

1 **Title:** Mercury concentrations in marine species from the Aleutian Islands: spatial and biological  
2 determinants

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22 **Abstract:** Several species found in the Bering Sea show significant spatial variation in total mercury  
23 concentrations ([THg]) longitudinally along the Aleutian Island chain. We assessed [THg] in other  
24 members of the Bering Sea food web to better understand the factors shaping regional differences. [THg]  
25 and stable carbon and nitrogen isotope ratios ( $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values) were measured in muscle tissue from  
26 1052 fishes and cephalopods from parts of the Bering Sea and North Pacific Ocean adjacent to the  
27 Aleutian Islands. The spatial distribution of the samples enabled regional comparisons for 8 species of  
28 fish and one species of cephalopod. Four species showed higher mean length-standardized [THg] in the  
29 western Aleutian Islands management area. [THg] in yellow Irish lord were very different relative to  
30 those observed in other species and when included in multi-species analyses drove the overall regional  
31 trends in mean [THg]. Multi-species analyses excluding measurements for yellow Irish lord showed mean  
32 length-standardized [THg] was greater in the western Aleutian Islands than in the central Aleutian Islands  
33 management area. Linear regression of [THg] and  $\delta^{15}\text{N}$  values showed a significant and positive  
34 relationship across all species, varying between regions and across species. Isotopic space of all species  
35 was significantly different between the western Aleutian Islands and central Aleutian Islands, driven  
36 largely by  $\delta^{13}\text{C}$  values. Stable isotope values observed follow the same regional trend of lower trophic  
37 taxa reported in the literature, with significantly lower  $\delta^{13}\text{C}$  values in the western Aleutian Islands. We  
38 conclude that there are regional differences in carbon and nitrogen stable isotope ecology, as well as  
39 species-specific feeding ecology that influence [THg] dynamics in part of the marine food web along the  
40 Aleutian Island chain. These regional differences are likely contributors to the observed regional  
41 variations of [THg] in some high-level predators found in these regions.

42 **Keywords:** Muscle, North Pacific; mercury; stable isotopes of C and N; fish

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44 **Highlights**

- 45 • [THg],  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values measured in 1052 fish and cephalopods from Alaska
- 46 • Unadjusted muscle [THg] vary by species, length, and region along Aleutian Islands
- 47 • Mean muscle [THg] of four species higher in the western Aleutian Islands
- 48 • Mean  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values for each species vary by region
- 49 • All species had lower mean  $\delta^{13}\text{C}$  values in the western Aleutian Islands

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54 **1.0 Introduction**

55 Mercury (Hg), a global contaminant that originates from natural and anthropogenic sources, is  
56 transported via long-range atmospheric transport and deposited in a wide range of ecosystems regardless  
57 of proximity to some point sources. Hg becomes more bioavailable after it is methylated and forms  
58 monomethylmercury that increases via biota uptake and biomagnification through the aquatic food web  
59 reaching concentrations of toxicological concern in some higher trophic organisms, such as Steller sea  
60 lions (*Eumetopias jubatus*), Pacific halibut (*Hippoglossus stenolepis*), and humans (Atwell et al., 1998;  
61 Bentzen et al., 2016; Fox et al., 2014; Rea et al., 2013). Monomethylmercury in fish can be a human and  
62 wildlife health issue (Hamade, 2014) as it is a neurotoxin that can cross the blood brain barrier and act as  
63 an endocrine disrupter (Walker et al., 2012) and cause oxidative stress (Lushchak, 2011).

64 Various ecological and physiological processes drive differences in total Hg concentrations  
65 ([THg]) within and between species in the same ecosystem (Jarman et al., 1996; Ward et al., 2010;  
66 Willacker et al., 2013). The specific location (e.g., niche) where organisms feed and inhabit can have a  
67 significant influence on Hg exposure and accumulation. Proximity to Hg sources, or access to different  
68 food webs with different prey assemblages, can provide potential for varying degrees of bioaccumulation  
69 and biomagnification that affect observed tissue concentrations of Hg (Cyr et al., 2017). In marine  
70 ecosystems, Hg sources (Laurier et al., 2004), ocean currents, upwellings (Gill and Fitzgerald, 1987), and  
71 water mass distributions with different geochemical properties (Lehnherr, 2014; Selin, 2009) can  
72 varyingly influence how Hg is transported in open oceans and reflected in upper trophic organisms.  
73 Additionally, biological and chemical oceanographic differences across marine ecosystems can provide  
74 varying conditions for the methylation of Hg, based on factors such as pH (Kelly et al., 2003), the  
75 presence of dissolved cations and organic matter (Boening, 2000; Douglas et al., 2012), and temperature  
76 (Johnson et al., 2016). Together, these factors can provide regionally specific conditions that can  
77 influence how Hg enters and moves through the food chain in a complicated fashion.

78 A notable example of a regional influence on [THg] is in biota of the Bering Sea and the North  
79 Pacific along the Aleutian Islands, from east to west. A pattern of higher [THg] has been documented in  
80 Steller sea lions and Pacific halibut from the western Aleutian Islands, Alaska, when compared to  
81 populations of those species found in other regions of the Aleutian Islands and Alaska (Bentzen et al.,  
82 2016; Rea et al., 2013). A similar geographic pattern was observed in tissues of bald eagles (*Haliaeetus*  
83 *leucocephalus*) and glaucous-winged gulls (*Larus glaucescens*), where [THg] increases from east to west  
84 across the Aleutians Islands (Anthony et al., 2007; Ricca et al., 2008). Significantly, Aleutian Island  
85 aggregations of Steller sea lions belonging to the western distinct population segment and some rookeries  
86 west of Samalga Pass in the western distinct population segment continue to decline in abundance  
87 (Atkinson et al., 2008; National Research Council, 2003; Fritz et al., 2014; NMFS, 2013). This pinniped  
88 management dilemma combined with the observations of [THg] in biota of the region led to this study to  
89 determine if higher [THg] is related to fish consumption.

90 Measurements of stable isotope ratios of carbon and nitrogen ( $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values, respectively;  
91 and together represent isotopic space) provide valuable ecological research tools that add a layer of  
92 dietary information for comparisons of [THg] within and among food webs (Cyr et al., 2017; McGrew et  
93 al., 2014; Power et al., 2002).  $\delta^{15}\text{N}$  values can be used as a proxy to infer trophic position (Peterson and  
94 Fry, 1987), while  $\delta^{13}\text{C}$  values can be used to differentiate the source of the primary production for a food  
95 web (Fry, 2006; Peterson and Fry, 1987).  $\delta^{13}\text{C}$  values can therefore be used to infer regional differences  
96 by differentiating the influence from different carbon sources (Budge et al., 2008; Wang et al., 2014),  
97 such as the degree of influence from marine versus terrestrial inputs (Fry, 2006; McGrew et al., 2014) or  
98 pelagic versus benthic sources (Boyle et al., 2012; Doi et al., 2010). Fish [THg] are often, but not always,  
99 correlated with increasing  $\delta^{15}\text{N}$  values, and can vary with spatial influences, as elucidated by using  $\delta^{13}\text{C}$   
100 values (Cyr et al., 2017). Together, stable isotopes of carbon and nitrogen can provide information on  
101 some of the ecological influences on [THg] within and among species and locations of fish.

102 Fish and invertebrates sampled from across the Aleutian Islands provide an ideal opportunity to  
103 examine regional influences on [THg] in a food web relative to wildlife management zones and  
104 oceanographic demarcations. Here we report [THg],  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values, and associated biological data  
105 for 1052 specimens of marine fishes and cephalopods, representing 19 species, from an area of the Bering  
106 Sea and North Pacific Ocean adjacent to the Aleutian Islands. The primary goal of this research was to  
107 analyze patterns and comparisons of muscle [THg] in a diverse group of fish and cephalopod species from  
108 the waters along the central and western portion of the Aleutian Islands to assist with understanding  
109 observations of [THg] in upper trophic level vertebrates (fish, avian, mammalian). In particular, we  
110 examined how geographic location (Steller sea lion management zones and Amchitka Pass, a distinct  
111 oceanographic demarcation) shapes the patterns of length (proxy for age), ecology (including  $\delta^{13}\text{C}$  and  
112  $\delta^{15}\text{N}$  values), and muscle [THg] of species examined.

## 113 **2.0 Methods**

114 *2.1 Sampling.* Specimens were obtained from commercial fish trawls (target and bycatch species)  
115 in 2013, 2014, and 2015; and donated subsamples from NOAA research cruises SF201401 and  
116 MS201401 in 2014 and 2015. Sampling spanned summer and winter seasons. Sampling regions along the  
117 Aleutian Islands were based on Steller sea lion management zones (western Aleutian Islands and central  
118 Aleutian Islands), located in the North Pacific Ocean and in the southern and western portion of the  
119 Bering Sea, between  $51.26^\circ\text{N}$  and  $53.22^\circ\text{N}$ , and  $171.89^\circ\text{W}$  and  $173.76^\circ\text{E}$  (Figure 1, York et al., 1996).  
120 Samples were also given a second regional assignment based on the longitudinal catch location in relation  
121 to Amchitka Pass,  $179.98^\circ\text{W}$  (Figure 1) that is considered a significant ecological boundary in the  
122 Bering Sea. Fish were collected from commercial trawls, frozen whole at  $-20^\circ\text{C}$  and shipped to the  
123 University of Alaska Fairbanks for analyses. Muscle samples provided from federal research cruises were  
124 subsampled from fish measured in the field, frozen at sea and shipped to University of Alaska Fairbanks  
125 for analyses.

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127           2.2 *Sample processing*. For specimens collected from commercial catches, fork length (in mm),  
128 and wet mass (in g) were recorded on thawed fish. Mantle length was measured for cephalopods.  
129 Approximately five grams of muscle (skin removed) was sampled from the left side of the fish from the  
130 area posterior to the pectoral fin, and ventral to the dorsal fin, or from the mantle for cephalopods.  
131 Specimens collected from federal research cruises were sampled on board the vessel, and in the lab a 5  
132 gram subsample was taken from the larger subsample. All samples were freeze dried (Labcono, FreeZone  
133 4.5 Liter) for a minimum of 48 h to remove all moisture and homogenized using a stainless-steel ball  
134 grinder (Retsch, CryoMill). Percent water content was calculated after drying using the following  
135 formula:

$$\left(\frac{\text{wet weight} - \text{dry weight}}{\text{wet weight}}\right) * 100$$

136  
137           2.3 *Total Hg analysis*. [THg] were measured in freeze dried samples using a Milestone DMA-80  
138 instrument, in accordance with U.S. EPA method #7473-EPA30B, 2007 SW 846, and reported as ng/g  
139 wet weight (ww) based on percent water values (Harley et al., 2015). The method detection limit for  
140 [THg] determination for muscle was 5 ng/g ww, calculated by the following formula:

$$\left(\frac{0.5 (g)}{\text{Sample mass dry weight (g)}}\right) * (1 - \text{sample \% H}_2\text{O})$$

141           Quality assurance and quality control measures included analyses of method blanks, Standard  
142 Reference Materials of similar matrices, and check standards. All samples were analyzed in triplicate.  
143 Samples for which the coefficient of variation among replicates was >10% were re-analyzed until <10%  
144 was reached, with the mean value for all replicates used for statistical comparisons. The standard  
145 reference materials used were DORM-4 (National Resource Council Canada; 410.0 ± 55.0 ng/g), and  
146 Lake Superior Fish (LSF, National Institute of Standards and Technology, Standard Reference Material®  
147 1946; 433.0 ± 9.0 ng/g ww). Mean percent recoveries (± SD) for each SRM were: 100 ng/g (liquid  
148 standard), 96.7 ± 10.9%; DORM-4, 93.5 ± 7.9%; LSF, 100.3 ± 10.1%.

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150           2.4 *Stable carbon and nitrogen isotope analysis.* Stable carbon and nitrogen isotope ratios of all  
151 samples were measured at the Alaska Stable Isotope Facility at the University of Alaska Fairbanks  
152 following the methods described in Cyr et al. (2017). Between 0.2 and 0.5 mg of freeze-dried  
153 homogenized muscle samples were analyzed using continuous-flow isotope ratio mass spectrometry (CF-  
154 IRMS, Thermo DeltaVPlus interfaced with a Costech ESC 4010 elemental analyzer via a ConFloIV  
155 system). Stable isotope ratios were expressed in  $\delta$  notation as parts per thousand (‰) relative to  
156 international standards (Vienna PeeDee Belemnite – VPDB for carbon and AIR for nitrogen):

$$\delta X\text{‰} = \left[ \left( \frac{R_{\text{Sample}}}{R_{\text{Standard}}} \right) - 1 \right] * 1000$$

157 where  $X$  is the element of interest,  $R$  is the ratio of the heavy to light isotope measured for that element,  
158 *Sample* is the sample of interest, and *Standard* is the standard used, VPDB for carbon or AIR for nitrogen.  
159 Reference checks using peptone (No. P-7750 meat-based protein, Sigma Chemical Company, Lot #76f-  
160 0300) were run every 10<sup>th</sup> sample, and blanks every 20<sup>th</sup> sample, with instrument precision typically <0.2  
161 ‰ for both carbon and nitrogen.

162

163           2.5 *Lipid extraction and correction.* Lipids are depleted in <sup>13</sup>C relative to <sup>12</sup>C compared with  
164 protein and carbohydrates, and can affect stable carbon isotope data by lowering the  $\delta^{13}\text{C}$  values with  
165 increasing lipid content of a sample (DeNiro and Epstein, 1977; Sweeting et al., 2006). High C:N values  
166 present difficulties with mathematical lipid-correction formulas, and can cause significant variability  
167 between the relationship of  $\Delta\delta^{13}\text{C}$  in relation to the C:N<sub>Bulk</sub> (Hoffman and Sutton, 2010; Post et al., 2007).  
168 A large proportion (92%) of the samples in this study had atomic C:N ratios above levels considered  
169 representative for protein, ~3.7 (Post et al., 2007; Sweeting et al., 2006), and differed between species,  
170 individuals and regions (Supplemental Figure 1). To account for this influence, we lipid-extracted a  
171 subset of two hundred and forty-five samples from eight of the species following a method modified from  
172 Folch et al. (1957). Briefly, 1.0 - 2.0 g of freeze-dried sample muscle was rinsed three times in a mixture



173 of 2:1 chloroform: methanol for 15 minutes, air dried overnight, then freeze-dried for 24 hours. These  
174 lipid-extracted samples were analyzed for  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values as described above and recalculated C:N  
175 ratios.

176 We determined that the influence of lipids in samples with  $\text{C:N}_{\text{Bulk}} > 10$  was considerable and  
177 introduced more variability and uncertainty than mathematical lipid-correction could reasonably account  
178 for (Supplemental Figures 1, 2 and 3), and subsequently removed ten samples with  $\text{C:N}_{\text{Bulk}} > 10$  from  
179  $\delta^{13}\text{C}$  analyses. The mean C:N ratio for all lipid-extracted samples was  $3.7 \pm 0.1$  ‰ (Supplemental Table  
180 1), and thus enabled the use of the  $\delta^{13}\text{C}_{\text{Lipid-extracted}}$  values to generate a mathematical correction formula.  
181 We followed the methods detailed in Post et al. (2007) by using their formula for % lipid:

$$\% \text{ lipid} = -20.54 + 7.24 * \text{C:N}_{\text{Bulk}}$$

182 followed by the equation to determine  $\Delta\delta^{13}\text{C}$  with our data:

$$\Delta\delta^{13}\text{C} = 0.45 + (0.094 * \% \text{ lipid})$$

183 and then combined the equations to generate the final correction formula (Supplemental Table 1,  
184 Supplemental Figure 3,  $R^2 = 0.80$ ,  $p \ll 0.001$ ) to mathematically correct the  $\delta^{13}\text{C}_{\text{Bulk}}$  values for the  
185 remainder of the dataset:

$$\delta^{13}\text{C}_{\text{Lipid-corrected}} = \delta^{13}\text{C}_{\text{Bulk}} - 1.48 + 0.65 * \text{C:N}_{\text{Bulk}}$$

187  
188 *2.6 Length standardization of [THg].* To account for regional or allometric effects on [THg] and  
189 allow for inter- and intra-species and regional comparisons, a length-standardization of [THg] was  
190 performed following the methods detailed in Eagles-Smith et al. (2016). Briefly, we standardized the log-  
191 transformed [THg] using the median length for each species by generating a linear mixed effects model  
192 with length and species as fixed covariates, and region as a random effect. We added the residuals from  
193 each fish back into the model to generate predictive [THg] values for each fish that were then back  
194 transformed to obtain the final length-standardized [THg] for each fish.

195

196           2.7 *Statistical analysis.* All analyses were performed using R statistical computing software (R  
197 Core Team 2015). All data were checked for normality by visual inspection of normal quantile-quantile  
198 plots, and residual plots (Ciancio et al., 2008), and using either the Shapiro-Wilk's test or the Kruskal-  
199 Wallis test. [THg] were log transformed for regression analysis to comply with normality assumptions.  
200 To ensure statistical power for comparisons, we restricted our regional and inter-species comparisons to  
201 the 9 species that had >10 individuals per region. Species in these comparisons were divided into two  
202 major groups: 1) darkfin sculpin (*Malacocottus zonurus*), Pacific cod (*Gadus macrocephalus*), yellow  
203 Irish lord (*Hemilepidotus jordani*), and arrowtooth flounder (*Atheresthes stomias*) were considered  
204 piscivorous based on the percentage of diet composed of fish (Yang and Nelson, 1999; Yang, 2003); and  
205 2) walleye pollock (*Gadus chalcogrammus*), Atka mackerel (*Pleurogrammus monopterygius*), northern  
206 rockfish (*Sebastes polyspinis*), Pacific Ocean perch (*Sebastes alutus*), and one invertebrate, the magistrate  
207 armhook squid (*Berryteuthis magister*) were considered primarily zooplanktivorous based on the  
208 percentage of diet composed of invertebrates (Yang and Nelson, 1999; Yang, 2003)

209           Summary statistics for [THg] are represented as geometric mean  $\pm$  standard deviation (SD), all  
210 other summary statistics are represented as arithmetic mean  $\pm$  SD. We conducted analysis of variance to  
211 determine the overall influence of species, region, and the interaction of species and region. Following  
212 this, regional differences for individual species' mass, length, unadjusted [THg], and length-standardized  
213 [THg] were using t-tests with a Holm correction to control for familywise errors. Results were considered  
214 significant at  $\alpha \leq 0.05$ . Rank order position for each species and region was determined using analysis of  
215 variance followed by the Tukey's Honestly Significant Difference *post hoc* test. Determining the  
216 influence of  $\delta^{15}\text{N}$  values,  $\delta^{13}\text{C}$  values, the interaction of  $\delta^{15}\text{N}$  or  $\delta^{13}\text{C}$  values and region on [THg] was  
217 determined using generalized linear models (GLM) for each species. Differences in the isotopic space  
218 between seasons and regions for each species were tested using the Hotelling's  $T^2$  test, comparing the  
219 mean  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values in multivariate space (Ciancio et al., 2008; Colombini et al., 2011).

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221

## 222 3.0 Results

223 *3.1 Data summary.* A total of 1052 samples from the western Aleutian Islands and the central  
224 Aleutian Islands, representing 19 species of marine fishes and cephalopods were analyzed for [THg], and  
225  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values. Across all species, mass and length varied considerably, ranging from 15 to 16800  
226 g ( $1,303.1 \text{ g} \pm 1,963.4$ ), and from 8 to 111 cm ( $41.3 \text{ cm} \pm 16.9$ ), respectively. Unadjusted [THg] for all  
227 fish and cephalopods ranged from 7.5 to 1,578.3 ng/g ww ( $149.47 \pm 173.74$ ).  $\delta^{15}\text{N}$  (‰) values ranged  
228 from 5.9 to 15.0 ‰ ( $11.1 \pm 1.5$  ‰), bulk  $\delta^{13}\text{C}$  values ranged from -26.5 to -17.6 ‰ ( $-20.9 \pm 1.5$  ‰), and  
229 lipid-corrected  $\delta^{13}\text{C}$  values ranged from -22.7 to -16.5 ‰ ( $-19.5 \pm 1.2$  ‰). Statistical comparisons  
230 between central Aleutian Islands and western Aleutian Islands for the nine species with >10 individuals  
231 per region are shown in Table 1. Summary statistics for all other species sampled and analyzed are  
232 provided in Supplemental Table 2.

233  
234 *3.2 Stable isotopes.* The seasonal isotopic space for each species and region was significantly  
235 different within the western Aleutian Islands in armhook squid, Atka mackerel, northern rockfish, Pacific  
236 cod, and walleye pollock, and within the central Aleutian Islands in all species except armhook squid and  
237 darkfin sculpin (Supplemental Table 3). These differences were small ( $< \sim 1$  ‰) and within analytical error  
238 and aquatic trophic level variation (Vander Zanden and Rasmussen, 2001), so we considered them not  
239 biologically significant. As a result, we pooled the stable isotope measurements within each species and  
240 region for our study.

241 All species occupied significantly different isotopic spaces between the central Aleutian Islands  
242 and western Aleutian Islands (Table 2, Figure 2). This is likely driven by the differences in mean lipid-  
243 corrected  $\delta^{13}\text{C}$  values between regions ( $\Delta$ central Aleutian Islands-western Aleutian Islands) that was  
244 different for all species (Table 1), whereas only 4 species had statistically different (Table 1,  $\alpha \leq 0.05$ )  
245 mean  $\delta^{15}\text{N}$  values between the western Aleutian Islands and central Aleutian Islands.

246

247           3.3 *Regional comparisons.* An analysis of variance model indicated that species and the  
248 interaction of species and region significantly contributed to the variability in length standardized [THg],  
249 while region alone was not significant ( $\alpha \leq 0.05$ ). Unadjusted [THg] were statistically higher in the  
250 western Aleutian Islands for arrowtooth flounder, Pacific cod, Pacific Ocean perch, and walleye pollock  
251 (4 of 9 species), whereas unadjusted [THg] were statistically higher in the central Aleutian Islands for  
252 yellow Irish lord and Atka mackerel ( $\alpha \leq 0.05$ ; Table 1). There was no difference in length-standardized  
253 [THg] between western Aleutian Islands and central Aleutian Islands across all fish combined ( $\alpha \leq 0.05$ ).  
254 Since this statistic is likely driven by the magnitude of [THg] in yellow Irish lord from the central  
255 Aleutian Islands (Figure 3), a further comparison for all species combined except yellow Irish showed the  
256 mean length-standardized [THg] was significantly higher in the western Aleutian Islands ( $\alpha \leq 0.001$ ).  
257 Intraspecies comparisons of mean length-standardized [THg] were higher in the western Aleutian Islands  
258 for arrowtooth flounder, Pacific cod, Pacific Ocean perch, and walleye pollock (four of nine species,  
259 Table 1, Figure 3). Only Atka mackerel and yellow Irish lord had higher mean length-standardized [THg]  
260 in the central Aleutian Islands (Table 1, Figure 3).

261           Regional comparisons based on secondary regional assignments using Amchitka Pass as the  
262 divide between western Aleutian Islands and central Aleutian Islands revealed that both mean unadjusted  
263 [THg] and length-standardized [THg] of all species combined were significantly greater in the western  
264 Aleutian Islands ( $\alpha \leq 0.001$ ). Intraspecies comparisons demonstrated that all species except Atka  
265 mackerel had greater mean length-standardized [THg] in the western Aleutian Islands, and statistically,  
266 the mean length-standardized [THg] of arrowtooth flounder, northern rockfish, Pacific cod, Pacific Ocean  
267 perch, walleye pollock, and yellow Irish lord were significantly greater in the western Aleutian Islands ( $\alpha$   
268  $\leq 0.05$ ). Isotopic space was different between the western Aleutian Islands and central Aleutian Islands  
269 for all species ( $\alpha \leq 0.01$ ), with consistently lower  $\delta^{13}\text{C}$  values in the western Aleutian Islands.

270           The overall descending rank order among species for mean length-standardized [THg] was  
271 yellow Irish lord > darkfin sculpin  $\geq$  Pacific cod > arrowtooth flounder  $\geq$  northern rockfish  $\geq$  Pacific

272 Ocean perch  $\geq$  walleye pollock  $\geq$  Atka mackerel  $\geq$  armhook squid ( $>$  indicates statistical difference,  $\alpha <$   
273 0.05, and  $\geq$  indicates a greater mean value but no statistical difference). Regionally, in the western  
274 Aleutian Islands, the descending rank order for length-standardized [THg] was darkfin sculpin  $\geq$  Pacific  
275 cod  $\geq$  arrowtooth flounder  $\geq$  yellow Irish lord  $\geq$  Pacific Ocean perch  $\geq$  walleye pollock  $\geq$  northern  
276 rockfish  $\geq$  Atka mackerel  $\geq$  armhook squid (Figure 3). In the central Aleutian Islands, the descending rank  
277 order for length-standardized [THg] was yellow Irish lord  $>$  darkfin sculpin  $\geq$  Pacific cod  $>$  northern  
278 rockfish  $\geq$  arrowtooth flounder  $\geq$  Pacific Ocean perch  $\geq$  Atka mackerel  $\geq$  walleye pollock  $\geq$  armhook  
279 squid (Figure 3).

280

281 *3.4 [THg] in relation to trophic position.* Unadjusted [THg] increased with increasing  $\delta^{15}\text{N}$  values  
282 ( $R^2 = 0.31$ ,  $p < 0.001$ ) across all species examined. Assessing this relationship within each species and  
283 region indicated 6 of the species had a significant, positive slope in the western Aleutian Islands; while 4  
284 of the species had a significant, positive slope in the central Aleutian Islands (Table 3). Using GLM the  
285 unadjusted [THg] in Pacific cod, Pacific Ocean perch, darkfin sculpin, northern rockfish, and yellow Irish  
286 lord were significantly influenced by  $\delta^{15}\text{N}$  values; and by  $\delta^{13}\text{C}$  values in arrowtooth flounder, Pacific cod,  
287 Pacific Ocean perch, walleye pollock, darkfin sculpin, and yellow Irish lord (Table 4). Unadjusted [THg]  
288 was significantly influenced by the interaction of  $\delta^{15}\text{N}$  values and region in walleye pollock, while the  
289 interaction of  $\delta^{13}\text{C}$  values and region significantly influenced [THg] in arrowtooth flounder, Pacific cod,  
290 walleye pollock, northern rockfish, and yellow Irish lord (Table 4).

291

## 292 **4.0 Discussion**

293 *4.1 Overview.* We sought to gain a better understanding of the influence of region and feeding  
294 ecology on observed [THg] in fish and cephalopod species by investigating a portion of the food web of  
295 the Bering Sea and North Pacific Ocean along the Aleutian Islands. We found a pattern of higher muscle  
296 [THg] in many of the fish from the western Aleutian Islands compared to the central Aleutian Islands. We  
297 also found a consistent pattern of differences in the isotopic space ( $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values) between the

298 western Aleutian Island and central Aleutian Island regions for all species (Table 4.2, Figure 4.2). Finally,  
299 we demonstrated that fish muscle [THg] is highly variable and influenced by known drivers of Hg tissue  
300 concentrations.

301

302 *4.2 Isotopes and feeding ecology.* The seasonal ranges of stable isotope values measured in our  
303 fish suggest negligible biological differences and allowed us to pool the isotope data by year. The  
304 decision to do this was further supported by related feeding ecology work from the Aleutian Islands that  
305 demonstrated that the seasonal variation in stable isotopes in Steller sea lion vibrissa was 2 - 5 ‰, and 3 -  
306 7 ‰ respectively for  $\delta^{13}\text{C}$  values and  $\delta^{15}\text{N}$  (Rea et al., 2015), far greater than the isotope variations  
307 measured in our fish. Doll et al. (2018) also noted the magnitude of the seasonal variation of Steller sea  
308 lion vibrissa, together with the minimal seasonal variation of their prey isotope values, and subsequently  
309 pooled their prey isotope values together seasonally.

310 The regional patterns of isotopic space were mostly driven by the differences in  $\delta^{13}\text{C}$  values  
311 rather than  $\delta^{15}\text{N}$  values. In both bulk  $\delta^{13}\text{C}$  data and lipid-corrected  $\delta^{13}\text{C}$  data, mean  $\delta^{13}\text{C}$  values in the  
312 western Aleutian Islands were lower than those from the central Aleutian Islands for each species (Table  
313 4.1, Figure 4.2). These measured differences in  $\delta^{13}\text{C}$  values are consistent with the literature, where Schell  
314 et al. (1998) determined that the  $\delta^{13}\text{C}$  values of both euphausiids and chaetognaths were lower in the  
315 western portion of the Aleutians. We suspect this is indicative of a difference in baseline stable isotope  
316 values, notably  $\delta^{13}\text{C}$  values, between the western Aleutian Islands and the central Aleutian Islands. If  
317 baseline differences occur at the level of primary production, assuming other isotope factors were roughly  
318 equal in the food web, that difference would be transferred up through the food web, resulting in  
319 differences in upper trophic level prey.

320 Across all fishes combined, muscle [THg] increased positively and significantly with increasing  
321  $\delta^{15}\text{N}$  values, indicating biomagnification through the food web, an expected finding well supported in the  
322 literature (Atwell et al., 1998; Coelho et al., 2013; Power et al., 2002). The parameters of this correlation  
323 however, varied strongly by species and regions. Most notably,  $\delta^{15}\text{N}$  values were generally lower in the

324 western Aleutian Islands than in the central Aleutian Islands, but muscle [THg] were generally higher in  
325 the western Aleutian Islands than in the central Aleutian Islands, a pattern opposite of expected  
326 assumptions based on known biomagnification dynamics of Hg. This pattern is also consistent with  
327 previous findings, in which Pacific halibut from the western Aleutian Islands had the lowest  $\delta^{15}\text{N}$  values,  
328 yet the highest muscle [THg] (Bentzen et al., 2016). The  $\delta^{15}\text{N}$  values of Pacific cod, walleye pollock,  
329 arrowtooth flounder, and yellow Irish lord were all lower than those reported by Gorbatenko et al. (2008)  
330 from Cape Olutorski and Cape Navarin in the western Bering Sea. These patterns suggest regional  
331 differences in [THg] along the Aleutian Islands likely originate at the base of the food chain and are  
332 amplified through feeding ecology, but not driven strictly by feeding ecology.

333 Consistent trends in differences of the biology and chemistry of the biota from central or eastern  
334 and western areas of the Aleutian Islands reinforce the idea that oceanographic and ecological conditions  
335 are likely driving differences in [THg] in biota between the western Aleutian Islands and central Aleutian  
336 Islands, with these differences amplified through the food web. Feeding ecology is likely a major driver  
337 of the observed differences in [THg] of higher trophic level prey, because modeling research has  
338 demonstrated that regional deposition of [THg] across the Bering Sea and Aleutian Islands does not differ  
339 regionally (Strode et al., 2008), and Hg contributions from various sources to the Bering Sea are similar  
340 (Sunderland et al., 2009). We highlight feeding ecology, because fish have limited capacity for  
341 redistributing, demethylating or eliminating THg or MeHg<sup>+</sup> that allows for efficient accumulation and  
342 retention of THg or MeHg<sup>+</sup> in muscle tissue (Amlund et al., 2007; Trudel and Rasmussen, 1997). These  
343 factors, in conjunction with the efficient assimilation of Hg in the fish gut allow biomagnification of Hg  
344 through the aquatic food web (Atwell et al., 1998; Coelho et al., 2013). Thus, higher trophic level fish  
345 tend to have higher [THg] than lower trophic fish and prey items. Our study supports this generalization  
346 as three of the species with the largest differences between western Aleutian Islands and central Aleutian  
347 Islands, arrowtooth flounder, Pacific cod, and yellow Irish lord are known to be higher trophic predators  
348 that feed primarily on fish or crustaceans (Yang, 2003). Darkfin sculpin had some of the highest mean  
349 muscle [THg], and their diet includes at least some shrimp. These four species also had the highest  $\delta^{15}\text{N}$

350 values (Table 4.1, Figure 4.2), and occupy trophic levels ranging from 3.8 to 4.5 (Aydin et al., 2007;  
351 Gorbatenko et al., 2008; Marsh et al., 2012). All other species in this study feed almost exclusively on  
352 some type of zooplankton (Yang, 2003), and occupy trophic levels ranging from 3.0 to 3.8, a full trophic  
353 level lower on the food web, providing less potential for biomagnification.

354

355 *4.3 Geographic trends.* Our data demonstrate a general trend of higher muscle [THg] in western  
356 Aleutian Island fish compared to central Aleutian Island fish. Four of the nine species had statistically  
357 greater mean length-standardized [THg] in the western Aleutian Islands than in the central Aleutian  
358 Islands. This geographical trend in [THg] is consistent with the literature regarding other marine-based  
359 taxa, such as Steller sea lion (Rea et al., 2013), Pacific halibut (Bentzen et al., 2016), glaucous-winged  
360 gulls (Ricca et al., 2008), and bald eagles (Anthony et al., 2007). The strong influence of yellow Irish lord  
361 on the overall regional patterns of [THg] are unique and discussed in subsequent paragraphs.

362 Amchitka Pass on the western end of the central Aleutian Islands (Figure 4.1) is considered a  
363 discrete ecological divide in the Bering Sea, with chemical and biological oceanographic processes  
364 differing from the west to the east of the pass (Logerwell et al., 2005). The consistent differences in  
365 [THg] we found by using Amchitka Pass as the boundary for western Aleutian Islands and central  
366 Aleutian Islands are likely related to the consistent differences in isotopic spaces, where specific chemical  
367 oceanographic differences east and west of Amchitka Pass have caused differences in isotope baseline  
368 values, and then feeding ecology influences the accumulation of THg in each food web.

369

370 *4.4 Yellow Irish lord.* Although yellow Irish lord were similar in size between the western  
371 Aleutian Islands and central Aleutian Islands regions, the mean length-standardized [THg] in the central  
372 Aleutian Islands was over two times higher than the western Aleutian Islands, a regional trend that is  
373 opposite to the one detected for the other species (Table 4.1, Figure 4.3). This may be related to trophic  
374 level, as yellow Irish lord in the central Aleutian Islands had higher  $\delta^{15}\text{N}$  values than those from the  
375 western Aleutian Islands. Yellow Irish lord feed on fish and benthic crustaceans (Yang, 2003) that are



376 higher trophic level prey than the zooplankton prey that many of the other fish species in this study  
377 consume. Yellow Irish lord could also simply be older fish. Long-lived fishes will eventually reach the  
378 asymptote of their length-at-age relationship, when they will continue to age and continue to accumulate  
379 Hg, but not exhibit a related increase in length (Eagles-Smith et al., 2014). The mean length-standardized  
380 [THg] of yellow Irish lord in the western Aleutian Islands was comparable to the mean length-  
381 standardized [THg] of several other species in this study, while the length-standardized [THg] of those  
382 from the central Aleutian Islands were much greater than any other species measured in this study. Based  
383 on the longer mean fork length for yellow Irish lord in the central Aleutian Islands, we suspect that these  
384 fish may be at or approaching the asymptote of their growth curve, approximately 46 cm for males and  
385 females combined (TenBrink and Buckley, 2013), indicating they are older individuals, but this age is not  
386 reflected in their length, and has allowed a greater amount of time for the accumulation of Hg in their  
387 muscle tissue (Eagles-Smith et al., 2014; Lange et al., 1994). Additionally, the maximum age of yellow  
388 Irish lord in the western Aleutian Islands has been reported to be less than more easterly portions of the  
389 Bering Sea (TenBrink and Aydin, 2009), further indicating the yellow Irish lord caught in the central  
390 Aleutian Islands may have been older individuals. Using only the length of a fish to understand Hg  
391 feeding ecology and accumulation dynamics would miss this level of important detail, consequently  
392 causing inappropriate comparisons of fish that are years apart in age. Age data greatly improves the  
393 ability to understand situations like this, and we recommend age estimations be conducted for fish that  
394 exhibit extreme relationships of [THg] and other metrics such as species, fish length,  $\delta^{15}\text{N}$  values, or  
395 location.

396

397 *4.5 Food web dietary exposure.* The muscle [THg] of many of the fish from the central Aleutian  
398 Islands are in general agreement with other published studies of Pacific cod (Burger et al., 2014, 2007;  
399 Burger and Gochfeld, 2007), yellow Irish lord (Burger et al., 2014, 2007), and Atka mackerel (Alaska,  
400 2017; Burger et al., 2007). The muscle [THg] for several species of fish from the western Aleutian Islands  
401 however, were higher than those reported in the literature, where the mean [THg] measured in Pacific cod

402 were nearly 150 ng/g ww greater than those measured by Burger et al. (2014), or the State of Alaska Hg  
403 biomonitoring study from 2001 to 2016 (Alaska, 2017). The [THg] of arrowtooth flounder from the  
404 central Aleutian Islands were comparable to those identified in Oregon by Childs and Gaffke (1973),  
405 while those from the central Aleutian Islands were higher than those measured in arrowtooth flounder  
406 from the Bering Sea by Gerber et al. (2012), and those from western Aleutian Islands were higher than  
407 values reported in any literature we identified. Our [THg] for walleye pollock and yellow Irish lord were  
408 also higher than those reported by the State of Alaska (Alaska, 2015), and were similar to reports of  
409 halibut across Alaska (Bentzen et al., 2016). Although these comparisons demonstrate the high degree of  
410 variability of fish [THg], given the consistency of the patterns, they also reinforce the overarching  
411 regional pattern of increased [THg] in the western Aleutian Islands.

412         These results point to influential regional food web differences resulting in mean muscle [THg]  
413 differences along the Aleutian chain for most species we examined. Where mean [THg] are different  
414 between the western Aleutian Islands and the central Aleutian Islands, the differences are substantial,  
415 such as 92 ng/g ww difference for Pacific cod (57 % increase), or 140 ng/g ww difference for arrowtooth  
416 flounder (180 % increase). It is also important to note that regardless of region, yellow Irish lord  
417 contained elevated [THg] compared to the other species measured. These differences may translate to  
418 different Hg exposure potential between the western Aleutian Islands and central Aleutian Islands regions  
419 for Steller sea lion and other top predators occupying those areas. These differences prove to be more  
420 directly associated with Amchitka Pass, as a biogeographical divide, when compared to Steller sea lion  
421 resource management zones.

422

## 423 **5.0 Conclusions**

424         We compiled an extensive set of [THg] and stable isotope data for 19 species found in the Bering  
425 Sea and North Pacific along the Aleutian Islands. These data enabled us to generate and validate a  
426 mathematical lipid-correction formula for Bering Sea and North Pacific fishes with  $C:N_{\text{Bulk}} < 10$ . We  
427 showed a general trend of elevated [THg] in fish from the western Aleutian Islands, compared to the

428 central Aleutian Islands, when using the management regions defined for Steller sea lion. When  
429 considering a regional delineation based on ecosystem characteristics, we note that Amchitka Pass offers  
430 a clearer and more consistent explanation for the observed trends in  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values and [THg] than  
431 those based on marine mammal species management zones. We document a consistent trend of lower  
432  $\delta^{13}\text{C}$  values in the western Aleutian Islands, providing a starting point to assess differences in primary  
433 production and feeding ecology, and subsequently the increase of [THg] through multiple trophic levels.  
434 These findings, in conjunction with previous research indicates potential regional differences in feeding  
435 ecology, resulting in both inter- and intraspecies biomagnification differences that drive higher [THg] in  
436 the western Aleutian Islands. Further research on this topic should include more comprehensive food web  
437 analysis of [THg] and stable isotopes (C and N), from primary production to top predators of the same  
438 location. Additionally, these measurements and analyses will be useful to others interested in Hg and  
439 other contaminants in this region and for determining consumption advice for humans. As such, these  
440 data have been shared with the State of Alaska Department of Environmental Conservation for this  
441 purpose.

442

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462 **References**

- 463 Alaska, S. of, 2017. Total Mercury in Alaska's Fish. Anchorage, Alaska.
- 464 Alaska, S. of, 2015. 2001-2012 Total mercury in Alaska Fish [WWW Document]. URL  
465 <http://dhss.alaska.gov/dph/Epi/eph/Pages/fish/default.aspx> (accessed 2.15.18).
- 466 Amlund, H., Lundebye, A.K., Berntssen, M.H.G., 2007. Accumulation and elimination of methylmercury  
467 in Atlantic cod (*Gadus morhua* L.) following dietary exposure. *Aquat. Toxicol.* 83, 323–330.  
468 doi:10.1016/j.aquatox.2007.05.008
- 469 Anthony, R.G., Miles, a K., Ricca, M. a, Estes, J. a, 2007. Environmental contaminants in bald eagle  
470 eggs from the Aleutian archipelago. *Environ. Toxicol. Chem.* 26, 1843–1855. doi:10.1897/06-  
471 334R.1
- 472 Atkinson, S., DeMaster, D.P., Calkins, D.G., 2008. Anthropogenic causes of the western Steller sea lion  
473 *Eumetopias jubatus* population decline and their threat to recovery. *Mamm. Rev.* 38.
- 474 Atwell, L., Hobson, K.A., Welch, H.E., 1998. Biomagnification and bioaccumulation of mercury in an  
475 arctic marine food web: Insights from stable nitrogen isotope analysis. *Can. J. Fish. Aquat. Sci.* 55,  
476 1114–1121. doi:10.1139/cjfas-55-5-1114
- 477 Aydin, K., Gaichas, S., Ortiz, I., Kinzey, D., Friday, N., 2007. A comparison of the Bering Sea, Gulf of  
478 Alaska, and Aleutian Islands Large Marine Ecosystems through food web modeling. NOAA Tech.  
479 Memo. NMFS-AFSC-, 298.
- 480 Bentzen, R., Castellini, J.M., Gerlach, R., Dykstra, C., O'Hara, T., 2016. Mercury concentrations in  
481 Alaska Pacific halibut muscle relative to stable isotopes of C and N and other biological variables.  
482 *Mar. Pollut. Bull.* 113, 110–116. doi:10.1016/j.marpolbul.2016.08.068
- 483 Boening, D.W., 2000. Ecological effects, transport, and fate of mercury: A general review. *Chemosphere*  
484 40, 1335–1351. doi:10.1016/S0045-6535(99)00283-0
- 485 Boyle, M.D., Ebert, D.A., Cailliet, G.M., 2012. Stable-isotope analysis of a deep-sea benthic-fish  
486 assemblage: evidence of an enriched benthic food web. *J. Fish Biol.* 80, 1485–1507.  
487 doi:10.1111/j.1095-8649.2012.03243.x

488 Budge, A.M., Wooller, M.J., Springer, A.M., Iverson, S.J., McRoy, C.P., Divoky, G.J., 2008. Tracing  
489 carbon flow in an arctic marine food web using fatty acid-stable isotope analysis. *Oecologia* 157,  
490 117–129. doi:10.1007/s00442-008-1053-7

491 Burger, J., Gochfeld, M., 2007. Risk to consumers from mercury in Pacific cod (*Gadus macrocephalus*)  
492 from the Aleutians: Fish age and size effects. *Environ. Res.* 105, 276–284.  
493 doi:10.1016/j.envres.2007.05.004

494 Burger, J., Gochfeld, M., Jeitner, C., Burke, S., Stamm, T., Snigaroff, R., Snigaroff, D., Patrick, R.,  
495 Weston, J., 2007. Mercury levels and potential risk from subsistence foods from the Aleutians. *Sci.*  
496 *Total Environ.* 384, 93–105. doi:10.1016/j.scitotenv.2007.05.004

497 Burger, J., Gochfeld, M., Jeitner, C., Pittfield, T., Donio, M., 2014. Heavy metals in fish from the  
498 Aleutians: Interspecific and locational differences. *Environ. Res.* 131, 119–130.  
499 doi:10.1016/j.envres.2014.02.016

500 Childs, E. a., Gaffke, J.N., 1973. Mercury content of Oregon groundfish. *Fish. Bull.* 71, 713–717.

501 Ciancio, J.E., Pascual, M.A., Botto, F., Frere, E., Iribarne, O., 2008. Trophic relationships of exotic  
502 anadromous salmonids in the southern Patagonian Shelf as inferred from stable isotopes. *Limnol.*  
503 *Oceanogr.* 53, 788–798. doi:10.4319/lo.2008.53.2.0788

504 Coelho, J.P., Mieirol, C.L., Pereira, E., Duarte, A.C., Pardal, M.A., 2013. Mercury biomagnification in a  
505 contaminated estuary food web: Effects of age and trophic position using stable isotope analyses.  
506 *Mar. Pollut. Bull.* 69, 110–115. doi:10.1016/j.marpolbul.2013.01.021

507 Colombini, I., Brilli, M., Fallaci, M., Gagnarli, E., Chelazzi, L., 2011. Food webs of a sandy beach  
508 macroinvertebrate community using stable isotopes analysis. *Acta Oecologica* 37, 422–432.  
509 doi:10.1016/j.actao.2011.05.010

510 Council, N.R., 2003. The decline of the Steller sea lion in Alaskan waters: untangling food webs and  
511 fishing nets. National Academy of Sciences, Washington, D.C.

512 Cyr, A., Sergeant, C.J., Lopez, J.A., O’Hara, T., 2017. Assessing the influence of migration barriers and  
513 feeding ecology on total mercury concentrations in Dolly Varden (*Salvelinus malma*) from a

514           glaciated and non-glaciated stream. *Sci. Total Environ.* 580, 710–718.  
515           doi:10.1016/j.scitotenv.2016.12.017

516 DeNiro, M.J., Epstein, S., 1977. Mechanism of carbon isotope fractionation associated with lipid  
517           synthesis. *Science* 197, 261–263. doi:10.1126/science.327543

518 Doi, H., Kikuchi, E., Shikano, S., Takagi, S., 2010. Differences in nitrogen and carbon stable isotopes  
519           between planktonic and benthic microalgae. *Limnology* 11, 185–192. doi:10.1007/s10201-009-  
520           0297-1

521 Doll, A.C., Taras, B.D., Stricker, C.A., Rea, L.D., O’Hara, T.M., Cyr, A.P., Mcdermott, S., Loomis,  
522           T.M., Fadley, B.S., Wunder, M.B., 2018. Temporal records of diet diversity dynamics in individual  
523           adult female Steller sea lion (*Eumetopias jubatus*) vibrissae. *Oecologia* 188, 263–275.  
524           doi:10.1007/s00442-018-4173-8

525 Douglas, T.A., Loseto, L.L., MacDonald, R.W., Outridge, P., Dommergue, A., Poulain, A., Amyot, M.,  
526           Barkay, T., Berg, T., Chetelat, J., Constant, P., Evans, M., Ferrari, C., Gantner, N., Johnson, M.S.,  
527           Kirk, J., Kroer, N., Larose, C., Lean, D., Nielsen, T.G., Poissant, L., Rognerud, S., Skov, H.,  
528           Sørensen, S., Wang, F., Wilson, S., Zdanowicz, C.M., 2012. The fate of mercury in Arctic terrestrial  
529           and aquatic ecosystems, a review. *Environ. Chem.* 9, 321–355. doi:10.1071/EN11140

530 Eagles-Smith, C.A., Willacker, J.J., Flanagan Pritz, C.M., 2014. Mercury in fishes from 21 national parks  
531           in the Western United States- Inter and intra-park variation in concentrations and ecological risk:  
532           U.S. Geological Survey Open- File Report 2014-105.

533 Eagles-Smith, C.A., Ackerman, J.T., Willacker, J.J., Tate, M.T., Lutz, M.A., Fleck, J.A., Stewart, A.R.,  
534           Wiener, J.G., Evers, D.C., Lepak, J.M., Davis, J.A., Flanagan Pritz, C., 2016. Spatial and temporal  
535           patterns of mercury concentrations in freshwater fish across the Western United States and Canada.  
536           *Sci. Total Environ.* doi:10.1016/j.scitotenv.2016.03.229

537 Folch, J., Lees, M., Stanley, G.H.S., 1957. A simple method for the isolation and purification of total  
538           lipids from animal tissues. *J Biol Chem.* doi:10.1007/s10858-011-9570-9

539 Fox, A.L., Hughes, E.A., Trocine, R.P., Trefry, J.H., Schonberg, S. V., McTigue, N.D., Lasorsa, B.K.,

540 Konar, B., Cooper, L.W., 2014. Mercury in the northeastern Chukchi Sea: Distribution patterns in  
541 seawater and sediments and biomagnification in the benthic food web. *Deep. Res. Part II Top. Stud.*  
542 *Oceanogr.* 102, 56–67. doi:10.1016/j.dsr2.2013.07.012

543 Fritz, L.W., Towell, R., Gelatt, T.S., Johnson, D.S., Loughlin, T.R., 2014. Recent increases in survival of  
544 western steller sea lions in Alaska and implications for recovery. *Endanger. Species Res.* 26, 13–24.  
545 doi:10.3354/esr00634

546 Fry, B., 2006. *Stable Isotope Ecology*. Springer Science + Business Media, LLC, New York.

547 Gerber, L.R., Karimi, R., Fitzgerald, T.P., 2012. Sustaining seafood for public health. *Front. Ecol.*  
548 *Environ.* 10, 487–493. doi:10.1890/120003

549 Gill, G.A., Fitzgerald, W.F., 1987. Mercury in surface waters of the open ocean. *Global Biogeochem.*  
550 *Cycles* 1, 199–212.

551 Gorbatenko, K.M., Kiyashko, S.I., Lazhentsev, a. Y., Nadtochii, V. a., Savin, a. B., 2008. Benthic-  
552 pelagic trophic interactions within the fish assemblage in the western Bering Sea shelf area  
553 according to stomach content analysis and ratios of C and N stable isotopes. *Russ. J. Mar. Biol.* 34,  
554 497–506. doi:10.1134/S1063074008070092

555 Hamade, A.K., 2014. *Fish Consumption Advice for Alaskans: A Risk Management Strategy To Optimize*  
556 *the Public’s Health*.

557 Harley, J., Lieske, C., Bhojwani, S., Castellini, J.M., Lopez, J.A., O’Hara, T.M., 2015. Mercury and  
558 methylmercury distribution in tissues of sculpins from the Bering Sea. *Polar Biol.* 38, 1535–1543.  
559 doi:10.1007/s00300-015-1716-x

560 Hoffman, J.C., Sutton, T.T., 2010. Lipid correction for carbon stable isotope analysis of deep-sea fishes.  
561 *Deep Sea Res. Part I Oceanogr. Res. Pap.* 57, 956–964. doi:10.1016/j.dsr.2010.05.003

562 Jarman, W.M., Hobson, K. a., Sydeman, W.J., Bacon, C.E., McLaren, E.B., 1996. Influence of Trophic  
563 Position and Feeding Location on Contaminant Levels in the Gulf of the Farallones Food Web  
564 Revealed by Stable Isotope Analysis. *Environ. Sci. Technol.* 30, 654–660. doi:10.1021/es950392n

565 Johnson, N.W., Mitchell, C.P.J., Engstrom, D.R., Bailey, L.T., Coleman Wasik, J.K., Berndt, M.E., 2016.



566 Methylmercury production in a chronically sulfate-impacted sub-boreal wetland. Environ. Sci.  
567 Process. Impacts 18, 725–734. doi:10.1039/C6EM00138F

568 Kelly, C. a, Rudd, J.W.M., Holoka, M.H., 2003. Effect of pH on mercury uptake by an aquatic bacterium:  
569 implications for Hg cycling. Environ. Sci. Technol. 37, 2941–2946. doi:10.1021/es026366o

570 Lange, T. R., Royals, H.E., Connor, L.L., 1994. Mercury Accumulation in Largemouth Bass  
571 (*Micropterus salmoides*) in a Florida lake. Arch. Environ. Contam. Toxicol. 27, 466–471.

572 Laurier, F.J.G., Mason, R.P., Gill, G.A., Whalin, L., 2004. Mercury distributions in the North Pacific  
573 Ocean - 20 Years of observations. Mar. Chem. 90, 3–19. doi:10.1016/j.marchem.2004.02.025

574 Lehnherr, I., 2014. Methylmercury biogeochemistry: a review with special reference to Arctic aquatic  
575 ecosystems. Environ. Rev. 22, 229–243. doi:10.1139/er-2013-0059

576 Lehnherr, I., St. Louis, V.L., Hintelmann, H., Kirk, J.L., 2011. Methylation of inorganic mercury in polar  
577 marine waters. Nat. Geosci. 4, 298–302. doi:10.1038/ngeo1134

578 Logerwell, E.A., Aydin, K., Barbeaux, S., Brown, E., Connors, M.E., Lowe, S., Orr, J.W., Ortiz, I.,  
579 Reuter, R., Spencer, P., 2005. Geographic patterns in the demersal ichthyofauna of the Aleutian  
580 Islands. Fish. Oceanogr. 14, 93–112.

581 Lushchak, V.I., 2011. Environmentally induced oxidative stress in aquatic animals. Aquat. Toxicol. 101,  
582 13–30. doi:10.1016/j.aquatox.2010.10.006

583 Marsh, J.M., Hillgruber, N., Foy, R.J., Marsh, J.M., Hillgruber, N., Temporal, R.J.F., Marsh, J.M.,  
584 Hillgruber, N., Foy, R.J., 2012. Temporal and Ontogenetic Variability in Trophic Role of Four  
585 Groundfish Species — Walleye Pollock , Pacific Cod , Arrowtooth Flounder , and Pacific Halibut  
586 — around Kodiak Island in the Gulf of Alaska. Trans. Am. Fish. Soc. 8487.  
587 doi:10.1080/00028487.2012.667042

588 McGrew, A.K., Ballweber, L.R., Moses, S.K., Stricker, C. a, Beckmen, K.B., Salman, M.D., O’Hara,  
589 T.M., 2014. Mercury in gray wolves (*Canis lupus*) in Alaska: increased exposure through  
590 consumption of marine prey. Sci. Total Environ. 468–469, 609–13.  
591 doi:10.1016/j.scitotenv.2013.08.045

592 NMFS, 2013. Draft status review of the eastern distinct population segment of Steller sea lion  
593 (*Eumetopias jubatus*), Protected Resources Division, Alaska Region, National Marine Fisheries  
594 Service, 709 W 9th St., Juneau, AK 99802.

595 Peterson, B.J., Fry, B., 1987. Stable isotopes in ecosystem studies. *Ann. Rev. Ecol. Syst* 18, 293–320.  
596 doi:10.1146/annurev.es.18.110187.001453

597 Post, D.M., Layman, C. a., Arrington, D.A., Takimoto, G., Quattrochi, J., Montaña, C.G., 2007. Getting  
598 to the fat of the matter: models, methods and assumptions for dealing with lipids in stable isotope  
599 analyses. *Oecologia* 152, 179–189. doi:10.1007/s00442-006-0630-x

600 Power, M., Klein, G.M., Guiguer, K.R.R.A., Kwan, M.K.H., 2002. Mercury accumulation in the fish  
601 community of a sub-Arctic lake in relation to trophic position and carbon sources. *J. Appl. Ecol.* 39,  
602 819–830. doi:10.1046/j.1365-2664.2002.00758.x

603 Rea, L.D., Castellini, J.M., Correa, L., Fadely, B.S., O’Hara, T.M., 2013. Maternal Steller sea lion diets  
604 elevate fetal mercury concentrations in an area of population decline. *Sci. Total Environ.* 454–455,  
605 277–282. doi:10.1016/j.scitotenv.2013.02.095

606 Rea, L.D., Christ, A.M., Hayden, A.B., Stegall, V.K., Farley, S.D., Stricker, C.A., Mellish, J.A.E.,  
607 Maniscalco, J.M., Waite, J.N., Burkanov, V.N., Pitcher, K.W., 2015. Age-specific vibrissae growth  
608 rates: A tool for determining the timing of ecologically important events in Steller sea lions. *Mar.*  
609 *Mammal Sci.* 31, 1213–1233. doi:10.1111/mms.12221

610 Ricca, M.A., Keith Miles, A., Anthony, R.G., 2008. Sources of organochlorine contaminants and mercury  
611 in seabirds from the Aleutian archipelago of Alaska: Inferences from spatial and trophic variation.  
612 *Sci. Total Environ.* 406, 308–323. doi:10.1016/j.scitotenv.2008.06.030

613 Schell, D.M., Barnett, B.A., Vinette, K.A., 1998. Carbon and nitrogen isotope ratios in zooplankton of the  
614 Bering, Chukchi and Beaufort seas. *Mar. Ecol. Prog. Ser.* 162, 11–23. doi:10.3354/meps162011

615 Selin, N.E., 2009. Global Biogeochemical Cycling of Mercury: A Review. *Annu. Rev. Environ. Resour.*  
616 34, 43–63. doi:10.1146/annurev.enviro.051308.084314

617 Strode, S.A., Jaegle, L., Jaffe, D.A., Swartzendruber, P.C., Selin, N.E., Holmes, C., Yantosca, R.M.,

618 2008. Trans-Pacific transport of mercury. *J. Geophys. Res.* 113, 1–12. doi:10.1029/2007JD009428

619 Sunderland, E.M., Krabbenhoft, D.P., Moreau, J.W., Strode, S. a., Landing, W.M., 2009. Mercury  
620 sources, distribution, and bioavailability in the North Pacific Ocean: Insights from data and models.  
621 *Global Biogeochem. Cycles* 23, n/a-n/a. doi:10.1029/2008GB003425

622 Sweeting, C.J., Polunin, N.V.C., Jennings, S., 2006. Effects of chemical lipid extraction and arithmetic  
623 lipid correction on stable isotope ratios of fish tissues. *Rapid Commun. Mass Spectrom.* 20, 595–  
624 601. doi:10.1002/rcm.2347

625 TenBrink, T.T., Aydin, K.Y., 2009. Life history traits of sculpins in the eastern Bering Sea and Aleutian  
626 Islands.

627 TenBrink, T.T., Buckley, T.W., 2013. Life-History Aspects of the Yellow Irish Lord (*Hemilepidotus*  
628 *jordani*) in the Eastern Bering Sea and Aleutian Islands. *Northwest. Nat.* 94, 126–136.  
629 doi:10.1898/12-33.1

630 Trudel, M., Rasmussen, J.B., 1997. Modeling the elimination of mercury by fish. *Environ. Sci. Technol.*  
631 31, 1716–1722. doi:10.1021/es960609t

632 Vander Zanden, J., Rasmussen, J.B., 2001. Variation in d15 N and d13 C trophic fractionation:  
633 Implications for aquatic food web studies. *Limnol. Oceanogr.* 46, 2061–2066.

634 Walker, C.H., Sibly, R.M., Hopkin, S.P., Peakall, D.B., 2012. *Principles of Ecotoxicology*, 4th ed. CRC  
635 Press, Boca Raton, FL.

636 Wang, S.W., Budge, S.M., Gradinger, R.R., Iken, K., Wooller, M.J., 2014. Fatty acid and stable isotope  
637 characteristics of sea ice and pelagic particulate organic matter in the Bering Sea: Tools for  
638 estimating sea ice algal contribution to Arctic food web production. *Oecologia* 174, 699–712.  
639 doi:10.1007/s00442-013-2832-3

640 Ward, D.M., Nislow, K.H., Folt, C.L., 2010. Bioaccumulation syndrome: Identifying factors that make  
641 some stream food webs prone to elevated mercury bioaccumulation. *Ann. N. Y. Acad. Sci.* 1195,  
642 62–83. doi:10.1111/j.1749-6632.2010.05456.x

643 Willacker, J.J., von Hippel, F.A., Ackerly, K.L., O’Hara, T.M., 2013. Habitat-specific foraging and sex

644 determine mercury concentrations in sympatric benthic and limnetic ecotypes of threespine  
645 stickleback. *Environ. Toxicol. Chem.* 32, 1623–1630. doi:10.1002/etc.2213

646 Yang, M.-S., Nelson, M.W., 1999. Food habits of the commercially important groundfishes in the Gulf of  
647 Alaska in 1990, 1993, and 1996, NOAA Technical Memorandum.

648 Yang, M.S., 2003. Food habits of important groundfishes in the Aleutian Islands in 1994 and 1997, AFSC  
649 Processed Report.

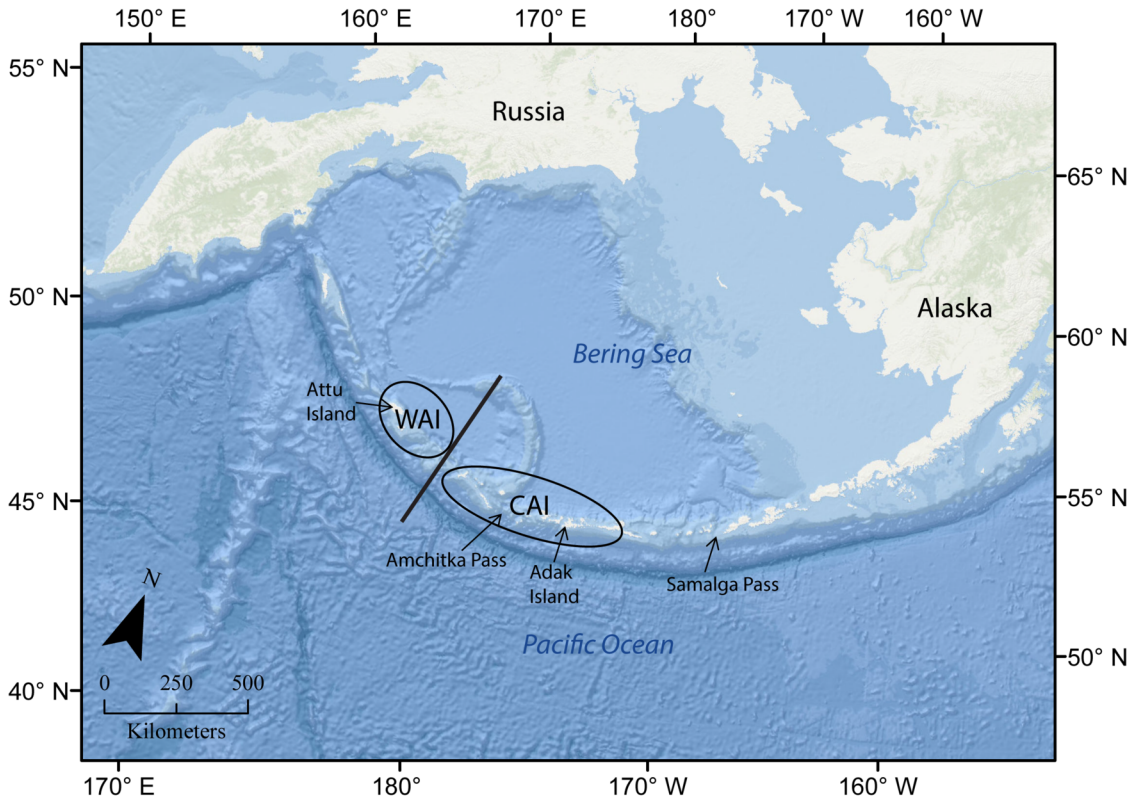
650 York, A.E., Merrick, R.L., Loughlin, T.R., 1996. An analysis of the Steller sea lion metapopulation in  
651 Alaska, in: McCullough, D.R. (Ed.), *Metapopulations and Wildlife Conservation*. Island Press,  
652 Washington, D.C., p. Pp. 259-292.

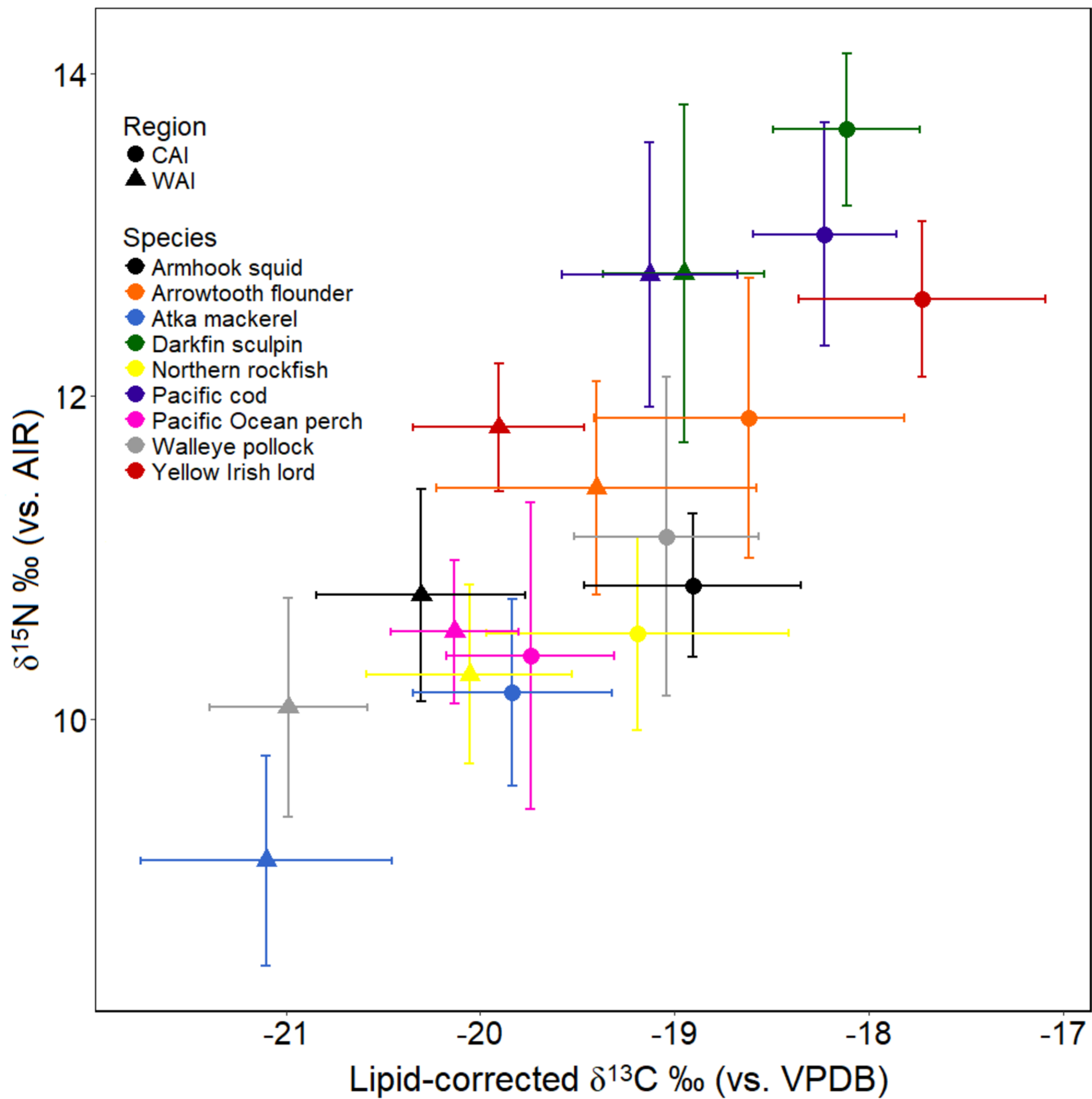
**Figure 1.** Map depicting the approximate extent of sample collection, within the context of Steller sea lion management regions. WAI is Western Aleutian Islands, and CAI is Central Aleutian Islands.

**Figure 2.** Mean  $\delta^{15}\text{N}$  and lipid-corrected  $\delta^{13}\text{C}$  values ( $\pm 1$  standard deviation) of muscle samples for each fish and invertebrate species, categorized by western Aleutian Islands (WAI) and central Aleutian Islands (CAI). VPDB is the isotopic standard Vienna Pee-Dee Belemnite, AIR is the isotopic standard atmospheric air.

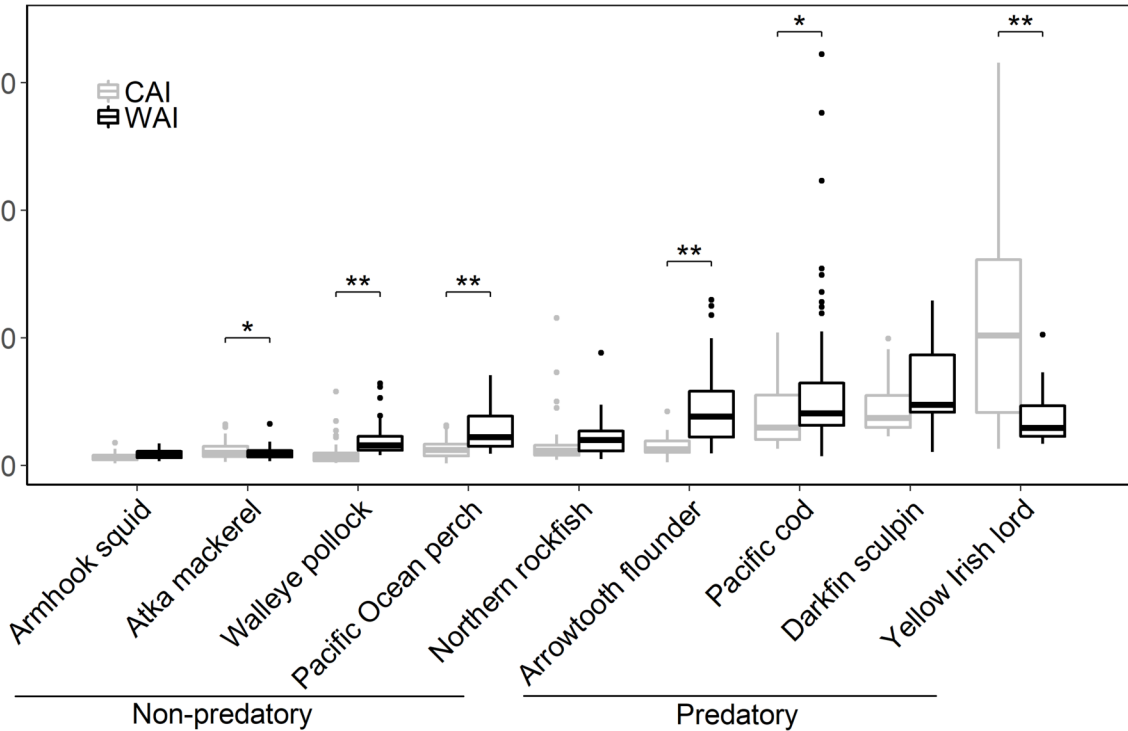
**Figure 3.** Box and whisker plot representing the length-standardized muscle [THg] for each fish and invertebrate species, characterized by region. Data presented on a log scale on the y-axis. Bold horizontal lines inside each box represent median values, bottom and top edges of boxes represent 25th and 75th percentiles, respectively, and the ends of the vertical solid lines represent  $\pm 1.5$  \* interquartile range. Length-standardized [THg] beyond this range are displayed as individual points. \* denotes significance level between regions for the species indicated,  $\alpha \leq 0.01$ ; \*\*, denotes significance  $\alpha \leq 0.001$ . WAI is Western Aleutian Islands, and CAI is Central Aleutian Islands.

**Figure 4.** [THg] and  $\delta^{15}\text{N}$  values for muscle samples, individual species' regression slopes. [THg] and  $\delta^{15}\text{N}$  values for muscle samples, individual species' regression slopes. WAI is Western Aleutian Islands, and CAI is Central Aleutian Islands. AIR is the isotopic standard atmospheric air.





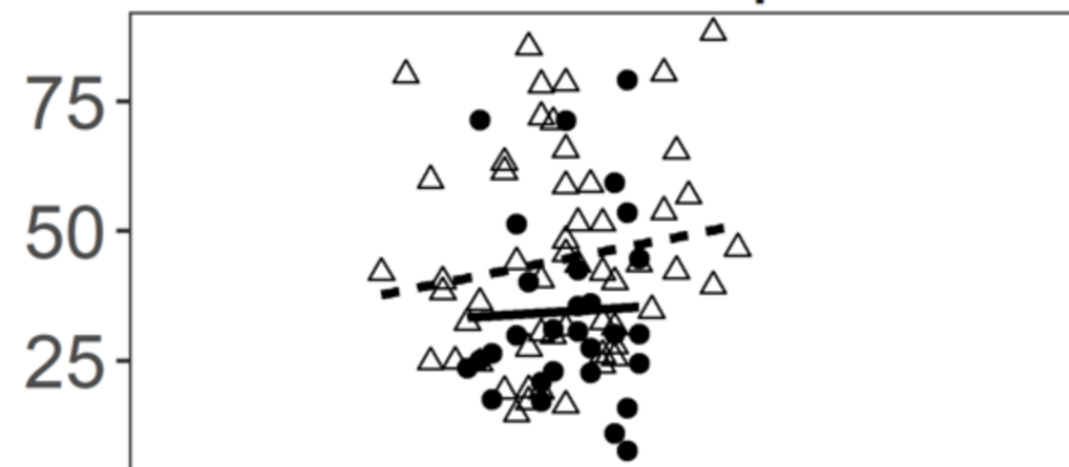
Length-standardized  
Total mercury (ng/g ww)



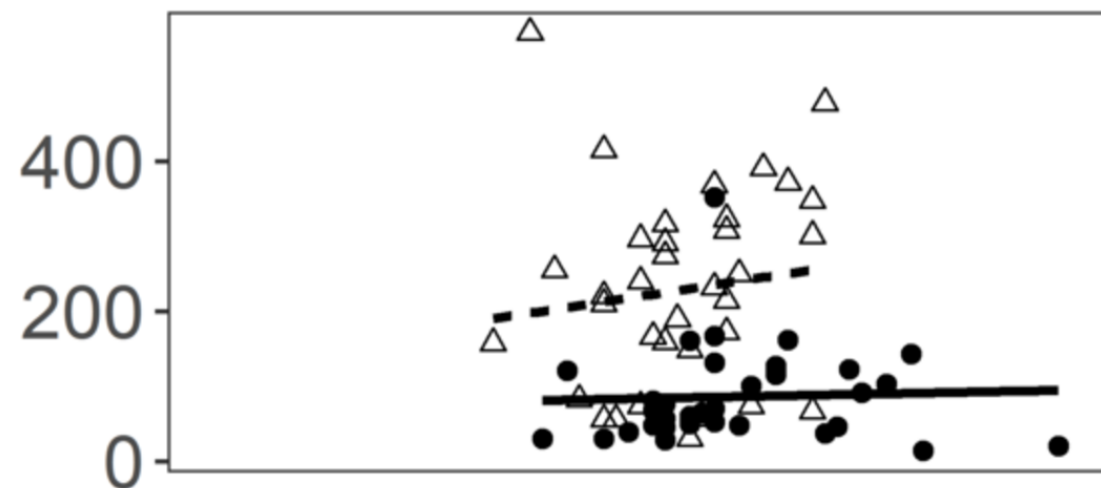


Muscle total mercury ng/g ww

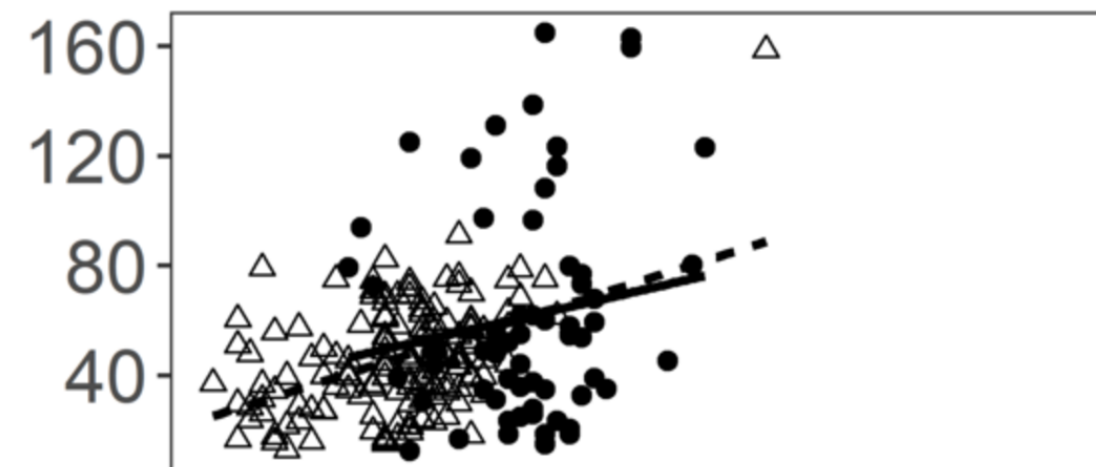
Armhook squid



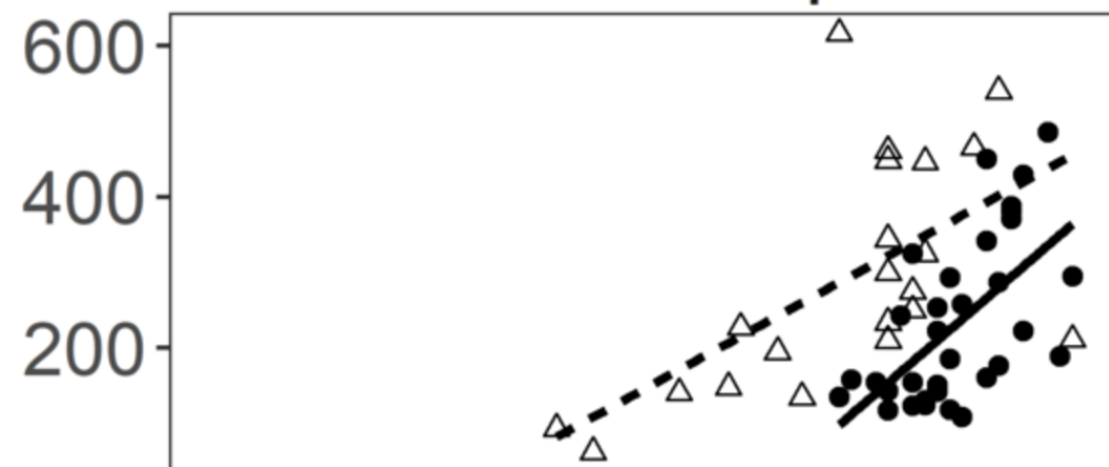
Arrowtooth flounder



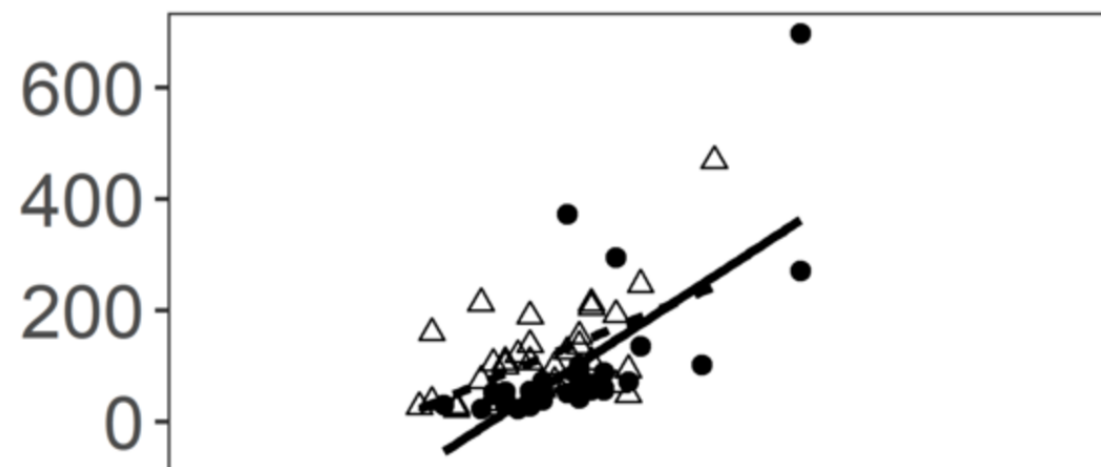
Atka mackerel



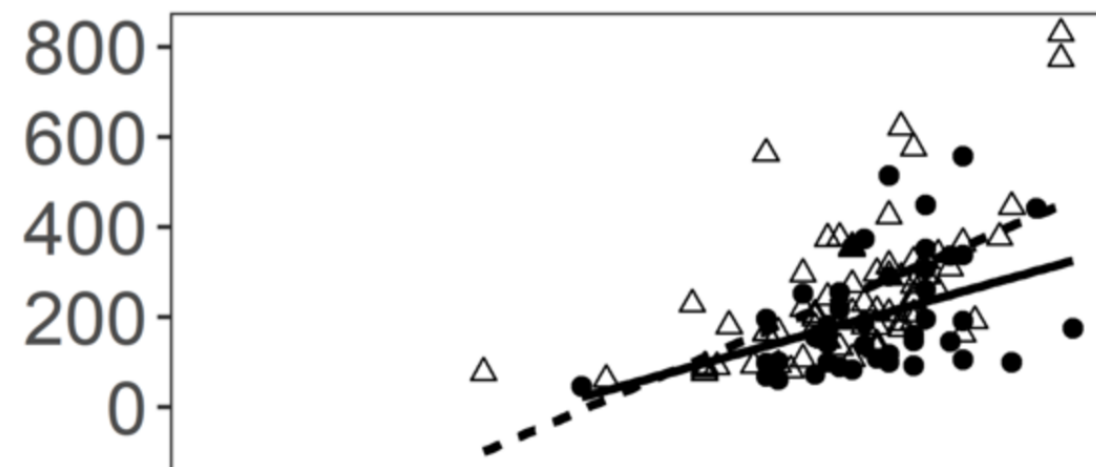
Darkfin sculpin



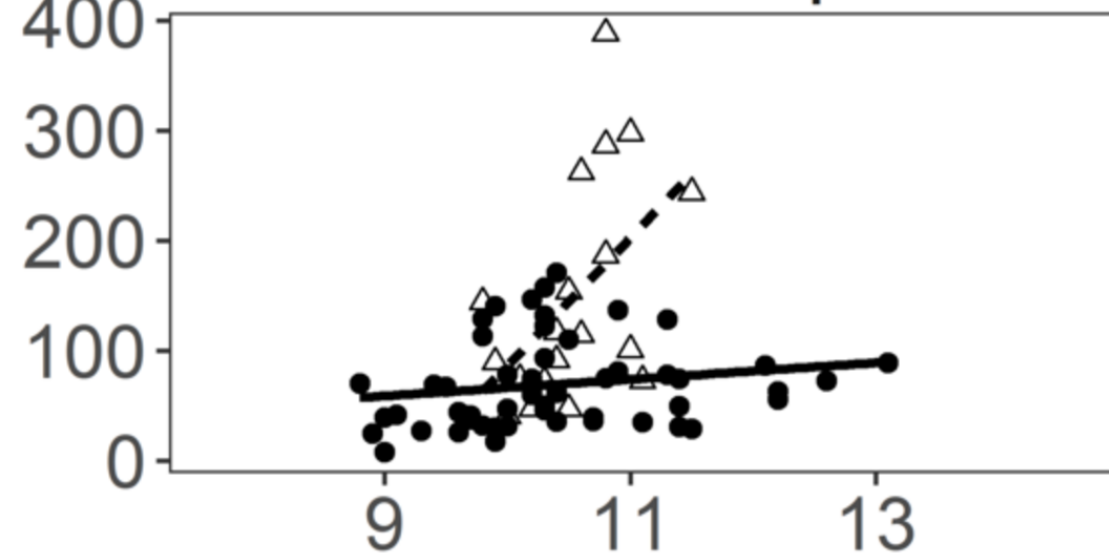
Northern rockfish



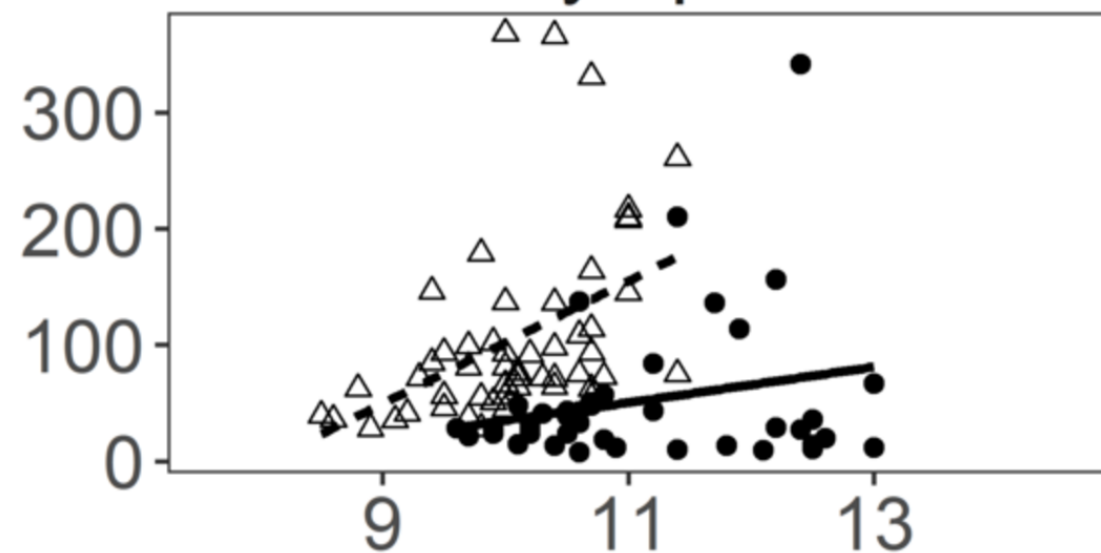
Pacific cod



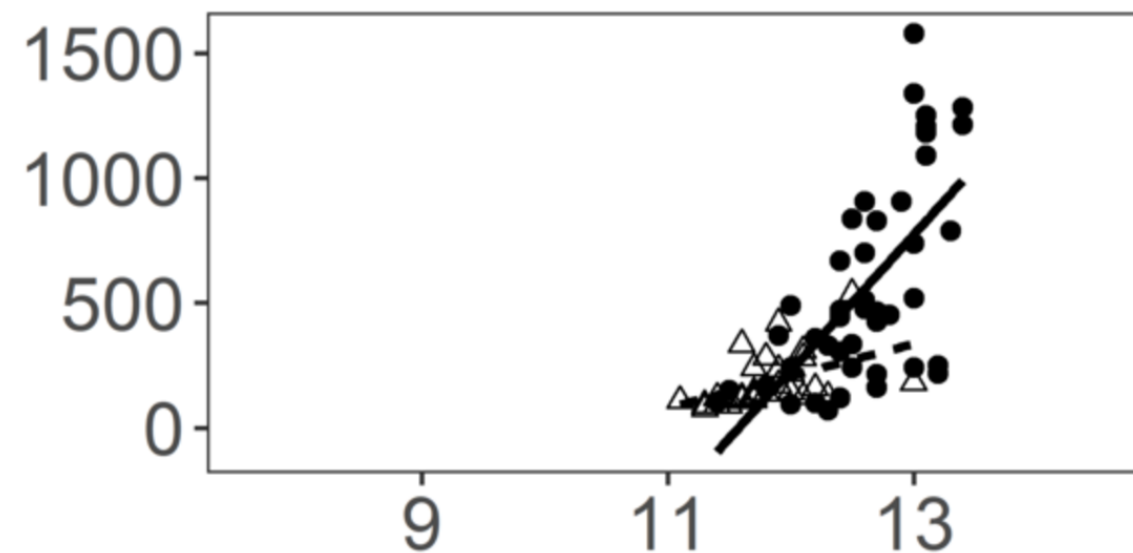
Pacific Ocean perch



Walleye pollock



Yellow Irish lord



Region

● CAI

△ WAI

$\delta^{15}\text{N}$  ‰ (vs. AIR)

**Table 1.** Total mercury concentrations ([THg]) and stable nitrogen and carbon isotope values for western Aleutian Islands (WAI) and central Aleutian Islands (CAI) fishes and cephalopods. Sample sizes (N) for each region, fork length (cm), mass (g), [THg] as measured (ng/g ww), length-standardized [THg] in ng/g ww,  $\delta^{15}\text{N}$  values, bulk  $\delta^{13}\text{C}$  values, and lipid-corrected  $\delta^{13}\text{C}$  values for each species in the dataset. Data are means  $\pm$  SD, geometric mean for [THg].

Species	Region	N	Fork Length (cm)	Mass (g)	Unadjusted [THg] ng/g ww	Length- standardized [THg] ng/g ww	$\delta^{15}\text{N}$ (‰)	$\delta^{13}\text{C}$ (‰)	Lipid- corrected $\delta^{13}\text{C}$ (‰)
Armhook squid	CAI	30	22.3 $\pm$ 3.2	414.7 $\pm$ 141.6	29.1 $\pm$ 18.4	28.9 $\pm$ 18.9	10.8 $\pm$ 0.5	<b>-20.4 <math>\pm</math> 0.5</b>	<b>-18.9 <math>\pm</math> 0.6</b>
<i>Berryteuthis magister</i>	WAI	67	25.1 $\pm$ 6.2	471.6 $\pm$ 144.4	42.1 $\pm$ 22.1	39.4 $\pm$ 18.4	10.8 $\pm$ 0.7	-21.7 $\pm$ 0.5	-20.3 $\pm$ 0.5
Arrowtooth flounder	CAI	41	48.1 $\pm$ 10.2	1,226.8 $\pm$ 824.6	69.8 $\pm$ 64.0	62.4 $\pm$ 39.3	11.9 $\pm$ 0.9	<b>-20.4 <math>\pm</math> 1.7</b>	<b>-18.6 <math>\pm</math> 0.8</b>
<i>Atheresthes stomias</i>	WAI	39	<b>59.6 <math>\pm</math> 17.2</b>	<b>2,705.1 <math>\pm</math> 1,971.0</b>	<b>188.8 <math>\pm</math> 127.9</b>	<b>184.4 <math>\pm</math> 161.8</b>	11.4 $\pm$ 0.7	-22.3 $\pm$ 2.0	-19.4 $\pm$ 0.8
Atka mackerel	CAI	73	<b>40.2 <math>\pm</math> 7.1</b>	<b>913.7 <math>\pm</math> 456.8</b>	<b>50.5 <math>\pm</math> 38.0</b>	<b>51.1 <math>\pm</math> 37.4</b>	<b>10.2 <math>\pm</math> 0.6</b>	<b>-22.0 <math>\pm</math> 1.2</b>	<b>-19.8 <math>\pm</math> 0.5</b>
<i>Pleurogrammus monopterygius</i>	WAI	148	36.8 $\pm$ 3.1	520.9 $\pm$ 239.4	42.8 $\pm$ 20.8	42.7 $\pm$ 19.4	9.1 $\pm$ 0.6	-22.8 $\pm$ 1.0	-21.1 $\pm$ 0.6
Darkfin sculpin	CAI	34	<b>20.2 <math>\pm</math> 3.3</b>	<b>158.8 <math>\pm</math> 74.3</b>	207.6 $\pm$ 109.7	208.0 $\pm$ 99.5	<b>13.7 <math>\pm</math> 0.5</b>	<b>-19.2 <math>\pm</math> 0.4</b>	<b>-18.1 <math>\pm</math> 0.4</b>
<i>Malacocottus zonurus</i>	WAI	21	16.9 $\pm$ 2.9	71.4 $\pm$ 46.3	250.2 $\pm$ 153.6	251.1 $\pm$ 158.3	12.8 $\pm$ 1.0	-20.0 $\pm$ 0.4	-19.0 $\pm$ 0.4
Northern rockfish	CAI	41	32.4 $\pm$ 3.9	485.4 $\pm$ 193.1	63.7 $\pm$ 121.0	64.3 $\pm$ 104.0	10.5 $\pm$ 0.6	<b>-20.3 <math>\pm</math> 0.7</b>	<b>-19.2 <math>\pm</math> 0.8</b>
<i>Sebastes polyspinis</i>	WAI	36	31.3 $\pm$ 3.4	496.4 $\pm$ 236.4	87.1 $\pm$ 86.8	87.6 $\pm$ 80.6	10.3 $\pm$ 0.6	-21.2 $\pm$ 0.7	-20.1 $\pm$ 0.5
Pacific cod	CAI	44	59.2 $\pm$ 11.0	2,915.9 $\pm$ 2,050.4	170.2 $\pm$ 127.6	162.5 $\pm$ 114.7	13.0 $\pm$ 0.7	<b>-19.3 <math>\pm</math> 0.4</b>	<b>-18.2 <math>\pm</math> 0.4</b>
<i>Gadus macrocephalus</i>	WAI	88	<b>68.9 <math>\pm</math> 25.5</b>	<b>4,475.0 <math>\pm</math> 4,372.7</b>	<b>225.3 <math>\pm</math> 163.3</b>	<b>236.6 <math>\pm</math> 279.5</b>	12.8 $\pm$ 0.8	-20.2 $\pm$ 0.6	-19.1 $\pm$ 0.5
Pacific Ocean perch	CAI	53	36.5 $\pm$ 2.8	700.0 $\pm$ 174.3	58.1 $\pm$ 39.9	57.5 $\pm$ 36.6	10.4 $\pm$ 1.0	<b>-21.4 <math>\pm</math> 0.9</b>	<b>-19.7 <math>\pm</math> 0.4</b>
<i>Sebastes alutus</i>	WAI	20	<b>39.5 <math>\pm</math> 3.2</b>	<b>935.0 <math>\pm</math> 306.6</b>	<b>116.1 <math>\pm</math> 100.2</b>	<b>118.8 <math>\pm</math> 87.3</b>	10.5 $\pm$ 0.4	-21.8 $\pm$ 0.8	-20.1 $\pm$ 0.3
Walleye pollock	CAI	41	54.5 $\pm$ 4.6	<b>1,412.2 <math>\pm</math> 321.9</b>	33.7 $\pm$ 64.3	34.1 $\pm$ 87.3	<b>11.1 <math>\pm</math> 1.0</b>	<b>-20.0 <math>\pm</math> 0.5</b>	<b>-19.0 <math>\pm</math> 0.5</b>
<i>Gadus chalcogrammus</i>	WAI	52	52.3 $\pm$ 7.5	1,051.9 $\pm$ 435.0	<b>85.7 <math>\pm</math> 80.9</b>	<b>88.2 <math>\pm</math> 62.6</b>	10.1 $\pm$ 0.7	-22.0 $\pm$ 0.4	-21.0 $\pm$ 0.4
Yellow Irish lord	CAI	45	<b>39.8 <math>\pm</math> 3.6</b>	780.0 $\pm$ 225.2	<b>410.3 <math>\pm</math> 408.7</b>	<b>412.2 <math>\pm</math> 390.7</b>	<b>12.6 <math>\pm</math> 0.5</b>	<b>-18.9 <math>\pm</math> 0.6</b>	<b>-17.7 <math>\pm</math> 0.6</b>
<i>Hemilepidotus jordani</i>	WAI	29	36.4 $\pm$ 5.2	627.6 $\pm$ 332.6	160.6 $\pm$ 108.9	162.9 $\pm$ 103.3	11.8 $\pm$ 0.4	-21.0 $\pm$ 0.4	-19.9 $\pm$ 0.4

Bold text indicates significant difference between WAI and CAI for the means of the given metric within a species, the larger value in bold, significance level  $\alpha \leq 0.05$

WAI and CAI are Steller sea lion management sub-regions

**Table 2.** Differences in the lipid-corrected  $\delta^{13}\text{C}$  values regionally ( $\Delta\text{CAI-WAI}$ ), and the regional difference (P values) for isotopic space comparisons for each species. Significance determined by Hotelling's  $T^2$  test comparing mean  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  values in multivariate space.

<b>Species</b>	<b><math>\Delta\text{CAI-WAI}</math> Lipid-corrected <math>\delta^{13}\text{C}</math> (‰)</b>	<b>Isotopic space <math>\delta^{15}\text{N}</math> ‰ and Lipid- corrected <math>\delta^{13}\text{C}</math> (‰)</b>
Armhook squid	1.4*	0.001
Arrowtooth flounder	0.8*	0.001
Atka mackerel	1.3*	0.001
Darkfin sculpin	0.8*	0.001
Northern rockfish	0.9*	0.001
Pacific cod	0.9*	0.001
Pacific Ocean perch	0.4*	0.01
Walleye pollock	1.9*	0.001
Yellow Irish lord	2.2*	0.001

WAI is western Aleutian Islands, CAI is central Aleutian Islands Steller sea lion management sub-regions  
 \* indicates a significant difference between WAI and CAI in mean lipid-corrected  $\delta^{13}\text{C}$  values,  $\alpha < 0.05$ .

**Table 3.** Variance explained ( $R^2$ ) and significance (P value) for linear regression of unadjusted total mercury concentrations ([THg]) and  $\delta^{15}\text{N}$  values for western (WAI) and central (CAI) Aleutian Islands.

<b>Species</b>	<b>WAI</b>	<b>CAI</b>
Armhook squid	0.03 (0.2)	0.00 (0.86)
Atka mackerel	<b>0.17 (0.001)</b>	0.02 (0.28)
Arrowtooth flounder	0.01 (0.52)	0.00 (0.84)
Pacific cod	<b>0.51 (0.0001)</b>	<b>0.24 (0.001)</b>
Pacific Ocean perch	<b>0.27 (0.02)</b>	0.08 (0.06)
Walleye pollock	0.3 (0.3)	0.00 (0.6)
Darkfin sculpin	<b>0.55 (0.001)</b>	<b>0.36 (0.001)</b>
Northern rockfish	<b>0.35 (0.001)</b>	<b>0.62 (0.0001)</b>
Yellow Irish lord	<b>0.28 (0.01)</b>	<b>0.40 (0.0001)</b>

Data displayed as:  $R^2$  (P value)

Bold text indicates significant relationship for [THg] and  $\delta^{15}\text{N}$  values for each region, significance level  $\alpha \leq 0.05$

WAI and CAI are Steller sea lion management sub-regions

**Table 4.** Significance (P value) from general linear models for the influence of  $\delta^{15}\text{N}$  values, lipid-corrected  $\delta^{13}\text{C}$  values, the interaction of  $\delta^{15}\text{N}$  values and region, and the interaction of lipid-corrected  $\delta^{13}\text{C}$  values and region on unadjusted total mercury concentrations ([THg]) for each species.

<b>Species</b>	<b><math>\delta^{15}\text{N}</math> (‰)</b>	<b>Lipid-corrected <math>\delta^{13}\text{C}</math> (‰)</b>	<b>Region* <math>\delta^{15}\text{N}</math> (‰)</b>	<b>Region * lipid-corrected <math>\delta^{13}\text{C}</math> (‰)</b>
Armhook squid	n.s.	n.s.	n.s.	n.s.
Atka mackerel	n.s.	n.s.	n.s.	n.s.
Arrowtooth flounder	n.s.	0.01	n.s.	0.01
Pacific cod	0.001	0.001	n.s.	0.001
Pacific Ocean perch	0.05	0.01	n.s.	n.s.
Walleye pollock	n.s.	0.01	0.05	0.001
Darkfin sculpin	0.001	0.001	n.s.	n.s.
Northern rockfish	0.001	n.s.	n.s.	0.01
Yellow Irish lord	0.001	0.001	n.s.	0.01

n.s. indicates not significant.