Validating Blueline Tilefish *Caulolatilus microps* Ages in the U.S. South Atlantic using Bomb Radiocarbon (F¹⁴C)

Kevin R. Spanik* and Joseph C. Ballenger

Marine Resources Research Institute, Marine Resources Division, South Carolina Department of Natural Resources, Charleston, SC, USA

217 Ft Johnson Rd Charleston SC 29412

* Corresponding author: email - <u>spanikk@dnr.sc.gov</u>; phone - 843-953-9857

-

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1111/jfb.15496

This article is protected by copyright. All rights reserved.

Validating Blueline Tilefish *Caulolatilus microps* Ages in the U.S. South Atlantic using Bomb Radiocarbon (F¹⁴C)

Abstract

Age validation is a critical component of an age-based stock assessment and subsequent species management. Our study used bomb radiocarbon analysis to validate age estimates of Blueline Tilefish *Caulolatilus microps*, a species for which regional stock assessment scientists have identified age validation as a high priority. We compared a *C. microps* F¹⁴C chronology to F¹⁴C chronologies for finfish of the U.S. South Atlantic Bight (SAB) and the northwest Atlantic (NWA). The high degree of correspondence in the chronologies exhibited for *C. microps* and other species of the SAB suggests a differential ¹⁴C uptake pattern in the SAB slope waters that is likely the result of local hydrological processes that delay ¹⁴C reaching the environments inhabited by these species. Our study was able to validate *C. microps* ages up to 25 years in the SAB, with strong evidence suggesting they are living to at least 50 years old.

Keywords: age validation, Blueline Tilefish, *Caulolatilus microps*, bomb radiocarbon

Introduction

Age estimates are critical for the population demographics calculations, agebased life history parameters (e.g., growth, mortality, stock productivity), and are required for age-structured population dynamics models. As such, age validation is a critical component of an age-based stock assessment and subsequent species management. The primary goal of any age validation study is to determine whether age estimates produced are, on average, correct for the species in question, with any associated error randomly distributed and no systematic bias (Francis et al. 2010). Therefore, the focus of age validation is to examine the bias of age determinations, not the precision of independent reads (Francis et al. 2010).

Validating age estimates can be a difficult and time-consuming task. However, there are a multitude of age validation methods available, including marginal increment analysis, radiometric dating, tag and re-capture, chemical tagging, and bomb radiocarbon analysis (Campana 2001). Bomb radiocarbon analysis is an accurate age validation method that has been successfully applied to studies of fishes around the world (Campana et al. 2008; Hamel et al. 2008; Horn et al. 2010; Andrews et al. 2015; Campana et al. 2016), including the southeastern coast of the united states (Lytton et al. 2016; Filer and Sedberry 2008; Friess and Sedberry 2011).

Two pieces of information are necessary to use bomb radiocarbon to validate an age: 1) a test data set that includes estimated ages and the associated radiocarbon values and 2) an accepted reference data set of ages and radiocarbon values for another species with a similar life history from the region and environment of interest (Francis et al. 2010). An implicit assumption of age validation using bomb radiocarbon is that the test and reference species occupy the same, or similar, environments with respect to ¹⁴C availability, so that the carbon incorporated into the carbonate structures of the two

species in the same year will contain the same proportion of ¹⁴C (Francis et al. 2010). In previous work, the test and reference data sets show a similar rapid increase in bomb radiocarbon (¹⁴C) levels beginning in the same year, with the timing at a specific locale being a function of local hydrographic processes (Francis et al. 2010). The timing of the ¹⁴C rise in oceanic waters relative to the peak observed in the atmosphere is later because of the time required for ¹⁴C to migrate from the atmosphere to specific marine region of interest (Francis et al. 2010). Due to the mixing rates of oceanic surface and deep waters, deeper oceanic waters have been shown to experience further delays in the timing of ¹⁴C rise (see Horn et al. 2010, Grammer et al. 2015, Campana et al. 2016).

Blueline Tilefish (*Caulolatilus microps*, Goode & Bean 1978) are a long-lived, commercially important, deepwater demersal species for which a recent stock assessment identified validation of an aging methodology as a high priority (SEDAR 2017). *C. microps* are patchily distributed at depths of 48-236 m, over irregular bottom characterized by ledges, boulders, or rubble piles along the outer continental shelf, shelf break, and upper Northwest Atlantic slope (Dooley 1978; Ross and Huntsman 1982; Parker and Mays 1998). A commercial fishery for *C. microps* along the southeastern United States coast began developing in the mid-1980s (Parker and Mays 1998; Harris et al. 2004). From 1993 to 2005, total annual landings of *C. microps* averaged 64.7 mt (range: 32.6-115 mt), before steeply increasing to a peak of 333 mt in 2008^a. *C. microps* landings moderated to a degree between 2009 and 2014, averaging 137 mt (range 63-214 mt), and have declined to an average of 38 mt (range 32-50 mt) between 2015 and 2021 as significant management regulation changes in the U.S. South Atlantic region based on the stock status determination that the stock was undergoing overfishing and currently overfished (SEDAR 2013, SEDAR 2017).

Deepwater species including *C. microps* (Harris et al. 2004; SEDAR 2013), are some of the most difficult species to age (Fenton et al. 1991; Peres and Haimovici 2004; Friess and Sedberry 2011; Lytton et al. 2016). Aging difficulty affects stock assessment accuracy and uncertainty and can lead to difficulties when managing a stock to prevent overfishing. Reducing assessment uncertainty for *C. microps* is important since deepwater species are notoriously susceptible to overfishing and slow rates of recovery because of general species longevity, slow growth, and slow maturation rates (Clark 2001; Clarke et al. 2003).

The goal of our study was to validate age estimates of *C. microps* (i.e., confirm assumed annual increment counts) from the U.S. South Atlantic Bight (SAB) by comparing onset of bomb-produced radiocarbon ($F^{14}C$) (Campana et al. 2008). We compare a *C. microps* bomb radiocarbon chronology to reference chronologies for finfish of the SAB and the northwest Atlantic (NWA). The findings of our study demonstrate the importance of appropriate reference chronology selection for bomb radiocarbon analysis and will have impacts on methods used to age *C. microps* for stock assessments in the U.S. South Atlantic.

Materials and Methods

The Marine Resource Monitoring, Assessment and Prediction program (MARMAP) of the South Carolina Department of Natural Resources (SCDNR) collects *C. microps* otoliths annually via routine fishery-independent sampling. The MARMAP program used a variety of sampling gears to collect *C. microps* including short bottom longlines, kali poles, experimental traps, and commercial bandit reels baited with either squid or clupeids. All otoliths in our study were collected by MARMAP between 1983 and 1997, from waters offshore of South Carolina (between 32.1 and 32.8°N latitude) in depths ranging from 57 to 210 m ($\overline{X} \pm SD = 180 \pm 6.9 m$).

Prior to selection of otoliths for use in age validation, two readers independently examined sectioned left otoliths to provide age estimates using a Leica M125 dissecting microscope under transmitted light at a magnification of 40 to 100X, without knowledge of fish size, capture date, or age estimated by the other reader. If two sections were available, readers used the section containing the core unless there was an obvious reason not to utilize that section, such as damage to the otolith. Increment counts were determined based on the counts of all opaque growth increments along the medial surface of the transverse otolith section dorsal to the sulcus (Figure 1). SCDNR age readers identified the first increment as the first opaque zone with a clear translucent zone separating it from the core; often this increment would appear as a doublet. If a doublet was not present, readers considered the first increment to be the first opaque zone with a clear translucent zone separating it from the core. We often encountered the groups of tightly compacted increments described by Harris et al. (2004), which typically occurred within the first few increments. The inner increments (up to ~ 6) were considered broad and diffuse, with outer (>6) increments becoming more regularly spaced, but tightly grouped. The inner increments were more clearly defined at 40-60X magnification, while outer increments were best read at 60-100X magnification. In many of the otolith sections, readability decreased at various points along the chosen reading axis, forcing readers to shift to a new reading axis by following a growth increment out along a lateral plane.

After initial increment counts, we selected twenty specimens for bomb radiocarbon age validation based on birth year as determined from increment count and year of capture (Table 1). We embedded the right sagittal otolith of each selected specimen in resin and obtained a single 1 mm thick transverse section through the core using a low-speed Isomet[®] saw. We washed the resultant section with deionized water

and allowed it to dry overnight, and then removed the extraneous otolith material surrounding the core using a Dremel[®] model 732 tool with a carbide-cutting wheel. To prevent cross-contamination we used a new carbide-cutting wheel for each otolith and performed core removal under a ventilation system. We removed additional surface contaminants by sonicating the extracted otolith cores for two 30-second time intervals in deionized water, followed by acid leaching in 10% HNO₃ for 30 seconds, and rinsing with deionized water. Post drying over-night, each otolith core was then measured to the nearest 0.01 mg to ensure enough material had been obtained for bomb radiocarbon analysis (>5 mg).

We shipped the resulting otolith cores in new sterile plastic 5 mL vials to the National Ocean Services Accelerator Mass Spectrometry (NOSAMS) facility at the Woods Hole Oceanographic Institution. Otolith core samples were then analyzed for $\Delta 14$ C using accelerator mass spectrometry (AMS) following standard methods (additional information can be found online: www.whoi.edu/nosams/radiocarbon-datacalculations). NOSAMS staff then calculate the fraction modern ($F^{14}C$), which is a measurement of the deviation of a sample's radiocarbon content from that of the 'modern' standard (Donahue et al. 1990; Reimer et al. 2004). 'Modern' is defined as 95% of the radiocarbon concentration (in AD 1950) of NBS Oxalic Acid I, normalized to $\delta^{13}C_{VPDB}$ ($\delta^{13}CVPDB = -19\%$; Karlen et al. 1964; Olsson 1970). A correction to $F^{14}C$ is made to normalize the sample result to a $\delta^{13}C_{VPDB}$ value of -25%, assuming a quadratic mass fractionation dependency using simultaneously measured ¹³C/¹²C ratios on the AMS system. An advantage of using $F^{14}C$, in contrast to $\Delta^{14}C$ as generally reported in fish bomb radiocarbon age validation studies, is that F¹⁴C does not change with time (i.e., does not depend on the year of measurement; Stenstrom et al. 2011). The resultant statistic, F¹⁴C, when plotted against birth year provides a measure of the

increase in ¹⁴C due to uptake of ¹⁴C from nuclear bomb testing in the 1950s through early 1970s compared to ¹⁴C levels found in 19th century wood.

In addition to the 20 samples selected for our study, 20 historical samples were included in our analyses from an unpublished bomb radiocarbon study performed in the early 2000's at the SCDNR (Table 1) that were also processed at the NOSAMS facility. We excluded one of these historical sample from further analyses due to lack of consensus among age readers (Table 1). We did not rely exclusively on the historical samples for the current age validation study because most of the estimated birth years fell between 1970 and 1986. When performing an age validation study using bomb radiocarbon it is best to use samples with presumed birth years that fall primarily during the period of drastic increase in atmospheric and surface water ¹⁴C concentrations due to nuclear testing, which occurred between 1958 and 1970 (Kalish, 1993).

A well-established ¹⁴C reference set of known age fish, coral, or other carbonate structures is not currently available for the U.S. South Atlantic region. However, there are three independent bomb radiocarbon studies of deepwater U.S. South Atlantic finfish species (Wreckfish (*Polyprion americanus*), Red Bream (*Beryx decadactylus*) and Barrelfish (*Hyperoglyphe perciformes*) that have been conducted using fish collected from the Charleston Bump area of the South Atlantic Bight (SAB) (Filer and Sedberry 2008; Friess and Sedberry 2011; Lytton et al. 2016). Considering the similar uptake pattern of ¹⁴C observed in these three independent studies and proximity to our study area, we chose to combine the radiocarbon data for Wreckfish, Red Bream and Barrelfish and use it as a SAB radiocarbon reference chronology (Table S1).

We also used a traditional finfish radiocarbon reference chronology from the Northwest Atlantic (NWA) for comparison due to its proximity to the study area in the same ocean basin and is influenced by the same water mass, and because it was used as a reference chronology in the studies that generated the SAB chronology. The NWA reference chronology is comprised of bomb radiocarbon data from Haddock (*Melanogrammus aeglefinus*), Red Fish (*Sebastes* spp.), yellowtail flounder (*Limanda ferruginea*) and bivalves collected in the NWA (Table S2; Campana et al. 2008).

We estimated the year of initial F¹⁴C increase using:

$$C_T = C_p - 0.9(C_P - C_L)$$

where C_T is the value corresponding to the 10% threshold contribution of F¹⁴C (Campana et al. 2008). C_P and C_L represent the peak and lowest values found within a chronology. The year of initial increase (Y_T) is then estimated as the first year the chronology exceeds C_T . Deviations in estimates of Y_T from test and reference data sets indicates potential biases in age estimates of the test data. All analyses were performed using the R Statistical Program version 3.3.1. Because all authors of reference data sets reported results in $\Delta^{14}C$, we used the R function

"AbsoluteFractionModern_from_Delta14C" available in the *SoilR* package (Sierra et al. 2012) to convert these to F¹⁴C measures.

Ethics statement

C. microps were collected under a letter of Acknowledgement from the National Oceanographic and Atmospheric Administration (NOAA) National Marine Fisheries Service and biological samples were obtained using methods of euthanasia consistent with recommendations from the American Fisheries Society.

(https://doi.org/10.47886/9781934874394.ch8)

Results

The *C. microps* otoliths used in the current bomb radiocarbon analysis (n=39) had increment counts ranging from 3 to 30, and estimated birth years (assuming one

increment formed per year) ranged from 1952-1987 (Table 1). The average percent error between SCDNR age readers for these otoliths was 6.94% with an average CV of 9.82%, and there was no apparent aging bias. Otolith core values of F¹⁴C ranged from 0.939 ($\Delta^{14}C = -62.49\%_0$) for a fish with an estimated birth year of 1961 to 1.099 ($\Delta^{14}C = 92.10\%_0$) for a fish with an estimated birth year of 1973 (Table 1). All values were well within the range of previously published F¹⁴C levels reported for finfish reference chronologies of the SAB (Table S1) and the NWA (Table S2).

Levels of $F^{14}C$ in otolith cores from *C. microps* increased prominently beginning in the early 1960s, peaked in the mid- to late-1970s, and declined toward the end of the time series. The $F^{14}C$ chronology of *C. microps* was similar to the SAB $F^{14}C$ chronology (Figure 2A). Compared to the NWA chronology (Figure 2B), there is a clear difference in uptake patterns, with an apparent delay in both peak $F^{14}C$ values and the onset of $F^{14}C$ increase for *C. microps*.

The onset of $F^{14}C$ increase for *C. microps* in our study was estimated to occur in 1960 (Table 2). Similarly, the estimated onset of $F^{14}C$ for the SAB reference chronology was 1959 (Table 2). For the NWA reference chronology, however, the onset of $F^{14}C$ increase was estimated to occur several years earlier in 1956 (Table 2).

The F¹⁴C chronologies for each of the deep-water finfish studies within the SAB reference were similar to that of *C. microps* in the current study, as evidenced by high overlap of the 95% confidence intervals for each species based on generalized additive model smoothers (Figure 3A). The updated SAB bomb radiocarbon reference chronology including the new data from the current study is shown in Figure 3B.

The uptake pattern of $F^{14}C$ in *C. microps* in our study was comparable with chronologies reported in studies of other species and areas, (e.g. Campana et al. 2008; Filer and Sedberry 2008; Friess and Sedberry 2011; Lytton et al. 2016) and supports interpreted increment counts as annual growth indicators in our *C. microps* otoliths. If the aging methodology had not been successful at identifying consistent structures, there would have been no observable uptake pattern or coherent time-series for *C. microps* in our study (Campana 1997). The outlier with an estimated birth year of 1952 and $F^{14}C$ value of 1.03 may be due to processing error where not all of the otolith material was removed to the core, or aging error attributed to the difficulty in aging deepwater species. However, it is not unreasonable to expect a level of stochasticity in the mechanisms by which different fish encounter different $F^{14}C$ levels due to the complex process in which atmospheric bomb $F^{14}C$ enters the marine environment and mixes to depth (Kalish, 1995).

One possible scenario to explain the discrepancy in estimating onset of $F^{14}C$ increase for *C. microps* between the two reference chronologies is that the aging methodology employed in the current study for *C. microps* results in biased age estimates, with age estimates representing under-ages relative to true age. Under this scenario, the obvious explanation for aging bias in *C. microps* derives from the difficulty of aging deepwater species (see Fenton et al. 1991; Peres and Haimovici 2004; Andrews et al. 2015; Campana et al. 2016), with the difficulty of increment identification in *C. microps* being reviewed in previous work (Ross and Huntsman 1982; Harris et al. 2004). General species longevity, slow growth, and slow maturation rates of deepwater species (Clark 2001; Clarke et al. 2003) leads to the formation of tightly compacted increments in older individuals. In addition, deepwater environments

-Author Manuscrip

are characterized by a lack of seasonality of the physical features of the environment (e.g., water temperature), necessitating that the formation of growth increments be driven by other factors (Swan and Gordon 2001). Ross and Huntsman (1982) suggested that water temperature was not the primary driver for increment formation in *C. microps*, as the annual range of bottom temperatures along the shelf-edge zone was just equivalent to the minimum range that could induce decreased growth in fish. They suggested increments in *C. microps* otoliths instead arise from a physiological rhythm or annual feeding cycle correlated with photoperiod. Alternatively, growth increments observed in deepwater finfish may reflect seasonal changes in prey availability derived from the surface environment (Swan and Gordon 2001). If under-aging is occurring, misinterpreted annuli are likely within the first several broad and diffuse increments that were grouped and interpreted as a single annulus by the SCDNR age readers in the current study.

A second possible scenario to explain the discrepancy in estimating onset of $F^{14}C$ increase for *C. microps* between the two reference chronologies is that currently employed aging methodology for *C. microps* results in un-biased age estimates. In this scenario, the lack of correspondence in the $F^{14}C$ chronology for *C. microps* and reference chronologies from the NWA is the result of regional differences in hydrological processes with respect to ¹⁴C. The apparent aging bias exhibited by *C. microps* relative to a geographically disparate reference chronology is not an uncommon occurrence for radiocarbon studies performed on deepwater fishes from the SAB region. Filer and Sedberry (2008), Friess and Sedberry (2011), and Lytton et al. (2016) all hypothesized that the apparent under-aging relative to a Haddock reference chronology (Campana 1997) in their respective studies may have resulted from differences in oceanographic conditions derived from localized upwelling events, regional differences

in onset of ¹⁴C increases in surface waters, and differential surface to depth mixing rates of water masses experienced in the two regions. A growing body of research is documenting the period of delay one would expect for the bomb signal to penetrate from surface to increasingly deeper waters. In the southwest Pacific Ocean, Grammer et al. (2015) suggested 5 to 10 years for radiocarbon to reach depths ~400-500 m, with initial increase not beginning until 1963 and not peaking until the early 1980s. In the northeast Atlantic, Nydal (1993) suggested a period of 18 years for F¹⁴C penetration to a depth of 1000 m; assuming a linear increase in depth through time, signal penetration to 200 m depth would take 3.6 years. Similarly, the bomb signal took 14 years to penetrate to a depth of 800 m in the Indian Ocean (Rubin and Key 2002); given the same calculations presented above, 3.5 years would be required to penetrate to a depth of 200 m. Finally, within the northwest Atlantic Campana et al. (2016) suggests a 9-year delay for the F¹⁴C signal to reach a depth of 390-450 m, relative to the same NWA reference chronology used in the current study. Friess and Sedberry (2011) could not attribute the observed phase-shift to this phenomenon because juvenile Red Bream have a long pelagic stage and, therefore, the F¹⁴C signal from the core in these otoliths would be expected to be similar to surface levels. While it is known that juveniles of Caulolatilus sp. inhabit demersal adult burrows as a means for predator avoidance (Able et al., 1987), habitat use by larval and age 0 C. microps is not precisely understood. Captive reared juveniles of the closely related tilefish species *Lopholatilus chamaeleonticeps* have been observed settling to the bottom and beginning to dig by 1.5 cm standard length (Fahay 1983), and analysis of δ 13C values and δ 15N values of eye lens protein suggest long single location residency throughout the lifespan (Vecchio et al., 2021).

The consistent offset between the NWA reference chronology and radiocarbon data from SAB deep-water species *C. microps* (the current study), Red Bream (Friess

and Sedberry 2011), Barrelfish (Filer and Sedberry 2008), and Wreckfish (Lytton et al. 2016) suggests unique environmental or oceanographic conditions in the study region. It is improbable that growth increment counts in each of these studies led to under-aging by the same proportional amount relative to the NWA reference chronology. A more likely explanation is that the NWA chronology was inappropriate for use as a reference curve for these species. Thus, we agree with the assessment of Campana et al. (2016) that it appears inevitable that the bomb signal would be delayed in reaching the depth of *C. microps* in the present study, with the delay resulting in the appearance of an apparent under-aging bias relative to the reference chronology from shallower waters in the NWA. This phenomenon is further supported by the appearance of slight over-aging (~1 year) of the *C. microps* in our study compared to the Red Bream, Barrelfish and Wreckfish included in the SAB reference chronology which all generally inhabit slightly deeper waters (375-700 m).

A strength of age validation using the bomb radiocarbon technique is that it allows for the validation of a maximum age because one can validate the age of individual fish up to the year of onset of increases in ¹⁴C to the fish's environment. Inspection of the ages of fish with estimated birth years in the early 1960's ($F^{14}C$ onset suggested introduction of ¹⁴C into the environment due to nuclear testing in 1960) in the current study indicates that *C. microps* ages are validated up to 25 years in the SAB. There were many specimens that we estimated to be 40-50 years old, but we did not select for bomb radiocarbon sampling because their presumed birth years did not fall within the period of drastic increase in atmospheric and surface water ¹⁴C concentrations due to nuclear testing. Because we validated annual increment formation, have consistency with our age estimates both between and within readers, and due to the prevalence of *C. microps* with ages of 40-50 years old in MARMAP

samples, we feel confident that *C. microps* live to at least 50 years, which is 10 years older than the maximum age of 40 years used in the most recent stock assessment (SEDAR 50).

In conclusion, the SAB reference chronology presented here provides reasonable evidence of a differential ¹⁴C uptake pattern in SAB deeper waters (~200-700 m deep) that is likely the result of local hydrological processes causing a delay of ¹⁴C reaching the local environments inhabited by these species. The results from C. microps in our study both agree with the properties of this unique regional chronology and contribute additional data that can be added to a SAB reference curve. We have provided methodology to accurately interpret annual growth increments of C. microps otoliths that will lead to more consistency in aging this deepwater species. We provide evidence that C. microps live to at least 50 years old (even older with under-aging error plausible), which would be a new benchmark maximum age for stock assessments in the region. The conflicting results of the aging methodology validation, depending on the reference chronology used for comparison, highlights the importance of using a reference chronology from the same, or similar, environments with respect to ¹⁴C availability. Cross-basin (Kastelle et al., 2016) and even within-basin (Wischniowski et al., 2015) comparisons have revealed chronologies with distinctly different properties between a validation and a reference data set, demonstrating how misapplication may lead to aging error. While our study provides a more appropriate bomb radiocarbon reference chronology for use within deeper waters of the SAB, the development of a true region-specific chronology in the SAB consisting of known-age specimens remains necessary to unequivocally distinguish between aging error and effects of oceanographic processes (Piner et al., 2005) and should be a future research goal. This chronology, along with additional information on the early life-history of C. microps

and other deep-water species would provide invaluable resources to confirm the results of the current study and further aid in the precision and accuracy of aging deep-water species in the region.

Acknowledgements

We thank the many folks who have participated in the MARMAP program sampling efforts over the years that have collected Blueline Tilefish. We thank Dr. Patrick Harris for the historical analyses of Blueline Tilefish otoliths for bomb radiocarbon. We thank Adam Lytton for his work preparing otolith core sections and with data analysis. We thank Drs. Marcel Reichert, Julie Vecchio, and Wally Bubley for internal reviews of this manuscript prior to submission that greatly improved the clarity of the work. We also would like to acknowledge the efforts of external reviewers that greatly improved the final article. This research was supported by NOAA grant NA11NMF4540174 (MARMAP) and MARFIN Grant No. NA17FF2870. This research represents contribution number XXXXX from the South Carolina Department of Natural Resources' Marine Resources Division.

Contributions

J.C.B. performed the majority of the data analyses. K.R.S. was highly involved in the research, and contributed to data generation, data analysis, and manuscript preparation

References

Able, K.W., D.C. Twichell, C.B. Grimes, and R.S. Jones. 1987. Tilefishes of the genus Caulolatilus construct burrows in the sea floor Bulletin of Marine Science 40(1): 1-10.

- Andrews, J. H., J. H. Choat, R. J. Hamilton, and E. E. DeMartini. 2015. Refined bomb radiocarbon dating of two iconic fishes of the Great Barrier Reef. Marine and Freshwater Research 66:305-316.
- Campana, S. E. 1997. Use of radiocarbon from nuclear fallout as a dated marker in the otoliths of Haddock, *Melanogrammus aeglefinus*. Marine Ecology Progress Series 150: 49-56.
- Campana, S. E. 2001. Accuracy, precision and quality control in age determination, including a review of the use and abuse of age validation methods. Journal of Fish Biology 59:197-242.
- Campana, S. E., A. E. Valentin, S. E. MacLellan, and J. B. Groot. 2016. Imageenhanced burnt otoliths, bomb radiocarbon and the growth dynamics of Redfish (*Sebastes mentella* and *S. fasciatus*) off the eastern coast of Canada. Marine and Freshwater Research 67:925-936.
- Campana, S. E., J. M. Casselman, and C. M. Jones. 2008. Bomb radiocarbon chronologies in the Arctic, with implications for the age validation of Lake Trout (*Salvelinus namaycush*) and other Arctic species. Canadian Journal of Fisheries and Aquatic Sciences 65:733-743.

Clark, M. 2001. Are deep-water fisheries sustainable? - the example of Orange Roughy (*Hoplostethus atlanticus*) in New Zealand. Fisheries Research 51:123-135.

Clarke, M. W., C. J. Kelly, P. L. Connolly, and J. P. Molloy. 2003. A life history approach to the assessment and management of deep-water fisheries in the Northeast Atlantic. Journal of the Northwest Atlantic Fisheries Society 31:401-411.

- Cook, M., G. R. Fitzhugh, and J. S. Franks. 2009. Validation of Yellowedge Grouper, *Epinephelus flavolimbatus*, age using nuclear bomb-produced radiocarbon. Environmental Biology of Fishes 86:461-472.
- Donahue, D. J., T. W. Linick, and A. J. T. Jull. 1990. Isotope-ratio and background corrections for accelerator mass-spectrometry radiocarbon measurements. Radiocarbon 32:135-142.
- Dooley, J. K. 1978. Systematic revision and comparative biology of the tilefishes (Perciformes: Branchiostegidae and Malacanthidae). Doctoral Dissertation, University of North Carolina, Chapel Hill, NC.

- Fenton, G. E., S. A. Short, and D. A. Ritz. 1991. Age determination of Orange Roughy, *Hoplostethus atlanticus* (Pisces: Trachichthyidae) using ²⁰¹Pb:²²⁶Ra disequilibria. Marine Biology 109:1972-202.
- Filer, K. R., and G. R. Sedberry. 2008. Age, growth and reproduction of the Barrelfish *Hyperoglyphe perciformis* (Mitchill) in the western North Atlantic. Journal of Fish Biology 72:861-882.
- Francis, R. I. C. C., S. E. Campana, and H. L. Neil. 2010. Validation of fish ageing methods should involve bias estimation rather than hypothesis testing: a proposed approach for bomb radiocarbon validations. Canadian Journal of Fisheries and Aquatic Sciences 67:1398-1408.
- Friess, C., and G. R. Sedberry. 2011. Age, growth, and spawning season of Red Bream (*Beryx decadactylus*) off the southeastern United States. Fishery Bulletin 109:20-33.
- Grammer, G. L., S. J. Fallon, C. Izzo, R. Wood, and B. M. Gillanders. 2015. Investigating bomb radiocarbon transport in the southern Pacific Ocean with otolith radiocarbon. Earth and Planetary Science Letters 424:59-68.
- Harris, P. J., D. M. Wyanski, and P. T. P. Mikell. 2004. Age, growth, and reproductive biology of Blueline Tilefish along the southeastern coast of the United States, 1982-1999. Transactions of the American Fisheries Society 133:1190-1204.
- Horn, P. L., H. L. Neil, L. J. Paul, and P. Marriott. 2010. Age validation and growth of Bluenose *Hyperoglyphe antarctica* using the bomb chronometer method of radiocarbon aging. Journal of Fish Biology 77:1552-1653.
- Kalish, J., M. 1993. Pre- and post-bomb radiocarbon in fish otoliths. Earth and Planetary Science Letters 114: 549-554.
- Kalish, J., M. 1995. Radiocarbon and fish biology. Recent development in fish otolith research. Edited by D.H. Secor, J.M. Dean, and S.E. Campana. University of South Carolina Press, Columbia, SC. pp. 637-653.
- Karlen, I., I. U. Olsson, P. Kallburg, and S. Kilici. 1964. Absolute determination of the activity of two ¹⁴C dating standards. Arkiv Geofysik 4:465-471.
- Kastelle, C. R., D. K. Kimura, and B. J. Goetz. 2008. Bomb radiocarbon validation of Pacific Ocean Perch (*Sebastes alutus*) using new statistical methods. Canadian Journal of Fisheries and Aquatic Sciences 65:1101-1112.

- Kastelle, C. R., T. E. Helser, S.G. Wischniowski, T. Loher, B. J. Goetz, and L.A. Kautzi. 2016. Incorporation of bomb-produced ¹⁴C into fish otoliths: A novel approach for evaluating age validation and bias with an application to yellowfin sole and northern rockfish. Ecologial Modelling 320:79-91.
- Lytton, A. R., J. C. Ballenger, M. J. M. Reichert, and T. I. Smart. 2016. Age validation of the North Atlantic stock of Wreckfish (*Polyprion americanus*), based on bomb radiocarbon (¹⁴C), and new estimates of life history parameters. Fishery Bulletin 114:77-88.
- Nydal, R. 1993. Application of bomb C-14 as a tracer in the global carbon cycle. Trends in Geophysical Research 2:355-364.
- Olsson, I. U. 1970. The use of Oxalic acid as a standard. Pages 17 *in* I. U. Olsson, editor. Radiocarbon Variations and Absolute Chronology. Nobel Symposium, 12th Proc., John Wiley & Sons, New York.
- Parker, R. O. J., and R. W. Mays. 1998. Southeastern U.S. deep-water reef fish assemblages, habitat characteristics, catches, and life history summaries. NOAA Technical Report NMFS 138, Seattle, WA.
- Peres, M. B., and M. Haimovici. 2004. Age and growth of southwestern Atlantic Wreckfish *Polyprion americanus*. Fisheries Research 66:157-169.
- Piner, K. R., and S. G. Wischniowski. 2004. Pacific Halibut chronology of bomb radiocarbon in otoltihs from 1944 to 1981 and a validation of ageing methods. Journal of Fish Biology 64:1060-1071.
- Reimer, P. J., T. A. Brown, and R. W. Reimer. 2004. Discussion: Reporting and calibration of post-bomb C-14 data. Radiocarbon 46:1299-1304.
- Ross, J. L., and G. R. Huntsman. 1982. Age, growth, and mortality of Blueline Tilefish from North Carolina and South Carolina. Transactions of the American Fisheries Society 111:585-592.
- Rubin, S. I., and R. M. Key. 2002. Separating natural and bomb-produced radiocarbon in the ocean: The potential alkalinity method. Global Biogeochemical Cycles 16.
- SEDAR. 2013. SEDAR 32 South Atlantic Blueline Tilefish Stock Assessment Report. SEDAR, North Charleston, SC. 378 pp. <u>http://sedarweb.org/docs/sar/S32_SA-BLT_SAR_FINAL_11.26.2013.pdf</u>.
- SEDAR. 2017. SEDAR 50 South Atlantic Blueline Tilefish Stock Assessment Report. SEDAR, North Charleston, SC. 542 pp. http://sedarweb.org/sedar-50

- Sierra, C. A., M. Mueller, and S. E. Trumbore. 2012. Models of soil organic matter decomposition: the SoilR package, version 1.0. Geoscientific Model Development 5:1045-1060.
- Stenstrom, K. E., G. Skog, E. Georgiadou, J. Genberg, and A. Johansson. 2011. A guide to radiocarbon units and calculations. Lund University, Department of Physics, Division of Nuclear Physics Interal Report LUNFD6 (NFFR-3111)/1-17/(2011), http://www.hic.ch.ntu.edu.tw/AMS/A%20guide%20to%20radiocarbon%20units %20and%20calculations.pdf.
- Swan, S. C., and J. D. M. Gordon. 2001. A review of age estimation in macrourid fishes with new data on age validation of juveniles. Fisheries Research 51:177-195.
- Vecchio, J. L., Ostroff, J. L., & Peebles, E. B. 2021. Isotopic characterization of lifetime movement by two demersal fishes from the northeastern Gulf of Mexico. Marine Ecology Progress Series, 657, 161-172.

Tables

Table 1: Summary of radiocarbon $(F^{14}C)$ results from Blueline Tilefish (<i>Caulolatilus microps</i>) otoliths collected off the southeast coast of the
United States via fishery-independent research of the MARMAP program. NOSAMS accession = identification number assigned by the Woods
Hole National Ocean Sciences Accelerator Mass Spectrometry Facility (13XXXX = contemporary sample; 4XXXX = historical sample);
SCDNR-ID = collection and specimen number for individual fish (project code is P05 for all); Kali = Kali Pole; Bandit = Bandit Reel; LBLL =
MARMAP long bottom longline gear; SBLL = MARMAP short bottom longline gear; Trap = MARMAP experimental fish trap; $\sigma = F^{14}C$ -sigma
value provided by NOSAMS; Age SE = the estimated age estimate standard error based on independent reads of the same fish by different
individuals experienced in aging Blueline Tilefish. Sample age = estimated age of the bomb radiocarbon sample, which is assumed to by 0.5 for
all fish as we are using the core material from the otolith; Sample year = birth year (determined from increment count and year of capture)
corrected for the "sample age" of the sample (e.g. Birth Year + Sample Age).

NOSAMS	SCDNR		Coll.	Latitude	Depth	TL					Age	Sample	Birth	Sample
accession	ID	Gear	year	(°N)	(m)	(mm)	F ¹⁴ C	σ	δ ¹³ C	Age	SE	age	year	year
136730	821608-1	Kali	1982	32.73	210	597	0.9388	0.0025	-2.12	21	2.9059	0.5	1961	1961.5
136731	821601-3	Kali	1982	32.73	210	655	0.9395	0.0027	-2.34	30	2.5899	0.5	1952	1952.5
136738	821612-5	Kali	1982	32.73	198	778	0.9422	0.0019	-2.94	22	1.9726	0.5	1960	1960.5
136731	821608-3	Kali	1982	32.73	210	589	0.9429	0.0024	-3.30	13	0.9689	0.5	1969	1969.5
136746	861436-4	Kali	1986	32.72	210	649	0.9476	0.0027	-2.10	25	0.9966	0.5	1961	1961.5
134740	834005-6	Bandit	1983	32.73	185	628	0.9509	0.0020	-2.30	18	1.2150	0.5	1965	1965.5

0	136744	851646-4	Kali	1985	32.63	177	588	0.9535	0.0022	-2.39	20	0.9184	0.5	1965	1965.5
	136737	821612-4	Kali	1982	32.73	198	580	0.9580	0.0021	-1.88	22	1.2857	0.5	1960	1960.5
\bigcirc	136748	861447-1	Kali	1986	32.63	185	550	0.9647	0.0019	-3.46	18	1.2000	0.5	1968	1968.5
S	136745	851919-1	Kali	1985	32.62	190	600	0.9669	0.0026	-2.74	24	0.9619	0.5	1961	1961.5
\square	136733	821609-1	LBLL	1982	32.73	196	714	0.9697	0.0028	-1.95	21	2.2887	0.5	1961	1961.5
	45054 ^a	834047-1	Kali	1983	32.73	198	584	0.9718	0.0042	-2.05					
	136743	840690-2	Kali	1984	32.73	201	526	0.9721	0.0021	-1.97	20	1.1429	0.5	1964	1964.5
σ	136739	821631-2	Kali	1982	32.73	209	596	0.9814	0.0021	-2.45	19	1.1429	0.5	1963	1963.5
\leq	136734	821609-2	Kali	1982	32.73	196	595	0.9894	0.0025	-3.68	16	2.0777	0.5	1966	1966.5
	136741	834053-4	LBLL	1983	32.73	207	722	1.0128	0.0022	-2.34	21	1.2509	0.5	1962	1962.5
<u> </u>	136735	821610-2	LBLL	1982	32.73	194	597	1.0190	0.0024	-2.63	9	1.3171	0.5	1973	1973.5
\bigcirc	136742	834061-1	Kali	1983	32.72	198	583	1.0240	0.0021	-2.56	16	1.0627	0.5	1967	1967.5
Ň	136747	861437-5	Kali	1986	32.73	194	567	1.0332	0.0021	-2.78	18	0.8921	0.5	1968	1968.5
	136732	821609-1	Kali	1982	32.73	196	778	1.0374	0.0030	-3.33	30	4.3716	0.5	1952	1952.5
	45063	861426-1	Kali	1986	32.74	190	574	1.0415	0.0041	-1.88	14	0.9368	0.5	1972	1972.5

\bigcirc	45059	851911-1	Kali	1985	32.54	201	478	1.0422	0.004	-2.83	13	0.5654	0.5	1972	1972.5
	45056	834062-1	LBLL	1983	32.73	207	494	1.0471	0.0052	-2.84	13	0.8081	0.5	1970	1970.5
\bigcirc	136736	821612-2	LBLL	1982	32.73	198	682	1.0509	0.0020	-3.22	15	0.7377	0.5	1967	1967.5
'n	45060	851918-5	Kali	1985	32.63	188	496	1.0548	0.0033	-3.10	10	0.7377	0.5	1975	1975.5
5	45057	840704-7	Kali	1984	32.74	199	528	1.0550	0.0037	-2.52	12	0.7825	0.5	1972	1972.5
	45062	861423-1	Kali	1986	32.73	201	640	1.0552	0.004	-3.53	9	1.3638	0.5	1977	1977.5
	45064	861432-5	Kali	1986	32.71	198	410	1.0583	0.005	-2.77	14	1.5000	0.5	1972	1972.5
σ	45055	834053-5	LBLL	1983	32.73	207	531	1.0595	0.0037	-3.50	10	0.6901	0.5	1973	1973.5
>	45061	851921-1	Kali	1985	32.63	181	532	1.0608	0.0037	-3.30	16	0.7190	0.5	1969	1969.5
	45068	981284-1	Trap	1998	32.84	174	609	1.0616	0.0036	-1.49	30	1.3801	0.5	1968	1968.5
	45069	971688-2	Trap	1997	32.75	183	563	1.0661	0.0039	-2.83	10	0.3401	0.5	1987	1987.5
\bigcirc	45066	971707-2	SBLL	1997	32.64	185	523	1.0686	0.0042	-3.10	12	0.4206	0.5	1985	1985.5
_	45065	971676-1	SBLL	1997	32.74	181	595	1.0703	0.0042	-4.22	10	0.4286	0.5	1987	1987.5
	45067	971708-3	Trap	1997	32.63	190	515	1.0750	0.0042	-2.78	11	0.5084	0.5	1986	1986.5
5	45050	831690-9	Bandit	1983	32.09	73	341	1.0795	0.0044	-4.48	3	0.4041	0.5	1980	1980.5

-

45051	831690-5	Bandit	1983	32.09	73	594	1.0828	0.0038	-3.49	9	0.5714	0.5	1974	1974.5
45052	831691-1	Trap ^b	1983	32.07	71	576	1.0874	0.0043	-3.66	3	1.2037	0.5	1980	1980.5
45053	831691-10	Trap ^b	1983	32.07	71	433	1.0901	0.0043	-4.79	4	0.6801	0.5	1979	1979.5
45058	851580-1	Bandit	1985	32.36	57	438	1.0992	0.0037	-4.10	12	0.8650	0.5	1973	1973.5

a – A consensus age for this specimen could not be determined, thus it was excluded from all further bomb radiocarbon analyses.

b –MARMAP Florida trap used to capture specimen

Table 2: Estimation of the onset of $F^{14}C$ increase using the methodology of Campana et al. (2008) for Blueline Tilefish and for each of the finfish reference chronologies considered. C_P and C_L represent the peak and lowest values found within a chronology. C_T is the value corresponding to the 10% threshold contribution of $F^{14}C$. The year of initial increase (Y_T) is then estimated as the first year the chronology exceeds C_T .

Set	C _P	C _L	C _T	Y _T ^a
Blueline Tilefish	1.0992	0.9388	0.9548	1960
SAB	1.1044	0.9322 ^b	0.9494	1959, 1965°
NWA	1.0680	0.9282	0.9422	1956

a – We did not consider any estimate of Y_T that was earlier than 1955; the onset of the F¹⁴C increase did not begin prior to 1955.

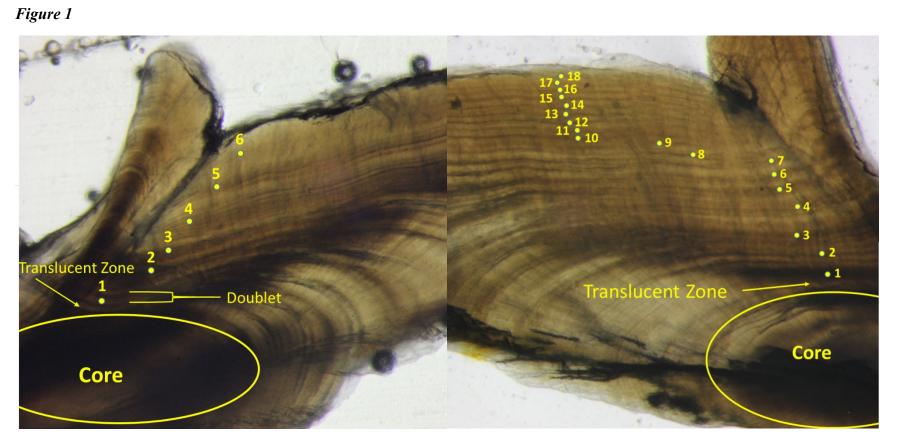
 $b-We \ excluded \ an \ outlier \ value \ of \ F^{14}C \ of \ 0.9109 \ (see \ Table \ S1) \ measured \ for \ Barrelfish \ from the \ calculation.$

c – First year that $F^{14}C$ exceeded C_T occurred in 1959; despite samples from other years between 1959 and 1965, $F^{14}C$ did not exceed C_T again until 1965.

-Author Manuscrip

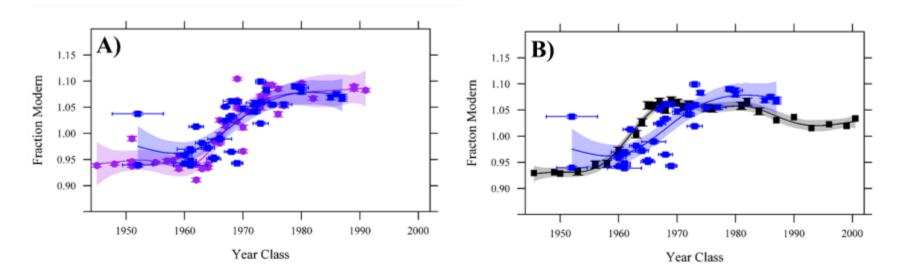
Figures

___ CLD uthor Manus



C aD \geq _ uth





 \bigcirc U Manu

Figure 3

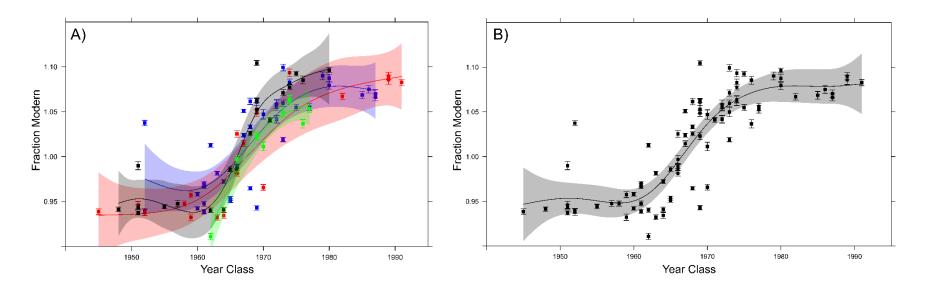


Figure Captions

Figure 1: Blueline tilefish otolith transverse section. Left panel – otolith thin section depicting a doublet that serves as the 1st increment as aged via the SCDNR; the outer increments appear as broad diffuse fields and were grouped. Right panel – otolith thin section lacking a clear doublet at the 1st increment; the first increment was considered the first opaque zone with a clear translucent zone separating it from the core along its entire length; subsequent increments appear as broad diffuse bands out to ~7 and then become more tightly compact and regularly spaced. Each dot represents and increment with its respective count.

Figure 2: Comparing the Blueline Tilefish $F^{14}C$ (blue) chronology to the A) SAB (purple) and B) NWA (black) reference chronologies. Horizontal and vertical error bars represent the standard error estimate for age estimates and the $F^{14}C \sigma$ from the bomb radiocarbon analysis, respectively. Shaded polygons represent 95% confidence intervals of $F^{14}C$ estimates based on generalized additive model smoothers.

Figure 3: A) Individual SAB species $F^{14}C$ chronologies for Wreckfish (black), Red Bream (red), Barrelfish (green), and Blueline Tilefish (blue) and B) updated SAB reference chronology including data from the current study. Horizontal and vertical error bars represent the standard error estimate for age estimates and the $F^{14}C \sigma$ from the bomb radiocarbon analysis, respectively. Shaded polygons represent 95% confidence intervals of $F^{14}C$ estimates based on generalized additive model smoothers. Tables

Table 1: Summary of radiocarbon $(F^{14}C)$ results from Blueline Tilefish (<i>Caulolatilus microps</i>) otoliths collected off the southeast coast of the
United States via fishery-independent research of the MARMAP program. NOSAMS accession = identification number assigned by the Woods
Hole National Ocean Sciences Accelerator Mass Spectrometry Facility (13XXXX = contemporary sample; 4XXXX = historical sample);
SCDNR-ID = collection and specimen number for individual fish (project code is P05 for all); Kali = Kali Pole; Bandit = Bandit Reel; LBLL =
MARMAP long bottom longline gear; SBLL = MARMAP short bottom longline gear; Trap = MARMAP experimental fish trap; $\sigma = F^{14}C$ -sigma
value provided by NOSAMS; Age SE = the estimated age estimate standard error based on independent reads of the same fish by different
individuals experienced in aging Blueline Tilefish. Sample age = estimated age of the bomb radiocarbon sample, which is assumed to by 0.5 for
all fish as we are using the core material from the otolith; Sample year = birth year (determined from increment count and year of capture)
corrected for the "sample age" of the sample (e.g. Birth Year + Sample Age).

NOSAMS	SCDNR		Coll.	Latitude	Depth	TL					Age	Sample	Birth	Sample
accession	ID	Gear	year	(°N)	(m)	(mm)	F ¹⁴ C	σ	δ ¹³ C	Age	SE	age	year	year
136730	821608-1	Kali	1982	32.73	210	597	0.9388	0.0025	-2.12	21	2.9059	0.5	1961	1961.5
136731	821601-3	Kali	1982	32.73	210	655	0.9395	0.0027	-2.34	30	2.5899	0.5	1952	1952.5
136738	821612-5	Kali	1982	32.73	198	778	0.9422	0.0019	-2.94	22	1.9726	0.5	1960	1960.5
136731	821608-3	Kali	1982	32.73	210	589	0.9429	0.0024	-3.30	13	0.9689	0.5	1969	1969.5
136746	861436-4	Kali	1986	32.72	210	649	0.9476	0.0027	-2.10	25	0.9966	0.5	1961	1961.5
134740	834005-6	Bandit	1983	32.73	185	628	0.9509	0.0020	-2.30	18	1.2150	0.5	1965	1965.5

0	136744	851646-4	Kali	1985	32.63	177	588	0.9535	0.0022	-2.39	20	0.9184	0.5	1965	1965.5
- <u> </u>	136737	821612-4	Kali	1982	32.73	198	580	0.9580	0.0021	-1.88	22	1.2857	0.5	1960	1960.5
$\overline{\bigcirc}$	136748	861447-1	Kali	1986	32.63	185	550	0.9647	0.0019	-3.46	18	1.2000	0.5	1968	1968.5
$\widetilde{\mathbf{O}}$	136745	851919-1	Kali	1985	32.62	190	600	0.9669	0.0026	-2.74	24	0.9619	0.5	1961	1961.5
<u> </u>	136733	821609-1	LBLL	1982	32.73	196	714	0.9697	0.0028	-1.95	21	2.2887	0.5	1961	1961.5
	45054 ^a	834047-1	Kali	1983	32.73	198	584	0.9718	0.0042	-2.05					
	136743	840690-2	Kali	1984	32.73	201	526	0.9721	0.0021	-1.97	20	1.1429	0.5	1964	1964.5
σ	136739	821631-2	Kali	1982	32.73	209	596	0.9814	0.0021	-2.45	19	1.1429	0.5	1963	1963.5
\leq	136734	821609-2	Kali	1982	32.73	196	595	0.9894	0.0025	-3.68	16	2.0777	0.5	1966	1966.5
	136741	834053-4	LBLL	1983	32.73	207	722	1.0128	0.0022	-2.34	21	1.2509	0.5	1962	1962.5
<u> </u>	136735	821610-2	LBLL	1982	32.73	194	597	1.0190	0.0024	-2.63	9	1.3171	0.5	1973	1973.5
\bigcirc	136742	834061-1	Kali	1983	32.72	198	583	1.0240	0.0021	-2.56	16	1.0627	0.5	1967	1967.5
	136747	861437-5	Kali	1986	32.73	194	567	1.0332	0.0021	-2.78	18	0.8921	0.5	1968	1968.5
	136732	821609-1	Kali	1982	32.73	196	778	1.0374	0.0030	-3.33	30	4.3716	0.5	1952	1952.5
	45063	861426-1	Kali	1986	32.74	190	574	1.0415	0.0041	-1.88	14	0.9368	0.5	1972	1972.5

0	45059	851911-1	Kali	1985	32.54	201	478	1.0422	0.004	-2.83	13	0.5654	0.5	1972	1972.5
	45056	834062-1	LBLL	1983	32.73	207	494	1.0471	0.0052	-2.84	13	0.8081	0.5	1970	1970.5
\bigcirc	136736	821612-2	LBLL	1982	32.73	198	682	1.0509	0.0020	-3.22	15	0.7377	0.5	1967	1967.5
$\widetilde{\Omega}$	45060	851918-5	Kali	1985	32.63	188	496	1.0548	0.0033	-3.10	10	0.7377	0.5	1975	1975.5
	45057	840704-7	Kali	1984	32.74	199	528	1.0550	0.0037	-2.52	12	0.7825	0.5	1972	1972.5
	45062	861423-1	Kali	1986	32.73	201	640	1.0552	0.004	-3.53	9	1.3638	0.5	1977	1977.5
	45064	861432-5	Kali	1986	32.71	198	410	1.0583	0.005	-2.77	14	1.5000	0.5	1972	1972.5
σ	45055	834053-5	LBLL	1983	32.73	207	531	1.0595	0.0037	-3.50	10	0.6901	0.5	1973	1973.5
\leq	45061	851921-1	Kali	1985	32.63	181	532	1.0608	0.0037	-3.30	16	0.7190	0.5	1969	1969.5
	45068	981284-1	Trap	1998	32.84	174	609	1.0616	0.0036	-1.49	30	1.3801	0.5	1968	1968.5
<u> </u>	45069	971688-2	Trap	1997	32.75	183	563	1.0661	0.0039	-2.83	10	0.3401	0.5	1987	1987.5
\bigcirc	45066	971707-2	SBLL	1997	32.64	185	523	1.0686	0.0042	-3.10	12	0.4206	0.5	1985	1985.5
	45065	971676-1	SBLL	1997	32.74	181	595	1.0703	0.0042	-4.22	10	0.4286	0.5	1987	1987.5
+	45067	971708-3	Trap	1997	32.63	190	515	1.0750	0.0042	-2.78	11	0.5084	0.5	1986	1986.5
	45050	831690-9	Bandit	1983	32.09	73	341	1.0795	0.0044	-4.48	3	0.4041	0.5	1980	1980.5

45051	831690-5	Bandit	1983	32.09	73	594	1.0828	0.0038	-3.49	9	0.5714	0.5	1974	1974.5
45052	831691-1	Trap ^b	1983	32.07	71	576	1.0874	0.0043	-3.66	3	1.2037	0.5	1980	1980.5
45053	831691-10	Trap ^b	1983	32.07	71	433	1.0901	0.0043	-4.79	4	0.6801	0.5	1979	1979.5
45058	851580-1	Bandit	1985	32.36	57	438	1.0992	0.0037	-4.10	12	0.8650	0.5	1973	1973.5

a – A consensus age for this specimen could not be determined, thus it was excluded from all further bomb radiocarbon analyses.

b –MARMAP Florida trap used to capture specimen

Table 2: Estimation of the onset of $F^{14}C$ increase using the methodology of Campana et al. (2008) for Blueline Tilefish and for each of the finfish reference chronologies considered. C_P and C_L represent the peak and lowest values found within a chronology. C_T is the value corresponding to the 10% threshold contribution of $F^{14}C$. The year of initial increase (Y_T) is then estimated as the first year the chronology exceeds C_T .

Set	C _P	C _L	C _T	Y _T ^a
Blueline Tilefish	1.0992	0.9388	0.9548	1960
SAB	1.1044	0.9322 ^b	0.9494	1959, 1965°
NWA	1.0680	0.9282	0.9422	1956

a – We did not consider any estimate of Y_T that was earlier than 1955; the onset of the F¹⁴C increase did not begin prior to 1955.

b – We excluded an outlier value of $F^{14}C$ of 0.9109 (see Table S1) measured for Barrelfish from the calculation.

c – First year that $F^{14}C$ exceeded C_T occurred in 1959; despite samples from other years between 1959 and 1965, $F^{14}C$ did not exceed C_T again until 1965.

Author Manuscript