Contrasting climate velocity impacts in warm and cool locations show that effects of marine warming are worse in already warmer temperate waters

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Warming worse in already warmer temperate waters: a meta-analysis of the impact of climate trends and velocities on 38 species of demersal marine fishes

Impacts of climate velocity on demersal fish biomass depend on initial climate conditions: a metaanalysis across 38 species in the northeast Pacific

Running head: Contrasting climate velocity impacts

Abstract

- 2 Species responses to climate change are often measured at broad spatiotemporal scales, which can miss the fine-scale changes that are most relevant to conservation and fisheries management.
- 4 We develop a scale-able geostatistical approach to assess how juvenile and adult fish distributions have been shaped by changes in bottom temperature and dissolved oxygen over a recent decade
- 6 of warming in the northeast Pacific. Across 38 demersal fishes, biomass trends were associated
- negatively with warming and positively with dissolved oxygen, but when trends in both biomass
- 8 and climate were converted to velocities—the speed and direction a population would have to move to maintain consistent conditions—the effect of temperature change differed depending on
- local conditions. In the warmest locations, warming velocities were associated with negative bi-
- otic velocities for 19 of 69 species-maturity combinations, and yet were almost always associated
- with stable or positive biotic velocities in the coolest locations (64 of 69). These spatially consis-
- tent biomass declines (negative biotic velocities) in the warmest locations and increases in cooler
- locations suggest a redistribution of species with the potential for new ecological and fisheries
 - interactions. After controlling for temperature, the more spatially consistent effects of dissolved
- oxygen were often negative, suggesting a mechanism other than hypoxia avoidance—potentially
 - changes in primary production. Our approach identifies the species and locations that are most
 - sensitive to observed changes in the environment at any scale, thus facilitating future vulnerability
- assessments.

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- Keywords: biotic velocity, climate change, context dependence, dissolved oxygen, groundfish,
- 22 spatiotemporal distribution models

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Introduction

Managing the impacts of a rapidly changing climate on ecological communities, particularly those that provide food for humans, is a critical challenge facing society (e.g., Doney et al. 2012). An increase in atmospheric CO2 is not only causing increases in both mean ocean temperature and the frequency of extreme heat waves (Frölicher et al. 2018), but is also affecting patterns of circulation, productivity, and marine chemistry (Pörtner et al. 2019). Combined, these environmental changes can impact the distribution and abundance of many ecologically and commercially important fish species, leading to local loss of some species, colonizations, and changes in species interactions and bycatch composition (e.g., Pinsky and Fogarty 2012, García Molinos et al. 2016, Morley et al. 2018). Furthermore, such changes can cause the efficiency of fishing to increase if population density increases faster than range expansion during population growth, or temporarily maintain catch rates despite population decline if organisms move towards preferred habitat as it becomes available (the basin model of density-dependent habitat selection theory; MacCall 1990, Thorson et al. 2016b). Because traditional stock assessment methods, fisheries regulations, and choices regarding habitat protection generally assume stationary species distributions, new methods that anticipate and incorporate the effects of climate change on species distributions will be crucial for successful resource management in the future (Hare et al. 2010, Bell et al. 2020).

Species responses to climate change are often studied along range edges (Parmesan and Yohe 2003, Sunday et al. 2015, Fredston et al. 2020, Fredston-Hermann et al. 2020) or as aggregate indices (e.g., at species or region levels; Pinsky et al. 2013, Thorson et al. 2016a, Morley et al. 2018). The centre of gravity is perhaps the most commonly used measure of changes in distribution for marine fishes (e.g., Perry et al. 2005, Rindorf and Lewy 2006, Dulvy et al. 2008, Nye et al. 2009, Adams et al. 2018, Rooper et al. 2020). However, these approaches can overlook fine-scale spatial variation that may be important for understanding species responses (Oldfather et al. 2020) and distribution shifts may take longer to manifest at aggregate scales. Aggregate measures such as the centre of gravity are also challenging to interpret and apply in cases where both surveys and management actions are constrained by geographic or political boundaries that partition the distribution of a species. Indeed, laboratory experiments have demonstrated that thermal tolerances and optimums can differ sub-regionally (e.g., Pörtner et al. 2008) and there is evidence that warm range edges have shifted further north than expected and cold range edges contracted southward

(in the northern hemisphere), which suggest roles for competition, predator-prey interactions, and/or density dependent habitat selection (Fredston *et al.* 2020). Despite this, projections of future species' distributions often assume that responses to climate variables are consistent across space and time (e.g., Morley *et al.* 2018). While coarse-scale changes can be informative for long-term planning, changes in local abundances at finer spatial scales will likely occur more quickly due to the shorter dispersal distances involved and may be more informative in steering local conservation actions.

Local velocities are commonly used to quantify changes at finer spatial scales than captured in population-wide indices (e.g., center of gravity; Brito-Morales *et al.* 2018). A local climate velocity represents the movement of an isocline—a boundary along which a climate metric is constant. More intuitively, a climate velocity gives the speed and direction a population must move to maintain a constant climate condition (e.g., temperature) (Loarie *et al.* 2009). Gradient-based estimates of climate velocity are calculated as a trend in a climate metric through time (e.g., temperature trend), which can be positive or negative, divided by the local gradient in space comprised of a magnitude and direction (see Methods Eq. 12; Table 1; Burrows *et al.* 2011). These velocities scale local climate trends to emphasize locations where climate is relatively consistent across a neighbourhood of cells. Alternatively, analog-based velocities are estimated using search algorithms that identify nearest climate matches within a user-defined threshold of change from the reference cell conditions (Hamann *et al.* 2015). While analog-based velocity estimates can be more geographically precise, the choice of thresholds and other statistical properties (e.g., clumpiness) make them less useful than gradient-based local velocities for meta-analysis (Ordonez and Williams 2013).

Changes in abundance, density, or probability of species occurrence can also be expressed as velocities (e.g, Serra-Diaz et al. 2014, Comte and Grenouillet 2015, Alabia et al. 2018). When applied to species distribution models, these are referred to as biotic velocities and can be thought of as the minimum distance one would have to move to maintain an equivalent degree of habitat suitability (Carroll et al. 2015, Comte and Grenouillet 2015). Similar to climate velocity, a positive local biotic velocity is associated with an increase in habitat suitability at the focal location and a negative value represents a decline in suitability. The magnitude of the velocity estimates the distance to the nearest location that is predicted to match the original probability of occurrence or abundance. Because changes in climate may cause shifts in fish population density before range shifts

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based on presence-absence are clearly detectable, abundance and biomass-based models of species distributions are potentially more sensitive to local change than simple occupancy estimates.

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Bottom-trawl fisheries tend to capture a taxonomically and ecologically diverse suite of fishes. For example, the groundfish bottom-trawl fishery in Canadian Pacific waters encounters >100 species (Anderson et al. 2019), many of which are managed via an individual transferable quota system with 100% at-sea and dockside monitoring (Turris 2000, Wallace et al. 2015, DFO 2019). Random depth-stratified fishery-independent bottom trawl surveys have been fitted with conductivity, temperature, depth (CTD), and dissolved oxygen (DO) sensors since 2008. In addition to estimates of biomass density for each species captured in the surveys, data on size distributions and reproductive maturity are collected for many species (Anderson et al. 2019). Collectively, these species occupy a large range of depths, especially along the shelf edge where short movements can result from large environmental changes, and vary in their potential for behavioural responses to climate. For example, some species are migratory or highly mobile (e.g., Sablefish (Anoplopoma fimbria, Anoplopomatidae) and many flatfish species (Pleuronectidae)), while others are relatively sedentary (e.g., many species of rockfish, Sebastes spp. (Sebastidae)). Furthermore, a warmer ocean is expected to hold less oxygen, while becoming more stratified (Levin and Le Bris 2015), and groundfish species occupying different depths vary in their sensitivity to hypoxia (Keller et al. 2017).

Here, we explore the extent to which groundfish distributions in the northeast Pacific have been shaped by local temperature and DO trends and velocities over a decade spanning a relatively cool period through a recent marine heat wave (Okey et al. 2014, Frölicher and Laufkötter 2018). We do this by quantifying broad patterns and species-level relationships between climatic and biotic change in order to answer the following questions: (1) Are local changes in bottom temperature or DO correlated with changes in local groundfish densities, and are these effects stronger in already warm or low-oxygen regions? For example, has local warming had a larger effect in locations that are already at the warm-extreme of a species' local distribution? (2) How do these relationships differ between the spatial contexts captured by gradient-based velocities vs. their component trends? (3) How do these effects vary between species and are they correlated with life-history characteristics such as age and growth rate; or ecological traits such as depth range, latitude, trophic level, foraging zone, or sociality? We address these questions by using

spatiotemporal models applied to a decade of survey-derived climate and species density data, and then assess relationships between velocities of biotic and climatic change for 38 commonly encountered species using a geostatistically explicit hierarchical analysis that controls for change in both temperature and DO.

Methods

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Survey data

We analyzed biomass density distributions and morphometric data for 38 species of groundfish that were regularly encountered by fisheries-independent bottom-trawl surveys and are widely distributed within Canadian Pacific waters (DFO 2020, Table S1). The surveys were stratified 142 within four regions, two of which were surveyed in odd years (Hecate Strait and Queen Charlotte Sound) and two in even years (West Coast Vancouver Island and West Coast Haida Gwaii) since at least 2005. Each region was sampled over the same month-long period between late May and early August in each survey year (Fig. S1). These surveys share similar random depth-stratified designs, fishing gear and fishing protocols (Sinclair et al. 2003). Combined, they covered most of the upper continental slope and shelf in Pacific Canada, and resulted in a mean of 326 samples per year (range from 200 to 436). We only included tows of > 15 minutes duration and converted total biomass of each species to a biomass density based on the speed, distance covered, and net opening (e.g., Williams et al. 2018, Anderson et al. 2019). Biological sampling protocols varied among species, depending on size of catch and commercial importance. In general, catches of between 10-50 fish were sexed, weighed and measured individually, while larger catches were subsampled and, for commercially important species, maturity data and otoliths for aging were collected.

Estimating spatiotemporal variation in maturity-specific biomass density

Because ontogenetic shifts in habitat, particularly depth, are well documented for groundfish species (e.g., Mindel *et al.* 2016, Barbeaux and Hollowed 2018, Li *et al.* 2019), we estimated biomass densities separately for mature and immature size classes whenever possible. Maturity was not assessed for certain Chondrichthyans, or when catches were particularly low. To do this, we first estimated length at 50% maturity as defined by gonadal development stages using ogives fit as sex-

specific logistic regressions to individual specimens (see Supporting Methods). To split the estimated biomass density per tow into mature and immature components, we calculated the summed biomass of all measured fish that were above (for mature) or below (for immature) the length-at-50% maturity threshold, divided by the total biomass of all measured fish, and multiplied this ratio by the estimated biomass density for each tow. For each tow that resulted in too small a catch for detailed measurements to have been taken, we applied the mean ratio from all measured tows to estimate mature biomass (applied to a median of 9% [range 1–40%] of each species' total sampled biomass). For species without any maturity data, we assumed that the total biomass estimate represented the mature population, because mature individuals are larger, and therefore likely to be numerically dominant (* in Table S1). However, it is possible that biomass sampled in some areas was actually dominated by immature individuals, particularly for the skate species, whose mean lengths fall close to the sizes at maturity found in Love (2011).

We modelled spatiotemporal biomass density separately for mature and immature fish of each species using spatial GLMMs. Environmental variables, such as temperature, may be included in these models explicitly and can be used to assess the vulnerability or tolerance of a given species to change (Godefroid *et al.* 2019). However, such approaches require strong assumptions (e.g., that effects are constant through both time and space). Instead, we use a climate-agnostic version of a species distribution model with a spatiotemporal random effect structure to estimate local variability in biomass density change. Our models relied on spatial random effects to capture unmeasured components of habitat suitability and allow suitability to change through time without making assumptions about the shape of species-specific responses to possible climatic and geographic covariates (e.g., Shelton *et al.* 2014, Ward *et al.* 2015, Thorson *et al.* 2015b, 2017). We modelled each species and maturity class separately because, although these density patterns are correlated, explicitly modelling those relationships among groundfish species has not been shown to dramatically improve precision of overall estimates (Thorson and Barnett 2017), and maintaining independent estimates should make identification of shared climate responses more conservative.

We modelled biomass density with a Tweedie distribution and a log link because densities contain both zeros and positive continuous values (Tweedie 1984, Dunn and Smyth 2005, Anderson

et al. 2019):

$$Y_{s,t} \sim \text{Tweedie}(\mu_{s,t}, p, \phi), 1 (1)$$

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$$\mu_{s,t} = \exp\left(\alpha_t + \gamma_{1,t} D_{s,t} + \gamma_{2,t} D_{s,t}^2 + \omega_s + \epsilon_{s,t}\right),\tag{2}$$

$$\gamma_{1,t} \sim \text{Normal}\left(\gamma_{1,t-1}, \sigma_{\gamma_1}^2\right),$$
(3)

$$\gamma_{2,t} \sim \text{Normal}\left(\gamma_{2,t-1}, \sigma_{\gamma_2}^2\right),$$
(4)

$$\omega \sim \text{MVNormal}(\mathbf{0}, \Sigma_{\omega}),$$
 (5)

$$\epsilon_t \sim \text{MVNormal}(\mathbf{0}, \Sigma_{\epsilon}),$$
 (6)

where $Y_{s,t}$ represents the biomass density at point in space s and time t, μ represents the mean biomass density, p represents the Tweedie power parameter, and ϕ represents the Tweedie dispersion parameter. The parameter α_t represents the mean effect for each year, and $\gamma_{1,t}$ and $\gamma_{2,t}$ represent time-varying coefficients associated with depth (D) and depth-squared covariates (D^2) , respectively, which both follow a random walk constrained by $\sigma_{\gamma_1}^2$ and $\sigma_{\gamma_2}^2$. The initial values $\gamma_{1,t}$ and $\gamma_{2,t}$ at t=1 share an implied Uniform $(-\infty,\infty)$ prior. We considered alternative covariates not described here (see Supporting Methods) The parameters ω_s and $\epsilon_{s,t}$ represent spatial and spatiotemporal random effects that were assumed drawn from Gaussian Markov random fields (e.g., Latimer et al. 2009, Cressie and Wikle 2011, Lindgren et al. 2011) with covariance matrices Σ_{ω} and Σ_{ϵ} that were constrained by Matérn covariance functions (Cressie and Wikle 2011). The covariance matrices for a given maturity-species combination shared a common κ parameter that controls the rate of decay of spatial correlation with distance (Cressie and Wikle 2011).

We modelled the spatial components as random fields using a triangulated mesh with vertices selected using a k-means algorithm (via the k-means function in R; e.g., Shelton *et al.* 2014) at a specified number of locations, known as knots, used to approximate the spatial variability in observations. We used 500 knots for mature density, 400 for immature density, and 300 for less well-sampled species (Bocaccio *Sebastes paucispinis*, Shortraker Rockfish *S. borealis*, and immature Redstripe Rockfish *S. proriger*). Based on estimated values of the spatial surface at these knot

locations, we used bilinear interpolation to approximate a continuous spatial field (Rue *et al.* 2009, Lindgren *et al.* 2011).

We fit our models in R version 3.6.1 (R Core Team 2019) with the R package sdmTMB (Anderson et al. 2019, 2020), which interfaces automatic differentiation and the Laplace approximation in the TMB (Template Model Builder) R package (Kristensen et al. 2016) with the SPDE (Stochastic Partial Differential Equation) approximation to Gaussian Markov fields from the INLA (Integrated Nested Laplace Approximation) R package (Rue et al. 2009) to find the value of the fixed effects that minimizes the the marginal negative log likelihood. We confirmed that the non-linear optimizer had converged by checking that the Hessian matrix was positive definite and the maximum absolute gradient across fixed effects was < 0.005.

Climate trends and velocities

Bottom temperature and dissolved oxygen (DO) levels have been collected on most tows during the synoptic bottom trawl surveys since 2008, using Seabird Electronics SBE 19 profilers. From these measurements, we predicted seafloor climate using an approach similar to the one described above for biomass density except we used a Gaussian observation model, 800 knots, and allowed the spatiotemporal random fields to follow an autoregressive (AR1) process:

$$C_{s,t} \sim \text{Normal}\left(\mu_{s,t}, \sigma^2\right),$$
 (7)

$$\mu_{s,t} = \mathbf{X}_{s,t}\boldsymbol{\beta} + \omega_s + \mathbf{x}_{s,t},\tag{8}$$

$$\omega \sim \text{MVNormal}(0, \Sigma_{\omega}),$$
 (9)

$$x_{t=1} \sim \text{MVNormal}(\mathbf{0}, \Sigma_{\epsilon}),$$
 (10)

$$x_{t>1} = \rho x_{t-1} + \sqrt{1-\rho^2} \epsilon_t, \ \epsilon_t \sim \text{MVNormal}(0, \Sigma_{\epsilon}).$$
 (11)

Here $C_{s,t}$ represents the climate variable (bottom temperature or log DO) in space s and time t, μ represents the mean, and σ represents the observation error standard deviation. The symbol $X_{s,t}$ represents a vector of predictors (described below) and β represents a vector of corresponding parameters. The spatial random effects ω_s were defined as in Eq. 5 whereas the spatiotemporal random effects were structured to follow a stationary AR1 process with first-order correlation ρ . Because DO is known to be influenced by both water temperature and seasonal biological

processes, we included a quadratic effect for temperature and a linear effect for day of year along with estimated means for each year. Although not shown above for simplicity, we again allowed the quadratic depth covariates to follow a random walk through time as in Eq. 3. Our bottom temperature model fixed effects included only depth and estimated means for each year, because including day of year did not improve model fit based on AIC (Akaike Information Criterion). Because bottom temperature data (but not DO) have been collected in synoptic surveys since 2003, we included these earlier data in the temperature model to provide more information for estimating the fixed spatial random field ω_s .

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For all climate and biomass models, we then projected the model predictions onto a 4×4 km grid (UTM 9 projection) representing the survey domain. We excluded all cells with predicted conditions outside the range of conditions observed during sampling (99% quantiles of 3.07 to 11.3 °C and 0.28 to 7.06 ml/L DO). We then calculated gradient-based velocities of change and constituent local trends and spatial gradients for each cell (see Supporting Methods for discussion of alternative velocity calculations). Gradient velocities (V) were calculated as a ratio of the temporal trend (linear regression slope of each cell's climate time series) divided by gradient in space g of variable

$$V_{\rm s} = \left(\Delta A_{\rm s}/\Delta t\right)/g_{\rm s},\tag{12}$$

where A is any temporally varying feature of focal cell s. Depending on the portion of the survey grid considered, we calculated the trend through time for biennial time-steps between 2008 and 2018 (6 surveys across 11 years) or 2009 and 2017 (5 surveys across 9 years). In order to compare between survey areas with different sampling years, we converted values for all cells to a rate of change, or trend, per decade (Table 1). The spatial gradient g_s was calculated as the vector sum of the mean north-south and east-west gradients based on a 3x3 cell neighbourhood (Burrows et al. 2011); however, the values of A from which a spatial gradient is calculated can be based on any particular subset of times t, or the mean of all Δt . The input information related to a cell and any cell near enough to share spatial information in a gradient or in the overall spatial models (this includes all response and predictor variables) all share the same sample years.

The magnitudes of gradients g_s strongly influence the distribution of velocities V. Most prior applications of gradient-based velocities have used g_s calculated from the mean cell conditions of the entire period analyzed (e.g., Burrows *et al.* 2011, Molinos *et al.* 2019). Estimated velocities will

tend to be larger (as $g_s \to 0$, $V_s \to \infty$) when more estimates (in this case sample years) are averaged for the cells included in the g_s calculation, because a larger sample reduces the variability between the mean values of adjacent cells. Furthermore, the gradients most relevant to the actual distance a population would need to travel are those present after changes have begun to occur. Given that samples were collected only once every two years and that there is variability among species in terms of when dispersal occurs and how long it takes, we used the last two sample periods in our estimates of spatial gradients (2015–2018). This time period begins the first survey season following the onset of the 2014–2016 marine heat wave in the north-eastern Pacific (Peterson *et al.* 2015) and is approximately the end point of the transition to warmer conditions in the Bering sea (Alabia *et al.* 2018). We calculated spatial gradients using the vocc R package (Brown and Schoeman 2018), which produces identical results when applied to the same data as the newer package VoCC (García Molinos *et al.* 2019). Finally, we collapsed the most extreme velocity estimates to their 0.005 and 0.995 quantiles to reduce the impact of outliers from the resulting heavy-tailed distributions.

Linking biotic changes with climate

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To explore the relationship between change in estimated local climate and percent changes in estimated biomass densities for each 4 × 4 km grid cell, we used similar spatial GLMMs to control for spatial autocorrelation among cells. Our models estimated the rate of change in biomass (*Y*) of each maturity class of each species as a function of local climate change (temperature and DO) and an interaction between the mean climate of each cell and its local rate of change:

$$\Delta Y_{k[s]} \sim \text{Normal}\left(\mu_{k[s]}, \sigma^2\right),$$
 (13)

$$\mu_{k[s]} = \beta_{0,k[s]} + \beta_{1,k[s]} \overline{T}_s + \beta_{2,k[s]} \Delta T_s + \beta_{3,k[s]} \overline{T}_s \Delta T_s + \beta_{4,k[s]} \overline{O}_s + \beta_{5,k[s]} \Delta O_s + \beta_{6,k[s]} \overline{O}_s \Delta O_s + \beta_{7,k[s]} \overline{Y}_{k[s]} + \omega_{k[s]},$$

$$(14)$$

$$\omega_k \sim \text{MVNormal}(\mathbf{0}, \Sigma_\omega), \text{ for } k = 1, \dots, K,$$
 (15)

$$\beta_{r,k} \sim \text{Normal}\left(\eta_{r,k}, \sigma_{\beta_r}^2\right), \text{ for } k = 1, \dots, K \text{ and } r = 0, \dots, 7,$$
 (16)

where \overline{T}_s and ΔT_s represent the mean temperature and decadal trend in temperature for spatial location s. A row of data represents a given spatial grid cell s and species-maturity k combination. The symbols \overline{O}_s and ΔO_s represent mean dissolved oxygen and decadal trend in dissolved oxygen,

and the symbol $\overline{Y}_{k[s]}$ represents log biomass density for species-maturity k. Parameters β_0 through β_7 (indexed by r) represent coefficients that are allowed to vary as random effects across species with means $\eta_{r,k}$ and variances $\sigma^2_{\beta_r}$. We accounted for spatial autocorrelation through the spatial random effects $\omega_{k[s]}$, which follow a Matérn Gaussian Markov random field as described above.

We fit model configurations where both biomass and climate were calculated as either raw temporal trends or gradient-based velocities. The trend-based models assessed whether biomass changes were correlated with changes in climate at the 4×4 km grid cell scale. The velocity-based models assessed whether changes in biomass, especially those with low local variability in biomass, were correlated with the predicted speed of climate isoclines within the 12×12 km neighbourhood of cells. We did not include both trends and velocities in the same model because both the units and spatial scales captured are different.

For each maturity class of each species, we included all grid cells that encompassed 95% of the mean total biomass across all sample years and the mean log biomass density of each cell as a covariate to reduce the influence of changes occurring only at either the highest or lowest densities for a particular species. These models used a 600 knot mesh, Gaussian observation errors when estimating trends, and Student-t observation errors (with a degrees of freedom fixed at 7) to account for heavy-tailed residuals when estimating velocities. We scaled all covariates by their standard deviations. We centered local average temperature, DO, and log biomass density by their overall means, but kept temperature and DO trend variables uncentered to maintain interpretability. We tested additional covariates, including local changes in fishing intensity, but we have not included them in the final models because they did not change our conclusions (see Supplementary Methods).

Simulation study

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We conducted a simulation study to assess: (1) the ability of the geostatistical models to cope with the high levels of spatial covariance inherent in spatial grid-based climate and biomass estimates, and (2) to what extent similarities in climate and biotic spatial gradients were responsible for the observed patterns in the velocity-based models. We simulated biomass trends for each species as random fields using the true variance and spatial correlation parameters estimated for each species. Next, we assessed how well our trend-based model accounted for spatial autocorrelation

among grid-based estimates by re-fitting the trend-based model using four unique iterations of the simulated data and contrasting the effect sizes and number of species that showed a significant relationship with climate trends in the observed vs. simulated models. If the spatial random effects were effective in preventing type I errors, the trend-based models using simulated data should not show a significant effect of climate more than expected by chance. In the case of velocities, we used the ratio of the simulated biomass trends to the observed spatial gradients in biomass to simulate biotic velocities (henceforth, 'time-null' velocities). This approach maintains the relationships between spatial gradients in biomass and climate that are likely to occur because both species abundances and climate on the seafloor are correlated with depth. Rather than being a test of spatial autocorrelation, these time-null velocities were used to test how important the gradient component was to the results of the fitted velocity model.

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Null models based on simulated biotic trends and observed climate trends showed fewer significant relationships at the species level than would be expected by chance (Figures S5, S6, and S7), confirming that the spatial random effects in our models (e.g. Figure S8) successfully controlled for the spatial autocorrelation. In contrast, models predicting time-null biotic velocities did produce more significant effects then would be expected by chance (Figures S₉, S₁₀, and S₁₁ vs. S₁₂). These associations were likely due to the simulated velocities being based on the observed spatial gradients (Figure S13); however, comparisons between the velocity model and time-null models suggest that some patterns cannot be accounted for by similarities in the spatial gradients and can be reliably attributed to variation in temporal trends (differences between areas encompassed in black vs. grey violins for interaction terms in Figure 2b). This is in contrast to the complete overlap in black and grey violins for DO velocity in Figure 2b, which indicates that any set of species with identical overall distributions and population variability, but completely random biomass trends, would be likely to show just as many significant species-specific effects. Taken together, these simulations suggest that the velocity model effectively combines both the temporal and spatial dimensions of biotic and climate change, which provides support for our choice to focus on this approach.

Life-history and ecological correlates of climate sensitivity

To assess potential ecological mechanisms and the extent to which the temporal and spatial scales considered were appropriate for the different species, we used mixed-effect models to assess concordance between species' life-history traits and ecology and the estimated effect of climate velocity. We first assessed the independent effects of mean population age (among immature populations only) and occupied depth (mean and range). We then tested for independent relationships between climate sensitivity and ecological groupings (including range limits, foraging zones, trophic level, and sociality), while controlling for the depth total ranges occupied (see Supplementary Methods).

Results

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Climate trends and velocities

We estimated bottom temperature and DO values biennially between late May and early August during 2008–2018 or 2007–2019, depending on the surveyed area. Seafloor temperature varied from $4.6\,^{\circ}\text{C}$ to $10.2\,^{\circ}\text{C}$ (95% quantile range) across 4×4 km grid cells that had a mean depth within the 99th quantile range of sampled depths (23 to 1112 m). For the same range of survey depths, DO ranged between 0.7 ml/L and 5.7 ml/L (95% quantiles). For both temperature and DO, the highest values were associated with the shallowest depths, while the lowest values were associated with the deepest locations (Figure 1b, c).

Over this period, summer seafloor temperature increased by an average of 0.6 °C per decade across the entire region (95% quantile range of -0.2 to 1.8 °C per decade; Figure 1d). In contrast, the direction of change in seafloor DO was more variable (95% quantiles: -2.8 to 0.6 ml/L per decade; Figure 1e). Warming tended to be most pronounced in the already warmer locations—mean of 1.3 °C/decade in cells shallower than 50 m (Figure S14b). Likewise, the greatest decreases in DO occurred in the shallowest locations (mean: -2 ml/L per decade); however, the highest variability in DO trend (95% quantiles: -1.7 to 0.6 ml/L per decade) occurred between 50 and 200 m depths (Figure S14e). There was a tendency for the shallowest depths to be occupied by groundfish species that have narrower depth ranges (e.g., Southern Rock Sole, *Lepidopsetta billineta*, vs. Dover Sole, *Microstomus pacificus* (Pleuronectidae); Figure S14g, Table S1).

When these local climate trends were placed in their geographic context by converting to gradient-based velocity estimates (Eq. 12), they implied that a population would have to move an average of 10.5 km/decade (mean of absolute values) to maintain its starting thermal environment and an average of 11 km/decade to maintain initial DO levels. Temperature velocities averaged positive, representing warming conditions (mean: 10.1; 95% interquantile range of -12 to 87; Figure 1f), while DO velocities averaged negative, representing declining DO levels (-6.26; -91 to 24; Figure 1g). Most locations with high climate velocities occurred in patches throughout Queen Charlotte Sound and Hecate Strait (dark red patches in Figure 1f). The most negative DO velocities occurred in shallower portions of Hecate Strait (largest green patch in Figure 1g). The largest velocities tended to be found across a broader range of depths than the largest climate trends (Figure S14).

Linking biotic changes with climate

Geostatistical models linking climatic (Figure 1d–g) and biotic trends (Figures S17 and S18) or velocities (Figures S19 and S20) resolved different aspects of biotic change (Figures S8 and S12). The effect of temperature velocity on biotic velocity was weakly positive across species (β : 0.28 km/decade with 95% CI -0.04 to 0.60; point range for "T change" shown in Figure 2b), despite a significant overall 0.55% decline in biomass (-0.87 to -0.22) per 1 SD increase in warming (0.8 °C per decade) based on local temperature trend only (point range for "T change" shown in Figures 2a). However, mean local temperature influenced the effect of temperature velocity on biotic velocity (β : -1.09, -1.48 to -0.70; "T interaction" in Figure 2b), such that when temperature velocity was high in the warmest parts of a species' range, local biomass was more likely to decline and exhibit larger negative or smaller positive biotic velocities. When temperature velocity was high in the coolest parts of a species range, local biomass was more likely to increase and to result in larger positive biotic velocities (or to decrease less and result in less negative biotic velocities).

Interactions between mean climate and climate velocity for each maturity class of each species can be illustrated as the predicted relationships between climate and biotic velocities at different mean local conditions (e.g., in Figures 3c and S15a, the blue and red lines are the predicted relationships for locations at the 0.025 and 97.5 quantile of mean local temperatures, respectively). For Redbanded Rockfish (*Sebastes babcocki*, Sebastidae) the horizontal blue line indicates stable biomass

(small absolute biotic velocities), while the red line with a negative slope means that biomass was more likely to be declining across a local area where conditions were warmest on average and getting warmer across more of the surrounding area (Figure 3b). The slopes of all predicted relationships (e.g., as illustrated in Figure 3c and Figure S15a) are plotted for all species-maturity combinations in Figure 4. Consistent with the overall interaction, the majority of species-maturity combinations with significant interactions between local mean temperature and temperature velocity had negative interactions (31 of 33 coloured dots and lines with red coefficients to the left of blue coefficients in Figure 4a). Over a third of these cases predicted a positive effect of increased temperature velocities for both the warmest and coolest locations, but that the relationship was more strongly positive in the cooler locations (13 of 31 species-maturity combinations with negative interactions).

To assess whether these relationships predicted that specific species' biotic velocities were increasing or decreasing under different climate velocities, the lines displayed in Figures 3c and S15a can be "sliced" at either the minimum temperature velocity experienced by each species (left end of lines) or at the maximum (right end of lines). The expected biotic velocity was near zero for most species-maturity combinations in locations experiencing minimum climate velocity (Figure 5a) regardless of mean temperature. However, in the warmest locations experiencing maximum climate velocity, the expected biotic velocity was strongly negative for 19 of 69 species-maturity combinations (Figure 5b). Meanwhile, in cooler locations experiencing the same high climate velocity, biotic velocities were often positive (e.g., Pacific Halibut, *Hippoglossus stenolepis*, Pleuronectidae).

After controlling for temperature, the average effect of DO velocity on biotic velocity was negative across species (β : -0.48 km/decade, -0.82 to -0.15; point and range for "DO change" in Figure 2b) despite there being a positive effect of DO trend on biomass trend (β : 0.34 % increase in biomass, 0.16 to 0.52; point and range for "DO change" in Figure 2a). Thus, while increasing DO was associated with increases in biomass at a local scale, DO velocity was not on average correlated with biotic velocities. However, unlike for temperature, DO velocity did not interact with mean DO availability consistently across species (β : 0.25, -0.05 to 0.55). Only two species (those with green point ranges on the positive side of the x-axis in Figure 4b) showed the expected interaction where locations with lower mean DO levels experiencing positive DO velocities were associated with decreases in biotic velocity and/or negative DO velocities were associated with decreases in

biotic velocity (e.g., immature Lingcod, *Ophiodon elongatus*, Hexagrammidae; Figure 3). In contrast, several species experienced declines in biotic velocity when DO velocity increased across the range of mean DO levels (black point ranges on negative side of x-axis in Figure 4b). Including DO in the model along with temperature did not substantially alter the effects of temperature for any species (Figure S16).

Life-history and ecological correlates of climate sensitivity

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We examined possible relationships between responses to climate velocities and each species' taxonomy, traits, and depth distributions. Relationships with biotic velocities that were negative at high temperatures or positive at low temperatures occurred in members of both the largest families represented in our analysis, Sebastidae (rockfish) and Pleuronectidae (righteye flounders) suggesting no strong patterns of similarity among species belonging to the same genus or family (Figures 4a and S12; see also non-significant family-level effects from hierarchical model, Figure S21). However, the effects of temperature velocities at high temperatures were most negative for Chondrichthyan biotic velocities (-1.8 km/decade per SD in temperature velocity, same units apply elsewhere) and rockfish species occupying shelf habitats (-1.3), neutral for continental slope rockfish (0.1) and flatfish (-0.1), and most positive for sablefish (1.0; mean across red values in Figure 4a).

Life-history failed to explain substantial variation in climate sensitivity in the warmest loca-442 tions, although more negative effects tended to be clustered in shallow depths and among younger immature populations (Figure S22). However, the positive effects of temperature velocities on biotic velocities in the coolest locations were strongest in species occupying a larger range of depths $(\beta: 0.57, 0.21 \text{ to } 0.92)$ and for immature populations with younger mean age $(\beta: -1.0, -2.0 \text{ to } 0)$. 446 Ecological factors were somewhat better at accounting for negative effects in the warmest locations. The effects of temperature velocity at high mean temperatures differed significantly between 448 species depending on diet (lower biotic velocities in zooplantivores than species at higher trophic levels; β : -0.96, -1.44 to -0.48), and use of foraging zones (higher biotic velocities in demersal 450 species relative to benthopelagic; β: 0.98, 0.47 to 1.49) after accounting for mean depth occupied (Figure S24 top row). The strongest negative effects of warming temperature velocities (estimated for the warmest parts of a species distribution) were for species occurring at intermediate depths, whereas most species with mean encounter depths deeper than 290 m appeared to increase in biotic velocity with more positive temperature velocities (Figure S25c).

In contrast, DO velocities at low mean DO locations only showed a strong positive effect on Lingcod biotic velocities (0.7) and negative effects were strongest for both continental slope rock-fish (-1.1) and flatfish (-1.4; mean across green values in Figure 4b). These negative relationships represent declining biotic velocity with increasing DO, or visa versa, and tended to be stronger both for species occupying deeper locations on average (β: -0.43, -0.73 to -0.13; Figure S25d) and a larger range of depths (β: -0.30, -0.6 to 0). It is notable, however, that the effect of trends in DO on percent change in biomass were also negative at these depths despite being mostly positive at intermediate depths (Figure S25b). At these intermediate depths (the mean occupied depth for species in this analysis of about 175 m), the effects of DO velocity at low DO was also most negative for species foraging at higher trophic levels (β: -0.33, -0.81 to 0.16), in the demersal zone (β: -0.79, -1.32 to -0.26), and with more solitary habits (β: -0.63, -1.21 to -0.06; Figure S24 bottom row).

Discussion

Using novel geostatistical models fit to bottom temperature, DO, and demersal fish biomass from 468 scientific trawl surveys, we related trends and velocities between climatic and biotic variables across 38 species. Local declines in demersal fish biomass were, on average, associated with warm-470 ing trends and decreases in DO. However, after converting trends to velocities, a clear interaction between temperature velocity and mean bottom temperature emerged. On average, and for roughly half the species-maturity combinations, temperature velocity had a more negative effect on biotic velocity in already warm locations than in relatively cool locations. Converting these 474 interaction effects into expected values, approximately one quarter of species-maturity combinations experienced declines (negative biotic velocities) in the warmest locations when experiencing 476 maximum warming. In contrast, locations experiencing minimal warming or cool locations experiencing maximum warming experienced stable or increasing biotic velocities. Characteristics such as trophic level, foraging zone, and sociality—as well as potentially confounding variables such as commercial fishing effort or catch-explained little of the observed effects. Although DO velocity results were more equivocal, planktivores responded more positively to DO velocity under low DO conditions (Figure S24f) than species with more diverse or higher trophic-level diets.

This suggests that the prevalence of strong negative relationships between DO and biotic velocities

(Figure 2b) might be explained by increases in primary production, causing decreases in DO due to increased rates of decomposition in benthic environments (Keister et al. 2020).

Scale and context dependence

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Our analysis is the first, to our knowledge, to explore how the interaction between climate velocities and local mean climate conditions affect fine-scale biotic velocities, and the first to contrast patterns between trend and velocity indices. Relationships between climate and biotic velocities have also been detected in marine species in the Arctic (Alabia et al. 2018) and between local climate velocities and species range shifts in tropical to subarctic zones along the Japanese archipelago (Kumagai et al. 2018) and along several portions of the continental shelf of North America (Pinsky et al. 2013, Fredston-Hermann et al. 2020). At a global scale, the impact of temperature change in marine environments appears to be highly dependent on baseline conditions, whether measured in range shifts relative to temperature velocity (e.g., Lenoir et al. 2020) or species richness and abundance in response to temperature trends (e.g., Antão et al. 2020). Specifically, this latter meta-analysis of patterns in marine taxa found that abundance was positively correlated with warming, except in the warmest of locations (Antão et al. 2020). The wide geographic scope of these analyses suggest that our focus on both local climate velocity and baseline environmental conditions may be broadly relevant to explaining climate change induced range shifts in marine taxa in regions around the globe.

While our trend model indicated an overall negative effect of rising temperatures on local fish density, the velocity model was consistent with the overall pattern in showing that most negative relationships occurred only in already warm locations (Figure 2). Large climate velocities reflect more spatially uniform environments—where a population would need to move greater distances to maintain constant climate—and likewise, small velocities reflect more spatially heterogeneous environments (Loarie *et al.* 2009). As a result, more spatially uniform regions have greater weight in the velocity model than in the trend model, and this stretching and compressing of trend values based on spatial heterogeneity likely explains why the negative interactive effect of mean temperature was only detected for velocities. Overall, we focused primarily on the velocity results since

they represent a more ecologically meaningful measure than trends alone, given that they account for the local reality species face if tracking a constant environment (Brito-Morales *et al.* 2018) or prey that themselves track the environment.

For many species, we do not know how far individuals travel on a daily or seasonal basis, so 514 uncertainty remains as to the extent to which the modelled spatial resolution is appropriate for each of the species in this analysis. While the trend- and velocity-based models capture slightly different spatial scales (4 \times 4 km focal cell vs. 12 \times 12 km encompassed when considering patterns among neighbouring cells as well), both resolutions are finer than what is often used for analyses of climate change in the marine environment (Oestreich et al. 2020, Pinsky et al. 2020), substantial environmental changes can occur at even finer scales, and these local-scale effects may be especially important for species with high site fidelity (e.g., Yelloweye Rockfish Sebastes ruberrimus, Hannah and Rankin 2011). However, in order to detect the impact of climate change on rockfish (many of which have generation times > 20 yrs), one would require either data in excess of 20 years, or to contrast patterns of change between age classes. Indeed, responses to environmental change can be expected to differ between species, depending on the life history of species including physiological tolerances, lifespan, and dispersal patterns (Massiot-Granier et al. 526 2018). Furthermore, reaching reproductive maturity frequently results in shifts in dispersal patterns, habitat selectivity, physiological tolerances (Laurel et al. 2007), and therefore represents a potentially important break point for understanding the impacts of climate change. Given the relatively short timescale encompassed in our analysis (one decade), we expected to find stronger 530 patterns in shorter-lived/immature portions of populations and more pelagic species. Within immature populations, those with a younger mean age were found to exhibit the most extreme responses to temperature velocity (positive in coolest locations and negative in warmest locations; Figure S22c); however, immature populations did not have overall stronger responses than mature 534 populations (Figure S23b). Ecological responses were somewhat complicated by interactions with depths occupied; but, contrary to expectation, more extreme responses tended to belong to dem-536 ersal foraging and solitary species, rather than those with more pelagic or schooling behaviours 538 (Figure S24c, g, h).

Aggregate metrics such as the centre of gravity have also demonstrated that demersal fishes use both shifts in latitude and depth to track changes in ocean temperatures (e.g., Perry et al. 2005,

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Dulvy et al. 2008, Li et al. 2019), but evidence that range edges on the North American continental shelf have shifted further north than expected or even contracted southward, suggest roles for competition, climate-independent mortality and/or density-dependent habitat selection (Li et al. 2019, Fredston et al. 2020). Indeed, fishing pressure on the Atlantic shelf was found to be more important than average bottom temperature for predicting centre of gravity for five groundfish species, despite temperature being more correlated with variance in biomass (Adams et al. 2018). This latter result suggests that there was spatial variability in temperature, or responses to temperature, which were not fully captured by the centre of gravity (VanDerWal et al. 2013). Fine-scale local effects may contribute to the relatively greater influence of temperature relative to fishing pressure in our analysis. Groundfish species in the eastern Bering Sea also do not show a strong correlation even for local climate and biotic velocities, but no interaction with mean conditions was reported (Alabia et al. 2018). Another potential explanation for stronger negative effects on Canadian Pacific groundfish is that species here are closer to the southern ends of their distributions and may therefore be closer to the warm end of their temperature tolerances, especially in the warmest locations. However, although we found the strongest negative biotic velocities in these warmer locations, species nearer to their northern range limit (designated by N under 'Range limit' in Table S1) were not more likely to show a positive response to temperature.

Limitations and implications

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There are a number of limitations to our analysis. First, our analysis cannot separate fish population movement in response to climate from a host of other possible explanations. For example, local changes in biomass density can be a result of movement, local population growth, age cohorts beyond our two maturity categories, changes in average body size (Shackell *et al.* 2010, Laurel *et al.* 2007), or effects of fishing not captured by the metrics of total catch or hours fished. Indeed, some of the hypothesized effects of warming climate and lower DO on fishes include higher metabolism and ability to store fat, reduced productivity, and slower growth resulting in generally smaller fish (Klein *et al.* 2017, Madeira *et al.* 2017). Furthermore, changes in local density may be correlated with climate, not because of groundfish thermal preference, but because groundfish seek prey or avoid predators that have themselves shifted their distribution in response to climate. Second, there are limitations to our data. The CTD climate data from Canadian Pacific trawl surveys are

only available from 2008 onward, the surveys occur in one seasonal period (May to August) and cover a given region only once every two years. Also, some of the species (e.g., shallower rockfish species) may be better sampled by longline gear than trawl gear. This spatial and temporal scope will miss overlap in major life-history events for some species (e.g., Sablefish; Beamish 2008), or seasonal movements (e.g., Pacific Halibut; Loher 2011). Importantly, the input data for our meta-analytic model are predictions from our first-stage geostatistical models. Third, it is possible that climate conditions may themselves affect survey catchability. For example, groundfish may flee gear more slowly or aggregate to avoid low DO conditions (Craig 2012), thereby making fish more catchable and biasing observations.

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This analysis suggests multiple future research directions. First, future efforts may aim to identify common spatial patterns across species using spatial dimension reduction tools such as spatial factor analysis (Thorson et al. 2015a). Areas where species overlap in their response would represent important areas for conservation (Brito-Morales et al. 2018), but also areas where competition may be expected to increase or new fisheries interactions may occur. Second, some rockfish are better sampled by longline survey gear and future analyses could use such survey data, or combine survey data from multiple gear types (Webster et al. 2020), to develop a composite density estimate. Third, our analysis used CTD data, which was only available for spring or summer from 2008 onwards and required a statistical model to extrapolate to the full region. An alternative would be a ROMS (Regional Ocean Modeling System) model (Peña et al. 2019), which could extend the temporal scope, allow for accounting of climate at other times of the year (e.g., temperature during spawning; Laurel and Rogers 2020), allow for inclusion of variables not typically measured with survey data (e.g., primary production), and allow for forward projections. Preliminary investigations indicated a strong correlation between our CTD projections and recently updated ROMS bottom temperatures. With the greater spatial and temporal extent that ROMS data will provide, calculation of more geographically precise analog-based climate velocities could be used to further refine the identification of areas important for conservation (Brito-Morales et al. 2018).

The spatial shifts we identified could have a number of management consequences. First, we observed changes over a decade and such redistribution is likely to compound over time. Redistribution can impact fishing opportunities and conservation of rarer species when "choke" species (species with limited quota that co-occur with species of fishing interest) limit fishing opportu-

nities (e.g., Forrest *et al.* 2020). Redistribution can also impact nations or peoples with relatively small defined spatial fishing rights and have consequences for marine spatial planning. For example, a marine reserve designed to protect a particular at-risk population may no longer be as effective after a local redistribution of abundance. Lastly, shifting distributions can affect calculation of indices of abundance, and estimates of stock size and stock status, which in turn may impact harvest recommendations (Szuwalski and Hollowed 2016, Karp *et al.* 2019).

Climate change is expected to have large impacts on fish stocks and their management, particularly with respect to changes in species distribution (e.g., Tommasi et al. 2017, Karp et al. 2019, Free et al. 2019). Legislation and policy in jurisdictions around the world (e.g., the US Magnuson-Stevens Act, Canada's Fisheries Act, the European Marine Strategy Framework Directive) require that environmental conditions affecting fish stocks be accounted for in management decisions such as setting sustainable catch limits and developing rebuilding plans. However, there is often a mismatch between scale of climate predictions, the scale at which species respond, and the scale of management decisions (Stock et al. 2011, Maureaud et al. 2021). For example, the populations analyzed in this study are managed at the mesoscale, with catch limits often determined for individual substocks (DFO 2019). The metrics presented in our paper represent fine-scale indicators of response to a changing environment, which are useful for assessing risk and conservation planning (Brito-Morales et al. 2018). Analyses such as ours can be incorporated into frameworks for improving advice for the management of fisheries under climate change (e.g. Plagányi et al. 2011, Punt et al. 2014, Karp et al. 2019).

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Data Availability Statement

The biological data that used in this study are openly available through https://open.canada.ca at https://open.canada.ca/data/en/dataset/a278d1af-d567-4964-a109-ae1e84cbd24a and associated environmental measurements are available from https://www.pac.dfo-mpo.gc.ca/stats/indexeng.html. All of the code underlying this analysis is available at https://github.com/pbs-assess/gfvelocities.

References

- Adams, C.F., Alade, L.A., Legault, C.M., O'Brien, L., Palmer, M.C., Sosebee, K.A. and Traver,
 M.L. (2018) Relative importance of population size, fishing pressure and temperature on the
 spatial distribution of nine Northwest Atlantic groundfish stocks. *PLOS ONE* 13, e0196583.
 doi:10.1371/journal.pone.0196583.
- Alabia, I.D., Molinos, J.G., Saitoh, S.I., Hirawake, T., Hirata, T. and Mueter, F.J. (2018) Distribution shifts of marine taxa in the Pacific Arctic under contemporary climate changes. *Diversity and Distributions* 24, 1583–1597. doi:10.1111/ddi.12788.
- Anderson, S.C., Keppel, E.A. and Edwards, A.M. (2019) A reproducible data synopsis for over 100 species of British Columbia groundfish. *DFO Can. Sci. Advis. Sec. Res. Doc.* 2019/041. URL http://www.dfo-mpo.gc.ca/csas-sccs/Publications/ResDocs-DocRech/ 2019/2019_041-eng.html.
- Anderson, S.C., Ward, E.J., English, P.A. and Barnett, L.A.K. (2020) sdmTMB: Spatiotemporal Species

 Distribution GLMMs with 'TMB'. URL https://github.com/pbs-assess/sdmTMB. R package version 0.0.11.9000.
- Antão, L.H., Bates, A.E., Blowes, S.A., Waldock, C. et al. (2020) Temperature-related biodiversity change across temperate marine and terrestrial systems. Nature Ecology and Evolution 4, 927–933. doi:10.1038/s41559-020-1185-7.
- Barbeaux, S.J. and Hollowed, A.B. (2018) Ontogeny matters: Climate variability and effects on fish distribution in the eastern Bering Sea. *Fisheries Oceanography* 27, 1–15. doi:10.1111/fog.12229.

- Beamish, R.J. (2008) Impacts of climate and climate change on the key species in the fisheries in the North Pacific. *North Pacific Marine Science Organization Scientific Report* 35.
- Bell, R.J., Odell, J., Kirchner, G. and Lomonico, S. (2020) Actions to promote and achieve climateready fisheries: summary of current practice. *Marine and Coastal Fisheries* 12, 166–190. doi:10.1002/mcf2.10112.
- Bizzarro, J., Yoklavich, M. and Wakefield, W. (2017) Diet composition and foraging ecology of U.S.
 Pacific Coast groundfishes with applications for fisheries management. *Environmental Biology* of Fishes 100. doi:10.1007/s10641-016-0529-2.
- Brito-Morales, I., García Molinos, J., Schoeman, D.S., Burrows, M.T. *et al.* (2018) Climate Velocity

 Can Inform Conservation in a Warming World. *Trends in Ecology & Evolution* 33, 441–457.

 doi:10.1016/j.tree.2018.03.009.
- Brown, C. and Schoeman, D. (2018) vocc: Functions for Calculating the Velocity of Climate Change.

 URL https://github.com/cbrown5/vocc. R package version 0.1.1.
- Burrows, M.T., Schoeman, D.S., Buckley, L.B., Moore, P. *et al.* (2011) The pace of shifting climate in marine and terrestrial ecosystems. *Science* 334, 652–655. doi:10.1126/science.1210288.
- Carroll, C., Lawler, J.J., Roberts, D.R. and Hamann, A. (2015) Biotic and Climatic Velocity
 Identify Contrasting Areas of Vulnerability to Climate Change. *PLOS ONE* 10, e0140486.
 doi:10.1371/journal.pone.0140486.
- Comte, L. and Grenouillet, G. (2015) Distribution shifts of freshwater fish under a variable climate:

 Comparing climatic, bioclimatic and biotic velocities. *Diversity and Distributions* 21, 1014–1026.

 doi:10.1111/ddi.12346.
- Craig, J.K. (2012) Aggregation on the edge: effects of hypoxia avoidance on the spatial distribution of brown shrimp and demersal fishes in the Northern Gulf of Mexico. *Marine Ecology Progress* Series 445, 75–95. doi:10.3354/mepso9437.
- Cressie, N.A.C. and Wikle, C.K. (2011) *Statistics for Spatio-Temporal Data*. Wiley Series in Probability and Statistics. Wiley, Hoboken, N.J.

- DFO (2019) Pacific region integrated fisheries management plan, groundfish, effective february 21, 2019, version 1.1. URL http://www.pac.dfo-mpo.gc.ca/fm-gp/mplans/ground-fond-ifmp-pgip-sm-eng.html.
- DFO (2020) Groundfish Synoptic Bottom Trawl Surveys Open Government Portal. URL https://open.canada.ca/data/en/dataset/a278d1af-d567-4964-a109-ae1e84cbd24a.
- Doney, S.C., Ruckelshaus, M., Emmett Duffy, J., Barry, J.P. *et al.* (2012) Climate change impacts on marine ecosystems. *Annual Review of Marine Science* 4, 11–37. doi:10.1146/annurev-marine-041911-111611.
- Dulvy, N.K., Rogers, S.I., Jennings, S., Stelzenmüller, V., Dye, S.R. and Skjoldal, H.R. (2008) Climate change and deepening of the North Sea fish assemblage: A biotic indicator of warming seas.

 *Journal of Applied Ecology 45, 1029–1039. doi:10.1111/j.1365-2664.2008.01488.x.
- Dunn, P.K. and Smyth, G.K. (2005) Series evaluation of Tweedie exponential dispersion model densities. *Statistics and Computing* 15, 267–280. doi:10.1007/s11222-005-4070-y.
- Forrest, R.E., Stewart, I.J., Monnahan, C.C., Bannar-Martin, K.H. and Lacko, L.C. (2020) Evidence for rapid avoidance of rockfish habitat under reduced quota and comprehensive at-sea monitoring in the British Columbia Pacific halibut fishery. Can. J. Fish. Aquat. Sci. 77, 1409–1420. doi:10.1139/cjfas-2019-0444.
- Fredston, A., Pinsky, M., Selden, R., Szuwalski, C., Thorson, J., Halpern, B. and Gaines, S. (2020)

 Range edges of North American marine species are tracking temperature over decades. Preprint:
- 698 https://doi.org/10.22541/au.160331933.33155622/v1.
- Fredston-Hermann, A., Selden, R., Pinsky, M., Gaines, S.D. and Halpern, B.S. (2020) Cold range edges of marine fishes track climate change better than warm edges. *Global Change Biology* **26**, 2908–2922. doi:10.1111/gcb.15035.
- Free, C.M., Thorson, J.T., Pinsky, M.L., Oken, K.L., Wiedenmann, J. and Jensen, O.P. (2019)
 Impacts of historical warming on marine fisheries production. *Science* **363**, 979–983.

 doi:10.1126/science.aau1758.

- Frölicher, T.L., Fischer, E.M. and Gruber, N. (2018) Marine heatwaves under global warming. *Nature* 560, 360. doi:10.1038/s41586-018-0383-9.
- Frölicher, T.L. and Laufkötter, C. (2018) Emerging risks from marine heat waves. *Nature Commu*nications 9, 650. doi:10.1038/s41467-018-03163-6.
- García Molinos, J., Schoeman, D.S., Brown, C.J. and Burrows, M.T. (2019) *VoCC: The Velocity of Cli-*mate Change and related climatic metrics. URL https://doi.org/10.5281/zenodo.3382092.
 R package version 1.0.0.
- García Molinos, J., Halpern, B., Schoeman, D., Brown, C. et al. (2016) Climate velocity and the future global redistribution of marine biodiversity. Nature Climate Change 6, 83–88.

 doi:10.1038/nclimate2769.
- Godefroid, M., Boldt, J.L., Thorson, J.T., Forrest, R. *et al.* (2019) Spatio-temporal models provide new insights on the biotic and abiotic drivers shaping Pacific Herring (*Clupea pallasi*) distribution. *Progress in Oceanography* 178, 102198. doi:10.1016/j.pocean.2019.102198.
- Hamann, A., Roberts, D.R., Barber, Q.E., Carroll, C. and Nielsen, S.E. (2015) Velocity of climate change algorithms for guiding conservation and management. *Global Change Biology* 21, 997–1004. doi:10.1111/gcb.12736.
- Hannah, R.W. and Rankin, P.S. (2011) Site fidelity and movement of eight species of Pacific rockfish at a high-relief rocky reef on the Oregon coast. *North American Journal of Fisheries Management* 31, 483–494. doi:10.1080/02755947.2011.591239.
- Hare, J.A., Alexander, M.A., Fogarty, M.J., Williams, E.H. and Scott, J.D. (2010) Forecasting the dynamics of a coastal fishery species using a coupled climate-population model. *Ecological Applications* 20, 452-464. doi:10.1890/08-1863.1.
- Karp, M.A., Peterson, J.O., Lynch, P.D., Griffis, R.B. *et al.* (2019) Accounting for shifting distributions and changing productivity in the development of scientific advice for fishery management. *ICES Journal of Marine Science* 76, 1305–1315. doi:10.1093/icesjms/fsz048.
- Keister, J.E., Winans, A.K. and Herrmann, B. (2020) Zooplankton Community Response to Seasonal Hypoxia: A Test of Three Hypotheses. *Diversity* 12, 21. doi:10.3390/d12010021.

- Keller, A.A., Ciannelli, L., Wakefield, W.W., Simon, V., Barth, J.A. and Pierce, S.D. (2017) Species-specific responses of demersal fishes to near-bottom oxygen levels within the California Current large marine ecosystem. *Marine Ecology Progress Series* 568, 151–173. doi:10.3354/meps12066.
- Klein, E.S., Smith, S.L. and Kritzer, J.P. (2017) Effects of climate change on four New England groundfish species. *Reviews in Fish Biology and Fisheries* 27, 317–338. doi:10.1007/s11160-016-9444-z.
- Kristensen, K., Nielsen, A., Berg, C.W., Skaug, H. and Bell, B.M. (2016) TMB: Automatic Differentiation and Laplace Approximation. *J. Stat. Softw.* **70**, 1–21. doi:10.18637/jss.vo70.io5.
- Kumagai, N.H., Molinos, J.G., Yamano, H., Takao, S., Fujii, M. and Yamanaka, Y. (2018) Ocean currents and herbivory drive macroalgae-to-coral community shift under climate warming. *PNAS* 115, 8990–8995. doi:10.1073/pnas.1716826115.
- Latimer, A.M., Banerjee, S., Sang Jr, H., Mosher, E.S. and Silander Jr, J.A. (2009) Hierarchical models facilitate spatial analysis of large data sets: a case study on invasive plant species in the northeastern United States. *Ecology Letters* 12, 144–154. doi:10.1111/j.1461-0248.2008.01270.x.
- Laurel, B.J. and Rogers, L.A. (2020) Loss of spawning habitat and prerecruits of Pacific cod during a Gulf of Alaska heatwave. Canadian Journal of Fisheries and Aquatic Sciences 77, 644-650.
 doi:10.1139/cjfas-2019-0238.
- Laurel, B.J., Stoner, A.W. and Hurst, T.P. (2007) Density-dependent habitat selection in marine flatfish: the dynamic role of ontogeny and temperature. *Marine Ecology Progress Series* 338, 183–192. doi:10.3354/meps338183.
- Lenoir, J., Bertrand, R., Comte, L., Bourgeaud, L., Hattab, T., Murienne, J. and Grenouillet, G. (2020)

 Species better track climate warming in the oceans than on land. *Nature Ecology and Evolution*4, 1044–1059. doi:10.1038/s41559-020-1198-2.
- Levin, L.A. and Le Bris, N. (2015) The deep ocean under climate change. *Science* **350**, 766–766 768. doi:10.1126/science.aado126. URL https://www.sciencemag.org/lookup/doi/10.1126/science.aad0126.

- Li, L., Hollowed, A.B., Cokelet, E.D., Barbeaux, S.J. *et al.* (2019) Subregional differences in ground-fish distributional responses to anomalous ocean bottom temperatures in the northeast Pacific. *Global Change Biology* 25, 2560–2575. doi:10.1111/gcb.14676.
- Lindgren, F., Rue, H. and Lindström, J. (2011) An explicit link between Gaussian fields and Gaussian

 Markov random fields: The stochastic partial differential equation approach. J. R. Stat. Soc. Ser.

 B Stat. Methodol. 73, 423–498. doi:10.1111/j.1467-9868.2011.00777.x.
- Loarie, S.R., Duffy, P.B., Hamilton, H., Asner, G.P., Field, C.B. and Ackerly, D.D. (2009) The velocity of climate change. *Nature* 462, 1052–1055. doi:10.1038/nature08649.
- Loher, T. (2011) Analysis of match-mismatch between commercial fishing periods and spawning ecology of Pacific halibut (*Hippoglossus Stenolepis*), based on winter surveys and behavioural data from electronic archival tags. *ICES Journal of Marine Science* 68, 2240–2251. doi:10.1093/icesjms/fsr152.
- Love, M.S. (2011) Certainly more than you want to know about the fishes of the Pacific Coast: a postmodern experience. Really Big Press.
- Love, M., Yoklavich, M. and Thorsteinson, L. (2002) *The Rockfishes of the Northeast Pacific*. University of California Press.
- MacCall, A.D. (1990) Dynamic geography of marine fish populations. Washington Sea Grant Program Seattle, WA.
- Madeira, C., Mendonça, V., Leal, M.C., Flores, A.A., Cabral, H.N., Diniz, M.S. and Vinagre, C. (2017)
 Thermal stress, thermal safety margins and acclimation capacity in tropical shallow waters—An
 experimental approach testing multiple end-points in two common fish. *Ecological Indicators* 146 158. doi:https://doi.org/10.1016/j.ecolind.2017.05.050.
- Massiot-Granier, F., Lassalle, G., Almeida, P.R., Aprahamian, M. et al. (2018) A generic method to assess species exploratory potential under climate change. Ecological Indicators 90, 615–623.
 doi:10.1016/j.ecolind.2018.03.047.
- Masson, D. and Fine, I. (2012) Modeling seasonal to interannual ocean variability of coastal British

 Columbia. *Journal of Geophysical Research: Oceans* 117. doi:https://doi.org/10.1029/2012JC008151.

- Maureaud, A.A., Frelat, R., Pécuchet, L., Shackell, N. *et al.* (2021) Are we ready to track climatedriven shifts in marine species across international boundaries? A global survey of scientific bottom trawl data. *Global Change Biology* 27, 220–236. doi:https://doi.org/10.1111/gcb.15404.
- Mindel, B.L., Webb, T.J., Neat, F.C. and Blanchard, J.L. (2016) A trait-based metric sheds new light on the nature of the body size-depth relationship in the deep sea. *Journal of Animal Ecology* 85,
 427–436. doi:10.1111/1365-2656.12471.
- Molinos, J.G., Schoeman, D.S., Brown, C.J. and Burrows, M.T. (2019) VoCC: An R package for calculating the velocity of climate change and related climatic metrics. *Methods in Ecology and Evolution* 10, 2195–2202. doi:10.1111/2041-210X.13295.
- Morley, J.W., Selden, R.L., Latour, R.J., Frölicher, T.L., Seagraves, R.J. and Pinsky, M.L. (2018) Projecting shifts in thermal habitat for 686 species on the North American continental shelf. *PLOS* ONE 13, e0196127. doi:10.1371/journal.pone.0196127.
- Nye, J.A., Link, J.S., Hare, J.A. and Overholtz, W.J. (2009) Changing spatial distribution of fish stocks in relation to climate and population size on the Northeast United States continental shelf. *Marine Ecology Progress Series* 393, 111–129. doi:10.3354/mepso8220.
- Oestreich, W.K., Chapman, M.S. and Crowder, L.B. (2020) A comparative analysis of dynamic management in marine and terrestrial systems. *Frontiers in Ecology and the Environment* 18, 496–504. doi:https://doi.org/10.1002/fee.2243.
- Okey, T.A., Alidina, H.M., Lo, V. and Jessen, S. (2014) Effects of climate change on Canada's Pacific marine ecosystems: a summary of scientific knowledge. *Reviews in Fish Biology and Fisheries* 24, 519–559. doi:10.1007/s11160-014-9342-1.
- Oldfather, M.F., Kling, M.M., Sheth, S.N., Emery, N.C. and Ackerly, D.D. (2020) Range edges in heterogeneous landscapes: Integrating geographic scale and climate complexity into range dynamics. *Global Change Biology* 26, 1055–1067. doi:10.1111/gcb.14897.
- Ordonez, A. and Williams, J.W. (2013) Projected climate reshuffling based on multivariate climate-availability, climate-analog, and climate-velocity analyses: Implications for community disaggregation. *Climatic Change* 119, 659–675. doi:10.1007/s10584-013-0752-1.

- Parmesan, C. and Yohe, G. (2003) A globally coherent fingerprint of climate change impacts across natural systems. *Nature* 421, 37–42. doi:10.1038/nature01286.
- Perry, A.L., Low, P.J., Ellis, J.R. and Reynolds, J.D. (2005) Climate change and distribution shifts in marine fishes. *Science* 308, 1912–1915. doi:10.1126/science.1111322.
- Peterson, W., Robert, M. and Bond, N. (2015) The warm blob-conditions in the northeastern Pacific Ocean. *PICES Press* 23, 36.
- Peña, M.A., Fine, I. and Callendar, W. (2019) Interannual variability in primary production and shelf-offshore transport of nutrients along the northeast Pacific Ocean margin. *Deep Sea Research Part II: Topical Studies in Oceanography* 169-170, 104637. doi:10.1016/j.dsr2.2019.104637.
- Pinsky, M.L., Rogers, L.A., Morley, J.W. and Frölicher, T.L. (2020) Ocean planning for species on the move provides substantial benefits and requires few trade-offs. *Science Advances* 6, eabb8428. doi:10.1126/sciadv.abb8428.
- Pinsky, M.L. and Fogarty, M. (2012) Lagged social-ecological responses to climate and range shifts in fisheries. *Climatic Change* 115, 883–891. doi:10.1007/s10584-012-0599-x.
- Pinsky, M.L., Worm, B., Fogarty, M.J., Sarmiento, J.L. and Levin, S.A. (2013) Marine taxa track local climate velocities. *Science* 341, 1239–1242. doi:10.1126/science.1239352.
- Plagányi, É.E., Weeks, S.J., Skewes, T.D., Gibbs, M.T. *et al.* (2011) Assessing the adequacy of current fisheries management under changing climate: A southern synopsis. *ICES Journal of Marine*Science 68, 1305–1317. doi:10.1093/icesjms/fsro49.
- Pörtner, H.O., Roberts, D.C., Masson-Delmotte, V., Zhai, P. et al. (2019) *IPCC special report on the*ocean and cryosphere in a changing climate. Cambridge University Press.
- Punt, A.E., A'mar, T., Bond, N.A., Butterworth, D.S. *et al.* (2014) Fisheries management under climate and environmental uncertainty: Control rules and performance simulation. *ICES Journal* of Marine Science 71, 2208–2220. doi:10.1093/icesjms/fsto57.
- Pörtner, H., Bock, C., Knust, R., Lannig, G., Lucassen, M., Mark, F. and Sartoris, F. (2008) Cod and climate in a latitudinal cline: physiological analyses of climate effects in marine fishes. *Clim.*Res. 37, 253-270. doi:10.3354/croo766.

- R Core Team (2019) *R: A Language and Environment for Statistical Computing.* R Foundation for Statistical Computing, Vienna, Austria.
- Rindorf, A. and Lewy, P. (2006) Warm, windy winters drive cod north and homing of spawners keeps them there. *Journal of Applied Ecology* 43, 445–453. doi:https://doi.org/10.1111/j.1365-2664.2006.01161.x.
- Rooper, C.N., Ortiz, I., Hermann, A.J., Laman, N., Cheng, W., Kearney, K. and Aydin, K. (2020)
 Predicted shifts of groundfish distribution in the Eastern Bering Sea under climate change, with
 implications for fish populations and fisheries management. *ICES Journal of Marine Science* doi:10.1093/icesjms/fsaa215.
- Royer, T.C. (1998) Coastal processes in the northern North Pacific. *The Global Coastal Ocean : Regional Studies and Synthesis* Publisher: John Wiley.
- Rue, H., Martino, S. and Chopin, N. (2009) Approximate Bayesian inference for latent Gaussian models by using integrated nested Laplace approximations. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)* 71, 319–392. doi:10.1111/j.1467-9868.2008.00700.x.
- Serra-Diaz, J.M., Franklin, J., Ninyerola, M., Davis, F.W., Syphard, A.D., Regan, H.M. and Ikegami,
 M. (2014) Bioclimatic velocity: The pace of species exposure to climate change. *Diversity and Distributions* 20, 169–180. doi:10.1111/ddi.12131.
- Shackell, N.L., Frank, K.T., Fisher, J.A.D., Petrie, B. and Leggett, W.C. (2010) Decline in top predator body size and changing climate alter trophic structure in an oceanic ecosystem. *Proceedings of the Royal Society B: Biological Sciences* 277, 1353–1360. doi:10.1098/rspb.2009.1020.
- Shelton, A.O., Thorson, J.T., Ward, E.J. and Feist, B.E. (2014) Spatial semiparametric models improve estimates of species abundance and distribution. *Canadian Journal of Fisheries and Aquatic Sciences* 71, 1655–1666. doi:10.1139/cjfas-2013-0508.
- Sinclair, A., Schnute, J., Haigh, R., Starr, P., Rick Stanley, Jeff Fargo and Workman, G. (2003) Feasibility of multispecies groundfish bottom trawl surveys on the BC coast. DFO Canadian Science
 Advisory Secretariat (CSAS) Research Document 2003/049.

- Stock, C.A., Alexander, M.A., Bond, N.A., Brander, K.M. *et al.* (2011) On the use of IPCC-class models to assess the impact of climate on Living Marine Resources. *Progress in Oceanography* 88, 1–27. doi:10.1016/j.pocean.2010.09.001.
- Sunday, J.M., Pecl, G.T., Frusher, S., Hobday, A.J. *et al.* (2015) Species traits and climate velocity explain geographic range shifts in an ocean-warming hotspot. *Ecology Letters* **18**, 944–9530. doi:10.1111/ele.12474.
- Szuwalski, C.S. and Hollowed, A.B. (2016) Climate change and non-stationary population processes in fisheries management. *ICES Journal of Marine Science* 73, 1297–1305. doi:10.1093/icesjms/fsv229.
- Thorson, J.T. and Barnett, L.A.K. (2017) Comparing estimates of abundance trends and distribution shifts using single- and multispecies models of fishes and biogenic habitat. *ICES Journal of Marine Science* 74, 1311–1321. doi:10.1093/icesjms/fsw193.
- Thorson, J.T., Fonner, R., Haltuch, M.A., Ono, K. and Winker, H. (2017) Accounting for spatiotemporal variation and fisher targeting when estimating abundance from multispecies fishery data. *Canadian Journal of Fisheries and Aquatic Sciences* 74, 1794–1807. doi:10.1139/cjfas-2015-0598.
- Thorson, J.T., Pinsky, M.L. and Ward, E.J. (2016a) Model-based inference for estimating shifts in species distribution, area occupied and centre of gravity. *Methods in Ecology and Evolution* 7, 990–1002. doi:10.1111/2041-210X.12567.
- Thorson, J.T., Rindorf, A., Gao, J., Hanselman, D.H. and Winker, H. (2016b) Density-dependent changes in effective area occupied for sea-bottom-associated marine fishes. *Proceedings of the Royal Society B: Biological Sciences* **283**, 20161853. doi:10.1098/rspb.2016.1853.
- Thorson, J.T., Scheuerell, M.D., Shelton, A.O., See, K.E., Skaug, H.J. and Kristensen, K. (2015a) Spatial factor analysis: a new tool for estimating joint species distributions and correlations in species range. *Methods in Ecology and Evolution* 6, 627–637. doi:https://doi.org/10.1111/2041-210X.12359.

- Thorson, J.T., Shelton, A.O., Ward, E.J. and Skaug, H.J. (2015b) Geostatistical delta-generalized linear mixed models improve precision for estimated abundance indices for West Coast ground-fishes. *ICES Journal of Marine Science* 72, 1297–1310. doi:10.1093/icesjms/fsu243.
- Tommasi, D., Stock, C.A., Hobday, A.J., Methot, R. *et al.* (2017) Managing living marine resources in a dynamic environment: The role of seasonal to decadal climate forecasts. *Progress in Oceanog-raphy* **152**, 15–49. doi:10.1016/j.pocean.2016.12.011.
- Turris, B.R. (2000) A comparison of British Columbia's ITQ fisheries for groundfish trawl and sablefish: Similar results from programmes with differing objectives, designs and processes.
 FAO Fisheries Technical Paper, 254–261.
- Tweedie, M.C.K. (1984) An index which distinguishes between some important exponential families. In: Statistics: Applications and New Directions. Proceedings of the Indian Statistical Institute Golden Jubilee International Conference (eds. J.K. Gosh and J. Roy). Indian Statistical Institute,

 Calcutta, pp. 579–604.
- VanDerWal, J., Murphy, H.T., Kutt, A.S., Perkins, G.C., Bateman, B.L., Perry, J.J. and Reside, A.E. (2013) Focus on poleward shifts in species' distribution underestimates the fingerprint of climate change. *Nature Climate Change* 3, 239–243. doi:10.1038/nclimate1688.
- Wallace, S., Turris, B., Driscoll, J., Bodtker, K., Mose, B. and Munro, G. (2015) Canada's Pacific groundfish trawl habitat agreement: A global first in an ecosystem approach to bottom trawl
 impacts. *Marine Policy* 60, 240–248. doi:10.1016/j.marpol.2015.06.028.
- Ward, E.J., Jannot, J.E., Lee, Y.W., Ono, K., Shelton, A.O. and Thorson, J.T. (2015) Using spatiotemporal species distribution models to identify temporally evolving hotspots of species cooccurrence. *Ecological Applications* 25, 2198–2209. doi:10.1890/15-0051.1.
- Webster, R.A., Soderlund, E., Dykstra, C.L. and Stewart, I.J. (2020) Monitoring change in a dynamic environment: spatiotemporal modelling of calibrated data from different types of fisheries surveys of pacific halibut. Canadian Journal of Fisheries and Aquatic Sciences 77, 1421–1432. doi:10.1139/cjfas-2019-0240.

- Williams, D.C., Nottingham, M.K., Olsen, N. and Wyeth, M.R. (2018) Summary of the Queen Charlotte Sound synoptic bottom trawl survey, July 6 August 8, 2015. DFO Can. Manuscr. Rep. Fish.
 Aquat. Sci. 3136, viii + 64 p.
- Yang, M.S., Dodd, K., Hibpshman, R. and Whitehouse, A. (2006) Food habits of groundfishes in the

 Gulf of Alaska in 1999 and 2001. *NOAA Technical Memorandum*, 13.

Tables

Table 1: Gradient-based velocity metrics and their definitions. Climatic variables are temperature and dissolved oxygen (DO); biotic variables are biomass density for the mature and immature components of a species. Climatic and biotic variables are represented generically by A. Our analysis treated gradient-based velocity as a scalar, using only the magnitude component of the velocity vector.

Term	Notation	Definition	Magnitude	Sign		
Local trend	$m_s^A = \frac{\Delta A_s}{\Delta t}$	Change in local biotic or climatic scalar <i>A</i> per decade	Temporal rate of change in <i>A</i>	Increasing (+) or decreasing (–) local trend in <i>A</i>		
Spatial gradient	$ec{g}_s^A$	Vector sum (magnitude, angle) of mean north or south and east or west gradients of A in a 3×3 cell spatial neighbourhood	Spatial rate of change in <i>A</i>	Vector magnitude and angle always positive (+)		
Gradient-based velocity	$\vec{V}_s^A = \frac{m_s^A}{\vec{g}_s^A}$	Vector velocity (magnitude, angle) from local trend of <i>A</i> divided by vector local spatial gradient of <i>A</i>	Speed of travel predicted to maintain initial <i>A</i>	Increasing (+) or decreasing (–) based on the local trend in <i>A</i>		

Figures

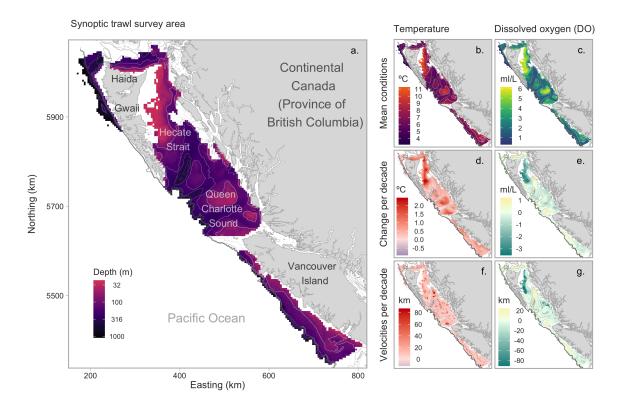
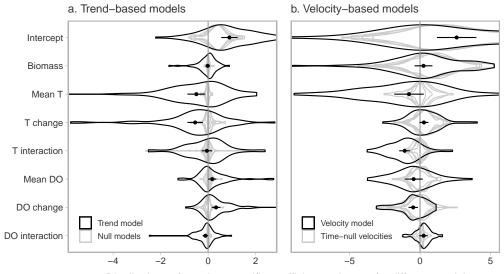


Figure 1: Maps of study area (a), predicted mean conditions (b, c), decadal trends (d, e), and decadal velocities (f, g) of bottom temperature and dissolved oxygen (DO) for 2008–2018 off the coast of British Columbia, Canada (UTM zone 9). Values are estimated using geostatistical spatiotemporal models of CTD sensor data collected during late-spring/early-summer groundfish trawl surveys. Bathymetry lines at every 100 m are overlaid in shades of grey that increase with depth.



Distributions of species-specific coefficient estimates for different models

Figure 2: Distribution of species-specific (random effect) coefficients from the model fitted to observed data (black violins) compared with coefficients from four simulated null models (gray violins). Each "violin" is based on a single model including all species: (a) trend-based models where climate and biotic change variables are all local trends and (b) velocity models where climate and biotic change variables are all velocities. Black points with ranges represent the observed-data global (fixed effect) coefficient estimates with 95% CIs. Null models used fitted covariate values, but simulated response data. Simulated time-null velocities used these same simulated trends divided by the real spatial gradients. The x-axes have been truncated slightly for interpretability.

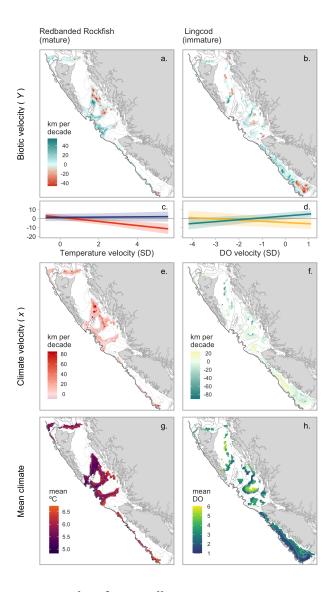


Figure 3: Maps and interaction plots for two illustrative species representing the most frequent relationship with temperature (left column) and the expected response with DO (right column). Mature Redbanded Rockfish had stable, near-zero, biotic velocities regardless of amount of warming in the coolest regions it occupied, and decreases in biomass when temperatures increased in warmer regions (a,c,e,g). Immature Lingcod biotic velocities increased with positive DO velocities in low mean DO locations only (b,d,f,h). In panel c, a blue line represents predicted biotic velocity (Y, y-axis) for different temperature velocities (x, x-axis) in the coolest locations (0.025 quantile of those occupied by 95% of the estimated biomass of each species) and a red line represents the same for the warmest locations (0.975 quantile). Likewise, for predictions at different DO velocities, green represents the lower quantile of mean DO and yellow the higher (d). Both the colours and slopes illustrated correspond with those in Figure 4. The maps include biotic velocity estimates for all locations that cumulatively account for 95% of the estimated biomass of each species (a, b), and the same climate estimates as in Figure 1, but trimmed to include only the values for the same locations as the biotic velocities for each species predicted relationships.

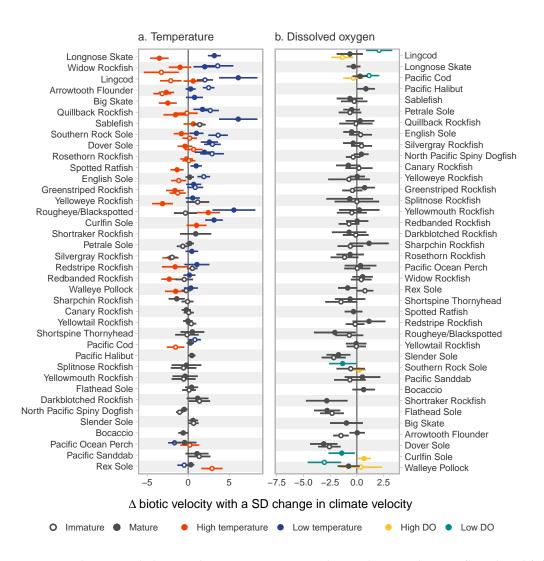


Figure 4: Mean climate and climate change interact in predicting biotic velocities (km/decade) for 38 groundfish species. Coloured dot-whiskers indicate slopes and 95% CIs of the predicted biotic velocities with 1 SD of change in climate velocity for the low and high 95% quantiles of mean local climate (i.e., the slopes of lines in interaction plots like those in Figure 3c, d). Species are ordered by the difference between the slopes at the highest and lowest quantiles of mean climate such that the more intuitive results are at the top: increases in climate velocity have a more positive impact on biotic velocity when starting at a low mean temperature (a) or DO level (b). Open circles indicate patterns for immature fish and closed circles represent individuals large enough to have a 50% chance of having reached reproductive maturity, or belonging to species for which maturity data was not available. Black dot-whiskers represent the slopes for each maturity class when the interaction is not significant. See Table S1 for scientific names.

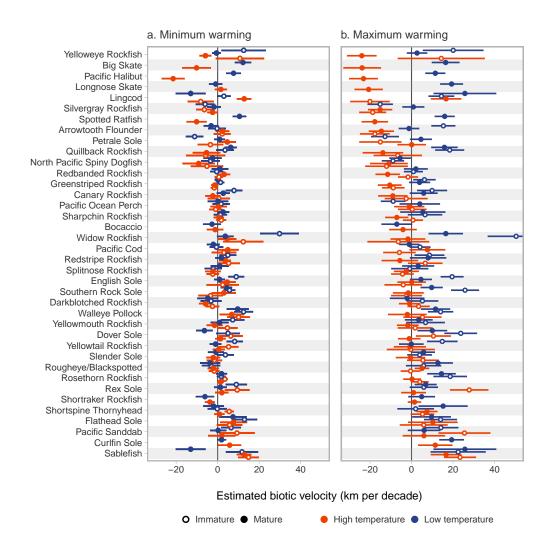


Figure 5: Estimates of biotic velocities (km/decade with 95% CI) for 38 groundfish species under different mean temperatures (blue and red represent low and high 95% quantiles) occupied and at (a) the minimum and (b) maximum temperature velocities experienced for each species. Open circles indicate patterns for immature fish and closed circles represent individuals large enough to have a 50% chance of being reproductively mature, or belonging to species for which maturity data were not available. Species are ordered by the minimum estimates at the maximum climate velocity experienced for each species. Therefore, species most likely to experience population declines with increasing temperatures are at the top.

Supporting Information

Methods

S1. Maturity and length-weight models

When maturity data were collected for a species in all years for all surveys (> 250 samples across all years and > 40 each year), we included a random intercept for year to allow for temporal change in size at maturity:

$$M_i \sim \text{Bernoulli}(p_i),$$
 (1)

$$p_i = \text{logit}^{-1} \left(\alpha + \alpha_t + \beta L_i \right), \tag{2}$$

$$\alpha_t \sim \text{Normal}\left(0, \sigma_\alpha^2\right),$$
 (3)

where M_i represents the mature (1) or immature (0) status of fish i, p_i represents the probability of maturity of fish i, α represents a global intercept, α_t represents a year-specific deviation for year t that is allowed to vary with a variance of σ_{α}^2 , β represents a coefficient, and L_i represents the length of fish i. We used year-specific 50% maturity probability thresholds to split observed catches into maturity classes. For species not meeting the above thresholds, we used a random effect of tow instead of year, and split catches based on the global estimate.

For all catches exceeding the species-specific threshold count of individual fish on a survey, either a random subset of roughly 50 fish or all individuals were measured for both length and weight. We therefore filled in occasional missing weights as

$$\log(W_i) \sim \text{Student-t}(3, \log(a) + b \log(L_i), \sigma),$$
 (4)

with W_i and L_i representing the weight and length for fish i, a and b being the species-specific parameter estimates of the length-weight relationship $W_i = aL_i^b$, σ representing the observation error scale, and 3 representing a degrees of freedom parameter fixed to allow for outlying residuals.

S2. Alternative spatiotemporal models of fish biomass density

We tried including the proportion of muddy sediment and proportion of sediment with any rocks as covariates in biomass density models, but the values we used were likely not specific enough to the trawl path sampled, and therefore not helpful to our models. Certainly more spatially precise estimates would be possible with refined substrate variables, but likely irrelevant to this analysis given the resolution and uncertainty in our climate variables. Furthermore, the spatial random effects can account for spatial factors that are constant across time, like substrate type, while

spatiotemporal random effects account for factors that vary from year to year spatially such as bottom temperature, water circulation patterns, species interactions, and species movement.

S3. Velocity calculation considerations

An alternative to gradient-based velocity calculations is to search for analogous conditions in the second time period and calculate the shortest actual distance to an analogous cell. This method requires a threshold of change below which cells are deemed analogous which is often derived from historical variability in climate. We calculated analog-distance velocities that were qualitatively similar to gradient-based velocities by using twice the minimum standard deviation of each climate variable as our threshold and time periods defined by prior-to and post onset of the same marine heatwave. However, models including these values tended to have more difficulty converging, so we relied instead on the gradient-based values and the component values for all our analyses (Figs 1 and S13).

S4. Alternative model configurations for linking biotic changes with climate

We tested additional covariates that we chose not to include in the final model. We calculated average hours fished by commercial bottom trawl for each 4 × 4 km grid cell per year between 2008 and 2018, as well as the percent change in time spent fishing during this time (Figure S26). The log of mean fishing intensity, the trend in fishing intensity, and the interaction between the two were all included as covariates in a climate trend model, but none of these variables showed any more significant negative relationships than might be expect by chance (Figure S27). We tested for an overall effects of maturity (treating mature fish as the intercept) on the main effects of all climate variables (all two-way interactions), and on the interactions (model with 3-way interactions). For each maturity class of each species, the model estimated an independent random spatial field either with or without genus or family as taxonomic grouping factors (Figure S21); however, final models do not include any grouping factors because they did not change any of the model estimates qualitatively. Finally, we attempted to assess the importance of the gradient component in velocity estimates by adding temperature gradient to a model of biomass change in response to temperature trends, and including a three-way interaction between temperature trend, mean, and gradient. None of the important effects we focus on in the results (Figure S23) were changed qualitatively by any of these added variables.

S₅. How climate sensitivity varies with life history and ecology

We calculated weighted means and interquartile ranges of depths from maturity-specific biomass densities for all survey catches. Because climate conditions experienced vary with depth and a narrower species depth range might increase other ecological effects, we include these variables

as covariates. From the individual fish caught in these trawl surveys, we calculated the maximum recorded body mass of each species and average age (estimated from otoliths or fin clips when available) for each maturity class of each species Given that age data were available for only a subset of species, we first assessed the independent effects of mean population ageand occupied depth (mean and range) on species responses to climate (Figure S22). These models also included an interaction with whether the response slope estimated was at the high or low extreme of mean conditions, and a random intercept for species (to capture the non-independence of the estimates for high and low climate extremes). Finally, to reduce leverage of extreme slope estimates, these extreme values were collapsed down to be equal to the 0.005 and 0.995 quantiles of all slope estimates. We then tested for relationships between ecological groupings and slope estimates (at both the high or low extreme of mean conditions) across both maturity classes, while controlling for the mean depth occupied. These models also used collapsed slope estimates (see previous paragraph), included a fixed effect of the mean climate (high or low) that the response slope was estimated for, and a random intercepts for both species and maturity classes nested within species.

The species and life history stages also varied with respect to their trophic level, behaviour, and overarching distribution (Table S1). Species were classified into either zooplanktivores or higher trophic level feeders based on the majority of their diet components from available literature (Love et al. 2002, Yang et al. 2006, Bizzarro et al. 2017). Species were classified as solitary and demersal (flatfishes, Lingcod, Sablefish, skates and some rockfishes) or bentho-pelagic and schooling (cods and some rockfishes). Rockfishes were classified based on Love et al. (2002). British Columbia occupies a unique oceanographic position at the bifurcation of the North Pacific Current (Royer 1998, Masson and Fine 2012). As such, the distribution of some groundfish species is limited to the north of British Columbia (such as for many of the rockfish species). Although species can occur in Alaska, British Columbia demarcates a northern boundary of relatively high abundance. It was expected that some of the groundfish species in this analysis might be responsive to these larger scale species distribution patterns. Thus, each species was qualitatively categorized based on bottom trawl survey catches in British Columbia, Washington State and Alaska as either occurring in the middle (high catches of the species in all three areas), northern (lowered catches of the species in Southeast Alaska) or southern (lowered catches in Washington State) portions of the species range. There were only three species in the southern portion of their range (Walleye Pollock, Flathead Sole and Arrowtooth Flounder), so in the analysis these were combined with the species in the middle of their distribution.

Table S1: Ecological data for 38 species analyzed, ordered by frequency of occurrence on trawl survey sets between 2008-2018. Classification codes: N = near northern range limit, S = near to centre of range or southern limit, L = zooplanktivore, H = all higher trophic diets.

Common name	Scientific name	Family	Sets	Depth occupied (m) Immature Mature		Mean age (years)		Range limit	Trophic level		Foraging zone	Sociality		
				Mean	IQR	Mean	IQR	Imm.	Mat.		Imm.	Mat.		
Arrowtooth Flounder	Atheresthes stomias	Pleuronectidae	85%	166	69	168	73	3	10	S	L	Н	Demersal	Solitary
Rex Sole	Glyptocephalus zachirus	Pleuronectidae	82%	139	55	165	67	-	-	S	H	Н	Demersal	Solitary
Spotted Ratfish	Hydrolagus colliei	Chimaeridae	80%	-	-	97	69	-	-	N	-	Н	Demersal	Schooling
Dover Sole	Microstomus pacificus	Pleuronectidae	74%	167	65	219	145	7	14	S	H	Н	Demersal	Solitary
Sablefish	Anoplopoma fimbria	Anoplopomatidae	55%	232	145	295	220	2	14	S	H	Н	Demersal	Solitary
North Pacific Spiny Dogfish	Squalus suckleyi	Squalidae	54%	115	61	124	74	-	-	N	L	Н	Benthopelagic	Schooling
Pacific Ocean Perch	Sebastes alutus	Sebastidae	52%	239	56	265	58	6	24	S	L	L	Benthopelagic ⁺	Schooling ⁺
Pacific Cod	Gadus macrocephalus	Gadidae	52%	121	69	125	67	2	4	S	Н	Н	Benthopelagic	Schooling
Silvergray Rockfish	Sebastes brevispinis	Sebastidae	46%	181	51	198	60	11	25	N	L	L	Benthopelagic	Solitary
Walleye Pollock	Gadus chalcogrammus	Gadidae	46%	133	56	140	73	-	-	S	L	L	Benthopelagic	Schooling
Slender Sole	Lyopsetta exilis	Pleuronectidae	44%	156	50	157	52	_	-	N	Н	Н	Demersal	Solitary
English Sole	Parophrys vetulus	Pleuronectidae	44%	72	46	89	43	-	_	N	Н	Н	Demersal	Solitary
Redbanded Rockfish	Sebastes babcocki	Sebastidae	41%	257	70	230	62	10	28	S	L	L	Benthopelagic	Solitary
Petrale Sole	Eopsetta jordani	Pleuronectidae	41%	117	46	120	48	5	9	N	L	Н	Demersal	Solitary
Pacific Halibut*	Hippoglossus stenolepis	Pleuronectidae	40%	-	-	110	88	-	-	S	-	Н	Demersal	Solitary
Shortspine Thornyhead	Sebastolobus alascanus	Sebastidae	40%	329	91	352	102	-	_	S	Н	Н	Demersal	Solitary
Lingcod	Ophiodon elongatus	Hexagrammidae	34%	126	51	145	70	3	7	S	Н	Н	Demersal	Solitary
Longnose Skate*	Raja rhina	Rajidae	32%	-	-	222	131	-	-	S	-	Н	Demersal	Solitary
Flathead Sole	Hippoglossoides elassodon	Pleuronectidae	32%	133	45	131	45	-	-	S	Н	Н	Demersal	Solitary
Sharpchin Rockfish	Sebastes zacentrus	Sebastidae	29%	213	50	235	25	7	16	S	L	L	Benthopelagic	Schooling
Greenstriped Rockfish	Sebastes elongatus	Sebastidae	26%	182	45	173	40	9	26	N	L	L	Demersal	Solitary
Rougheye/Blackspotted**	Sebastes aleutianus**	Sebastidae	24%	356	50	371	60	16	36	S	Н	Н	Demersal	Solitary
Redstripe Rockfish	Sebastes proriger	Sebastidae	23%	134	65	182	50	5	15	S	L	L	Benthopelagic	Schooling
Southern Rock Sole	Lepidopsetta bilineata	Pleuronectidae	22%	46	32	50	33	4	8	S	Н	Н	Demersal	Solitary
Yellowtail Rockfish	Sebastes flavidus	Sebastidae	22%	116	43	148	36	6	15	N	L	L	Benthopelagic	Schooling
Rosethorn Rockfish	Sebastes helvomaculatus	Sebastidae	20%	228	65	249	69	-	-	N	Н	Н	Demersal	Solitary
Canary Rockfish	Sebastes pinniger	Sebastidae	19%	147	53	164	34	8	18	N	L	L	Benthopelagic	Schooling
Pacific Sanddab	Citharichthys sordidus	Paralichthyidae	18%	76	35	83	26	_	_	N	Н	Н	Demersal	Solitary
Yellowmouth Rockfish	Sebastes reedi	Sebastidae	15%	217	25	232	46	11	33	N	L	L	Benthopelagic	Schooling
Splitnose Rockfish	Sebastes diploproa	Sebastidae	14%	250	39	294	33	7	19	N	L	L	Benthopelagic	Schooling
Darkblotched Rockfish	Sebastes crameri	Sebastidae	11%	256	91	306	48	_	_	N	L	L	Demersal	Solitary
Curlfin Sole***	Pleuronichthys decurrens	Pleuronectidae	10%	-5-	-	58	28	_	_	N	-	H	Demersal	Solitary
Bocaccio***	Sebastes paucispinis	Sebastidae	10%	_	-	162	38	_	22	N	_	Н	Benthopelagic	Solitary
Ouillback Rockfish	Sebastes maliger	Sebastidae	10%	65	21	76	37	6	20	N	L	Н	Demersal	Solitary
Yelloweye Rockfish	Sebastes ruberrimus	Sebastidae	9%	149	39	156	36	-	_	S	L	Н	Demersal	Solitary
Big Skate*	Beringraja binoculata	Rajidae	9%		-	-5° 75	58	_	_	S	-	Н	Demersal	Solitary
Widow Rockfish	Sebastes entomelas	Sebastidae	9%	74	82	180	141	2	21	N	L	L	Benthopelagic	Schooling
Shortraker Rockfish***	Sebastes borealis	Sebastidae	5%	-	_	393	153	_	63	S	-	H	Demersal	Solitary

⁺Immature individuals use the alternate strategy

^{*}Species for which maturity data not collected

^{**}Complex that includes Rougheye Rockfish Sebastes aleutianus and a formerly unrecognized species, Blackspotted Rockfish Sebastes melanostictus

^{***}Species not captured frequently enough when immature to fit the spatial model

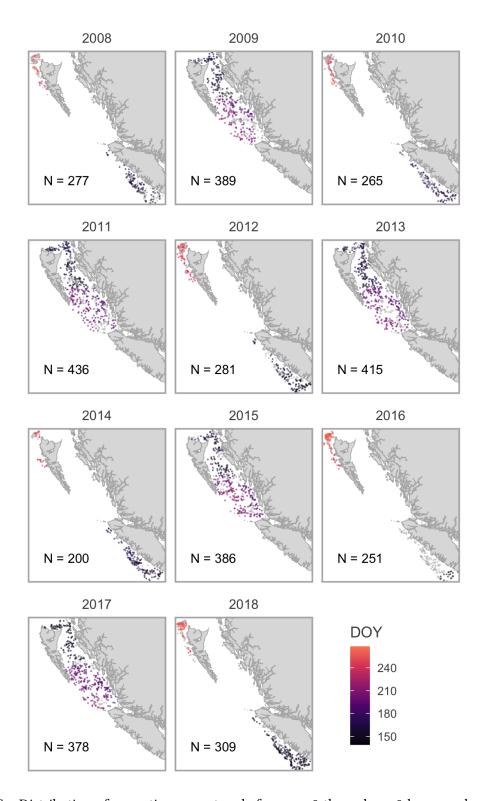


Figure S1: Distribution of synoptic survey trawls from 2008 through 2018 by year, day of year (DOY), and in space (UTM coordinates correspond to those in Figure 1). N is the total trawls analyzed for each year. Grey points are trawls that lack DO sensor data.

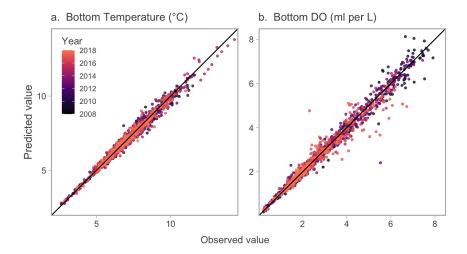


Figure S2: Predictions from models of bottom temperature (a) and DO (b) vs. observed sensor measurements on the trawl surveys.

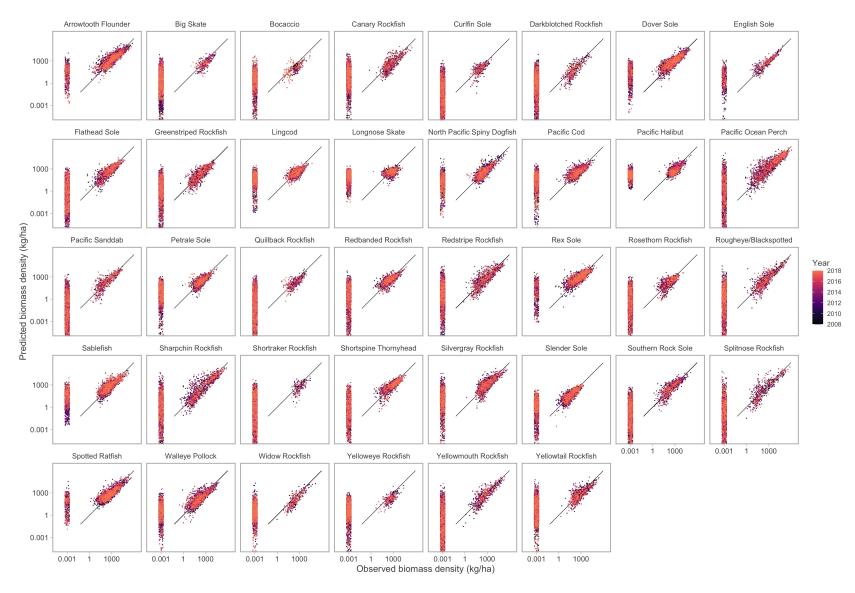


Figure S₃: Predicted vs. observed mature biomass densities. Observed absences have been assigned 0.001 kg/ha and given some horizontal random jitter to improve visibility of data points and the 1:1 line is truncated to not intersect these adjusted values.

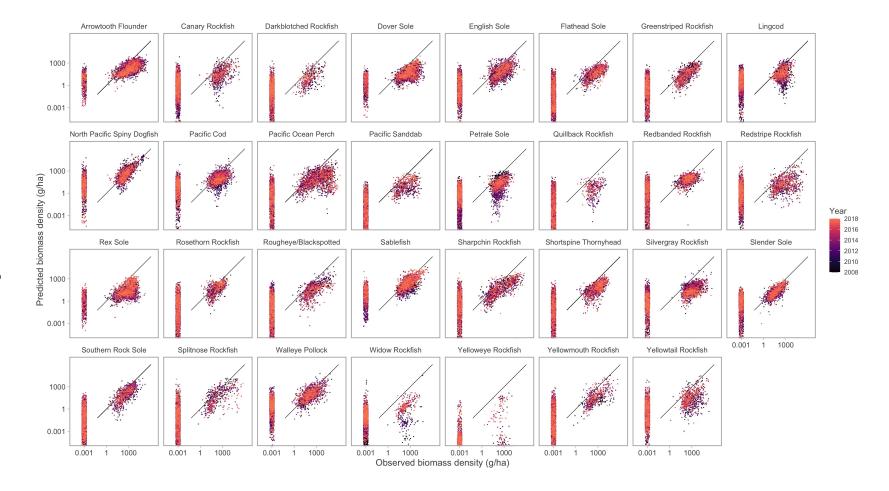


Figure S₄: Predicted vs. observed **immature** biomass densities. Same treatment as Figure S₃ except that units are g per hectare instead of kg per hectare. Relatively few positive samples for Widow and Yelloweye Rockfish result in relatively poor model fit to the positive sample component.



Figure S₅: Species-specific coefficient estimates from one example of a simulated trends model (null #1) ordered by the estimated effect of temperature trend.

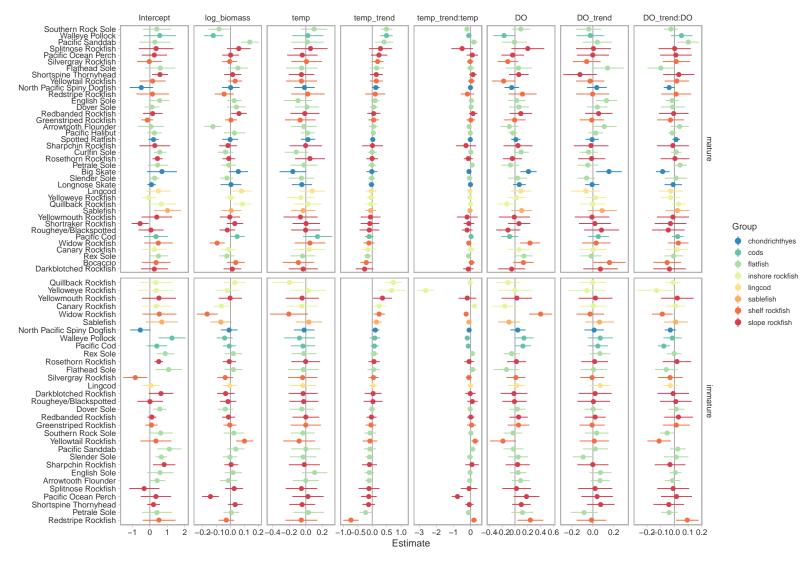


Figure S6: Species-specific coefficient estimates from one example of a simulated trends model (null #2) ordered by the estimated effect of temperature trend.

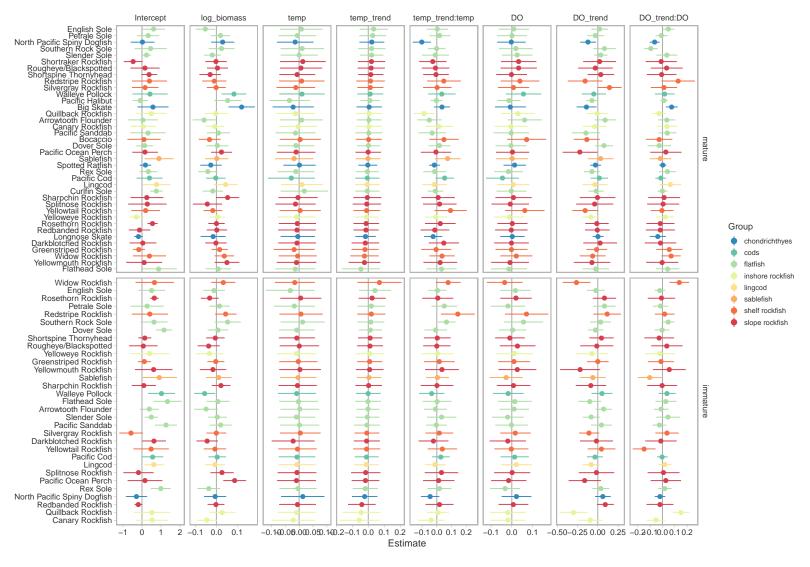


Figure S7: Species-specific coefficient estimates from one example of a simulated trends model (null #3) ordered by the estimated effect of temperature trend.

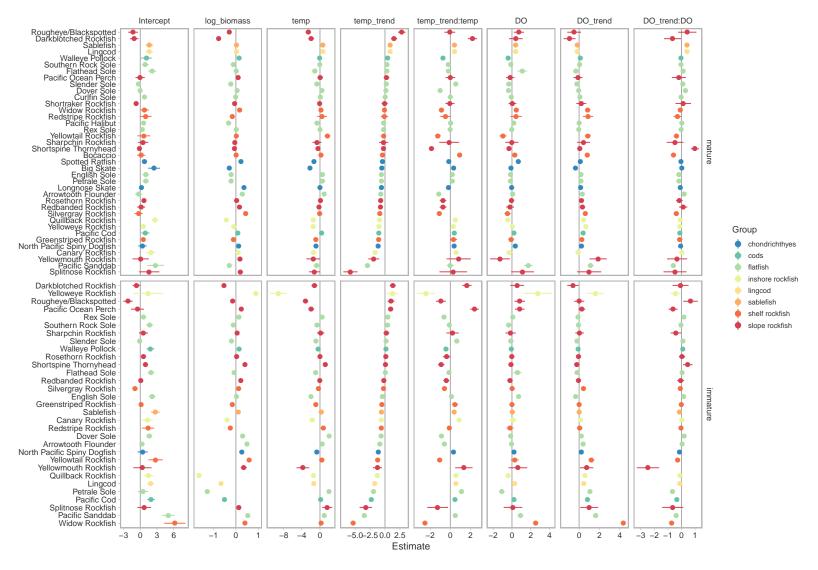


Figure S8: Species-specific coefficient estimates from trend model ordered by the estimated effect of temperature trend for a cell with an average biomass density and climate conditions and with no change in DO.

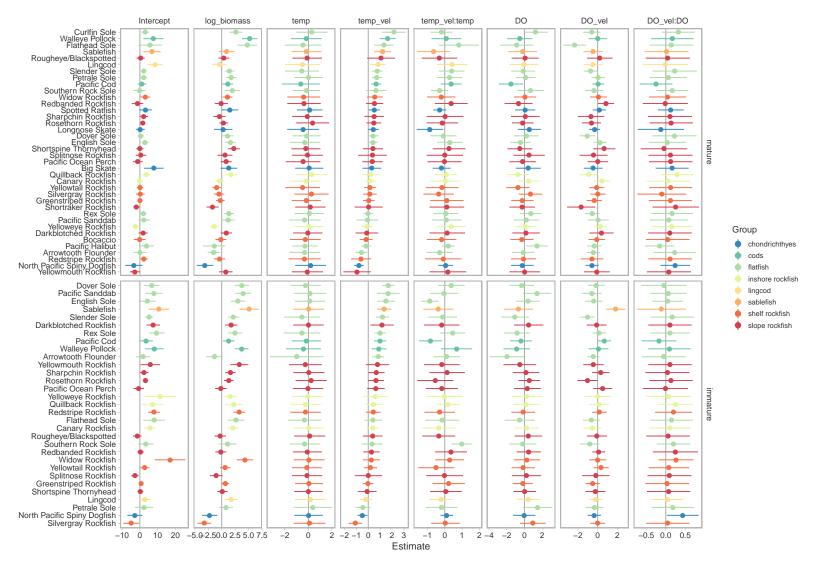


Figure S9: Species-specific coefficient estimates from one example of a simulated velocity model (null #1) ordered by the estimated effect of temperature velocity. The simulated biotic velocities used for this model were derived from simulated trends and true spatial gradients.

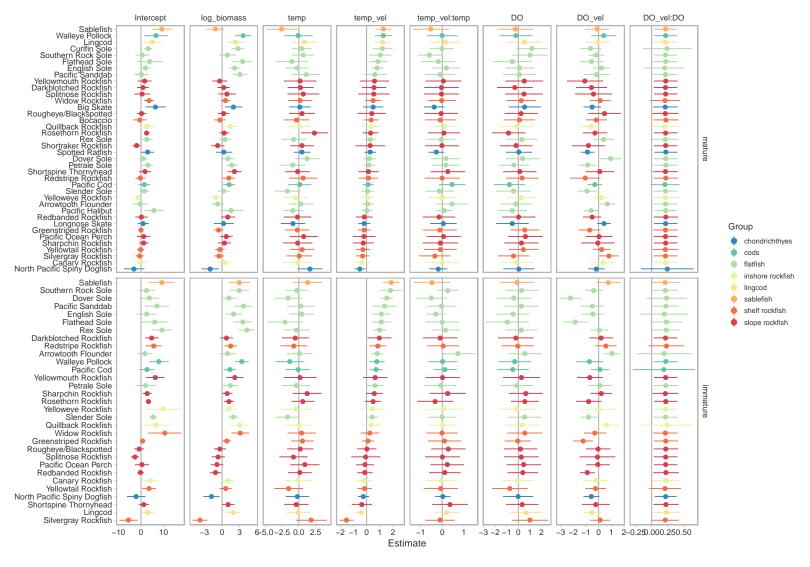


Figure S10: Species-specific coefficient estimates from one example of a simulated velocity model (null #2) ordered by the estimated effect of temperature velocity. The simulated biotic velocities used for this model were derived from simulated trends and true spatial gradients.

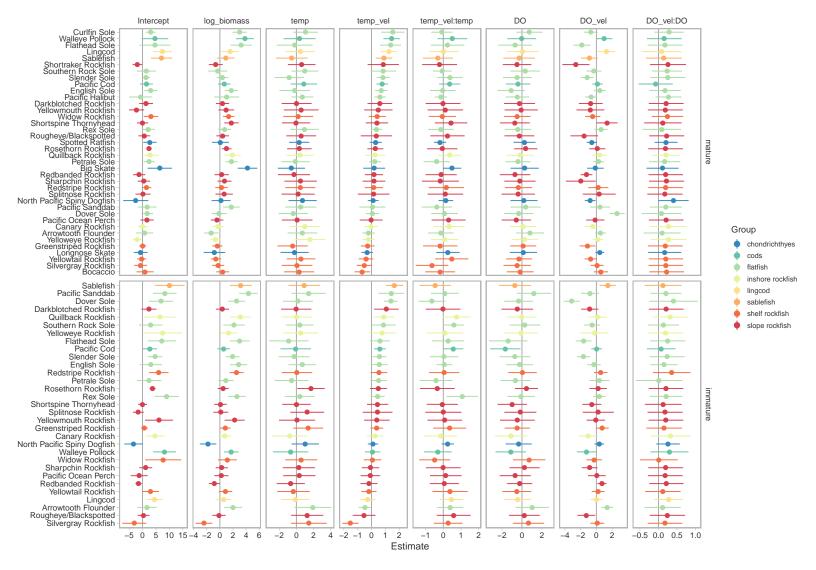


Figure S11: Species-specific coefficient estimates from one example of a simulated velocity model (null #3) ordered by the estimated effect of temperature velocity. The simulated biotic velocities used for this model were derived from simulated trends and true spatial gradients.

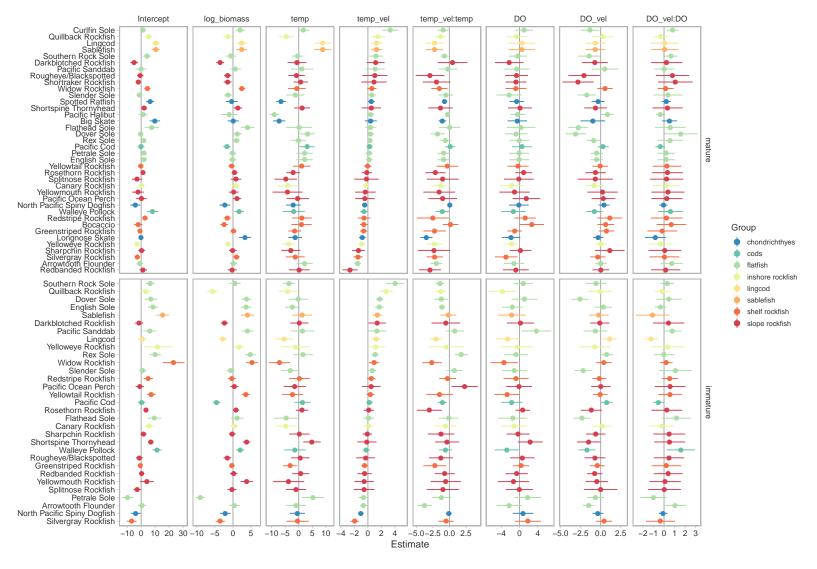


Figure S12: Species-specific coefficient estimates from velocity model ordered by the estimated effect of temperature velocity, which represents the change in the velocity of biomass change for a cell with an average biomass density and climate conditions and with no change in DO.

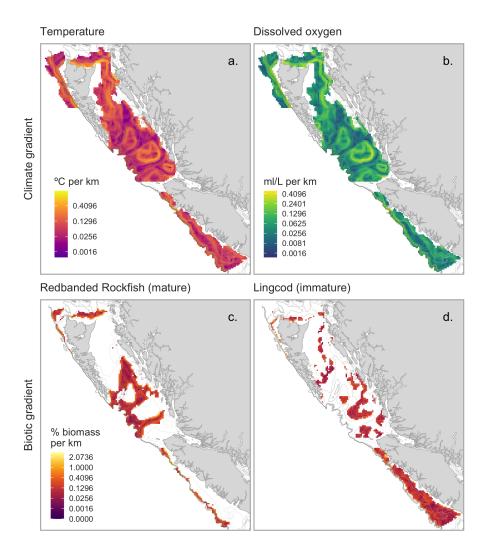


Figure S13: Maps of the gradient component of velocity calculations for both climate and biomass. Biotic gradient panels are for example species.

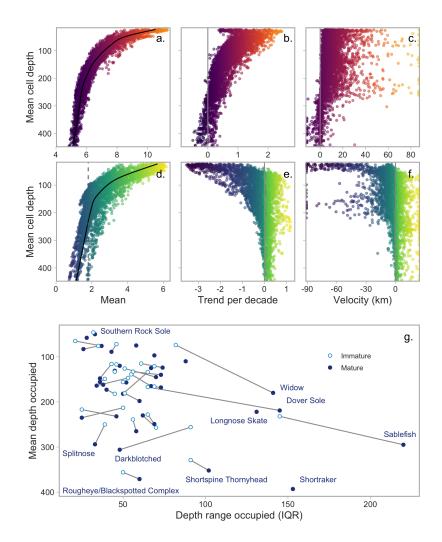


Figure S14: The climate conditions of each cell across depths (y-axes of upper panels) are indicated by both colours and the values on the x-axes of the 6 upper plots. Mean depth occupied by groundfish species co-varies with interquartile depth range occupied (g) and many of these species move deeper (lower on y-axis) with maturity (classes linked by grey lines) often with a corresponding increase in depth range (to right on x-axis).

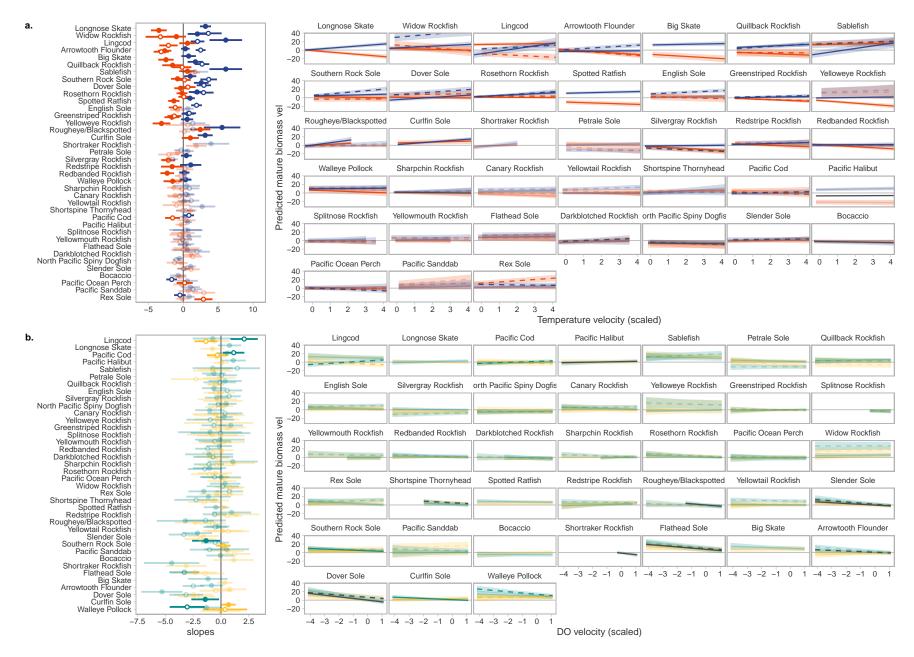


Figure S15: Slopes of predicted biotic **velocity** based on interaction between (a) seafloor temperature **velocity** and the mean temperature, or (b) seafloor DO **velocity** and mean DO levels. Species are ordered by the difference between their slopes at the highest and lowest quantiles of mean climate. Colours and symbols are the same as in Figure 4, except that now paler shades indicate that the difference between slopes was not significant and black lines represent these cases where the slope overall slope was still significant.

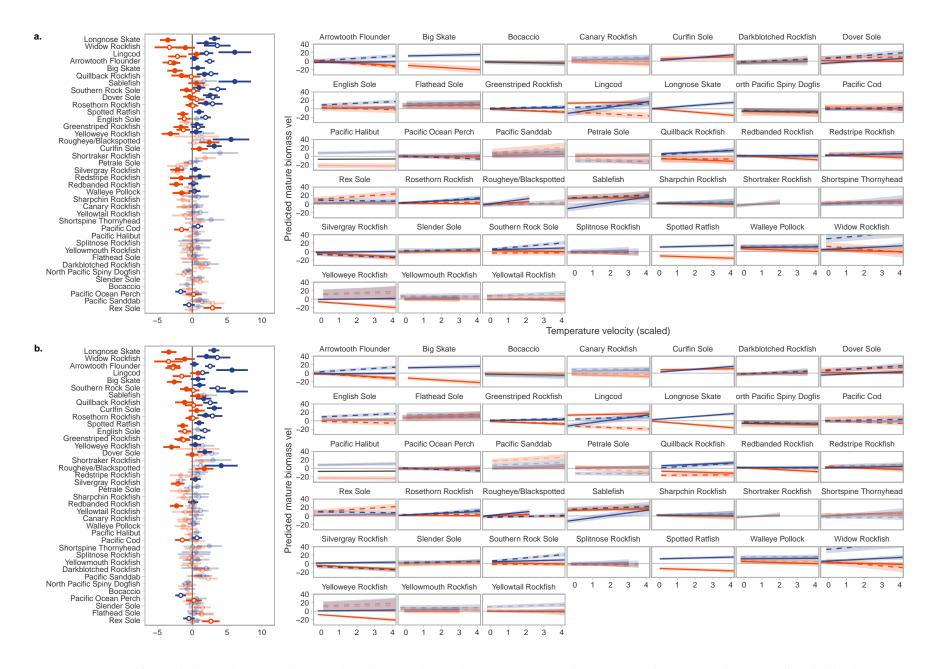


Figure S16: Here, (a) we duplicate the same slopes and predicted relationships as in Figure 4, but this time the species plots are ordered alphabetically for ease of comparison with (b) results from a similar model that lacks all of the DO variables. Differences in the ordering of the species in the left hand panels show that the precise values do change slightly, however the patterns and significance of effects are not changed by including DO in the model.



Figure S17: Biomass trends for mature fish populations (top and bottom 5th quantiles are not included in the colour scale, but are coloured by the nearest scale value).

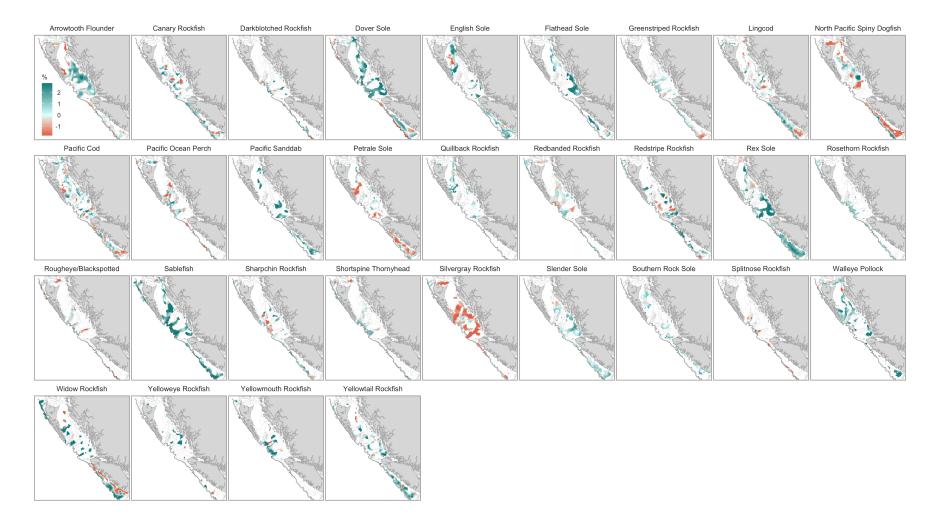


Figure S18: Biomass trends for immature fish populations on same scale as Figure S17.



Figure S19: Biotic velocities for mature fish populations.

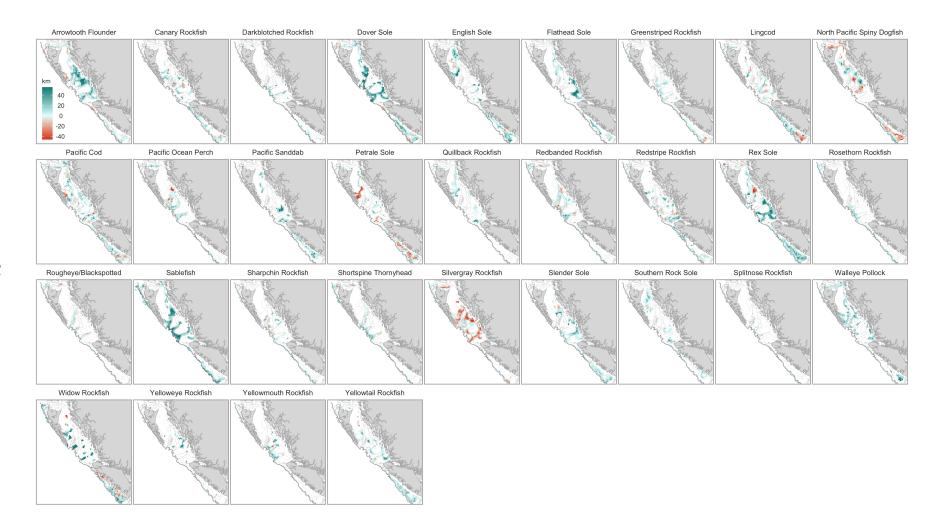


Figure S20: Biotic velocities for immature fish populations.

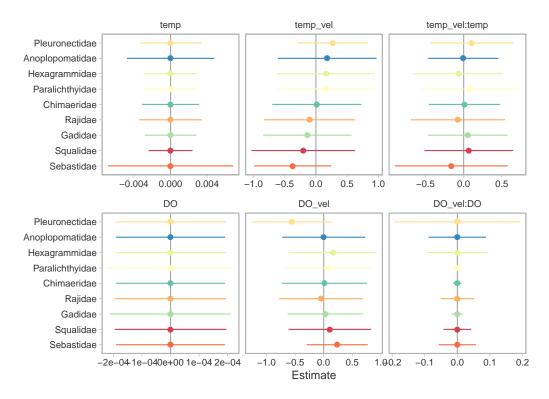


Figure S21: Family-level coefficient estimates for climate variables all have confidence intervals overlapping zero. Families are ordered by decreasing biotic velocity with increasing temperature velocity. This model includes the same climate variables as the "Velocity" models in Figures 2b and S23.

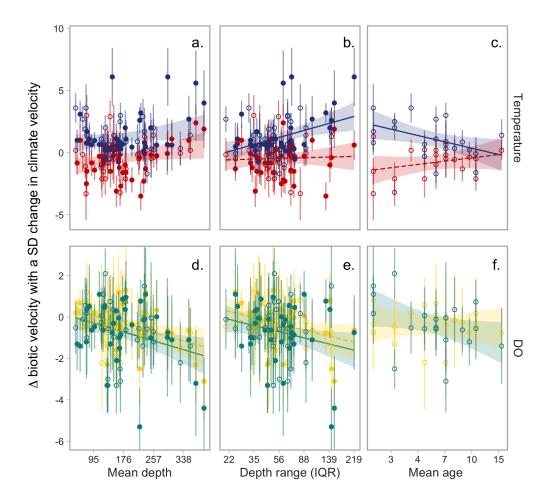


Figure S22: Occupied range of depths and immature population age correlate with the effect of warming temperature velocities on biotic velocities at low temperatures (a-c) and/or DO levels (e). Mean depth occupied only correlated with effects of DO (d). Each panel illustrates a separate model with a random effect of species and a fixed effect of mean local climate as a high or low 95th quantile for temperature or DO levels (colours and shapes match Figure 4). Points with ranges represent each species' raw slope estimates from the spatial models and their CI. Regression lines indicate significant relationships and corresponding uncertainties are based only on fixed-effects, and do not account for uncertainty in the slope estimates.

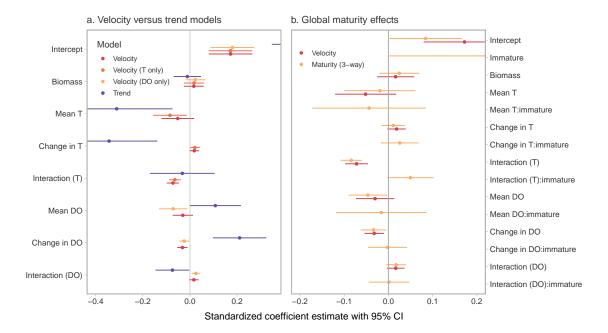


Figure S23: Global coefficient estimates for all fixed effects in a range of model configurations. All 'velocity' models predicted biotic velocity in response to climate changes calculated as gradient-based velocities (trend/spatial gradient). 'Trend' model predicts % change in biomass in response to climate changes measured as trends. Unless noted in parentheses, both temperature (T) and dissolved oxygen (DO) are included in each model. 'Interactions' are between the change in each climate variable (included in parentheses) and the mean conditions for that same climate variable. All models incorporate some degree of density-dependence by including the mean estimated biomass across all years. The model with a maturity effect (b. yellow dot-whisker-climate velocities and means interact with maturity) treats mature populations as the intercept.

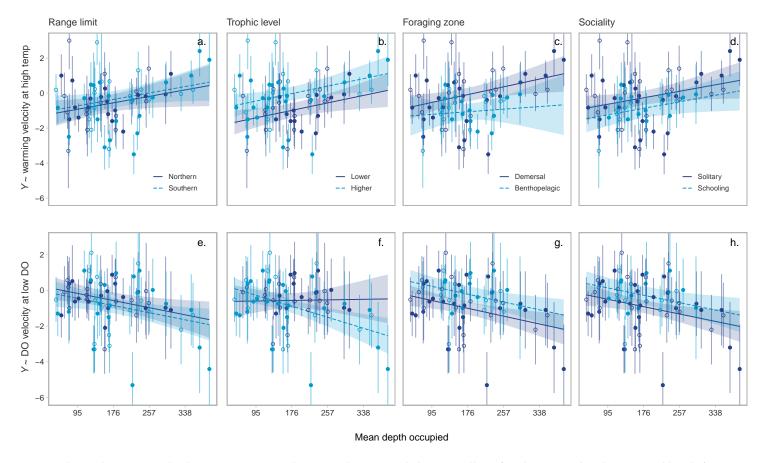


Figure S24: Ecological traits overlaid on responses to climate velocities, while controlling for the mean depth occupied by different maturity classes of each species. Points represent each species' raw slope estimates and their CI from the velocity model each coloured based on species and maturity-class specific ecology, with open circles for immature, and closed circles for mature biomass. Regression lines are derived from mixed-effect models with random intercepts for species and maturity classes nested within species; however, uncertainties are based only on fixed-effects and do not account for uncertainty in the slope estimates. In addition to the variables illustrated, each model contains slopes for both high and low mean conditions, but only the raw data and the estimated relationship for the stronger effect (highest temperatures and lowest DO) are plotted here. An interaction between ecology and mean depth is included only when statistically significant.

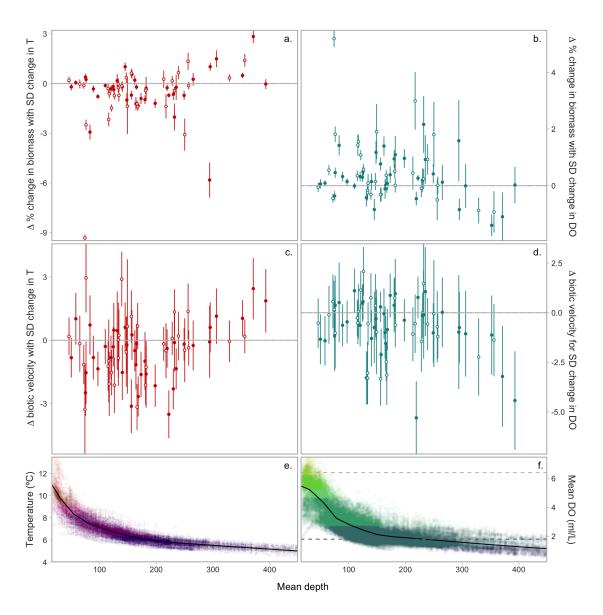


Figure S25: The slopes of the relationship between changes in biomass and climate, when calculated for the environmental extremes expected to have the greatest physiological impact—(b) highest temperature locations and (c) lowest DO locations—do not change predictably with mean occupied depth. Line segments on points represent 95% CIs on slope estimates from spatial models. Bottom panels illustrate relationships (smoothed solid lines are GAMs) between estimated temperature (d) and DO (e) with mean depth in each 4x4 km survey grid cell (dots). Dashed lines (e) represent complete saturation of seawater at 1 bar, 10 degree C, 35 salinity (upper), or threshold for mildly hypoxic conditions (< 1.8 ml/L; lower).

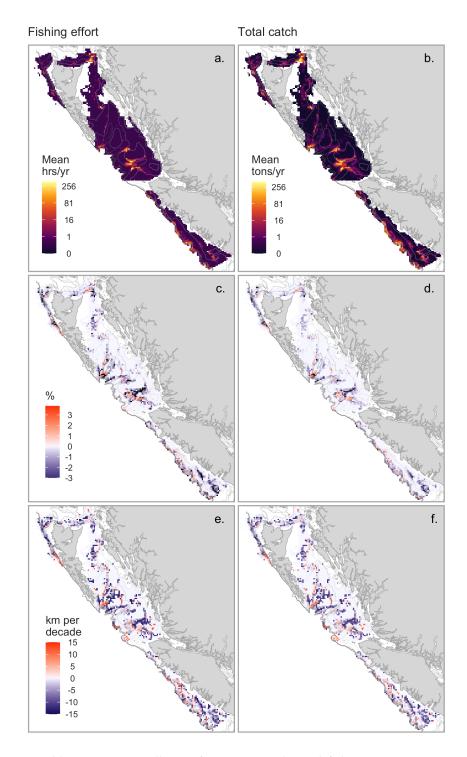


Figure S26: Variables capturing cell-specific commercial trawl fishing intensity: mean hours of commercial trawl fishing conducted (a) and mean tons of fish caught (b) per year, the % change in each estimate of fishing pressure (c, d), and the velocity of change in fishing pressure (e, f) between 2008 and 2018 for each 4×4 km cell within the survey footprint.

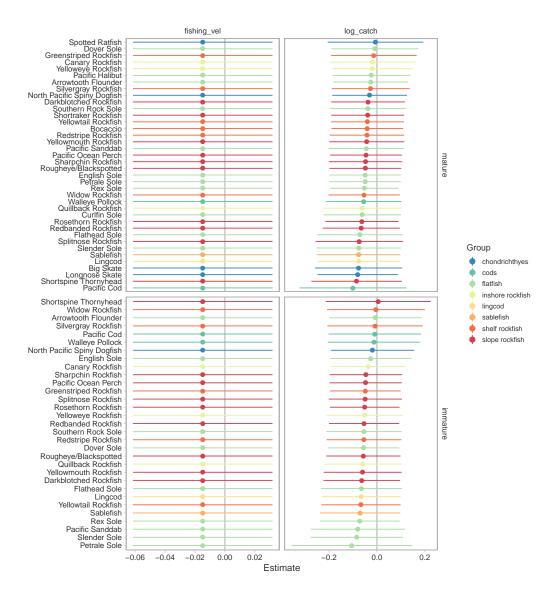


Figure S27: Species-specific coefficient estimates for variables accounting for fishing effort. This model includes the same climate variables as the "Velocity" models in Figures 2b and S23, as well as two variables capturing cell-specific commercial trawl fishing intensity: the velocity of change in mean hours of fishing that occurred annually between 2008 and 2018 (fishing_vel) and the mean tons of fish captured annually across those years (log_catch). Other variables illustrated in Figure S26 were explored, including interactions between mean and change values, but the rest were not estimate-able in models including the full set of climate variables.

Reviews

Editor Comments to Author:

I have chosen to go for major revision, despite only one of the three reviewers recommending this course of action, as I think this will give you greater scope for dealing with the issues that have been raised.

As you know Fish and Fisheries asks authors to submit papers with wide geographical scope. Your study is limited to a relatively small area off the west coast of Canada. You do mention the wider context in your discussion but it would good to see this made more explicit by making comparisons with other areas of the world where your work could be relevant. The papers you mention that deal with other areas must be based on results from other areas and it would be useful if you could be more specific about these so the reader can see more clearly the wider significance of your results.

We were pleased to learn of the three helpful and positive reviews of our manuscript. We have addressed all reviewer comments in our response below.

Regarding the above comment about making more specific comparisons with other areas of the world where the work could be relevant, we have added new text from line 489 to line 501 in the discussion.

Reviewer: 1

Comments to the Author Comments on the manuscript, "Contrasting climate velocity impacts in warm and cool locations show that effects of marine warming are worst in already warmer temperate waters" (English et al.)

General Comments A1: The authors examined biotic velocity patterns (from biomass distributions of marine species) within the Canadian basin in response to local climate trends and velocities. Overall, the paper is well-written and the results were clearly presented. It was interesting to see the differences in species-specific responses to changes in local climate, depending on whether a region experiences warming or cooling trends over time. I also liked that the authors recognized potential caveats of their study in the discussion, which partly addressed my first concern on the use of predicted species-specific biomass in linking biological changes with the local climate. I would think that using the actual biomass estimates as a response to the GLMMs would be a more straightforward way of capturing the relationship between these variables. On this note, I wonder as to how the observed and predicted species biomass (for mature and immature classes) compared overall.

We understand the concern with using predicted biomass in our second-level model, as we note in the discussion. We used modelled biomass estimates as a response because we needed a biomass estimate in each grid cell to calculate biotic velocities for comparison to equivalent climate velocities.

Regarding observed and predicted species biomass: we have added Figs S₃ and S₄ of predicted vs. observed plots for each species-maturity combination. Despite model convergence, a couple immature species have a relatively poor fit to the positive biomass component due to low numbers of positive samples, which we note in the caption. We note that the immature components for these two species (Widow Rockfish and Yelloweye Rockfish) either follow the same pattern as the mature component in the biotic-climate velocity models with greater uncertainty (Widow Rockfish, Fig. 4) or had insignificant effects (Yelloweye Rockfish, Fig. 4).

A2: Also, as to the computation of biotic and climatic velocities, were model spatial predictions done annually between 2008 and 2018 (I supposed they were to allow for the calculation of pixel-wise temporal trends)? This is quite unclear and it would help to briefly mention/clarify this in the text. In the computation of the spatial gradients, the authors used the last two sample periods (2015-2018) but I am not quite clear on which temporal trends were used to calculate the velocities? I see that in particular, temporal trends were computed for biennial time-steps depending on the location of the survey LL.241-245 (now line 247). I would suspect that depending on the temporal subset, the temporal trends would be different, which would also be reflected in the subsequent computed values of velocities. How were these differences in the temporal trend between time slices (and/or portions of the surveyed region) handled in the analyses?

Model spatial predictions were made biennially between 2008–2018 or 2007–2017, depending on the surveyed area. The temporal trends were for these same timesteps (stated on line 247), but how differences between survey area are addressed is now explained more clearly with sentences on line 249: "In order to compare between survey areas with different sampling years, we converted values for all cells to a rate of change, or trend, per decade" and line 254 "input information related to a cell and any cell near enough to share spatial information in a gradient or in the overall spatial models (this includes all response and predictor variables) all share the same sample years." We have not attempted to interpolate for missing year-locations and instead to focus on the patterns detected in those that were sampled.

Specific comments

A3: L.97: Similarly should read Similar

Change made. See line 99.

A4: L.113: in should read from

Change made. See line 113.

A5: L.134: In this subsection, it is good to have the information on survey sites (number of sites sampled) and a map that will show the sampling stations, perhaps in the supplementary materials.

We have added Fig. S1, which shows the distribution of survey samples in both time and space, and the mean and range of sample numbers per year are now given on line ??.

A6: L.243: should it be 11 years between 2008 and 2018? And 9 years between 2009 and 2017?

The reviewer is correct, thanks. Change made. See line 247.

Figure 1a, please include the geographical coordinates on the map for reference.

Change made.

Reviewer: 2

Comments to the Author

B1: Thanks for the opportunity to review this work from English et al. This work presents an interesting evaluation and application of temporal trends of temperature and oxygen across several demersal marine fishes, using in some cases, climate velocity. My main concern within this this work is that the gap that this manuscript is trying to fill is a little bit unclear. For example, lines 79-92 is just justification/explanation of the different climate velocity metrics. The conclusion is that local velocity is better because analog velocity has a problem with the threshold (this is a problem of definition not the metric itself). Another is in lines 93-103, which introduces the concept of biotic velocity but unclear to me why should be included here. Same with lines 104-116. Totally disconnected from the previous paragraphs. I would have expected that you take the concepts mentioned before and bring the gap together of why groundfish...etc. Lines 111-116 IS the gap of this paper in my opinion and has been expressed in only 5 lines.

We describe the broad gap we are trying to fill from line 66 to line 80, preceding the explanations on line 81 through line 105 that introduce the concepts underpinning our approach to filling this gap. The gap identified by the reviewer (on line 113 to line 118) is more of a description of the case study on which we test our approach.

B2: Another major issue with this manuscript is related to climate velocity and Oxygen. Brito-Morales et al. 2018 provide a good example of why this could be not appropriate from an ecological point of view (at least in marine systems). Climate velocity, in ecological terms, indicates how much and in which direction a species distribution would need to move to remain in the same climatic environment. So, are species' distribution really "following" oxygen levels? What is the performance curve of preference for oxygen for species in general? (For example, Figure II in Brito-Morales et al. 2018) If oxygen declines, species' distributions will tend to shift to their "preference level" and, more importantly, if the conditions are restored species would tend to shift back to their original condition? There is a reason why climate velocity in marine systems has been used mainly with temperature and in terrestrial systems with temperature and rainfall. The answer is in Box 3 (Methodological Considerations When Applying Climate Velocity, Which Environmental Variables?) in Brito-Morales et al. 2018. I'm not saying that the authors' approach is wrong, it is just that I do not understand the ecological reason why oxygen is a good predictor of species range shifts. The authors have not provided any fundamental analyses to justify this inclusion in the paper.

We agree that our justification for considering an effect of DO on fish distributions was omitted from the this manuscript and thank the reviewer for pointing this out. We have add this to line 118 where we now note that "a warmer ocean is expected to hold less oxygen, while becoming more stratified (Levin and Le Bris 2015), and some groundfish species in the Eastern Pacific have been shown to respond to these changes (Keller *et al.* 2017)". While a climate-analog approach using a hypoxia threshold would be interesting to explore, we think it is beyond the scope of this paper. Instead, we use this analysis to test whether including evidence of any local changes (trends or velocity) in DO are correlated with the change in species biomass, after accounting for changes in temperature. Further analyses that explore potential thresholds of response are planned; however, relatively few locations in this analysis actually drop below expected hypoxia thresholds. What we are testing for here is actually whether a reduction in DO might also be correlated with habitat selection or population growth rates through non-lethal or indirect mechanisms (e.g., slower metabolic rates, predator avoidance, changes in decomposition rates). We note aspects of this in the discussion on line 483, line 563, and line 566.

We have added a supplemental figure depicting the species-level predictions in response to temperature from a model with just temperature and from one with both variables Figure S16 and refer to it on line 427 where we now say "Including DO in the model along with temperature did not substantially alter the effects of temperature for any species".

More details comments:

B₃: 79-81. That's not totally true. Local climate velocity has been more used in medium-coarse resolution because its tends to be much variable at finer scales due to the temporal component (denominator).

We have clarified on line 81 that population-wide indices refers to single value for a whole population, such as centre of gravity, which is a much coarser scale than a "medium-coarse resolution". We address issues with variability in the denominator (the spatial component) in the paragraph starting on line 257.

B4: 118. (now line 122) It says "velocity" but should be "climate velocity"?

We think the current phrasing "shaped by local temperature and DO trends and velocities" is clearest because it refers specifically to temperature and DO and couldn't be understood as meaning other climate variables not listed.

B5-6: 125, 130, n... Be consistent with terminology. Should be "climate velocity" instead of "velocity"

We think that because we are looking at both biotic and climatic velocities, "gradient-based velocities" is appropriate on line 129 and "velocities of biotic and climatic change" on line 133 is also most accurate.

B7: 217 (now line 221) "We predicted" but is it possible to use survey data to compare how well the predictions are with "real" information? I think this is an important step in using prediction. There

is a good free available dataset at finer resolution that could be used to compare the predictions and has been omitted by authors.

Agreed. We have added supplemental Figure S2 of predictions vs. observed data for climate variables. However, we are not sure what finer resolution free data is being referred to. Sea surface temperature data exists, but so far as we know, similar spatial and temporal resolution bottom temperature and DO layers are not freely available. In the discussion starting on line 587, we also acknowledge that "An alternative would be a ROMS (Regional Ocean Modeling System) model (Peña et al. 2019), which could extend the temporal scope, allow for accounting of climate at other times of the year (e.g., temperature during spawning; Laurel and Rogers 2020), allow for inclusion of variables not typically measured with survey data (e.g., primary production), and allow for forward projections. Preliminary investigations indicated a strong correlation between our CTD projections and recently updated ROMS bottom temperatures. With the greater spatial and temporal extent that ROMS data will provide, calculation of more geographically precise analog-based climate velocities could be used to further refine the identification of areas important for conservation (Brito-Morales et al. 2018)." The ROMS model output, however, was not yet available when this analysis was conducted.

B8: 237 (now line 242) gradient-based velocity or more commonly "Local climate velocity" in my opinion does not fit well at finer resolution. It is more for medium-coarse resolution (see Brito-Morales et al. 2018 SOM). Also, in terms of climatic variables local climate velocity "has been favoured by ecologists when gradients are smooth and where there is one main variable driving change" (Brito-Morales et al. 2018). This is study is not that case. Another "climate-analog velocity has usually been used with multiple variables" (Brito-Morales et al. 2018). To me, this would be an adequate metric to applied in this paper. Also, check Garcia Molinos et al. 2017 GCB paper in Japan MPAs using climate-analog velocity.

We discuss climate-analog velocity and explain why we do not use it on line 92 "the choice of thresholds and other statistical properties (e.g., clumpiness) make them less useful than gradient-based local velocities for meta-analysis (Ordonez and Williams 2013)". Indeed we did explore their application in this context and identified these two primary obstacles: 1. without a much longer time series of environmental variables from which to identify relevant thresholds of environmental change, any threshold choice becomes quite arbitrary, and 2. statistical meta analyses that control for spatial autocorrelation worked well on the gradient-based velocities by employing a Student-t observation distribution, but the analog climate velocities produced bizarre statistical distributions ("clumpy" and extremely heavy tailed) that we couldn't adequately fit models to.

B10: 260 (now line 269) "We calculated spatial gradients using the vocc R package (Brown and Shoeman)". The authors should not be using this package as it has been replaced by "García Molinos, J., Schoeman, D. S., Brown, C. J. and Burrows, M. T. (2019). VoCC: The Velocity of Climate Change and related climatic metrics. R package version 1.0.0. "I strongly recommend performing new update calculations based on the previous paper/Gituhub repository.

This update came out after we had invested effort writing custom functions that were tightly integrated with the data formats accepted and produced by the older version. Therefore, we opted to use a subset of the data used in the VoCC vignette to test that both sets of functions produce

identical results; fortunately, they do. We have added text to line 269 also directing the reader to the newer package version and now state that we tested to ensure they produced the same results.

B11: 294 (now line 305) Simulation study section. I would move this section to Supplemental as it is disconnected from the others and was difficult to follow up.

We feel that the simulation study is critical to justifying the validity of our analysis, only takes two paragraphs to explain the method and results, and is illustrated in a key figure (2). Therefore, we would prefer to leave it in the main text. However, if the editor feels differently, we would be happy to move this section to the supplement.

B12: 356-359 (now line 427) Local climate velocity is a vector so it is important to know also the direction (not just how much but also where!). This is why using O2 is not so good idea in terms of having 2 different local climatic variables in climate velocity estimates (see main comment). For instance, are vectors from both variables in opposite directions? how do they cancel each other if that it's true? Does the same direction mean you added those vectors (and magnitudes) together? What would that mean for species/fish context? (which is the more relevant aspect for the paper) Perhaps using analog climate velocity for multivariate purposes (Garcia Molinos et al. 2017 GCB is a good place to start). To me, these results do not mean too much if the context is just to describe climate velocity estimates. Please check also Hamann et al. 2015 for more context and Brito-Morales et al. 2018 Box 3 Combining Environmental Variables section.

We considered this, but if temperature and DO levels both go into the same velocity calculation, it won't help us understand which factor is more likely to be driving the observed patterns. We think a combined analogue velocity would be interesting, but outside the scope of this paper. We did, however, test to see if including DO was impacting our conclusions regarding the effects of temperature, and it does not. We have added a supplemental figure depicting the species-level predictions in a model with just temperature and one with both variables Figure S16 and refer to it on line 427.

B14: 493-498 (now line 514) This just confirm what I've been suggesting in previous comments: I do not think that local climate velocity is a good estimate metric at finer resolutions or even with multiple environmental variables. I strongly recommend authors to climate-analog velocity and a proper, perhaps structured and well cited, framework about why Oxygen is a variable than can be used with climate velocity to predict species' range shifts.

Please see our responses to B2, B3 and B12 above.

B₁₅: 538 The authors constantly through the ms refers to "individual movement", which is not appropriate for climate velocity in my opinion. Climate velocity is a metric that is related with population movement and NOT individual movement. Seems to me that authors have not realised that this is a major ecological point of view and should be noted and corrected in the actual version of the ms.

We have modified all references to "individual", "organism", and "fish" movement to refer to population or fish-population movement.

Reviewer: 3

Comments to the Author

C1: This is a complicated paper. It presents some important results that will help to refine current ideas about the spatial adjustments of fish communities in response to climate shifts. The manuscript is a little hard to read in places where the authors use jargon. An example is l.52 where they refer to "climate-induced hyperstability-catch rates". I suspect that many readers will have no idea what this is. Line 73 uses the term "depredation", which is correct, but I had to look it up: URL.

"Climate-induced hyperstability-catch rates" has been replaced by "such changes can cause the efficiency of fishing to increase if population density increases faster than range expansion during population growth, or temporarily maintain catch rates despite population decline if organisms move towards preferred habitat as it becomes available" on line 53. We replaced "depredation" by "predator-prey interactions" on line 71. We have tried to reduce jargon wherever possible throughout the manuscript.

C2: In an effort to be specific, yet brief, about their results, the authors resort to complex sentences. One example is the repeated use of this phrasing: "result in smaller negative or larger positive biotic velocities." Another example is on l.399 (now line 411): "the expected biotic velocity was strongly negative for a number of species-maturity combinations (19 of 69) in the warmest locations when experiencing maximum climate velocity and tended to be positive in cooler locations experiencing the same high climate velocity". These sentence constructions are hard to digest because the point is lost in their complex structure.

In the results line 387 now reads "local biomass was more likely to increase and to result in larger positive biotic velocities (or to decrease less and result in less negative biotic velocities)" The sentence on line 411 now reads "in the warmest locations experiencing maximum climate velocity the expected biotic velocity was strongly negative for 19 of 69 species-maturity combinations (Figure 5b). Meanwhile, in cooler locations experiencing the same high climate velocity, biotic velocities were often positive"

We acknowledge that these are still somewhat complex statements (although hopefully in easier to digest now), but we are trying to balance simplicity with specificity and accuracy in the results section. We do interpret these results using more general language in the abstract and in the discussion (e.g., line 474).

C3: A surprising result is the very small distances that fish would need to move to maintain their starting thermal or oxygen conditions. l.356-359 (now line 366). "... an organism would have to move an average of 10.5 km/decade (mean of absolute values) to maintain its starting thermal environment and an average of 11 km/decade to maintain initial DO levels." I'm surprised that such small shifts are measurable given the imprecision of trawl surveys. The reported shifts are about 1km per year ... I can't help wondering whether simply repeating a trawl survey might yield movements of this magnitude, without needing to invoke community adjustment to environmental shifts. The authors are detecting a very small effect using sophisticated statistics operating on

imprecise data. The analyses are impressive, but are these very small spatial shifts a real effect of climate? And what are the implications of such small shifts. The authors miss the opportunity to point this out in the Discussion (see below).

It's important to note that the 10.5 km/decade value is an *average* amount of climate velocity across the whole coast. Some locations experience much more extreme temperature climate velocities (up to \sim 80 km per decade; Figure 1f). When it comes to animal abundances (as opposed to the temperature measurements underlying the above), we agree that there is a large amount of observation error in this or any trawl survey and this error is a very valid concern. The fact that "simply repeating a trawl survey might yield movements of this magnitude" is partly the reason for (1) the hierarchical approach across many species and (2) the simulation study. By simulating similar magnitudes of biotic change (with similar spatial correlations), we showed on line 322 that "Null models based on simulated biotic trends and observed climate trends showed fewer significant relationships at the species level than would be expected by chance (Figures S5, S6, and S7)."

C4: In the Discussion, l.575 (now line 606) states: "Climate change is expected to have large impacts on fish stocks and their management, particularly with respect to changes in species distribution (e.g., Tommasi et al. 2017, Karp et al. 2019, Free et al. 2019)". l.588 (states that "Shifts in distribution, such as those reported here, can have implications for calculation of indices of abundance, and estimates of stock size and stock status, which in turn may impact harvest recommendations (Szuwalski and Hollowed 2016, Karp et al. 2019)." I think given the very small spatial shifts that this study demonstrates, the authors should really turn this discussion focus around. Specifically, while large spatial shifts are expected, what this fine scale study demonstrates is that small, spatial shifts may occur due to complex interactions between species-specific fish distributions and both climate trends and climate velocities. Whereas large shifts in distributions would demand adjustment of management strategies, the small shifts detected in this study indicate no such adjustment is needed in the near-term, while providing a sophisticated set of analytical tools for sensitive detection of larger shifts if and when they occur.

We agree with the statement that "small, spatial shifts may occur due to complex interactions between species-specific fish distributions and both climate trends and climate velocities" and think that our discussion gets at this point. We do not necessarily agree that "the small shifts detected in this study indicate no such adjustment [of management strategies] is needed." We note that:

- We are only examining a single decade and such changes will compound over time.
- A redistribution of species can have major consequences for fishing opportunities and conservation of rarer species when "choke" species (species with limited quota that co-occur with species of fishing interest) limit fishing opportunities. For example, on the British Columbia coast, Bocaccio and Yelloweye Rockfish have limited fishing opportunities for more desired species with higher quotas (e.g., Halibut).
- A redistribution of species can have major consequences when fishing is geographically fixed, such as for small nations or Indigenous peoples with defined spatial fishing regions.

• A redistribution of species can have major consequences for marine spatial planning. For example, a marine reserve designed to protect a particular at-risk population may no longer be as effective after a local redistribution of abundance.

We now include these points in a new paragraph starting on line 596 in the discussion.

I think the paper should be published. Papers that are true to the data, even in the face of dominant paradigms, are good for science. But the authors should not shy away from conclusions supported by their data, in preference to what might have been expected prior to analysis.

Thank you, we agree.