

1 **Positional and ontogenetic variation in vertebral centra morphology in five batoid species**

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13 Running head: Morphology of batoid vertebral columns

14

15 **Abstract**

16 An increasing number of studies on elasmobranchs have shown that band pair counts in vertebral
17 centra do not accurately reflect age. Research in sharks indicates that the number of band pairs
18 vary with body size and centrum morphology is related to structural needs. A study of this kind
19 has not been undertaken on batoids, thus we examined the relationship between band pair
20 deposition and morphology of centra along the vertebral column, and ontogenetically, for five
21 batoid species (little skate, *Leucoraja erinacea*, winter skate, *Leucoraja ocellata*, barndoor skate,
22 *Dipturus laevis*, Atlantic stingray, *Dasyatis sabina*, and round ray, *Urotrygon halleri*). Centrum
23 morphology and band pair count varied along the vertebral column in all individuals of all
24 species except in young of the year. Variation in band pair counts among centra within
25 individuals supports the hypothesis that band pair formation is related to somatic growth and
26 body shape rather than to an annual cycle.

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28 Additional keywords: band pair counts, skates and rays, somatic growth, age

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30

31 **Introduction**

32 The vertebral centra of elasmobranchs have characteristic alternating opaque and
33 translucent bands (termed a band pair) that have been assumed to represent one year of growth
34 and used to estimate age (Ridewood 1921; Haskell 1948; Ishiyama 1951; Cailliet *et al.* 2006).
35 Annual band pair formation was assumed because more band pairs were found in the centra of
36 larger individuals (Ridewood 1921). Subsequent studies demonstrated positive relationships
37 between increasing body size and both centrum size and number of band pairs across individuals

38 (Ishiyama 1951; Jones and Geen 1977; Cailliet and Goldman 2004) supporting the use of
39 vertebral bands pairs for age estimates.

40 Over time issues have become apparent in the use of vertebral band pairs for age estimates.
41 Several species do not deposit band pairs annually, instead, depositing them relative to somatic
42 growth (Natanson and Cailliet 1990; Tanaka 1990; Natanson *et al.* 2008). Other studies
43 demonstrate decreased band pair deposition in older and larger individuals showing band pair
44 counts underestimate age (see review by Harry 2018). Identifying these instances is critical to
45 avoid inaccurate age estimates.

46 Another issue relates to the positive relationship between the number of band pairs and the
47 size of the centrum. This positive relationship is true among individuals where larger individuals
48 have larger centra with more band pairs (Cailliet and Goldman 2004), but it is also true within an
49 individual; where centrum size and number of band pairs varies along the vertebral column
50 (Natanson *et al.* 2018). Several studies have shown that small (young of the year [YOY])
51 individuals often have the same number of band pairs throughout their vertebral column and
52 similar centrum sizes however, band pair counts and centra size increasingly vary along the
53 vertebral column in larger, maturing individuals of the same species (Natanson and Cailliet 1990;
54 Natanson *et al.* 2008; Huvaneers *et al.* 2013; Natanson *et al.* 2018). This has been shown in
55 several species belonging to different families. If band pair deposition varies along the vertebral
56 column with centrum size then band pair count cannot be related to age. Natanson *et al.* (2018)
57 concluded that the number of band pairs vary along the column in direct correlation with the
58 girth of the fish where the centra were taken and any relationship with time is not causative, but
59 correlative through the somatic growth rate.

60 A majority of the research on elasmobranch vertebral column morphology and frequency
61 of band pair deposition have been conducted on sharks, not batoids (Harry 2018; Natanson *et al.*
62 2018). Ageing of batoids is subject to the same assumptions (Ishiyama 1951; Cailliet *et al.*
63 2006), but there are only three confirmed batoid species demonstrating age underestimation
64 (Natanson 1993; McPhie and Campana 2009; Pierce and Bennett 2009; James 2020). This is
65 likely due to fewer validation studies on batoids (Harry 2018) rather than the absence of age
66 underestimation.

67 The goal of the present study was to investigate whether vertebral band pairs are related
68 to somatic growth and/or ontogeny in batoids. We measured centrum dimensions and counted
69 the band pairs in individual centra along the columns of various-sized individuals of five batoid
70 species (little skate, *Leucoraja erinacea*, winter skate, *Leucoraja ocellata*, barndoor skate,
71 *Dipturus laevis*, Atlantic stingray, *Dasyatis sabina*, and round ray, *Urotrygon helleri*; James
72 2018). We then related centrum morphology to the number of band pairs of each centrum along
73 the vertebral column and examined these relationships by size, sex, and species. Variation of
74 band pair counts along the vertebral column of an individual calls into question the assumption
75 that band pair formation is directly related to time, and raises concerns for using band pair counts
76 for age estimates.

77

78 **Methods**

79 Skates were obtained opportunistically from commercial fishermen off the coast of
80 Rhode Island and Massachusetts, USA. Atlantic stingrays were obtained from Seal Beach,
81 California, and round rays from Indian River, Florida. Total length (TL; straight-line distance
82 from snout tip to tail tip), and disc width (DW; straight-line distance from wing tip to wing tip)

83 were measured to the nearest 0.1 cm on all individuals. By convention, TL is used in analyses for
84 skates, while DW is used in analyses for stingrays (Francis 2006). Sex and maturity status were
85 determined by visually inspecting gonad condition (Ebert 2005).

86 Forty-two little skates (14 immature [small], 15 near size-at-maturity [medium], and 13
87 mature [large]), six winter skates and six barndoor skates (two small, two medium, and two large
88 of each), nine Atlantic stingrays (three small, three medium, and three large), and ten round rays
89 (two small, four medium, and four large) were collected and measured for analysis of centrum
90 morphology (Table 1; James 2018). Band pair counts were conducted along the vertebral
91 columns of a subset of nine little skates, and six each of Atlantic stingray and round rays (Table
92 1). The subset for each species included small, medium, and large individuals. All winter skates
93 and barndoor skates were analyzed for the number of band pairs (Table 1). Males and females
94 were evenly represented where possible (Table 1).

95

96 *Centrum Morphology*

97 The vertebral column was extracted from each fish starting with the first vertebra behind
98 the synarcual cartilage and ending at the 80th vertebra. One round ray had sustained a tail injury
99 and vertebrae were only available to the 48th centrum.

100 Each vertebral centrum was measured to the nearest 0.1 mm in three dimensions: dorso-
101 ventral diameter (DVD), lateral diameter (LD) and rostro-caudal length (LEN), using Vernier
102 calipers following Natanson *et al.* (2018) (Figure 1). Each measurement was divided by TL for
103 the skate species and DW for the stingray species to standardize data across sizes for direct
104 comparison. Standardized data were plotted against centrum number for each individual, noting
105 the centrum number at the transition from abdominal cavity to tail.

106 Multiple generalized additive models (GAMs) were fit to each species using mgcv
107 package in R (Wood 2011; R Core Team 2017) to assess whether each centrum measurement
108 (DVD, LD, and LEN) was similar within a species, by sex, by size, or by individual (James
109 2018). Four GAM variations were run for each measurement for each species: all data pooled,
110 data grouped by sex, size class, and individual. Three GAM iterations were run: different
111 intercepts only, different smoothing functions only, and different intercepts and smoothing
112 functions for each sex, size class, and individual scenario. For each GAM variation the number
113 of knots was specified to be larger than the estimated degrees of freedom using the gam.check
114 function of mgcv in R. Model fit was assessed with Akaike's Information Criterion (AIC)
115 (Haddon 2001).

116

117 *Band Pair Counts*

118 To determine if band pair number varied along the vertebral column of an individual,
119 every fifth centrum was processed histologically to visually enhance the band pairs (as per
120 Natanson *et al.* 2007). Centrum sections were viewed under a dissecting microscope (Nikon
121 SMZ1500[®], Melville, NY, USA¹) using reflected light and images were captured with a digital
122 camera (Nikon DSR12, Tokyo, Japan) and image processing software (NIS Elements, v. 4.40,
123 Nikon, Tokyo, Japan). Two band pair counts were made for each individual by a primary (KCJ)
124 and a secondary reader using editing software (Adobe Photoshop CC (Adobe Systems, San Jose,
125 CA, USA)). The birth band was identified as the first fully-formed band beyond the focus and
126 was associated with an angle change in the corpus calcareum of the centrum (Casey *et al.* 1985;
127 Cailliet and Goldman 2004). Each sample was assigned a unique ID number so that the reader

¹ Use of Trade Names does not imply endorsement from the NMFS.

128 had no knowledge of the size, sex, or location along the vertebral column. Coefficient of
129 variation (CV) was calculated within and between readers to assess repeatability of counts and
130 precision (Chang 1982). Values <10% were considered acceptable. Bias, as a result of either
131 systematic or random error, was assessed using the Evans-Hoenig's (1998) test of symmetry.
132 Band pair counts of zero are excluded from precision and bias analyses. Intra-reader precision
133 and bias were compared between the first and second count of each reader while inter-reader
134 precision and bias was compared between the second band pair counts. If the second band pair
135 count differed by three or more band pairs, the centrum was examined together and a consensus
136 count was reached. Final band pair counts were assigned from the primary reader's second count
137 or the consensus count.

138 Final band pair count was plotted by centrum number for each individual. The mean band
139 pair count and 95% confidence interval (CI) of the mean was calculated for each individual to
140 test if band-pair count varied significantly among centra along the vertebral column. If more than
141 5% of the band pair counts fell outside of the 95% CI then band pair counts were significantly
142 different within an individual. A mixed-effects model was used to determine if there was a
143 correlation between band pair count and the three centrum measurements for each species with
144 individual included as a random effect.

145

146 **Results**

147 *Centrum Morphology*

148 Centrum morphology varied along the vertebral column in all species (Figure 2-6; James
149 2018). The transition between abdominal and caudal centra occurred at the 24th to the 47th
150 centrum depending on the species (Table 1). For little and winter skates DVD and LD increased

151 from the head and peaked at the level of the mid-abdominal cavity (approximately in line with
152 the pectoral fin tips) then decreased through the transition from abdominal to caudal centra and
153 continued to decrease in the caudal centra (Figures 2 and 3). Abdominal centra were wider than
154 they were tall ($LD > DVD$; ovoid), while caudal centra were circular ($LD = DVD$; Figures 2 and
155 3). Rostro-caudal length in little and winter skates increased from the head to the transition from
156 abdominal to caudal centra where the LEN decreased sharply; LEN was constant among the
157 caudal centra (Figures 2 and 3).

158 While trends in size of DVD and LD along the vertebral column in the barndoor skate
159 were similar as in winter and little skates, the centra shape was different along the column.
160 Barndoor skate centra were circular along the entire length of the column (Figure 4). In the
161 barndoor skate, LEN followed a similar trend as in little and winter skates until approximately
162 the 45th centrum, where LEN was greater than DVD and LD (Figure 4).

163 Atlantic stingray and round ray had similar centra morphologies, which differed from the
164 skate species. In the rays, DVD and LD increased from the head, were constant along the
165 abdominal cavity, and decreased in the caudal centra (Figures 5 and 6). Atlantic stingray and
166 round ray centra were slightly ovoid along the abdominal cavity. Rostro-caudal length increased
167 from the head until the transition from abdominal to caudal vertebrae after which LEN quickly
168 decreased, but the decrease was less dramatic than that seen in the skate species. In both ray
169 species, LEN was constant along the tail.

170 For all species studied, the centrum morphology along the vertebral column was best
171 described by individual variation (Figure 7). The best-fit GAMs modeled each individual with its
172 own intercept and smoothing function for all species and measurements rather than by sex or
173 species (Supplemental Table 1; James 2018). The only exception was the LEN measurements in

174 Atlantic stingrays, which was best modeled by each individual with its own intercept, but the
175 same smoothing function for all individuals (Supplemental Table 1). However, all models
176 applied to the DVD and LD measurements of little skate, winter skate, barndoor skate, and round
177 ray fit the data well with adjusted $r^2 > 0.79$ (Supplemental Table 1). The LEN measurement for
178 these species and all measurements for Atlantic stingray had lower adjusted r^2 values ranging
179 from 0.28 to 0.91 (Supplemental Table 1).

180

181 *Band Pair Counts*

182 The number of centra counted per batoid ranged from 11-17 (Table 1). Intra-reader CV
183 ranged between 6.9 - 14.9% for primary reader and 6.4 - 12.5% for the secondary reader (Table
184 2), while the inter-reader CV ranged from 10.1 - 21.4% (Table 2) depending on species. Of 478
185 samples, 13.6% were read by consensus (Table 2). Intra-reader bias was detected for barndoor
186 skate (primary reader) and for Atlantic stingray (secondary reader) using the Evans-Hoenig
187 (1998) test of symmetry. Inter-reader bias was detected only for barndoor skate (Table 2).

188 Significant differences in band pair counts were found along the column of all individuals
189 except in YOY. Excluding YOY, 17.6 – 100% of the band pair counts fell outside of the 95% CI
190 (Figures 2-6; James 2018). The band pair counts for the three skate species were roughly
191 correlated with the pattern of the DVD and LD measurements (Figures 2-4). Band pair counts for
192 the Atlantic stingray and the round ray did not exhibit a trend along the vertebral column, but
193 still showed significant differences among different centra within an individual (Figures 5 and 6).
194 The largest range in band pair counts within an individual was seven band pairs for little skate,
195 eight band pairs for winter skate, 11 band pairs in barndoor skate, five band pairs in Atlantic
196 stingray, and six band pairs for round ray. Abdominal centra typically had higher band pair

197 counts than caudal centra. The two smallest Atlantic stingray specimens examined were YOY
198 and did not have band pairs (Figure 5).

199 The number of band pairs was related to centrum morphology in all species, except
200 winter skate. Dorso-ventral diameter, LD, and LEN were significantly correlated with the band
201 pair counts of little skate, barndoor skate, and round ray (Table 3; James 2018). Atlantic stingray
202 were significantly correlated with DVD and LD, but not with LEN. Winter skate did not have
203 any significant correlations.

204

205 **Discussion**

206 Variable band pair counts among centra within an individual has now been observed in
207 15 species representing 9 elasmobranch families (Natanson and Cailliet 1990; Natanson *et al.*
208 2008; Huveneers *et al.* 2013; Natanson *et al.* 2018; current study). The presence of this variation
209 suggests that the mechanism that regulates the formation of band pairs is not related to time.
210 When differences in band pair counts between more anterior and more posterior centra were
211 detected in previous studies, it was suggested that band pairs in smaller, caudal centra were more
212 difficult to interpret (Brown and Gruber 1988; Officer *et al.* 1996; Natanson *et al.* 2006; Piercy *et*
213 *al.* 2006). In this study we did not find it difficult to interpret band pairs in smaller centra, and we
214 confirmed that the number of band pairs within an individual varies along the vertebral column
215 in batoids. Band pair counts that vary among centra along the vertebral column of an individual
216 cannot accurately reflect a single age estimate (Natanson *et al.* 2018). The positive relationship
217 observed between band pair count and centra morphology for four of the five batoid species
218 examined in this study makes band pair counts unreliable as a tool for ageing in these species.

219 The hypothesis that structural needs of the individual may regulate the formation of band
220 pairs is suggested on the basis that mineralization of the centra enhances skeletal strength and
221 mechanical support (Kemp and Westrin 1979; Clement 1992; Porter *et al.* 2006; 2007).
222 Regardless of species, larger centra have more band pairs indicating that band pair deposition is a
223 structural requirement of the individual and not related to time (Natanson *et al.* 2018). The
224 positive correlation of body girth measurements to centrum size led Natanson *et al.* (2018) to
225 suggest that differences in deposition patterns are correlated with body type and swimming mode
226 among species. This was supported by Thomson and Simanek's (1977) five categories of body
227 and tail type, in which species of similar body shapes and swimming styles also had similar
228 centrum morphology and Ingle *et al.* (2018) who found that larger centra with more band pairs
229 had lower toughness and stiffness than smaller centra with fewer band pairs. More band pairs
230 (present in abdominal vertebrae) may support body mass, while fewer band pairs (present in
231 anterior and posterior vertebrae) with higher toughness and stiffness allow absorption of more
232 energy and elastic recoil to facilitate swimming (Ingle *et al.* 2018; Natanson *et al.* 2018). Batoids
233 possess vastly different swimming styles than many sharks, however the Atlantic angel shark
234 uses a swimming mode that is an intermediate between caudal fin propulsion and paired fin
235 propulsion (Wilga and Lauder 2004). These dorso-ventrally flattened sharks also demonstrate a
236 relationship between body shape and centrum morphology (Natanson *et al.* 2018), so it is
237 reasonable to extend this relationship to batoids. The body girth measurements used by Natanson
238 *et al.* (2018) did not translate to a batoid body plan (James, unpub. data) so a different approach
239 will have to be used to investigate the relationship between body shape and centrum
240 morphology.

241 Centrum morphologies in these five batoid species were roughly similar to the centrum
242 morphology of sharks (Natanson *et al.* 2018). The Atlantic angel shark had the most similar
243 centrum morphology to the batoids with the largest centrum in the middle of the abdominal
244 cavity (approximately in line with the tips of the pectoral fins), while the largest centrum for the
245 other shark species was at the end of the abdominal cavity (Natanson *et al.* 2018). The Atlantic
246 angel shark also had ovoid centra (Natanson *et al.* 2018) similar to the abdominal centra of
247 batoids. In carcharhinids and lamnids, centra were circular, except for the abdominal centra of
248 very large lamnids (i.e. shortfin mako; Natanson *et al.* 2018). We suggest that ovoid centra are
249 characteristic of dorso-ventrally compressed elasmobranchs and may be related to undulation of
250 pectoral fins as a swimming strategy, a benthic lifestyle, or the ability to bend dorso-ventrally,
251 not just laterally as in most sharks.

252 Similarities in centrum morphology exist across and within species. Individual variation
253 was the best descriptor of centrum morphology for batoids, however most GAMs fit the data
254 well (Supplemental Table 1). The good fit of these models may be in part due to the uniformity
255 of morphology of caudal vertebrae (e.g. Figure 7). In contrast, abdominal vertebrae display high
256 individual variation (e.g. Figure 7) suggesting that the conditions an individual experiences
257 influences growth. In Atlantic salmon, *Salmo salar* Linnaeus 1758, centra within an individual
258 grew at different rates depending on the photoperiod regime (Fjelldal *et al.* 2005). Based on our
259 results, we suggest that factors affecting individual body growth (food availability, temperature,
260 population density, and genetics [McDowall 1994]) may also affect individual centrum
261 morphology in batoids.

262 The paradigm of annual band pair deposition within centra of elasmobranchs has been
263 disproven in many species (Natanson and Cailliet 1990; Tanaka 1990; Francis *et al.* 2007;

264 Huvaneers *et al.* 2013; Harry 2018; Natanson *et al.* 2018). Here we add to that body of literature
265 with five batoids species, supporting the idea that band pair number is related to somatic growth
266 and/or the structural needs of the individual in elasmobranchs as a group (Natanson and Cailliet
267 1990; Tanaka 1990; Natanson *et al.* 2008; Natanson *et al.* 2018). We also reinforce the call for
268 caution when using band pair counts as a proxy of age without validation (Beamish and
269 MacFarlane 1983). Future work investigating the impact of inaccurate ages on stock-assessment
270 model results and determining an alternate method to age elasmobranchs should be at the
271 forefront of elasmobranch ageing.

272

273 **Conflicts of Interest**

274 The authors declare no conflicts of interest.

275

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285 James (2018).

286 **References**

287 Beamish, R. J., and McFarlane, G. A. (1983). The forgotten requirement for age validation in
288 fisheries biology. *Transactions of the American Fisheries Society* **112**, 735-743.

289

290 Brown, C. A., and Gruber, S. H. (1988). Age assessment of the Lemon shark, *Negaprion*
291 *brevirostris*, using tetracycline validated vertebral centra. *Copeia* **1988**, 747-753.

292

293 Cailliet, G. M., and Goldman, K. J. (2004). Ch 14 Age determination and validation in
294 Chondrichthyan fishes. *Biology of sharks and their relatives* 2004 CRC Press LLC

295

296 Cailliet, G. M., Smith, W. D., Mollet, H. F., and Goldman, K. J. (2006). Age and growth studies
297 of chondrichthyan fishes: the need for consistency in terminology, verification,
298 validation, and growth function fitting. *Environmental Biology of Fishes* **77**, 211-228.

299

300 Casey, J. G., Pratt, H. L., and Stillwell, C. E. (1985). Age and growth of the sandbar shark
301 (*Carcharhinus plumbeus*) from the Western North Atlantic. *Canadian Journal of*
302 *Fisheries and Aquatic Sciences* **42**, 963-975.

303

304 Chang, W. Y. B. (1982). A statistical method for evaluating the reproducibility of age
305 determination. *Canadian Journal of Fisheries and Aquatic Sciences* **39**, 1208-1210.

306

307 Clement, J. G. (1992). Re-examination of the fine structure of endoskeletal mineralization in
308 Chondrichthyans: Implications for growth, ageing and calcium homeostasis. Australian
309 Journal of Marine and Freshwater Research **43**, 157-181.

310

311 Ebert, D. A. (2005). Reproductive biology of skates, *Bathyraja* (Ishiyama), along the eastern
312 Bering Sea continental slope. Journal of Fish Biology **66**, 618-649.

313

314 Evans, G. T., and Hoenig, J. M. (1998). Testing and viewing symmetry in contingency tables,
315 with applications to readers of fish ages. Biometrics **54**, 620-629.

316

317 Fjelldal, P. G., Nordgarde, U., Berg, A., Grotmol, S., Totland, G. K., Wargelius, A., and Hansen,
318 T. (2005). Vertebrae of the trunk and tail display different growth rates in response to
319 photoperiod in Atlantic salmon, *Salmo salar* L., post-smolts. Aquaculture **250**, 516-524.

320

321 Francis, M.P. (2006). Morphometric minefields—towards a measurement standard for
322 chondrichthyan fishes. Environmental Biology of Fishes **77**, 407-421.

323

324 Francis, M. P., Campana, S. E., and Jones, C. M. (2007). Age under-estimation in New Zealand
325 Porbeagle Sharks (*Lamna nasus*): is there an upper limit to ages that can be determined
326 from shark vertebrae? Marine and Freshwater Research **58**, 10-23.

327

328 Haddon, M. (2001). Modeling and quantitative measures in fisheries. Chapman & Hall/CRC
329 Press, Boca Raton FL

330

331 Harry, A. V. (2018). Evidence of systemic age underestimation in shark and ray ageing studies.

332 Fish and Fisheries **2018**, 1-16.

333

334 Haskell, W. L. (1948). An investigation of the possibility of determining the age of sharks

335 through annuli as shown in cross-sections of vertebrae. Annual Report of the Marine

336 Laboratory of the Texas Game, and Fish Commission **FY 1948-49**, 212-217.

337

338 Huveneers, C., Stead, J., Bennett, M. B., Lee, K. A., and Harcourt, R. G. (2013). Age and growth

339 determination of three sympatric wobbegong sharks: How reliable is growth band

340 periodicity in Orectolobidae? Fisheries Research **147**, 413-425.

341

342 Ingle, D. I., Natanson, L. J., and Porter, M. E. (2018). Mechanical behavior of shark vertebral

343 centra at biologically relevant strains. Journal of Experimental Biology **221**, jeb188318.

344

345 Ishiyama, R. (1951). Studies on the rays and skates belonging to the family Rajidae, found in

346 Japan and adjacent regions. 2. On the age-determination of Japanese Black-Skate *Raja*

347 *fusca*. Bulletin of the Japanese Society for the Science of Fish **16**, 112-118.

348

349 James, K.C. (2018). Analysis of band pair formation in elasmobranch vertebrae with

350 implications for fisheries management. PhD Thesis. University of Rhode Island. Open

351 Access Dissertations. Paper 760.

352

353 James, K.C. (2020). Vertebral growth and band-pair deposition in sexually mature little skates

354 *Leucoraja erinacea*: is adult band-pair deposition annual? *Journal of Fish Biology* **96**, 4-

355 13.

356

357 Jones, B. C., and Geen, G. H. (1977). Age determination of an elasmobranch (*Squalus acanthias*)

358 by x-ray spectrometry. *Journal of the Fisheries Research Board of Canada* **34**, 44-48.

359

360 Kemp, N. E., and Westrin, S. K. (1979). Ultrastructure of calcified cartilage in the endoskeletal

361 tesserae of sharks. *Journal of Morphology* **160**, 75-102.

362

363 McDowall, R. M. (1994). On size and growth in freshwater fish. *Ecology of Freshwater Fish* **3**,

364 67-79.

365

366 McPhie, R. P., and Campana, S. E. (2009). Bomb dating and age determination of skates (family

367 Rajidae) off the eastern coast of Canada. *ICES Journal of Marine Science* **66**, 546-560.

368

369 Natanson, L. J. (1993). Effect of temperature on band deposition in the Little Skate, *Raja*

370 *erinacea*. *Copeia* **1993**, 199-206.

371

372 Natanson, L., Kohler, N., Ardizzone, D., Cailliet, G., Wintner, S., and Mollet, S. (2006).

373 Validated age and growth estimates for the shortfin mako, *Isurus oxyrinchus*, in the North

374 Atlantic Ocean. *Environmental Biology of Fishes* **77**, 367-383.

375

376 Natanson, L. J., and Cailliet, G. M. (1990). Vertebral growth zone deposition in Pacific Angel
377 Sharks. *Copeia* **1990**, 1133-1145.

378

379 Natanson, L. J., Skomal, G. B., Hoffmann, S., Porter, M., Goldman, K. J., and Serra, D. (2018).
380 Age and growth of elasmobranchs: do band pairs on vertebral centra record age? *Marine*
381 and Freshwater Research

382

383 Natanson, L. J., Sulikowski, J. A., Kneebone, J. R., Tsang, P. C. (2007). Age and growth
384 estimates for the Smooth Skate, *Malacoraja senta*, in the Gulf of Maine. *Environmental*
385 *Biology of Fishes* **80**, 293-308.

386

387 Natanson, L. J., Wintner, S., Johansson, F., Piercy, A. N., Campbell, P., De Maddalena, A.,
388 Gulak, S. J., Human, B., Fulgosi, F. C., Ebert, D. A., Hemida, F., Mollen, F. H., Vanni,
389 S., Burgess, G. H., Compagno, L. J. V., and Wedderburn-Maxwell, A. (2008).
390 Ontogenetic vertebral growth patterns in the Basking Shark, *Cetorhinus maximus*. *Marine*
391 *Ecology Progress Series* **361**, 267-278.

392

393 Officer, R. A., Gason, A. S., Walker, T. I., and Clement, J. G. (1996). Sources of variation in
394 counts of growth increments in vertebrae from gummy shark, *Mustelus antarcticus*, and
395 school shark, *Galeorhinus galeus*: implications for age determination. *Canadian Journal*
396 *of Fisheries and Aquatic Sciences* **53**, 1765-1777.

397

398 Pierce, S. J., and Bennett, M. B. (2009). Validated annual band-pair periodicity and growth
399 parameters of blue-spotted maskray *Neotrygon kuhlii* from south-east Queensland,
400 Australia. *Journal of Fish Biology* **75**, 2490-2508.

401

402 Piercy, A. N., Ford, T. S., Levy, L. M., and Snelson, F. F. (2006). Analysis of variability in
403 vertebral morphology and growth ring counts in two Carcharhinid sharks. *Environmental
404 Biology of Fishes* **77**, 401-406.

405

406 Porter, M. E., Beltran, J. L., Koob, T. J., and Summers, A. P. (2006). Material properties and
407 biochemical composition of mineralized vertebral cartilage in seven elasmobranch
408 species (Chondrichthyes). *The Journal of Experimental Biology* **209**, 2920-2928.

409

410 Porter, M. E., Koob, T. J., and Summers, A. P. (2007). The contribution of mineral to the
411 material properties of vertebral cartilage from the smooth-hound shark *Mustelus
412 californicus*. *The Journal of Experimental Biology* **210**, 3319-3327.

413

414 R Core Team. (2017). R: A language and environment for statistical computing. R Foundation
415 for Statistical Computing, Vienna, Austria URL <https://www.R-project.org/>.

416

417 Ridewood, W. G. (1921). On the calcification of the vertebral centra in sharks and rays.
418 *Philosophical Transactions of the Royal Society B* **210**, 311-407.

419

420 Tanaka, S. (1990). Age and growth studies on the calcified structures of newborn sharks in
421 laboratory aquaria using tetracycline. In *Elasmobranchs as Living Resources: Advances*
422 in the Biology, Ecology, Systematics, and the Status of Fisheries NOAA Technical
423 Report **90**, 189-202.

424

425 Thomson, K. S., and Simanek, D. E. (1977). Body form and locomotion in sharks. *American*
426 *Zoologist* **17**, 343-354.

427

428 Wilga, C. A. D., and Lauder, G. V. (2004). Biomechanics of locomotion in sharks, rays, and
429 chimeras. Carrier, J. C., Musick, J. A., and Heithaus, M. R. (eds.) *Biology of Sharks and*
430 *Their Relatives* 2004 CRC Press LLC

431

432 Wood, S. N. (2011). Fast stable restricted maximum likelihood and marginal likelihood
433 estimation of semiparametric generalized linear models. *Journal of the Royal Statistical Society B* **73**, 3-36.

435

436 **Table and Figure Captions**

437

438 Table 1. Individuals of five batoid species used in this study. For little skate *Leucoraja erinacea*,
439 the first nine individuals were analyzed for band pair counts. For winter skate *Leucoraja*
440 *ocellata*, barndoor skate *Dipturus laevis*, Altantic stingray *Dasyatis sabina*, and round ray
441 *Urobatis halleri*, the first six individuals were analyzed for band pair counts. All individuals
442 were measured for centrum morphology. For brevity, only condensed data are presented for the
443 additional 33 little skates used for centrum morphology. TL is total length, DW is disc width, L
444 is large, M is medium in the Size column and Male in the Sex column, S is for small, F is for
445 female.

446

447 Table 2. Bias and precision of band-pair counts within and between readers. Asterisk indicates
448 significant bias.

449

450 Table 3. Linear mixed-effects model comparing band-pair counts with the three centrum
451 measurements with individual included in the model as a random effect. Asterisk indicates
452 significant correlation.

453

454 Figure 1. Diagram of a vertebral centrum and the three measurements: dorso-ventral diameter
455 (DVD), lateral diameter (LD), and rostro-caudal length (LEN). R is rostral and C is caudal.

456

457 Figure 2. Band-pair count and standardized centrum measurements along the vertebral column
458 for nine little skates analyzed for band pairs.

459

460 Figure 3. Band-pair count and standardized centrum measurements along the vertebral column
461 for the six winter skates analyzed for band pairs.

462

463 Figure 4. Band-pair count and standardized centrum measurements along the vertebral column
464 for six barndoor skates analyzed for band pairs.

465

466 Figure 5. Band-pair count and standardized centrum measurements along the vertebral column
467 for six Atlantic stingrays analyzed for band pairs.

468

469 Figure 6. Band-pair count and standardized centrum measurements along the vertebral column
470 for six round rays analyzed for band pairs.

471

472 Figure 7. Example of generalized additive model variations with (a) all data pooled, data pooled
473 (b) by sex, (c) by size class, and (d) by individual fit to barndoor skate data.

474

475 Supplemental Table 1. Generalized additive model results for 10 different models to best
476 describe centrum morphology along the vertebral column for five batoid species. k is the number
477 of knots used in each model.

Table 1. Individuals of five batoid species used in this study. For little skate *Leucoraja erinacea*, the first nine individuals were analyzed for band pair counts. For winter skate *Leucoraja ocellata*, barndoor skate *Dipturus laevis*, Altantic stingray *Dasyatis sabina*, and round ray *Urotrygon halleri*, the first six individuals were analyzed for band pair counts. All individuals were measured for centrum morphology. For brevity, only condensed data are presented for the additional 33 little skates used for centrum morphology. TL is total length, DW is disc width, L is large, M is medium in the Size column and Male in the Sex column, S is for small, F is for female.

Species	TL (cm)	DW (cm)	Size	Sex	Maturity	Transition ^a	Centra Counted ^b	Individual ID
Little Skate	49.0	29.0	L	F	Mature	22	13	LE03
	48.5	29.5	L	M	Mature	22	12	LE01
	48.0	27.0	L	F	Mature	24	17	LE07
	42.8	25.2	M	F	Immature	27	17	LE06
	41.5	23.8	M	M	Mature	26	17	LE05
	39.6	24.0	M	F	Immature	27	17	LE19
	26.1	14.5	S	M	Immature	26	17	LE04
	25.6	15.5	S	F	Immature	21	14	LE02
	23.4	13.7	S	F	Immature	22	15	LE20
	43.0-	24.6-	5 F, 5					
	47.5	27.3	L	M	1 Immature, 9 Mature	22-25	N/A	N/A
	38.3-	22.8-	5 F, 7					
	42.4	25.4	M	M	11 Immature, 2 Mature	23-26	N/A	N/A
	25.1-	15.2-	5 F, 6					
	32.6	19.8	S	M	Immature	21-25	N/A	N/A
Winter Skate	80.0	49.9	L	F	Mature	27	17	LO04
	75.3	47.8	L	M	Mature	30	17	LO07
	63.3	39.1	M	F	Immature	26	17	LO08
	61.7	40.5	M	M	Immature	27	16	LO09
	43.2	26.9	S	M	Immature	31	17	LO06
	37.1	22.0	S	F	Immature	29	16	LO05
Barndoor Skate	130.0	92.6 ^c	L	M	Mature	25	17	DL04

	117.4	80.9	L	F	Mature	25	17	DL16
	107.5	76.7 ^c	M	F	Mature	27	14	DL01
	90.0	64.3 ^c	M	M	Immature	27	16	DL03
	52.3	37.6	S	F	Immature	31	17	DL05
	49.0	36.3	S	M	Immature	24	17	DL06
Atlantic Stingray	48.9	26.5	L	F	Mature		15	DS24
	44.8	26.9	L	F	Mature	36	17	DS26
	40.0	18.0	M	F	Immature	49	16	DS32
	36.6	17.8	M	F	Immature	46	17	DS30
	34.2	13.4	S	F	Immature	46	16	DS27
	25.0	13.3	S	F	Immature	47	15	DS33
	59.8	30.3	L	F	Mature	31	N/A	DS31
	42.6	16.0	M	F	Immature	46	N/A	DS29
Round Ray	30.0	11.0	S	M	Immature	46	N/A	DS25
	36.7	20.4	L	M	Mature	31	17	UH01
	21.3	20.0	L	F	Mature	32	11	UH08
	30.5	16.7	M	F	Mature	32	17	UH05
	30.0	16.5	M	M	Mature	34	16	UH10
	24.0	13.3	S	M	Immature	34	17	UH09
	22.8	13.1	S	F	Immature	41	17	UH07
	34.2	19.8	L	M	Mature	34	N/A	UH02
	33.4	18.6	L	M	Mature	30	N/A	UH03
	29.9	17.8	M	F	Mature	31	N/A	UH04
	27.2	16.1	M	F	Mature	31	N/A	UH06

^aVertebral number where the transition between abdominal and caudal vertebrae occurs.

^bNumber of centra counted for each individual. N/A means band-pair counts were not determined.

^cDisc Width was estimated using Total Length to Disc Width relationship from Gedamke 2006.

Table 2. Bias and precision of band pair counts within and between readers for each of the five batoid species. n is the total number of centra counted.

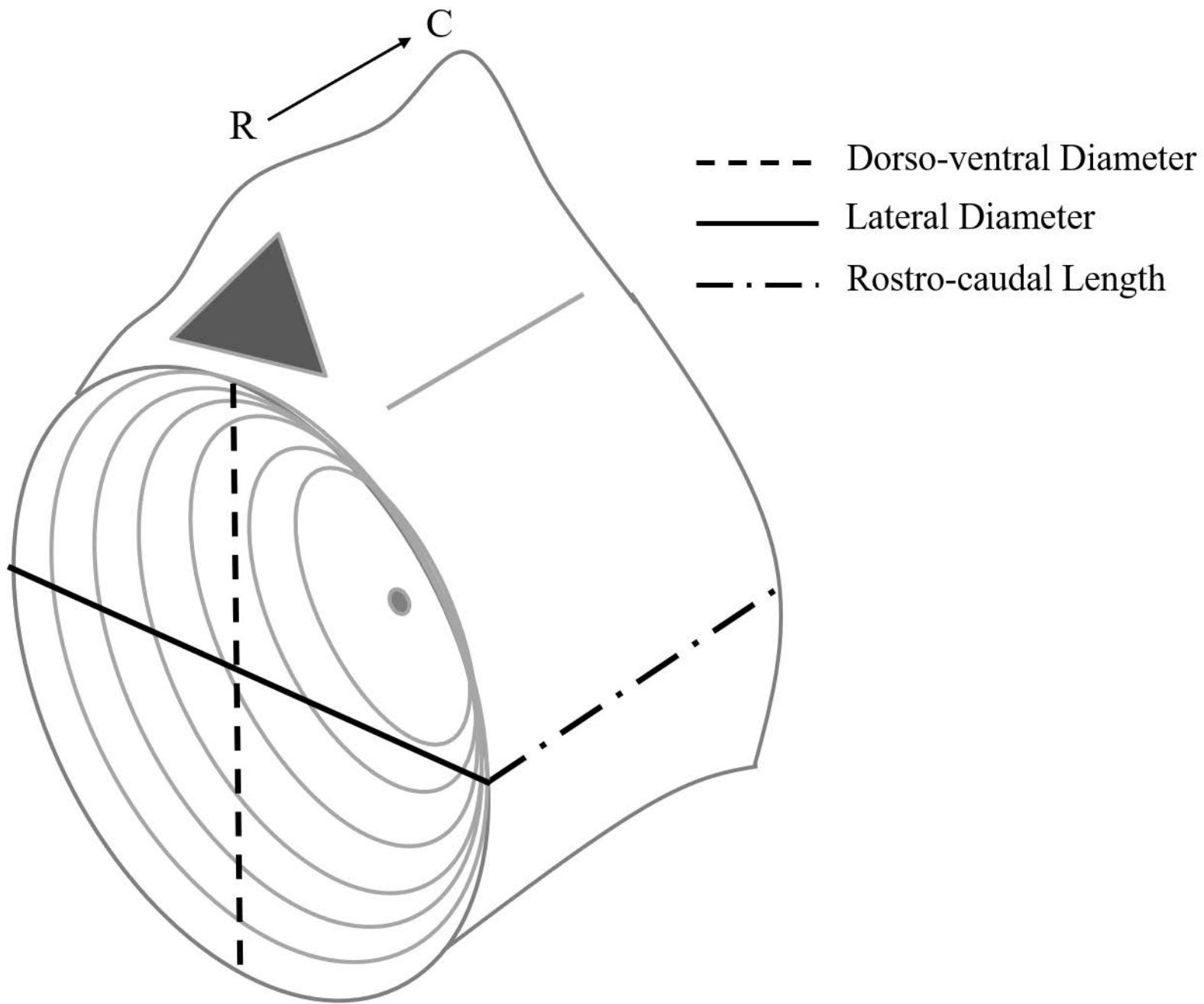
Species	Evans-Hoenig (1998) Bias test					
	n ^a	df	χ^2	p	CV	Consensus ^b
Little Skate	Primary reader	4	8.71	0.069	9.08	
	Second reader	3	2.38	0.497	11.89	
	Inter-reader	130	7	11.09	0.135	16
Winter Skate	Primary reader	6	11.30	0.800	6.86	
	Second reader	5	7.69	0.174	8.80	
	Inter-reader	99	6	7.22	0.301	10.08
Barndoor Skate	Primary reader	5	11.49	0.042	*	9.35
	Second reader	4	3.42	0.490		6.37
	Inter-reader	95	5	12.63	0.027	*
Atlantic Stingray	Primary reader	3	2.50	0.475		14.88
	Second reader	2	9.76	0.008	*	12.54
	Inter-reader	63	4	8.43	0.077	21.35
Round Ray	Primary reader	4	4.06	0.398		9.81
	Second reader	3	2.44	0.486		9.93
	Inter-reader	91	4	3.31	0.507	11.83
						15

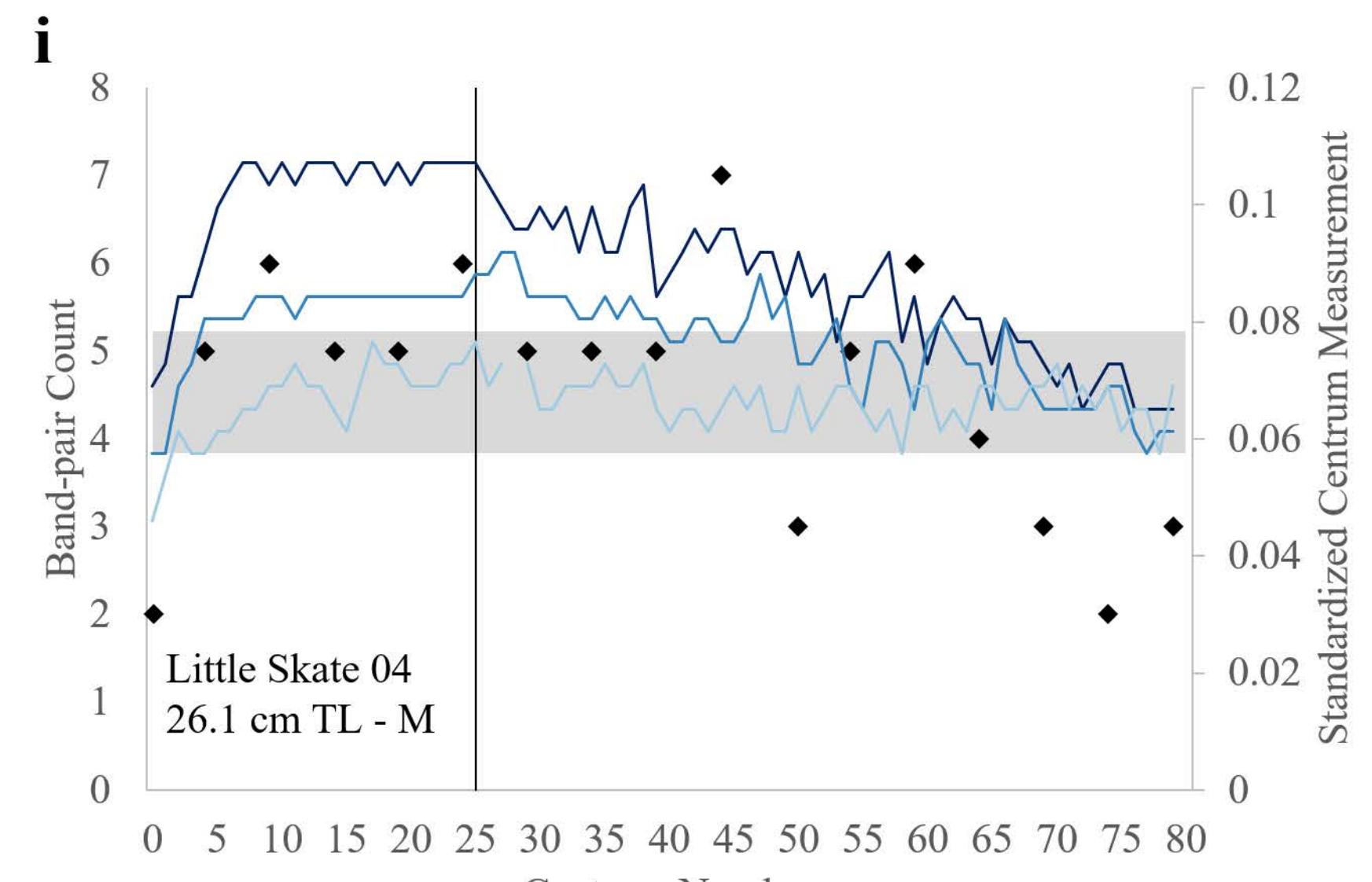
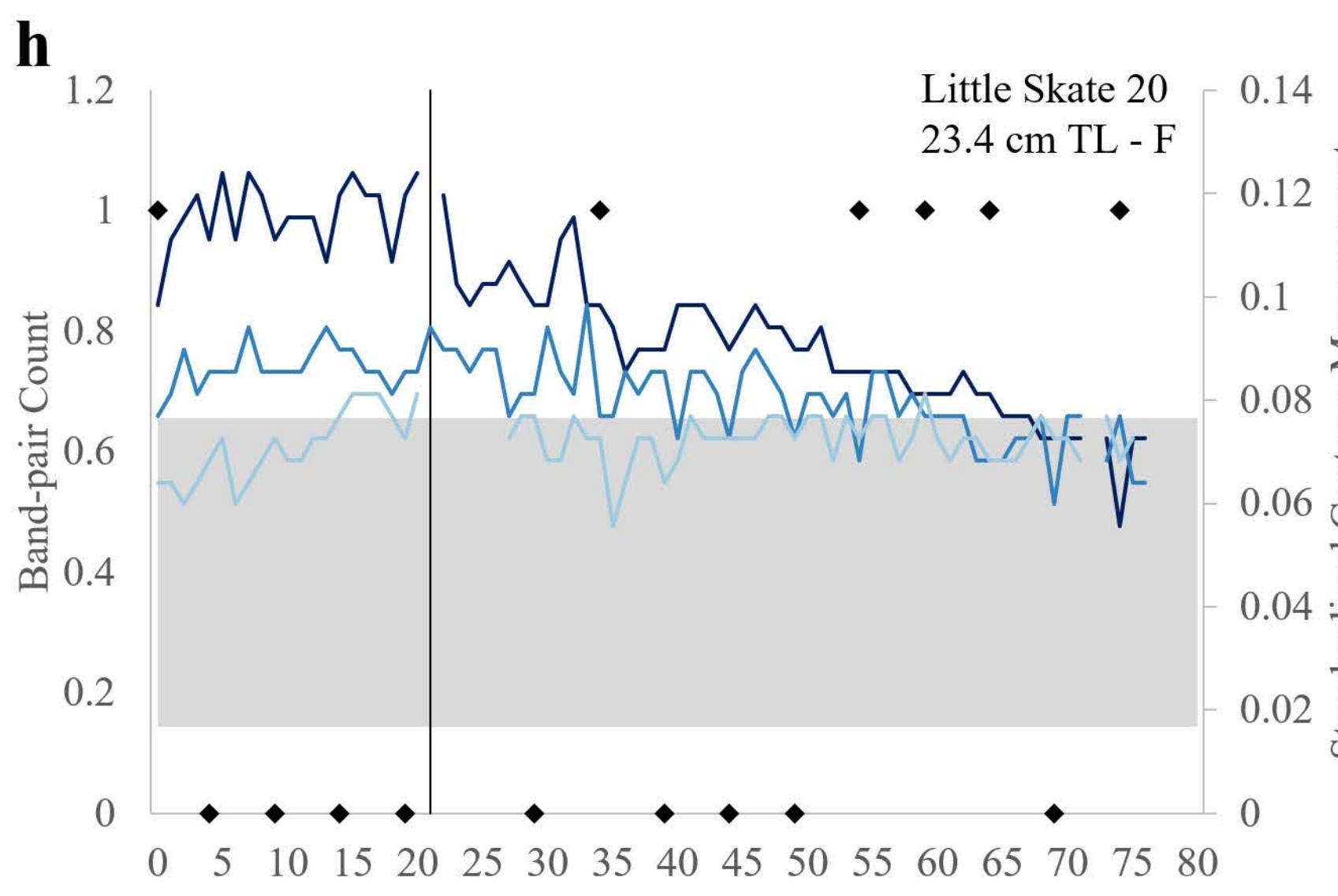
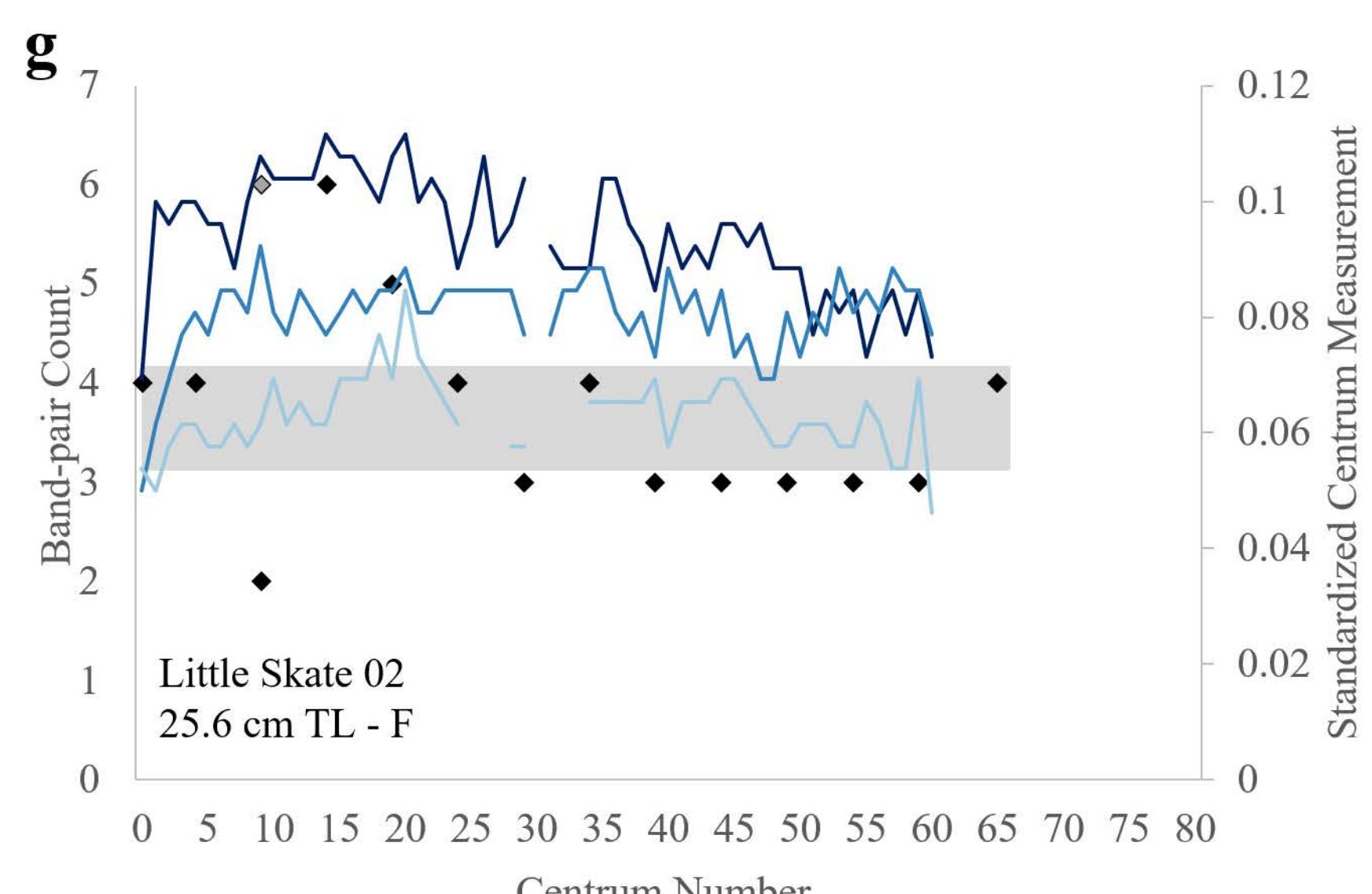
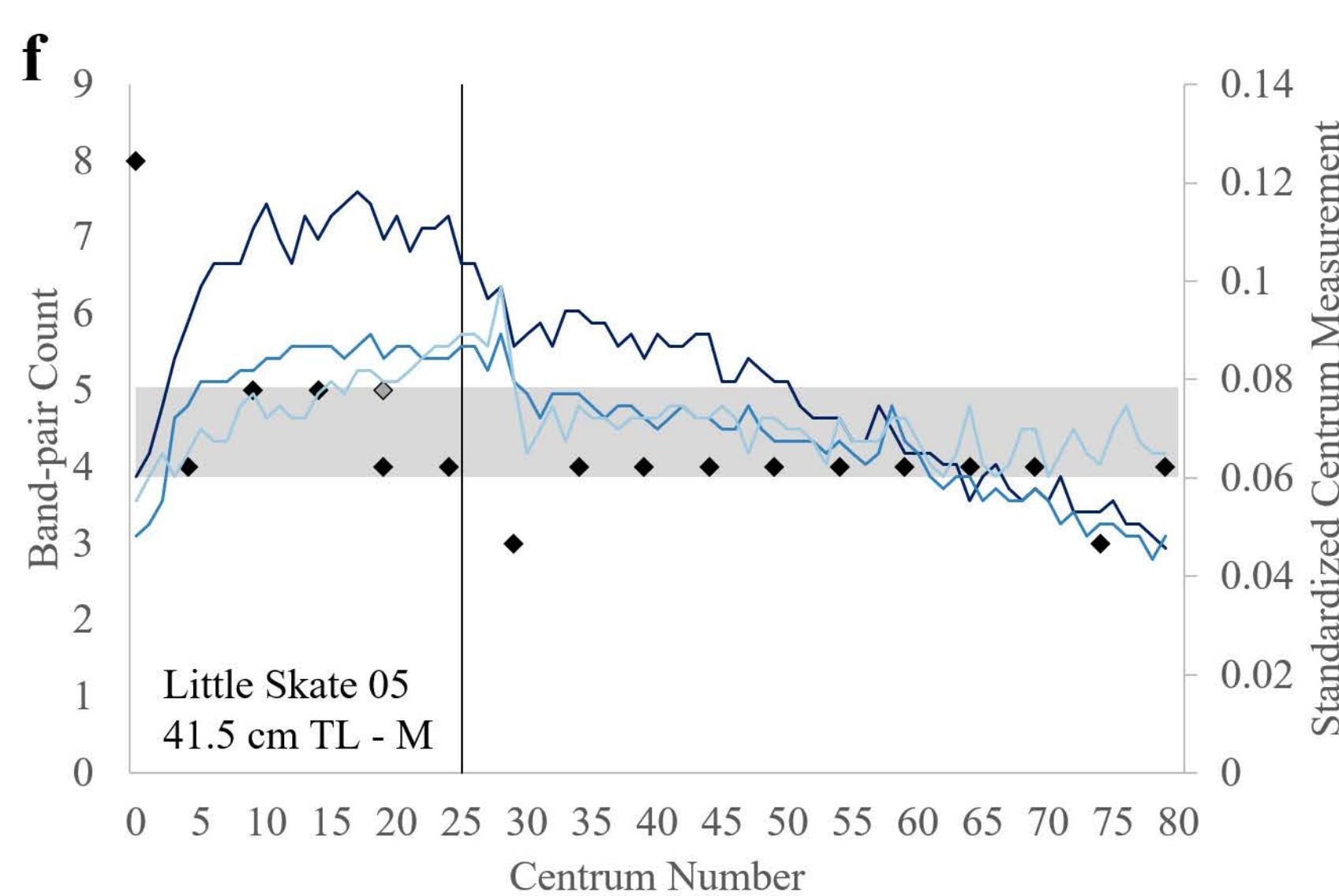
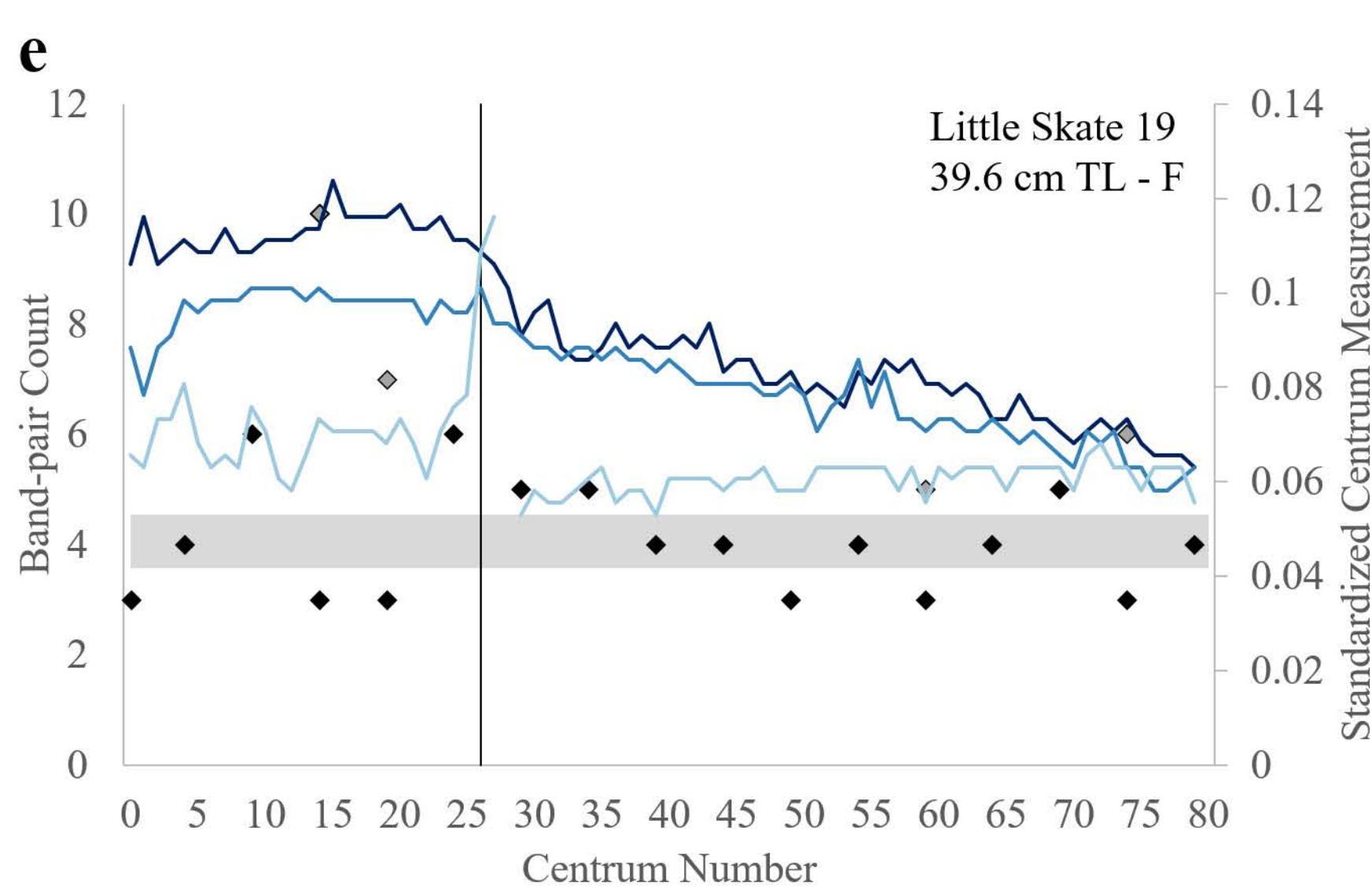
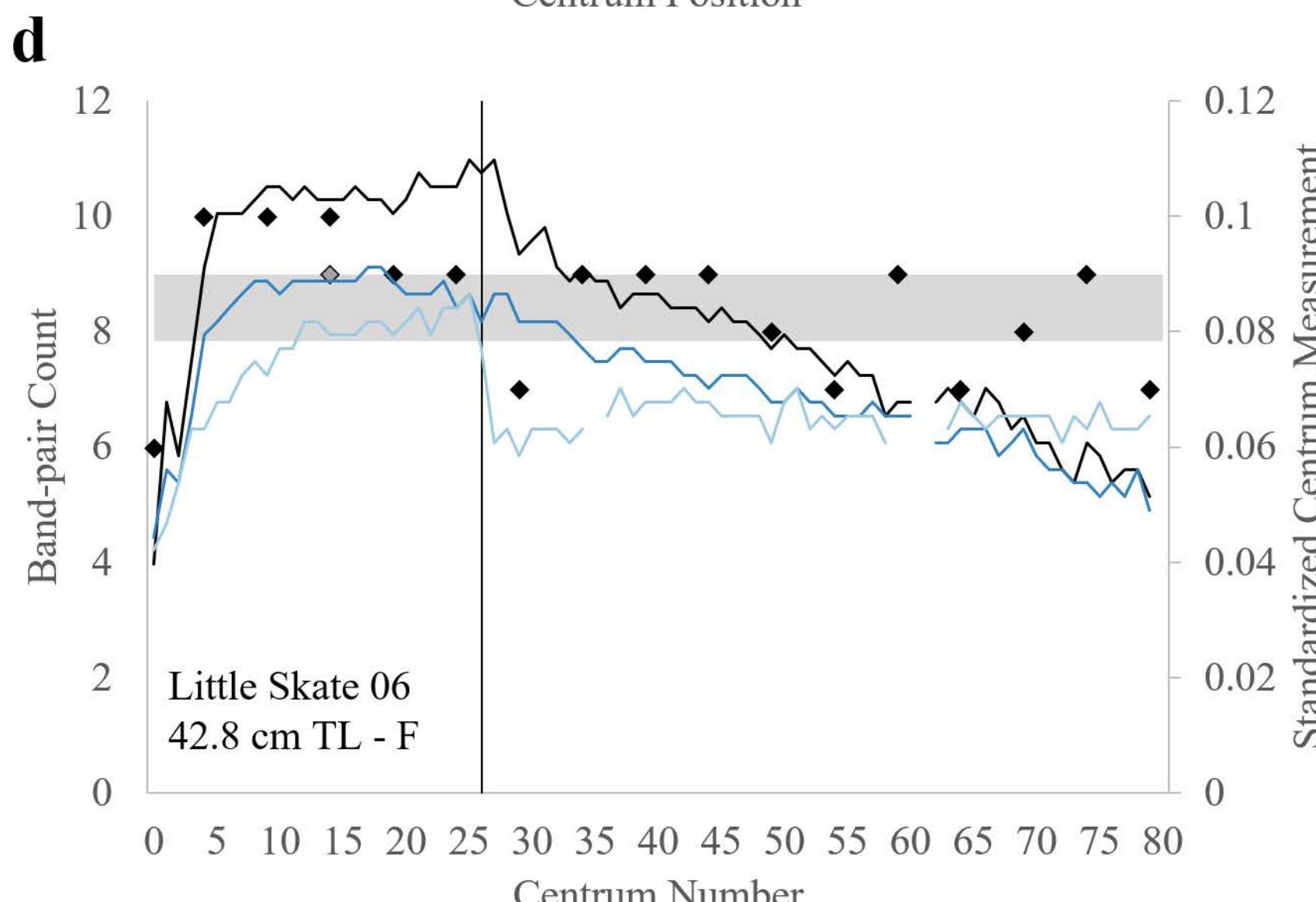
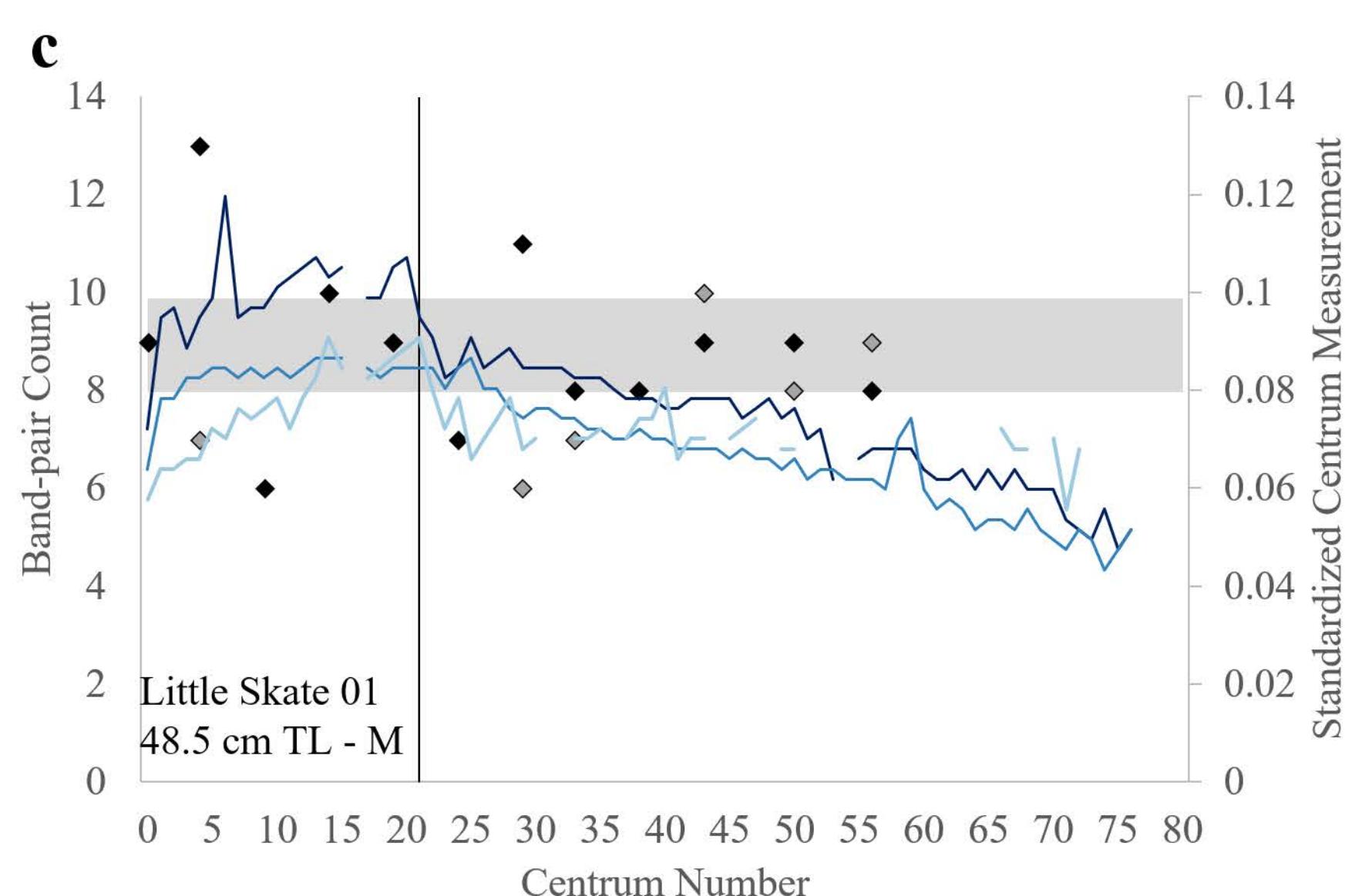
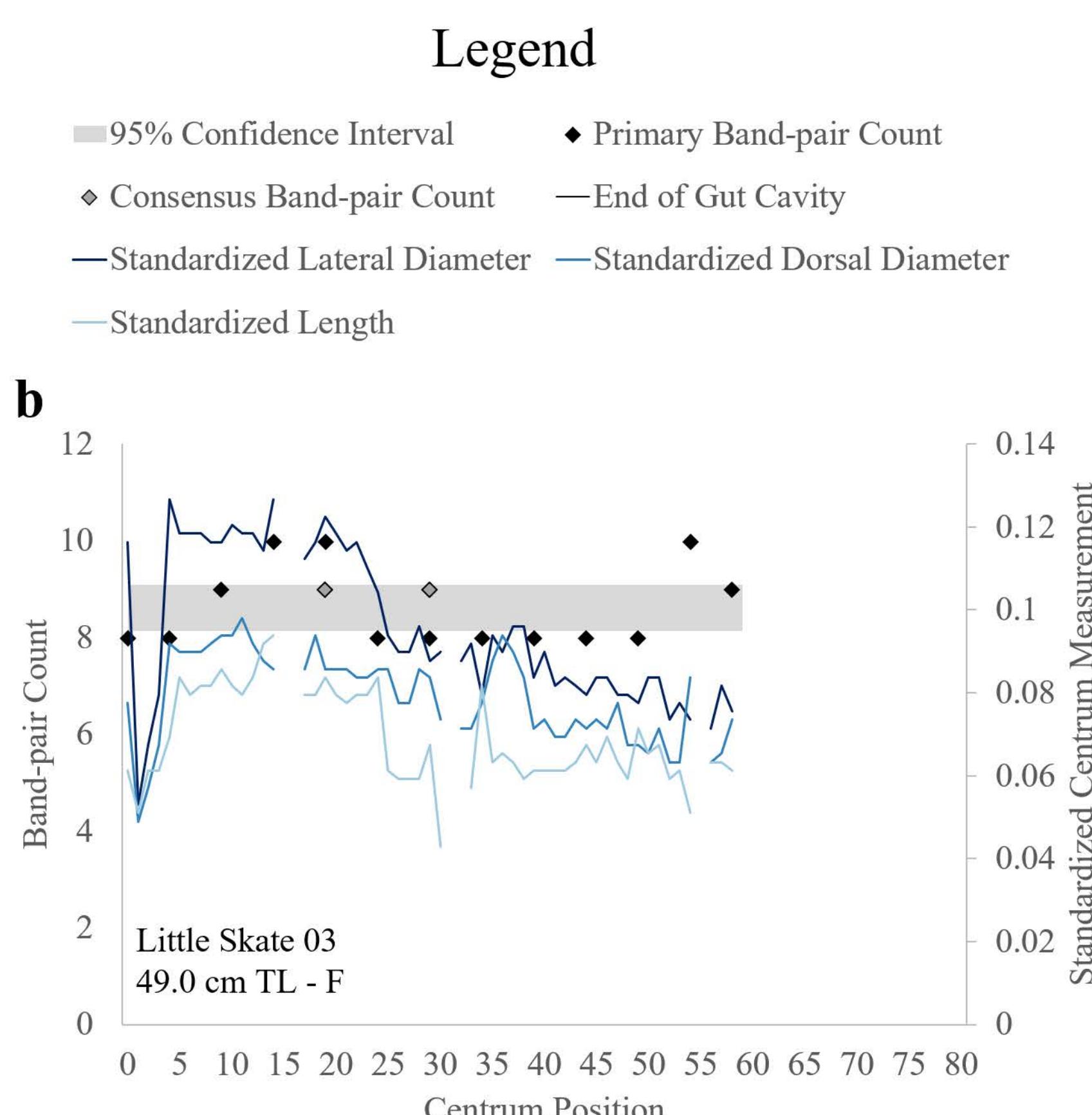
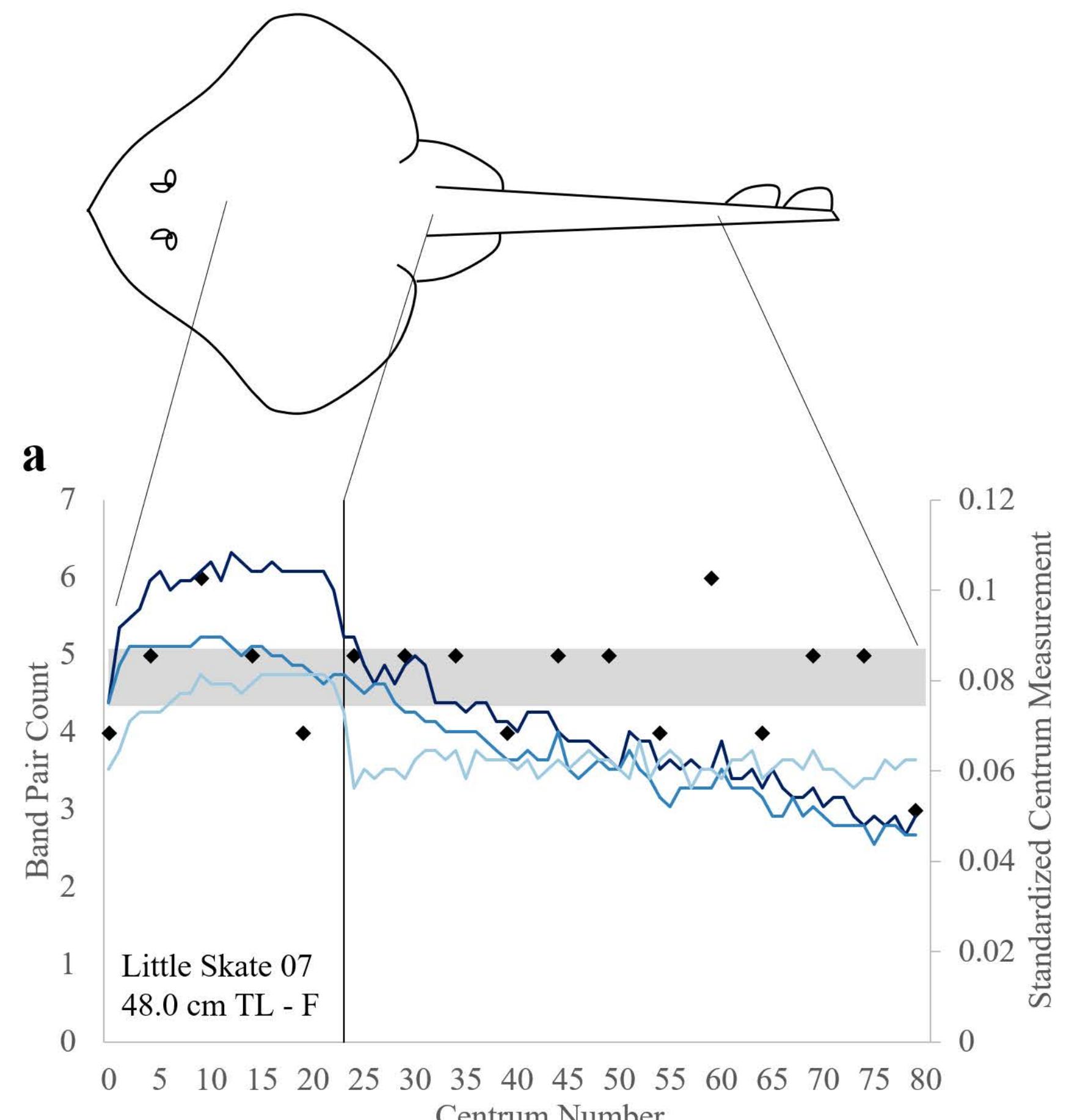
^aBand pair counts of zero are excluded from bias and precision analyses

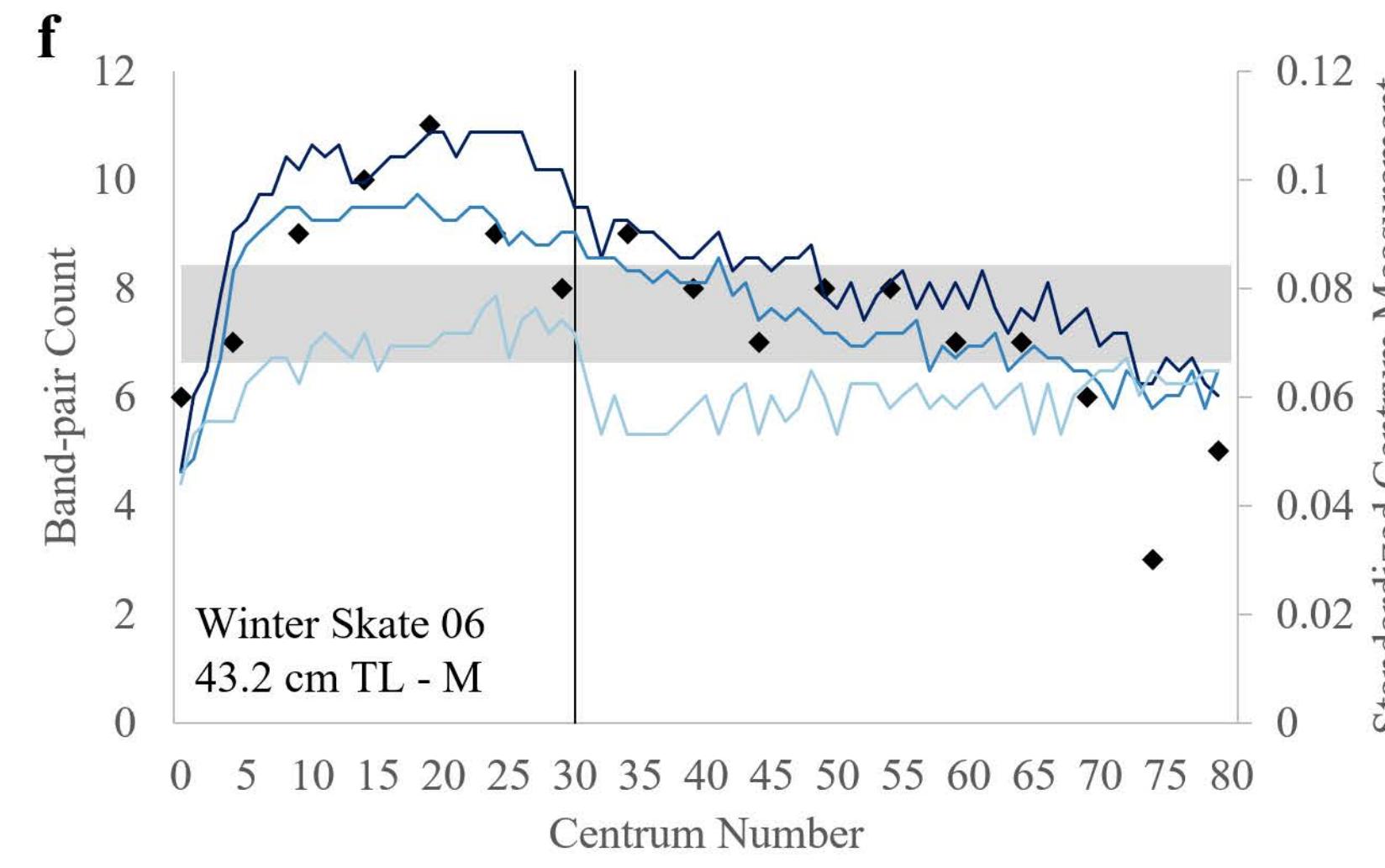
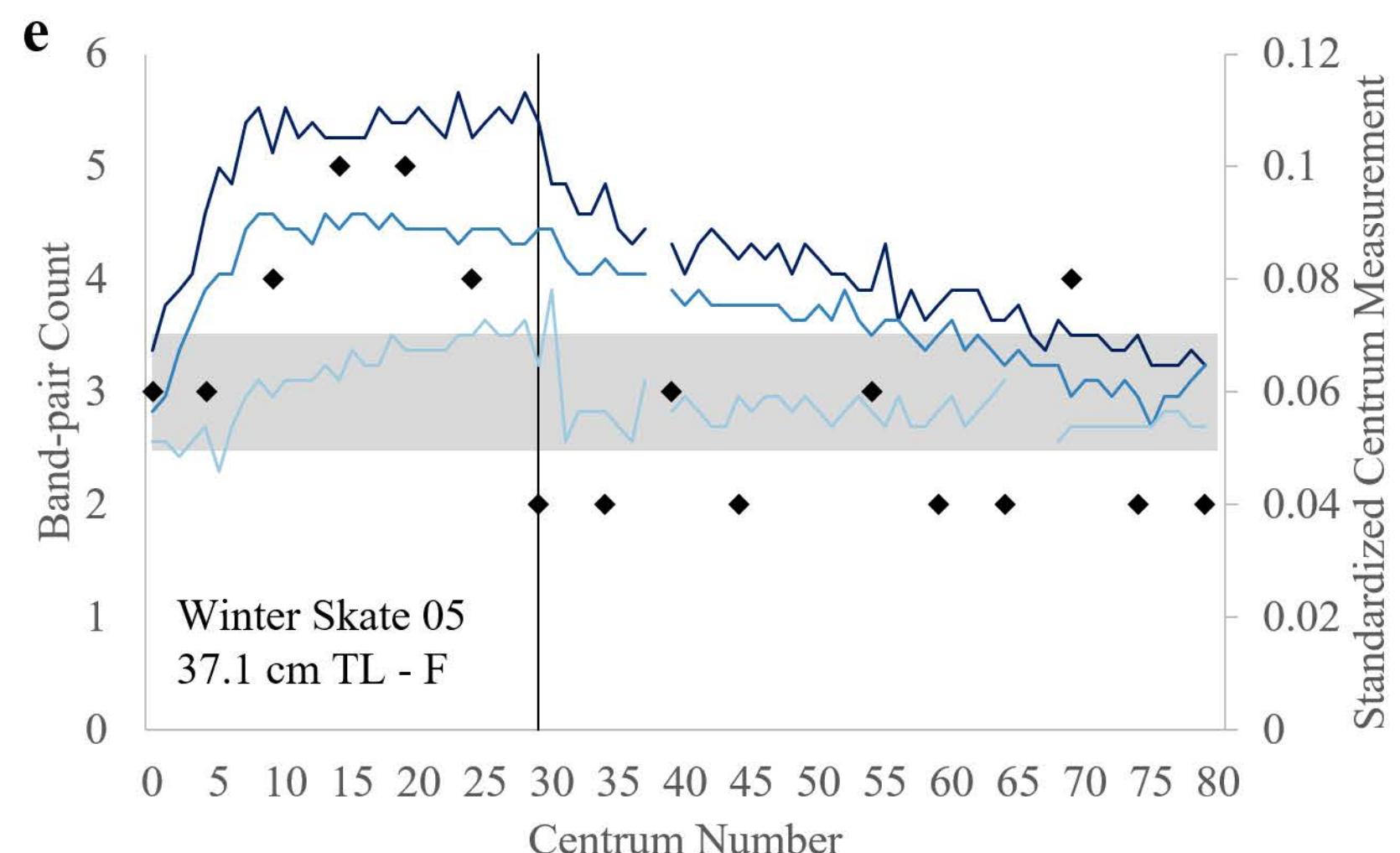
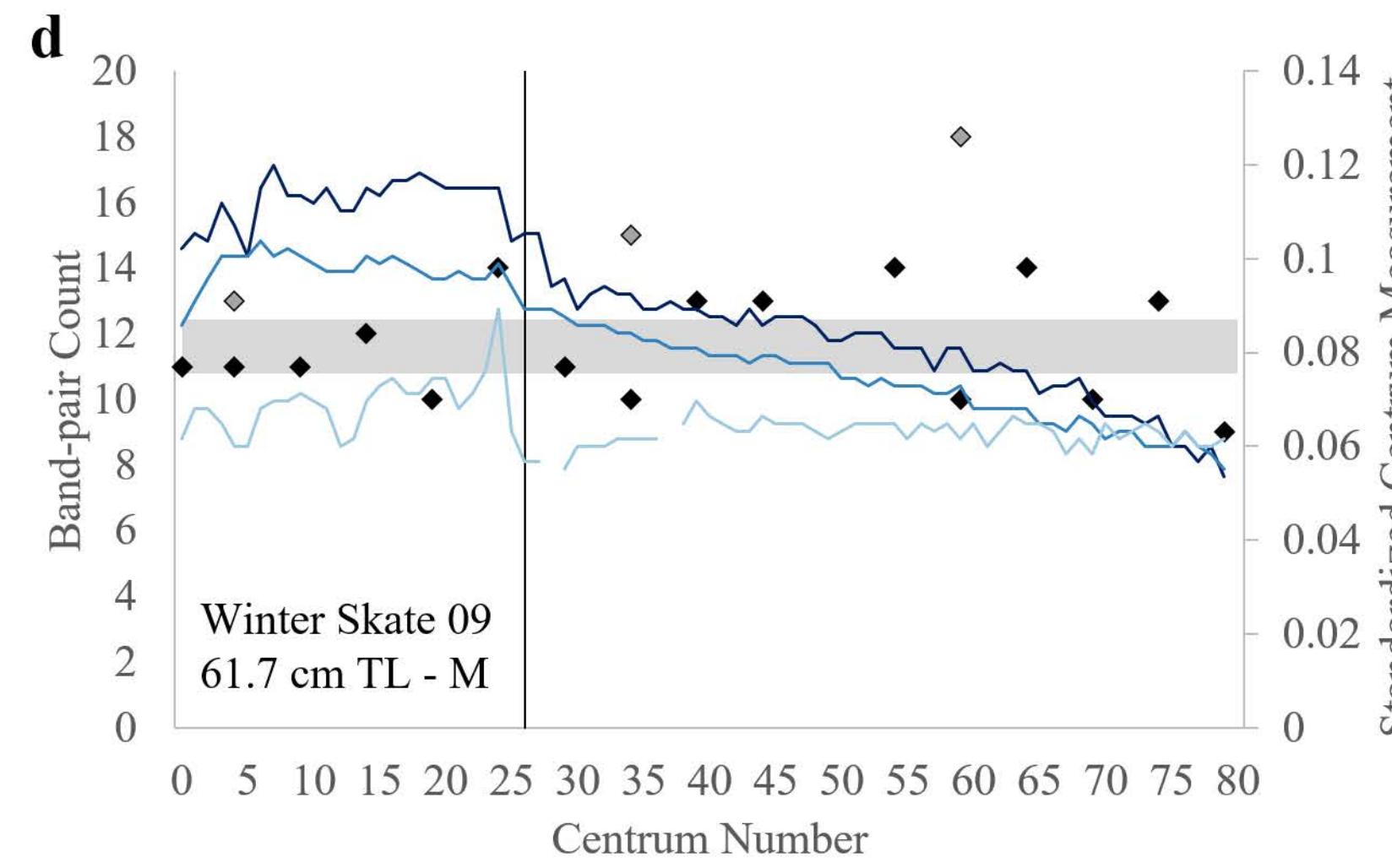
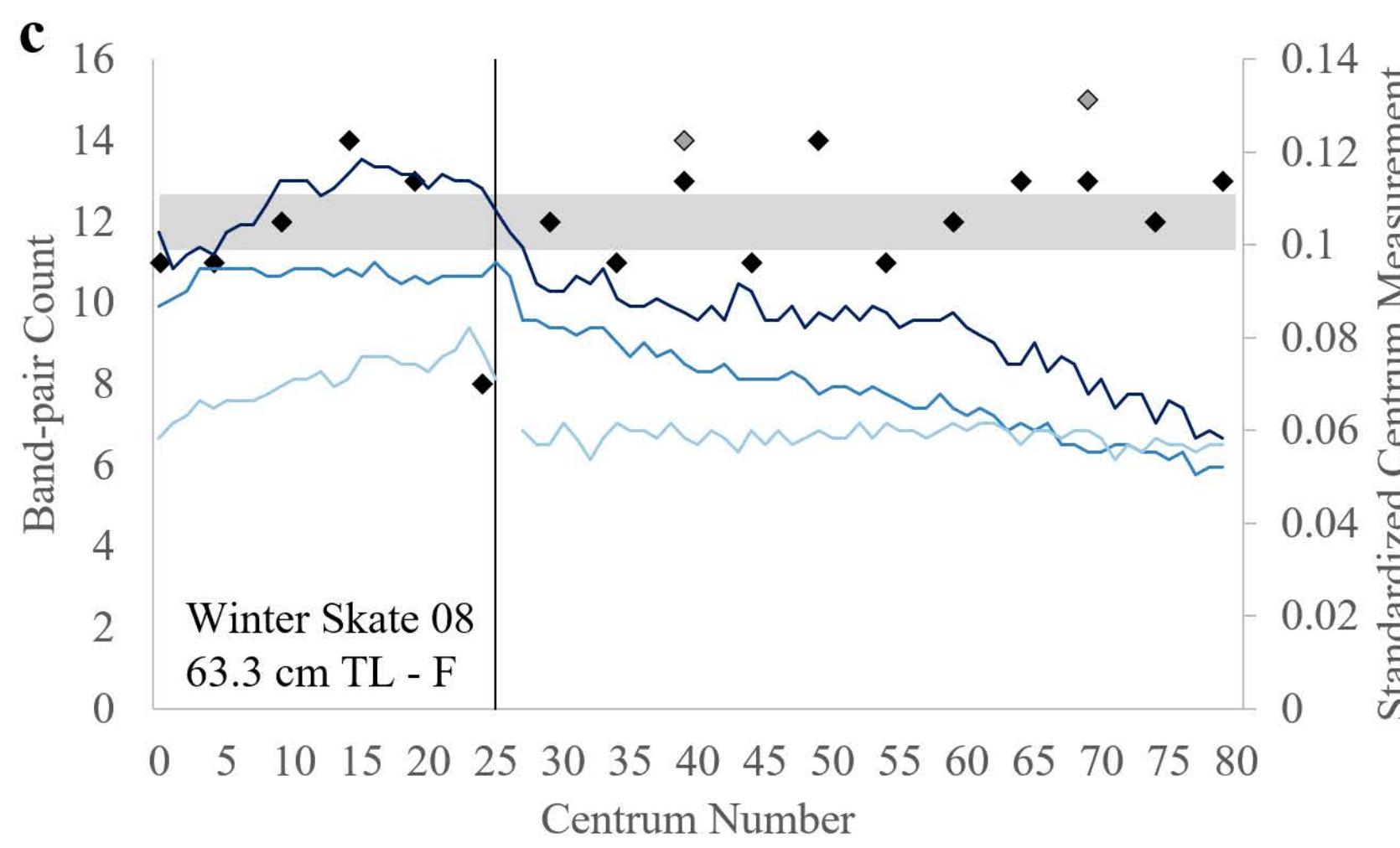
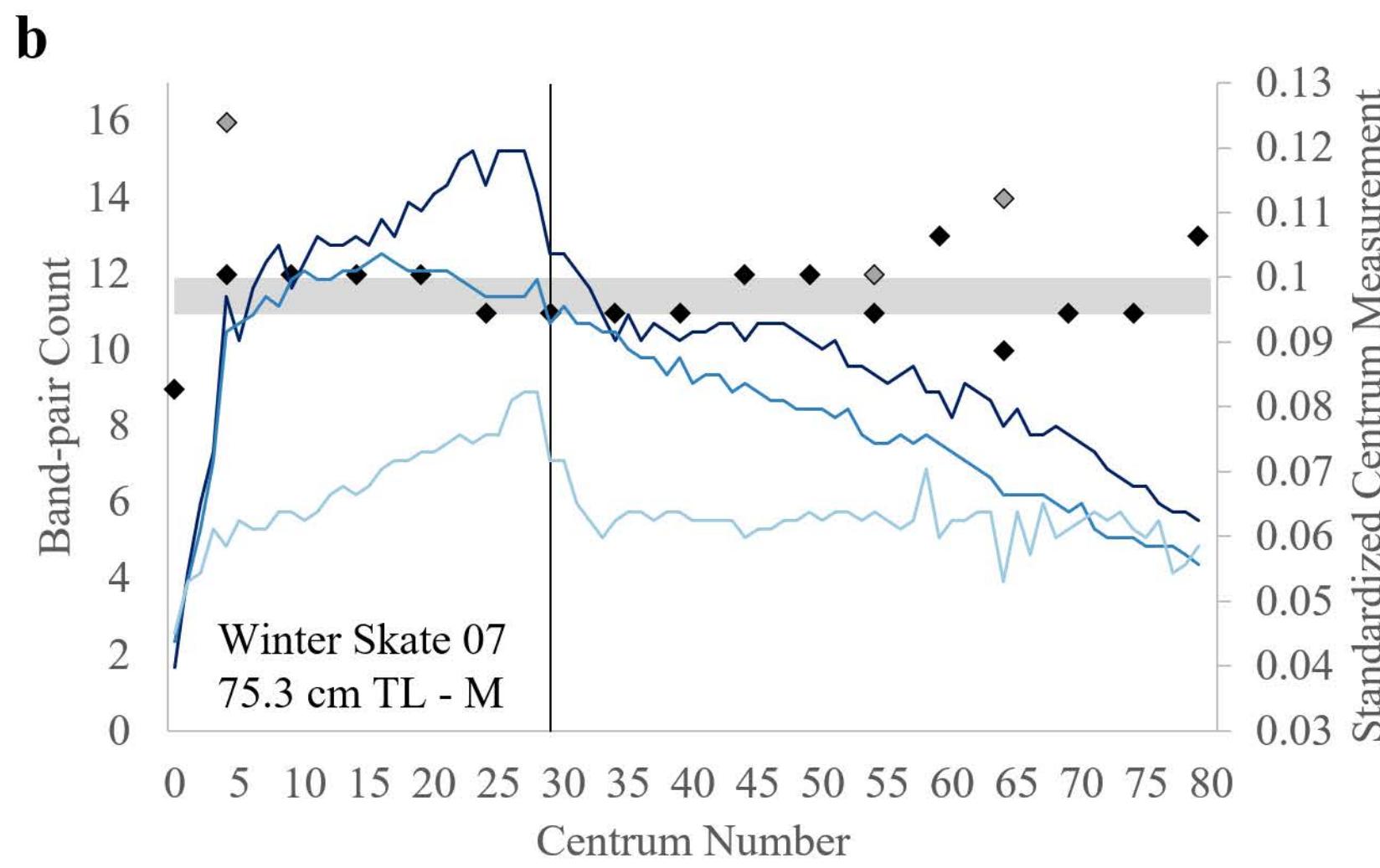
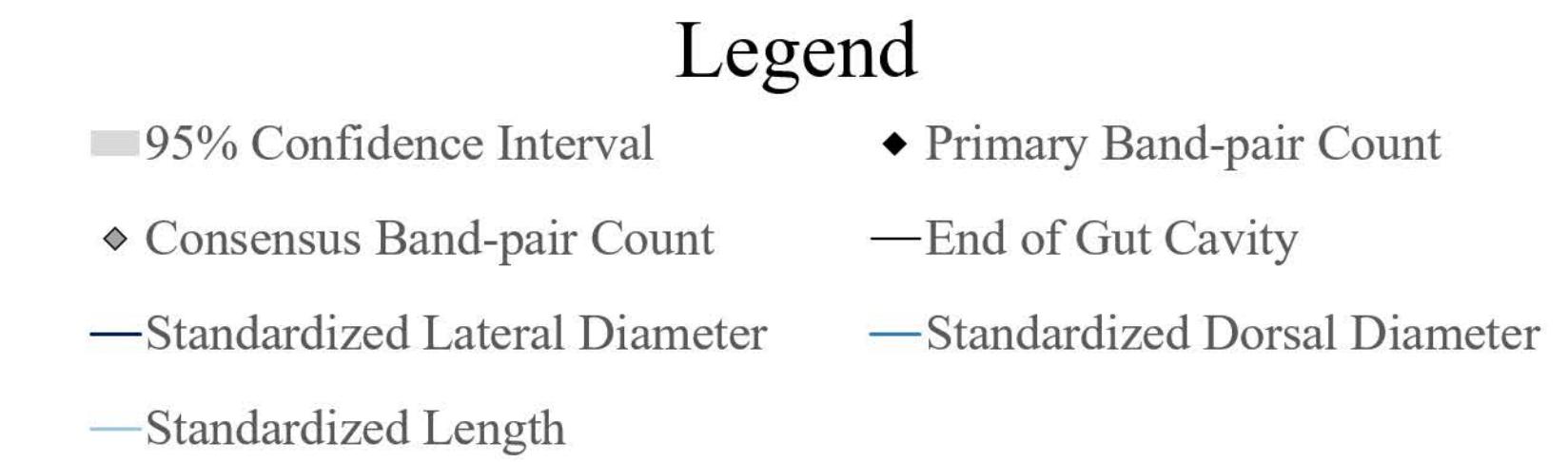
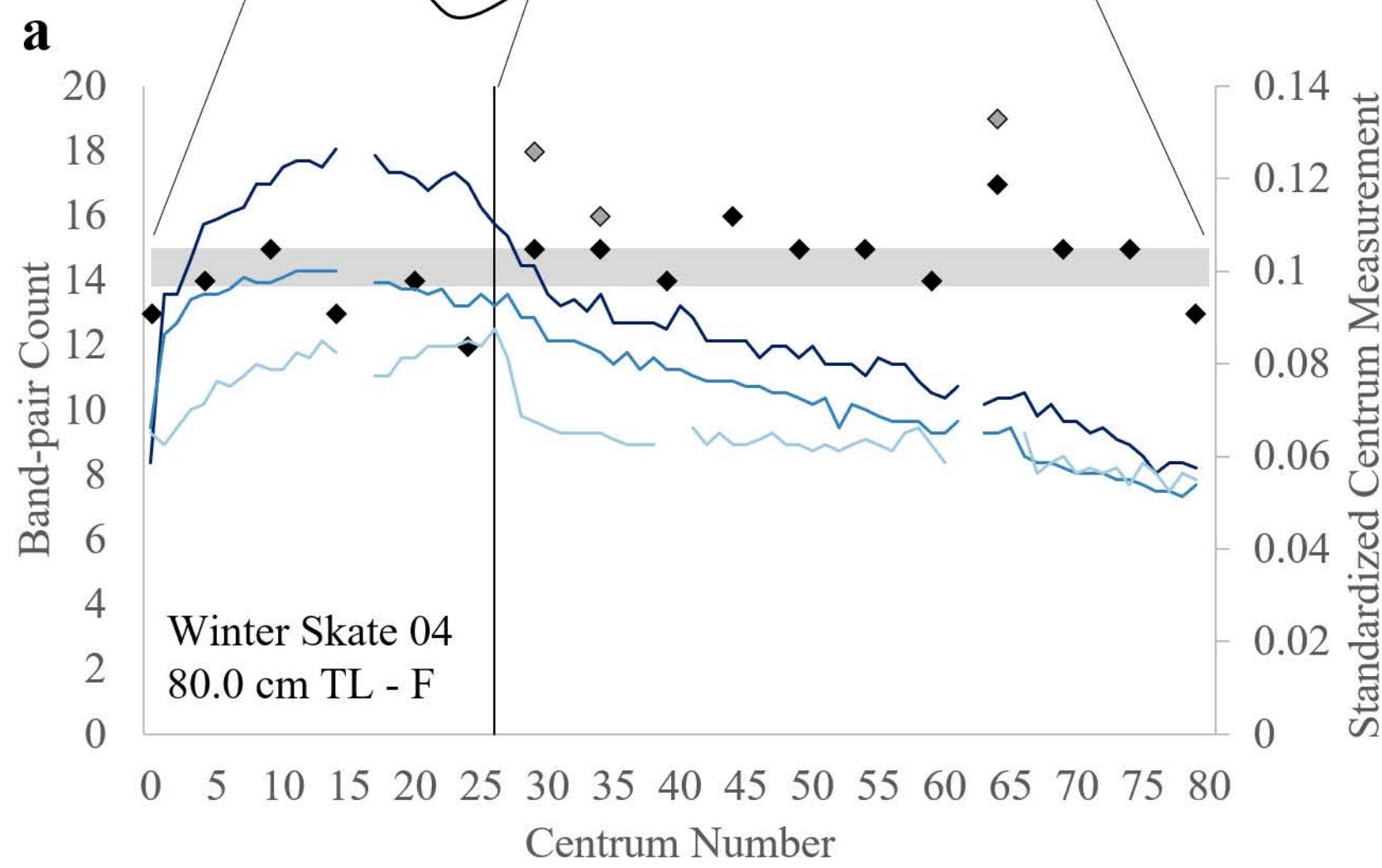
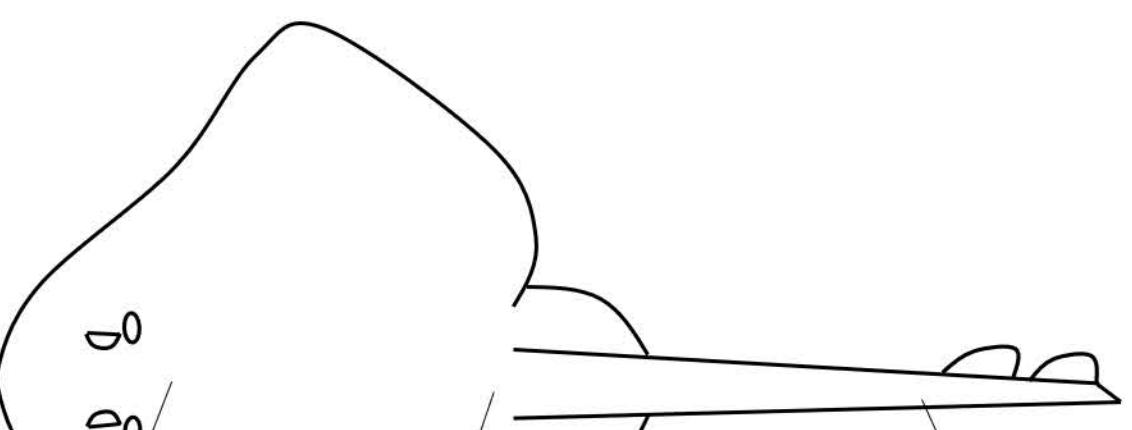
^bNumber of centra counted by both readers together for consensus.

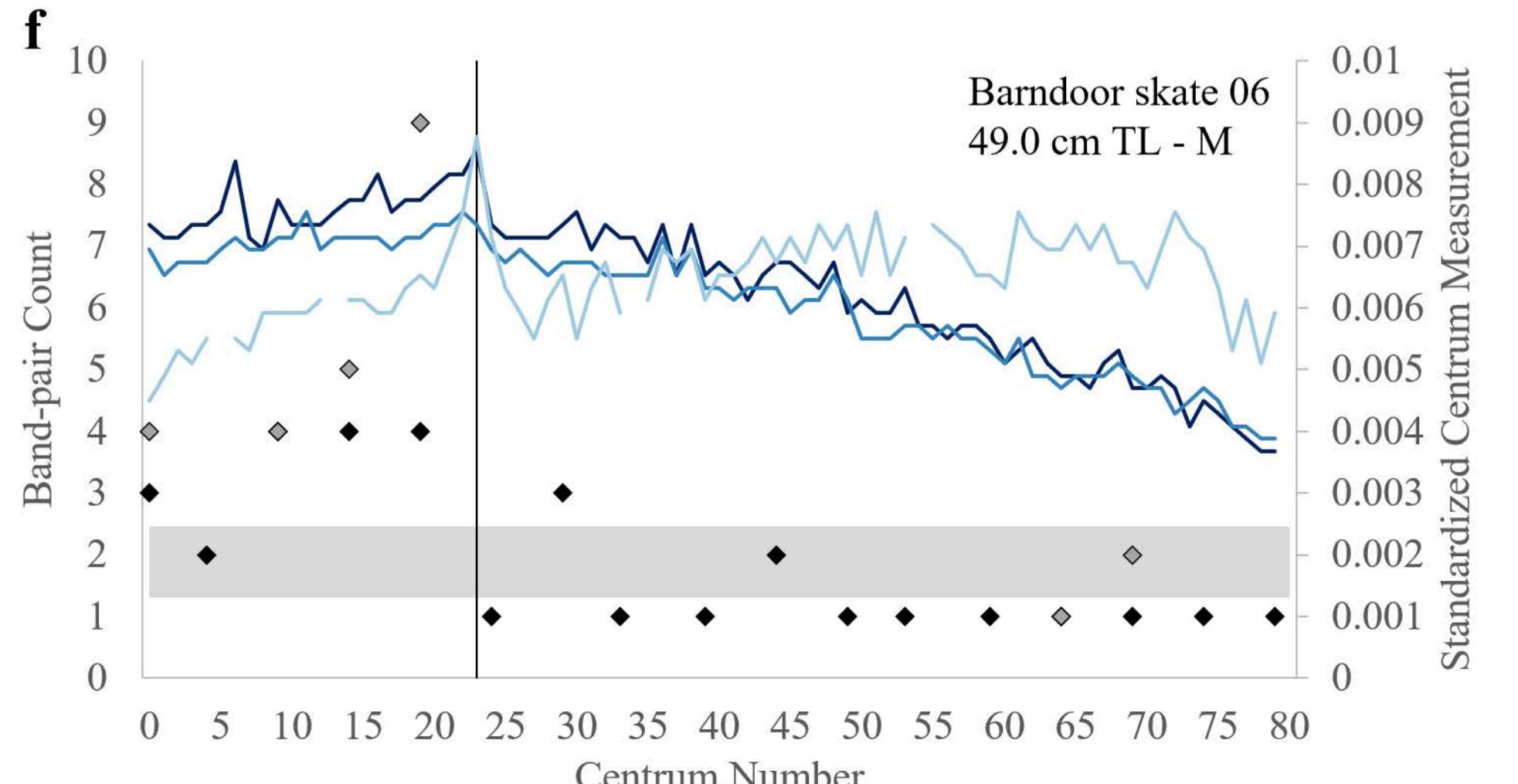
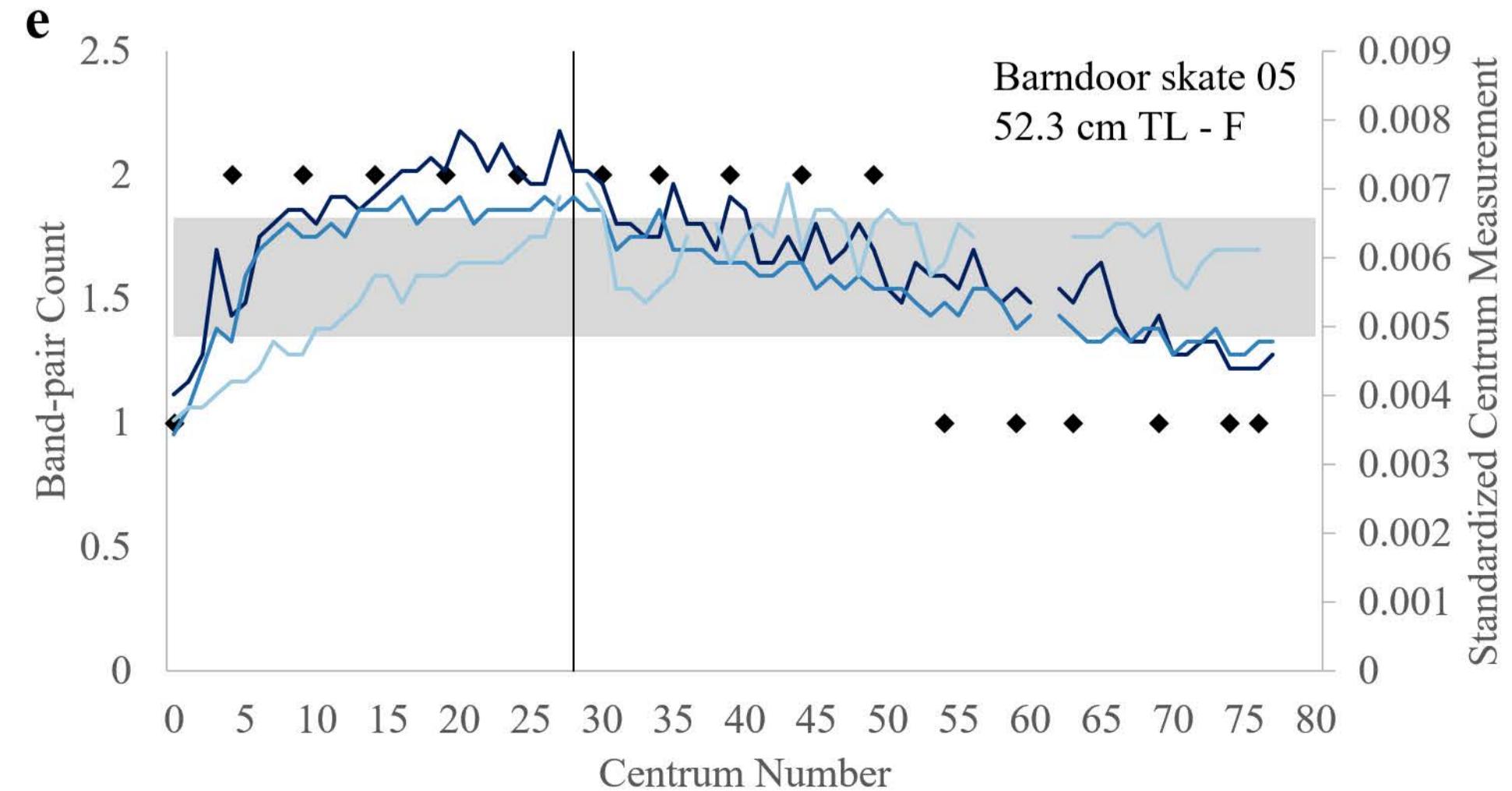
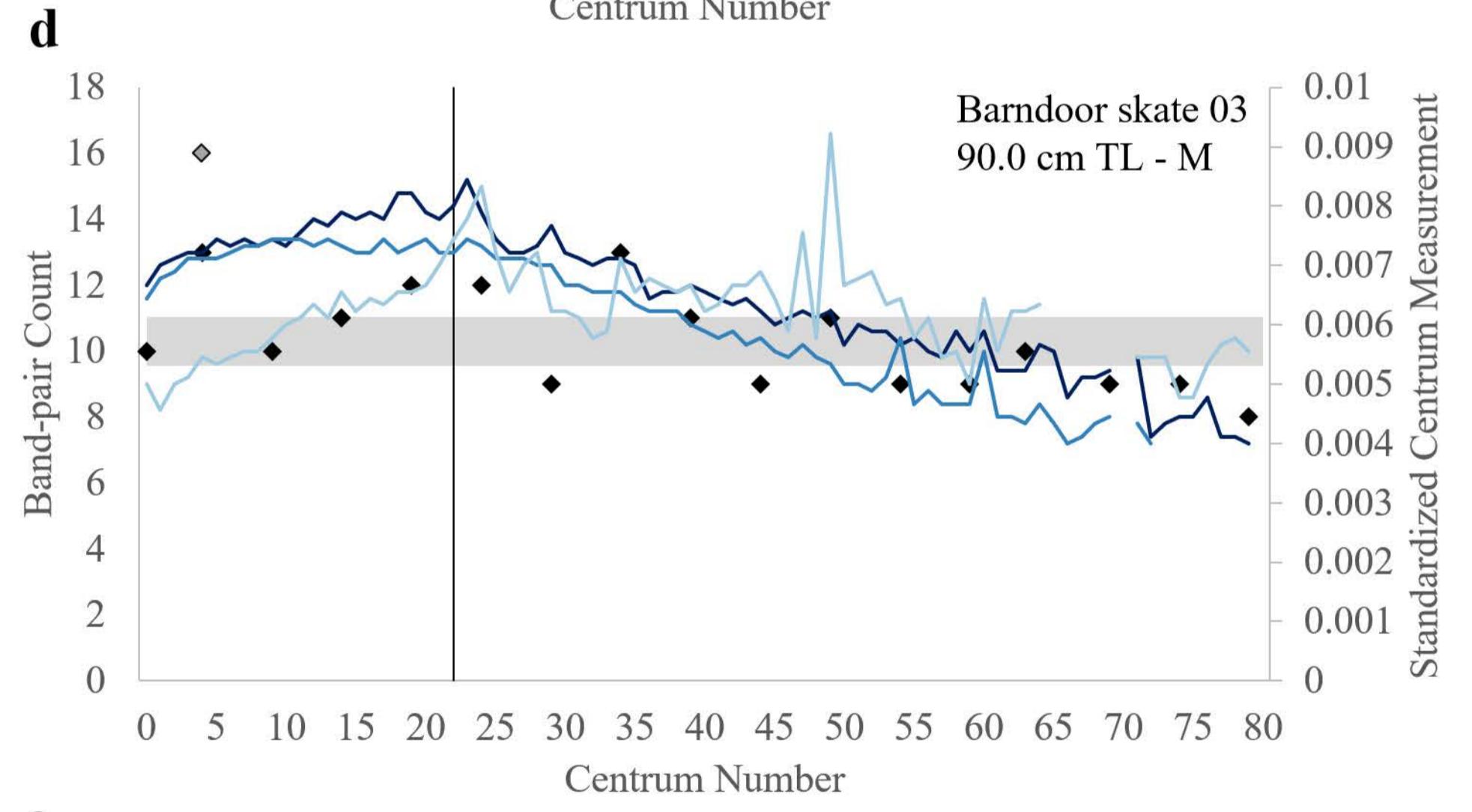
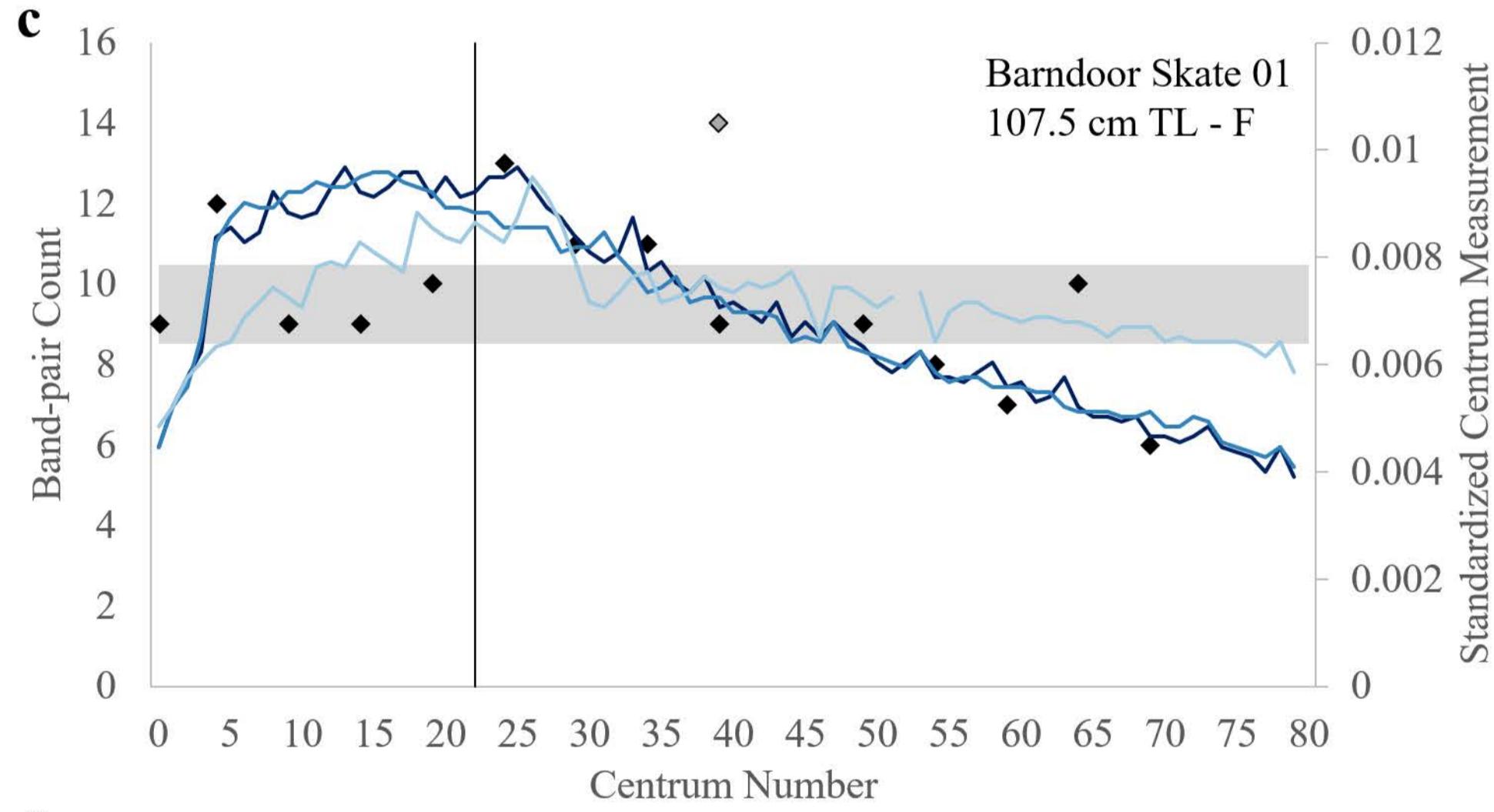
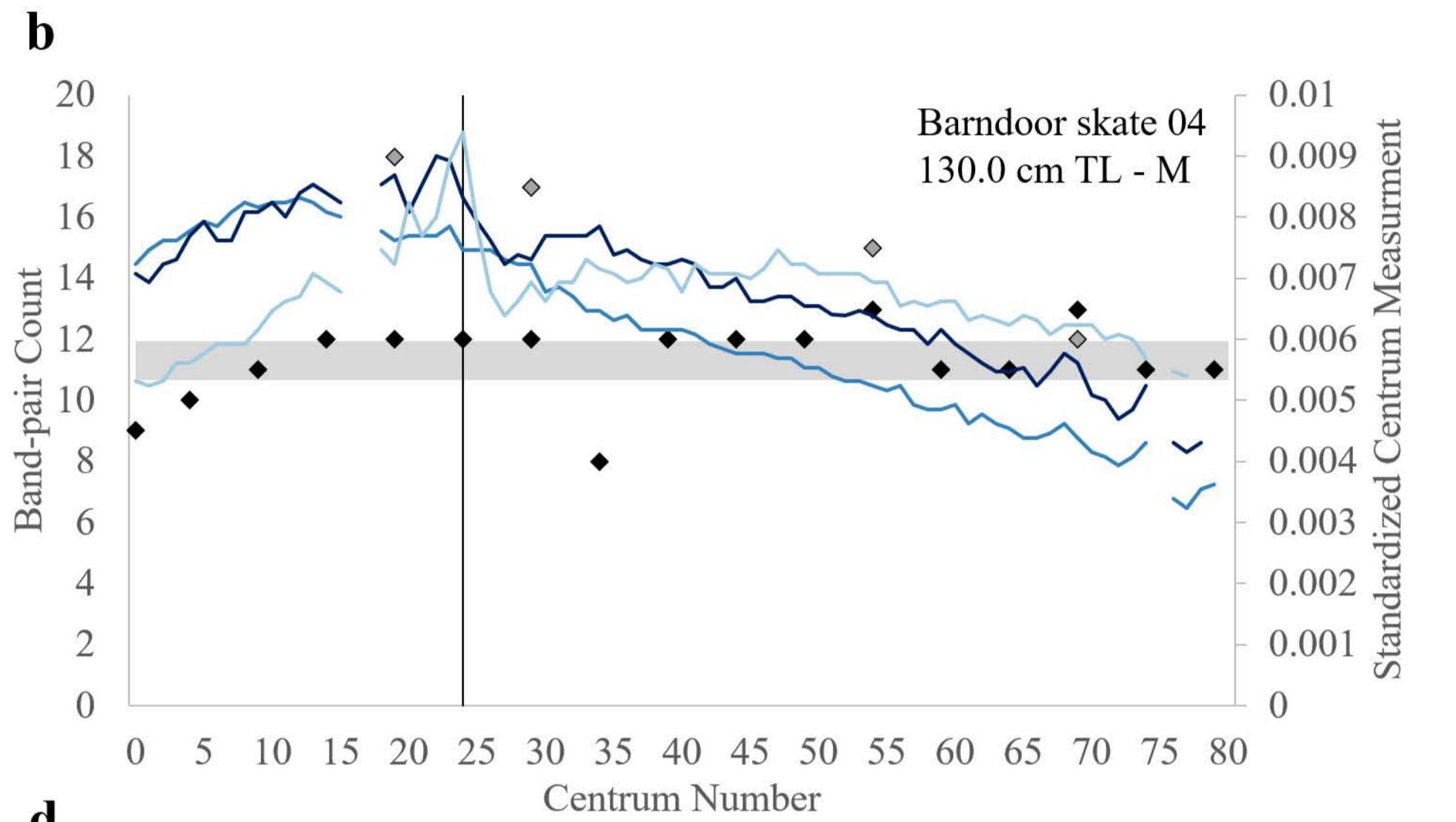
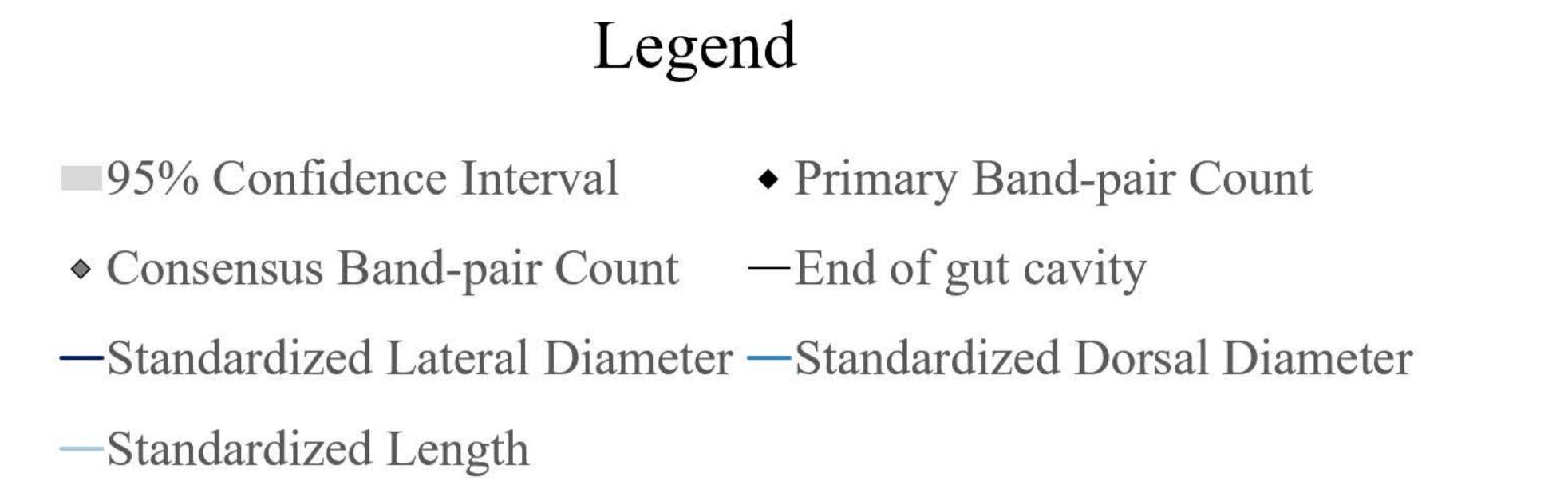
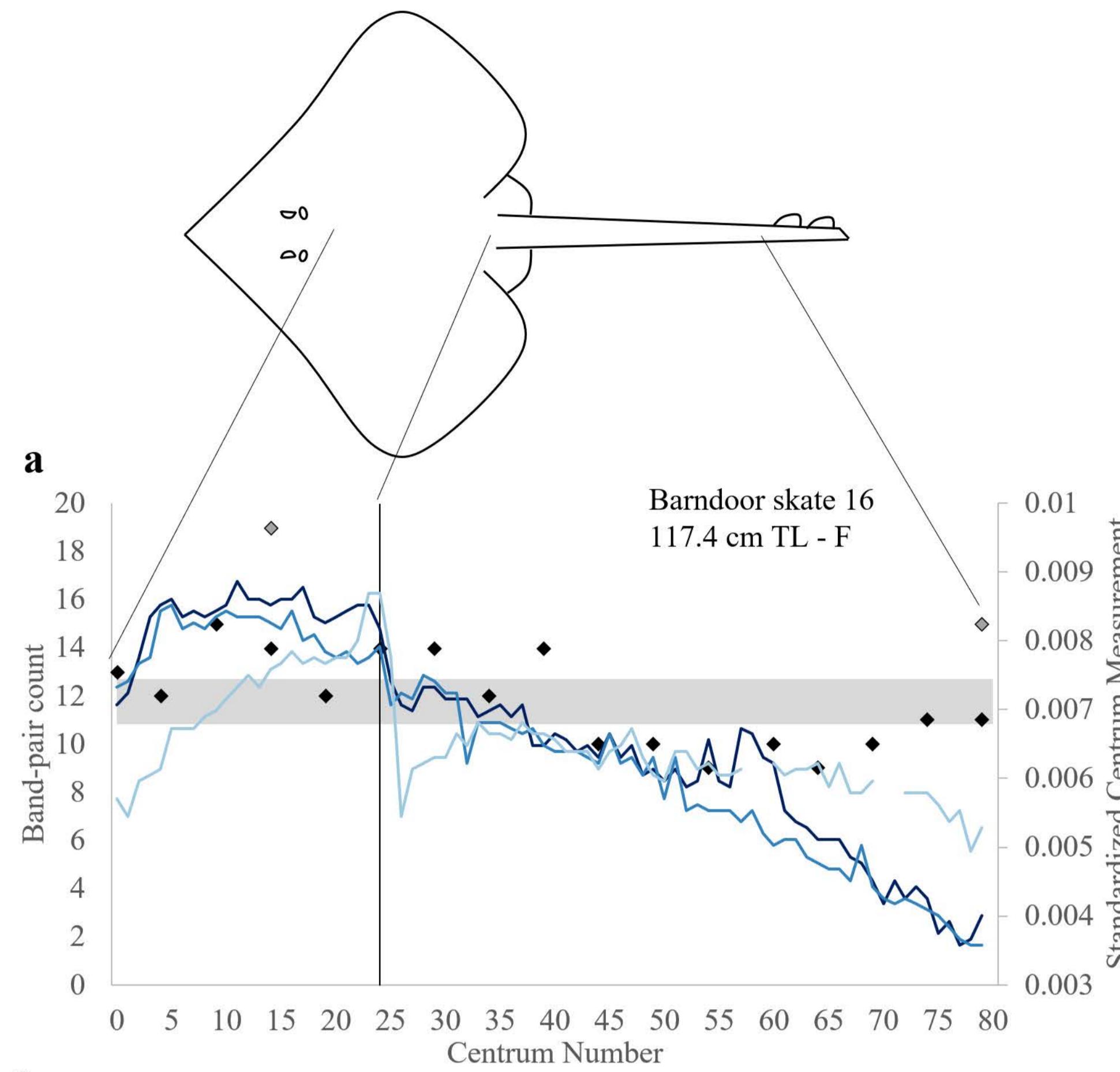
Table 3. Linear mixed-effects model comparing band pair counts with the three centrum measurements with individual included the model as a random effect. Asterisk indicates significant correlation.

Species	Measurement	Estimate	S.E.	df	t-value	p-value
Little Skate	Dorsal					
	Diameter	0.591	0.218	129	2.716	0.0075 *
	Lateral					
	Diameter	0.435	0.140	129	3.101	0.0024 *
Winter Skate	Length	0.624	0.300	127	2.078	0.0398 *
	Dorsal					
	Diameter	0.123	0.184	93	0.667	0.5066
	Lateral					
	Diameter	0.092	0.146	93	0.628	0.5316
Barndoor Skate	Length	-0.325	0.364	92	0.894	0.3737
	Dorsal					
	Diameter	0.572	0.148	91	3.864	0.0002 *
	Lateral					
	Diameter	0.646	0.154	91	4.205	0.0001 *
Atlantic Stingray	Length	0.666	0.244	90	2.732	0.0076 *
	Dorsal					
	Diameter	0.865	0.326	88	2.649	0.0096 *
	Lateral					
	Diameter	0.961	0.210	88	4.574	0.0000 *
Round Ray	Length	0.915	0.499	87	1.833	0.0703
	Dorsal					
	Diameter	0.807	0.278	88	2.909	0.0046 *
	Lateral					
	Diameter	0.625	0.264	88	2.371	0.0199 *
	Length	0.900	0.431	86	2.085	0.0400 *



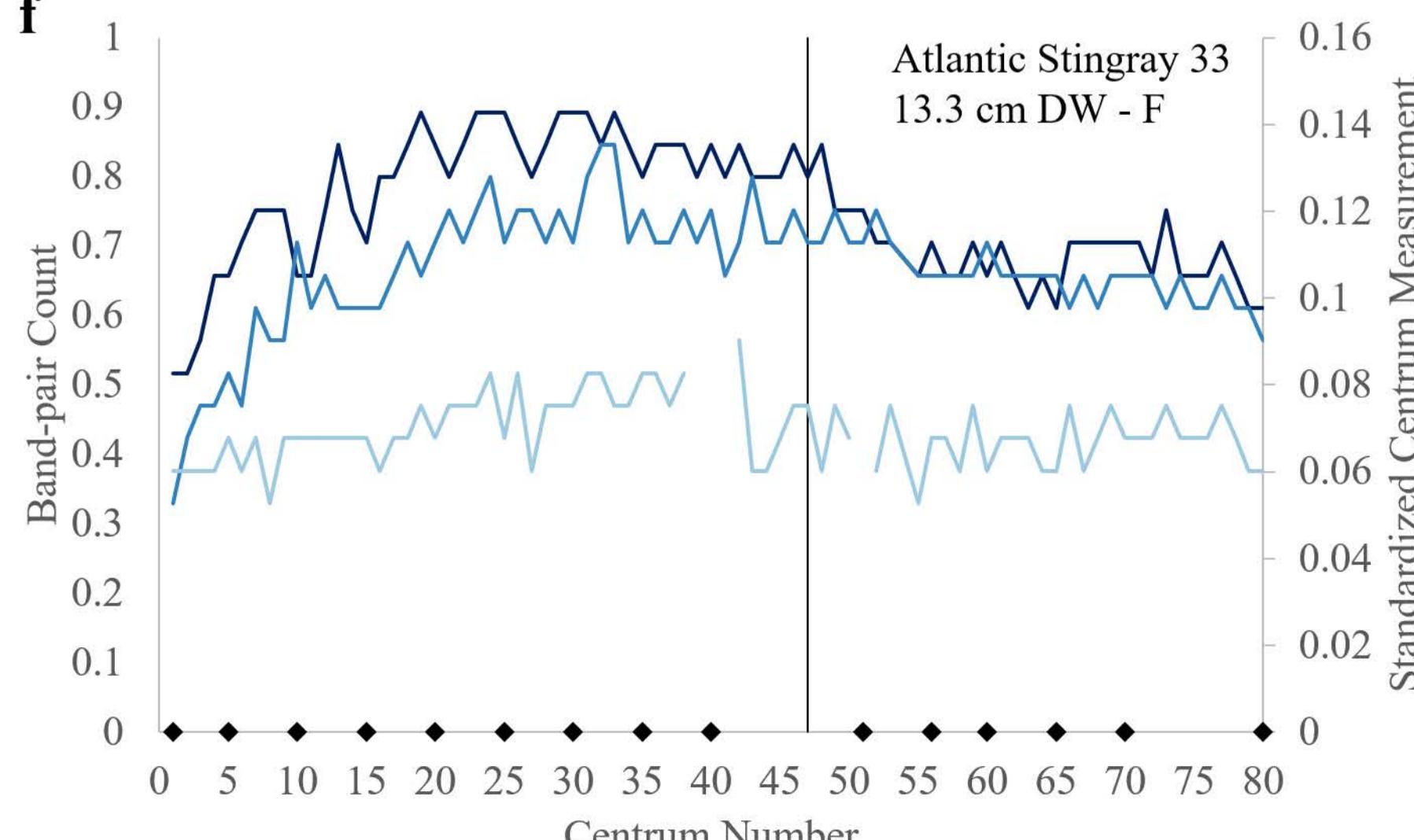
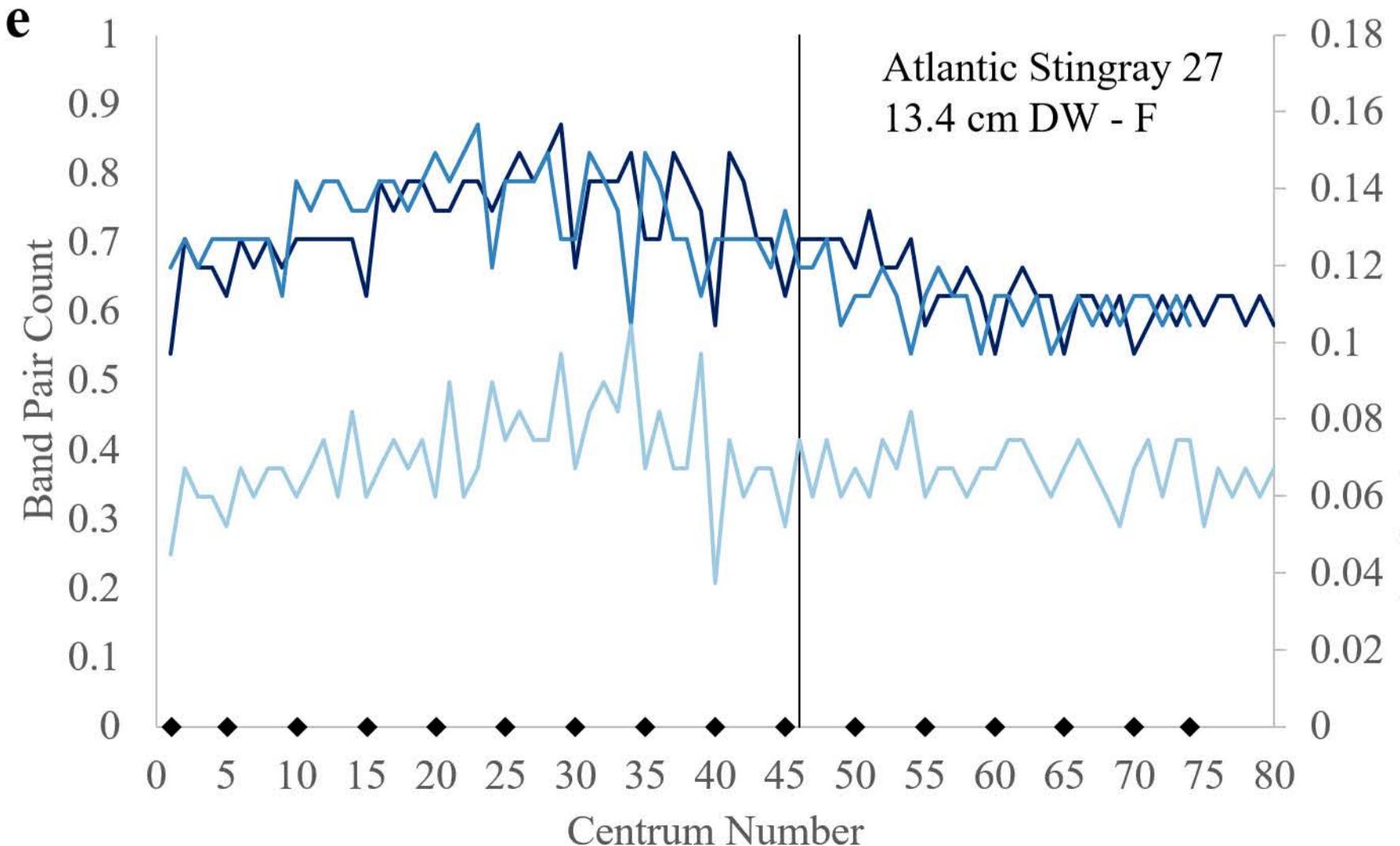
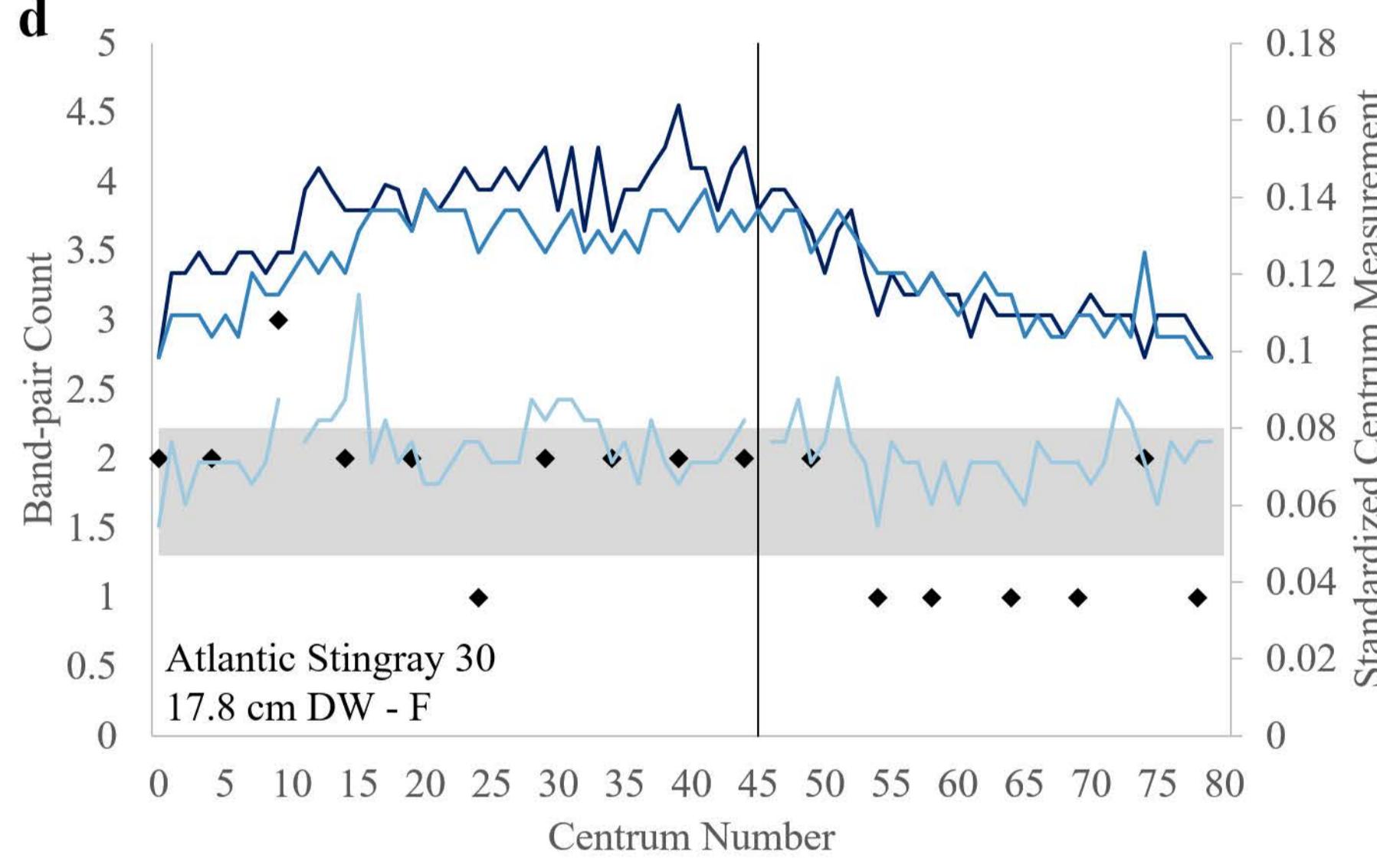
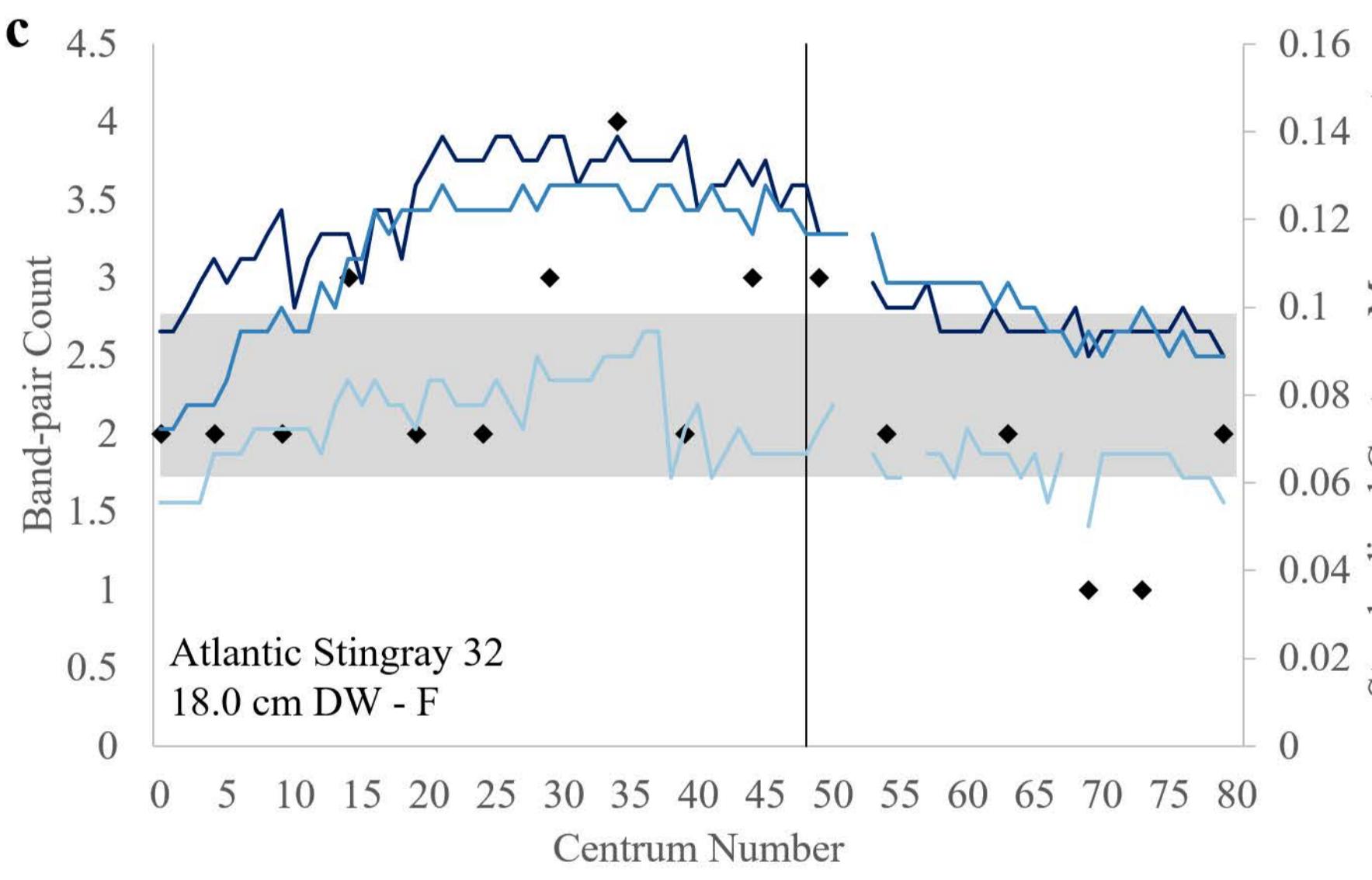
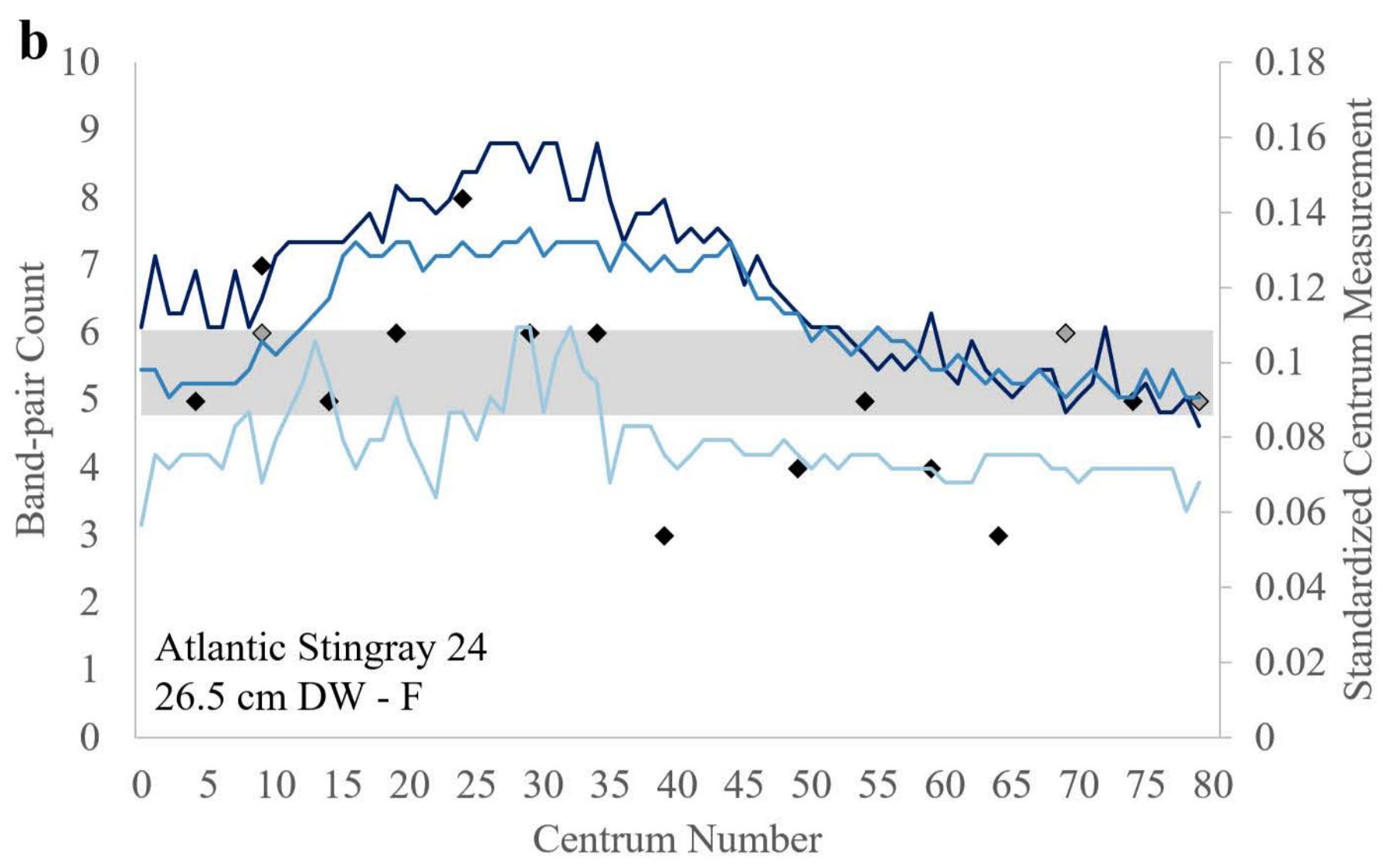
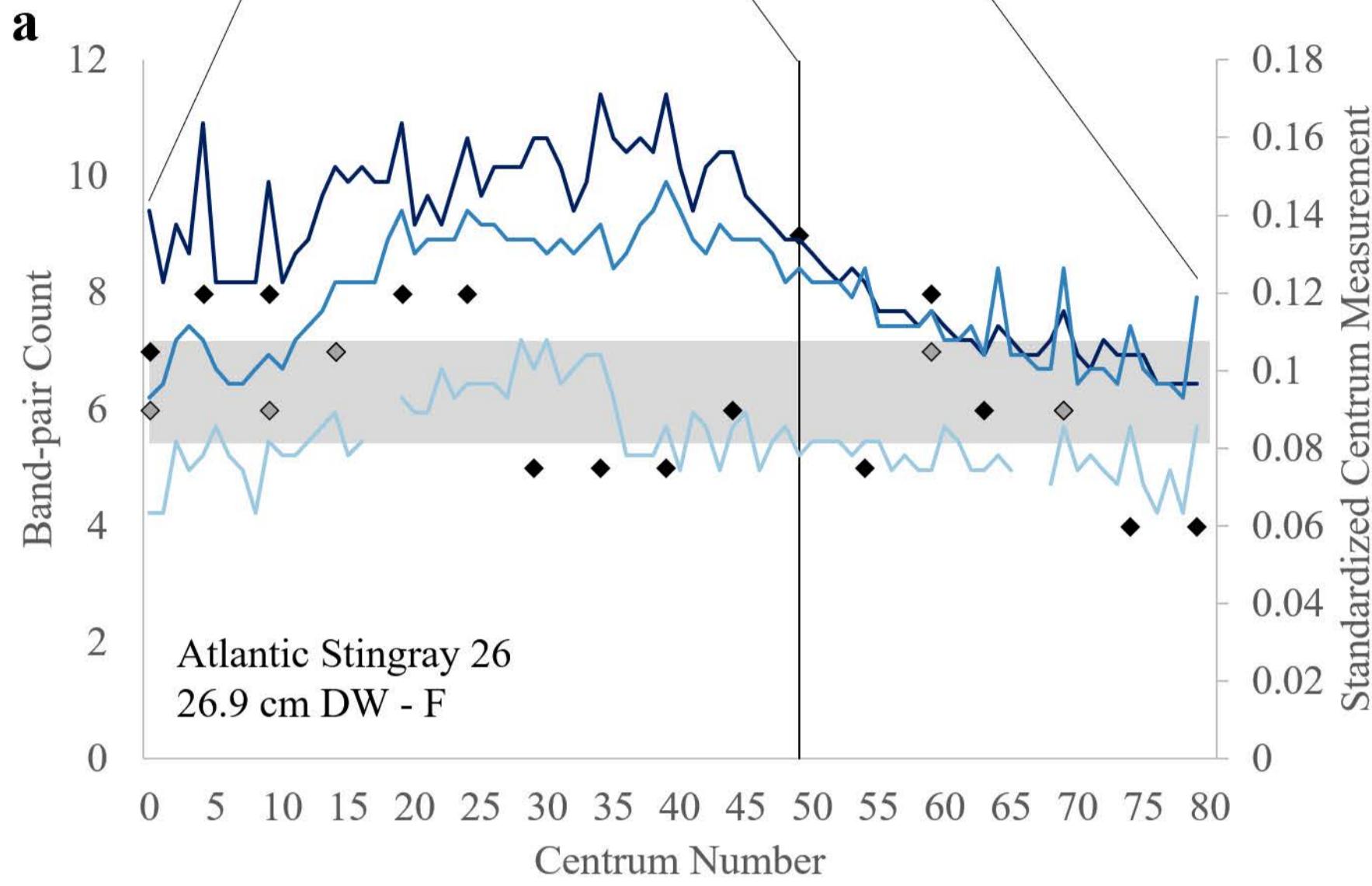


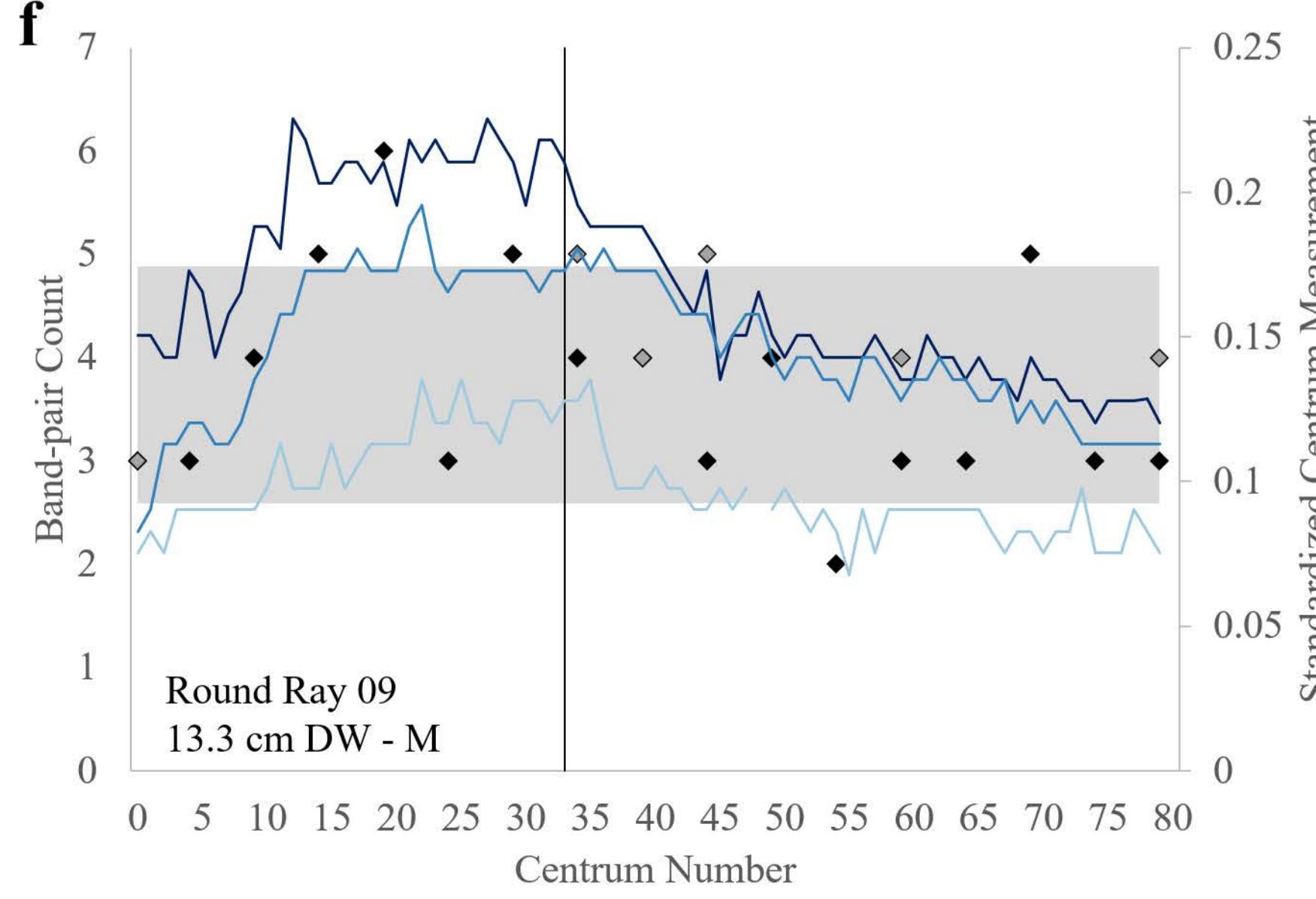
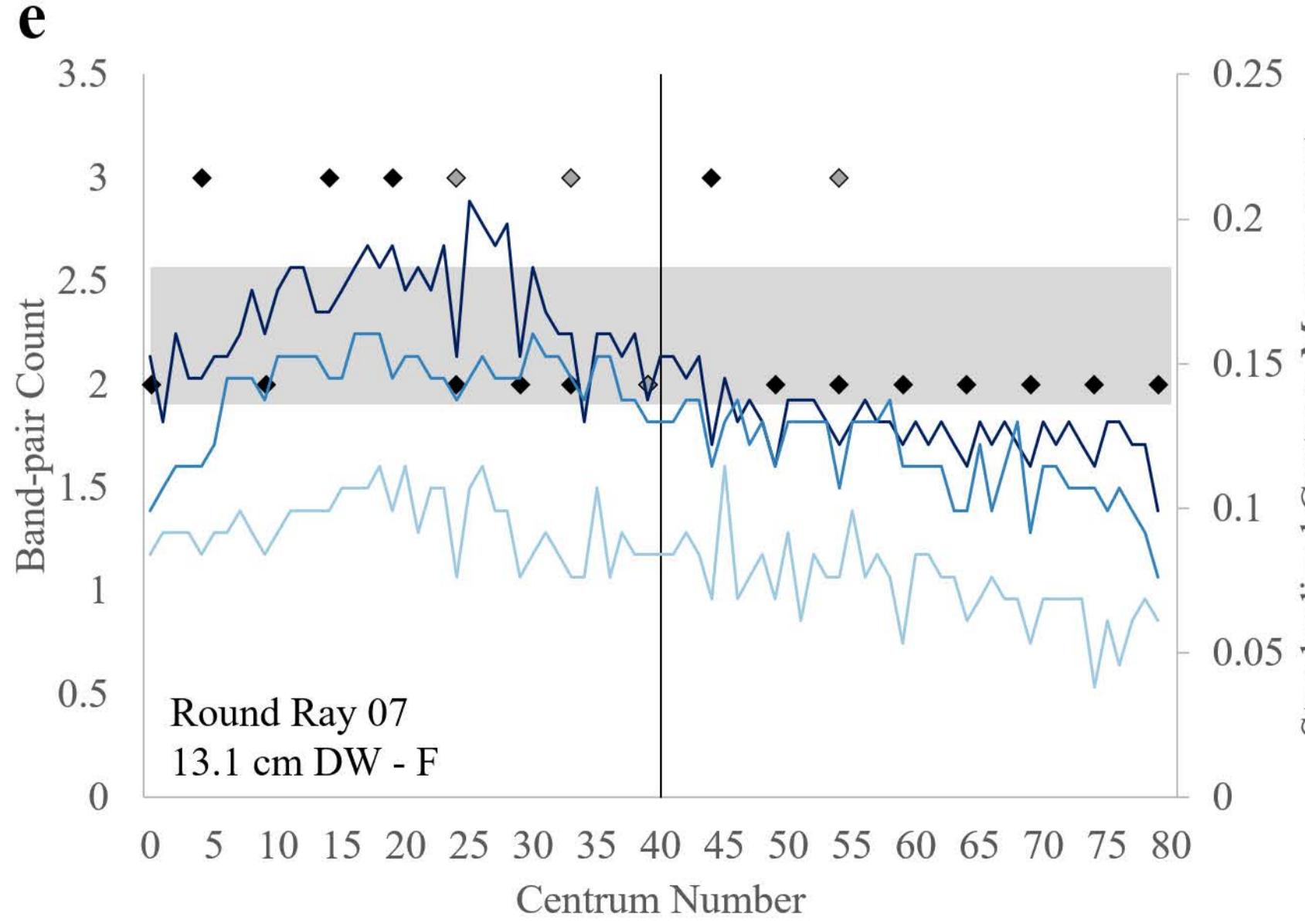
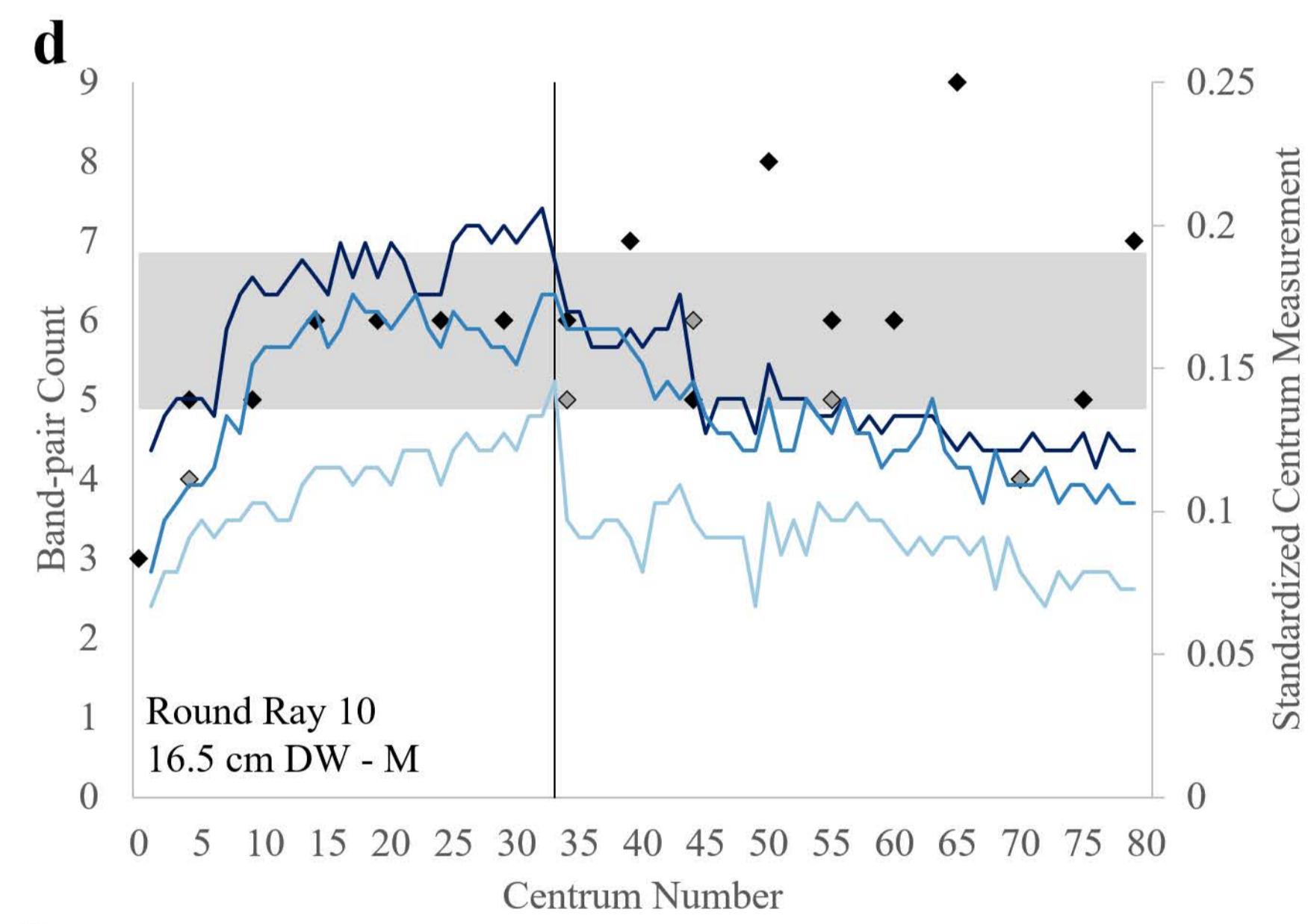
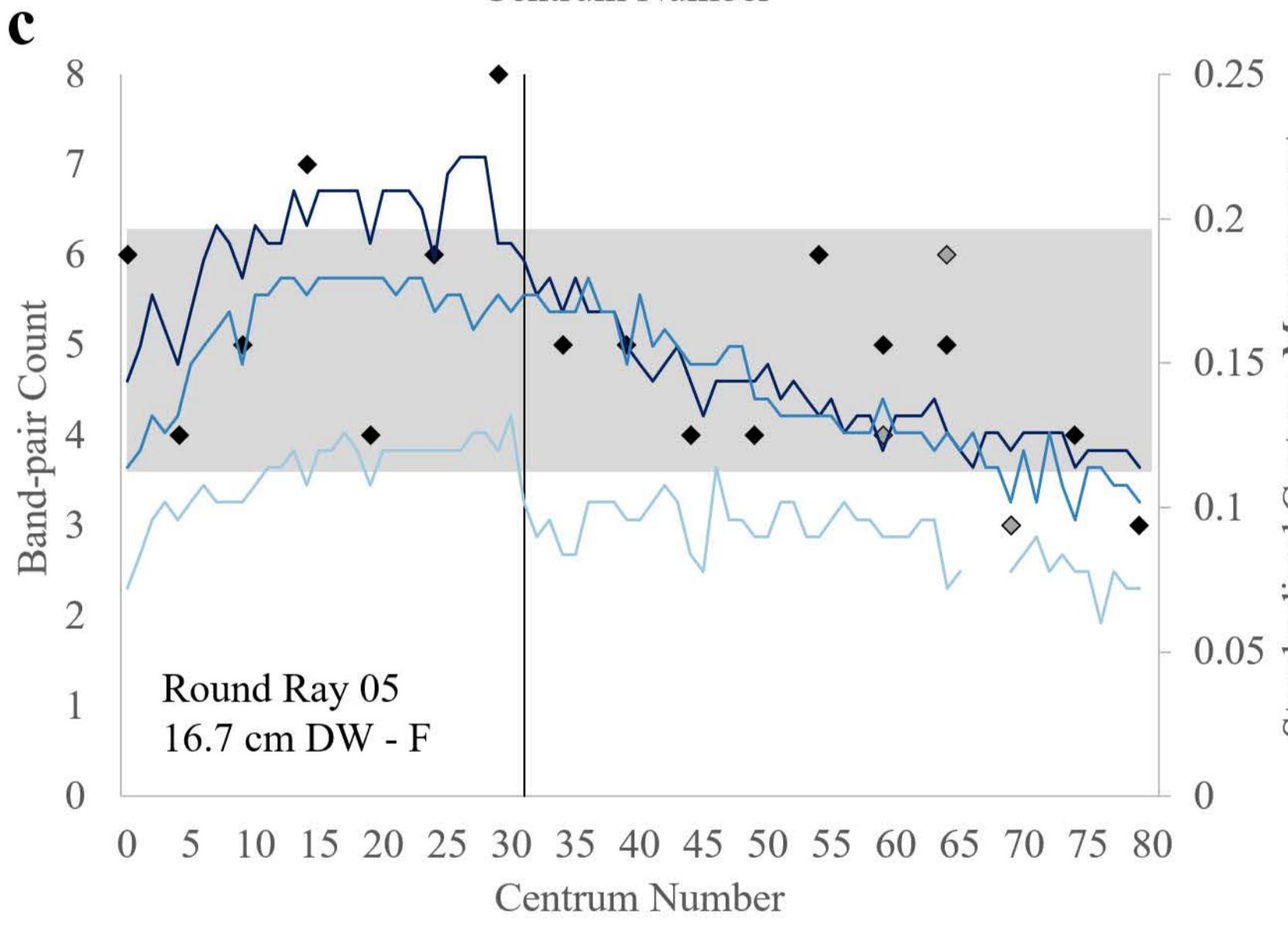
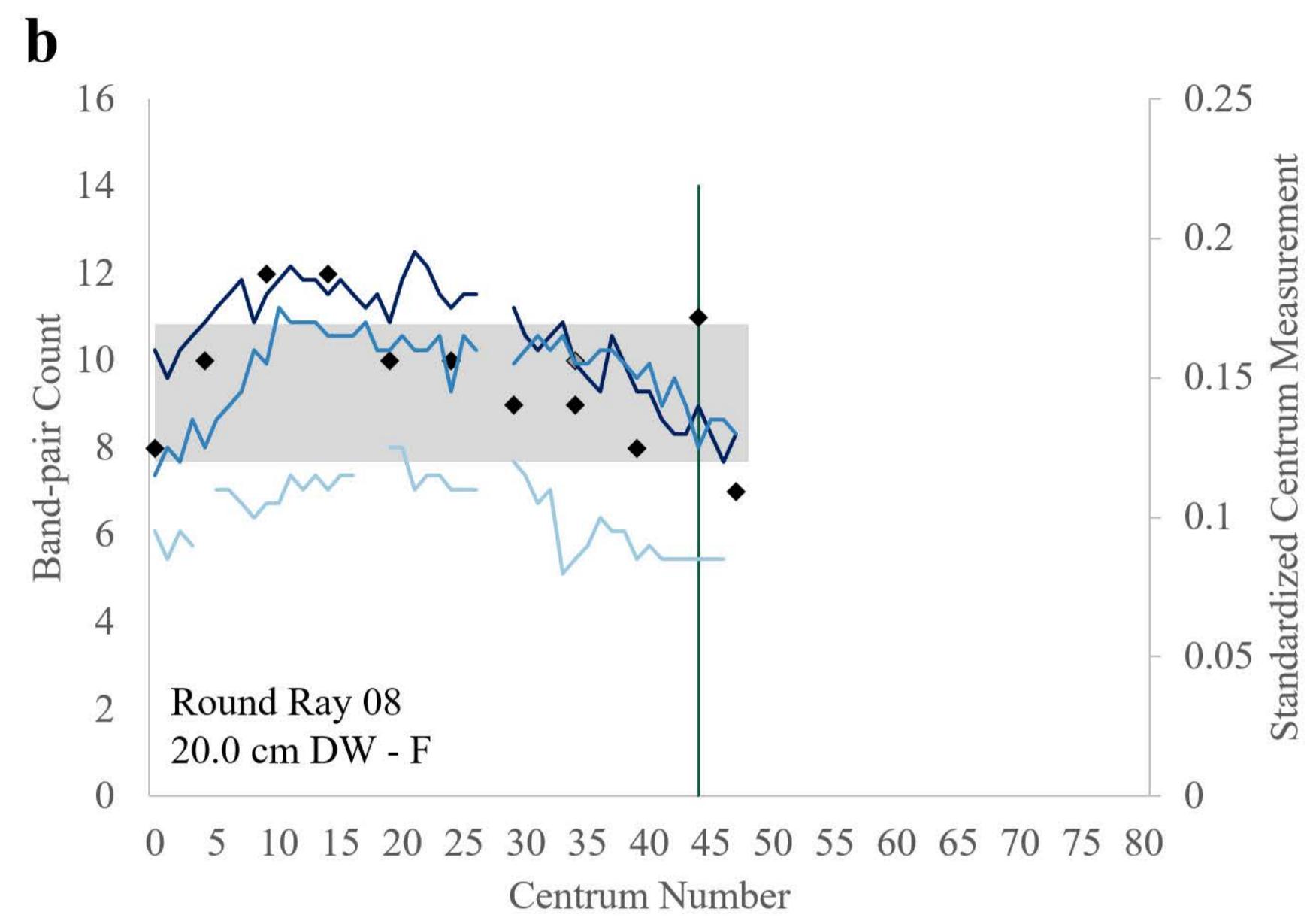
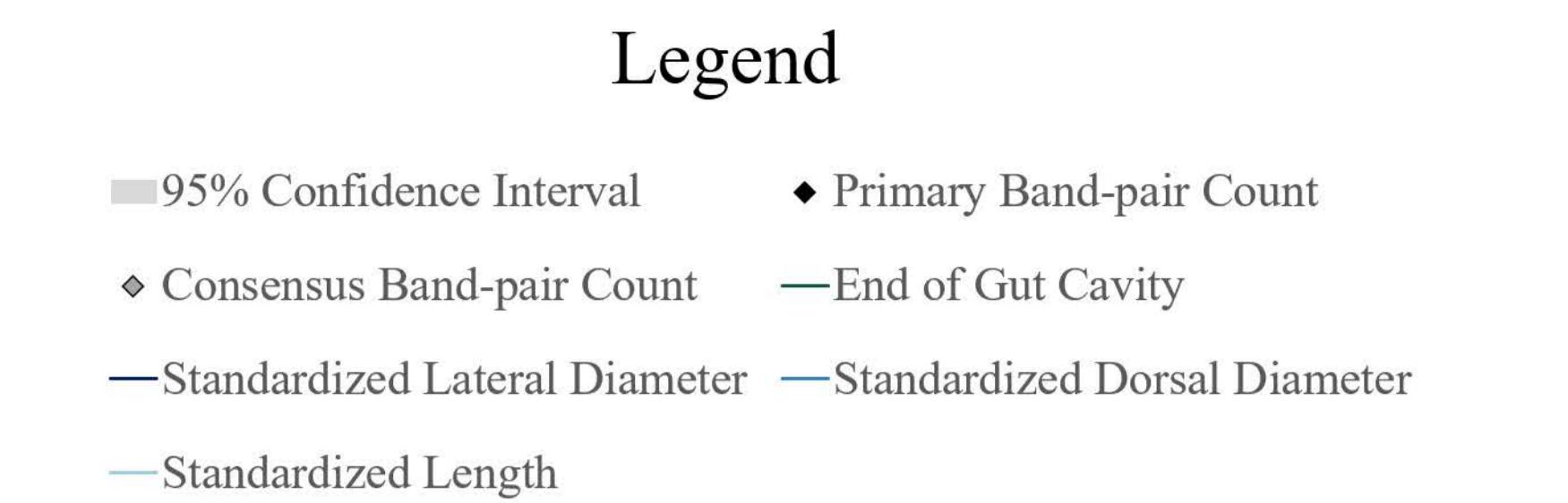
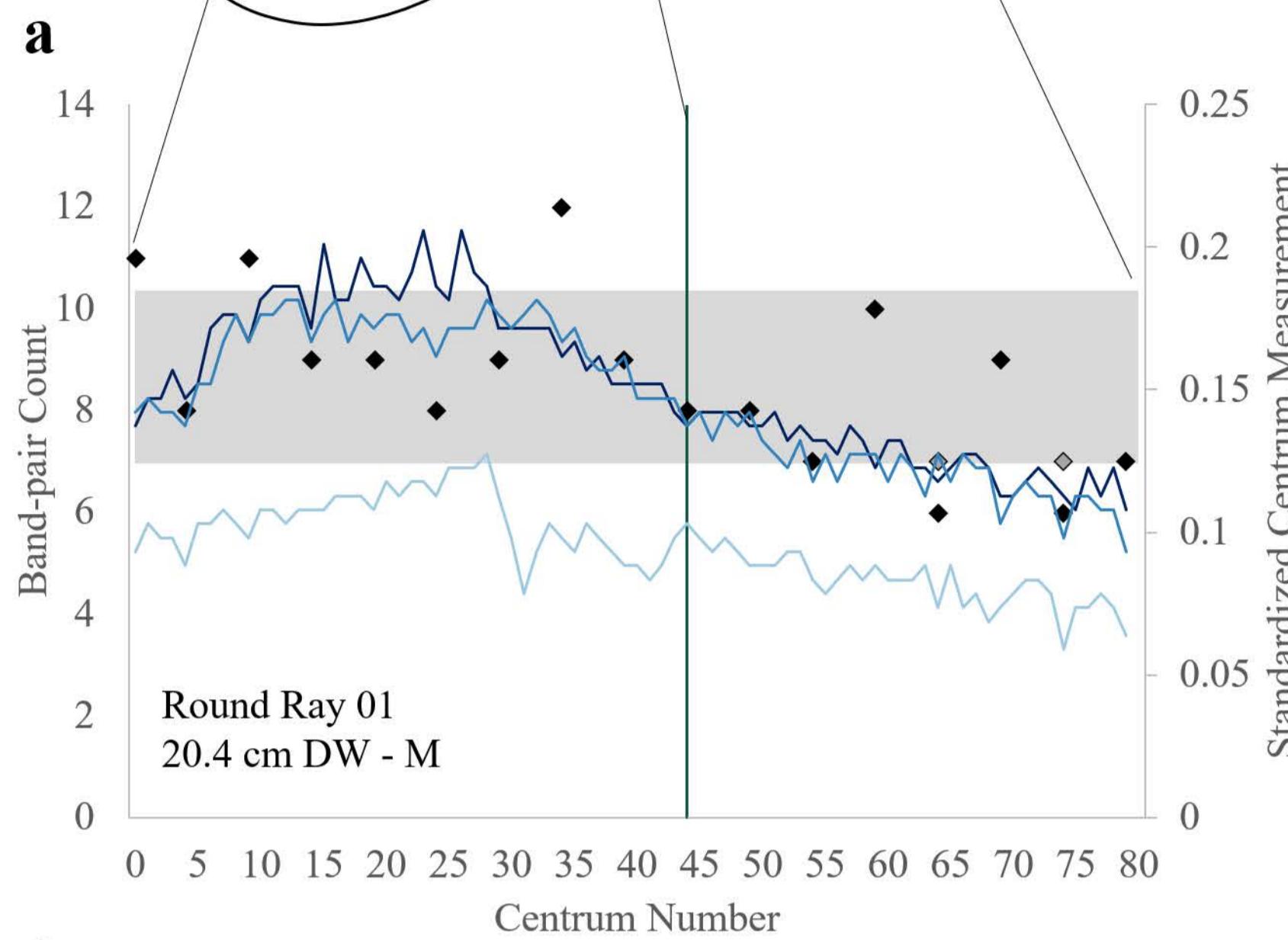
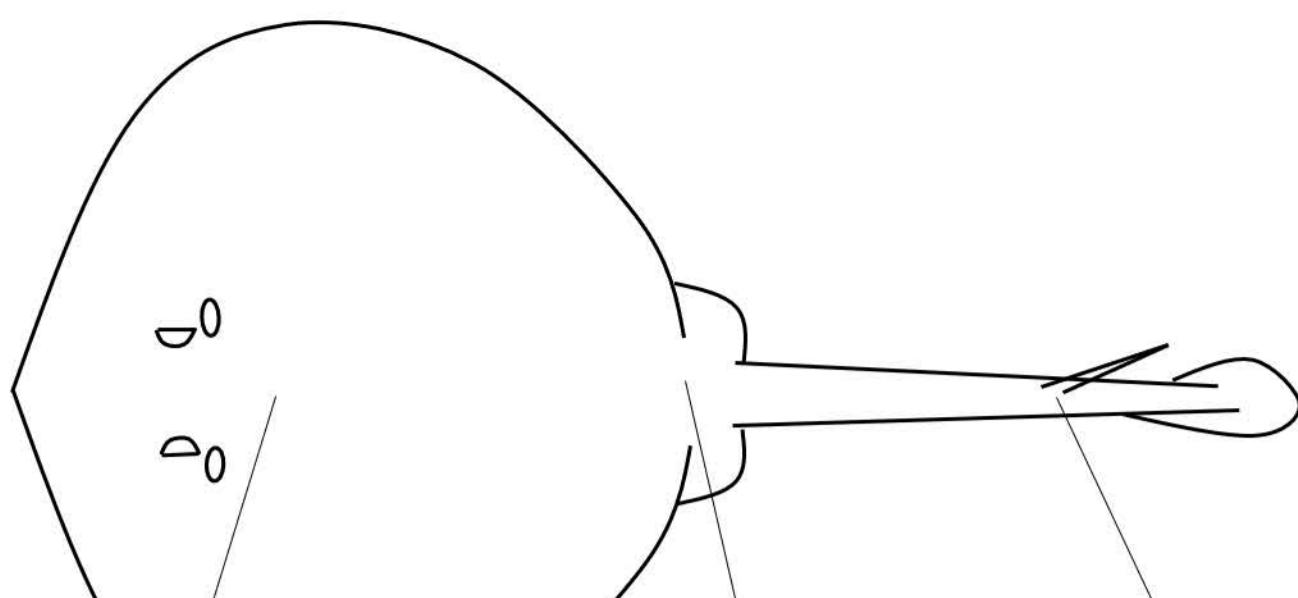


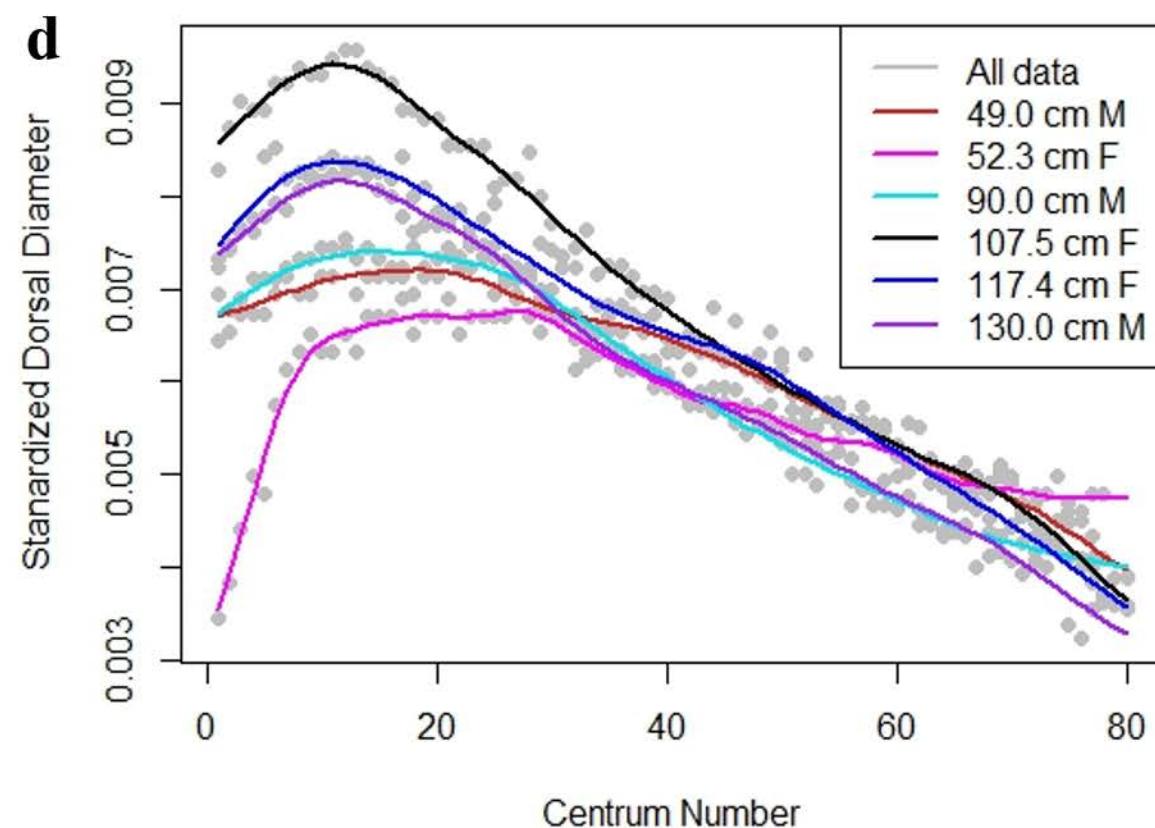
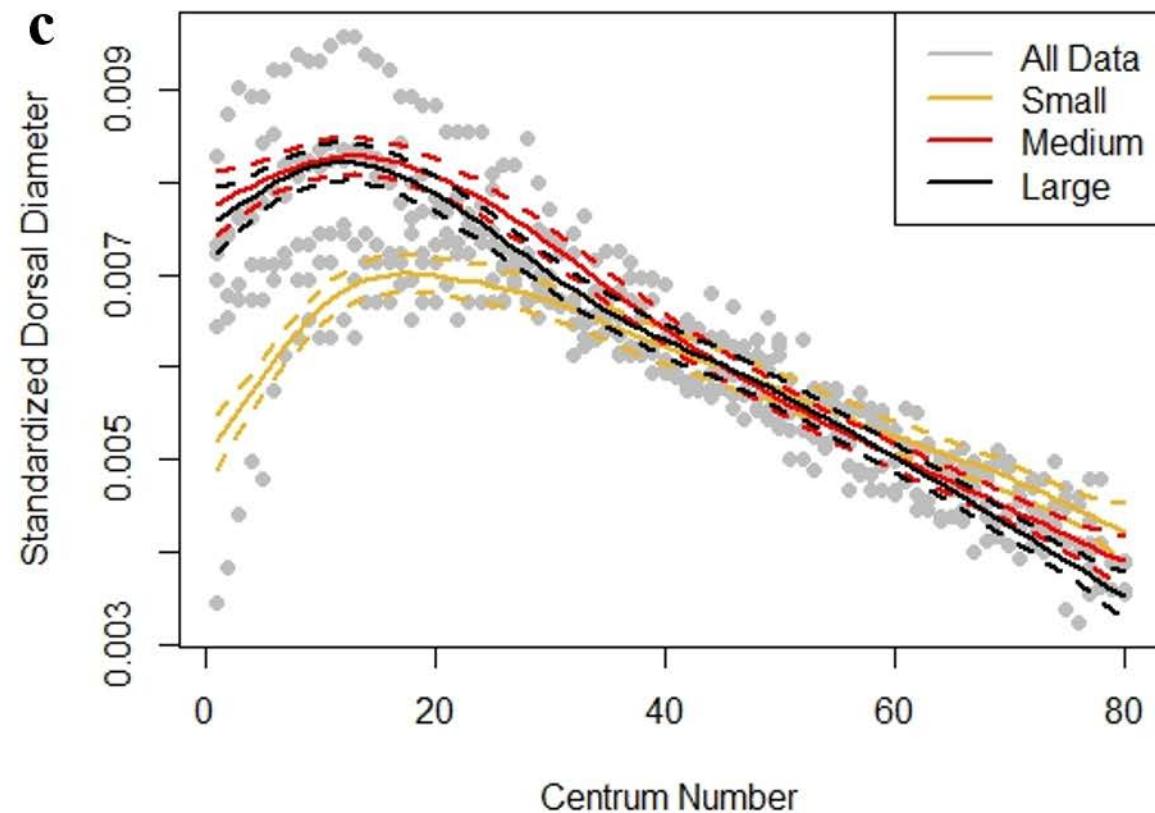
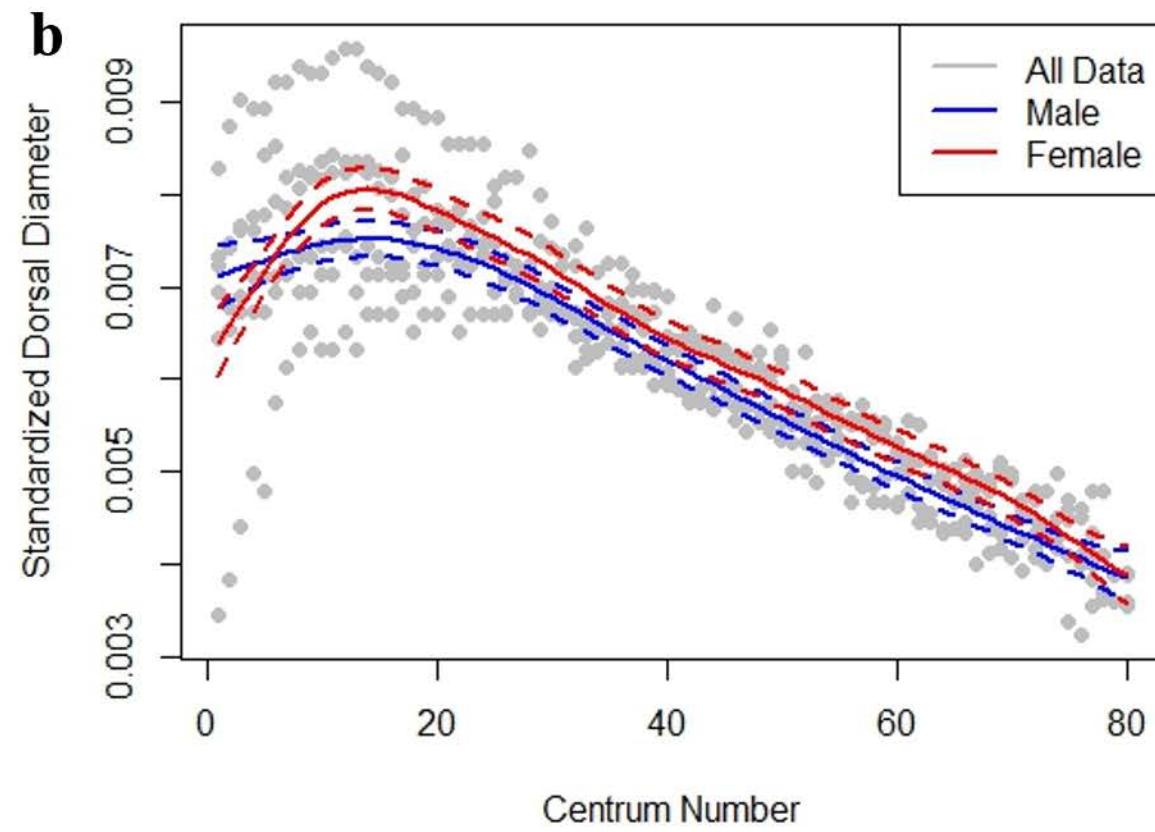
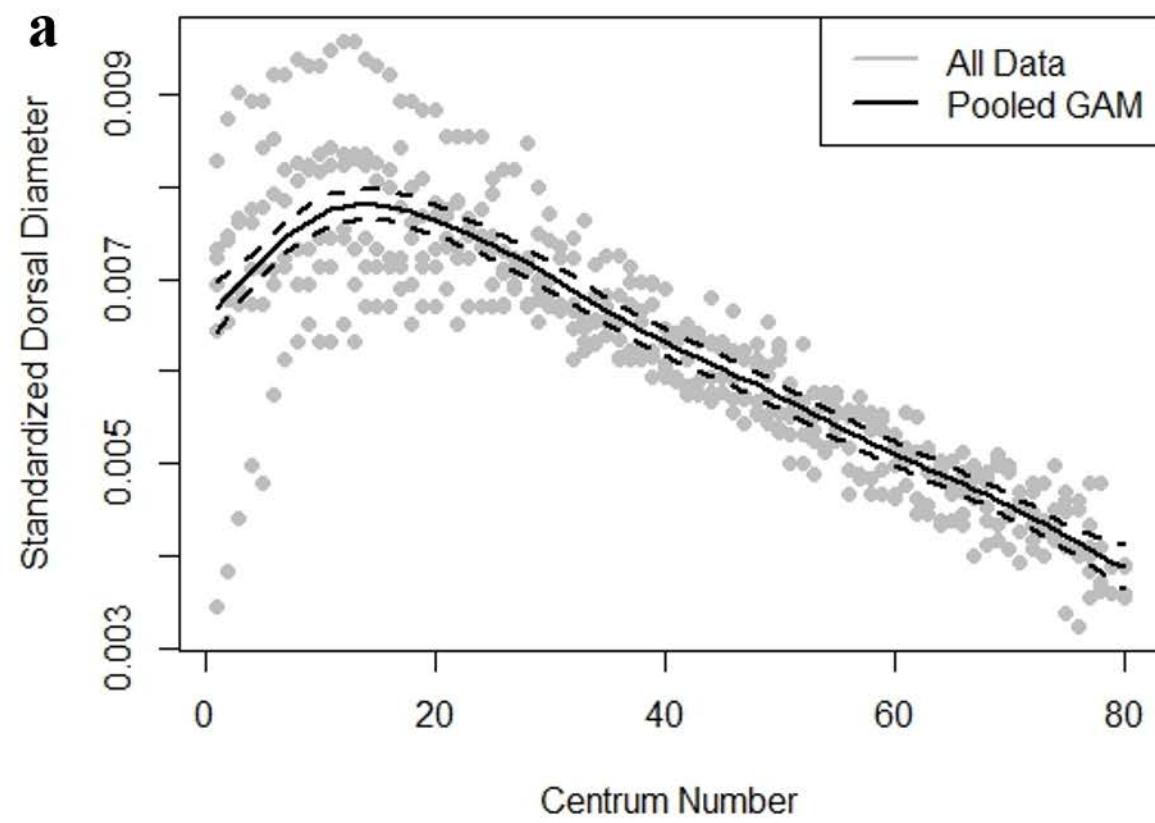


Legend

- 95% Confidence Interval
- ◆ Primary Band-pair Count
- ◆ Consensus Band-pair Count
- End of Gut Cavity
- Standardized Lateral Diameter
- Standardized Dorsal Diameter
- Standardized Length







Supplementary Table 1. Generalized additive model results for 10 different models to best describe centrum morphology along the vertebral column for five batoid species. k is the number of knots used in each model.

Species	Model	Dorso-ventral Diameter				Lateral Diameter				Rostro-caudal Length					
		Deviance	k	Adjusted r ²	AIC	Deviance	k	Adjusted r ²	AIC	Deviance	k	Adjusted r ²	AIC	AAIC	
Little Skate	Pooled	3277.71	15	0.84	-25275.12	4449.48	3273.56	18	0.88	-23443.01	4433.9	3088.55	15	0.45	
	BySex	Different Smoother	3265.75	18	0.84	-25263.06	4461.54	3261.56	18	0.88	-23453.14	4423.8	3072.59	18	0.47
		Different Intercept	3275.28	18	0.84	-25276.17	4448.43	3272.56	18	0.88	-23443.65	4433.3	3085.32	18	0.46
BySize	Different Smoother and Intercept	3264.76	18	0.84	-25261.29	4463.31	3260.57	18	0.88	-23454.93	4422	3071.55	18	0.48	
	Different Smoother	3256.68	18	0.85	-25330.62	4393.98	3250.89	18	0.89	-23733.30	4143.6	3060.25	18	0.52	
	Different Intercept	3274.29	18	0.84	-25278.37	4446.23	3271.6	18	0.88	-23440.82	4436.1	3084.36	18	0.47	
ByIndividual	Different Smoother and Intercept	3254.69	18	0.85	-25329.92	4394.68	3248.95	18	0.89	-23734.11	4142.8	3058.12	18	0.53	
	Different Smoother	3034.36	18	0.9	-26446.93	3277.67	2963.43	18	0.93	-24876.66	3000.3	2756.94	18	0.65	
	Different Intercept	3234.89	18	0.9	-26696.52	3028.08	3232.78	18	0.92	-24667.08	3209.9	3045.69	18	0.58	
Winter Skate	Different Smoother and Intercept	2901.33	18	0.96	-29724.60	0	2813.94	18	0.97	-27876.94	0	2644.62	18	0.78	
	Pooled	462.05	15	0.86	-3614.77	926.251	462.03	16	0.86	-3451.75	691.53	454.63	13	0.52	
	BySex	Different Smoother	452.01	18	0.87	-3650.95	890.077	451.25	18	0.88	-3489.35	653.93	449.49	10	0.54
BySize	Different Intercept	459.59	20	0.87	-3663.15	877.869	460.36	18	0.86	-3449.98	693.3	450.41	18	0.86	
	Different Smoother and Intercept	450.31	18	0.89	-3708.53	832.493	450.26	18	0.88	-3487.91	655.37	440.36	18	0.88	
	Different Smoother	443.47	15	0.93	-3896.50	644.521	439.29	17	0.92	-3677.35	465.93	441.87	10	0.58	
ByIndividual	Different Intercept	459.99	16	0.87	-3638.81	902.213	459.56	17	0.88	-3481.86	661.42	451.96	14	0.6	
	Different Smoother and Intercept	442.44	14	0.93	-3925.36	615.662	434.22	20	0.93	-3724.76	418.52	439.14	10	0.66	
	Different Smoother	412.76	17	0.95	-4084.52	456.506	397.4	20	0.96	-3999.54	143.74	426.11	12	0.65	
Barndoor Skate	Different Intercept	456.29	17	0.89	-3701.02	840.003	456.59	17	0.88	-3479.35	663.93	451.37	10	0.64	
	Different Smoother and Intercept	378.1	23	0.98	-4541.02	0	385	20	0.97	-4143.28	0	394.28	17	0.83	
	Pooled	459.00	10	0.79	-5574.04	1085.87	462.9	15	0.83	-5725.93	670.57	442.45	10	0.32	
BySex	Different Smoother	453.69	18	0.79	-5574.10	1085.81	458.24	10	0.83	-5731.21	665.29	437.25	10	0.33	
	Different Intercept	457.93	10	0.8	-5594.87	1065.04	462.8	10	0.83	-5723.23	673.28	441.47	10	0.32	
	Different Smoother and Intercept	453.24	10	0.8	-5596.25	1063.66	457.26	10	0.83	-5730.57	665.94	436.28	10	0.33	
BySize	Different Smoother	448.64	10	0.87	-5799.38	860.529	452.42	10	0.87	-5840.35	556.15	432.06	10	0.47	
	Different Intercept	457.05	10	0.82	-5632.87	1027.04	461.75	15	0.86	-5808.95	587.56	440.27	10	0.4	
	Different Smoother and Intercept	446.04	10	0.89	-5852.63	807.275	449.73	10	0.89	-5922.32	474.19	429.11	10	0.55	
ByIndividual	Different Smoother	433.82	10	0.92	-5955.65	704.26	434.07	10	0.93	-6090.14	306.37	418.69	10	0.52	
	Different Intercept	453.72	10	0.89	-5807.76	852.147	457.46	10	0.89	-5875.56	520.94	437	10	0.57	
	Different Smoother and Intercept	406.41	18	0.98	-6659.91	0	416.74	15	0.96	-6396.50	0	388.06	18	0.82	
Atlantic Stingray	Pooled	695.09	10	0.59	-4384.39	950.027	694.26	15	0.69	-4296.73	606.96	676.88	10	0.29	
	BySex	Different Smoother	691.18	15	0.61	-4420.81	913.605	690.54	10	0.69	-4295.07	608.62	673.52	10	0.69
	Different Intercept	693.28	15	0.59	-4382.33	952.085	694.03	10	0.69	-4307.25	596.44	677.05	10	0.69	
BySize	Different Smoother and Intercept	690.12	15	0.61	-4418.96	915.457	689.51	10	0.69	-4307.12	596.57	672.5	10	0.69	
	Different Smoother	681.98	15	0.64	-4465.12	869.296	683.35	10	0.72	-4375.49	528.2	666.12	15	0.3	
	Different Intercept	691.94	15	0.68	-4538.10	796.308	692.29	10	0.69	-4304.36	599.33	670.6	18	0.47	
ByIndividual	Different Smoother and Intercept	678.48	15	0.72	-4634.37	700.044	681.32	10	0.73	-4379.01	524.68	660.07	15	0.46	
	Different Smoother	658.12	10	0.66	-4487.95	846.458	654.97	10	0.74	-4386.82	516.87	648.52	10	0.28	
	Different Intercept	686.68	10	0.81	-4891.77	442.645	686.89	15	0.82	-4661.20	242.49	665.7	15	0.52	
Round Ray	Different Smoother and Intercept	622.79	15	0.91	-5334.41	0	631	15	0.88	-4903.69	0	629.71	15	0.52	
	Pooled	750.84	16	0.81	-4794.71	538.898	751.51	15	0.84	-4662.83	468.25	743.36	15	0.66	
	BySex	Different Smoother	745.95	10	0.81	-4806.81	526.799	742.96	15	0.85	-4710.33	420.76	736.55	15	0.67
BySize	Different Intercept	752.88	10	0.82	-4831.30	502.313	749.58	18	0.85	-4680.77	450.32	742.35	15	0.66	
	Different Smoother and Intercept	744.91	10	0.82	-4843.31	490.301	741.88	15	0.86	-4732.16	398.92	735.51	15	0.68	
	Different Smoother	734.47	15	0.82	-4824.02	509.591	736.98	15	0.85	-4685.57	445.52	730.4	15	0.66	
ByIndividual	Different Intercept	749.22	15	0.81	-4808.90	524.709	749.47	15	0.85	-4698.94	432.14	741.32	15	0.67	
	Different Smoother and Intercept	732.75	14	0.82	-4841.70	491.916	738.3	10	0.86	-4720.63	410.46	728.5	15	0.67	
	Different Smoother	685.33	17	0.86	-4924.13	409.486	698.01	11	0.87	-4763.74	367.35	695.76	15	0.7	
Different Intercept	741.4	16	0.87	-5037.33	296.281	742.03	15	0.89	-4937.57	193.52	733.99	15	0.72		
	Different Smoother and Intercept	656.47	23	0.92	-5333.61	0	680.26	12	0.92	-5131.09	0	681.04	15	0.77	
	Different Smoother	509.75	15	0.82	-5333.61	0	5131.09	15	0.92	-5097.25	0	510.47	15	0.77	