

## **Supporting Information for**

## Constraining nonlinear time series modeling with the metabolic theory of ecology

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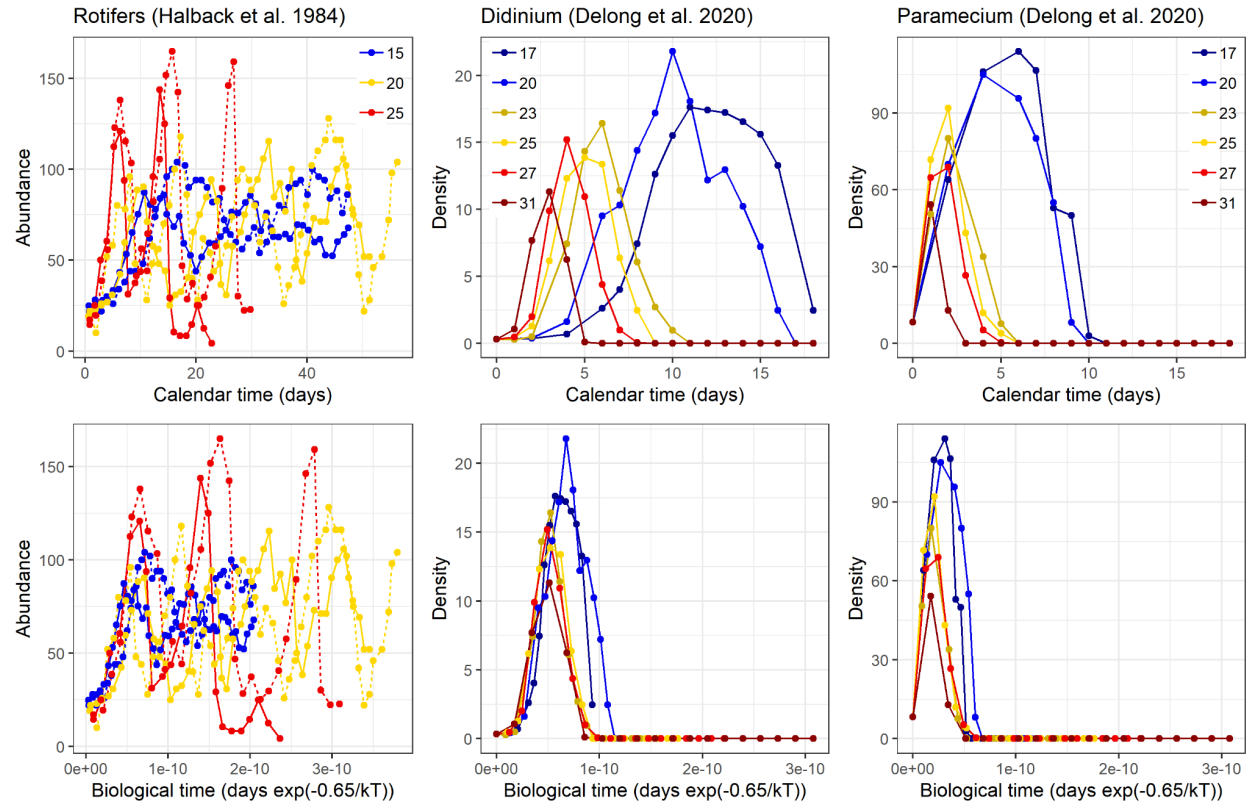
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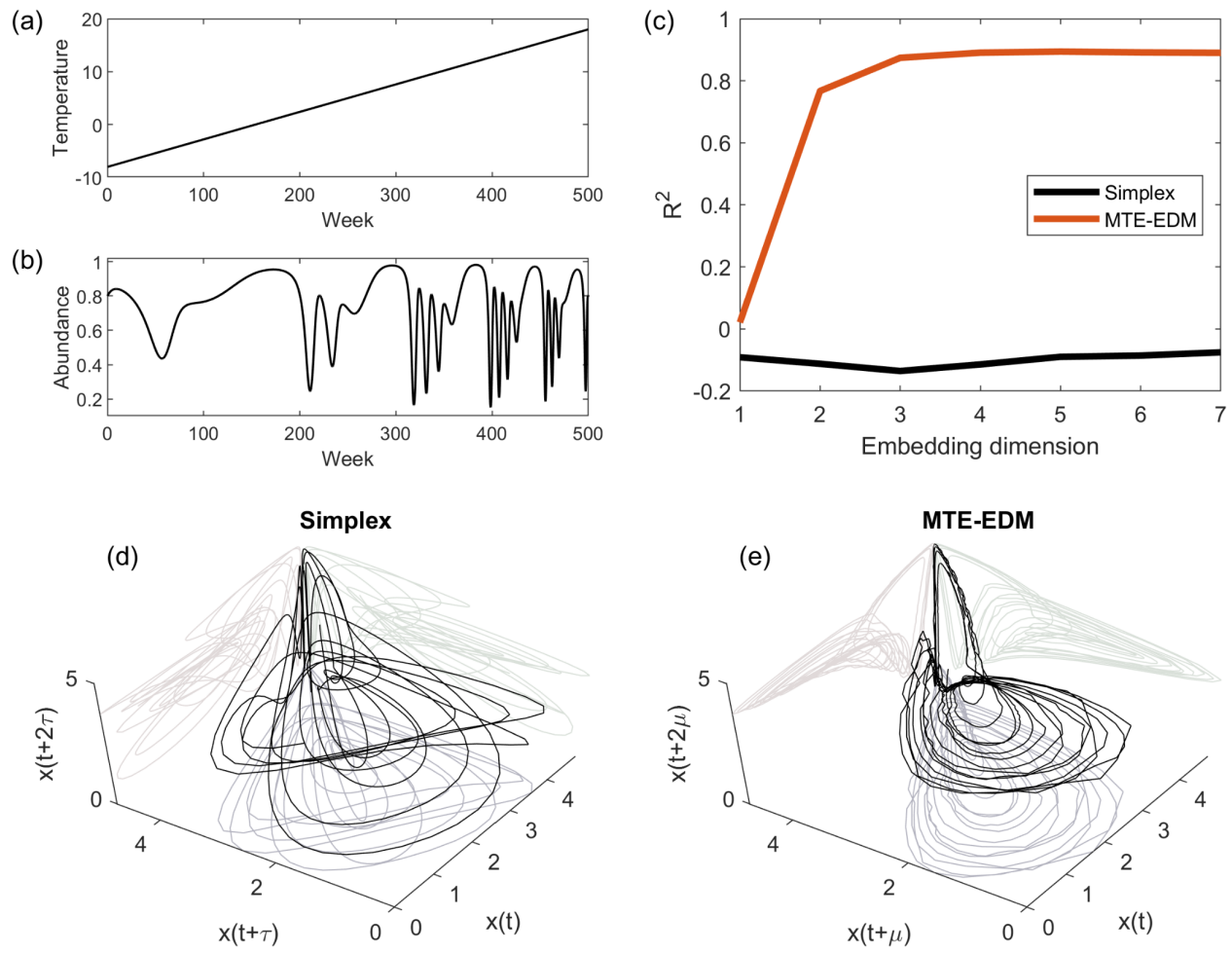
### **This PDF file includes:**

Figures S1 to S5

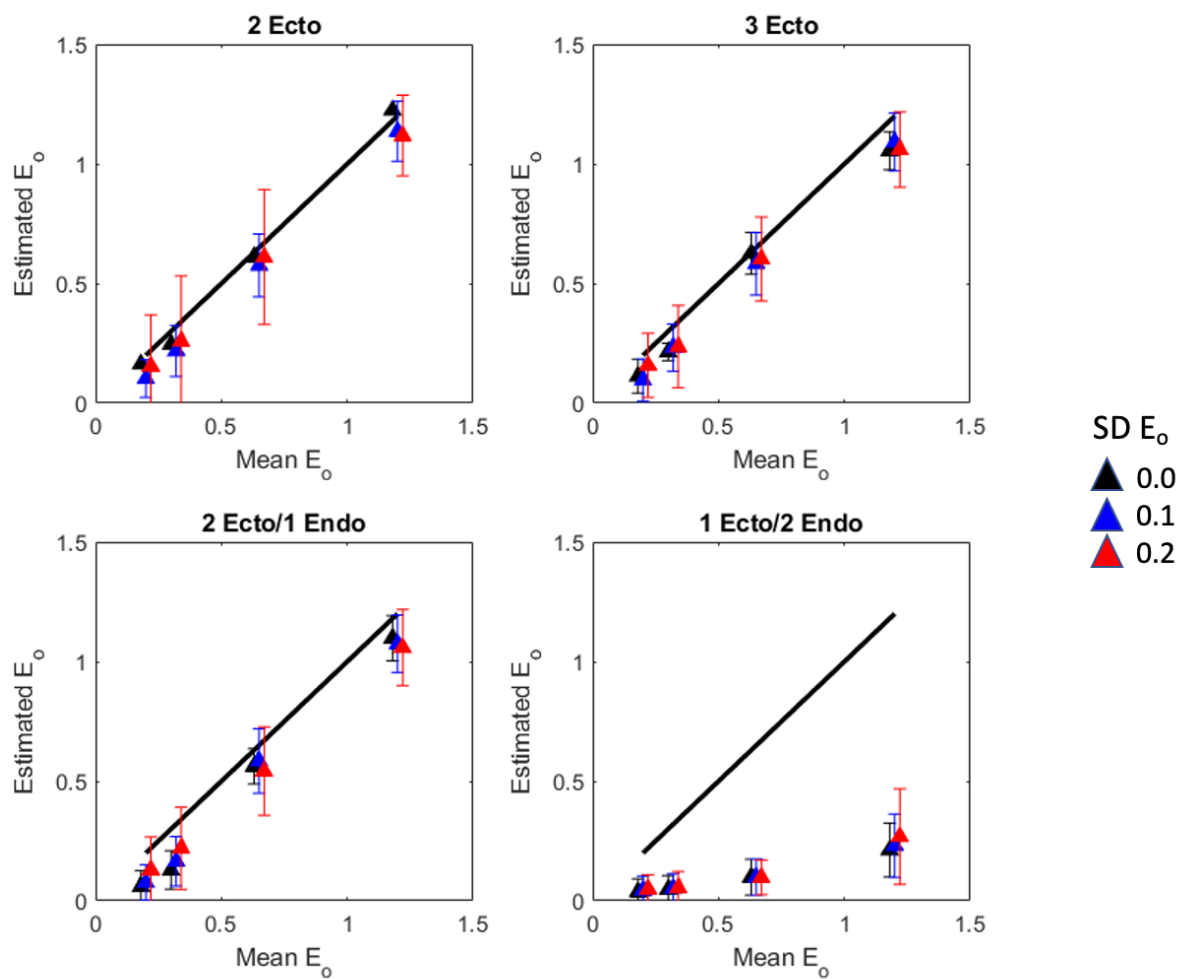
Tables S1 to S2



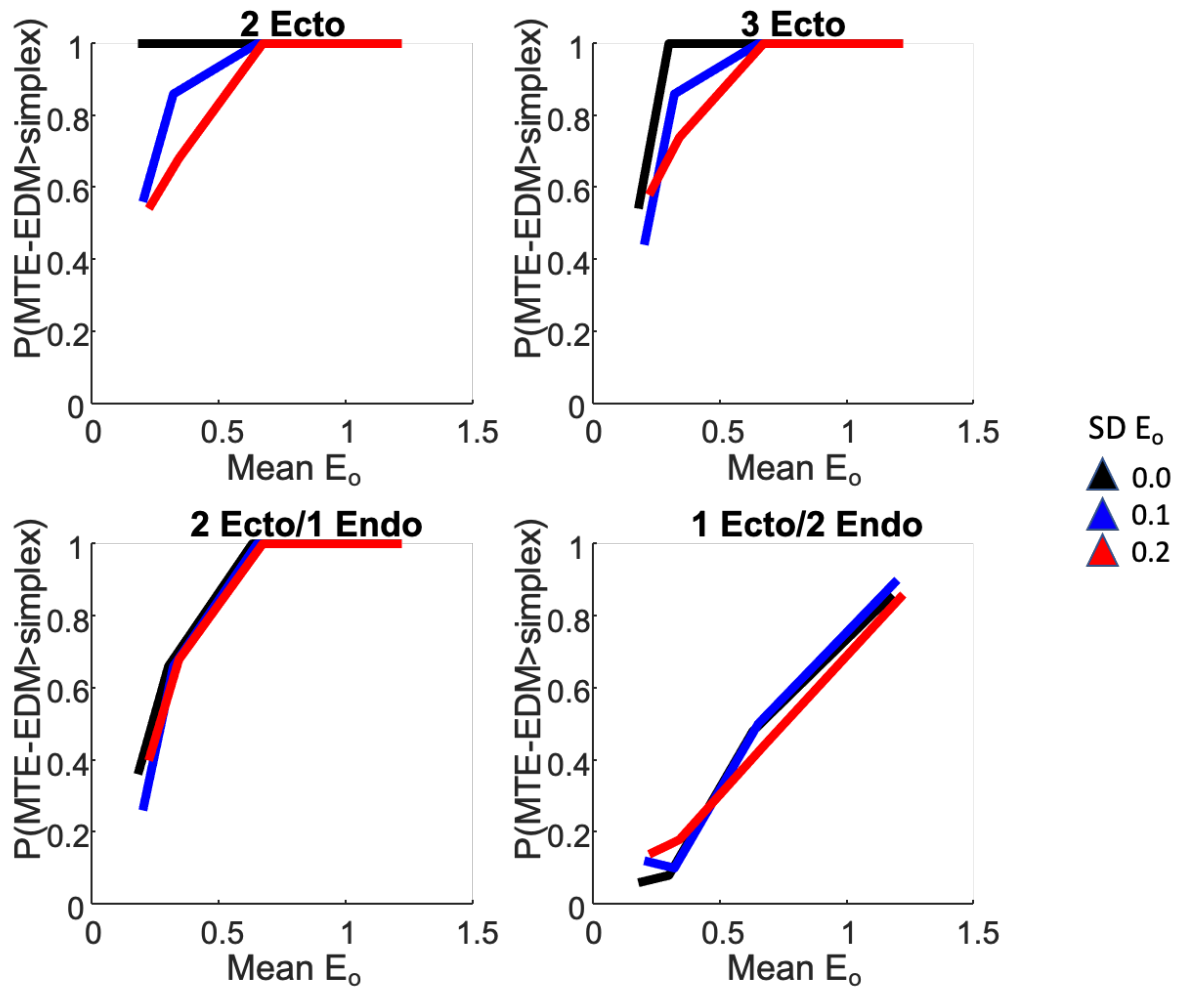
**Figure S1.** Time series from constant temperature laboratory experiments, plotted against calendar time and metabolic time.



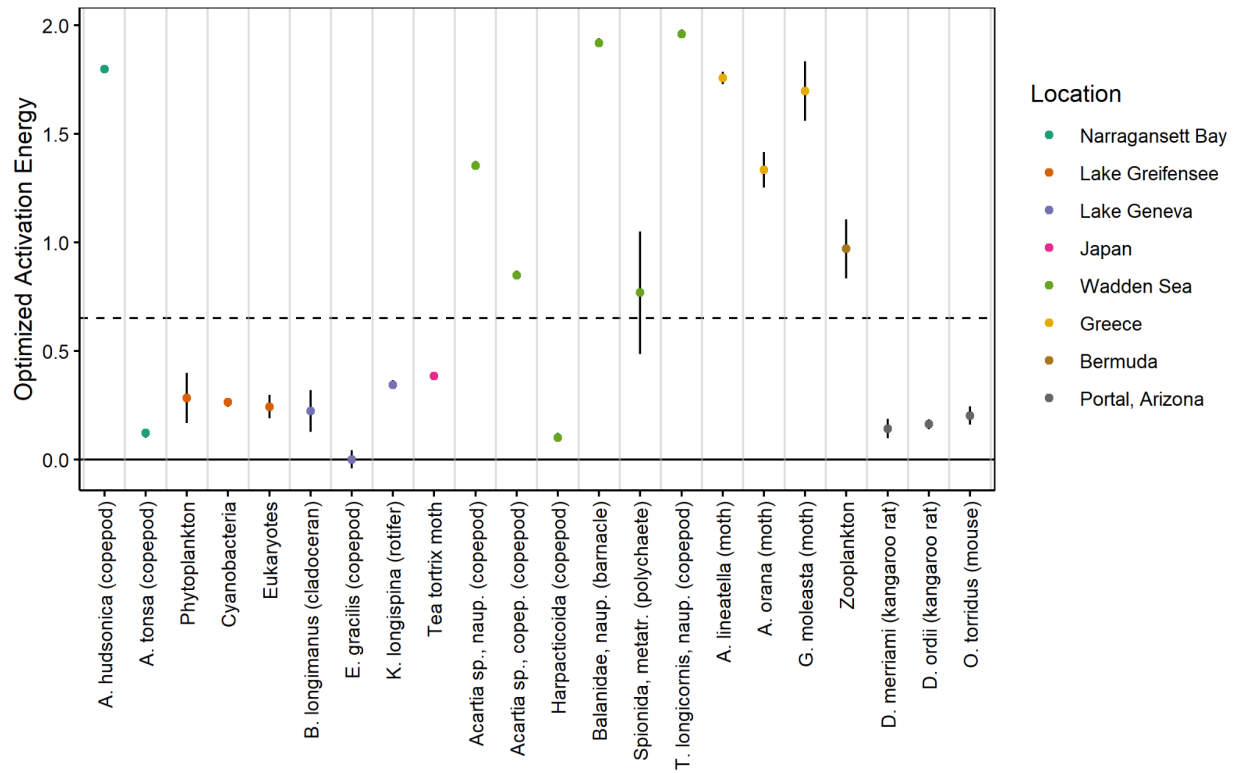
**Figure S2.** Simulated population dynamics with a linear temperature trend. Panel (a) shows temperature and (b) abundance time series. (c) Leave-one-out prediction  $R^2$  for Simplex and MTE-EDM with different embedding dimensions. (d) Reconstructed attractor in delay-coordinate space using a fixed calendar timestep or (e) using a fixed metabolic timestep. The reconstructed attractor in delay-coordinate space using a fixed metabolic timestep (e) is much closer to the true attractor, permitting more accurate forecasts.



**Figure S3:** Estimated versus true activation energy across four scenarios of communities of ecto- and endotherms: 2 ectotherms, 3 ectotherms, 2 ectotherms and 1 endotherm, and 1 ectotherm and 2 endotherms. The estimated activation energy can be recovered despite different levels of the mean activation energy ( $E_0$ ) and variation in  $E_0$  (three levels of SD shown in black, blue and red). Only in the community dominated by endotherms, the estimated activation energy is biased low.



**Figure S4:** Probability that MTE-EDM outperforms Simplex across four scenarios of communities of ecto- and endotherms: 2 ectotherms, 3 ectotherms, 2 ectotherms and 1 endotherm, and 1 ectotherm and 2 endotherms. When the community consists only of ectotherms (panels A-C), probability of significant improvement increases with the mean activation energy (i.e. greater sensitivity to temperature on average) and decreasing with variability among species. If the community is dominated by endotherms, the improvement probability increases only linearly with the mean activation energy.



**Figure S5.** Optimized activation energies for each empirical time series. Error bars are approximate standard deviations. The horizontal dashed line is the UTD value (0.65). The Simplex model (no temperature dependence) corresponds to an activation energy of 0.

**Table S1.** Results of ordinary least squares regressions of natural log-transformed cycle period vs. Arrhenius temperature. P-values for parameters are given in square brackets, 95% confidence intervals in parentheses.

| Study                   | Population                     | Slope<br>(Activation Energy)   | Intercept               | R <sup>2</sup> | N  |
|-------------------------|--------------------------------|--------------------------------|-------------------------|----------------|----|
| DeLong & Lyon (2020)    | <i>Didinium</i>                | 0.68 [0.005]**<br>(0.39,0.97)  | -24.3<br>(-35.6,-12.9)  | 0.95           | 5  |
|                         | <i>Paramecium</i>              | 0.67 [0.003]**<br>(0.37,0.97)  | -24.4<br>(-36.2,-12.7)  | 0.91           | 6  |
| Halbach (1984)          | <i>Brachionus calyciflorus</i> | 0.57 [0.011]*<br>(0.22,0.93)   | -20.2<br>(-34.3,-6)     | 0.83           | 6  |
| Fussman et al. (2014)   | <i>T. pyriformis</i>           | -0.76 [0.026]*<br>(-1.4,-0.1)  | 31.8<br>(6.3,57.3)      | 0.40           | 12 |
|                         | <i>P. fluorescens</i>          | -0.39 [0.071]<br>(-0.83, 0.04) | 18.0<br>(0.98, 34.9)    | 0.21           | 16 |
| Laughton & Knell (2019) | “Good” food                    | 0.72 [0.42]<br>(-1.28, 2.72)   | -25.2<br>(-101.7, 51.2) | 0.09           | 9  |
|                         | “Bad” food                     | -0.66 [0.46]<br>(-2.68, 1.35)  | 27.4<br>(-49.7, 104.6)  | 0.08           | 9  |

\*\* p<0.01, \* p<0.05

**Table S2.** Citations for empirical datasets

| <b>Dataset</b>           | <b>Citation<br/>(abundance)</b> | <b>Citation<br/>(temperature)</b> |
|--------------------------|---------------------------------|-----------------------------------|
| Narragansett Bay         | (1)                             | (1)                               |
| Lake Greifensee          | (2)                             | (2)                               |
| Lake Geneva              | (3)                             | (3)                               |
| Tea Tortrix Moth - Japan | (4)                             | (4)                               |
| Wadden Sea               | (5-6)                           | (7)                               |
| Greece                   | (8)                             | (8)                               |
| Bermuda                  | (9)                             | (9)                               |
| Portal, Arizona          | (10)                            | (10)                              |

(1) Smayda, T.J. & the Bunker C community (1959-1997). Narragansett Bay Plankton Time Series. Graduate School of Oceanography, URI. Data available at: NABATS.org

(2) Thomas, M.K.; Fontana, S.; Reyes, M.; Kehoe, M.; Pomati, F. (2019). Data from: The predictability of a lake phytoplankton community, over time-scales of hours to years, Dryad, Dataset, <https://doi.org/10.5061/dryad.r4454>

(3) Rimet F, Anneville O, Barbet D, Chardon C, Crépin L, Domaizon I, Dorioz J-M, Espinat L, Frossard V, Guillard J, Goulon C, Hamelet V, Hustache J-C, Jacquet S, Lainé L, Montuelle B, Perney P, Quetin P, Rasconi S, Schellenberger A, Tran-Khac V, Monet G. (2020) The Observatory on LAKes (OLA) database: Sixty years of environmental data accessible to the public. J Limnol. 79. <https://doi.org/10.4081/jlimnol.2020.1944>

(4) Nelson, W.A.; Bjornstad, O.N.; Yamanaka, T. (2013). Data from: Recurrent insect outbreaks caused by temperature-driven changes in system stability, Dryad, Dataset, <https://doi.org/10.5061/dryad.n11d4>

(5) Martens, P. (2007). Abundance of zooplankton at times series station List Reede in 1999-2006. Alfred Wegener Institute - Wadden Sea Station Sylt, PANGAEA, <https://doi.org/10.1594/PANGAEA.646280>; <https://doi.org/10.1594/PANGAEA.646281>; <https://doi.org/10.1594/PANGAEA.646282>; <https://doi.org/10.1594/PANGAEA.646283>; <https://doi.org/10.1594/PANGAEA.646284>; <https://doi.org/10.1594/PANGAEA.646285>; <https://doi.org/10.1594/PANGAEA.646286>; <https://doi.org/10.1594/PANGAEA.646287>

(6) Martens, P. (2011). Abundance of zooplankton at times series station List Reede in 2007-2008. Alfred Wegener Institute - Wadden Sea Station Sylt, PANGAEA, <https://doi.org/10.1594/PANGAEA.756061>; <https://doi.org/10.1594/PANGAEA.756062>

(7) van Beusekom, J. (2010). Hydrochemistry time series at station List Reede in 1999-2007. Alfred Wegener Institute - Wadden Sea Station Sylt, PANGAEA <https://doi.org/10.1594/PANGAEA.745142>; <https://doi.org/10.1594/PANGAEA.745143>; <https://doi.org/10.1594/PANGAEA.745144>; <https://doi.org/10.1594/PANGAEA.745145>; <https://doi.org/10.1594/PANGAEA.745146>; <https://doi.org/10.1594/PANGAEA.745147>; <https://doi.org/10.1594/PANGAEA.745148>; <https://doi.org/10.1594/PANGAEA.745149>; <https://doi.org/10.1594/PANGAEA.745150>; <https://doi.org/10.1594/PANGAEA.745151>

(8) Damos, P. (2016). Using multivariate cross correlations, Granger causality and graphical models to quantify spatiotemporal synchronization and causality between pest populations. BMC Ecology, 16, 33. <https://doi.org/10.1186/s12898-016-0087-7>

(9) Bermuda Atlantic Time Series: [http://batsftp.bios.edu/BATS/bottle/bval\\_bottle.txt](http://batsftp.bios.edu/BATS/bottle/bval_bottle.txt)

(10) S. K. Morgan Ernest, Glenda M. Yenni, Ginger Allington, Ellen K. Bledsoe, Erica M. Christensen, Renata M. Diaz, Keith Geluso, Jacob R. Goheen, Qinfeng Guo, Edward Heske, Douglas Kelt, Joan M. Meiners, Jim Munger, Carla Restrepo, Douglas A. Samson, Michele R. Schutzenhofer, Marian Skupski, Sarah R. Supp, Kate Thibault, Shawn Taylor, Ethan White, Hao Ye, Diane W. Davidson, James H. Brown, Thomas J. Valone. The Portal Project: a long-term study of a Chihuahuan desert ecosystem. bioRxiv 332783; doi: <https://doi.org/10.1101/332783>