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8 **Validity of an external sex determination method in Atlantic Sturgeon (*Acipenser***  
9 ***oxyrinchus oxyrinchus*)**

10 **Running Title:** Determining sex from urogenital morphology in sturgeon

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23 the University of New England. This manuscript represents Marine Science Center contribution  
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## 25 **Summary**

26 The validity of the shape of the urogenital opening was tested as a predicable means to determine  
27 the sex of Atlantic sturgeon captured from in the Saco River estuary, Maine. Evaluation of 121  
28 individuals ranging in size from 107 to 182 cm fork length were compared to non-lethal  
29 radioimmunoassay determined sex data previously examined for these individuals within this  
30 estuary. The results suggested that using the shape of the urogenital opening is not a reliable  
31 means to determine sex as only 51% of Atlantic sturgeon were correctly identified female.  
32 Additionally, there was no significant difference in correctness relative to 10 cm fork length size  
33 classes.

## 34 **Introduction**

35 An important basic dataset to obtain for any fish population is the sex of individuals, as it  
36 can allow for an evaluation of the sex ratio, indicating temporal and spatial sexual segregation or  
37 aggregation (e.g. Taubert, 1980b; Buckley & Kynard, 1985b; Collins & Smith, 1997). However,  
38 many fish do not exhibit external sexual dimorphism throughout their life stages, making  
39 baseline reproductive data difficult to obtain in some species. The sturgeon family,  
40 *Acipenseridae*, is a world-wide threatened and endangered group (IUCN, 2010) that lacks  
41 external sexual dimorphism (Vecsei, Litvak, Noakes, Rien, & Hochleithner, 2003). Since  
42 sturgeon populations are critically low, large datasets on sex ratios cannot viably be obtained via  
43 lethal methods (i.e. gross dissection).

44 With many fish populations declining in both marine and freshwater environments,  
45 research methods that use nonlethal assessment allow for data to be collected in large sample  
46 sizes without hindering the population (Chiotti, Boase, Hondorp, & Briggs, 2016). In response,  
47 non-lethal methodology development and implementation in fish research and management is a  
48 rapidly advancing area (e.g. Vecsei et al., 2003; Bryan et al., 2007; Chiotti et al., 2016).  
49 Moreover, non-lethal methods can be used in tandem with some other close research areas (i.e.

50 feeding ecology and spatial usage) to determine essential fish habitat (Novak, Carlson, Wheeler,  
51 Wippelhauser, & Sulikowski, 2017).

52 Many non-lethal methods have been assessed in sturgeon including: endoscopy for  
53 inspection and collection of gonadal tissue (e.g. Wildhaber et al., 2005), sex steroid hormone  
54 levels (e.g. Webb et al., 2002), ultrasonography (e.g. Moghim, Vajhi, Veshkini, & Masoudifard,  
55 2002), morphological features and measurements (e.g. Maltsev & Merkulov, 2006; Podushka,  
56 2008), and a physical attempt to secrete eggs or milt from an adult fish (e.g. Heise, Bringolf,  
57 Patterson, Cope, & Ross, 2009). These techniques have varying levels of invasiveness, accuracy,  
58 and cost. Endoscopy for gonadal tissue collection an invasive technique requiring  
59 anesthetization, an incision in the abdominal cavity, and the insertion of an endoscope  
60 (Wildhaber et al., 2005). This method requires means to recover the fish for a period, as well as a  
61 high level of skill from the examiners to not only perform the surgery but histologically analyze  
62 the sampled tissue. Sex steroid hormone analysis to determine sex is a non-lethal method, using  
63 circulating levels of hormone ratios to determine sex (Webb, Feist, Foster, Schreck, &  
64 Fitzpatrick, 2002). This method is minimally invasive and requires a low amount of skill to  
65 collect a sample, but a high degree of time and expense to process samples in a laboratory  
66 setting. Ultrasonography is also non-invasive; however it requires proper equipment and training  
67 to not only correctly locate the gonad, but determine sex from imaging. While this method has  
68 varying degrees of success, when done properly, the mature gonads of each sex give off key  
69 signatures in an ultrasound images (Moghim et al., 2002).

70 Other more easily examined external characteristics have also been assessed to sex  
71 various species of sturgeon. Methods such as urogenital region morphometry (Billard, 2002) and  
72 craniological measurements (Maltsev & Merkulov, 2006) have been performed with some  
73 success, however Chebanov and Galich (2011) warned that these methods are not recommended  
74 given uncertainty despite ease. Although Atlantic sturgeon are not considered externally sexually  
75 dimorphic, it was suggested (Vecsei et al., 2003) that the urogenital opening shape may serve as  
76 an indicator of sex, thus, providing a fast and nonlethal method to differentiate males from  
77 females in Atlantic sturgeon. Results by Vecsei et al. (2003) indicated that a “Y” shaped  
78 urogenital opening indicated a male (Figure 1a), and while an “O” shaped urogenital opening

79 indicated a female (Figure 1b). Although an 82% accuracy was observed, only 17 fish were  
80 evaluated.

81 In Wheeler, Novak, Wipplehauser, and Sulikowski (2016), Atlantic sturgeon in the Saco  
82 River estuary (SRE) in Maine were sexed using a combination of circulating blood hormones  
83 techniques. Results showed 93% of the 288 fish sampled were sexed, demonstrating the validity  
84 of this non-lethal method in this endangered species (Wheeler et al., 2016). The goal of this  
85 study herein was to use the previously determined Atlantic sturgeon sex data from Wheeler et al.  
86 (2016) to assess the validity of the external morphological feature described in Vecsei et al.  
87 (2003) using a more robust sample size.

## 88 **Materials and Methods**

### 89 *Sampling*

90 This research was part of a larger study assessing Atlantic sturgeon in the Saco River  
91 estuary. The details of capture and sampling methods can be found in Wheeler et al. (2016).  
92 After routine sampling, the urogenital opening was inspected for shape and photographed. After  
93 the sampling procedure, sturgeon were allowed to recover in a net-pen prior to release.

### 94 *Sex determination*

95 The sex of Atlantic sturgeon captured in the SRE was determined in vivo via a  
96 comparison of two calculations using circulating levels of sex steroid hormones. The full details  
97 of the analysis can be found in Wheeler et al. (2016). Briefly, testosterone (T) and 17 $\beta$ -estradiol  
98 (E<sub>2</sub>) was quantified via radioimmunoassay (RIA) methods modified from Tsang and Callard  
99 (1987) and Sulikowski, Tsang, and Howell (2004). The ratios of these hormones in circulation  
100 were compared to mean hormone concentrations from Van Eenennaam et al. (1996) in addition  
101 to application in discriminant function analysis from Webb et al. (2002). Sex output from these  
102 two methods correlated in 93% of Atlantic sturgeon, showing the validity of this method as one  
103 non-lethal sex determining method (Wheeler et al., 2016). Therefore, we were able to directly  
104 compare morphological shape of the urogenital region herein to steroid hormone sex  
105 determination from previous work. Images obtained during sampling from urogenital  
106 photography were read by two readers in conjunction without prior knowledge of the specimen's

107 sex as determined via RIA. Each reader assigned sex individually which was then discussed if  
108 determinations conflicted and a final sex status was given to each specimen. If an individual did  
109 not strictly adhere to either shape it was not included in the statistical data analysis. Here forward  
110 we use the term relative correctness to describe the relationship between urogenital morphology  
111 sex and Wheeler et al. (2016) RIA sex, as we do not have dissection or histological validation of  
112 the sex of any individuals in the SRE.

### 113 **Statistical Analysis**

114  
115 A logistic regression with forward selection was performed to assess if relative  
116 correctness was influenced by 10 cm fork length size classes (ranging from 100-190 cm) as well  
117 as months sampled (May-November 2013 & 2014) (*SYSTAT (Systat Software, San Jose, CA)*).

### 118 **Results**

119 Atlantic sturgeon used within this study ranged from 106-182 cm fork length (n=140), with 121  
120 individuals adhering to a Y or O shape. Overall, sex determination was correct to Wheeler *et al.*,  
121 (2016) sex data in 51% of fish, ranging from 0.0-76.9% correctness when assessed by 10 cm size  
122 classes (Figure 2). The resulting logistic regression was not significant (AIC=167.130; df=6;  
123  $p=0.107$ ), therefore we could not conclude any influence of 10 cm size classes or months on  
124 relative correctness (Figure 2 & 3).

### 125 **Discussion**

126 Our findings suggest assessment of urogenital shape in SRE sturgeon was highly  
127 variable. Some individuals did have a defined “Y” or “O” shape (Figure 1) and other lacked  
128 adherence to one shape or the other (Figure 4). However, this is in contrast to Vecsei et al.  
129 (2003), which found this method to be 82% accurate in live wild Atlantic sturgeon, when  
130 validated with gross dissection.

131 Other work has found urogenital shape as a good indicator of sex. For example, this  
132 technique correctly sexed 91% of males and 94% of females in mature Northern pike (*Esox*  
133 *lucius*) (Casselman 1974), with 3-4 month old young of the year being sexed correctly 72% of  
134 the time. Similarly, using a modified method applied to muskellunge (*Esox masquinongy*), sex

135 was determined correctly in 92-98% of juveniles, and 100% in adults (Lebeau & Pageau, 1989).  
136 Finally, high accuracies of this technique applied in yellow perch (*Perca flavescens*) have also  
137 been reported ranging from 82.7-97.4% depending on total length (Malison, Held, & Kaatz,  
138 2011). In some cases, discrepancies were reported due to reader skill levels (Guerrero, 1982).

139 Despite the aforementioned findings and based on the difficulties observed in the current  
140 study, the use of urogenital shape as an indicator of sex in Atlantic sturgeon appears complex.  
141 However, it is important to note the current study's limitations, where sex was not confirmed via  
142 dissection or histology, and maturity stages of our individuals were not known. In the future, this  
143 method needs further validation across maturity stages and seasons for the *Acipenseridae* family  
144 before application in the field.

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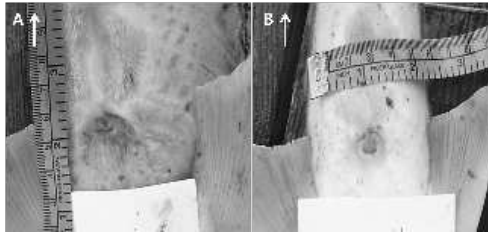
## 220 **Figure Legends**

221 Figure 1: Urogenital opening morphology of a male “Y” shape (**A**) and female “O” shape (**B**) in  
222 Atlantic sturgeon caught in the Saco River estuary. The posterior ends are represented by the  
223 arrows - as we have two specimens and two arrows.

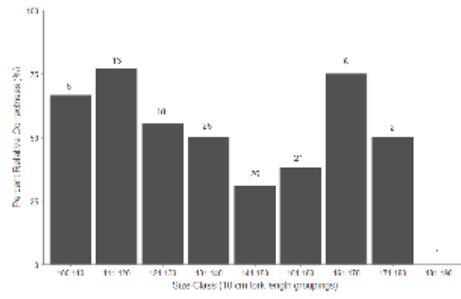
224  
225 Figure 2. The percent relative correctness of urogenital morphology sex to RIA sex data from  
226 Wheeler et al. (2016) over 10 cm fork length size classes. There was no statistical significance  
227 between any of the groups at  $\alpha \leq 0.05$ . N values are represented by numbers above each bar.

228  
229 Figure 3. The percent relative correctness of urogenital morphology sex to RIA sex data from  
230 Wheeler et al. (2016) over months of sampling in 2013 and 2014 combined. There was no  
231 statistical significance between any of the groups at  $\alpha \leq 0.05$ . N values are represented by  
232 numbers above each bar.

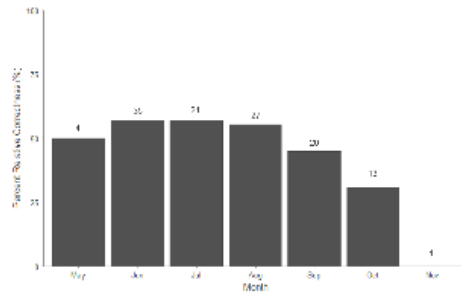
233 Figure 4: Urogenital opening morphology of an Atlantic sturgeon with unidentifiable sex  
234 externally. Note the lack of adherence to a “Y” or “O” shape. The posterior end is represented by  
235 the arrow.



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