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Article type : Article

Examining the severity of roof-hooking injuries in dolphinfish: a comparison between computed tomography and gross necropsy

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ABSTRACT

We describe hook trauma to the roof of the mouth in dolphinfish *Coryphaena hippurus* and compare computed tomography (CT) scanning to gross necropsy (GN) as a technique for diagnosing hooking injury in fish. Forty-two dolphinfish carcasses spanning a range of hook injuries were collected and CT scanned, and 33 of these were evaluated using GN. Specimens were hooked either in the roof of the mouth, the eye via the roof or upper jaw, or the jaw (control

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1002/nafm.10252](https://doi.org/10.1002/nafm.10252)

30 group). In 75% of roof-hooked individuals, GN revealed nondisplaced to comminuted fractures
31 of the bones of the suspensorium, hematomas in and laceration of the extraocular muscles, and/or
32 damage to the optic nerve. These injuries have the potential to compromise vision and therefore
33 decrease post-release survival rates of obligate sight-feeding species such as dolphinfish. We
34 evaluated the effectiveness of CT scanning to diagnose injury and found that CT could
35 efficiently and accurately identify fractures and some soft tissue damage, but some injuries found
36 in GN (e.g. optic nerve damage) were not observed on CT scans. Based on our findings, it is
37 likely that mortality is greater in dolphinfish when hooked in the roof of the mouth than in the
38 jaw. This study demonstrates a novel technique that was effective at diagnosing hooking injuries
39 associated with the roof of the mouth.

40

41 INTRODUCTION

42 Evaluation of population status requires knowledge of mortality numbers of fish that are
43 caught, whether due to harvest or catch-and-release (C&R). However, the fate of discarded fishes
44 is often unknown (Davis 2002) and disregarding post-release mortality can lead to uncertainty in
45 stock assessments (Williams 2002, Pollock and Pine 2007). It is important to understand the
46 anatomical effects of hooking and to make use of diagnostic techniques that are best-suited for
47 specific species and their respective injuries. This can allow for more informed C&R mortality
48 rate estimates and allow anglers to make more informed decisions when choosing whether to
49 retain their catch. With increased popularity in C&R, it is valuable to provide information that
50 promotes sustainable angling practices (Brownscombe et al. 2017).

51 The recreational fishery in the U.S. South Atlantic region targets dolphinfish *Coryphaena*
52 *hippurus* using hook-and-line gear, and dolphinfish are most often hooked in the jaw, followed
53 by roof, gill, eye, gut, and body, respectively (C.S. Mikles, personal observation). Dolphinfish
54 are pelagic piscivores that are primarily reliant on sight for foraging (Loew and McFarland
55 1990). Given their abundance and aggressive feeding behavior, dolphinfish have been and
56 continue to be one of the top-ranked recreational fisheries in numbers caught within the U.S.
57 South Atlantic (Rose and Hassler 1969; NOAA 2012). For multiple reasons, including ethical
58 angling, size, and bag limits, dolphinfish in this region are often released after capture (Carter
59 and Liese 2012). Discard mortality of dolphinfish has not been estimated for this fishery or for
60 other fisheries directed toward it throughout its worldwide range.

61 Many studies that have analyzed C&R mortality rates often incorporate hooking location
62 into these estimates. Hooking location has been established to be the most important contributor
63 to post-release mortality among reviews of C&R studies across species (Muoneke and Childress
64 1994, Bartholomew and Bohnsack 2005). Hook trauma has been assessed in other recreationally
65 caught fishes but has yet to be characterized in dolphinfish.

66 Post-release mortality for shallow (jaw, roof, eye) and deep hooking locations (gut and
67 gills) has been studied in several species. Jaw hooking is generally considered to be a location
68 associated with low mortality, averaging between 0-10% (Grover et al. 2002, James et al. 2007,
69 Lyle et al. 2007, Veiga et al. 2011, Campbell et al. 2014). Independent of morphological
70 differences, deep hooking in the gut and gills is often associated with a higher mortality rate than
71 shallow hooking locations (Warner 1976, Domeier et al. 2003, Rudershausen et al. 2014). On the
72 other hand, published estimates of mortality on eye- and roof-hooked fish are more variable,
73 possibly because of the difficulties in observing the degree of injury. Across a number of species,
74 damage to the eye from hooking injuries likely contributes to post-release mortality due to
75 difficulties associated with feeding and predator avoidance (Prince et al. 2002, DuBois and
76 Dubielzig 2004, Cooke and Sneddon 2007). The extent of injuries and rate of catch and release
77 mortality in dolphinfish as a result of hooking injury to each of these anatomical sites is
78 unknown.

79 Many studies that assess C&R mortality rates do not describe roof hooking or distinguish
80 it from jaw hooking (Murphy et al. 1995, Taylor et al. 2001, Stachura et al. 2012, Bergmann et
81 al. 2014). This may be due to a perception that roof-hooked fish have mortality rates similar to
82 fish hooked in the jaw or to the difficulty in observing damage to this location without necropsy
83 (Belle 1997). Mortality for this hooking location ranges from being used as a control (i.e.
84 assuming 0% mortality) in Chinook salmon *Oncorhynchus tshawytscha* (Grover et al. 2002) to
85 rates as high as 80% in pelagic fishes (Faltermann and Graves 2002). This variability is likely due
86 to differences in morphology and feeding behavior across species. Given such variation it is
87 important to assess injuries to the roof of the mouth on a species-specific basis. The roof of the
88 mouth in dolphinfish lies in close proximity to the bottom of the eye, which led us to explore
89 different techniques to examine injuries in these tissues.

90 The objectives of this study were to describe roof-hooking injuries in dolphinfish and
91 compare computed tomography (CT) scanning to gross necropsy (GN) as a technique for

92 diagnosing hooking injury in fish. CT scanning has applications in aquatic veterinary medicine to
93 diagnose disease (Garland et al. 2002) but this technique has not been used in published work to
94 investigate the effects of hooking damage in fish and to better understand the impacts of C&R.
95 We hypothesized that hooking location influences the level of injury and predicted that roof-
96 hooked fish will have greater injury relative to jaw-hooked fish.

97 **METHODS**

98 **Carcass Collection and Hook Location Assignment**

99 Dolphinfish carcasses were collected from May through July of 2016 and 2017 at the
100 Morehead City, North Carolina waterfront. Fish were collected opportunistically from a cleaning
101 operation that fillets fish landed by recreational charter boats between 2 and 10 hours after they
102 are boated; therefore, exact gear type, hook size, and landing methods were unknown. While
103 fishing practices differ among boats, the charter fleet typically angles dolphinfish by trolling with
104 J hooks with dead natural baits and/or artificial lures, or by bailing with circle hooks with dead
105 natural bait (Rudershausen et al. 2012). Additionally, dolphinfish are gaffed or brought on board
106 without gaffing; after boating, dolphinfish are put directly on ice. None of the dolphinfish
107 retained for CT scanning were gaffed in the head. The fork lengths of the fish were measured,
108 and carcasses were examined for hooking location and external damage. The hooking location
109 was determined through observation of wounds left by hooks or from hooks left in place.

110 Dolphinfish retained for CT scanning were hooked either in the jaw, eye, or roof of the
111 mouth (Table 1). Fish hooked in the jaw served as controls for CT analysis and injury
112 characterization. Having very minimal injury (see below), jaw hooked fish also controlled for
113 differences in angling practices and any potential damage inflicted on fish prior to collection.

114 **Computed Tomography (CT) Scanning**

115 The carcasses collected at the Morehead City waterfront were frozen at -20°C. All heads
116 were scanned at the North Carolina State University College of Veterinary Medicine using the
117 Siemens SOMATOM Sensation 16 (Siemens Medical Solutions, Malvern, PA), with a veterinary
118 small adult ear setting at a slice thickness of 0.75 mm and a reconstruction increment of 0.4 mm.
119 After CT scanning, heads were stored and thawed at 4°C for subsequent gross necropsy (below).

120 The scanned images were examined in Horos® DICOM medical image viewer
121 (<https://www.horosproject.org>). The CT scans were first naïvely evaluated by the image analyzer
122 with no knowledge of the hooking location or the extent of injury. The CT scans were

123 reevaluated a second time after we became more familiar with the images and internal anatomy
124 as well as cross-referencing with dockside hooking location designations. Cross-referencing is an
125 important tool to diagnose conditions and understand the extent of injuries (Cockcroft and
126 Holmes 2003).

127 The CT scans were examined in both transverse and coronal sections. Within the CT scan
128 images, bone structure appears white (radio-opaque, mineral opacity), soft tissue and hematoma
129 grey (soft tissue opacity), and air-influx black (gas opacity, radiolucent). Gas is expected in the
130 oral and gill cavities since they are exposed to air after the fish is boated, but air is also
131 introduced through fractures in the bone and can be traceable from the fracture site. Gas as an
132 artifact is sometimes present and can be attributed to air introduced by decapitation or
133 decomposition; control fish served to represent the effects of decapitation and freezing/thawing.
134 Assessing the degree of bilateral symmetry between injured and uninjured sides of the same
135 individual can be used as an internal control since hooking injury occurred only to one side of
136 each individual that we collected.

137 Roof anatomical evaluations focused on the palate, or suspensorium, which in
138 dolphinfish is a delicate structure composed of a series of thin bones and cartilages (Fig 2a, b).
139 Specifically, the endopterygoid, ectopterygoid, metapterygoid, palatine, and quadrate form the
140 suspensorium (Hilton 2011). The endopterygoid is the bone most susceptible to fracture caused
141 by hook trauma, due to its thin dorsal shelf that supports and protects the orbital cavity.

142 We designated three CT injury categories based on our interpretations of the scans: CT 1
143 - no visible damage or trauma to the suspensorium or to the orbit (Fig. 1a), CT 2 - fracture to
144 bone(s) forming the suspensorium, paired with gas influx continuous from the fracture site
145 extending only into the base of the orbit (gas confined to the orbital floor) (Fig. 1b), and CT 3 –
146 severe fracture (displaced or comminuted) to bone(s) forming the suspensorium, paired with gas
147 influx continuous from the fracture site extending past the orbital floor dorsally to the level of the
148 optic nerve, extraocular muscles, or the eye (Fig. 1c).

149 **Gross Necropsy (GN) and Comparison to CT Scans**

150 We performed GNs to identify the extent of damage caused by hooking and to compare
151 to CT scans. GNs were necessary because soft tissue damage in CT scans was evaluated in terms
152 of gas path and volume and adjacent bone fractures, rather than observing the injuries *in situ*.
153 The eye and surrounding structures were examined from a ventral perspective using the

154 following procedure in order to preserve the integrity of the tissues damaged by hooking. First,
155 the opercula and gill arches on both sides of the fish were removed. The lower jaw was removed
156 at the articulation between the maxilla and quadrate and the dentary, exposing the length of the
157 roof of the mouth. The mucosa of the roof was examined for any signs of potential hooking
158 damage (e.g. laceration that penetrates the mucosa), and the outer (ventral) layer was removed,
159 exposing the superficial muscle and the endopterygoid. The muscle and surface of the
160 endopterygoid were examined and then carefully removed, exposing the orbit and the extraocular
161 muscles. The interior surface of the eye and the extraocular muscles were evaluated for damage,
162 then the extraocular muscles were carefully removed. The optic nerve was evaluated for damage
163 or laceration, then the conjunctiva was cut and the surrounding muscles and mucosa removed to
164 evaluate the state of the eye and optic nerve further. Finally, the uninjured contralateral orbit was
165 examined as an internal control.

166 We established three GN injury categories: GN 1 - no visible damage or trauma to the
167 suspensorium or to the orbit, any laceration is minimal and superficial, GN 2 - visible laceration
168 of the mucosa and fracture to the endopterygoid, damage to muscle is superficial, and GN 3 -
169 visible laceration of the mucosa, and fracture to the endopterygoid and/or the ectopterygoid,
170 paired with damage to at least one of the extraocular muscles and/or the optic nerve.

171 The percentage agreement between CT and GN categories were determined to assess the
172 ability to predict GN injuries from CT scans. Additionally, a Kolmogorov-Smirnov goodness of
173 fit test for discrete ordinal data (Zar 1996) was used to compare the observed CT counts to
174 expected counts (based on GN results) for fish hooked in the roof of the mouth. To provide a
175 qualitative measure of the relative injuries between jaw, eye and roof-hooked dolphinfish, we
176 calculated a weighted average of injury by hooking location using the number of fish assigned to
177 GN and CT scores. The weighted average was:

178
$$\frac{(n.1 * 1) + (n.2 * 2) + (n.3 * 3)}{\sum n},$$

179 where n is number of fish in one of the three GN or CT injury categories and 1, 2, and 3 are the
180 GN or CT injury scores. We performed a Kruskal-Wallis test with Dunn post-hoc tests to
181 determine the relationship between hooking location and injury scores assigned through CT and
182 GN (Sokal and Rohlf 1995). Significance was assessed at $\alpha = 0.05$. Statistical analyses were
183 conducted in R using the packages “dplyr” and “FSA” (Wickham 2018, Ogle 2018, R Core
184 Team 2018).

185 **RESULTS**

186 **Carcass Collection and Hook Location Assignment**

187 Forty-two dolphinfish carcasses were collected across the three hooking locations (roof
188 of the mouth, eye via the roof or upper jaw, and jaw), and fish ranged in fork length from 480-
189 985 mm (Table 1). The 14 control fish examined in the laboratory had no injuries to the roof or
190 the eye, confirming the dockside assignment of jaw hooking.

191 **Computed Tomography (CT) Interpretations**

192 The CT scans showed no evidence of fractured bones and gas intrusion in the control
193 fish; however, these injuries were observable on CT scans in 23 out of 28 non-control fish
194 (Figure 1). Fractures to the bones of the suspensorium could be identified and were categorized
195 as non-displaced, displaced, or comminuted. Fractures were observed in the endopterygoid more
196 frequently than in any other bones of the suspensorium. Gas artifact could be seen in the eyes
197 and surrounding the orbital cavity in both non-control individuals and control fish, and can be
198 attributed to the effects of decapitation, decomposition and freezing/thawing. However, the
199 volume of gas artifact was minimal and distinguishable from gas influx from a fracture site in
200 non-control fish, since it appeared random and scattered instead of intruding directly from an
201 epithelial location.

202 We assigned 14 controls, five roof, and one eye hooked fish to CT 1 (Table 2; Figure 1a).
203 There were six roof and three eye hooked fish assigned to the CT 2 condition where scans
204 identified non-displaced fractures to the bones of the suspensorium and minimal gas intrusion
205 (Table 2; Figure 1b). Scans from fish in CT 3 showed displaced or comminuted fractures to the
206 bones of the suspensorium paired with obvious gas intrusion that was traceable to the level of the
207 orbital floor (Figure 1c); this condition was found in five roof and eight eye hooked fish (Table
208 2). The highest proportion of fish in CT 3 were hooked in the eye, followed by roof.

209 **Gross Necropsy (GN) and Comparison to Computed Tomography (CT) Scanning**

210 Gross necropsies were performed on all roof- and eye-hooked fish, and five control fish
211 (total=33). The remainder of the control fish were examined for external damage but not
212 dissected. Damage seen by dissection was a result of hook injury, all fish were assigned one of
213 the three gross necropsy injury categories (Table 2). All 14 control fish fell into GN 1; full
214 dissections of all control fish were not necessary to determine their placement in GN 1 as nine
215 control fish not fully dissected were consistent with five control fish that were fully dissected.

216 The proportions of fish in GN1, GN2, and GN3 for roof- and eye-hooked fish were similar to the
217 proportions in the CT categories (Table 2). In a few instances, the CT categories overestimated
218 or underestimated the degree of damage as determined by GN but had a total percent agreement
219 of 80.9% with GN. Discrepancies mostly occurred between categories 2 and 3; the CT
220 interpretation underestimated the damage in five cases and overestimated it in three. For roof-
221 hooked fish, there was no statistical difference in injury score assignment between the CT and
222 GN (K-S test: $d_{max} = 2$, $p > 0.50$).

223 For fish in GN 1 (n=19), all damage was superficial. Lacerations to the mucosa in the
224 roof of the mouth were observed but no fracture was observed, and no damage occurred to the
225 superficial muscle. The CT assessment scored 1 and the GN scored 2 for only one individual,
226 which had a chip fracture in the endopterygoid, along with slight damage to the surrounding
227 superficial muscle.

228 For fish in GN 2 (n=9), fractures of varying types and severities to the endopterygoid
229 were observed but soft tissue damage did not extend past the superficial muscle, which was often
230 bruised or torn. Of the nine fish scored as CT 2, five of these were also scored as GN 2.

231 For fish in GN 3 (n=14), fractures of varying types and severities occurred to the
232 endopterygoid and ectopterygoid. The superficial muscle was damaged, and damage occurred to
233 the extraocular muscles and/or to the optic nerve. Hematoma was often present in the orbital
234 floor. One roof-hooked fish and three eye-hooked fish sustained injuries to the optic nerve. Of
235 the 13 fish scored as CT 3, 10 were also scored as GN 3.

236 Of the 16 fish hooked in the roof, four fell into GN 1, five in GN 2, and seven in GN 3.
237 Of the 12 fish hooked in the eye (globe or fornix) via the roof or upper jaw, one was placed in
238 GN 1, four in GN 2, and seven in GN 3. All control fish were placed in GN 1. Twelve out of 16
239 (75%) roof-hooked fish sustained a combination of fractures to the suspensorium, laceration of
240 extraocular muscles, and/or optic nerve damage. Assessment by CT never diagnosed damage
241 where none was detected by gross necropsy. There was no identifiable consistent manner in
242 which the endopterygoid was fractured, except that the thinnest parts of the bone were most
243 susceptible to damage.

244 The weighted average GN injury scores for jaw-, roof-, and eye-hooked dolphinfish were
245 1.0, 2.2, and 2.5, respectively, and were 1.0, 2.0, and 2.6 for CT scores. The results of the
246 Kruskal-Wallis test showed differences in the severity of hooking injuries among jaw-, roof-, and

247 eye-hooked fish for CT ($\chi^2 = 22.6$, $p < 0.001$) and GN techniques ($\chi^2 = 22.3$, $p < 0.001$). Post-
248 hoc analyses for each categorization revealed differences between jaw and roof ($p < 0.002$) and
249 jaw and eye ($p < 0.001$) for both GN and CT techniques; however, there were no differences in
250 injury scores between roof and eye ($p > 0.05$) for either technique. Thus, roof-hooked
251 dolphinfish have injury levels that are closer to eye-hooked relative to jaw-hooked dolphinfish.

252 **DISCUSSION**

253 The effects of roof hooking in contributing to C&R mortality have seldom been studied
254 compared to the total number of C&R mortality estimates. Our prediction that dolphinfish
255 hooked in the roof of the mouth would sustain injuries that are more severe than jaw-hooked fish
256 was supported by computed tomography (CT) and gross necropsy (GN) findings. While our
257 sample of 16 roof-hooked fish is modest, the extent and variability in injury was extensive.

258 There was a high percentage (75%) of roof-hooked dolphinfish with damage to the bones
259 of the suspensorium, extraocular muscles, and/or optic nerve. The same injuries were observed
260 92% of the time in eye-hooked fish. CT scans and GN results had similar findings when the
261 bones of the suspensorium were fractured; however, there was ambiguity in determining the
262 extent of soft tissue damage with CT. CT scans more accurately show bone structure, so
263 differences in fracture severity were easily discernable to categorize fish in either CT 2 or 3.
264 Tracing the path of gas influx provided some indication of the extent of damage present, but
265 results from gross necropsies were more definitive. For roof- and eye-hooked fish, internal
266 damage to the musculature, nerve pathways, and the orbit can vary in severity and is not
267 necessarily correlated with the severity of fracture. The gross necropsies served to validate the
268 diagnoses from the scans, and also provided more specific information on soft tissue damage.
269 The diagnoses from the CT agreed with the GN around 80% of the time. We have demonstrated
270 the use of CT for comparing the severity of hooking injuries across hooking locations that are
271 difficult to observe and have not been previously studied in dolphinfish.

272 The endopterygoid and superficial muscle provide a thin layer of protection between the
273 oral and orbital cavities and are not suited to withstand hook damage. Our understanding of these
274 injuries provides insight into the potential for post-release survival. The injuries we describe can
275 result in severe eye damage and potentially impair vision. For example, damage to the lens or the
276 sclera, intraocular hemorrhage, and enucleation were designated as injuries most likely to result
277 in long-term visual impairment of stream trout (DuBois and Dubielzig 2004). Fish hooked in

278 eye-associated tissues will likely suffer a degree of vision loss, which has been associated with
279 higher mortality (Warner 1976, Pauley and Thomas 1993). If selectively harvesting, anglers may
280 consider choosing to keep individuals with greater hooking damage (Brownscombe et al. 2017).
281 Given the importance of sight-feeding to dolphinfish, we recommend retaining individuals of
282 legal size with eye or roof-hooking over fish hooked in the jaw. Additionally, trolling with circle
283 hooks would reduce the amount of deep (e.g. eye and roof) hooking (Rudershausen et al. 2012).

284 Dolphinfish hooked in the roof of the mouth sustained higher degrees of damage than jaw
285 hooked fish. Thus, hooking in the roof of the mouth would likely result in higher mortality than
286 jaw hooking based on the injuries that we observed. Fractures and muscle damage often cause
287 blood loss, and these hook injuries can create pathways through which seawater and pathogens
288 may be introduced to vital areas. Depending on the severity and location of hooking damage, the
289 presence of bleeding is often linked to post-release mortality, as it is dependent on the perfusion
290 of vasculature and critical organs (Arlinghaus et al. 2007). Numerous studies have found that
291 bleeding, along with hooking location, are the most important factors when assessing mortality
292 of angler-caught fish (Nuhfer and Alexander 1992, Meka 2004, Weltersbach and Strehlow 2013,
293 Gargan et al. 2014). We did not observe bleeding immediately after angling, although hematoma
294 was often present in the orbital cavity in roof-hooked fish with medium (GN2) or high degrees of
295 damage (GN3). While the degree of physical trauma can be a good predictor of mortality
296 (Domeier et al. 2003, Skomal 2007), we recommend a more quantitative estimate of C&R
297 mortality by hooking location in dolphinfish using experimental caging (Grover et al. 2002,
298 Gutowsky et al 2015), large-scale mark-recapture study (Pine et al. 2003, Rudershausen et al.
299 2014), telemetry (Capizanno et al. 2016), or use of accelerometer loggers (Brownscombe et al.
300 2013, Lennox et al. 2018).

301 Of the studies that have examined injuries and mortality for roof-hooked fish, the results
302 have been mixed and are likely species-specific. Roof hooking has been observed and described
303 in other pelagic fishes (Faltermann and Graves 2002, Prince et al. 2002, Prince et al. 2007).
304 Faltermann and Graves (2002) assessed hooking mortality among pelagic fishes and determined a
305 discard mortality rate of 80% for fish hooked in the roof of the mouth; however, the sample size
306 was small (n=5), and the mortality rate determined for jaw-hooked fish (corner and lower jaw)
307 was also notably high (48.9%). The injuries to roof-hooked dolphinfish were very similar to
308 those described by Prince et al. (2002, 2007) for roof-hooked Atlantic sailfish *Istiophorus*

309 *platypterus*. Hooking in the roof of the mouth resulted in lacerations to the rear palate and
310 hemorrhaging of the eye in sailfish (Prince et al. 2007). The authors classified hooking in the
311 roof to be an undesirable location that may lead to post-release mortality due to latent injuries to
312 the eye. The resemblance of roof hooking injuries between our study and Prince's (2002, 2007)
313 findings are likely a result of similarities in anatomy, as both dolphinfish and sailfish have an
314 insubstantial palate. Among more distantly related fishes inhabiting different environments,
315 results for roof hooking were increasingly varied. For example, in cutthroat trout *Oncorhynchus*
316 *clarkii* individuals hooked in the jaw had an estimated mortality rate of 6%, while those hooked
317 in the roof of the mouth showed a mortality of 29%. (Pauley and Thomas 1993). In pumpkinseed
318 *Lepomis gibbosus* with molariform teeth, roof hooking was insignificant in discard mortality
319 estimates (Cooke et al. 2003), and in Chinook salmon, the roof of the mouth was designated as a
320 location with minimal injury and treated as a control for mortality estimates (Grover et al. 2002).
321 We recommend future research on hook injuries for fishes known to have mouth and eye
322 morphologies similar to dolphinfish and sailfish.

323 We observed severe injuries to a peripheral hooking location that outwardly does not
324 appear to result in severe injury. This has also been the case for roof injuries to bluefin tuna
325 *Thunnus thynnus*, in which the same injury could only be characterized by performing gross
326 necropsies (Belle 1997). In sharks, hooking damage to the basihyal was suggested to result in
327 high mortality, which was unexpected (Danylchuk et al. 2014). Serious injuries from hooking are
328 likely found in other fishes and is an area worthy of future research. Increased use of these
329 diagnostic tools for specific species and fisheries will aid to the understanding of hooking
330 injuries to different locations and allow anglers to make more informed decisions when
331 practicing catch-and-release.

332 Our research is unique in that it used detailed necropsy and medical imaging to reveal
333 cryptic hooking injuries. CT scanning may be a tool that C&R researchers choose to use in future
334 studies given the agreement between approaches and the time savings of CT scanning.
335 Additionally, CT scanning could be used as a first approach to identify severely-injured fish for
336 GNs. GNs were more insightful but required considerably more time than scanning. However,
337 GN validated the CT interpretation and revealed the mechanism and character of the respective
338 injuries.

339 While this study was specific to dolphinfish, we demonstrate a novel application of CT
340 techniques that are becoming more accessible with improved technology, free imaging software,
341 and scientific interest of scanning fish. In tandem with detailed necropsies, CT offers an
342 enhanced technique to characterize injuries that provides insight into potential risk for post-
343 release mortality. The application of similar methods to other fish species with similar anatomies
344 could expand our current understanding of the various injuries caused by hooking.

345

346 **ACKNOWLEDGEMENTS**

347 Funding for this work was provided by NOAA Saltonstall-Kennedy grant
348 #NA15NMF4270346 and a North Carolina State University CMAST summer fellowship. We
349 thank Brendan Runde, Jeffery Merrell, Claire Pelletier, and Kelsey Moore for assistance with
350 dolphinfish sampling, and Kate Archibald for assistance with gross necropsy.

351

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495

496 **Figure Descriptions**

497 Figure 1. Computed Tomography (CT) scan diagnoses of dolphinfish *Coryphaena hippurus* by
498 category. Each increment of scale bar on leftmost side of the image represents 1 cm.

499 1a. CT 1 – No visible damage or trauma to the suspensorium or to the orbit. Individual was
500 hooked in the jaw, and served as a control. Gas present in small quantities bilaterally in and
501 behind the eyes is attributed to decapitation and/or decomposition. Arrows indicate the intact
502 bone structure of the endopterygoid in transverse (left) and coronal (right) sections.

503 1b. CT 2 – Fracture to bone(s) forming the suspensorium paired with asymmetrical gas influx
504 continuous from the fracture site extending into the base of the orbit (gas is confined to the
505 orbital floor). Individual was hooked in the roof of the mouth. Arrows indicate fracture site of
506 endopterygoid (oblique, displaced). Gas is continuous from the oral cavity to the base of the
507 orbital floor.

508 1c. CT 3 – Severe fracture (displaced or comminuted) to bone(s) forming the suspensorium,
509 paired with asymmetrical gas influx continuous from the fracture site extending past the orbital
510 floor to the level of the optic nerve, extraocular muscles, or the eye. Individual was hooked in the
511 roof of the mouth. Left arrow indicates gas influx while right arrow indicates the fracture site of
512 endopterygoid (comminuted). Gas is continuous from the oral cavity past the orbital floor,
513 including around the globe.

514

515 Figure 2. Suspensorium of dolphinfish *Coryphaena hippurus* in lateral view (a) and ventral view
516 (b). Abbreviations: ecp=ectopterygoid; enp=endopterygoid; mpt=metapterygoid; pal=palatine;
517 para=parasphenoid; q=quadrate; vom=vomer.

518

519 Figure 1a.



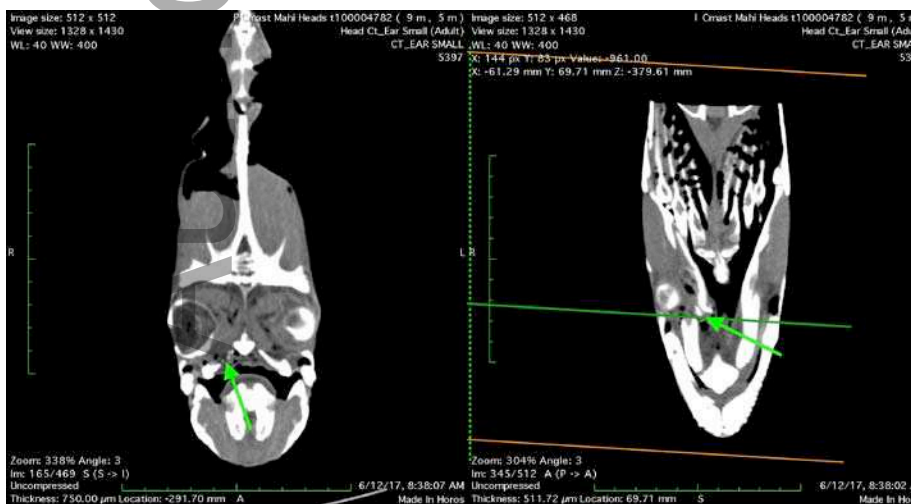
520

521 Figure 1b.

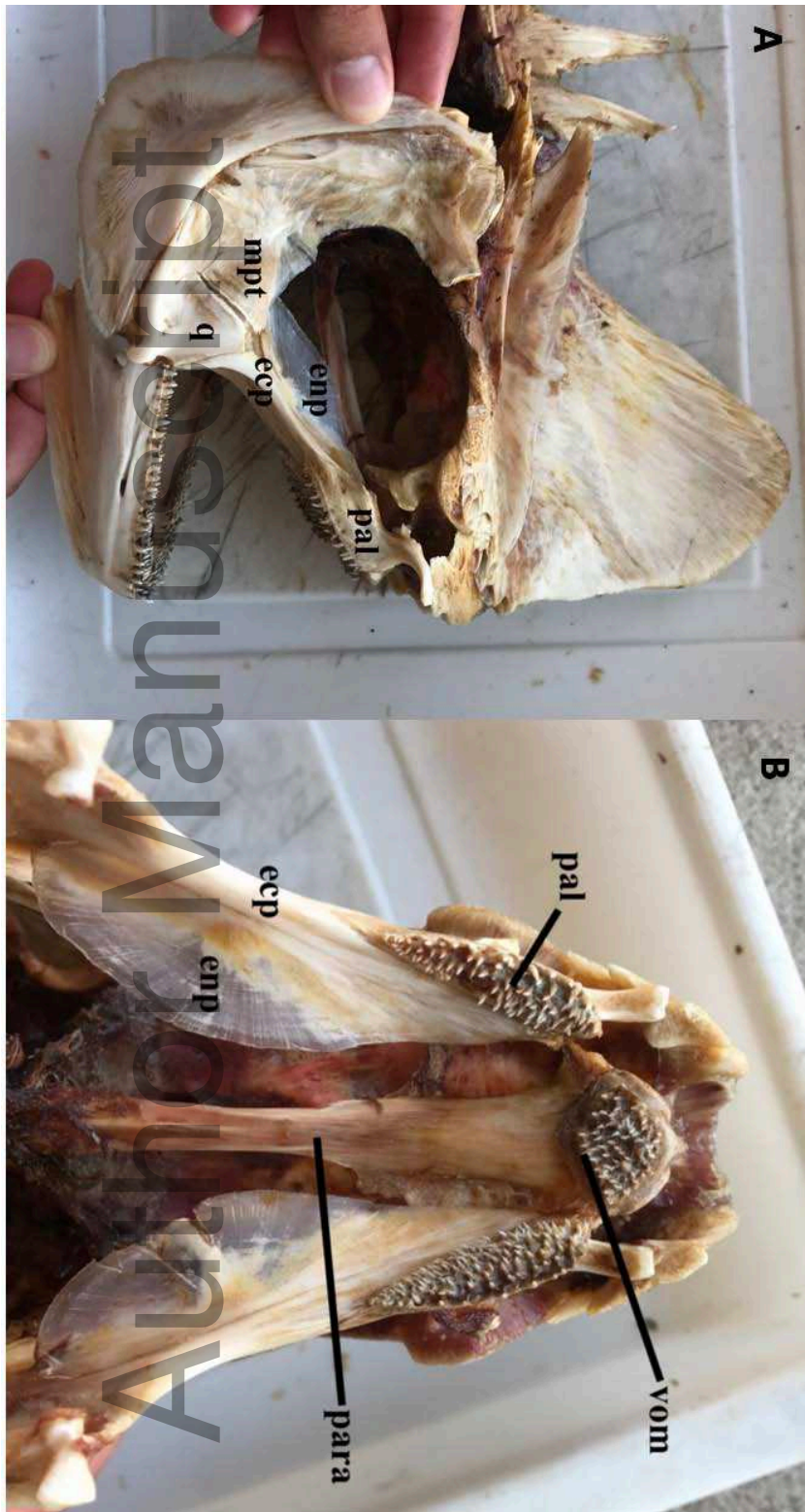


522

523 Figure 1c.



524



526

527 **Table Descriptions**

528 Table 1. Number of fish collected per dockside-designated hooking location, and mean and range
529 of fork lengths of all fish and locations. Four fish were not measured or included in these
530 averages.

531 Table 2. Computed tomography (CT) and gross necropsy (GN) categorization of all fish based
532 on dockside-designated hooking locations.

533

534 Table 1.

Hook location	Number of Fish	Average fork length (mm)	Range (mm)
All fish	42	679	480-985
Roof of the mouth	16	746	504-985
Eye	12	748	505-950
Jaw	14	584	480-880

535

536 Table 2.

	Roof (16)	Eye (12)	Jaw (14)	Total (42)
CT 1	5	1	14	20
CT 2	6	3	0	9
CT 3	5	8	0	13
GN 1	4	1	14	19
GN 2	5	4	0	9
GN 3	7	7	0	14

537