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37 Abstract

As ecosystem engineers, oysters create and maintain structured habitat and can influence 38 trophodynamics and benthic-pelagic coupling in the surrounding landscape. The physical reef 39 structure and associated biotic parameters can affect the availability of food resources for 40 oysters. Oysters and potential composite food sources — suspended particulate organic matter 41 (SPOM) and surface sediment organic matter (SSOM) — were assessed using a dual stable 42 isotope (δ^{13} C, δ^{15} N) approach at three reef types (natural, restored, and unconsolidated) 43 44 seasonally for two years to determine if changes in physical and/or biotic parameters affected the relative availability and/or use of food resources by oysters. SPOM was more depleted in ¹³C (-45 46 24.2 ± 0.6 %, mean \pm SD) than SSOM (-21.2 ± 0.8 %). SPOM composition is likely dominated by autochthonous phytoplankton production, while SSOM includes trapped phytoplankton and 47 benthic microalgae. SSOM was used by oysters in increasing proportions relative to SPOM over 48 49 time at all reef types. This temporal trend is likely due to increased oyster biomass over time, promoting enhanced microphytobenthos growth through feedback effects related to oyster 50 51 biodeposits. Structural differences between reef types observed in this study had no effect on food resource availability and use by oysters, indicating strong bentho-pelagic coupling likely 52 due to shallow depths as well as strong and consistent winds. This study provides insights for 53 restoration of oyster reefs as it highlights that food resources used by oysters remain similar 54 among reef types despite changes in abiotic and biotic parameters among habitats and over time. 55 **Keywords:** 56 food source; oyster reef; bentho-pelagic coupling; stable isotopes; suspended particulate organic 57 matter; sediment organic matter; Gulf of Mexico 58

60 **1. Introduction**

Eastern oysters, *Crassostrea virginica*, are ecosystem engineers that create and maintain 61 structured biogenic habitat in estuarine areas and provide resources for other organisms. Oyster 62 reefs serve as nursery grounds for a number of commercially and ecologically important species 63 by enhancing survival and abundance (Beck et al. 2001). Oyster reefs substantially augment 64 production (>200 g m⁻² y⁻¹) for associated fish and mobile crustaceans (Peterson et al. 2003, zu 65 66 Ermgassen et al. 2015). Oysters help regulate nutrient dynamics and mediate benthic-pelagic coupling through their suspension-feeding activities (Beseres Pollack et al. 2013, Dame et al. 67 1984, Newell 2004). Oysters filter large amounts of particulate matter from the water column 68 69 and excrete particles as biodeposits, thereby transferring organic matter from the water column to the benthos (Dame et al. 1984, Dame 2012, Newell 2004, Prins et al. 1998). Moreover, the three-70 dimensional structure of oyster reefs provides a large range of microhabitats, increasing the 71 72 diversity of organisms and supporting complex trophic webs (Bateman and Bishop 2017, Grabowski and Peterson 2007, Summerhayes et al. 2009). 73 74 Oysters can rely on a large diversity of food resources, whose availability relates to both temporal and spatial variation. Food resources present in estuarine water columns can include 75 phytoplankton, detritus, bacteria, and resuspended materials including benthic microalgae 76 (Cranford et al. 2005, Dame 2012, Grant et al. 1997, Leal et al. 2008, Riera and Richard 1996). 77 The composition of suspended particulate organic matter can vary spatially with relative location 78 within an estuary (i.e., with relative influences of riverine and oceanic inputs; Leal et al. 2008, 79 Quan et al. 2012, Riera and Richard 1996) and with relative proximity to other habitats (e.g., salt 80 81 marshes, tidal flats; Kang et al. 2003, Quan et al. 2012, Riera and Richard 1996, Winemiller et al. 2007). Availability of food resources also varies temporally, with daily, seasonal and perhaps 82

longer-term climatic cycles (Cranford and Hill 1999, Grant et al. 1997, MacIntyre and Cullen 83 1996, Prins et al. 1998, Reyna et al. 2017). Tidal currents, wave action, high winds and human 84 activities can cause resuspension of bottom sediments and organic matter, further influencing 85 food resource composition available to suspension-feeders at varying spatial and temporal scales 86 (Grant et al. 1997, Miller et al. 1996, Prins et al. 1998). Additionally, oysters can promote the 87 growth of microphytobenthos through their biodeposits, effectively transferring nutrients to the 88 89 sediments where they can fuel microalgae growth (Hargrave et al. 2008, Haven and Morales-90 Alamo 1966, Leal et al. 2008, Prins et al. 1998).

The physical structure of the habitat (e.g. sediment grain size, vertical relief) and the 91 92 biotic parameters (e.g., oyster density, biomass) themselves can affect the availability of food resources for oysters (Dame et al. 1984), and therefore the production of oyster reefs (Soniat et 93 al. 2004). Indeed, resuspension of sediment and organic matter is higher in open, bare habitats or 94 95 where sediment grain size is small—increasing the availability of benthic food resources for oysters-than in enclosed or structured habitats or coarse sediment (Grant et al. 1997, Sundbäck 96 1984). However, extraordinarily high sediment resuspension can also have negative effects on 97 suspension-feeders. High turbidity caused by resuspended sediments can decrease filtration 98 ability and dilute food concentrations (Dame 2012, Grant et al. 1990, Newell and Langdon 99 1996). Higher biomass of oysters, leading to a higher three-dimensional complexity, may 100 decrease water flow speed over the reef and increase deposition of particles from the water 101 column (Colden et al. 2016). 102

103 The vertical relief of reefs may also influence the availability of sediment food sources to 104 oysters. Because large amounts of suspended sediment can contribute to the burial of oyster 105 reefs, approaches for reef restoration have moved to constructing higher vertical relief reefs to

prevent sedimentation and burial (Bartol et al. 1999, Grabowski 2004, Lenihan 1999, Powers et
al. 2009, Schulte et al. 2009). However, high vertical relief could subsequently alter the degree
of benthic-pelagic coupling and flow of organic matter through reef ecosystems (Fréchette et al.
109 1989, Quan et al. 2012). Thus, as movement continues toward designing and building higher
vertical relief reefs, it is important to understand how the relative availability of different food
sources may change with reef height.

112 The aim of this study is to compare the food resources used by oysters in several oyster 113 reef types to determine if changes in physical and/or biotic parameters affect (1) the availability of potential food sources to oysters, and (2) the use of pelagic vs. benthic food sources by 114 115 oysters. We examined the relative contributions of water column and benthic food resources to subtidal oysters (Crassostrea virginica Gmelin) in a shallow Texas estuary using a dual stable 116 isotope ($\delta^{13}C$, $\delta^{15}N$) approach. Analysis of stable isotope composition, particularly carbon 117 118 isotope composition, can be used to elucidate the different food resources utilized by consumers in natural settings (Fry and Sherr 1984, Fry 2006, Peterson 1999). Three reef types with 119 120 different characteristics were compared: (1) a natural reef, (2) a restored reef, and (3) an unconsolidated reef. Field sampling was conducted over a two year period to examine how the 121 use of food resources by oysters may differ among these reef types depending on their structure, 122 as well as during development of the restored reef. 123

124 **2.** Materials and methods

125 2.1 Study Area and Experimental Design

126 The study is a 2-way design, where the first independent variable compares three reef127 types (natural, restored, unconsolidated), and the second independent variable compares response

over six time periods. The dependent variables are C and N stable isotope compositions inpotential water column and sediment composite food resources and oysters.

The Mission-Aransas Estuary is a bar-built system in the Texas Coastal Bend (Fig. 1a). 130 The estuary occupies approximately 463 km² and is composed of several shallow (average 2 m 131 depth) bays (Armstrong 1987). The area has a semi-arid, subtropical climate with infrequent rain 132 events. Evaporation typically exceeds precipitation (62.7 cm year⁻¹ net water loss) (Armstrong 133 1987). Two rivers feed the system: the Mission River flows through Mission Bay into Copano 134 135 Bay and the Aransas River flows directly into Copano Bay. Average freshwater inflow to the estuary is low (< 500 million m³ year⁻¹) (Armstrong 1987, Montagna et al. 1996). The estuary is 136 137 microtidal (0.15 m tidal range) and water movement is predominantly wind-influenced (Evans and Morehead Palmer 2012). Mixing with the Gulf of Mexico occurs via a tidal inlet at the 138 southern end of the system. High winds characteristic of the region maintain well-mixed 139 140 conditions throughout the system (Evans and Morehead Palmer 2012). Average water residence time is 360 days, but can be as long as three years (Beseres Pollack et al. 2011, Montagna et 141 al.1996). Oyster reefs are common in the estuary (Fig. 1a). Reefs are primarily subtidal and are 142 more prominent in areas of low to moderate salinity. 143

The study was conducted in Copano Bay (Fig. 1b; 28.135 °N, 97.053 °W), where average
depth is 1.1 m. Oysters and potential composite food resources — suspended particulate organic
matter (SPOM) and surface sediment organic matter (SSOM) — were sampled at three reef
types, which had the following characteristics (see also supplementary material 1):
Natural reef: Low vertical relief (< 0.1 m), with densities (mean ± standard deviation)

of adult and spat oysters of 196 ± 134 and 859 ± 922 individuals.m⁻², respectively.

- Unconsolidated reef: Dense shell hash and mud with scattered oysters (97 \pm 35 adult individuals.m⁻², 652 \pm 912 spat individuals.m⁻²). No vertical relief (0 m; oysters directly on the sediment).

- Restored reef: Eight higher vertical relief (0.3 m) reef mounds (600 m² each)
constructed in summer 2011 with a crushed concrete base and reclaimed oyster shell veneer.
Oyster densities were 280 ± 164 and 1462 ± 1595 individuals.m⁻² for adults and for spat,
respectively.

Sampling commenced in February 2012 (approximately six months post-reef
construction) and continued seasonally through September 2013 for a total of six sampling
periods (February 2012, June 2012, September 2012, March 2013, June 2013 and September
2013).

161 2.2 CONTINUOUS ENVIRONMENTAL DATA

Salinity, chlorophyll-*a* (chl-*a*, μ g L⁻¹) and wind speed (m s⁻¹) were collected near the study site (Copano Bay East station; 28.132 °N, 97.034 °W) as part of the National Estuarine Research Reserve System monitoring program (NERRS 2014). All data were collected for the entire study period, starting in July 2011 when reef construction began and continuing until October 2013 when the study was complete.

167 2.3 SAMPLING AND PREPARATION OF WATER CHLOROPHYLL-*A* AND COMPOSITE FOOD SOURCE168 SAMPLES

Bottom water samples were collected just above the sediment-water interface at each reef type using a horizontal van Dorn water sampler for quantification of chl-*a* and stable isotope analysis (δ^{13} C, δ^{15} N) of SPOM. Water was filtered on Whatman GF/F glass fiber filters (0.7 µm porosity) to collect SPOM for chl-*a* analysis. Filters were stored at -20 °C until analysis. SPOM

173 samples for stable isotope analyses were sieved on a 300- μ m screen to eliminate large detrital 174 particles and zooplankton, and then filtered on two precombusted Whatman GF/F glass fiber 175 filters (0.7 μ m porosity). Filters were frozen at -20 °C and freeze-dried. Carbonates were 176 removed from filters for δ^{13} C and %C analyses by contact with HCl fumes in a vacuum-enclosed 177 system. Untreated freeze-dried filters were used for δ^{15} N and %N analyses.

Three cylindrical sediment cores (37.4 cm^2) were collected by divers from each reef type 178 for stable isotope analysis (δ^{13} C, δ^{15} N) of SSOM. Cores were collected and handled with 179 180 minimal disturbance and the top 2 cm were sliced and stored at -20 °C until processing. Samples were thawed and sieved wet on a 500 μ m mesh screen to eliminate macrofauna, shell pieces and 181 182 large detrital particles. Sieved sediment was freeze-dried and ground using a mortar and pestle. Carbonates were removed from sediment for δ^{13} C and %C analyses using 2 mol L⁻¹ HCl. HCl 183 was added drop-by-drop until cessation of bubbling. Samples were then dried at 60 °C using a 184 185 dry block heater under air flow. Dried samples were re-homogenized into ultrapure water using an ultrasonic bath, freeze-dried and ground again. Untreated ground sediment was used for $\delta^{15}N$ 186 and %N analyses. 187

188 2.4 SAMPLING AND PREPARATION OF OYSTERS

Oysters were collected from each reef type by divers and stored on ice for transport to the laboratory. They were cleaned of epibionts and kept alive for up to 36 h in filtered seawater to allow for gut content evacuation (Dubois et al. 2007). Oysters were frozen at -20 °C and then dissected to collect digestive gland material. We aimed to collect three spat oyster (≤ 25 mm shell height) samples and three adult oyster (> 25 mm) samples from each reef type during each sampling event. Spat oysters were pooled in groups (2-4 individuals) when possible to obtain enough material for stable isotope analysis. Low oyster density in the unconsolidated reef often

limited the number of oysters collected. In total, 52 adult oysters and 44 spat groups (111

197 individual spat) were analyzed, ranging in size from 25.4 to 76.8 mm (47.2 \pm 13.4 mm, mean \pm

standard deviation (SD)) and from 11.5 to 25.0 mm (18.4 ± 3.4 mm), respectively.

199 Digestive gland samples were freeze-dried and ground to a homogenous powder. 200 Digestive glands contain high amounts of lipids, which are depleted in ¹³C relative to other 201 tissues due to the different biochemical pathways involved in their synthesis (DeNiro and 202 Epstein 1977). As a result, the δ^{13} C value of a raw sample does not only reflect the food 203 resources of a consumer. Thus, lipids were extracted from samples for δ^{13} C and %C analyses 204 using two successive extractions with cyclohexane. Samples were then dried at 45 °C and 205 ground again. Untreated ground samples were used for δ^{15} N and %N analyses.

206 2.5 Chlorophyll-*A* and stable isotope ratio measurements

207 Chl-a was extracted from filters overnight using a non-acidification technique and read 208 on a Turner Trilogy fluorometer (Turner Designs, Sunnyvale, USA) (Welschmeyer 1994, EPA method 445.0). Elemental and stable isotope compositions were determined using an elemental 209 analyzer (Flash EA 1112, Thermo Scientific, Milan, Italy) coupled to an isotope ratio mass 210 spectrometer (Delta V Advantage with a Conflo IV interface, Thermo Scientific, Bremen, 211 Germany). Analyses were conducted at the Littoral, Environment and Societies Joint Research 212 Unit stable isotope facility (University of La Rochelle, France). Results are expressed in δ 213 notation as deviations from standards (Vienna Pee Dee Belemnite for δ^{13} C and N₂ in air for δ^{15} N) 214 following the formula: $\delta^{13}C$ or $\delta^{15}N = [(R_{sample}/R_{standard}) - 1] \times 10^3$, where R is ${}^{13}C/{}^{12}C$ or ${}^{15}N/{}^{14}N$. 215 Calibration was done using reference materials (USGS-24, IAEA-CH6, -600 for carbon; IAEA-216 N2, -NO-3, -600 for nitrogen). Analytical precision was < 0.15 % based on the analyses of 217 acetanilide (Thermo Scientific) used as laboratory internal standard. 218

219 2.6 Statistical analyses

Statistical analyses were performed using R 2.12.2 (R Foundation for Statistical 220 Computing 2011). Comparisons between measured parameters among sampling dates and reef 221 types were conducted using nonparametric procedures (Zar 2010). Kruskal-Wallis tests 222 (kruskal.test) were used to compare isotope compositions and C:N ratios of SPOM, SSOM and 223 ovsters, and bottom water chl-a concentrations and C:chl-a ratios of SPOM. Kruskal–Wallis 224 225 tests were followed by multiple comparisons of means (*kruskalmc* in *R pgirmess* package) 226 (Giraudoux 2011). Isotope compositions of oysters and food sources were compared considering trophic fractionation factors of 0.3 ± 1.3 % (mean \pm SD) for δ^{13} C values and of 2.3 ± 1.6 % for 227 δ^{15} N values (Vander Zanden and Rasmussen 2001). Theoretical oyster food resource use was 228 computed by subtracting the trophic fractionation values from observed δ^{13} C and δ^{15} N values of 229 oysters to correct for fractionation. Contributions of SPOM and SSOM as food resources to 230 231 oysters were estimated by solving mixing models within a Bayesian framework (siarmcmcdirichlety4 in R siar package) (Parnell et al. 2010, Parnell and Jackson 2014). Models 232 were run for 500,000 iterations and the first 50,000 iterations were discarded. Means and 233 credibility intervals (CI) of 0.95 and 0.25 were computed and displayed on figures. 234 235 3. Results 236 **3.1 CONTINUOUS ENVIRONMENTAL PARAMETERS** Monthly average salinity ranged from 25.2 to 38.8, with lowest values observed during 237

May 2012 (Fig. 2). Monthly chl-*a* concentrations ranged from 2.0 to 16.1 μ g L⁻¹, and varied throughout the study (Fig. 2). Monthly average wind speed ranged from 1.1 to 7.3 m s⁻¹, with

- lowest values observed from August through October 2012 (Fig. 2).

241 3.2 Composition of Potential food resources

| 242 | Mean bottom water chl-a concentrations of collected SPOM samples ranged from 0.7 \pm |
|-----|---|
| 243 | 0.6 to $4.3 \pm 1.3 \ \mu g \ L^{-1}$ (mean $\pm SD$) (Fig. 2). Chl- <i>a</i> concentrations were similar among reef types |
| 244 | ($P > 0.173$), except during March 2013, when chl- <i>a</i> was higher at the unconsolidated reef |
| 245 | compared to the restored reef ($P = 0.024$). Across all reef types, chl- <i>a</i> concentration was lower |
| 246 | in February 2012 than in September 2012 and 2013 ($P < 0.001$). Mean C:chl- <i>a</i> ratios ranged |
| 247 | from 58.8 ± 9.2 to 328.8 ± 67.5 ; ratios were similar among reef types and were greater in |
| 248 | February 2012 than in September 2012 and 2013 (<i>P</i> = 0.023; Fig. 3). |
| 249 | Mean δ^{13} C values of SPOM ranged from -24.8 ± 0.3 % (mean ± SD) to -23.5 ± 0.3 %; |
| 250 | mean δ^{15} N values ranged from 7.3 ± 0.3 to 8.7 ± 0.2 ‰ (Fig. 4). Mean δ^{13} C and δ^{15} N values of |
| 251 | SPOM were similar among reef types. δ^{13} C values were different among some sampling periods |
| 252 | (<i>P</i> = 0.016), but no clear temporal patterns were evident for δ^{13} C values. The δ^{15} N values were |
| 253 | generally more enriched during September sampling periods ($P = 0.010$). Mean C:N ratios of |
| 254 | SPOM ranged from 6.4 ± 0.5 to 12.8 ± 2.5 and were similar among reef types. C:N ratios of |
| 255 | SPOM were lowest during September sampling periods in both 2012 and 2013 compared to all |
| 256 | other sampling periods (Fig. 3), though not significantly different ($P = 0.086$). |
| 257 | Mean δ^{13} C values of SSOM ranged from -22.6 ± 1.0 to -20.1 ± 0.2 % (mean ± SD) (Fig. |
| 258 | 4), and followed a general pattern of enrichment over time at all reef types ($P < 0.001$). Mean |
| 259 | δ^{15} N values ranged from 7.5 ± 0.3 to 8.5 ± 0.5 % (Fig. 4). Mean δ^{13} C and δ^{15} N values of SSOM |
| 260 | were similar among reef types ($P > 0.066$ and $P > 0.099$, respectively). Mean C:N ratios of |
| 261 | SSOM ranged from 7.7 ± 0.5 to 10.6 ± 1.3 and were similar among reef types ($P > 0.077$). δ^{15} N |
| 262 | values and C:N ratios were similar among sampling periods ($P = 0.186$ and $P = 0.228$, |
| 263 | respectively). |

264 3.3 STABLE ISOTOPE COMPOSITION OF OYSTERS

| 265 | Oyster mean δ^{13} C values ranged from -23.3 ± 0.2 to -20.1 ± 0.5 % (mean ± SD) and |
|-----|---|
| 266 | mean $\delta^{15}N$ values ranged from 8.2 ± < 0.1 to 11.1 ± 0.6 % (Fig. 4). Mean $\delta^{13}C$ and $\delta^{15}N$ values |
| 267 | were similar among reef types except in the first sampling period (February 2012); oysters from |
| 268 | the natural reef were more enriched in both ¹³ C ($P = 0.010$) and ¹⁵ N ($P = 0.044$) than oysters at |
| 269 | unconsolidated reef. Overall, spat and adult oysters had similar δ^{13} C values (<i>P</i> = 0.478), while |
| 270 | spat were more ¹⁵ N-enriched (10.2 ± 1.0 %) compared to adult oysters (9.5 ± 1.0 %) ($P =$ |
| 271 | 0.003). Oysters were more enriched in both ¹³ C and ¹⁵ N during June and September 2013 than |
| 272 | during other months ($P < 0.001$ for both δ^{13} C and δ^{15} N values). Theoretical oyster food resource |
| 273 | use, corrected for trophic fractionation by subtracting 0.3 $\%$ from $\delta^{13}C$ values and 2.3 $\%$ from |
| 274 | δ^{15} N values of oysters, overlaps predominantly with SSOM food resources (Fig. 4). |
| 275 | 3.4 MIXING MODEL ESTIMATIONS OF FOOD SOURCE CONTRIBUTIONS TO OYSTERS |
| 276 | Mixing model results revealed a general trend of decreasing SPOM and increasing SSOM |
| 277 | food resource contributions to oysters over time, among all reef types (Fig. 5). In February 2012, |
| 278 | SPOM contributed from 20 to 98 % (95% CI range) and SSOM contributed from 2 to 80 %. By |
| 279 | the end of the study (September 2013), the relative importance of food sources to oysters |
| 280 | switched with SPOM contributions ranging from 2 to 54 $\%$ and SSOM from 47 to 98 $\%.$ |
| 281 | Slightly higher proportions of SPOM contribution was observed in September 2012 than in |
| 282 | adjacent sampling dates. The same temporal patterns of food source contributions were observed |
| 283 | among the three reef types. |

284 **4. Discussion**

285 4.1 ORIGIN AND COMPOSITION OF FOOD RESOURCES

Average δ^{13} C values of SPOM were more depleted (-24.2 ± 0.6 %) than SSOM (-21.2 ± 286 0.8 ‰) throughout the study (mean \pm SD). The range of δ^{13} C values observed from SPOM (-287 25.1 to -23.6 %) fall within ranges typical of river-estuarine phytoplankton (-30 to -24 %), and 288 C₃ marsh (-26 to -23 ‰) and terrestrial (-30 to -23 ‰) plants (Fry and Sherr 1984). Recent work 289 (2010-2011) in the Mission-Aransas Estuary determined δ^{13} C values for C₃ marsh plants (-29 to -290 291 27 %) and estuarine SPOM (-27 to -21 %) (Lebreton et al. 2016). C:N ratios less than 20 and C:chl-a ratios less than 200 indicate a dominant influence of fresh organic matter (e.g., fresh 292 293 phytoplankton) rather than degraded matter (e.g., highly decayed phytoplankton, terrestrial or saltmarsh plant detritus) (Cifuentes et al. 1988, Leal et al. 2008). Considered together, the δ^{13} C 294 values of SPOM, and low C:N and C:chl-a ratios observed throughout much of the study indicate 295 296 a dominant influence of autochthonous phytoplankton production on SPOM (Fry and Sherr 297 1984, Lebreton et al. 2016).

Mean δ^{13} C values of SSOM (ranging from -22.6 to -20.1 %) indicate potential influences 298 from a variety of sources: trapped phytoplankton, benthic microalgae, as well as a mixture of 299 material of marsh (C₄ plants) and of riverine and terrestrial (C₃ plants) origin (Fry and Sherr 300 1984). C:N ratios remain low (generally < 10), indicative of fresh organic matter (Kang et al. 301 2003) and thus indicate that marsh, riverine and terrestrial sources of organic matter play only a 302 minor role in the composition of SSOM. We infer that SSOM is largely composed of fresh 303 304 organic matter sources, such as trapped phytoplankton (-24 %; Fry and Sherr 1984), but also benthic microalgae, which are more ¹³C-enriched, with δ^{13} C values of -17 ‰ in Fry and Sherr 305 (1984), and ranging from -21.7 to -12.0 % in Lebreton et al. (2016). The distinct ranges of δ^{13} C 306

307 values observed for SSOM and SPOM further indicate that SSOM is not only composed of

308 settled SPOM, but likely contains a large amount of benthic microalgae.

309 4.2 Spatio-temporal variations of spom and ssom compositions

At the spatial scale, the origin and composition of SPOM and of SSOM were generally similar among reef types (natural, restored, unconsolidated), demonstrating that differences of structure (i.e., vertical relief, bottom composition, oyster biomass and densities) among reefs do not affect the quantity and the quality of the food sources available for oysters at the local scale (i.e., reef scale). Rezek et al. (2017) demonstrated that changes of food web structure in oyster reefs under development are related to changes of benthic macrofauna community but not to changes in food source uses by macrofauna.

317 At the temporal scale, SPOM was of higher quality in late summer (i.e., September) as 318 indicated by the lower C:N and C:chl-a ratios and the higher chl-a concentrations in the bottom 319 water. This indicates that the proportion of autochthonous phytoplankton in the bottom water SPOM is higher during this period. Local primary productivity in the water column was 320 321 probably enhanced during this period due to seasonally low wind speeds (NCDC 2017), leading to lower resuspension of sediment-trapped organic and to higher penetration of light into the 322 water column. Lower primary productivity during winter months, and thus less fresh organic 323 matter in the SPOM pool, explains the higher C:chl-a ratios observed among all reef types during 324 February 2012. The lack of clear relationship between chl-a concentrations measured in the 325 bottom water (i.e., our samples) and in the water column (i.e., NERRS measurements) indicates 326 327 that SPOM composition in bottom water can differ from that in the water column, highlighting the importance of sampling SPOM as close as possible to the oyster reefs. 328

In the sediment, a temporal increase of δ^{13} C values was observed for SSOM, with values 329 most depleted in ¹³C during February 2012 and becoming more enriched through September 330 2013 among all reef types. This pattern likely reflects an increasing influence of benthic 331 microalgae, which are enriched in ¹³C, and/or a decreasing influence of more ¹³C-depleted 332 organic matter (i.e., trapped phytoplankton) on SSOM across the period. As this development is 333 observed at the scale of the three reef types, it can be related to a large change, at the scale of the 334 335 ecosystem, or to a change at a local scale (i.e., one reef type) affecting the functioning of the 336 whole ecosystem. We therefore suggest two hypotheses to explain this change: (1) At the scale of the ecosystem, a gradual decrease of SPOM trapping into the sediment. Nevertheless, such a 337 338 trend would probably be linked with a similar trend of some abiotic parameters, such as wind velocity or turbidity, which is not the case, making this hypothesis less likely. (2) At a local 339 scale, increasing oyster biomass at the restored reef may have enhanced growth of 340 341 microphytobenthos through feedback effects related to oyster biodeposits. As suspensionfeeders, oysters can have a strong influence on the degree of benthic-pelagic coupling within a 342 343 system (Dame 2012, Newell 2004, Prins et al. 1998). Oysters remove suspended particles from the water column, including those resuspended from the sediment, and deposit feces and 344 pseudofeces, or biodeposits, onto the sediment, thereby linking organic matter and nutrient flows 345 between pelagic and benthic habitats (Hsieh et al. 2000, Newell 2004, Prins et al. 1998, Quan et 346 al. 2012). Feces and pseudofeces excreted by oysters may stimulate benthic production as this 347 organic matter is remineralized by bacteria and releases nutrients that can be used by 348 349 microphytobenthos (Miller et al. 1996, Prins et al. 1998). Indeed, after the introduction and intensification of oyster culture, benthic primary production has been shown to increase 350 (Leguerrier et al. 2004), as has enrichment of sediments due to biodeposits, resulting in high 351

sediment chl-*a* concentrations (Castel et al. 1989). In our study, the restoration of the reef does
affect the SSOM composition in this reef type, as well as in the adjacent habitats (i.e., natural
and unconsolidated reefs) highlighting the importance of the connectivity between these different
habitats at the ecosystem scale. This highlights the potential benefits of reef restoration, not only
for the oyster reef itself, but also for the adjacent systems.

357 4.3 Oyster utilization of water column and sediment food resources

The range of δ^{13} C values observed in oysters overlaps mostly with SSOM δ^{13} C values, 358 359 indicating that oysters use food resources from both the SPOM and SSOM pools, regardless of reef type. The strong role of SSOM as a food resource for oysters is confirmed by the similar 360 shift of δ^{13} C values between oysters and SSOM. As SSOM becomes more ¹³C-enriched at all 361 reef types over the duration of the study-probably due to an increase of benthic microalgae into 362 SSOM composition-so do oysters. Additionally, mixing model results indicate that the 363 364 contribution of SSOM to ovsters increases over the duration of the study across all reef types, and that proportions of SSOM as a food source for oysters are similar among the three reef types. 365 366 SSOM becomes accessible to oysters via resuspension during water movement (Kang et al. 2003, Miller et al. 1996). Minimal tidal ranges and freshwater inflows, coupled with consistently 367 strong winds in the region, indicate that wind-induced waves are the predominant forcing factor 368 causing resuspension in this system (Armstrong 1987, Evans and Morehead Palmer 2012). 369 The restored reef in this study was constructed with relatively high vertical relief (0.3 m). 370 Oyster reef restoration as a whole has become increasingly focused on providing vertical 371 structure (i.e., >0.2 m above the bottom) to minimize sedimentation and improve temporal 372 sustainability (Powers et al. 2009, Schulte et al. 2009). The vertical biogenic structure interacts 373 with hydrodynamic boundary-layer flow over the bottom (Butman et al. 1994, Green et al. 1998), 374

375 influencing resuspension and food availability (Fréchette et al. 1989, Grant et al. 1990, Quan et 376 al. 2012, Wildish and Kristmanson 1984), larval settlement (Hills et al. 1999, Lapointe and Bourget 1999), and secondary production (Lenihan 1999). In contrast, the unconsolidated and 377 natural reefs in the current study had relatively low vertical relief (0m; < 0.1 m, respectively). 378 Although it could be predicted that SSOM food resources are more accessible to suspension 379 feeders lower in the water column (Hsieh et al. 2000), results indicate food resource use was 380 381 similar between reef types, even during periods of low wind. Temporal trends observed in 382 SSOM of increased δ^{13} C values over time are reflected in oyster δ^{13} C values across reef types, indicating that small-scale differences in vertical relief are not sufficient to limit consumption of 383 384 SSOM in Copano Bay. This highlights the very strong bentho-pelagic coupling in this ecosystem, probably due to shallow depths as well as strong and consistent winds. Further 385 research is warranted to examine whether oysters on higher-vertical relief reefs ($\geq 1-2$ m; e.g. 386 387 Dunn et al. 2014, Puckett and Eggleston 2012) or from reefs restored in deeper or less wind driven systems become decoupled from SSOM food resources compared to lower-vertical relief 388 389 counterparts.

390 4.4 TEMPORAL INCREASE OF SSOM AS A FOOD RESOURCE FOR OYSTERS

There are two probable explanations for the consistent temporal change in relative contribution of SPOM and SSOM food resources to oysters among reef-types: (1) a decrease of the role of SPOM, potentially connected to changes in environmental drivers, and/or (2) an increase of the role of SSOM, linked to feedback effects of oysters on SSOM composition and availability. Composition of organic matter resources can be influenced by various environmental factors, and may also affect the assimilation of food resources by consumers (Fry 2006, Michener and Kaufman 2007).

398 In the Copano-Aransas estuary, SPOM composition is highly connected to environmental drivers (Lebreton et al. 2016, Mooney and McClelland 2012, Reyna et al. 2017). The higher 399 concentrations in chl-a and better quality (i.e., higher C:N and C:chl-a ratios) of SPOM in late 400 summer, likely related to lower wind speeds, may affect its use by oysters. Indeed, in September 401 2012, the proportions of SPOM as a food source to oysters is slightly higher than the general 402 observed pattern. Nevertheless, this slight discrepancy is relatively minor compared to the 403 404 general temporal pattern observed, which shows a general increase of the SSOM as a food source 405 over the course of the study, highlighting the importance of benthic food resources into the diet of oysters (Dame 2012, Riera and Richard 1996). 406

407 The alternative explanation for the higher use of SSOM over time is that increasing oyster biomass at the restored reef enhanced growth of microphytobenthos through feedback 408 effects related to oyster biodeposits (see section 4.2). Enhanced benthic production can provide 409 410 important food sources to suspension-feeders such as ovsters and other benthic organisms (Leal et al. 2008, Miller et al. 1996, Ubertini et al. 2012), particularly in shallow systems where SSOM 411 412 is resuspended on a regular basis. In addition, South Texas embayments are characterized by very low freshwater inflows, limiting nutrient inputs (Bianchi et al. 1999), and therefore 413 production of phytoplankton (Mooney and McClelland 2012). As a result, the feedback of 414 oysters on benthic primary production may be critical in supporting oysters and reef-associated 415 fauna (Leguerrier et al. 2004, Prins et al. 1998, Ubertini et al. 2012). Drought conditions 416 persisted throughout the study period, probably limiting phytoplankton production (Reyna et al. 417 418 2017). Thus, the increasing influence of SSOM on oysters over the study period may be related 419 to observed increases in oyster biomass following reef restoration, coincident with prolonged drought (Blomberg 2015). 420

421 **5.** Conclusion

Our study demonstrated that, in a shallow and wind-driven ecosystem like the Mission-422 Aransas Estuary, differences in oyster reef structure do not affect the composition of food 423 resources available to oysters and their use by oysters. This is probably related to the regular 424 resuspension of the organic matter stored in the sediment and to the strong connectivity between 425 the different studied reef types. Similar studies should be carried out in ecosystems 426 427 characterized by different physical properties (e.g., deeper systems, higher stratification) to 428 determine how these physical properties can affect flows of organic matter in natural and restored oyster reefs. In terms of restoration ecology, it may also be of strong interest to 429 430 determine how vertical relief is affecting availability of the food resources from the benthos.

Another major conclusion of our study is that oysters rely on large amounts of sediment organic matter as food resources, highlighting the importance of bentho-pelagic coupling in oyster reefs, and supporting the positive feedback role of oyster reefs on the production of benthic microalgae and/or trapping of organic matter. Better determination of the composition of sediment organic matter in and around oyster reefs would provide significant knowledge toward a better understanding of the role of this material in oyster reefs, and flows of organic matter in these habitats.

438

439 Acknowledgements

B.N.B. was supported by the National Oceanic and Atmospheric Administration, Office
of Education Educational Partnership Program (Award NA11SEC4810001). The contents of this
manuscript are solely the responsibility of the award recipient and do not necessarily represent
the official views of the U.S. Department of Commerce, National Oceanic and Atmospheric

Administration. Research trip of B.N.B. at the UMR LIENSs and stable isotope analyses were
funded in the framework of the "CNRS research and higher education chair" granted to B.L.
Reef construction was funded by the Gulf of Mexico Foundation-NOAA Community Based
Restoration Partnership (Award GCRP#10-01). We thank Texas A&M University – Corpus
Christi and the Harte Research Institute for support, and the many volunteers who assisted in the
field and lab. We thank three anonymous reviewers whose comments have helped improve this
manuscript. We declare no competing interests.

451

452 Author Contributions

453 Conceived and designed the experiments: BNB, BL, JBP. Performed the experiments: BNB, BL,

454 TAP, GG, JBP. Analyzed the data: BNB, BL, GG. Contributed reagents/materials/analysis tools:

455 BL, JBP, PAM. Wrote the paper: BNB. Extensive editorial comments, insight into results: BL,

456 TAP, GG, JBP, PAM.

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653 **Figure Captions:**

654

Figure 1. Study area. (a) Location of study site within the Mission-Aransas Estuary, Texas,USA. (b) Extent of sampling area among three reef types.

657

Figure 2. Continuous environmental observations over the course of the study (July 2011

through October 2013): monthly average wind speed (m s^{-1}), salinity and monthly grab samples

of chlorophyll-a (µg L⁻¹) from the Mission-Aransas National Estuarine Research Reserve

661 (MANERR) Copano Bay East station. Bottom water chlorophyll-a (µg L⁻¹) samples are

662 indicated with dots during each sampling period. Reef construction is indicated with the dark

663 gray bar; sampling periods are indicated with light shaded bars.

664

Figure 3. C:N and C:chl-*a* ratios (mean \pm SD) observed for suspended particulate organic matter sampled across the study area during each sampling period.

667

Figure 4. δ^{13} C and δ^{15} N values (mean ± SD) of suspended particulate organic matter (SPOM),

surface sediment organic matter (SSOM) and oysters (all sizes) during each sampling date. (a)

670 Unconsolidated reef. (b) Natural reef. (c) Restored reef. Grey areas represent the range of

671 theoretical oyster food source stable isotope composition accounting for trophic fractionation.

672

Figure 5. Seasonal comparison of contributions (%) of potential food resources (SPOM:

- 674 suspended particulate organic matter, SSOM: surface sediment organic matter) to oyster diets
- resulting from the mixing model SIAR. (a) Unconsolidated reef. (b) Natural reef. (c) Restored

reef. Mean, 0.25 and 0.95 credibility intervals are shown with horizontal bar, grey box andwhiskers, respectively.









