Lean, and J.D. Reist. 2016. Using thermal limits to assess establishment of fish dispersing to high-latitude and high-elevation watersheds. Can.J.Fish. Aquat. Sci 73:1750-1758

Ξ

Ebbin, S.A. 2002. What's Up? The Transformation of Upstream-Downstream Relationships on Alaska's Kuskokwim River. Polar Geography 26, 147–166. https://doi.org/10.1080/789610136.

Echave, J.D., C.V. Manhard, W.W. Smoker, M.D. Adkison, and A.J. Gharrett. 2017. Out crosses between seasonally different segments of a Pacific salmon population reveal local adaptation. Environmental Biology of Fishes 100:1469–1481. <u>https://doi.org/10.1007/</u> s10641-017-0657-3.

Echave, K., M. Eagleton, E. Farley, and J. Orsi. 2012. A Refined Description of Essential Fish Habitat for Pacific Salmon Within the U. S. Exclusive Economic Zone in Alaska. NOAA Technical Memorandum NMFS-AFSC-236

Eggers, D.M., R.L. Bachman, and J. Stahl. 2010. Stock status and escapement goals for Chilkat Lake sockeye salmon in Southeast Alaska. Alaska Department of Fish and Game, Fishery Manuscript No. 10-05, Anchorage.

Eiler, J.H., M.M. Masuda, T.R. Spencer, R.J. Driscoll, and C.B. Schreck. 2014 Distribution, Stock Composition and Timing, and Tagging Response of Wild Chinook Salmon Returning to a Large, Free-Flowing River Basin. Transactions of the American Fisheries Society 143:1476–1507.

Eiler, J.H., A.N. Evans, and C.B. Schreck. 2015. Migratory patterns of wild Chinook salmon *Oncorhynchus tshawytscha* returning to a large, free-flowing river basin. PLoS One 10:1–33.

Eiler, J.H., M.M. Masuda, and A.N. Evans. 2023. Swimming depths and water temperatures encountered by radio-archival-tagged Chinook Salmon during their spawning migration in the Yukon River basin. Transactions of the American Fisheries Society 152:51–74.

Eisner, L., N. Hillgruber, E. Martinson, and J. Maselko. 2012. Pelagic fish and zooplankton species assemblages in relation to water mass characteristics in the northern Bering and southeast Chukchi seas. Polar Biology 36:87–113. <u>https://doi.org/10.1007/</u> s00300-012-1241-0.

Eldridge, W.H., J.J. Hard, and K.A. Naish. 2010. Simulating fishery-induced evolution in chinook salmon: the role of gear, location, and genetic correlation among traits. Ecological Applications 20(7):1936-1948.

Elison, T.B., K.L. Schaberg, and D.J. Bergstrom. 2012. Kuskokwim River Salmon Stock Status and Kuskokwim Area Fisheries, 2012. A Report to the Alaska Board of Fisheries. Alaska Department of Fish and Game, Special Publication No. 12-27, Anchorage.

Esenkulova, S., C. Neville, E. DiCicco, and I. Pearsall. 2022. Indications that algal blooms may affect wild salmon in a similar way as farmed salmon. Harmful Algae 118:102310. <u>https://doi.org/10.1016/j.hal.2022.102310</u>.

Eskelin, A., and A.W. Barclay. 2022. Eastside set gillnet Chinook salmon harvest composition in Upper Cook Inlet, Alaska, 2021. Alaska Department of Fish and Game, Fishery Data Series No. 22-06, Anchorage.

Eskelin, T., and A.W. Barclay. 2022. Russian River early-run sockeye salmon run timing into the Kenai River, 2018–2020. Alaska Department of Fish and Game, Fishery Data Series No. 22-33, Anchorage.

Espinasse, B., B.P.V. Hunt, B.P. Finney, J.K. Fryer, A.V. Bugaev, and E.A. Pakhomov. 2020. Using stable isotopes to infer stock-specific high-seas distribution of maturing sockeye salmon in the North Pacific. Ecology and Evolution 10:13555–13570. https:// doi.org/10.1002/ece3.7022

Estensen, J.L., S.N. Schmidt, S. Garcia, C.M. Gleason, B.M. Borba, D.M. Jallen, A.J. Padilla, K.M. Hilton. 2018. Annual Management Report Yukon Area, 2016, Fishery Management Report No.18-14.

Evenson, D.F., C. Habicht, M. Stopha, A.R. Munro, T.R. Meyers, and W.D. Templin. 2018. Salmon hatcheries in Alaska – A review of the implementation of plans, permits, and policies designed to provide protection for wild stocks. Alaska Department of Fish and Game, Special Publication No. 18-12, Anchorage.

F

Fall, J.A., R.T. Stanek, B. Davis, L. Williams, and R. Walker. 2004. Cook Inlet Customary and Traditional Subsistence Fisheries Assessment. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 285. Juneau.

Farley, E.V., J.M. Murphy, B.W. Wing, J.H. Moss, and A. Middleton. 2005. Distribution, migration pathways, and size of western Alaska juvenile salmon along the eastern Bering Sea shelf. Alaska Fishery Research Bulletin 11:15–26.

Farley, E.V., Jr., J.M. Murphy, M.D. Adkison, L.B. Eisner, J.H. Helle, J.H. Moss, and J. Nielsen. 2007. Early marine growth in relation to marine-stage survival rates for Alaska Sockeye salmon. Fishery Bulletin 105:121-130.

Farley, E.V., and J.H. Moss. 2009. Growth rate potential of juvenile chum salmon on the eastern Bering Sea shelf: an assessment of salmon carrying capacity. North Pacific Anadromous Fish Commission Bulletin 5:265-277.

Farley, E.V., and M. Trudel. 2009. Growth Rate Potential of Juvenile Sockeye Salmon in Warmer and Cooler Years on the Eastern Bering Sea Shelf. Journal of Marine Biology 2009:1–10. https:// doi.org/10.1155/2009/640215

Farley, E.V., A. Starovoytov, S. Naydenko, R. Heintz, M. Trudel, C. Guthrie, L. Eisner, and J.R. Guyon. 2011. Implications of a warming eastern Bering Sea for Bristol Bay sockeye salmon. ICES Journal of Marine Science 68(6):1138-1146

Farley, E.V., Jr., T.D. Beacham, and A.V Bugaev. 2018. Ocean ecology of sockeye salmon. Pages 319-389 in R.J. Beamish, editor. The ocean ecology of Pacific salmon and trout. American Fisheries Society, Bethesda, Maryland.

Farley, E.V., Jr., J.M. Murphy, K. Cieciel, E.M. Yasumiishi, K. Dunmall, T. Sformo, and P. Rand. 2020. Response of Pink salmon to climate warming in the northern Bering Sea. Deep-Sea Research II. https://doi.org/10.1016/j.dsr2.2020.104830.

Farley, E.V., Jr., E.M. Yasumiishi, J.M. Murphy, W. Strasburger, F. Swell, K. Howard, S. Garcia, and J.H. Moss. 2024. Critical periods in the marine life history of juvenile western Alaska chum salmon in a changing climate. Marine Ecology Progress Series 726:149-160. https://doi.org/10.3354/meps14491.

Feddern, M.L., E.R. Schoen, R. Shaftel, C.J. Cunningham, C. Chythlook, B.M. Connors, A.D. Murdoch, V.R. vonBiela, B. Woods. 2023. Kings of the North: Bridging disciplines to understand the effects of changing climate on Chinook salmon in the Arctic-Yukon-Kuskokwim Region. Fisheries 48(8):317-356. doi: 10.1002/fish.10923.

Fellman, J.B., Hood, E., Dryer, W., Pyare, S., 2015. Stream Physical Characteristics Impact Habitat Quality for Pacific Salmon in Two Temperate Coastal Watersheds. Plos One 10, e0132652. https:// doi.org/10.1371/journal.pone.0132652

Fellman, J.B., S. Nagorski, S. Pyare, A.W. Vermilyea, D. Scott, and E. Hood. 2014. Stream temperature response to variable glacier coverage in coastal watersheds of Southeast Alaska. Hydrological Processes 28:2062–2073. https://doi.org/10.1002/hyp.9742

Fergusson, E. A., M.V. Sturdevant, and J.A. Orsi. 2013. Trophic relationships among juvenile salmon during 16-year time series of climate variability in Southeast Alaska. North Pacific Anadromous Fish Commission Technical Report 9:112–117

Fergusson, E., T. Miller, M.V. McPhee, C. Fugate, and H. Schultz. 2020. Trophic responses of juvenile Pacific salmon to warm and cool periods within inside marine waters of Southeast Alaska. Progress in Oceanography 186. <u>https://doi.org/10.1016/j.pocean.2020.102378</u>.

Fergusson, E., J. Murphy, and A. Gray. 2021. Southeast Alaska coastal monitoring survey: salmon trophic ecology and bioenergetics, 2019. NPAFC Doc. 1949. 40 pp. National Oceanic and Atmospheric Administration (NOAA), National Marine Fisheries Service (NMFS), Alaska Fisheries Science Center, Auke Bay Laboratories, Ted Stevens Marine Research Institute (Available at https://npafc.org).

Fergusson, E., W. Strasburger, J. Murphy, A. Piston, S. Heinl, and A. Gray. 2022. Southeast Alaska Coastal Monitoring Survey May–July 2021. NPAFC Doc. 2021. 44 pp. National Oceanic and Atmospheric Administration (NOAA), National Marine Fisheries Service (NMFS), Alaska Fisheries Science Center, Auke Bay Laboratories, Ted Stevens Marine Research Institute, and Alaska Department of Fish and Game (Available at https://npafc.org).

Finkle, H., K. Schaberg, M. Foster, and T. Polum. 2022. Review of Salmon Escapement Goals in the Chignik Management Area, 2020. Alaska Department of Fish and Game, Fishery Manuscript No. 22-05.

Finkle, H., K. Schaberg, M. Foster, M. Wattum, and T. Polum. 2022. Review of Salmon Escapement Goals in the Alaska Peninsula and Aleutian Islands Management Areas, 2020. Alaska Department of Fish and Game, Fishery Manuscript No. 22-06.

Finkle, H. 2023. Chignik watershed sockeye salmon run reconstruction and escapement goals. Oral Report to the Alaska Board of Fisheries 2023. Alaska Department of Fish and Game.

Finnegan, S.P., N.J. Svoboda, S.L. Schooler, and J.L. Belant. 2023. Phenological overlap of terrestrial and marine food resources did not reduce salmon consumption by Kodiak brown bears. Global Ecology and Conservation 45:e02506.

Fish, T.M., and A.W. Piston. 2022. Hugh Smith Lake sockeye salmon stock assessment, 2021. Alaska Department of Fish and Game, Fishery Data Series No. 22-18, Anchorage.

Flannery, B.G., P.A. Cran, J.H. Eiler, D. Beacham, N.A. DeCovich, W.D. Templin, O.L. Schlei and J.K. Wenburg. 2012. Comparison of Radiotelemetry and Microsatellites for Determining the Origin of Yukon River Chinook Salmon. North American Journal of Fisheries Management 32(4):720-730. Flannery, B.G., J.K. Wenburg, and A.J. Gharrett. 2007. Evolution of mitochondrial DNA variation within and among Yukon River chum salmon populations. Transactions of the American Fisheries Society 136:902–910.

Flannery, B.G., J.K. Wenburg, and A.J. Gharrett. 2007. Variation of amplified fragment length polymorphisms in Yukon River chum salmon: population structure and application to mixed-stock analysis. Transactions of the American Fisheries Society 136:911–925.

Flemming, S.M., N.L. Zeiser, S.C. Heinl, C.S. Jalbert, and S.E. Miller. 2022. Stock assessment study of Chilkoot Lake sockeye salmon, 2020–2021. Alaska Department of Fish and Game, Fishery Data Series No. 22-31, Anchorage.

Fox, E.K.C., T.D. Lawson, and M.D. Keyse. 2022. 2022 South Alaska Peninsula salmon annual management report and 2021 subsistence fisheries in the Alaska Peninsula, Aleutian Islands, and Atka-Amlia Islands management areas. Alaska Department of Fish and Game, Division of Commercial Fisheries, Fishery Management Report No. 22-32, Anchorage.

Friedland, K.D., R.V. Walker, N.D. Davis, K.W. Myers, G.W. Boehlert, S. Urawa, and Y. Ueno. 2001. Open-ocean orientation and return migration routes of chum salmon based on temperature data from data storage tags. Marine Ecology Progress Series 216:235–252.

Frost, T.J., E.M. Yasumiishi, B.A. Agler, M.D. Adkison, and M.V. McPhee. 2021. Density-dependent effects of eastern Kamchatka pink salmon (*Oncorhynchus gorbuscha*) and Japanese chum salmon (*O. keta*) on age-specific growth of western Alaska chum salmon. Fisheries Oceanography 30:99–109. <u>https://doi.org/10.1111/</u>fog.12505

Fukushima, M., and W.W. Smoker. 1998. Spawning Habitat Segregation of Sympatric Sockeye and Pink Salmon. Transactions of the American Fisheries Society 127:253–260. <u>https://doi.org/10.157</u> 7/1548-8659(1998)127<0253:SHSOSS>2.0.CO;2

Fukushima, M., S.G. Taylor, and W.W. Smoker. 1997. Fry and smolt production of pink and sockeye salmon in the Auke Lake system, Southeast Alaska. Acta Hydrobiologica Sinica 21:1–21.

G

Garcia, S., and F. Sewall. 2021. Diet and Energy Density Assessment of Juvenile Chinook Salmon from Northeastern Bering Sea Trawl. Alaska Department of Fish and Game, Fishery Data Series No. 21-05.

Garvin, M.R., C.M. Kondzela, P.C. Martin, B. Finney, J. Guyon, W.D. Templin, N. DeCovich, S. Gilk-Baumer, and A.J. Gharrett. 2013. Recent physical connections may explain weak genetic structure in western Alaskan chum salmon (*Oncorhynchus keta*) populations. Ecology and Evolution 3(7):2362-2377.

Gazey, W.J., and K.K. English. 2000. Assessment of sockeye and pink salmon stocks in the northern boundary area using run reconstruction techniques, 1982-95. Canadian Technical Report of Fisheries and Aquatic Sciences. 2320: 132p.

Geiger, H.J., W.W. Smoker, L.A. Zhivotovsky, and A.J. Gharrett. 1997. Variability of family size and marine survival in pink salmon (*Oncorhychus gorbuscha*) has implications for conservation biology and human use. Canadian Journal of Fisheries and Aquatic Sciences 54:2684–2690.

Geiger, H.J. 2003. Sockeye salmon stock status and escapement goal for ReDoubt Lake in Southeast Alaska. Alaska Department of Fish and Game, Regional Information Report No. 1J03-01.

Geiger, H.J., I. Wang, P. Malecha, K. Hebert, W.W. Smoker, and A.J. Gharrett. 2007. What causes variability in pink salmon family size? Transactions of the American Fisheries Society 136:1688–1698.

Gharrett, A.J., and S.M. Shirley. 1985. A genetic examination of spawning methodology in a salmon hatchery. Aquaculture 47:245–256.

Gharrett, A.J., and W.W. Smoker. 1991. Two generations of hybrids between even- and odd-year pink salmon (*Oncorhynchus gorbuscha*): a test for outbreeding depression? Canadian Journal of Fisheries and Aquatic Sciences 48:1744–1749.

Gharrett, A., J. Joyce, and W. Smoker. 2013. Fine-scale temporal adaptation within a salmonid population: mechanism and consequences. Molecular Ecology 22:4457–4469. <u>https://doi.</u> org/10.1111/ mec.12400.

Gilk, S.E., I.A. Wang, C.L. Hoover, W.W. Smoker, and S.G. Taylor. 2004. Outbreeding depression in hybrids between spatially separated pink salmon, *Oncorhynchus gorbuscha*, populations: marine survival, homing ability, and variability in family size. Environmental Biology of Fishes 69:287–297.

Gilk, S.E., W.D. Templin, D.B. Molyneaux, T. Hamazaki, and J.A. Pawluk. 2005. Characteristics of fall chum salmon Oncorhynchus keta in the Kuskokwim River drainage. Alaska Department of Fish and Game, Fishery Data Series No. 05-56, Anchorage.

Gilk, S.E., D.B. Molyneaux, T. Hamazaki, J.A. Pawluk and W.D. Templin. 2009. Biological and genetic characteristics of fall and summer chum salmon in the Kuskokwim River, Alaska. Pages 161-179 in Krueger, C.C. and C.E. Zimmerman, editors. Pacific Salmon: ecology and management of western Alaska's populations. American Fisheries Society Symposium 70, Bethesda, MD.

Gilk-Baumer, S., S.M. Turner, C. Habicht, and S.C. Heinl. 2013. Genetic stock identification of McDonald Lake sockeye salmon in selected Southeast Alaska fisheries, 2007-2009. Alaska Department of Fish and Game, Fishery Manuscript Series No. 13-04, Anchorage. Gilk-Baumer, S.E., S.D. Rogers Olive, D.K. Harris, S.C. Heinl, E.K. C. Fox, and W.D. Templin. 2015. Genetic mixed stock analysis of sockeye salmon harvests in selected northern Chatham Strait commercial fisheries, Southeast Alaska, 2012-2014. Alaska Department of Fish and Game, Fishery Data Series No. 15-03, Anchorage.

Gilk-Baumer et al. 2017. Genetic stock composition of the commercial harvest of Chinook salmon in Copper River District, 2013-2017. Alaska Department of Fish and Game, Fishery Manuscript Series No. 17-09, Anchorage.

Gisclair, B.R., 2009. Salmon bycatch management in the Bering Sea walleye pollock fishery: threats and opportunities in western Alaska. American Fisheries Society Symposium 70:799–816.

Graham, C.J., T.M. Sutton, M.D. Adkison, M.V. McPhee, and P.J. Richards. 2019. Evaluation of growth, survival, and recruitment of Chinook Salmon in Southeast Alaska Rivers. Transactions of the American Fisheries Society. 148:243–259. https://doi.org/10.1002/tafs.10148

Granath, K.L., W.W. Smoker, A.J. Gharrett, and J.J. Hard. 2004. Effects on embryo development time and survival of intercrossing three geographically separate populations of Southeast Alaska coho salmon *Oncorhynchus kisutch*. Environmental Biology of Fishes 69:299–306.

Gray, A.K., W.T. Mccraney, C.T. Marvin, C.M. Kondzela, H.T. Nguyen, and J.R. Guyon. 2011. Genetic Stock Composition Analysis of Chum Salmon Bycatch Samples from the 2007 Bering Sea Groundfish Fisheries. U.S. Department of Commerce, NOAA Technical Memorandum. NMFS-AFSC-221, 29 p.

Grebmeier, J.M., J.E. Overland, S.E. Moore, E.V. Farley, E.C. Carmack, L.W. Cooper, K.E. Frey, J.H. Helle, F.A. McLaughlin, and S.L. McNutt. 2006. A Major Ecosystem Shift in the Northern Bering Sea. Science 311:1461–1464.

Grebmeier, J.M., L.W. Cooper, H.M. Feder, and B.I. Sirenko. 2006. Ecosystem dynamics of the Pacific-influenced Northern Bering and Chukchi Seas in the Amerasian Arctic. Progress In Oceanography 71:331–361. https://doi.org/10.1016/j.pocean.2006.10.001

Griffiths, J.R. D.E. Schindler, L.S. Balistrieri, and G.T. Ruggerone. 2011. Effects of simultaneous climate change and geomorphic evolution on thermal characteristics of a shallow Alaskan lake. Limnology and Oceanography 56(1):193-205

Griffiths, J.R., D.E. Schindler, G.T. Ruggerone, and J.D. Bumgarner. 2014. Climate variation is filtered differently among lakes to influence growth of juvenile sockeye salmon in an Alaskan watershed. Oikos 0:1-12

Godduhn, A.R., D.M. Runfola, C.R. McDevitt, J. Park, G. Rakhmetova, J.S. Magdanz, H.S. Cold, and C.L. Brown. 2020. Patterns and trends of subsistence salmon harvest and use in the Kuskokwim River Drainage, 1990-2016. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 468.

Guthrie, C.M., R.L. Wilmot. 2004. Genetic structure of wild chinook salmon populations of Southeast Alaska and northern British Columbia. Environmental Biology of Fishes 69:81–93.

Guthrie, C., H. Nguyen, and J. Guyon. 2014. Genetic Stock Composition Analysis of Chinook Salmon Bycatch Samples from the 2012 Bering Sea and Gulf of Alaska Trawl Fisheries. NOAA Technical Memorandum 33.

Guthrie, C.M.G., H.T. Nguyen, M. Marsh, and J.R. Guyon. 2019. Genetic Stock Composition Analysis of Chinook Salmon Bycatch Samples from the 2017 Gulf of Alaska Trawl Fisheries. U.S. Department of Commerce, NOAA Technical Memorandum NMFS-AFSC-390, 30 p.

Guyon, J.R., C.M. Guthrie III, and H. Nguyen. 2010. Genetic stock composition analysis of Chinook salmon bycatch samples from the 2007 "B" season and 2009 Bering Sea trawl fisheries. Report to the North Pacific Fishery Management Council.

Guyon, J.R., C. Kondzela, T. McCraney, C. Marvin, and E. Martinson. 2010. Genetic Stock Composition Analysis of Chum Salmon Bycatch Samples from the 2005 Bering Sea Groundfish Fisheries, Report to the North Pacific Fishery Management Council.

Η

Habicht, C., L.W. Seeb, and J.E. Seeb. 2007. Genetic and ecological divergence defines population structure of sockeye salmon populations returning to Bristol Bay, Alaska, and provides a toold for admixture analysis. Transactions of the American Fisheries Society 136:82-94. DOI: 10.1577/T06-001.1

Habicht, C., L.W. Seeb, K.W. Myers, E.V. Farley Jr., and J.E. Seeb. 2010. Summer-Fall distribution of stocks of immature sockeye salmon in the Bering Sea as revealed by single-nucleotide polymorphisms. Transactions of the American Fisheries Society 139(4):1171-1191.

Hagerman, G., R. Ehresmann, and L. Shaul. 2018. Annual Management Report for the 2017 Southeast Alaska/Yakutat Salmon Troll Fisheries. Alaska Department of Fish and Game, Fishery Management Report No. 18-02, Anchorage.

Hagerman, G., M. Vaughn, and J. Priest. 2021. Annual management report for the 2020 Southeast Alaska/Yakutat salmon troll fisheries. Alaska Department of Fish and Game, Fishery Management Report No. 21-17.

Hagerman, G.T., D.K. Harris, J.T. Williams, D.J. Teske, B.W. Elliott, N.L. Zeiser, and R.S. Chapell. 2022. Northern Southeast Alaska Chinook salmon stock status and action plan, 2022. Alaska Department of Fish and Game, Regional Information Report No. 1J22-17, Douglas, Alaska.

Halas, G. and M. Cunningham. 2019. Nushagak River Chinook Salmon: Local and Traditional Knowledge and Subsistence Harvests. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 453, Anchorage.

Halupka, K.C., M.F. Willson, M.D. Bryant, F.H. Everest, and A.J. Gharrett. 2003. Conservation of population diversity of Pacific salmon in southeast Alaska. North American Journal of Fisheries Management 23:1057–1086.

Hamazaki, T., M. Evenson, S.J. Fleischman, and K.L. Schaberg. 2012. Spawner-recruit analysis and escapement goal recommendation for Chinook salmon in the Kuskokwim River drainage. Alaska Department of Fish and Game, Fishery Manuscript Series No. 12-08, Anchorage.

Hamazaki, T. and N. DeCovich. Application of the genetic mark-recapture technique for run size estimation of Yukon River Chinook salmon. 2014. North American Journal of Fisheries Management 34:276-286.

Hamilton, K., and L.A. Mysak. 1986. Possible Effects of the Sitka Eddy on Sockeye Oncorhynchus nerka) and Pink Salmon (*Oncorhynchus gorbuscha*) Migration off Southeast Alaska. Canadian Journal of Fisheries and Aquatic Sciences 43:498–504.

Hard, J.J., and W.R. Heard. 1999. Analysis of straying variation in Alaskan hatchery chinook salmon (*Oncorhynchus tshawytscha*) following transplantation. Canadian Journal of Fisheries and Aquatic Sciences 56:578–589. https://doi.org/10.1139/f98-199.

Hard, J.J. 2004. Evolution of Chinook salmon life history under size-selective harvest. In A. Hendry and S. Sterns (eds), Evolution Illuminated: Salmon and their relatives, p. 315 – 337. Oxford, NY.

Hard, J.J., M.R. Gross, M. Heino, R. Hilborn, R.G. Kope, R. Law, and J.D. Reynolds. 2008. Evolutionary consequences of fishing and their implications for salmon. Evolutionary Applications 1(2):388-408. doi:1111.j.1752-4571.2008.00020.x.

Haynie, A.C., and H.P. Huntington. 2016. Strong connections, loose coupling: the influence of the Bering Sea ecosystem on commercial fisheries and subsistence harvests in Alaska. Ecology and Society 21. https://doi.org/10.5751/ES-08729-210406.

Heard, W.R., 1978. Probable case of streambed overseeding - 1967 pink salmon, *Oncorhynchus gorbuscha*, spawners and survival of their progeny in Sashin Creek, Southeastern Alaska. Fishery Bulletin 76:569–582.

Heard, W.R., 1998. Do hatchery salmon affect the North Pacific Ocean ecosystem? North Pacific Anadromous Fish Commission Bulletin 1:405–411.

Heard, W.R., J.A. Orsi, A.C. Wertheimer, M.V. Sturdevant, J.M. Murphy, D.G. Mortensen, B.L.Wing, and A.G. Celewycz. 2001. A synthesis of research on early marine ecology of juvenile Pacific salmon in southeast Alaska. North Pacific Anadromous Fisheries Commission Technical Report 2:3–6.

Heard, W.R., 2012. Overview of salmon stock enhancement in southeast Alaska and compatibility with maintenance of hatchery and wild stocks. Environmental Biology of Fishes 94:273–283. https://doi.org/10.1007/s10641-011-9855-6

Heard, W.R., S.G. Taylor, and J.A. Orsi. 2013. Survival and Early Marine Migration of Enhanced Age-0 Sockeye Salmon Smolts Raised in Freshwater and Seawater at Auke Creek, Southeast Alaska. North Pacific Anadromous Fish Commission, Technical Report 9:235–238.

Hebert, K.P., P.L. Goddard, W.W. Smoker, and A.J. Gharrett. 1998. Quantitative genetic variation and genotype by environment interaction of embryo development rate in pink salmon (*Oncorhynchus gorbuscha*). Canadian Journal of Fisheries and Aquatic Sciences 55:2048–2057.

Heinl, S.C., J.F. Koerner, and D.J. Blick. 2000. Portland Canal chum salmon coded wire tagging project, 1988-1995. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report No. 1J00-16, Juneau.

Heinl, S.C., and A.W. Piston. 2009. Standardizing and automating the Southeast Alaska pink salmon escapement index. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report No. 1J09-06, Douglas.

Heinl, S.C., R.L. Bachman, and K. Jensen. 2011. Sockeye salmon stock status and escapement goals in Southeast Alaska. Alaska Department of Fish and Game, Special Publication No. 11-20, Anchorage.

Henry, E., 2021. Redoubt Lake Sockeye Monitoring and Lake Fertilization 2021 Annual Report. U.S. Department of Agriculture, Forest Service, Sitka, Alaska.

Hillgruber, N., and C.E. Zimmerman. 2009. Estuarine Ecology of Juvenile Salmon in Western Alaska: A Review. Pages 183–199. American Fisheries Society.

Hiroko I., A.R. Brenner, and A. Godduhn. 2013. Socioeconomic patterns in subsistence salmon fisheries: historical and contemporary trends in five Kuskokwim River communities and overview of the 2012 season. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 382.

Hoffman, R., and T. Thynes. 2022. Klukshu River sockeye salmon stock status and action plan, 2022. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 1J22-10, Douglas

Hoffman, S.H. 1982. Northern Southeastern Alaska pink salmon (*Oncorhynchus gorbuscha*) tagging investigations, 1977-1980. Alaska Department of Fish and Game, Division of Commercial Fisheries, Information Leaflet No. 196, Juneau.

Hoffman, S.H. 1983. Southern Southeastern Alaska pink salmon (*Oncorhynchus gorbuscha*) tagging investigations, 1981. Alaska Department of Fish and Game, Division of Commercial Fisheries, Technical Data Report No. 92, Juneau. Hoffman, S.H., L. Talley, and M.C. Seibel. 1983. 1982 U.S./Canada research pink and sockeye salmon tagging, interception rates, migration patterns, run timing, and stock intermingling in southern Southeast Alaska and Northern British Columbia. [in]: Final Report 1982 salmon research conducted in Southeast Alaska by the Alaska Department of Fish and Game in conjunction with joint U.S.-Canada Interception investigations. Contract No. NASO-82-00134.

Hoffman, S.H., L. Talley, and M.C. Seibel. 1984. U.S./ Canada cooperative pink and sockeye salmon tagging, interception rates, migration pattern, run timing, and stock intermingling research in southern Southeast Alaska and northern British Columbia, 1982. Alaska Department of Fish and Game Technical Data Report No. 110.

Hoffman, S.H., L. Talley, and M.C. Seibel. 1985. 1984 pink and chum salmon tagging, national contribution rates, migration patterns, run timing, and stock intermingling research in southern Southeast Alaska and northern British Columbia [in]: Final Report. 1984 salmon research conducted in Southeast Alaska by the Alaska Department of Fish and Game in conjunction with National Marine Fisheries Service Auke Bay Laboratory for joint U.S.-Canada Interception Studies. Contract No. WASC-84-00179.

Holen, D., J.A. Fall, and R. La Vine. 2011. Customary and traditional use worksheet: salmon, Copper River District, Prince William Sound Management Area. Alaska Department of Game Division of Subsistence Special Publication No. BOF 2011-06. Anchorage.

Holen, D., S.M. Hazell, and G. Zimpelman, editors. 2015. The Harvest and Use of Wild Resources in Selected Communities of the Copper River Basin and East Glenn Highway, Alaska, 2013. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 405. Anchorage.

Hollowed, A.B., S.J. Barbeaux, E.D. Cokelet, E. Farley, S. Kotwicki, P.H. Ressler, C. Spital, and C.D. Wilson. 2012. Effects of climate variations on pelagic ocean habitats and their role in structuring forage fish distributions in the Bering Sea. Deep Sea Research Part II: Topical Studies in Oceanography 65–70:230–250. https://doi. org/10.1016/j.dsr2.2012.02.008

Hollowell, G., E.O. Otis, and E. Ford. 2023. 2022 Lower Cook Inlet area salmon annual management report. Alaska Department of Fish and Game, Fishery Management Report No. 23-04, Anchorage.

Honea, J.M., J.C. Jorgensen, M.M. McClure, T.D. Cooney, K. Engie, D. Holzer, and R. Hilborn. 2009. Evaluating habitat effects on population status: influence of habitat restoration on springrun Chinook salmon. Freshwater Biology 54(7):1576-1592. doi:10.1111/j.1365-2427.2009.02208.x

Horne-brine, M.H., D. Warnke, and L. Dubois. 2011. Salmon Age and Sex Composition and Mean Lengths for the Yukon River Area, 2009. Alaska Department of Fish and Game, Fishery Data Series No. 11-16

Howard, K.G., S.J. Hayes, and D.F. Evenson. 2009. Yukon River Chinook salmon stock status and action plan 2010; a report to the Alaska Board of Fisheries. Alaska Department of Fish and Game, Anchorage.

Howard, K.G., and D.F. Evenson. 2010. Yukon River Chinook salmon comparative mesh size study. Alaska Department of Fish and Game, Fishery Data Series No. 10-92

Howard, K.G., J.M. Murphy, L.I. Wilson, J.H. Moss, and E.V. Farley, Jr. 2016. Size-selective mortality of Chinook salmon in relation to body energy after the first summer in nearshore marine habitats. North Pacific Anadromous Fish Commission Bulletin 6:1-11.

Howard, K.G., K.M. Miller, and J. Murphy. 2017. Estuarine Fish Ecology of the Yukon River Delta, 2014–2015. Alaska Department of Fish and Game, Fishery Data Series No. 17-16, Anchorage.

Howard, K.G., and V. von Biela. 2023. Adult spawners: a critical period for subarctic Chinook salmon in a changing climate. Global Change Biology 29:1759-1773. doi: 10.1111/gcb.16610

Howe, E.L., and S. Martin. 2009. Demographic Change, Economic Conditions, and Subsistence Salmon Harvests in Alaska's Arctic-Yu-kon-Kuskokwim Region. American Fisheries Society Symposium 70:433–461.

Howe, N.S., M.C. Hale, C.D. Waters, S.M. Schaal, K.R. Shedd, and W.A. Larson. 2024. Genomic evidence form domestication selection in three hatchery populations of Chinook salmon, *Oncorhynchus tshawytscha*. Evolutionary Applications 17(2):e13656.

Hunt Jr, G.L., P. Stabeno, G. Walters, E. Sinclair, R.D. Brodeur, J.M. Napp, and N.A. Bond. 2002. Climate change and control of the southeastern Bering Sea pelagic ecosystem. Deep Sea Research Part II: Topical Studies in Oceanography 49:5821–5853. https://doi.org/10.1016/S0967-0645(02)00321-1

Hunt, G.L., K.O. Coyle, L.B. Eisner, E.V. Farley, R.A. Heintz, F. Mueter, J.M. Napp, J.E. Overland, P.H. Ressler, S. Salo, and J. Stabeno. 2011. Climate impacts on eastern Bering Sea foodwebs : a synthesis of new data and an assessment of the Oscillating Control Hypothesis. ICES Journal of Marine Science 68:1230–1243.

Hunt, G.L., P.H. Ressler, G.A. Gibson, A. De Robertis, K. Aydin, M.F. Sigler, I. Ortiz, E.J. Lessard, B.C. Williams, A. Pinchuk, and T. Buckley. 2015. Euphausiids in the eastern Bering Sea: A synthesis of recent studies of euphausiid production, consumption and population control. Deep-Sea Research Part II: Topical Studies in Oceanography 134:204–222. https://doi.org/10.1016/j. dsr2.2015.10.007

Huntington, H.P., S.L. Danielson, F.K. Wiese, M. Baker, P. Boveng, J.J. Citta, A. De Robertis, D.M. S. Dickson, E. Farley, J.C. George, K. Iken, D.G. Kimmel, K. Kuletz, C. Ladd, R. Levine, L. Quakenbush, P. Stabeno, K.M. Stafford, D. Stockwell, and C. Wilson. 2020. Evidence suggests potential transformation of the Pacific Arctic ecosystem is underway. Nature Climate Change 10:342-348. <u>https://doi.org/10.1038/s41558-020-0695-2</u>. Hutchinson-Scarbrough, L. and D. Koster. 2021. Subsistence Harvest Assessment of Salmon and Local Traditional Knowledge of Chinook Salmon in the Chignik Management Area, 2014–2016. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 462, Anchorage

Hutchinson-Scarbrough, L., D. Gerkey, G. Halas, C. Larson, L. A. Sill, J.M. Van Lanen, and M. Cunningham. 2020. Subsistence Salmon Networks in Select Bristol Bay and Alaska Peninsula Communities, 2016. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 459, Anchorage

Hyatt, K.D., and K.L. Mathias. 2005. Evaluation of hatchery versus wild sockeye salmon fry growth and survival in two British Columbia Lakes. North American Journal of Fisheries Management 25:745-762.

Hyer, K.E., and C.J. Schleusner. 2005. Chinook salmon age, sex, and length analysis from selected escapement projects on the Yukon River. Alaska Department of Fish and Game, Alaska Fisheries Technical Report Number 87.

lanelli, J.N. and D.L. Stram. 2015. Estimating impacts of the pollock fishery bycatch on western Alaska Chinook salmon. ICES Journal of Marine Science 72(4):1159-1172.

lanelli, J.N., J. Gauvin, D.L. Stram, K. Haflinger, and P. Stabeno. 2010. Temperature/depth data collections on Bering Sea groundfish vessels to reduce bycatch. North Pacific Research Board Final Report 731, 58 p.

Inman, S.C., J. Esquible, M.L. Jones, W.R. Bechtol, and B. Connors. 2021. Opportunities and impediments for use of local data in the management of salmon fisheries. Ecology and Society 26(2):26.

Irvine, J.R., R.W. Macdonald, R.J. Brown, L. Godbout, J.D. Reist, and E.C. Carmack. 2009. Salmon in the Arctic and how they avoid lethal low temperatures. North Pacific Anadromous Fish Comm Bulletin 5:39–50.

Ishida, Y., T. Azumaya, M. Fukuwaka, and N. Davis. 2002. Interannual variability in stock abundance and body size of Pacific salmon in the central Bering Sea Interannual variability in stock abundance and body size of Pacific salmon in the central Bering Sea. Progress in Oceanography 55:223–234. <u>https://doi.org/10.1016/</u>S0079-6611(02)00080-0

J

Jaenicke, H.W., A.G. Celewycz, J.E. Bailey, and J.A.Orsi. 1985. Paired open beach seines to study estuarine migrations of juvenile salmon. Marine Fisheries Review 46:62–67.

Jaenicke, H.W., and A.G. Celewycz. 1994. Marine distribution and size of juvenile Pacific salmon in southeast Alaska and northern British Columbia. Fishery Bulletin 92:79–90.

Jalbert, C.S., S.C. Heinl, A. Reynolds Manney, and K.R. Shedd. 2022. Commercial harvest of Klawock Lake sockeye salmon in the District 103 and 104 purse seine fisheries, Southeast Alaska, 2018–2021. Alaska Department of Fish and Game, Fishery Data Series No. 22-34, Anchorage.

Jallen, D.M., S.K.S. Decker, and T. Hamazaki. 2017. Subsistence and personal use salmon harvests in the Alaska portion of the Yukon River drainage, 2013. Alaska Department of Fish and Game, Fishery Data Series No. 17-08, Anchorage.

Jasper J.R., C. Habicht, S. Moffitt, R. Brenner, J. Marsh, B. Lewis, E. Creelman Fox, Z. Grauvogel, S.D. Rogers Olive, and W.S. Grant. 2013. Source-sink estimates of genetic introgression show influence of hatchery strays on wild chum salmon populations in Prince William Sound, Alaska. PLoS ONE 8(12):e81916.

Jasper, J.R., N. DeCovich, and W.D. Templin. 2012. Western Alaska Salmon Stock Identification Program Technical Document 4: Status of the SNP baseline for chum salmon. (PDF 1,013 kB) Alaska Department of Fish and Game, Regional Information Report 5J12-09, Anchorage

Jasper, J.R., and D.F Evenson. 2006. Length-girth, length-weight, and fecundity of Yukon River Chinook salmon Oncorhynchus tshawytscha., Alaska Department of Fish and Game, Fishery Data Series No. 06-70, Anchorage.

Johnson, G., C. Kondzela, J. Whittle, K. Miller, and J. Guyon. 2019. Genetic Characterization of Juvenile Chum Salmon (*Oncorhynchus keta*) Migrating out of the Yukon River Delta. Technical Report 51–53. <u>https://doi.org/10.23849/npafctr15/51.53</u>.

Johnson, G.C., and P.J. Stabeno. 2004. The Bering Slope Current system revisited. Journal of Physical Oceanography 34:384– 398. https://doi.org/10.1175/1520-0485(2004)034<0384 :TBSCSR>2.0.CO;2

Johnson, S.W., M.L. Murphy, D.J. Csepp, P.M. Harris, and J.F. Thedinga. 2003. A survey of fish assemblages in eelgrass and kelp habitats of southeastern Alaska. U. S. Department of Commerce, NOAA Technical Memorandum. NMFS-AFSC-139, 39 p.

Johnson, S.W., and J.F. Thedinga. 2005. Fish use and size of eelgrass meadows in southeastern Alaska: a baseline for long-term assessment of biotic change. Northwest Science 79:141–155.

Johnson, T.A., S.C. Heinl, and H.J. Geiger. 2005. McDonald Lake: Stock status report. Alaska Department of Fish and Game, Fishery Manuscript No. 05-07, Anchorage.

Jones, B., and M. Kukkonen. 2017. Local and Traditional Knowledge of Abundance of Chinook Salmon in the Kenai River. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 431, Anchorage

Jones, B.E. and D. Koster. 2018. Subsistence Harvests and Uses of Salmon in Tyonek, 2015 and 2016. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 439, Anchorage. Jones, B., M. Cunningham, and D. Koster. 2019. Subsistence Harvest Assessment and Biological Sampling of Chinook Salmon in the Togiak River Drainage. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 454, Anchorage.

Jones, B., and M. Cunningham. 2020. The Harvest and Use of Salmon by Residents of King Salmon, Naknek, and South Naknek, Alaska, 2017 and 2018. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 470, Anchorage.

Jones, B., and M. Cunningham. 2020. The Harvest and Use of Wild Resources in Port Heiden, Alaska, 2018. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 465: Anchorage.

Jones, E.L., T.J. Quinn, , and B.W. Van Alen. 1998. Observer Accuracy and Precision in Aerial and Foot Survey Counts of Pink Salmon in a Southeast Alaska Stream. North American Journal of Fisheries Management 18:832–846. https://doi.org/10.1577/1548-8675(1 998)018<0832:OAAPIA>2.0.CO;2

Jorgensen, J.C., C. Nicol, C. Fogel, and T.J. Beechie. 2021. Identifying the potential of anadromous salmonid habitat restoration with life cycle models. PLOS One 16(9): e0256792. <u>https://doi.org/10.1371/journal.pone.0256792</u>

Josephson, R., A. Wertheimer, D. Gaudet, E. Knudsen, B. Adams, D. Bernard, S. Heinl, A. Piston, and W. Templin. 2021. Proportions of Hatchery Fish in Escapements of Summer-Run Chum Salmon in Southeast Alaska, 2013-2015. North American Journal of Fisheries Management 41:724-738.

Joy, P. J., S.B. Haught, R.E. Brenner, S. Miller, J.W. Erickson, J.W. Savereide, and T.R. McKinley. 2021. Escapement goal review of Copper and Bering Rivers and Prince William Sound Pacific salmon stocks, 2020. Alaska Department of Fish and Game, Fishery Manuscript No. 21-02, Anchorage.

Joy, P., J.W. Savereide, M. Tyers, and S.J. Fleischman. 2021. Run reconstruction, spawner–recruit analysis, and escapement goal recommendation for Chinook salmon in the Copper River. Alaska Department of Fish and Game, Fishery Manuscript No. 21-01, Anchorage.

Κ

Kaeriyama, M., M. Nakamura, R. Edpalina, J.R. Bower, H. Yamaguchi, R.V. Walker, and K.W. Myers. 2004. Change in feeding ecology and trophic dynamics of Pacific salmon (*Oncorhynchus* spp.) in the central Gulf of Alaska in relation to climate events. Fisheries Oceanography 13(3):197-207.

Kaga, T., S. Sato, T. Azumaya, N.D. Davis, and M. Fukuwaka. 2013. Lipid content of chum salmon *Oncorhynchus keta* affected by pink salmon *O. gorbuscha* abundance in the central Bering Sea. Marine Ecology Progress Series 478:211–221. <u>https://doi.org/10.3354/</u> meps10179.

Kaplan, I.C., S.K. Gaichas, C.C. Stawitz, P.D. Lynch, K.N. Marshall, J.J. Deroba, M. Masi, J.K.T. Brodziak, K.Y. Aydin, K. Holsman, H. Townsend, D. Tommasi, J.A. Smith, S. Koenigstein, M. Weijerman, and J. Link. 2021. Management strategy evaluation: allowing the light on the hill to illuminate more than one species. Frontiers in Marine Science doi:10.3389/fmars.2021.624355.

Karpenko, V.I., A.F. Volkov, and M.V. Koval. 2007. Diets of Pacific salmon in the Sea of Okhotsk, Bering Sea, and Northwest Pacific Ocean. North Pacific Anadromous Fish Commission Bulletin 4:105–116.

Keating, J.M., D. Koster, and J.M. Van Lanen. 2020. Recovery of a Subsistence Way of Life: Assessments of Resource Harvests in Cordova, Chenega, Tatitlek, Port Graham, and Nanwalek, Alaska since the Exxon Valdez Oil Spill. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 471, Anchorage.

Khen, G.V., A.V. Zavolokin, Г.B. Хен, and A.B. Заволокин. 2015. Change in the water circulation and its implications for distribution and abundance of salmons in the western Bering Sea. TINRO News 95–115.

Kishi, M.J., M. Kaeriyama, H. Ueno, and Y. Kamezawa. 2010. The effect of climate change on the growth of Japanese chum salmon (*Oncorhynchus keta*) using a bioenergetics model coupled with a three-dimensional lower trophic ecosystem model (NEMURO). Deep-Sea Research Part II-Topical Studies in Oceanography 57:1257–1265. https://doi.org/10.1016/j.dsr2.2009.12.013.

Kitada, S., and H. Kishino. 2023. Effects of climate, release practices, predation, and competition on declining hatchery-enhanced chum salmon. bioRxiv. doi: https://doi.org/10.1101/2023.09.18.558187

Kline, T.C., J.J. Goering, O.A. Mathison, P.H. Poe, and P.L. Parker. 1990. Recycling of elements transported upstream by runs of Pacific salmon: I. 15-N and 13-C evidence in Sashin Creek, southeastern Alaska. Canadian Journal of Fisheries and Aquatic Sciences 47:136–144.

Knudsen, E.E., P.S. Rand, K.B. Gorman, D.R. Bernard, and W.D. Templin. 2021. Hatchery-origin stray rates and total run characteristics for Pink Salmon and Chum Salmon returning to prince william sound, Alaska, in 2013–2015. Marine and Coastal Fisheries, 13(1):41-68.

Kocan, R., and P. Hershberger. 2006. Differences in Ichthyophonous prevalence and infection severity between upper Yukon River and Tanana River chinook salmon, *Oncorhynchus tshawytscha* (Walbaum) stocks. Journal of Fish Diseases 29:497–503.

Kocan, R., P. Hershberger, and J. Winton. 2004. Ichthyophoniasis: An emerging disease of Chinook salmon in the Yukon River. Journal of Aquatic Animal Health, 16(2):58-72, doi: 10.1577/H03-068.1.

Kocan R., P. Hershberger, G. Sanders, and J. Winton. 2009. Effects of temperature on disease progression and swimming stamina in Ichthyophonus-infected rainbow trout, *Oncorhynchus mykiss* Walbaum. Journal of Fish Diseases 32:835–843.

Kohan, M.L., F.J. Mueter, J.A. Orsi, and M.V. McPhee. 2017. Variation in size, condition, and abundance of juvenile chum salmon (*Oncorhynchus keta*) in relation to marine factors in Southeast Alaska. Deep Sea research Part II, <u>https://doi.org/10.1016/j</u>. dsr2.2017.09.005

Kondzela, C.M., C.M. Guthrie, S.L. Hawkins, C.D. Russell, and J.H. Helle. 1994. Genetic relationships among chum salmon populations in southeast Alaska and northern British Columbia. Canadian Journal of Fisheries and Aquatic Sciences 51(Suppl. 1):50-64.

Kondzela, C.M., M. Garvin, R. Riley, J. Murphy, J. Moss, S. Adam Fuller, and A. Gharrett. 2009. Preliminary genetic analysis of juvenile Chum salmon from the Chukchi Sea and Bering Strait. North Pacific Anadromous Fish Commission Bulletin 5:25-27.

Kondzela, C.M., J.A. Whittle, C.T. Marvin et al. 2016. Genetic analysis identifies consistent proportions of seasonal life history types in Yukon River juvenile and adult Chum salmon. North Pacific Anadromous Fish Commission Bulletin 6:439-450

Kondzela, C.M., J.A. Whittle, D. Yates, S.C. Vulstek, H.T. Nguyen, and J.R. Guyon. 2016. Genetic stock composition analysis of Chum Salmon from the prohibited species catch of the 2014 Bering Sea Walleye Pollock trawl fishery and Gulf of Alaska groundfish fisheries. U. S. Department of Commerce, NOAA Tech Memo. https://doi. org/10.7289/V5/TM-AFSC-314

Kondzela, C.M., J.A.Whittle, S.C. Vulstek, H.T. Nguyen, and J.R. Guyon. 2017. Genetic stock composition analysis of Chum Salmon from the prohibited species catch of the 2015 Bering Sea Walleye Pollock trawl fishery and Gulf of Alaska groundfish fisheries. 49. https://doi.org/10.7289/V5/TM-AFSC-314

Kovach, R.P., A.J. Gharrett, and D.A. Tallmon. 2015. Genetic change for earlier migration timing in a pink salmon population. Proceedings of the Royal Society B: Biological Sciences 279:3870-3878. doi:10.1098/rspb.2012.1158.

Kovach, R.P., S.C. Ellison, S. Pyare, and D.A. Tallmon. 2015. Temporal patterns in adult salmon migration timing across Southeast Alaska. Global Change Biology 2:1821–1833.

Krkosek, M., R. Hilborn, R.M. Peterman, and T. Quinn. 2011. Cycles, stochasticity and density dependence in pink salmon population dynamics. Proceedings of the Royal Society B: Biological Sciences 278:2060–2068.

Krueger, C.C., and C.E. Zimmerman, editors. 2009. Pacific salmon: ecology and management of western Alaska's populations. American Fisheries Society, Symposium 70, Bethesda, Maryland.

L

La Vine, R., M. Kukkonen, B. Jones, and G. Zimpelman. 2013. Subsistence harvests and uses of wild resources in Copper Center, Slana/Nabesna Road, Mentasta Lake, and Mentasta Pass, Alaska, 2010. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 380. Anchorage, Alaska.

LaCroix, J.J., A.C. Wertheimer, J.A. Orsi, M.V. Sturdevant, E.A.Fergusson, and N.A. Bond. 2009. A top-down survival mechanism during early marine residency explains coho salmon year-class strength in southeast Alaska. Deep-Sea Research Part II: Topical Studies in Oceanography 56:2560–2569. <u>https://doi.org/10.1016/j</u>. dsr2.2009.03.006

Ladd, C., 2014. Seasonal and interannual variability of the Bering Slope Current. Deep Sea Research Part II: Topical Studies in Oceanography 109:5–13. https://doi.org/10.1016/j.dsr2.2013.12.005

Landingham, J., M. Sturdevant, and R. Brodeur. 1998. Feeding habits of juvenile Pacific salmon in marine waters of southeastern Alaska and northern British Columbia. Fishery Bulletin 96:285–302.

Langdon, S.J. 2006. Traditional knowledge and harvesting of salmon by Huna and Hinya Tlingit. US Fish and Wildlife Service, Office of Subsistence Management, Fisheries Resource Monitoring Program, Final Report (Project No. 02-104), Anchorage.

Langdon, S.J., 2006. Tidal pulse fishing: Selective traditional Tlingit salmon fishing techniques on the west coast of the Prince of Wales Archipelago., in: Traditional Ecological Knowledge and Natural Resource Management. University of Nebraska Press, pp. 21–46.

Langdon, S.J., 2007. Sustaining a relationship: inquiry into the emergence of a logic of engagement with salmon among the Southern Tlingits, in: Native Americans and the Environment: Perspectives on the Ecological Indian. University of Nebraska Press.

Langdon, S.J., 2018. Approaching Leviathan: Efforts to Establish Small-Scale, Community Based Commercial Salmon Fisheries in Southeast Alaskan Indigenous Communities, in: Fisheries, Quota Management and Quota Transfer, MARE Publication Series. Springer International Publishing, pp. 197–214.

Langdon, S. 2021. K'iis Xaadas relations with sockeye salmon: contemporary efforts at constructing a neo-traditional regime of stewardship. Maritime Studies 20:157-173.

Larsen, C.F., R.J. Motyka, J.T. Freymueller, K.A. Echelmeyer, and E.R. Ivins. 2005. Rapid viscoelastic uplift in southeast Alaska caused by post-Little Ice Age glacial retreat. Earth and Planetary Science Letters 237:548–560.

Larsen, C.F., R.J. Motyka, A.A. Arendt, K.A. Echelmeyer, and P.E. Geissler. 2007. Glacier changes in southeast Alaska and northwest British Columbia and contribution to sea level rise. Journal of Geophysical Research, Vol. 112, F01007, doi:10.1029/2006JF000586.

Larson, W.A., F.M. Utter, K.W. Myers, W.D. Templin, J.E. Seeb, C.M. Guthrie III, A.V. Bugaev, and L.W. Seeb. 2013. Single-nucleotide polymorphisms reveal distribution and migration of Chinook salmon (*Oncorhynchus tshawytscha*) in the Bering Sea and North Pacific Ocean. Canadian Journal of Fisheries and Aquatic Sciences 70:128-141.

Larson W.A., L.W. Seeb, M.V. Everett, R.K. Waples, W.D. Templin, and J.E. Seeb. 2014. Genotyping by sequencing resolves shallow population structure to inform conservation of Chinook salmon (*Oncorhynchus tshawytscha*). Evolutionary Applications 7(3):355-369.

Larson, W.A., J.E. Seeb, C.E. Pascal, W.D. Templin, and L.W. Seeb. 2014. Single-nucleotide polymorphisms (SNPs) identified through genotyping-by-sequencing improve genetic stock identification of Chinook salmon (*Oncorhynchus tshawytscha*) from western Alaska. Canadian Journal of Fisheries and Aquatic Sciences 71(5):698-708.

LaVine, R., M.J. Lisac and P. Coiley-Kenner. 2007. Traditional ecological knowledge of 20th century ecosystems and fish populations in the Kuskokwim Bay Region. U.S. Fish and Wildlife Service, Office of Subsistence Management, Fisheries Resource Monitoring Program (Project no. FIS 04 – 351) Anchorage, Alaska.

Lee, E., T. Dann, and H. Hoyt. 2021. Yukon River Chinook Genetic Baseline Improvements. Yukon River Panel Restoration and Enhancement Fund Final Report, URE-163-19N.

Leon, J.M., and M.V. McPhee., 2013. Freshwater Growth and Recruitment in Two Western Alaskan Populations of Chinook Salmon. North Pacific Anadromous Fish Commission Technical Report 9:229–231.

Levi, T., J.M. Allen, D. Bell, J. Joyce, J.R. Russell, D.A. Tallmon, S.C. Vulstek, C. Yang, D.W. Yu. 2019. Environmental DNA for the enumeration and management of Pacific salmon. Molecular Ecology Resources 19:597–608. https://doi.org/10.1111/1755-0998.12987

Lewis B., W.S. Grant, R.E. Brenner, and T. Hamazaki. 2015. Changes in size and age of Chinook salmon *Oncorhynchus tshawytscha* returning to alaska. PLoS ONE 10(6): e0130184. 17 pp. doi:10.1371/ journal.pone.0130184.

Lewis, B.A., and T.P. Zadina. 2002. The history of the subsistence and commercial fisheries, stock assessment and enhancement activities, and watershed disturbances in the Klawock Lake drainage on Prince of Wales Island. Alaska Department of Fish and Game, Regional Information Report No. 1J01-39 Revised.

Liller, Z.W., A.R. Brodersen, and J.N. Clark. 2013. Salmon Age, Sex, and Length Catalog for the Kuskokwim Area, 2010 and 2011. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 3A13-01, Anchorage.

Litzow, M.A., F.J. Mueter, and A.J. Hobday. 2014. Reassessing regime shifts in the North Pacific: incremental climate change and commercial fishing are necessary for explaining decadal-scale biological variability. Global Change Biology 20:38–50. https://doi. org/10.1111/gcb.12373 Litzow, M.A., 2017. Indications of hysteresis and early warning signals of reduced community resilience during a Bering Sea cold anomaly. Marine Ecology Progress Series 571:13–28.

Loewen, M., and L. Henslee. 2017. The 2016 Chignik River sockeye salmon smolt outmigration. Alaska Department of Fish and Game, Fishery Data Series No. 17-11, Anchorage.

Loring, P.A., and C. Gerlach. 2010. Food Security and Conservation of Yukon River Salmon: Are We Asking Too Much of the Yukon River? Sustainability 2:2965–2987. https://doi.org/10.3390/su2092965

Lum, J.L., and L. Fair. 2018. Chilkat River and King Salmon River king salmon stock status and action plan, 2018. Alaska Department of Fish and Game, Regional Information Report No. 1J18-05, Douglas.

Μ

MacNeil, M.A., N.A.J. Graham, J.E. Cinner, N.K. Dulvy, P.A. Loring, S. Jennings, N.V.C. Polunin, A.T. Fisk, and T.R. McClanahan. 2010. Transitional states in marine fisheries: adapting to predicted global change. Philosophical Transactions of the Royal Society B: Biological Sciences 365:3753–3763.

Magdanz, J.S. 1981. Northern Bering Sea subsistence report. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 4.

Magdanz, J.S. and C. Utermohle. 1997. The subsistence salmon fishery in the Norton Sound, Port Clarence and Kotzebue Districts, 1994. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 237, Juneau.

Magdanz, J.S., S. Tahbone, K.R. Kamletz. and A. Ahmasuk. 2005. Subsistence salmon fishing by residents of Nome, Alaska, 2001. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 274, Juneau.

Magdanz, J.S., E Trigg, A. Ahmasuk, P. Nanouk, D.S. Koster and K.R. Kamletz. 2005. Patterns and trends in subsistence salmon harvests, Norton Sound and Port Clarence, 1994-2003. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 294, Juneau.

Magdanz, J.S., S. Tahbone, A. Ahmasuk, D.S. Koster and B.L. Davis. 2007. Customary trade and barter in subsistence fish in the Seward Peninsula area, Alaska. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 328, Juneau.

Magdanz, J.S., H. Smith, N. Braem and P. Fox and D.S. Koster. 2011. Patterns and trends in subsistence fish harvests, Northwest Alaska, 1994-2004. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 366, Kotzebue.

Mahnken, C., G. Ruggerone, W. Waknitzl, and T. Flagg. 1998. A Historical Perspective on Salmonid Production from Pacific Rim Hatcheries. North Pacific Anadromous Fish Commission Bulletin 1:38–53. Malick, M.J., M.D. Adkison, and A.C. Wertheimer. 2009. Variable effects of biological and environmental processes on coho salmon survival in southeast Alaska. Transactions of the American Fisheries Society 138: 846-860.

Malison, R.L., K.V. Kuzishchin, and J.A. Stanford. 2016. Do beaver dams reduce habitat connectivity and salmon productivity in expansive river floodplains? PeerJ 4:e2403; doi/ 10.7717/peerj.2403

Manhard, C.V., J.E. Joyce, W.W. Smoker, and A.J. Gharrett. 2016. Ecological factors influencing lifetime productivity of pink salmon (*Oncorhynchus gorbuscha*) in an Alaskan stream. Canadian Journal of Fisheries and Aquatic Sciences 74:1325–1336.

Manhard, C.V., J.E. Joyce, and A.J. Gharrett. 2017. Evolution of phenology in a salmonid population: a potential adaptive response to climate change. Canadian Journal of Fisheries and Aquatic Sciences 74:1519–1527. https://doi.org/10.1139/cjfas-2017-0028.

Manhard, C.V., J.E. Joyce, W.W. Smoker, and A.J. Gharrett. 2017. Ecological factors influencing lifetime productivity of pink salmon (*Oncorhynchus gorbuscha*) in an Alaskan stream. Canadian Journal of Fisheries and Aquatic Sciences 74:1325–1336. https:// doi. org/10.1139/cjfas-2016-0335.

Manhard, C.V., M.D. Adkison, J.J. Hard, W.W. Smoker, and A.J. Gharrett. 2018. Local adaptation of phenology revealed in outcrosses between spawning segments of a salmonid population. Molecular Ecology 27:4698–4710. https://doi.org/10.1111/mec.14908.

Manishin, K.A., C.J. Cunningham, P.A.H. Westley, and A.C. Seitz. 2021. Can late stage marine mortality explain observed shifts in age structure of Chinook salmon? PLoS ONE 16:1–14. <u>https://doi.org/10.1371/journal.pone.0247370</u>.

Mantua, N.J., N.G. Taylor, G.T. Ruggerone. K.W. Myers, et al. 2009. The salmon MALBEC Project: A North Pacific-scale study to support salmon conservation planning. North Pacific Anadromous Fish Commission Bulletin 5:333-354.

Manzer, J.I. 1964. Preliminary observations on the vertical distribution of Pacific salmon (Genus *Oncorhynchus*) in the Gulf of Alaska. Journal of the Fisheries Board of Canada 21(5):891-903. <u>https://</u> doi.org/10.1139/f64-086

Marchioni, M.A., J.F. Fall, B. Davis, and G. Zimpleman. 2016. Kodiak City, Larsen Bay and Old Harbor: An Ethnographic Study of Traditional Subsistence Salmon Harvests and Uses. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 418, Anchorage.

Marston, B., and A. Frothingham. 2022. Upper Cook Inlet commercial fisheries annual management report, 2021. Alaska Department of Fish and Game, Fishery Management Report No. 22-16, Anchorage.

Martin, D.J., C.J. Whitmus, L.E. Hachmeister, E.C. Volk, and S.L. Schroder. 1987. Distribution and abundance of juvenile salmon and other fishes in the Yukon Delta. Final Report, Outer Continental Shelf Environmental Assessment Program Research Unit 660. 152p.

Martinson, E.C., J.H. Helle, D.L. Scarnecchia, and H.H. Stokes. 2008. Density-dependent growth of Alaska sockeye salmon in relation to climate-ocean regimes, population abundance, and body size, 1925 to 1998. Marine Ecology Progress Series 370:1 - 18.

Matarese, A.C., D.M. Blood, S.J. Picquelle, and J.L. Benson. 2003. Atlas of abundance and distribution patterns of ichthyoplankton from the northeast Pacific Ocean and Bering Sea ecosystems based on research conducted by the Alaska Fisheries Science Center (1972-1996). NOAA Professional Paper NMFS 1, 281 p.

McConnell, C.J., P.A.H. Westley, and M.V. McPhee. 2018. Differences in fitness-associated traits between hatchery and wild chum salmon despite long-term immigration by strays. Aquaculture Environment Interactions 10:99–113. https://doi.org/10.3354/aei00261.

McCraney, W.T., E.V. Farley, C.M. Kondzela, S.V. Naydenko, A.N. Starovoytov, and J.R. Guyon. 2012. Genetic stock identification of overwintering chum salmon in the North Pacific Ocean. Environmental Biology of Fishes 94:663-668.

McDevitt, C., D. Koster, D. Runfola, M. Horne-Brine, and J. Esquible-Hussion. 2020. Subsistence fisheries harvest monitoring report, Kuskokwim Management Area, Alaska, 2018. ADF&G Division of Subsistence, Technical Paper No. 467.

McDevitt, C., D. Koster, D. Runfola, M. Horne-Brine, and J. Esquible. 2021. Subsistence fisheries harvest monitoring report, Kuskokwim Management Area, Alaska, 2019. ADF&G Division of Subsistence, Technical Paper No. 475.

McDevitt, C., D. Koster, D. Runfola, M. Horne-Brine, and J. Esquible. 2021. Subsistence Fisheries Harvest Monitoring Report, Kuskokwim Fisheries Management Area, Alaska, 2020. ADF&G Division of Subsistence, Technical Paper No. 483.

McDevitt, C., and D. Koster. 2022. Subsistence fisheries harvest monitoring report, Kuskokwim Fisheries Management Area, 2021. ADF&G Division of Subsistence, Technical Paper No. 489.

McGregor, A.J., B.W. Van Alen, and V. Alen. 1987. Abundance, age, and sex compositions of Chinook, Sockeye, Coho, and Chum salmon catches and escapements in Southeast Alaska by: Commissioner. ADF&G Technical Data Report 200.

McKinley, T., N. DeCovich, J.W. Erickson, T. Hamazaki, R. Begich, and T.L. Vincent. 2020. Review of salmon escapement goals in Upper Cook Inlet, Alaska, 2019. Alaska Department of Fish and Game, Fishery Manuscript No. 20-02, Anchorage. McKinney, G.J., P.D Barry, C. Pascal, J.E. Seeb, L.W. Seeb, and M.V. McPhee. 2022. A New Genotyping-in-Thousands-by-Sequencing Single Nucleotide Polymorphism Panel for Mixed-Stock Analysis of Chum Salmon from Coastal Western Alaska. North American Journal of Fisheries Management 42:1134–1143. <u>https://doi.org/10.1002/</u> nafm.10805

McMillan, J.R., B. Morrison, N. Chambers, G. Ruggerone, L. Bernatches, J. Stanford, and H. Neville. 2023. A global synthesis of peer-reviewed research on the effects of hatchery salmonids on wild salmonids. Fisheries Management and Ecology DOI:10.1111/ fme.12643

McPhee, M.V., T.H. Tappenbeck, D.C. Whited, and J.A. Stanford. 2009. Genetic diversity and population structure in the Kuskokwim River drainage support the recurrent evolution hypothesis for sockeye salmon life histories. Transactions of the American Fisheries Society 138:1481–1489.

McPhee, M.V., M.S. Zimmerman, T.D. Beacham, B.R. Beckman, J.B. Olsen, L.W. Seeb, and W.D. Templin. 2009. A hierarchical framework to identify influences on Pacific salmon population abundance and structure in the Arctic-Yukon-Kuskokwim region. American Fisheries Society Symposium 70:1177–1198.

McPhee, M.V., J.M. Leon, L.I. Wilson, J.E. Siegel, and B.A. Agler. 2016. Changing growth and maturity in western Alaskan chinook salmon, *Oncorhynchus tshawytscha*, Brood Years 1975-2005. North Pacific Anadromous Fish Commission Bulletin 6:307–327.

McPhee, M., J. Siegel, and M. Adkison. 2019. Is a Warming Bering Sea Leading to Smaller Chinook Salmon? North Pacific Anadromous Fish Commission Technical Report 15:117–119. <u>https://doi.</u> org/10.23849/npafctr15/117.119.

Meka, J.M., C.E. Zimmerman, R.A. Heintz, and S.W. Wang. 2005. Body condition and feeding ecology of Kuskokwim River chum salmon (*Oncorhynchus keta*) during freshwater outmigration 1–61.

Meredith, B.L., N.D. Frost, K.S. Reppert, and G. Hagerman. 2022. Unuk and Chickamin Chinook salmon stock status and action plan, 2022. Alaska Department of Fish and Game, Alaska Department of Fish and Game, Regional Information Report No. 1J22-13, Douglas.

Mikow, E., B. Retherford, A. Goddhun and M.L. Kostick. 2016. Exploring the Subsistence Fisheries of Point Lay and Wainwright, Alaska. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 419, Fairbanks.

Mikow, E.H., B. Retherford, D.M. Runfola, and D. Gonzalez. 2019. Local traditional knowledge of salmon freshwater ecology in the middle and upper Kuskokwim River. ADF&G Division of Subsistence, Technical Paper No. 450, Fairbanks.

Miller, K., R. Shaftel, and D. Bogan. 2020. Diets and Prey Items of Juvenile Chinook (*Oncorhynchus tshawytscha*) and Coho Salmon (*O. kisutch*) on the Yukon Delta. U.S. Dep. Commer., NOAA Tech. Memo. NMFS-AFSC-410:1–54. https://doi.org/10.13140/ RG.2.2.20435.60961

Miller, K.B. and C.M. Weiss. 2023. Disentangling population level differences in juvenile migration phenology for three species of salmon on the Yukon River. Journal of Marine Science and Engineering 11, 589. https:// doi.org/10.3390/jmse11030589

Miller, S.E., J.M. Murphy, S.C. Heinl, A.W. Piston, E.A. Fergusson, R.E. Brenner, W.W. Strasburger, and J.H. Moss. 2022. Southeast Alaska pink salmon forecasting models. Alaska Department of Fish and Game, Fishery Manuscript No. 22-03, Anchorage.

Miller, S.E., M. Adkison, and L. Haldorson. 2012. Relationship of water column stability to the growth, condition, and survival of pink salmon (*Oncorhynchus gorbuscha*) in the northern coastal Gulf of Alaska and Prince William Sound. Canadian Journal of Fisheries and Aquatic Sciences 69:955–969. https://doi.org/10.1139/f2012-031

Minobe, S. 2002. Interannual to interdecadal changes in the Bering Sea and concurrent 1998/99 changes over the North Pacific Progress in Oceanography 55:45–64.

Molyneaux, D.B., D.L. Folletti, L.K. Brannian, and G. Roczicka. 2005. Fishery Data Series No . 05-45 Age, Sex, and Length Composition of Chinook Salmon from the 2004 Kuskokwim River Subsistence Fishery Final Report for Project 04-353 USFWS Office of Subsistence Management by.

Molyneaux, D.B., D.L. Folletti, and A.R. Brodersen. 2008. Salmon age, sex and length catalog for the Kuskokwim area, 2007., Regional Information Report. Alaska Department of Fish and Game, Division of Commercial Fisheries, Anchorage, AK.

Moncrieff, C., C. Brown, and L. Sill. 2009. Natural indicators of salmon abundance and timing, Yukon River., AYK Sustainable Salmon Initia-tive Final Report. Bering Sea Fisherman's Association, Anchorage.

Mortensen, D., A. Wertheimer, S. Taylor, and J. Landingham. 2000. The relation between early marine growth of pink salmon, *Oncorhynchus gorbuscha*, and marine water temperature, secondary production, and survival to adulthood. Fishery Bulletin 98:319–335.

Moss, J.H., J.M. Murphy, E.V. Farley, Jr., L.B. Eisner, and A.G. Andrews. 2009. Juvenile pink and chum salmon distribution, diet, and growth in the northern Bering and Chukchi seas. North Pacific Anadromous Fish Commission Bulletin 5:191-196.

Mossop, B. and M.J. Bradford. 2011. Importance of large woody debris for juvenile Chinook salmon habitat in small boreal forest streams in the upper Yukon River basin, Canada. Canadian Journal of Forest Research 34(9):1955-1966. https://doi.org/10.1139/x04-066

Mueter, F.J., and M.A. Litzow. 2008. Sea ice retreat alters the biogeog-raphy of the Bering Sea continental shelf. Ecological applications: a publication of the Ecological Society of America 18:309–20.

Mueter, F.J., B. Planque, G.L. Hunt, I.D. Alabia, T. Hirawake, L. Eisner, P. Dalpadado, M. Chierici, K.F. Drinkwater, N. Harada, P. Arneberg, and S.I. Saitoh. 2021. Possible future scenarios in the gateways to the Arctic for Subarctic and Arctic marine systems: II. prey resources, food webs, fish, and fisheries. ICES Journal of Marine Science 78(9):3017–3045.

Mundy, P.R., and D.F. Evenson. 2011. Environmental controls of phenology of high-latitude Chinook salmon populations of the Yukon River, North America, with application to fishery management. ICES Journal of Marine Science 68(6):1155-1164.

Munro, A.R., C. Habicht, T.H. Dann, D.M. Eggers, W.D. Templin, M J. Witteveen, T.T. Baker, K.G. Howard, J.R. Jasper, S.D. Rogers Olive, H.L. Liller, E.L. Chenoweth, and E.C. Volk. 2012. Harvest and harvest rates of chum salmon stocks in fisheries of the Western Alaska Salmon Stock Identification Program (WASSIP), 2007–2009. Alaska Department of Fish and Game, Special Publication No. 12-25, Anchorage

Munro, A.R. 2023. Summary of Pacific salmon escapement goals in Alaska with a review of escapements from 2014 to 2022. Alaska Department of Fish and Game, Fishery Manuscript No. 23-01, Anchorage.

Murphy, J.M., W.D. Templin, E.V. Farley, Jr., and J.E. Seeb. 2009. Stock-structured distribution of western Alaska and Yukon juvenile Chinook salmon (*Oncorhynchus tshawytscha*) from United States BASIS surveys, 2002–2007. North Pacific Anadromous Fish Commission Bulletin 5:51–59.

Murphy, J., K. Howard, L. Eisner, A. Andrews, W. Templin, C. Guthrie, K. Cox, and E. Farley. 2013. Linking Abundance, Distribution, and Size of Juvenile Yukon River Chinook Salmon to Survival in the Northern Bering Sea. North Pacific Anadromous Fish Commission Technical Report 9:25–30.

Murphy, J.M., E.V. Farley, J.N. Ianelli, and D.L. Stram. 2016. Distribution, Diet, and Bycatch of Chum Salmon in the Eastern Bering Sea. North Pacific Anadromous Fish Commission Bulletin 6:219–234. https://doi.org/10.23849/npafcb6/219.234.

Murphy, J.M., K.G. Howard, J.C. Gann, K.C. Cieciel, W.D. Templin, and C.M. Guthrie III. 2017. Juvenile Chinook salmon abundance in the northern Bering Sea: implications for future returns and fisheries in the Yukon River. Deep Sea Research Part II: Topical Studies in Oceanography 135: 156-167. <u>https://doi.org/10.1016/j</u>. dsr2.2016.06.002.

Murphy, J. M., E.A. Fergusson, A. Piston, S. Heinl, A. Gray, and E. Farley. 2019. Southeast Alaska pink salmon growth and harvest forecast models. North Pacific Anadromous Fish Commission Technical Report No. 15:75-91.

Murphy, J., S. Garcia, J. Dimond, J. Moss, F. Sewall, W. Strasburger, E. Lee, T. Dann, E. Labunski, T. Zeller, A. Gray, C. Waters, D. Jallen, D. Nicolls, R. Conlon, K. Cieciel, K. Howard, B. Harris, N. Wolf, and E. Farley. 2021. Northern Bering Sea surface trawl and ecosystem survey cruise report, 2019. U.S. Dep. Commer., NOAA Tech. Memo. NMFS-AFSC-423, 124 p.

Murphy, M.L., 1984. Primary production and grazing in freshwater and intertidal reaches of a coastal stream, southeast Alaska. Limnology and Oceanography 29:805–815.

Murphy, M.L., J. Heifetz, J.F. Thedinga, S.W. Johnson, and K.V. Koski. 1989. Habitat utilization by juvenile Pacific salmon (Oncorhynchus) in the glacial Taku River, southeast Alaska. Canadian Journal of Fisheries and Aquatic Sciences 46:1677–1685.

Myers, K.W., R.V. Walker, and N. Davis. 2001. Ocean Distribution and Migration Patterns of Yukon River Chinook Salmon. Final Report to: Yukon River Drainage Fisheries Association. School of Aquatic and Fisheries Sciences, University of Washington, Seattle, SAFS-UW-0109.

Myers, K.W., N.D. Davis, R.V. Walker, and J.L. Armstrong. 2006. Migration studies of salmon in the Bering Sea. Final Report, NOAA contract No. NA17RJ1232 AM021. University of Washington.

Myers, K.W., R.V. Walker, N.D. Davis, J.L. Armstrong, and M. Kaeriyama. 2009. High seas distribution, biology, and ecology of AYK salmon: direct information from high seas tagging experiments, 1954 - 2006. American Fisheries Society Symposium 70:201-239.

Myers, K., R. Walker, N. Davis, J. Armstrong, W. Fournier, N. Mantua, and J. Raymond-Yakoubian. 2010. Climate-Ocean Effects on AYK Chinook Salmon. Page 249. SAFS-UW-1003, School of Aquatic and Fishery Sciences, University of Washington, Seattle.

Myers, K., J. Irvine, E. Logerwell, S. Urawa, S. Naydenko, A. Zavolokin, and N. Davis. 2016. Pacific Salmon and Steelhead: Life in a Changing Winter Ocean. North Pacific Anadromous Fish Commission Bulletin 6:113–138. https://doi.org/10.23849/ npafcb6/113.138.

Ν

Nagasawa, K. 1998. Predation by salmon sharks on Pacific salmon in the North Pacific Ocean. North Pacific Anadromous Fish Commission Bulletin 1:419-433.

Nagasawa, K. 2023. Catch of coho salmon (*Oncorhynchus kisutch*) infected with the freshwater parasite Salvelinema walkeri (Nematoda: *Cystidicolidae*) in the Gulf of Alaska in the early winter. Species Diversity 28:41-146. DOI: 10.12782/specdiv.28.141

Nakatani, R.E., G.J. Paulik, and R. Van Cleve. 1975. Pink salmon (*Oncoryhnchus gorbuscha*) tagging experiments in S. E. Alaska, 1938–1942 and 1945. National Oceanic and Atmospheric Administration, National Marine Fisheries Service, Special Scientific Report, Fisheries Series 686, Seattle. National Research Council. 2005. Developing a Research and Restoration Plan for Arctic-Yukon-Kuskokwim (Western Alaska) Salmon. Washington, DC: The National Academies Press. https:// doi.org/10.17226/11080.

Naves, L.C., M.F. Turek, and W.E. Simeone. 2010. Subsistence– personal use salmon harvest, Southeast–Yakutat Management Region, 1996–2006. Alaska Department of Fish and Game Division of Subsistence Technical Paper No. 350, Anchorage.

NBTC (Northern Boundary Technical Committee). 2005. Stock Composition Estimates and individual stock assignments based on genetic microsatellites and scale patterns for test mixtures of Alaskan and Canadian sockeye salmon. TCNB (05)-2.

Neilsen, J.L., and G.T. Ruggerone. 2004. Top-down and bottom-up linkages among climate, growth, competition, and production of sockeye salmon populations in Bristol Bay, Alaska, 1955-2000. PICES 13th Annual Meeting book of abstracts. 24

Neuswanger, J.R., M.S. Wipfli, M.J. Evenson, N.F. Hughes, A.E. Rosenberger, and B. Jonsson, , 2015. Low productivity of Chinook salmon strongly correlates with high summer stream discharge in two Alaskan rivers in the Yukon drainage. Canadian Journal of Fisheries and Aquatic Sciences 1–13. https://doi.org/10.1139/cjfas-2014-0498Nielsen, J.L., and G.T. Ruggerone. 2009. Climate change and a dynamic ocean carrying capacity: growth and survival of Pacific salmon at sea. American Fisheries Society Symposium 71:77-96.

Nielsen, J.L., G.T. Ruggerone, and C.E. Zimmerman. 2012. Adaptive strategies and life history characteristics in a warming climate: salmon in the Arctic? Environ Biology of Fishes 96:1187-1226

North Pacific Anadromous Fish Commission. 2023. The status and trends of Pacific salmon and steelhead trout stocks with linkages to their ecosystem. North Pacific Anadromous Fish Commission Technical Report 19. 256 pp. https://doi.org/10.23849/LOEX7610 NPFMC LKTKS Search Engine https://lktks.npfmc.org/

0

O'Neill, D., 2012. The fall of the Yukon kings, *in*: Banerjee, S. (Ed.), Arctic Voices: Resistance at the Tipping Point. Seven Stories Press, NY, 25p.

Ohlberger, J., D.E. Schindler, R.J. Brown, J.M.S. Harding, M.D. Adkison, A.R. Munro, L. Horstmann, and J. Spaeder. 2020. The reproductive value of large females: consequences of shifts in demographic structure for population reproductive potential in Chinook salmon. Canadian Journal of Fisheries and Aquatic Sciences dx.doi.org/10.1139/cjfas-2020-0012

Ohlberger, J., E.J. Ward., R.E. Brenner, M.E. Hunsicker, et al. 2022. Non-stationary and interactive effects of climate and competition on pink salmon productivity. Global Change Biology. DOI:10.1111/ gcb.16049.

Oke, K.B., F. Mueter, and M.A. Litzow. 2019. Warming leads to opposite patterns in weight-at-age for young versus old age classes of Bering Sea walleye pollock. Canadian Journal of Fisheries and Aquatic Sciences 79:1655-1666. dx.doi.org/10.1139/cjfas-2021-0315.

Olsen, J.B., P.A. Crane, B.G. Flannery, K. Dunmall, W.D. Templin, and J.K. Wenburg. 2010. Comparative landscape genetic analysis of three Pacific salmon species from subarctic North America. Conservation Genetics 12:223-241.

Olsen, J.B., T.D. Beacham, M. Wetklo, L.W. Seeb, C.T. Smith, B.G. Flannery, J.K. Wenburg. 2010. The influence of hydrology and waterway distance on population structure of Chinook salmon *Oncorhynchus tshawytscha* in a large river. Journal of Fish Biology 76:1128-1148.

Orsi, J.A., and A.C. Wertheimer. 1995. Marine vertical distribution of juvenile Chinook and coho salmon in southeastern Alaska. Transactions of the American Fisheries Society 124:159–169.

Orsi, J. A., and H.W. Jaenicke. 1996. Marine distribution and origin of pre-recruit Chinook salmon *Oncorynchus tshawytscha*, in southeastern Alaska. Fishery Bulletin 94:482-497

Orsi, J.A., M.V. Sturdevant, J.M. Murphy, D.G. Mortensen, and B.L. Wing. 2000. Seasonal habitat use and early marine ecology of juvenile Pacific salmon in Southeastern Alaska. North Pacific Anadromous Fish Commission Bulletin No. 2:111–122.

Orsi, J.A., A.C. Wertheimer, M.V. Sturdevant, E.A. Fergusson, D.G. Mortensen, and B.L. Wing. 2004. Juvenile chum salmon consumption of zooplankton in marine waters of southeastern Alaska: a bioenergetics approach to implications of hatchery stock interactions. Reviews in Fish Biology and Fisheries 14:335-359.

Orsi, J., A. Wertheimer, M. Sturdevant, E. Fergusson, and B. Wing. 2009. Insights from a 12-year biophysical time series of juvenile Pacific salmon in Southeast Alaska: the Southeast Alaska Coastal Monitoring Project (SECM). Alaska Fisheries Science Center Quarterly Report July–September 2009.

Orsi, J.A., Fergusson, E.A., Sturdevant, M.V., Wing, B.L., Wertheimer, A.C., Heard, W.R., 2009. Annual survey of juvenile salmon, ecologically-related species, and environmental factors in the marine waters of southeastern Alaska, May - August 2008. North Pacific Anadromous Fish Commission Doc. 1181, 72 pp.

Orsi, J., E. Fergusson, and J.E. Joyce. 2013. Connecting the "Dots" Among Coastal Ocean Metrics and Pacific Salmon Production in Southeast Alaska, 1997-2012. North Pacific Anadromous Fish Commission, Technical Report 9.

Oslund, S., S. Ivey, and D. Lescanec. 2020. Area Management Report for the sport fisheries of northern Cook Inlet, 2017–2018. Alaska Department of Fish and Game, Fishery Management Report No. 20-04, Anchorage. Otis, E.O., J.W. Erickson, C. Kerkvliet, and T. McKinley. 2016. A review of escapement goals for salmon stocks in Lower Cook Inlet, Alaska, 2016. Alaska Department of Fish and Game, Fishery Manuscript Series No. 16-08, Anchorage.

Otis, E.O., G.J. Hollowell, and E.G. Ford. 2018. Observations of pink salmon hatchery proportions in selected Lower Cook Inlet escapements, 2014-2017. Alaska Department of Fish and Game, Special Publication No. SP18-11, Anchorage.

Ovando, D., C. Cunningham, P. Kuriyama, C. Boatright, and R. Hilborn. 2022. Improving forecasts of sockeye salmon (*Oncorhynchus nerka*) with parametric and nonparametric models. Canadian Journal of Fisheries and Aquatic Sciences, 99:1-13.

Oxman, D.S., Smoker, W.W., Gharrett, A.J., 2013. Developmental progression of gill rakers as a post-hatch developmental marker in pink salmon, *Oncorhynchus gorbuscha*. Environmental Biology of Fishes 96:677–689. https://doi.org/10.1007/s10641-012-0058-6

Ρ

Paige, A.W., S. Churchill, N. Ratner, M. Turek, and P. Coiley-Kenner. 2009. Local knowledge, harvest patterns, and community use of salmon in Wrangell, Alaska. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 323, Juneau.

Parker-Stetter, S.L., J.K. Horne, E.V. Farley, D.H. Barbee, A.G. Andrews, L.B. Eisner, and J.M. Nomura. 2013. Summer distributions of forage fish in the eastern Bering Sea. Deep-Sea Research Part II: Topical Studies in Oceanography 94:211–230. https://doi.org/10.1016/j.dsr2.2013.04.022.

Pella, J.J., and H.J Geiger. 2009. Sampling considerations for estimating geographic origins of Chinook salmon bycatch in the Bering Sea pollock fishery., Special Publication No. SP 09-08. Alaska Department of Fish and Game, Anchorage.

Peltz, L., and J.P. Koenings. 1989. Evidence for temperature limitation of juvenile sockeye salmon, (*Oncorynchus nerka*), growth in Hugh Smith Lake, Alaska. FRED Reports No. 90

Pestal, G., C.J. Schwarz and R.A. Clark. 2020. Taku River Sockeye Salmon Stock Assessment Review and Updated 1984-2018 Abundance Estimates. Pacific Salmon Comm. Tech. Rep. No. 43: 118 p.

Petrou, E.L., L. Hauser, R.S. Waples, J.E. Seeb, W.D. Templin, D. Gomez-Uchida and L.W. Seeb. 2013. Secondary contact and changes in coastal habitat availability influence the nonequilibrium population structure of a salmonid (*Oncorhynchus keta*). Molecular Ecology 22(23):5848-5860.

Petrou, E.L., J.E. Seeb, L. Hauser, M.J. Witteveen, W.D. Templin, and L.W. Seeb. 2014. Fine-scale sampling reveals distinct isolation by distance patterns in chum salmon (*Oncorhynchus keta*) populations occupying a glacially dynamic environment. Conservation Genetics 15(1):229-243.

Piston, A.W., S.C. Heinl, H.J. Geiger, and T.A. Johnson. 2006. Hugh Smith Lake sockeye salmon adult and juvenile studies, 2003-2005. Alaska Department of Fish and Game, Fishery Data Series No. 06-51, Anchorage.

Piston, A.W., and S.C. Heinl. 2012a. Hatchery Chum Salmon Straying in Southeast Alaska, 2011. Alaska Department of Fish and Game, Fishery Data Series No. 12-45, Anchorage.

Piston, A.W., and S.C. Heinl. 2012b. Hatchery Chum Salmon Straying Studies in Southeast Alaska, 2008–2010. Alaska Department of Fish and Game, Fishery Manuscript Series No. 12-01, Anchorage.

Piston, A.W., and S.C. Heinl. 2020. Chum salmon stock status and escapement goals in southeast Alaska through 2019. Alaska Department of Fish and Game, Special Publication No. 20-10, Anchorage.

Piston, A.W., and S.C. Heinl. 2020. Pink salmon stock status and escapement goals in southeast Alaska through 2019. Alaska Department of Fish and Game, Special Publication No. 20-09, Anchorage.

Piston, A.W. 2021. District 104 purse seine fishery harvest pattern analysis. Pacific Salmon Commission Technical Report No. 44: 127 p.

Poetter, A. D., and A. Tiernan. 2017. 2016 Kuskokwim area management report. Alaska Department of Fish and Game, Fishery Management Report No. 17-50, Anchorage.

Polum, T., M. Witteveen, M. Stratton, and M. Evans. 2019. Report on selected sport fisheries of the Kodiak Management Area, 2008–2017. Alaska Department of Fish and Game, Fishery Management Report No. 19-04, Anchorage.

Polum, T. 2023. Report on selected sport fisheries of the Alaska Peninsula–Aleutian Islands Management Area, 2012–2021. Alaska Department of Fish and Game, Fishery Management Report No. 23-01, Anchorage.

Porter, T.J., S.W. Schoenemann, L.J. Davies, E.J. Steig, S. Bandara, and D.G. Froese. 2019. Recent summer warming in northwestern Canada exceeds the Holocene thermal maximum. Nature Communications 10(1):1631.

Priest, J., S.C. Heinl, and L.D. Shaul. 2021. Coho salmon stock status in southeast Alaska: a review of trends in productivity, harvest, and abundance through 2019. Pacific Salmon Commission Technical Report No. 45.

Punt, A.E., D.S. Butterworth, C.L. de Moor, J.A.A. De Oliveira, and M. Haddon. 2016. Management strategy evaluation: best practices. Fish and Fisheries 17:303–334. https://doi.org/10.1111/faf.12104

R

Ramos, J. and R. Mason. Traditional Ecological Knowledge of Tlingit People Concerning the Sockeye Salmon Fishery of the Dry Bay Area. Fisheries Information Service Project 01-091. 79 p. Ransbury, S.R., N.L. Zeiser, J.A. Bednarski, S.C. Heinl, C.S. Jalbert, and S.E. Miller. 2021. Stock assessment study of Chilkat Lake and River sockeye salmon, 2017–2020. Alaska Department of Fish and Game, Fishery Manuscript Series No. 21-06, Anchorage.

Ratner, N.C. and J.A. Dizard. 2006. Local knowledge, harvest patterns, and community use of sockeye salmon in Hoonah, Alaska. ADF&G Division of Subsistence, Technical Paper No. 307.

Ratner, N.C., P. Brown, J. Rowan, D. Yates, M. Smith, J.A. Dizard, A. Paige, and M.F. Turek. 2006. Local knowledge, customary practices, and harvest of sockeye salmon from the Klawock and Sarkar rivers, Prince of Wales Island, Alaska. ADF&G Division of Subsistence, Technical Paper No. 308.

Raymond-Yakoubian, B., and J. Raymond-Yakoubian. 2015. "Always taught not to waste": Traditional Knowledge and Norton Sound/ Bering Strait Salmon Populations." 2015 Arctic-Yukon-Kuskokwim Sustainable Salmon Initiative Project 1333 Final Product. Kawerak. Inc. Social Science Program: Nome, AK.

Raymond-Yakoubian, J., 2009. Climate-Ocean Effects on Chinook Salmon: Local Traditional Knowledge Component. Final report to the Arctic Yukon Kuskokwim Sustainable Salmon Initiative for project 712. Kawerak. Inc. Social Science Program: Nome, AK.

Raymond-Yakoubian, J., B. Raymond-Yakoubian, and C. Moncrieff. 2017. The incorporation of traditional knowledge into Alaska federal fisheries management. Marine Policy 78:132-142.

Ream, J.T. and J. Merriam. 2017. Local and traditional knowledge of Stikine River Chinook salmon: a local perspective on a vital commercial, sport, and subsistence fish. Alaska Department of Fish and Game Division of Subsistence, Technical paper No. 430, Anchorage.

Reese, C., N. Hillgruber, M. Sturdevant, A. Wertheimer, W. Smoker, and R. Focht. 2009. Spatial and temporal distribution and the potential for estuarine interactions between wild and hatchery chum salmon (*Oncorhynchus keta*) in Taku Inlet, Alaska. Fishery Bulletin 107:433–450.

Reid, A.J., L.E. Eckert, J.F. Lane, N. Young, S.G. Hinch, C.T. Darimont, S.J. Cooke, N.C. Ban, and A. Marshall. 2020. "Two-Eyed Seeing": An Indigenous framework to transform fisheries research and management. Fish and Fisheries 22:243-261, doi:10.1111/faf.12516.

Reimer, A.M., A. Rogers, and J.N. Sanchirico. 2024. Adaptive systems for climate-ready fisheries management. Resources for the Future, Working Paper 24-06.

Reimer, A.M., and N.A. DeCovich. 2020. Susitna River Chinook salmon run reconstruction and escapement goal analysis. Alaska Department of Fish and Game, Fishery Manuscript No. 20-01, Anchorage.

Rich, W.H. 1927. Salmon-tagging experiments in Alaska, 1924 and 1925. Bulletin of the United States Bureau of Fisheries 42:109–146.

Rich, W.H., and A.J. Suomela. 1929. Salmon-tagging experiments in Alaska, 1926. Bulletin of the United States Bureau of Fisheries 43(Part 2):71–104.

Riddell, B.E., K.G. Howard, and A.R. Munro. 2022. Salmon returns in the North Pacific in relation to expedition observations (and next steps). In L. Fitzpatrick, tech. editor. Virtual Conference on Winter Ecology of Pacific Salmon and Results from the Two Gulf of Alaska Expeditions, North Pacific Anadromous Fish Commission Technical Report 18:115–139. (Available at https://npafc.org)

Riffe, R., and B. Mercer. 2006. Effects of habitat and predator–prey interactions on stocked sockeye fry in Tatsamenie Lake. Alaska Department of Fish and Game, Fishery Manuscript No. 06-02, Anchorage.

Rogers, O., S.D., E.K.C. Fox, and S.E. Gilk-Baumer. 2018. Genetic baseline for mixed stock analyses of sockeye salmon harvested in SEAK for Pacific salmon treaty applications, 2018. Alaska Department of Fish and Game, Fishery Manuscript No. 18-03

Rogers, D.E., and G.T. Ruggerone. 1993. Factors affecting marine growth of Bristol Bay sockeye salmon. Fisheries Research 18:89-103.

Ruggerone, G.T., and D.E. Rogers. 1992. Predation on sockeye salmon fry by juvenile coho salmon in the Chignik Lakes, Alaska: Implications for salmon management. North American Journal of Fisheries Management 12:87-102.

Ruggerone, G.T., R. Hanson, and D.E. Rogers. 2000. Selective predation by brown bears foraging on spawning sockeye salmon. Canadian Journal of Zoology 78:974-981.

Ruggerone, G.T. 2003. Rapid natural habitat degradation and consequences for sockeye salmon production in the Chignik Lakes system, Alaska. SAFS-UW-0309, 133pgs.

Ruggerone, G.T., M. Zimmermann, K.W. Myers, J.L. Nielsen, and D.E. Rogers. 2003. Competition between Asian pink salmon and Alaskan sockeye salmon in the North Pacific Ocean. Fisheries Oceanography 12(3):209-219.

Ruggerone, G.T. 2004. Pre-season forecast of sockeye salmon migration timing in Bristol Bay, Alaska based on oceanographic and biological variables. North Pacific Research Board Report 39pgs

Ruggerone, G.T., and J.L. Nielsen. 2004. Evidence for competitive dominance of Pink salmon over other salmonids in the North Pacific Ocean. Reviews in Fish Biology and Fisheries 14:371-390.

Ruggerone, G.T., E. Farley, J. Nielsen, and P. Hagen. 2005. Seasonal marine growth of Bristol Bay sockeye salmon in relation to competition with Asian pink salmon and the 1977 ocean regime shift. Fishery Bulletin 103:355-370.

Ruggerone, G.T., J.L. Nielsen, and J. Bumgarner. 2007. Linkages between Alaskan sockeye salmon abundance, growth at sea, and climate, 1955 - 2002. Deep-Sea Research II 54:2776-2793.

Ruggerone, G.T., and J.L. Nielsen. 2009. A review of growth and survival of salmon at sea in response to competition and climate change. American Fisheries Society Symposium 70:241-265.

Ruggerone, G.T., J.L Nielsen, and B.A. Agler. 2009. Linking marine and freshwater growth in western Alaska Chinook salmon *Oncorhynchus tshawytscha*. Journal of Fish Biology 75:1287–1301.

Ruggerone, G.T., J.L. Nielsen, and B.A. Agler. 2009. Climate, growth and population dynamics of Yukon River Chinook salmon. North Pacific Anadromous Fish Commission Bulletin 5:279–285

Ruggerone, G.T., R.M. Peterman, B. Dorner, and K.W. Myers. 2010. Magnitude and trends in abundance of hatchery and wild pink salmon, chum salmon, and sockeye salmon in the North Pacific Ocean. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 2:306-328.

Ruggerone, G.T., B.A. Agler, J.L. Nielsen. 2012. Evidence for competition at sea between Norton Sound chum salmon and Asian hatchery chum salmon. Environmental Biology of Fishes 94:149-163.

Ruggerone, G.T., B.A. Agler, L. Wilson, and E.V. Farley, Jr. 2013. Size-selective mortality of Bristol Bay sockeye smolts in relation to smolt characteristics, ocean conditions, and sockeye salmon productivity. North Pacific Anadromous Fish Commission Technical Report 9:210-213.

Ruggerone, G.T., and B.M. Connors. 2015. Productivity and life history of sockeye salmon in relation to competition with pink and sockeye salmon in the North Pacific Ocean. Canadian Journal of Fisheries and Aquatic Sciences 72:818-833.

Ruggerone, G.T., B.A. Agler, B.M. Connors, E.V. Farley, Jr., et al. 2016. Pink and sockeye salmon interactions at sea and their influence on forecast error of Bristol Bay sockeye salmon. North Pacific Anadromous Fish Commission Bulletin 6:349-361.

Ruggerone, G.T., J.R. Irvine, and B. Connors. 2021. Did recent heatwaves and record high pink salmon abundance lead to a tipping point that caused record declines in North Pacific salmon abundance and harvest in 2020? North Pacific Anadromous Fish Commission, Technical Report 17:78-82.

Ruggerone, G.T., A.M. Springer, G.B. van Vliet, B. Connors, J.R. Irvine, L.D. Shaul, M.R. Sloat, and W.I. Atlas. 2023. From diatoms to killer whales: impacts of pink salmon on North Pacific ecosystems. Marine Ecology Progress Series 719:1-40.

Runfola, D.M., H. Ikuta, A.R. Brenner, J.J. Simon, J. Park, D. S. Koster, and M. Kostick. 2017. Bethel subsistence, 2012: wild resource harvests and uses, land use patterns, and subsistence economy in the hub community of the Yukon–Kuskokwim Delta, Alaska. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 393, Fairbanks.

Runfola, D.M., C.R. McDevitt, and C.L. Brown. 2018. Overview of the development and implementation of the Kuskokwim River household subsistence king salmon permit system, 2018. Alaska Department of Fish and Game Division of Subsistence, Special Publication No. -006.

Runfola, D.M. and D. Koster. 2019. Inseason estimation of subsistence salmon fishing effort and harvest in the lower Kuskokwim River, 2015–2018. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 449, Fairbanks.

Runfola, D.M., L.S. Naaktgeboren, and D. Koster. 2019. Inseason subsistence salmon harvest assessments in 9 communities of the middle Kuskokwim River, 2015–2018. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 455.

Russell, C., J. Botz, and J. Morella. 2021 Prince William Sound area salmon fisheries -- a report to the Alaska Board of Fisheries, 2021. Alaska Department of Fish and Game, Special Publication No. 21-09, Anchorage.

Russell, C.W. 2023. North Alaska Peninsula commercial salmon annual management report, 2022. Alaska Department of Fish and Game, Fishery Management Report No. 23-03, Anchorage.

S

Salomone, P.G., K. Courtney, G.T. Hagerman, P.A. Fowler, and P.J. Richards. 2022. Stikine River and Andrew Creek Chinook salmon stock status and action plan, 2021. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report No. 1J22-15, Douglas.

Sato, S., S. Moriya, T. Azumaya, and H. Nagoya. 2009. Stock Distribution Patterns of Chum Salmon in the Bering Sea and North Pacific Ocean during the Summer and Fall of 2002 – 2004. North Pacific Anadromous Fish Commission Bulletin 5:29–37.

Scannell, H., J. Botz, K. Gatt, J. Morella, J. Buza, and R. Ertz. 2023. 2021 Prince William Sound area finfish management report. Alaska Department of Fish and Game, Fishery Management Report No. 23-06, Anchorage.

Schaberg, K.L., Z.W. Liller, and D.B. Molyneaux. 2010. A Mark – Recapture study of Kuskokwim River coho, chum, sockeye, and Chinook salmon, 2001 – 2006. Alaska Department of Fish and Game, Fishery Data Series No. 10-32, Anchorage.

Schaberg, K.L., Z.W. Liller, D.B. Molyneaux, B.G. Bue, and L. Stuby. 2012. Estimates of Total Annual Return of Chinook Salmon to the Kuskokwim River, 2002 – 2007. Alaska Department of Fish and Game, Fishery Data Series No. 12-36, Anchorage.

Schaberg, K., T. McKinley and H. Finkle. 2023. Review of Salmon Escapement Goals in the Alaska Peninsula, Aleutian Islands and Chignik Management Areas. Oral Report to the Alaska Board of Fisheries 2023. Scheuerell, M.D., R. Hilborn, M.H. Ruckelshaus, K.K. Bartz, K.M. Lagueux, A.D. Haas, and K. Rawson. 2006. The Shiraz model: a tool for incorporating anthropogenic effects and fish-habitat relationships in conservation planning. Canadian Journal of Fisheries and Aquatic Sciences 63: 1596-1607. doi:10.1139/F06-056

Schindler, D., C. Krueger, P. Bisson, M. Bradford, B. Clark, J. Conitz, K. Howard, M. Jones, J. Murphy, K. Myers, M. Scheuerel, E. Volk, and J. Winton. 2013. Arctic -Yukon-Kuskokwim Chinook Salmon Research Action Plan: Evidence of Decline of Chinook Salmon Populations and Recommendations for Future Research. Page 70. AYK Sustainable Salmon Initiative, Anchorage, AK.

Schoch, G.C., D.M. Albert, and C.S. Shanley. 2014. An Estuarine Habitat Classification for a Complex Fjordal Island Archipelago. Estuaries and Coasts 37:160–176. https://doi.org/10.1007/ s12237-013-9622-3

Schuster, M., M.D. Booz, and A.W. Barclay. 2021. Chinook salmon sport harvest genetic stock and biological compositions in Cook Inlet salt waters, 2014–2018. Alaska Department of Fish and Game, Fishery Manuscript No. 21-04, Anchorage.

Schwanke, C.J., and M.J. Piche. 2023. Run timing and spawning distribution of Copper River Chinook salmon, 2019–2021. Alaska Department of Fish and Game, Fishery Data Series No. 23-14, Anchorage.

Seeb, J.E., C. Habicht, W.D. Templin, J.B. Shaklee, L.W. Seeb, and F.M. Utter. 1999. Allozyme and mtDNA variation describe ecologically important genetic structure of even-year pink salmon inhabiting Prince William Sound, Alaska. Ecology of Freshwater Fish 8:122-140.

Seeb, L.W., C. Habicht, W.D. Templin, K E. Tarbox, R.Z. Davis, L.K. Brannian, and J.E. Seeb. 2000. Genetic diversity of sockeye salmon (*Oncorhynchus nerka*) of Cook Inlet, Alaska, and its application to restoration of populations affected by the Exxon Valdez oil spill. Transactions of the American Fisheries Society 129:1223-1249.

Seeb, L.W., P.A. Crane, C.M. Kondzela, R.L. Wilmot, S. Urawa, N.V. Varnavskaya, and J.E. Seeb. 2004. Migration of Pacific Rim chum salmon on the high seas: insights from genetic data. Environmental Biology of Fishes 69:21-36.

Seeb, L.W., A. Antonovich, M. Banks, T. Beacham, R. Bellinger, S. Blankenship, M. Campbell, N. DeCovich, J.C. Garza, C. Guthrie, T. Lundrigan, P. Moran, S. Narum, J. Stephenson, J. Supernault, D. Teel, W.D. Templin, J.K. Wenburg, S. Young, and C.T. Smith. 2007. Development of a standardized DNA database for Chinook salmon. Fisheries 32(11):540–552

Seeb, L.W., N.A. DeCovich, A.W. Barclay, C.T. Smith, and W.D. Templin. 2009. Timing and origin of Chinook salmon stocks in the Copper River and adjacent ocean fisheries using DNA markers. Alaska Department of Fish and Game, Fishery Data Series No. 09-58, Anchorage.

Seeb, L.W., W.D. Templin, S. Sato, S. Abe, K. Warheit, J.Y. Park, and J.E. Seeb. 2011. Single nucleotide polymorphisms across a species' range: implications for conservation studies of Pacific salmon. Molecular Ecology Resources, 11:195–217. doi: 10.1111/j.1755-0998.2010.02966.x

Seeb, L.W., J.E. Seeb, C. Habicht, E.F. Farley, Jr., and F.M. Utter. 2011. Single-nucleotide polymorphic gentoypes reveal patterns of early juvenile migration of sockeye salmon in the eastern Bering Sea. Transactions of the American Fisheries Society 140(3):734-748.

Seitz, A.C., M.B. Courtney, M.D. Evans, and K. Manishin. 2019. Pop-up satellite archival tags reveal evidence of intense predation on large immature Chinook salmon (*Oncorhynchus tshawytscha*) in the North Pacific Ocean. Canadian Journal of Fisheries and Aquatic Sciences, 76(9):1608-1615.

Sergeant, C.J., J.R. Bellmore, C. McConnell, and J.W. Moore. 2017. High salmon density and low discharge create periodic hypoxia in coastal rivers. Ecosphere 8, e01846. https://doi.org/10.1002/ ecs2.1846

Sergeant, C.J., J.R. Bellmore, R.A. Bellmore, J.A. Falke, F.J. Mueter, and P.A.H. Westley. 2023. Hypoxia vulnerability in the salmon watersheds of Southeast Alaska. Science of The Total Environment 896, 165247. https://doi.org/10.1016/j.scitotenv.2023.165247

Shanley, C.S., and D.M. Albert. 2014. Climate Change Sensitivity Index for Pacific Salmon Habitat in Southeast Alaska. PLoS ONE 9, e104799. https://doi.org/10.1371/journal.pone.0104799

Shaul, L.D., R Ericksen, K. Crabtree, and J. Lum. 2013. Beyond the estuary: an extension of the nomad life history strategy in coho salmon. North Pacific Anadromous Fish Commission Technical Report No. 9:171–175.

Shaul, L.D., and H.J. Geiger. 2016. Effects of climate and competition for offshore prey on growth, survival, and reproductive potential of coho salmon in Southeast Alaska. North Pacific Anadromous Fish Commission Bulletin 6:329-347. doi: 10.23849/npafcb6/329.347

Shaul, L.D., J.A. Bednarski, J.T. Williams, and B.W. Elliott. 2019. Stock status and review of factors affecting coho salmon returns and escapements in Southeast Alaska. Alaska Department of Fish and Game, Regional Information Report No. 1J19-12, Douglas.

Shedd, K., T.H. Dann, C. Habicht, and W.D. Templin. 2014. Defining reproductive success: which fish count? Alaska Hatchery Research Program Technical Document No. 1.

Shedd, K., T.H. Dann, C. Habicht, and W.D. Templin. 2014. Parentage SNP selection - SEAK chum. Alaska Hatchery Research Program Technical Document No. 2.

Shedd, K.R., T.H. Dann, H.A. Hoyt, M.B. Foster, and C. Habicht. 2016. Genetic baseline of North American sockeye salmon for mixed stock analyses of Kodiak Management Area commercial fisheries, 2014–2016. Alaska Department of Fish and Game, Fishery Manuscript Series No. 16-03, Anchorage. Shedd, K., M. Foster, M. Wattum, T. Polum, M. Witteveen, M. Stratton, T. Dann, H. Hoyt, and C. Habicht. 2016. Genetic Stock Composition of the Commercial and Sport Harvest of Chinook Salmon in Westward Region, 2014-2016. Alaska Department of Fish and Game, Fishery Manuscript Series No. 16-11

Shedd, K.R., D.L. Leonard, and J.V. Nichols. 2022. Mixed stock analysis of Chinook salmon harvested in Southeast Alaska commercial troll and sport fisheries, 2019. Alaska Department of Fish and Game, Fishery Data Series No. 22-20, Anchorage

Shedd, K.R., E.A. Lescak, C. Habicht, E.E. Knudsen, T.H. Dann, H.A. Hoyt, D.J. Prince, and W.D. Templin. 2022. Reduced relative fitness in hatchery-origin Pink Salmon in two streams in Prince William Sound, Alaska. Evolutionary Applications, 15(3):429–446. <u>https://doi.org/10.1111/eva.13356</u>.

Siddon, E.C., T. Kristiansen, F.J. Mueter, K.K. Holsman, R.A. Heintz, and E.V. Farley. 2013. Spatial match-mismatch between juvenile fish and prey provides a mechanism for recruitment variability across contrasting climate conditions in the eastern Bering Sea. PloS One 8, e84526. https://doi.org/10.1371/journal.pone.0084526

Siddon, E. 2022. Ecosystem Status Report 2022: Eastern Bering Sea, Stock Assessment and Fishery Evaluation Report, North Pacific Fishery Management Council, 1007 W. 4rd Ave., Suite 400, Anchorage, AK 99501.

Siegel, J.E., M.V. McPhee, and M.D. Adkison. 2017. Evidence that marine temperatures influence growth and maturation of western Alaskan Chinook Salmon *Oncorhynchus tshawytscha*. Marine and Coastal Fisheries 9:441–456. <u>https://doi.org/10.1080/19425120</u>. 2017.1353563

Siegel, J.E., M.D. Adkison, and M.V. McPhee. 2018. Changing maturation reaction norms and the effects of growth history in Alaskan Chinook salmon. Marine Ecology Progress Series 595 ;187–202. https://doi.org/10.3354/meps12564

Sigler, M., M. Renner, and S. Danielson. 2011. Fluxes, fins, and feathers: relationships among the Bering, Chukchi, and Beaufort seas in a time of climate change. Oceanography 24:250–265.

Sigler, M., A. Hollowed, K. Holsman, S. Zador, A. Haynie, A. Himes-Cornell, P. Mundy, S. Davis, J. Duffy-Anderson, T. Gelatt, B. Gerke, and P. Stabeno. 2016. Alaska Regional Action Plan for the Southeastern Bering Sea. NOAA Technical Memorandum NMFS AFSC i–50. http://dx.doi.org/10.7289/V5/TM-AFSC-336

Sigler, M.F., F.J. Mueter, B.A. Bluhm, M.S. Busby, E.D. Cokelet, S.L. Danielson, A.D. Robertis, L.B. Eisner, E.V. Farley, K. Iken, K.J. Kuletz, R.R. Lauth, E.A. Logerwell, and A.I. Pinchuk. 2016. Late summer open water zoogeography of the northern Bering and Chukchi seas. Deep Sea Research Part II: Topical Studies in Oceanography. https://doi.org/10.1016/j.dsr2.2016.03.005

Sill, L.A., G. Halas, and D. Koster. 2019. Copper River Chinook salmon: the intersection of commercial fisheries and the subsistence way of life in Cordova, Alaska. Alaska Department of Fish and Game, Division of Subsistence, Technical Paper No. 444, Anchorage.

Sill, L.A. and J.M. Van Lanen. 2022. Local and traditional knowledge of Chilkat Chinook Salmon. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 463, Anchorage.

Simeone, W.E. and K. James. 2002. Copper River Subsistence Evaluation 2000 & Traditional Knowledge Project, Part One. Alaska Department of Fish and Game, Division of Subsistence, Final Report No. FIS 00-040, Anchorage, Alaska.

Simeone, W.E., and E.McC. Valentine, 2007. Ahtna knowledge of long-term changes in salmon runs in the Upper Copper River drainage, Alaska. Alaska Department of Fish and Game, Division of Subsistence Technical Paper No. 324. Juneau.

Sisk, J., 1991. The Southeastern Alaska Salmon Industry: Historical Overview and Current Status 1–15. Southeast Alaska Conservation Assessment, Chapter 9.5. 15 p.

Sloat, M.R., G.H. Reeves, and K.R. Christiansen. 2016. Stream network geomorphology mediates predicted vulnerability of anadromous fish habitat to hydrologic change in southeast Alaska. Global Change Biology 604–620. https://doi.org/10.1111/gcb.13466

Smith, C.T., R.J. Nelson, C.C. Wood, and B.F. Koop. 2001. Glacial biogeography of North American Coho salmon (*Oncorhynchus kisutch*). Molecular Ecology 10: 2775-2785.

Smith, C.T., W.D. Templin, J.E. Seeb, and L.W. Seeb. 2005. Single Nucleotide Polymorphisms Provide Rapid and Accurate Estimates of the Proportions of U.S. and Canadian Chinook Salmon Caught in Yukon River Fisheries. North American Journal of Fisheries Management 25:944–953. https://doi.org/10.1577/m04-143.1

Smoker, W.W., A.J. Gharrett, M.S. Stekoll, and J.E. Joyce. 1994. Genetic analysis of size in an anadromous population of pink salmon. Canadian Journal of Fisheries and Aquatic Sciences 51:S9–S15.

Smoker, W.W., A.J. Gharrett, and M.S. Stekoll. 1998. Genetic variation of return date in a population of pink salmon: a consequence of fluctuating environment and dispersive selection? Alaska Fishery Research Bulletin 5:46–54.

Smoker, W.W., A.J. Gharrett, M.S. Stekoll, and S.G. Taylor. 2000. Genetic variation of fecundity and egg size in anadromous pink salmon (*Oncorhynchus gorbuscha*) Walbaum. Alaska Fishery Research Bulletin 7(1):44-50.

Somerville, M.A., and T.R. Hansen. 2021. Fishery management report for the recreational, personal use, and subsistence fisheries of the Upper Copper/Upper Susitna Management Area, 2019. Alaska Department of Fish and Game, Fishery Management Report No. 21-07, Anchorage. Springer, A.M., C.P. McRoy, and K.R. Turco. 1989. The paradox of pelagic food webs in the northern Bering Sea—II. Zooplankton communities. Continental Shelf Research 9:359–386. https://doi. org/10.1016/0278-4343(89)90039-3

Springer, A.M., and C.P. McRoy. 1993. The paradox of pelagic food webs in the northern Bering Sea: Patterns of primary production. Continental Shelf Research 13:575–599. https://doi. org/10.1016/0278-4343(93)90095-f

Springer, A.M., G.B. van Vliet. 2014. Climate change, pink salmon, and the nexus between bottom-up and top-down forcing in the subarctic Pacific Ocean and Bering Sea. Proceedings of the National Academy of Sciences 111:E1880–E1888. <u>https://doi.org/10.1073/</u>pnas.1319089111

St. Saviour, A., A.W. Barclay, and N. Logelin. 2020. Northern Cook Inlet Chinook salmon marine harvest stock composition, 2016–2017. Alaska Department of Fish and Game, Fishery Data Series No. 20-27, Anchorage.

Stachura, M.M., T.E. Essington, N.J. Mantua, A.B. Hollowed, M.A. Haltuch, P.D. Spencer, T.A. Branch, and M.J. Doyle. 2014. Linking Northeast Pacific recruitment synchrony to environmental variability. Fisheries Oceanography 23:389–408. https://doi.org/10.1111/ fog.12066

Stopha, M. 2012. An evaluation of the Port Graham salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J12-28, Anchorage.

Stopha, M. 2012. An evaluation of the Trail Lakes salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J12-21, Anchorage.

Stopha, M. and J. Musslewhite. 2012. An evaluation of the Tutka Bay Lagoon salmon hatchery for consistency with statewide policies and prescribed management practices. Alaska Department of Fish and Game Division of Commercial Fisheries, Regional Information Report 5J12-05, Anchorage.

Stopha, M. 2013. An evaluation of the Armin F. Koernig salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J13-11, Anchorage.

Stopha, M. 2013. An evaluation of the Cannery Creek salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J13-06, Anchorage.

Stopha, M. 2013. An evaluation of the Eklutna salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J13-02, Anchorage.

Stopha, M. 2013. An evaluation of the Gulkana salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J13-05, Anchorage.

Stopha, M. 2013. An evaluation of the Main Bay salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J13-07, Anchorage.

Stopha, M. 2013. An evaluation of the Solomon Gulch salmon hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J13-04, Anchorage.

Stopha, M. 2013. An evaluation of the Wally Noerenberg Hatchery for consistency with statewide policies and prescribed management practice. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report 5J13-10, Anchorage.

Stram, D.L., and J.N. Ianelli. 2009. Eastern Bering Sea Pollock Trawl Fisheries: Variation in Salmon Bycatch over Time and Space. American Fisheries Society Symposium 70:827–850.

Stram, D.L., and J.N. Ianelli. 2015. Evaluating the efficacy of salmon bycatch measures using fishery-dependent data. ICES Journal of Marine Science 72:1173–1180. https://doi.org/10.1093/icesjms/ fsu168

Sturdevant, M.V., M.F. Sigler, and J.A. Orsi. 2009. Sablefish predation on juvenile Pacific salmon in the coastal marine waters of Southeast Alaska in 1999. Transactions of the American Fisheries Society, 138(3):675-691.

Sturdevant, M.V., E. Fergusson, N. Hillgruber, C. Reese, J. Orsi, R. Focht, A. Wertheimer, and B. Smoker. 2012. Lack of trophic competition among wild and hatchery juvenile chum salmon during early marine residence in Taku Inlet, Southeast Alaska. Environmental Biology of Fishes 94: 101–116. <u>https://doi.org/10.1007/</u> s10641-011-9899-7.

Sturdevant, M.V., R. Brenner, E.A. Fergusson, J.A. Orsi, and W.R. Heard. 2013. Does Predation by Returning Adult Pink Salmon Regulate Pink Salmon or Herring Abundance? North Pacific Anadromous Fish Commission Technical Report No. 9:153-164.

Su, Z., and M.D. Adkison. 2002. Optimal in-season management of pink salmon (*Oncorhynchus gorbuscha*) given uncertain run sizes and seasonal changes in economic value. Canadian Journal of Fisheries and Aquatic Sciences 59:1648–1659. https://doi. org/10.1139/ f02-133

T.

Tadokoro, K., Y. Ishida, N.D. Davis, S. Ueyanagi, and T. Sugimoto. 1996. Change in chum salmon (*Oncorhychus keta*) stomach contents associated with fluctuation of pink salmon (*O. gorbuscha*) abundance in the central subarctic Pacific and Bering Sea. Fisheries Oceanography 5:89–99.

Tarpey, C.M., J.E. Seeb, G.J. McKinney, W.D. Templin, A.V. Bugaev, S. Sato, and L.W. Seeb. 2017. SNP data describe contemporary population structure and diversity in allochronic lineages of pink salmon (*Oncorhynchus gorbuscha*). Canadian Journal of Fisheries and Aquatic Sciences Online 15 August 2017. doi:10.1139/ cjfas-2017-0023.

Templin, W.D., 2001. The history of propagation and transportation of Chinook salmon *Oncorhynchus tshawytscha* stocks at hatcheries in Southeast Alaska, 1972 – 1998. Alaska Department of Fish and Game, Regional Information Report 5J01-05, 48 pp.

Templin, W.D., J.E. Seeb, J.R. Jasper, A.W. Barclay, and L.W. Seeb. 2011. Genetic differentiation of Alaska Chinook salmon: the missing link for migratory studies. Molecular ecology resources 11 Suppl 1, 226–246. https://doi.org/10.1111/j.1755-0998.2010.02968.x

Thedinga, J.F., A.C. Wertheimer, R.A. Heintz, J.M. Maselko, and S.D. Rice. 2000. Effect of stock, coded-wire tagging, and transplant on straying of pink salmon (*Oncorhynchus gorbuscha*) in southeastern Alaska. Canadian Journal of Fisheries and Aquatic Sciences 57:2076–2085.

Theriault, V., G.R. Moyer, L.S. Jackson, M.S. Blouin, and M.A. Banks. 2011. Reduced reproductive success of hatchery coho salmon in the wild: insights into most likely mechanisms. Molecular Ecology, 20(9):1860-1869.

Tiegs, S.D., D.T. Chaloner, P. Levi, J. Rüegg, J.L. Tank, and G.A. Lamberti. 2008. Timber harvest transforms ecological roles of salmon in southeast Alaska rain forest streams. Ecological Applications 18:4–11. https://doi.org/10.1890/07-0655.1

Tiernan A., T. Elison, T. Sands, and J. Head. 2022. Overview of the Bristol Bay commercial salmon fishery, 2019–2022: a report to the Alaska Board of Fisheries. Alaska Department of Fish and Game, Special Publication No. 22-17, Anchorage.

Trainor, A. B.M. McDavid, L.A. Sill, and L.S. Naaktgeboren. 2019. Local and Traditional Knowledge of Freshwater Life Stages of Chinook and Chum Salmon in Anvik, Huslia, Allakaket, and Fort Yukon. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 447, Fairbanks.

Trainor, A., D. Gerkey, B.M. McDavid, H.S. Cold, J. Park and D.S. Koster. 2021. How Subsistence Salmon Connects Households and Communities: an exploration of Salmon Production and Exchange Networks in Three Communities on the Yukon River, 2018-2019. Alaska Department of Fish and Game Division of Subsistence, Technical Paper No. 481, Fairbanks.

Trudel, M. and M.E. Thiess. 2007. Regional Variation in the Marine Growth and Energy Accumulation of Juvenile Chinook Salmon and Coho Salmon along the West Coast of North America. American Fisheries Society Symposium 57:205–232.

Twardek W.M., N.W.R. Lapointe, and S.J. Cooke. 2022. High egg retention in Chinook Salmon *Oncorhynchus tshawytscha* carcasses sampled downstream of a migratory barrier. Journal of Fish Biology 100:715–726.

Twardek, W.M. 2022. Evaluating the consequences of physical barriers on fish during long-distance upstream migrations through rivers. Thesis Carleton University.

U

United States and Canada. 2006. Potential causes of size trends in Yukon River Chinook salmon populations. Regional Information Report No. 3A06-07. Yukon River Joint Technical Committee.

Urawa, S., K. Nagasawa, L. Margolis, and A. Moles. 1998. Stock identification of chinook salmon (*Oncorhynchus tshawytscha*) in the north Pacific Ocean and Bering Sea by parasite tags. North Pacific Anadromous Fisheries Commission Bulletin 1:199–204.

Urawa, S., S. Sato, P.A. Crane, and B. Agler. 2009. Stock-specific ocean distribution and migration of chum salmon in the Bering Sea and North Pacific Ocean. North Pacific Anadromous Fish Commission Bulletin 5:131–146.

Utter, F.M., M.V. McPhee, and F.W. Allendorf. 2009. Population genetics and the management of Arctic-Yukon-Kuskokwim salmon populations. American Fisheries Society Symposium 70:97-123.

V

Vallion, A.C., A.C. Wertheimer, W.R. Heard, and R.M. Martin. 1981. Summary of data and research pertaining to the pink salmon population at Little Port Walter, Alaska, 1964-80. NWAFC Processed Report 81-10.

von Biela, V.R., L. Bowen, S.D. McCormick, M.P. Carey, D.S. Donnelly, S. Waters, A.M. Regish, S.M. Laske, R.J. Brown, S. Larson, S. Zuray, and C.E. Zimmerman. 2020. Evidence of prevalent heat stress in Yukon River Chinook salmon. Canadian Journal of Fisheries and Aquatic Sciences. 77(12):1878-1892. <u>https://doi.org/10.1139/</u> cjfas-2020-0209

Vega, S.L., T.M. Sutton, and J.M. Murphy. 2016. Marine-entry timing and growth rates of juvenile Chum salmon in Alaska waters of the Chukchi and northern Bering seas. Deep-Sea Research II 135:137-144.

Vega, S.L., J.M. Head, T. Hamazaki, J.W. Erickson, and T.R. McKinley. 2022. Review of salmon escapement goals in Bristol Bay, Alaska, 2021. Alaska Department of Fish and Game, Fishery Manuscript No. 22-07, Anchorage.

Volkov, A.F. 2022. Appendicularia in the Bering, Okhotsk, Chukchi Seas and North Pacific and their significance for feeding nekton. Izv. Tikhookean. Nauchno-Issled. Inst. Rybn. Khoz. Okeanogr., 202(2):390–408. DOI: 10.26428/1606-9919-2022-202- 390-408. EDN: BXOLJN.

von Biela V.R., C.J. Sergeant, M.P. Carey, Z. Liller, C. Russell, S. Quinn-Davidson, P.S. Rand, P.A.H. Westley, C.E. Zimmerman. 2022. Premature Mortality Observations among Alaska's Pacific Salmon During Record Heat and Drought in 2019. Fisheries 47:157–168.

Vulstek, S.C., J.R. Russell, J.E. Joyce, and A.K. Gray. 2022. 2017 Auke Creek research station report: data summary and historical trends from 1980 to 2017. NOAA Technical Memorandum NMFS-AFSC-436.

W

Wadle, J., and J. Baumer. 2023. Chignik River Sockeye Salmon Stock of Concern Action Plan. Oral Report to Alaska Board of Fisheries. Alaska Department of Fish and Game.

Walker, R., K. Myers, N.D. Davis, K.Y. Aydin, K.D. Friedland, H.R. Carlson, G.W. Boehlert, S. Urawa, Y. Ueno, and G. Anma. 2000. Diurnal variation in thermal environment experienced by salmonids in the North Pacific as indicated by data storage tags. Fisheries Oceanography 9: 171–186.

Walsey, V., and J. Brewer. 2018. Managed out of existence: over-regulation of Indigenous subsistence fishing of the Yukon River. GeoJournal 83:1169–1180. <u>https://doi.org/10.1007/</u> s10708-018-9879-y

Wang, I.A., E.H. Leder, W.W. Smoker, and A.J. Gharrett. 2006. Timing of development during epiboly in embryos of second-generation crosses and backcrosses between odd- and even-broodyear pink salmon, *Oncorhynchus gorbuscha*. Environmental Biology of Fishes 75:325–332.

Wang, J., J. Zhang, E. Watanabe, M. Ikeda, K. Mizobata, J.E. Walsh, X. Bai, and B. Wu. 2009. Is the dipole anomaly a major driver to record lows in arctic summer sea ice extent? Geophysical Research Letters 36:1–5. https://doi.org/10.1029/2008GL036706

Wang, M., J.E. Overland, and P. Stabeno. 2012. Future climate of the Bering and Chukchi Seas projected by global climate models. Deep-Sea Research Part II: Topical Studies in Oceanography 65–70, 46–57. https://doi.org/10.1016/j.dsr2.2012.022

Waples, R., 2009. Conserving the evolutionary legacy of Arctic-Yukon-Kuskokwim salmon. in Krueger, Charles & Zimmerman, Christian. 2009. Pacific salmon: ecology and management of western Alaska's populations. pp 125–139.

Wechter, M.E., B.R. Beckman, A.G. Andrews, A.H. Beaudreau, and M.V. McPhee. 2017. Growth and condition of juvenile chum and pink salmon in the northeastern Bering Sea. Deep-Sea Research II 135:145-155.

Weingartner, T.J., S.L. Danielson, and T.C. Royer. 2005. Freshwater variability and predictability in the Alaska Coastal Current. Deep-Sea Research Part II: Topical Studies in Oceanography 52:169–191. https://doi.org/10.1016/j.dsr2.2004.09.030

Weitkamp, L.A., J.A. Orsi, K.W. Myers, and R.C. Francis. 2011. Contrasting Early Marine Ecology of Chinook Salmon and Coho Salmon in Southeast Alaska: Insight into Factors Affecting Marine Survival, Marine and Coastal Fisheries, 3(1):233-249, DOI:10.1080 /19425120.2011.588919.

Welch, D.W., A.D. Porter, and E.L. Rechisky. 2020. A synthesis of the coast-wide decline in survival of West Coast Chinook salmon. Fish and Fisheries DOI:10.1111/faf.12514.

Wells, B.K., D.D. Huff, B.J. Burke, R.D. Brodeur, J.A. Santora, J.C. Field, K. Risherson, N.J. Mantua, K.L. Fresh, M.M. McClure. W.H. Satterthwaite, F. Darby, S.J. Kim, R.W. Zabel, and S.T. Lindley. 2020. Implementing ecosystem-based management principles in the design of a salmon ocean ecology program. Frontiers in Marine Science doi: 10.3389/fmars.2020.00342.

Wertheimer, A.C., J.A. Orsi, E.A. Fergusson, and M.V. Sturdevant. 2013. Forecasting pink salmon harvest in Southeast Alaska from juvenile salmon abundance and associated biophysical parameters: 2012 returns and 2013 forecast. North Pacific Anadromous Fish Commission Document 1486. 23 pp.

Wessel, M.L., W.W. Smoker, and J.E. Joyce. 2006. Variation of morphology among juvenile chinook salmon of hatchery, hybrid, and wild origin. Transactions of the American Fisheries Society 135:333–340. https://doi.org/10.1577/T04-078.1

Whittle, J.A., C.M. Kondzela, H.T. Nguyen, K. Hauch, D. Cuadra, and J.R. Guyon. 2018. Genetic stock composition analysis of Chum Salmon from the prohibited species catch of the 2016 Bering Sea Walleye Pollock trawl fishery and Gulf of Alaska groundfish fisheries. 56. https://doi.org/10.7289/V5/TM-AFSC-314

Whittle, J.A., Kondzela, C.M., Watson, J.T., Barry, P.D., Nguyen, H.T., Yasumiishi, E.M., Nicolls, D., Larson, W.A., 2021. Genetic Stock Composition Analysis of Chum Salmon from the Prohibited Species Catch of the 2018 Bering Sea Walleye Pollock Trawl Fishery and Gulf of Alaska Groundfish Fisheries. U. S. Dep. Commer., NOAA Tech. Memo. NMFS-AFSC-, 81 p.

Williams, D.L., and C.A. Shelden. 2010. Kogrukluk River weir salmon studies, 2008. Alaska Department of Fish and Game, Fishery Data Series No. 10-24, Anchorage.

Williams, L., P. Coiley-Kenner, and D. Koster. 2010. Subsistence harvests and uses of salmon, trout, and char in Akhiok, Larsen Bay, Old Harbor, Ouzinkie, and Port Lions, Alaska, 2004 and 2005. Alaska Department of Fish and Game, Division of Subsistence, Technical Paper No. 329, Anchorage.

Williams, D.L., and C.A. Shelden. 2011. Kogrukluk River salmon studies, 2010. Alaska Department of Fish and Game, Fishery Data Series No. 11-49, Anchorage.

Wilson, L. 2023. Alaska salmon fisheries enhancement annual report 2022. Alaska Department of Fish and Game, Division of Commercial Fisheries, Regional Information Report No. 5J23-04, Juneau.

Wipfli, M.S., 1997. Terrestrial invertebrates as salmonid prey and nitrogen sources in streams: contrasting old-growth and young-growth riparian forests in southeastern Alaska, U.S.A. Canadian Journal of Fisheries and Aquatic Sciences 54:1259–1269. https://doi.org/10.1139/f97-034.

Wipfli, M.S., J. Hudson, and J. Caouette. 1998. Influence of salmon carcasses on stream productivity: response of biofilm and benthic macroinvertebrates in southeastern Alaska. Canadian Journal of Fisheries and Aquatic Sciences, https://doi.org/10.1139/f98-031.

Wipfli, M.S., J.P. Hudson, D.T. Chaloner, and J.P. Caouette. 1999. Influence of salmon spawner densities on stream productivity in Southeast Alaska. Canadian Journal of Fisheries and Aquatic Sciences 56:1600–1611. https://doi.org/10.1139/f99-087.

Wipfli, M.S., and D.P. Gregovich. 2002. Export of invertebrates and detritus from fishless headwater streams in southeastern Alaska: Implications for downstream salmonid production. Freshwater Biology 47:957–969. https://doi.org/10.1046/j.1365-2427.2002.00826.x.

Witherell, D., D. Ackley, and C. Coon. 2002. An Overview of Salmon Bycatch in Alaska Groundfish Fisheries. Alaska Fishery Research Bulletin 9:53–64.

Witteveen, M.J., and K. Shedd. 2016. Chinook salmon genetic sampling along the Alaska Peninsula and adjacent areas results, 2012–2014. Alaska Department of Fish and Game, Fishery Data Series No. 16-25, Anchorage.

Wolfe, R.J., and J. Spaeder. 2009. People and salmon of the Yukon and Kuskokwim drainages and Norton Sound in Alaska: fishery harvests, culture change, and local knowledge systems. American Fisheries Society Symposium 70:349-379.

Woodgate, R.A., K. Aagaard, and T.J. Weingartner. 2006. Interannual changes in the Bering Strait fluxes of volume, heat and freshwater between 1991 and 2004. Geophysical Research Letters 33:2–6. https://doi.org/10.1029/2006GL026931

Woody, C.A., editor. 2018. Bristol Bay Alaska: natural resources of the aquatic and terrestrial ecosystems. J. Ross Publishing, Plantation, Florida. 589 pp.

Woody, C.A., J. Olsen, J. Reynolds, and P. Bentzen. 2000. Temporal variation in genotypic and phenotypic traits of two sockeye salmon populations. Transactions of the American Fisheries Society 129:1031-1043.

Υ

Yasumiishi, E.M., K.R. Criddle, N. Hillgruber, F.J. Mueter, and J.H. Helle. 2015. Chum salmon (*Oncorhynchus keta*) growth and temperature indices as indicators of the year-class strength of age-1 walleye pollock (*Gadus chalcogrammus*) in the eastern Bering Sea. Fisheries Oceanography 24:242–256. <u>https://doi.org/10.1111/</u> fog.12108

Yasumiishi, E.M., E.V. Farley, G.T. Ruggerone, B.A. Agler, and L.I. Wilson. 2016. Trends and factors influencing the length, compensatory growth, and size-selective mortality of juvenile Bristol Bay, Alaska, sockeye salmon at sea. Marine and Coastal Fisheries: Dynamics, Management, and Ecosystem Science 8:315-333. DOI: 10.1080/19425120.2016.1167793

Yasumiishi, E.M., E.V. Farley, Jr., J. Maselko, K.Y. Aydin, et al. 2019. Differential north-south response of juvenile Chinook salmon marine growth to ecosystem change in the eastern Bering Sea, 1974 - 2010. ICES Journal of Marine Science, doi:10.1093/icesjms/ fsz166

YRJTC. 2006. Potential causes of size trends in Yukon River Chinook salmon populations. RIP No. 3A06-07

Ζ

Zimmermann, H.H., et al. 2023. Marine ecosystem shifts with deglacial sea-ice loss inferred from ancient DNA shotgun sequencing. Nature Communications: 14, Article number: 1650. https://doi. org/10.1038/s41467-023-36845-x

Zuray, S., R. Kocan, and P. Hershberger. 2012. Synchronous Cycling of Ichthyophoniasis with Chinook Salmon Density Revealed during the Annual Yukon River Spawning Migration. Transactions of the American Fisheries Society 141:615–623. <u>https://doi.org/10.108</u> 0/00028487.2012.683476

Appendix 4: Public Testimony

Public testimony was gathered through the Alaska Salmon Research Task Force website and also via email exchange with the Alaska Fisheries Science Center Communications staff and Ed Farley.

Testimony from the website.

Public Input on Alaska Salmon Research Task Force Report

Website questionnaire

In response to unprecedented declines in chum and Chinook salmon on the Yukon and Kuskokwim rivers, the Alaska Salmon Research Task Force was created by an act of Congress.

The purpose of the Alaska Salmon Research Task Force is to compile science and Traditional Knowledge, to identify what is known about salmon in Alaska, data gaps, and needed research. This information will be used to develop a collaborative science plan for sustainable salmon management in Alaska. We greatly value your input in the development of this plan.

Task Force meetings are open to the public. Time has been reserved at the end of each monthly Task Force meeting for public comment. In addition, the Task Force will be holding a 2-day public meeting on November 14-15, 2023 in Anchorage. Public comment will be accepted from 1-4:30 pm (Alaska Time) on both days. Participants will have the option to join in person, virtually or via conference call.

If you would like to provide written comment(s) to inform the Task Force discussions and the development of its collaborative science plan, please provide information via this feedback form. The deadline for sub mitting input is March 15, 2024.

EmailValid email What is your name (First and Last)? Short answer text

What information or data (e.g., salmon migration, seasonal distribution, abundance and trends etc.) would you like to make the Task Force aware of?

Long answer text

What are the data/research gaps you believe should be addressed for understanding productivity trends in Alaska salmon?

Long answer text

What additional research is needed to improve understanding of Alaska salmon productivity and abundance trends?

Long answer text

What does a coordinated salmon research strategy that addresses increased variability or declines in Pacific salmon returns to Alaska look like to you?

Long answer text

Anything else you want to share with the Task Force? Final thoughts, comments, questions or requests for special accommodations.

Long answer text

Public Response to Website questionnaire

10/14/2023 5:38:16 jonathan.salmon@igiugig.gov Jonathan Salmon

Lack of Chums. No one has noticed that the Kvichak River and Bristol Bay is also missing Chum Salmon. The oversight due to the abundance of Sockeye. There is a key component missing when one begins to turn up missing. I think salmon spawning grounds need to be researched as well as smolt out migration. I can't speak to what the research would like like. But I do think that it has to include local voice. There is work being done with a phone app to be able to submit information and I have used that. I will come up with more after meeting with a team from the Kvichak.

10/17/2023 22:28:45 bobshavelson@gmail.com Bob Shavelson

The stream temperature data collected by Cook Inletkeeper and others is vital to understand if state water quality standards are being met, or not, and if they are not, the state should enforce the Clean Water Act and develop TMDL's and other controls for thermal inputs. The biggest gaps to address are the effects of hatchery fish on the food and habitat needs of wild fish in the ocean, and the impacts of factory trawlers and catcher processors have on wild stocks. What effect do hatchery fish in the north pacific have on wild Alaskan King and other salmon? What effects does trawling have on wild Alaskan salmon? Among other things, it means discarding the antiquated and unworkable fallacy of maximum sustainable yield, and devising a management regime for all fisheries that recognizes the inherent limits of natural capacity. The state likes to boast of it's "world class management regime," yet Alaska is repeating the very same mistakes that have lead to the declines and closures of fisheries across the planet. Now, as temperatures rise, a truly sustainable management program must embrace a precautionary approach to fish management which does not allow profits to drive decision making.

10/24/2023 16:10:41

mclinton2@alaska.edu Morag Clinton

I think there should be mention of emerging disease and the impacts of altered host/pathogen dynamics in the face of climate change. Many publications note altered host pathogen dynamics in varied environmental conditions, with warming considered to increase host susceptibility to disease in many instances (REF1). Warm waters and other consequences of climate change are known stressors to Pacific salmon, likely with knock-on immunological consequences for disease susceptibility. This is likely to be particularly pronounced in response to novel diseases in Alaska waters, for which likely lack native immunity to. For example, Tetracapsuloides bryosalmonae, an emerging disease in Alaskan Pacific salmon, even reduces the thermal tolerance of infected fish (REF2).

While there is currently little literature regarding the impact of disease on Alaskan productivity trends outside of a hatchery setting, that does not mean it is not happening. It just means we are not studying it.

REF1 = https://digitalcommons.unl.edu/parasitologyfacpubs/606

REF2 = https://doi.org/10.1111/1365-2435.12701

We need to be conducting 1. Disease surveillance research that is proactive, no reactive. We cannot safeguard Alaska salmon stocks if every emerging disease is a surprise, but if we identify emerging disease in a timely manner and plan accordingly, we can have responses in place for disease events. 2. We need to be studying the immunological function and response of Pacific salmon to specific disease agents in different climate scenarios to understand the potential impacts of these diseases. These studies need to use Alaskan fish. 3. We need to be identifying 'at risk' populations of fish and carefully monitoring them for disease. 4. We need to be including impact of disease in management modelling so events such as a disease outbreak do not impact escapement. It needs varied perspectives including strong Indigenous Alaskan participation to ensure all voices are heard.

10/25/2023 11:13:10 cewan@chitina.org Corina Ewan

seasonal distribution The first run of fish gets to Copper River (Ahtna) people in July The first run of fish used to get to Copper River people by May every year. Now the first run comes in July. This depletes Ahtna's traditional way of sharing; leaving elders, disabled people and young ones to be hungry because there is not enough time or fish to prepare and share. Gives hope for increase salmon returns to Alaska.

The commercial fishery needs to meet the subsistence fishery half way on sustaining our fish for all who are involved. Make committee/board seats half commercial and half subsistence users so they can educate each other and sustain our fish.

10/25/2023 12:09:07 tribaladministrator@chitina.org Dean Barlip

Salmon Migration Commercial Fishermen vessels are allowed to be too close to the mouth of major inland rivers. They are taking fish that the US GOV under ANICLA VIII promised to protect. There have been numerous villages that had historically low fishing numbers. I believe a large portion of this falls to the need for more fish to be allowed upriver for subsistence and sustaining the species.

10/31/2023 15:36:13 kbayconservation@gmail.com Penelope Haas

We believe that this study should encompass not only data gaps, but an analysis of what data is and isn't being used to make management decisions. So for example, from our perspective, there is mountain of data out there on hatchery impacts and climate trends that ADF&G is ignoring. Not only do they not use this data, the do not have a system in place to determine what data they do and do not use or how it should be used. The problem here is not a data gap per se as much as a problem of policies not reflecting data. To build in that, we need both greater transparency in decision-making--the public should be able to see what data is being used (just like how ADF&G shows escapement numbers)--and a better data organizational system so previously-collected data is easier to access. We need more studies on the near-shore impacts of hatchery releases on wild populations.

10/31/2023 16:52:29 ferrariangela@ymail.com Angela Ferrari

The amount of salmon bycatch reported by NOAA weekly Commercial trawler salmon bycatch. NOAAs weekly bycatch reports are staggering for salmon and other aquatic species. All aspects need to be considered as well as the effect on fish from commercial trawlers. The amount of bycatch going on in Alaskan waters is having a very negative effect on our fish and marine mammals. Overfishing and bycatch is causing declines in fish stocks as well as starvation for some species.

"Bycatch is under reported and has not been an addressed issue to the decline of salmon. Chum salmon do not even have a cap on them which has adversely affected the Yukon Delta area and all 56 communities along the Kuskokwim river. Observers are not on 100% of the vessels like they need to be. Furthermore, most observers end up working for the fisheries after the observation

position is over and are approached for positions with the fisheries while observing. So there is a question of observer honesty and accuracy there as well for a lot of us. Another detrimental aspect to commercial trawling is the decimation of habitat for many species as well as salmon. It takes years for a trawled ecosystem to recover. They do not need to drag the bottom of the ocean. Recent studies in the waters by Kodiak have raised this concern."

11/1/2023 11:07:16 pattydavedavenport@hotmail.com David Davenport

The King Salmon return to the Goodpaster River in the eastern interior of Alaska has continually declined over the years and the Pogo gold mine since the start of operation is allowed to dump there treated waste water outflow back into the river. This cannot be good for the ecosystem and salmon smolt, fry, eggs, etc. They have over 600 employees in the camp and they have a sewage settling pond that has overflowed several times from flooding back into the river. That is untreated human waste! Need more study in prime spawning grounds. More water quality testing on a regular basis by independent contractors not in the pocket of large corporate mineral developmers. YRDFA, a mix of diversity of knowledgeable user groups being listened to more by government agencies. Stop polluting or rivers, more commercial fishery by catch laws and stronger fines. More EPA oversight

11/1/2023 12:53:42 rachel.christiansen@wsu.edu Rachel Zander

Take a look at what Bristol Bay has done. Talk to biologist like Tim Sands - there you will find your answers and research priorities that should be applied to the YK. See above. You already know the answer to this. And if NOAA doesn't, which I know it does, then go ask all the ADF&G biologists.

More ADF&G oversight and leas industry involvement and oversight. Why are trawl representatives allowed on a SALMON RESEARCH task force? You have folks from the At Sea Processors Association, Trident, and a CVRF executive (AKA the CDQ with the heaviest pollock investment in the Bering Sea) on this task force - what do you think will happen? You have some of the same people here that have been on the NPFMC for years. You think you're going to come up with new solutions or answers? We aren't stupid. We know where this is heading. It is yet another waste of taxpayer dollars. It is -a risk mitigation and public relations campaign masquerading as a "research" task force.

If NOAA really wants to solve these problems, then look outside NOAA personnel. Recruit more ADF&G biologists, involve non-CDQ tribal representatives who are further inland and more greatly affected by the starvation and theft of salmon in the Bering Sea. Involve SALMON commercial interests - NOT POLLOCK. Once NOAA realizes the regulatory capture it faces with the pollock industry and strives to change that, the public is never going to trust this tax-payer funded organization under the Department of Commerce. Unless we rewrite the MSA and stop prioritizing OPTIMUM YIELD, things are never going to change, even if salmon stocks collapse, because pollock interest will continue to persist and pervade this organization like the cancer that it is."

11/1/2023 14:36:39 jschroeder1@une.edu Jayme Schroeder

Trawl bycatch I believe the data is already abundantly clear. Industrial scale trawling is destroying salmon stocks and prey species. It is a poor fishing practice that destroys the entire ecosystem to harvest a mediocre resource. It needs to stop. Stop trawling and let the stocks return. That would be quite the study!

A study that can make a change. We know that trawling is a major concern for salmon bycatch. Why is it allowed when subsistence fishing is shut down? It's criminal.

Please look critically at the numbers. Bycatch might be a small percentage of overall trawl catch but it is massive tonnages. We have to stop supporting and subsidizing such a wasteful fishery.

11/2/2023 9:05:56 james.savereide@alaska.gov James Savereide

High-water events Neuswanger et al (2015) showed a strong correlation between summer discharge and low productivity The same methodology should be applied to other import salmon streams because in my experience we have seen more high-water events in the last 15 years versus the 15 before that. I really think we just need to take more action but politics is in the way. Thanks for the opportunity to comment.

11/3/2023 16:28:20 mdadkison@alaska.edu Milo Adkison

I think the focus on variability and declines is too narrow. The human dimensions aspect, particularly adaptation to changes in salmon production, is more important than understanding why the changes are happening. I'd like to see some consideration of management approaches that facilitate adaptation, and don't lock stakeholders into dependence on a single variable stock either through regulation or significant financial barriers to diversification

11/3/2023 19:20:44 sparrowfisheriesllc@gmail.com Jaymi Bethea

Salmon migration and bycatch The true bycatch count on the Golf and Bering Sea trawlers A change to 2 observers on each trawler so crewmen can't throw bycatch over behind the observers backs. Shutting down trawl fisheries if they excess allowed bycatch rates for salmon, this allowable bycatch amount needs to be lowered. Why are we wasting huge amounts of resources with the allowable bycatch in the Bering and Golf trawl fisheries, they need to be reanalyzed.

11/3/2023 19:52:50 lunatefisheries@gmail.com Hayden Linscheid

The large amount of salmon bycatch due to trawlers What's happening to salmon while out at sea, where are they disappearing to more observers and regulations for the trawl fleet Shutting trawlers down I believe (and have seen) the damages trawlers cause to salmon and all species while harvesting cheap ground fish and pollock

11/5/2023 16:12:30 northernstar2017@gmail.com Ben Phillips

1. There is strong correlation between the eastern population growth of Stellar Sea lions and the decline of Chinook ocean survival rates across Alaska. In other words, the yearly rate of increase of the population of Stellar Sea Lions matches with the yearly rate of decline of chinook ocean survival rates and returning chinook adults. However, just because there may happen to be a correlation as previously described, that by itself does not prove that Stellar Sea Lions are to blame. Several other criteria must be met to show causation.

2. For instance, is the current Stellar population capable of consuming that much prey? Short answer: yes. A Stellar Sea lion eats 5%-8% of its body weight per day. That translates to 100-160 lbs of prey per day for a 2,000# Stellar, or 36,500-58,400 lbs per year. Multiply that by the number of Stellar Sea lions. Even if Chinook only make up a small fraction of a Stellar Sea lions diet, because of the sheer size of the current population they are capable of consuming enough chinook as prey to account for the chinook decline, even if it is only a small percentage.

3. Do Stellar foraging areas contain chinook? Short answer: yes. Most, if not all, of the current Stellar rookeries are located inside zones of high chinook abundance. This may also explain the smaller average size of returning adult chinook. 4. During the chinook life cycle while at sea, if chinook have to constantly make the choice between food and being eaten by Stellars, a prey fear response is triggered and the end result is smaller returning adult chinook and higher mortality rates (even if a chinook survives by constantly avoiding predators, it is more likely to be malnourished or become a mortality statistic. Referred to as non contact fear driven "additive mortality" caused by predation.)

So, we have 4 correlations regarding Stellars and Chinook decline. Just about as good as it gets in predictive fisheries management. A strong argument that Stellar Sea lions are most likely the cause of Chinook decline in Alaska. They are a growing fleet, fishing 365 days a year, without rules or limits. Based on the above figures, Stellar Sea lions are most likely currently consuming more chinook than all commercial, sport and subsistence fisheries in Alaska combined. By the way, citing work by Dr Walters and Dr Peter Olesiuk, that is already the case in BC. To say that pinnipeds are not a problem in Alaska for Chinook stocks today is just plain bad management. Just so I am clear: chinook salmon will not recover in Alaskan unless the pinniped population size is addressed.

Now there are 3 ways ADFG can respond to this:

Way #1: Continue like it is and keep cutting back or closing human fishing and hope that stocks somehow magically recover with pre-Anthropocene pinniped population levels.(chinook stocks won't recover this way. Continuing this current management will most likely lead to pinnipeds replacing all human take of chinook).

Way #2: ADFG gets congressional approval to lethally remove enough Stellars to allow for chinook recovery. (Yes this would work but be very difficult politically and would be wildly unpopular and most likely tank Alaska Seafood retail sales)

Way #3: State of Alaska takes over MMPA management of pinnipeds. The MMPA allows for this, but it has never been done. ADFG would expand its current pinniped management partnership with specific Alaska indigenous people to include all the indigenous people of Alaska and First Nations People of the Yukon and British Columbia and officially recognize the important role subsistence take of pinnipeds historically played in maintaining a heathy Salmon ecosystem by harvesting pinnipeds. ADFG would allow year around subsistence take, set harvest guidelines, etc (this is already being done in B.C.). ADFG could also issue a set amount of pinniped 'deprivation tags' to the partner indigenous managers that could be sold by the partner managers to Alaska Residents for subsistence (as needed for management population control) and to licensed commercial fishermen (for damage control).

Please see the 2021 Lecture given by Dr. Carl Walters of the University of British Columbia on "the role of pinnipeds in recent BC fisheries collapses". It gives credibility to my statements and cites peer reviewed published work on this subject. This is a Covid-19 protocol zoom meeting lecture recorded in 2021 and posted on YouTube. See: https://www.youtube.com/watch?v=4ZF_0npi8M0
Also, I would like to bring attention to the work of the Washington Acadamy of Science on pinniped predation of salmon. Gov. Jay Inslee commissioned the orca task force to find best available science to recover the endangered Southern Resident Killer Whale (SRKW). As part of that process, the Washington Acadamy of Science was contracted to find the best way to increase prey (chinook salmon). Three reports were issued by the Acadamy and they all dealt with pinniped impacts. The most recent (Nov 2022) is here: https://washacad.org/wp-content/uploads/2022/11/ Pinniped-Predation-on-Salmonids-in-the-Washington-Portionsof-the-Salish-Sea-and-Outer-Coast-1.pdf What was the pinniped population in 1880? How about 1920? How about at statehood? Just before the MMPA was passed? Where did NOAA managers come up with the target population models being used today for the eastern and western DPS of the Stellar Sea lion. An 8th grader could do the math using the same criteria Dr. Peter Olisiuk used (in his peer reviewed published work) to back-figure BC stellar populations to all the way back 1880. I did just that and came up with a very different, much lower figure than what NOAA managers are using. I would like to see a more scientifically supported method used to determine ocean carrying capacity of Stellar sea lions that INCUDES consideration of sustainable Alaska HUMAN fishing and ALL species of Stellar prey, especially chinook. #1 Priority: address predation by pinnipeds. There needs to be an open and frank discussion on the effects of the value (versus science based) MMPA, the original legislative intent of the MMPA, and how the current problems with pinnipeds were originally contemplated by congress. The truth is the result of the MMPA is not a success at all for Chinook. It is currently an unmitigated disaster for chinook. Further, it turns out to be a mistake to remove one of the co-evolved predators of pinnipeds from the ecosystem: MAN. Return the balance and Chinook will thrive. It's called science based, opposed to value based, science.

11/9/2023 15:36:46 gabe@yukonsalmon.org Gabe Canfield

Marine and Freshwater harvests include space for bycatch studies but do not include cross-species intercept fisheries such as the Area M fishery in Western Alaska. Looking at the intra-species harvest impacts are an important focus factor. In Hatchery-Wild salmon interactions, there are concerns in our communities among genetic biodiversity loss when hatchery fish are introduced. Research into genetics of hatchery salmon needs to be continued and expanded before any future increases to hatchery production moves forward. In freshwater habitat changes, the notable impact of climate change on freshwater habitat not only in streams but on banks and water tables in watersheds should be prioritized. On the Yukon River, wildfires impacted the upper Yukon in summer of 2023 and the lower Yukon in summer of 2022, and in some places where fires had never before been seen, impacting areas along the salmon-rearing habitat. Additionally, as climate change continues, plants migrate to areas where they have never populated before and create different and changing environments along banks, with more or less shade for rearing habitats changing the landscape. There is room for significant study in this arena.

There is currently no section on man-made disasters and impacts such as mine-tailings, oil spills, dam breakages, military superfund sites, construction over salmon habitat, water quality, or other human impacts. Research is required as a part of this process in prevention rather than cleanup of damaging environmental impacts. Each of these individual impacts could have their own research focus and study, and the inclusion of these impacts are a necessary part of the picture. Not just research but action must be taken at this critical stage. Research that focuses on prevention rather than disaster response in extreme salmon decline.

11/13/2023 23:51:50 gkvsons@alaska.net Gale K. Vick

The Task Force is likely aware of all available data. It is the unknown and unavailable data we need to be concerned with. There is often a grey area between suggestions for management vs. suggestions for research. We should look at both and see where they intersect. Timeliness is critical for so many managerial decisions but too often managers will take not action with the excuse we need more robust data. Our regulatory bodies, however, have requirements that they act on the best available data. This results in a very lop-sided decision making process if that data is not connecting the right dots. We see examples of this all the time when decisions to shut down one sector is not consistent with shutting down a related sector simply because we have not adequately identified or examine the correlation between data sets. One sector alone cannot bear the entire burden of salmon decline or stock rebuilding.

If we start with a true ""gravel to gravel"" concept the absolute has to be in following the path of a single salmon through its life cycle. That pathway will tell us where the data gaps are. But I am sure the Task Force knows this.

With that in mind, here are my thoughts:

(1) Develop comprehensive plan for identifying and monitoring AND protecting all primary and secondary salmon spawning habitat in the AYK

(2) Develop Run reconstruction for Kuskokwim River salmon, all species

(3) Research straying of hatchery Chinook from Whitehorse into Alaska streams

(4) Conduct complete and on-going genetics study of Alaska Peninsula, Area M intercept fisheries - all species, utilizing real-time data analysis to indicate relationship to AYK stocks (5) Increase monitoring of fish harvest in Area M, both on fishing grounds and within processing plants, all species, all sectors to increase confidence in harvest numbers

(6) Examine mortality on tagging studies, when and where the most mortality is likely

(7) Assess confidence level in juvenile salmon studies - in-river, estuary and marine and then increasing opportunity to get real data by using citizen's science, particularly in remote areas

(8) Conduct a real assessment of GOA hatchery impacts of pink and chum salmon production on wild salmon stocks

(9) Conduct a cost-benefit analysis of GOA hatcheries, assessing the true cost of each fish that is harvested

(10) Assess what stock abundance and harvest opportunity might occur if we got rid of all hatcheries

(1) Intensive research on gear impacts for both harvest, bycatch and discard

(2) More comprehensive water quality monitoring, especially around high human impact areas. Monitoring for PFAS, mining affluence, permafrost melting, storm drain run-off, temperature fluctuations, and so much more. And then a reporting system the public can access on a regular basis

(3) Genetics, genetics, genetics - assessing the genetics of all intercept and bycatch

(4) Fecundity, fecundity, fecundity - assessing the ASL of all spawning habitat Most salmon habitat in Alaska is remote and poorly understood. Developing a comprehensive Citizen's Science / Community Based Monitoring program would help acquire needed data as well as educating stakeholders. Additionally, co-management agreements, if monitored correctly, can go a long way in both protecting wild stocks and identifying en route and terminal problems.

(1) Far greater communication and coordination between state and federal management agencies

(2) Greater dialogue with sectors about joint solutions

(3) True ""gravel to gravel"" management for anadromous species

(4) Maximum usage of citizen's science and local and traditional knowledge and documentation linked to central data collection that can be accessed by all managers, across jurisdictions

(5) Increased co-management with stakeholders, tribal entities

(6) Increased communications system for stakeholders

(7) Enumeration of all suspected impacts on salmon decline and how to assign responsibility for those factors that we can change vs. those we cannot (8) Vastly increase public awareness of spawning and rearing habitat and create riparian zone regulations near that habitat to prevent or mitigate human impacts, in all seasons

(9) Consideration of increased salmon protected areas "One year to conduct assessment of research needs on salmon in Alaska is both unrealistic and ultimately not significantly useful to change. This is an on-going process and assessing and conducting needed research is part of the process to guide managers and stakeholders. There are many research venues, such as AYK-SSI, NPRB, PWSSC and others, that need to be linked in ways that go beyond funding opportunity.

Our salmon mess was created by a fragmented system that still gives agency managers plenty of opportunity to blame other sectors and not focus on the obvious. But one sector, one region alone cannot fix a systemic problem. We are literally all in this together.

We need historical reference going back at least 100 years. It's far too easy to put salmon decline blame on regime shifts but we've had these forever. Comparing data on climate along with managerial decisions of the time, harvest and escapement numbers is extremely helpful so that we do NOT shift our baselines, comparing data in very limited time periods. A huge factor in this is hatchery production.

Stakeholders often feel overwhelmed by conflicting research reporting and need a way to digest and interpret research data that is meaningful to the way that they address the managers and Legislature. Stakeholders know one true thing: the allocative battles between sectors relies on independent peer-reviewed research that can give all us a much clearer (and emerging) picture of impacts at all levels.

It seems we are going to have to micro-manage our salmon from now on if we are truly going to rebuild stocks. If that means ALL sectors and regions within a salmon's life cycle must stand down for so many years, then we should bite the bullet and do it. Wild salmon are too important to our ecosystem to allow them to disappear as they have done in so many other places around the world. Alaska has absolutely no excuses.

Here is the final story. Our Canadian counterparts on the Yukon River have been telling us for 20 years or more that not meeting escapement and ASL goals was going to put the rest of the Yukon, indeed, the entire line of passage, into the same crushing dilemma. We didn't listen. Now we are. And so should the rest of the state. Before it is too late to save discrete stocks, age classes and average sizes for salmon. Sadly, we are on a fast track for all of those losses.

11/14/2023 13:16:20 bear@alaska.net Nancy Hillstrand

marine competition from hatchery fish, and other predator prey assemblages. FOOD WEB biotic component included as critical in all habitat assessments, illegal foreign fishing fleets within 200 mile limit, targeting of Chinook smolt as a delicacy seen at Japanese fish markets Tokyo, comprehensive culvert failures assessment completed and begin following modern engineering practices and promote the dynamically stable ecological functions of the stream system at the specific road crossing location. Update obsolete memorandum of Agreement between ADFG and ADOT for modern engineering practices on culvert repair and maintenance. Mixed stock harvests must have mor monitoring and rapid immediate reporting. Hatchery fish in migrations require equation to estimate a statistically precautionary number of pounds or kilos of food per body weight. What metric will be standard by species...2% 4%

The term met or exceeded is likely inaccurate in PWS pinks and chums, SE chums and LCI pinks. Otolith sampling is far from robust. "Otolith adjusted" numbers are not always accurately reported or used so wild fish component is masked in harvest in escapement. How to evaluate this for statistically precautionary numbers since statistically robust is most often not attainable. Genetic sampling is a must that must use the latest and greatest means of getting this information ASAP. Verify and clarify the word "production". Is this natural production or hatchery production or use a different term to describe what you are talking about. The state of Alaska has become so hatchery centric that the concept of natural production is getting lost.

May "receive" impacts

Page 17 Include predators drawn into hatcheries. Predation must be expanded to entire food web for ecosystem approach

Page 21 gaps - Goals have been arbitrarily combined or lowered to give illusion of meeting escapement goals - serious shifted baselines. Get historical perspective going back as far as escapement goals have been set as a range to see how these have c hanged and been altered. These changes need to be corrected as declines are masked from this arbitrary practice. Also escapement goals are supplemented with stray hatchery fish bloating numbers also diverging from actual escapement. Very few systems are monitored so the picture of impact is very obscure.

Document currents fish are traveling in and when to the best of our knowledge that can be added to as we learn more. Foreign countries harvest is savvy on this information.

Use the anadromous waters Atlas or if NOAA has a better more modern mapper use this and create layers to depict information without so many words. Policy makers do not have or make time to read. Document nursery areas for prey species eaten by Chinook, sand lance, herring, amphipods where are the hot spot hunting grounds... with the best information we have now to be added on to as we learn more. Is harvest in fisheries taking too mujch of these species? Like squid. Watch Chinese, Japanese, American markets... are they going up or down? Is this poundage taken from currents used? These hot spot nurseries need to be protected as strongly as the Chinook themselves Page 7 graphs need to be in context so units can have relative scale comparisons. Comm harvest millions of fish can be at a glance relatively compared using same scale unit so abundance can be at a glance evaluated by scale. For instance pink salmon chrt goes to 200 million, chum to 25 million, sockeye to 60, coho to 10 and Chinook to 800,000. For the general public or policy makers this can be very misleading.

Perform cost analysis on any habitat altering projects first that contain restoration funding to show full cost of poor obsolete engineering practices and use contract bonds that hold companies that do damage responsible so their engineering gets elevated beyond cheapest method."

11/15/2023 12:08:45 katharine.miller@noaa.gov Katharine Miller

Appendix # of the Oct. 23 Draft Report - Research Efforts by Agencies, NOAA This section is missing a description of the 10 yr + research effort on juvenile migration from the Yukon River that has been occurring since 2014. Freshwater Juvenile Migration Research: We track changes in stock-group specific phenology, size, diets, and condition of juvenile Chinook salmon as they migrate from the Yukon River. Changes in phenology and size of chum and coho are also investigated. Research is conducted in collaboration with the Yukon Delta Fisheries Development Association. This short but growing time series is being used to evaluate freshwater smolt/spawner ratios, and to explore how phenology, growth and condition vary in relation to environmental factors. We are also exploring how phenology synchrony between juvenile salmon and freshwater prey/ocean conditions may be affected by environmental changes.

12/7/2023 8:36:42 madeline@crrcalaska.org Madeline Lee

Tribally-led salmon monitoring projects: English Bay Lakes sockeye salmon in Lower Cook Inlet, Copper River Chinook salmon, and Eshamy sockeye salmon in PWS Influence of hatcheries on productivity; Abundance of receding glaciers creating new Pacific salmon habitat; Impacts on reproductive processes from diseases such as IVN in sockeye salmon and Ichthyophonus in Chinook salmon possibly caused by hatchery release, over-crowding at low water levels, and higher water temperatures; Anadromous Waters Catalog stream surveys to assess new spawning grounds for Pacific salmon; Long-term Pacific salmon monitoring with in

stream video weirs to collect historical abundance trends Tribes as co-managers and co-producers with agencies and universities using a two-eyed seeing approach, which has been shown to provide a path for coexistence in fisheries research and management, serving as a bridge for Indigenous knowledge and Western science (Reid et al. 2020). Chugach Regional Resources Commission is at the forefront of Tribally-led Pacific salmon monitoring and research. We hope to hear from you about ways to collaborate on future research initiatives that impact our region.

1/28/2024 21:19:08 dallasak789@hotmail.com Ron carmon

1901, i found the rules on cookinlet. And i believe we should go back to them rules. I believe them rules, was not in effecr long enough to save our community. Or long enough, to create a influnce of propsphecy. Them rules clearly say fish from cookinlet Belongs to the fishing port of kenai alaska. Research proves, you cant change , what mother nature designed. You mess with , nature it will self destruck. Coastal conservation, brought 6.4 million guides to alaska. Funded by the 20 thousand box stores selling fishing gear.

They do not care about fish. Kenai needs fish, And kenai protected it self with laws. But laws been beoken. We aaked, noaa, the fed, the state,adfg. To protect . The fish, the people, The fishing grounds.

It getting clearer every day. Obama new word, stake holder. Is a investment firm, stakeholder capital investment org. I bought a fishing permit, to create a investment for me, my community, my state. A agreement, im a shareholder. Noaa your a share holder. Not a stake holder. Thats a obama word. You didnt build nothing, We did. Meaning goverment built this america. No npfmca, noaa, adfg, you are shareholders. Not stake holders.

The fishery said back in 1901. Legal fishing is from the mouth of the river to 120 miles out. Not a intersept fisheey for seine boats in Kodiak So lets fo back to 1901 rules. The fishing grounds, are the river mouth, to 120 miles out. Not the eez zone. It fish belongs to the port of kenai.

2/3/2024 17:43:05 dallasak789@hotmail.com Ron Carmon

Salmon ,salmon,salmon! What i like to rem ind you off For 40 of my 50 years of fishing. The trend has been to take away the oppertunity to comerical fish. Upper cookinlet fisherman, trend is stop us , stop us, stop us. From making a living , letting us fish. Sport fishing in creased by 6 million guides 546 million dollars of goes to the dipnet fishery, the city of kenai revenue has increased to 31 million . With the dip net fishery. Guides, fish the resourse for free, They net 100 grand in 3 months.

Commerical fishing, reduced to under 25 grand a year The salmon, kingsalmon been reduced to less than 30000 fish. Why ?guides fish the spawning grounds. And 24 /7 catch and release. All data suggest, corperate greed, Bass pro, the 20 thousand box store. Has ruined the fishery in the cookinlet. The answer lies, in governent , coruption is killing the runs of salmon. 24/7fishing, snagging in the spawning grounds And 6 .4 million guides, bringing the whole world to alaska to fish our fish. The boats loaded with 5 people, they take 6 fish, 7 days a week x 300 days a year. Do the math.

Not enough fish for the commerical fisherman. Science, will lie, and politic in creases. Adfg, are corupt, they will distort the science. And they do it dailya Adfg is paid, by commerical fishing. To protect our fish for the port of kenai. We paid them, to give our fisherman, the ecconemcy of our community, The state a income. And adfg sold us out, because bass pro, pays 216 million a year, to adfg retirement. The taak force, being paid also by bass pro.

These fish 1901 rules, say the fish belong to the people of the port of kenai. Not the sporting good stores.

3/12/2024 13:06:23 memiller@nps.gov Mark Miller

See comments regarding data/research gaps in the next section below. The following assessment of data/research gaps for Copper River sockeye and king salmon was prepared by the National Park Service, Wrangell-St. Elias National Park & Preserve, informed in part by collaborative discussions with staff of the Ahtna Intertribal Resource Commission.

Management of mixed-stock salmon fisheries in the Copper River Basin should be viewed as a balancing act that provides salmon harvest opportunities for stakeholders participating in commercial, personal use, recreational, and subsistence fisheries, while most importantly, ensuring that we maintain the population diversity that is necessary for sustaining current and future resiliency of Copper River sockeye and king salmon populations in relation to harvest and changing environmental conditions. Observations in the Basin are giving rise to concerns about potential effects of inriver mortality on stock-specific escapement, with implications for overall population diversity and resilience, as well as food security of harvesters in upper portions of the Copper River drainage. These observations and concerns lead to the following research needs, to –

1. Estimate stock-specific sockeye, Chinook, and coho salmon escapements into tributaries of the Copper River with a high degree of certainty (e.g., projects using weirs, sonar, and/or mark-recapture methods).

2. Develop and implement escapement-monitoring methods to create long-term indices that inform improved understanding of changes in pre-spawn mortality rates of tributary-specific adult sockeye and Chinook salmon in the Copper River upstream of the Miles Lake sonar.

3. As a means of ensuring population diversity in relation to harvest and changing environmental conditions, establish a long-term program for acquiring genetic stock composition data for Copper River salmon harvested by the Copper River District commercial fishery and inriver personal use and subsistence fisheries, and apply these data and other appropriate information to assess status and trends of exploitation rates for distinct spawning stocks of Copper River salmon and ensure, to the extent practicable, that exploitation is proportional across stocks and does not place distinct stocks at elevated risk of extirpation. This work must be designed and implemented in a manner that estimates and distinguishes exploitation rates for Copper River salmon stocks from Copper River delta and Prince William Sound stocks.

4. Use stock-specific escapement monitoring, paired with basin-wide radio telemetry and genetic sampling of unique stocks to understand effects of changing environmental conditions (driven by climate change or other factors) and increasing harvest pressures on migration timing and survival rates of adult salmon.

5. Estimate and monitor ""escapement quality"" via measures such as body size and energy content (important factors affecting migratory success), diversity and loads of pathogens, fecundity, age, and sex ratio, to help inform salmon management in the Copper River.

6. Collect baseline information on juvenile sockeye salmon abundance, outmigration, timing, condition, and mortality across unique sub-watersheds of the Copper River.

7. Assess juvenile sockeye salmon abundance in key rearing lakes to fine-tune spawner/recruit relationships from tributary specific adult spawning escapements.

8. Increase the frequency of monitoring necessary for evaluating effects of changing environmental conditions, stock-specific salmon population dynamics, and downstream harvest pressures on aspects of food security and well-being for vulnerable populations of upriver harvesters." See comments regarding a coordinated salmon research strategy in the next section below. STA-BILITY of food availability and access over time is a key dimension of food security, as defined by bodies including the World Bank, the United Nations Food and Agriculture Organization (https:// www.fao.org/3/al936e/al936e00.pdf), and the Inuit Circumpolar Council-Alaska (https://iccalaska.org/wp-icc/wp-content/ uploads/2016/03/Food-Security-Report-Brochure.pdf). Thus, a coordinated salmon research strategy that addresses increased variability or declines in Pacific salmon returns to Alaska must explicitly include components that address the issue of food security for rural Alaskans who are most dependent on salmon as a traditional food source (e.g., Nesbitt and Moore 2016, doi: 10.1111/1365-2664.12717). From this perspective, a legitimate and necessary component of the salmon research strategy would focus on ways in which downstream harvest-management strategies may be adjusted to mitigate food security impacts on upstream subsistence users of salmon. Many thanks to the talented and dedicated group of Alaska fisheries experts who have devoted their time and expertise to this task, with added special thanks to the NOAA Fisheries team who have led the effort.

3/15/2024 17:16:37 chair@kritfc.org Jonathan Samuelson (Chair of KRITFC)

KRITFC's Commissioners, Elders, and In-Season Managers have a wealth of knowledge about Kuskokwim salmon life histories, abundance, migration patterns and distribution; connections with weather, climate, and the ecosystem; and importance for regional communities' food security and culture. Some of this information has been provided through our 2021 and 2022 Situation Reports and our KRITFC-USFWS Salmon Management Strategies (available here: www.kuskosalmon.org/documents), but most is not documented in written format. This could be provided upon further request.

The Task Force should consider the salmon contributions to the 2022 and 2023 Eastern Bering Sea Ecosystem Status Reports, specifically those on Factors Affecting Yukon and Kuskokwim River Chinook (2022, pp. 24-26) and Chum (2023, pp. 131-133) Salmon Lifecycles. These reports are available here: https://www.fisheries.noaa.gov/alaska/ecosystems/ecosystem-status-reports-gulf-alaska-bering-sea-and-aleutian-islands

Additionally, the Task Force should consider the preliminary Draft Environmental Impact Statement and Social Impact Assessment for reducing non-Chinook (chum) salmon bycatch, published March 11, 2024, for the North Pacific Fishery Management Council's review. These are available here (see item C2; other salmon-related documents forthcoming: https://meetings.npfmc.org/Meeting/ Details/3039

Gaps include:

-Indigenous, Traditional, and Local Knowledge about salmon is not fully documented throughout Pacific salmon's migratory range, and it is not included equitably in research (or policy) priorities.

-Understanding the breadth of impacts of salmon declines to well-being in Western Alaska communities and ecologies.

-Genetic breakdowns of salmon caught as bycatch/intercept in marine fisheries are not known at a fine scale, making management and protection of stocks at highest risk of extirpation difficult.

-Understandings of climate impacts on salmon migrations and of distributions to more northern parts of the state are nascent.

-Effects of hatchery releases on wild salmon stocks, particularly those from Alaskan hatcheries.

Additional research is needed to address the following:

-IK/TK/LK: Additional funding, personnel, time, and importance needs to be allocated to documenting IK/TK/LK about salmon, including historical narratives of salmon and current causes of decline/abundance. These efforts should be led by Tribal entities and Knowledge holders, but collaborative amongst partners.

-Social impacts research: The breadth of salmon declines' impacts to food security, nutrition and physical health, mental health, social cohesiveness, culture and language/knowledge transfers, and traditional ways of life is not fully known.

-Marine habitat: We do not fully understand the impacts from fisheries (including pelagic and bottom trawl gear and vessel contact with shallow shelfs) and from climate change on salmon's marine habitat. These should be studied and modeled to understand the threats that salmon experience in their ocean "nursery." (However, trawl gear should not be used to study habitat impacts.)

-Freshwater habitat: We do not fully understand how changing in-river freshwater habitats, including erosion, fluctuating water levels and temperatures, decreases in snow/increases in ice coverage, changing break-up timings, and interactions with other species in the ecosystem, are affecting spawners, eggs, fry, and smolt.

-Freshwater productivity: There are few in-river monitoring projects that capture trends in the productivity of salmon-spawning tributaries.

-Genetic research and specificity: Policy makers, Tribal citizens, and public stakeholders need enhanced genetic information on which stocks are most heavily encountered as bycatch/intercept. Genetic sampling and analysis should occur in real-time to meaningfully protect salmon caught in bycatch/intercept fisheries.

-Northward salmon migrations: It is unclear why, how, and to what extent salmon are migrating northward (above Kotzebue), and how that will impact current Western Alaska salmon distribution and fisheries.

-Migration studies: Additional studies to understand migrations of Western Alaska salmon would be useful to understand gravel-to-gravel salmon distribution and would inform policy to protect these salmon throughout their migratory pathways, especially in the oceans.

-Hatchery interactions: More information is needed to understand how hatchery fish, including and especially those from Alaskan hatcheries, are impacting wild stocks' prey availability, health/ diseases, and distribution.

KRITFC emphasizes that efforts to narrow these data gaps and pursue research under these (or other) themes should include Tribal entities as partners and collaborators from the outset of the project development and funding application process. Local and Tribal people should be employed in these projects as much as possible. A coordinated salmon research strategy should consider gravel-to-gravel impacts to salmon health and viability. It should account for salmon throughout their life cycle and across management and research jurisdictions. This strategy would be inherently ecosystem-based in nature and account for interactions and relationships between salmon and all other species and ecosystem elements they encounter, including weather and climate, predators, prey, similar pelagic foragers, seabirds, vegetation, and humans – those that harvest salmon intentionally (i.e., subsistence fishers) and unintentionally (i.e., bycatch). Western scientific, social scientific, and Traditional research would be supported and included in this strategy.

Tribes and Tribal organizations, NGOs, state agencies, federal agencies, and international entities should band together to understand salmon, and what is happening to salmon. These research efforts should be led by Salmon People – primarily Alaska Native people and organizations – and should be centered on Indigenous data, knowledge, and wisdom as well as Western science. Stable, non-competitive funding should support these freshwater-to-marine research efforts, and substantial amounts of this funding (i.e., more than 50%) should be dedicated to Tribal governments and (inter)Tribal organizations to employ Tribal citizens in monitoring work.

Further, this gravel-to-gravel salmon research strategy should connect with policy to protect and restore salmon to abundance. Research efforts should not just gather information (i.e., about climate impacts, habitat changes, or fine-scale genetics) for the sake of understanding, but to inform Tribal, state, and federal decision-makers of the best pathways forward to recovering salmon stocks. Informing and effecting meaningful policy and management actions should be the primary focus for the research strategy. KRITFC appreciates that this Task Force has endeavored to be expansive and inclusive. Accepting a wide variety of experts to the Task Force, an unlimited number of members to the AYK Working Group, and regular public comment is encouraging to our Tribes.

We emphasize that partnerships amongst all research and management entities are key for expanding our understanding of salmon. Tribal entities are engaged and ready to collaborate with federal, state, and NGO entities to protect our salmon and ways of life – neither of which can be separated from the other. "

Public input via email

On Thu, Apr 11, 2024 at 8:12 PM N.J. Hillstrand <halibuts@ gmail.com> wrote:

Added Yukon factor?

B.C. salmon could be declining due to increase in use of road salt | CTV News

https://bc.ctvnews.ca/is-road-salt-killing-pacific-salmon-thats-what-a-new-b-c-study-is-trying-to-find-out-1.6144331

From: Wood, Mike E (DFG) <mike.wood@alaska.gov> Date: Fri, Apr 26, 2024 at 8:46 AM Subject: Comments for Task Force from BOF member Mike Wood

To: afsc.communications@noaa.gov <afsc.communications@noaa.gov>

Dear Task Force,

Thank You for the opportunity to comment on this amazing task force and its mission. I am heartened to see the depth of complexity and seriousness of identifying ALL the issues surrounding the health and viability of salmon for Alaska and ocean wide. I have reread the priorities identified by all stakeholders and agree wholeheartedly with the issues identified.

As a member of the Alaska board of Fisheries i took special note of lines 122-151. I strongly desire the information discovered by this task force in order to make the most informed decisions, The complexity of all the variables can be overwhelming. In particular i desire how adjustments to all the factors can be tweaked by managers and government officials to slightly nudge this huge tanker of momentum in the correct direction. In particular i hope Genetic Stock Identification is increased in areas of harvest in mixed stock fisheries without escapement goals to help identify harvest able surplus. Also the complexity of Hatchery contribution ocean wide, locally and the impacts to specific areas far from hatchery release.

I will end here, but I want you to know how much i personally appreciate the approach of the Task Force and hope that the findings will be used as soon as possible by all user groups and managers and industry to better affect the outcome of salmon and communities that rely on them as quickly as possible.

Thank You,

Sincerely, Mike Wood

From: Tessa Frost <tfrost@ssraa.org> Date: Mon, Apr 29, 2024 at 12:04 PM Subject: Comments on Draft Task Force Report To: <afsc.communications@noaa.gov> Cc: Susan Doherty <sdoherty@ssraa.org>

Hello,

After reviewing the draft Alaska Salmon Research Task Force report from 4/1/24. I have the following comment.

In light of the recent barrage of anti-hatchery claims and BOF proposals, I think it's prudent to note that a majority of the interactions between western Alaska salmon and hatchery salmon are a result of the abundance of non-Alaska hatchery salmon. For example, results from GSI from the 2023 Bering Sea B-pollock fishery determined that Asian chum (NE and SE Asia) comprised 68.8% of the total chum bycatch, compared to 18.7% that were of E GOA/PNW origin (Table 1, Barry et al. 2024).

Please consider adding this "non-Alaska hatchery chum" verbiage to the report so people are more educated about the country of origin the WAK salmon are interacting with, and to shed light on the fact that reducing Alaska hatchery salmon releases likely will not have an impact on WAK salmon returns.

thank you,

Tessa

Tessa Frost (Minicucci)

Research & Evaluation Manager Southern Southeast Regional Aquaculture Association (SSRAA) 14 Borch St., Ketchikan, AK 99901 Office: 907-228-4388

From: akhunt via _NMFS AFSC Communications <afsc.communications@noaa.gov> Date: Tue, Apr 30, 2024 at 10:15 PM Subject: From Virgil Umphenour Corrections To: <afsc.communications@noaa.gov>

line 142 since 2000 line 143 recently 2020-2023 line 146 no subsistence fishing for chum or chinook salmon in district 5&6upper Yukon & tanana rivers 2020-2023.

Virgil Umphenour May 1, 2024 From: Bill Templin

Ed, Here are the two reviews that I received.

Cheers, Bill

Reviewer #1

_ _ _ _

Hello,

Thanks for the opportunity to read the report. There are a few things that publications should catch like ADF&G writes inriver as 1 word, font changes etc. So, I stayed away from those types of edits. Below are just a few observations and 1 comment:

- The legend in figure 1 doesn't match the description
- The legend in figure 2 doesn't match the description
- If you could match the colors of Figure 2 in the WG report to the life-cycle figure in your report you would have similarity between the documents
- The reference section needs a thorough review because of repeat citations, incomplete citations, and citations with all capital letters etc.

 Hopefully more effective monitoring of salmon indicator stocks (line 108) leads to more consistent base funding of escapement projects on the Chena and Salcha Rivers, the two most important spawning tributaries on the Alaska side of the Yukon River.

Reviewer #2

That report came together nicely, and the length is certainly appropriate such governmental documentation! I don't have the time to go over it all in detail, but I went thru enough to get some thoughts together. So here are just a few points to consider.

- It might be worth emphasizing that we must develop adaptive research / management strategies to keep pace with ever changing impacts of environmental change.
- Under "Freshwater habitat changes" consider addressing the need to explore the possibility of range expansion and colonization of new habitats. This was mentioned obliquely in "Freshwater Harvest" but I think it warrants more directed attention.
- In "Salmon Health and Condition", a very specific reference is made regarding Thiamine deficiencies which seems a bit specific relative to the content provided under the other headers, so it comes off as a little odd in context. Maybe a more general explanation indicating a need to link physiological responses to environmental change and its impact of fish health. Also consider the need to address the effects of ocean acidification on health, behavior, prey availability/quality, etc. I didn't see it mentioned anywhere, and given its potential impact, it should be.
- Under "New Technologies", it is worth mentioning development of biochronologies and life history reconstruction.
- The section "More Effective Monitoring of Indicator Stocks" got me thinking about the need to explore the possibilities of new runs getting established as new habitats become available (see #2 above). Is this report a forum for discussing expansions to current surveys and such? It should be.
- All research recommendations and initiatives require financial support, and a list of potential sources were provided in Appendix 6, but that is buried deep in the report. Consider referencing the issue and potential solutions in the executive summary as funding is essential for making these recommendations a reality. Given the current economy, limited funds for research, and the competitive nature of these funding sources, you may want to consider offering up solutions that are outside our typical comfort zone licensing fees, test fisheries, tourism taxes, citizen science, volunteers, donations, etc. Given ongoing inability of the federal government to agree on long term budgets only emphasizes the need to explore more unique options. I am not sure this is the venue to discuss finances, but then again, what is?

Appendix 5: NPAFC Science Plan

The 2023–2027 North Pacific Anadromous Fish Commission Science Plan Primary Goal and Research Objectives

The primary goal of the 2023–2027 Science Plan is to: "Establish a research framework to develop a mechanistic understanding of the impact of changing climate on salmon abundance and distribution trends in the North Pacific Ocean." (1) Improve knowledge of the relative biomass, distribution, migration, and fitness of Pacific salmon in the ocean (Present Knowledge); (2) Understand causes and anticipate changes in the production of Pacific salmon and the marine ecosystems producing them (Forward Action).

Improved understanding of the mechanisms that regulate the distribution and abundance of Pacific salmon will promote the conservation of anadromous populations in the North Pacific Ocean, allow for better projections, or at least include realistic uncertainty given climate change, of Pacific salmon production trends in the future, and enhance the sustainable fisheries management, food security, and economic security in member nations.

The timing of the NPAFC 2023–2027 Science Plan overlaps with the proposed implementation of Basin-scale Events to Coastal Impacts (BECI; 2021–2030). It is anticipated that a BECI science plan will be finalized at the PICES Annual Meeting during fall 2023.

NPAFC Research Themes

(1) Status of Pacific salmon and steelhead trout (Present Knowledge);

(2) Pacific salmon and steelhead trout in a changing North Pacific Ocean (Forward Action);

- (3) New technologies;
- (4) Management systems;
- (5) Integrated information systems.

Theme 1. Status of Pacific Salmon and Steelhead Trout (Present Knowledge)

Outcome: The present status of salmon and their environments is documented and reported.

The purpose of this theme is to document and effectively report on the present status of salmon and their habitats. The NPAFC collates annual statistics on catch, escapement, and hatchery releases of Pacific salmon around the Pacific Rim. There is an ongoing need to maintain and improve monitoring of spawning escapement, catch, smolt production and other biological information for potential use in the projecting salmon return strength or ocean survival. Long-term time series are particularly valuable in understanding linkages between climate and Pacific salmon production. Data on hatchery fish should be maintained separately from data on wild fish as much as practicable. Biological information such as age composition of a population, body size, fecundity and egg size are monitored whenever feasible.

1. Status of Key Salmon Populations

Monitor Key Populations—Continue reporting on ongoing monitoring programs for key salmon populations and identify new sampling opportunities. Identify additional key populations that can be monitored to provide status information for co-existing salmon populations and their ecosystems.

Stock Assessments—Monitor current and emerging stock assessment methods in cooperation with partners potentially including ICES, NASCO and the Pacific Salmon Commission.

Report on Status of Salmon in the NPO—Report annually on the Status of Salmon in the NPO. Consider utilizing the Interactive Mapping System developed within the NPAFC Working Groupon Salmon Marking. Could be northern hemispheric in scope in cooperation with Atlantic and Arctic partners.

Data Quality—Improve the quantification and documentation of uncertainty associated with existing and new data time series and maintain wild and hatchery salmon data separately in the timeseries.

New Baseline Information—Provide a data review and annual Pacific salmon hatchery and wild abundance data updates to Ruggerone and Irvine (2018). These methods could be drawn from those described in Ruggerone et al. (2010) and Ruggerone and Irvine (2018) and adapted as needed. These data would be reviewed and provided by NPAFC member countries as part of the WGSA annual workplans. These data would be managed and warehoused by the NPAFC, similar to the catch and hatchery release statistics: <u>https://npafc.org/statistics/</u>. The long-term goal will be to make it possible for each party to easily estimate the annual wild and hatchery abundance and biomass of salmon in the North Pacific Ocean.

2. Monitor Salmon in the Ocean

Gathering information on the marine ecology of Pacific salmon is critical to our understanding of how climate variability impact ecosystem function, salmon fitness, distribution, migration and survival. Anadromous salmon migrate in the ocean to maximize their growth and survival. Their seasonal migration and distribution patterns are stock specific, and fundamental migration routes may be genetically fixed. Increasing information on seasonal ocean migration and distribution of key salmon populations contributes to: planning effective ocean monitoring surveys, better climate modelling and projecting, better management to avoid incidental salmon bycatch, and efficient enforcement activities to protect salmon in the ocean.

Therefore, the recommendation is to:

- Continue integrated ecosystem marine survey monitoring activities currently conducted by Parties within respective exclusive economic zones and the Convention area to collect observations on the biological and physical oceanographic characteristics and observations on size-at-age, external traits, gonads, health and condition (e.g., energy density/lipid, thiamine deficiency, parasites and diseases), stomach contents, and potential population impacts.
- Monitor northward expansion of salmon into Arctic regions (e.g. northern Bering Sea; Chukchi Sea; Beaufort Sea).

Theme 2. Pacific Salmon and Steelhead Trout in a Changing North Pacific Ocean (Forward Action)

Outcome: The effects of natural environmental variability and human factors affecting salmon distribution and abundance are understood and quantified.

Climate change may result in significant variability and overall declines in the carrying capacity and usable habitat (distribution) of Pacific salmon in the North Pacific Ocean, potentially leading to expanded use of the Arctic Ocean, at least seasonally. An improved understanding of linkages between environmental changes and Pacific salmon production will help to plan for the economic consequences of these changes. The objectives are to understand and quantify the effects of environmental variability and anthropogenic factors affecting salmon distribution and abundance, and to project future changes with improved models.

(2-1) Pacific Salmon Distribution/ Migration, Climate and Ocean Changes

In recent years, there have been shifts in the distribution of salmon in northern regions, but some declines at the southern edges of their distribution along the Asian and North American continents. These geographical shifts in salmon abundance may be related to climate-induced changes in habitat/environments operating at regional and local scales. What are the relevant mechanisms influencing shifts spatial distribution and migration? What is driving Pacific salmon movement into the Arctic?

(2-2) Pacific Salmon Density Dependence, Carrying Capacity, Climate and Ocean Changes

With the potential of limited food resources in the ocean, it is important to understand the implications of habitat use by Pacific salmon populations at various levels of abundance, the productive capacity of habitats for each life stage, and the potential implications of density dependent effects.

There is a need to understand odd/even year differences in survival/growth of salmon species that has been correlated with pink salmon abundance. Is this a top-down effect? Or are there other explanations that may help explain this correlation?

There is a need for more comprehensive studies on the role of salmon in pelagic communities, the food availability for salmon and the nutritional quality of prey organisms.

Understand inter-and intra-specific competition among salmon at sea.

(2-3) Pacific Salmon Critical Periods, Climate and Ocean Changes

Variation in the early marine survival of Pacific salmon has been hypothesized to have a major role in determining the numbers of adults that return to spawn. However, there has been limited evidence to support this hypothesis. We need to understand the causes of mortality at each stage of the salmon life cycle and evaluate whether any particular life history period is critical.

Ocean Entry—Juvenile abundance, timing and body size at ocean entry may be important parameters that are critical to understanding and quantifying mortality at sea. Examine how these parameters are associated with salmon survival or brood year strength.

Growth—Increased energy efficiency for growth of juveniles in the early marine period may be a key to their survival and optimization of hatchery production.

Prey Organisms—Identify which prey organisms are important for salmon growth at each stage and region, and examine if the abundance of prey organisms limits salmon production.

Salmon Health—Examine effects of pathogens and stressors on the growth and survival of salmon in the ocean.

(2-4) Modelling the Future for Salmon

Reliable projection models of future salmon distribution, abundance and survival is important for sustainable resource management and for projecting future variations in production due to changing climate. Researchers and analysts should consider developing statistical models as well as ecosystem models coupled with biophysical models to estimate the impact of climate change on salmon populations, and to create future scenarios for salmon distribution and abundance.

Explain the unequal stock/species specific response of Pacific salmon to climate change. E.g., why are Asian pink salmon and Bristol Bay sockeye thriving under contemporary conditions while other species/stocks are not doing as well?

Model projections of impacts of climate change on salmon production and make progress in understanding unexplained variability in salmon abundance, migration, growth, size-at-age, and survival.

Theme 3. New Technologies

Outcome: New technologies and analytical methods are advanced and applied to salmon research.

Novel stock and fish identification methods including new molecular techniques, hatchery mass marking, and intelligent tags continue to be developed, and these tools are integral to comprehensive and cost-effective monitoring and mechanistic studies to facilitate the formulation of effective models predicting the distribution and abundance of salmon populations. Although considerable progress has been made in both the basic understanding of population differentiation of mixed marine salmonid assemblages and in genetic research technologies, this knowledge is still insufficient to understand the spatial distribution of different populations in the ocean and the differences in their responses to changing environmental conditions. Implementing genetic methods to differentiate mixed marine salmonid assemblages and to expand the database of reference samples are increasingly needed.

- *Molecular Identification*—Develop effective molecular techniques and baselines to identify the geographical origin of individual fish/population.
- *Genomics*—Use genomic technology for the rapid assessment of the physiological health status and cause of the condition of salmon.
- Environmental DNA (eDNA)—Develop eDNA methods for the rapid and non-lethal estimation of salmon distribution and potentially abundance.
- Mass Marking—Develop mass marking techniques to identify hatchery salmon in mixtures of populations. Thermal otolith marking is a successful tool for mass marking, but available mark patterns are limited.

- Intelligent Tags—Develop tagging methods to investigate the habitat conditions, predators and navigation mechanism of salmon migrating in the ocean.
- Salmon Observation Systems—Improve tracking technologies to increase knowledge of stock-specific patterns of migration and survival at each life stage.
- Remote Sensing/Autonomous Vehicles—Application of remote sensing technologies such satellite imagery and ocean gliders/ saildrones outfitted with sensors such as acoustics and new camera technologies to understand changes in the biophysical environment experienced by salmon.

The NPAFC should focus on the following new tools and activities to improve salmon identification:

- Conduct additional pink salmon genetic baseline studies to address questions regarding GSI and range expansion.
- Expand our understanding of eDNA methods, appropriate use and application.
- Develop and standardize Pacific salmon genetic data and analysis methods for a comprehensive coastwide genetic baseline database.

Theme 4. Management Systems

Outcome: Information required for effective management systems is available and applied to enforcement activities, social systems, and salmon movements into the Arctic.

The objective is to provide scientific advice that effectively informs management systems including their cultural and socioeconomic aspects. Enforcement of NPAFC's convention measures that prohibit directed fishing for anadromous fish within the Convention Area is the responsibility of NPAFC's Committee on Enforcement (ENFO). The CSRS is increasingly playing a role in providing information on the probable distribution of Pacific salmon at different times of the year, and therefore likely locations of illegal, unreported, and unregulated (IUU) fishing.

Climate change adaptation is a social process, and this is one of the key challenges facing salmon, ecosystems and humans moving forward. At this time, we allocate very little research effort towards the sociology of adaptation process in societies. The role of NPAFC to form collaborations on salmon, will need to consider how shifting salmon distributions and abundances, among other species, may disrupt the NPAFC role and connections moving forward. This involves shifting away from status quo ways of managing fisheries and salmon, with transformational shifts in our management systems.

With a northward shift in salmon distributions given climate change, the impacts on coastal communities will be large and will require NPAFC to engage with other fields of knowledge, such as the social sciences, to facilitate connections, agreements, collaborations between Nations. The absence of this field of expertise in NPAFC represents a critical gap that will be required to meet the huge challenges of climate change.

Theme 5. Integrated Information Systems

Outcome: Freely available information systems mobilize and synthesize historic and current data about salmon and their environment.

It is essential that salmon and ecosystem data are readily available for researchers and for machine learning AI applications. Therefore, the goal is to build upon data mobilization approaches developed during the IYS. This includes: (1) the development of data mobilization approaches that ensure adherence to the FAIR principles with data Findable, Accessible, Interoperative and Re-usable; (2) the application of a "federated" approach to integrating data sets from the respective parties in common agreed to data formats, e.g. the Global Ocean Observatory System standards (GOOS); (3) improve the ability to share information and collaborate on research efficiently using a modern web-based framework. Data assembled as part of the other themes are to be linked in a central data system.

Cooperative Research Approaches and Implementation of the Science Plan Pertinent approaches to cooperative research under the 2023–2027 Science Plan will include compilation and synthesis of existing data and metadata to generate and test specific hypotheses, integration of ecological monitoring research in the ocean using research vessels and/or remote sensing, conceptual and quantitative modeling, process-oriented field and laboratory studies, and retrospective analyses. Scientific results from cooperative studies using these approaches will progressively reduce major gaps in knowledge with respect to the research themes, as well as make significant contributions to BECI research in collaboration with other relevant partners such as ENFO, PICES, ICES and NASCO. New scientific information will also contribute to effective enforcement activities by member nations to protect Pacific salmon from IUU fishing in the Convention Area. Progress on research themes or issues of the 2023-2027 Science Plan will be reviewed annually during the NPAFC Annual Meeting. Symposia, workshops, and other science meetings will be scheduled during this time as appropriate.

Appendix 6: Alaska Bycatch Review Task Force Report

Final Report — November 2022

Salmon Recommendations

Much of the salmon research identified was similar for both the Bering Sea/Aleutian Island and the Gulf of Alaska. Listed below is the research identified for Western Alaska salmon and research which is unique for the Gulf of Alaska.

Western Alaska Salmon

Research Goals

- Research to improve our ability to determine the stock of origin of chum and Chinook salmon taken as bycatch.
- Research to reduce bycatch through improved understanding of distribution and migration of Western Alaska chum and Chinook salmon stocks migration patterns to better predict and therefore avoid bycatch "hot spots" in the BSAI region.ng.

Research that helps us understand the relative importance of particular mechanisms for driving abundance of Western Alaska Chinook and chum.

a) Improved information on marine migration patterns and its relation to fishery locations and

i. The projects AFSC mentioned that Sabrina Garcia (Chinook salmon) and Wes Larson (chum salmon) are leading in the Bering Sea: Model ocean distribution and migration of AK Chum and Chinook salmon stocks in the Bering Sea to predict distribution and hotspots.

ii. A tagging project of immature chum salmon in the North Pacific Ocean to help us understand their destination, timing, and maturity.

iii. A synthesis of marine migration information from fishery-dependent data sources, marine surveys, and tagging studies, and how these patterns have changed with a changing climate.

b) Improved information on the characteristics of fishery catches.

i. There are still improvements that can be made in the ability to assess age, and specifically stock-specific age of Chinook and chum salmon caught in any marine fisheries.

c) Improved information to help understand fishery impacts

i. Improved Adult Equivelant (AEQ) modeling through 'stock specific' chinook and chum salmon bycatch. Particularly for western Alaska chum salmon, AEQ analyses are limited by:

- age classification data gaps in adult chum abundance across all of the western Alaska stock group. Studies that improve the ability to estimate abundance of all chum salmon in the western Alaska stock reporting group. Continued genetics work is needed.
- the ability to break up that reporting group. This might be remedied by using technologies that go beyond genetic assignment alone (use of pathogens, stable isotopes, etc.)

Research that can provide an additional (non-adult) abundance estimate

This will be really powerful for helping triangulate which life stages are most important for determining good or poor productivity. The committee recommends that research should span the life-cycle of the salmon species.

a) Understand critical survival periods for western Alaska salmon through integrated ecosystem assessment surveys including expansion of the northern Bering Sea pelagic trawl survey into the near shore waters north of the Yukon River including Norton Sound.

i. Similar research is being planned in the southern Bering Sea to have a more comprehensive assessment of Western Alaska Chinook and chum.

NOTE: Neither of these projects are funded beyond 2023.

ii. Ecosystem indicators: summer sea temperature, phytoplankton/zoo plankton community structure; salmon and pelagic fish catch per unit effort, distribution, energy density for fitness, size, stomach contents. These indicators are being utilized to understand climate impact on the northern Bering Sea ecosystem, fish fitness and survival. The recent information from the northern Bering Sea pelagic trawl survey suggests that the marine heat wave within the NBS during 2016 to 2019 negatively affected juvenile Chum salmon fitness (shift to low quality prey, increased metabolic rates due to higher SST), likely leading to high winter mortality. The data suggest that Chinook salmon abundance is impacted by factors affecting them in freshwater and early marine residence.

b) Studies that help understand how ocean/climate conditions impact future runs

i. Marine pelagic trawl surveys in the northern and southern Bering Sea can help us address this (see above).

ii. NOAA and ADF&G are collaborating on using International Year of the Salmon (IYS) catches and samples to examine immature AYK chum salmon in the North Pacific Ocean during winter. (This is not yet funded.)

iii. Immature salmon surveys (like the IYS surveys) in the Bering Sea and North Pacific Ocean. There is currently no funding support for charter vessel to conduct the survey, collecting and processing samples or paying for gear and supplies.

c) Studies that help us understand the role of diet, health, and disease on the survival and spawning success of Western AK Chinook and chum

i. Understanding vectors of Ichthyophonus infection for Yukon Chinook salmon, and whether it is causing significant en route mortality during the spawning migration

ii. Understanding diet, nutrition, and condition of Western AK Chinook and chum stocks at juvenile (marine pelagic trawl surveys in the northern and southern Bering Sea – see above), immature (IYS surveys, industry catches, etc.), and adult life stages (returning samples from lower river test fisheries- pilot work started for Yukon Chinook, but only funded through 2022).

Gulf of Alaska Chinook Salmon

Conduct annual genetic and spatial assessment of Gulf of Alaska (GOA) Chinook salmon. This recommendation is intended to include, in addition to the genetic assessment that is currently taking place, that efforts should be made to produce estimates of both the spatial and temporal bycatch of Alaska stocks of Chinook salmon, as well as characterizations of the age, sex and size of the bycatch of Chinook salmon identified as stocks of Alaska origin. If further progress can be made towards identifications of stock of origin of Alaska Chinook salmon taken as bycatch, that too should be pursued.

a) Studies that help us understand the relative role of marine interceptions and bycatch.

i. Improved information on marine migration patterns and its relation to fishery locations and timing. Extend the distribution and timing projects using bycatch data in the Bering Sea to include the western GOA.

ii. Improved demographic information that will enable assessment of stock specific impacts.

- Collect samples to improve demographic information such as stock, age, sex, size and maturity for Chinook and chum salmon caught in any marine fisheries.
- Improved information to help understand fishery impacts through AEQ or similar analyses.

b) Research that can provide an additional (non-adult) abundance estimate. This is useful for helping triangulate which life stages are most important for determining productivity.

i. Juvenile salmon surveys: a survey occurs annually in the eastern GOA to monitor Southeast Alaska salmon stocks (Southeast Coastal Monitoring project).

 ADF&G will pilot a juvenile salmon survey in the western Gulf of Alaska in 2023. This will align with surveys in the northern and southern Bering Sea and Southeast Alaska to give a comprehensive assessment of Alaska Chinook and chum salmon early in the marine life stages.

Note: neither the GOA nor the Bering Sea projects are funded beyond 2023

c) Studies that help us understand how ocean/climate conditions impact future runs.

i. Marine pelagic trawl surveys in the Bering Sea and Gulf of Alaska (including western/central Alaska and SEAK surveys).

ii. Immature salmon surveys (like the IYS surveys) in the Bering Sea, Gulf of Alaska, and North Pacific Ocean.

Appendix 7: Alaska Salmon Research Collaboration

Federal, State, Tribal, Non Governmental Organizations, and others engaged in Alaska salmon research

State

Alaska Department of Fish and Game https://www.adfg.alaska.gov/

The Alaska Department of Fish and Game maintains active and comprehensive management and research programs to ensure fish and wildlife populations are "utilized, developed, and maintained on the sustained yield principle," in accordance with Alaska's Constitution.

Salmon Ocean Ecology Program

https://www.adfg.alaska.gov/index.cfm? adfg=salmonoceanecology.main#:~:text=Our% 20Mission,salmon%20in% 20the%20marine% 20environment.

The Salmon Ocean Ecology Program supports statewide salmon fisheries management through the assessment and monitoring of salmon in the marine environment. Our goals are to understand the marine life of Alaskan salmon, use this information to assist fishery management decision-making, and help answer pressing questions about marine processes that influence the abundance and characteristics of our salmon populations.

• Alaska Sustainable Salmon Fund

http://www.akssf.org/

The Alaska Sustainable Salmon Fund (AKSSF) program, administered by the **Alaska Department of Fish and Game**, manages the State of Alaska's allocations from the federal Pacific Coastal Salmon Recovery Fund (PCSRF). PCSRF was established by Congress in 2000 to protect, restore, and conserve Pacific salmon and steelhead populations and their habitats. AKSSF allocates its funds annually through competitive calls for proposals (CFPs) that open in the spring (usually mid-April). Eligible projects conserve habitat, restore habitat, monitor subsistence salmon populations, investigate the causes of Chinook declines, and conduct climate impact studies to identify resilient salmon habitat. Please see the AKSSF Objectives document for more information.

Gravel to Gravel Research

https://www.adfg.alaska.gov/index.cfm?adfg=graveltogravel. main

Chinook salmon (*Oncorhynchus tshawytscha*) are an iconic fish in Alaska and they support the cultural well-being, economy, and livelihood of many families and communities. However, Chinook salmon abundance across Alaska has declined over the past two decades. Chinook salmon numbers returning to many Alaska rivers began to decline in 2007, at times requiring painful restrictions on fisheries that harvest these stocks. While populations of Chinook salmon in Alaska have historically alternated between periods of higher and lower production, the fishery restrictions caused by this extended period of low runs has created social and economic hardship across many communities in both rural and urban Alaska.

There is an urgent need for an adaptive and comprehensive plan to guide research efforts to support the resilience of Alaska's Chinook salmon. In October 2022, ADF&G scientists developed a conceptual research framework called "Gravel to Gravel"; to address this issue. This document outlines a coordinated effort by the State of Alaska that will remain adaptive to changing situations and stakeholder input, as we work to both understand the decline and provide the knowledge necessary to support the development of creative solutions that will support the resiliency of Alaska's iconic Chinook salmon into the future. This document provides the following:

- 1) a brief update on the status of Chinook salmon and the fisheries that depend on them in Alaska;
- 2) a review of prior Chinook salmon research initiatives upon which this research program will build a knowledge base; and
- 3) a framework for developing a cooperative and inclusive research program to determine root causes of the sustained decline as well as possible management and policy actions based on these findings.

Download the Plan (PDF 4,415 kB)

Alaska Bycatch Advisory Council

https://www.adfg.alaska.gov/index.cfm? adfg=bycatchtaskforce. main

On March 10, 2023, the Commissioner of Fish and Game established the Bycatch Advisory Council to advise the department on ways to implement the recommendations contained within the final report of the Alaska Bycatch Review Task Force (see Appendix 4).

Federal

NOAA Fisheries – Alaska Fisheries Science Center

Marine Ecology of Pacific Salmon

https://www.fisheries.noaa.gov/alaska/ecosystems/ alaska-ecosystem-monitoring-and-assessment

Pacific salmon play an important role in Alaska's marine ecosystems and are a valuable commercial, recreational, and subsistence resource. NOAA Fisheries scientists forecast salmon harvests, assess the impact of commercial fisheries on salmon, and evaluate how salmon populations respond to environmental changes. The information we provide helps managers make science-based decisions to ensure sustainable fish populations, fisheries, and fishing communities.

 Yukon River Freshwater Juvenile Migration Research in collaboration with YDFDA

https://www.fisheries.noaa.gov/feature-story/community-steps-continue-yukon-river-salmon-research-during-pandemic

We track changes in stock-group specific phenology, size, diets, and condition of juvenile Chinook salmon as they migrate from the Yukon River. Changes in phenology and size of chum and coho are also investigated. Research is conducted in collaboration with the Yukon Delta Fisheries Development Association. This short but growing time series is being used to evaluate freshwater smolt/spawner ratios, and to explore how phenology, growth and condition vary in relation to environmental factors. We are also exploring how phenology synchrony between juvenile salmon and freshwater prey/ocean conditions may be affected by environmental changes.

Little Port Walter

https://www.fisheries.noaa.gov/about/little-port -walter-research-station

NOAA's Little Port Walter Research Station was established in 1934 and is the oldest year-round biological field station in Alaska. The station is also the second rainiest place in North America, receiving an average of roughly 5.97 meters (235 inches) of precipitation per year. The station is located on the southeastern side of Baranof Island, approximately 140 kilometers south of Juneau, in a pristine and biologically strategic location (Latitude: 56.38 N, Longitude: 134.65 W) on U.S. Forest Service land in the Tongass National Forest. It is located on Chatham Strait near a major entrance to the inside waters of southeast Alaska and is just inshore from the Alaska Coastal Current. This location makes it possible to address a wide range of research questions and objectives. The station supports both seasonal and year-round research, with the onsite population ranging from two people in the winter up to 25 in the summer depending on research needs. To date, research here has resulted in 216 peer-reviewed scientific journal publications, as well as dozens of graduate student theses, technical memos, and annual reports.

Auke Creek

https://www.fisheries.noaa.gov/about/auke-creek -research-station Located 12 miles from downtown Juneau, Auke Creek Research Station is operated by the Auke Bay Laboratories (ABL) Salmon Ocean Ecology study program on a cooperative basis with University of Alaska Fairbanks (UAF), the Alaska Department of Fish and Game (ADF&G), and the University of Alaska Southeast (UAS). The long time series of observations on the seven anadromous fish species made at the Station's counting weir is not available elsewhere in Alaska. First hand evidence of changes in fish populations in response to climate change is provided by the biological and environmental information generated at this Station. Its information is also used by ADF&G to guide harvest management decisions on commercial and recreational fisheries in the region. An experimental hatchery located near the mouth of the stream provides insights into the genetic basis for many aspects of the behavior of anadromous fish species, and it has been used to train three generations of graduate students in genetics and salmonid biology. The accessibility of the Station by road from the urban area of Juneau makes it a popular scientific educational resource for Juneau Public Schools and the general public.

U.S. DOI Bureau of Land Management (BLM) – Gravel to Gravel Keystone Initiative

https://www.blm.gov/programs/aquatic-resources/alaska/ gravel-gravel-keystone-initiative

The Department of the Interior is investing more than \$16 million over the next four years from President Biden's **Bipartisan Infrastructure Law** to improve the resilience of ecosystems and salmon in Alaska's Yukon, Kuskokwim and Norton Sound region as part of the Gravel to Gravel Keystone Initiative. This initial multi-million-dollar investment serves as a catalyst for additional in-vestments in Gravel to Gravel, which is made up of three elements:

- 1. Investments to improve resiliency of Pacific salmon
- 2. Renewed commitment to strengthening relationships through co-stewardship
- 3. Responding to ecosystem threats to food security.

While the BLM is working across all three elements of Gravel to Gravel, we are heavily focused on improving watershed resiliency through assessment and restoration. And we, in the BLM are doing what we can where we can with the provided funding to make a positive and significant impact for the communities and ecosystems of the Yukon, Kuskokwim, and Norton Sound region.

Department of Interior, Office of Subsistence Management

https://www.doi.gov/subsistence/osm

The Office of Subsistence Management is housed within the U.S. Fish and Wildlife Service, and provides administrative support to the Federal Subsistence Board and the Federal Subsistence Regional Advisory Councils. The staff of the Office of Subsistence Management includes fish and wildlife biologists, anthropologists,

technical and administrative staff, and liaisons to the Alaska Department of Fish and Game and the Alaska Native community. The staff provides support for the regulatory process and the Fisheries Resource Monitoring Program.

U.S. Fish and Wildlife Service – Gravel to Gravel Initiative

https://www.fws.gov/page/gravel-to-gravel-keystone -iniative

With Gravel to Gravel investments, the U.S. Fish and Wildlife Service is actively supporting and funding a variety of projects that will ensure safe, resilient, and equitable futures for our people, salmon, land, and waters. We are working to shape this Initiative with local and regional partners, including the Tanana Chiefs Conference, Association of Village Council Presidents, Kawerak, Inc., the Kuskokwim Intertribal Fish Commission, the Yukon River Intertribal Fish Commission, the Bureau of Land Management, USGS, National Park Service, the Yukon River Drainage Fisheries Association, State of Alaska, and nonprofit partners like Trout Unlimited. Importantly, the initiative is not a one-and-done effort. Gravel to Gravel-funded projects will build upon previous work and partnerships, while catalyzing the future of our service in Alaska - leveraging new funding, and strengthening fresh relationships, as we continue our work in serving Alaska's people, ecosystems, and wildlife.

Tribal

Arctic Yukon Kuskokwim Sustainable Salmon Initiative

https://www.aykssi.org

In response to salmon declines, Bering Sea Fishermen's Association and regional Native organizations (Association of Village Council Presidents, Kawerak, Inc., and Tanana Chiefs Conference) joined with state and federal agencies to create the AYK SSI, a proactive science-based program working cooperatively to identify and address the critical salmon research needs facing this region. The AYK SSI is the largest example of co-management of research-funding addressing salmon within the Pacific Rim and one of the largest, most successful programs of its kind in North America. OUR MISSION is to collaboratively develop and implement a comprehensive research plan to understand the causes of the declines and recoveries of AYK salmon.

Bristol Bay Regional Seafood Development Association

https://www.bbrsda.com/

Our mission is to maximize the value of the Bristol Bay fishery for the benefit of our members. One of the primary activities of the BBRSDA is funding (actually co-funding, in most cases) projects designed to strengthen our fishery. Across a broad range of disciplines – economic research, quality-improvement, fisheries science and marketing, among others – these programs are where most of our members' annual 1% assessment goes. BBRSDA's participation in approved projects – as well as the responsibilities of funded entities – are defined in contracts with the grantees (ADFG, research institutions, fishermen, service providers, municipalities, etc.).

Chugach Regional Resources Commission https://crrcalaska.org/

Since 1984, our organization's mission is to promote Tribal sovereignty and the protection of our subsistence lifestyle through the development and implementation of Tribal natural resource management programs to assure the conservation, sound economic development, and stewardship of the natural resources in the traditional use areas of the Chugach region.

We are the leading inter-Tribal fish and wildlife commission. The CRRC is founded as a community-based, Alaska Native, natural resource management organization. Community resiliency and self-determination are central threads interwoven throughout all our activities.

Kuskokwim River Inter-Tribal Fish Commission

https://www.kuskosalmon.org/

Thirty-three federally-recognized Tribes working together toward unified salmon co-management, research, and monitoring as we protect Kuskokwim salmon and traditional ways of life. Formed in 2015, the Kuskokwim River Inter-Tribal Fish Commission (KRITFC) works to develop a meaningful role for tribes and rural residents engaged in the management of Kuskokwim fisheries from the headwaters to the sea.

The Salmon Project https://salmonproject.org/

The Salmon Project is a celebration of wild salmon's place at the heart of Alaskan life, and the diverse ways it is present in our values, our culture, and our landscape.

About

From 2012 to 2019, The Salmon Project shone a spotlight on the role salmon has in all Alaskans' lives, reinforcing our culture and identities, and showing how our individual choices affect this incredible resource. By telling the story of our shared Salmon Love, we laid a foundation for a statewide movement to ensure that Alaskans' lives will always be salmon lives. Our project moved out of its active stage at the end of 2019. Many of our projects and initiatives are evergreen.

Our Vision

To create a future where Alaskans are united in an:

- Awareness of the economic, environmental, social and cultural importance of salmon for ourselves, and for all Alaskans, including those whose connection to salmon is different than our own.
- **2. Understanding** of the challenges facing Alaska's wild salmon resource.
- Commitment to collective decisions and personal actions that will ensure future generations of Alaskans live with an abundance of wild salmon.

Yukon Delta Fisheries Development Association

https://ydfda.org/

Our mission is to create a self-sustaining, independent fishing company that will create income and employment opportunities for Yukon Delta residents.

Yukon River Drainage Fisheries Association https://yukonsalmon.org/

A wide array of issues affect Yukon River fisheries. Therefore YRDFA concentrates its work in a number of program areas to achieve its mission. These include: Policy Advocacy - A wide range of agencies and boards impact Yukon River management, from the State of Alaska Board of Fish to the federal North Pacific Management Council and the international Yukon River Salmon Agreement. YRDFA participates in dialogues with all these agencies, representing the interests of Yukon River communities in state, federal, and international forums. Conservation & Restoration -YRDFA works to protect wild salmon stocks and the habitats upon which they depend. Through biological research and participation in management, YRDFA works on behalf of Yukon River fishers. Cultural Preservation - Traditional ecological knowledge is a vital source of information about salmon populations and a rich part of Yukon River cultures.

YRDFA documents Local and Traditional Knowledge (LTK) about the salmon, the river, and the people. YRDFA works to incorporate LTK into Yukon fisheries management and works to protect the subsistence rights that are the foundation of Alaskan Native culture. Economic Opportunity - Inhabitants of communities in the Yukon River drainage possess valuable knowledge and skills that can be indispensable to the success of local projects. YRDFA strengthens fishing communities by bringing jobs and training to communities, and increasing participation in fisheries management. YRDFA also works to increase market opportunities and values for Yukon River salmon. Information Sharing - YRDFA is the only consistent forum for ongoing dialogue and information-sharing between all parties with interests in the Yukon River fishery. YRDFA plays a key role in keeping fishermen and women informed and relaying their opinions to managers. YRDFA brings together fishers from throughout the watershed to reach consensus on policy positions that are good for the salmon, the people, and the river.

Yukon River Inter-Tribal Fish Commission

https://www.tananachiefs.org/tribal-resources-stewardship -program/fish-commission/

The Yukon River Inter-Tribal Fish Commission (YRITFC) with Tanana Chiefs Conference (TCC) was founded in 2014 when Yukon River Tribes came together in St. Mary's and formed the Fish Commission in response to low king salmon returns. YRITFC works with a variety of partners to oversee 28 federally recognized villages.

Universities

University of Alaska Fairbanks - Center for Salmon and Society

https://www.uaf.edu/cfos/research/center-for-salmon-society/ index.php

The Center for Salmon and Society seeks to engage all salmon-connected Alaskans in objective forums to foster dialogue using the best available science to identify trade-offs inherent in complex natural resource decisions.

University of Alaska Fairbanks - Tamamta Program

https://www.tamamta.org/

Tamamta, a Yup'ik and Sugpiaq word meaning 'all of us', is centered on elevating 14,000+ years of Indigenous stewardship and bridging Indigenous and Western sciences to transform graduate education and research in fisheries and marine sciences.

This program is motivated by the deep inequities that persist in the education and resource management systems in Alaska. While Alaska Natives make up nearly 20% of the state's population, less than 3% of students and less than 1% of faculty in our college are Alaska Native. There is a near complete absence of Alaska Native people in state or federal resource management bodies, and a near complete absence of Indigenous knowledge being taught or guiding management. We want to change this!

In the Tamamta Program we are trying to transform our whole approach to education, research, and management. Our team builds on years of cross-cultural and cross-disciplinary work to address pressing questions of equity and sustainability of life and relations in Alaska. Our fellows will use a co-production of knowledge (bridging Indigenous and Western knowledge) approach to explore key questions in our fisheries and marine and ocean systems.

University of Washington Alaska Salmon Program https://alaskasalmonprogram.org/

Our program focuses on all aspects of the ecology and evolution of Pacific salmon in the watersheds of western Alaska, the Bering Sea, and the Gulf of Alaska. We actively pursue discovery science in an era of rapid global change to produce data and knowledge for managing and conserving regional ecosystems and their fisheries, and provides insights relevant to fisheries and watershed management across the globe. Our educational mission provides research opportunities for undergraduate and graduate students, and we seek to engage with regional K-12 programs, other citizens, and management agencies to improve our collective understanding of these remarkable ecosystems.

Regional Fishery Management Organizations

North Pacific Anadromous Fish Commission https://www.npafc.org/

The North Pacific Anadromous Fish Commission (NPAFC) is an international inter-governmental organization established by the Convention for the Conservation of Anadromous Stocks in the North Pacific Ocean. The Convention was signed on February 11, 1992, and took effect on February 16, 1993. The member countries are Canada, Japan, the Republic of Korea, the Russian Federation, and the United States of America. As defined in the Convention, the primary objective of the NPAFC is to promote the conservation of anadromous stocks in the Convention Area. The Convention Area is the international waters of the North Pacific Ocean and its adjacent seas north of 33° North beyond the 200-mile zones (exclusive economic zones) of the coastal States. For the purposes of NPAFC, anadromous stocks are the stocks of these species that migrate into the Convention Area.

Scientific Research

The Convention authorizes fishing for anadromous fish in the Convention Area for scientific purposes under national and joint research programs approved by the NPAFC. The taking of anadromous fish for scientific purposes must be consistent with the needs of the research program and provisions of the Convention and be reported to the Commission. Scientific research is conducted under the **Science Plan**, which may include investigations on species ecologically related to anadromous stocks. The member countries cooperate in collecting, reporting, and exchanging biostatistical data, biological samples, fisheries data, and organizing scientific communications, such as seminars, workshops, exchanges of scientific personnel, and publications. The members provide catch, enhancement, and other technical information and material pertaining to areas adjacent to the Convention Area from which anadromous stocks migrate into the Convention Area.

Pacific Salmon Commission https://www.psc.org/

The Pacific Salmon Commission is the body formed by the govern-ments of Canada and the United States in 1985 to implement the Pacific Salmon Treaty. It is our shared responsibility to conserve the Pacific Salmon in order to achieve optimum production and to divide the harvests so that each country reaps the benefits of its investment in salmon management. The Pacific Salmon Commission oversees two Endowment Funds established in 1999 to support projects in Canada and the United States that develop improved information for resource manage-ment; rehabilitate and restore marine and freshwater salmon habitats; and, enhance wild stock production through low tech-nology techniques.

PICES - Basin-Scale Events to Coastal Impacts (BECI)

https://beci.info/

BECI (Basin Scale Events to Coastal Impacts) is an ambitious project to develop an international ocean intelligence system for the North Pacific Ocean that uses enhanced observations, numerical modeling, and data analytics infrastructure to provide timely and targeted information on the impacts of current and future climate events on ocean ecosystems. BECI was proposed by the North Pacific Anadromous Fish Commission (NPAFC) and the North Pacific Marine Science Organization (PICES) which was endorsed by the United Nations Decade of Ocean Science and Sustainable Development (UNDOS) in 2021.

Non-governmental Organizations

Arctic Beaver Observations Network https://sites.google.com/alaska.edu/a-bon/

The Arctic Beaver Observation Network (A-BON) is a group of scientists, indigenous groups, land managers, and local observers who are concerned about the expansion of beaver populations into Arctic landscapes. This collaboration began in 2020 and assembles a broad range of perspectives from Alaska, Canada, Europe, and Russia to coordinate research and observations related to beaver colonization of the Arctic and the impacts it is having on ecosystems and people.

Alaska Fish Habitat Partnerships https://www.alaskafishhabitat.org/

Our Philosophy Working together to protect, maintain, restore and enhance fish habitat throughout Alaska. *Our History* Alaska's first partnership to be formally recognized by the National Fish Habitat Partnership board was the Mat-Su Basin Salmon Habitat Partnership in 2006, followed by the Southwest Alaska Partnership in 2008, the Kenai Peninsula Partnership in 2010, and the Southeast Alaska Partnership in 2014. The Western Native Trout Initiative and Pacific Lamprey Partnership serve larger geographies that include Alaska.

Alaska's Partnerships Rainbow Trout at Fish Creek, Matanuska-Susitna Valley, Alaska. USFWS/K.Mueller Operating under the banner of the **National Fish Habitat Partner-ship**, Alaska's recognized fish habitat partnerships are working on behalf of Alaska's wild, native fish and their habitats. These six recognized partnerships are part of a national network of locally-driven, voluntary, and non-regulatory collaboratives. Active partnerships made up of diverse interests are increasingly necessary to sustain Alaska's locally and globally important fisheries – especially in geographic areas where habitat overlays a mosaic of private, state, tribal and federal lands and threats to fish habitat are at play.

Alaska Sealife Center https://www.alaskasealife.org/

Our Science Mission The overall goal of our Science Program is to develop an under-standing of the role of marine mammals, birds and fish in the arctic and subarctic marine ecosystems, and to generate scientific knowledge relevant to resource management and policy. Our projects focus on Alaska marine life and environments, but reach globally with international collaborations. The Center's unique geo-graphic location, marine cold water research facilities, live animal collections, and specialized staff allows us to use a combination of experimental and field research to:

- Investigate physiological and ecological processes affecting marine animal population dynamics.
- Conduct controlled experiments to understand factors affecting reproductive success and fitness in marine species.
- Monitor marine animal responses to environmental variability and stressors.
- Evaluate human impacts on our marine environment and animal populations.
- Develop tools to support recovery and restoration of marine resources.

North Pacific Research Board https://nprb.org/

Supporting Research in the North Pacific Alaska supports some of the most diverse and abundant marine ecosystems in the world. Species large and small depend upon the state's unique marine environments for survival from coastal kelp beds to the undercarriage of multi-year Arctic sea ice. For coastal communities and those that utilize Alaska marine resources, this dependence extends beyond food or survival to traditional, cultural, and economic values that are shared by many-even to those outside the state. Alaska generates more than 40% of the commercial fish landings in the United States. It is no wonder that with over 44,000 miles of coastline stretching from the frigid, exposed waters of the Beaufort Sea to the sheltered narrows of the Inside Passage, a shared responsibility and knowledge is required to support the long-term health of these ecosystems. Understanding Alaska's marine ecosystems is a collaborative effort. Using science and local traditional knowledge, scientists from all over the world are studying the oceanography, plants, and animals of the North Pacific, Bering Sea, and the Arctic Ocean through funding support by the North Pacific Research Board (NPRB). NPRB's mission is to build a clear understanding of these ecosystems thereby enabling effective management and sustainable use of marine resources.

Prince William Sound Science Center https://pwssc.org/

In Prince William Sound, wild and hatchery-bred pink and chum salmon are important commercial fisheries. Pink salmon is the largest of any commercial fishery and is serviced primarily by the purse seine fishery. Commercial, recreational and subsistence harvests of salmon profoundly affect the economic and cultural fabric of Prince William Sound communities. Coho, sockeye, Chinook, pink, and chum salmon support valuable fisheries in the region. The economic impact of these fisheries is critical to many small coastal communities here, and around the globe. Yet, the interactions between wild and hatchery fish are little understood. Our research focuses primarily on two species of Pacific salmon found in Alaska, both of which evolved from their ancient rainbow trout ancestors. They start their lives as freshwater fish, then change and develop the ability to live and grow in the ocean where they mature. As native fish evolve and interact with hatchery fish, there are inevitable impacts. We seek to understand those impacts in order to help maintain the unique identity and health of every species.

Salmon and People Project

https://global.si.edu/projects/salmon-and-people-project

Project Highlights The Kenai Lowlands region of the Kenai Peninsula in south central Alaska covers roughly 9400 square km. The watersheds of this region support abundant salmon that underpin sport fisheries and millions of dollars in economic activity related to commercial salmon fisheries. The food security and cultural identity

of many residents all depend on abundant and reliable salmon populations. Over the past 15+ years, the Smithsonian Environmental Research Center's Dennis Whigham has collaborated with Coowe Walker of the Kachemak Bay National Estuarine Research Reserve (KBNERR), Ryan King of Baylor University (BU) and Mark Rains of the University of South Florida (USF). The collaboration has resulted in research that shows the important ecological relationships between elements of the landscape like local plants and how many young salmon there are in the streams. The continued existence of resilient salmon populations, partic-ularly on lands like the Kenai Lowlands that lack state or federal conservation status, will require Alaskans to decide what invest-ments they need to make to ensure they can sustainably support the goods and services provided by the landscape.

State of Alaska's Salmon and People https://alaskasalmonandpeople.org/

The mission of the State of Alaska's Salmon and People project is to create an equitable decision-making platform for all stakeholders through information synthesis, collaboration, and stakeholder engagement.

Appendix 8: Task Force Members

Alaska Salmon Research

Task Force Members and Affiliations





Dr. Andrew Munro is a fisheries scientist and statewide stock assessment scientist for the Alaska Department of Fish and Game. He has been a member of the North Pacific Fishery Management Council's Scientific and Statistical Committee (SSC) since 2019. He has expertise in fish biology, stock assessment, and salmon. Munro is also well-versed in the Council's history with Pacific salmon conservation efforts in the federal fisheries, and the management needs for understanding the Pacific salmon life cycle in Alaska. Munro also chairs a Working Group on Stock Assessment for the North Pacific Anadromous Fish Commission. He also participates on technical panels for the Arctic-Yukon-Kuskokwim Sustainable Salmon Initiative. He has a PhD in Fish and Wildlife Biology from Montana State University, an MS in Biology, with an emphasis on Marine and Freshwater Ecology from East Stroudsburg University, and a BS in Biology, with a concentration in ecology and a minor in chemistry, from Ursinus College in Collegeville, Pennsylvania.

Ed Farley (Chair), NOAA Fisheries, Alaska Fisheries Science Center

Dr. Farley has extensive experience and expertise in marine ecology of Pacific salmon. He also commercially fished in Bristol Bay from 1982 to 1997. Farley holds a PhD in Fisheries from the University of Alaska Fairbanks. His dissertation focused on early marine ecology of Bristol Bay sockeye salmon to better understand mechanisms in their marine life history that impact pro-duction. Farley has worked for NOAA Fisheries at the Alaska Fisheries Science Center since 1997. He is currently the program manager of the Ecosystem Monitoring and Assessment Program. The program is focused on understanding the impacts of climate change on ecosystems and fish fitness and survival. He developed and implemented the Alaska Fisheries Science Center's salmon early marine ecology surveys in the eastern Bering Sea in 1999. The data from these surveys are used to forecast adult returns of Yukon River Chinook salmon and to understand how the rapidly warming Bering Sea is impacting early marine growth and survival of Bristol Bay sockeye salmon and western Alaska Chum and Chinook salmon. He is the chair of the Science Sub Committee of the North Pacific Anadromous Fish Commission (NPAFC). This sub committee is charged with conservation of Pacific salmon in the North Pacific Ocean. Since 1997, Farley has developed and led NPAFC efforts to understand production dynamics of Bering Sea salmon stocks (Asian and North American) through the Bering Aleutian Salmon International Survey (BASIS) Program and to understand winter ecology of Pacific salmon through the International Year of the Salmon Program. Farley was the lead author on the ocean ecology of sockeye salmon chapter in the Ocean Ecology of Pacific Salmon and Trout. This document was published in 2018. It is where much of the NPAFC marine research has been summarized.









Bill Templin, Alaska Department of Fish and Game

Dr. Bill Templin is currently the chief fishery scientist for salmon at the Alaska Department of Fish and Game, Division of Commercial Fisheries. In this capacity, he is responsible for overseeing the division's statewide salmon research and stock assessment programs and helping ensure that research is well integrated with fisheries management. He has 29 years of experience conducting and overseeing fisheries research on commercially important fish and shellfish species in Alaska. For nine of these years, he served as the principal geneticist, in charge of the department's genetics program. During this time, he has represented the State of Alaska in various statewide, national and international settings including the Pacific Salmon Commission, North Pacific Anadromous Fish Commission, and the Intergovernmental Consultative Committee on Fisheries.

Andrew Piston, Pacific Salmon Commission

Andrew "Andy" Piston currently supervises the Southeast Alaska pink and chum salmon research programs, Ketchikan area sockeye salmon research programs, and other regional salmon stock assessment projects for the Alaska Department of Fish and Game. He is responsible for monitoring the escapement, production, survival and harvest patterns, and overall health of Southeastern Alaska's pink and chum salmon stocks. He is also responsible for developing recommendations and scientific advice for managers, the Alaska Board of Fisheries, Pacific Salmon Commission, and other organizations. He works cooperatively with NOAA Fisheries to implement the Southeast Coastal Monitoring Project and to produce joint pre-season pink salmon harvest forecasts. He has been involved with salmon research projects in Southeast Alaska since 1994. He has served on the Northern Boundary Panel of the Pacific Salmon Commission since 2016, when he was appointed by the Governor of Alaska. Previously, Piston served as the co-chair of the Northern Boundary Technical Committee. He has been a technical committee member since 2010.

Oscar Evon, Native Village of Kwigillingok

Mr. Evon was born and raised in Kwigillingok, Alaska. He is a subsistence fisherman and member of the Native Village of Kwigillingok. He is the Director of Regional Affairs for the Coastal Villages Region Fund. He served as a board member from 2000-2009, eventually becoming Board of Directors president and chair. He also acted as the fund's director of programs. Evon's previous roles include Tribal administrator and COVID-19 coordinator for the Native Village of Kwigillingok, office manager for Alaska Moravian Bible Seminary, and community outreach coordinator for E3 Alaska. He serves on the North Pacific Fishery Management Council's Salmon Bycatch Committee.



Jacob Ivanoff (vice Chair), Native Village of Unalakleet

Mr. Ivanoff is a resident of Unalakleet, Alaska. He has personal knowledge of, and direct experience with, subsistence harvest and uses of salmon in rural Alaska. He is highly educated, receiving knowledge from Elders and others about salmon and through various positions he has held professionally over the years. Ivanoff is affiliated with the Native Village of Unalakleet, with the Tribe, and with other salmon-related entities in the Bering Strait region. Ivanoff is chair of the Alaska Department of Fish and Game's South Norton Sound Advisory Committee.









Karla Jensen, Native Village of Pedro Bay

Ms. Jensen is a Services Specialist 1 with the Pedro Bay Village Council. She is a Board member for the United Tribes of Bristol Bay. The Board of Directors consists of representatives from each of United Tribes of Bristol Bay's 15 member Tribal governments. United Tribes of Bristol Bay is a Tribal consortium working to protect the traditional Yup'ik, Dena'ina, and Alutiiq ways of life in Southwest Alaska that depend on the pristine Bristol Bay watershed and all it sustains, most notably Bristol Bay's wild salmon. As a political division of our member Tribal governments, their work is primarily focused in three areas: Tribal consultation with government agencies on issues affecting the Alaska Native way of life; grassroots organizing in the local, statewide, and national arena; and youth empowerment and organizing in the Bristol Bay region.

Caroline Brown, Alaska Department of Fish and Game

Ms. Brown is the statewide Subsistence Research Director. She is responsible for coordinating all ethnographic research and policy recommendations on subsistence practices for the Subsistence Section of Alaska Department of Fish and Game. Prior to this role, Brown was the Northern Regional Program Manager from 2017-2020 and the lead subsistence resource specialist for interior Alaska from 2002-2017. Brown holds an MA degree in anthropology from the University of Chicago where she was also a PhD candidate. Over the last 20 years, Brown has conducted multiple projects involving the documentation and analysis of local, traditional and Indigenous knowledge throughout Alaska. She also serves as the alternate U.S. co-chair on the U.S./Canada Yukon River Panel.

Justin Leon, Kuskokwim River Inter-Tribal Fish Commission

Mr. Leon serves as the Fisheries Biologist for the Kuskokwim River Inter-Tribal Fish Commission. Before this, he served as the Senior Tribal Climate Resilience Liaison for the Alaska Region for the Native American Fish and Wildlife Society. Through his roles as the Alaska Tribal Liaison with Native American Fish and Wildlife Society and Fisheries Biologist with the Kuskokwim River Inter-Tribal Fish Commission, Leon has garnered experience working with Alaska Native Tribes and Tribal citizens, and bridging local and Indigenous Knowledge with western science. Leon has a BS from the University of Georgia in wildlife management. He moved to Alaska after graduating in 2008 and has made Alaska home since then. He obtained his MS in fisheries from the University of Alaska Fairbanks, where he focused on Chinook salmon in the Yukon and Kuskokwim rivers. After graduate school, he spent 10 years as a fishery biologist for the Alaska Department of Fish and Game. As a fishery biologist, he worked with crab and salmon research, and wild fisheries stock management in Northwest Alaska, the Alaskan interior, and the Aleutian Islands.

Michelle Stratton, Alaska Marine Conservation Council/Fisherman

Ms. Stratton works for the Alaska Marine Conservation Council. She was born and raised in Palmer, Alaska, and grew up set netting for salmon with her family on the west side of Cook Inlet. She began her career as a technician for the Alaska Department of Fish and Game before earning her MS degree in Fisheries Science from the University of Alaska Fairbanks. At the same time, she was working eight years as an ADF&G fisheries biologist. In her position at Alaska Marine Conservation Council, Stratton devotes much of her time toward fisheries research and education, helping build connections between Alaska's fishing communities and the scientific processes that support them. As a lifelong subsistence hunter and fisherman, Stratton has a passion for fisheries biology and its role in sustaining the thriving food systems and wild places that she has lived within most of her life. She also is an owner-operator of a set net site on the south end of Kodiak Island.



Mike Flores, Charter Boat Fisherman

Mr. Flores has over 30 years of experience owning and operating a large charter fishing business on the Kenai Peninsula. He has recently completed service on the Alaska Bycatch Task Force. Mr. Flores is serving on the Charter Halibut Management Committee of the North Pacific Fisheries Management Council. He is familiar with the processes and procedures of Alaska State boards and committees.



Austin Estabrooks, At-sea Processors Association

Mr. Estabrooks is a natural resource analyst for the At-sea Processors Association (APA). He has worked on various aspects of salmon bycatch mitigation undertaken by the pollock catcher processor (CP) fleet operating in the Bering Sea. He is the primary author of the Incentive Plan Agreement (IPA) under which the CP fleet operates to meet the objectives of Amendment 91 and 110. He is responsible for monitoring in-season bycatch of both Chinook and chum to help identify hot-spots for avoidance as well as working closely with Auke Bay geneticists to identify longer term spatio-temporal trends of chum salmon stock distributions as reflected in the bycatch. He has conducted at-sea experiments with salmon lights and salmon excluder devices using deploy and retrieve cameras to quantify escapement. Through the Pollock Conservation Cooperative Research Center, Mr. Estabrooks also advises APA on funding extensive research on Alaska salmon. This includes recent projects developing species distribution models for Chinook salmon and investigating Yukon chum salmon early life history. Prior to APA, he spent nearly five years in the Bering Sea and Gulf of Alaska working as a North Pacific Groundfish observer, where he collected systematic genetic samples of salmon bycatch in the pollock fishery.



Tom Carpenter, Commercial Fisherman

Mr. Tom Carpenter hails from Cordova, Alaska. In 2022, he was appointed by Alaska Governor Mike Dunleavy to serve on the Alaska Board of Fisheries. He is retired from the U.S. Coast Guard and has participated in various fisheries throughout his career. He has served as a crewman on a seiner and a gillnetter before buying his own boat. He also purchased and operated a sporting goods store in Cordova. Carpenter has served for over 22 years on the Copper River/ Prince William Sound Advisory Committee.



Steve Reifenstuhl, retired, Northern Southeast Regional Aquaculture Association

Mr. Reifenstuhl has over 45 years of experience in Alaska salmon fisheries management, research (salmon biology and ecology, post-secondary education), salmon habitat restoration (cooperative projects with U.S. Fish and Wildlife Service) and salmon hatchery production. Among his various roles and accomplishments, he was a founding board member of Alaska's Salmon Hatchery/ Wild research program, served on the S.E. Regional Advisory Council, North Pacific Research Board Advisory Panel and was a board member of United Fishermen of Alaska. He recently retired as general manager of the Northern Southeast Regional Aquaculture Association.



Megan McPhee, University of Fairbanks, Alaska

Dr. Megan McPhee is an associate professor at the College of Fisheries and Oceans, University of Alaska, Fairbanks, located at the Juneau Fisheries Center. She is a fisheries ecologist who focuses on the ecology, evolutionary biology, and population structure of Pacific salmon. Relevant research topics include marine ecology of chum salmon, connections between climate, growth rate, and age/size at maturity in western Alaska Chinook salmon and Southeast Alaska steelhead; effects of competition on growth of western Alaska chum salmon in the North Pacific, genetic stock identification of western Alaska chum salmon, and hatchery-wild interactions in Southeast Alaska. She sits on the steering committees of the Southeast Alaska Fish Habitat Partnership and the International Year of the Salmon. She is also an associated faculty with the Tamamta program at UAF, which seeks to elevate the role of Indigenous knowledge in fisheries education and research.



Megan Williams, Arctic Program, Ocean Conservancy/ University of Alaska Fairbanks

Dr. Megan Williams is a fisheries scientist with Ocean Conservancy. Her education and professional experiences have focused on fisheries interactions with apex predators and predator ecology in Alaska. She has worked extensively on bycatch issues and climate readiness in fisheries management at both state and federal levels since 2010. Her current work focuses on integrating western science and Traditional/ Indigenous Knowledge to understand cumulative threats to salmon and rural communities and to identify climate resilient solutions. She also serves as the chair of the Alaska Scientific Review Group that advises NOAA Fisheries and the U.S. Fish and Wildlife Service on the best available science and subsistence information to be included in annual Marine Mammal Stock Assessment Reports in Alaska.





Tommy Sheridan, University of Alaska Fairbanks

Mr. Sheridan is the associate director for the University of Alaska Fairbanks (UAF) Alaska Blue Economy Center. He is also currently serving as community site coordinator for the Alaska Regional Collaboration for Innovation and Commercialization (ARCTIC) Program through UAF's Alaska Center for Energy and Power to establish Cordova, Alaska as a Community Innovation Hub. He spent eight years working for Northern Southeast Regional Aquaculture Association in the salmon hatchery industry, six years working for Alaska Department of Fish and Game as a commercial salmon fisheries manager, and three years working for Silver Bay Seafoods in the seafood processing sector. He has graduate level education in fisheries and fisheries management, and has taught undergraduate fisheries courses in both Alaska and Oregon. He has served as a board member for Prince William Sound Aquaculture Corporation, Alaska Fisheries Development Foundation, Prince William Sound Science Center, and Sitka Sound Science Center. He was appointed to the Governor's Alaska Bycatch Review Task Force (ABRT) in 2021, and currently serves on the North Pacific Anadromous Fish Commission as one of two US Commissioners.

Noëlle Yochum, Alaska Pacific University/ Trident Seafoods

Dr. Noëlle Yochum is affiliated faculty with Alaska Pacific University and is the Senior Manager of Fishing Innovation and Sustainability for Trident Seafoods. Prior to this, Dr. Yochum led the Conservation Engineering group at the Alaska Fisheries Science Center (NOAA Fisheries). In both capacities, Dr. Yochum's focus is on collaborative research with industry and scientific partners to find innovative ways to evaluate and mitigate incidental impacts of fishing, including bycatch, bycatch mortality, and effects on fish habitat. This work is done through field and laboratory research to understand fish behavior and to improve fishing gear design and practices. In addition to work in Alaska and the U.S. west coast, she has conducted related work on the U.S. east coast and abroad.



Katie Howard, Alaska Department of Fish and Game

Dr. Kathrine "Katie" Howard is a statewide fishery scientist with the Alaska Department of Fish and Game and serves as the Salmon Ocean Ecology Lead. She holds a PhD and an MS degree in zoology from the University of Hawaii and a BS in biology and a BA in English from the University of Idaho. Katie began at ADF&G in 2009 as the Yukon Area Research biologist and was quickly promoted to the Arctic-Yukon-Kuskokwim regional research biologist before becoming the AYK fisheries scientist and eventually holding a statewide scientist position overseeing the Salmon Ocean Ecology Program. Katie has been involved with multiple initiatives and projects that include collaborative marine surveys to assess juvenile Chinook and chum salmon stocks, North Pacific Anadromous Fish Commission, and International Year of the Salmon.

Appendix 9: AYK WG Members

Names and primary affiliations of the Arctic-Yukon-Kuskokwim Working Group of the Task Force.

Task Force Members

Name	Primary Affiliation	
Andrew Munro	Alaska Department of Fish and Game	
Austin Estabrooks	At-Sea Processors Association	
Bill Templin	Alaska Department of Fish and Game	
Caroline Brown	Alaska Department of Fish and Game	
Ed Farley	NOAA Fisheries, Alaska Fisheries Science Center	
Jacob Ivanoff	Native Village of Unalakleet	
Justin Leon	Kuskokwim River Inter-Tribal Fish Commission	
Katie Howard (co-chair of WG)	Alaska Department of Fish and Game	
Megan McPhee	University of Alaska Fairbanks	
Megan Williams	Ocean Conservancy	
Michelle Stratton	Alaska Marine Conservation Council/ Fisherman	
Noelle Yochum	Trident Seafoods	
Oscar Evon	Coastal Villages Region Fund (CVRF)	
Steve Reifenstuhl	Salmon Biologist/Consultant (Retired)	
Tom Carpenter	Commercial Fisherman	
Public Members		
Adolph Lupie	Lower Kuskokwim River	
Andy Bassich	Upper Yukon River	
Bill Alstrom	Lower Yukon River	
Brooke Woods	Yukon River	
Charlie Lean	Norton Sound	
Charlie Wright		
Courtney Weiss	Yukon Delta Fisheries Development Association (YDFDA)	

Task Force Members

Name	Primary Affiliation	
Curry Cunningham	University of Alaska – Fairbanks	
Dan Gillikin	Native Village of Napaimute, Aniak, AK	
Daniel Schindler (co-chair of WG)	University of Washington	
Hannah Heimbuch	Commercial Fisher	
James Nicori	Kuskokwim region	
Jennifer Hooper	Association of Village Council Presi- dents, Yukon-Kuskokwim Delta Region	
Jessica Black	Associate vice chancellor for rural, community education and Native education. University of Alaska - Fairbanks	
Joe Spaeder	Arctic-Yukon-Kuskokwim Sustainable Salmon Initiative	
Mark McNeley	Native Village of Nelson Lagoon	
Martin Andrew	Kuskokwim region	
Marvin Okitkun	Kotlik, lower Yukon Delta	
Michelle Quillin	Tanana Chiefs Conference, Fairbanks	
Patrick Barry	NOAA Alaska Fisheries Science Center	
Ragnar Alstrom	Executive Director, Yukon Delta Fisheries Development Association (YDFDA)	
Renae Ivanoff	NSEDC's Fisheries Research and Development Director	
Scott Gende	National Park Service	
Serena Fitka	<pre>/ukon Delta Fisheries Development Association (YDFDA)</pre>	
Todd Sformo	North Slope Borough - Department of Wildlife Management	
Vanessa von Biela	U.S. Geological Survey, Alaska Science Center	
Virgil Umphenour	Upper Yukon River	

Appendix 10: AYK WG Hypotheses and Research Questions

List of all hypotheses and research questions considered by the AYK WG for understanding the causes of recent declines in AYK chum salmon and Chinook salmon. Individual hypotheses and questions were developed as components of each of the major research themes (possible explanations) specified by the Task Force. Individual scores are the total number of points allocated to each hypothesis or question by the AYK WG, which were summed within each theme (total score = TS) to develop weights for prioritizing research across the Task Force research themes.

Research Theme*	Individual score	Specific question or hypothesis	
Spawner Health TS = 102	28	Quantify causes, magnitude, and consequences of en route and pre-spawn mortality. What are interactions with climate and changing ocean conditions? Roles of disease and parasites	
	23	Improved understanding of spawner quality (age, sex ratios, size, genetic divesity, nutritional and health condition of spawners) and how changes to spawner quality impact reproductive success and stock productivity	
	9	Identifying vectors of salmon disease, and conditions leading to changes in disease prevalence and intensity (e.g., environmental conditions and/or changes to migratory patterns)	
	13	Interactive effects of multiple stressors on spawning adults during their freshwater migration (e.g., climate conditions)	
	3	Sublethal impacts, such as straying or other behavior changes, during poor migration conditions (i.e. warm temperatures, low water) on migrating fish	
	19	Identifying conditions/thresholds where genetic population size is so critically low that stocks are at particular risk for extirpation, and assessing whether any existing stocks meet those criteria	
	6	Programmatic review to determine whether inriver stock assessments have adequate precision and accuracy for estimating abundance	
Freshwater Harvest (Commercial and Sport) TS = 35	9	What are legacy effects of historical FW fisheries (e.g., gear effects, harvest rates) on current population status and demographics?	
	6	What is role of unobserved fishery-mortality (e.g., drop-outs from small mesh nets) on mortality rates, fishery selection, and ultimately population productivity?	
	4	What is the extent that FW harvest can further depress weak stock components while targeting productive stocks (related to management under uncertainty). Can we improve stock resolution and harvest specificity?	
	12	Are the current BEG's/SEG's spawner recruit relationships used to estimate surplus fish available for harvest still valid in light of new theories about where mortality may be occurring i.e. fresh vs marine waters and does it matter?	
	4	Ecological consequences of harvesting to achieve the upper limits of escapement goal range, compared to lower limits of escapement goal range?	
	0	Is there any down side (related to future recruitment) to focusing harvest on "Jack" Chinook salmon?	

*Possible explanation for AYK salmon decline - (TS = total score)

Research Theme*	Individual score	Specific question or hypothesis	
Freshwater Predators TS = 19	9	Are changes in freshwater predators reducing FW survival of juvenile salmon?	
	1	Do changing environmental conditions (temp, flow, turbidity) have an influence on predation success and/or the abundance and size of predators?	
	3	How do beavers indirectly affect predation on juvenile salmon by altering habitat?	
	0	Are changes in harvest of FW predators changing predation on juvenile salmon?	
	6	Do climate-induced changes in coho salmon alter predation regimes on other salmon in FW	
Marine Predators TS = 24	12	What is the role of changes in marine apex predators (orcas, sharks, sea lions) on demographic structure (eg. body sizes and ages) and abundance of AYK salmon? What are abundance trends, diets and spatial distribution of predator populations?	
	5	What are impacts of marine predators on pre-adult and smolt life stages?	
	7	Are there interactions between changing climate and marine predators that affect ecology of AYK salmon?	
Freshwater conditions for	14	Are changes in watershed habitat productivity and capacity reducing fitness and abundance of smolts leaving watersheds?	
eggs and juvenile rearing and migration TS = 51	15	Is the hydrology (discharge, magnitude, duration, bank full flows) changing and what is the effect? Loss of complexity, disconnected habitats, channel morphology. Effects on FW food webs and growth and survival of juvenile salmon. Effects on egg incubation?	
13 = 51	6	Have changes in marine-derived nutrients (MDN) from declining salmon populations, changes in fish and wildlife management, reduced the productivity of freshwater nursery habitats?	
	5	Is the spread of beavers into areas that had not previously had beavers changing the quality and quantity of habitat for salmon?	
r	0	How do fires, permafrost degradation, and localized logging in riparian zone for firewood influence habitat value?	
	11	Climate effects on floods, spring breakup, thermal refuges, flows, temperatures, permafrost?	
Marine food	47	Are changing marine food webs and climate reducing quantity and quality of food for smolts?	
limitation TS = 131	65	Is competition with hatchery pinks and chums, Bristol Bay sockeye, reducing marine survival of AYK salmon?	
	19	Are changes in nearshore habitat conditions affecting smolt survival of AYK salmon	
Climate change	35	Interactions between climate change and all other factors	
(freshwater and marine) TS = 62	0	What are the impacts of increasing presence of salmon in the high Arctic on resident non-salmon species?	
	8	Are large scale climate variation linked between freshwater and marine ecosystems in the AYK region? If so how do we separate them, do we need to?	
	16	Climate-driven changes in sea ice and impacts for plankton phenology (match/mismatch)	
Marine Harvest TS = 156	42	What are 1) catch rates and 2) stock composition of fish in state, federal and foreign fisheries? How do these compared to escapement estimates in AYK watersheds. Need more effective monitoring of all 3 components	
	49	Is bycatch in US federal fisheries causing declines of AYK salmon?	
	38	Are interceptions in other Alaska state fisheries causing declines of AYK salmon?	
	15	Is foreign IUU (illegal, unregulated, unreported) fisheries causing declines of AYK salmon?	
	21	Are genetic markers adequate for understanding impacts of bycatch and interceptions on population dynamics?	

*Possible explanation for AYK salmon decline - (TS = total score)

Related Publications

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www.mafc.org).	(SSC), the Committee on Scientific Research and Statistics (CSRS) (Available at	
	www.mafc.org).	

North Pacific Anadromous Fish Commission Science Plan 2023—2027

https://www.npafc.org/science-plan/



Alaska Bycatch Review Task Force Final Report – November 2022

https://www.adfg.alaska.gov/static/fishing/PDFs/ bycatchtaskforce/abrt_final_report.pdf



Understanding the factors that limit Alaska Chinook salmon productivity

https://www.adfg.alaska.gov/static/ fishing/pdfs/research/graveltogravel/ chinookgraveltogravel_researchplan.pdf





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