

1 TEMPORAL AND SPATIAL COMPARISONS OF OCEAN QUAHOG (*ARCTICA*
2 *ISLANDICA*) GROWTH AND LIFESPAN ON THE MID-ATLANTIC CONTINENTAL
3 SHELF DURING INSHORE TRANSGRESSIONS OF THEIR RANGE FROM THE
4 NEOGLACIAL THROUGH THE TWENTIETH CENTURY

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Abstract

Arctica islandica provide long-term records of climate change on the U.S. northeast continental shelf transgressing and regressing across the shelf numerous times synchronously with cold and warm climatic periods. The availability of *A. islandica* in the death assemblage over a wide geographic and temporal range makes this species well suited for documenting both spatial and temporal influences of climate change in the Mid-Atlantic through the correlation of growth rates in response to changing water conditions. This study focuses on comparing regional growth of subfossil ocean quahogs obtained offshore of the Delmarva Peninsula (Delmarva), and living during the cold periods since the Holocene Climate Optimum, with living *A. islandica* from offshore New Jersey, offshore Long Island, and Georges Bank. These populations exhibited different growth rates, with subfossil individuals from Delmarva death assemblages, representing previous Holocene cold periods, having growth rates as greater than or equal the growth rates of living individuals. Moreover, the growth rates for subfossil *A. islandica* from Delmarva that were alive from 1740-1940 were more rapid than contemporaneous individuals of the same age alive today. Higher growth rates for *A. islandica* from off Delmarva suggest that conditions supported near maximum growth during the cold periods after the Holocene Climate Optimum, possibly due to increased food supply in water shallower than that inhabited today. Unlike many bivalves, evidence for range recession of *A. islandica* as bottom water temperatures warm is found first in juvenile abundance, suggesting that recruitment ceases long before the population's demise: range recession in this species is a 100+ year process determined by the survivorship of the oldest and largest individuals. This study is the largest spatial and temporal growth comparison of *A. islandica* ever recorded and the first record of the process by which this species' inshore range regresses as temperatures rise.

50 **1.1 Introduction**

51 Organism growth is controlled by ontogeny, genetics, and the environment (Hemeon et
52 al., 2021a). Comparing growth over time between cohorts within and between populations
53 allows one to isolate the effects of a species' surrounding environments to determine the
54 influence of environmentally-driven changes in growth (Black et al., 2008; Peharda et al., 2019;
55 Hemeon et al., 2023). By comparing temperature regimes and climatic events to known periods
56 of growth, beneficial, neutral, or detrimental effects on growth can be inferred from
57 chronological growth increments recorded in the hard parts of an organism (Richardson, 2001;
58 Killam and Clapham, 2018). For example, annual lines (annuli) deposited in the shells of bivalve
59 molluscs (similar to the rings in a tree) capture a record of environmental information from the
60 surrounding habitat (Peterson et al., 1985; Richardson, 2001; Kraeuter et al., 2007; Ridgeway et
61 al., 2011; Peharda et al., 2019). The distance between sequential annuli varies within a shell,
62 recording the suitability of environmental conditions during each growth period. These annuli
63 result from a changing rate of carbonate deposition during periods of slower or faster growth
64 relative to the continuing deposition of organic matrix. Hence, the sampling of bivalves from a
65 range of spatial, temporal, and environmental settings can provide consistent and accurate
66 records of the individual's chronological age and local climate (Jones et al., 1984; Austad, 1996;
67 Brey et al., 2011; Ridgeway et al., 2011; Shirai et al., 2018).

68 *Arctica islandica*, commonly called the ocean quahog, is a boreal clam with a habitat
69 range extending along the mid-Atlantic continental shelf and throughout most boreal seas in the
70 northern hemisphere. Ocean quahogs have a well-known sensitivity to temperature, with an
71 upper thermal limit of ~15°C, and an extensive lifespan often exceeding 300 years in age with
72 the oldest aged at 507 years (Butler et al., 2013). These characteristics make *A. islandica* well

73 suited for documenting the spatial and temporal influence of climate change, and previous
74 investigations of this clam species span the majority of the North Atlantic, including the Mid-
75 Atlantic region (Witbaard, 1996; Dahlgren et al., 2000; Witbaard and Bergman, 2003;
76 Wanamaker et al., 2011; Butler et al., 2013; Reynolds et al., 2017; Hemeon et al., 2021a).

77 On the U.S. Mid-Atlantic continental shelf, *A. islandica* is found at latitudes farther south
78 than the normal boreal provincial boundary (Hale, 2010). This unusual southern extension in
79 range is a result of the Cold Pool, a body of cold bottom water trapped by thermal stratification
80 during the late spring to early fall which maintains mean summer temperatures typically at
81 13.5°C or lower, with fall temperatures rarely exceeding 16°C when stratification breaks down
82 (Lentz, 2017; Chen et al., 2018; Chen and Curchitser, 2020; Friedland et al., 2022).

83 As a result of the environmental information archived in the growth record of *A.*
84 *islandica*, many studies have been conducted examining the determinants of growth in this long-
85 lived species, with particular emphasis on temperature and food supply (Schöne et al., 2005;
86 Wanamaker et al., 2009; Begum et al., 2010; Vihtakari et al., 2016; Ballesta-Artero et al., 2017).
87 Ocean quahogs are unique in their tendency for continuous growth into old age which limits the
88 application of growth models that describe asymptotic growth at old age, typical of most bivalve
89 species (e.g., McCuaig and Green, 1983; Devillers et al., 1998; Luquin-Cavarrubias et al., 2016).
90 Hemeon et al. (2023) determined that the best-fit growth model for ocean quahogs is a modified
91 Tanaka model (MT) when compared to both the traditional Tanaka model (Tanaka, 1982, 1988)
92 proposed by Pace et al. (2017a) and the von Bertalanffy model commonly used (e.g., Solidoro et
93 al., 2000; Appleyard and DeAlteris, 2001; Kilada et al., 2009; Brey et al., 2011; Chute et al.,
94 2016). The original Tanaka growth model, a power growth function, fits animals with
95 indeterminate growth (Pace et al., 2017a), which is the case for ocean quahogs, but even so, it

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96 tends to underestimate growth of *A. islandica* at old age. Consequently, Hemeon et al. (2021a)
97 modified the traditional Tanaka model (Tanaka 1982, Pace et al. 2017b) by adding an additional
98 parameter, *g*, which permits the model to better estimate length at old age.

99 A primary objective of this study is to compare growth dynamics in existing and past
100 ocean quahog populations from the U.S. Mid-Atlantic continental shelf. LeClaire et al. (2022)
101 documented the presence of subfossil *A. islandica* shells in death assemblages sampled on the
102 continental shelf off the Delmarva Peninsula inshore of the species' present depth range. These
103 encompassed animals that lived during all four major cold events since the Holocene Climate
104 Optimum (60-4400 cal years BP), including the two cold period events during the Neoglacial,
105 the Dark Ages Cold Period, and the Little Ice Age. Given this spatial and temporal distribution of
106 ocean quahogs, LeClaire et al., (2022) proposed that the inshore boundary of the Cold Pool had
107 transgressed and regressed across the continental shelf a number of times in the past and that
108 during the transgressions associated with colder climates *A. islandica* occupied the inner to
109 middle portion of the continental shelf inshore of their range today. The objective of this study is
110 to compare the growth rates of subfossil ocean quahogs both spatially and temporally amongst
111 themselves and with recently documented growth rates for living *A. islandica* obtained from
112 three different regions in the U.S. Mid-Atlantic: New Jersey, Long Island, and Georges Bank
113 (Hemeon et al., 2021a; Hemeon et al., in press; Sower et al., 2023b).

114 Notably, Pace et al., (2017b), Hemeon et al. (2023), and Sower et al., (2022, 2023b)
115 document the extraordinary variability in *A. islandica* growth rates as a function of birth year for
116 *A. islandica* living in these Mid-Atlantic populations. Clams born in a given birth year integrate
117 the subsequent climatology over their lifetimes, developing a growth curve that can differ from
118 *A. islandica* born in other birth years. These differences document the sensitivity of the species

119 to temperature change, a sensitivity that produces a differential growth dynamic based on the
120 variance in temperature exposure for each cohort. Hence, comparisons can describe differences
121 in growth over space and time, revealing the influence of biogeographic location and climate
122 change on ocean quahog growth. Subsequent findings offer potential to identify the temperature
123 history of the past populations alive during historical climate events, such as those identified by
124 LeClaire et al., (2022), based on comparisons to extant populations.

125 **1.2 Materials and Methods**

126 **1.2.1 Sample processing**

127 Growth data for living ocean quahogs were obtained from three different studies
128 including animals collected off New Jersey (39 50.40'N, 72 49.20' W; Sower et al., 2023a),
129 Georges Bank (40 43.66'N, 67 48.32'W; Hemeon et al., 2021a), and Long Island (40 5.95'N, 73
130 00.74' W; Hemeon et al., 2023a) in 2017 and 2019 (Fig 1). Growth data from subfossil shells
131 were obtained from locations reported by LeClaire et al., (2022), inshore and South of the Cold
132 Pool (Figs 1-3). The living ocean quahogs were collected within the extent of the Cold Pool
133 between 1959-2021 (Fig 3). Live and dead ocean quahogs were collected along the continental
134 shelf using hydraulic dredges towed by commercial clam vessels. For live animals, a standard
135 commercial ocean quahog hydraulic dredge was used (see Poussard et al., 2021). For subfossil
136 collections, the dredge was lined with 1-inch-square wire on the bottom surface and knife shelf.
137 Material collected by the dredge was sorted by clam species, live ocean quahog samples were
138 shucked, and both live and subfossil shells were retained and archived for later analysis.
139 Samples selected from the archive were processed through a standard procedure described by
140 Pace et al. (2017a) and Hemeon et al. (2021b). Each shell was cut along the maximum growth
141 axis using a Kobalt wet tile saw. After being cut, shells were then ground using abrasive paper of

142 increasing grit gauge (240, 320, 400, and 600) and polished with a polycrystalline diamond
143 suspension fluid (6 μm and 1 μm diamond sizes) (Pace et al., 2017a; Hemeon et al., 2021b) to
144 obtain a mirror finish. Shell hinges were imaged using a high-definition Olympus DP73 digital
145 microscope camera. For subfossil shells and a subsample of live-collected shells (see LeClaire et
146 al., 2022), after photographing the hinge, shell powder samples were extracted from the central
147 portion of the interior edge of the shell near the hinge and umbo, using a Dremel tool to obtain
148 carbonate that represented the youngest part of the shell hinge (i.e., carbonate deposited during
149 the earliest growth). The powder sample (>10 mg) was sent to the W. M. Keck Carbon Cycle
150 Accelerator Mass Spectrometry Laboratory at the University of California, Irvine for analysis.
151 Radiocarbon dates were calibrated according to the methods in LeClaire et al. (2022).

152 **1.2.2 Aging**

153 Following the methods described in Pace et al. (2017a), images of the shell hinge plate
154 were annotated using ImageJ software to estimate age. Reader precision was evaluated using the
155 double-blind technique of Hemeon et al. (2021b) who provide a detailed evaluation of reader
156 precision in the aging of *A. islandica* also applicable to this study. Accuracy was evaluated by
157 comparing radiocarbon dated specimens of known date-of-death with reader ages as described by
158 LeClaire et al. (2022). Annual growth increments were measured in pixels by the ObjectJ plugin
159 for ImageJ (Pace et al., 2018; Hemeon et al., 2021b). Annual lines observed on the shell hinge
160 plate are proportional to increments on the outer shell valve (Mann, unpubl. data); therefore,
161 annual growth increments on the hinge plate were converted to annual growth increments of the
162 total shell length using the ratio of the hinge plate length to total length after conversion of pixel
163 dimension to mm (Pace et al., 2018; Hemeon et al., 2021b). Supplementary information on *A.*
164 *islandica* shell processing procedures can be found at:

165 https://www.vims.edu/research/units/labgroups/molluscan_ecology/publications/topic/ocean_quahog_arctica/index.php

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168 **1.2.3 Growth Model**

169 The growth increments for each shell were cumulatively summed to create a growth
170 curve for that specific individual. Using the Akaike information criterion (AIC), Hemeon et al.
171 (2021a) determined that the modified Tanaka model (MT) was the best-fit growth model for
172 ocean quahog growth (Eq 1):

$$173 \quad \text{Eq (1)} \quad L_t = d + \frac{1}{\sqrt{f}} \log(2f(t - c) + 2\sqrt{f^2(t - c)^2 + fa}) + gt^{2.5}$$

174 where L_t is the shell length in mm at a given age in years (t). The MT model parameters can be
175 understood as follows. Parameter c (yr) denotes the age at maximum growth rate. At the age of
176 maximum growth, c , the growth rate is $1/\sqrt{a}$. Therefore, parameter a ($\text{yr}^2 \text{mm}^{-2}$) describes the
177 maximum growth rate which will occur at age c . Parameter f (yr^{-2}) controls the rate at which
178 growth declines with increasing age. For older animals, growth rate reduces to $1/(t\sqrt{f})$.
179 Parameters d (mm) and g ($\text{mm yr}^{-2.5}$) are scalars of size, with g influencing the rate of growth
180 rate decline with increasing age determined by parameter f . All MT model parameters except d ,
181 were forced to be ≥ 0 during model convergence to prevent the estimation of negative square
182 roots and logarithms.

183 For comparison with data from Hemeon et al. (2021a, 2023b) and Sower et al. (2023b),
184 dead shells reported in LeClaire et al., (2022) that were born after 1700 were divided into 20-
185 year groups defined by birth year.

186 **1.2.4 Death Dates**

187 The age-at-death of each subfossil shell collected off Delmarva and reported in LeClaire

188 et al. (2022) was added to the radiocarbon age (assumed to approximate the birth year of the
189 animal), to determine the year of death.

190 **1.2.5 Temperature Model**

191 Using data from Hemeon et al. (2021a, 2023a), animals collected from Georges Bank and
192 Long Island were compared to bottom water temperatures for these collection locations. Bottom
193 temperature data were obtained from the simulations of the Northwest Atlantic Ocean in the
194 Regional Ocean Modelling System (ROMS-NWA; Shchepetkin and McWilliams, 2005) for the
195 period 1959-1992 and from the Global Ocean Physics Reanalysis (Glorys12v1 reanalysis;
196 Lellouche et al., 2021) for the period 1993-2017. Based on the methodology developed by du
197 Pontavice et al. (2022), bottom temperature from ROMS-NWA used for the period 1959-1992
198 were bias-corrected using the monthly climatologies of observed bottom temperature from the
199 Northwest Atlantic Ocean regional climatology (NWARC) (Seidov et al., 2016). The extent of
200 the Cold Pool was estimated between 1959 and 2021 (Fig 3) calculated using a high-resolution
201 long-term bottom temperature product described in du Pontavice et al. (2023).

202 A generalized additive model (GAM) was fit to the yearly maximum values obtained
203 from the bottom temperature dataset compared to the yearly growth of animals to describe the
204 relationship between growth and temperature. Several temperature metrics were investigated;
205 yearly maximum temperature was determined to have the largest and most significant effect on
206 growth. The majority of temperatures between 1958-2017 fall within the range of temperatures
207 that ocean quahogs are able to tolerate (5-15 °C). Consequently, using the maximum temperature
208 enabled a focus on the temperatures at the edge or outside of this clam's temperature tolerance.
209 The GAMs were fit in R Statistical Software (v4.1.2; R Core Team 2021) using the mgcv
210 package (v1.8-34; Wood, 2011). Growth was the response variable with maximum yearly

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211 temperature, sex, location, and animal age in that growth year used as predictor variables to
212 explain the differences in growth. The model was tested with `gam.check()` to determine the
213 appropriate number of basis functions, consequently setting the number for maximum
214 temperature and age to 15 ($k=15$):

$$\text{Eq (2) Growth Increment} = s(\text{maximum temperature}, k = 15) + s(\text{age}, k = 15) + \\ \text{sex} + \text{location}.$$

217 Model variables were also checked for collinearity using the `concurvity()` function and
218 determined not be strongly correlated to one another.

219 **1.3 Results**

220 Comparison of the MT growth curves for the subfossil shells collected off Delmarva with
221 birth dates contemporaneous with living animals from New Jersey, Long Island, and Georges
222 Bank born after 1700 CE showed that the subfossil ocean quahogs from Delmarva grew faster
223 than those from all other regions, with animals from Georges Bank representing the second
224 fastest growth rates (Fig 4). Growth curves from Long Island and New Jersey were extremely
225 similar at young and middle ages, but older New Jersey animals were larger than Long Island
226 animals at the same age (Fig 4). When comparing populations with contemporaneous birth dates
227 (1700-2017 CE) from Georges Bank, Long Island, New Jersey, and Delmarva, maximum growth
228 rate (i.e., smaller a) was highest for Delmarva (Table 1), followed by Georges Bank and then by
229 New Jersey and Long Island. The age at maximum growth rate (c), was youngest at Georges
230 Bank, with Long Island, New Jersey and Delmarva demonstrating increasing age at maximum
231 growth rate (Table 2). All of these ages at maximum growth fell within the first 4 years of life,
232 however. The rate of decline of growth rate with age was lowest at Delmarva (higher f), then
233 New Jersey, Long Island, and Georges Bank. A scale of body size, (d) was largest at Georges

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234 Bank, then Delmarva, Long Island, and New Jersey (Table 1).

235 Figures 5 depicts temporal and spatial trends in growth from 1740-1940 CE at Delmarva,
236 Georges Bank, and Long Island respectively broken into 20-yr groupings by birthdate. Parameter
237 values are provided in Table 2. The New Jersey population was not grouped as growth rates were
238 similar to Long Island and as a result was excluded from Figure 5. Without exception, growth
239 rates for the vicennial groups were as high or higher for the subfossil Delmarva shells than for
240 live-collected animals from Long Island and Georges Bank.

241 Modified Tanaka growth curves (Fig 6) for animals born during major climate events in
242 the Holocene illustrate the temporal changes in growth during these major events including the
243 Little Ice Age (LIA) (207-462 cal years BP), Medieval Warm Period (MWP) (877cal years BP),
244 Dark Ages Cold Period (DACP) (1167-1223 cal years BP), Roman Warm Period (RWP) (2,447
245 cal years BP), Neoglacial part 1 (Neo1) (2813-3093 cal years BP), Neoglacial part 2 (Neo2)
246 (3418-3542 cal years BP), and the Meghalayan stage boundary (Meg) (3817-4302 cal years BP).
247 For more on these time periods, see Wanner et al. (2011), Auger et al. (2019), and LeClaire et al.
248 (2022). Parameter values for the Modified Tanaka growth curves for each of these time periods
249 are provided in Table 3. The growth of subfossil ocean quahogs collected from Delmarva was
250 always greater than or equal to the growth of ocean quahogs from Long Island, Georges Bank,
251 and New Jersey, regardless of the time period in which they lived. Growth was fastest during the
252 Roman Warm Period and the Dark Ages Cold Period, followed by the Medieval Warm Period.
253 Growth during the Meghalayan stage boundary was almost the same as during the earliest part of
254 the Neoglacial, with the late Neoglacial shells growing slightly faster. Almost no difference is
255 present during Neo1 and Neo2 while the differences are substantial during the Dark Ages Cold
256 Period and Roman Warm Period. Growth during the Little Ice Age and between 1740-1960 CE

257 were very similar.

258 Subfossil shells from Delmarva lived to between 16-247 years of age (Fig 7), an age
259 range similar to that recorded for present-day living populations by Hemeon et al. (2021a, 2023a)
260 and Sower et al. (2023a). Age-at-death was well distributed across radiocarbon birth years
261 between 60-4400 years ago (Fig 7), suggesting little variability in age-at-death within this
262 admittedly small sample size. Analyzing the subfossil Delmarva *A. islandica* that died in the past
263 200 years, younger animals became less common closer to present time, with their near
264 disappearance starting about 120 years before present (Fig 8).

265 The generalized additive model (GAM) showed a negative relationship between the
266 annual maximum temperature of the Cold Pool at the Long Island and Georges Bank collection
267 locations and the growth of *A. islandica* living there during the 1958-2017 time period (Fig 9).
268 The ANOVA (conducted via the `summary()` function) verified the smooth terms (maximum
269 temperature and age) and parametric coefficients (sex and location) used in the GAM (Eq 2)
270 were all significant ($p < 2e^{-16}$), explaining 40.3% of the deviance ($R^2=0.403$). Results are
271 consistent with the known differential in growth rate between Long Island and Georges Bank
272 (Hemeon et al., 2023b), and the well-established dimorphism between the growth rate of males
273 and females (Hemeon et al., 2023a; Sower et al., 2022, 2023a). The GAM revealed a modest
274 downward trend in growth rate across the temperature range of 10-15 °C, but a substantial
275 decline as maximum temperature exceeded 15°C, consistent with the known thermal limit for
276 ocean quahogs of ~16°C.

277 **1.4 Discussion**

278 **1.4.1 Regional and temporal comparisons in growth rates**

279 The growth models change over time between the 20-yr groupings, with growth rates at

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280 age increasing from the mid-to-late 19th century into the 20th century as reported by Hemeon et
281 al. (2021a, 2023b) and Sower et al. (2022, 2023b). Regional differences in growth rate of ocean
282 quahogs were also observed for living *A. islandica* across the three study regions. As Hemeon et
283 al., (2023b) reported, Georges Bank *A. islandica* exhibit faster growth rates than Long Island. As
284 observed by Sower et al. (2023b), growth of ocean quahogs from New Jersey and Long Island
285 had very similar rates, nearly identical until animals reached an old age (Fig 4, Table 1).

286 The growth models show different growth rates for these living populations and the
287 subfossil *A. islandica* from Delmarva with contemporary birth dates as well as older Delmarva
288 clams from earlier Holocene cold climate events. Unexpectedly, Delmarva subfossil ocean
289 quahogs collected inshore of the present-day distribution of the species grew as fast or faster than
290 animals from these other studied regions. Previous to this study, both Hemeon et al., (2023b) and
291 Pace et al., (2018) found ocean quahogs from Georges Bank to have the highest growth rates in
292 the Mid-Atlantic among living populations so far studied; however, even growth rates at Georges
293 Bank were equaled or exceeded by growth rates obtained from subfossil shells off Delmarva in
294 this study.

295 Delmarva subfossil ocean quahogs born post-1700 had equal or higher growth rates
296 compared to living animals born after 1700 from all three regions, and this higher rate of growth
297 was consistently present regardless of vicennial birth group. Continuously higher growth rates
298 for the population born between 1740-1960 inshore off Delmarva support the inference that the
299 Delmarva populations lived under conditions that may have maximized growth prior to the
300 demise of this population which likely was concluded in the 1970s. Additionally, the differences
301 between growth curves during the major climate events in the Holocene predict a period for
302 optimal growth between Medieval Warm Period and Roman Warm Period (Fig 6). During the

303 Long Island Age, the growth curves are similar compared to the modern period (except for
304 animals at old age) while animals living during Medieval and Roman Warm Period had growth
305 rates much greater than in the other periods and regions (Fig 6). These observed higher growth
306 rates for subfossil populations off Delmarva suggest that environmental conditions supporting
307 optimal growth were present during the lifetime of these clams. Presumably, bottom
308 temperatures supported maximum growth before reaching intolerable levels that eventually led
309 to the death of all inshore ocean quahogs prior to the 1980s (Fig 8,9).

310 **1.4.2 Why were growth rates so high during past cold periods?**

311 For most bivalves, including *A. islandica*, growth rates increase with increasing
312 temperature until an optimal temperature is exceeded and growth rates decline. This parabolic
313 relationship of physiology and temperature is well described (Woodin et al., 2013) and
314 physiologically based in the relationship of filtration rate and respiration rate in bivalves: the two
315 metabolic energetic processes that are primary determinants of scope for growth (Ren and Ross,
316 2001; Hofmann et al., 2006; Munroe et al., 2013). The GAM model (Fig 9) shows the influence
317 of present-day yearly maximum temperatures at the Long Island and Georges Bank sites, with a
318 clear break point at 15 °C. This temperature is often exceeded for periods during the early fall
319 when water-column stratification breaks down, the increasing frequency of which being a
320 product of the rapid warming of this region of the Mid-Atlantic (Lucey and Nye, 2010; Pershing
321 et al., 2015; Saba et al., 2016; Kavanaugh et al., 2017). Presumably, present-day high
322 temperatures cause *A. islandica* growth to slow either due to a physiological constraint or to a
323 cessation of feeding due to estivation to escape the highest temperatures when the Cold Pool
324 decays in the early fall. Burrowing behavior in the species is well described (Ragnarsson and
325 Thórarinsdóttir, 2002; Strahl et al., 2011) and the species can survive without oxygen for

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326 extended periods of time (Taylor, 1976a,b; Oeschger and Storey, 1993). Whether due to
327 physiological constraints or estivation, the growing season is effectively shortened. If such
328 extreme conditions were present for subfossil shells off Delmarva, one would not expect to
329 observe the higher growth rates consistently present for Delmarva animals relative to rates
330 observed for living animals from Long Island and Georges Bank.

331 Accordingly, one could infer that increased temperatures relative to today seem an
332 unlikely explanation for the increased growth rate, and this is further supported by the fact that
333 growth rates for dead *A. islandica* born in the 19th and early 20th century off Delmarva are
334 higher than contemporaneous clams still living at Long Island and Georges Bank. Yet, evidence
335 clearly indicates that growth rates for Long Island and Georges Bank were lower in the 19th
336 century than observed today (Pace et al., 2018; Hemeon et al., 2023a). Consequently, a simple
337 increase in temperature appears insufficient to support the observed higher growth rates for
338 subfossil shells collected off Delmarva that lived throughout the Holocene. Shells from ocean
339 quahogs spawned during most cold and warm events during the Holocene were growing as fast
340 or at faster rates than the present populations in Georges Bank, Long Island, and New Jersey.

341 Increases in food supply also produce increasing rates of growth (Schöne et al., 2005;
342 Mette et al., 2016; Ballesta-Artero et al., 2017). As the fast rate of growth for subfossil Delmarva
343 shells indicates that conditions off Delmarva were most suitable for growth for the resident *A.*
344 *islandica* population at that time, and that increased temperature was a necessary, but not a
345 sufficient, explanation, an additional increased food supply would seem to be a co-occurring
346 requisite. Little is known about the influence of warming temperatures on phytoplankton
347 production in the studied region (Friedland et al., 2018, 2020b), though considerable attention
348 has been given to the influence of warming temperatures on phytoplankton production in general

349 (e.g., Oviatt, 2004; Richardson and Schoeman, 2004; Osman et al., 2019; Lotze et al., 2019). In
350 particular, detailed knowledge of the influence of the inshore-offshore depth gradient on
351 phytoplankton production is limited in the studied region (Yoder et al., 2002; Mouw and Yoder,
352 2005; Munroe et al., 2013). Despite the lack of studies, phytoplankton production has been
353 shown to be insufficient to meet the food supply requirements of the largest bivalve in the
354 region, the Atlantic surfclam *Spisula solidissima* (Munroe et al. 2013). Munroe et al., (2013)
355 argue that benthic primary production is a critical supplement to phytoplankton production to
356 support biomass of the Atlantic surfclam on the Mid-Atlantic continental shelf.

357 Whereas not enough is known about the food requirement of ocean quahogs to blithely
358 extrapolate from the surfclam case, the subfossil ocean quahogs with radiocarbon dates from
359 known cold periods since the Holocene Climate Optimum were collected at study sites off
360 Delmarva with depths considerably shallower than the present range of this species south of
361 Long Island. Sea level at the earliest cold times recorded by the Delmarva animals was slightly
362 lower than today (Engelhart et al., 2011), indicating animals would have lived at depths even
363 shallower than presently recorded depths at these sites. *Artica islandica* have no shallow-water
364 depth restriction, and are found at much shallower depths than those off Delmarva, including
365 locations well inshore off Long Island, as well as elsewhere in the North Atlantic (Zettler et al.,
366 2001; Ragnarsson and Thórarinsdóttir, 2002; Strahl et al., 2011; Begum et al., 2019). Hence, the
367 depth range of past occupation off Delmarva is not unusual for the species. Shallow depths are
368 higher in the photic zone where benthic primary production is enhanced (Munroe et al. 2013)
369 permitting speculation that the increased growth rates observed were due not only to optimal
370 thermal conditions, but also to greater food availability.

371 **1.4.3 The dynamics of range recession in ocean quahogs**

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372 Ocean quahog growth at Georges Bank was comparable to growth off Delmarva during
373 early Holocene cold periods (Meg and Neol,2) and somewhat slower than subsequent cold
374 periods (the DACP and LIA). Throughout the Meghalayan and Neoglacial, the Delmarva growth
375 curves are similar to those of the living Georges Bank populations in the modern period (Fig 6)
376 suggesting that thermal conditions during the Neoglacial were similar to those in Georges Bank
377 after 1700. Notably, the distribution of the Delmarva subfossil shells strongly suggests that the
378 Cold Pool, a key oceanographic feature of the region, has waxed and waned across the
379 continental shelf off the U.S. east coast consistent with known cold and warm periods in the past
380 and produced the transgressions and regressions of the boreal community on the continental shelf
381 herein exemplified by the ocean quahog (Fig 3). The long-term history of the Cold Pool is not
382 well understood, but the sensitivity of the regional footprint of the Cold Pool to recent warming
383 temperatures is well described (Friedland et al. 2020a, 2022).

384 The occupation of the Delmarva inner continental shelf by ocean quahogs represents
385 periods of transgression of habitable cold water inshore until conditions became unfavorable.
386 The death assemblages produced contain a range of sizes and ages of animals, from small and
387 young to large and old. Death assemblage compositions tend to be biased in favor of small
388 (young) animals because the process of natural mortality usually adds a declining number of
389 animals as the cohort ages and the individuals increase in size (Hallam, 1967; Cummins et al.,
390 1986; Tomašových, 2004); whereas, taphonomic processes normally bias the assemblage against
391 the preservation of the smallest size classes (Staff et al., 1986; Powell and Stanton, 1996;
392 Kidwell, 2001). Larger animals were selected for processing due to gear limitations, but the use
393 of a lined dredge permitted routine collection of animals <20 years old so that the age
394 distribution of the samples provides a good representation of the expected age distribution of the

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395 entire population at death except for the very youngest individuals; oldest ages >150 yr are
396 consistent with the age range of present-day living populations (Hemeon et al. 2023a, 2023b;
397 Sower et al., 2023a). Consequently, ages-at-death are likely older than an unbiased sampling of
398 the entire size spectrum of the assemblage would otherwise provide, but a bias favoring young
399 animals would still be anticipated by cohort mortality dynamics and despite the counterweighing
400 bias of taphonomy, as these animals are young, but not necessarily small (Figures 4-5).
401 Consequently, increased numbers of animals at younger age would still be expected, and
402 particularly as the most recent cohorts added to the death assemblage tend to be most numerous
403 (Olszewski, 2004; Kosnik et al., 2009; Tomašových et al., 2014, 2018).

404 Surprisingly, the opposite was observed (Fig 7). Considering the subfossil animals that
405 lived contemporaneously with living animals (in the past 200 years), young animals become less
406 common after approximately 120 years before present (Fig 8). This shift in age distribution
407 shows the age-at-death increasing through time with the animals of oldest age-at-death dying
408 most recently, coinciding with the warm period during the 1930s-1950s and the 1960s-1970s
409 cooling (Nixon et al., 2004; Bellucci et al., 2017; McClenachan et al., 2019). The few most
410 recent new recruits observed in this study coincidentally date from this most recent period of
411 cooling. The near absence of younger animals in the last 100 years of the timeseries overall,
412 however, implies that recruitment decreased dramatically around 120 years ago, with the
413 remaining deaths of older animals representing the slow demise of a non-recruiting population
414 originally produced primarily by recruitment in the 19th century.

415 Older animals likely have a higher tolerance to high temperatures as a result of their
416 ability to burrow deeper into the sediment (Taylor, 1976a; Ragnarsson and Thórarinsdóttir, 2002;
417 Strahl et al., 2011) to escape the higher temperatures that occur routinely in the Cold Pool when

418 stratification breaks down in the fall before winter temperatures arrive. Temperature conditions
419 similar to the annual maximum temperatures depicted in Figure 8 result in a negative trend in
420 growth and likely forecast a future cessation of growth, followed by death as higher temperatures
421 remain for longer periods in the fall. Though direct observations are lacking, one might consider
422 that survivorship would be very low for newly-settled animals under thermal conditions present
423 through much of the 20th into the 21st century. This interpretation supports the conclusion that the
424 times of death for recently dead shells collected off Delmarva are the product of an increase in
425 bottom temperature that limited survivorship of newly-settled *A. islandica* followed by the
426 eventual demise of the older animals as continuing yearly mortality took its toll.

427 The analysis of age-at-death depicted in Figure 7 provides a unique mechanism that
428 might be utilized to observe the rate of range recession in *A. islandica*. Unlike many bivalves,
429 such as Atlantic surfclams (e.g., Narváez et al., 2015), large ocean quahogs are less susceptible
430 to increasing temperatures as a result of burrowing behavior that limits exposure to unfavorable
431 conditions. Evidence of inshore range boundary recession for this species will likely be found
432 first in the abundance of the juvenile animals in the population. Most importantly, evidence from
433 the death assemblages sampled here strongly indicates that an *A. islandica* range shift on the
434 trailing edge of the range is a 100+ year process rather than subdecadal as in many bivalve
435 species such as the Atlantic surfclam (Jones et al., 2010; Hofmann et al., 2018; Baden et al.,
436 2021). In this, the observed range of the ocean quahog in the Mid-Atlantic Bight may more
437 closely reflect the range core of the past rather than present-day and more importantly obscure
438 the ongoing effects of warming temperatures in the continuing presence of apparently robust
439 populations wherein recruitment may have failed long ago.

440 **1.5 Conclusions**

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441 Death assemblages are proving to be an important repository of information for the
442 influence of climate change on the living community over timespans that exceed those of modern
443 benthic surveys (Black et al., 2009; Wanamaker et al., 2009; Meadows et al., 2019; Powell et al.,
444 2020). Ocean quahogs represent a long-term record of climate change on the U.S. northeast-coast
445 continental shelf. This species transgressed and regressed across the shelf numerous times in
446 accordance with cold and warm climatic periods. The growth rates of this species provide a
447 unique view of climatic conditions during the times of occupation. Living ocean quahogs from
448 multiple regions across the North-Atlantic shelf (Georges Bank, Long Island, New Jersey) were
449 compared to subfossil shells collected from a region off Delmarva inshore of the present range.
450 These populations exhibited different growth rates, with subfossil clams from Delmarva growing
451 the fastest. Moreover, ocean quahog growth compared between regions and 20-year groupings
452 from 1740 to 1940 showed both regional and temporal differences in growth, indicating that
453 ocean quahogs that once occupied the inner-to-middle continental shelf off Delmarva continued
454 to grow faster than animals from living populations taken from other regions throughout this
455 timespan. Individuals representing each of the cold periods after the Holocene Climate Optimum,
456 occupying the studied region off Delmarva, grew as fast or faster than individuals presently
457 living in the Mid-Atlantic region probably due to optimal temperatures accompanied by
458 increased food supply in shallow water. *Arctica islandica* growth changed throughout the
459 Holocene, reflecting the changes in environmental conditions throughout the epoch with periods
460 of the Neoglacial most closely representing modern-day growth rates suggestive of colder
461 temperature conditions inshore during that time period.

462 Examining subfossil ocean quahogs from Delmarva with a shorter time-since-death, that
463 is dying nearer present-day, the age range at death for ocean quahogs with the most recent

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Tables

Table 1 Modified Tanaka growth parameters (Eq. 1) from living populations from Georges Bank, Long Island, and New Jersey, and the subfossil Delmarva animals with contemporaneous birth dates.

Group	Parameter	Georges Bank		Long Island		New Jersey		Delmarva	
		Estimate	SE	Estimate	SE	Estimate	SE	Estimate	SE
Contemporary Population	<i>a</i>	7.36E-03	7.34E-04	1.57E-02	7.29E-04	1.69E-02	6.07E-04	7.91E-03	9.17E-04
	<i>c</i>	7.62E-01	7.62E-02	1.77E+00	6.88E-02	2.40E+00	6.55E-02	3.76E+00	1.72E-01
	<i>d</i>	8.78E+01	2.03E-01	7.73E+01	1.53E-01	7.61E+01	1.62E-01	8.00E+01	8.41E-01
	<i>f</i>	3.00E-03	2.82E-05	3.90E-03	3.48E-05	4.34E-03	4.30E-05	6.99E-03	4.21E-04
	<i>g</i>	6.04E-06	4.29E-07	5.07E-06	2.66E-07	9.34E-06	3.86E-07	4.59E-05	5.68E-06

Table 2 Modified Tanaka growth parameters and standard error (SE) for 20-year groupings by birth for Georges Bank (GB), Long Island (LI), and the subfossil Delmarva animals of contemporaneous birth dates.

Cohort	Parameter	Georges Bank Population		Long Island Population		Delmarva Population	
		Estimate	SE	Estimate	SE	Estimate	SE
1740	a	6.55E-03	9.64E-03	9.91E-03	9.63E-03	8.54E-03	5.13E-03
	c	0.00E+00	8.07E-01	0.00E+00	5.85E-01	2.94E+00	1.03E+00
	d	8.10E+01	1.45E+00	7.09E+01	7.68E-01	8.72E+01	4.89E+00
	f	2.19E-03	1.34E-04	2.91E-03	1.26E-04	4.60E-03	1.21E-03
	g	9.14E-06	1.01E-06	1.46E-05	5.97E-07	7.29E-06	2.76E-05
1760	a			4.46E-02	2.83E-02	7.21E-03	1.41E-03
	c			0.00E+00	1.22E+00	2.96E+00	2.75E-01
	d			7.36E+01	1.29E+00	7.99E+01	1.22E+00
	f			2.56E-03	1.76E-04	6.52E-03	5.63E-04
	g			1.58E-05	9.69E-07	3.08E-05	6.19E-06
1780	a	1.14E-02	6.26E-03	1.46E-02	8.31E-03	3.15E-03	2.99E-03
	c	0.00E+00	4.09E-01	0.00E+00	4.77E-01	1.50E+00	8.12E-01
	d	8.42E+01	6.49E-01	6.98E+01	6.71E-01	1.03E+02	4.56E+00
	f	2.61E-03	8.21E-05	2.84E-03	1.01E-04	2.94E-03	5.23E-04
	g	7.67E-06	6.34E-07	2.02E-05	6.73E-07	0.00E+00	1.49E-05
1800	a	5.74E-03	6.39E-03	1.33E-02	5.85E-03	8.08E-03	1.05E-03
	c	0.00E+00	5.14E-01	0.00E+00	3.52E-01	2.42E+00	2.20E-01
	d	8.78E+01	1.01E+00	7.24E+01	5.46E-01	9.22E+01	1.17E+00
	f	2.53E-03	1.15E-04	2.83E-03	7.92E-05	3.73E-03	1.97E-04
	g	6.91E-06	1.17E-06	2.10E-05	6.53E-07	3.64E-05	6.29E-06
1820	a	9.56E-03	4.31E-03	2.50E-02	9.59E-03	1.04E-02	1.60E-03
	c	0.00E+00	3.24E-01	0.00E+00	4.73E-01	3.48E+00	2.86E-01
	d	8.95E+01	6.58E-01	7.54E+01	6.61E-01	8.42E+01	1.36E+00
	f	2.41E-03	6.72E-05	2.99E-03	1.07E-04	5.73E-03	4.83E-04
	g	9.31E-06	8.87E-07	2.49E-05	9.34E-07	2.95E-05	8.94E-06
1840	a	9.48E-03	4.35E-03	4.37E-03	4.27E-03	1.31E-02	3.02E-03
	c	2.64E-01	3.63E-01	0.00E+00	3.12E-01	2.63E+00	4.63E-01
	d	9.11E+01	8.51E-01	7.77E+01	6.29E-01	8.59E+01	1.85E+00
	f	2.52E-03	8.96E-05	3.21E-03	1.04E-04	3.90E-03	3.57E-04
	g	9.38E-06	1.50E-06	2.60E-05	1.24E-06	3.53E-05	7.55E-06
1860	a	1.12E-02	2.63E-03	4.62E-03	3.72E-03	1.55E-02	6.00E-03
	c	9.50E-01	2.62E-01	1.51E-01	3.05E-01	2.77E+00	9.63E-01
	d	8.82E+01	7.35E-01	8.00E+01	7.24E-01	8.98E+01	4.74E+00
	f	2.91E-03	9.38E-05	3.16E-03	1.11E-04	3.67E-03	7.63E-04
	g	1.91E-05	1.87E-06	3.13E-05	1.78E-06	0.00E+00	2.97E-05
1880	a	1.01E-02	1.67E-03	1.43E-02	3.30E-03	3.60E-03	1.32E-03
	c	1.08E+00	1.84E-01	8.66E-01	2.89E-01	2.20E+00	9.14E-01
	d	8.69E+01	5.93E-01	8.33E+01	8.02E-01	1.58E+02	2.41E+01
	f	3.11E-03	8.17E-05	2.99E-03	1.06E-04	1.36E-03	5.30E-04
	g	1.79E-05	2.04E-06	3.01E-05	2.59E-06	0.00E+00	9.72E-04
1940	a	1.06E-02	9.45E-04	1.25E-02	5.62E-04	1.70E-02	3.78E-02
	c	2.21E+00	1.65E-01	2.74E+00	9.12E-02	0.00E+00	6.27E+01
	d	8.31E+01	9.85E-01	7.39E+01	5.22E-01	7.14E+02	2.65E+03
	f	4.27E-03	1.93E-04	5.60E-03	1.63E-04	7.03E-05	5.00E-04
	g	7.39E-05	1.30E-05	9.39E-05	8.46E-06	0.00E+00	1.18E-02

SE, standard error

Table 3 Modified Tanaka model parameters for subfossil shells from Delmarva grouped by climate event before present time (BP), including the Little Ice Age (LIA) (207-462 cal years BP), Medieval Warm Period (MWP) (877 cal years BP), Dark Ages Cold Period (DACP) (1167-1223 cal years BP), Roman Warm Period (RWP) (2,447 cal years BP), Neoglacial part 1 (Neo1) (2813-3093 cal years BP), Neoglacial part 2 (Neo2) (3418-3542 cal years BP), and the Meghalayan stage boundary (Meg) (3817-4302 cal years BP).

Climate Event	Parameter	Delmarva	
		Estimate	SE
Modern	<i>a</i>	7.91E-03	9.17E-04
	<i>c</i>	3.76E+00	1.72E-01
	<i>d</i>	8.00E+01	8.41E-01
	<i>f</i>	6.99E-03	4.21E-04
	<i>g</i>	4.59E-05	5.68E-06
Little Ice Age	<i>a</i>	8.77E-03	4.07E-03
	<i>c</i>	1.74E+00	6.79E-01
	<i>d</i>	1.00E+02	2.98E+00
	<i>f</i>	2.79E-03	3.20E-04
	<i>g</i>	0.00E+00	1.02E-05
Medieval Warm Period	<i>a</i>	3.89E-03	6.43E-04
	<i>c</i>	4.10E+00	1.19E-01
	<i>d</i>	8.32E+01	1.14E+00
	<i>f</i>	1.02E-02	8.89E-04
	<i>g</i>	6.67E-05	3.41E-05
Dark Ages Cold Period	<i>a</i>	1.03E-03	7.28E-03
	<i>c</i>	0.00E+00	2.57E+00
	<i>d</i>	1.59E+02	4.74E+01
	<i>f</i>	1.04E-03	7.05E-04
	<i>g</i>	0.00E+00	2.03E-03
Roman Warm Period	<i>a</i>	4.17E-03	1.81E-03
	<i>c</i>	1.60E+00	8.25E-01
	<i>d</i>	1.23E+02	1.74E+01
	<i>f</i>	2.18E-03	8.41E-04
	<i>g</i>	0.00E+00	1.19E-03
Neoglacial Part 1	<i>a</i>	3.53E-03	5.72E-03
	<i>c</i>	1.01E+00	8.77E-01
	<i>d</i>	8.60E+01	2.69E+00
	<i>f</i>	3.75E-03	5.43E-04

	<i>g</i>	0.00E+00	6.41E-06
	<i>a</i>	3.67E-03	9.30E-03
Neoglacial Part 2	<i>c</i>	0.00E+00	1.02E+00
	<i>d</i>	9.36E+01	2.77E+00
	<i>f</i>	1.97E-03	1.89E-04
	<i>g</i>	0.00E+00	2.80E-06
	<i>a</i>	1.11E-02	2.93E-03
Meghalayan	<i>c</i>	3.24E+00	4.49E-01
	<i>d</i>	7.16E+01	1.58E+00
	<i>f</i>	6.61E-03	7.83E-04
	<i>g</i>	7.74E-05	7.33E-06

Figures

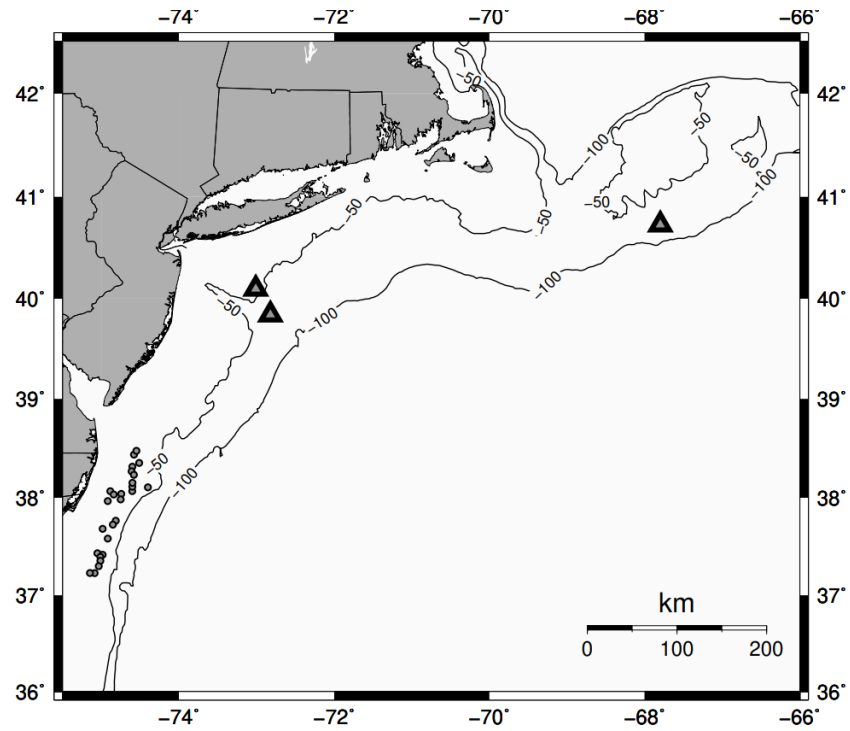


Figure 1. Map of the *Arctica islandica* sample collection locations in the Northwestern Atlantic. Grey triangles represent locations where live ocean quahogs were collected offshore of Long Island, New Jersey, and Georges Bank. Grey circles represent subfossil ocean quahog collection sites from offshore of the Delmarva Peninsula, magnified in Figure 2. Bathymetric contour depths in meters.

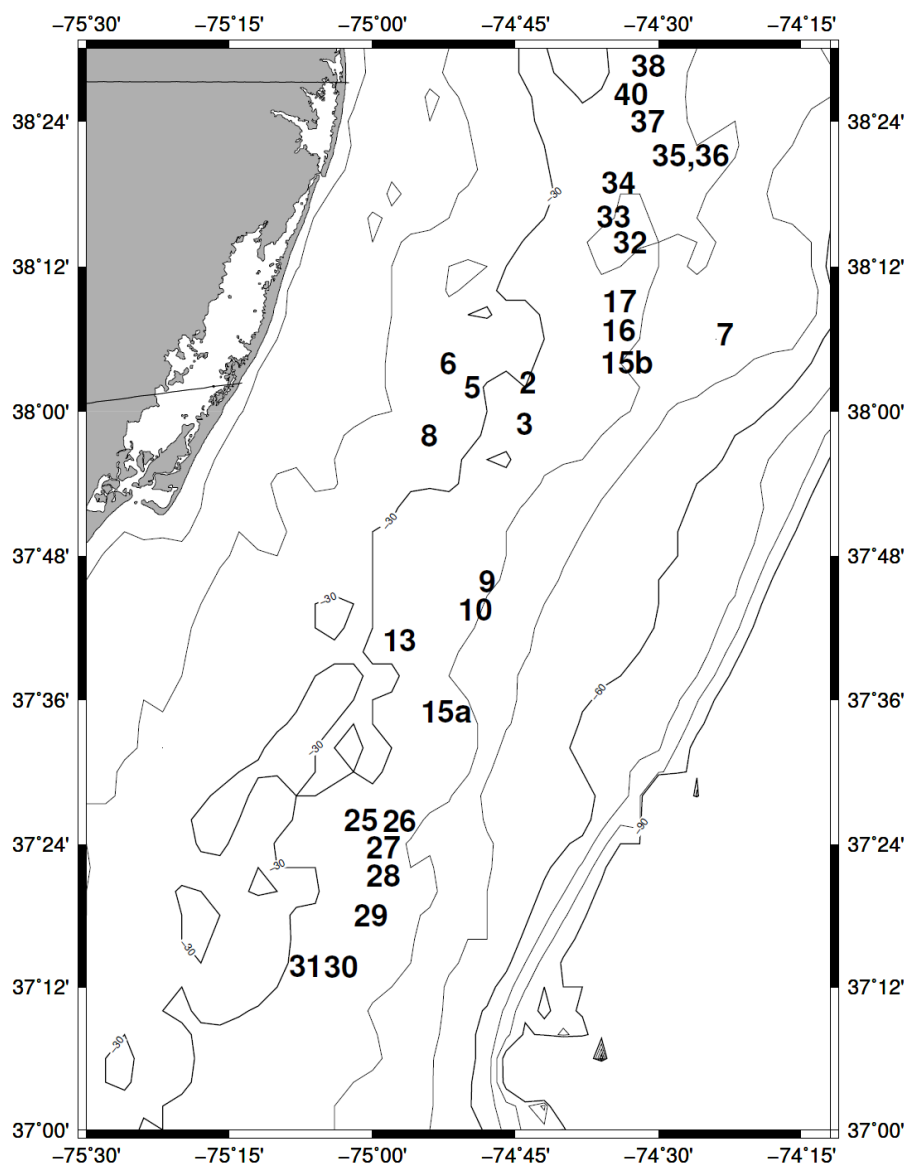


Figure 2. Subfossil ocean quahog collection sites from offshore of the Delmarva (DMV) Peninsula. Stations are identified as numbered during the survey. Bathymetric contour depths in meters. Additional details including the geographic distribution of shells of different times-since-death are provided by LeClaire et al. (2022).

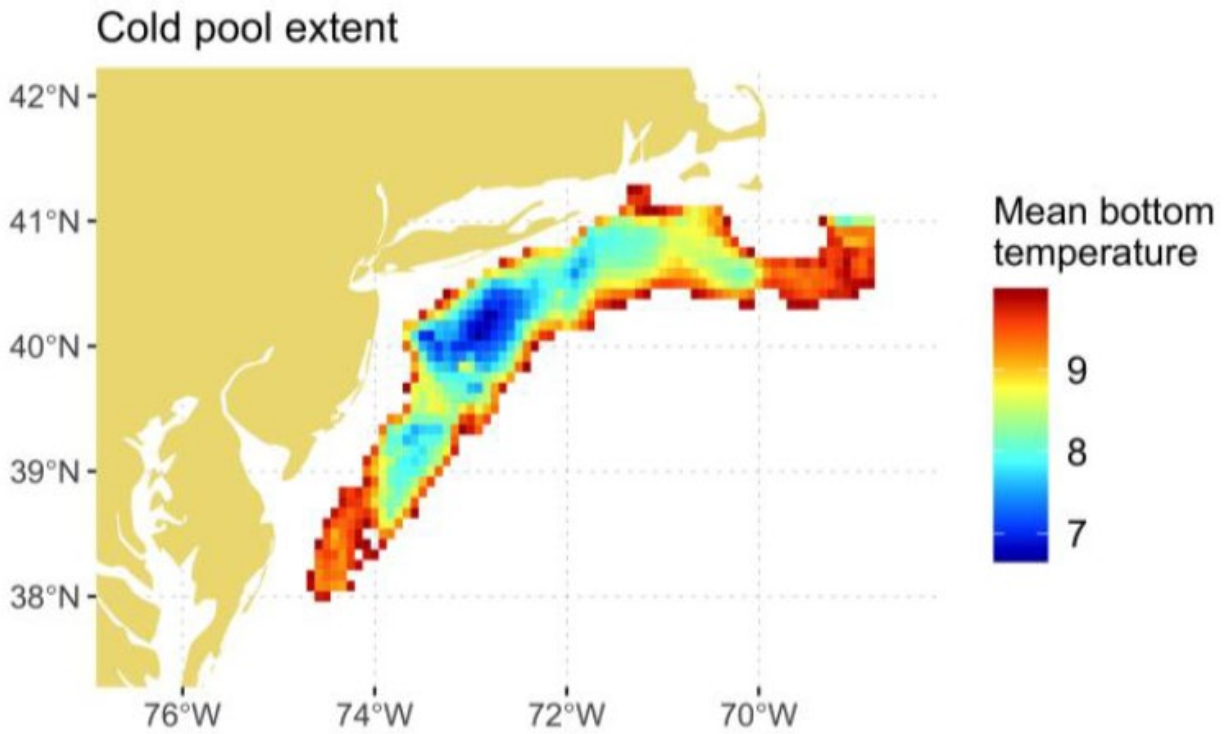


Figure 3 Cold Pool extent for the period 1959-2021. The mean bottom temperature and extent were calculated using a high-resolution long-term bottom temperature product described in du Pontavice et al. (2023).

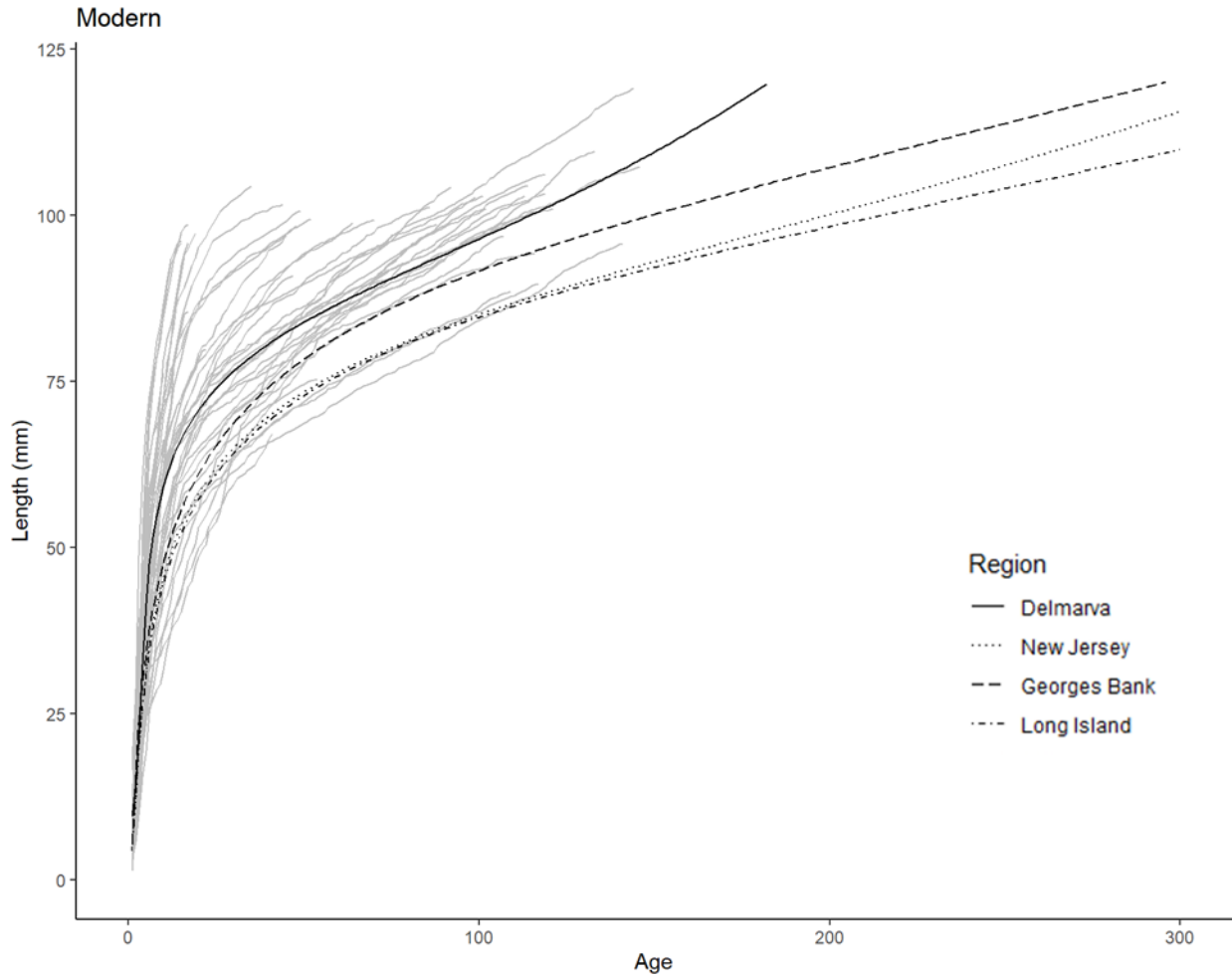


Figure 4 Growth curves for animals with birth dates contemporaneous with living ocean quahogs collected from the Mid-Atlantic continental shelf. Black lines represent Modified Tanaka growth curves for ocean quahogs born after 1700 BCE compared across regions. Average growth curves for subfossil shells from Delmarva (solid line) and living animals from New Jersey (dotted line), Georges Bank (dashed line), and Long Island (dot-dashed line) are presented along with individual growth curves for each animal in the subfossil Delmarva sample (light grey lines).

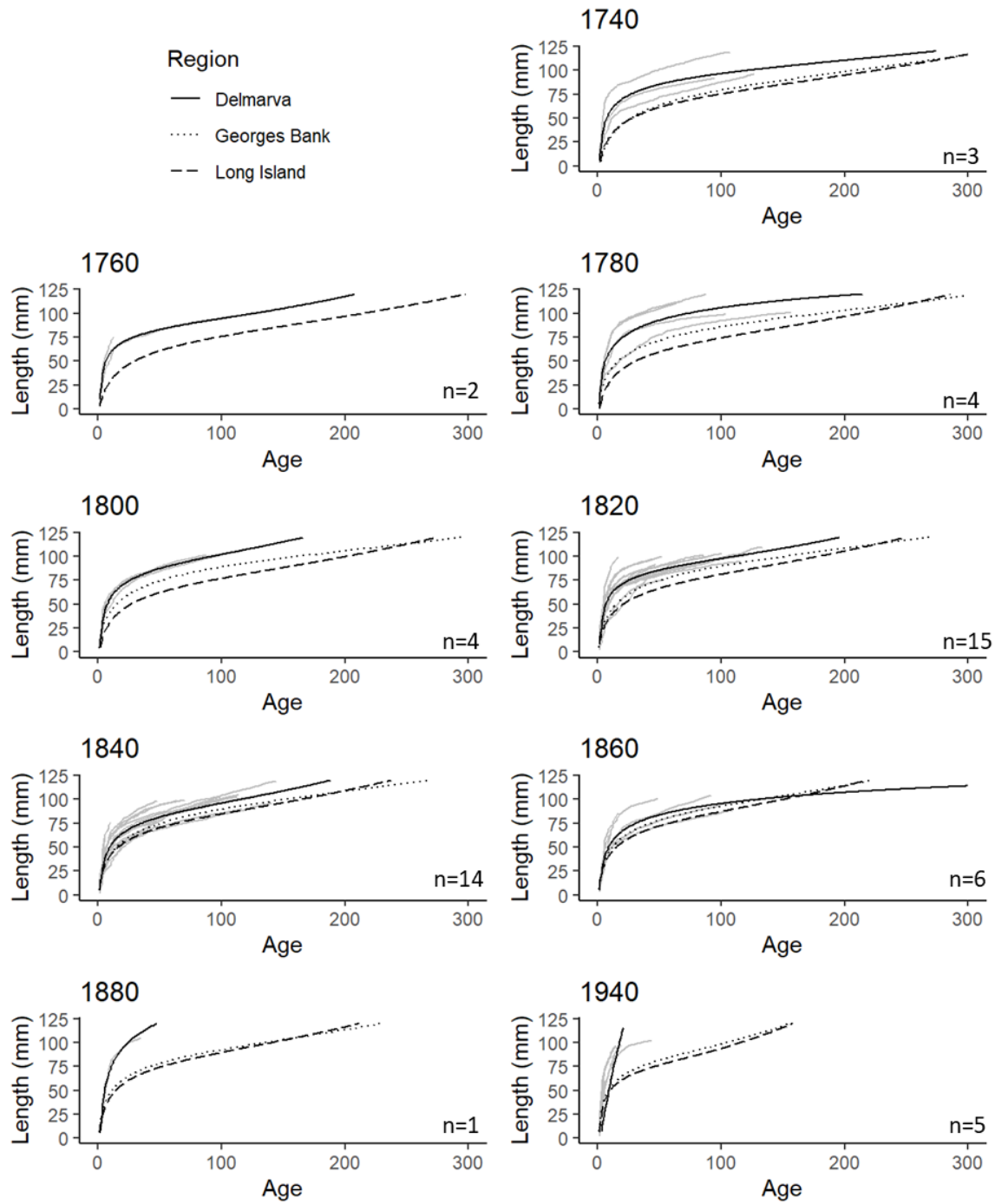


Figure 5 Growth curves for 20-yr groups defined by birthdate for ocean quahogs born since 1700. Black lines represent average Modified Tanaka curves compared across regions and 20-year groupings by birth date including subfossil shells from Delmarva (solid line) and living animals from Long Island (dashed line) and Georges Bank (dotted line). Light grey lines represent the individual growth curves for each animal in the subfossil contemporaneous Delmarva sample.

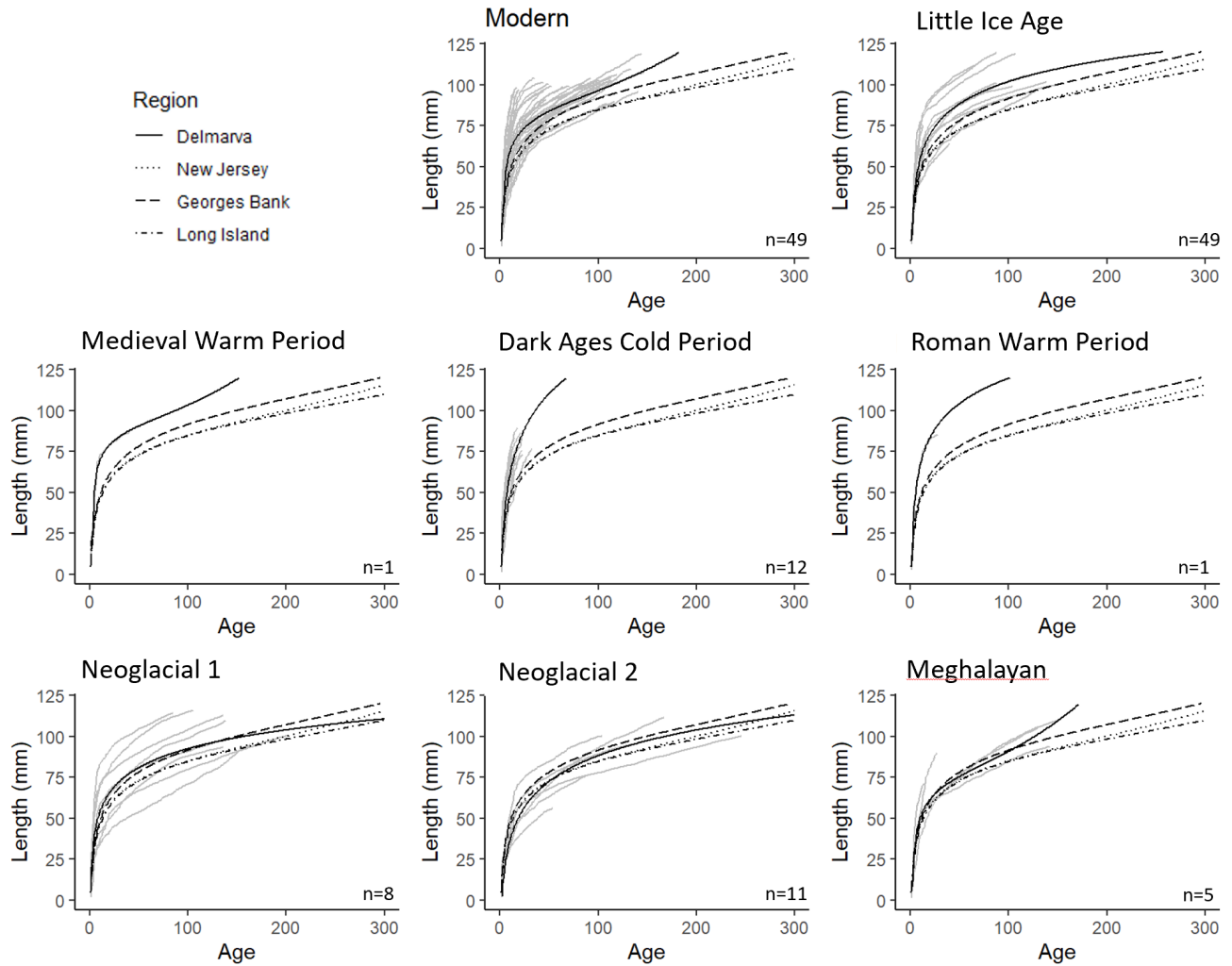


Figure 6 Growth curves for ocean quahogs born during selected time intervals of the Holocene. Black lines represent average Modified Tanaka growth curves for subfossil shells born off Delmarva during major climate events in the Holocene: Modern (60-203 cal years BP), Little Ice Age (LIA) (207-462 cal years BP), Medieval Warm Period (MWP) (877 cal years BP), Dark Ages Cold Period (1167-1223 cal years BP), Roman Warm Period (RWP) (2447 cal years BP), Neoglacial part 1 (Neo1) (2813-3093 cal years BP), Neoglacial part 2 (Neo2) (3418-3542 cal years BP), Meghalayan stage boundary (Meg) (3817-4302 cal years BP). Solid lines represent the average growth curve for subfossil shells sampled off the Delmarva Peninsula. Curves from New Jersey, Georges Bank, and Long Island represent living animals born after 1700 (Modern) and used for comparison in the graphs depicting growth during the Holocene climate events. The dotted line represents the growth for living *A. islandica* off the coast of New Jersey, the long dashes represent Georges Bank, and the dot-dash line represents Long Island. Light grey lines represent the individual growth curves for each animal in the subfossil Delmarva sample born in the designated climate period.

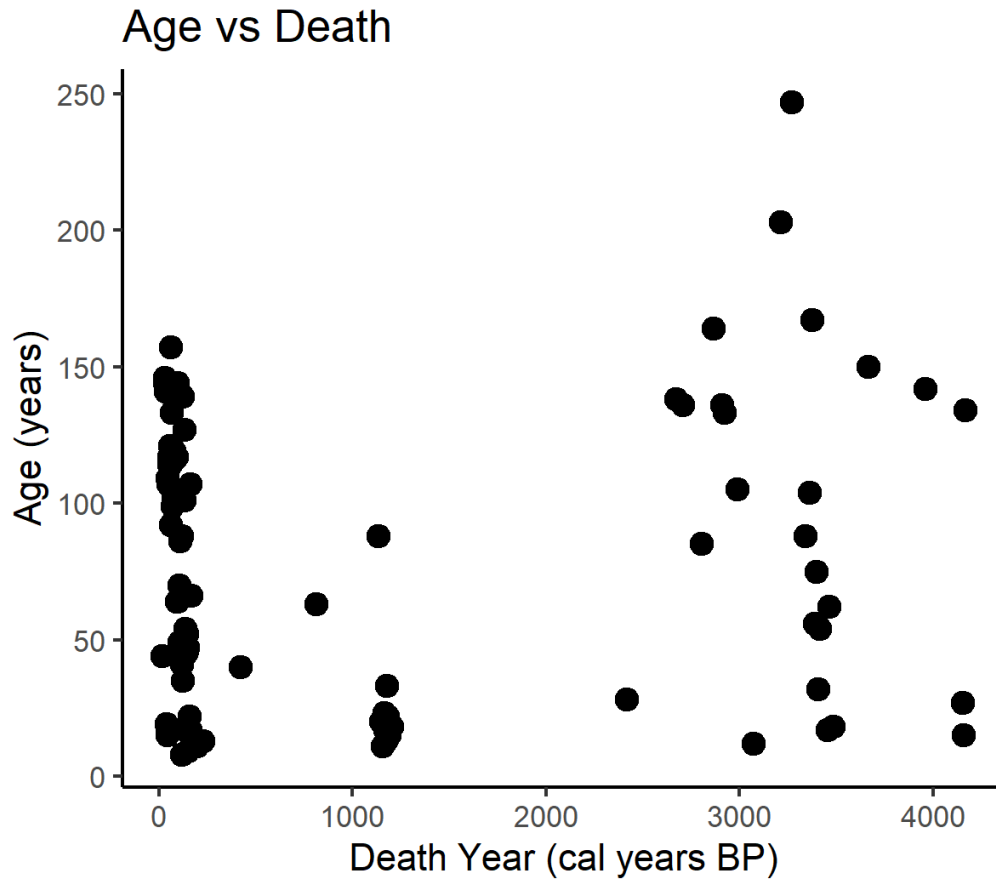


Figure 7 Age-at-death of subfossil ocean quahogs compared to the time (death year) that the animal died. Death years are in years before present, focusing on animals that died between 62-4400 years ago.

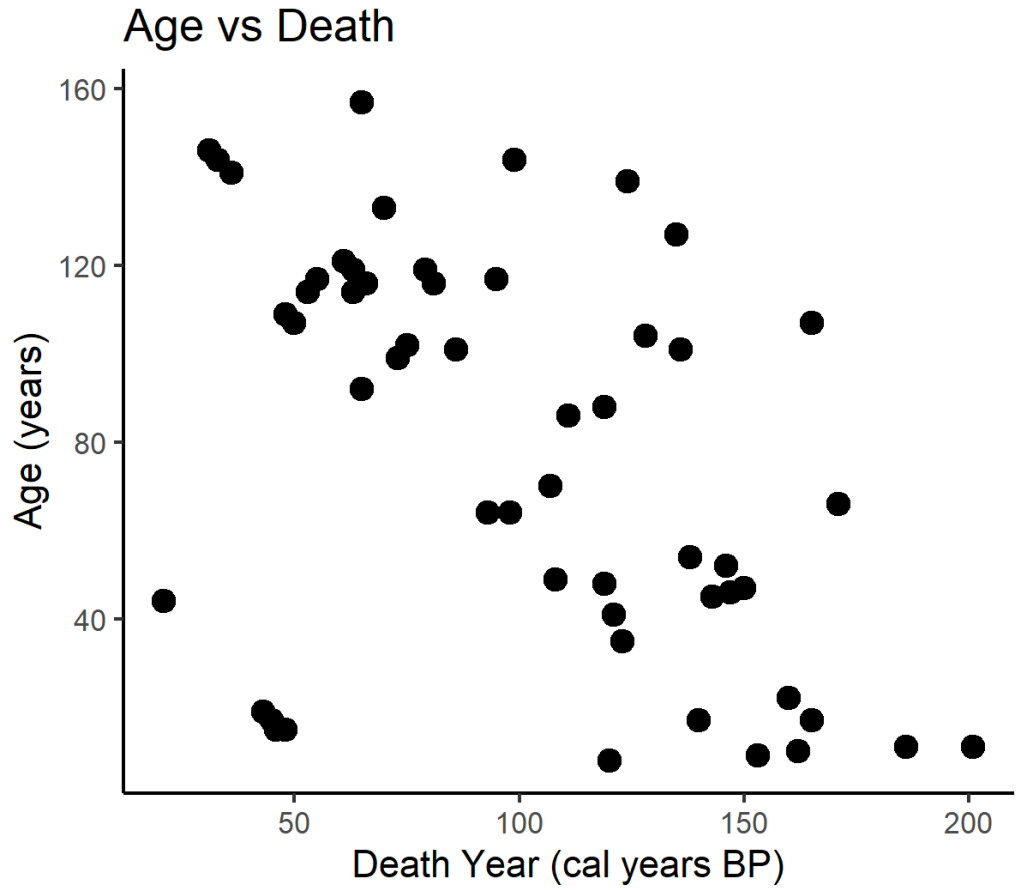


Figure 8 Age of subfossil ocean quahogs at death compared to the time (death year) that the animal died. Death years are in years before present, focusing on animals that died in the past 200 years.

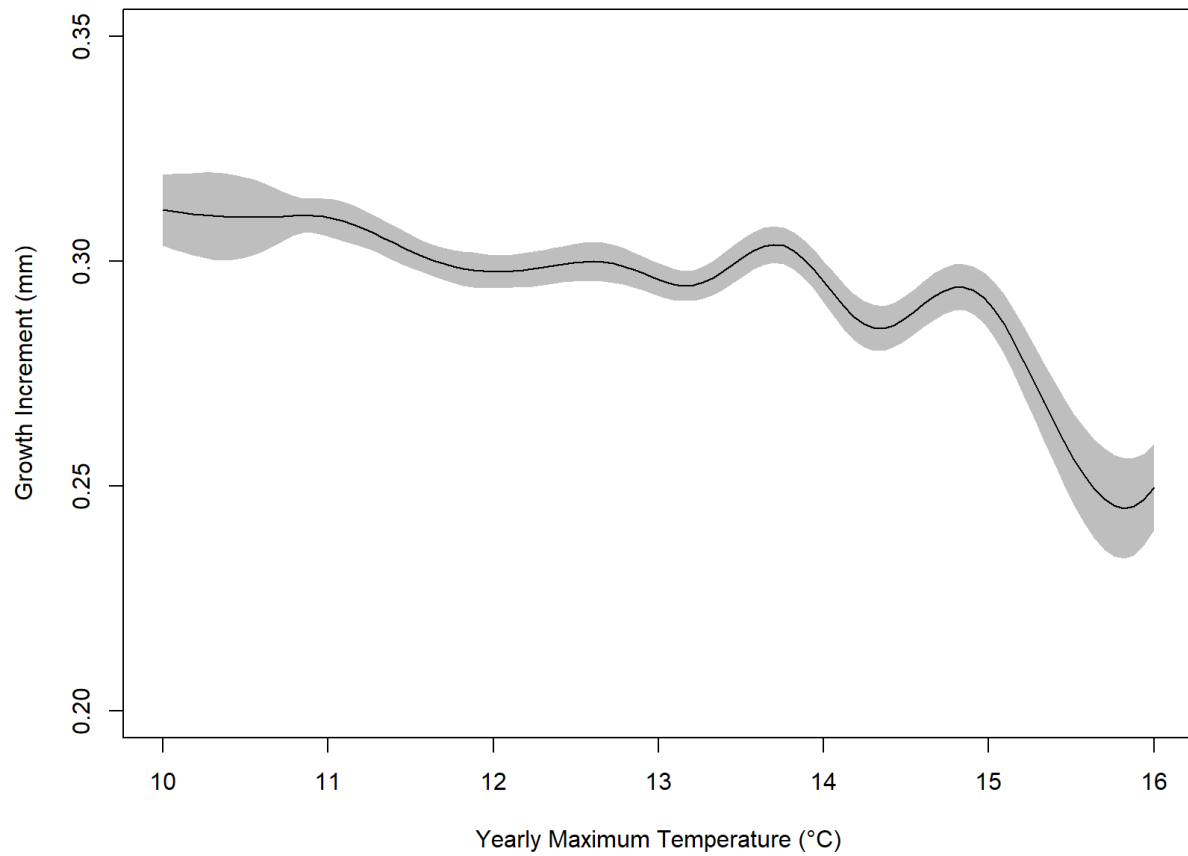


Figure 9 General additive model of yearly growth increment as a function of yearly maximum temperature between 1958-2017 (ROMS-NWA; du Pontavice et al., 2022). Grey bar represents the 95% confidence interval surrounding the GAM.