

# **NOAA Technical Memorandum NMFS**

**JANUARY 2023**

# **DISTRIBUTION, BIOMASS, AND DEMOGRAPHICS OF COASTAL PELAGIC FISHES IN THE CALIFORNIA CURRENT ECOSYSTEM DURING SPRING 2021 BASED ON ACOUSTIC-TRAWL SAMPLING**

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NOAA-TM-NMFS-SWFSC-675

U.S. DEPARTMENT OF COMMERCE National Oceanic and Atmospheric Administration National Marine Fisheries Service Southwest Fisheries Science Center

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#### **Recommended citation**

Zwolinski, Juan P., Josiah S. Renfree, Kevin L. Stierhoff, and David A. Demer. 2023. Distribution, biomass, and demographics of coastal pelagic fishes in the California Current Ecosystem during spring 2021 based on acoustic-trawl sampling. U.S. Department of Commerce, NOAA Technical Memorandum NMFS-SWFSC-675. <https://doi.org/10.25923/zvzf-3306>

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## <span id="page-7-0"></span>**Executive Summary**

This report provides: 1) a detailed description of the acoustic-trawl method used by NOAA's Southwest Fisheries Science Center (SWFSC) for direct assessments of the dominant coastal pelagic fish species (CPS), i.e.: Pacifc Sardine *Sardinops sagax*, Northern Anchovy *Engraulis mordax*, Pacifc Mackerel *Scomber japonicus*, and Jack Mackerel *Trachurus symmetricus* in the California Current Ecosystem (CCE) of the west coast of North America; and 2) estimates of the biomasses, distributions, and demographics of those CPS encountered in the survey area between 20 March and 13 April 2021. The core survey region, which was sampled by NOAA Ship *Reuben Lasker* (hereafter, *Lasker*), spanned most of the continental shelf between San Diego and San Francisco, California (CA). Throughout the core region, *Lasker* sampled along transects oriented approximately perpendicular to the coast, from the shallowest navigable depth  $(\sim 30 \text{ m})$  to offshore distances of ~75 nmi in the Southern CA Bight (SCB) and ~35 nmi between Point Conception and San Francisco. To estimate the CPS biomasses in the nearshore region, where sampling by *Lasker* was deemed inefficient, unsafe, or both, fishing vessel  $(F/V)$  *Long Beach Carnage* sampled to  $\sim$ 10 m depth along 5-nmilong transects spaced 5 nmi apart between Point Conception and San Diego, and around Santa Cruz and Santa Catalina Islands in the SCB.

The biomasses, distributions, and demographics for each species and stock are for the survey area and period, and therefore may not represent the entire populations.

The estimated biomass of the central stock of Northern Anchovy was  $1,370,303$  t (CI<sub>95%</sub> = 1,076,749 -1,959,163 t,  $CV = 16\%$ ). In the core region, sampled by *Lasker*, biomass was 1,358,587 t ( $CI_{95\%} = 1,070,094$ ) - 1,940,986 t, CV = 17%). In the nearshore region, sampled by *Long Beach Carnage*, biomass was 11,716 t ( $CI_{95\%} = 6,655$  - 18,176 t, CV = 26%), which was 0.86% of the total biomass. The central stock ranged from approximately Monterey Bay to San Diego, and its distribution of standard length (*LS*) ranged from 8 to 16 cm with a mode between 10 and 12 cm.

The estimated biomass of the southern stock of Pacifc Sardine in U.S. waters, distributed from approximately Point Conception to San Diego, was 24,547 t ( $CI_{95\%} = 7,697 - 38,339$  t,  $CV = 29\%$ ). Only 6% of the total biomass, 1,504 t (CI<sub>95%</sub> = 556 - 2,959 t, CV = 40%), was in the core region. The remaining 94% (23,043 t,  $CI_{95\%} = 7,140 - 35,380$  t,  $CV = 31\%$ , was in the nearshore region. Its  $L<sub>S</sub>$  distribution was predominately between 12 to 15 cm with a mode at 10 cm.

The core survey area included some "good" and "optimal" habitat for the northern stock of Pacifc Sardine, particularly north of Point Conception. However, the Pacifc Sardine sampled in the SCB were exclusively in the nearshore region and around the northern Channel Islands, which had unsuitable habitat due to excessive chlorophyll-a concentration. Therefore, the Pacifc Sardine observed in the spring 2021 survey were attributed to the southern stock.

The estimated biomass of Pacific Mackerel was  $92.3 \text{ t}$  (CI<sub>95%</sub> = 7.43 - 221 t, CV = 64%), all observed in the nearshore region. Pacifc Mackerel ranged from approximately Oceanside to San Diego and fork length  $(L_F)$  ranged from 14 to 25 cm with a mode at 18 cm.

The estimated biomass of Jack Mackerel was 16,882 t (CI<sub>95%</sub> = 3,783 - 41,209 t, CV = 57%), all in the core region. Jack Mackerel ranged from approximately Point Conception to San Diego and *L<sup>F</sup>* ranged from 9 to 23 cm with a mode at 13 cm.

The total estimated biomass of four stocks (four species) within the survey area was 1,411,825 t. Of this, 97% (1,370,303 t) was attributed to the central stock of Northern Anchovy. Contributions by other stocks were southern stock of Pacifc Sardine (1.7%), Jack Mackerel (1.2%), and Pacifc Mackerel (0.01%).

## <span id="page-8-0"></span>**1 Introduction**

In the California Current Ecosystem (CCE), five coastal pelagic fish species (CPS; i.e.: Pacific Sardine *Sardinops sagax*, Northern Anchovy *Engraulis mordax*, Jack Mackerel *Trachurus symmetricus*, Pacifc Mackerel *Scomber japonicus*, and Pacifc Herring *Clupea pallasii*) comprise the bulk of the forage fsh assemblage. The biomasses of these populations can change by an order of magnitude within a few years and represent important prey for marine mammals, birds, and larger migratory fshes (Field *et al.*, 2001), and are targets of commercial fsheries.

During summer and fall, the northern stock of Pacifc Sardine typically migrates north to feed in the productive coastal upwelling areas of Oregon (OR), Washington (WA), and Vancouver Island (Zwolinski *et al.*, 2012, and references therein). The predominantly piscivorous adult Pacifc and Jack Mackerels also migrate north in summer, but go farther ofshore to feed (Zwolinski *et al.*, 2014, and references therein). In the winter and spring, the northern stock of Pacifc Sardine stock typically migrates south to its spawning grounds, generally of central and southern CA (Demer *et al.*, 2012) and occasionally of OR and WA (Lo *et al.*, 2011). These migrations vary in extent with population size; fsh age and length; and oceanographic conditions. For example, the transition zone chlorophyll front (TZCF, Polovina *et al.*, 2001) may delineate the ofshore and southern limits of Pacifc Sardine and Pacifc Mackerel habitats (e.g., Demer *et al.*, 2012; Zwolinski *et al.*, 2012), and juveniles may have nursery areas in and ofshore of the SCB, downstream of upwelling regions. In contrast, Northern Anchovy spawn predominantly during winter, mostly within the SCB where seasonal down-welling increases retention of their eggs and larvae (Bakun and Parrish, 1982). Pacifc Herring spawn in intertidal beach areas (Love, 1996). The northern stock of Northern Anchovy is located of WA and OR and the central stock is located off Central and Southern CA. Whether a species migrates or remains in an area depends on its stock size, reproductive and feeding behaviors, and afnity to certain oceanographic or seabed habitats.

Acoustic-trawl method (ATM) surveys, which combine information collected with echosounders and nets, were introduced to the CCE more than 48 years ago to survey CPS of the west coast of the U.S. (Mais, 1974, 1977; Smith, 1978). Following a two-decade hiatus, the ATM was reintroduced in the CCE in spring 2006 to sample the then abundant Pacifc Sardine population (Cutter and Demer, 2008). Since then, this sampling efort has continued and expanded through annual or semi-annual surveys (Zwolinski *et al.*, 2014). Beginning in 2011, the ATM estimates of Pacifc Sardine abundance, age structure, and distribution have been incorporated in the annual assessments of the northern stock (Hill *et al.*, 2017). Additionally, ATM survey results are applied to estimate the abundances, demographics, and distributions of epipelagic and semi-demersal fshes (e.g., Swartzman, 1997; Williams *et al.*, 2013; Zwolinski *et al.*, 2014) and plankton (Hewitt and Demer, 2000).

This document describes, in detail, the ATM as presently used by NOAA's Southwest Fisheries Science Center (SWFSC) to survey the distributions and abundances of CPS and their oceanographic environments (e.g., Cutter and Demer, 2008; Demer *et al.*, 2012; Zwolinski *et al.*, 2014). In general terms, the contemporary ATM combines information from satellite-sensed oceanographic conditions, calibrated multi-frequency echosounders, probe-sampled oceanographic conditions, pump-sampled fsh eggs, and trawl-net catches of juvenile and adult CPS. The survey area is initially planned with consideration to the expected distribution of a priority stock or stock assemblage, in this case, the central stock of Northern Anchovy (**Fig. [1](#page-9-0)**). As time permits, the survey area is further expanded to encompass as much of the potential distribution as possible for other CPS present off the U.S. West Coast.

Along transects in the survey area, multi-frequency split-beam echosounders transmit sound pulses downward beneath the ship and receive echoes from animals and the seabed in the path of the sound waves. Measurements of sound speed and absorption from conductivity-temperature-depth (CTD) probes allow accurate compensation of these echoes for propagation losses. The calibrated echo intensities, normalized to the range-dependent observational volume, provide indications of the target type and behavior (e.g., Demer *et al.*, 2009).

<span id="page-9-0"></span>

Figure 1: Distribution of eggs of Northern Anchovy sampled by the CUFES during spring surveys from 2017 through 2019. Eggs, indicating the presence of spawning Northern Anchovy, were used to delineate the spring 2021 survey area.

Echoes from marine organisms are a function of their body composition, shape, and size relative to the sensing-sound wavelength, and their orientation relative to the incident sound waves (Cutter *et al.*, 2009; Demer *et al.*, 2009; Renfree *et al.*, 2009). Variations in echo intensity across frequencies, known as echo spectra, indicate the taxonomic groups contributing to the echoes. The CPS, with highly refective swim bladders, create high intensity echoes of sound pulses at all echosounder frequencies (e.g., Conti and Demer, 2003). In contrast, krill, with acoustic properties closer to those of the surrounding sea-water, produce lower intensity echoes, particularly at lower frequencies (e.g., Demer *et al.*, 2003). The echo energy attributed to CPS, based on empirical echo spectra (Demer *et al.*, 2012), are apportioned to species using trawl-catch proportions (Zwolinski *et al.*, 2014).

Animal densities are estimated by dividing the summed intensities attributed to a species by the lengthweighted average echo intensity (the mean backscattering cross-section) from animals of that species (e.g., Demer *et al.*, 2012). Transects with similar densities are grouped into post-sampling strata that mimic the natural patchiness of the target species (e.g., Zwolinski *et al.*, 2014). An estimate of abundance is obtained by multiplying the average estimated density in the stratum by the stratum area (Demer *et al.*, 2012). The associated sampling variance is calculated using non-parametric bootstrap of the mean transect densities. The total abundance estimate in the survey area is the sum of abundances in all strata. Similarly, the total variance estimate is the sum of the variance in each stratum.

In spring 2021, the ATM survey performed aboard NOAA Ship *Reuben Lasker* (hereafter, *Lasker*) was augmented with coordinated sampling by a fshing vessel (F/V *Long Beach Carnage*) to estimate the biomasses of CPS in nearshore regions where sampling by *Lasker* was not possible or safe.

Presented here are: 1) a detailed description of the ATM used to survey CPS in the CCE off the west coast of North America; and 2) estimates of the abundance, biomass, size structure, and distribution of CPS, specifcally the central stock of Northern Anchovy, southern stock of Pacifc Sardine, Pacifc Mackerel, and Jack Mackerel for the core and nearshore survey regions. Additional details about the survey may be found in the survey report (Zwolinski *et al.*, 2023).

## <span id="page-10-0"></span>**2 Methods**

## <span id="page-10-1"></span>**2.1 Sampling**

#### <span id="page-10-2"></span>**2.1.1 Design**

The spring 2021 survey was conducted principally using *Lasker*. The sampling domain, or core region, between San Francisco and San Diego, was defned by the expected distribution of the central stock of Northern Anchovy (**Fig. [1](#page-9-0)**), which also includes a portion of the potential habitat of the northern stock of Pacifc Sardine (**Fig. [2a](#page-11-0)**), predominantly north of Point Conception. Habitat unsuitable for the northern stock may be suitable for the southern stock of Pacifc Sardine (Demer and Zwolinski, 2014). East to west, the sampling domain extends from the coast to at least the 1,000 ftm (~1830 m) isobath (**Fig. [3](#page-12-0)**). Considering the expected distribution of the target species, the acceptable uncertainty in biomass estimates, and the available ship time (25 days at sea, DAS), the principal survey objective was the estimations of biomass for the central stock of Northern Anchovy in the survey region. Additionally, biomass estimates were sought for the other CPS in the survey domain, including: Pacifc Sardine, Pacifc Mackerel, and Jack Mackerel.

<span id="page-11-0"></span>

Figure 2: Distribution of a) potential habitat for the northern stock of Pacifc Sardine (Zwolinski *et al.*, 2011), which is <sup>a</sup> function of b) chlorophyll-a concentration and c) sea surface temperature (SST). The images are 8-day composites including data from <sup>23</sup> March to <sup>1</sup> April, 2021. The potential habitat categories are based on data from spring CalCOFI surveys conducted between 1998 and 2009 and refect the probability of fnding at least one Pacifc Sardine egg on <sup>a</sup> standard CUFES sample. The good and optimal habitat areas collectively include 90% of the Pacifc Sardine biomass, and the bad and unsuitable areas contain the remaining 10%.

In the core region north of Point Conception, the transects were spaced 20 nmi apart, and extend from the shallowest navigable depth  $({\sim}30 \text{ m})$  to either a distance of  ${\sim}35 \text{ nm}$  offshore or to the 1,000-fathom isobath, whichever is farthest (**Fig. [3](#page-12-0)**). South of Point Conception, the transects were spaced 15 nmi apart, and were as long as 70 nmi. Throughout the core region, where CPS were observed within the westernmost 3 nmi of the transect, that transect and the next one to the north were extended in 5-nmi increments until no CPS were observed in the echograms in the last 3 nmi of the extension. The aim was to reach the end of the transects with a practically zero biomass density.

<span id="page-12-0"></span>Additional sampling was conducted in the nearshore region along 5-nmi-long transects spaced 5 nmi apart between Point Conception and San Diego using *Long Beach Carnage* equipped with multi-frequency echosounders (magenta lines, **Fig. [3](#page-12-0)**). The goal of the nearshore sampling was to estimate the abundance and biomass of CPS close to shore, in shallow water, or both, where *Lasker* could not safely navigate or trawl.



Figure 3: Planned compulsory (black lines) transect lines sampled by *Lasker*; nearshore transect lines sampled by *Long Beach Carnage* (magenta lines); and 50-, 200-, 500-, and 2,000-m isobaths (gray lines).

#### <span id="page-13-0"></span>**2.1.2 Acoustic**

#### **2.1.2.1 Acoustic equipment**

On *Lasker*, multi-frequency Wide-Bandwidth Transceivers (Simrad 18-, 38-, 70-, 120-, 200-, and 333-kHz EK80 WBTs; Kongsberg) were confgured with split-beam transducers (Simrad ES18-11, ES38B, ES70- 7C, ES120-7C, ES200-7C, and ES333-7C, respectively; Kongsberg). The transducers were mounted on the bottom of a retractable keel or "centerboard" (**Fig. [4](#page-13-1)**). The keel was retracted (transducers ~5-m depth) during calibration, and extended to the intermediate position (transducers ~7-m depth) during the survey. Exceptions were made during shallow water operations, when the keel was retracted; or during times of heavy weather, when the keel was extended (transducers  $\sim$ 9-m depth) to provide extra stability and reduce the efect of weather-generated noise. In addition, acoustic data were also collected using a multibeam echosounder (Simrad ME70; Kongsberg), multibeam sonar (Simrad MS70, Kongsberg), and scanning sonar (Simrad SX90; Kongsberg). Transducer position and motion were measured at 5 Hz using an inertial motion unit (Applanix POS-MV; Trimble).

On *Long Beach Carnage*, the SWFSC's multi-frequency echosounders (Simrad 38-, 70-, 120-, and 200-kHz EK60 GPTs; Kongsberg) were confgured with the SWFSC's multi-frequency transducer array (MTA4) with split-beam transducers (Simrad ES38-12, ES70-7C, ES120-7C and ES200-7C; Kongsberg) mounted on the bottom of a pole(**Fig. [5](#page-14-0)**).

<span id="page-13-1"></span>

Figure 4: Echosounder transducers mounted on the bottom of the retractable centerboard on *Lasker*. During the survey, the centerboard was extended, typically positioning the transducers  $\sim$ 2 m below the keel at a water depth of  $\sim$ 7 m.

<span id="page-14-0"></span>

Figure 5: Transducers (Top-bottom: Simrad ES200-7C, ES120-7C, ES38-12, and ES70-7C, Kongsberg) in a pole-mounted multi-transducer array (MTA4) installed on the *Long Beach Carnage*.

#### **2.1.2.2 Echosounder calibrations**

#### **2.1.2.2.1** *Lasker*

The echosounder systems aboard *Lasker* were calibrated on 1 March while the vessel was docked at 10th Avenue Marine Terminal (32.6956 ◦N, -117.15278 ◦W) using the standard sphere technique (Demer *et al.*, 2015; Foote *et al.*, 1987). Each WBT was calibrated in both CW (continuous wave or narrowband mode) and FM mode (i.e., frequency modulation or broadband mode). The principle reference target was a 38.1-mm diameter sphere made from tungsten carbide (WC) with 6% cobalt binder material (WC38.1; *Lasker* sphere #1). Calibrations of WBTs in FM mode used both the WC38.1 and a 25-mm diameter WC sphere. A CTD was cast to measure temperature and salinity versus depth, to estimate sound speeds at the transducer and sphere depths, and the time-averaged sound speed and absorption coefficients for the range between them. The theoretical target strength  $(TS; dB \text{ re } 1 \text{ m}^2)$  of the sphere was calculated using the Standard Sphere Target Strength Calculator<sup>[1](#page-14-1)</sup> and values for the sphere, sound-pulse, and seawater properties. The sphere was positioned throughout the main lobe of each of the transducer beams using three motorized downriggers, two on one side of the vessel and one on the other. The GPTs and WBTs were confgured using the calibration results via the control software (Simrad EK80 v1.12.2; Kongsberg; **Table [1](#page-15-0)**). Calibration results for WBTs in FM mode are presented in the survey report (Stierhoff *et al.*, 2020a).

<span id="page-14-1"></span><sup>1</sup><https://swfscdata.nmfs.noaa.gov/AST/SphereTS/>

<span id="page-15-0"></span>Table 1: Wide-Bandwidth Transceiver (Simrad EK80 WBT; Kongsberg) information, pre-calibration settings, and beam model results following calibration (below the horizontal line). Prior to the survey, on-axis gain  $(G_0)$ , beam angles and angle offsets, and  $S_A$  Correction  $(S_A \text{corr})$  values from calibration results were entered into the WBT control software (Simrad EK80 v1.12.2; Kongsberg).

		Frequency (kHz)					
	Units	18	38	70	120	200	333
Model		<b>ES18</b>	<b>ES38-7</b>	<b>ES70-7C</b>	<b>ES120-7C</b>	<b>ES200-7C</b>	ES333-7C
Serial Number		2106	337	233	783	513	124
Transmit Power $(p_{\text{et}})$	W	1000	1000	600	<b>200</b>	90	35
Pulse Duration $(\tau)$	ms	1.024	1.024	1.024	1.024	1.024	1.024
Eq. Two-way Beam Angle $(\Psi)$	$dB$ re $1$ sr	$-17$	$-20.7$	$-20.7$	$-20.7$	$-20.7$	$-20.7$
On-axis Gain $(G_0)$	$dB$ re $1$	23.1	26.27	27.63	26.79	27.14	26.59
$S_{\rm a}$ Correction ( $S_{\rm a}$ corr)	$dB$ re $1$	$-0.0267$	0.0471	$-0.0081$	$-0.037$	$-0.0755$	$-0.1435$
<b>RMS</b>	dВ	0.039	0.047	0.04	0.032	0.079	0.11
3-dB Beamwidth Along. $(\alpha_{-3dB})$	deg	10.37	6.41	6.73	6.6	6.57	6.55
3-dB Beamwidth Athw. $(\beta_{-3dB})$	$\deg$	10.42	6.42	6.71	6.6	6.57	6.62
Angle Offset Along. $(\alpha_0)$	deg	$-0.06$	$\Omega$	$-0.03$	$-0.02$	$-0.03$	0.06
Angle Offset Athw. $(\beta_0)$	$\deg$	$-0.02$	$-0.04$	$-0.03$	0.01	$\overline{0}$	0.03

#### **2.1.2.2.2** *Long Beach Carnage*

The echosounders were calibrated using a WC38.1 sphere in a tank at the SWFSC. Beam model results were entered into the GPT-control software and are presented in **Table [2](#page-15-1)**.

<span id="page-15-1"></span>Table 2: General Purpose Transceiver (Simrad EK60 GPT; Kongsberg) beam model results estimated from a tank calibration of echosounders aboard *Long Beach Carnage* using a WC38.1. Prior to the survey, calibrated on-axis gain  $(G_0)$ , beam angles and angle offsets, and  $S_a$  Correction  $(S_a$ corr) values were entered into the GPT-control software (Simrad EK80 v1.12.2; Kongsberg).



#### **2.1.2.3 Data collection**

Computer clocks were synchronized with the GPS clock (UTC) using synchronization software (NetTime[2](#page-16-1)). The 18-kHz GPT, operated by a separate PC from the other echosounders, was programmed to track the seabed and output the detected depth to the ship's Scientifc Computing System (SCS). The echosounders were controlled by the ER60 Adaptive Logger (EAL<sup>[3](#page-16-2)</sup>, Renfree and Demer, 2016). The EAL optimizes the pulse interval based on the seabed depth, while avoiding aliased seabed echoes, and was programmed such that once an hour the echosounders would operate in passive mode and record three pings, for obtaining estimates of the background noise level. The echosounders collected data continuously throughout the survey, but transect sampling was conducted only during daylight hours, approximately between sunrise and sunset.

Measurements of volume backscattering strength  $(S_V; dB \rvert H \rvert^2 m^{-3})$  and *TS* (dB re 1 m<sup>2</sup>), indexed by time and geographic positions provided by GPS receivers, were logged to 60 m beyond the detected seabed range or to a maximum of 350 m, and stored in Simrad .raw format with a 50-MB maximum fle size. During daytime, the echosounders were set to operate in CW mode to remain consistent with echo integration methods used during prior surveys and to reduce data volume; at nighttime, echosounders were set to FM mode to improve *T S* estimation and species diferentiation for CPS near the surface. For each acoustic instrument, the prefx for the fle names is a concatenation of the survey name (e.g., 2103RL), the operational mode (CW or FM), and the logging commencement date and time from the EK80 software. For example, fle generated by the Simrad EK80 software for a WBT operated in CW mode is named 2103RL-CW-D20210401-T125901.raw.

To minimize acoustic interference, transmit pulses from the EK80, ME70, MS70, SX90, and acoustic Doppler current profler (ADCP; Ocean Surveyor Model OS75, Teledyne RD Instruments) were triggered using a synchronization system (K-Sync, Simrad). The K-Sync trigger rate, and thus echosounder ping interval, was modulated by the EAL using the 18-kHz seabed depth provided by the SCS. During daytime, the ME70, SX90, and ADCP were operated continuously, while the MS70 was only operated at times when CPS were present. At nighttime, only the EK80 and ADCP were operated. All other instruments that produce sound within the echosounder bandwidths were secured during daytime survey operations. Exceptions were made during stations (e.g., plankton sampling and fish trawling) or in shallow water when the vessel's command occasionally operated the bridge's 50- and 200-kHz echosounders (Furuno), the Doppler velocity log (Sperry Marine Model SRD-500A), or both. Data from the ME70, MS70, and SX90 are not presented in this report.

#### <span id="page-16-0"></span>**2.1.3 Oceanographic**

#### <span id="page-16-4"></span>**2.1.3.1 Conductivity and temperature versus depth (CTD) sampling**

Conductivity and temperature were measured versus depth to  $350 \text{ m}$  (or to within  $\sim 10 \text{ m}$  of the seabed when less than 350 m) with calibrated sensors on a CTD rosette (Model SBE911+; Seabird) or underway probe [UnderwayCTD (UCTD); Oceanscience] cast from the vessel. At least one cast was planned along each acoustic transect. These data were used to calculate the harmonic mean sound speed (Demer *et al.*, 2015) for estimating ranges to the sound scatterers, and frequency-specific sound absorption coefficients for compensating signal attenuation of the sound pulse between the transducer and scatters (Simmonds and MacLennan, 2005) (see **Section [2.2.2](#page-21-2)**). These data also indicated the depth of the surface mixed layer, above which most epipelagic CPS reside during the day, which aids in the determination of the integration-stop depth and removal of non-CPS backscatter during acoustic data processing (see **Section [2.2.4](#page-22-0)**).

#### **2.1.3.2 Scientifc Computer System sampling**

While underway, information about the position and direction (e.g., latitude, longitude, speed, course over ground, and heading), weather (air temperature, humidity, wind speed and direction, and barometric pressure), and sea-surface oceanography (e.g., temperature, salinity, and fuorescence) were measured continuously and logged using *Lasker*'s Scientifc Computer System (SCS). Data from a subset of these sensors, logged with a standardized form at 1-min resolution, are available on NOAA's ERDDAP data server<sup>[4](#page-16-3)</sup>.

<span id="page-16-1"></span><sup>2</sup><http://timesynctool.com>

<span id="page-16-2"></span><sup>3</sup>[https://www.fsheries.noaa.gov/west-coast/science-data/ek80-adaptive-logger](https://www.fisheries.noaa.gov/west-coast/science-data/ek80-adaptive-logger)

<span id="page-16-3"></span><sup>4</sup><https://coastwatch.pfeg.noaa.gov/erddap/tabledap/fsuNoaaShipWTEG.html>

#### <span id="page-17-0"></span>**2.1.4 Fish-eggs**

During the day, fsh eggs were sampled using a continuous underway fsh egg sampler (CUFES, Checkley *et al.*, 1997), which collects particles at a rate of  $~640$  l min<sup>-1</sup> from an intake at  $~3$ -m depth on the hull of the ship. Fish eggs in the sampled water were sieved by a 505-*µ*m mesh. Pacifc Sardine, Northern Anchovy, Jack Mackerel, and Pacifc Hake (*Merluccius productus*) eggs were identifed to species, counted, and logged. Eggs from other species (e.g., Pacifc Mackerel and fatfshes) were also counted and logged as "other fsh eggs." Typically, the duration of each CUFES sample was 30 min, corresponding to a distance of 5 nmi at a speed of 10 kn. Because the duration of the egg phase is less than three days for most CPS, the egg distributions inferred from CUFES indicated the presence of actively spawning fsh within roughly 10 nmi, depending on advection and dispersal, and were used in combination with CPS echoes to select trawl locations.

#### <span id="page-17-1"></span>**2.1.5 Trawl**

After sunset, CPS schools tend to ascend and disperse and are less likely to avoid a net (Mais, 1977). Therefore, trawling was conducted during the night to better sample the fsh aggregations dispersed near the surface to obtain information efficiently about species composition, lengths, and weights.

#### **2.1.5.1 Sampling gear**

The net, a Nordic 264 rope trawl (NET Systems, Bainbridge Island, WA; **Fig. [6a](#page-19-0),b**), was towed at the surface for 45 min at a speed of 3.5-4.5 kn. The net has a rectangular opening with an area of approximately  $300 \text{ m}^2$  ( $\sim$ 15-m tall x 20-m wide), a throat with variable-sized mesh and a "marine mammal excluder device" to prevent the capture of large animals, such as dolphins, turtles, or sharks while retaining target species (Dotson *et al.*, 2010), and an 8-mm square-mesh cod-end liner (to retain a large range of animal sizes). The trawl doors were foam-flled and the trawl headrope was lined with foats so the trawl towed at the surface.

#### **2.1.5.2 Sampling locations**

Up to three nighttime (i.e., 30 min after sunset to 30 min before sunrise) surface trawls, typically spaced 10-nmi apart, were conducted in areas where echoes from CPS schools were observed earlier that day. Each evening, trawl locations were selected by an acoustician who monitored CPS echoes and a member of the trawl group who measured the densities of CPS eggs in the CUFES. The locations were provided to the watch officers who charted the proposed trawl sites.

Trawl locations were selected using the following criteria, in descending priority: CPS schools in echograms that day; CPS eggs in CUFES that day; and the trawl locations and catches during the previous night. If no CPS echoes or CPS eggs were observed along a transect that day, the trawls were alternatively placed nearshore one night and ofshore the next night, with consideration given to the seabed depth and the modeled distribution of CPS habitat. Each morning, after the last trawl or 30 min prior to sunrise, *Lasker* resumed sampling at the location where the acoustic sampling stopped the previous day.

#### **2.1.5.3 Sample processing**

If the total volume of the trawl catch was five 35-l baskets  $\sim$ 175 l) or less, all target species were separated from the catch, sorted by species, weighed, and enumerated. If the volume of the entire catch was more than fve baskets, a fve-basket random subsample that included non-target species was collected, sorted by species, weighed, and enumerated; the remainder of the total catch was weighed. In these cases, the weight of the entire catch was calculated as the sum of the subsample and remainder weights. The weight of the *e*-th species in the total catch  $(C_{T,e})$  was obtained by summing the catch weight of the respective species in the subsample  $(C_{S,e})$  and the corresponding catch in the remainder  $(C_{R,e})$ , which was calculated as:

$$
C_{R,e} = C_R * P_{w,e},\tag{1}
$$

where  $P_{w,e} = C_{S,e}/\frac{s}{1}C_{S,e}$ , is the proportion in weight of the *e*-th species in the subsample. The number of specimens of the *e*-th species in the total catch  $(N_{T,e})$  was estimated by:

$$
N_{T,e} = \frac{C_{T,e}}{\overline{w}_e},\tag{2}
$$

where  $\overline{w}_e$  is the mean weight of the *e*-th species in the subsample. If available, a random sample of 50 specimens of Jack and Pacifc Mackerel, and 75 specimens of Pacifc Sardine and Northern anchovy were removed randomly from the subsample. Individual measurements were made of standard length (*LS*) for Pacifc Sardine and Northern Anchovy, and fork length (*L<sup>F</sup>* ) for Pacifc Herring and Jack and Pacifc Mackerels, in cm, and total weight  $(w)$ , in g. In addition, sex and maturity were recorded for all specimens following the methods described in Dorval et al. (2022). Otoliths were removed from all 50 Pacifc Sardine in the subsample; for other CPS species, 25 otoliths were removed uniformly from the range of sizes present for aging (Schwartzkopf *et al.*, 2022). The combined catches in up to three trawls per night (i.e., trawl cluster) were used to estimate the proportions of species contributing to the nearest samples of acoustic backscatter.

#### **2.1.5.4 Quality assurance and quality control**

At sea, trawl data were entered into a database (Microsoft Access). During and following the survey, data were further scrutinized and verifed, or corrected. Missing length (*Lmiss*) and weight (*Wmiss*) measurements were estimated as  $W_{miss} = \beta_0 L^{\beta_1}$  and  $L_{miss} = (W/\beta_0)^{(1/\beta_1)}$ , respectively, where values for  $\beta_0$  and  $\beta_1$  are species- and season-specifc parameters of the length-versus-weight relationships described in Palance et al. (Palance *et al.*, 2019). To identify measurement or data-entry errors, length and weight data were graphically compared (**Fig.** [7\)](#page-20-1) to measurements from previous surveys and models of season-specifc lengthversus-weight from previous surveys (Palance *et al.*, 2019). Outliers were fagged, reviewed by the trawl team, and mitigated. Catch data were removed from aborted trawl hauls, or hauls otherwise deemed unacceptable.

<span id="page-19-0"></span>

Figure 6: Schematic drawings of the Nordic 264 rope trawl a) net and b) cod-end.

<span id="page-20-1"></span>

Figure 7: Specimen length-versus-weight from the current survey (colored points, by sex) compared to those from previous SWFSC surveys during the same season (gray points, all sexes) and models (dashed lines, Palance *et al.*, 2019).

#### <span id="page-20-0"></span>**2.1.6 Purse-seine**

A purse seine net, ~290-m-long and ~22-m-deep net with ~17.5 mm mesh size, was set from *Long Beach Carnage* to provide information about size, age, and species composition of fshes observed in the echosounders in the nearshore region. All specimens collected were frozen and later processed by the CA Department of Fish and Wildlife (CDFW).

A minimum of one set per day was planned during daylight hours. In the event of abundant CPS or an unsuccessful daytime set, a set was made at night. For each set, three dip net samples, spatially separated as much as possible, were collected, and specimens were frozen for later analysis by CDFW biologists. The total weight (tons) of the school was estimated by the captain. After the survey, each dip net sample was sorted, weighed, and counted to provide a combined weight and count for each species. Next, all three dip net samples were combined and up to 50 specimens were randomly sampled to provide a combined weight for each set. Lengths (mm; *L<sup>S</sup>* for Pacifc Sardine and Northern Anchovy and *L<sup>F</sup>* for all others) and weights (g) were measured for up to 50 randomly selected specimens of each species. Macroscopic maturity stages were determined visually, and otoliths were extracted for aging.

#### <span id="page-21-0"></span>**2.2 Data processing**

#### <span id="page-21-1"></span>**2.2.1 Acoustic and oceanographic data**

The calibrated echosounder data from each transect were processed using commercial software [\(Echoview](https://www.echoview.com/) v12.0, Echoview Software Pty Ltd.) and estimates of the sound speed and absorption coefficient were calculated with contemporaneous data from CTD probes cast while stationary or underway (UCTD, see **Section [2.1.3.1](#page-16-4)**). Data collected along the daytime transects at speeds  $\geq$  5 kn were used to estimate CPS densities. Nighttime acoustic data were assumed to be negatively biased due to diel-vertical migration (DVM) and disaggregation of the target species' schools (Cutter and Demer, 2008).

#### <span id="page-21-2"></span>**2.2.2 Sound speed and absorption calculation**

Depth derived from pressure in CTD casts was used to bin samples into 1-m depth increments. Sound speed in each increment  $(c_w, i, m s^{-1})$  was estimated from the average salinity, density, and pH [if measured, else  $pH = 8$ ; Chen and Millero (1977); Seabird (2013). The harmonic sound speed in the water column ( $\bar{c}_w$ , m  $s^{-1}$ ) was calculated over the upper 70 m as:

$$
\overline{c}_w = \frac{\sum_{i=1}^N \Delta r_i}{\sum_{i=1}^N \Delta r_i / c_{w,i}},\tag{3}
$$

where *∆r* is the depth of increment *i* (Seabird, 2013). Measurements of seawater temperature (*tw*, ◦C), salinity  $(s_w, \text{psu})$ , depth, pH, and  $\bar{c}_w$  are also used to calculate the mean frequency-specific absorption coefficients ( $\overline{\alpha}_a$ , dB m<sup>-1</sup>) over the entire depth range using equations in Francois and Garrison (1982), Ainslie and McColm (1998), and Doonan et al. (2003). Both  $\bar{c}_w$  and  $\bar{\alpha}_a$  are later used to estimate ranges to the sound scatterers, to compensate the echo signal for spherical spreading and attenuation during propagation of the sound pulse from the transducer to the scatterer range and back (Simmonds and MacLennan, 2005). The CTD rosette, when cast, also provides measures of fuorescence and dissolved oxygen concentration versus depth, which may be used to estimate the vertical dimension of Pacifc Sardine potential habitat (Zwolinski *et al.*, 2011), particularly the depth of the upper-mixed layer where most epipelagic CPS reside. The latter information is used to inform echo classifcation (see **Section [2.2.3](#page-21-3)**).

#### <span id="page-21-3"></span>**2.2.3 Echo-classifcation**

Echoes from schooling CPS were identifed using a semi-automated data processing algorithm implemented using Echoview software (v12.0). The flters and thresholds were based on a subsample of echoes from randomly selected CPS schools. The aim of the flter criteria is to retain at least 95% of the noise-free backscatter from CPS schools while rejecting at least 95% of the non-CPS backscatter (**Fig. [8](#page-22-1)**). Data from *Lasker* and *Long Beach Carnage* were processed using the following steps:

- 1. Match geometry of the 70-, 120-, 200-, and 333-kHz *S<sup>v</sup>* to the 38-kHz *Sv*;
- 2. Remove passive-mode pings;
- 3. Estimate and subtract background noise using the background noise removal function (De Robertis and Higginbottom, 2007) in Echoview (**Figs. [8b](#page-22-1), e**);
- 4. Average the noise-free *S<sup>v</sup>* echograms using non-overlapping 11-sample by 3-ping bins;
- 5. Expand the averaged, noise-reduced  $S<sub>v</sub>$  echograms with a 7 pixel x 7 pixel dilation;
- 6. For each pixel, compute:  $S_{v,200\text{kHz}} S_{v,38\text{kHz}}$ ,  $S_{v,120\text{kHz}} S_{v,38\text{kHz}}$ , and  $S_{v,70\text{kHz}} S_{v,38\text{kHz}}$ ;
- 7. Create a Boolean echogram for  $S_v$  differences in the CPS range:  $-13.85 < S_{v,70kHz} S_{v,38kHz}$ 9*.*89 and − 13*.*5 *< Sv,*120kHz − *Sv,*38kHz *<* 9*.*37 and − 13*.*51 *< Sv,*200kHz − *Sv,*38kHz *<* 12*.*53;
- 8. Compute the 120- and 200-kHz Variance-to-Mean Ratios (*V MR*120kHz and *V MR*200kHz, respectively, Demer *et al.*, 2009) using the difference between noise-filtered  $S_v$  (Step 3) and averaged  $S_v$  (Step 4);
- 9. Expand the  $VMR_{120kHz}$  and  $VMR_{200kHz}$  echograms with a 7 pixel x 7 pixel dilation;
- 10. Create a Boolean echogram based on the *VMR*s in the CPS range:  $VMR_{120kHz} > -65$  dB and  $VMR_{200kHz} > -65$  dB. Diffuse backscattering layers have low *VMR* (Zwolinski *et al.*, 2010) whereas fsh schools have high *V MR* (Demer *et al.*, 2009);
- 11. Intersect the two Boolean echograms to create an echogram with "TRUE" samples for candidate CPS schools and "FALSE" elsewhere;
- 12. Mask the noise-reduced echograms using the CPS Boolean echogram (**Figs. [8c](#page-22-1), f**);
- 13. Create an integration-start line 5 m below the transducer  $(\sim 10 \text{ m depth});$
- 14. Create an integration-stop line 3 m above the estimated seabed (Demer *et al.*, 2009), or to the maximum logging range (e.g., 1000 m), whichever is shallowest;
- 15. Set the minimum *S<sup>v</sup>* threshold to -60 dB (corresponding to a density of approximately three 20-cm-long Pacific Sardine per  $100 \text{ m}^3$ ;
- 16. Integrate the volume backscattering coefficients  $(s_V, m^2 m^{-3})$  attributed to CPS over 5-m depths and averaged over 100-m distances;
- 17. Output the resulting nautical area scattering coefficients  $(s_A; m^2 \text{ nmi}^{-2})$  and associated information from each transect and frequency to comma-delimited text (.csv) fles.

When necessary, the start and stop integration lines were manually edited to exclude reverberation due to bubbles, to include the entirety of shallow CPS aggregations, or to exclude seabed echoes.

#### <span id="page-22-0"></span>**2.2.4 Removal of non-CPS backscatter**

In addition to echoes from target CPS, echoes may also be present from other pelagic fsh species (e.g., Pacifc Saury, *Cololabis saira*), or semi-demersal fsh such as Pacifc Hake and rockfshes (*Sebastes* spp.). When analyzing the acoustic-survey data, it was therefore necessary to manually flter "acoustic by-catch," i.e., backscatter not from the target species. Fish echoes in proximity to rocky seabed, and difuse backscatter were excluded from the CPS analysis using a custom interactive script for R (**Fig. [9](#page-23-2)**).

<span id="page-22-1"></span>

Figure 8: Two examples of echograms depicting CPS schools (red) and plankton aggregations (blue and green) at 38 kHz (top) and 120 kHz (bottom). Example data processing steps include the original echogram (a, d), after noise subtraction and bin-averaging (b, e), and after fltering to retain only putative CPS echoes  $(d, f)$ .

<span id="page-23-2"></span>

Figure 9: (Top) Integrated echo energy versus depth for all "CPS-like" targets and, (Bottom) integrated echo energy versus depth excluding likely rockfshes and near-surface reverberation (images created in R, R Core Team, 2021). Both images show the seabed (black line), and the fnal upper (red line) and lower (blue line) integration depths by interval.

#### <span id="page-23-0"></span>**2.2.5 Quality assurance and quality control**

The largest 38-kHz integrated backscattering coefficient values  $(s_A, m^2 \text{ nmi}^{-2})$  were graphically examined to identify potential errors in the integrated data from Echoview processing (e.g., when a portion of the seabed was accidentally integrated). If found, errors were corrected and data were re-integrated prior to use for biomass estimation.

#### <span id="page-23-1"></span>**2.2.6 Echo integral partitioning and acoustic inversion**

For fishes with swimbladders, the acoustic backscattering cross-section of an individual ( $\sigma_{bs}$ , m<sup>2</sup>) depends on many factors but mostly on the acoustic wavelength and the swimbladder size and orientation relative to the incident sound pulse. For echosounder sampling in this survey,  $\sigma_{bs}$  is a function of the dorsal-surface area of the swimbladder and was approximated by a function of fish length, i.e.:

$$
\sigma_{bs} = 10^{\frac{m \log_{10}(L) + b}{10}},\tag{4}
$$

where *m* and *b* are frequency and species-specific parameters that are obtained theoretically or experimentally (see references below). *TS*, a logarithmic representation of  $\sigma_{bs}$ , is defined as:

$$
TS = 10 \log_{10}(\sigma_{bs}) = m \log_{10}(L) + b. \tag{5}
$$

*TS* has units of dB re 1 m<sup>2</sup> if defined for an individual, or dB re 1 m<sup>2</sup> kg<sup>-1</sup> if defined by weight. The following equations for  $TS_{38kHz}$  were used in this analysis:

<span id="page-24-0"></span>
$$
TS_{38\text{kHz}} = -14.90 \times \log_{10}(L_T) - 13.21
$$
, for Pacific Sardine; (6)

<span id="page-24-2"></span>
$$
TS_{38\text{kHz}} = -11.97 \times \log_{10}(L_T) - 11.58561
$$
, for Pacific and Round Herring; (7)

<span id="page-24-3"></span>
$$
TS_{38kHz} = -13.87 \times \log_{10}(L_T) - 11.797
$$
, for Northern Anchovy; and (8)

<span id="page-24-1"></span>
$$
TS_{38\text{kHz}} = -15.44 \times \log_{10}(L_T) - 7.75
$$
, for Pacific and Jack Mackerels, (9)

where the units for total length  $(L_T)$  is cm and  $TS$  is dB re 1 m<sup>2</sup> kg<sup>-1</sup>.

Equations [\(6\)](#page-24-0) and [\(9\)](#page-24-1) were derived from echosounder measurements of  $\sigma_{bs}$  from in situ fish, and measures of *L<sup>T</sup>* and *W* from concomitant catches of South American Pilchard (*Sardinops ocellatus*) and Horse Mackerel (*Trachurus trachurus*) of South Africa (Barange *et al.*, 1996). Because mackerels have similar *T S* (Peña, 2008), Equation [\(9\)](#page-24-1) is used for both Pacifc and Jack Mackerels. For Pacifc Herring, Equation [\(7\)](#page-24-2) was derived from that of Thomas et al. (2002) measured at 120 kHz with the following modifcations: 1) the intercept used here was calculated as the average intercept of Thomas et al.'s spring and fall regressions; 2) the intercept was compensated for swimbladder compression after Zhao et al. (2008) using the average depth for Pacifc Herring of 44 m; and 3) the intercept was increased by 2.98 dB to account for the change of frequency from 120 to 38 kHz (Saunders *et al.*, 2012). Equation [\(7\)](#page-24-2) was also used for Round Herring. For Northern Anchovy, Equation [\(8\)](#page-24-3) was derived from that of Kang et al. (2009), after compensation of the swimbladder volume (Ona, 2003; Zhao *et al.*, 2008) for the average depth of Northern Anchovy observed in summer 2016 (19 m, Zwolinski *et al.*, 2017).

To calculate  $TS_{38\text{kHz}}$ ,  $L_T$  was estimated from measurements of  $L_S$  or  $L_F$  using linear relationships between length measurements derived from specimens collected in the CCE (Palance *et al.*, 2019): for Pacifc Sardine,  $L_T$  = 0.3574 + 1.149 $L_S$ ; for Northern Anchovy,  $L_T$  = 0.2056 + 1.1646 $L_S$ ; for Pacific Mackerel,  $L_T$  =  $0.2994 + 1.092L_F$ ; for Jack Mackerel  $L_T = 0.7295 + 1.078L_F$ ; and for Pacific Herring  $L_T = -0.105 + 1.2L_F$ . Since length-length conversions were not available for Round Herring, *L<sup>T</sup>* was approximated from *L<sup>F</sup>* using the equation for Pacifc Herring. Individual weights can be estimated from *L<sup>T</sup>* using the length-weight relationships described in Palance et al. (2019).

The proportions of species in a trawl cluster were considered representative of the proportions of species in the vicinity of the cluster. Therefore, the proportion of the echo-integral from the  $e$ -th species  $(P_e)$  in an ensemble of *s* species can be calculated from the species catches  $N_1, N_2, ..., N_s$  and the respective average backscattering cross-sections  $\sigma_{bs_1}, \sigma_{bs_2}, \ldots, \sigma_{bs_s}$  (Nakken and Dommasnes, 1975). The acoustic proportion for the *e*-th species in the *a*-th trawl  $(P_{ae})$  is:

$$
P_{ae} = \frac{N_{ae} \times \overline{w}_{ae} \times \overline{\sigma}_{bs,ae}}{e = 1(N_{ae} \times \overline{w}_{ae} \times \overline{\sigma}_{bs,ae})},\tag{10}
$$

where  $\overline{\sigma}_{bs,ae}$  is the arithmetic counterpart of the average target strength  $(\overline{TS}_{ae})$  averaged for all  $n_{ae}$  individuals of species *e* in the random sample of trawl *a*:

$$
\overline{\sigma}_{bs,ae} = \frac{\frac{n_{ae}}{i=1} 10^{(TS_i/10)}}{n_{ae}},\tag{11}
$$

and  $\overline{w}_{ae}$  is the average weight:  $\overline{w}_{ae} = \frac{n_{ae}}{i=1} w_{aei}/n_{ae}$ . The total number of individuals of species e in a trawl a  $(N_{ae})$  is obtained by:  $N_{ae} = \frac{n_{ae}}{w_{s,ae}} \times w_{t,ae}$ , where  $w_{s,ae}$  is the weight of the  $n_{$ randomly, and  $w_{t,ae}$  is the total weight of the respective species' catch.

catches across the *h* trawls in the cluster:  $N_{ge} = \frac{h_g}{a=1} N_{ae}$ . The backscattering cross-section for species *e* The trawls within a cluster were combined to reduce sampling variability (see **Section [2.2.7](#page-25-0)**), and the number of individuals caught from the *e*-th species in a cluster  $g(N_{ge})$  was obtained by summing the in the *g*-th cluster with *a* trawls is then given by:

$$
\overline{\sigma}_{bs,ge} = \frac{\frac{h_g}{a=1} N_{ae} \times \overline{w}_{ae} \times \overline{\sigma}_{bs,ae}}{\frac{s_g}{a=1} N_{ae} \times \overline{w}_{ae}},\tag{12}
$$

where:

$$
\overline{w}_{ge} = \frac{\frac{h_g}{a=1} N_{ae} \times \overline{w}_{ae}}{\frac{h_g}{a=1} N_{ae}},\tag{13}
$$

and the proportion  $(P_{qe})$  is;

<span id="page-25-3"></span>
$$
P_{ge} = \frac{N_{ge} \times \overline{w}_{ge} \times \overline{\sigma}_{bs,ae}}{s_{e=1}(N_{ge} \times \overline{w}_{ge} \times \overline{\sigma}_{bs,ge})}.
$$
\n(14)

#### <span id="page-25-0"></span>**2.2.7 Trawl clustering and species proportioning**

The catches of trawls that occurred on the same night were combined into a trawl cluster. Biomass densities  $(\rho)$  were calculated for 100-m transect intervals by dividing the integrated area backscatter coefficients for each CPS species by the mean backscattering cross-sectional area (MacLennan *et al.*, 2002) estimated in the nearest trawl cluster. Survey data were post-stratifed to account for spatial heterogeneity in sampling efort and biomass density in a similar way to that performed for Pacifc Sardine (Zwolinski *et al.*, 2016).

For a generic 100-m long acoustic interval, the area backscattering coefficient for species  $e: s_{A,e} = s_{A,cps} \times P_{ge}$ , where  $P_{ge}$  is the species acoustic proportion of the nearest trawl cluster (Equation [\(14\)](#page-25-3)), was used to estimate the biomass density (*ρw,e*) (MacLennan *et al.*, 2002; Simmonds and MacLennan, 2005) for every 100-m interval, using the size and species composition of the nearest (space and time) trawl cluster (**Fig. [10](#page-26-0)**):

$$
\rho_{w,e} = \frac{s_{A,e}}{4\pi \overline{\sigma}_{bs,e}}.\tag{15}
$$

The biomass densities were converted to numerical densities using:  $\rho_{n,e} = \rho_{w,e}/\overline{w}_e$ , where  $\overline{w}_e$  is the corresponding mean weight. Also, for each acoustic interval, the biomass or numeric densities are partitioned into length classes according to the species' length distribution in the respective trawl cluster.

#### <span id="page-25-1"></span>**2.3 Data analysis**

#### <span id="page-25-2"></span>**2.3.1 Post-stratifcation**

The transects were used as sampling units (Simmonds and Fryer, 1996). Because each species does not generally span the entire survey area (Demer and Zwolinski, 2017; Zwolinski *et al.*, 2014), the sampling domain was stratifed for each species and stock. Strata were defned by uniform transect spacing (sampling intensity) and either presences (positive densities and potentially structural zeros) or absences (real zeros) of species biomass. Each stratum has: 1) at least three transects, with approximately equal spacing, 2) fewer than three consecutive transects with zero-biomass density, and 3) bounding transects with zero-biomass density (**Fig. [11](#page-27-0)**). This approach tracks stock patchiness and creates statistically-independent, stationary, post-sampling strata (Johannesson and Mitson, 1983; Simmonds *et al.*, 1992). For Northern Anchovy, we defne the separation between the northern and central stock at Cape Mendocino (40.4 ◦N).

<span id="page-26-0"></span>

Figure 10: a) Polygons enclosing 100-m acoustic intervals from *Lasker* assigned to each trawl cluster, with its number located at the center of mass of all trawls in that cluster; and b) the acoustic proportions of CPS in trawl clusters, and the location of trawl cluster <sup>10</sup> with no CPS (black point).

<span id="page-27-0"></span>

Figure 11: Biomass density  $(\log(t \text{ nm}^2+1))$  versus latitude (easternmost portion of each transect) and strata used to estimate biomass and abundance (shaded regions; outline indicates stratum number) for each species and survey vessel (labels above plots; *RL* = *Lasker*). Strata with no outline were not included because of too few specimens  $( $10$  individuals), trawl clusters  $( $2$  clusters), or both. Blue number labels correspond$$ to transects with positive biomass  $(\log_{10}(t+1) > 0.01)$ . Point fills indicate transect spacing (nmi).

#### <span id="page-28-0"></span>**2.3.2 Estimation of biomass and sampling precision**

For each stratum and stock, the biomass  $(\hat{B}; \text{kg})$  of each species was estimated by:

$$
\hat{B} = A \times \hat{D},\tag{16}
$$

where *A* is the stratum area (nmi<sup>2</sup>) and  $\hat{D}$  is the estimated mean biomass density (kg nmi<sup>-2</sup>):

<span id="page-28-3"></span>
$$
\hat{D} = \frac{\sum_{l=1}^{k} \overline{\rho}_{w,l} c_l}{\sum_{l=1}^{k} c_l},\tag{17}
$$

where  $\bar{\rho}_{w,l}$  is the mean biomass density of the species on transect *l*,  $c_l$  is the transect length, and *k* is the total number of transects. The variance of  $\hat{B}$  is a function of the variability of the transect-mean densities and associated lengths. Treating transects as replicate samples of the underlying population (Simmonds and Fryer, 1996), the variance was calculated using bootstrap resampling (Efron, 1981) based on transects as sampling units. Provided that each stratum has independent and identically-distributed transect means (i.e., densities on nearby transects are not correlated, and they share the same statistical distribution), bootstrap or other random-sampling estimators provide unbiased asymptotic estimates of variance.

The 95% confidence intervals ( $CI_{95\%}$ ) for the mean biomass densities  $(\hat{D})$  were estimated as the 0.025 and 0.975 percentiles of the distribution of 1000 bootstrap survey-mean biomass densities. Coefficient of variation (CV, %) values were obtained by dividing the bootstrapped standard error by the mean estimate (Efron, 1981). Total biomass in the survey area was estimated as the sum of the biomasses in each stratum, and the associated sampling variance was calculated as the sum of the variances across strata.

#### <span id="page-28-1"></span>**2.3.3 Abundance- and biomass-at-length estimation**

The numerical densities by length class (**Section [2.2.7](#page-25-0)**) were averaged for each stratum in a similar way as that used for biomass in Equation [\(17\)](#page-28-3), and multiplied by the stratum area to obtain abundance per length class.

#### <span id="page-28-2"></span>**2.3.4 Percent contribution of abundance per cluster**

The percent contribution of each cluster to the estimated abundance in a stratum (**Appendix [A](#page-56-1)**) was calculated as:

$$
\frac{\sum_{i=1}^{l} \overline{\rho}_{ci}}{\sum_{c=1}^{C} \sum_{i=1}^{l} \overline{\rho}_{ci}},\tag{18}
$$

where  $\overline{\rho}_{ci}$  is the numerical density in interval *i* represented by the nearest trawl cluster *c*.

## <span id="page-29-0"></span>**3 Results**

## <span id="page-29-1"></span>**3.1 Sampling efort and allocation**

The spring 2021 survey spanned the area between San Francisco and San Diego, and was completed in 25 DAS, between 20 March and 14 April 2021. In the core area, sampled by *Lasker*, acoustic sampling was conducted along 26 daytime east-west transects that totaled 1,639 nmi. Catches from a total of 49 nighttime surface trawls were combined into 19 trawl clusters. As many as two post-survey strata were defned considering transect spacing and the densities of echoes attributed to each species. In the nearshore survey area, acoustic sampling by *Long Beach Carnage* was conducted along 61 daytime, east-west transects totaling 191 nmi. As many as fve post-survey strata were defned considering transect spacing and the densities of echoes attributed to CPS. Biomasses and abundances were estimated for each species in the core and nearshore survey areas.

## <span id="page-29-2"></span>**3.2 Acoustic backscatter**

Acoustic backscatter ascribed to CPS was observed throughout the core survey area, but was most prevalent south of Monterey Bay, especially ofshore of the SCB, south of the northern Channel Islands (**Fig. [12a](#page-30-0)**). Multiple transects were extended (**Appendix [B](#page-59-0)**) to ensure zero-biomass intervals at their ofshore ends. Acoustic backscatter ascribed to CPS was also observed throughout the nearshore survey area, but was most prevalent between Long Beach and Oceanside (**Fig. [12a](#page-30-0)**). Greater than 90% of the biomass for each species was apportioned using catch data from trawl clusters within 30 nmi (**Fig. [13](#page-31-0)**).

## <span id="page-29-3"></span>**3.3 Egg densities and distributions**

Northern Anchovy eggs were abundant in CUFES samples south of Monterey Bay, but were most abundant and coincident with the acoustic backscatter in the ofshore portions of transects south of Point Conception (**Fig. [12b](#page-30-0)**). A few samples in the SCB contained Pacifc Sardine eggs, mostly along the southeast coast of Santa Catalina Island (**Fig. [12b](#page-30-0)**). No Jack Mackerel eggs were observed.

## <span id="page-29-4"></span>**3.4 Trawl catch**

Northern Anchovy was the predominant species in all but one of the trawl clusters (**Fig. [12c](#page-30-0)**). Relatively small numbers of Pacifc Sardine were collected ofshore of the northern Channel Islands and east of San Nicolas Island (too few to view in **Fig. [12c](#page-30-0)**). Jack Mackerel were present in several trawls ofshore in the SCB, and one near Long Beach, and small numbers of Pacifc Mackerel were caught west of San Nicolas Island (also too few to view in **Fig. [12c](#page-30-0)**). Overall, the 49 trawls captured a combined 4,293 kg of CPS (4,290 kg of Northern Anchovy, 1.32 kg of Pacifc Sardine, 0.556 kg of Pacifc Mackerel, 1.13 kg of Jack Mackerel, and no Pacifc or Round Herring).

#### <span id="page-29-5"></span>**3.5 Purse seine catch**

Pacifc Sardine were predominant, by weight, in purse seine samples collected in the SCB by *Long Beach Carnage* (**Fig. [12c](#page-30-0)**). Overall, the 10 seines resulted in a combined sample of 15.7 kg of CPS (2.5 kg of Northern Anchovy, 12.3 kg of Pacifc Sardine, 0.944 kg of Pacifc Mackerel, and no Jack Mackerel, Pacifc Herring, or Round Herring).

<span id="page-30-0"></span>

Figure 12: Spatial distributions of: a) 38-kHz integrated backscattering coefficients ( $s_A$ , m<sup>2</sup> nmi<sup>-2</sup>; averaged over 2000-m distance intervals and from 5 to 350 <sup>m</sup> deep) observed by *Lasker* and *Long Beach Carnage* and ascribed to CPS; b) CUFES egg density (eggs <sup>m</sup>-3) for Northern Anchovy, Pacifc Sardine, and Jack Mackerel; and c) acoustic proportions of CPS in trawl clusters (black outline) and purse seines (white outline; black points indicate clusters or purse seines with no CPS).

<span id="page-31-0"></span>

Figure 13: Proportion (top) and cumulative proportion (bottom) of biomass versus distance to the nearest positive trawl cluster. Dashed vertical lines (bottom) represent the cluster distance where cumulative biomass equals 90%.

## <span id="page-32-0"></span>**3.6 Biomass distribution and demographics**

#### <span id="page-32-1"></span>**3.6.1 Northern Anchovy**

#### **3.6.1.1 Central stock**

The total estimated biomass of the central stock of Northern Anchovy in U.S. waters was  $1,370,303$  t (CI<sub>95%</sub>)  $= 1,076,749 - 1,959,163$  t,  $CV = 16\%)$ . In the core region, sampled by *Lasker*, the biomass was 1,358,587 t  $(CI_{95\%} = 1,070,094 - 1,940,986$  t,  $CV = 17\%$ ; Table [3](#page-32-2)); the stock was distributed from north of Monterey Bay to San Diego (**Fig. [14a](#page-34-0)**). *L<sup>S</sup>* ranged from 8 to 16 cm with a mode between 10 and 13 cm (**Table [4](#page-33-0)**, **Fig.** [15](#page-35-0)). In the nearshore region, sampled by *Long Beach Carnage*, the biomass was 11,716 t (CI<sub>95%</sub> = 6,655 - 18,176 t, CV = 26%; **Table [3](#page-32-2)**), mostly distributed between Ventura and San Diego (**Fig. [14b](#page-34-0)**). Biomass in the nearshore region comprised 0.86% of the total biomass. The length distribution was similar to that in the core region (**Table [4](#page-33-0)**, **Fig. [15](#page-35-0)**).

<span id="page-32-2"></span>Table 3: Biomass estimates (metric tons, t) and their precision (upper and lower 95% confdence intervals, CI<sub>95%</sub>; and coefficients of variation, CVs) for the central stock of Northern Anchovy (*Engraulis mordax*) in the core and nearshore survey regions. Stratum areas are nmi<sup>2</sup>.



<span id="page-33-0"></span>Table 4: Abundance versus standard length (*LS*, cm) for the central stock of Northern Anchovy (*Engraulis mordax*) in the core and nearshore survey regions.

	Region				
$L_S$	$\rm Core$	Nearshore			
1	0	0			
$\overline{2}$	0	0			
3	0	0			
4	0	$\Omega$			
5	0	0			
6	0	0			
7	0	3,595,242			
8	762,238,601	321,952,670			
9	9,632,798,691	896,905,365			
10	29,286,125,416	243,563,227			
11	17,923,875,332	38,018,647			
12	17,321,267,534	$\overline{37},572,450$			
13	11,181,245,156	24,013,255			
14	4,512,360,465	$\overline{1,}302,202$			
$\overline{15}$	906,295,154	156,840			
16	200,632,184	0			
17	0	0			
18	0	0			
19	$\overline{0}$	0			
20	0	0			

<span id="page-34-0"></span>

Figure 14: Biomass densities of the central stock of Northern Anchovy (*Engraulis mordax*), per strata, in the a) core and b) nearshore survey regions. The blue numbers represent the locations of trawl clusters or purse seines with at least one Northern Anchovy. The gray line represents the vessel tracks.

<span id="page-35-0"></span>

Figure 15: Abundance versus standard length (*LS*, upper panel) and biomass (t) versus *L<sup>S</sup>* (lower panel) for the central stock of Northern Anchovy (*Engraulis mordax*) in the core and nearshore survey regions.

#### <span id="page-36-0"></span>**3.6.2 Pacifc Sardine**

#### **3.6.2.1 Southern stock**

The total estimated biomass of the southern stock of Pacific Sardine in U.S. waters was  $24,547$  t (CI<sub>95%</sub>)  $= 7,697 - 38,339$  t,  $CV = 29.3\%$ ; see **Section [4.1.2.1](#page-48-4)** for more details on the stock classification). In the core region, biomass was 1,[5](#page-36-1)04 t ( $CI_{95\%}$  = 556 - 2,959 t, CV = 40%; Table 5), and was distributed from approximately Point Conception to Oceanside (**Fig. [16a](#page-38-0)**). *L<sup>S</sup>* ranged from 12 to 20 cm with a mode at 13 cm (**Table [6](#page-37-0)**, **Fig.** [17](#page-39-0)). In the nearshore region, biomass was 23,043 t ( $CI_{95\%} = 7,140 - 35,380$  t, CV = 31%; **Table [5](#page-36-1)**), comprising 94% of the total biomass. It was distributed between Point Conception and San Diego, but most of it was near Santa Barbara, between Long Beach and Oceanside, and around Santa Cruz Island. These areas did not contain northern stock habitat (Zwolinski *et al.*, 2011), so none were ascribed to the northern stock. The length distribution was similar to that in the core region (**Table [6](#page-37-0)**, **Fig. [17](#page-39-0)**).

<span id="page-36-1"></span>Table 5: Biomass estimates (metric tons, t) and their precisions (upper and lower 95% confdence intervals, CI95%; and coefcients of variation, CVs) for the southern stock of Pacifc Sardine (*Sardinops sagax*) in the core and nearshore survey regions. Stratum areas are nmi<sup>2</sup>.



<span id="page-37-0"></span>Table 6: Abundance versus standard length (*LS*, cm) for the southern stock of Pacifc Sardine (*Sardinops sagax*) in the core and nearshore survey regions.

	Region				
	Core	Nearshore			
$rac{L_S}{1}$	0	0			
$\overline{2}$	$\overline{0}$	$\overline{0}$			
$\overline{3}$	$\overline{0}$	$\overline{0}$			
$\overline{4}$	$\overline{0}$	$\overline{0}$			
5	$\overline{0}$	$\overline{0}$			
$\overline{6}$	$\overline{0}$	$\overline{0}$			
7	$\overline{0}$	$\overline{0}$			
8	$\overline{0}$	$\overline{0}$			
9	$\overline{0}$	$\overline{0}$			
10	0	$\overline{0}$			
$\overline{11}$	$\overline{0}$	47,243,617			
$\overline{12}$	1,155,897	265,755,034			
13	8,516,604	265,788,913			
$\overline{14}$	5,996,830	106,827,536			
$\overline{15}$	0	89,632,839			
16	1,155,897	30,809,885			
17	6,522,487	0			
18	62,495	17,290,884			
19	0	9,620,145			
$\overline{20}$	5,754,512	0			
21	$\overline{0}$	$\overline{0}$			
22	$\overline{0}$	$\overline{0}$			
23	$\overline{0}$	$\overline{0}$			
$\overline{24}$	$\overline{0}$	$\overline{0}$			
25	$\overline{0}$	$\overline{0}$			
26	$\overline{0}$	$\overline{0}$			
$\overline{27}$	$\overline{0}$	$\overline{0}$			
28	$\overline{0}$	$\overline{0}$			
29	$\overline{0}$	$\overline{0}$			
$\overline{30}$	$\overline{0}$	$\overline{0}$			

<span id="page-38-0"></span>

Figure 16: Biomass densities of the southern stock of Pacifc Sardine (*Sardinops sagax*), per strata, in the a) core and b) nearshore survey regions. The blue numbers represent the locations of trawl clusters or purse seines with at least one Pacifc Sardine. The gray lines represents the vessel tracks.

<span id="page-39-0"></span>

Figure 17: Estimated abundance (upper panel) and biomass (lower panel) versus standard length (*LS*, cm) for the southern stock of Pacifc Sardine (*Sardinops sagax*) in the core and nearshore survey regions.

#### <span id="page-40-0"></span>**3.6.3 Pacifc Mackerel**

In the nearshore region, the biomass of Pacific Mackerel in U.S. waters was 92.3 t ( $CI_{95\%} = 7.43 - 221$  t, CV = 64%; **Table [7](#page-40-1)**, **Fig. [18b](#page-42-0)**), distributed from San Clemente to San Diego. *L<sup>F</sup>* ranged from 14 to 25 cm with a mode at 18 cm (**Table [8](#page-41-0)**, **Fig. [19](#page-43-0)**).

<span id="page-40-1"></span>Table 7: Biomass estimates (metric tons, t) and their precisions (upper and lower 95% confdence intervals, CI<sub>95%</sub>; and coefficients of variation, CVs) for Pacific Mackerel (*Scomber japonicus*) in nearshore region. No Pacifc Mackerel were caught in the core region. Stratum areas are nmi<sup>2</sup>.



<span id="page-41-0"></span>Table 8: Abundance versus fork length (*L<sup>F</sup>* , cm) for Pacifc Mackerel (*Scomber japonicus*) in the nearshore survey region.



<span id="page-42-0"></span>

Figure 18: Biomass densities of the Pacifc Mackerel (*Scomber japonicus*), per strata, in the nearshore survey regions. The blue numbers represent the locations of purse seines with at least one Pacifc Mackerel. The gray line represents the vessel track.

<span id="page-43-0"></span>

Figure 19: Estimated abundance (upper panel) and biomass (lower panel) versus fork length ( $L_F$ , cm) for Pacifc Mackerel (*Scomber japonicus*) in the core and nearshore survey regions.

#### <span id="page-44-0"></span>**3.6.4 Jack Mackerel**

The total estimated biomass of Jack Mackerel in U.S. waters was  $16,882$  t (CI<sub>95%</sub> = 3,783 - 41,209 t, CV = 57%), all in the core region (**Table [9](#page-44-1)**), and was distributed from approximately Point Conception to San Diego (**Fig. [20a](#page-46-0)**). *L<sup>F</sup>* ranged from 9 to 23 cm with a mode at 13 cm (**Table [10](#page-45-0)**, **Fig. [21](#page-47-0)**).

<span id="page-44-1"></span>Table 9: Biomass estimates (metric tons, t) and their precisions (upper and lower 95% confdence intervals, CI<sub>95%</sub>; and coefficients of variation, CVs) for Jack Mackerel (*Trachurus symmetricus*) in the core region. There were no Jack Mackerel caught in the nearshore region. Stratum area is nmi<sup>2</sup>.



$L_F$	Abundance	$L_{F}$	Abundance
1	0	31	0
$\overline{2}$	$\boldsymbol{0}$	$\overline{32}$	$\boldsymbol{0}$
$\overline{3}$	$\overline{0}$	33	$\overline{0}$
$\overline{4}$	$\overline{0}$	$\overline{34}$	$\overline{0}$
$\overline{5}$	$\boldsymbol{0}$	35	$\boldsymbol{0}$
$\overline{6}$	$\overline{0}$	$\overline{36}$	$\overline{0}$
7	$\overline{0}$	37	$\overline{0}$
8	$\overline{0}$	$\bar{3}8$	$\overline{0}$
9	$\overline{510}$	$\overline{3}9$	$\overline{0}$
$\overline{10}$	$\overline{510}$	40	$\boldsymbol{0}$
$\overline{11}$	1,019	41	$\overline{0}$
12	48,283	42	$\boldsymbol{0}$
13	264,447,693	43	$\overline{0}$
14	20,475,908	44	$\overline{0}$
15	$\overline{85,91}1,281$	45	$\boldsymbol{0}$
$\overline{16}$	3,314,276	46	$\overline{0}$
17	16,055,929	$\overline{4}$ 7	$\boldsymbol{0}$
18	0	48	$\overline{0}$
19	16,055,929	49	$\overline{0}$
20	16,055,929	50	0
$\overline{21}$	$\overline{16}$ ,055,929	51	$\overline{0}$
22	0	52	$\boldsymbol{0}$
$\overline{23}$	5,493,875	$\overline{53}$	$\overline{0}$
24	0	54	$\overline{0}$
25	$\overline{0}$	$\overline{55}$	$\overline{0}$
$\overline{26}$	$\overline{0}$	$\overline{56}$	$\overline{0}$
27	$\boldsymbol{0}$	57	$\boldsymbol{0}$
$\overline{2}8$	$\overline{0}$	58	$\overline{0}$
29	$\overline{0}$	59	$\overline{0}$
$\bar{3}0$	$\overline{0}$	60	$\boldsymbol{0}$

<span id="page-45-0"></span>Table 10: Abundance versus fork length (*L<sup>F</sup>* , cm) for Jack Mackerel (*Trachurus symmetricus*) in the core survey region.

<span id="page-46-0"></span>

Figure 20: Biomass densities of Jack Mackerel (*Trachurus symmetricus*), per strata, in the core survey region. The blue numbers represent the locations of trawl clusters with at least one Jack Mackerel. The gray line represents the vessel track.

<span id="page-47-0"></span>

Figure 21: Estimated abundance (upper panel) and biomass (lower panel) versus fork length ( $L_F$ , cm) for Jack Mackerel (*Trachurus symmetricus*) in the core survey region. No Jack Mackerel were caught in the nearshore region.

## <span id="page-48-0"></span>**4 Discussion**

The principal objective of the Spring 2021 CPS survey was to assess the central stock of Northern Anchovy in U.S. waters. Then, as possible, estimates were also sought for Pacifc Sardine, Pacifc Mackerel, and Jack Mackerel in the survey area. With the benefts of favorable weather and few problems, *Lasker* surveyed from San Diego to San Francisco. All transects south of Point Conception, where the biomass of Northern Anchovy was expected to be greatest, were spaced 15-nmi apart to reduce the variance. North of Point Conception, transects were spaced 20-nmi apart to maximize the extent of the survey area within the available 25 DAS.

## <span id="page-48-1"></span>**4.1 Biomass and abundance**

#### <span id="page-48-2"></span>**4.1.1 Northern Anchovy**

#### **4.1.1.1 Central stock**

The estimated biomass of the central stock of Northern Anchovy in the core survey region was 1,358,587 t  $(CI_{95\%} = 1,070,094 - 1,940,986 t)$  in spring 2021, which is 60% more than the 810,634 t  $(CI_{95\%} = 587,317 - 1,0800 t)$ 1,066,265) observed in summer 2019 (Stierhof *et al.*, 2020b). In spring 2021, the distribution of *L<sup>S</sup>* had one mode between 10 and 13 cm, likely corresponding to a 2020 cohort.

Although most *Lasker* transects had practically zero biomass density at their western ends, the declines are abrupt. The distribution of Northern Anchovy eggs from the spring 2021 CalCOFI survey (unpublished data) also shows this pattern, which indicates that the abundance of Northern Anchovy farther ofshore of the survey area is likely small (**Appendix [B](#page-59-0)**).

#### <span id="page-48-3"></span>**4.1.2 Pacifc Sardine**

#### <span id="page-48-4"></span>**4.1.2.1 Southern stock**

The core survey area included some "good" and "optimal" habitat for the northern stock of Pacifc Sardine, particularly north of Point Conception (**Fig. [2a](#page-11-0)**). However, the Pacifc Sardine sampled in the SCB were exclusively in the nearshore region and around the northern Channel Islands, which had unsuitable habitat due to excessive chlorophyll-a concentration (**Fig. [2b](#page-11-0)**, **Fig. [22](#page-49-0)**). Therefore, the Pacifc Sardine observed in the spring 2021 survey, with a biomass of 24,547 t ( $CI_{95\%} = 7,697$  - 38,339 t), were attributed to the southern stock. Pacifc Sardine eggs were absent from the nearshore, suggesting that the Pacifc Sardine, with a modal length between 12 and 13 cm, were immature, age-0 fish that likely recruited locally to the southern stock because their forecast growth matched the 16 to 18 cm lengths of Pacifc Sardine sampled in the SCB in summer survey (Stierhoff *et al.*, 2023). Furthermore, it is likely that few or none of them migrated and recruited to the northern stock because there were no Pacifc Sardine with comparable sizes in northern stock habitat sampled of OR and WA during summer 2021 (**Fig. [23](#page-50-0)**).



<span id="page-49-0"></span>

Figure 22: The spatial distribution of Pacifc Sardine during the spring survey overlaid on the potential habitat for the northern stock (Zwolinski *et al.*, 2011), which is principally defned by a non-linear-combination of SST and the logarithm of chlorophyll-a concentration. Based on spring CalCOFI data from 1998 to 2009, the "optimal" and "good" areas include at least 90% of the northern stock Pacifc Sardine biomass. During March 2021, the nearshore waters of the SCB included only "unsuitable" or "bad" habitat for the northern stock.

<span id="page-50-0"></span>

Figure 23: Relative abundance versus standard length (*LS*) for Pacifc Sardine (*Sardinops sagax*) estimated during the spring 2021 (top) and the summer 2021 survey (Stierhoff *et al.*, 2023), separated by putative southern (middle) and northern stocks (bottom).

#### <span id="page-51-0"></span>**4.1.3 Pacifc Mackerel**

There was a negligible amount of Pacifc mackerel observed by *Lasker* and *Long Beach Carnage*. See the summer 2021 CCE biomass report (Stierhoff *et al.*, 2023) for a better indication of the status of this stock.

#### <span id="page-51-1"></span>**4.1.4 Jack Mackerel**

The estimated biomass of Jack Mackerel in the core survey region was  $16,882$  t (CI<sub>95%</sub> = 3,783 - 41,209 t), which is less than 5% of the biomass estimated in the summer 2019 survey. See the summer 2021 CCE biomass report (Stierhof *et al.*, 2023) for a better indication of the status of this stock.

#### <span id="page-51-2"></span>**4.2 Ecosystem dynamics: Forage fsh community**

The ATM has been used worldwide to monitor the biomasses and distributions of pelagic and mid-water fsh stocks worldwide (e.g., Coetzee *et al.*, 2008; Karp and Walters, 1994; Simmonds *et al.*, 2009). In the CCE, ATM surveys have been used to directly assess Pacifc Hake (Edwards *et al.*, 2018; JTC, 2014), rockfshes (Demer, 2012a, 2012b, 2012c; Starr *et al.*, 1996), Pacifc Herring (Thomas and Thorne, 2003), and CPS (Hill *et al.*, 2017; Mais, 1974, 1977). Focused initially, in 2006, on Pacifc Sardine (Cutter and Demer, 2008), the SWFSC's ATM surveys of CPS in the CCE have evolved to assess the fve most abundant forage-fsh species (Zwolinski *et al.*, 2014): Pacifc Sardine, Northern Anchovy, Jack Mackerel, Pacifc Mackerel, and Pacifc Herring. The proportions of these stocks that are in water too shallow to be sampled by NOAA ships are estimated using samples collected from fishing vessels (Stierhoff *et al.*, 2020b, this study) and USVs (Stierhoff *et al.*, 2020b). Also, concurrent satellite- and ship-based measures of their biotic and abiotic habitats are used to provide an ecosystem perspective.

The 2021 spring survey area encompassed the expected distribution of Northern Anchovy and spanned the waters between San Diego and San Francisco, out to 150 km of Southern CA. In the SCB, the sampling from *Lasker* was augmented by nearshore transects sampled by *Long Beach Carnage*. The reduced latitudinal extent of the spring survey, relative to previous years' summer surveys, only allowed an accurate assessment of the entire central stock of Northern Anchovy. Other CPS stocks were sampled partially, if at all. The estimated biomass of 1,358,587 t ( $CV = 16\%$ ) indicates that the central stock continued to grow since summer 2019, when it was estimated at 810,634 t (CV = 0.13, Stierhof *et al.*, 2020b). Compared to Northern Anchovy, other CPS comprised an insignifcant fraction of the overall biomass in the region. Noteworthy is the virtual absence of spawning Pacifc Sardine. Prior to 2014, northern stock Pacifc Sardine dominated the CPS fsh assemblage, particularly during the spring in the SCB (Zwolinski *et al.*, 2014). In spring 2021, southern stock Pacifc Sardine were the most abundant CPS in the SCB, mostly located within 3 nmi of the coast and, based on their lengths and virtually absent spawning, they were mostly sexually immature age-0 fsh.

## <span id="page-51-3"></span>**Acknowledgements**

The authors greatly appreciate that the ATM surveys require an enormous efort by multiple groups of people at the SWFSC, particularly the Advanced Survey Technologies group (Gabriel Johnson and Scott Mau) and biological sampling team (N. Bowlin, B. Erisman, M. Human, B. Schwartzkopf, A. Thompson, A. Freire, L. Giuseffi, V. Hermanson, K. James, O. Snodgrass, W. Watson); the officers and crew of FSV *Reuben Lasker*; and the Fisheries Resources Division administrative staf. We thank Capts. Rich Ashley and Tom Brinton (F/V *Long Beach Carnage*) for their coordination and cooperation during the nearshore sampling, and to Joel van Noord for leading the at-sea processing of purse seine specimens. We thank Diane Pleschner-Steele for contracting the *Long Beach Carnage* to conduct the nearshore survey of the SCB. We thank Dianna Porzio, Trung Nguyen, Kelly Kloos, and Trevor Stocking (CA Department of Fish and Wildlife) for their assistance processing specimens from purse-seine set from the Long Beach Carnage. Finally, reviews by Peter Kuriyama and Annie Yau improved this report.

## <span id="page-52-0"></span>**References**

- Ainslie, M. A., and McColm, J. G. 1998. A simplifed formula for viscous and chemical [absorption](https://doi.org/Doi%2010.1121/1.421258) in sea [water.](https://doi.org/Doi%2010.1121/1.421258) Journal of the Acoustical Society of America, 103: 1671–1672.
- Bakun, A., and Parrish, R. H. 1982. Turbulence, transport, and pelagic fsh in the California and Peru current systems. California Cooperative Oceanic Fisheries Investigations Reports, 23: 99–112.
- Barange, M., Hampton, I., and Soule, M. 1996. Empirical [determination](https://doi.org/DOI%2010.1006/jmsc.1996.0026) of the in situ target strengths of three loosely [aggregated](https://doi.org/DOI%2010.1006/jmsc.1996.0026) pelagic fsh species. ICES Journal of Marine Science, 53: 225–232.
- Checkley, D. M., Ortner, P. B., Settle, L. R., and Cummings, S. R. 1997. A continuous, underway fsh egg sampler. Fisheries Oceanography, 6: 58–73.
- Chen, C. T., and Millero, F. J. 1977. Speed of sound in seawater at high [pressures.](https://doi.org/Doi%2010.1121/1.381646) Journal of the Acoustical Society of America, 62: 1129–1135.
- Coetzee, J. C., Merkle, D., Moor, C. L. de, Twatwa, N. M., Barange, M., and Butterworth, D. S. 2008. Refned estimates of South African pelagic fsh biomass from [hydro-acoustic](https://doi.org/Doi%2010.2989/Ajms.2008.30.2.1.551) surveys: Quantifying the efects of target strength, signal [attenuation](https://doi.org/Doi%2010.2989/Ajms.2008.30.2.1.551) and receiver saturation. African Journal of Marine Science, 30: 205–217.
- Conti, S. G., and Demer, D. A. 2003. [Wide-bandwidth](https://doi.org/Doi%2010.1016/S1054-3139(03)00056-0) acoustical characterization of anchovy and sardine from reverberation [measurements](https://doi.org/Doi%2010.1016/S1054-3139(03)00056-0) in an echoic tank. ICES Journal of Marine Science, 60: 617–624.
- Cutter, G. R., and Demer, D. A. 2008. California Current Ecosystem Survey 2006. Acoustic cruise reports for NOAA FSV *Oscar Dyson* and NOAA FRV *David Starr Jordan*. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-SWFSC-415: 98 pp.
- Cutter, G. R., Renfree, J. S., Cox, M. J., Brierley, A. S., and Demer, D. A. 2009. Modelling [three-dimensional](https://doi.org/DOI%2010.1093/icesjms/fsp040) directivity of sound scattering by Antarctic krill: Progress towards biomass estimation using [multibeam](https://doi.org/DOI%2010.1093/icesjms/fsp040) [sonar.](https://doi.org/DOI%2010.1093/icesjms/fsp040) ICES Journal of Marine Science, 66: 1245–1251.
- De Robertis, A., and Higginbottom, I. 2007. A [post-processing](https://doi.org/10.1093/icesjms/fsm112) technique to estimate the signal-to-noise ratio and remove [echosounder](https://doi.org/10.1093/icesjms/fsm112) background noise. ICES Journal of Marine Science, 64: 1282–1291.
- Demer, D. A. 2012a. 2007 survey of rockfshes in the Southern California Bight using the collaborative optical-acoustic survey technique. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-SWFSC-498: 110.
- Demer, D. A. 2012b. 2004 survey of rockfshes in the Southern California Bight using the collaborative optical-acoustic survey technique. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-SWFSC-497: 96.
- Demer, D. A. 2012c. 2003 survey of rockfshes in the Southern California Bight using the collaborative optical-acoustic survey technique. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-SWFSC-496: 82.
- Demer, D. A., Berger, L., Bernasconi, M., Bethke, E., Boswell, K., Chu, D., Domokos, R., *et al.* 2015. Calibration of acoustic instruments. ICES Cooperative Research Report No. 326: 133 pp.
- Demer, D. A., Conti, S. G., De Rosny, J., and Roux, P. 2003. Absolute [measurements](https://doi.org/Doi%2010.1121/1.1542648) of total target strength from [reverberation](https://doi.org/Doi%2010.1121/1.1542648) in a cavity. Journal of the Acoustical Society of America, 113: 1387–1394.
- Demer, D. A., Kloser, R. J., MacLennan, D. N., and Ona, E. 2009. An [introduction](https://doi.org/DOI%2010.1093/icesjms/fsp146) to the proceedings and a synthesis of the 2008 ICES [Symposium](https://doi.org/DOI%2010.1093/icesjms/fsp146) on the Ecosystem Approach with Fisheries Acoustics and [Complementary](https://doi.org/DOI%2010.1093/icesjms/fsp146) Technologies (SEAFACTS). ICES Journal of Marine Science, 66: 961–965.
- Demer, D. A., and Zwolinski, J. P. 2014. [Corroboration](https://doi.org/DOI%2010.1093/icesjms/fst135) and refnement of a method for diferentiating landings from two stocks of Pacifc sardine (*Sardinops sagax*) in the [California](https://doi.org/DOI%2010.1093/icesjms/fst135) Current. ICES Journal of Marine Science, 71: 328–335.
- Demer, D. A., and Zwolinski, J. P. 2017. A method to [consistently](https://doi.org/10.1080/02755947.2016.1264510) approach the target total fshing fraction of Pacifc sardine and other [internationally](https://doi.org/10.1080/02755947.2016.1264510) exploited fsh stocks. North American Journal of Fisheries Management, 37: 284–293.
- Demer, D. A., Zwolinski, J. P., Byers, K. A., Cutter, G. R., Renfree, J. S., Sessions, T. S., and Macewicz, B. J. 2012. Prediction and confrmation of seasonal migration of Pacifc sardine (*Sardinops sagax*) in the California Current Ecosystem. Fishery Bulletin, 110: 52–70.
- Doonan, I. J., Coombs, R. F., and McClatchie, S. 2003. The [absorption](https://doi.org/10.1016/S1054-3139(03)00120-6) of sound in seawater in relation to the estimation of [deep-water](https://doi.org/10.1016/S1054-3139(03)00120-6) fsh biomass. ICES Journal of Marine Science, 60: 1047–1055.
- Dorval, E., Schwartzkopf, B. D., James, K. C., Vasquez, L., and Erisman, B. E. 2022. Sampling methodology for estimating life history parameters of coastal pelagic species along the U.S. Pacifc Coast. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-660: 46 pp.
- Dotson, R. C., Grifth, D. A., King, D. L., and Emmett, R. L. 2010. Evaluation of a marine mammal

excluder device (MMED) for a Nordic 264 midwater rope trawl. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-SWFSC-455: 19.

- Edwards, A. M., Taylor, I. G., Grandin, C. J., and Berger, A. M. 2018. Status of the Pacifc hake (whiting) stock in U.S. and Canadian waters in 2018. Prepared by the Joint Technical Committee of the U.S. and Canada Pacifc Hake/Whiting Agreement, National Marine Fisheries Service and Fisheries and Oceans Canada. Report. Pacifc Fishery Management Council.
- Efron, B. 1981. Nonparametric standard errors and confdence intervals. Canadian Journal of Statistics, 9: 139–158.
- Field, J. C., Francis, R. C., and Strom, A. 2001. Toward a fsheries ecosystem plan for the northern California Current. California Cooperative Oceanic Fisheries Investigations Reports, 42: 74–87.
- Foote, K. G., Knudsen, H. P., Vestnes, G., MacLennan, D. N., and Simmonds. E., J. 1987. Calibration of acoustic instruments for fsh density estimation: A practical guide. ICES Cooperative Research Report, 144: 69 pp.
- Francois, R. E., and Garrison, G. R. 1982. [Sound-absorption](https://doi.org/Doi%2010.1121/1.388170) based on ocean measurements. Part 1: Pure water and [magnesium-sulfate](https://doi.org/Doi%2010.1121/1.388170) contributions. Journal of the Acoustical Society of America, 72: 896–907.
- Hewitt, R. P., and Demer, D. A. 2000. The use of acoustic sampling to estimate the dispersion and [abundance](https://doi.org/Doi%2010.1016/S0165-7836(00)00171-5) of [euphausiids,](https://doi.org/Doi%2010.1016/S0165-7836(00)00171-5) with an emphasis on Antarctic krill, *Euphausia superba*. Fisheries Research, 47: 215–229.
- Hill, K. T., Crone, P. R., and Zwolinski, J. P. 2017. Assessment of the Pacifc sardine resource in 2017 for U.S. Management in 2017-18. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-576: 264 pp.
- Johannesson, K., and Mitson, R. 1983. Fisheries acoustics. A practical manual for aquatic biomass estimation. FAO Fisheries Technical Paper.
- JTC. 2014. Status of the Pacifc Hake (whiting) stock in U.S. and Canadian waters in 2014 with a management strategy evaluation. Report.
- Kang, D., Cho, S., Lee, C., Myoung, J. G., and Na, J. 2009. Ex situ [target-strength](https://doi.org/10.1093/icesjms/fsp042) measurements of Japanese anchovy (*Engraulis japonicus*) in the coastal [Northwest](https://doi.org/10.1093/icesjms/fsp042) Pacifc. ICES Journal of Marine Science, 66: 1219–1224.
- Karp, W. A., and Walters, G. E. 1994. Survey assessment of semi-pelagic Gadoids: the example of walleye pollock, *Theragra chalcogramma*, in the Eastern Bering Sea. Marine Fisheries Review, 56: 8–22.
- Lo, N. C. H., Macewicz, B. J., and Grifth, D. A. 2011. [Migration](https://doi.org/DOI%2010.5343/bms.2010.1077) of Pacifc sardine (*Sardinops sagax*) of the West Coast of United States in [2003-2005.](https://doi.org/DOI%2010.5343/bms.2010.1077) Bulletin of Marine Science, 87: 395–412.
- Love, M. S. 1996. Probably More Than You Want to Know About the Fishes of the Pacific Coast. Really Big Press, Santa Barbara, CA.
- MacLennan, D. N., Fernandes, P. G., and Dalen, J. 2002. A consistent approach to [defnitions](https://doi.org/DOI%2010.1006/imsc.2001.1158) and symbols in fsheries [acoustics.](https://doi.org/DOI%2010.1006/imsc.2001.1158) ICES Journal of Marine Science, 59: 365–369.
- Mais, K. F. 1974. Pelagic fsh surveys in the California Current. State of California, Resources Agency, Dept. of Fish and Game, Sacramento, CA: 79 pp.
- Mais, K. F. 1977. Acoustic surveys of Northern anchovies in the California Current System, 1966-1972. International Council for the Exploration of the Sea, 170: 287–295.
- Nakken, O., and Dommasnes, A. 1975. The application of an echo integration system in investigations of the stock strength of the Barents Sea capelin 1971-1974. ICES C.M., B:25: 20.
- Ona, E. 2003. An expanded target-strength relationship for herring. ICES Journal of Marine Science, 60: 493–499.
- Palance, D., Macewicz, B., Stierhof, K. L., Demer, D. A., and Zwolinski, J. P. 2019. Length conversions and mass-length relationships of fve forage-fsh species in the California current ecosystem. Journal of Fish Biology, 95: 1116–1124.
- Peña, H. 2008. In situ [target-strength](https://doi.org/DOI%2010.1093/icesjms/fsn043) measurements of Chilean jack mackerel (*Trachurus symmetricus murphyi*) collected with a scientific [echosounder](https://doi.org/DOI%2010.1093/icesjms/fsn043) installed on a fishing vessel. ICES Journal of Marine Science, 65: 594–604.
- Polovina, J. J., Howell, E., Kobayashi, D. R., and Seki, M. P. 2001. The transition zone [chlorophyll](https://doi.org/10.1016/S0079-6611(01)00036-2) front, a dynamic global feature defning [migration](https://doi.org/10.1016/S0079-6611(01)00036-2) and forage habitat for marine resources. Progress in Oceanography, 49: 469–483.
- R Core Team. 2021. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. [https://www.R-project.org/.](https://www.R-project.org/)
- Renfree, J. S., and Demer, D. A. 2016. Optimising transmit interval and logging range while avoiding aliased

seabed echoes. ICES Journal of Marine Science, 73: 1955–1964.

- Renfree, J. S., Hayes, S. A., and Demer, D. A. 2009. [Sound-scattering](https://doi.org/DOI%2010.1093/icesjms/fsp069) spectra of steelhead (*Oncorhynchus mykiss*), coho (*O. kisutch*), and chinook (*O. [tshawytscha](https://doi.org/DOI%2010.1093/icesjms/fsp069)*) salmonids. ICES Journal of Marine Science, 66: 1091–1099.
- Saunders, R. A., O'Donnell, C., Korneliussen, R. J., Fassler, S. M. M., Clarke, M. W., Egan, A., and Reid, D. 2012. Utility of 18-kHz acoustic data for [abundance](https://doi.org/10.1093/icesjms/fss059) estimation of Atlantic herring (*Clupea harengus*). ICES Journal of Marine Science, 69: 1086–1098.
- Schwartzkopf, B. D., Dorval, E., James, K. C., Walker, J. M., Snodgrass, O. E., Porzio, D. L., and Erisman, B. E. 2022. A summary report on the life history information on the central subpopulation of Northern Anchovy *(Engraulis mordax)* for the 2021 stock assessment. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-659: 76 pp.
- Seabird. 2013. Seasoft V2 SBE Data Processing Manual Revision 7.22.4. Sea-Bird Electronics, Washington, USA.
- Simmonds, E. J., and Fryer, R. J. 1996. Which are better, random or [systematic](https://doi.org/DOI%2010.1006/jmsc.1996.0004) acoustic surveys? A [simulation](https://doi.org/DOI%2010.1006/jmsc.1996.0004) using North Sea herring as an example. ICES Journal of Marine Science, 53: 39–50.
- Simmonds, E. J., Gutierrez, M., Chipollini, A., Gerlotto, F., Woillez, M., and Bertrand, A. 2009. [Optimizing](https://doi.org/DOI%2010.1093/icesjms/fsp118) the design of acoustic surveys of Peruvian [Anchoveta.](https://doi.org/DOI%2010.1093/icesjms/fsp118) ICES Journal of Marine Science, 66: 1341–1348.
- Simmonds, E. J., and MacLennan, D. N. 2005. Fisheries Acoustics: Theory and Practice, 2nd Edition. Blackwell Publishing, Oxford.
- Simmonds, E. J., Williamson, N. J., Gerlotto, F., and Aglen, A. 1992. Acoustic survey design and analysis procedures: A comprehensive review of good practice. ICES Cooperative Research Report, 187: 1–127.
- Smith, P. E. 1978. Precision of sonar mapping for pelagic fsh assessment in the California Current. ICES Journal of Marine Science, 38: 33–40.
- Starr, R. M., Fox, D. S., Hixon, M. A., Tissot, B. N., Johnson, G. E., and Barss, W. H. 1996. Comparison of submersible-survey and hydroacoustic-survey estimates of fsh density on a rocky bank. Fishery Bulletin, 94: 113–123.
- Stierhof, K. L., Renfree, J. S., Rojas-González, R. I., Vallarta-Zárate, J. R. F., Zwolinski, J. P., and Demer, D. A. 2023. Distribution, biomass, and [demographics](https://doi.org/10.25923/77kp-ww39) of coastal pelagic fshes in the California Current Ecosystem during summer 2021 based on [acoustic-trawl](https://doi.org/10.25923/77kp-ww39) sampling. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-676: 86 pp.
- Stierhof, K. L., Zwolinski, G. E., J. P. Johnson, Renfree, J. S., Mau, S. A., Murfn, D. W., Sessions, T. S., and Demer, D. A. 2020a. Report on the 2019 California Current [Ecosystem](https://doi.org/10.25923/dg6w-q452) (CCE) Survey (1907RL), 13 June to 9 [September](https://doi.org/10.25923/dg6w-q452) 2019, conducted aboard NOAA Ship *Reuben Lasker*, fshing vessels *Lisa Marie* and *Long Beach Carnage*, and three [unmanned](https://doi.org/10.25923/dg6w-q452) sailboats. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-625: 49 pp.
- Stierhof, K. L., Zwolinski, J. P., and Demer, D. A. 2020b. [Distribution,](https://doi.org/10.25923/nghv-7c40) biomass, and demography of coastal pelagic fshes in the California Current Ecosystem during summer 2019 based on [acoustic-trawl](https://doi.org/10.25923/nghv-7c40) sampling. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-626: 80 pp.
- Swartzman, G. 1997. Analysis of the summer [distribution](https://doi.org/10.1006/jmsc.1996.0160) of fsh schools in the Pacifc Eastern Boundary [Current.](https://doi.org/10.1006/jmsc.1996.0160) ICES Journal of Marine Science, 54: 105–116.
- Thomas, G. L., Kirsch, J., and Thorne, R. E. 2002. Ex situ target strength measurements of Pacifc herring and Pacifc sand lance. North American Journal of Fisheries Management, 22: 1136–1145.
- Thomas, G. L., and Thorne, R. E. 2003. [Acoustical-optical](https://doi.org/10.1016/S0990-7440(03)00044-5) assessment of Pacifc Herring and their predator [assemblage](https://doi.org/10.1016/S0990-7440(03)00044-5) in Prince William Sound, Alaska. Aquatic Living Resources, 16: 247–253.
- Williams, K., Wilson, C. D., and Horne, J. K. 2013. Walleye pollock (*Theragra [chalcogramma](https://doi.org/10.1016/j.fishres.2013.01.016)*) behavior in [midwater](https://doi.org/10.1016/j.fishres.2013.01.016) trawls. Fisheries Research, 143: 109–118.
- Zhao, X., Wang, Y., and Dai, F. 2008. Depth-dependent target strength of anchovy (*Engraulis japonicus*) measured in situ. ICES Journal of Marine Science, 65: 882–888.
- Zwolinski, J. P., Bowlin, N. M., Erisman, B. E., James, K. C., Johnson, G. E., Mau, S. A., Murfn, D. W., *et al.* 2023. Report on the 2021 Spring CPS Survey (2103RL), 20 March to 13 April 2021, [conducted](https://doi.org/10.25923/xepm-9790) aboard NOAA Ship *Reuben Lasker* and fshing vessel *Long Beach [Carnage](https://doi.org/10.25923/xepm-9790)*. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-674: 30 pp.
- Zwolinski, J. P., Demer, D. A., Byers, K. A., Cutter, G. R., Renfree, J. S., Sessions, T. S., and Macewicz, B. J. 2012. Distributions and abundances of Pacifc sardine (*Sardinops sagax*) and other pelagic fshes

in the California Current Ecosystem during spring 2006, 2008, and 2010, estimated from acoustic-trawl surveys. Fishery Bulletin, 110: 110–122.

- Zwolinski, J. P., Demer, D. A., Cutter Jr., G. R., Stierhof, K., and Macewicz, B. J. 2014. Building on Fisheries Acoustics for Marine Ecosystem Surveys. Oceanography, 27: 68–79.
- Zwolinski, J. P., Demer, D. A., Macewicz, B. J., Cutter, G. R., Elliot, B. E., Mau, S. A., Murfn, D. W., *et al.* 2016. Acoustic-trawl estimates of northern-stock Pacifc sardine biomass during 2015. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-559: 15 pp.
- Zwolinski, J. P., Demer, D. A., Macewicz, B. J., Mau, S. A., Murfn, D. W., Palance, D., Renfree, J. S., *et al.* 2017. Distribution, biomass and demography of the central-stock of Northern anchovy during summer 2016, estimated from acoustic-trawl sampling. U.S. Dep. Commer., NOAA Tech. Memo., NMFS-SWFSC-572: 18 pp.
- Zwolinski, J. P., Emmett, R. L., and Demer, D. A. 2011. [Predicting](https://doi.org/DOI%2010.1093/icesjms/fsr038) habitat to optimize sampling of Pacifc sardine (*[Sardinops](https://doi.org/DOI%2010.1093/icesjms/fsr038) sagax*). ICES Journal of Marine Science, 68: 867–879.
- Zwolinski, J. P., Oliveira, P. B., Quintino, V., and Stratoudakis, Y. 2010. Sardine [potential](https://doi.org/DOI%2010.1093/icesjms/fsq068) habitat and [environmental](https://doi.org/DOI%2010.1093/icesjms/fsq068) forcing of western Portugal. ICES Journal of Marine Science, 67: 1553–1564.

## <span id="page-56-0"></span>**Appendix**

## <span id="page-56-1"></span>**A Length distributions and percent contribution to biomass by species and cluster**

## <span id="page-56-2"></span>**A.1 Northern Anchovy**

Standard length (*LS*) frequency distributions of Northern Anchovy (*Engraulis mordax*) per nighttime trawl cluster, annotated with the number of individuals caught and their percentage contributions to the abundance in each stratum.



## <span id="page-57-0"></span>**A.2 Pacifc Sardine**

Standard length (*LS*) frequency distributions of Pacifc Sardine (*Sardinops sagax*) per nighttime trawl cluster, annotated with the number of individuals caught and their percentage contributions to the abundance in each stratum.



## <span id="page-58-0"></span>**A.3 Jack Mackerel**

Fork length (*L<sup>F</sup>* ) frequency distributions of Jack Mackerel (*Trachurus symmetricus*) per nighttime trawl cluster, annotated with the number of individuals caught and their percentage contributions to the abundance in each stratum.



## <span id="page-59-0"></span>**B Investigation of Northern Anchovy ofshore of the surveyed area**

The biomass of the central stock of Northern Anchovy occupied mostly the central to western sections of the survey area (**Fig. [14a](#page-34-0)**). Two lines of reason indicate that the estimated stock biomass was not substantially biased due to any unsampled Northern Anchovy farther ofshore. First, and most signifcantly, there was practically zero biomass on the western ends of all the survey transects (**Fig. [24](#page-59-1)**). Second, there were few Northern Anchovy eggs west of the Spring 2021 CPS Survey area during the Spring 2021 CalCOFI<sup>[5](#page-59-2)</sup> Survey (data available via ERDDAP[6](#page-59-3)), less than a month later (**Fig. [24](#page-59-1)**).

<span id="page-59-1"></span>

Figure 24: Distribution of Northern Anchovy inferred from the Spring 2021 CPS and CalCOFI surveys. From the CPS survey, the red circles represent the smallest biomass densities that sum to 5% of the total biomass, and the green circles represent the largest biomass densities that sum to 95% of the biomass. From the CalCOFI survey, less than one month later, the blue circles indicate the smallest densities of Northern Anchovy eggs that sum to 5% of the total Northern Anchovy eggs, and the "+" symbols indicate the largest densities of Northern Anchovy eggs that sum to 95% of the total Northern Anchovy eggs.

<span id="page-59-2"></span><sup>5</sup>[https://calcof.org/](https://calcofi.org/)

<span id="page-59-3"></span><sup>6</sup><https://coastwatch.pfeg.noaa.gov/erddap/tabledap/erdCalCOFIcufes.html>