

1 **Techniques for improving estimates of maturity ogives in groundfish using**
2 **double-reads and measurement error models**

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23 **Abstract**

24 The reproductive output of a population depends upon physiological factors, including maturation
25 rates and fecundity –at-size and –at-age, as well as the rate at which post-maturation females fail
26 to spawn (i.e. skipped spawning). These rates are increasingly included in stock assessment
27 models, and are thought to change over time due to harvest and environmental factors. Thus, it is
28 important to accurately estimate maturation and skipped spawning rates while also including
29 information on imprecision. For this task, we developed a new double-read and measurement-error
30 modeling protocol for estimating maturity that is based on the use of multiple histological reads of
31 ovaries to account for reader error caused by poorly prepared slides, nuclear smear, and early yolk
32 development. Application to three U.S. West Coast groundfishes (Pacific hake *Merluccius*
33 *productus*, darkblotched rockfish *Sebastes crameri*, and canary rockfish *Sebastes pinniger*)
34 indicates that reader uncertainty is strongly predictive of reader error rates. Results also show
35 differences in rates of skipped spawning among species, which should be further investigated. We
36 recommend that future maturity studies record reader certainty, use models that incorporate
37 covariates into the analysis, and conduct an initial double reader analysis. If readers exhibit little
38 variation, then double reads may not be necessary. In addition, slide quality should also be
39 recorded, so that future studies do not confuse this with reader imprecision. This improved protocol
40 will assist in estimating life history, as well as environmental, and anthropogenic effects on
41 maturity.

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43 **Keywords:** maturity ogive; skipped spawning; histological error; reader certainty; groundfish;
44 measurement error model

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47 **1. Introduction**

48 Reproduction is a basic demographic process in population dynamics, and density-independent
49 and -dependent changes in reproductive success contribute to a large portion of variability in
50 marine populations. Timing of sexual maturity will vary among individuals according to individual
51 size, age, season, location, and other factors (Jørgensen and Fiksen, 2006). Maturity is a function
52 of length and age, and fast growing fish generally mature at much younger ages. Growth rates slow
53 during reproduction, with immature fish growing faster than mature fish, and increase following
54 skipped spawning events (Folkvord et al., 2014). These factors, paired with errors in correctly
55 identifying maturity stages, influence our ability to accurately estimate the probability of sexual
56 maturity as a function of length and/or age (termed the “maturity ogive”). In addition, since many
57 individuals do not spawn annually following initial maturation, the average skipped spawning rate
58 may be poorly estimated. Skipped spawning and reader error remain understudied and often go
59 unaccounted for in fisheries models (Rideout et al., 2000; Kennedy et al., 2011). Understanding
60 how shifting marine environments and anthropogenic impacts affect these life history processes
61 and our ability to accurately quantify them is vital to the accuracy of stock recruitment models
62 used in fisheries management (Burton, 1999; Kennedy et al., 2011).

63 In recent years, improvements in survey design and statistical methods have revolutionized
64 how other demographic processes are estimated. For example, estimation of growth curves now
65 uses methods that explicitly account for misreading of individual ages (e.g., Cope and Punt, 2007),
66 while estimates of natural mortality involve state-space modeling of individual tagged fishes (e.g.,
67 Schaub and Royle, 2013). Estimating reader error variability for aging fish is routinely based on
68 double reads of a single otolith by different individuals, along with validation methods that confirm
69 the accuracy of the results or indicate the degree of bias (Campana, 2001; Hamel, 2008; Punt et

70 al., 2008). Hierarchical models are used to process the results of double-reads, allowing error to
71 propagate and be accurately reported (Royle and Dorazio, 2008) when estimating emergent
72 biological process such as shifts in size- or age-at-maturity and skipped spawning. Research has
73 revealed maturity ogives shift over time, e.g. Atlantic cod (Olsen et al., 2004). Skipped spawning
74 and incidence of high intensity atresia (breakdown and reabsorption of vitellogenic oocytes) are
75 correlated with nutrient availability and temperature, factors that are highly variable over space
76 and time (Rideout et al., 2006). Including spatial and temporal variables in hierarchical maturity
77 models allows differences in sampling to be ‘controlled’ when estimating decadal changes in
78 maturity. Recent research has proposed using mixed-effects models or covariates to smooth time-
79 variation in the maturity ogive given sparse annual sampling of fish maturity (Cadigan et al., 2014;
80 Wright et al., 2011; Xu et al., 2015).

81 Reader variability is often calculated for age reading, but to the authors’ knowledge it has not
82 been incorporated into maturity models. In studies on age, re-aging or double reading otoliths is
83 considered important for data reliability (Hare, 2007). Maturity studies based on macroscopic
84 examination of ovaries found high variability in the determination of mature and immature fish
85 (Costa, 2009). Histological analysis allows for greater accuracy and insight into skipped spawning
86 events, but the effect of reader error on variability of microscopic determination of maturity states
87 is unclear. Accounting for variability in measurements of maturity (either using histological or
88 macroscopic methods) also allows analysts to more accurately characterize uncertainty regarding
89 estimates of the maturity ogive. Uncertainty in these maturity analyses can then be incorporated in
90 stock assessment models and resulting management advice (Stewart et al., 2013).

91 Here, we use a measurement error model to estimate maturity ogives for three species
92 inhabiting the California Current system: Pacific hake (*Merluccius productus*), darkblotched

93 rockfish (*Sebastes crameri*), and canary rockfish (*Sebastes pinniger*), and also incorporate a double
94 read analysis. These species were selected based upon the availability of samples, variations in life
95 history, and importance along the U.S. West Coast. The objectives of this study are to: (1) to
96 identify and reduce the main sources of reader error in estimating maturity of fish, (2) to add
97 uncertainty in maturity reads and reader precision to model analysis and (3) to select the best fit
98 maturity ogive model for each species. We noted that reader uncertainty was a strong predictor of
99 reader error rates, implying that reader certainty should be documented during histological
100 analysis.

101 **2. Methods**

102 *2.1 Study species*

103 The life history strategies for the three species studied varied greatly, but all are commercially
104 important either economically or as catch-limiting species along the West Coast. Pacific hake
105 occur from 25°N to 55°N on the West Coast of North America, and are typically found from
106 southern California waters to the Queen Charlotte Sound. The coastal Pacific hake stock is the
107 most abundant groundfish species in the California Current system. A study conducted from 1990-
108 92 visually assessed maturity for female Pacific hake, with length-at-50%-maturity estimated as
109 37.8 cm (Dorn and Saunders, 1997). Spawning reportedly takes place off the coast of California
110 from January to March, but more recent studies suggest spawning may occur in multiple batches
111 throughout the year (Taylor et al., 2015). It is uncertain whether batches spawned outside of the
112 winter spawning season are viable. Pacific hake are estimated to live up to 20 years of age (Hesler
113 and Alade, 2012).

114 Canary rockfish (*Sebastes pinniger*) are distributed from Baja California to the Gulf of Alaska
115 with the highest concentrations between British Columbia, Canada and central California to

116 Bodega Bay (Love et al., 2002; Miller and Lee, 1972). This species has an estimated lifespan of
117 75 - 95 years and a maximum female size of 61 cm; though, females are rarely observed over 30
118 years of age. Recent histological analysis of ovaries collected by the Oregon Department of Fish
119 and Wildlife (ODFW) and the West Coast groundfish bottom trawl survey (WCGBTS) estimated
120 the length-at-50%-maturity to be approximately 42 cm (Thorson and Wetzel, 2015). Canary
121 rockfish, like all *Sebastes* are viviparous, live bearing fish, and spawn in the winter months (Love,
122 1996; Thorson and Wetzel, 2015).

123 Darkblotched rockfish (*Sebastes crameri*) are found in the southeast Bering Sea and Aleutian
124 Islands to Santa Catalina Island, California, and are commercially important from Northern
125 California to the Canadian border. Darkblotched rockfish have an estimated maximum lifespan of
126 105 years and size of 58 cm (Gertseva and Thorson, 2014; Love, 1996). Length-at-50%-maturity
127 was recently assessed for darkblotched rockfish using ovaries collected during the 2011 – 2012
128 WCGBT survey, and found to be 30 cm (Frey et al., 2015). Similar to canary rockfish, they are
129 viviparous and spawning occurs during the winter months (Nichol and Pikitch, 1994; Frey et al.,
130 2015).

131 *2.2 Data collection and Histological Analysis*

132 Pacific hake, canary rockfish, and darkblotched rockfish were sampled during the Northwest
133 Fisheries Science Center's (NWFSC) annual fishery-independent bottom trawl survey off the U.S.
134 West Coast from May – October (see Bradburn et al., 2011 for sampling methods). A random
135 subsample of each species was selected from the catch at each site to determine sex, fork length
136 (cm), weight (kg), and age (yr). Age was subsequently determined from extracted otoliths using
137 the break-and-burn method (Beamish and Chilton, 1982). From 2009 - 2011, ovaries were
138 collected opportunistically from females with associated age samples. Since 2012, collections were

139 based on length bins to ensure a broad size range of fish for inclusion in maturity analysis. Ovaries
140 were stored in cloth sampling bags and stored in 10% neutral buffered formalin (sodium
141 bicarbonate). Pacific hake (n = 329), darkblotched rockfish (n = 307), and canary rockfish (n =
142 131) ovaries were histologically examined by two independent readers (R1 and R2). Four hake
143 samples were excluded from the model analysis due to missing ages.

144 Tissue samples from individual ovaries were embedded in paraffin, thin-sectioned to 4- μ m,
145 mounted on slides, and stained with hematoxylin and eosin (H&E) stain (Sheehan and Hrapchak,
146 1980). To determine maturity, each prepared ovary section was examined using a Leica DM1000
147 binocular microscope at 40x – 400x magnification, equipped with a Leica DFC295 camera and
148 imaging software (Leica Microsystems LAS EZ 4.0).

149 Ovaries with oocytes containing dark-stained vitellogenin yolk (eosin positive) and more
150 advanced stages (hydration and/or embryonic development) were classified as mature. We used
151 an oocyte development table to assess each sample following similar methods developed to stage
152 maturity for *Sebastes aleutianus* and *S. borealis* (McDermott, 1994), *S. crameri* (Nichol and
153 Pikitch, 1994), and *Pleurogrammus monoptergius* (McDermott and Lowe, 1997). Oocyte
154 developmental stage 4 and maturity stage 3 were defined as mature for all three species (Tables
155 S1 & S2). Samples estimated with over 25% oocytes in an atretic state were marked as high
156 intensity atresia following McDermott (1994). Oocytes exhibiting atresia were not used for
157 maturity determination; therefore, only non-atretic oocytes were used in maturity analysis. Ovary
158 samples were identified as mature solely by the presence of yolk and characteristics associated
159 with more advanced maturity stages, for the purposes of identifying fish that would be contributing
160 to the overall spawning biomass in any one year (Fig. S1 & S2). Determination of maturity was
161 independent of size and age. Post spawning samples (termed “spent”) were characterized by the

162 presence of a large number of post ovulatory follicles (POFs) and did not necessarily contain
163 yolked oocytes, but were also classified as mature. Resting fish, if encountered would be recorded
164 as mature, but the sample period (late May – October) does not overlap with the normal resting
165 periods for the species examined here.

166 *2.3 Double Reads and Certainty*

167 Two readers determined the maturity for each species examined. Reader 1 (R1) had three years
168 of experience conducting maturity analysis on multiple groundfish species, and recorded the
169 maturity status, oocyte stages present, the proportion of atretic oocytes, presence of (POFs), oocyte
170 diameters, and certainty of the maturity determination for each sample. Certainty of the
171 determination was recorded as a 1 (certain) or 0 (uncertain). Reader 2 (R2) had no experience
172 determining histological maturity, and was trained by R1 for two weeks previous to conducting
173 this research project. R2 recorded maturity and certainty level for each ovary sample. Both readers
174 examined each sample twice to confirm their results and level of certainty, but recorded only the
175 final reads. Results were not discussed prior to completion of data collection to ensure that any
176 disagreements in maturity assignment were statistically independent between the two readers.

177 *2.4 Model development*

178 We developed a model that synthesizes information from two readers while flexibly estimating
179 the maturity ogive and form of reader error. Only double maturity reads with both length and age
180 data were used in this model. Both readers followed the same protocols and are treated identically
181 in the model. A basic assumption of the model was that fish mature as a function of age and length
182 before reaching an asymptotic level. The asymptote is less than 1.0 if a fraction of fully mature
183 individuals skipped spawning. This definition of maturity thus includes both the process for first
184 reaching maturity and subsequent rates of skipped-spawning. Specifically, the probability that

185 ovary O_i for individual i is either mature ($O_i=1$) or immature ($O_i=0$) follows a Bernoulli
 186 distribution:

$$187 \quad O_i \sim \text{Bernoulli}(p_i) \quad (1)$$

188 where *Bernoulli* (p_i) is a Bernoulli distribution with estimated probability of maturity p_i for ovary
 189 i . The probability of maturity, p_i , is defined as:

$$190 \quad p_i = \Phi(\beta_0 + \beta_A A_i + \beta_L L_i + \beta_D D_i) \cdot \beta_{max} \quad (2)$$

191 with β_0 the intercept, β_A the effect of age on maturity, β_L the effect of length on maturity, β_{max} the
 192 estimated asymptotic maturity rate, β_D the effect of calendar date on maturity probability. Φ is the
 193 logit-function where $\Phi(X) = e^X / (1 + e^X)$ which transforms the linear predictors of the model to a
 194 response variable (i.e., expected proportion mature) that is bounded between 0 and 1, such that
 195 β_{max} is the maximum for p_i .

196 We then specified that the j -th read of ovary i , $\tilde{O}_{i,j}$, has some probability of incorrectly defining
 197 the maturity state:

$$198 \quad \tilde{O}_{i,j} \sim \text{Bernoulli}(O_i(1 - q_i) + (1 - O_i)q_i) \quad (3)$$

199 where q_i is the probability of reader error for ovary i :

$$200 \quad q_i = 0.5\Phi(\eta_0 + \eta_U U_{i,j}) \quad (4)$$

201 where η_0 is the intercept for reader-error probability, and η_U is the effect of reader uncertainty on
 202 reader error probability. Equation 4 uses 0.5 times the logit-transformation to ensure that the
 203 probability of reader error is bounded between 0 and 0.5, i.e., that reader error is never more likely
 204 to be wrong than right. Uncertainty $U_{i,j}$ is noted by reader j for ovary i and equals 1 whenever the
 205 reader expresses uncertainty about the state of maturity for a sample, and 0 otherwise.

206 Parameters for this model are estimated by maximum marginal likelihood:

$$207 \quad \hat{\boldsymbol{\beta}}, \hat{\boldsymbol{\eta}} = \operatorname{argmax}_{\boldsymbol{\beta}, \boldsymbol{\eta}} (L(\boldsymbol{\beta}, \boldsymbol{\eta} | \tilde{\boldsymbol{O}}_{i,j})) \quad (5)$$

208 where:

$$209 \quad L(\boldsymbol{\beta}, \boldsymbol{\eta} | \tilde{\boldsymbol{O}}) = \prod_{i=1}^{n_i} \sum_{O_{i=0}}^1 \left(\Pr[O_i | \boldsymbol{\beta}] \prod_{j=1}^{n_j} \Pr[\tilde{O}_{i,j} | O_i, \boldsymbol{\eta}] \right) \quad (6)$$

210 The model treats the ‘true’ state of each ovary as unknown, and thus estimates the state as a random
211 effect. This is done by ‘integrating across’ the true but unknown state for each ovary (Thorson and
212 Minto, 2015). This integral is computationally feasible because each ovary has only two possible
213 states (mature or immature), and hence the integral is accomplished using the summand term:

$$214 \quad \sum_{O_{i=0}}^1 \left(\Pr[O_i | \boldsymbol{\beta}] \prod_{j=1}^{n_j} \Pr[\tilde{O}_{i,j} | O_i, \boldsymbol{\eta}] \right) \quad (7)$$

215 where $\Pr[O_i | \boldsymbol{\beta}]$ is the prior probability of each state and $\Pr[\tilde{O}_{i,j} | O_i, \boldsymbol{\eta}]$ is the probability of each
216 observation conditional on that unknown state. Standard errors for each parameter are derived from
217 the maximum likelihood estimate, and for estimated parameters they were derived using the delta
218 method. These computations were done using the R statistical platform (R Development Core
219 Team, 2014). A Nelder-Mead nonlinear minimization algorithm (i.e., the *nlm* function in the
220 base package of the R statistical environment) was used to identify values of fixed effects that
221 maximize the likelihood function.

222 For each species, we fit the model including or excluding all combinations of age, length, date,
223 and an asymptotic maturity rate less than 1.0, as well as the effect of reader certainty on reader
224 error probability. The best fit model was selected using the Akaike Information Criterion (AIC)
225 (Burnham and Anderson, 2002).

226 **3. Results**

227 Estimated maturity ogives differ among species (Table 1). Most significantly, maturity
228 asymptotes were 1.0 (i.e., no skipped spawning) for canary rockfish, 0.96 (SE = 0.144) for
229 darkblotched, and 0.81 (SE = 0.131) for Pacific hake. No models with asymptotes equal to 1.0
230 were considered plausible models (i.e. $\Delta AIC < 10$) for Pacific hake. However, for darkblotched
231 rockfish a number of models with asymptotes of 1.0 exhibited ΔAIC s ranging from 3.24 to 7.18,
232 suggesting that evidence for skipped spawning was weak for this species. Species also differ in
233 terms of which factors contribute to the maturity ogive: maturity for hake is best predicted by age,
234 while maturity in darkblotched and canary rockfishes is best predicted by length. Similarly, the
235 date sampling occurred was positively and significantly related to maturity in both hake and
236 darkblotched rockfish, but was negative for canary rockfish (Table 1).

237 Additionally, there are large differences in reader error probabilities when reads are recorded
238 as “certain” or “uncertain” (Table 1). For all three species, reads marked as “certain” had <2%
239 probability of exhibiting disagreement between readers. By contrast, the model predicts that
240 maturity reads marked as “uncertain” had an 11% error probability for Pacific hake, 18% error
241 probability for darkblotched rockfish, and 27% error probability for canary rockfish (Table 1).
242 This implies that maturity readers are generally capable of identifying when their maturity
243 determination is less likely to be correct.

244 The readers had varying degrees of disagreement among the three species assessed. For
245 darkblotched rockfish, R1 and R2 disagreed on maturity assignments for 9 of 307 samples (3.2%
246 disagreement), when both readers were certain (n = 4), when only R2 was uncertain (n = 3), and
247 when both readers were uncertain (n = 2) (Table 2). Similarly, R1 and R2 disagreed on maturity
248 for Pacific hake 9 out of 329 samples (2.7 %), when both readers recorded certainty (n = 6), and

249 when R2 was uncertain (n = 3) (Table 2). For canary rockfish, the readers disagreed 11 out of 131
250 samples (8.4 %), when both readers were certain (n = 3), when both were uncertain (n = 2), and
251 when R2 was uncertain (n = 6) (Table 2). R1 (experienced) recorded uncertainty for < 2% of the
252 samples, while R2 (inexperienced) recorded uncertainty for about 11% of all maturity samples
253 read.

254 Inspection of the estimated maturity ogive revealed very different results for each species.
255 Hake rapidly matured between ages 1 and 2 (Fig. 1). This was reflected in the age at 10% maturity
256 ($A_{10} = \sim 1$ year), 50% maturity ($A_{50} = \sim 2$ years) and 90% maturity ($A_{90} = \sim 4 - 6$ years), with standard
257 errors increasing with age (Fig. 1). Darkblotched rockfish by contrast had a relatively smooth
258 maturity ogive as a function of length, with 50% maturity (L_{50}) occurring at ~ 30 cm, and a range
259 from 10% maturity (L_{10}) at 25 – 26 cm to 90% mature (L_{90}) at 34 – 35 cm. Canary rockfish had
260 the shallowest ogive slope, with a length at 50% maturity of 43 - 44 cm, a range of size at maturity
261 from 35 - 36 cm (L_{10}) to 51 - 53 cm (L_{90}) and greater imprecision, due to a small sample size.

262 **4. Discussion**

263 *4.1 Error in Maturity Reads*

264 It is important to identify all sources of potential error in maturity determinations. This is
265 especially true for fisheries managers comparing results for life history models based on historical
266 data with estimates using current length/age at maturity information. Methods typically vary
267 between historical and current maturity studies. Often older estimates were based on macroscopic
268 maturity determinations. Several studies have evaluated macroscopic versus histological maturity
269 determinations, and found variable agreement (Costa, 2009; McBride et al., 2013). Often
270 disagreements occur in fish that are shifting from immature to mature and in partially spent fish.
271 To the authors' knowledge, no previous studies of maturity in fish have evaluated histological

272 reader error. Our study shows that there is variation between readers, similar to the findings of
273 otolith reader comparison studies (e.g., Punt et al., 2008). If modeled correctly, data from multiple
274 readers may be able to more accurately predict the maturity state of fish and provide less uncertain
275 estimates of maturity status. We conclude that 1) level of certainty is an important component in
276 maturity studies conducted using one or more readers, 2) reader error may be attributed to slide
277 quality and if so should be recorded as such, 3) reader error may also be linked to samples in early
278 stages of vitellogenesis, when yolk is beginning to form, and 4) best fit models using selected
279 covariates with the lowest AIC value can provide more accurate estimates of length- or age-at-
280 50%-maturity for use in fisheries management.

281 Three primary explanations for variation and uncertainty between readers in maturity analysis
282 include: poor slide quality, nuclear smear, and determination of early yolk formation. Poor slides
283 (Fig. 2a) often result when the stain is either too dark or light, if contamination from other fish
284 samples occurs, and if ovarian tissue is folded during thin sectioning or mounting. When dull
285 blades are used to section an ovary, the nucleus can be smeared across portions of the oocyte. We
286 define this as nuclear smear (Fig. 2b) and note that it is problematic if cell structures critical for
287 maturity determination are obscured. Finally, determining maturity is difficult for some fish with
288 oocyte development stages III and IV, since very few yolk droplets may be present (Fig. 2c). When
289 yolk is present in only one or two oocytes, a reader may miss this early stage of development. An
290 experienced reader may be able to recognize other characteristics that are associated with maturing
291 fish ovaries (larger oocytes, thicker ovarian wall tissue, etc.), which could explain some of the
292 variation we observed among the two readers.

293 Our results show that while skill level is important when estimating uncertainty, the data are
294 valuable even with readers of different skill, provided each reader records their level of certainty.

295 R1, with greater experience, marked more reads as “certain” than R2 and both readers produced
296 consistently agreeable reads on slides marked as “certain”. By incorporating uncertainty in model
297 results, readers with variable levels of experience produce valuable data for determining maturity
298 ogives.

299 *4.2 Suggested Future Research Design*

300 Given the results of this study, we suggest that future research include: documentation of reader
301 certainty/uncertainty and its relationship to slide quality, double reads by a single reader or two
302 independent readers when beginning a study on a new species, or if there is high variation between
303 two readers, notes on slide quality and other factors used in estimating maturity status, estimates
304 of atresia, and seasonal variation in sample collections. This study highlights the importance of
305 recording both uncertainty and why a sample was defined as uncertain. If a maturity determination
306 is uncertain because of poor slide quality, then it should not be used in reader imprecision
307 estimates. We recommend that double reads be conducted on either all or a subset of samples to
308 produce requisite data for comparisons between reads. In addition, reader uncertainty could vary
309 as improvements are made in maturity sampling. Reader uncertainty and variation among two
310 readers’ maturity analysis should also be evaluated for samples collected in and out of the
311 spawning season. The authors acknowledge that double reads may be time consuming, and
312 providing up to date information from one reader is higher priority than from two readers.
313 However, conducting double reader analysis when a new reader is used will standardize techniques
314 for maturity determinations. Our study assumed both readers were equally able to identify if fish
315 were mature. Alternative models could be developed where one reader is assumed correct but the
316 other reader’s results are variable. Information and notes that indicate any difficulty in assigning
317 maturity to a sample would be a valuable addition for analyses. Estimating the proportion of

318 oocytes in atresia for each sample should be recorded, so that mass atresia events can be accounted
319 for.

320 Our results show that calendar date was positively associated with the maturity for
321 darkblotched rockfish and Pacific hake, but negatively related to canary rockfish. We speculate
322 that differences in maturity within a given season are caused by both an increase in the proportion
323 of fishes that have matured over time, and a change in the proportion of atresia. However, we
324 conclude that the effects of sampling seasonality are species dependent, and should be investigated
325 at the species level. When targeting the spawning season for a given species is not possible, we
326 recommend that date be included as a covariate in future models. Our samples were collected
327 primarily from May to October, outside of the observed spawning season for canary and
328 darkblotched rockfishes (January to March). Identifying the presence of early yolk is essential to
329 predicting sexual maturity outside of the spawning season, the higher level of uncertainty from the
330 two readers for canary rockfish (8.4%) may be related to when sampling occurred, since some
331 samples were collected within a few months following the spawning season. Several of the mature
332 ovaries collected in late May were in the early vitellogenesis stage, when it is more difficult to see
333 early yolk development. Additionally, we have noted that the observed resting/recovering period
334 varies greatly among the studied species. An in-depth histological analysis of ovary development
335 throughout the year would likely enhance the accuracy of the maturity estimates predicted by the
336 model. Previous research indicates temporal variation in fish reproduction, including onset of
337 sexual maturity and spawning events (Lowerre-Barbieri et al., 2011). In particular, interannual
338 changes in skipped spawning imply that spawning output varies among years, and could contribute
339 to observed variability in recruitment. Including skipped spawning events in stock-recruitment
340 models is critical, but identifying this may not be possible throughout the entire annual

341 reproductive period (Jørgensen et al., 2006). Recent advances in state-space modeling of
342 maturation schedules may be useful when accounting for interannual variability in skipped-
343 spawning rates (Cadigan et al. 2014; Xu et al. 2015).

344 *4.3 Management Implications*

345 The U.S. West Coast groundfish fishery supports an industry with more than 90 species of fish
346 in Washington, Oregon, and California (Pacific States Marine Fisheries Commission, 2012).
347 Current management regulations are based on stock assessments that include catch and landings
348 data, age and length records, and recruitment estimates, among many other model parameters. Due
349 to the complex life histories of many of these fishes, it is also important to consider the
350 reproductive physiology and strategies of these species. Understanding the ages and/or lengths
351 over which a stock's members transition from immature to mature status is fundamental to
352 identifying the proportion of the stock that can be sustainably harvested in a given year.
353 Histological maturity determinations provide an accurate estimate for use in stock recruitment
354 relationships, as well as provide information on abortive maturation and skipped spawning rates,
355 spawning output, and estimated fecundity. Failing to include skipped spawning events in future
356 recruitment estimates may lead to an overestimation of the actual spawning output (Rideout et al.,
357 2005), and may also contribute to the substantial variation seen in spawning output and
358 reproductive success for marine species (Morgan et al., 2011; Thorson et al., 2014). Furthermore,
359 histological data on maturation and atresia will allow scientists to detect changes in age- and
360 length-at-50%-maturity over time and across varying environmental conditions, and allow
361 fisheries managers to make decisions based on these changes.

362 Evolving life traits in response to low nutrient availability is thought to be species-specific,
363 and each species in this study varies greatly in life history. Skipped spawning is theorized to be

364 unlikely for species that are short lived, because the likelihood of surviving many spawning
365 seasons is low (Rideout et al., 2005). Heavily exploited species with longer life spans and slower
366 growth, such as canary and darkblotched rockfishes, are of particular concern to fisheries
367 management. Studies show these long-lived species are already more vulnerable to
368 overexploitation and less able to compensate for warming ocean temperatures; thus a shift in
369 bioenergetics may compromise reproductive success (Perry et al., 2005). Based on model
370 estimates, we saw skipped spawning rate of ~ 19 % for Pacific hake and ~ 4% for darkblotched
371 rockfish. Other studies found rates ranging from 2% to 35% (Jørgensen et al., 2006). The best fit
372 model for canary rockfish (based on AIC values), had an asymptote of 1.0, equivalent to no skipped
373 spawning. However, plausible models for canary rockfish (AIC < 10) suggest that skipped
374 spawning rates as high as 14% may occur. This is further apparent in the maturity ogive. The
375 relatively small sample size for this species coupled with the low number of large individuals may
376 have influenced these results. In addition, canary and darkblotched rockfishes reportedly spawn
377 during the winter months. Since samples were collected from May – October this would not be the
378 best time to estimate skipped spawning rates. Additional samples closer to the actual spawning
379 season are needed to confirm the presence or absence of skipped spawning.

380 Short lived species, such as Pacific hake, are unlikely candidates for large skipped spawning
381 events (Rideout et al., 2005). Hake are batch spawners that reportedly spawn during the winter
382 months. However, some samples collected in this study, from May – October, were in spawning
383 condition with hydrated oocytes and POFs present. However, the high rates of atresia observed,
384 suggest that recent batches may have been aborted. This may be a more common event for batch
385 spawners and should not be treated as a ‘normal’ skipped spawning event. The relatively high rates

386 of mass atresia (skipped spawning ~ 19%) observed in Pacific hake should be investigated further,
387 and modeled differently from non-batch spawners such as darkblotched and canary rockfishes.

388 In recent years, stock assessment scientists and fisheries managers expressed a need for up-to-
389 date maturity data for inclusion in annual species-specific stock assessments. Since shifts in
390 environmental conditions impact life history processes for marine fish, alterations in size and age
391 at maturity should be monitored closely and tied with changing oceanographic conditions.
392 Maturity information should be collected periodically for all three species studied here to
393 accurately estimate maturity ogives and skipped spawning rates over time. If minimal training and
394 indication of certainty level can be achieved, maturity data may be collected with enhanced spatial
395 and temporal coverage from fishery dependent and independent collections; thus increasing the
396 availability of current life history information and monitoring of skipped spawning for U.S. West
397 Coast fishes.

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409 **References**

- 410 Beamish, R.J., and Chilton, D.E. 1982. Preliminary evaluation of a method to determine the age
411 of sablefish (*Anoplopoma fimbria*). Canadian Journal of Fisheries and Aquatic Sciences,
412 39: 277–287.
- 413 Bradburn, M., Keller, A.A., and Horness, B.H. 2011. The 2003 to 2008 U.S. West Coast bottom
414 trawl surveys of groundfish resources off Washington, Oregon, and California: Estimates
415 of distribution, abundance, length and age composition. U.S. Department of Commerce,
416 NOAA Technical Memorandum, NMFS-NWFSC-114, 323.
- 417 Burnham, K.P., and Anderson, D.R. 2002. Model selection and multimodel inference: A practical
418 information-theoretic approach. Springer, New York.
- 419 Burton, M.P.M. 1999. Notes on potential errors in estimating spawning stock biomass:
420 determining the effects of non-participating adults for some groundfish species. Journal of
421 Northwest Atlantic Fishery Science, 25: 205–213.
- 422 Cadigan, N.G., Morgan, M.J., and Brattey, J. 2014. Improved estimation and forecasts of stock
423 maturities using generalized linear mixed models with auto-correlated random effect. Fish.
424 Manag. Ecol. 21: 343–356.
- 425 Campana, S.E. 2001. Accuracy, precision and quality control in age determination, including a
426 review of the use and abuse of age validation methods. Journal of Fish Biology, 59: 197–
427 242.
- 428 Cope, J.M., and Punt, A.E. 2007. Admitting ageing error when fitting growth curves: an example
429 using the von Bertalanffy growth function with random effects. Canadian Journal of
430 Fisheries and Aquatic Sciences, 64: 205–218.

431 Costa, A. M. 2009. Macroscopic vs. microscopic identification of the maturity stages of female
432 horse mackerel. ICES Journal of Marine Science, 66: 509–516.

433 Dorn, M.W., and Saunders, M. 1997. Status of the coastal Pacific whiting stock in U.S. and Canada
434 in 1997., *In* Appendix: Status of the Pacific Coast Groundfish Fishery Through 1997 and
435 Recommended Biological Catches for 1998: Stock Assessment and Fishery Evaluation.
436 Pacific Fishery Management Council, Portland, OR.

437 Folkvord, A., Jørgensen, C., Korsbrekke, K., Nash, R.D.M., Nilsen, T., and Skjaeraasen, J.E. 2014.
438 Trade-offs between growth and reproduction in wild Atlantic cod. Canadian Journal of
439 Fisheries and Aquatic Sciences, 71: 1106–1112.

440 Frey, P.F., Head, M.A., Keller, A.A. 2015. Maturity and growth of darkblotched rockfish, *Sebastes*
441 *crameri*, along the U.S. west coast. Environ. Biol. Fish, 98: 2353–2365.

442 Gertseva, V. V. and Thorson, J.T. 2014. Status of the darkblotched rockfish resource off the
443 continental U.S. Pacific Coast in 2013. Pacific Fisheries Management Council, Portland,
444 OR.

445 Hamel, O.S. 2008 Status and future prospects for the darkblotched rockfish resource in waters off
446 Washington, Oregon and California as assessed in 2007. Pacific Fishery Management
447 Council, Portland, OR.

448 Hare, S.R. 2007. Variability and precision in the aging of halibut otoliths. International Pacific
449 Halibut Commission Report of Assessment and Research Activities, 2008: 231–250.

450 Hesler, T.E., and Alade, L. 2012. A retrospective of the hake stocks off the Atlantic and Pacific
451 coasts of the United States: Uncertainties and challenges facing assessment and
452 management in a complex environment. Fisheries Research, 114: 2–18.

- 453 Jørgensen, C., and Fiksen, Ø. 2006. State-dependent energy allocation in cod (*Gadus morhua*).
454 Canadian Journal of Fisheries and Aquatic Sciences, 63: 186–199.
- 455 Jørgensen, C., Ernande, B., Fiksen, Ø., and Dieckmann, U. 2006. The logic of skipped spawning
456 in fish. Canadian Journal of Fisheries and Aquatic Sciences, 63: 200–211.
- 457 Kennedy, J., Nash, R.D. Slotte, A., and Kjesbu, O.S. 2011. The role of fecundity regulation and
458 abortive maturation in the reproductive strategy of Norwegian spring-spawning herring
459 (*Clupea harengus*). Marine Biology, 158: 1287–1299.
- 460 Love, M. 1996. Probably more than you want to know about the fishes of the Pacific Coast. 2nd
461 edn. Really Big Press, Santa Barbara, CA. 335 pp.
- 462 Love, M.S. Yoklavich, M., and Thorsteinson, L.K. 2002. The rockfishes of the Northeast Pacific.
463 University of California Press, Berkeley, CA. 405 pp.
- 464 Lowerre-Barbieri, S.K., Ganas, K., Saborido-Rey, F., Murua, H., and Hunter, J.R. 2011
465 Reproductive timing in marine fishes: variability, temporal scales, and methods. Marine
466 and Coastal Fisheries, Management, and Ecosystem Science, 3: 71–91.
- 467 McBride, R.S., Vidal, T.E., and Cadrin, S.X. 2013. Changes in size and age at maturity of the
468 northern stock of Tilefish (*Lopholatilus chamaeleonticeps*) after a period of overfishing.
469 Fisheries Bulletin, 111: 161–174.
- 470 McDermott, S.F. 1994. Reproductive biology of rougheye and shortraker rockfish, *Sebastes*
471 *aleutianus* and *Sebastes borealis*. M.S. thesis, Univ. Washington, Seattle, WA. 81pp.
- 472 McDermott, S.F., and Lowe, S.A. 1997. The reproductive cycle and sexual maturity of Atka
473 mackerel, *Pleurogrammus monopterygius*, in Alaska waters. Fisheries Bulletin, 95: 321–
474 333.

475 Miller, D.J., and Lee R.N. 1972. Guide to the coastal marine fishes of California. California Dept.
476 Fish and Game. Fisheries Bulletin, 157: 1–235.

477 Morgan, M.J., Perez-Rodriguez, A., Saborido-Rey, F., and Marshall, C.T. 2011. Does increased
478 information about reproductive potential result in better prediction of recruitment? *Can. J.*
479 *Fish Aquat. Sci.* 68: 1361–1368.

480 Neuheimer, A. B., and GrønkJaer, P. 2012 Climate effects on size-at-age: growth in warming
481 waters compensates for earlier maturity in an exploited marine fish. *Global Change*
482 *Biology*, 18: 1812–1822.

483 Nichol, D.G., and Pikitch, E.K. 1994. Reproduction of darkblotched rockfish off the Oregon Coast.
484 *Coastal Transactions of the American Fisheries Society*, 123: 469–481.

485 Olsen, E.M., Heino, M., Lilly, G.R., Morgan, M.J., Brattey, J., Ernande, B., and Diechmann, U.
486 2004. Maturation trends indicative of rapid evolution preceded the collapse of northern
487 cod. *Nature*, 428: 932–935.

488 Pacific States Marine Fisheries Commission. 2012, 6th Annual Report. Published for the Congress
489 of the United States and the Governors and Legislatures of Washington, Oregon,
490 California, Alaska and Idaho.

491 Punt, A.E., Smith, D.C., Krusic Golub, K., and Robertson, S. 2008. Quantifying age-reading error
492 for use in fisheries stock assessments, with application to species in Australia’s southern
493 and eastern scalefish and shark fishery. *Canadian Journal of Fisheries and Aquatic*
494 *Sciences*, 65: 1991–2005.

495 R Development Core Team. 2014. R: a language and environment for statistical computing. R
496 Foundation for Statistical Computing, Vienna, Austria.

497 Rideout, R.M., Rose, G.A., and Burton, M.P. 2005 Skipped spawning in female iteroparous fishes.
498 Fish and Fisheries, 6: 50–72.

499 Royle, J.A., and Dorazio, R.M. 2008. Hierarchical modeling and inference in ecology: the analysis
500 of data from populations, metapopulations and communities. Academic Press, San Diego,
501 CA. xviii, 444 pp.

502 Schaub, M., and Royle, J.A. 2013. Estimating true instead of apparent survival using spatial
503 Cormack-Jolly-Seber models. *Methods in Ecology and Evolution*, 5: 1316–1326.

504 Sheehan, D.C., Hrapchak, B.B. 1980. Theory and practice of histotechnology, 2nd edn. Battelle
505 Press, Columbus, OH.

506 Stewart, I.J., Hicks, A.C., Taylor, I.G., Thorson, J.T., Wetzel, C., and Kupschus, S. 2013. A
507 comparison of stock assessment uncertainty estimates using maximum likelihood and
508 Bayesian methods implemented with the same model framework. *Fisheries Research*, 142:
509 37–46.

510 Taylor, I.G., Grandin, C., Hicks, A.C., Taylor, N., and Cox, S. 2015. Status of the Pacific Hake
511 (whiting) stock in U.S. and Canadian waters in 2015. Prepared by the Joint Technical
512 Committee of the U.S. and Canada Pacific Hake/Whiting Agreement; National Marine
513 Fishery Service; Canada Department of Fisheries and Oceans. 159p.

514 Thorson, J.T., Jensen, O.P., and Zipkin, E.F. 2014. How variable is recruitment for exploited
515 marine fishes? A hierarchical model for testing life history theory. *Can. J. Fish. Aquat. Sci.*
516 71: 973–983. doi: 10.1139/cjfas-2013-0645.

517 Thorson, J.T., and Minto, C. 2015. Mixed effects: a unifying framework for statistical modelling
518 in fisheries biology. *ICES J. Mar. Sci. J. Cons.* 72: 1245–1256. doi: 10.1139/icesjms/fsu
519 213.

520 Wright, P.J., Millar, C.P. and Gibb, F.M. 2011. Intrastock differences in maturation schedules of
521 Atlantic cod, *Gadus morhua*. ICES J. Mar. Sci. J. Cons. 68: 1918–1927.

522 Xu, X., Cantoni, E., Flemming, J.M., and Field, C. 2015. Robust state space models for estimating
523 fish stock maturities. Can. J. Stat. 43: 133–150.

524

525

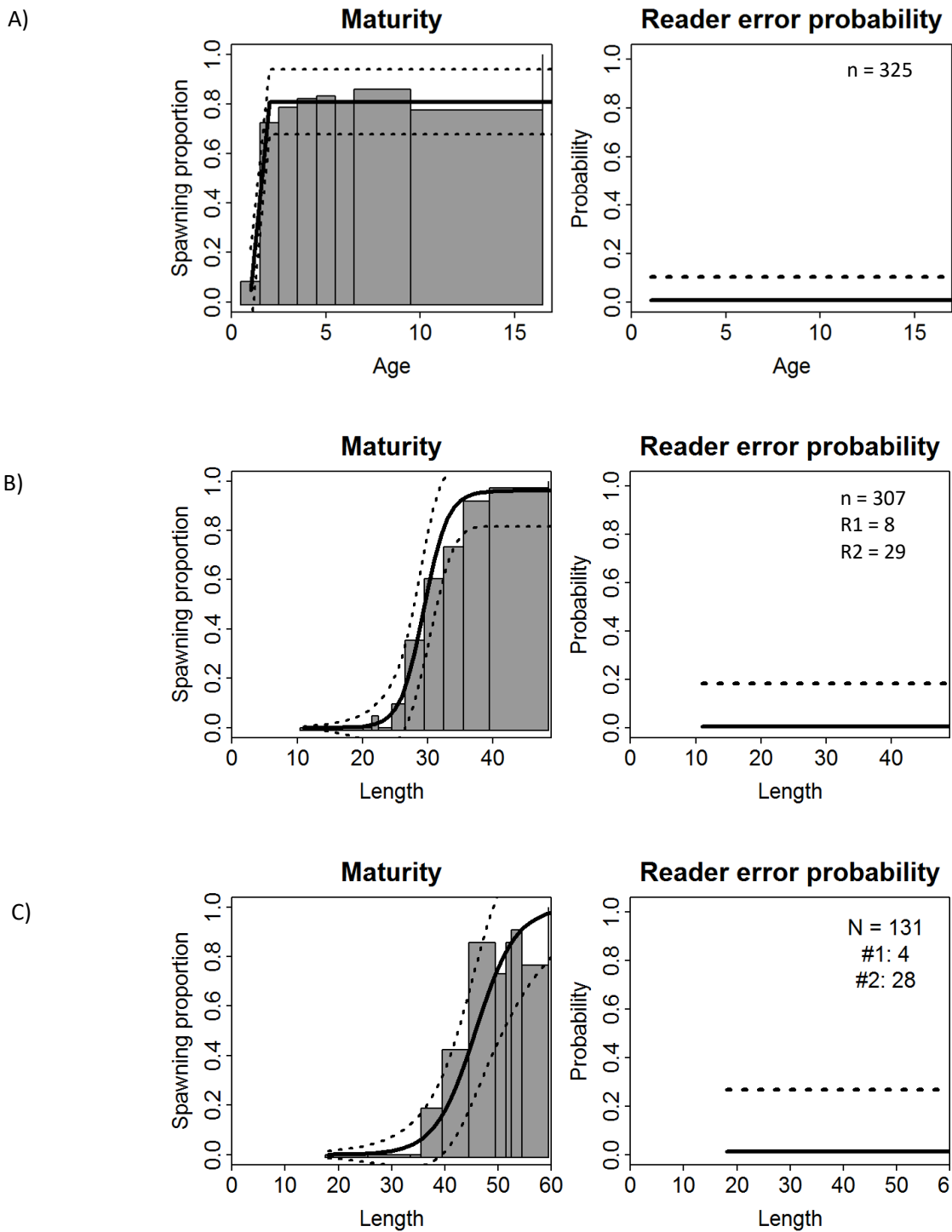


Fig. 1 – Estimated maturity ogive as a function of age (yr) or length (cm) with ± 1 standard error (dashed line) (left panel), and reader error probability (right panel) for A) Pacific hake, B) darkblotched rockfish and C) canary rockfish. Grey bars in the left panel represent the proportion mature for each 10th percentile bin of observed ages/lengths. Total sample size is in the right panel along with the number of “uncertain” maturity reads for readers R1 and R2, and the reader error probability for “uncertain” (dashed line) and “certain” samples (solid line).

Table 1. Parameter estimates representing the intercept, the effect of age (yr), length (cm), and calendar date (Julian day) on the probability of maturity (in logit-space), as well as the asymptotic probability of maturity (i.e. the skipped spawning rate for mature individuals). Also shown are the probability of reader error for reads marked as ‘certain’ or ‘uncertain’, and finally the difference in the Akaike Information Criterion (AIC) among models (the best model has $\Delta AIC=0$, and we only show models for which $\Delta AIC < 10$) for Pacific hake, darkblotched rockfish, and canary rockfish. Standard errors are shown in parenthesis. NA indicates that a parameter was fixed in a given model: age, length, and date effect would be fixed at 0 and asymptote fixed at 1.0.

Intercept	Maturity ogive			Asymptote	Error Probability		ΔAIC
	Age effect	Length effect	Date effect		‘Certain’	‘Uncertain’	
Hake							
-27.922 (97.961)	20.566 (9.895)	NA	1.447 (0.112)	0.809 (0.131)	0.007 (0.049)	0.105 (0.018)	0
-29.733 (100.706)	20.146 (10.026)	0.092 (0.479)	1.446 (0.112)	0.809 (0.131)	0.007 (0.049)	0.105 (0.018)	1.91
-27.870 (109.993)	20.513 (10.485)	NA	1.451 (0.112)	0.810 (0.131)	0.011 (0.054)	NA	4.43
-29.879 (101.546)	20.292 (10.068)	0.092 (0.479)	1.451 (0.112)	0.810 (0.131)	0.011 (0.054)	NA	6.34
Darkblotched							
-17.517 (1.904)	NA	0.555 (0.245)	3.191 (0.546)	0.961 (0.144)	0.006 (0.050)	0.182 (0.011)	0
-17.174 (2.367)	0.048 (0.448)	0.534 (0.329)	3.193 (0.542)	0.961 (0.143)	0.006 (0.050)	0.184 (0.011)	1.98
-14.992 (1.448)	NA	0.470 (0.211)	-0.755 (0.095)	NA	0.007 (0.052)	0.203 (0.012)	3.24
-14.955 (1.549)	0.001 (0.130)	0.498 (0.234)	NA	0.958 (0.155)	0.006 (0.049)	0.193 (0.011)	4.65
-13.173 (1.183)	NA	0.431 (0.200)	NA	NA	0.007 (0.052)	0.217 (0.011)	5.14
-15.009 (1.453)	0.001 (0.071)	0.470 (0.212)	-0.755 (0.095)	NA	0.007 (0.052)	0.204 (0.012)	5.28
-13.186 (1.186)	0.001 (0.071)	0.432 (0.200)	NA	NA	0.007 (0.052)	0.218 (0.011)	7.18
Canary							
-13.319 (1.698)	NA	0.265 (0.183)	-1.330 (0.126)	NA	0.015 (0.077)	0.267 (0.019)	0
-12.777 (1.942)	0.043 (0.283)	0.241 (0.234)	-1.425 (0.228)	NA	0.015 (0.077)	0.267 (0.019)	1.85
-12.808 (2.013)	0.043 (0.284)	0.241 (0.237)	6.90 (15.516)	0.999 (0.125)	0.015 (0.077)	0.267 (0.019)	3.85
-20.463 (5.043)	NA	0.495 (0.356)	NA	0.858 (0.191)	0.015 (0.079)	0.258 (0.022)	6.30
-20.458 (5.045)	0.001 (0.133)	0.495 (0.357)	NA	0.858 (0.191)	0.015 (0.079)	0.258 (0.022)	8.30
-10.617 (1.309)	NA	0.240 (0.170)	NA	NA	0.016 (0.080)	0.270 (0.021)	9.93

Table 2. Disagreements in assessment of maturity state between readers R1 and R2 for canary rockfish, darkblotched rockfish, and Pacific hake, all samples included in analysis. Disagreements are expressed based on the level of uncertainty recorded by each reader for each sample with **a** the overall number of samples with disagreements relative to **b** the total number of samples for each level of certainty (e.g. when both readers were certain for canary rockfish, they disagreed on 2.9% of the samples, and when both readers were uncertain they disagreed 100% of the time¹).

Certainty between readers	Proportion (a/b) : Percent Disagreement		
	Canary n = 131	Darkblotched n = 307	Hake n = 329
Both certain	3/103 : 2.9% ¹	4/276 : 1.4%	6/303 : 2%
R1 certain, R2 uncertain	6/25 : 24%	3/23 : 13%	3/23 : 13%
Both uncertain	2/2 : 100%	2/5 : 40%	0/2 : 0%