¹ Apex marine predators and ocean health: proactive

² screening of halogenated organic contaminants

³ reveals ecosystem indicator species

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15 ABSTRACT

Despite decades-long bans on the production and use of certain chemicals, many halogenated organic compounds (HOCs) are persistent and can bioaccumulate in the marine environment with the potential to cause physiological harm to marine fauna. Highly lipid-rich tissue (e.g., marine mammal blubber) functions as a reservoir for HOCs, and selecting ideal indicator species is a priority for retrospective and proactive screening efforts. We selected five marine mammal species as possible indicators for the Southern California Bight (SCB) and

| 22 | applied a non-targeted analytical method paired with an automated data reduction strategy to | | | | |
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| 23 | catalog a broad range of known, known but unexpected, and unknown compounds in their | | | | |
| 24 | blubber. A total of 194 HOCs were detected across the study species (n = 25 individuals), 81% | | | | |
| 25 | of which are not routinely monitored, including 30 halogenated natural products and 45 | | | | |
| 26 | compounds of unknown structure and origin. The cetacean species (long-beaked common | | | | |
| 27 | dolphin, short-beaked common dolphin, and Risso's dolphin) averaged 128 HOCs, whereas | | | | |
| 28 | pinnipeds (California sea lion and Pacific harbor seal) averaged 47 HOCs. We suspect this | | | | |
| 29 | disparity can be attributed to differences in life history, foraging strategies, and/or enzyme- | | | | |
| 30 | mediated metabolism. Our results support proposing (1) the long- and short-beaked common | | | | |
| 31 | dolphin as apex marine predator sentinels for future and retrospective biomonitoring of the SCB | | | | |
| 32 | ecosystem and (2) the use of non-targeted contaminant analyses to identify and prioritize | | | | |
| 33 | emerging contaminants. The use of a sentinel marine species together with the non-targeted | | | | |
| 34 | analytical approach will enable a proactive approach to environmental contaminant monitoring. | | | | |
| 35 | | | | | |
| 36 | Keywords: nontargeted mass spectrometry; halogenated organic contaminants; marine | | | | |
| 37 | mammals; bioaccumulation | | | | |
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| 39 | Highlights: | | | | |
| 40 | • organic contaminants were identified in Southern California marine mammal blubber | | | | |
| 41 | • cetaceans accumulated more contaminants than pinnipeds | | | | |
| 42 | • common dolphins are proposed as sentinels for emerging contaminants in this region | | | | |
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45 1. INTRODUCTION

46 The Southern California Bight (SCB), in the eastern North Pacific Ocean, hosts notable 47 biodiversity in its coastal and pelagic habitats. A multitude of species are exposed to legacy and 48 ongoing contamination by bioaccumulative compounds such as dichlorodiphenyltrichloroethane 49 (DDT) related compounds, polychlorinated biphenyls (PCBs), and polybrominated diphenyl 50 ethers (PBDEs) (Dodder et al., 2012; Maruya and Schiff, 2009). Negative health effects such as 51 endocrine disruption (Brouwer et al., 1989), reproductive impairment (Gilmartin et al., 1976; 52 Roos et al., 2012; Schwacke et al., 2002), and immune system suppression (Hall et al., 1992; 53 Ross, 2002; Schwacke et al., 2012) experienced by marine mammals in other geographic 54 regions, but less studied in the SCB, have been attributed to chronic organic contaminant 55 exposure, potentially increasing susceptibility to infectious disease. HOCs have also been 56 speculated to enhance sensitivity of certain marine mammal species to poisoning by domoic acid 57 (Tiedeken and Ramsdell, 2010), a potent neurotoxin produced by harmful algal blooms that are 58 increasing in magnitude and frequency in the SCB (Gulland and Hall, 2007). Thus, apex 59 predators (i.e., cetaceans and pinnipeds) in this region are integral to the detection of previously 60 unrecognized environmental contaminants in addition to providing exposure data for possible 61 contaminant-related health impacts.

A comprehensive assessment of contaminant load is required to evaluate the toxicological risk associated with contaminant exposure in wildlife, which may vary depending on the complexity of the contaminant mixture (Desforges et al., 2017). Marine mammals serve as effective indicators of marine pollution due to their high trophic position, large stores of lipidrich blubber, and relatively long lifespans (Bossart, 2011; Ross, 2000). These characteristics can lead to relatively high concentrations and diverse classes of bioaccumulative HOCs from both

68 anthropogenic and natural sources. Routine targeted screening for tissue HOCs consists of 69 defined lists of compounds that may only account for a proportion of the contaminant load. As a 70 result, many uncharacterized compounds with potential to cause physiological harm may be 71 missed by routine research and monitoring (Shaul et al., 2015). A non-targeted analytical method 72 employing comprehensive two-dimensional gas chromatography coupled with time-of-flight 73 mass spectrometry (GC×GC/TOF-MS) is capable of expanding the analysis to include thousands 74 of both known and previously unidentified chemical constituents through enhanced 75 chromatographic resolution and narrower chromatographic peak widths leading to enhanced 76 sensitivity compared to single dimension GC systems (Hoh et al., 2012). The analytical method 77 was combined with searches against reference mass spectra, de novo interpretation, and matches 78 to authentic standards. The certainty was categorized based on the method of identification (e.g., 79 matches to authentic standards had the highest confidence) (Hoh et al., 2012). Recent non-80 targeted studies of dolphins and seabirds have revealed that a substantial proportion of 81 compounds are not routinely monitored (Alonso et al., 2017; Hoh et al., 2012; Millow et al., 82 2015; Shaul et al., 2015). Prior analyses of blubber samples from eight Pacific common 83 bottlenose dolphins (Tursiops truncatus) identified 280 unmonitored and/or unknown 84 compounds (86%) among 327 total HOCs (Shaul et al., 2015). The bottlenose dolphin study 85 established the utility of this analytical method for detecting an expanded range of HOCs in 86 wildlife tissues and identifying priority compounds for further investigation. 87 In this study, our primary objective was to evaluate apex marine sentinels for monitoring 88 bioaccumulative HOCs using non-targeted analysis. We created an inventory of anthropogenic 89 and naturally occurring bioaccumulative HOCs for each of five marine mammal species as

90 candidate sentinels for the SCB: eastern North Pacific long-beaked common dolphins (Delphinus

91 bairdii), short-beaked common dolphins (Delphinus delphis), Risso's dolphins (Grampus 92 griseus), California sea lions (Zalophus californianus) and Pacific harbor seals (Phoca vitulina). 93 A secondary technical objective of this study was the development of software to automatically 94 identify HOCs from their fragmentation mass spectra, enabling data reduction and sample size 95 expansion (Code 1 and 2, Supplemental Information). This was necessary in order to evaluate 96 chemical accumulation trends across species. Additional objectives were to identify unexpected 97 compounds that may pose an elevated risk based on abundance and frequency of detection, and 98 prioritize unknowns for potential source and structure determination.

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100 2. MATERIALS AND METHODS

101 Study species were selected based on established habitat range within the SCB, known 102 availability of archived specimens, and documented susceptibility or suspected resistance to 103 domoic acid toxicosis (Danil et al., 2010; Goldstein et al., 2008). Blubber samples were collected 104 between 1990 and 2014, from individuals incidentally killed during fishing activities in the SCB 105 or those that stranded dead. Five blubber samples from each of the five species were analyzed. 106 Samples were restricted to males with priority given to mature (using length as a proxy) by-107 caught individuals (Table S2) to reduce the variability caused by 1) including females known to 108 offload contaminants, and 2) age and health status (stranded individuals are generally sick). 109 Blubber samples were archived at -20 °C. Approximately 20 g of frozen, full-depth blubber was 110 sub-sampled and processed following protocols outlined by Shaul et al. (2015). Final extracts 111 were analyzed on a Pegasus 4D GC×GC/TOF-MS equipped with an Agilent 6890 gas 112 chromatograph using instrumental parameters optimized for marine mammal blubber by Hoh et 113 al. (2012).

| 114 | Data were processed using the LECO ChromaTOF mass spectrometer data system |
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| 115 | (version 4.51.6.0 optimized for Pegasus) and an automated data handling procedure. Details are |
| 116 | described in SI-1 and Figure S4. Briefly, custom data reduction software was developed based on |
| 117 | the algorithm described by Pena-Abaurrea et al., 2014, which examined mass spectra for ion |
| 118 | intensity ratios characteristic of halogenation. Additional rules and a cross-checking procedure |
| 119 | were applied to reduce the false positive rate. If the same mass spectrum was present in > 2 |
| 120 | samples, the cross-checking procedure required a manual search for the compound in the |
| 121 | remaining 23 samples. |
| 122 | On average, 479 halogenated mass spectra per sample were selected by the algorithm (out |
| 123 | of an average of 9210 chromatographic features per sample), rate of 80% to 85% compared to a |
| 124 | fully manual process (Table S3). Core components of the R script are provided in SI-1, and the |
| 125 | full R script is provided at https://github.com/OrgMassSpec/IdentifyHalogenatedSpectra. |
| 126 | Chromatographic peaks and the corresponding mass spectra selected by the algorithm were |
| 127 | manually reviewed, and confirmed halogenated compounds were structurally identified through |
| 128 | searches against existing mass spectral libraries generated from prior analysis of marine |
| 129 | mammals using the same method (Mackintosh et al., 2016; Shaul et al., 2015), searches against |
| 130 | the NIST 2014 Electron Impact Mass Spectral Library, matches to authentic reference standards, |
| 131 | or manual interpretation following the earlier methods. The mass spectra of unknown |
| 132 | compounds are provided in SI-2. All identified compounds were also identified in the |
| 133 | aforementioned prior studies and are provided in their publications (Mackintosh et al., 2016; |
| 134 | Shaul et al., 2015). |
| 135 | |

3. RESULTS AND DISCUSSION

3.1 Species Contaminant Profile Comparisons. Cetacean blubber contained markedly more
compounds than pinniped blubber, with the long-beaked common dolphin, short-beaked
common dolphin, and Risso's dolphin averaging 133, 128, and 124 HOCs, respectively. These
averages were two- to three-fold higher than average number of HOCs detected in the California
sea lion (53 HOCs) and harbor seal (40 HOCs; Table 1).

142 As a structural class, PCBs had the largest number of individual compounds, and were 143 between 28% to 32% of the total number of identified compounds in cetaceans, and 43% to 45% 144 in pinnipeds (Table 1). For the purpose of identifying which species had the highest number of 145 individual compounds, PCBs are included in the discussion below. However, parent PCB 146 compounds are not included in contaminant profile comparisons because they are well-147 characterized in the literature and frequently monitored. We included PCB metabolites 148 (methylsulfonyl-PCBs) in profile comparisons because they are typically unmonitored. A total of 149 194 different HOCs, excluding PCBs, from various anthropogenic, natural, mixed, and unknown 150 sources were identified across all 25 samples, 157 (or 81% of total) of which are not included in 151 routine environmental monitoring surveys.

152 Hierarchical clustering of samples based on similarity in anthropogenic HOC prevalence 153 identified two primary, separate clusters comprising cetacean and pinniped samples, revealing 154 that cetacean contaminant profiles were more diverse compared to pinnipeds (Figure S1). The 155 heat map shows this difference is not limited to a few compounds, but is consistent across the 156 majority of individual contaminants. The most frequently detected compounds included $p_{,p}$ '-157 DDE, trans nonachor, tris(4-chlorophenyl)methane (4,4',4"-TCPM), BDE-47, BDE-100, and 158 BDE-99, all of which were detected in all 25 samples. Two potential confounding factors 159 influencing contaminant accumulation were animal length (as a proxy for age) and collection

year (Table S2). Contaminant profiles within species were not associated with either of these twofactors (data not shown).

162 The HOCs detected in this study were organized into 35 classes based on similarity of 163 molecular structure (Table 1; structural class descriptions in Table S1). DDT-related compounds 164 constituted the most abundant structural class in both of the cetacean and pinniped groups 165 (Figure 1), as well as for each species (Figure S2), which is consistent with the established 166 contaminant signature found in marine mammals from this region (Blasius and Goodmanlowe, 167 2008; Shaul et al., 2015). The TCPM structural class consisted of eight isomers and was the 168 second-most abundant. It was detected at levels comparable to the chlordane-related and PBDE 169 classes, both of which are legacy HOCs of concern in the SCB (Dodder et al., 2012; Maruya and 170 Schiff, 2009). Following its discovery in Puget Sound and Baltic Sea seals around 1990 (Walker 171 et al., 1989; Zook et al., 1992), 4,4',4''-TCPM was determined to be an impurity from the 172 technical preparation of pesticides such as DDT (Buser, 1995) and dicofol (de Boer, 1997). 173 TCPM and its derivative, tris(4-chlorophenyl)methanol (TCPMOH), were measured in marine 174 mammals from North America and Asia in the early 2000s, and were determined to have high 175 biomagnification potential (Kajiwara, K. Kannan, M. Muraoka, M., 2001; Kannan et al., 2004; 176 Minh et al., 2000; Watanabe et al., 2000). Although previously found to be one to three orders of 177 magnitude greater in sea lions off the Northern and Central California coast compared to species 178 from other locations (Kajiwara, K. Kannan, M. Muraoka, M., 2001; Kannan et al., 2004), TCPM 179 and TCPMOH appear to have lost recognition as a pervasive contaminant in the North Pacific 180 over the past decade. The parent compound, 4,4',4"-TCPM, was detected in every blubber 181 sample in this study. Notably, five additional TCPM isomers were present in nearly all dolphin 182 samples (Figure 2) and constitute a relevant portion of the accumulated structural class.

183 However, within the SCB, these isomers have only been observed by prior non-targeted studies

184 of bottlenose dolphins (Mackintosh et al., 2016; Shaul et al., 2015). The consistent ratio of

185 TCPM isomers observed across dolphin samples suggests similar exposure to a consistent

186 mixture of isomers, constituting a persistent and unmonitored

187 chemical mixture that could permeate trophic levels throughout the region.

188 3.2 Halogenated Natural Products. The 30 identified halogenated natural products (HNPs) 189 belonged to six different structural classes and accounted for 15% of the HOCs. These 190 compounds are produced by algae or sponges and have a biomagnification potential similar to 191 anthropogenic HOCs (Pangallo and Reddy, 2010). Despite their global distribution and profusion 192 in marine food webs and biota, the health implications of exposure to HNPs are largely unknown 193 (Shaul et al., 2015). We observed a more diverse array of HNPs in the dolphin species compared 194 to the pinnipeds, with dimethyl bipyrroles (DMBPs) the most prevalent and abundant class 195 (Table 1, Figure 1). 1,1'-Dimethyl-tetrabromo-dichloro-2,2'-bipyrrole (DMBP Br₄Cl₂) was the 196 most common individual natural product, and was detected in all but one sea lion and four harbor 197 seal samples (Figure S2). This compound was previously detected in California sea lion blubber 198 at concentrations rivaling that of BDE 47 (Stapleton et al., 2006). Heptachlorinated 1-methyl 199 1',2-bipyrrole (MBP 7Cl) was the second-most common HNP and was found in all cetacean 200 samples and three sea lion samples. This widespread natural product, also referred to as Q1, has 201 been identified in Atlantic marine mammals (Teuten et al., 2006).

HNP profiles have been shown to be unique to different marine species and ecotypes
within a geographic region (Dorneles et al., 2010; Hauler et al., 2013; Pangallo and Reddy, 2010;
Shaul et al., 2015). The species in this study displayed nearly unique HNP profiles (Figure 3).
California sea lions clustered separately from harbor seals, and all common dolphins grouped

separately from the Risso's dolphins (with the exception of two short-beaked common dolphins),
suggesting that foraging strategies and locations may be discernable using HNP profiles. In
contrast, anthropogenic profiles were unique between cetaceans and pinnipeds, but species
within these taxonomic groups were not different (Figure S1).

210 The difference between HNP and anthropogenic contaminant clustering may be due to 211 the origin of these compounds to the local marine environment. Legacy anthropogenic 212 compounds originated from terrestrial sources and now contaminate broad areas of marine 213 sediment in the SCB (Dodder et al., 2012; Maruya and Schiff, 2009). HNPs may have spatially 214 distinct ecological profiles based on algal and sponge community composition, leading to unique 215 bioaccumulation patterns based on the habitats and foraging strategies of the different species. 216 HNPs have important potential applications beyond toxicological assessments, including the 217 differentiation of ecotypes or populations within species (Dorneles et al., 2010; Shaul et al., 218 2015), evaluating trophic relationships (Pangallo and Reddy, 2010), and could enhance indirect 219 chemical methods for determining diet composition of free-ranging marine mammal species 220 (Bowen and Iverson, 2012).

221 **3.3 Unknown Contaminants.** Based on isotopic profiles in their mass spectra, 45 observed 222 compounds were determined to contain halogens, but their complete structural identities could 223 not be determined through mass spectral searches or de novo interpretation (mass spectra in SI-2 224 and accessible online at https://github.com/OrgMassSpec/SpecLibMarineUnknown2018). Ten 225 of the halogenated unknown compounds have not, to our knowledge, been previously identified 226 in marine biota; they are first 10 spectra described in SI-2. The number of unknowns accounted 227 for 24% of the total number of identified HOCs and outnumbered each of the DDT-related 228 (n=24), chlordane-related (n=14) and PBDE (n=16) structural classes. These unidentified

229 compounds could be emerging contaminants from current anthropogenic activities, previously 230 undiscovered legacy pollutants, or halogenated natural products. For example, the unknown-8-2 231 isomer (SI-2, page 46) was relatively abundant in cetacean blubber and a significant regression 232 was found with Σ DDT-Related (R²=0.79, p<0.001), Σ Chlordane-Related (R²=0.74, p<0.001), 233 Σ TCPM (R²=0.82, p<0.001, and the TCPMOH Isomer 2 metabolite (R²=0.88, p<0.001); Figure 234 4), indicating the unknown compound may have a similar history of anthropogenic use in this 235 region. Unknown-8-2 did not have a significant regression with Σ PBDE, a compound class with 236 a different history compared to the other anthropogenic contaminants, including later peak use 237 (Dodder et al., 2012). There was a significant, but less robust regression with $\Sigma DMBP$ (R²=0.4, 238 p < 0.001), indicating unknown-8-2 may not be a natural product.

239 3.4 Differences in Cetacean vs. Pinniped Profiles. Pinnipeds appear to accumulate fewer 240 compounds across a smaller number of structural classes, however the compounds that do 241 accumulate in pinniped blubber are found at similar abundance to those in cetaceans (Figure 1). 242 Although California sea lions feed on the same pelagic schooling fish as common dolphins, and 243 even regularly feed in industrialized embayments (Meng et al., 2009), the multitude of HOCs in 244 their contaminant profile is reduced (53 average total HOCs in sea lions versus 133 and 128 245 average total HOCs in long- and short-beaked common dolphins, respectively; Table 1). Possible 246 explanations for this discrepancy are likely physiological, such as differences in life history 247 strategies and enzyme-mediated metabolism. Pinnipeds undergo periods of fasting during annual 248 breeding and molting seasons (Ling, 1970). These processes are energetically costly, enhancing 249 blubber turnover and mobilizing contaminants to the bloodstream (Peterson et al., 2014). Small 250 cetacean species maintain a homogenous HOC profile across blubber depth indicating more 251 stable concentrations over time (Méndez-Fernandez et al., 2016). Cetaceans also have a lower

252 capacity for metabolizing PCBs compared to other terrestrial and marine mammals, such as seals 253 (Routti et al., 2008; Tanabe et al., 1988), due to lower or absent CYP2B liver enzyme activity 254 (Boon et al., 1997). An enhanced capacity for metabolizing and excreting certain organic 255 contaminants was proposed to explain the much lower levels of TCPM and TCPMOH observed 256 in pinnipeds compared to cetaceans (Minh et al., 2000). Enhanced metabolism has also been 257 proposed as an explanation for the absence of HNPs from pinniped blubber despite presence in 258 prey items (Pangallo and Reddy, 2010) and the lack of methoxy-BDEs detected in California sea 259 lions (Stapleton et al., 2006). The expanded range of HOCs evaluated in this study revealed 260 additional structural classes that appear to align with this phenomenon (Figure 1). 261 **3.5 Best Sentinel Determination.** Based upon the multitude and abundance of detected HOCs, 262 cetacean species are more effective regional bioindicators for cataloging emerging and unknown 263 HOCs. The following discussion excludes PCBs because they were not previously included in 264 the associated bottlenose dolphin study (Shaul et al., 2015). Whereas 103 anthropogenic and 265 naturally occurring unmonitored compounds, and 45 unknowns, were detected in the cetacean 266 samples from the current study (94% of all unmonitored compounds from both cetaceans and 267 pinnipeds), only 46 typically unmonitored HOCs from anthropogenic and natural sources, and 12 268 unknowns, were detected in pinniped blubber. Within the cetaceans, the long-beaked common, 269 short-beaked common, and Risso's dolphin samples averaged 90, 89, and 89 HOCs per sample, 270 respectively. In contrast, an average of 209 HOCs per sample (n=8) was detected in Pacific 271 common bottlenose dolphins in the prior study using the same blubber sampling and 272 instrumental method (Shaul et al., 2015). The bottlenose dolphins were collected from the same 273 region, during approximately the same timeframe (1995-2010). The data processing methods 274 were different. Halogenated compounds in the bottlenose dolphins were identified by a fully

manual process. For the current project, as described in detail in SI-1, we used an automated
process (with manual cross-checking between samples as a final step). Direct comparison of the
two processes determined the automated process detection rate of halogenated compounds was
80% to 85% of the manual process. The difference in detection rates is not large enough to
account for the difference in the number of HOCs detected in bottlenose dolphins compared to
the other species.

281 This indicates the bottlenose is the most contaminated marine mammal among the six 282 species studied in the SCB. However, bottlenose dolphins do not often strand, limiting 283 availability of full-depth blubber samples from this species, and hampering ongoing alternate 284 research that relies on live biopsy samples (Figure S3). Archived specimens are also 285 exceptionally rare for the Risso's dolphin, which typically range offshore. Therefore, based on 286 similar HOC profiles and readily available sample material, it is recommended that long- and 287 short-beaked common dolphins (which contained 80% of the unmonitored compounds found 288 among all species in this study) serve as effective sentinels for fulfilling the two main priorities 289 of the non-targeted work in the SCB: retrospective evaluation of contaminant trends and 290 proactive screening for emerging contaminants of concern.

3.6 Domoic Acid Susceptibility. Species with observed sensitivity to domoic acid toxicosis (i.e., long-beaked common dolphins, short-beaked common dolphin, and California sea lions) did not contain unique contaminant signatures compared to those that have not been documented to experience mass mortality events following blooms (i.e., common bottlenose dolphins, Risso's dolphins, and harbor seals (Gulland and Hall, 2007; Lefebvre et al., 1999). In fact, domoic acid-related stranding events were not observed for the most contaminated species, the Pacific common bottlenose dolphin. This suggests contaminants do not play a profound role in the

domoic acid sensitivity exhibited by particular SCB marine mammal populations at the species
level. This result reinforces existing speculation that prey preference likely has the greatest
impact on the risk of poisoning by domoic acid (Lefebvre et al., 1999). The Northern anchovy
has been implicated as the primary vector of exposure and is a preferred prey item for California
sea lions, long-, and short-beaked common dolphins (Lowry and Carretta, 1999), all of which
experience domoic acid toxicosis (Danil et al., 2010).

304 3.7 Implications for Contaminant Monitoring. The population of the five coastal counties 305 bordering the SCB is projected to increase to over 20 million people by 2020 (Maruya and 306 Schiff, 2009), thus there is a demand for vigilance in monitoring of both legacy and emerging 307 contaminants in this ecologically and economically valuable coastal zone. Therefore, as new 308 compounds are released into the environment, non-targeted contaminant analysis allows 309 researchers and managers to comprehensively screen for emerging compounds of concern, fast-310 tracking them for possible source, structure, and toxicity testing. Further evaluation and 311 characterization of the unknown compounds highlighted in this study is imperative for assessing 312 seafood safety and possible distribution of such contaminants on a global scale, particularly 313 when the abundance of these compounds rivals that of many legacy pollutants. Additionally, 314 chemicals identified in this study may serve as unique tracers to gather information about the 315 physiology and ecology of marine mammal species that serve as sentinels of marine 316 environmental quality.

317

318 SUPPLEMENTARY DATA

319 The following is the supplementary data related to this article:

320 Supporting Information Part 1 (SI-1) and Supporting Information Part 2 (SI-2).

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- 326 N.G.D., E.H., S.J.C., D.W.W., K.D., and K.A.M. designed the research plan, S.J.C., D.W.W.,
- and K.D. contributed sample material, J.M.C performed the sample preparation, J.M.C., N.G.D.,
- and E.H. performed the instrumental and data analysis, N.G.D. wrote the custom software, and
- 329 J.M.C., N.G.D., E.H., S.J.C., D.W.W., K.D., and K.A.M wrote the paper.
- 330 Notes
- 331 The authors declare no conflict of interest.
- 332

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- 344 **REFERENCES**
- 345

- Alonso, M.B., Maruya, K.A., Dodder, N.G., Lailson-Brito, J., Azevedo, A., Santos-Neto, E.,
 Torres, J.P.M., Malm, O., Hoh, E., 2017. Nontargeted Screening of Halogenated
 Organic Compounds in Bottlenose Dolphins (Tursiops truncatus) from Rio de
- 351 Janeiro, Brazil. Environ. Sci. Technol. 51, 1176–1185.
- 352 https://doi.org/10.1021/acs.est.6b04186
- Blasius, M.E., Goodmanlowe, G.D., 2008. Contaminants still high in top-level carnivores in
 the Southern California Bight: Levels of DDT and PCBs in resident and transient
 pinnipeds. Mar. Pollut. Bull. 56, 1973–1982.
- 356 https://doi.org/10.1016/j.marpolbul.2008.08.011
- Boon, J.P., Van der Meer, J., Allchin, C.R., Law, R.J., Klungsøyr, J., Leonards, P.E.G., Spliid, H.,
 Storr-Hansen, E., Mckenzie, C., Wells, D.E., 1997. Concentration-dependent changes
 of PCB patterns in fish-eating mammals: structural evidence for induction of
 cytochrome P450. Arch. Environ. Contam. Toxicol. 33, 298–311.
- Bossart, G.D., 2011. Marine Mammals as Sentinel Species for Oceans and Human Health.
 Vet. Pathol. 48, 676–690. https://doi.org/10.1177/0300985810388525
- Bowen, W.D., Iverson, S.J., 2012. Methods of estimating marine mammal diets: A review of
 validation experiments and sources of bias and uncertainty. Mar. Mammal Sci. 29,
 719–754. https://doi.org/10.1111/j.1748-7692.2012.00604.x
- Brouwer, A., Reijnders, P.J.H., Koeman, J.H., 1989. Polychlorinated biphenyl (PCB)contaminated fish induces vitamin A and thyroid hormone deficiency in the
 common seal (Phoca vitulina). Aquat. Toxicol. 15, 99–106.
- Buser, H.R., 1995. DDT, a Potential Source of Environmental Tris(4-chlorophenyl)methane
 and Tris(4-chlorophenyl)methanol. Environ. Sci. Technol. 29, 2133–2139.
- Danil, K., Chivers, S.J., Henshaw, M.D., Thieleking, J.L., Daniels, R., Leger, J.A.S., 2010.
 Cetacean strandings in San Diego County, California, USA. J. Cetacean Res. Manag. 11, 163–184.
- de Boer, J., 1997. Environmental Distribution and Toxicity of Tris(4 Chlorophenyl)Methanol and Tris(4-Chlorophenyl)Methane, in: Reviews of
 Environmental Contamination and Toxicology. pp. 95–106.
- Desforges, J.-P., Levin, M., Jasperse, L., De Guise, S., Eulaers, I., Letcher, R.J., Acquarone, M.,
 Nordøy, E., Folkow, L.P., Hammer Jensen, T., Grøndahl, C., Bertelsen, M.F., St. Leger, J.,
 Almunia, J., Sonne, C., Dietz, R., 2017. Effects of Polar Bear and Killer Whale Derived
 Contaminant Cocktails on Marine Mammal Immunity. Environ. Sci. Technol. 51,
 11431–11439. https://doi.org/10.1021/acs.est.7b03532
- Dodder, N.G., Maruya, K.A., Lauenstein, G.G., Ramirez, J., Ritter, K.J., Schiff, K.C., 2012.
 Distribution and sources of polybrominated diphenyl ethers in the southern california bight. Environ. Toxicol. Chem. SETAC 31, 2239–2245.
 https://doi.org/10.1002/etc.1957
- Dorneles, P.R., Lailson-Brito, J., Dirtu, A.C., Weijs, L., Azevedo, A.F., Torres, J.P.M., Malm, O.,
 Neels, H., Blust, R., Das, K., Covaci, A., 2010. Anthropogenic and naturally-produced
 organobrominated compounds in marine mammals from Brazil. Environ. Int. 36,
 60–67. https://doi.org/10.1016/j.envint.2009.10.001

- 390 Gilmartin, W.G., Delong, R.L., Smith, A.W., Sweeney, J.C., LAPPE, B.W.D., Risebrough, R.W., 391 Griner, L.A., Dailey, M.D., Peakall, D.B., 1976. Premature parturition in the California 392 sea lion. J. Wildl. Dis. 12, 104–115.
- 393 Goldstein, T., Mazet, J.A.K., Zabka, T.S., Langlois, G., Colegrove, K.M., Silver, M., Bargu, S., Van 394 Dolah, F., Leighfield, T., Conrad, P.A., Barakos, J., Williams, D.C., Dennison, S., 395 Haulena, M., Gulland, F.M.D., 2008. Novel symptomatology and changing
- 396 epidemiology of domoic acid toxicosis in California sea lions (Zalophus
- 397 californianus): an increasing risk to marine mammal health. Proc. R. Soc. Lond. B 398 Biol. Sci. 275, 267–276.
- 399 Gulland, F.M.D., Hall, A.J., 2007. Is Marine Mammal Health Deteriorating? Trends in the 400 Global Reporting of Marine Mammal Disease. EcoHealth 4, 135–150. https://doi.org/10.1007/s10393-007-0097-1 401
- 402 Hall, A.J., Law, R.J., Wells, D.E., Harwood, J., Ross, H.M., Kennedy, S., Allchin, C.R., Campbell, 403 L.A., Pomerov, P.P., 1992. Organochlorine levels in common seals (Phoca vitulina) 404 which were victims and survivors of the 1988 phocine distemper epizootic. Sci. 405 Total Environ. 115, 145–162.
- 406 Hauler, C., Martin, R., Knölker, H.-J., Gaus, C., Mueller, J.F., Vetter, W., 2013. Discovery and 407 widespread occurrence of polyhalogenated 1,1'-dimethyl-2,2'-bipyrroles (PDBPs) in 408 marine biota. Environ. Pollut. 178, 329-335. 409
 - https://doi.org/10.1016/j.envpol.2013.03.025
- 410 Hoh, E., Dodder, N.G., Lehotay, S.J., Pangallo, K.C., Reddy, C.M., Maruya, K.A., 2012. 411 Nontargeted comprehensive two-dimensional gas chromatography/time-of-flight 412 mass spectrometry method and software for inventorying persistent and 413 bioaccumulative contaminants in marine environments. Environ. Sci. Technol. 46,
- 414 8001-8008. https://doi.org/10.1021/es301139q
- Kajiwara, K. Kannan, M. Muraoka, M., N., 2001. Organochlorine Pesticides, Polychlorinated 415 416 Biphenyls, and Butyltin Compounds in Blubber and Livers of Stranded California Sea 417 Lions, Elephant Seals, and Harbor Seals from Coastal California, USA. Arch. Environ. Contam. Toxicol. 41, 90–99. https://doi.org/10.1007/s002440010224 418
- 419 Kannan, K., Kajiwara, N., Le Boeuf, B., Tanabe, S., 2004. Organochlorine pesticides and 420 polychlorinated biphenyls in California sea lions. Environ. Pollut. 131, 425–434. 421 https://doi.org/10.1016/j.envpol.2004.03.004
- 422 Lefebvre, K.A., Powell, C.L., Busman, M., Doucette, G.J., Moeller, P.D., Silver, J.B., Miller, P.E., 423 Hughes, M.P., Singaram, S., Silver, M.W., others, 1999. Detection of domoic acid in 424 northern anchovies and California sea lions associated with an unusual mortality 425 event. Nat. Toxins 7, 85-92.
- 426 Ling, J.K., 1970. Pelage and molting in wild mammals with special reference to aquatic 427 forms. O. Rev. Biol. 16-54.
- 428 Lowry, M.S., Carretta, J.V., 1999. Market squid (Loligo opalescens) in the diet of California 429 sea lions (Zalophus californianus) in southern California (1981-1995). Calif. Coop. 430 Ocean. Fish. Investig. Rep. 196–207.
- 431 Mackintosh, S.A., Dodder, N.G., Shaul, N.J., Aluwihare, L.I., Maruya, K.A., Chivers, S.J., Danil, 432 K., Weller, D.W., Hoh, E., 2016. Newly Identified DDT-Related Compounds
- 433 Accumulating in Southern California Bottlenose Dolphins. Environ. Sci. Technol. 50,
- 434 12129–12137. https://doi.org/10.1021/acs.est.6b03150

- Maruya, K.A., Schiff, K., 2009. The extent and magnitude of sediment contamination in the
 Southern California Bight, in: Geological Society of America Special Papers.
 Geological Society of America, pp. 399–412.
- 438 Méndez-Fernandez, P., Galluzzi Polesi, P., Taniguchi, S., de O. Santos, M.C., Montone, R.C.,
 439 2016. Validating the use of biopsy sampling in contamination assessment studies of
 440 small cetaceans. Mar. Pollut. Bull. 107, 364–369.
- 441 https://doi.org/10.1016/j.marpolbul.2016.04.021
- Meng, X.-Z., Blasius, M.E., Gossett, R.W., Maruya, K.A., 2009. Polybrominated diphenyl ethers
 in pinnipeds stranded along the southern California coast. Environ. Pollut. 157,
 2731–2736. https://doi.org/10.1016/j.envpol.2009.04.029
- Millow, C.J., Mackintosh, S.A., Lewison, R.L., Dodder, N.G., Hoh, E., 2015. Identifying
 bioaccumulative halogenated organic compounds using a nontargeted analytical
 approach: seabirds as sentinels. PloS One 10, e0127205.
 https://doi.org/10.1371/journal.pone.0127205
- Minh, T.B., Watanabe, M., Tanabe, S., Miyazaki, N., Jefferson, T.A., Prudente, M.S.,
 Subramanian, A., Karuppiah, S., 2000. Widespread contamination by tris (4-
- 450 Sublamanian, A., Kaluppian, S., 2000. Widespread containnation by this (4-451 chlorophenyl) methane and tris (4-chlorophenyl) methanol in cetaceans from the 452 North Pacific and Asian coastal waters. Environ. Pollut. 110, 459–468.
- Pangallo, K.C., Reddy, C.M., 2010. Marine Natural Products, the Halogenated 1'-Methyl-1,2'bipyrroles, Biomagnify in a Northwestern Atlantic Food Web. Environ. Sci. Technol.
 44, 5741–5747. https://doi.org/10.1021/es101039d
- Peterson, S.H., Hassrick, J.L., Lafontaine, A., Thomé, J.-P., Crocker, D.E., Debier, C., Costa, D.P.,
 2014. Effects of Age, Adipose Percent, and Reproduction on PCB Concentrations and
 Profiles in an Extreme Fasting North Pacific Marine Mammal. PLoS ONE 9, e96191.
 https://doi.org/10.1371/journal.pone.0096191
- Roos, A.M., Bäcklin, B.-M.V.M., Helander, B.O., Rigét, F.F., Eriksson, U.C., 2012. Improved
 reproductive success in otters (Lutra lutra), grey seals (Halichoerus grypus) and sea
 eagles (Haliaeetus albicilla) from Sweden in relation to concentrations of
 organochlorine contaminants. Environ. Pollut. 170, 268–275.
- 464 https://doi.org/10.1016/j.envpol.2012.07.017
- 465 Ross, P.S., 2002. The Role of Immunotoxic Environmental Contaminants in Facilitating the
 466 Emergence of Infectious Diseases in Marine Mammals. Hum. Ecol. Risk Assess. Int. J.
 467 8, 277–292. https://doi.org/10.1080/20028091056917
- 468 Ross, P.S., 2000. Marine Mammals as Sentinels in Ecological Risk Assessment. Hum. Ecol.
 469 Risk Assess. Int. J. 6, 29–46. https://doi.org/10.1080/10807030091124437
- Routti, H., Letcher, R.J., Arukwe, A., van Bavel, B., Yoccoz, N.G., Chu, S., Gabrielsen, G.W.,
 2008. Biotransformation of PCBs in Relation to Phase I and II XenobioticMetabolizing Enzyme Activities in Ringed Seals (Phoca hispida) from Svalbard and
 the Baltic Sea. Environ. Sci. Technol. 42, 8952–8958.
 https://doi.org/10.1021/es801682f
- Schwacke, L.H., Voit, E.O., Hansen, L.J., Wells, R.S., Mitchum, G.B., Hohn, A.A., Fair, P.A., 2002.
 Probabilistic risk assessment of reproductive effects of polychlorinated biphenyls
 on bottlenose dolphins (Tursiops truncatus) from the Southeast United States coast.
- Environ. Toxicol. Chem. 21, 2752–2764. https://doi.org/10.1002/etc.5620211232
 Schwacke, L.H., Zolman, E.S., Balmer, B.C., De Guise, S., George, R.C., Hoguet, J., Hohn, A.A.,
 Kucklick, J.R., Lamb, S., Levin, M., Litz, J.A., McFee, W.E., Place, N.J., Townsend, F.I.,

| 481 | Wells, R.S., Rowles, T.K., 2012. Anaemia, hypothyroidism and immune suppression |
|-----|---|
| 482 | associated with polychlorinated biphenyl exposure in bottlenose dolphins (Tursiops |
| 483 | truncatus). Proc. Biol. Sci. 279, 48–57. https://doi.org/10.1098/rspb.2011.0665 |
| 484 | Shaul, N.J., Dodder, N.G., Aluwihare, L.I., Mackintosh, S.A., Maruya, K.A., Chivers, S.J., Danil, |
| 485 | K., Weller, D.W., Hoh, E., 2015. Nontargeted biomonitoring of halogenated organic |
| 486 | compounds in two ecotypes of bottlenose dolphins (Tursiops truncatus) from the |
| 487 | Southern California Bight. Environ. Sci. Technol. 49, 1328–1338. |
| 488 | https://doi.org/10.1021/es505156q |
| 489 | Stapleton, H.M., Dodder, N.G., Kucklick, J.R., Reddy, C.M., Schantz, M.M., Becker, P.R., Gulland, |
| 490 | F., Porter, B.J., Wise, S.A., 2006. Determination of HBCD, PBDEs and MeO-BDEs in |
| 491 | California sea lions (Zalophus californianus) stranded between 1993 and 2003. Mar. |
| 492 | Pollut. Bull. 52, 522–531. https://doi.org/10.1016/j.marpolbul.2005.09.045 |
| 493 | Tanabe, S., Watanabe, S., Kan, H., Tatsukawa, R., 1988. Capacity and Mode of PCB |
| 494 | Metabolism in Small Cetaceans. Mar. Mammal Sci. 4, 103–124. |
| 495 | Teuten, E.L., Pedler, B.E., Hangsterfer, A.N., Reddy, C.M., 2006. Identification of highly |
| 496 | brominated analogues of Q1 in marine mammals. Environ. Pollut. 144, 336–344. |
| 497 | https://doi.org/10.1016/j.envpol.2005.10.052 |
| 498 | Tiedeken, J.A., Ramsdell, J.S., 2010. Zebrafish Seizure Model Identifies p,p'-DDE as the |
| 499 | Dominant Contaminant of Fetal California Sea Lions That Accounts for Synergistic |
| 500 | Activity with Domoic Acid. Environ. Health Perspect. 118, 545–551. |
| 501 | Walker, W., Jarman, W.M., de Lappe, B.W., Tefft, J.A., 1989. Identification of |
| 502 | Tris(chlorophenyl)methanol in Blubber of Harbor Seals from Puget Sound. |
| 503 | Chemosphere 18, 1799–1804. |
| 504 | Watanabe, M., Kannan, K., Takahashi, A., Loganathan, B.G., Odell, D.K., Tanabe, S., Giesy, J.P., |
| 505 | 2000. Polychlorinated biphenyls, organochlorine pesticides, tris (4-chlorophenyl) |
| 506 | methane, and tris (4-chlorophenyl) methanol in livers of small cetaceans stranded |
| 507 | along Florida coastal waters, USA. Environ. Toxicol. Chem. 19, 1566–1574. |
| 508 | Zook, D.R., Buser, H.R., Bergqvist, PA., Rappe, C., Olsson, M., 1992. Detection of |
| 509 | Tris(chlorophenyl)methane and Tris(4-chlorophenyl)methanol in Ringed Seal |
| 510 | (Phoca hispida) from the Baltic Sea. Ambio 21, 557–560. |
| 511 | |









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TCPM







| | No. HOCs in class | (No. not typically monitored) | Source | Long-beaked common dolphin (<i>D. capensis</i> , n=5) Average no. HOCs | Short-beaked common dolphin (D. delphis, n=5) Average no. HOCs | Risso's dolphin (G. griseus, n=5) Average no. HOCs | California sea lion (Z. californianus, n=5) Average no. HOCs | Harbor seal (P. vitulina, n=5) Average no. HOCs |
|-------------------------------------|----------------------|-------------------------------------|---------------|--|--|---|--|---|
| Structural class | | | | | | | | |
| B/CDE | 1 | 1 | unknown | 0 | 0 | 0 | 0 | 0 |
| Brominated anisole | 1 | 1 | mixed | 1(1) | 0 | 0 | 0 | 0 |
| Chlordane-related | 14 | 9 | anthropogenic | 8 (3) | 8 (4) | 7 (3) | 5 (2) | 1 (0) |
| Chlorinated benzene | 1 | 1 | anthropogenic | 1(1) | 1(1) | 1(1) | 0 | 1 (1) |
| Chlorinated styrene | 1 | 1 | anthropogenic | 1(1) | 0 | 0 | 0 | 0 |
| DDT-related | 24 | 16 | anthropogenic | 13 (7) | 12 (5) | 10 (4) | 4 (2) | 3 (1) |
| Dichlorobenzophenone | 1 | 1 | anthropogenic | 0 | 0 | 0 | 0 | 0 |
| Dieldrin | 1 | 0 | anthropogenic | 0 | 1 (0) | 0 | 0 | 0 |
| DMBP | 16 | 16 | natural | 8 (8) | 12 (12) | 13 (13) | 2(2) | 0 |
| HCH-related | 2 | 0 | anthropogenic | 1 (0) | 1 (0) | 1 (0) | 1 (0) | 0 |
| Heptachlor-related | 3 | 2 | anthropogenic | 1(1) | 2(1) | 1 (0) | 0 | 0 |
| MBP | 6 | 6 | natural | 4 (4) | 4 (4) | 3 (3) | 2 (2) | 0 |
| MeO-B/CDE | 1 | 1 | natural | 0 | 0 | 1(1) | 0 | 0 |
| MeO-BDE | 3 | 3 | natural | 2 (2) | 2 (2) | 2 (2) | 0 | 0 |
| MeO-CDE | 1 | 1 | unknown | 0 | 0 | 0 | 0 | 0 |
| MeO-PBB | 1 | 1 | natural | 1(1) | 1(1) | 1(1) | 0 | 0 |
| Methylenebistrichloroanisole | 1 | 1 | anthropogenic | 1(1) | 1(1) | 1(1) | 0 | 0 |
| Methylsulfonyl-PCB | 7 | 7 | anthropogenic | 2 (2) | 1 (1) | 1(1) | 1(1) | 3 (3) |
| Mirex-related | 5 | 4 | anthropogenic | 3 (2) | 3 (2) | 2(1) | 1 (0) | 1 (0) |
| PBB | 10 | 9 | anthropogenic | 4 (3) | 4 (3) | 4 (3) | 1 (0) | 2 (2) |
| PBDE | 16 | 6 | anthropogenic | 10(1) | 9 (2) | 9(1) | 8 (0) | 6 (0) |
| PBHD | 3 | 3 | natural | 1(1) | 1(1) | 2 (2) | 0 | 0 |
| PCT | 6 | 6 | anthropogenic | 2(2) | 1(1) | 3 (3) | 0 | 0 |
| Pyrrolidinecarbonyl chloride | 1 | 1 | anthropogenic | 0 | 0 | 0 | 0 | 0 |
| TCPM | 8 | 8 | anthropogenic | 6 (6) | 6 (6) | 6 (6) | 2 (2) | 1(1) |
| ТСРМОН | 2 | 2 | anthropogenic | 1(1) | 1(1) | 1(1) | 0 | 0 |
| Toxaphene | 13 | 5 | anthropogenic | 4 (0) | 7 (0) | 5 (0) | 0 | 0 |
| Unknown | 22 | 22 | unknown | 5 (5) | 5 (5) | 6 (6) | 0 | 0 |
| Unknown-3 | 2 | 2 | unknown | 1(1) | 1(1) | 0 | 0 | 0 |
| Unknown-4 | 12 | 12 | unknown | 6 (6) | 4 (4) | 4 (4) | 1(1) | 0 |
| Unknown-5 | 2 | 2 | unknown | 1(1) | 1 (1) | 2 (2) | 0 | 0 |
| Unknown-6 | 3 | 3 | unknown | 1(1) | 1 (1) | 0 | 0 | 0 |
| Unknown-7 | 2 | 2 | unknown | 1(1) | 1(1) | 1(1) | 0 | 1(1) |
| Unknown-8 | 2 | 2 | unknown | 1 (1) | 1(1) | 1(1) | 0 | 0 |
| Average PCBs | - | - | anthropogenic | 43 | 39 | 35 | 24 | 17 |
| Average Total HOCs (excluding PCBs) | 194 | - | mixed | 90 | 89 | 89 | 29 | 22 |
| (Range) | | | | (77 - 98) | (77 - 102) | (70 - 108) | (14 - 55) | (11 - 32) |

Contaminant Biomagnification

GC×GC-TOF Non-Targeted Blubber Analysis of Multiple Marine Mammal Candidate Sentinels

Pinnipeds Cetaceans

Contaminants

Profile Comparison

Short-Beaked Common Dolphins, photo taken under NOAA Fisheries Permit 1409