FUNCTIONAL AND PHYLOGENETIC DIVERSITY OF SHARKS IN THE NORTHEASTERN PACIFIC

3 Zachary A. Siders^{1*}, Lauren B. Trotta², Fabio P. Caltabellotta^{1,3}, Katherine B. Loesser⁴, 4 Benjamin Baiser², Robert N. M. Ahrens⁵

6 Fisheries and Aquatic Sciences Program, School of Forest, Fisheries, and Geomatics Sciences, University of Florida, Gainesville, FL

Department of Wildlife Ecology and Conservation, University of Florida, Gainesville, FL Coastal Oregon Marine Experiment Station, Oregon State University, Newport, OR 10 4Department of Oceanography and Coastal Sciences, Louisiana State University, Baton

11 Rouge, LA

12 ⁵ Fisheries Research and Monitoring Division, Pacific Islands Fisheries Sciences Center, National Oceanic and Atmospheric Administration, Honolulu, HI

*Corresponding Author:

Zachary A. Siders

zsiders@ufl.edu

Fisheries and Aquatic Sciences

19 University of Florida

20 7922 NW 71st Street

Gainesville, FL 32605

Acknowledgements:

 This work would not be possible without support from University of Florida Biodiversity Institute, College of Agricultural and Life Sciences, and College of Liberal Arts and Sciences. Museum and private collections helped inform much of this data and we would like to thank Dr. Gordon Hubbell and Dr. Andrea Kroetz for providing access to their private collections. We would also like to thank the Florida Museum of Natural History and Dr. Gavin Naylor, the Smithsonian Institution's National Museum of Natural History and Dr. Jeff Clayton, and National Oceanic and Atmospheric Administration Southeast Science Center Panama City Laboratory and Dr. Enric Cortés for providing access to collections. No permits were necessary for this work and the authors have no conflicts of interest to report.

Biosketch:

 Zach Siders is a quantitative ecologist. He builds and applies integrative tools to discern patterns and understand processes across the ecological hierarchy and inform resilient management strategies.

 Lauren Trotta is an ecologist applying phylogenetic approaches to better characterize and understand drivers of diversity in communities.

Fabio P. Caltabellotta is a quantitative marine fisheries scientist. He uses advanced

- data analysis tools and innovative approaches to improve marine fisheries resource
- management and conservation.

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record.](http://dx.doi.org/10.1111/jbi.14383) Please cite this article as doi: [10.1111/jbi.14383](http://dx.doi.org/10.1111/jbi.14383)

This article is protected by copyright. All rights reserved.

 Benjamin Baiser is a community ecologist and macroecologist. He takes empirical and theoretical approaches with a wide-range of organisms and systems to explore how species composition, traits, phylogenetic diversity, and interaction structure change over space and time.

 Katherine Loesser is a graduate student studying estuary food web dynamics. She contributed to this work as part of her undergraduate thesis.

 Robert Ahrens is a Fisheries Research Biologist at the NOAA Pacific Islands Fisheries Science Centers. He leads management strategy evaluations related to the management of fisheries and protected species.

Author contributions:

 ZAS, LBT, and FPC conceived the ideas. ZAS designed the methodology. LBT built the phylogeny. KBL and FPC collated the trait values from the literature. KBL and ZAS did the photogrammetry. ZAS, KBL, and FPC collected pictographs. ZAS did the analysis. All authors contributed to synthesis and interpretation of results, the writing, and review.

Running Title: Shark biodiversity in the northeastern Pacific

ABSTRACT

 Aim: The expansive spatial scale of pelagic communities and the difficulty in acquiring pelagic species' functional traits has stymied an understanding of marine community dynamics. We assembled and analyzed a shark trait database and community phylogeny to identify the major axes of trait variation that define shark functional groups. We tested whether membership to biophysical macroecological strata is related to these functional or phylogenetic relationships.

Location: Northeastern Pacific, 180-255ºE and 0-50ºN

Major taxa studied: Sharks (Class: Chondrichthyes, superorder: Selachimorpha)

 Methods: We built a community phylogeny and collected habitat, reproductive, somatic growth, trophic, and dentition traits. We used Principal Coordinates Analyses (PCoA) to identify axes of trait variation and hierarchical clustering to classify functional groups. We tested whether functional or phylogenetic relatedness determined species' membership to strata from five macroecological gradients: latitude, habitat type, 85 thermal, carbon source, and bathymetry.

 Results: We assembled 38 traits from 1,225 records from 130 sources, 260 pictographs from seven sources, and 631 teeth from 79 jaw specimens. Life history, *r* versus *K* selection, was responsible for the biggest division in the functional dendrogram. Vertical habitat preference, growth rates, diet, and dental morphology generated further divisions between *r*- or *K*-selected species. Vertical habitat preference, carbon source,

 and biochemical habitat type were significantly dispersed or clustered on the functional dendrogram or phylogram.

 Main conclusions: Habitat and reproductive traits were the most important trait suites driving shark functional diversity. Through ordination and clustering, we were able to associate major axes of trait variation to the membership of shark functional groups. The phylogram approximated well the functional dendrogram's backbone but was a poor substitute for the trait diversity at the tips. Given the long evolutionary history of sharks and coincident expansive trait diversity, merging functional and phylogenetic approaches was necessary to capture the dimensions of shark biodiversity.

103 Keywords: dimensions of biodiversity, elasmobranchs, museum collections, photogrammetry, pictographs, traits

INTRODUCTION

 The pelagic ocean covers over 50% of the planet's surface, but little has been explored of its community ecology (though see Grady et al., 2019; Pimiento et al., 2020). Pelagic species, in particular marine megafauna, can range thousands of kilometers and, as a result, pelagic "communities" span immense spatial scales, equal to or greater than the scale of many terrestrial macroecological gradients. These aspects impede the collection of specimens for measuring traits and reduce the application of common community ecology quantitative approaches (Schlägel et al., 2020). Thus, we do not have a baseline understanding of the functional and phylogenetic diversity of many marine communities especially those in the pelagic ocean. Neglecting this vast ecosystem stifles our understanding of the ecological mechanisms determining community structure and function (Villéger et al., 2017).

 In the pelagic ocean, sharks are the most abundant large-bodied species and exert strong top-down control on ecosystems as apex predators, mesopredators, and cannibals (Kitchell et al., 2002). Unlike other large-bodied pelagic fishes (e.g., tunas and billfishes) that evolved in the Miocene (~17 mya), most extant shark lineages evolved far earlier in the early Jurassic (Sorenson et al., 2014) and the first pelagic sharks evolved far earlier in the Early Cretaceous (~140 mya). This long evolutionary history coincides with an immense breadth of shark functional ecology. Sharks have evolved to inhabit nearly every marine habitat (Compagno et al., 2005), exhibit a variety of reproductive strategies from oviparity to various modes of viviparity (Parsons et al., 2008), range in length from 20 cm to 18 m (Compagno et al., 2005), and are consumers at almost every trophic level of the ocean food web (Motta & Huber, 2012). Further, sharks are frequently caught in fisheries (Karp et al., 2011; WPRFMC, 2019) and, as a

 result, many species have measured traits. Together, their ecological role, trait diversity, and relatively thorough sampling makes shark assemblages a robust choice for exploring pelagic community ecology.

 Species' traits can provide insight into species functions, interactions, distributions, and mechanisms that structure and maintain the diversity of communities and the function of ecosystems (Cadotte et al., 2011; McGill et al., 2006). Further, traits are the phenotypic expression of genetic diversity and combining trait-based approaches with phylogenetic approaches provides multi-faceted information about the evolutionary structure and resultant functioning of ecological assemblages (Mazel et al., 2017; Tucker et al., 2018; Violle et al., 2007). In a macroecological context, traits can provide insight into intra- and inter-specific distributions of niche breadth along ecological gradients (Pigot et al., 2016; Read et al., 2018) as well as the structure of the trait morphospace in an evolutionary lineage or community (Blonder et al., 2014; Price et al., 2019). Typically, the influence of a macroecological gradient on a community is assessed by comparing the taxonomic, functional, or phylogenetic diversity for a set of locales spanning the gradient. However, the open ocean's sheer size, extreme depths, strong seasonal changes, and the ability of pelagic species to move across these dimensions reduce the applicability of a defined local community. Instead, macroecological gradients (e.g., latitude, depth, temperature) can be divided into strata and used to assign species' memberships from a regional pool using natural history observations. The functional and phylogenetic relationships between species that co- occur within a given strata can then be a used to assess the influence of the macroecological gradient on community structure (Li et al., 2019).

 The ocean has classic latitudinal gradients as well as gradients coincident with increasing depth, such as pressure, light, productivity, temperature, and oxygen, that exert selective pressures on species traits (e.g., the evolution of bioluminescence, Martini & Haddock, 2017), as well as influence community assembly processes (Martini et al., 2019; Yasuhara et al., 2012). Previous studies have found that, at macro-scales, lower latitudes and higher surface temperatures are generally correlated with increased ectotherm diversity, while higher latitudes and lower surface temperatures are generally correlated with lower ectotherm diversity (Worm et al., 2005) and higher endotherm diversity (Grady et al., 2019). Macro-scale gradients result from changes in the carbon source, assessed using stable isotopes, and can reflect a species preference for neritic, transitional, or pelagic habitats (Davenport & Bax, 2002; Miller et al., 2008). At mesoscales, habitat type – as defined by biochemistry (Raes et al., 2018), static features (e.g., seamounts and hydrothermal vents) (Morato et al., 2010), or physical forcing (e.g., eddies) (Carvalho et al., 2019) – can drive differences in oceanic community composition.

 The Northeastern Pacific (NEP) is an ideal location to explore pelagic biodiversity 168 at macroecological scales as it covers 60° of latitude, several oceanic biomes (Sayre et al., 2021), a range of sea surface temperatures from tropical to Nearctic conditions, neritic and pelagic habitats, and depths from the surface to over 8000 m. This region is characterized by the Northeast Pacific Subtropical Gyre, an anticyclonic current formation. Strong upwelling occurs in the east along the California coast while numerous anticyclonic upwelling eddies occur far from the coast (Sun et al., 2019). Slow zonal currents form a 40º latitudinal swath of warm surface temperatures

 facilitating the expansion of tropical and sub-tropical species into higher latitudes while the California Current and cold-core pelagic eddies provide suitable habitat for temperate species at lower latitudes. Numerous oceanic seamounts, including the Hawaiian Archipelago, provide opportunities for mixing between epi-, meso-, and bathypelagic species in the same spatial area.

 The goal of this study was to compare the community functional and phylogenetic diversity of sharks occupying the NEP. Our objectives were to 1) collate a 182 trait database for sharks within the NEP using primary collection and literature search, 2) describe the major axes of functional diversity in trait suites, 3) characterize shark functional groups and their associated phylogenetic diversity, 4) identify the principal traits that drive the separation of shark functional groups, and 5) assess whether membership to macroecological strata is related to the functional or phylogenetic relationships between species.

MATERIALS AND METHODS

 Our general methods were to 1) build a community phylogeny, 2) collect and compile traits from a variety of sources, 3) conduct multivariate analyses (i.e., ordination, hierarchical clustering) to find major axes of trait variation among sharks and identify shark functional groups, 4) test whether functional or phylogenetic relatedness determined species' membership to macroecological strata.

Regional Pool

 We compiled the regional species list for the NEP, 180-255ºE and 0-50ºN , from published (Ebert et al. 2017) and online sources on Hawaiian and Californian ocean biodiversity (e.g.,<http://dlnr.hawaii.gov/sharks/hawaii-sharks/species-list/> and [https://swfsc.noaa.gov/textblock.aspx?ParentMenuId=123&id=971\)](https://swfsc.noaa.gov/textblock.aspx?ParentMenuId=123&id=971). For phylogenetic

 analyses, we selected outgroup species from three lineages sister to Selachimorpha (sharks) also found in this region of the NEP: Chimaeriformes, Myliobatiformes, and Rajiformes.

Phylogenetic Reconstruction

 As none of the recent Selachimorpha phylogenetic trees (Sorenson et al., 2014; Stein et al., 2018) had complete coverage our regional pool, we built a community phylogeny (or purpose-built phylogeny, sensu Li et al., 2019) to reconstruct the evolutionary relationships between the co-occurring shark species. Briefly, we used four mitochondrial loci *COI*, *CytB*, *NADH2*, and *16S*, and one nuclear locus, *Rag1*, to reconstruct the phylogenetic relationships for this dataset (full details in Supplemental Information 1). For each species, we downloaded sequences from GenBank (Supplemental Table 1), aligned sequences using MAFFT v7.388 (Katoh & Standley, 2013) plugin in Geneious Prime 2019.1.3 [\(https://www.geneious.com\)](https://www.geneious.com/), and, using PartitionFinder v. 2.1.1 (Lanfear et al., 2012), determined the optimal model of evolution for each locus. We reconstructed phylogenetic relationships for individual gene trees (Supplemental Figure 1) as well as all concatenated loci using Randomized Axelerated Maximum Likelihood (RAxML) (Stamatakis, 2006, 2014). We applied an ordinal level topological constraint to our concatenated RAxML analysis that reflects widely accepted relationships between Selachimorpha lineages deep in the tree (Sorenson et al., 2014) and is common practice in phylogenetic tree building in Selachimorpha (Sorenson et al., 2014; Stein et al., 2018).

Functional Traits

 We desired to capture a broad suite of traits reflecting the habitat use, life history strategies, diet preferences, and morphological variation associated with the functional ecology of the sharks in the regional pool (Violle et al., 2007). To do such, we compiled traits (Supplemental Table 2)(Figure 1) for the species of interest using literature search, pictographs (Supplemental Table 3), or enumeration and photogrammetry (Supplemental Table 4). Pictographs of each shark species' lateral profile (or body shape) were used to capture differences in relative external morphology (e.g., proportions, fin length, angle, and placement) as a supplement to the information on maximum length. Enumeration of shark dentition was used to capture differences in the number of teeth while photogrammetry was used to capture differences in the relative morphology of a species' tooth crown. Each pictograph or photograph was converted to a silhouette and its outline was reconstructed with Elliptical Fourier Analysis (EFA). The resulting Fourier coefficients were used then decomposed using Principal Components Analyses and the resulting principal component (PC) scores were used as traits in downstream analyses. The remaining continuous trait values were from multiple sources for each trait and were combined using a weighted average with the number of citations as weights. All traits were standardized to approximate a standard normal distribution for analyses. The complete trait set was then divided by biological theme into trait suites: habitat, reproductive, trophic, somatic growth, and dental morphology (Supplemental Table 2). Additional details on the trait collation and collection, weighting scheme, and standardization are available in Supplemental Information 2.

Principal Coordinates Analysis

 To identify and describe the major axes of shark functional diversity in each trait suite and all suites combined, we conducted a Principal Coordinates Analysis (PCoAs)

 on the Gower dissimilarity of the given trait suite. For each trait suite PCoA, all species 246 that had 50% or more coverage of the suite in question were included. The trait loading ranks were calculated as a vector sum of the first and second PCo loadings weighted by the percent variance explained by each PCo. All PCoAs were summarized in plots of 249 the first and second PCo scores for each species and the trait coverage within the suite was calculated. Additional details on the PCoA are available in Supplemental Information 3.

Hierarchical Clustering

 We used hierarchical clustering with the complete linkage method on the overall Gower trait dissimilarity matrix to construct a functional dendrogram. We identified the optimal number of functional groups in the functional dendrogram using the gap statistic and the first standard error maximum criterion (Maechler et al., 2019). Using the optimal number of clusters, cluster membership was assigned to each species. The mean trait value was calculated for each cluster removing any species with missing values. We assessed the phylogenetic clustering of each trait and the phylogenetic diversity of each functional group. For the former, the phylogenetic signal of each trait was evaluated by calculating Pagel's lambda using the *phytools* package (Revell, 2012). For the latter, three phylogenetic diversity metrics, phylogenetic distance (PD), the mean pairwise distance (MPD), and the mean nearest taxon distance (MNTD), were calculated for each functional group using the *picante* package (Kembel et al., 2010) using 999 265 iterations and an $\alpha = 0.05$ to assess significant standardized effect sizes (Z-scores) (Webb et al., 2002) using a tip-swapping null model. Additional details on the hierarchical clustering and optimal cluster search are available in Supplemental Information 3.

Macroecological relationships

 To test macroecological relationships of functional and phylogenetic diversity for shark assemblages, we defined strata for five gradients: latitudinal gradient, habitat type based on biochemistry (Sayre et al., 2021) (Supplemental Table 5), thermal gradient, carbon gradient, and bathymetric gradient. These strata were defined using independent sources on species' ranges, ecological marine units, sea surface temperature, and stable isotopes as well as from the depth ranges defined during our trait collection (strata definitions available in Supplemental Information 3). We then tested the phylogenetic and functional dispersion for each stratum across each of the macroecological gradients. The permutation-based standardized effect size was calculated using *picante* package (Kembel et al., 2010) for three diversity metrics: functional or phylogenetic distance (FD/PD), MPD, and MNTD using a tip-swapping null 281 model with 999 permutations and at an $\alpha = 0.05$.

RESULTS

Regional Pool

 The regional shark list included 56 species, but four species had either zero sequences (i.e., *Apristurus spongiceps*, *Centrophorus tessellatus*, *Etmopterus villosus*) or a few low quality sequences (i.e., *Cephaloscyllium ventriosum*) available on Genbank and thus were excluded from the regional pool, reducing the total to 52 species.

Phylogeny

 In addition to the regional pool, six additional species were included in the phylogeny as an outgroup (Supplemental Information 1). For all species (ingroup + outgroup) our dataset had 81.4% coverage across all sequences for the regional pool and the outgroup species (Supplemental Table 1). The final phylogram has 57 internal nodes, 12 of which were constrained (Figure 1). For unconstrainted nodes, bootstrap values ranged from 27 to 100 with and average bootstrap value of 84.84% (Supplemental Figure 2).

Functional Traits

 Each trait included at least 58% of species in the regional pool (Figure 1). 298 Generally, age at maturity and von Bertalanffy k values were missing together and 299 missing most among the traits. Similarly, diet compositions and $\delta^{15}N$ were missing together and missing the second most often among the traits. Minimum and maximum depth of occurrence, maximum length, and reproductive mode all had 100% coverage. Maximum depth, reproductive mode, size at birth, number of offspring, maximum length, δ^{15} N, and proportion of diet containing mollusks, crustaceans, zooplankton and plants all had strong phylogenetic signal (Figure 1).

Literature Traits

 Fourteen traits came solely from literature-derived sources (73% of all records), while four were informed by additional data collection through the photogrammetry process. A total of 1,225 trait value records were compiled from 130 unique sources. The median number of records per species was 26 from a median of 8 sources, with a minimum of 5 records (3 sources) and a maximum of 103 records (28 sources) (Supplemental Figure 3 & 4). On average, the literature-derived trait coverage was 78% per species with a minimum of 14% and a maximum of 100% (Supplemental Figure 5). Four traits had 100% coverage (Figure 1) (Supplemental Figure 6).

Pictograph Traits

 Body shape profiles (Figure 1) were compiled for 51 species (Supplemental Table 3). In total, seven sources were used to compile body shape silhouettes (Supplemental Figure 7) with a median of 5 sources per species, a minimum of 2 sources, a maximum of 7, and a total of 260 pictographs. Landmarks located at the apical point of the rostrum, the first dorsal fin, and the upper caudal fin lobe (Supplemental Figure 8) were needed to sufficiently align the silhouettes (Supplemental Figure 9). After harmonic calibration, 15 harmonics were used in EFA of body shape morphology (Supplemental Figure 10).

Photogrammetric Traits

 Tooth crown morphology was compiled for 35 species from 80 specimens for a total of 631 teeth (Figure 2, Supplemental Table 4). A minimum of one jaw specimen per species was obtained, a maximum of 12, and a median of 2. Teeth were aligned with centering and scaling (Supplemental Figure 11). After harmonic calibration, 16 harmonics were used in EFA of tooth crown morphology (Supplemental Figure 12).

Trait Processing

 For the Principal Components Analysis on the EF pictograph coefficients, body shapes varied principally in terms of fin erectness where species with the "classic" shark profile such as Carcharhinids and Lamnids separated from species with more "torpedo" profiles such as Squaliforms for PC1 (Supplemental Figure 13). Within the "classic" shark profiles, further separation along PC2 occurred as a function of whether the pectoral, dorsal, and caudal fins extended perpendicular to the anterior-posterior axis, as in Longfin Mako Shark (*Isurus paucus*), or Oceanic Whitetip Shark (*Carcharhinus*

 longimanus), or the caudal fin extended parallel to the anterior-posterior axis, as in the Alopiids (Supplemental Figure 13).

 For the PCA on EF photogrammetric coefficients, teeth shapes separated whether the crown height was longer than the crown width or vice versa for PC1 (Supplemental Figure 14). Along PC2, convex teeth, as in Lamnids and Carcharhinus species, separated from concave teeth, as in Gulper Shark (*Centrophorus granulosus*) and in Gray Smoothhound Shark (*Mustelus californicus*). A large cluster of species was identified that all roughly possessed the classic shark tooth, with roughly identical crown heights and widths as well as slightly concave (Supplemental Figure 14).

Principal Coordinates Analysis of Trait Variation

 The habitat PCoA scores split into two clusters (Figure 3A) defined by minimum 348 depths at and below the surface, while within those clusters, species separated by $\delta^{13}C$ and maximum depth (Supplemental Figure 15, Supplemental Table 6). Reproductive PCoA scores separated by life history strategy (Figure 3B). Species with large size at birth, late ages at maturity, low number of offspring, and high maternal investment separated from species with small sizes at birth, early ages at maturity, high number of offspring, and less maternal investment (Supplemental Figure 16). Similarly, the somatic growth PCoA scores separated larger, slower growing species from smaller, faster growing species (Figure 3C; Supplemental Figure 17). Nitrogen stable isotopic 356 signature (δ^{15} N), a trait integrating across the diet categories, was the strongest loading for the trophic trait suite PCoA (Supplemental Figure 18; Supplemental Table 5), with clear separation between apex predators like Great White Shark (*Carcharodon carcharias*), and zooplanktivores like Basking Shark (*Cetorhinus maximus*) (Figure 3D).

 In the dental trait suite PCoA, species with higher dorsal and ventral symphyseal and row counts separated from the rest of the species (Supplemental Figure 19). Teeth PC scores grouped tightly by PC axis across all eight teeth positions (Figure 3E). In the overall trait suite, the PCoA scores strongly separated by life history strategy, grouping species by reproductive traits, and minimum depth of occurrence (Figure 3F) (Supplemental Figure 20). Four of the five reproductive traits and two of the three habitat traits were in the top ten ranked loadings of the overall suite (Supplemental Table 5**).** Across PCoAs, species with trait coverage, that surpassed the exclusionary threshold, but was still low tended to have extreme PCoA scores, but not all in the same direction (Figure 3).

Hierarchical clustering of functional groups

 We identified ten as the optimal number of functional groups from the shark functional dendrogram (Figure 4). Group A was all Lamniforms with the exception of Blue Shark (*Prionace glauca*; Carcharhiniformes) and was a cluster of viviparous, mostly oophagous, pelagic species that on average exhibited a "classic" shark lateral profile, grew slowly, matured late, were high on the food chain and more specialized on cephalopods than teleosts with spear-shaped teeth (Figure 4B). Group B contained large, pelagic Lamniforms with large sizes at birth, very slow growth rates, late ages at maturity, and high tooth counts, at least for Megamouth (*Megachasma pelagios*) and Basking sharks. Group C contained mostly Lamniforms except for the Sphyrnids and had traits similar to group A but with higher nitrogen signatures from diets more generalized on teleosts, cephalopods, and marine mammals, and with tooth crowns equally wide and high. Group D was all Carcharhinid epipelagic, neritic, placentotrophic species that exhibit a "classic" shark lateral profile and were mostly teleosts specialists

 with tooth crowns equally wide and high. Group E was a small mesopelagic cluster of viviparous oophagic species with mostly missing values.

 Group F contained small-sized, pelagic, viviparous lecithotrophic mostly Squaliform species but with entirely different tooth crown shapes, whose convex crowns were angled laterally. Group G had only two species, Whale Shark (*Rhincodon typus*) and Sixgill Shark (*Hexanchus griseus*), with extremely slow life history strategies relative to the regional pool. Group H had the most taxonomic heterogeny with species from four shark orders that are viviparous lecithotrophic, diet generalists with tooth crowns wider than high. Group I were crustacean feeding specialists with a fast life history strategy relative to the regional pool and included only Carcharhiniforms with the exception of Horn Shark (*Heterodontus francisci*; Heterodontiformes). Group J was small-sized, viviparous lecithotrophic, mesopelagic Squaliform species. No shark functional groups were significantly over-dispersed (Figure 4B). Overall, groups B, C, D, and J were significantly under-dispersed across the phylogeny using the PD metric. Further, groups C and D were significantly under-dispersed as measured by MNTD (i.e., near the tips of the tree), while groups C, D, J, and I were under-dispersed according to MPD (i.e., deep in the phylogeny).

 Overall, the functional diversity of the regional pool split broadly into two groups along an axis of *r* (F-J groups) versus *K* (A-E groups) life history strategy: lecithotrophic viviparous and oviparous species with many, small offspring or placentotrophic and oophagous viviparous species with few, large offspring (Pianka, 1970) (Figure 4B). Differences in bathymetric habitat selection, somatic growth rates (e.g., age at maturity and Brody growth coefficient), and dietary specialization further split *r*-selected species.

407 In the *K*-selected group, carbon source (i.e., δ^{13} C), zooplanktivory, tooth counts, and tooth crown shape further split apart members. Species to species differences at the tips or within a functional cluster tended to result from differences in continuous traits. 410 For example, in cluster D, differences in δ^{13} C gave rise to four subgroups likely reflecting affinity for coastal zones. These comprised of an extremely coastal species, Whitetip Reef Shark (*Triaenodon obesus*), coastal reef shark species, such as Gray Reef (*Carcharhinus amblyrhynchos*) and Blacktip Reef Sharks (*C. melanopterus*), continental slope species, such as Spinner Shark (*C. falciformis*), and, lastly pelagic species, such as Oceanic Whitetip Shark.

Macroecological signal

 In the macroecological analysis, only Leopard Shark (*Triakis semifasciata*) was excluded from all analyses using AquaMaps distributions (latitude, thermal, and habitat 419 type), and species without δ^{13} C values were excluded from the carbon gradient test. The composition (Supplemental Table 7) of the latitudinal strata, habitat types, thermal strata, carbon source strata, and bathymetric strata had few significant instances of over– or under-dispersion on the functional dendrogram or the phylogeny across all diversity metrics (Figure 5). However, the epipelagic assemblage was over-dispersed for FD, the bathypelagic assemblage was under-dispersed for MNTD, the neritic assemblage was under-dispersed for both FD and MPD, and the Northern Subtropical Epipelagic habitat assemblage (EMU 11) was under-dispersed for MNTD. For the phylogeny, only neritic assemblages were significantly under-dispersed for MPD and only North Pacific and Beaufort Sea Epipelagic habitat assemblages were significantly over-dispersed for PD and MPD.

 Of the five macroecological gradients we tested, bathymetric habitat selection and carbon source had the most non-random dispersions relative to the functional dendrogram and phylogeny (Figure 5F-G). Many species occur in the epipelagic resulting in significant over-dispersion while bathypelagic species mostly come from Squaliformes with similar functional ecology (group F) resulting in significant under- dispersion (Figure 4). This can be clearly seen in the habitat trait suite PCoA where the strongest clustering between species was along minimum depth preference (Figure 3A) as well as by depth selection falling in the top three rankings of the habitat and overall 438 trait suite PCoAs (Table 1). Preference for neritic waters, identified by higher δ^{13} C values, also strongly matched the functional and phylogenetic structure. Differences in carbon source tended to structure a few of the functional groups broadly, with group B 441 comprised of strongly pelagic species (low δ^{13} C values) and group D comprised of neritic species (Figure 4A); however, most of the functional groups did not strongly align to a particular end of the carbon gradient axis (Figure 5F). On the phylogeny, neritic species were generally from Carcharhinidae and were clustered together deep in the functional dendrogram and phylogeny (under-dispersed for MPD), but not at the tips (random for MNTD) (Figure 5G).

DISCUSSION

 By combining literature sources, pictographs, and primary photogrammetric collection, we were able to produce a robust, novel trait database for sharks in the Northeastern Pacific. Combining these trait data with a phylogeny, we described major axes of functional diversity, tested shark traits for phylogenetic signal, defined shark functional groups, and assessed the phylogenetic and functional relatedness of shark

 species that co-occurred across a suite of macroecological gradients. Principally, we found the diversity *within* a functional group was poorly explained by phylogenetic distance and that the community phylogeny was a proxy for differences *between* functional groups. This is empirical support of simulation studies that have suggested that phylogenetic diversity may not encapsulate all the functional diversity in a community (e.g., Mazel et al., 2017; Tucker et al., 2018). Across the macroecological gradients we tested, we found bathymetry to be the strongest driver of community structure, which parallels similar findings in teleosts both nearshore (Fujita et al., 1995; Moranta et al., 1998) and offshore (reviewed by Sutton, 2013).

 Likely as a result of the broad species' ranges of most species in our regional pool, we did not find support for latitudinal or thermal gradients as other marine megafauna studies have found (Grady et al., 2019; Worm et al., 2005)(Figure 5). We posit that the scale of the contrast between the surface and vertical gradients drives this phenomenon. For example, the surface thermal gradient in the NEP spans roughly 467 4000 km and varies from as low as 10°C in the California Current to as high as 30°C in the Pacific North Equatorial Current. In contrast, the vertical thermal gradient is equally as large within the first 500 m from the surface. Along with increasing pressures and decreasing dissolved oxygen, depth is likely to be a stronger functional diversity driver than latitude in the pelagic ocean (Rosa et al., 2008; Yasuhara et al., 2012). In addition, our finding of weak support for functional and phylogenetic clustering based on carbon source (Figure 5F-G) is similar to other studies that have documented changes as a function of distance from shore for large taxa oceanic communities (Davenport & Bax, 2002; Morato et al., 2010).

 Relative to other community ecology datasets, many NEP shark species are large, mobile, marine organisms with close to circumglobal distributions (Supplemental Table 7). Defining a community is difficult in these circumstances without fine-scale occurrence data (Schlägel et al., 2020), which is often strongly biased against rare species that can disproportionately contribute to functional diversity (Leitão et al., 2016). Another consideration is that the habitat matrix is three-dimensional and strongly dynamic. Increases in depth coincide with increases in pressure and decreases in light, productivity, temperature, and oxygen generating sub-surface habitats that can result in different assemblages (Farré et al., 2016; Martini et al., 2019). An inexact analog is migratory birds whose community membership changes along their migratory pathways (Chen et al., 2018; Zuckerberg et al., 2016). We use "inexact" purposefully as the seasonal dynamics within a given locale in the NEP can be equivalent to the creation and erosion of elevational gradients and 20º shifts in latitudinal gradients within a year as eddies form and dissipate, current strength waxes and wanes, and sub-surface features stabilize and abate. Subdividing these seasonal and vertical components more explicitly may be necessary to draw better comparisons to between macroecological gradients relevant to the pelagic ocean to those commonly studied in terrestrial, freshwater, or neritic ecosystems.

 An obvious caveat to our conclusions is the role of missing data within the trait matrix. Not surprisingly, traits derived from more labor-intensive collection, such as growth rates, standardized diets, and age at maturity, were missing for the greatest number of species. Similarly, rare and deep-water species, such as Frilled Shark (*Chlamydoselachus anguineus*), were missing the greatest number of traits (Figure 1).

 Compared to community studies of other taxa, many shark species in our regional pool are exploited either directly or indirectly through fisheries bycatch (WPRFMC, 2019). For example, Filetail Catshark (*Pseudotriakis microdon*) has been collected in fisheries- independent surveys (Keller et al., 2014), but has not had a formal study on age and growth. Thus, for some species, the hurdle to improving trait coverage is not the collection of specimens but generating usable data from those collections.

 Our use of pictographs for body morphology and photogrammetry for tooth crown morphology represent one method to generate functional traits for these data-poor species. These digitization efforts were designed to provide holistic traits with reasonable species coverage (51/52 and 35/52 species, respectively). It appears this holism was accomplished as the broad splitting of the functional dendrogram along the *r*-*K* axis was reflected in the body shapes and tooth crown morphologies. The contrast between species with elongated first dorsal and pectoral fins as well as a high aspect ratio caudal fin (groups A-D) and species with more pronounced second dorsal and anal fins as well as lower aspect ratio caudal fins (groups F-J)(Figure 4) is consistent with the groups from the seminal pictograph analyses on shark body shapes by Thomson and Simanek (1977). Similarly, groups A-E had tooth crowns with centered apexes, often long crowns, and flat root margins while F-J groups had off-centered apexes, short crowns, and, for three of the five groups, bulbous root margins. We were impressed with this parallelism across trait axes as the pictographs and photographs were analyzed in a scale-free way. We recommend other functional ecology studies attempt similar efforts for difficult to acquire traits based on their ease of implementation (at least for the pictographs) and success herein.

 In terms of impact of trait coverage on our analyses, species with lower coverage tended to be on the fringes of the PCoAs; especially those in trait suites with a mixture high and low coverage (e.g., habitat, reproductive, somatic growth, and overall) (Figure 3 and Supplemental Figures 15-20). Low trait coverage played a role in determining the functional groups with groups E and H seemingly defined by it (Figure 4). A reasonable assumption would be that species with many missing trait values would likely be reassigned to other clusters or grouped into a larger cluster with improved data coverage. This is important for considering downstream applications, such as calculating functional diversity at finer spatiotemporal scales, because the dendrogram structure is partially influenced by coverage, and is likely to change as new data are included (Kim et al., 2018).

 For our phylogenetic analyses, we chose to use a phylogram over a chronogram. This allowed us to focus on comparing trait diversity to feature diversity rather than divergence time between species (Jantzen et al., 2019). Phylogenies are often used as proxies for functional diversity based on the assumption that, when using a robust phylogeny and including numerous functional traits, the two should be highly correlated; though this is not always the case (Mazel et al., 2017; Tucker et al., 2018). In this study, we sought to explicitly relate the multiple facets of functional diversity to the phylogenetic information derived from our highly sampled, well-resolved community phylogeny (Gerhold et al., 2015). We found that individual traits tend to either have very high or very low phylogenetic signal (Figure 1). Across functional groups, half of them were clustered deep within the phylogeny and 40% were clustered overall. Despite this strong similarity in the functional dendrogram and phylogram's backbone structure, only

 20% of functional groups had species clustered near the branch tips. Thus, for our shark species, the phylogram is a proxy for extreme differences in functional traits derived from evolutionary divergence deep in the tree but, not for the diversity within a functional group occurring near the functional dendrogram tips.

 Lastly, conducting PCoAs alongside the functional dendrogram analyses facilitated a better understanding of how the structure of the functional dendrogram was generated. Distinct clustering and strongly loading traits in the habitat and reproductive trait suite PCoAs reflected deep splits and influential traits in the functional dendrogram while equivocal clusters in somatic, trophic, and dentition trait suite PCoAs reflected shallower splits. Additionally, by conducting PCoAs on the suites independently, we could identify major axes of variation for each suite and compare to how those axes integrated in the functional dendrogram, in the functional groups, and in the overall PCoA. This is entirely expected as the functional dendrogram and PCoAs used the same data but, by facilitating different visualization, yielded complementary and confirmative approaches.

 Trait datasets have a multitude of applications within functional ecology and community phylogenetics (Cadotte et al., 2011). Applying trait filling approaches to our trait matrix is a likely necessary step for pursuing some of these applications. The strong signal of bathymetry and carbon source gradients apparent in the PCoAs and functional groups are an opportunity to explore the correlates of macroecological turnover in taxonomic, functional, and phylogenetic diversity in the pelagic ocean. Lastly, there are opportunities to apply these diversity enumerations to the management of sharks for the numerous fisheries operating in the NEP. The listing of Oceanic

 Whitetip Shark and Scalloped Hammerhead Shark (*Sphryna lewini*) under the Endangered Species Act places considerable pressure on United States fisheries to find solutions to reduce shark-fishing interactions (Karp et al., 2011; WPRFMC, 2019). This trait database and community phylogenetics analysis can be a complementary way to categorize bycatch risk across shark species, define appropriate surrogates for risk analyses of data-deficient species, and provide holistic metrics of shark biodiversity impacts.

REFERENCES

- Blonder, B., Lamanna, C., Violle, C., & Enquist, B. J. (2014). The n-dimensional
- hypervolume: The n-dimensional hypervolume. *Global Ecology and*

Biogeography, *23*(5), 595–609. https://doi.org/10.1111/geb.12146

- Cadotte, M. W., Carscadden, K., & Mirotchnick, N. (2011). Beyond species: Functional diversity and the maintenance of ecological processes and services: Functional diversity in ecology and conservation. *Journal of Applied Ecology*, *48*(5), 1079– 1087. https://doi.org/10.1111/j.1365-2664.2011.02048.x
- Carvalho, A. da C. de O., Mendes, C. R. B., Kerr, R., Azevedo, J. L. L. de, Galdino, F., & Tavano, V. M. (2019). The impact of mesoscale eddies on the phytoplankton community in the South Atlantic Ocean: HPLC-CHEMTAX approach. *Marine Environmental Research*, *144*, 154–165.

https://doi.org/10.1016/j.marenvres.2018.12.003

- Chen, C., Holyoak, M., Si, X., Wang, Y., & Ding, P. (2018). Do seasonal species assemblages differ in their biogeography? Evidence from the spatial structure of bird communities on land‐bridge islands. *Journal of Biogeography*, *45*(2), 473–
- 483. https://doi.org/10.1111/jbi.13112
- Compagno, L. J. V., Dando, M., & Fowler, S. L. (2005). *Sharks of the World*. Princeton University Press.
- https://press.princeton.edu/books/paperback/9780691120720/sharks-of-the- world
	- Davenport, S. R., & Bax, N. J. (2002). A trophic study of a marine ecosystem off
	- southeastern Australia using stable isotopes of carbon and nitrogen. *Canadian*
- *Journal of Fisheries and Aquatic Sciences*, *59*(3), 514–530.
- https://doi.org/10.1139/f02-031
- Farré, M., Tuset, V. M., Cartes, J. E., Massutí, E., & Lombarte, A. (2016). Depth-related trends in morphological and functional diversity of demersal fish assemblages in the western Mediterranean Sea. *Progress in Oceanography*, *147*, 22–37.
- https://doi.org/10.1016/j.pocean.2016.07.006
- Fujita, T., Inada, T., & Ishito, Y. (1995). Depth-gradient structure of the demersal fish community on the continental shelf and upper slope off Sendai Bay, Japan.
- *Marine Ecology Progress Series*, *118*, 13–23.
- https://doi.org/10.3354/meps118013
	- Gerhold, P., Cahill, J. F., Winter, M., Bartish, I. V., & Prinzing, A. (2015). Phylogenetic patterns are not proxies of community assembly mechanisms (they are far better). *Functional Ecology*, *29*(5), 600–614. https://doi.org/10.1111/1365- 2435.12425
- Grady, J. M., Maitner, B. S., Winter, A. S., Kaschner, K., Tittensor, D. P., Record, S.,
- Smith, F. A., Wilson, A. M., Dell, A. I., Zarnetske, P. L., Wearing, H. J., Alfaro, B.,
- & Brown, J. H. (2019). Metabolic asymmetry and the global diversity of marine
- predators. *Science*, *363*(6425), eaat4220.
- https://doi.org/10.1126/science.aat4220
- Jantzen, J. R., Whitten, W. M., Neubig, K. M., Majure, L. C., Soltis, D. E., & Soltis, P. S.
- (2019). Effects of taxon sampling and tree reconstruction methods on
- phylodiversity metrics. *Ecology and Evolution*, *9*(17), 9479–9499.
- https://doi.org/10.1002/ece3.5425

 $\overline{\zeta}$

 $\big($

 \blacksquare

 $\big($

 Proceedings of the Royal Society B: Biological Sciences, *283*(1828), 20160084. https://doi.org/10.1098/rspb.2016.0084

Li, D., Trotta, L., Marx, H. E., Allen, J. M., Sun, M., Soltis, D. E., Soltis, P. S., Guralnick,

- R. P., & Baiser, B. (2019). For common community phylogenetic analyses, go ahead and use synthesis phylogenies. *Ecology*, *100*(9).
- https://doi.org/10.1002/ecy.2788
- Maechler, M., Rousseeuw, P., Struyf, A., Hubert, M., & Hornik, K. (2019). *cluster: Cluster Analysis Basics and Extensions*.
- Martini, S., & Haddock, S. H. D. (2017). Quantification of bioluminescence from the surface to the deep sea demonstrates its predominance as an ecological trait. *Scientific Reports*, *7*(1), 45750. https://doi.org/10.1038/srep45750
- Martini, S., Kuhnz, L., Mallefet, J., & Haddock, S. H. D. (2019). Distribution and quantification of bioluminescence as an ecological trait in the deep sea benthos. *Scientific Reports*, *9*(1), 14654. https://doi.org/10.1038/s41598-019-50961-z
- Mazel, F., Mooers, A. O., Riva, G. V. D., & Pennell, M. W. (2017). Conserving
- Phylogenetic Diversity Can Be a Poor Strategy for Conserving Functional
	- Diversity. *Systematic Biology*, *66*(6), 1019–1027.
- https://doi.org/10.1093/sysbio/syx054
- McGill, B. J., Enquist, B. J., Weiher, E., & Westoby, M. (2006). *Rebuilding community ecology from functional traits*. 8.
- Miller, T. W., Brodeur, R. D., & Rau, G. H. (2008). Carbon stable isotopes reveal relative contribution of shelf-slope production to the Northern California Current pelagic

 \rightarrow

 ζ

 \mathbf{r} \sim

S

 $\overline{}$

 \rightarrow

-

×

S

- Sayre, R. G., Wright, D. J., Breyer, S. P., Butler, K. A., Graafeiland, K. V., Costello, M. J., Harris, P. T., Goodin, K. L., Guinotte, J. M., Basher, Z., Kavanaugh, M. T., Halpin, P. N., Monaco, M. E., Cressie, N., Aniello, P., Frye, C. E., & Stephens, D. (2021). A Three-Dimensional Mapping of the Ocean Based on Environmental Data. *Oceanography*, *30*(1), 90–103. Schlägel, U. E., Grimm, V., Blaum, N., Colangeli, P., Dammhahn, M., Eccard, J. A., Hausmann, S. L., Herde, A., Hofer, H., Joshi, J., Kramer‐Schadt, S., Litwin, M., Lozada‐Gobilard, S. D., Müller, M. E. H., Müller, T., Nathan, R., Petermann, J. S., Pirhofer‐Walzl, K., Radchuk, V., … Jeltsch, F. (2020). Movement‐mediated community assembly and coexistence. *Biological Reviews*, *95*(4), 1073–1096. https://doi.org/10.1111/brv.12600 Sorenson, L., Santini, F., & Alfaro, M. E. (2014). The effect of habitat on modern shark diversification. *Journal of Evolutionary Biology*, *27*(8), 1536–1548. https://doi.org/10.1111/jeb.12405 Stamatakis, A. (2006). RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics*, *22*(21), 2688–2690. https://doi.org/10.1093/bioinformatics/btl446 Stamatakis, A. (2014). RAxML version 8: A tool for phylogenetic analysis and post- analysis of large phylogenies. *Bioinformatics*, *30*(9), 1312–1313. https://doi.org/10.1093/bioinformatics/btu033
	- Stein, R. W., Mull, C. G., Kuhn, T. S., Aschliman, N. C., Davidson, L. N. K., Joy, J. B.,
	- Smith, G. J., Dulvy, N. K., & Mooers, A. O. (2018). Global priorities for conserving

 the evolutionary history of sharks, rays and chimaeras. *Nature Ecology & Evolution*, *2*(2), 288–298. https://doi.org/10.1038/s41559-017-0448-4 Sun, W., Dong, C., Tan, W., & He, Y. (2019). Statistical Characteristics of Cyclonic Warm-Core Eddies and Anticyclonic Cold-Core Eddies in the North Pacific Based on Remote Sensing Data. *Remote Sensing*, *11*(2), 208. https://doi.org/10.3390/rs11020208 Sutton, T. T. (2013). Vertical ecology of the pelagic ocean: Classical patterns and new perspectives: vertical ecology of the pelagic ocean. *Journal of Fish Biology*, *83*(6), 1508–1527. https://doi.org/10.1111/jfb.12263 Thomson, K. S., & Simanek, D. E. (1977). Body Form and Locomotion in Sharks. *American Zoologist*, *17*(2), 343–354. https://doi.org/10.1093/icb/17.2.343 Tucker, C. M., Davies, T. J., Cadotte, M. W., & Pearse, W. D. (2018). On the relationship between phylogenetic diversity and trait diversity. *Ecology*, *99*(6), 1473–1479. https://doi.org/10.1002/ecy.2349 Villéger, S., Brosse, S., Mouchet, M., Mouillot, D., & Vanni, M. J. (2017). Functional ecology of fish: Current approaches and future challenges. *Aquatic Sciences*, *79*(4), 783–801. https://doi.org/10.1007/s00027-017-0546-z Violle, C., Navas, M.-L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., & Garnier, E. (2007). Let the concept of trait be functional! *Oikos*, *116*(5), 882–892. https://doi.org/10.1111/j.0030-1299.2007.15559.x Webb, C. O., Ackerly, D. D., McPeek, M. A., & Donoghue, M. J. (2002). Phylogenies and Community Ecology. *Annual Review of Ecology and Systematics*, *33*(1), 475–505. https://doi.org/10.1146/annurev.ecolsys.33.010802.150448

 $\overline{}$

DATA AVAILABILITY STATEMENT

 All scripts and data to reproduce the data and supplementary information can be found at [https://osf.io/eszvb/.](https://osf.io/eszvb/)

- 779 **TABLE 1**
- 780 Percent of total variance explained by each principal coordinate (PCo) for each trait
- 781 suite and the top three loadings from the Principal Coordinates Analysis of NE Pacific
- **782** shark functional traits.

785 **FIGURE 1**

788 Summary of the phylogram (left-side) and trait table (right-side) for the 52 shark species 789 in the NE Pacific regional pool. Different colors in the phylogram identify different shark 790 orders. All traits are blocked by suite (habitat, reproduction, somatic growth, and trophic)

ー

 with darker colors indicating higher scaled values for continuous traits. The four reproductive modes are colored individually. The proportion of the trait filled and the 793 phylogenetic signal of the trait, measured by Pagel's λ , are along the bottom with darker colors indicating values closer to 1 (i.e., complete coverage or strong phylogenetic 795 signal for a particular trait). The mean body shape from pictographs for each species is Shown on the right.

 \geq

798

 Summary of the dental traits and subset of the phylogram (left-side) for NE Pacific shark species with jaw specimens. The number of teeth replicates (ranging from 8 to 96), the dorsal row count, ventral row count, dorsal symphyseal count, and ventral symphyseal count are shown in colored circles on the left, warmer colors indicate higher 803 trait values. The mean tooth shape is drawn in black for each dorsal (D) and ventral (V) **position one through four adjacent to the symphysis.**

Ē.

É

 \geq

806 807

808 The first and second principal coordinate from a Principal Coordinates Analysis (PCoA) 809 of each trait suite, habitat (A), reproductive (B), somatic growth (C), trophic (D), dental 810 (E), and the overall trait matrix (F) for the NE Pacific shark regional pool. For each 811 PCoA, the percent variance explained by the principal coordinate is shown in the axis 812 labels, the coordinates of each shark species included in the analysis is shown in the

 circles colored by the percent trait coverage for each suite, and the loadings are shown in the gray arrows. Loadings are provided to show the relative variation in trait loading across suites, see Supplemental Figures 15-20 for loading labels. The scales of the coordinates are shown along the lower and left axes while the scales of the loadings are 817 shown along the right and top axes. Ecological interpretations of the PCoAs are provided above the bottom axis for PCo1, to the right of the left axis for PCo2, and

823 The functional dendrogram for the NE Pacific shark regional pool from the full trait 824 matrix (A) with the ten functional groups identified by the gap statistic and first standard 825 error max criterion denoted by each color block. The cluster means for each functional

-SCH udgw. 821 822 group are shown for each trait (B) with darker colors indicating high trait values. For the pictograph and photogrammetric silhouettes, the mean silhouette is shown. The Z-score for phylogenetic distance (PD), mean pairwise distance (MPD), and mean nearest taxon

distance (MNTD) is also shown for each cluster, warmer colors indicate under-

dispersion while cooler colors indicate overdispersion and asterisks indicate significant

835 Macroecological strata (A-E) and their clustering on the functional dendrogram (F) and 836 the phylogram (G) for the NE Pacific shark regional pool. A species membership to 837 specific (A) latitudinal strata (tropics, subtropics, temperate, and arctic), (B) habitat type

 based on ecological marine unit (EMU) classification, or (C) thermal strata based on average sea surface temperature were determined by their distribution defined by a species' 50% probability of presence. Membership to (D) carbon source gradient strata was based on a species' carbon stable isotope signature and to (E) bathymetric strata were based on a species minimum and maximum depth. The standardized effect size (Z-score) of the functional distance (FD), phylogenetic distance (PD), mean pairwise distance (MPD), and mean nearest taxon distance (MNTD) was determined from permutation using a tip-swapping null model using the functional dendrogram (F) or the 846 phylogeny (G). Significant Z-scores were denoted by asterisks ($\alpha = 0.05$). All maps are