



# Assessing prevalence and correlates of blue-colored flesh in lingcod (*Ophiodon elongatus*) across their geographic range

Aaron W. E. Galloway<sup>1</sup> · Anne H. Beaudreau<sup>2</sup> · Michael D. Thomas<sup>1</sup> · Bonnie L. Basnett<sup>3</sup> · Laurel S. Lam<sup>3,7</sup> · Scott L. Hamilton<sup>3</sup> · Kelly S. Andrews<sup>4</sup> · Julie B. Schram<sup>1,5</sup> · Jessica Watson<sup>6</sup> · Jameal F. Samhoury<sup>4</sup>

Received: 25 February 2021 / Accepted: 5 July 2021 / Published online: 17 August 2021  
© The Author(s) 2021

## Abstract

Intraspecific variation in external and internal pigmentation is common among fishes and explained by a variety of biological and ecological factors. Blue-colored flesh in fishes is relatively rare but has been documented in some species of the sculpin, greenling, and perch families. Diet, starvation, photoprotection, and camouflage have all been suggested as proximate mechanisms driving blue flesh, but causal factors are poorly understood. We evaluated the relative importance of biological and spatial factors that could explain variation in blue coloration in 2021 lingcod (*Ophiodon elongatus*) captured across their range in the northeastern Pacific, from southeast Alaska to southern California. The probability of having blue flesh was highest for fish that were female, caught in shallower water, and smaller in body size. The incidence of blueness varied by region (4–25% of all fish) but was also confounded by differences in sex ratios of fish caught among regions. We analyzed the multivariate fatty acid composition of a subset of 175 fish from across the sampling range to test for differences in trophic biomarkers in blue lingcod. Lingcod fatty acid composition differed between regions and flesh colors but not between sexes. Blue-fleshed fish had lower concentrations of total fatty acids, 18:1 $\omega$ -9, 16:1 $\omega$ -7, 18:1 $\omega$ -7, and  $\omega$ -6 fatty acids, suggesting differences in energetics and energy storage in blue fish. While our data indicate potential links between diet and blue flesh in lingcod, important questions remain about the physiological mechanisms governing blueness and its biological consequences.

---

Responsible Editor: A. E. Todgham.

---

Reviewed by undisclosed experts.

---

✉ Aaron W. E. Galloway  
agallow3@uoregon.edu

<sup>1</sup> Oregon Institute of Marine Biology, University of Oregon, Charleston, OR, USA

<sup>2</sup> School of Marine and Environmental Affairs, University of Washington, Seattle, WA, USA

<sup>3</sup> Moss Landing Marine Laboratories, San Jose State University, Moss Landing, Moss Landing CA, USA

<sup>4</sup> Conservation Biology Division, Northwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Seattle, WA, USA

<sup>5</sup> University of Alaska Southeast, Juneau, AK, USA

<sup>6</sup> Oregon Department of Fish and Wildlife, Newport, OR, USA

<sup>7</sup> Pacific States Marine Fisheries Commission, Northwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Seattle, WA, USA

## Introduction

External pigmentation in fishes is under evolutionary pressure and has consequences for fitness, mediated through effects of coloration on survival and reproduction (Leclercq et al. 2010). While fish also vary in their internal (flesh) coloration, less is known about the mechanisms and selective pressures influencing pigmentation under the skin. Flesh coloration in fish is known to be affected by both intrinsic (hormones and nervous system) and extrinsic factors (chemicals and food; Prince 1916; Tunison et al. 1947). Color production in fishes is generally either due to pigmentation or is structurally based (Price et al. 2008), the latter being more common in blue-colored fishes (Fujii 1993). Variation in flesh color has been most extensively studied for species that are raised in aquaculture settings, where deviation from expected color will affect the market value; for example, salmon are known to vary in their flesh redness both as a function of diet and genetics (Torrissen and Naevdal 1988).

Blue coloration in tissues and serum has been noted in a range of fishes (reviewed in Nomura 1967), both in freshwater and marine systems. In sculpins (Cottidae), it has been

shown that blue coloration is due to the bile pigment biliverdin, a byproduct of heme catabolism; the coloration can be found in the serum (Bada 1970; Low and Bada 1974) and the tissues (Yamaguchi et al. 1976). Bard et al. (1996) described an unusual series of “green” tunas (Scombridae; multiple genera) landed in ports of West Africa, which had confirmed biliverdin in blue-stained bone structure. Some eels (*Anguilla japonica* and *A. rostrata*; Yamaguchi et al. 1966; Ellis and Poluhowich 1981) and wrasses (*Labrus bergylta*; Clark et al. 2016) are known to have blue–green color serum and high biliverdin concentrations in plasma. Blue flesh (i.e., muscle tissue) has also been reported in the humphead wrasse (*Cheilinus undulatus*; Yamaguchi and Matsura 1969), garfish (*Belone belone*; Fontaine 1941), and Pacific saury (*Cololabis saira*; Nomura 1967). Blue-morphs of lakewalling walleye (*Sander vitreus*) are caused by biliverdin-bound fluorescent proteins in the mucus layer of the fish, which has been suggested as a response to UV radiation (Schaefer et al. 2014; Ghosh et al. 2016). Blue blood and tissues, as a result of high biliverdin concentrations, has been documented in other vertebrates, including lizards (*Prasinochaema* and other genera Rodriguez et al. 2018), frogs (*Chiromantis*; Grismer et al. 2007), and worms (*Manduca sexta*; Goodman et al. 1985).

Fishers and ecologists alike have long puzzled over the striking bright blue or blue–green coloration within the tissues of some benthic fishes in the greenling (Hexagrammidae) and sculpin families (Yamaguchi et al. 1976; Love 2011). In several of these species that support recreational and commercial fisheries in the northeastern Pacific—lingcod (*Ophiodon elongatus*), rock greenling (*Hexagrammos lagocephalus*), and cabezon (*Scorpaenichthys marmoratus*)—the coloration of the flesh is at times bright enough that it can be observed on the outside of the fish (Fig. 1). In addition, blue-fleshed fish can be particularly valued in the marketplace (Chuck’s Seafood, Charleston, Oregon, personal communication to AWEG; Bland 2014). However, there are no published quantitative analyses that examine incidence, and proximate or ultimate factors that cause blue coloration in the greenling family. Lingcod provide a particularly valuable opportunity for understanding the ecological factors explaining the prevalence of blue flesh in fish across a large geographic range, due to their cosmopolitan distribution throughout the northeastern Pacific Ocean, from the Gulf of Alaska to central Baja Mexico (Miller and Lea 1972).

Lingcod reside in diverse habitats from the shoreline to a depth of at least 427 m (Miller and Geibel 1973), and typically exhibit a high degree of site fidelity (Starr et al. 2004; Greenley et al. 2016). Lingcod make a seasonal migration from deeper to shallower nearshore waters where they spawn in late fall through winter (Phillips 1958; Starr et al. 2004).

As lingcod age and grow, they tend to move to deeper, moderate- to high-relief rocky habitats (Cass et al. 1990; Bassett et al. 2018). Females grow faster than males and reach larger sizes, usually inhabiting deeper depths (Miller and Geibel 1973). Lingcod are generalist predators that consume a diversity of pelagic and demersal fishes and mobile invertebrates (Beaudreau and Essington 2007, 2009), including cephalopods and crustaceans that contain respiratory proteins (hemocyanins) that are blue when oxygenated (van Holde et al. 2001). Blue flesh in lingcod is a frequently discussed topic in the fishing community and is often attributed to differences in the diets of individual fish; however, this assertion has not, to our knowledge, been tested formally.

A valuable method for understanding diets in marine fishes is through quantification of the concentrations and composition of fatty acids in a consumer’s tissues (Parrish 2013; Pethybridge et al. 2018). Individual fatty acids and key classes of physiologically important fatty acids are commonly used as trophic tracers (i.e., ‘biomarkers’) for inferring feeding relationships among predators and prey (Budge et al. 2002; Dalsgaard et al. 2003). The integration of fatty acids into the tissues of fishes is complex and depends largely upon their diet (Xu et al. 2020) (exogenous sources) but also depends on individual species physiology (endogenous processes), which is also governed by environmental conditions (reviewed in Arts and Kohler 2009; Parrish 2013; Xu et al. 2020). Essential fatty acids (EFAs) such as docosahexaenoic (DHA; 22:6 $\omega$ -3), eicosapentaenoic (EPA; 20:5 $\omega$ -3), and arachidonic (ARA; 20:4 $\omega$ -6) fatty acids are important for the nutritional physiology of fishes (Sargent et al. 1999; Copeman et al. 2002; Bell and Sargent 2003; Parrish 2013). Moreover, the ratio of  $\omega$ -3: $\omega$ -6 fatty acids, and total fatty acids can be used as indices of fish condition (Pethybridge et al. 2015). A common application of fatty acids as trophic indicators in fishes is the comparative analysis of conspecifics between life stages, with different coloration, or based on differences in the collection habitats, or across spatial or environmental gradients (Budge et al. 2002; Pethybridge et al. 2015; Copeman et al. 2016). This kind of comparative approach is often used for species in which consumer-specific fatty acid trophic modification is unknown (Galloway and Budge 2020), and does not aim to infer consumer diet composition, but rather assumes that differences in fatty acids of conspecifics between treatments or across a geographical gradient indicates dissimilarities in trophic ecology.

We used data from multiple field collection efforts to broadly document spatial and demographic patterns in the prevalence of blue coloration in lingcod throughout their range along the U.S. west coast and to investigate potential physiological and environmental correlates of blue flesh

**Fig. 1** Examples of blue-fleshed fish and serum. **a** Blue-fleshed rock greenling (*Hexagrammos lagocephalus*; Aleutian Islands, AK; Photo by Scott Gabara). **b**, **c** Blue flesh and mouth tissues of a lingcod (Port Orford, OR; Photos by Aaron Galloway); **d** Blue serum from lingcod (San Juan Islands, WA; Photo by Aaron Dufault); **e** Representative range of color variants— from the left, individuals 1, 3, and 4 were designated ‘brown’ and individuals 2 and 5 were designated ‘blue’ (Photo by Laurel Lam)



in the species. We performed an initial pilot study based on lingcod caught in southern Oregon in 2016 and tested whether the fatty acids differed between blue and non-blue (hereafter, ‘brown’) fish. We then extracted the fatty acids from a subset of the lingcod sampled in 2015–2017 over a broad geographic range, from southeastern Alaska to southern California, to determine whether blue and brown lingcod could be distinguished based on their fatty acid profiles. We used the full latitudinal dataset to evaluate the relative importance of influential fatty acids (as determined by the trophic tracer analyses), geographic factors, and biological variables in determining the probability of blue flesh in lingcod.

## Methods

### Field and laboratory methods

#### Southern Oregon survey

We collected muscle tissue, using an 8 mm biopsy punch, from the left dorsal area of lingcod ( $n = 72$ ) in the Redfish Rocks Marine Reserve and Humbug Comparison Area near Port Orford, Oregon. The samples were collected by the Oregon Department of Fish and Wildlife Marine Reserves Program on six dates in June and July 2016 using longline and hook-and-line sampling methods

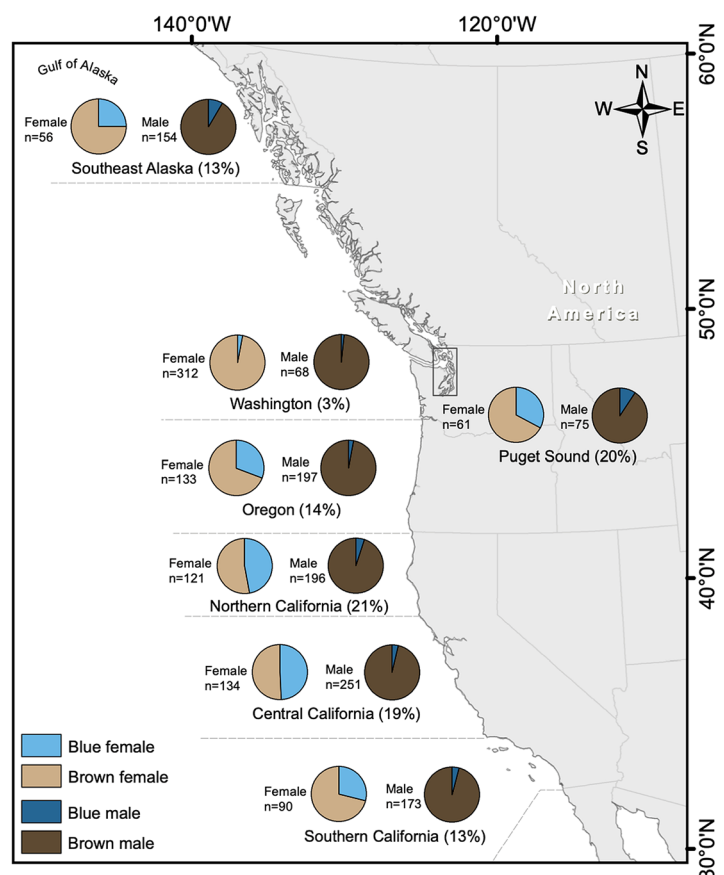
described in Huntington and Watson (2017). In the field, fork length (cm), sex (male/female), muscle tissue coloration (blue, brown), fish condition comments, bottom depth, and collection site coordinates were recorded. We stored all fish tissues on ice until they could be frozen at the lab (e.g., < 6 h after collection) in an  $-80^{\circ}\text{C}$  freezer. This dataset was used to identify emergent patterns in the sex ratio, fatty acid composition, and comparisons of total  $\omega$ -6 fatty acids (see methods below) between blue and brown fish. This initial analysis thus identified hypotheses to test with the independent and much larger fatty acid dataset from the latitudinal survey.

### Latitudinal survey

Lingcod were collected in 2015–2017 from chartered commercial passenger fishing vessels from Yakutat, Alaska to San Diego, California, USA ( $54^{\circ}30'$  N– $34^{\circ}30'$  N). The following seven regions were broadly identified (Fig. 2): southeast Alaska ( $54^{\circ}30'$  N– $59^{\circ}48'$  N), Puget Sound, coastal Washington ( $46^{\circ}16'$  N– $49^{\circ}$  N), Oregon ( $42^{\circ}$  N– $46^{\circ}16'$  N), northern California ( $38^{\circ}02'$  N– $42^{\circ}$  N), central California ( $34^{\circ}30'$  N– $38^{\circ}02'$  N), and southern California ( $32^{\circ}32'$  N– $34^{\circ}30'$  N). Three to four fishing ports were selected per

region (24 ports total) based on location and accessibility of charter vessels. Regions were generally selected based on major biogeographic features (point conception, Cape Mendocino, Columbia River, and Puget Sound). We collected lingcod using hook-and-line (jigging with artificial lures). Hook-and-line gear is an effective method of capturing fishes in high-relief, rocky habitat that cannot be sampled using nets (e.g., trawls) and is commonly used in lingcod studies (e.g., Beaudreau and Essington 2007, 2009). Like all gear types, hook-and-line can potentially introduce selection bias, such as the potential to capture more actively feeding fish and selecting for post-settlement juvenile and adult lingcod (> 20 cm total length), as smaller lingcod have not yet settled into rocky reef habitats. Acknowledging these limitations, we aimed to capture a representative sample of lingcod across a wide range of size, sex, and age classes by sampling shallow (< 61 m) and deep (61–168 m) nearshore and offshore rocky reefs equally. We fished for an average of 2.5 days out of each port to obtain the targeted sample size of 75–100 lingcod per port. Additional samples were provided by the Alaska commercial longline fishery, a rockfish bycatch study in Puget Sound (K. Andrews, unpublished data), the Oregon Department of Fish and Wildlife Marine Reserves Program (Huntington et al. 2014), and the

**Fig. 2** Map summarizing the regional proportions of blue and brown lingcod by sex. The percentage of all fish that were blue is shown for each region in parentheses. Additional details about the sample sizes in each category and numbers included in the fatty acids analysis are reported in Table S1



California Collaborative Fisheries Research Program (Wendt and Starr 2009).

Location information (i.e., latitude, longitude, bottom depth, relief, and time of day) was collected at the start and stop of each fishing drift. Total length (cm), weight (kg), sex, and color were recorded for each individual immediately upon capture. Samples were then placed on ice or frozen until dissection. Sex was determined externally (by the presence or absence of a conical papilla) and internally (by the presence or absence of ovaries or testes). Color was visually assessed both externally and internally by two authors to ensure consistency (BLB, LSL). We determined external color based on skin pigmentation immediately after each fish was landed, then verified external color by examining the buccal cavity and dorsal fin membrane color. This secondary check was done because mottling patterns on the skin can confound paler blue hues, while the buccal cavity and fin membrane are typically uniform in color and offer a cleaner presentation of the blue coloration (Fig. 1). Internal color provided a third check on coloration and was determined based on muscle tissue during the time of dissection, between 8 and 36 h after capture. In externally blue fish, the internal color of the flesh is also blue; however, in the more common externally brown fish, the internal color of the flesh is white/gray. While the intensity of blue pigmentation was observed as a gradient, color was recorded as binary (blue vs. brown; Fig. 1e) to avoid subjective determination of color intensity. The multiple methods (skin, buccal cavity, fin membrane, and flesh) used to assess and verify color improved confidence in our designation of external and internal color. During the full dissection process, we recorded liver, gonad, whole stomach weight, and stomach content weight (g). A piece of muscle tissue was collected from the anterior dorsal region on the left side for each individual and placed in a 1.8 mL cryogenic vial and frozen at  $-20^{\circ}\text{C}$  until fatty acids extraction in the spring of 2018. A total of 2157 fish with color, sex, and size information were recorded.

### Lab analysis of fatty acids

As it was not possible to extract fatty acids from all latitudinal survey lingcod tissue samples, we selected a subset of 175 samples for fatty acid analysis (Table S1) using a stratified random sampling approach. Since an initial analysis of the southern Oregon data indicated that blue coloration varied by sex, we subsampled to achieve approximately equal proportions of sex and coloration (blue or brown) within each of the seven regions sampled in the latitudinal survey. A minimum of 5 lingcod were sampled from each stratum (i.e., combination of geographical region, sex, and flesh

color), except for the case of blue male lingcod in Washington, where we did not have any representative fish tissues to draw from (Table S1). All samples were lyophilized, while frozen, for 48 h, to achieve complete dryness. Total fatty acid extraction procedures followed Thomas et al. (2020), using a  $\sim 20$  mg dry weight tissue aliquot. Briefly, all lipids were extracted from fish tissues using a 2:1:0.75 mixture of chloroform:methanol:NaCl (0.9%). An unmethylated internal standard (19:0) was added to each sample to quantify extraction and methylation efficiency. Samples were sonicated to facilitate cell lysis and centrifuged to isolate the lipid-containing organic phase. An aliquot of the organic phase was removed for gravimetric determination of total lipid content ( $\text{mg lipid g}^{-1}$  dry wt.) following Kainz et al. (2017). The remaining portion of the organic phase was evaporated to dryness and 1 mL toluene and 2 mL methanolic sulfuric acid (1%) were added immediately before incubating samples at  $90^{\circ}\text{C}$  for 90 min to form fatty acid methyl esters. After incubation, purified samples were neutralized with 1.5 mL  $\text{KHCO}_3$  (2%) and extracted twice with n-hexane. Analysis of fatty acid methyl esters followed procedures described in Taipale et al. (2013) using a Shimadzu QP2020 gas chromatograph-mass spectrometer at the Oregon Institute of Marine Biology.

### Data analysis

We used multiple datasets and analytic methods to address the study objectives. A visual summary of datasets used, study questions, statistical methods, and key results is shown in Figure S1.

### Data preparation

Prior to analyses, we prepared the data as follows. Out of the 2157 lingcod from the latitudinal survey, there was agreement between external and internal color designations for 1934 fish (89.7%). There were discrepancies between external and internal color for 136 fish (6.3%); specifically, 2.4% of the fish were assigned brown externally and blue internally, whereas 3.9% of the fish were assigned blue externally and brown internally. These 136 fish were excluded from further analysis. Twenty-two fish were missing an external color designation (1.0%) and 65 fish were missing an internal color designation (3.0%), due to accidental omission while recording field data. Rather than reducing the sample by 87 fish, we assigned the 22 fish of unknown external color to their observed internal color. All further analyses were performed using external color designations. A total of 2021 lingcod with complete size, sex, and color information were available for analysis.

## Fatty acids

A total of 46 unique fatty acids were identified and used in our analyses of both fatty acid datasets. For the Southern Oregon survey, we compared the sex ratio of blue and brown fish using a Chi-square test and analyzed fatty acids of 72 lingcod (41 male and 31 female) in 2017. These preliminary analyses inspired the larger and more robust latitudinal lingcod fatty acids effort, designed to account for blueness, sex, and region (described below). We compared multivariate fatty acid composition using two-way permutational multivariate analysis of variance (PERMANOVA; 9999 permutations, Type III SS,  $p$  values calculated from unrestricted permutation of the raw data; Anderson et al. 2008) on the factors ‘color’ and ‘sex’. Concentrations of key a-priori determined fatty acid categories [total polyunsaturated fatty acids (PUFA), total  $\omega$ -6 fatty acids, the  $\omega$ -3: $\omega$ -6 fatty acid ratio, total long-chain PUFA [LCPUFA; sum of 20:4 $\omega$ -6 (ARA), 20:5 $\omega$ -3 (EPA), 22:6 $\omega$ -3 (DHA)], and total lipids] were log-transformed and compared with two-tailed  $t$  tests.

To further examine differences in fatty acid composition and concentrations at a larger geographic scale (i.e., combinations of region, sex, and color), we extracted and analyzed 175 latitudinal fatty acid samples. Two of these samples were later excluded due to their extreme outlier status when visualized in multivariate space; on further inspection, these samples had anomalous and unreasonable levels of several fatty acids, likely due to extraction process errors. We, therefore, performed subsequent multivariate analyses on 173 samples using 3-way PERMANOVA, using log-transformed fatty acid concentration data  $\{\log[X + 1]\}$ , with the factors ‘region’ (random factor), ‘sex’, and ‘color’ (both fixed factors). We focused analyses on the fatty acid concentration data because concentrations are more informative from a physiological standpoint. Table S2 reports the mean and SD of all fatty acids summarized by lingcod flesh color. We tested the homogeneity of multivariate dispersions (PERMDISP) of factors analyzed in the PERMANOVA. We used multivariate similarity percentages (SIMPER) on the transformed fatty acid concentration data to identify the fatty acids that were most influential in separating the blue and brown fish. In tables, we report the untransformed, actual mean concentrations of fatty acids for ease of interpretation, expressed as mg fatty acids  $g^{-1}$  dry weight. We visualized the multivariate group dispersion patterns using a non-metric multidimensional scaling (nMDS) analysis (Euclidean distance), with individual fatty acids correlated with the ordination axes shown as vector overlays. Individual and summary categories of fatty acids that were identified in the initial southern Oregon regional collections, in the literature (e.g., Pethybridge et al. 2015), or the SIMPER analysis on the full west coast latitudinal collections were compared

with two-sample  $t$  tests and boxplots. We used Primer v6 (Clarke and Gorley 2006) with the PERMANOVA + add-on (Anderson et al. 2008) for multivariate analyses. All tests of statistical significance were two-tailed and generally used  $\alpha = 0.05$ . For the latitudinal univariate fatty acid comparisons, the significance level of the tests was adjusted using a Bonferroni-correction for multiple tests (where  $\alpha = 0.05/10$  tests = 0.005). Univariate fatty acid analyses were run in R version 4.1.0 (R Development Core Team 2021).

## Probability of blue coloration: all surveys

Lingcod were caught across a similar range of depths for all regions except Washington, where fishing locations were deeper on average than other regions (Figure S2). To avoid the confounding effects of region and depth, we restricted analyses to fish caught at depths shallower than 125 m, resulting in removal of an additional 223 fish. To test hypotheses regarding potential drivers of blue flesh, we performed separate logistic regression analyses on the prepared data ( $n = 1798$  lingcod; hereafter ‘full dataset’) and a subset of the full dataset for which fatty acid concentrations were measured ( $n = 163$  lingcod; hereafter ‘FA only dataset’). For the full dataset, we used logistic regression to estimate the probability of blue coloration as a function of geographic region, depth, length, and sex. Region and depth were included in the models to evaluate whether latitudinal and depth distribution differences among individuals accounted for variation in coloration. Sex and length were included as proxies for sex- and size-based differences in diet and physiological status (e.g., reproductive hormones, growth hormones, and demographic shifts in diets) that are potential determinants of blue flesh. For the FA only dataset, five full models were constructed, each including region, depth, length, sex, and one of the following fatty acid variables: total fatty acid concentration,  $\omega$ -3: $\omega$ -6 ratio, 18:1 $\omega$ -9, 16:1 $\omega$ -7, or 18:1 $\omega$ -7 (concentrations). Total fatty acid concentration was included because of the significant differences found between blue and brown fish. The ‘ $\omega$ -3: $\omega$ -6’ factor was included because of hypotheses generated from the independent southern Oregon dataset and because  $\omega$ -6 fatty acids, particularly ARA, may be an important indicator of nutritional status in fishes (Sargent et al. 1999; Bell and Sargent 2003). The other three biomarkers were included because they were identified from the SIMPER analysis as the most influential fatty acids explaining differences between blue and brown fish (i.e., each contributing > 10%; see “Results”). Because of the relatively low sample size in the FA only dataset, including all potential fatty acid predictors in the full model would have resulted in a very large number of candidate models and can result in overfitting the data without contributing meaningful biological inference (Burnham and Anderson 2002).

For each of the datasets, multiple logistic regression models were fitted to the binary response of blue (1) or brown (0), with a logit link function using the 'glm' function in the R package 'MASS' (R Development Core Team 2021). Since the units of measurement and scales of predictor variables varied substantially, we standardized the factors prior to fitting the regression models (Gelman 2008) using the 'rescale' function in the R package 'arm' (R Development Core Team 2021). Rescaling predictor variables also allows for direct comparison of estimated coefficients (Gelman 2008). The sets of candidate models included the full models and all possible combinations of potential predictors, as described above for each dataset. Model selection was performed using Akaike's Information Criterion corrected for small sample size (AICc), where models with AICc values within two units of the lowest AICc across all models were included in the set of top models (Burnham and Anderson 2002). We evaluated the relative importance of explanatory factors using parameter weights, calculated as the sum of Akaike model weights across all models including a given parameter (Burnham and Anderson 2002). Parameter weights range from 0 to 1, where values approaching 1 signify a strong weight of evidence that the predictor should be included in the best-fit model (Burnham and Anderson 2002).

## Results

After removing fish with no length measurement, unidentified sex or color, and colors with mismatched external and internal color designations, we were left with 2,021 fish for analysis (Table S1) in the latitudinal survey dataset. Of these, 286 (14%) were blue in color. Most blue fish were female (81%) and the proportion of blue lingcod within each region ranged from 3% (Washington coast) to 21% (northern California) (Fig. 2).

### Southern Oregon survey

The large majority (90%) of the lingcod in the initial southern Oregon survey with blue flesh were female, differing strongly from an even sex ratio (Pearson's Chi-squared test;  $\chi^2 = 20.18$ ,  $df = 1$ ,  $p < 0.0001$ ). Of females, 55% were blue-fleshed and 45% were brown, whereas only 5% of males were blue. Multivariate fatty acid composition in the southern Oregon dataset did not differ by color, sex, or their interaction (2-way PERMANOVA; color: Pseudo  $F = 0.83$ ,  $df = 1$ ,  $p = 0.41$ ; sex: Pseudo  $F = 2.09$ ,  $df = 1$ ,  $p = 0.11$ ; color x sex: Pseudo  $F = 0.96$ ,  $df = 1$ ,  $p = 0.36$ ; no nMDS presented). The southern Oregon lingcod dataset was limited in the sample size of blue fish. Since almost all blue fish were female, preliminary analyses of univariate fatty acid

summary categories focused on female fish. Blue female lingcod had a higher  $\omega$ -3: $\omega$ -6 ratio ( $t$  test = 2.23  $df = 26.70$ ,  $p = 0.035$ ), lower total log( $\omega$ -6) fatty acids ( $t$  test = -2.25,  $df = 21.93$ ,  $p = 0.034$ ), and lower log(20:4 $\omega$ -6) (ARA) content ( $t$  test = 2.11,  $df = 21.52$ ,  $p = 0.046$ ), but these initial comparisons were not statistically significant when Bonferroni-correcting for multiple tests in the Oregon dataset. These preliminary findings with small sample sizes were used as a pilot study to inform the later analysis and are, therefore, interpreted with caution.

### Latitudinal survey fatty acids

Lingcod multivariate fatty acid concentrations differed significantly between regions and fish color, but not sex; there were no significant interactions among any terms and the full model explained ~26% of the total variation (3-way PERMANOVA; Table S3). Multivariate fatty acid dispersion did not differ among regions (PERMDISP;  $F_{6, 166} = 1.39$ ,  $P$  (perm) = 0.57) or sexes (PERMDISP;  $F_{1, 171} = 0.93$ ,  $P$  (perm) = 0.45), but blue and brown fish differed in their dispersion (PERMDISP;  $F_{1172} = 9.42$ ,  $P$  (perm) = 0.012). The nMDS ordination (Fig. 3) showed generally lower multivariate fatty acid dispersion within blue-fleshed lingcod, with tighter grouping on the left side of the plot (except for 3 outliers, which were included in the PERMDISP test).

SIMPER identified that 18:1 $\omega$ -9, 16:1 $\omega$ -7, 18:1 $\omega$ -7, 20:5 $\omega$ -3, 22:6 $\omega$ -3, 16:0, 14:0, 20:1 $\omega$ -9, 20:4 $\omega$ -6, 18:0, and 22:5 $\omega$ -3, in that order, were the most important fatty acids for differentiating blue and brown lingcod (see SIMPER results, Table 1). Blue fish (pooled across sexes and regions) had significantly lower concentrations of log 16:1 $\omega$ -7 ( $t$  test = 4.09,  $df = 169.24$ ,  $p = 0.0001$ ), log 18:1 $\omega$ -9 ( $t$  test = 4.63,  $df = 171$ ,  $p < 0.0001$ ), log 18:1 $\omega$ -7 ( $t$  test = 4.35,  $df = 168.72$ ,  $p < 0.0001$ ), log 20:4 $\omega$ -6 ( $t$  test = 4.05,  $df = 166.51$ ,  $p = 0.0001$ ), log 20:5 $\omega$ -3 ( $t$  test = 4.37,  $df = 164.39$ ,  $p < 0.0001$ ), log total  $\omega$ -6 ( $t$  test = 4.67,  $df = 159.81$ ,  $p < 0.0001$ ), and log total fatty acids ( $t$  test = 3.47,  $df = 170.37$ ,  $p = 0.0007$ ) (Fig. 4). There were no differences in the concentrations of log 22:6 $\omega$ -3 ( $t$  test = 0.42,  $df = 166.99$ ,  $p = 0.67$ ) or log total lipids ( $t$  test = 1.57,  $df = 160.37$ ,  $p = 0.12$ ; not pictured). Blue fish had a significantly higher  $\omega$ -3: $\omega$ -6 ratio ( $t$  test = 4.31,  $df = 170.18$ ,  $p < 0.0001$ ) than brown fish (Fig. 4).

### Probability of blue coloration

We modeled the probability of blue flesh as a function of region, capture depth, total length, and sex using the full dataset. The best model included all of these variables (Table 2), which all had high parameter weights (> 0.8). Sex had the greatest effect on probability of blue coloration, followed by length, depth, and region (Table S4). Based on

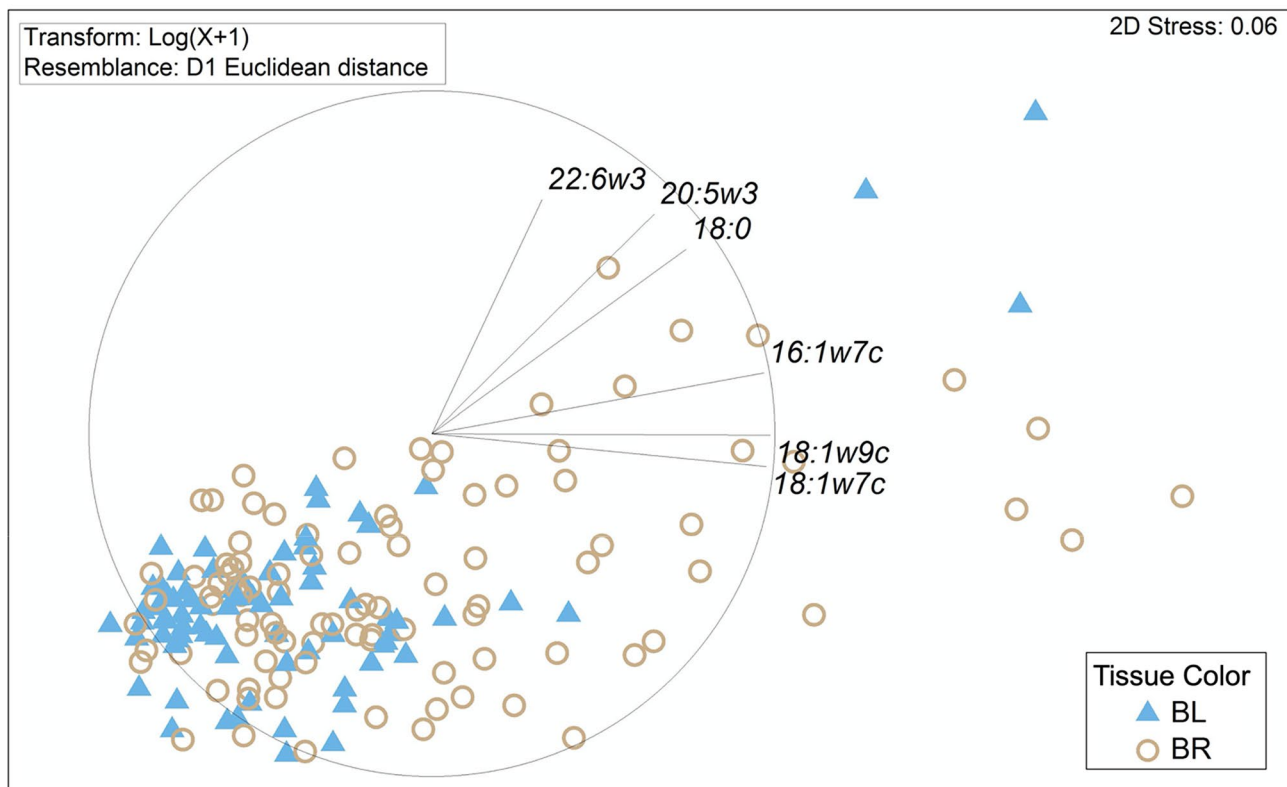
the signs of estimated coefficients from the best-fit model, lingcod that were female, relatively smaller, and relatively shallower in bottom depth at capture had a higher probability of blue coloration (Fig. 5). There was no apparent latitudinal trend in the proportion of lingcod showing blue coloration and regional differences were driven largely by the Washington coast, where the predicted probability of blue coloration was smallest, and fewer samples were collected in shallower waters near the coast.

Fatty acid concentrations were relatively unimportant in determining the probability of blue coloration (Table 2). Analysis of the fatty acid-only dataset revealed that eight models performed equivalently ( $\Delta AIC \leq 2$ ) and, when considered together, included all potential predictors. However, consistent with the analysis of the full dataset, the most parsimonious model ( $\Delta AIC = 0$ ) included only sex, length, and depth (Table 2). Sex and length were the most influential predictors (parameter weights = 1) with the greatest effect sizes, and depth was a relatively important predictor with parameter weight = 0.78 (Table S4). As in the full model, lingcod that were female, relatively smaller, and residing in relatively shallower depths had a higher predicted probability of blue coloration compared to males, relatively larger individuals, and those residing in relatively deeper depths

(Table S4). Region, total fatty acid concentration,  $\omega$ -3: $\omega$ -6 (selected a priori), 18:1 $\omega$ -9, 16:1 $\omega$ -7, and 18:1 $\omega$ -7 (the top three fatty acids from the SIMPER) were relatively unimportant in explaining variation in occurrence of blue lingcod, with effect sizes not different from zero (Table 2) and parameter weights < 0.3 (Table S4).

## Discussion

The question of why some lingcod are blue has long been of interest to seafood workers, fishers, scientists, and naturalists. More than an ecological curiosity, this phenomenon has commercial implications and possible evolutionary consequences that deserve further investigation. Our study was the first to use an extensive dataset from across the species range to improve our understanding of spatial and demographic patterns in the prevalence of blue fish and potential dietary and physiological drivers of blue flesh. We found that the probability of having blue flesh was highest for fish that were female, caught in shallower water, and smaller in total length. Sex is a previously overlooked factor for blue color in lingcod, given that > 80% of blue fish across this large geographic range were female. Incidence of blueness

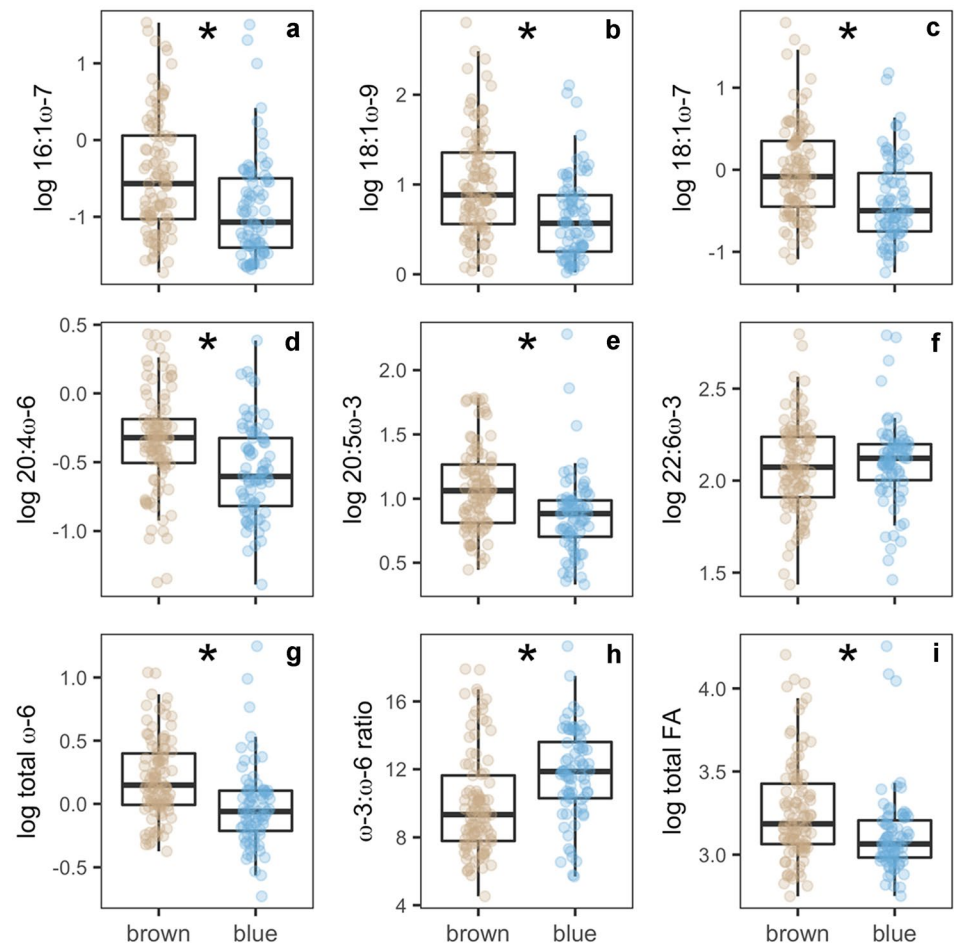


**Fig. 3** Non-metric multidimensional scaling (nMDS) plot showing multivariate dispersion of blue- and brown-fleshed lingcod. The analysis includes 173 fatty acid samples from the West Coast latitudi-

nal dataset. Data are Log(X+1) transformed fatty acid concentrations (mg FA g<sup>-1</sup> dry weight)



**Fig. 4** Data (colored points) are jittered within each category to reduce overplotting. Fatty acids were included based on SIMPER results and a priori hypotheses. **a–c** Key mono-unsaturated fatty acids; **d–f** key polyunsaturated essential fatty acids; **g** the sum of all  $\omega$ -6 fatty acids; **h** the  $\omega$ -3: $\omega$ -6 fatty acid ratio; and **i** total fatty acids. All comparisons differ significantly except for panel f (indicated by “\*”; Bonferroni-corrected *t* tests, where  $\alpha < 0.005$ )



**Table 1** Multivariate similarity percentages (SIMPER) analysis on the  $\log(X + 1)$  transformed fatty acid concentration data (mg FA  $\text{g}^{-1}$  dry wt.) was used to identify the fatty acids that were most influential in separating the blue and brown fish visualized in the nMDS (Fig. 3)

| Fatty acid       | mg Fatty acid $\text{g}^{-1}$ dry wt. (untransformed) |      |       |      | (Transformed)     |        |
|------------------|---|------|-------|------|-------------------|--------|
|                  | Blue  |      | Brown |      | SIMPER statistics |        |
|                  | mean  | SD   | mean  | SD   | % Contrib         | Cum. % |
| 18:1 $\omega$ -9 | 2.14  | 1.35 | 3.34  | 2.55 | 24.52             | 24.52  |
| 16:1 $\omega$ -7 | 0.56  | 0.69 | 0.90  | 0.87 | 15.31             | 39.84  |
| 18:1 $\omega$ -7 | 0.80  | 0.53 | 1.20  | 0.93 | 12.20             | 52.04  |
| 20:5 $\omega$ -3 | 2.52  | 1.13 | 3.12  | 1.10 | 9.02              | 61.06  |
| 22:6 $\omega$ -3 | 8.38  | 2.03 | 8.30  | 2.14 | 5.85              | 66.91  |
| 16:0             | 4.44  | 1.32 | 5.03  | 1.48 | 5.74              | 72.65  |
| 14:0             | 0.20  | 0.29 | 0.33  | 0.32 | 5.40              | 78.04  |
| 20:1 $\omega$ -9 | 0.26  | 0.25 | 0.36  | 0.26 | 3.76              | 81.81  |
| 20:4 $\omega$ -6 | 0.61  | 0.22 | 0.76  | 0.28 | 3.12              | 84.93  |
| 18:0             | 1.48  | 0.43 | 1.62  | 0.40 | 2.73              | 87.66  |
| 22:5 $\omega$ -3 | 0.36  | 0.17 | 0.43  | 0.19 | 1.94              | 89.60  |

The untransformed fatty acid mean and SD concentrations for each category (blue and brown) are presented in the first four columns, followed by the SIMPER statistics. Fatty acids are listed in order of their ascending percent contribution (% Contrib.) in differentiating samples based on lingcod color. The Cum. % column identifies the cumulative % explained as each variable is added

**Table 2** Model diagnostics from analyses of the full dataset and the fatty acid '(FA) only' dataset

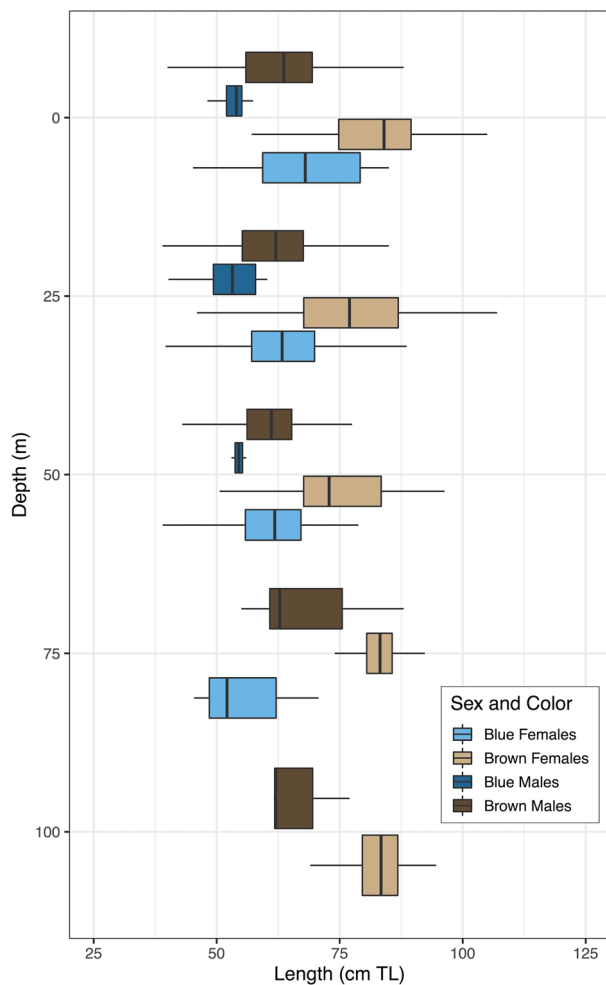
| Full dataset  |   |        |               |       |
|---|---|--------|---------------|-------|
| Model form  | K | AICc   | $\Delta$ AICc | Wi    |
| Pr{blue} ~ region + depth + length + sex                    | 6 | 1059.9 | 0.0           | 0.858 |
| Pr{blue} ~ depth + length + sex                             | 5 | 1063.5 | 3.6           | 0.142 |
| FA only dataset   |   |        |               |       |
| Model form  | K | AICc   | $\Delta$ AICc | Wi    |
| Pr{blue} ~ depth + length + sex                             | 5 | 177.9  | 0.0           | 0.134 |
| Pr{blue} ~ depth + length + sex + 18:1 $\omega$ -9          | 6 | 178.0  | 0.1           | 0.125 |
| Pr{blue} ~ depth + length + sex + 18:1 $\omega$ -7          | 6 | 178.4  | 0.6           | 0.100 |
| Pr{blue} ~ depth + length + sex + total FA                  | 6 | 178.9  | 1.1           | 0.079 |
| Pr{blue} ~ depth + length + sex + $\omega$ -3: $\omega$ -6  | 6 | 179.4  | 1.6           | 0.061 |
| Pr{blue} ~ region + depth + length + sex                    | 6 | 179.5  | 1.7           | 0.058 |
| Pr{blue} ~ depth + length + sex + 16:1 $\omega$ -7          | 6 | 179.6  | 1.7           | 0.056 |
| Pr{blue} ~ region + depth + length + sex + 18:1 $\omega$ -9 | 7 | 180.0  | 2.1           | 0.047 |

Only the set of models within 2 units of the lowest AICc is shown for each analysis. Using the full dataset, the probability of blue coloration (Pr{blue}) was modeled as a function of region, depth, lingcod length, and lingcod sex. Using the FA only dataset, Pr{blue} was modeled as a function of the same factors plus a fatty acid (FA) variable (18:1 $\omega$ -9, 16:1 $\omega$ -7, 18:1 $\omega$ -7,  $\omega$ -3: $\omega$ -6, or total FA). K is the number of estimated parameters, AICc is Akaike's information criterion corrected for small sample size,  $\Delta$ AICc is the AICc for each model minus the lowest AICc across all candidate models, and Wi is the Akaike model weight

was 14% coastwide and varied by region (3–21% of all fish) but was also confounded by sex ratios of fish caught among regions. In lingcod, smaller females are typically found inshore and migrate to deeper waters at larger sizes, returning only for reproduction (Low and Beamish 1978); however, some reproductively mature female lingcod remain in shallow nearshore habitats. For example, an acoustic telemetry study in central California found that small females had the highest residency within the nearshore array (Greenley et al. 2016). Considering our results showing that smaller, shallower, female fish are more likely to be blue, it is possible that remaining in nearshore environments is associated with a physiological response or a dietary difference that is less common in larger, deep dwelling females and cohabitating males. Recent research has shown that male blue lingcod also carry heavier burdens of metazoan parasites than do brown males, brown females, or blue females (Wood et al. in revision), and that those blue males are in poorer condition (low Fulton's K condition factor) than other sex-color combinations. It may be that starvation stress causes the release of compounds causing blueness and also weakens immune defenses, resulting in higher parasite burdens for males only because males are already immunocompromised by male sex hormones (Folstad and Karter 1992; Roberts et al. 2004; Foo et al. 2017).

Biliverdin has long been identified as the underlying cause of blue coloration of serum and other body parts in fishes (Fang and Bada 1990). We observed that lingcod with blue flesh reliably had visibly blue-colored serum. Biliverdin is found in the bile and serum of a diverse lineage of

animals. In mammals, it is rarely detectable, except during hepatic dysfunction, as it is enzymatically converted to yellow/orange bilirubin; however, it is the primary bile pigment in many avian, reptile, amphibian, and fish species (Cornelius 1984). In fishes, biliverdin may serve a role in external coloration (Gagnon 2006), starvation, or as a signal of stress (Love 1980). For example, attempts to replace traditional fish meal with plant-based proteins in aquaculture of red sea bream (*Pagrus major*) and yellowtail (*Seriola quinqueradiata*) have resulted in a taurine deficiency, linked to increased levels of biliverdin in hepatic fluids and resulting in a condition known as green liver (Takagi et al. 2010). Starvation has also been shown to lead to increased levels of serum biliverdin in a number of fish species (Fang 1987). For example, in Atlantic cod (*Gadus morhua*) it has been shown that with experimental starvation there is an excess production of blue-green bile in the gall bladder, which leaks into the intestines (Love 1980). In our initial exploratory analyses of the lingcod data, we did not identify total lipid content or the hepatosomatic index (HSI), which are common indices of condition or starvation, as important factors in predicting blueness; however, using a different subset of samples, Wood et al. (in revision) report that blue lingcod of both sexes have lower HSI than brown individuals. Other hypotheses for why biliverdin accumulates in the tissues have been advanced, but not tested. For example, Yamaguchi et al. (1976) posited that the biliverdin in blue-fleshed sculpins (*Pseudoblennius percoides*) might be stored in the flesh before being transported to skin and eggs for the purpose of camouflage, ultra-violet protection,



**Fig. 5** Distributions of lingcod length across 25 m depth bins from the full West Coast latitudinal dataset. Separate boxplots are shown for each combination of color and sex. Boxes show the interquartile range (IQR), with the median shown as a line within each box. Whiskers show  $\pm 1.5 \times \text{IQR}$ . Outliers were removed from the figure for ease of visualization

or through involvement with reproductive activities, when needed. We collected additional samples of lingcod and their serum in Puget Sound, Washington, ( $n = 70$ ) during opportunistic research efforts during spring–summer of 2018 and 2019 in an attempt to analyze biliverdin and serum coloration. Unfortunately, we were not able to replicate other fish serum biliverdin analysis methods, despite the very clear blue coloration of the serum of blue-fleshed fish (Figs. S3, S4). The serum pilot analysis is described in further detail in the Supplemental Materials. Further research is needed to characterize conditions under which biliverdin production and excretion is modulated in lingcod.

The association between fatty acid concentrations and lingcod blueness was nuanced. Multivariate fatty acid signatures differed between blue and brown lingcod, but an

nMDS visualization showed considerable variability and logistic regressions suggested relatively little support for individual fatty acids as predictors of blueness. SIMPER analyses identified that the most important fatty acids for differentiating blue and brown fish were 18:1 $\omega$ -9, 16:1 $\omega$ -7, 18:1 $\omega$ -7, 20:5 $\omega$ -3 (EPA), 22:6 $\omega$ -3 (DHA), and 16:0, which accounted for 73% of the cumulative variation in the SIMPER analysis. Most of these fatty acids had small absolute differences in terms of total mg fatty acids  $\text{g}^{-1}$  of tissue, and while most of these differences were statistically significant, without additional experimental work on lingcod physiology, it is unknown if these differences are biologically significant. The five fatty acid variables (18:1 $\omega$ -9, 16:1 $\omega$ -7, 18:1 $\omega$ -7,  $\omega$ -3: $\omega$ -6, and total FA) tested in logistic regression models were relatively unimportant in explaining the probability of blue flesh, when effects of sex, length, and depth were accounted for. Several of the most important fatty acids for differentiating blue and brown fish (16:0, 18:1 $\omega$ -9, and 20:1 $\omega$ -9) have also been described as the primary sources of metabolic energy in fishes (Tocher 2003). Concentrations of each of these fatty acids were lower in blue lingcod, which might indicate that blue fish are obtaining relatively more energy from catabolization of energy stores. The mono-unsaturated fatty acid 18:1 $\omega$ -9 (oleic acid) was the most important fatty acid in the comparison of blue and brown lingcod, based on both the multivariate and regression analyses. Oleic acid is not a particularly valuable trophic biomarker of specific sources because it is common in most marine organisms, and can commonly be synthesized de novo (Sargent et al. 2002; Tocher 2003). However, high levels of oleic acid or the ratio of 18:1 $\omega$ -9 to 18:1 $\omega$ -7 have been identified as possible markers for carnivory in some zooplankton (Brett 2009); it is unclear if this relationship translates to fishes. Evaluating all 11 fatty acids identified with SIMPER showed that the concentrations of all 4 mono-unsaturated and 3 saturated fatty acids were lower in blue fish. DHA and EPA, which are considered physiologically essential fatty acids in marine fishes (Tocher 2003), were the most important PUFAs in separating blue and brown lingcod, despite the fact that DHA did not differ significantly between blue and brown fish.

Total fatty acid content was significantly lower in blue fish (Fig. 4i), which have 17.5% lower mean total fatty acid concentrations (Table S2) compared to brown fish. Total fatty acids are sometimes used as an indicator of fish condition (e.g., Pethybridge et al. 2015); whether these observed differences are physiologically significant in lingcod is unknown. In addition, we found that blue-fleshed fish had significantly lower concentrations of  $\omega$ -6 fatty acids, which are precursors for the synthesis of eicosanoids, which modulate inflammation and facilitate the immune response in fishes (Bell and Sargent 2003; Arts and Kohler 2009). Excess eicosanoid synthesis in fishes is known to be initiated

by stress or other stimuli, triggering the oxygenation of these fatty acids through various pathways, resulting in ARA-derived eicosanoids which promote inflammation and EPA-derived anti-inflammatory eicosanoids (Arts and Kohler 2009). Blue fish had lower content of 20:4 $\omega$ -6 (ARA) and EPA, which are the primary fatty acids from which eicosanoids are derived (Sargent et al. 1999; Arts and Kohler 2009). Our observation of lower  $\omega$ -6 content in blue fish is still just an emergent coincident pattern; it is unclear if lower  $\omega$ -6 content is causatively linked to the blue flesh, particularly given the weak effects of fatty acids after accounting for size, sex, and depth.

The fatty acids of marine consumers are driven by a combination of diet, physiology, and environmental conditions (Dalsgaard et al. 2003; Xu et al. 2020). In this study, multivariate fatty acid composition differed among fish across regions and flesh colors, but not between sexes. Sex was the most important factor for predicting the probability of blue flesh in the logistic regression analysis, possibly swamping out effects of fatty acids. Further investigation of the role of sex steroids and other factors, such as parasite load, in explaining phenotypic variation in fish coloration may help to better understand why the prevalence of blue flesh varies among sexes. The multivariate results did not differ when we evaluated the fatty acids of only female lingcod. The lack of differences in fatty acids between males and females suggests that lingcod of both sexes have generally similar diets, or that both sexes experience the same physiological response to the underlying driver of blueness. The reason for the three blue outliers on the right side of the nMDS ordination is unknown. Data exploration did not identify any particular explanatory factor (fish length, depth captured, region, sex, etc.) that could account for their anomalous position relative to other blue fish. Despite this variation, the PERMDISP analysis still indicated that brown fish were more variable than blue fish in their fatty acid composition. The fatty acids of lingcod differed across the large geographic range we investigated, which may indicate differences in trophic partitioning of lingcod in the different major eco-regions of the Eastern Pacific. Region did not interact with color in the fatty acid analysis, indicating that blue fish were depleted in key fatty acids compared to brown fish in all regions sampled. Future work using additional tools for trophic inference, including stomach contents and stable isotope analysis, should leverage these initial findings to see if there are multiple lines of evidence to support the fatty acid results, which suggest divergent trophic ecology of the fish across regions. Our analysis was based on the commonly used total lipid extraction on muscle tissue, but analysis of only neutral lipids may be preferable for assessment of diet in a predator (Dalsgaard et al. 2003). We used muscle tissue, as this was consistently collected from the same part of the fish from all lingcod in the latitudinal study; but future work

could also look at other tissues (e.g., adipose tissue or liver) which might better reflect recent diets (Budge et al. 2006).

In addition to the scientific interest in the physiological, ecological, and evolutionary roles that heterogeneous coloration patterns play in animal populations, this topic is of profound interest in the general public. Albino individuals among fish and wildlife species garner media stories in the popular press (Doughton 2007; Sanchez 2019; Dickson 2020). For lingcod, blue individuals are the subject of many forum conversations, and there are many opinions on what causes this color variation among anglers (e.g., <http://www.theoutdoorline.com/forum/>; <https://www.sportfishingbc.com/forum/index.php>). We recently received a confirmed report of a very rare bright-pink lingcod color morph caught by a recreational angler out of Newport Oregon in 2020, which had pink external and internal flesh coloration (unpublished observation reported to AWEG by Oregon Department of Fish and Wildlife). In the marketplace, the coloration of fish fillets also plays a role in the price and demand of seafood products, with blue lingcod fillets “always selling faster than white counterparts” (Bland 2014). This interest has the potential to lend itself to community (‘citizen’) science opportunities. Integrating data collected by recreational anglers on the sex, size, color, depth of capture, and stomach contents for this popular sportfish would not only provide new data across unsampled areas, but would also provide a framework for collaboration between science and stakeholders, which could improve trust in the management decision-making process (Enquist et al. 2017; Venturelli et al. 2017; Andrews et al. 2019).

Our analysis has provided new insights into the prevalence and factors associated with blue flesh for a very large sample of lingcod, across their geographic range. This is, to our knowledge, the largest such analysis of fish flesh coloration ever published. Most published work on color variants in other fishes highlight relatively rare or novel occurrences. These initial results draw attention to several outstanding mysteries. Why is it that female fish are more likely to have blue flesh? Our analysis identifies that sex has been an otherwise overlooked factor that is relevant for blue color in lingcod, given our discovery that > 80% of blue fish are also female. Does reproductive status, which may affect males and females differently, affect the blue coloration? A common hypothesis among fishers and biologists that blue-colored flesh in lingcod is due to diet (Love 2011). Unfortunately, our findings do not resolve this mystery. The differences in fatty acids among blue and brown fish can be taken as support for this hypothesis; however, it is still possible differences in fatty acids are a reflection of some other physiological process besides diet, and this question will not be resolved without lab experiments.

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1007/s00227-021-03936-6>.

**Acknowledgements** We are grateful to G. Williams, K. Nichols, and G. Longo for helping with the logistics and coordination of fishing trips, and to the many people who helped collect these samples, including over 200 volunteer fishers, charter captains and deckhands from the following sportfishing vessels: F/V Chubasco, F/V Tigerfish, F/V New Sea Angler, F/V Hula Girl, F/V Malia Kai, F/V D&D, F/V Ocean Captain, F/V Betty Kay, F/V Tidewinds, F/V Telstar, F/V Samson, F/V Superstar, F/V Reel Steel, F/V Endeavor, F/V Stardust, F/V Toronado, and the F/V Outer Limits, F/V Perseverance, F/V Kingfisher, F/V Jackpot, F/V Sea Raven, F/V Fishfull Thinking II, F/V The Joker, F/V All-StarFishing, and student technicians T. Li, K. Critchlow, and B. Warren. We appreciate the constructive comments from two anonymous reviewers, C. Wood for reviewing an earlier version of the manuscript, and the helpful suggestions on study design and relevant literature from R. Yoshioka and C. Sergeant.

**Author contributions** Study conceptualization: AWEG and AHB; field data collection: BLB, LSL, SLH, KSA, JBS, JW, and JFS; methodology: all the authors; lab work: MDT, BLB, LSL, JBS, and KSA; formal analysis and investigation: AWEG, AHB, and MDT; writing—original draft preparation: AWEG and AHB; writing—review and editing: all the authors; funding acquisition: AWEG, SLH, and JFS.

**Funding** The work was supported by the following funding sources: a State Wildlife Grant (ODFW Marine Reserves Program Research Grant, facilitated by B. Huntington); the National Oceanographic and Atmospheric (NOAA) National Marine Fisheries Service (NMFS) Habitat Assessment Improvement Plan (#NA15-NMF4550353).

**Data availability** The datasets and code generated and analyzed during the current study are available from the corresponding author on reasonable request, as some of the data are still being evaluated by graduate student coauthors for other research.

## Declarations

**Conflict of interest** The authors have no relevant financial or non-financial interests to disclose beyond these grants. The scientific results and conclusions, as well as any views or opinions expressed herein, are those of the authors and do not necessarily reflect the views of NOAA or the Department of Commerce.

**Ethical approval** Lingcod sampling efforts: SJSU IACUC protocol #964, collections were permitted by NOAA SRP-31-2016 and SRP-21-2017, CDFW SCP-6477, ODFW ORSTP #20237 and 21074, and WDFW permit SCP-16-138 and SCP-17-024. Puget Sound serum collections: NOAA SRP 17062-5 M and WDFW ANDREWS 18-122.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

- Anderson MJ, Gorley RN, Clarke KR (2008) PERMANOVA+ for PRIMER: guide to software and statistical methods. PRIMER-E Ltd, Plymouth
- Andrews K, Nichols K, Harvey C, Tolimieri N, Obaza A, Garner R, Tonnes D (2019) All hands on deck: local ecological knowledge and expert volunteers contribute to the first delisting of a marine fish species under the endangered species act. *Citizen Sci* 4:37. <https://doi.org/10.5334/cstp.221>
- Arts MT, Kohler CC (2009) Health and condition in fish: the influence of lipids on membrane competency and immune response. *Lipids in aquatic ecosystems*. Springer, New York, pp 237–255
- Bada JL (1970) A blue-green pigment isolated from blood plasma of the arctic sculpin (*Myoxocephalus scorpioides*). *Experientia* 26:251–252. <https://doi.org/10.1007/BF01900075>
- Bard F-X, Amon Kothias JB, Panfili J, Hervé A (1996) Unusual occurrence of “green tunas” in Abidjan landings, 1993–1995. *Collectiv Volume Sci Papers* 45:126–129
- Bassett M, Lindholm J, Garza C, Kvitek R, Wilson-Vandenberg D (2018) Lingcod (*Ophiodon elongatus*) habitat associations in California: implications for conservation and management. *Environ Biol Fish* 101:203–213. <https://doi.org/10.1007/s10641-017-0692-0>
- Beaudreau AH, Essington TE (2007) Spatial, temporal, and ontogenetic patterns of predation on rockfishes by lingcod. *Trans Am Fish Soc* 136:1438–1452. <https://doi.org/10.1577/T06-236.1>
- Beaudreau AH, Essington TE (2009) Development of a new field-based approach for estimating consumption rates of fishes and comparison with a bioenergetics model for lingcod (*Ophiodon elongatus*). *Can J Fish Aquat Sci* 66:565–578. <https://doi.org/10.1139/F09-021>
- Bell JG, Sargent JR (2003) Arachidonic acid in aquaculture feeds: current status and future opportunities. *Aquaculture* 218:491–499. [https://doi.org/10.1016/S0044-8486\(02\)00370-8](https://doi.org/10.1016/S0044-8486(02)00370-8)
- Bland A (2014) Red fish, blue fish: where the fish flesh rainbow comes from. National Public Radio: <https://www.npr.org/sections/thesalt/2014/06/17/322920045/red-fish-blue-fish-where-the-fish-flesh-rainbow-comes-from>. Accessed on 9 Oct 2020
- Brett MT, Müller-Navarra DC, Persson J (2009) Crustacean zooplankton fatty acid composition. In: Arts MT, Brett MT, Kainz M (eds) *Lipids in Aquatic Ecosystems*. Springer, New York, pp 115–146.
- Budge SM, Iverson SJ, Bowen WD, Ackman RG (2002) Among- and within-species variability in fatty acid signatures of marine fish and invertebrates on the Scotian Shelf, Georges Bank, and southern Gulf of St. Lawrence. *Can J Fish Aquat Sci* 59:886–898
- Budge SM, Iverson SJ, Koopman HN (2006) Studying trophic ecology in marine ecosystems using fatty acids: a primer on analysis and interpretation. *Mar Mamm Sci* 22:759–801
- Burnham KP, Anderson DR (eds) (2002) *Model selection and multimodel inference: a practical information-theoretic approach*. Springer, New York
- Cass AJ, Beamish RJ, McFarlane GA (1990) Lingcod, *Ophiodon elongatus* Canadian Special Publication of Fisheries and Aquatic Sciences 109. 40 p.
- Clark W, Leclercq E, Migaud H, Nairn J, Davie A (2016) Isolation, identification and characterisation of ballan wrasse *Labrus bergylta* plasma pigment. *J Fish Biol* 89:2070–2084. <https://doi.org/10.1111/jfb.13106>
- Clarke KR, Gorley RN (2006) PRIMER v6: user manual/tutorial, 1st edn. PRIMER-E Ltd, Plymouth
- Copeman LA, Parrish CC, Brown JA, Harel M (2002) Effects of docosahexaenoic, eicosapentaenoic, and arachidonic acids on the early growth, survival, lipid composition and pigmentation of yellowtail

- flounder (*Limanda ferruginea*): a live food enrichment experiment. *Aquaculture* 210:285–304. [https://doi.org/10.1016/s0044-8486\(01\)00849-3](https://doi.org/10.1016/s0044-8486(01)00849-3)
- Copeman LA, Laurel BJ, Boswell KM, Sremba AL, Klinck K, Heintz RA, Vollenweider JJ, Helser TE, Spencer ML (2016) Ontogenetic and spatial variability in trophic biomarkers of juvenile saffron cod (*Eleginus gracilis*) from the Beaufort, Chukchi and Bering Seas. *Polar Biol* 39:1109–1126. <https://doi.org/10.1007/s00300-015-1792-y>
- Cornelius CE (1984) Biliverdin in biological systems. In: Ryder OA, Byrd ML (eds) *One medicine: a tribute to Kurt Benirschke, director center for reproduction of endangered species zoological society of San Diego and professor of pathology and reproductive medicine University of California San Diego from his students and colleagues*. Springer, Berlin, pp 321–334
- Dalsgaard J, St. John M, Kattner G, Müller-Navarra DC, Hagen W (2003) Fatty acid trophic markers in the pelagic marine environment. *Adv Mar Biol* 46:225–340
- Dickson C (2020) Rare white orca sighted off Alaskan coast. Canada Broadcast Corporation News. Accessed 2020-08-17
- Doughton S (2007) UW's albino ratfish is 1 in 7 million. *The Seattle Times*. Accessed 2020-01-12
- Ellis MJ, Poluhowich JJ (1981) Biliverdin concentrations in the plasmas of fresh and brackish water eels, *Anguilla rostrata*. *Comp Biochem Physiol A* 70A:587–589
- Enquist CA, Jackson ST, Garfin GM, Davis FW, Gerber LR, Littell JA, Tank JL, Terando AJ, Wall TU, Halpern B, Hiers JK, Morelli TL, McNie E, Stephenson NL, Williamson MA, Woodhouse CA, Yung L, Brunson MW, Hall KR, Hallett LM, Lawson DM, Moritz MA, Nydick K, Pairis A, Ray AJ, Regan C, Safford HD, Schwartz MW, Shaw MR (2017) Foundations of translational ecology. *Front Ecol Environ* 15:541–550. <https://doi.org/10.1002/fee.1733>
- Fang LS (1987) Study of the heme catabolism of fish. *Comp Biochem Physiol B* 88:667–673. [https://doi.org/10.1016/0305-0491\(87\)90361-0](https://doi.org/10.1016/0305-0491(87)90361-0)
- Fang LS, Bada JL (1990) The blue-green blood plasma of marine fish. *Comp Biochem Physiol B* 97:37–45. [https://doi.org/10.1016/0305-0491\(90\)90174-R](https://doi.org/10.1016/0305-0491(90)90174-R)
- Folstad I, Karter AJ (1992) Parasites, bright males, and the immunocompetence handicap. *Am Nat* 139:603–622. <https://doi.org/10.1086/285346>
- Fontaine M (1941) A propos du pigment vert de l'Orphie (*Belone belone* L.). *Bull Inst Oceanogr Monaco* 793:1–3
- Foo YZ, Nakagawa S, Rhodes G, Simmons LW (2017) The effects of sex hormones on immune function: a meta-analysis. *Biol Rev* 92:551–571. <https://doi.org/10.1111/brv.12243>
- Fujii R (1993) Cytophysiology of fish chromatophores. In: Jeon KW, Friedlander M, Jarvik J (eds) *International review of cytology*. Academic Press, Cambridge, pp 191–255
- Gagnon MM (2006) Serum biliverdin as source of colouration upon sexual maturation in male blue-throated wrasse *Notolabrus tetricus*. *J Fish Biol* 68:1879–1882. <https://doi.org/10.1111/j.0022-1112.2006.01033.x>
- Galloway AWE, Budge SM (2020) The critical importance of experimentation in biomarker-based trophic ecology. *Philos Trans R Soc B* 375:20190638. <https://doi.org/10.1098/rstb.2019.0638>
- Gelman A (2008) Scaling regression inputs by dividing by two standard deviations. *Stat Med* 27:2865–2873. <https://doi.org/10.1002/sim.3107>
- Ghosh S, Yu C-L, Ferraro DJ, Sudha S, Pal SK, Schaefer WF, Gibson DT, Ramaswamy S (2016) Blue protein with red fluorescence. *Proc Natl Acad Sci* 113:11513–11518. <https://doi.org/10.1073/pnas.1525622113>
- Goodman WG, Adams B, Trost JT (1985) Purification and characterization of a biliverdin-associated protein from the hemolymph of *Manduca sexta*. *Biochemistry* 24:1168–1175. <https://doi.org/10.1021/bi00326a017>
- Greenley A, Green K, Starr RM (2016) Seasonal and ontogenetic movements of lingcod (*Ophiodon elongatus*) in central California, with implications for marine protected area management. *Calif Cooperative Ocean Fisheries Investigations Rep* 57:71–88
- Grismer LL, Thy N, Chav T, Holden J (2007) A new species of *Chiromantis* Peters 1854 (Anura: Rhacophoridae) from Phnom Samkos in the Northwestern Cardamom Mountains, Cambodia. *Herpetologica* 63:392–400
- Huntington BE, Watson JL (2017) Tailoring ecological monitoring to individual marine reserves: comparing longline to hook-and-line gear to monitor fish species. *Marine and Coastal Fisheries* 9:432–440. <https://doi.org/10.1080/19425120.2017.1360419>
- Huntington BE, Watson J, Matteson K, McIntosh N, Wagman D, Pierson K, Don C, Fox D, Marion SR, Groth S (2014) Oregon marine reserves ecological monitoring report 2010–11. Newport, OR, USA. [https://oregonmarinereserves.com/content/uploads/2016/02/EcologicalMonitoring\\_Report\\_ODFW\\_2014.pdf](https://oregonmarinereserves.com/content/uploads/2016/02/EcologicalMonitoring_Report_ODFW_2014.pdf)
- Kainz MJ, Hager HH, Rasconi S, Kahilainen KK, Amundsen P-A, Hayden B (2017) Polyunsaturated fatty acids in fishes increase with total lipids irrespective of feeding sources and trophic position. *Ecosphere* 8:e01753. <https://doi.org/10.1002/ecs2.1753>
- Leclercq E, Taylor JF, Migaud H (2010) Morphological skin colour changes in teleosts. *Fish Fish* 11:159–193. <https://doi.org/10.1111/j.1467-2979.2009.00346.x>
- Love RM (1980) *The chemical biology of fishes*. Volume 2: advances 1968–1977. Academic Press, London
- Love MS (2011) *Certainly more than you want to know about the fishes of the Pacific Coast*, 1st edn. Really Big Press, Santa Barbara, CA, USA
- Low CJ, Beamish RJ (1978) A study of the nesting behavior of Lingcod (*Ophiodon elongatus*) in the Strait of Georgia, British Columbia. Fisheries and Marine Service Technical Report 843. Nanaimo, BC, Canada.
- Low PS, Bada JL (1974) Bile pigments in the blood serum of fish from the family Cottidae. *Comp Biochem Physiol A Physiol* 47:411–418. [https://doi.org/10.1016/0300-9629\(74\)90003-6](https://doi.org/10.1016/0300-9629(74)90003-6)
- Miller DJ, Geibel JJ (1973) Summary of blue rockfish and lingcod life histories; a reef ecology study; and giant kelp, *Macrocystis pyrifera*, experiments in Monterey Bay, California. *State Calif Dep Fish Game Fish Bull* 158:1–135
- Miller DJ, Lea RN (1972) Guide to the coastal marine fishes of California. *Fish Bull* 157:1–249
- Nomura T (1967) Bile pigments of fish. *Bulletin of the Japanese Society of Scientific Fisheries* 33:860–865. <https://doi.org/10.2331/suisan.33.860>
- Parrish CC (2013) Lipids in marine ecosystems. *Int Sch Res Notices*. <https://doi.org/10.5402/2013/604045> (Article ID 604045)
- Pethybridge HR, Parrish CC, Morriongiello J, Young JW, Farley JH, Gunasekera RM, Nichols PD (2015) Spatial patterns and temperature predictions of tuna fatty acids: tracing essential nutrients and changes in primary producers. *PLoS ONE* 10:e0131598. <https://doi.org/10.1371/journal.pone.0131598>
- Pethybridge HR, Choy CA, Polovina JJ, Fulton EA (2018) Improving marine ecosystem models with biochemical tracers. *Ann Rev Mar Sci* 10:199–228. <https://doi.org/10.1146/annurev-marine-121916-063256>
- Phillips JB (1958) A review of the lingcod, *Ophiodon elongatus*. *Calif Fish Game* 44:19–27
- Price AC, Weadick CJ, Shim J, Rodd FH (2008) Pigments, patterns, and fish behavior. *Zebrafish* 5:297–307. <https://doi.org/10.1089/zeb.2008.0551>
- Prince EE (1916) On the red color of the flesh in the salmon and trouts. *Trans Am Fish Soc* 46:50–61. [https://doi.org/10.1577/1548-8659\(1916\)46\[50:OTRCOT\]2.0.CO;2](https://doi.org/10.1577/1548-8659(1916)46[50:OTRCOT]2.0.CO;2)

- R Development Core Team (2021) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL: <http://www.R-project.org/>. Accessed on February 1, 2021
- Roberts ML, Buchanan KL, Evans MR (2004) Testing the immunocompetence handicap hypothesis: a review of the evidence. *Anim Behav* 68:227–239. <https://doi.org/10.1016/j.anbehav.2004.05.001>
- Rodriguez ZB, Perkins SL, Austin CC (2018) Multiple origins of green blood in New Guinea lizards. *Sci Adv* 4:eaa05017. <https://doi.org/10.1126/sciadv.aao5017>
- Sanchez C (2019) Unmasking the legend of Lake Merritt's extinct albino raccoons. Oakland Magazine, Oakland. <https://www.oaklandmagazine.com/July-2019/Unmaskingthe-Legend-of-Lake-Merritts-Extinct-Albino-Raccoons/>
- Sargent JR, Bell G, McEvoy L, Tocher DR, Estevez A (1999) Recent developments in the essential fatty acid nutrition in fish. *Aquaculture* 177:191–199
- Sargent JR, Tocher DR, Bell JG (2002) The lipids. In: Halver JE, Hardy RW (eds) *Fish Nutrition*, 3rd edn. Academic Press, San Diego CA, pp 181–257
- Schaefer WF, Schmitz MH, Blazer VS, Ehlinger TJ, Berges JA (2014) Localization and seasonal variation of blue pigment (sandercyanin) in walleye (*Sander vitreus*). *Can J Fish Aquat Sci*. <https://doi.org/10.1139/cjfas-2014-0139>
- Starr RM, O'Connell V, Ralston S (2004) Movements of lingcod (*Ophiodon elongatus*) in southeast Alaska: potential for increased conservation and yield from marine reserves. *Can J Fish Aquat Sci* 61:1083–1094. <https://doi.org/10.1139/f04-054>
- Taipale S, Strandberg U, Peltomaa E, Galloway AWE, Ojala A, Brett MT (2013) Fatty acid composition as biomarkers of freshwater microalgae: analysis of 37 strains of microalgae in 22 genera and in seven classes. *Aquat Microb Ecol* 71:165–178. <https://doi.org/10.3354/ame01671>
- Takagi S, Murata H, Goto T, Hatate H, Endo M, Yamashita H, Miyatake H, Ukawa M (2010) Necessity of dietary taurine supplementation for preventing green liver symptom and improving growth performance in yearling red sea bream *Pagrus major* fed non-fishmeal diets based on soy protein concentrate. *Fish Sci* 76:119. <https://doi.org/10.1007/s12562-009-0185-y>
- Thomas MD, Schram JB, Clark-Henry ZF, Yednock BK, Shanks AL, Galloway AWE (2020) Juvenile Dungeness crabs (*Metacarcinus magister*) selectively integrate and modify the fatty acids of their experimental diets. *Philos Transactions R Soc B* 375:20200038. <https://doi.org/10.1098/rstb.2020.0038>
- Tocher DR (2003) Metabolism and functions of lipids and fatty acids in teleost fish. *Rev Fish Sci* 11:107–184. <https://doi.org/10.1080/713610925>
- Torrissen OJ, Naevdal G (1988) Pigmentation of salmonids—variation in flesh carotenoids of Atlantic salmon. *Aquaculture* 68:305–310. [https://doi.org/10.1016/0044-8486\(88\)90244-X](https://doi.org/10.1016/0044-8486(88)90244-X)
- Tunison AV, Brockway DR, Maxwell JM, Dorr AL, Mccay CM (1947) Coloration of fish. *Prog Fish-Culturist* 9:53–61. [https://doi.org/10.1577/1548-8640\(1947\)9\[53:COF\]2.0.CO;2](https://doi.org/10.1577/1548-8640(1947)9[53:COF]2.0.CO;2)
- van Holde KE, Miller KI, Decker H (2001) Hemocyanins and invertebrate evolution. *J Biol Chem* 276:15563–15566. <https://doi.org/10.1074/jbc.R100010200>
- Venturelli PA, Hyder K, Skov C (2017) Angler apps as a source of recreational fisheries data: opportunities, challenges and proposed standards. *Fish Fish* 18:578–595. <https://doi.org/10.1111/faf.12189>
- Wendt DE, Starr RM (2009) Collaborative research: an effective way to collect data for stock assessments and evaluate marine protected areas in California. *fidm* 2009:315–324. <https://doi.org/10.1577/C08-054.1>
- Wood CL, Leslie KL, Greene A, Lam L, Basnett B, Hamilton S, Samhoury JF (in review) The weaker sex: Male lingcod (*Ophiodon elongatus*) with blue color polymorphism are more burdened by parasites than are other sex–color combinations
- Xu H, Turchini GM, Francis DS, Liang M, Mock TS, Rombenso A, Ai Q (2020) Are fish what they eat? A fatty acid's perspective. *Prog Lipid Res* 80:101064. <https://doi.org/10.1016/j.plipres.2020.101064>
- Yamaguchi K, Matura F (1969) A blue pigment from the muscle of a marine teleost, “Hiroa”, *Cheilinus undulatus* Ruppell. *Bull Jpn Soc Sci Fish* 35:920–926
- Yamaguchi K, Kochiyama Y, Hashimoto K, Matsuura F (1966) Studies on a blue-green serum pigment of eel—II. Identification of prosthetic group. *日本水産学会誌* NSUGAF 32:873–880. <https://doi.org/10.2331/suisan.32.873>
- Yamaguchi K, Hashimoto K, Matsuura F (1976) Identity of blue pigments obtained from different tissues of the sculpin, *Pseudoblennius percoides* Günther. *Comp Biochem Physiol B* 55:85–87

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.