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Good practices for estimating and using length-at-age in integrated  
stock assessments

HuiHua Lee<sup>1,2</sup>, Mark N. Maunder<sup>3,4</sup> and Kevin R. Piner<sup>5</sup>

<sup>1</sup>Corresponding author at: NOAA Fisheries, Southwest Fisheries Science Center, Fisheries Resources  
Division 8901 La Jolla Shores Drive, La Jolla Ca. USA  
E-mail address: huihua.Lee@noaa.gov

<sup>2</sup>NOAA Fisheries, Southwest Fisheries Science Center, Fisheries Resources Division 8901 La Jolla  
Shores Drive, La Jolla Ca. USA

<sup>3</sup>Inter-American Tropical Tuna Commission 8901 La Jolla Shores Drive, La Jolla Ca. USA

<sup>4</sup>Center for the Advancement of Population Assessment Methodology, Scripps Institution of  
Oceanography, La Jolla, CA, USA

<sup>5</sup>NOAA Fisheries, Southwest Fisheries Science Center, Fisheries Resources Division 8901 La Jolla  
Shores Drive, La Jolla Ca. USA (retired)

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## Abstract

Estimating growth (increase in size with age) is an integral component of fish population assessment. The use of integrated assessment models combined with the influence of misfitting size composition data on results have led to renewed interest in how growth is modeled in the assessment process. The types of data available to describe the growth process control how the length-at-age relationship will be estimated. Many factors contribute to the complexity of estimating length-at-age, including multiple sources of biological variability and difficulties in getting representative samples. The growth process in the population dynamics model is linked to all other processes and data but most directly influences the assessment model through 1) converting numbers into weight and vice versa, 2) productivity, and 3) modifying fits of size composition data. In some cases, an assessment may be insensitive to moderate levels of misspecification of the growth process, and therefore, relatively simple treatments may be adequate. However, in many cases, especially those where the fit of size composition is influential in estimating scale, a more thorough treatment of the growth process is needed. A complete treatment of growth will estimate the most important forms of biological variability, including individual, sex-specific, temporal, and spatial variability. Several types of sampling bias, including selectivity, length-stratified sampling, and spatial and measurement error, will likely also need to be addressed. When sufficient data are available, assessment authors should consider estimating the growth process as part of the integrated assessment model or consider empirical approaches for situations with high biological variability and sampling bias.

*Keywords:* Growth, Length-at-age, Integrated stock assessments

37

## 1. Introduction

38           The field popularly known as ‘age and growth’ has often served as the gateway for  
39 biologists into the broader arena of population assessment. Understanding fish growth patterns  
40 plays a key role in more than just the applied fields. Growth patterns have been used to assert  
41 evolutionary processes (Roff, 1984; 1993; 2000; Hendry et al., 2000; Haugen and Vøllestad,  
42 2001), to infer less easily observed life history traits (Pauly, 1980; Griffiths and Harrod, 2007),  
43 connectivity between areas (Helsler, 1996; Swain and Foote, 1999), and movement (McDaniel et  
44 al., 2016). However, it could be argued that growth studies play one of their most crucial roles in  
45 the population assessment of exploited stocks.

46           The increasing use of integrated assessment models (i.e., those that combine several  
47 sources of data into a single model) combined with the influence on assessment results from  
48 misfitting size composition data (Francis, 2016) has led to renewed interest in the applied  
49 understanding of fish growth. It was once presumed that because growth models are simple and  
50 data can be readily gathered, the estimation of this biological process is relatively uncomplicated.  
51 However, this assumption has proven to be far from reality as the difficulties faced in collecting  
52 representative growth data are the same as for any data sources used in stock assessments.  
53 Despite the importance of age and growth research, there has been surprisingly little change in  
54 the applied approaches used in assessments to depict growth. The von Bertalanffy Growth  
55 function (VBGF; Beverton and Holt, 1957; von Bertalanffy, 1957) remains (by far) the most  
56 commonly used growth model (Flinn and Midway, 2021). Relatively little attention has been  
57 paid by stock assessors to the many sources of variability in growth (Lorenzen, 2016). A  
58 workshop (Maunder et al., 2016) aimed at providing guidelines for treating the length-at-age

59 relationship in age-structured models (Francis, 2016) outlines many of the issues, but no clear  
60 consensus could be developed for using growth modeling in applied population assessments.

61 In this paper, we attempt to expand upon the conceptual framework of Francis (2016)  
62 and summarize the current level of understanding of fish growth. Additionally, we will provide  
63 some advice to assessment scientists about the process of fish growth, explicitly covering how  
64 the estimates of size-at-age affect integrated population dynamics modeling and methods of its  
65 estimation. We focus on length-at-age as it relates to integrated age-structured population models  
66 because of their widespread use and because length-at-age is especially important in fitting size  
67 composition data. However, many of the same considerations could apply to other types of  
68 assessment models and other forms of fish growth (e.g., length-weight).

## 69 **2. Biology**

70 The terms "growth" and "size-at-age" are often used interchangeably. However, for the  
71 sake of simplicity in the remainder of this paper, we will define "growth" as the action of  
72 changing length over time, "length-at-age" as an expression of that action, and the "growth  
73 process" as the mathematical relationship between length and age.

### 74 *2.1. Theoretical model*

75 The biological model underpinning growth generally assumes that nutrients are  
76 metabolized over time, making growth age-dependent. Fish are somewhat unique among  
77 vertebrates in that growth can be indeterminate in maximum length (Talbot, 1993) and quite  
78 variable (Lorenzen, 2016). Variability arises because local factors that affect nutrient availability,  
79 metabolism, or anabolism will ultimately affect the local expression of length-at-age.

80            *2.1.1. Nutrient availability*

81            Nutrient availability affects growth and maximum fish size (Yoneda and Wright, 2005;  
82 Auer et al., 2015; Kuzuhara et al., 2019), influenced by both biotic and abiotic factors (Eker-  
83 Develi et al., 2022). Ecological processes like hypoxia (Eby et al., 2005; Campbell and Rice,  
84 2014) impact actual nutrients available to a fish, and density-dependent competition affects  
85 growth in both juvenile (Lorenzen and Enberg, 2002) and adult stages (van Gemert and  
86 Andersen, 2018).

87            *2.1.2. Metabolism*

88            Abiotic factors, such as temperature, have been shown to influence individual  
89 metabolism (Campana et al., 1995; Gillooly et al., 2001). It is widely agreed that metabolism and  
90 growth peak within an optimal temperature range and decline as temperature departs from that  
91 range (Little and Seebacher, 2021). However, the effects of temperature on growth can be  
92 complex, as increased growth rates may cause early maturation, which may ultimately decrease  
93 the maximum length-at-age (Daufresne et al., 2009). Salinity (Bœuf and Payan, 2001), pH (Mota  
94 et al., 2018), and oxygen intake (Pauly, 2021) also affect metabolism (Bœuf and Payan,  
95 2001; Mota et al., 2018).

96            *2.1.3. Anabolism*

97            The process of synthesizing new tissue in fish is complex, variable, and influenced by  
98 multiple factors that interact with each other. Age is often the primary factor affecting the growth  
99 rate in fish (Denechaud et al., 2020). Maturation, which may also be influenced by age, often has  
100 a negative impact on somatic growth (Roff, 1983; 2000) as more metabolized nutrients are

101 allocated to reproduction (Minte-Vera et al., 2016). However, these effects can vary among  
102 individuals and are influenced by environmental factors (van Poorten and Walters, 2016).

103 *2.2. Does understanding biology even matter?*

104           The complexity and interactions of factors that simultaneously influence nutrient  
105 availability and metabolism make a mechanistic understanding of the drivers of growth and its  
106 variability difficult and beyond the scope of the typical stock assessment. Consequently, it is  
107 reasonable to ask if a mechanistic understanding of the growth process is necessary (van Poorten  
108 and Walters, 2016). Most population assessment work is descriptive, involving recreating the  
109 past population abundances. The role of fishing in determining those dynamics may not require a  
110 mechanistic understanding of growth drivers. In some cases, a reasonably accurate description of  
111 length-at-age may suffice. However, in cases where fine-scale estimates of growth are important  
112 and observations of length-at-age are incomplete or where we need to forecast beyond the  
113 observation period, a mechanistic understanding may improve predictions. This is particularly  
114 evident in long-term projections, where length-at-age may be related to potential environmental  
115 changes (Lee et al., 2018; Denechaud et al., 2020). However, a high bar should be placed on  
116 such studies for informing management, as the mechanistic growth drivers are likely more  
117 complicated than generally acknowledged. Therefore, the key question for applied work is what  
118 constitutes a "reasonably accurate" representation of length-at-age for the questions being asked  
119 and how we can achieve it.

120

### 3. Data

121 Before discussing how to or what is an accurate estimate of length-at-age, the most basic  
122 consideration governing the growth process estimation is what data is available. Although when  
123 estimated inside the population dynamics model, information on the process may come from  
124 indirect sources (Sippel et al., 2017), for this paper there are several categories of direct sources.  
125 These direct sources generally consist of size measurements and associated measures of time.  
126 Time can be either an age or an increment of time. In most instances where an age is associated  
127 with a length, that age is derived from counting and validating daily or annual rings from hard  
128 parts such as otoliths, fin rays, spines, or vertebrae, although other products of metabolism have  
129 also been used (Kilada and Driscoll, 2017). Increments of time are typically derived from capture  
130 and recapture events without necessarily knowing the age. The following is a brief discussion of  
131 the most common types of data used in routine stock assessment.

#### 132 *3.1. Length composition (length modes)*

133 Length composition, though not directly associated with age or time increment, can  
134 reveal cohort information through modal structures (Leigh and Hearn, 2000), which suggests  
135 time as a relative age. More complicated methods have been developed to decompose less  
136 obvious structures (usually larger sizes) into age groups (Pauly, 1987; Fournier et al., 1990;  
137 Laslett et al., 2004). Seasonal modal changes can offer insight into cohort growth rates.

#### 138 *3.2. Paired age-length observations*

139 The most commonly used data to estimate the length-at-age relation are paired  
140 observations of length and age. Francis (2016) described two ways these paired observations

141 could be treated, which relate to whether length or age can be considered representative of a  
142 given value of the other. The more traditional use of paired age-length data assumes that lengths  
143 are representative of a given age (random-at-age). Alternatively, the same type of data can be  
144 used with an estimation procedure that assumes that the ages are representative of a length  
145 (random-at-length). Random-at-length data is also sometimes referred to as conditional age-at-  
146 length.

### 147 *3.3. Mean length at age*

148 A variation of the paired length and age data is observations of mean length at age. This  
149 type of data is similar to random-at-age data, but it has the drawback that the growth pattern is  
150 summarized, and information on the individual variability is missing. The use of mean length-at-  
151 age is often because the information is taken from a report, and the individual data is not  
152 available. Alternatively, mean length-at-age could be used when the analyst does not want a  
153 large number of samples from small or intermediate-aged fish to have too much influence and  
154 wants to give equal weight to each age.

### 155 *3.4. Tagging*

156 Tagging offers direct growth observations (Francis, 1988; Hearn and Polacheck, 2003;  
157 Francis et al., 2016), through measurements taken between capture and recapture increments of  
158 time. However, tagging can be costly, suffer from sparse recaptures across regions/ages/size  
159 groups, and may lack age information. Concerns also exist about the stress effects on growth  
160 rates due to tagging, making them not representative of the actual population (Smircich and  
161 Kelly, 2014; Vollset et al., 2020). Typically, estimating the growth process and length-at-age  
162 requires some method or other data to derive the fish's age (e.g., Eveson et al., 2004).



163 **4. Why is estimating the growth process so complicated?**

164 With the range of data available and potentially no imperative to understand the  
165 mechanistic drivers, why does it remain so difficult to get a complete and reasonably accurate  
166 representation of a population's length-at-age? Many factors contribute to the complexity of  
167 estimating this process, including the biological variability in the growth pattern itself and a  
168 multitude of sampling biases in the data. What follows is only a partial list of some of the key  
169 types of biological variability and sampling biases.

170 *4.1. Biological variability/system processes*

171 Biological variability is the natural variation in length-at-age and should ideally be  
172 included in the assessment model as a system model process along with its associated variability  
173 (Maunder and Piner, 2015). When estimating length-at-age for an assessment, ideally, many  
174 sources of variability should be considered separately. However, in practice, these multiple  
175 sources of variability are often not specifically addressed with separate components of the  
176 system model process but are subsumed into other types of variability, model processes, or  
177 observation error (Maunder and Piner, 2017; Piner et al., 2018).

178 *4.1.1. Individual variability*

179 The most basic source of biological variability is the individual variability in the lengths  
180 for a given age, but it is not always fully considered. When it is estimated, there is no real  
181 consensus on how it should be modeled, such as whether it should be modeled as a constant  
182 (standard deviation) or proportional (coefficient of variation) across ages (or length) or how it  
183 changes with age (constant, linear, or a functional form) or the shape of the distribution of

184 lengths for a given age (normal, lognormal). Simulation studies indicate that even when  
185 individual variability is estimated, the variability may be underestimated (Schemmel et al., 2022)  
186 and almost always oversimplified.

187           Furthermore, the observed individual variability is not the same as that expressed in the  
188 system processes of the assessment model. In most integrated assessments, synthetic length  
189 distributions are created at discrete time step intervals (e.g., year, quarter) to compare to the  
190 observed length composition collected over that interval. Thus, the individual variability in the  
191 growth process of the model should include variability due to the continuous growth during the  
192 interval in addition to the natural variability. The shorter the model's time step interval and the  
193 slower the growth of fish, the smaller the contribution from growth over the interval will be to  
194 the individual variability parameters. Individual variability in spawning time and larval survival  
195 can also contribute to the variation (see 4.1.4 below).

#### 196           *4.1.2. Sexual dimorphism*

197           Even under the same environmental conditions, fish of the same species often show some  
198 sexual dimorphism. The onset of sexual differences in length-at-age often becomes apparent  
199 after maturation (Hüssy et al., 2012), with multiple hypotheses for this phenomenon (Slatkin,  
200 1984). Assessment and growth models that are not sex-specific (one-sex models) often implicitly  
201 include (intended or not) sex-specific differences in length-at-age as an additional contribution to  
202 individual variability.

#### 203           *4.1.3. Temporal (annual/cohort) variability*

204 Surprisingly, few studies aimed at length-at-age estimates for integrated assessment  
205 consider that the length-at-age relationship can change across years (although those for Cohort  
206 Analysis and Virtual Population Analysis applications often do). This temporal variability  
207 includes long-term trends or interannual variability. When temporal variability is estimated, it is  
208 usually attributed to either annual changes in the growth pattern or cohort growth differences  
209 (Thorson and Minte-Vera, 2016; Correa et al., 2021). Annual changes may occur when  
210 environmental conditions within a year cause growth pattern aberrations to more than one age  
211 class in the population. Cohort growth differences arise when cohorts display a different growth  
212 pattern that may also be associated with environmental factors, which may be spatially  
213 dependent or density-dependent. For instance, Denechaud et al. (2020) demonstrated density-  
214 dependent cohort-specific growth patterns for Atlantic cod. However, potential changes to the  
215 individual variability, rather than or in addition to changes in mean length-at-age, have received  
216 relatively less attention.

#### 217 *4.1.4. Timing of spawning/hatching*

218 The spawning seasons are not fixed points in time and exhibit variability in both the  
219 timing of peak spawning and the duration of spawning. When a fixed birthdate assumption is  
220 made in the assessment or growth model in conjunction with variability in peak spawning, it may  
221 appear as temporal differences in the length-at-age of young fish even when the growth pattern is  
222 unchanged. Durham and Wilde (2005) demonstrated this phenomenon, although the persistence  
223 of such apparent differences in length-at-age across the lifespan of the cohorts is uncertain.  
224 Variability in the duration of spawning likely impacts the spread of lengths for young fish.  
225 Because spawning timing and duration variability primarily affect young fish, these sources of  
226 variability may be subsumed in the individual variability parameter(s) of young ages. However,

227 it is not clear whether expanding the individual variability will lead to accurate estimates of  
228 young fish length-at-age when hatch dates vary, and the growth process is assumed to be time-  
229 invariant.

#### 230 *4.1.5. Seasonal variability*

231 Fish do not grow at the same rate throughout the year due to changes in nutrient  
232 availability, temperature, and other factors affecting metabolism (Pauly, 1990; Bacon et al.,  
233 2005). Length-at-age estimation models that account for seasonal effects have been available for  
234 decades (Pitcher and Macdonald, 1973). However, routine stock assessment rarely incorporates  
235 these seasonal growth effects.

#### 236 *4.1.6. Spatial variability*

237 Spatial differences in length-at-age can arise due to regional variations in growth  
238 patterns. Campana et al. (1995) demonstrated that Atlantic Cod (*Gadus morhua*) could exhibit up  
239 to a 3-fold difference in lengths-at-age across the stock distribution. Regional growth rates are  
240 likely related to either nutrient availability or factors affecting metabolism within specific  
241 geographical areas. In the case of Atlantic cod, the authors surmised that local temperatures  
242 likely contributed to the difference. However, it is unclear if these differences in growth rates are  
243 maintained if fish move from one region to another. Despite this potential source of variation,  
244 these spatial patterns in length-at-age are not routinely included in stock assessment models  
245 (including spatially explicit assessments). Instead, spatial variability may be implicitly  
246 approximated using estimates of other biological process variability, model processes, or  
247 observation error (e.g., McGarvey et al., 2007). Nevertheless, even knowing the contribution of

248 genetic and environmentally driven components still may not produce accurate estimates of  
249 length-at-age for all regions.

#### 250 *4.1.7. Fishing-induced variability*

251 The consequences of fishing have been shown to alter length-at-age at different spatial  
252 and temporal scales (Kristiansen and Svåsand, 1998; Hilborn and Minte-Vera, 2008; Andersen  
253 and Brander, 2009; Eikeset et al., 2016). Over shorter periods, high fishing mortality in  
254 conjunction with size-selective gears can lead to temporal changes in cohort mean length-at-age  
255 and its associated variability (Lee, 1912). The typical assessment with a time-invariant growth  
256 process does not account for this effect, as expected lengths in a time interval are a function of  
257 the growth process and its individual variability (typically assumed to be normally distributed),  
258 without keeping track of cumulative removals of faster-growing fish. Thus, the effect of size-  
259 selective removals on the population size structure is lost between time intervals in the model.  
260 Beyond short-term effects, decadal fishing pressures have been theorized to permanently alter  
261 life history characteristics such as maturation and growth processes (Audzijonyte et al., 2013).  
262 However, the common assertion that fishing will inevitably cause a slowing of growth due to the  
263 removal of faster-growing individuals has been challenged (Enberg et al., 2012). These decadal  
264 effects are likely to show up as temporal variability or trends in the growth process.

#### 265 *4.1.8. Shrinkage*

266 The general model of fish growth assumes growth over any unit of time. However,  
267 limited studies have documented that vertebrates can shrink in length under harsh environments.  
268 The mechanisms for such a survival strategy are not clear, but observational studies in fish  
269 (Huusko et al., 2011), amphibians (Bendik and Gluesenkamp, 2013), and laboratory studies of

270 crustaceans (Marinovic and Mangel, 1999) have documented length shrinkage. Despite its  
271 assumed rarity, in practical work, observations of fish shrinkage have likely been attributed to  
272 some form of observation error (e.g., data entry error) or rigor mortis.

#### 273 *4.2. Sampling Biases/observational processes*

274 Estimates of population lengths-at-age, along with their associated variability, are further  
275 complicated by various sampling biases. Within the assessment model, sampling bias is  
276 approximated by estimating observation processes (e.g., selectivity), accounting for bias in the  
277 observation error (e.g., the effective sample sizes used in the likelihood for length composition  
278 data), or absorbed by other model processes (e.g., natural mortality; Maunder and Piner, 2017).  
279 Sampling biases can affect the representativeness of estimated growth rates, observed lengths,  
280 observed ages, or a combination of all these factors.

##### 281 *4.2.1. Gear*

282 The effects of fishery gear selectivity (Maunder et al., 2014; Sampson, 2014) are perhaps  
283 the best study of observational biases. Fishing gear does not operate randomly with respect to the  
284 size/age of the fish captured. Selectivity bias in the growth parameter estimates has been well-  
285 studied, and several estimation methods are available to deal with this issue (Frater and  
286 Stefansson, 2020). However, the effects of fishermen-directed choices on the size/age of fish  
287 captured in fishing operations are much less studied. Purse seine fishermen have been shown to  
288 avoid capturing unwanted species or sizes of targeted species (Marçalo et al., 2019). These  
289 decisions can be based on regulations, market forces, or fishermen's preferences and can be  
290 adjusted within short time frames. In most assessments, these non-gear selection decisions are  
291 likely subsumed in the gear selectivity estimates. The extent to which this approach will account

292 for such effects is not well-studied. It has generally been regarded that gear effects are more  
293 consequential for length-at-age estimation methods assuming lengths are random-at-age, as pre-  
294 selection of age seems less likely than selection based on length.

#### 295 *4.2.2. Discarding*

296 Discarding (Rochet and Trenkel, 2005) is a post-operational decision that can affect the  
297 sizes of fish ultimately sampled from commercial fisheries (Catchpole et al., 2014). Both market  
298 forces and regulation often contribute to size-selective discarding (Aarts and Poos, 2009), which  
299 can influence the length-at-age similarly to gear selectivity. Some popular assessment models  
300 enable the estimation of retention curves to approximate the discarding process. However, this  
301 estimation requires observations of discarded fish or the imposition of strong assumptions about  
302 the discard process itself.

#### 303 *4.2.3. Spatial patterns in size/age*

304 Spatial patterns in observed length-at-age can arise even when the population exhibits a  
305 single common growth pattern across regions, but each region contains different segments of the  
306 population's length or age structure. These spatial patterns can be attributed to fish behaviors  
307 (McDaniel et al., 2016) or exploitation patterns (Frank et al., 2018). High and low regional levels  
308 of exploitation with size-selective gear can result in different observed lengths-at-age by area.  
309 Patterns in larval settling influence the observed regional length-at-age. Perhaps the most  
310 common cause of these observational spatial patterns is fish movements. Size-based movements  
311 (Nøttestad et al., 1999) are thought to arise from the differential swimming speeds associated  
312 with size, thus influencing observed lengths-at-age across migratory space. Age-based seasonal  
313 migrations were documented by McDaniel et al. (2016) in Pacific sardine (*Sardinop sagax*),

314 influencing regional ages-at-length. Similar to migration, age-specific habitat selection, possibly  
315 due to maturation, can make some segments of the age structure less available to fishing gear  
316 (Tao et al., 2021). Spatially explicit assessment models with an estimation of movement or  
317 potentially estimating the appropriate form of availability in a fleets-as-areas model have been  
318 shown to approximate these observational spatial effects (Lee et al., 2017).

#### 319 *4.2.4. Sampling*

320 Size-selective onboard or dockside sample collection can affect the observed length-at-  
321 age similar to size-selective gears. In some instances, researchers try to address the lack of data  
322 on certain poorly observed size classes (often the largest sizes) by adding additional samples to  
323 understand length-at-age better (Xu et al., 2016). In other instances, sampling strategies are used  
324 to improve estimates of length-at-age for stratified estimation methods. Random sampling, fixed  
325 sampling, and proportional sampling strategies have been studied via simulations, with methods  
326 fairing differently depending on factors such as fishery selectivity, sample size, and life history  
327 characteristics (Goodyear, 2019; Schemmel et al., 2022). Several growth estimation methods  
328 (Goodyear, 2019; Lusk et al., 2021) have been developed specifically for these stratified designs  
329 (Kimura, 1977). Although sampling methods can affect all length-at-age estimation methods,  
330 random-at-age methods are the most likely impacted as age selection strategies are less often  
331 used (Piner et al., 2016; Lee et al., 2017).

#### 332 *4.2.5. Measurement error*

333 Measurement errors can occur in both age estimates and length measurements. Aging  
334 error is generally regarded as the larger of the two sources of errors. Not all structures used to  
335 age fish are confirmed to produce annual rings, and the interpretability of these structures can be



336 quite variable (Cailliet et al., 2001; Campana, 2001). For example, age determination becomes  
337 difficult when annual marks are reabsorbed in fin spines (Rodríguez-Marín et al., 2012). Multiple  
338 methods have been developed to quantify the reliability/interpretability (aging error) for stock  
339 assessment purposes (Richards et al., 1992; Punt et al., 2008). However, in most applied  
340 situations, this interpretability is not the true error; instead, it is usually some measure of reader-  
341 to-reader variability. Even when that measure of interpretability is measured against the true age,  
342 correcting observed ages requires an estimation of the population age structure or an assumption  
343 that the observed ages are a random sample of the population.

344         Although generally thought less critical, error also exists in the length measurement  
345 (Page et al., 2004). In some examples, this error increases with fish size and may be related to  
346 human tendencies to prefer recording digits ending in 0 or 5 (Bunch et al., 2013). The issue of  
347 measurement error will be compounded when measurements are taken on difficult-to-measure  
348 fish (e.g., live releases for tagging, large sharks released alive) or when measurements are taken  
349 under challenging conditions (e.g., rough seas or using flexible tape measures). In cases where  
350 lengths are a conversion from one metric to another, such as fork length to total length (Hansen  
351 et al., 2020), the conversion itself can introduce additional variability and bias. Even when fish  
352 length is measured accurately, post-mortem shrinkage has been shown to occur in relatively short  
353 periods (Morison et al., 2003; Morison, 2004). Measurement and aging errors likely impact all  
354 length-at-age estimation methods.

355         Estimation of the growth process may include fitting data that requires conversion from  
356 one type of size measurement to another (e.g., length from weight). This conversion adds an  
357 additional source of variability and modeling complexity. The variability in the weight-length  
358 relationship, which includes similar components as those described for length-at-age, also needs

359 to be incorporated into all the model components. Some models also fit weight frequency data,  
360 often in conjunction with length composition data, but the variation in the weight-length  
361 relationship is often overlooked, causing a biased estimation of the variation in the length-at-age.  
362 Conversion from processed weight to whole weight, a common practice for some species, adds  
363 additional variation to the estimation process.

#### 364 4.2.6. *Learned behaviors/avoidance*

365 A nearly completely overlooked bias in some length-at-age estimation methods (and  
366 stock assessments in general) is a behavioral adaptation to fishing. Behavior has been described  
367 as a combination of innate (hard-wired) and learned responses to environmental stimuli (Kieffer  
368 and Colgan, 1992). Experimental evidence suggests that fish behaviors can be modified by  
369 repeated exposure to environmental stimuli. Yue et al. (2004) demonstrated that fish learn from  
370 impacts with fishing gears, and long-term memory of that event leads to avoidance behaviors.  
371 Similarly, largemouth bass (*Micropterus salmoides*) have demonstrated lure avoidance through  
372 experience-based learning rather than from conspecifics mimicking (Louison et al., 2019).  
373 Learned behaviors are likely an age-based process because the number of experiences leading to  
374 learning will likely increase with age. Other age-based patterns, such as age-related habitat  
375 choices (Tao et al., 2021) or movements (McDaniel et al., 2016), may be learned rather than  
376 innate behaviors. If the learned responses (e.g., avoidance) lead to a lower probability of capture,  
377 then assumptions about the representativeness of random-at-length data are in question.

378

## 5. How influential is growth on assessment results?

379 Before moving to estimating the growth process, it may be helpful to investigate its  
380 influence on assessment models (e.g., Wang et al., 2015; Stawitz et al., 2019; Correa et al., 2021)  
381 to help assess what might be a ‘reasonably accurate’ representation. In integrated population  
382 dynamics modeling, the growth process is linked to all other processes and data. However, to  
383 characterize its influence on assessment results, we are simplifying this complexity into the  
384 specific subunits of the model that are likely the most directly influenced by length-at-age  
385 estimates. Length-at-age most directly influences the assessment model through 1) converting  
386 weight into numbers and vice versa, 2) its contribution to the production function, and 3)  
387 modifying fits of size composition data. Relatively simple sensitivity diagnostics can help gauge  
388 the magnitude of assessment errors resulting from each growth process misspecification (see an  
389 example of each misspecification applied in Appendix A).

### 390 *5.1. Converting weight into numbers and vice versa*

391 Most integrated stock assessment models are formulated in terms of numbers of fish at  
392 each modeled age, but the catch is often recorded in weight. The total catch in weight needs to be  
393 converted into numbers-at-age in the age-structured models. The conversion is done through a  
394 series of population dynamic equations, including the selectivity curve, the growth curve (length-  
395 at-age relationship), the length-weight relationship, and the observed catch composition  
396 (assuming here it is size composition). If the growth process is misspecified, the incorrect  
397 numbers of fish are removed from the population, which will bias estimates of derived quantities.  
398 Indices of abundance are also often recorded in biomass and have to be scaled to the numbers in  
399 the stock assessment model. Similarly, the management quantities are often specified as biomass

400 and will be biased because the numbers-at-age often have to be converted back into some form  
401 of a population biomass measure (e.g., spawning biomass, maximum sustainable yield). We  
402 generally believe that for size classes when fish are growing, there is usually adequate  
403 information about length-at-age. However, it may be common that the length-at-age of the old  
404 fish is misspecified due to the low number of specimens and the difficulty in distinguishing  
405 growth zones in the otoliths of older fish. Because old fish often make up a small proportion of  
406 the catch and the biomass, the misspecification of growth in terms of converting numbers into  
407 weight and vice versa may be a relatively minor impact compared to the way the growth process  
408 interacts with other model sub-units. One exception is that, in an unexploited population, old  
409 individuals make up a larger proportion of the population. Therefore, estimates of depletion may  
410 be more impacted by bias in the length estimates for old individuals.

## 411 *5.2. Production function, estimation of absolute abundance, and management quantities*

412 The growth process also informs yield-per-recruit, which is a tradeoff between growth  
413 and natural mortality and, in conjunction with the stock-recruitment relationship, controls the  
414 shape of the production function. The absolute abundance, depletion, and productivity determine  
415 how the catch impacts the population and therefore determine how the model fits to the indices  
416 of relative abundance (Figure 1). Misspecified growth will result in biased estimates of absolute  
417 abundance or depletion level. The absolute abundance and productivity also determine the  
418 maximum sustainable yield and related management quantities. Therefore, a misspecified growth  
419 process will also bias estimates of MSY-based management quantities. A key question to assess  
420 is how much misspecification of the growth process affects absolute abundance estimates.

## 421 *5.3. Fits to length composition and estimation of absolute abundance*

422 Misspecification of the growth process may most influence the fit to length-composition  
423 data and its effect on estimates of abundance. The decline in the proportion of numbers-at-age in  
424 the population resulting from a combination of natural and fishing mortality is mediated through  
425 the selectivity to produce the catch-at-age (Figure 2). The smaller the proportion of old fish  
426 observed in the catch, given the selectivity curve and natural mortality, the higher the estimated  
427 exploitation rate. In models fit to length composition data, the catch-at-age is translated into  
428 catch-at-length through the estimated lengths-at-ages. So, the fewer large fish in the catch, the  
429 higher the exploitation rate. Estimates of exploitation rate and catch correlate to population  
430 abundance. Because the catch is usually assumed to be known with little or no error, if the  
431 growth curve is misspecified, the estimates of population abundance will also be biased. The  
432 effects of misspecification of the growth process are not confined to large fish, as fits to  
433 intermediate lengths will also influence estimates of abundance. The role of intermediate sizes is  
434 more difficult to conceptualize and may be less influential. However, these considerations should  
435 be evaluated for each specific situation.

#### 436 *5.4. Length-structured models*

437 Prior discussions have focused on the influence of the growth process on age-structured  
438 models, with emphasis on those that fit length composition data. Although beyond the scope of  
439 this work, length-structured models are also used for stock assessment (Punt et al., 2013). Those  
440 models are particularly used in situations where aging is impossible or when the fishing mortality  
441 significantly changes the length-at-age distribution (i.e., it is not normally distributed due to high  
442 fishing mortality and/or minimum legal size). In length-structured models, misspecification of  
443 the growth curve (e.g., the length transition matrix) can also influence the productivity of the  
444 stock and the fits to the indices of relative abundance and length composition data. Age and

445 length-structured models or hybrids (McGarvey et al., 2007; Methot and Wetzel, 2013; Punt et  
446 al., 2013) are impacted similarly.

## 447 **6. Methods of estimation**

448 Several decisions need to be made about the methods used to estimate the growth  
449 process and those decisions will be influenced by the available data, the complexity of biological  
450 variation and sampling biases, and consideration as to what is a reasonably accurate  
451 representation. The two basic decisions about methods are the functional forms of the growth  
452 process and its variability (structural choice) and the statistical methods (procedural choice) used  
453 to estimate it (Francis, 2016).

### 454 *6.1. Structural decisions*

455 One would think that the choice of the functional form of the growth process would be  
456 the simplest and most biologically based decision in applied work. In contrast, the types of  
457 biological variability to be estimated may be a somewhat more complex choice. Despite  
458 theoretical work suggesting that the VBGF may not always be the most appropriate functional  
459 form (Day and Taylor, 1997), a recent review of published growth studies (196) found that the  
460 VBGF is by far the most commonly used growth model (Flinn and Midway, 2021). Other  
461 popular functional forms included the three-parameter Gompertz model and the three-parameter  
462 logistic model. Newer growth models that explicitly consider the cost of reproduction have also  
463 been proposed (Minte-Vera et al., 2016). Undoubtedly, even more forms will be proposed and  
464 available for consideration. Several authors have made a case for some form of model averaging  
465 or multi-model inference (Katsanevakis, 2006) because the preferred functional form by model

466 selection criteria (e.g., AIC) may not always provide the most accurate predictions of length-at-  
467 age (Thorson and Simpfendorfer, 2009). However, Vincenzi et al. (2020) argued that with good  
468 data, the predicted length-at-age might not be functionally different when estimated using  
469 different approaches. It is also not clear how model averaged length-at-age estimates could be  
470 used in some popular stock assessment model packages. Examination of data, data fit and  
471 realism of results across spatial and temporal scales are important and should guide these  
472 structural choices.

473         The dominance of the VBGF for applied work is in part rooted in both tradition and the  
474 current limitations in choices of functional forms supported by widely used assessment packages.  
475 Issues related to assessment model support will likely diminish with the development of the next  
476 generation of assessment models (Punt et al., 2020), with more open code facilitating inclusion  
477 and experimentation with alternative functional forms. Perhaps the larger issue in applied  
478 assessment is the general practice of ignoring biological variability in the length-at-age  
479 (Lorenzen, 2016). This reluctance may be due to data limitations or estimation complexity as  
480 much as the availability of these features in assessment packages. Assessments often ignore  
481 variability in all life history traits and implicitly account for the error in other components of the  
482 model. This practice causes either additional observational error, process variability in  
483 recruitment, or selectivity. It is often unclear how this practice has influenced assessment results  
484 (Maunder and Piner, 2017).

## 485 *6.2. Procedural decisions*

486         After deciding on the functional form of the growth process and what variability to  
487 include, the next decision is what method will be used to estimate it. This research area has seen

488 the development of a host of new statistical methods to estimate length-at-age. The different  
489 procedures are usually developed in response to specific sampling biases, improved statistical  
490 methods, or to make use of alternate forms of data. We divide the methods into external  
491 estimation (the estimation is done external to the assessment model and results brought into the  
492 model) and internal estimation (the estimation is done as part of the assessment model itself).

### 493 *6.2.1. External estimates*

494 Most length-at-age estimates are derived from standalone methods. These approaches  
495 often are designed to account for at least one of the observational biases, forms of system  
496 variability, or types of data that can be used. They generally solve their specific issues by making  
497 a set of assumptions; thus, each approach may be good for one situation and inappropriate for  
498 another. Knowledge of the potential data biases is needed to evaluate which approach, given its  
499 assumption, may be most appropriate. The studies available are too many to summarize here.  
500 However, we give a few examples of approaches designed to solve some of the complex issues  
501 described in Section 4 or use data described in Section 3.

#### 502 *6.2.1.1. Length-only methods*

503 There are many methods for estimating length-at-age using length-frequency data  
504 (Schnute and Fournier, 1980; Mildenerger et al., 2017), with the Multifan program (Fournier,  
505 1990; Fournier et al., 1998) perhaps being the best known of the statistically rigorous  
506 approaches. The use of lengths only has not been without criticism (Wang, 1994), as the  
507 reliability of asserting an age with length-only data becomes more difficult as the growth slows  
508 and the modal structure becomes less apparent. Even younger ages can be difficult to determine  
509 as spawning seasons become more diffuse (Batts et al., 2019). As with all methods, careful



510 consideration of the assumptions of that method should be made. For example, Wang and  
511 Somers (1996) described an alternative method to estimate growth using length frequency for a  
512 short-lived prawn dealing with protracted spawning. However, the method overcomes the bias  
513 due to protracted spawning by assuming a known maximum size at recruitment.

#### 514 *6.2.1.2. Methods that deal with selectivity bias*

515 Because bias caused by size selection in fisheries has been known for more than 100  
516 years, methods that attempt to account for length-based selection are perhaps the most common.  
517 A seldom used but innovative approach described by Taylor et al. (2005) accounted for size-  
518 selective gears. Their method, however, must assume strong information on the fishing history  
519 and selectivity shape. The use of random-at-length data has also been shown to account for size  
520 selectivity but at the cost of approximately knowing the age structure of the sampled population  
521 (Piner et al., 2016).

#### 522 *6.2.1.3. Methods that use length-stratified samples*

523 The advantage of stratifying samples used to estimate length-at-age is ensuring good  
524 coverage of all sizes of fish. Several methods have been developed using these length-stratified  
525 samples (Perreault et al., 2020). However, the same issues related to assumptions about gear  
526 selection and spatial availability will apply.

#### 527 *6.2.1.4. Methods incorporate aging error*

528 Cope and Punt (2007) demonstrated a method incorporating aging error that assumes  
529 knowledge of the population's age structure and reader-to-reader ageing precision using random  
530 effects methods to estimate length-at-age. As with methods that use knowledge of the age-

531 structure of the population, any age-based biases that affect the representativeness of the age  
532 structure may bias estimates. As the approach assumes random-at-age, length-based sampling  
533 issues will also affect the estimates.

#### 534 *6.2.1.5. Meta-analysis*

535 Meta-analytical methods have been used in fisheries to estimate various life history  
536 characteristics (Thorson et al., 2014). They have been used specifically to estimate individual  
537 growth variability (Pilling et al., 2002), variability in growth between populations of a species  
538 (Helser and Lai, 2004), and estimate growth across closely related taxa (Helser et al., 2007). For  
539 applied work, the advantage of these methods is to improve parameter estimation for units with  
540 weaker data by borrowing information from the more data-rich populations. The estimation  
541 procedure used to estimate the length-at-age relationship will be subject to the same biases  
542 associated with that method, and improvements due to borrowing information will be only as  
543 good as the representativeness of those populations for the unit of interest.

#### 544 *6.2.1.6. Methods incorporating covariates*

545 Estimating the length-at-age relationship has included other explanatory variables such as  
546 water depth, spatial location, and cohorts (Helser et al., 2007; Kimura, 2008). The evaluation of  
547 covariates is often for larger ecological questions; however, it could be helpful for applied  
548 assessment where observations are sparse. In those cases, the covariate may help inform the  
549 estimate. However, in those situations where there is enough information to explore covariates,  
550 there was likely enough information to estimate length-at-age without the covariate. Perhaps,  
551 covariates may be most useful in forecasting future changes in growth in response to  
552 hypothesized changes in the environment.

553 *6.2.1.7. Methods that combine multiple data types*

554 We normally think of integrating multiple data types as the exclusive purview of the  
555 integrated population dynamics model, but several approaches have been developed to use  
556 multiple data types to estimate growth. Several papers have been developed that integrate the  
557 direct observation of growth from tagging and direct observations of length-at-age from paired  
558 age-length data (Eveson et al., 2004; Aires-da-Silva et al., 2015; Scherrer et al., 2021). Much like  
559 other growth estimation methods, specific sampling biases and biological sources of variability  
560 may still need to be addressed and may differ among the different data sets used in the analysis.

561 *6.2.1.8. Back calculation methods*

562 When paired age and length samples are particularly sparse, back-calculated lengths-at-  
563 age are sometimes used to estimate the growth process. Back-calculation typically involves  
564 establishing a relationship between fish size and otolith size and subsequently using that  
565 relationship to predict a younger fish size than observed. Often done to fill in missing ages, it has  
566 also been used to create repeated measures on the same fish. Jones (2000) developed methods  
567 accounting for the lack of independence of the repeated measures, and Francis (1990) created a  
568 review of available estimation methods and caveats.

569 *6.2.1.9. Random-at-length methods*

570 Although treating the paired age and length data as random-at-length is less commonly  
571 used, random-at-length estimation procedures are not affected by length sampling biases. Some  
572 evidence suggests that estimates of individual variability are improved when paired age-length  
573 data is treated as random at length (Piner et al., 2016). Lee et al. (2019) showed that age

574 sampling biases that are not properly accounted for will cause biased growth estimates. As  
575 lengths are conditioned on the ages of the population, this approach needs an estimate of the age  
576 structure of the population. Although approximate methods were developed (Piner et al., 2016),  
577 this approach depends on a good approximation of the population age structure and has generally  
578 been constrained to use within the population dynamics model (see Section 6.2.2).

#### 579 *6.2.1.10. Empirical transition matrices/age-length keys*

580 Although these approaches are not used to estimate the growth process, they are observed  
581 measures of the age distribution for a length or the length distribution for an age (less often). In  
582 applied work, these approaches are most commonly used for converting length composition into  
583 age composition, which is then used in the assessment model. Empirical approaches do not need  
584 to assume some functional relationship between length and age and are unaffected by selectivity  
585 issues in the conversion when only used to represent the fishery/survey unit. Ailloud and Hoenig  
586 (2019) provide a good overview of both forward (estimate ages for a length) and inverse  
587 (estimate lengths for an age) keys, as well as a method to combine both. Much like with growth  
588 estimation methods, this approach has a larger literature background that we can present here: an  
589 example is Babyn et al. (2021), who provided methods for estimating spatial keys. In this paper,  
590 we lump this method with empirical weight at age (discussed below 6.2.2.2) as empirical  
591 approaches.

### 592 *6.2.2. As part of the dynamic model*

#### 593 *6.2.2.1. Internal*

594           The growth process is increasingly estimated as part of the integrated population dynamic  
595 model (Zhu et al., 2016). In theory, estimation inside the assessment model offers many clear  
596 advantages over external methods. Integrated population dynamics models can incorporate many  
597 (if not all) data types and potentially draw information from indirect sources, such as the linkage  
598 between catch and indices via the production function (Sippel et al., 2017). Additionally,  
599 incorporating the growth process estimation as part of the assessment model may provide a better  
600 estimate of the total assessment uncertainty. The most often cited advantage of estimating length-  
601 at-age in these models is that they can estimate the relevant systems and observational processes  
602 needed to match both random-at-age and random-at-length assumptions. Spatial availability and  
603 age-based effects can be separated from length-based gear and sampling selection (Lee et al.,  
604 2017; Piner et al., 2018). Aging error and random-at-length data are most adeptly handled inside  
605 the population model as an estimate of the population age structure can be combined with  
606 measures of aging precision. In a separate application, Stewart and Piner (2007) used known age  
607 samples as data to estimate aging bias. Integrated assessment models can also incorporate (or  
608 estimate) much of the biological variability, such as individual, sexual dimorphism, and temporal  
609 variability. However, spatial patterns in the growth process and movement of fish with those  
610 patterns from one area to another are still somewhat of an unknown problem.

611           Despite the obvious advantages of estimating length-at-age as part of the assessment  
612 model, many applied assessments still specify this process based on external estimates. Although  
613 the external estimates of length-at-age must rely on assumptions that may be estimable in the  
614 assessment model, estimating the growth processes as part of the assessment is far from  
615 straightforward. The correct observation and system processes need to be estimated. Piner et al.  
616 (2018) demonstrated that estimating only length-based selection patterns in a fleets-as-areas

617 model, when gear selection is length-based and availability is age-based, will result in biased  
618 estimates of mean length-at-age. In this case the model may incorrectly approximate the age-  
619 based availability effect as length-based selection. Care should also be taken when adding  
620 additional data sets to the integrated model with the only purpose of better informing growth.  
621 These additional data sets require that different model processes (both observation and systems)  
622 be estimated linking that data to the dynamics, or bias can be introduced to the dynamics  
623 themselves. Lee et al. (2019) demonstrated that conditional age-at-length data (random-at-length)  
624 also have direct information on the age structure of the population. If the appropriate age-based  
625 observation or systems processes are not estimated (including potentially temporal variability in  
626 the growth process), that data provides bias not only on the growth process but the population  
627 age structure itself. Additionally, in assessments that estimate the growth process, it is common  
628 to assume at least one fleet is represented by an asymptotic selectivity to inform the model that  
629 the very largest fish are observed. However, the validity of this assumption may be questionable  
630 (Sampson, 2014), and erroneously invoking it may lead to biased results. In information-limited  
631 systems, estimating the growth process may lead to convergence issues and will likely increase  
632 run times substantially when fitting length composition data. These issues will be even more  
633 pronounced when several forms of biological variability in the growth process are also estimated.  
634 There is some hope that incorporating random effects into next generation assessment models  
635 will aid in the estimation of these time-varying processes.

#### 636 *6.2.2.2. Bypassing the growth process*

637 A number of integrated assessments bypass the growth process for some or all sub-  
638 components of the assessment model. In some sense, this could be seen as abandoning the  
639 conceptual approach of integrated modeling; however, depending on the circumstance it may

640 provide more reliable assessment results. As already mentioned, empirical age-length keys  
641 bypass the growth process in converting observed lengths to ages. The advantage of this  
642 empirical approach is that multiple sources of process variability that may be difficult to estimate  
643 may be adequately addressed with appropriate samples. Empirical weight-at-age approaches take  
644 this concept further by bypassing the length-at-age relationship entirely by inputting observed  
645 weights-at-age for each fleet as model structure. Empirical weight-at-age models are age-based  
646 (age composition data) and can be combined with the use of age-length keys if composition data  
647 are lengths. This approach to dealing with length-at-age has several advantages, especially for  
648 highly variable populations. Changes in lengths-at-age due to all relevant biological processes  
649 are embedded in the observed weights-at-age and thus do not need to be estimated. Using the  
650 empirical approach can also account for some aspects of spatial variation in length-at-age.  
651 Because the weight-at-age represents each fleet, size-based selectivity is already incorporated  
652 and doesn't need to be estimated (although an age-based selection is needed to link to the  
653 underlying numbers-at-age). However, aging error may be problematic in developing the  
654 weights-at-age observations because population numbers-at-age are still needed to correctly  
655 interpret aging error. This may be a more significant problem for very young fish as the aging  
656 error will be unidirectional and compounded if age-related weight gains are rapid. For example,  
657 aging errors will lead to some age 1+ fish being misclassified as age 0, which may substantially  
658 bias the weight of age 0 fish. In addition, to calculate the total population weight-at-age, one fleet  
659 or survey is usually assumed to be representative of the population, and size-selectivity may lead  
660 to biased population biomass. Although this approach is intended to be observation-based, some  
661 level of imputation/estimation of missing weights-at-age may be necessary. This replacing of  
662 missing or unrealistic weights-at-ages might be considered a form of growth process estimation

663 and may be subject to a separate set of assumptions. As with length-at-age estimation, these  
664 assumptions will be somewhat subjective; for example, within a cohort, fish may shrink from  
665 one year to the next, and decisions about shrinking (Huusko et al., 2011; Bendik and  
666 Gluesenkamp, 2013) versus sampling issues may have to be addressed. Finally, because much of  
667 the ‘estimation’ in empirical methods is done outside the assessment model, total assessment  
668 uncertainty will likely be reduced relative to internal estimation.

## 669 **7. Good practices**

670 The previous sections have highlighted our understanding of length-at-age and the  
671 difficulties in estimating and using this system model process. What follows is our general  
672 conclusions (in bullet points) regarding estimating and using the growth process in applied age-  
673 structured integrated assessment models. We provide guidance by the same sections as given  
674 above and include the underlying rationale. We also provide an additional section on expressing  
675 length-at-age uncertainty in the total assessment uncertainty.

### 676 *7.1. Data - Good Practices*

- 677 • Include all relevant data when the important sampling biases can be identified and adequately  
678 addressed.

#### 679 Rationale

680 In principle, increasing the amount and types of data used should add information to the  
681 estimation of the growth process and its variability. Different data types have been shown to  
682 have better information on either mean length, growth or individual variability, and for different  
683 ages. When estimating length-at-age in the integrated assessment model, care should be taken to



684 include only data where important sampling biases are identified and can be adequately  
685 addressed. Within the integrated assessment model, unmodelled observational processes will  
686 lead to biased estimates not only of the growth process but also the population dynamics. In  
687 cases where a single time-invariant growth curve is estimated in the assessment model, consider  
688 if adding multiple years of data such as random-at-length is needed as this may greatly increase  
689 the amount of observational and systems processes that need estimation.

## 690 *7.2. Biological variability - Good Practices*

- 691 • Estimate all relevant biological variability whenever possible, focusing on individual, sex-  
692 specific and temporal variability.
- 693 • Be explicit about what forms of variability will be subsumed into the individual variability  
694 parameters. If the individual variability is specified, care should be taken to match the  
695 specified variability with fish growth over the time step interval used in the assessment  
696 model.
- 697 • When fish growth is rapid and the growth process is estimated inside the assessment model,  
698 consider modeling smaller timestep intervals in the assessment.
- 699 • Investigate possible spatial variability and if regional growth patterns are found to be  
700 important to assessment results consider empirical methods, separate assessments for each  
701 area or spatially explicit assessments with regional growth processes.

## 702 Rationale

703 Individual, sex-specific, temporal, and potentially spatial variability are likely sources of  
704 biological variability for many stocks. Individual, sex-specific, and temporal variability offer no  
705 real impediments to estimation and should be investigated in most assessments. Estimating

706 individual and temporal variability inside the assessment model also offers some advantages with  
707 dealing with other forms of biological variability. If individual variability is estimated inside the  
708 assessment model, it will likely account for variability from the duration of spawning and growth  
709 within the time step interval of the assessment model. For fast growing fish, smaller model time  
710 step intervals can reduce the potential errors from continuous growth over the interval. If  
711 temporal variability is estimated, it will likely account for variability from spawning timing and  
712 fishing induced variability. Spatial variability due to regional growth patterns may present the  
713 most problems for the assessment model as a common growth process with inflated individual  
714 variability may not provide good predictions of regional length composition. In cases with  
715 impactful levels of biological spatial variability, empirical approaches should be considered. If  
716 movement between areas is low, separate assessments for each area or spatially explicit  
717 assessments with regional growth processes could be considered.

### 718 *7.3. Sampling biases/observational processes - Good Practices*

- 719 • When estimating the growth process internally, estimate all the appropriate observational  
720 and/or system processes linking data to the estimated dynamics. When using external growth  
721 estimation methods, choose the methods that best deal with the important sampling biases.
- 722 • As the number of important sampling biases increase, estimate the growth process as part of  
723 the integrated model.
- 724 • If sampling biases are difficult to address consider empirical approaches.

#### 725 Rationale

726 We consider the major sources of sampling bias common to most assessments are gear  
727 selectivity, length stratified sampling, spatial patterns, and measurement error. More than one

728 form of sampling bias can exist for any particular data set and may change over time.  
729 Observational processes may need to be estimated in age, length, or both to deal with the  
730 sampling bias. Because aging error requires some measure of population age structure, it will be  
731 most easily handled as part of the assessment model. As more and different types of important  
732 sampling biases are identified, estimate the growth process as part of the integrated assessment  
733 model or consider empirical approaches.

#### 734 *7.4. Influence of growth on assessment - Good Practices*

- 735 • Investigate how length-at-age estimates will influence the assessment models results through  
736 sensitivity diagnostics. The greater the influence of misspecification on results, the more  
737 accurate the estimates of both the growth process and its biological variability will be  
738 needed.
- 739 • The growth process is likely to be influential when composition data are lengths and they are  
740 used to provide information about abundance. In cases when growth is uncertain and there is  
741 other reliable information on absolute abundance, consideration should be given to managing  
742 the influence of length-composition data (e.g., data weighting or observational model  
743 processes estimated)
- 744 • In cases where misspecification of the growth process will affect assessment results but  
745 estimation of all relevant variability is not possible, consider empirical methods.

#### 746 Rationale

747 The sensitivity of assessment results to the misspecification of the growth process will  
748 influence what is considered reasonably accurate in the representation of the length-at-age and its  
749 associated variability. A determination of the minimum level of growth process complexity in

750 the stock does not set an upper bound to the level of growth complexity incorporated into the  
751 assessment. The maximum level of complexity will likely be influenced by data availability,  
752 knowledge of the system, and analyst time. There may also be a cost to the estimation of less  
753 consequential processes or variability as it could lead to issues in the estimation of more  
754 consequential processes and their variability (e.g., selectivity, M etc.).

755 For cases where misspecification of the growth process will have limited influence on the  
756 estimated dynamics, a time-invariant estimate of length-at-age may be the minimum resolution  
757 required to get a reliable assessment result. These situations will likely be when the population  
758 scale is not derived from fitting to length composition data. This could occur when a strong  
759 production function explains the depletion in an index of relative abundance caused by catch or  
760 when an absolute biomass survey is part of the assessment. Reduced model sensitivity to the  
761 growth process may be reinforced when catches come from young fish, age composition data are  
762 available, or when catches are recorded in numbers rather than weights. Model diagnostics and  
763 sensitivity type analyses will be helpful in the impact of the growth process misspecification.

764 For cases where fitting length composition is used to estimate population abundance, a  
765 more accurate and, therefore more complex/complete treatment of the growth process may be  
766 needed. This includes accounting for all forms of observational bias and estimating the relevant  
767 biological process variability. However, estimating all those observational and system processes  
768 may become challenging. Alternatively, empirical methods may be used if appropriate samples  
769 are present and with consideration of the issues mentioned.

## 770 *7.5. Methods*

### 771 *7.5.1. Structural Decisions - Good Practices*

- 772           • Use the most appropriate functional form for the data with consideration of what the  
773           assessment package used will support.

774           Rationale

775                       Structural decisions about the functional form of the growth process should be  
776           made based upon the best evidence but may have to conform to the forms available in the  
777           assessment models used. Care should be taken not to use a highly constrained growth  
778           model that is not appropriate because the typically abundant data for young fish can  
779           control the estimates of mean length-at-age for old fish and the fit to length frequency  
780           data represented by old fish (the large lengths) often influences the estimates of absolute  
781           abundance.

782           7.5.2. Procedural Decisions - Good Practices

- 783           • In situations where the growth process will not be influential on results an external  
784           estimate may be appropriate, but internal estimation should still be considered.
- 785           • In complex situations with multiple sources of biological variability and sampling  
786           biases, estimate the growth process as part of the integrated assessment model where  
787           possible.
- 788           • If the growth process is too complex to be estimated as part of the assessment model,  
789           consider empirical methods.
- 790           • If growth varies by area and movement among areas is low, consider assessing each  
791           area separately using the regional growth process or use empirical methods.

792           Rationale

793                   Although external methods of estimating length-at-age can be used to handle  
794 multiple forms of biological variability, internal estimation may be more appropriate for  
795 situations where accurate estimates of length-at-age and its variation is needed. Internal  
796 estimation will better account for multiple sources of sampling bias and add to the  
797 assessment estimate of uncertainty. Estimating temporal variability and individual  
798 variability will likely account for other sources like spawning timing and growth during  
799 the time step interval of the model. When external estimates are deemed sufficient, use  
800 the most appropriate method that deals best with the sampling bias identified. Consider  
801 empirical estimates when biological variability is important but too complex to  
802 adequately estimate inside the assessment model. Modeling of spatial difference in  
803 growth is complicated in age-structured population dynamics models and when  
804 movement rates are low it may be adequate to assess each region separately.

805 *7.6. Incorporating uncertainty in the growth process into characterizing assessment uncertainty*  
806 *- Good Practices*

- 807 • Estimate the growth process as part of the dynamic model.
- 808 • All key sources of biological variability should be appropriately accounted for and estimated.
- 809 • Consider a more flexible growth form to represent the uncertainty in the growth form beyond  
810 the VBGF.
- 811 • When the estimates of length-at-age influence model results and the growth process is  
812 estimated externally, consider an ensemble modeling approach or a joint prior on the growth  
813 parameters to incorporate growth process uncertainty.

- 814 • Further research is needed to understand the appropriate approaches to represent uncertainty  
815 when using empirical methods.

816 Rationale

817 Estimating key biological processes as part of the assessment model will likely provide a  
818 more complete estimate of the total uncertainty. This includes all the relevant sources of  
819 biological variability. Despite the popularity of the VB growth function, it often is relatively  
820 limited in dealing with growth patterns of very young fish, and given the general abundance of  
821 young fish samples can result in biased estimates of mean length-at-age for old fish. Developers  
822 of the next generation of assessment models should make it a priority to provide more flexible  
823 options. As empirical approaches become more popular with integrated assessment modelers,  
824 some thoughts should be given to how to incorporate uncertainty in the unmodeled growth  
825 processes into the total assessment uncertainty. When growth misspecification is a key source of  
826 uncertainty in assessment results and not estimated as part of the assessment model, ensemble  
827 approaches or the use of a joint prior on the growth parameters may be needed to incorporate this  
828 uncertainty into assessment results. The joint prior accounts for the common correlation between  
829 estimates of the asymptotic length and the growth rate parameter.

## 830 **8. Final Thoughts and Future Research**

831 The general recommendation of this work is to use either empirical approaches or to  
832 estimate the growth process and all its biological variability as part of the assessment model. The  
833 authors recognize that this is a ‘best case’ recommendation and many (if not most) assessments  
834 will not have the complete age data for the empirical approach or the information load to

835 estimate the growth process and all its complexity. Estimation of the growth process will be even  
836 more problematic due to likely parameter confounding if good practices recommend estimating  
837 other model processes (e.g., M, selectivity etc.) with both flexibility and including process  
838 variation.

839         If information constrains growth process estimation then perhaps the more important  
840 question for the typical assessment is what to do if reasonably accurate estimates of the length-  
841 at-age or complete age data are not available. Because so many assessments rely on size  
842 composition data, more research needs to focus on how best to approach modeling length  
843 composition data. This is especially crucial when size composition data are used to estimate  
844 abundance, as these models tend to be data rich and information poor. Even in information rich  
845 situations, more work needs to be done on the best approaches to deal with the seemingly  
846 inevitable misfit to length composition data arising from incomplete growth modeling. The  
847 relative roles of data weighting/observation error, estimation of alternative observational  
848 processes or alternative modeling approaches (like empirical weight-at-age) should be  
849 investigated. This research needs to be done with the understanding that variability in other  
850 system processes (e.g., natural mortality) and observation processes (e.g., selectivity) may be  
851 more important to estimate.



852

## **CRedit authorship contribution statement**

853

**Huihua Lee:** Writing – original draft preparation, Writing – review & editing

854

preparation, Analyses, Visualization.

855

**Kevin Piner:** Conceptualization, Writing – original draft preparation, Writing – review

856

& editing preparation.

857

**Mark Maunder:** Writing – review & editing preparation.

858

## **Declaration of Competing Interest**

859

The authors declare that they have no known competing financial interests or personal

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861

## **Data availability**

862

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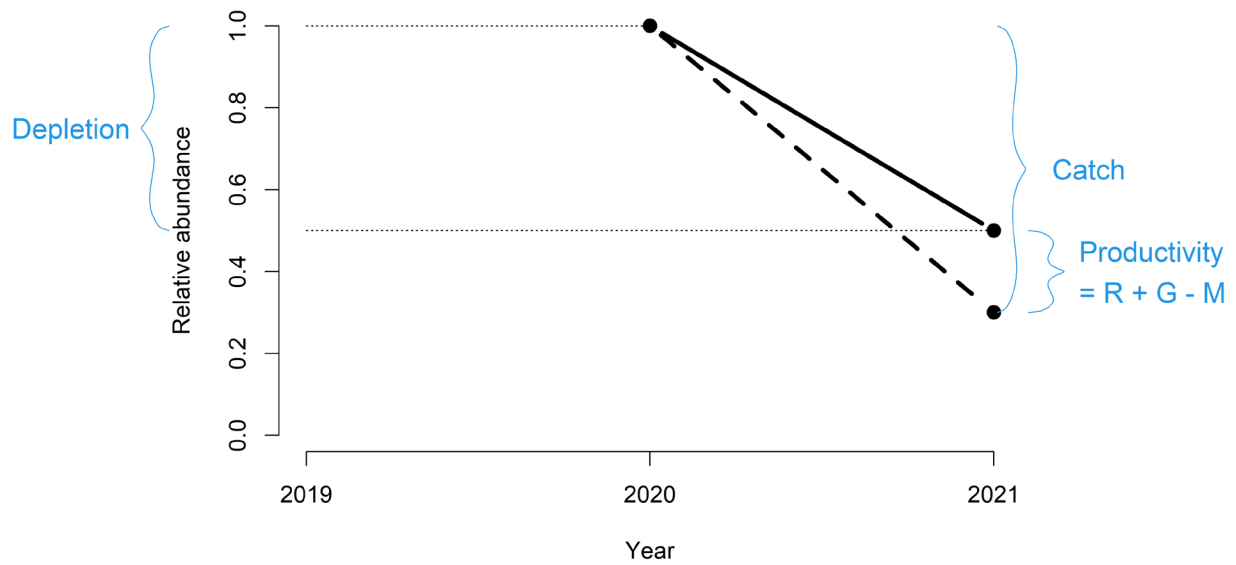
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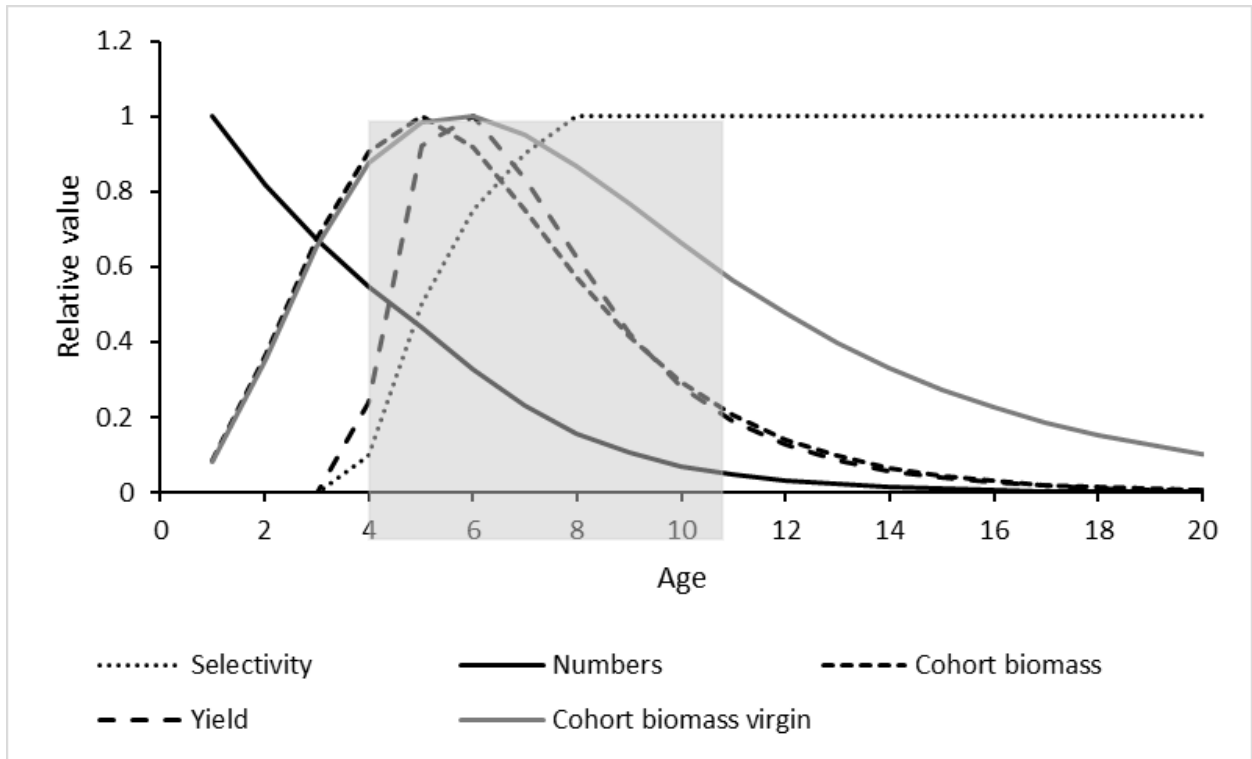
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1290

1291 Figure 1. Illustration of the absolute abundance information contained the systems model process  
 1292 of the production function, catch, and index of relative abundance. The solid and dashed lines  
 1293 represent depletion in the presence and the absence of biological processes, respectively.  $R$  is  
 1294 recruitment,  $G$  is growth, and  $M$  is natural mortality.

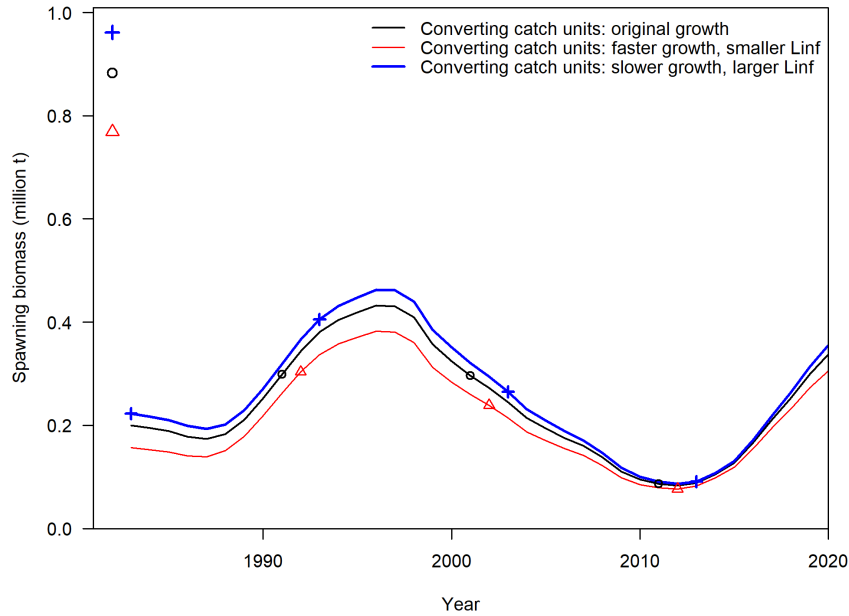




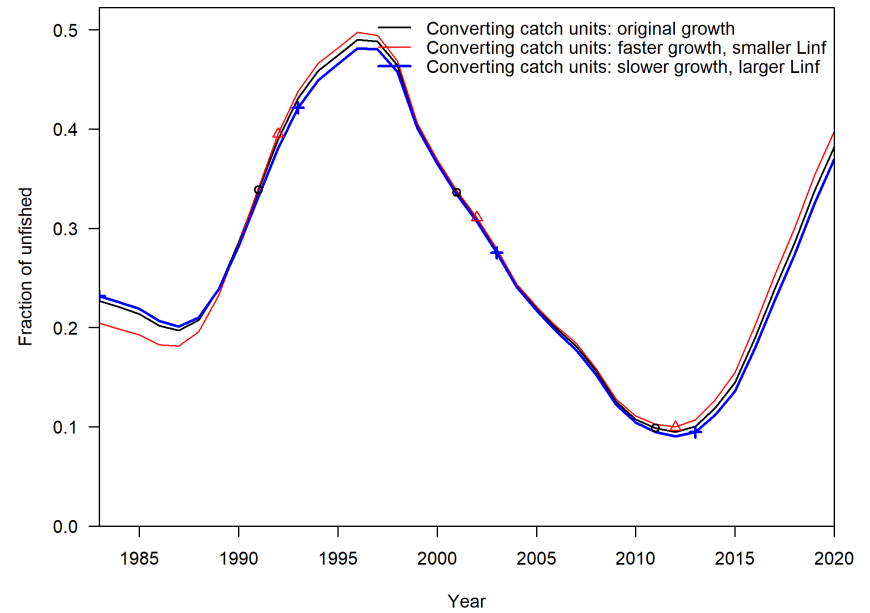
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1296 Figure 2. Illustration of the roles of both systems model and observational processes on the  
 1297 abundance of a cohort and catch-at -age.

A.

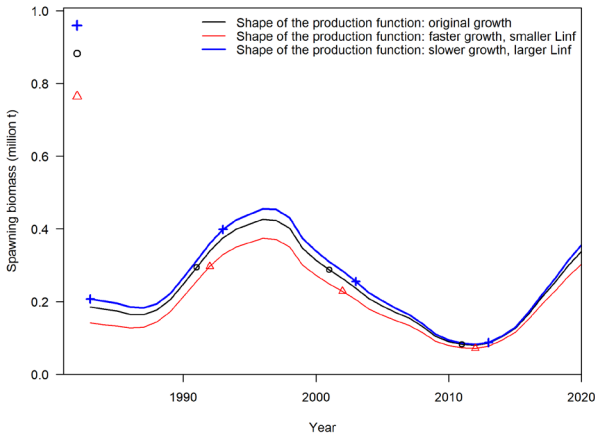


B.

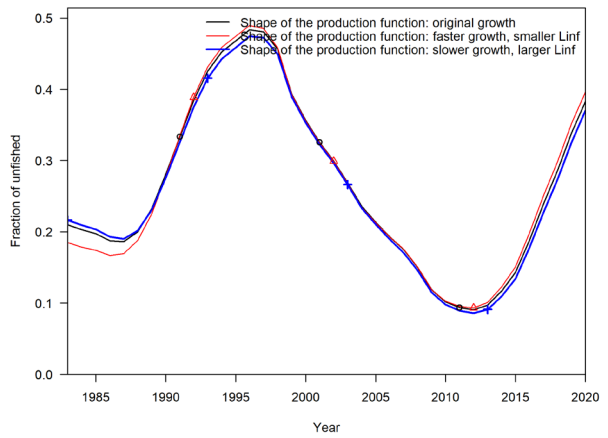


1298 Figure 4. Sensitivity diagnostic results from misspecifying the growth process on converting numbers into weight in the Pacific  
1299 Bluefin tuna stock assessment. Panel A is spawning biomass and B is fraction unfished (depletion level). The three lines in each plot  
1300 are ASPM models specifying faster growth and smaller Linf (red), original length-at-age (black), and slower growth and larger Linf  
1301 (blue). Unfished spawning biomass estimates for model runs are given in panel A as triangle, circle and cross, respectively.

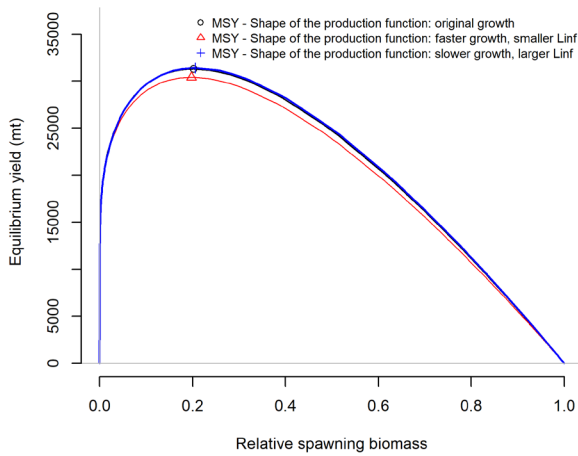
A.



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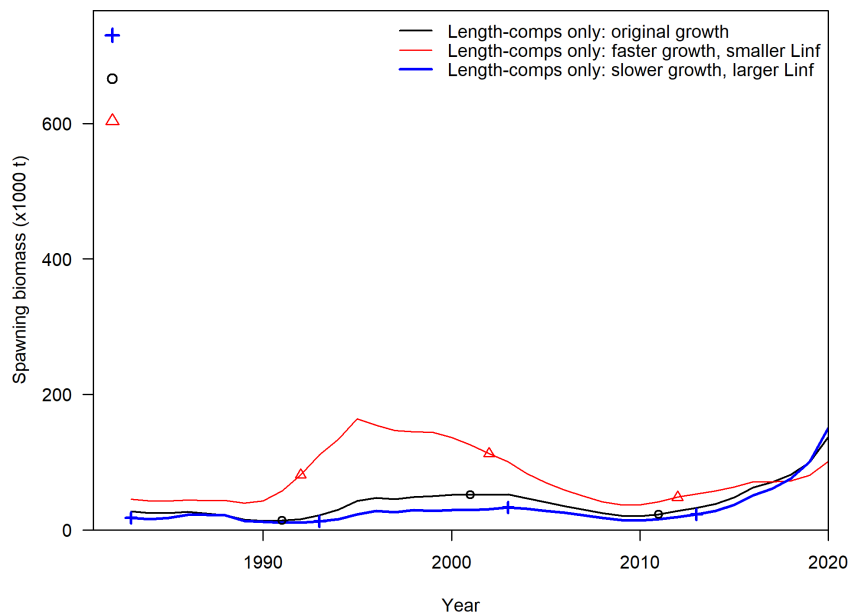
C.



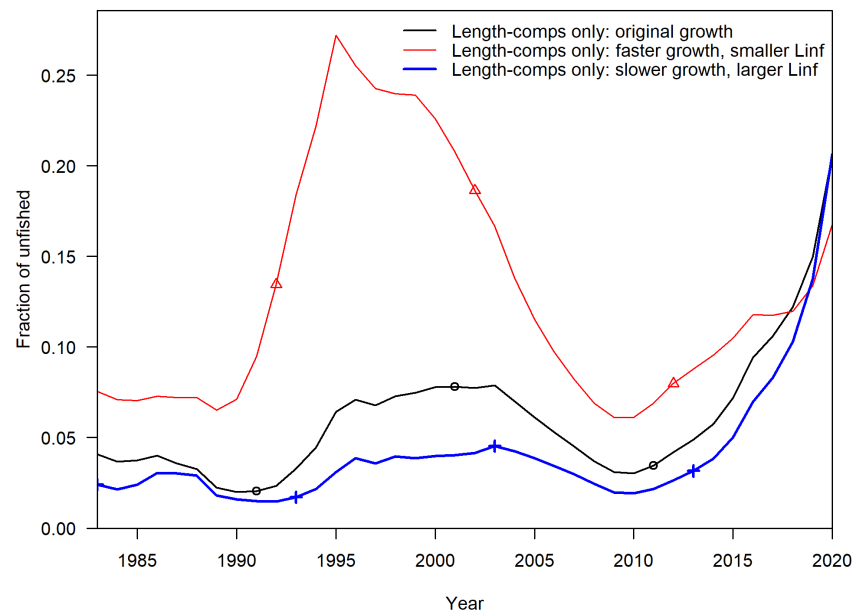
1303 Figure 5. Sensitivity diagnostic results from misspecifying the growth process on the shape of  
 1304 the production function and calculation of biomass from model estimates of numbers-at-age.  
 1305 Panel A is spawning biomass, B is fraction unfished (depletion level), and C is the production  
 1306 curve. The three lines in each plot are ASPM models specifying faster growing and smaller Linf

1307 (red), original length-at-age (black), and slower growth and larger  $L_{inf}$  (blue). Unfished  
1308 spawning biomass estimates for model runs are given in panel A as triangle, circle and cross,  
1309 respectively.

A.



B.



1311 Figure 6. Sensitivity diagnostic results from misspecifying the growth process in the Pacific Bluefin tuna stock assessment that fits to  
 1312 length compositions only. Panel A is spawning biomass and B is fraction unfishes (depletion level). The three lines in each plot are  
 1313 the models specifying faster growth and smaller Linf (red), original length-at-age (black), and slower growth and larger Linf (blue).  
 1314 Unfishes spawning biomass estimates for model runs are given in panel A as triangle, circle and cross, respectively.