New insights into impacts of anthropogenic nutrients on urban ecosystem processes on the Southern California coastal shelf: Introduction and synthesis

Meredith D.A. Howard^{1*}, Raphael M. Kudela², Karen McLaughlin¹

¹Southern California Coastal Water Research Project, 3535 Harbor Blvd. Suite 110, Costa Mesa, CA 92626 USA

²University of California, Santa Cruz, Ocean Science Department, 1156 High Street, Santa Cruz, CA 95064 USA

*Corresponding Author: <u>mhoward@sccwrp.org</u> 714-755-3263

1 Abstract

Anthropogenic nutrient inputs are one of the most important factors contributing to 2 eutrophication of coastal waters. Coastal upwelling regions are naturally highly variable, 3 exhibiting faster flushing and lower retention times than estuarine systems. As such, these 4 5 regions are considered more resilient to anthropogenic influences than other coastal waters. 6 Recent studies have shown our perception of the sustainability of these systems may be flawed and that anthropogenic nutrients can have an impact at local and regional spatial scales within 7 these larger upwelling ecosystems. Maintenance of an outfall pipe discharging wastewater 8 9 effluent to the Southern California Bight (SCB) provided an opportunity to study effects of anthropogenic nutrient inputs on a near-shore coastal ecosystem. The diversion of wastewater 10 effluent from a primary, offshore outfall to a secondary, near-shore outfall set up a large-scale, in 11 situ experiment allowing researchers to track the fate of wastewater plumes as they were "turned 12 off" in one area and "turned on" in another. In this introduction to a special issue, we synthesize 13 results of one such wastewater diversion conducted by the Orange County Sanitation District 14 (OCSD) during fall 2012. Anthropogenic nitrogen (N) from point-source discharges altered 15 biogeochemical cycling and the community composition of bacteria and phytoplankton. 16 17 Nitrification of ammonium to nitrate in wastewater effluent close to outfalls constituted a significant source of N utilized by the biological community that should be considered in 18 quantifying "new" production. The microbial-loop component of the plankton community played 19 20 a significant role, exemplified by a large response of heterotrophic bacteria to wastewater effluent that resulted in nutrient immobilization within the bacterial food web. This response, 21 combined with the photosynthetic inhibition of phytoplankton due to disinfection byproducts, 22 23 suppressed phytoplankton responses. Our findings have ramifications for future studies and

24	regulatory monitoring, emphasizing the need to consider chemical and biological responses to
25	wastewater effluent in assessing effects of anthropogenic nutrient inputs on urbanized coastal
26	ecosystems.
27	
28	
29	
30	Key words: wastewater; anthropogenic nutrients; microbial loop; phytoplankton;
31	biogeochemical cycle; nitrogen
32	

33 1. Introduction

Coastal upwelling regions along eastern boundary currents are among the most biologically 34 productive ecosystems in the world's ocean, supplying a significant fraction of global "new" 35 primary productivity, and representing ecologically and economically important habitats (Chavez 36 and Toggweiler, 1995; Capone and Hutchins, 2013). These regions are highly variable in time 37 38 and space, experiencing large ranges of nutrients, sea surface temperature (SST), dissolved oxygen (DO), carbon dioxide (CO₂), and pH (Chavez et al., 2003; Feely et al., 2008; Bograd et 39 al., 2008; Booth et al., 2014; Bograd et al., 2015). Some upwelling regions face increased 40 41 anthropogenic nutrient loading from publicly owned treatment works (POTWs), a potentially deleterious manifestation of human-accelerated global change (Howarth and Marino, 2006; 42 Scavia and Bricker, 2006). Despite a perception that anthropogenic nutrient inputs to these 43 regions are small compared to inputs from upwelling, recent evidence suggests such inputs may 44 significantly affect near-shore waters adjacent to urban areas (Capone and Hutchins, 2013; 45 Howard et al., 2014), with ramifications for the resilience of coastal ecosystems (Capone and 46 47 Hutchins, 2013; Bograd et al., 2015).

The Southern California Bight (SCB) is one of the most densely populated areas in the 48 49 United States, with four counties situated along the shoreline comprising nearly 25% of the 50 nation's coastal population (Culliton et al., 1990; Schiff et al., 2000). Associated with this dense urban area are large point-source nutrient inputs from POTWs, combined with dynamic 51 52 processes, such as upwelling, that impose natural variability characteristic of eastern boundary 53 currents (Chavez et al., 2003, 2009). As such, the SCB presents a suitable environment to 54 investigate responses to changes in point-source nutrient inputs. The SCB has a Mediterranean 55 climate with rainfall and runoff limited to storms occurring primarily in winter (Lyon and Stein,

56 2009). Consequently, 92% of total terrestrial N loading to coastal waters of SCB is from wastewater effluent, most of which is discharged directly into coastal waters via outfall pipes 57 (Sengupta et al., 2013). N contributions from outfall pipes are approximately equivalent to N 58 59 inputs from upwelling at spatial scales of 10s of kilometers (km), effectively doubling N loading to the shelf, altering composition of the N pool (effluent N is primarily ammonium (NH_4^+) ; 60 61 upwelled N is primarily nitrate (NO_3) , and the N:P ratio, as relatively little phosphorus is discharged in wastewater effluent (Howard et al., 2014). 62 Emerging evidence suggests that anthropogenic nutrient inputs significantly affect coastal 63 waters of SCB. The spatial extent and duration of phytoplankton blooms has increased during the 64 65 past decade, with chronic outbreaks in areas of SCB that receive anthropogenic nutrient inputs (Schnetzer et al., 2007; Nezlin et al., 2012; Schnetzer et al., 2013; Seubert et al., 2013). 66 Furthermore, observed distributions of chlorophyll a and phytoplankton in the near-shore cannot 67 68 be accounted for by upwelling alone (Kim et al., 2009; Corcoran et al., 2010; Nezlin et al., 2012; 69 Reifel et al., 2013). Accordingly, anthropogenic nutrient inputs represent an important water-70 quality consideration because stimulation of phytoplankton biomass and productivity may signal the occurrence of harmful algal blooms (HAB) and increased biomass to support respiration of 71 72 organic matter that contributes to reduced DO (Booth et al., 2014).

The Orange County Sanitation District (OCSD) is one of three large POTWs discharging secondarily treated wastewater effluent into the coastal ocean adjacent to the greater Los Angeles basin. OCSD serves a population of more than 2.6 million, treating, collecting, and disposing of sewage from two plants, with effluent delivered off Huntington Beach, California. Maintenance of the primary ocean outfall pipes provided an uncommon opportunity to study effects of wastewater effluent on near-shore coastal waters. During maintenance events, wastewater

effluent is diverted from a primary offshore outfall to a secondary near-shore outfall while the
principal outfall is inspected and repaired. Consequently, such diversions set up large-scale, *in situ* experiments that allow researchers to track the fate of wastewater plumes as they are "turned
off" in one area and "turned on" in another.

83 In this special issue, a series of papers presents the results of monitoring and research associated with the OCSD diversion during fall 2012. OCSD diverted approximately 528 x 10^{6} L 84 d⁻¹ of secondarily treated wastewater effluent for three weeks. While previous diversions were 85 conducted in SCB, OCSD had not conducted a diversion of this magnitude since 1972. The 86 Environmental Impact Report (EIR) summarized anticipated biological responses to nutrient 87 88 inputs, estimating the wastewater-effluent plume would contain up to 42 μ M NH₄⁺, potentially generating a phytoplankton bloom of 40-50 mg m⁻³ chlorophyll a (OCSD, 2011), based on 89 90 observed impacts of a diversion in Santa Monica Bay in 2006 (Reifel et al., 2013). To mitigate 91 the potential impacts of fecal indicator bacteria and other pathogens (Noble and Xu, 2004), OCSD used enhanced chlorination followed by dechlorination of the discharge, so the effect of 92 the plume would primarily be due to nutrient additions rather than intact microbial populations. 93 94 A substantial field program was coordinated to track the fate of the wastewater plume and to 95 ascertain effects of the diversion on nutrient inputs and biological responses. This 96 multidisciplinary program combined monitoring by a robotics and sensor network coupled to 97 predictive ocean modeling, field observations and experiments to assess effects of nutrient inputs 98 on biogeochemical cycling and ecosystem dynamics, and *in situ* and satellite observations to 99 identify and track plume location, transport, and mixing. This special issue summarizes our findings and discusses implications for N cycling and rate processes, and microbial responses to 100 101 inputs of wastewater effluent in an urbanized coastal region of the SCB.

102 2. Materials and Methods

103 2.1 Study site and design

Ocean outfalls for OCSD are located off the coast of Huntington Beach, California (Fig. 1). 104 From 11 September 2012 to 3 October 2012, OCSD diverted wastewater effluent from the 105 106 primary 120-inch-diameter outfall pipe located 5 miles offshore at a depth of 56 m, to a near-107 shore 78-inch diameter outfall pipe located 1 mile offshore at a depth of 16 m. Ship surveys, experiments, and modeling were combined to monitor effects of the diversion, and a variety of 108 instruments were deployed to track the plume and determine ecological effects in near-shore and 109 110 offshore environments. A robotics and sensor network included four autonomous underwater 111 vehicles, with three Teledyne Webb Slocum gliders and one REMUS, a buoyancy glider, a Liquid Robotics Surface Wave Glider, two water-quality buoys, an Environmental Sample 112 Processor (ESP), three autonomous wave-powered profiling moorings (Wirewalkers), and four 113 Acoustic Doppler Current Profilers (Fig. 1). 114

The Regional Ocean Modeling System (ROMS) used a multi-scale, three-dimensional 115 variational data assimilation method with a mesoscale atmospheric model to produce now-casts 116 every six hours and daily 72-h forecasts during the diversion (Rogowski et al., 2014). Model 117 118 outputs guided monitoring resources and ship surveys. ROMS incorporated real-time data streams, including both 2 km and 6 km HF radar surface current data, vertical profiles of 119 temperature and salinity from gliders and the MBARI M1 mooring, and SST from Advanced 120 Very High Resolution Radiometer (AVHRR), and Moderate Resolution Infrared 121 122 Spectroradiometer (MODIS) (Rogowski et al., 2014). Observational data were not assimilated 123 into now-casts and forecasts because they were not available in real time. Rogowski et al. (2014) 124 provides a detailed description of modeling efforts during and after the diversion study.

125 2.2 Shipboard Surveys

126 Shipboard surveys covered pre-, during, and post-diversion intervals, as described by Caron et al. (this issue) and summarized here. Weekly "plume-tracking" cruises aboard the M/V Nerissa 127 128 (OCSD) focused on water quality and enumeration of bacteria and phytoplankton for stations in 129 the vicinity of the discharge and at the long outfall pipe. An adaptive sampling design was used to position stations up-coast and down-coast, depending on daily current predictions from 130 131 ROMS, assuring sampling at a subset of stations in close proximity to the discharge. The sampling zones occupied on these regulatory-based surveys were defined in the California Ocean 132 Plan. 133

Corresponding "event" cruises were conducted weekly aboard the R/V Yellowfin (Southern 134 135 California Marine Institute) or the *M/V Nerissa*, occupying five stations on an onshore - offshore transect to document near-shore effects of wastewater effluent at the short outfall, and changes at 136 the long outfall resulting from cessation of discharge. A "regional" cruise evaluated 137 138 phytoplankton responses over a larger geographical area (Caron et al., this issue). Discrete 139 sampling and experiments included characterization of spatial and temporal changes in nutrient 140 concentrations, shifts in community composition of bacteria and phytoplankton, N cycling, and 141 isotopic-source tracking to determine the extent of wastewater effluent effects. A number of 142 experiments focused on phytoplankton responses to wastewater effluent and other nutrients, 143 including N-preference and NH₄⁺-inhibition. A retrospective analysis relied on satellite data from 144 multiple sensors to distinguish biophysical responses from the diverted effluent (Gierach et al., this issue). 145

146 **3. Results summary**

147 *3.1. Plume tracking*

Multiple approaches were used during the 2012 OCSD diversion to overcome technical 148 149 challenges associated with plume tracking, such as locating and monitoring the plume in near 150 real-time. Technologies included wave-powered profiling moorings (Wirewalkers) (Lucas and 151 Kudela, this issue; Kudela et al., this issue), buoys and moorings including bottom-mounted Acoustic Doppler Current Profilers (ADCP), two ocean buoys and an Environmental Sample 152 Processor (Caron et al., this issue; Lucas and Kudela, this issue), autonomous underwater 153 154 vehicles (AUVs) such as Teledyne Slocum gliders and a Liquid Robotics Waveglider (Seegers et al., this issue), and remote sensing (Gierach et al., this issue). From these observations, a 155 consistent plume signature was identified, which allowed for plume tracking and estimation of 156 157 plume dilution and dispersion.

158 Prior to the wastewater-effluent diversion, we expected the OCSD plume would be surfacetrapped, given the volume of discharge and shallow release depth. Consistent with this 159 160 expectation, visual observations confirmed the presence of a surface signature (Fig. 2). Vertical 161 profiles, however, revealed a complex mixing pattern, with plume water interacting with both 162 surface and bottom layers (Lucas and Kudela, this issue). This led to the presence of plume water 163 in both near-field (mixing between the effluent and ambient waters) and far-field (mixing where dilution occurs primarily with waters of the same density), and a significant component in the 164 165 subsurface (Lucas and Kudela, this issue).

Plume tracking by Lagrangian drifters showed plume waters mainly traveled in onshore and alongshore directions (Rogowski et al., 2014). Satellite imagery also proved useful to track the plume and biological responses (Gierach et al., this issue), revealing patterns consistent with

169 Lagrangian drifters. Gierach et al. (this issue) estimated that chlorophyll a and SST must vary by 1 mg m⁻³ and 0.5 °C, respectively in order to successfully identify and track the plume using 170 satellite imagery, while synthetic aperture radar (SAR) was useful when wind speeds were 3 to 8 171 m s⁻¹. Glider profiles (Seegers et al., this issue) suggested most of the plume was entrained in the 172 upper 20 m, but with a significant plume signature extending as deep as 40 m. Together, these 173 174 observations provide a consistent view of plume dynamics, with waters in the plume characterized by low SST, low salinity, and elevated chromophoric dissolved organic matter 175 (CDOM). The plume was predominantly surface-trapped and maintained in the upper ~ 20 m for 176 177 significant periods (days) in the vicinity of the outfall. In situ observations confirmed elevated CDOM in the plume as detected by satellites (Seegers et al., this issue; Gierach et al. this issue). 178 The distribution of the plume also agreed well with modeling outputs for plume dispersal 179 180 (Uchiyama et al., 2014).

181 Dilution of the plume was more difficult to estimate. We expected the plume would be diluted 30:1 in the near-field, primarily due to the design of diffusers. Data from Lagrangian 182 183 drifters showed dilution > 100:1 within 1 km of the outfall pipe (Rogowski et al., 2014). Nutrient 184 distributions were consistent with at least 100:1 dilution within the region (Caron et al., this issue), but surface mapping showed water of low salinity, and low variable fluorescence 185 occurred several km from the outfall (Kudela et al., this issue). These observations can be 186 reconciled by taking into account the subsurface lensing observed by Lucas and Kudela (this 187 188 issue). While the bulk of the plume was surface-trapped and rapidly diluted, a significant fraction 189 of wastewater effluent was likely entrained in intermediate layers, persisting until dissipated by 190 intermediate- and far-field processes, such as wind-driven mixing and internal waves. Weak 191 transport and rapid mixing resulted in a plume-influenced region extending several km around

the outfall pipe, and remaining in the general vicinity of the outfall pipe for several days (Lucas
and Kudela, this issue). This was sufficiently long for biological responses to develop,
exemplified by heterotrophic bacteria (Caron et al., this issue), and phytoplankton growth and
photosynthesis (Kudela et al., this issue).

196 The OCSD diversion was studied with intensive observations and modeling. It is unlikely 197 that a comparable study could be maintained by most management entities, highlighting the need 198 for a cooperative use of advanced technologies. Notably, results from gliders, drifters, shipboard observations, and satellite retrievals converged on similar estimates of dilution, while high-199 resolution vertical observations from Wirewalkers and spatial transects of properties such as 200 201 variable fluorescence identified more concentrated layers and lensing effects. This suggests that routine monitoring at the meso-scale using multiple observational approaches would capture the 202 203 bulk oceanographic responses to nutrient inputs, but could miss small-scale effects.

204 *3.2. Biological responses*

205 Biological responses during the OCSD diversion in 2012 differed from those in previous 206 studies. Prior to the OCSD diversion, a much smaller diversion was conducted from the 207 Hyperion Wastewater Treatment Plant in Santa Monica Bay, California for three days in 2006. Also, there was a major breach in San Diego, California in 1992, resulting in discharge of 208 209 untreated effluent for several months. Although information in the published literature about the 210 San Diego event is sparse, dinoflagellate blooms were reported around the periphery of an 211 effluent plume that was visible from airborne platforms (Casey et al., 1992; Tegner et al., 1995; Lange et al., unpublished data). The March 1993 issue of the journal, *Photogrammetric* 212 213 Engineering and Remote Sensing featured the San Diego breach on the cover, as reproduced here 214 (Fig. 3). The 2006 diversion in Santa Monica Bay produced dramatic biological responses,

215	including increases of potentially harmful dinoflagellates, Lingulodinium polyedrum and
216	Cochlodinium sp., subsequently identified as C. fulvescens (Howard et al., 2012) and Akashiwo
217	sanguinea (Reifel et al., 2013).
218	Given this historical context and anticipated impacts of the OCSD diversion described in the
219	EIR (OCSD, 2011), we anticipated a large phytoplankton response to nutrient inputs during the
220	2012 diversion. The EIR predicted a persistent phytoplankton bloom with chlorophyll a of 40-50
221	mg m ⁻³ in response to NH_4^+ in the discharge effluent (OCSD, 2011). But contrary to
222	expectations, chlorophyll a remained low throughout the diversion (Caron et al., this issue;
223	Kudela and Lucas, this issue; Kudela et al., this issue; Seegers et al., this issue). So what actually
224	happened following the diversion? Caron et al. (this issue) showed responses by various fractions
225	of the planktonic community, including counts of phytoplankton, microzooplankton,
226	picoeukaryotes, cyanobacteria, and heterotrophic bacteria. Overall, phytoplankton composition
227	shifted slightly during the diversion, with increased cyanobacteria and phototrophic,
228	picoplanktonic eukaryotes. In contrast to the Hyperion diversion in Santa Monica Bay, there was
229	little to no increase of potentially harmful dinoflagellates, despite the longer diversion (three
230	weeks vs. three days). While background concentrations of the toxic diatom, Pseudo-nitzschia
231	spp. were detected, Caron et al. (this issue) measured relatively low concentrations of domoic
232	acid ($\leq 0.05 \ \mu g \ L^{-1}$) associated with this genus. Total phytoplankton biomass as chlorophyll <i>a</i>
233	remained low throughout the diversion ($< 5 \text{ mg m}^{-3}$), while bacterial biomass and microbial
234	grazers showed strong responses, with heterotrophic bacteria increasing by an order-of-
235	magnitude at mid-diversion (Caron et al., this issue). There was some evidence of increased
236	chlorophyll a toward the end of the diversion (Lucas and Kudela, this issue), and subsurface

observations from gliders (Seegers et al., this issue) corroborated a moderate increase ofphytoplankton biomass at depth, although stimulation of a bloom did not occur.

Experimental manipulations using enclosures showed the phytoplankton community was 239 240 physiologically capable of using nutrients in the wastewater effluent (Seubert et al., this issue; 241 Kudela et al., this issue). A "worst-case scenario" of 1:10 dilution resulted in substantial growth (Seubert et al., this issue), while lower additions of effluent (such as 1:100 or 1:1000) or various 242 combinations of nutrients supported robust growth rates, often exceeding 1 division day⁻¹ 243 (Kudela et al., this issue). NH₄⁺ inhibition has been suggested to affect growth and productivity 244 of some phytoplankton taxa (Dugdale et al., 2006), particularly diatoms, but high NH₄⁺ 245 246 concentrations sufficient to be inhibitory were not detected (Seubert et al., this issue; Kudela et al., this issue). The results from N uptake kinetics experiments showed no obvious physiological 247 impairment attributed solely to N concentrations; NH_4^+ suppression of NO_3^- uptake was not 248 249 sufficient to account for the lack of biological response, and there was physiological capacity to respond to ambient N substrates (Kudela et al., this issue). 250 251 To explain the lack of phytoplankton stimulation despite an apparent physiological capacity

252 for a bloom to develop, Kudela et al. (this issue) considered effects of chlorination and 253 subsequent dechlorination, and the production of inhibitory disinfection by-products (DBP). 254 Wastewater effluent contained hypochlorite residuals above regulatory requirements, thereby 255 documenting over-chlorination. Kudela et al. (this issue) found that effluent treated with 256 hypochlorite that was incompletely neutralized with bisulfite led to a significant inhibition of phytoplankton photosynthesis lasting ~24 h, with suppressed growth continuing for at least 72 h. 257 Analogous inhibition of heterotrophic bacteria was not observed, consistent with the findings of 258 259 Caron et al. (this issue) and Seubert et al. (this issue). These results suggest DBP may have

precluded development of a phytoplankton bloom while allowing a strong response by the
bacterial assemblage. Caron et al. (this issue) propose that the presence of DBP on the
continental shelf for several days (Kudela et al., this issue; Lucas et al., this issue) was sufficient
to depress phytoplankton and stimulate bacteria, resulting in nutrient immobilization in the
bacterial food web, and thereby suppressing stimulation of a phytoplankton bloom that had been
anticipated prior to the diversion.

266 *3.3. Biogeochemical cycling*

Wastewater effluent in ocean outfalls represents a large point source of NH₄⁺; therefore, high 267 rates of NH₄⁺-oxidation were expected to occur near the outfall with ramifications for N cycling. 268 McLaughlin et al. (this issue) hypothesized NH₄⁺ in effluent would sustain high nitrification 269 rates, i.e., the sequential oxidation of NH_4^+ to NO_3^- via nitrite (NO_2^-) near the long-outfall pipe 270 during normal operations, and would be reduced to background rates when the pipe was out of 271 272 operation. The authors also expected effluent N to be incorporated into biomass of bacteria and phytoplankton around outfall pipes. The expected decrease of nitrification at the long-outfall 273 pipe, measured in bottle incubations as accumulation of ¹⁵N in the dissolved NO₃⁻ pool following 274 addition of isotopically enriched ¹⁵NH₄⁺, was observed following the diversion to the short 275 outfall, and "background" nitrification rates were consistent with those previously reported for 276 277 SCB (Ward, 1987). Furthermore, decreased nitrification at the long outfall was consistent with decreased NH₄⁺ emission during the diversion. 278

The importance of nitrification to biogeochemical cycling of N around the OCSD outfall was substantiated by nutrient concentrations revealing a "hot spot" of NH_4^+ and NO_2^- over the ocean outfall pipe under normal operations. The dual isotopic composition of dissolved NO_3^- ($\delta^{15}N_{NO3}$ and $\delta^{18}O_{NO3}$) indicated N-assimilation and denitrification rates were low relative to nitrification,

consistent with low phytoplankton biomass observed during the study period (Caron et al., this 283 284 issue; Seegers et al., this issue), possibly caused by DBP in wastewater effluent (Kudela et al., this issue). The isotopic composition of suspended particulate matter indicated low $\delta^{15}N_{PN}$ and 285 $\delta^{13}C_{PN}$ values around the outfall under normal operations, suggesting incorporation of "nitrified" 286 (low ¹⁵N NO₃⁻) and wastewater dissolved organic carbon into biomass. These results suggest that 287 288 effluent altered N cycling in close proximity to the outfall (McLaughlin et al., this issue). An implication of these findings is that rapid oxidation of effluent NH₄⁺ around outfalls contributes 289 significantly to the pool of NO₃⁻ in urban coastal areas. This unconventional source of NO₃⁻ in 290 the context of "new" production (sensu Dugdale and Goering, 1967) should be considered in 291 distinguishing upwelled and regenerated NO₃⁻ to support primary productivity. Thus, both NH₄⁺ 292 and transformed NO₃⁻ represent new sources of N. Resulting changes in the proportions of N 293 294 forms have implications for the community composition of phytoplankton, as preferences for specific N forms differ among taxonomic groups (Dortch and Conway, 1984; Dortch, 1990; 295 Kudela and Cochlan, 2000; Dugdale et al., 2007; Kudela et al., 2008; Collos and Harrison, 296 297 2014).

298 **4.** Conclusions

Eutrophication of coastal waters associated with anthropogenic nutrient inputs is welldocumented (Howarth, 2008; Paerl and Piehler, 2008), but is often overlooked for coastal upwelling regions. Several insights emerged from the diversion described here with implications for coastal ocean management. First, results indicate anthropogenic N inputs from point-source ocean outfalls impact biogeochemical processes, community composition of phytoplankton and bacteria, phytoplankton physiology, and N cycling within the vicinity of the outfall. Decreased nitrification rates at the long outfall during the diversion indicate that wastewater effluent altered

306 N cycling in close proximity to the outfalls. The geographic extent of this effect needs to be 307 characterized in future studies to assess if management actions to mitigate these effects are warranted. Second, rapid oxidation of effluent NH_4^+ to NO_3^- in close proximity to the outfalls 308 309 represents a significant source of "new" N to the biological community that needs to be 310 considered in quantifying biomass and productivity of phytoplankton in near-shore waters. 311 Changes in the inventory, form, and biogeochemical cycling of N affect the phytoplankton community composition and competition among algal taxa (Dortch and Conway, 1984; Dortch, 312 1990, Dugdale et al., 2007; Collos and Harrison, 2014) and development of phytoplankton 313 314 blooms (Kudela and Cochlan, 2000; Glibert et al., 2005; Beman et al., 2005; Glibert et al., 2006; 315 Howard et al., 2007; Cochlan et al., 2008; Heisler et al., 2008; Kudela et al., 2008), as N is the primary macronutrient limiting phytoplankton in coastal waters (Dugdale, 1967; Ryther and 316 317 Dunstan, 1971; Eppley et al., 1979; Howarth and Marino, 2006). Third, wastewater effluent elicited significant changes in microbial community composition, evident in a muted 318 319 phytoplankton response and large increase in abundance of heterotrophic bacteria, suggesting the 320 microbial loop plays a significant, perhaps under-appreciated role in urbanized coastal waters of 321 SCB. Finally, a convergence of dilution estimates from multiple observational modes highlights 322 the ability to track the bulk transport of effluent plumes, while observations of lensing and small patches of less-diluted effluent highlights the need for high-resolution sampling to quantify 323 spatial and temporal variability. 324

Our findings indicate wastewater effluent alters N cycling and processes and microbial community composition in the nearshore urbanized coastal waters of the SCB. Studies from the Hyperion Wastewater Treatment Plant diversion in Santa Monica Bay in 2006 suggested regulatory implementation of nutrient reductions to avoid bloom development and minimize

anthropogenic impacts (Reifel et al. 2013). However, our results do not support regulatory
changes at this time, as too few studies have been completed and the spatial extent of the relative
impact is currently unknown. Instead, we suggest additional studies that will determine the
spatial extent of the anthropogenic impacts and use modeling and scenario analysis to determine
if nutrient reductions will significantly reduce these impacts.

Routine regulatory requirements for monitoring point-source discharges through the National 334 335 Pollutant Discharge Elimination System (NPDES) permits for ocean monitoring of large POTW discharges in SCB do not include measurements of nutrients, biological productivity, or chemical 336 processes. Additionally, NPDES permit requirements for monitoring effluent differ amongst the 337 338 four large POTW permits in SCB. Given that wastewater effluent may constitute nutrient inputs comparable to those from coastal upwelling (Howard et al., 2014), and that wastewater effluent 339 340 influences N cycling in close proximity to outfalls (McLaughlin et al., this issue), a 341 comprehensive record of the total N loading that includes different forms of N would be 342 beneficial. Current NPDES requirements are not focused on nutrients; thus, regulatory 343 monitoring programs do not provide comprehensive data to evaluate long-term ecological changes associated with point-source nutrient discharges. These parameters, however, are critical 344 345 for identifying the impacts from point source discharges on coastal ocean ecosystems. Therefore, 346 we suggest more nutrient focused regulatory monitoring requirements for both effluent and receiving waters that should be incorporated into NPDES monitoring requirement programs. 347 348 Mitigation of pathogens by OCSD's enhanced chlorination and dechlorination of discharge proved successful (Rogowski et al., 2014), but had the unintended consequence of inhibiting 349 phytoplankton biomass and productivity, and possibly preventing development of a harmful algal 350 351 bloom, such as occurred in Santa Monica Bay following a shorter diversion. Future diversions

and normal operations of POTWs should consider possible effects of DBP on the biota, as
monitoring of residual chloride may not fully capture potential impacts on the biological
community.

The multi-dimensional approach to monitoring developed for this study provided several 355 356 important points for future diversions and/or management scenarios that require enhanced or extensive monitoring. First, now-casts and forecasts from ROMS combined with real-time 357 358 observations provided a comprehensive overview of oceanic conditions independent of sporadic in situ and remote sensing data. Accordingly, ROMS outputs were critical to management 359 decisions and to guide adaptive sampling. Second, satellite observations supported detection of 360 361 the wastewater plume over a larger spatial scale than would have been possible with in situ observations alone, but with some limitations due to cloud cover that affected availability of 362 363 imagery. We recommend the expenditure of effort to develop metrics to use satellite data for 364 tracking plumes of wastewater effluent and quantifying phytoplankton responses, as this approach offers important advantages in spatial and temporal resolution. A more comprehensive 365 assessment of point-source nutrient discharge for the SCB or other regions could also benefit 366 from an Observing System Simulation Experiment (OSSE) framework, which would allow for a 367 368 value-of-information approach to identify the most effective monitoring tools, and also assess 369 new technologies as they become available. For the SCB, this approach is not warranted at this time given the infrequent timing of diversions (decades) and the current lack of a regulatory 370 371 requirement for more sophisticated monitoring, but consideration should be given to this 372 recommendation for other locales or for the SCB if agencies are required in the future to take a 373 more holistic approach to wastewater management.

374

375 Acknowledgments

- We thank OCSD for in-kind support, and the captains and crews of the R/V Nerissa and R/V
- 377 Yellowfin. George Robertson (OCSD) was instrumental in making this project possible. This
- 378 work was supported by the National Science Foundation (NSF) RAPID award OCE1251573
- 379 (RMK), National Oceanic and Atmospheric Administration (NOAA) ECOHAB award
- 380 NA11NOS4780030 (RMK and MDH), and the Southern California Coastal Water Research
- 381 Project Authority (SCCWRP). This is NOAA ECOHAB publication # 819. We thank Abel
- 382 Santana (SCCWRP) for creation of the maps, Jeff Myers (NASA Ames Research Center) for the
- 383 San Diego image, and Mike McCarthy and Michael Mengel (OCSD) for photographs of the
- 384 surfacing effluent.

385 **References**

- Beman, J.M., Arrigo, K.R., Matson, P.A., 2005. Agricultural runoff fuels large phytoplankton
 blooms in vulnerable areas of the ocean. Nature 434, 211-214.
- Bograd, S.J., Castro, C.G., DiLorenzo, E., Palacios, D.M., Bailey, H., Gilly, W. Chavez, F.P.,
 2008. Oxygen declines and the shoaling of the hypoxic boundary in the California Current.
 Geophys. Res. Lett. 35, L12607.
- Bograd, S.J., Buil, M.P., DiLorenzo, E., Castro, C.G., Schroeder, I.D., Goericke, R., Anderson,
 C.R., Benitez-Nelson, C., Whitney, F.A., 2015. Changes in source waters to the Southern
 California Bight. Deep Sea Research Part II: Topical Studies in Oceanography 112, 42-52.
- Booth, A.T., Sutula, M., Micheli, F., Weisberg, S.B., Bograd, S.J., Steele, A., Schoen, J.,
 Crowder, L.B., 2014. Patterns and potential drivers of declining oxygen content along the
 southern California coast. Limnol. Oceanogr. 59, 1127-1138.
- Capone, D.G., Hutchins, D.A., 2013. Microbial biogeochemistry of coastal upwelling regimes in
 a changing ocean. Nature Geosci. 6, 711-717.
- Caron, D.A., Gellene, A.G., Smith, J., Seubert, E.L., Campbell, V. Sukhatme, G.S., Seegers, B.,
 Jones, B.H., Howard, M.D.A., Kudela, R.M., Hayashi, K., Ryan, J., Birch, J., Demir-Hilton,
 E., Yamahara, K., Scholin, C., Mengel, M., Robertson, G., 2016. Response of the
 phytoplankton and bacterial communities during a wastewater effluent diversion into
 nearshore coastal waters. Estuar. Coast. Shelf Sci. (accepted).
- Casey, R. Ciccateri, A., Dougherty, K., Gacek, L., Lane, S., Liponi, K., Leeds, R., Walsh, F.,
 1992. Oceanographic effects of the 1992 Point Loma sewage pipe spill. Proc., Annual Mtg.
 Geol. Soc. America 24, 26-29.
- 407 Chavez, F., Toggweiler, J.R.T., 1995. Physical estimates of global new production: The
- 408 upwelling contribution. In: Summerhayes, C.P., Emeis, K.C., Angel, M.V., Smith, R.L.,
- Zeitzschel, B. [Eds.], Upwelling in the oceans: Modern Processes and Ancient Records.
 Wiley, pp. 149–169.
- Chavez, F., Smith, J., Lluch-Cota, S.E., Niquen, M.C., 2003. From anchovies to sardines and
 back: multidecadal change in the Pacific Ocean. Science 299, 217-221.
- Chavez, F.P., Messie, M., 2009. A comparison of Eastern Boundary Upwelling Ecosystems.
 Prog. Oceanogr. 83, 80-96.
- Collos, Y., Harrison, P.J., 2014. Acclimation and toxicity of high ammonium concentrations to
 unicellular algae. Mar. Poll. Bull. 80, 8-23.
- Corcoran, A.A., Reifel, K.M., Jones, B.H., Shipe, R.F., 2010. Spatiotemporal development of
 physical, chemical, and biological characteristics of stormwater plumes in Santa Monica Bay,
 California (USA). J. Sea Res. 63, 129-142.
- 420 Culliton, T., Warren, M., Goodspeed, T., Remer, D., Blackwell, C., McDonough III, J., 1990.
- 421 Fifty Years of Population Changes Along the Nation's Coasts. Coastal Trends Series, Report
- 422 No. 2, National Oceanic and Atmospheric Administration, Strategic Assessment Branch.
- 423 Rockville, MD.

- 424 Dortch, Q., 1990. The interaction between ammonium and nitrate uptake in phytoplankton. Mar.
 425 Ecol. Prog. Ser. 61, 183-201.
- 426 Dortch, Q., Conway, H.L., 1984. Interactions between nitrate and ammonium uptake: variation
 427 with growth rate, nitrogen source and species. Mar. Biol. 79, 151-164.
- 428 Dugdale, R.C., 1967. Nutrient limitation in the sea: Dynamics, identification and significance.
 429 Limnol. Oceanogr. 12, 685-695.
- Dugdale, R.C., Goering, J.J., 1967. Uptake of new and regenerated forms of nitrogen in primary
 productivity. Limnol. Oceanogr. 12, 196-206.
- Dugdale, R.C., Wilkerson, F.P., Hogue, V.E., Marchi, A., 2006. Nutrient controls on new
 production in the Bodega Bay, California, coastal upwelling plume. Deep-Sea Research II
 53, 3049-3062.
- 435 Dugdale, R., Wilkerson, F., Hogue, V., Marchi, A., 2007. The role of ammonium and nitrate in
 436 spring bloom development in San Francisco Bay. Estuar. Coast. Shelf Sci. 73, 17-29.
- Eppley, R., Renger, E., Harrison, W., 1979. Nitrate and phytoplankton production in California
 coastal waters. Limnol. Oceanogr. 24, 483-494.
- Feely, R.A., Sabine, C.L., Hernandez-Ayon, J.M., Ianson, D., Hales, B., 2008. Evidence for
 upwelling of corrosive "acidified" water onto the continental shelf. Science 320, 1490.
- Gierach, M.M., Holt, B., Trinh, R., Pan, B., Rains, C., 2016. Satellite detection of wastewater
 diversion plumes in Southern California. Estuar. Coast. Shelf Sci. (accepted).
- Glibert, P., Seitzinger, S., Heil, C.A., Burkholder, J.M., Parrow, M.W., Codispoti, L.A., Kelly,
 V., 2005. The role of eutrophication in the global proliferation of harmful algal blooms.
 Oceanography 18, 198-209.
- Howard, M.D.A., Jones, A.C., Schnetzer, A., Countway, P.D., Tomas, C.R., Kudela, R.M.,
 Hayashi, K., Chia, P., Caron, D.A., 2012. Quantitative real-time PCR for *Cochlodinium fulvescens* (Dinophyceae), a potentially harmful dinoflagellate from California coastal
 waters. J. Phycol. 48, 384-393.
- Howard, M.D.A., Sutula, M., Caron, D.A., Chao, Y., Farrara, J.D., Frenzel, H., Jones, B.,
 Robertson, G., McLaughlin, K., Sengupta, A. 2014. Anthropogenic nutrient sources rival
 natural sources on small scales in the coastal waters of the Southern California Bight.
 Limnol. Oceanogr. 99, 285-297.
- Howarth, R.W., Marino, R., 2006. Nitrogen as the limiting nutrient for eutrophication in coastal
 marine ecosystems: Evolving views over three decades. Limnol. Oceanogr. 51, 364-376.
- Howarth, R.W., 2008. Coastal nitrogen pollution: A review of sources and trends globally and
 regionally. Harmful Algae 8, 14-20.
- Kim, H. J., A. J. Miller, J. McGowan, and M. L. Carter. 2009. Coastal phytoplankton blooms in
 the Southern California Bight. Prog. Oceanogr. 82, 137-147,
 doi:10.1016/j.pocean.2009.05.002
- Kudela, R.M., Howard, M.D.A., Hayashi, K., Beck, C., 2016. Evaluation of ammonium
 inhibition and nitrogen uptake kinetics during a wastewater diversion into nearshore coastal
 waters in Southern California. Estuar. Coast. Shelf Sci. (submitted).

- Kudela, R.M., Lucas, A.J., Negrey, K.H., Howard, M., McLaughlin, K., 2016. Death from
 below: Investigation of inhibitory factors in bloom development during a wastewater effluent
 diversion. Estuar. Coast. Shelf Sci. (accepted).
- Kudela, R.M., Cochlan, W.P., 2000. The kinetics of nitrogen and carbon uptake and the
 influence of irradiance for a natural population of *Lingulodinium polyedrum* (Pyrrophyta) off
 Southern California. Aquat. Microbial Ecol. 21, 31-47.
- Kudela, R.M., Lane, J.Q., Cochlan, W.P., 2008. The potential role of anthropogenically derived
 nitrogen in the growth of harmful algae in California, USA. Harmful Algae 8, 103-110.
- 472 Lyon, G. S., Stein, E.D., 2009. How effective has the Clean Water Act been at reducing pollutant
 473 mass emissions to the Southern California Bight over the past 35 years? Environ. Mon.
 474 Assess. 154, 413-426.
- 475 Lucas, A.J., Kudela, R.M., 2016. The fine-scale vertical variability of a wastewater plume in
 476 stratified coastal waters. Estuar. Coast. Shelf Sci. (accepted).
- McLaughlin, K., Nezlin, N.P., Howard, M.D.A., Beck, C.D.A., Kudela, R.M., Mengel, M.,
 Robertson, G.L., 2016. Tracking the fate of anthropogenic N from wastewater discharge into
 the coastal ocean, Southern California, USA. Estuar. Coast. Shelf Sci. (accepted).
- 480 Nezlin, N.P., Sutula, M.A. Stumpf, R.P. Sengupta, A., 2012. Phytoplankton blooms detected by
 481 SeaWiFS along the central and southern California Coast. J. Geophys. Res. 117, C07004.
- Noble, M., Xu, J.P., 2004. Huntington Beach Shoreline Contamination Investigation, Phase III,
 Final Report, Coastal circulation and transport patterns: the likelihood of OCSD's plume
 impacting Huntington Beach shoreline. US Geological Survey Open-File Report 2004-1019
 (http://pubs.usgs.gov/of/2004/1019/S).
- 486 Orange County Sanitation Division (OCSD), 2011. Outfall Land Section and OOBS Piping
 487 Rehabilitation Draft Environmental 718 Impact Report, Volume 2: Appendices
 488 (http://www.ocsd.com/Home/ShowDocument?id=10785).
- Paerl, H.W., Piehler, M.F., 2008. Nitrogen and marine eutrophication. In: Capone, D., Bronk, D.,
 Mulholland, M., Carpenter, E. [Eds.], Nitrogen in the marine environment. Elsevier, pp. 529567.
- 492 Reifel, K.M., Corcoran, A.A., Cash, C., Shipe, R., Jones, B.H., 2013. Effects of a surfacing
 493 effluent plume on a coastal phytoplankton community. Cont. Shelf Res. 60, 38-50.
- Rogowski, P., Terrill, E., Thomas, J., Rosenfeld, L., Largier, J., 2014. 2012 Orange County
 Sanitation District (OCSD) Outfall Diversion –Summary Report. Final Report prepared for
 the Orange County Sanitation District. March 25, 2014, 92 p., plus appendices.
- 497 Ryther, J., Dunstan, W., 1971. Nitrogen, phosphorus and eutrophication in the coastal marine
 498 environment. Science 171, 1008-1112.
- Scavia, D., Bricker, S.B., 2006. Coastal eutrophication assessment in the United States.
 Biogeochem. 79, 187-208.
- Schiff, K.C., Allen, M.J., Zeng, E.Y., Bay, S.M., 2000. Southern California. Marine Pollution
 Bulletin, 41, 76-93.

- Schnetzer, A., Miller, P., Schaffner, R., Stauffer, B., Jones, B., Weisberg, S., DiGiacomo, P.,
 Berelson, W., Caron, D., 2007. Blooms of *Pseudo-nitzschia* and domoic acid in the San
 Pedro Channel and Los Angeles harbor areas of the Southern California Bight, 2003-2004.
 Harmful Algae 6, 372-387.
- Schnetzer, A., Jones, B.H., Schaffner, R.A., Cetinic, I., Fitzpatrick, E., Miller, P.E., Caron, D.A.,
 2013. Coastal upwelling linked to toxic *Pseudo-nitzschia australis* blooms in Los Angeles
 coastal waters, 2005 2007. J. Plankton Res. 35, 1080-1092.
- Seegers, B.N., Teel., E.N., Kudela, R.M., Caron, D.A., Jones, B.H., 2016. Glider and remote
 sensing perspective of the upper layer response to an extended shallow coastal diversion of
 municipal wastewater effluent. Estuar. Coast. Shelf Sci. (accepted).
- Seubert, E.L., Gellene, A.G., Campbell, V., Smith, J., Robertson, G., Caron, D.A., 2016.
 Incubation experiments to determine the response of a natural plankton community to treated
 sewage effluent. Estuar. Coast. Shelf Sci. (submitted).
- Seubert, E.L., Gellene, A.G., Howard, M.D.A., Connell, P., Ragan, M., Jones, B.H., Runyan, J.,
 Caron, D.A., 2013. Seasonal and annual dynamics of harmful algae and algal toxins revealed
 through weekly monitoring at two coastal ocean sites off southern California, USA. Environ.
 Sci. Pollution Res. 20, 6878-6895.
- Sengupta, A., Sutula, M.A., McLaughlin, K., Howard, M.D.A., Tiefenthaler, L., Bitner, T.V.,
 2013. Terrestrial nutrient loads and fluxes to the Southern California Bight, USA. In: Schiff,
 K.C., Miller, K. (Eds.), Southern California Coastal Water Research Project 2013 Annual
 Report, edited by pp. 245-258, Southern California Coastal Water Research Project, Costa
 Mesa, CA.
- Tegner, M.J., Dayton, P.K., Edwards, P.B., Riser, K.L., Chadwick, D.B., Dean, T.A., Deysher,
 L., 1995. Effects of a large sewage spill on a kelp forest community: Catastrophe or
 disturbance? Mar. Environ. Res. 40, 181-224.
- Uchiyama, Y., Idica, E.Y., McWilliams, J.C., Stolzenbach, K.D., 2014. Wastewater effluent
 dispersal in Southern California Bays. Cont. Shelf Res. 76, 36-52.
- Ward, B.B., 1987. Nitrogen transformations in the Southern California Bight. Deep-Sea
 Research Part A 34, 785-805.

532 Figure Legends

- 533 Figure 1. Map of the study area illustrating instruments comprising the sensor network. The gray
- dashed line and gray dashed railroad lines show offshore and nearshore glider transects,
- respectively. Fixed platforms consist of water-quality moorings, including ADCPs (dark gray
- triangle), ADCPs only (open circles), Wirewalkers (white stars) and the ESP (gray square).
- Figure 2. Photographs of the wastewater effluent surfacing during the OCSD diversion. Courtesyof Mike McCarthy and Michael Mengel (OCSD).
- 539 Figure 3. Image of the effluent breach in San Diego, California on 25 February 1992 from the
- 540 NASA Airborne Ocean Color Imager (AOCI). Courtesy of Jeff Myers (NASA Ames Research541 Center).





543 Fig. 1



- 547 Fig. 2



556 Fig. 3