

## GOPEN ACCESS

**Citation:** Farr ER, Johnson MR, Nelson MW, Hare JA, Morrison WE, Lettrich MD, et al. (2021) An assessment of marine, estuarine, and riverine habitat vulnerability to climate change in the Northeast U.S. PLoS ONE 16(12): e0260654. https://doi.org/10.1371/journal.pone.0260654

Editor: Daniel E. Duplisea, Maurice Lamontagne Institute, CANADA

Received: July 6, 2021

Accepted: November 12, 2021

Published: December 9, 2021

**Copyright:** This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the <u>Creative</u> Commons CC0 public domain dedication.

**Data Availability Statement:** All relevant data are within the manuscript and its Supporting Information files.

**Funding:** Funding for this project was provided by the National Oceanic and Atmospheric Administration (NOAA) NMFS Office of Habitat Conservation and NMFS Office of Science and Technology. ECS Federal, Inc in support of NOAA NMFS Office of Science and Technology provided salary for authors MWN and MDL. PJA was funded by Award 20-10-B-281 from the National Marine RESEARCH ARTICLE

# An assessment of marine, estuarine, and riverine habitat vulnerability to climate change in the Northeast U.S.

Emily R. Farr<sup>1\*</sup>, Michael R. Johnson<sup>2</sup>, Mark W. Nelson<sup>3</sup>, Jonathan A. Hare<sup>4</sup>, Wendy E. Morrison<sup>5</sup>, Matthew D. Lettrich<sup>3</sup>, Bruce Vogt<sup>6</sup>, Christopher Meaney<sup>7</sup>, Ursula A. Howson<sup>8</sup>, Peter J. Auster<sup>9</sup>, Frank A. Borsuk<sup>10</sup>, Damian C. Brady<sup>11</sup>, Matthew J. Cashman<sup>12</sup>, Phil Colarusso<sup>13</sup>, Jonathan H. Grabowski<sup>14</sup>, James P. Hawkes<sup>15</sup>, Renee Mercaldo-Allen<sup>16</sup>, David B. Packer<sup>17</sup>, David K. Stevenson<sup>2</sup>

1 Office of Habitat Conservation, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Silver Spring, Maryland, United States of America, 2 Habitat and Ecosystem Services Division, Greater Atlantic Regional Fisheries Office, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Gloucester, Massachusetts, United States of America, 3 ECS, Under contract to the Office of Science and Technology, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Silver Spring, Maryland, United States of America, 4 Northeast Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Woods Hole, Massachusetts, United States of America, 5 Office of Sustainable Fisheries, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Silver Spring, Maryland, United States of America, 6 NOAA Chesapeake Bay Office, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Annapolis, Maryland, United States of America, 7 Gulf of Maine Coastal Program, U.S. Fish and Wildlife Service, Falmouth, Maine, United States of America, 8 Office of Renewable Energy Programs, Bureau of Ocean Energy Management, Sterling, Virginia, United States of America, 9 Mystic Aquarium & University of Connecticut, Groton, Connecticut, United States of America, 10 Region 3, U.S. Environmental Protection Agency, Wheeling, West Virginia, United States of America, 11 Darling Marine Center, University of Maine, Walpole, Maine, United States of America, 12 Maryland-Delaware-DC Water Science Center, U.S. Geological Survey, Baltimore, Maryland, United States of America, 13 Region 1, U.S. Environmental Protection Agency, Boston, Massachusetts, United States of America, 14 Marine Science Center, Northeastern University, Nahant, Massachusetts, United States of America, 15 Northeast Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Orono, Maine, United States of America, 16 Milford Laboratory, Northeast Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Milford, Connecticut, United States of America, 17 James J. Howard Marine Sciences Laboratory, Northeast Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Highlands, New Jersey, United States of America

\* efarr@manomet.org

## Abstract

Climate change is impacting the function and distribution of habitats used by marine, coastal, and diadromous species. These impacts often exacerbate the anthropogenic stressors that habitats face, particularly in the coastal environment. We conducted a climate vulnerability assessment of 52 marine, estuarine, and riverine habitats in the Northeast U.S. to develop an ecosystem-scale understanding of the impact of climate change on these habitats. The trait-based assessment considers the overall vulnerability of a habitat to climate change to be a function of two main components, sensitivity and exposure, and relies on a process of expert elicitation. The climate vulnerability ranks ranged from low to very high, with living habitats identified as the most vulnerable. Over half of the habitats examined in this study are expected to be impacted negatively by climate change, while four habitats are

Sanctuary Foundation. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

expected to have positive effects. Coastal habitats were also identified as highly vulnerable, in part due to the influence of non-climate anthropogenic stressors. The results of this assessment provide regional managers and scientists with a tool to inform habitat conservation, restoration, and research priorities, fisheries and protected species management, and coastal and ocean planning.

### Introduction

Climate change is impacting all aspects of marine ecosystems [1–4]. There has been substantial work to understand the effect of climate change on marine species and communities [5–9]. In comparison, the effect of climate change on marine, estuarine, and riverine habitats is not as well understood. In some cases, significant research has advanced our understanding of the effects of climate change on individual habitats, or a single component of habitat. For example, many studies have investigated shifting thermal habitat and its effects on species distributions [10–15]. Other studies have explored the impacts of projected changes in the climate on living habitats such as corals [16, 17], mollusks [18–21], seagrasses [22–24], and coastal wetlands [25–28]. However, these studies typically focus on a limited number of climate drivers (e.g., temperature, pH), and do not provide a comprehensive assessment of how climate change may affect habitats that support marine, coastal, and diadromous species (i.e., fish, invertebrates, and protected species). Warming waters in rivers, estuaries, and the ocean, in concert with ocean acidification, water column stratification, deoxygenation, and sea level rise (SLR) can interact with one another and with other stressors to cause complex and often unanticipated synergistic climate effects to species and habitats [4, 29–33].

Understanding how climate change will impact habitats across an ecosystem is necessary to inform decisions about habitat conservation, fisheries management, and coastal and offshore planning. Habitats have long been impacted by human activities such as land-use and land-cover change, point and non-point source pollution, dredging and filling, fishing, sand and gravel mining, oil and gas exploration, damming, and shoreline hardening [34–39]. There is a growing understanding that climate change also has affected, and will increasingly affect, riverine, estuarine, and marine habitats. The effects of climate change will exacerbate the vulnerability of species, habitats, and ecosystems that are already under stress from natural and anthropogenic stressors [4, 40-42].

Habitat requirements differ by species, and as climate change affects habitats, the indirect effects on species will be multifaceted [32, 41]. One of the most straightforward examples is the change in species distribution as a result of warming waters [43–45]. The water column is habitat for aquatic species, and most, if not all, have an optimal temperature range [29, 46, 47]; as water temperature increases, the optimal temperature for marine species would generally shift poleward and species distributions would follow.

There are numerous other effects of climate on habitat and thus numerous other indirect effects of climate on species that use those habitats, especially those (e.g., coastal wetlands, shellfish habitats, kelp forests, and corals) which provide important ecosystem services and functions for key life stages of marine and coastal organisms [1, 4, 48]. Over the past century, coastal wetlands have been affected by SLR, contributing to a cumulative loss of habitat [49–54]. Coastal wetland habitat supports juvenile growth and survival for many marine fish species [55] and their prey [56], and the climate-driven impairment of habitat may result in decreased population productivity for sensitive species. Marsh loss also threatens the

populations of many species of birds that depend on coastal wetland habitat for breeding, nesting, and wintering [57]. Increasing temperatures may shift the geographic range of kelp beds, with implications for the myriad fish, birds, invertebrates, and marine mammals that they support [58, 59]. Calcifying marine organisms, including mollusks, echinoderms, and corals, are particularly sensitive to changes in pH, carbonate ion concentration, and the saturation state of calcium carbonate minerals–collectively known as ocean acidification [20, 60–63]. Rising temperatures and ocean acidification are negatively impacting shallow and deep sea corals [16, 17, 64]. Loss of live coral cover is related to decreases in abundance of a number of fish species; the magnitude of decline has been associated with the dependence on live coral [64–67].

Although the effects of climate change on living habitats may be more pronounced, the effects on habitats with primarily non-living characteristics (e.g., sand, mud, rock, water column) cannot be overlooked. These habitats serve an important role in the reproduction of several groups of protected species and in the foraging of other species. For example, beaches that pinnipeds use for pupping and sea turtles use for nesting [68] may be affected by increased erosion and inundation from SLR, and the frontal features that aggregate prey for fish, seabirds, and marine mammals in the open ocean may shift in location and strength [69, 70]. SLR is also impacting the intertidal foraging [71] and coastal nesting [72] habitat used by shorebirds and seabirds.

To develop an ecosystem-scale understanding of the effect of climate change on habitats, we conducted a trait-based climate vulnerability assessment for marine, estuarine, and coastal riverine habitats in the Northeast U.S. Shelf Ecosystem, from Cape Hatteras through the Gulf of Maine. The coastal northeastern U.S. and the adjacent continental shelf are warming at a particularly rapid rate [73–75] that is expected to increase [76]. Observed rates of sea level rise in the Northeast have also been higher than the global mean, and are projected to increase [77]. This underscores the importance of understanding how these changes will impact the region's habitats. A trait-based climate vulnerability assessment is one method used to evaluate the potential risks of climate change to species or ecosystems [78-81]. An expert elicitation process was used to estimate climate sensitivities and exposure, which can provide broad, transparent, and relatively rapid evaluation of the vulnerability of multiple species [80, 82–84], habitats [85, 86], or ecosystems [87]. This approach facilitates assessment of a large geographic area with variability in the availability of data across habitats and space (e.g., habitat range and condition, physical and chemical thresholds for habitat impacts, and limitations in downscaled climate projections for nearshore areas). The main purpose of this assessment is to provide information for scientists and natural resource managers to identify research priorities and improve management decisions for these particular habitats and the species that rely on them. Further, we seek to begin to elucidate the many indirect effects of climate change on species through direct effects on habitats.

#### Methods

#### Method development

At a NOAA Fisheries workshop in July 2018, habitat specialists and managers reviewed literature of various existing climate vulnerability assessment (CVA) methodologies and decided to base this habitat CVA on a hybrid of the framework developed for NOAA's Fish Stock Climate Vulnerability Assessment (FSCVA) [81, 88], and a habitat vulnerability model developed for the Northeastern Association of Fish and Wildlife Agencies (NEAFWA) [85]. This hybrid approach adapted elements from each framework to design a methodology that could be applied to the full suite of marine, estuarine, and riverine habitats in the Northeast U.S. Using the overall structure of the NOAA's FSCVA allowed the results of this assessment to be easily synthesized with the vulnerability of specific species. The NEAFWA methodology provided attributes indicative of the response of terrestrial and non-tidal wetland habitats to climate change, which were adapted to marine, estuarine, and riverine habitats.

This assessment used a trait-based framework that incorporated two components: sensitivity and exposure. The general premise was that the overall climate vulnerability of a habitat is a function of the interplay between the sensitivity and the potential exposure to future change. Many vulnerability assessments also include an adaptive capacity component. However, adaptive capacity and sensitivity in biological systems are confounded; the same trait that infers high sensitivity may be viewed as low adaptive capacity, and vice versa [89]. Therefore, we incorporated adaptive capacity into the sensitivity component.

#### Habitat selection

In this assessment, habitat is defined as the coastal rivers, estuaries, and marine waters, from the bottom through the water column including the physical, geological, chemical, and biological components of an ecosystem that a species depends on to complete its life cycle-reproduction, development, growth, and survival [90, 91]. We included 52 habitats based on their importance to NOAA's trust resources (https://www.fisheries.noaa.gov/region/new-englandmid-atlantic#species) in the Northeast U.S.: 23 marine habitats, 19 estuarine habitats, and 10 riverine habitats (Table 1). Habitats that were present in multiple systems (e.g., submerged aquatic vegetation is present in all three systems) were assessed separately to capture differences in the climate and non-climate stressors on that habitat. Selected habitats were arranged in a hierarchical classification system based on the Federal Geographic Data Committee update [92] to the Cowardin Classification of Wetlands and Deepwater Habitats of the United States [93], with some modifications. For example, categories were included for water column habitats in the riverine, estuarine, and marine systems, which are not present in the Cowardin classification system. The resulting classification system allows for a comparison of the climate vulnerability of habitats across systems, sub-systems, classes, sub-classes, and geographic areas, which can reveal patterns and key drivers of vulnerability.

Definitions were developed for features and living and non-living characteristics of each habitat (S1 File). In order to explore patterns in the results, we also categorized the habitats into bottom substrate, living, water column, artificial structures, and invasive species. While each habitat was assigned to a single category based on its defining characteristics, some habitats could fit in multiple categories. For example, sand and mud bottom substrates also include living components (e.g., bivalve and gastropod infauna and epifauna communities), and invasive species (e.g., *Phragmites australis*) are also living habitats.

For most habitats, we assessed climate vulnerability across the full geographic range of the study area, with three exceptions: estuarine emergent native wetland, estuarine emergent invasive wetland, and deep sea coral & sponge. Estuarine emergent wetlands (both native and invasive sub-classes) were assessed separately in the Mid-Atlantic and New England because biogeographic differences in coastal wetlands and/or the rate of SLR could result in differential climate vulnerabilities for this habitat. Deep sea coral and sponge habitats were split between the Gulf of Maine and the areas farther offshore on the Northeast U.S. Continental Shelf, slope, submarine canyons, and seamounts to assess potential differences in climate vulnerability associated with coral and sponge habitat density, depth, biodiversity, population genetics, and anthropogenic drivers such as impacts from fishing gear [65, 94–98].

In addition, sub-systems, classes, and sub-classes were differentiated to capture expected differences in sensitivity or exposure to climate change within the region. For example, emergent wetlands in tidal and non-tidal portions of the riverine system were assessed separately to evaluate the potential effects of changes in salinity and/or SLR on these two habitat types.

System	Sub-system	Class	Sub-class	Common Name	Category	Geographic Area
Marine	Intertidal	Reef	Mollusk	Marine Intertidal Shellfish Reef	Living	Entire Area
Marine	Intertidal	Rocky Bottom	Bedrock, Rubble, Cobble, Gravel	Marine Intertidal Rocky Bottom	Bottom Substrate	Entire Area
Marine	Intertidal	Unconsolidated Bottom	Mud	Marine Intertidal Mud Bottom	Bottom Substrate	Entire Area
Marine	Intertidal	Unconsolidated Bottom	Sand	Marine Intertidal Sand Bottom	Bottom Substrate	Entire Area
Marine	Subtidal	Aquatic Bed	Kelp	Marine Kelp	Living	Entire Area
Marine	Subtidal & Intertidal	Aquatic Bed	Red, Green, Small-brown Algae	Marine Red, Green, Small-brown Algae	Living	Entire Area
Marine	Subtidal & Intertidal	Aquatic Bed	Rooted Vascular	Marine Submerged Aquatic Vegetation	Living	Entire Area
Marine	Subtidal	Reef	Mollusk	Marine Subtidal Shellfish Reef	Living	Entire Area
Marine	Subtidal <200 m	Rocky Bottom	Bedrock, Rubble, Cobble, Gravel	Marine Rocky Bottom <200 m	Bottom Substrate	Entire Area
Marine	Subtidal <200 m	Unconsolidated Bottom	Mud	Marine Mud Bottom <200 m	Bottom Substrate	Entire Area
Marine	Subtidal <200 m	Unconsolidated Bottom	Sand	Marine Sand Bottom <200 m	Bottom Substrate	Entire Area
Marine	Subtidal >200 m	Reef	Deep Sea Coral & Sponge	Deep Sea Coral & Sponge: Gulf of Maine	Living	Gulf of Maine
Marine	Subtidal >200 m	Reef	Deep Sea Coral & Sponge	Deep Sea Coral & Sponge: Seamounts and Canyons	Living	Seamounts & Canyons
Marine	Subtidal >200 m	Rocky Bottom	Bedrock, Rubble, Cobble, Gravel	Marine Rocky Bottom >200 m	Bottom Substrate	Entire Area
Marine	Subtidal >200 m	Unconsolidated Bottom	Mud	Marine Mud Bottom >200 m	Bottom Substrate	Entire Area
Marine	Subtidal >200 m	Unconsolidated Bottom	Sand	Marine Sand Bottom >200 m	Bottom Substrate	Entire Area
Marine	Subtidal & Intertidal	Reef	Mollusk Aquaculture	Marine Shellfish Aquaculture	Artificial	Entire Area
Marine	Subtidal & Intertidal	Rocky Bottom	Artificial Structures	Marine Artificial Structures	Artificial	Entire Area
Marine	Subtidal <20 m	Water Column	Shallow Inner Shelf	Marine Shallow Inner Shelf Water Column	Water Column	Entire Area
Marine	Subtidal <200 m	Water Column	Shelf Surface	Marine Shelf Surface Water Column	Water Column	Entire Area
Marine	Subtidal <200 m	Water Column	Shelf Bottom	Marine Shelf Bottom Water Column	Water Column	Entire Area
Marine	Subtidal >200 m	Water Column	Slope Surface	Marine Slope Surface Water Column	Water Column	Entire Area
Marine	Subtidal >200 m	Water Column	Slope Bottom	Marine Slope Bottom Water Column	Water Column	Entire Area
Estuarine	Intertidal	Emergent Wetland	Invasive Wetland	Estuarine Invasive Wetland: Mid-Atlantic	Invasive	Mid-Atlantic
Estuarine	Intertidal	Emergent Wetland	Invasive Wetland	Estuarine Invasive Wetland: New England	Invasive	New England
Estuarine	Intertidal	Emergent Wetland	Native Wetland	Estuarine Native Wetland: Mid-Atlantic	Living	Mid-Atlantic
Estuarine	Intertidal	Emergent Wetland	Native Wetland	Estuarine Native Wetland: New England	Living	New England
Estuarine	Intertidal	Reef	Mollusk	Estuarine Intertidal Shellfish Reef	Living	Entire Area
Estuarine	Intertidal	Rocky Bottom	Bedrock, Rubble, Cobble, Gravel	Estuarine Intertidal Rocky Bottom	Bottom Substrate	Entire Area
Estuarine	Intertidal	Rocky Bottom	Artificial Structures	Estuarine Intertidal Artificial Structures	Artificial	Entire Area
Estuarine	Intertidal	Unconsolidated Bottom	Mud	Estuarine Intertidal Mud Bottom	Bottom Substrate	Entire Area
Estuarine	Intertidal	Unconsolidated Bottom	Sand	Estuarine Intertidal Sand Bottom	Bottom Substrate	Entire Area
Estuarine	Subtidal	Aquatic Bed	Kelp	Estuarine Kelp	Living	Entire Area

#### Table 1. Classification of habitats included in the assessment.

(Continued)

System	Sub-system	Class	Sub-class	Common Name	Category	Geographic Area
Estuarine	Subtidal & Intertidal	Aquatic Bed	Red, Green, Small-brown Algae	Estuarine Red, Green, Small-brown Algae	Living	Entire Area
Estuarine	Subtidal & Intertidal	Aquatic Bed	Rooted Vascular	Estuarine Submerged Aquatic Vegetation	Living	Entire Area
Estuarine	Subtidal	Reef	Mollusk	Estuarine Subtidal Shellfish Reef	Living	Entire Area
Estuarine	Subtidal	Rocky Bottom	Bedrock, Rubble, Cobble, Gravel	Estuarine Subtidal Rocky Bottom	Bottom Substrate	Entire Area
Estuarine	Subtidal & Intertidal	Reef	Mollusk Aquaculture	Estuarine Shellfish Aquaculture	Artificial	Entire Area
Estuarine	Subtidal	Rocky Bottom	Artificial Structures	Estuarine Subtidal Artificial Structures	Artificial	Entire Area
Estuarine	Subtidal	Unconsolidated Bottom	Mud	Estuarine Subtidal Mud Bottom	Bottom Substrate	Entire Area
Estuarine	Subtidal	Unconsolidated Bottom	Sand	Estuarine Subtidal Sand Bottom	Bottom Substrate	Entire Area
Estuarine	Subtidal	Water Column	Well-mixed	Estuarine Water Column	Water Column	Entire Area
Riverine	Non-tidal	Emergent Wetland	Invasive Wetland	Riverine Non-tidal Invasive Wetland	Invasive	Entire Area
Riverine	Non-tidal	Emergent Wetland	Native Wetland	Riverine Non-tidal Native Wetland	Living	Entire Area
Riverine	Tidal	Emergent Wetland	Invasive Wetland	Riverine Tidal Invasive Wetland	Invasive	Entire Area
Riverine	Tidal	Emergent Wetland	Native Wetland	Riverine Tidal Native Wetland	Living	Entire Area
Riverine	Tidal & Non- Tidal	Aquatic Bed	Algae	Riverine Algae	Living	Entire Area
Riverine	Tidal & Non- Tidal	Aquatic Bed	Rooted Vascular	Riverine Submerged Aquatic Vegetation	Living	Entire Area
Riverine	Tidal & Non- Tidal	Rocky Bottom	Bedrock, Rubble, Cobble, Gravel	Riverine Rocky Bottom	Bottom Substrate	Entire Area
Riverine	Tidal & Non- Tidal	Unconsolidated Bottom	Mud	Riverine Mud Bottom	Bottom Substrate	Entire Area
Riverine	Tidal & Non- Tidal	Unconsolidated Bottom	Sand	Riverine Sand Bottom	Bottom Substrate	Entire Area
Riverine	Tidal & Non- Tidal	Water Column	Well-mixed	Riverine Water Column	Water Column	Entire Area

#### Table 1. (Continued)

See <u>S1 File</u> for a more detailed description of each habitat.

https://doi.org/10.1371/journal.pone.0260654.t001

Similarly, emergent wetland habitats for both the estuarine and riverine systems were further divided into native (e.g., *Spartina* spp.) and invasive species (e.g., non-native genotype of *Phragmites australis*). Although other prominent invasive plant species occur in the Northeast U.S. region, their populations are not large enough to form a distinct habitat type like the dense and pervasive stands of *Phragmites australis* present in estuaries and tidal and non-tidal portions of rivers and streams in the region. In the marine system, artificial reefs and intertidal and subtidal artificial hard bottoms (e.g., riprap for shoreline protection) were assessed under a single artificial category. However, because of the prevalence of hardened shoreline structures in the estuarine system, intertidal and subtidal artificial structures were assessed separately. All river and stream habitats used by diadromous species were included in the assessment, and were not differentiated by stream order.

#### Sensitivity

The sensitivity of habitats to climate change was evaluated using nine sensitivity attributes (Table 2 and S2 File) covering a diverse range of traits indicative of how a habitat will respond

Sensitivity Attribute	Assessment Definition
Habitat Condition	The ability of the habitat to support a natural, fully-functional ecological community of organisms and the associated/expected ecosystem services.
Habitat Fragmentation	The extent to which a previously contiguous habitat is subdivided into isolated patches or fragments due to anthropogenic causes.
Distribution and Range	The historic geographic extent of a habitat, including the leading (i.e., the expanding or colonizing) edge and trailing (i.e., contracting or declining) edge, if applicable, and the water depths for which the habitat naturally occurs.
Mobility/Ability to Spread or Disperse	The ability or capability of a habitat to disperse, move, or spread to areas beyond its existing location.
Resistance	The ability of a habitat to tolerate a stressor and persist while retaining its functionality when subjected to a disturbance.
Resilience	The ability of, and the time period for, a habitat to recover from a disturbance.
Sensitivity to Changes in Abiotic Factors	A measure of a habitat's ability to tolerate changes in chemical and physical characteristics of the environment (temperature, salinity, dissolved oxygen, carbonate chemistry, and synergistic effects).
Sensitivity and Intensity of Non- Climate Stressors	A measure of a habitat's response to existing non-climate stressors, as well as the intensity of those stressors (dredging/filling, pollution/eutrophication, invasive species, harmful algal blooms, shoreline hardening, and synergistic effects).
Dependency on Critical Ecological Linkages	The extent to which a habitat depends upon associated species to maintain its health or function as a habitat.

Table 2.	Sensitivity	attributes.
----------	-------------	-------------

See S2 File for more detailed descriptions of the sensitivity attributes.

https://doi.org/10.1371/journal.pone.0260654.t002

to future changes in climate. For example, Habitat Condition and Habitat Fragmentation both reflect the ability of a habitat to support a natural and fully-functioning ecological community of organisms, while Mobility/Ability to Spread or Disperse, Resilience, and Sensitivity to Changes in Abiotic Factors are attributes that measure how well a habitat may respond to changes in climate. Sensitivity and Intensity of Non-climate Stressors was included to assess the effects of a suite of anthropogenic stressors on a habitat, because such stressors have the potential to reduce the ecological function and the ability of habitats to withstand climaterelated stressors [4, 40-42]. This attribute includes non-climate stressors that may affect habitats in riverine (e.g., dams, water diversions), estuarine (e.g., navigational dredging, eutrophication, shoreline hardening), and marine (e.g., bottom-tending fishing gear, ocean energy development) systems. To ensure consistency and repeatability in the application of these sensitivity attributes, a written definition and scoring bins were developed for each attribute. The scoring bins characterize Low, Moderate, High, and Very High scores for each sensitivity attribute. The assessment used a five-tally scoring method, as described in the FSCVA framework, where each scorer could distribute their five tallies between any of the scoring bins to best describe the uncertainty and/or the geographic variability within the study area. Whereas it is possible that some of these sensitivity attributes may have a stronger effect on vulnerability than others, this relationship is unknown and may differ between habitats; therefore, the sensitivity attributes were all given equal weightings.

#### **Climate exposure**

We used eleven equally weighted exposure factors to indicate the magnitude and overlap of projected changes in climate with the distribution of habitats (Table 3 and S3 File). This methodology uses a wide range of exposure factors, any of which could increase the vulnerability of

Exposure Factor	Projection / Source
Sea Surface Temperature	Northwest Atlantic Regional Ocean Modeling System
Bottom Temperature	Northwest Atlantic Regional Ocean Modeling System
Sea Surface Salinity	Northwest Atlantic Regional Ocean Modeling System
Bottom Salinity	Northwest Atlantic Regional Ocean Modeling System
рН	Coupled Model Intercomparison Project Phase 5
Precipitation	Coupled Model Intercomparison Project Phase 5
Air Temperature	Coupled Model Intercomparison Project Phase 5
Streamflow (Floods and Droughts)	Demaria et al. (2016)
River Temperature	Letcher et al. (2016)
Sea Level Rise	Sweet et al. (2017)

Tuble St. Encodure fuetoro	Table 3.	Exposure	factors.
----------------------------	----------	----------	----------

See S3 File for more detailed descriptions of the Streamflow, River Temperature, and Sea Level Rise exposure factors.

https://doi.org/10.1371/journal.pone.0260654.t003

a habitat, instead of focusing on just a single variable (e.g., temperature). We used this approach because individual habitats are likely to respond differently to the various changes that are anticipated with climate change. This multiple stressor approach also allowed us to identify potential cumulative or compounding impacts of multiple exposure factors. It is important to note that not all exposure factors directly impact each of these habitats. Only the factors that were applicable to each habitat were scored. For example, exposure to SLR was scored for coastal, shallow-water habitats (e.g., emergent wetlands, intertidal mud and sand), but not offshore habitats (e.g., deep sea corals and sponges). Multiple temperature (surface, bottom, air, and river) and salinity (surface and bottom) factors were included to account for variation in conditions by depth and location. To avoid double counting within each habitat, only the most appropriate temperature or salinity exposure factor was scored.

The selected exposure factors represent the main climate-driven impacts to the function and viability of habitats in the Northeast U.S. For most of the exposure factors, estimates of projected change were taken from NOAA's Ocean Climate Change Web Portal (https://psl. noaa.gov/ipcc/). Projections were based on the Intergovernmental Panel on Climate Change, Representative Concentration Pathway 8.5 (RCP 8.5), which represents a scenario that assumes little to no stabilization of greenhouse gas emissions over the time horizon for the assessment [99, 100]. We used the results from the downscaled Northwest Atlantic Regional Ocean Modeling System (ROMS-NWA) for ocean temperature and salinity exposure factor projections. Precipitation, air temperature (which was used as a proxy for water temperature in intertidal habitats), and pH were taken from the Coupled Model Intercomparison Project Phase 5 (CMIP5) global model results. For precipitation, experts considered the projected change in the annual mean from CMIP5 as well as information on the projected change in the frequency and intensity of extreme precipitation events (S3 File). Reproducing the method used in NOAA's previous CVAs [81-83, 101], scoring of these model-derived exposure factors was based on a comparison of the future modeled mean to the historic variability as a change in standard deviations. However, unlike these previous CVAs, which were based on mid-century projections, the end-of-century time frame was applied in this study because the structures in many large-scale projects that may impact fish habitats (e.g., bridges and roads, hydropower dams, and major coastal, offshore, and urban development) can be in place over a 50 to 75-year time horizon [38]. In addition, for temporal scales less than 50 years, natural variability contributes substantial uncertainty in climate projections over local and regional spatial scales [102, 103]. For example, the change in global mean sea surface temperature for the

first half of the 21<sup>st</sup> century is similar under most emission scenarios and increasingly diverges later in the century [104]. The end-of-century time frame for ROMS-NWA is 2070–2099, and for CMIP is 2050–2099. The exposure scoring bins from the FSCVA were scaled to appropriately capture the greater changes expected by the end of the century, and to ensure comparability with the species vulnerability assessments.

Projections for SLR were based on the 1.0 m (Intermediate) mean global SLR scenario for 2100 [77], which resulted in a range of SLR between 1.2 and 1.4 m for the study area. Because other projections (ROMS, CMIP) present mean change over a broader period of time (i.e., the latter half of the 21st century), we chose a SLR projection matching the midpoint of the broader projections. The exposure score for SLR was based on both the magnitude of change in sea level and spatial overlap of the relevant habitats with projected change in sea level (S3 File). For example, intertidal habitats were assumed to have greater exposure to SLR than shallow water subtidal habitats.

Because the climate drivers in the riverine system are very different from those in the marine and estuarine systems, we developed a suite of additional exposure factors that are unique to riverine habitats. For example, river flow rates are driven by a complex array of factors (e.g. precipitation, evapotranspiration, groundwater contributions, land use, and anthropogenic water management) [105], such that precipitation alone is not an adequate proxy to capture the impact of climate change on stream and river flow. Therefore, we developed a scoring rubric for flooding and droughts based on a suite of regional streamflow projections [106] (S3 File).

Similarly, air temperature is not a linear predictor of river temperature, which is also influenced by landscape and environmental drivers such as riparian cover, snow melt, and groundwater [107]. River temperature projections were based on data retrieved from the U.S. Geological Survey's (USGS) Spatial Hydro-Ecological Decision System (SHEDS) Stream Temperature Model [107]. Using the +4°C air temperature scenario (an approximation of the endof-century projections in the Northeast U.S.), we used the projected mean summer stream temperature, historic mean summer stream temperature, and the variability of the historic mean stream temperature to develop scoring bins similar to our modeled climate projections for other exposure factors. These data were aggregated by the USGS Watershed Boundary Dataset 6-digit hydrological unit code (HUC6) basins and displayed on a map color coded with the appropriate scoring bins (S3 File). The SHEDS model includes temperature data for first, second, and third order streams only, largely due to greater potential influence of non-climate anthropogenic activities on temperatures in larger rivers. For the purposes of this assessment, we used the aggregated HUC6 stream temperature data for all riverine habitats.

#### Scoring protocol

The scoring for this assessment was based on the protocol described in the FSCVA framework [88]. Fifteen experts participated in the sensitivity scoring, with five experts scoring each of the three systems (marine, estuarine, and riverine). Experts from academia, state, and federal institutions were selected based on recommendations from regional NOAA Fisheries and fishery management council staff to cover a range of geographic and habitat expertise. We conducted a pilot scoring with a subset of scorers and habitats to gather feedback on the process, resulting in improvements in the guidance materials and sensitivity attribute definitions. Scoring was a two-step process similar to the Delphi approach [108, 109]. Experts were first given the opportunity to independently score each sensitivity attribute for each habitat based on the Sensitivity Attribute Definitions (S2 File), a synthesis of information and literature for each habitat. In addition, the experts assigned a data quality score for the sensitivity attributes for each habitat.

the availability of information and uncertainty (Table 4). At the end of this initial round, the scores from each expert were compiled and analyzed for discussion. A second round of scoring was completed at a facilitated workshop in February 2020, where the fifteen experts discussed their individual scores and were given the opportunity to adjust their scores based on group discussions. Many of the habitats were closely related between systems (e.g., intertidal rocky bottom in the marine and estuarine systems), so experts from other systems were encouraged to participate in the discussions for similar habitats. This two-step scoring process necessitated each expert to develop their own scores based on the materials provided and their expert opinion, and then leverage the knowledge and expertise of the other experts shared at the facilitated workshop to inform their final scores. This process alleviates geographic bias or differing interpretations of the materials used during scoring. However, experts were not compelled to change their scores, as consensus was not the aim.

The exposure factors for all habitats were scored by five experts with experience using climate projections in vulnerability assessments. Maps depicting the distribution of each habitat were generated primarily from the Northeast Data Portal (https://www.northeastoceandata. org/) and the Marine Cadastre (https://marinecadastre.gov/). For habitats whose distribution was not well mapped in the study area (e.g., red, green, and small brown algae), or occurred at too fine a scale for spatial comparison with climate projections (e.g., riverine bottom substrate), experts relied on a textual description of the habitat's distribution. Independently, each climate exposure expert visually integrated the habitat distribution maps with the climate projection maps and provided a score for each exposure factor based on the overlap between the habitat distribution and the magnitude of the change in exposure. If an exposure factor was not directly applicable to a habitat (e.g., river temperature is not directly relevant to offshore habitats), the factor was not scored. Each exposure factor was also given a data quality score by each expert to reflect any perceived uncertainties based on the availability and resolution of habitat mapping and the resolution of the modeled climate exposure projections. The resulting exposure scores were compiled and analyzed for discussion with the other scorers. The second round of exposure scoring was completed over a series of webinars where differences in scoring and nuances in the projections and habitat maps were discussed. Experts were then given an opportunity to adjust their scores based on these discussions.

#### Calculating vulnerability ranks

Overall vulnerability ranks for each habitat were calculated in a three-step process developed in the FSCVA framework. The scoring bins for the sensitivity attributes and exposure factors were assigned a numerical value (Low = 1, Moderate = 2, High = 3, Very High = 4). A weighted

Data Quality Score	Description		
3	Adequate Data. The score is based on data which have been observed, modeled or empirically measured for the habitat in question and come from a reputable source.		
2	Limited Data. The score is based on data which has a higher degree of uncertainty. The data used to score the attribute may be based on related or similar habitat, come from outside the study area, or the reliability of the source may be limited.		
1	Expert Judgment. The attribute score reflects the expert judgment of the reviewer and is based on their general knowledge of this attribute for the habitat or a related habitat.		
0	No Data. No information to base an attribute score on. Very little is known about this attribute for this habitat, and there is no basis for forming an expert opinion (to be used judiciously).		

#### Table 4. Data quality matrix.

Criteria used in sensitivity and exposure scoring to identify quality and availability of data.

https://doi.org/10.1371/journal.pone.0260654.t004

Logic Rule	Component Score	Habitat Sensitivity and Exposure Categorical Rank
3 or more attribute or factor means $\geq$ 3.5	4	Very High
2 or more attribute or factor means $\geq 3.0$	3	High
2 or more attribute or factor means $\geq 2.5$	2	Moderate
All other scores	1	Low

Table 5. Logic rule for calculatin	g climate exposure and	l sensitivity for each	ı habitat type

The scoring rubric is based on a logic model where a certain number of individual scores above a certain threshold are used to determine the overall climate exposure and sensitivity (Hare et al. 2016).

https://doi.org/10.1371/journal.pone.0260654.t005

mean for all the tallies across all expert's scores was calculated for each attribute and factor. These attribute and factor means were rolled up into a component score by applying the logic model described in Table 5. The purpose of this logic model is to identify the attributes of highest importance for a habitat. Averages were not used to calculate component scores because the low scores tend to devalue important drivers of climate vulnerability. Note that the criteria for the Very High component score is a difficult threshold to achieve and was designed to indicate that the habitat has multiple very large climate vulnerability drivers. Finally, the component scores were multiplied to provide a categorical vulnerability rank classified as: 1–3 low, 4–6 moderate, 8–9 high, and 12–16 very high.

This methodology utilizes discrete scoring thresholds at several stages while calculating overall vulnerability rank. To detect borderline cases, where the change in just a few tallies in one or more attributes could change the overall vulnerability rank, we conducted a bootstrap analysis for each habitat. The tallies for every attribute and factor were resampled, with replacement, which were then used to recalculate the attribute means, component scores, and vulnerability ranks. Each outcome was recorded and the process repeated 5,000 times. The proportion of the outcomes in each bootstrap vulnerability rank was then compared to the categorical vulnerability rank. This analysis helped identify borderline cases where there was a significant likelihood the habitat could have a higher or lower vulnerability rank. In addition, a leave-one-out sensitivity analysis was performed by leaving out the score for each sensitivity attribute or exposure factor to identify its influence on the overall vulnerability rank across habitats. The same analysis was performed to identify the influence of each scorer on the vulnerability ranks for the system they scored.

An additional analysis was performed on the riverine habitats based on feedback that the classification system may have obscured key interdependencies between riverine habitats. As part of this additional analysis, the individual tallies of the exposure and sensitivity from the riverine water column were combined with the tallies for riverine aquatic bed (algae and submerged aquatic bed), and streambed and bank rocky, sand, and mud bottom habitats. New sensitivity and exposure component scores were calculated based on these combined bed and water column tallies, then the logic model was applied to develop combined vulnerability ranks.

#### Direction of climate effect

After completing the final round of scoring in the February 2020 workshop, the habitat experts were queried on what the overall effect of climate change would be for each habitat (i.e., positive, neutral, or negative). As an example, positive climate effects could include expansion of range, improved condition of the habitat, or reduced habitat fragmentation. Negative climate impacts could include a contraction of range, reduced condition of the habitat, increased

fragmentation, or a loss of critical ecological linkages. Scoring was completed similarly to the sensitivity attribute scoring – each expert was given four tallies to spread amongst three scoring bins. The tallies for each habitat were summed across all experts' scores, and summary statistics were calculated. Positive tallies received a value of 1, neutral received 0, and negative received -1. If the weighted average of the tallies was greater than 0.5, the habitat was classified as likely to be positively affected by climate change; conversely, if the value was less than -0.5, the habitat was classified as likely to be negatively impacted by climate change.

## Habitat narratives

Summaries of the assessment results and information used to score each habitat's climate vulnerability were compiled into habitat narratives (S4 File). These narratives may be informative to end users, as they identify the details of the important drivers of climate sensitivity, exposure, and vulnerability ranks, the key data gaps, relevant non-climate stressors, and overall climate vulnerability of each habitat.

#### Results

#### Overall habitat climate vulnerability

The climate vulnerability ranks for the 52 habitats spanned from Low to Very High (Fig 1 and S5 File). The Low vulnerability rank contained the largest number of habitats (20 habitats; 38%), followed by High (14 habitats; 27%), Moderate (14 habitats; 27%), and Very High (five habitats; 10%).

The marine system had the highest proportion of habitats with High or Very High vulnerability ranks (nine habitats, 39%), followed by seven habitats (37%) in the estuarine system, and three habitats (30%) in the riverine system (Fig 2). The five habitats receiving Very High vulnerability ranks were all living habitats in the marine and estuarine systems. Fifteen of the top twenty most vulnerable habitats were in the living category according to the categorical ranking (Fig 3). Habitats in the artificial structures and invasive categories generally had the lowest vulnerability ranks, none of which ranked greater than Moderate.

The supplemental riverine analysis that combined riverine water column with other riverine habitats yielded two differences from the original results (Table 6). First, when combined with water column scores, riverine rocky bottom changed from Low to Moderate sensitivity and vulnerability. Second, riverine submerged aquatic vegetation changed from High to Moderate sensitivity and vulnerability. The change for submerged aquatic vegetation is counterintuitive given that both riverine water column and submerged aquatic vegetation had High vulnerability ranks in the original analysis, but can be explained by the way the logic model is used to calculate vulnerability. Specifically, different attributes drove the High sensitivity ranks for the two individual habitats such that when combined, the component scores no longer met the threshold for a High sensitivity rank.

#### Sensitivity

Sensitivity ranks for habitats ranged from Low to Very High. One half (26) of the habitats were ranked as Low, 15 were High (29%), nine were Moderate (17%), and only two were Very High (4%). The two habitats with Very High sensitivity ranks (deep sea coral and sponge habitats for both Gulf of Maine and the seamounts and canyons) had multiple High to Very High mean sensitivity attribute scores. The sensitivity attributes that had the strongest influence on climate vulnerability were Sensitivity to Changes in Abiotic Factors and Sensitivity and Intensity of Non-Climate Stressors (Fig 4A). Removal of the Sensitivity to Changes in Abiotic



**Fig 1. Overall climate vulnerability matrix.** Overall climate vulnerability rank is denoted by color: low (green), moderate (yellow), high (orange), and very high (red). Mar = Marine; Est = Estuarine; Riv = Riverine. Categorical ranks used for overall habitat vulnerability in the matrix, and order within each vulnerability cell based on bootstrap rank. Borderline cases from bootstrap results indicated with bold and italics: **bold**  $\geq$ 0.25 probability that the habitat's vulnerability rank is one rank higher than the categorical rank; *italics*  $\geq$ 0.25 probability that the habitat's vulnerability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank; *italics*  $\geq$ 0.25 probability rank is one rank lower than the categorical rank.

https://doi.org/10.1371/journal.pone.0260654.g001

Factors attribute in the leave-one-out sensitivity analysis changed the vulnerability ranks of six habitats, and removal of the Sensitivity and Intensity of Non-Climate Stressors attribute changed the vulnerability ranks of five habitats. Removal of Resilience and Habitat Fragmentation each changed the vulnerability ranks of two habitats. Due to the way the logic model calculates categorical ranks, in every case where the leave-one-out sensitivity analysis changed the vulnerability of a habitat, the vulnerability went down by one rank. Habitat Condition scored High for many of the marine and estuarine nearshore and riverine living habitats (Fig 4B). Dependency on Critical Ecological Linkages generally had the lowest scores of all the sensitivity attributes. Scorers debated the importance and meaning of this attribute, which may have contributed to its low scores. A sensitivity analysis to identify the influence of individual scorers found that no scorers had outsized influence on the vulnerability ranks. Two scorers had slightly greater influence than others, and removal of those two scorers only changed the vulnerability ranks of three habitats each. Overall, the leave-one-out analysis for scorers increased the vulnerability in 13 habitats and decreased the vulnerability in seven (each by one rank).

#### **Climate exposure**

Overall, the climate exposure ranks for habitats tended to be higher than their sensitivity ranks; 37 of the 52 habitats had higher exposure ranks, whereas only three had higher



**Fig 2. Climate vulnerability in the marine, estuarine, and riverine systems.** Number of habitats in the marine (A), estuarine (B), and riverine (C) systems with low, moderate, high, and very high vulnerability ranks. Certainty in rank is denoted by text font and text color: very high certainty (>95%, black font), high certainty (90–95%, black, italic font), moderate certainty (66–90%, white font), low certainty (<66%, white, italic font).

https://doi.org/10.1371/journal.pone.0260654.g002

sensitivity ranks (12 had the same sensitivity and exposure ranks). Although exposure ranks ranged from Low to Very High, most of the habitats received a High rank (30; 58%), followed by Very High (13; 25%), Moderate (8; 15%), and Low (1; 2%). The exposure factors that had the strongest influence on climate vulnerability were pH, Temperature (Air, Surface, Bottom, and River), and SLR (Fig 5A). Removal of pH in the leave-one-out sensitivity analysis changed the vulnerability ranks of 72% of the habitats for which that exposure factor was scored (23 of 32 habitats); removal of Temperature exposure factors changed the vulnerability ranks of 42% of habitats for which those exposure factors were scored (22 of 52 habitats); and removal of SLR changed the vulnerability ranks of 65% of the habitats for which that exposure factor was scored (13 of 20 habitats). In the riverine system, removal of Floods changed the vulnerability rank of five habitats (50%). Precipitation was not highly influential for any of the exposure ranks of all three systems. Bottom temperature had the lowest overall scores across all of the temperature exposure factors (Fig 5B). The sea level rise and two salinity exposure factors had a large spread, reflecting spatial variability in the study area.

#### **Bootstrap analysis**

For the majority of habitats, the bootstrap outcomes matched the categorical vulnerability ranks calculated from the experts' tallies. However, the bootstrap analysis identified ten habitats that had more than 25% of the outcomes in a higher or lower vulnerability rank, and all but two were influenced primarily by the exposure scores. This finding indicated that the climate vulnerability of these habitats are on the borderline between vulnerability ranks. Four of



#### Category

Fig 3. Relative climate vulnerability by category. Proportion of habitats in each category with low, moderate, high, and very high vulnerability ranks.

https://doi.org/10.1371/journal.pone.0260654.g003

#### Table 6. Supplemental riverine analysis.

<b>Riverine Habitat Combinations</b>	Rank			
	Sensitivity	Exposure	Vulnerability	
Algae + water column	Low	High	Low	
Submerged aquatic vegetation + water column	Moderate	High	Moderate	
Rocky bottom + water column	Moderate	High	Moderate	
Mud + water column	Moderate	High	Moderate	
Sand + water column	Moderate	High	Moderate	

https://doi.org/10.1371/journal.pone.0260654.t006



**Fig 4. Sensitivity attributes.** Results of sensitivity analysis for the influence of individual sensitivity attributes on overall vulnerability rank (A) and mean sensitivity scores across all habitats [boxes represent median and interquartile range (IQR), whiskers are values within 1.5\*IQR, and points are potential outliers] (B).

https://doi.org/10.1371/journal.pone.0260654.g004



**Fig 5. Exposure factors.** Results of sensitivity analysis for the influence of individual exposure factors on overall vulnerability rank (A) and mean exposure scores across all habitats (boxes represent median and interquartile range (IQR), whiskers are values within 1.5\*IQR, and points are potential outliers) (B). All four Temperature exposure factors (Air, Surface, Bottom, River) were analyzed together in the sensitivity analysis, since only one temperature exposure factor was scored for each habitat. The same was done for the two Salinity exposure factors (Surface, Bottom).

https://doi.org/10.1371/journal.pone.0260654.g005

these habitats had greater than 50% of the bootstrap outcomes in a different rank, indicating an extreme case of borderline categorical rank (i.e., deep sea coral and sponge: seamounts and canyons, riverine water column, estuarine intertidal rocky bottom, and riverine tidal native wetland). The habitat narratives discuss the specific details for each of these cases (S4 File).

## Direction of climate effect

Based on the experts' qualitative assessment of climate vulnerability, 54% (28) of the habitats are expected to be negatively impacted by climate change, 38% (20) are expected to be



Fig 6. Direction of climate effect by system. Proportion of habitats in each system expected to be positively, neutrally, and negatively affected by climate change.

https://doi.org/10.1371/journal.pone.0260654.g006

neutrally impacted by climate change, and only 8% (4) of the habitats are expected to be positively affected by climate change (S6 File). Most of the marine system habitats were classified as being negatively impacted by climate change, whereas potential impacts to estuarine and riverine system habitats were more mixed (Fig 6). When split by category, the majority of living and water column habitats were expected to be negatively impacted by climate change (Fig 7). The four invasive emergent wetland habitats were the only habitats that were expected to be positively affected by climate change.

#### Discussion

The results of this habitat climate vulnerability assessment revealed a wide range of climate vulnerabilities for marine, estuarine, and riverine habitats in the Northeast U.S. This assessment found that climate exposure had a greater influence than sensitivity on the overall climate vulnerability for most habitats. The climate models used in our assessment projected that a number of climate factors will deviate substantially from the historical variability and range. This is consistent with other climate studies for the Northeast U.S., including projected changes in sea surface temperature [76, 110, 111], sea levels [77], and ocean pH and aragonite saturation state [112, 113], and as reported in other climate vulnerability assessments for this region [81, 114]. Many of the habitats with high sensitivity to physical and chemical changes associated with climate change, including increasing temperature, ocean acidification, and SLR, ranked the highest in overall climate vulnerability. However, even habitats with moderate sensitivity will be vulnerable to climate change given the magnitude of the projected changes.

Projected change in temperature (surface, bottom, air, and riverine) was an important climate driver for nearly all of the habitats evaluated. While the CMIP5 and ROMS-NWA



Fig 7. Direction of climate effect by category. Number of habitats in each category expected to be positively, neutrally, and negatively affected by climate change.

https://doi.org/10.1371/journal.pone.0260654.g007

projections used in this assessment generally show more uniform warming across the U.S. Northeast, regional ocean observations, in the surface and bottom, indicate that the Gulf of Maine and Georges Bank have warmed faster than the Mid-Atlantic Bight [115]. This enhanced warming in New England waters is associated with a northern shift of the Gulf Stream and a weakening of the Labrador Current around the tail end of the Grand Banks [116–118], which is resolved in the high-resolution global climate model CM2.6 [76]. Therefore, marine habitats in portions of the Gulf of Maine, where Gulf Stream associated slope water enters at a greater proportion, and along the shelf break and flanks of Georges Bank may be more vulnerable to climate change than other areas. The CM2.6 model resolves this regional circulation change [76] and is consistent with recent observations indicating a faster and less uniform warming than the ROMS-NWA model, which may have caused the temperature scores for offshore habitats to be biased low in our assessment.

Ocean acidification was also a major factor in the climate exposure for marine and estuarine habitats based on the large projected decrease in pH (20–60 standard deviations from the historical mean). However, the impacts of higher concentrations of  $CO_2$  and acidification for some habitats are generally not well known, and could potentially be advantageous for habitats such as fleshy alga, submerged aquatic vegetation, and emergent wetlands [62, 113, 119–121]. SLR was also an important driver of vulnerability for all intertidal habitats in the marine and estuarine systems. The exposure scores for Droughts and Floods were less important for riverine habitats than temperature overall, which may be a result of the variable patterns of projected change in high and low flow events over the study area [106], whereas the trend for temperature is unidirectional.

#### Patterns in climate vulnerability across habitats

The majority of the habitats that were identified as highly or very highly vulnerable to climate change are living and occur in the estuarine and marine coastal environment, including emergent wetlands, shellfish reefs, subtidal kelp, and submerged aquatic vegetation, and several intertidal habitats in both the estuarine and marine systems. At least 50% of all commercially valuable fish and shellfish in the U.S. depend upon estuaries and nearby coastal waters during one or more life history stages [122], although other reports estimate the dependency as high as 85% [123]. In particular, estuaries provide critical nursery and settlement habitat for many species [122, 124, 125]. Destruction or even a reduction in the condition of important estuarine and coastal habitats can ripple through the food web and lead to decreased abundance of commercially important fish and shellfish species [126–130].

Coastal wetlands, submerged aquatic vegetation, and shellfish reefs minimize coastal flooding, erosion, and runoff, and provide protection to the coastal environment from storm surge and higher sea levels [131, 132]. In addition, because carbon sequestered in the soils of coastal wetlands can be stored for centuries to thousands of years, the loss of coastal wetlands will have significant implications for mitigating greenhouse gas emissions [133, 134]. Stored carbon often is released to the atmosphere when wetlands are destroyed or converted to a different habitat type [135–137], or through increased decomposition due to higher temperature [27, 30]. Although living habitats may have some capacity to adapt to climate change through long-term genetic change, or through short-term acclimatization and phenotypic plasticity, the rate of climate change could exceed the adaptive capacity of many aquatic habitats [32, 138, 139].

Climate change impacts coastal habitats in a multitude of ways. Estuarine and shallow coastal marine water temperatures are influenced by atmospheric warming and solar radiation to a greater degree than oceanic waters. Habitats in the intertidal and shallow subtidal zone are also vulnerable to increased inundation by higher sea levels, erosion, and more frequent and intense coastal storms, leading to physical disruption and conversion of the habitat to a different type (e.g., vegetated to unvegetated, intertidal to subtidal) [25, 52, 140, 141]. In addition, because coastal waters are subject to more sources of low-salinity, acidic waters from rivers and streams, and generally are less buffered than oceanic waters, they are potentially more susceptible to acidification than oceanic waters [31, 113]. More frequent and intense precipitation events can also decrease the salinity and dissolved oxygen conditions in estuarine waters for extended periods of time, such as occurred in the Chesapeake Bay in 2018 and 2019 [142], with implications for fish and shellfish.

Many coastal emergent wetlands in the Northeast U.S. have failed to keep pace with SLR over the past few decades [54, 143–146], leading to erosion and submergence of marsh platforms, and loss of coastal wetland habitat [52, 147]. The mean rate of SLR in the Northeast U. S. over the 20th and 21st centuries has been approximately 2–6 mm per year [148], and could increase to approximately 12–14 mm per year by the end of the century under a 1.0 m global SLR scenario [77]. Some studies in the Northeast U.S. have reported maximum vertical accretion rates for coastal emergent wetlands of around 5–7 mm per year [49, 145, 149, 150], suggesting that many coastal emergent wetlands may become inundated by rising sea levels in the second half of the 21st century. Furthermore, coastal areas hardened by shoreline structures will restrict the capacity of coastal wetlands to migrate inland with increasing SLR [25, 50, 151].

The coastal habitats that were identified as having the highest climate vulnerability are also those most often at risk from degradation due to coastal development. Coastal habitats are threatened by stormwater pollution, eutrophication and general water quality degradation, navigational dredging, shoreline hardening, dredging and filling for coastal development, and the spread of invasive species [37, 38, 42, 152, 153]. Climate-related impacts are likely to exacerbate historic and ongoing degradation of habitats that are already in poor condition from non-climate, anthropogenic impacts [4, 40, 41].

Deep sea corals and sponges found on both seamounts and canyons in the Mid-Atlantic and the Gulf of Maine were the fourth and sixth most vulnerable habitats for all systems, respectively. While deep sea corals and sponges are believed to have lower exposure to many non-climate, anthropogenic effects due to their water depths and distance from shore, these habitats are generally expected to be very sensitive to changes in climate [17, 154, 155], and to anthropogenic activities such as bottom-tending fishing gear. Current observations and historic records suggest that coral habitats were once more extensive in the Gulf of Maine and that current habitat represents refuges that have persisted in the face of intensive bottom fishing, while much of the habitat that was lost has not recovered [66, 96, 156].

Riverine habitats that support diadromous species are experiencing significant climate impacts, including changing hydrology and increasing water temperature. In the past century, stream discharges for rivers with near-natural streamflow in New England and the Mid-Atlantic have generally increased, as have the magnitude and frequency of floods [157–161]. Climate studies that incorporate hydrological models have projected increased variability in streamflow, with greater frequencies of both high-flow and low-flow events predicted for much of the Northeast region [106, 162]. Changes in streamflow magnitude, frequency, and timing can impact riverine habitats and the aquatic species that rely on them [163, 164]. Increases in high flow events can cause stream channel erosion and increased sediment, nutrient, and microbial pathogen delivery to streams, while droughts and decreases in low flow volume can expose aquatic organisms to high temperatures and low dissolved oxygen [105]. Stream temperatures are projected to increase significantly by the end of the century, with the largest increases in the southern Mid-Atlantic and northern New England [107]. These changes make cold- and cool-water rocky-bottom river systems in the Northeast U.S., and the species they support, particularly vulnerable, with implications to food web structure and energy flow in riverine communities [165–169]. In addition, riverine habitats have been historically altered by a host of non-climate perturbations, including damming, water diversion, mineral mining, and storm water pollution [170-173], which can exacerbate climate-related changes in temperature and streamflow.

We considered the vulnerability of several artificial and invasive habitats due to their prevalence in the region and the role they play in providing habitat for some species. The vulnerability ranks for all of these habitats were Moderate and Low. Artificial structures constructed in the subtidal zone include many different materials and purposes, from shoreline protection (e.g., groins, jetties), wrecks, and artificial fishing reefs. Although the materials used in artificial reefs and wrecks are often non-natural (e.g., concrete, steel), they also support biotic communities that can provide ecosystem benefits in some cases. Many of the organisms associated with these structures, particularly mollusks and other shell-forming organisms, are sensitive to changes in temperature and pH. Conversely, artificial structures such as riprap revetments and seawalls constructed for shoreline protection generally support less diverse communities and provide fewer ecological benefits compared to natural shoreline habitats [174–177], and can contain higher occurrences of marine exotic/invasive species compared to native material [178–180].

Four habitats were assessed under the invasive category: two estuarine (New England and Mid-Atlantic) and two riverine (tidal and non-tidal waters) emergent wetland habitat types. These habitats were the only ones in the assessment expected to be positively affected by climate change. The results here are consistent with other studies which suggest invasive species

(e.g., *Phragmites*) may be better adapted to anthropogenic stressors and the effects of climate change, and can out-compete native plant community habitats [42, 181, 182].

#### Management applications and future research

The climate vulnerability of habitats has important implications for the management and protection of fisheries and protected species. Loss, change, or degradation of habitats will impact the species that depend on them. For example, even when a physical habitat may appear to be unchanged, increasing water temperature can impact the water column surrounding it, which in turn can affect the distribution and abundance of associated species, with potential ecosystem-wide effects [37, 42, 183, 184].

Understanding habitat vulnerability can provide a more complete picture of the vulnerability of species. For example, the FSCVA [81] ranked winter flounder as very highly vulnerable to climate change due to low stock status in the southern part of its range and declining population productivity associated with increased nearshore temperature that has been linked to poor stock recruitment. Habitats important to winter flounder—including submerged aquatic vegetation, kelp, intertidal sand and mud, and tidal wetlands [124, 185–188]—are vulnerable to higher air and water temperature, SLR, and habitat fragmentation. The high climate vulnerability of these habitats, and high dependency of winter flounder on these habitats, suggests a potential critical nexus of climate vulnerability for this species. More broadly, the results from this assessment may help fisheries managers better understand ecosystem drivers of species vulnerability, particularly in cases where fish populations, at least in part, are not meeting fishery objectives due to factors other than fishing mortality [7, 39, 41, 42, 189–191].

The results of this assessment can be used by managers in several additional ways, including updates to designations for Essential Fish Habitat (EFH) and Habitat Areas of Particular Concern under the Magnuson-Stevens Fishery Conservation and Management Act, especially as species distributions shift into new areas. They can also be used to support EFH consultations, Endangered Species Act consultations and critical habitat designations, environmental assessments and environmental impact statements prepared under the National Environmental Policy Act, and updates to Fishery Management Plans. Information about climate vulnerability can also be used to prioritize investments in habitat conservation and restoration. Those involved in developing or implementing state wildlife action plans or place-based management plans like the National Estuary Program's Comprehensive Conservation Management Plans may find this assessment useful as it will help identify vulnerable habitats and assist in prioritizing efforts to mobilize conservation action and collaboration. Similarly, these results can inform coastal and ocean planning to minimize impacts on highly vulnerable habitats from nearshore and offshore development, and build coastal resilience. As an example, living shorelines that incorporate vegetation alone or in combination with hardened shoreline structures can serve as an alternative approach to traditional "gray" coastal infrastructure for risk reduction, and may provide additional social and ecological benefits [176, 192-194]. Protecting and conserving riverine, estuarine, and marine habitats not only provides productive, functioning habitat for fish, invertebrate, and protected species populations in the short-term, but also increases the climate resiliency of habitats in the long-term.

One of the goals of this assessment was to develop a framework for habitat climate vulnerability that can be applied to other regions. After applying this framework for habitats in the Northeast U.S. region, several elements may be improved upon in future assessments. Classifying the habitats into meaningful categories required balancing the need to keep the assessment to a manageable number of habitats, but avoiding generalized habitat types that would miss important nuances. For example, the marine artificial structures category included several types of structures (e.g., artificial reefs, wrecks, jetties, riprap, and living shorelines), making it difficult to provide a single sensitivity score. We also chose to score bottom substrate, living, and water column habitats separately, rather than assessing "ecological niches" made up of multiple habitat types (e.g., intertidal rocky bottom with attached kelp). In the riverine system, we did not separate habitats into different thermal regimes or sizes. These decisions may have had implications for the vulnerability ranks of some habitats. For instance, it is documented in the literature that cold water, rocky bottom stream habitats are highly vulnerable to climate change [149, 150, 153], so the low vulnerability rank for rocky bottom habitat may have missed important relationships between water column and substrate, and the different ecological roles of cold- and warm-water riverine habitats. Although we conducted supplemental analyses to better understand the vulnerability of riverine "ecological niche" habitats after the scoring process was complete, future assessments may consider defining riverine habitats differently.

Future assessments may consider additional exposure factors, and synergies between them. This climate vulnerability framework could benefit from a greater consideration of exposure to extreme weather events. While extreme precipitation events were considered, this assessment did not account for exposure to marine heatwaves or other events anticipated to occur as a result of climate change. Further, because climate projections were not readily available for water column stratification [110, 195] and hypoxia [196], we did not evaluate these climate exposure factors, but their impacts on habitat quality may warrant inclusion in future habitat CVAs. Lastly, a number of climate exposure factors, such as warming waters, ocean acidification, and deoxygenation, can interact with one another and with other stressors to cause complex and often unanticipated synergistic climate impacts on habitats in the Northeast U.S. [4, 30, 31, 33]. Assessing potential synergistic and additive climate impacts may be useful for future CVAs.

This assessment applied a broad approach to examining the vulnerability of marine, estuarine, and riverine habitats across the Northeast U.S. to climate change. Yet, climate change often impacts habitats at much smaller scales, and exposure and sensitivity may also vary between watersheds, estuaries, or basins. Thus, the climate vulnerability ranking of some habitats may have been higher or lower had we conducted this assessment on a smaller geographic scale. One of the challenges in a smaller-scale analysis is the resolution of climate models, underscoring the importance of downscaled climate models with better resolution in the nearshore and coastal environments. Finally, this assessment complements prior work to understand the vulnerability of fish and invertebrate species [81] and fishing communities in the Northeast U.S. [114] because climate vulnerabilities of these components of the system are closely linked. For instance, the vulnerability of a habitat influences the vulnerability of species, which in turn influences the vulnerability of communities that rely on those habitats and species. Future assessments should therefore examine the vulnerability of each of these components as a connected social-ecological system rather than as individual, independent parts.

## Supporting information

**S1 File. Habitat classification and definitions.** Descriptions and definitions of each habitat in the assessment.

(PDF)

**S2 File. Sensitivity attribute definitions.** Background, definitions, and scoring bins for each sensitivity attribute. (PDF)

**S3 File. Exposure factor descriptions.** Descriptions and scoring bins for the precipitation, sea level rise, streamflow (droughts and floods), and stream temperature exposure factors. (PDF)

**S4 File. Habitat narratives.** Detailed scoring results for and descriptions of the climate impacts on each habitat including the key drivers of vulnerability, data quality and gaps, and background information on the habitat. (PDF)

**S5 File. Bootstrap and categorical vulnerability natrix.** Table showing the bootstrap and categorical vulnerability ranks of each habitat. (XLSX)

**S6 File. Direction of climate effect.** Table showing the direction of climate effect scores for each habitat. (XLSX)

**Acknowledgments** 

We thank Vince Saba, Michael Alexander, James Scott, and Gaelle Hervieux for assisting with climate projections. We thank Renee Eaton for developing a spatial analysis of stream temperature data, and William Sweet for assisting with SLR projections. We thank Mathias Collins for serving as an expert scorer. We thank Rory Saunders, Page Valentine, Vince Guida, Doug Christel, Keith Hanson, Todd Lutte, and Alison Verkade for assisting in drafting habitat profiles and narratives. We thank Doug Rasher for providing information on kelp distribution, and Ben Letcher for providing stream temperature data. We thank Donna Johnson, Diane Borggaard, Brian Grieve, Rebecca Peters, Janelle Mueller, and Kathy Middleton for their contributions to this project. Finally, we thank Kara Meckley, Roger Griffis, Jessica Coakley, Michelle Bachman, Margaret M. (Peg) Brady, Lou Chiarella, Thomas Noji, Mark Monaco, Tony Marshak, and Kenric Osgood for providing guidance and support throughout the project. Finally, we thank all of the authors for their significant effort to develop the habitat narratives as supplementary material to the manuscript. Acknowledgment of the above individuals does not imply their endorsement of this work; the authors have sole responsibility for the content of this contribution. The scientific results and conclusions, as well as any views or opinions expressed herein, are those of the authors and do not necessarily reflect those of NOAA, the Department of Commerce, or the U.S. Fish and Wildlife Service.

## **Author Contributions**

**Conceptualization:** Michael R. Johnson, Mark W. Nelson, Jonathan A. Hare, Ursula A. Howson.

Formal analysis: Emily R. Farr, Michael R. Johnson, Mark W. Nelson.

- Investigation: Emily R. Farr, Michael R. Johnson, Mark W. Nelson, Wendy E. Morrison, Matthew D. Lettrich, Bruce Vogt, Christopher Meaney, Ursula A. Howson, Peter J. Auster, Frank A. Borsuk, Damian C. Brady, Matthew J. Cashman, Phil Colarusso, Jonathan H. Grabowski, James P. Hawkes, Renee Mercaldo-Allen, David B. Packer, David K. Stevenson.
- Methodology: Emily R. Farr, Michael R. Johnson, Mark W. Nelson, Jonathan A. Hare, Wendy E. Morrison, Matthew D. Lettrich, Bruce Vogt, Christopher Meaney.

Visualization: Emily R. Farr.

- Writing original draft: Emily R. Farr, Michael R. Johnson, Mark W. Nelson, Jonathan A. Hare.
- Writing review & editing: Wendy E. Morrison, Matthew D. Lettrich, Bruce Vogt, Christopher Meaney, Ursula A. Howson, Peter J. Auster, Frank A. Borsuk, Damian C. Brady, Matthew J. Cashman, Phil Colarusso, Jonathan H. Grabowski, James P. Hawkes, Renee Mercaldo-Allen, David B. Packer, David K. Stevenson.

#### References

- 1. Howard J, Babij E, Griffis R, Helmuth B, Himes-Cornell A, Niemier P, et al. Ocean and marine resources in a changing climate. Oceanography and Marine Biology: An Annual Review. 2013; 51:71–192.
- 2. Intergovernmental Panel on Climate Change. Summary for policymakers. Climate Change 2014. Impacts, adaptation, and vulnerability. Part A: Global and sectoral aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Field C, Barros V, Dokken D, Mach K, Mastrandrea M, Bilir T, et al., editors. Cambridge, United Kingdom and New York, NY, USA: Intergovernmental Panel on Climate Change, 2014.
- Nelson EJ, Kareiva P, Ruckelshaus M, Arkema K, Geller G, Girvetz E, et al. Climate change's impact on key ecosystem services and the human well-being they support in the US. Frontiers in Ecology and the Environment. 2013; 11(9):483–893. <u>https://doi.org/10.1890/120312</u>
- 4. Pershing AJ, Griffis RB, Jewett EB, Armstrong CT, Bruno JF, Busch DS, et al. Oceans and marine resources. Impacts, Risks, and Adaptation in the United States: Fourth National Climate Assessment. Reidmiller DR, Avery CW, Easterling DR, Kunkel KE, Lewis KLM, Maycock TK, et al., editors. Washington, DC, USA: U.S. Global Change Research Program, 2018.
- Blanchard JL, Jennings S, Holmes R, Harle J, Merino G, Allen JI, et al. Potential consequences of climate change for primary production and fish production in large marine ecosystems. Philosophical Transactions of the Royal Society B Biological Sciences. 2012; 367(1605):2979–89. https://doi.org/ 10.1098/rstb.2012.0231 PMID: 23007086
- Kjesbu OS, Bogstad B, Devine JA, Gjosaeter H, Howell D, Ingvaldsen RB, et al. Synergies between climate and management for Atlantic cod fisheries at high latitudes. Proceedings of the National Academy of Sciences USA. 2014; 111(9):3478–83. https://doi.org/10.1073/pnas.1316342111 PMID: 24550465
- 7. Pinsky M, Mantua N. Emerging adaptation approaches for climate-ready fisheries management. Oceanography. 2014; 27(4):146–59. https://doi.org/10.5670/oceanog.2014.93
- Kortsch S, Primicerio R, Fossheim M, Dolgov AV, Aschan M. Climate change alters the structure of arctic marine food webs due to poleward shifts of boreal generalists. Proceedings of the Royal Society B. 2015; 282(1814). https://doi.org/10.1098/rspb.2015.1546 PMID: 26336179
- Miller AS, Shepherd GR, Fratantoni PS. Offshore habitat preference of overwintering juvenile and adult black sea bass, *Centropristis striata*, and the relationship to year-class success. PLoS One. 2016; 11(1):e0147627. https://doi.org/10.1371/journal.pone.0147627 PMID: 26824350
- Hare JA, Alexander MA, Fogarty MJ, Williams EH, Scott JD. Forecasting the dynamics of a coastal fishery species using a coupled climate–population model. Ecological Applications. 2010; 20(2):452– 64. https://doi.org/10.1890/08-1863.1 PMID: 20405799
- Hare JA, Manderson JP, Nye JA, Alexander MA, Auster PJ, Borggaard DL, et al. Cusk (*Brosme brosme*) and climate change: assessing the threat to a candidate marine fish species under the US Endangered Species Act. ICES Journal of Marine Science. 2012; 69(10):1753–68. https://doi.org/10.1093/icesjms/fss160
- Jones MC, Cheung WWL. Multi-model ensemble projections of climate change effects on global marine biodiversity. ICES Journal of Marine Science. 2015; 72(3):741–52. https://doi.org/10.1093/ icesjms/fsu172
- Kleisner KM, Fogarty MJ, McGee S, Hare JA, Moret S, Perretti CT, et al. Marine species distribution shifts on the U.S. Northeast Continental Shelf under continued ocean warming. Progress in Oceanography. 2017; 153:24–36. https://doi.org/10.1016/j.pocean.2017.04.001
- Morley JW, Selden RL, Latour RJ, Frölicher TL, Seagraves RJ, Pinsky ML. Projecting shifts in thermal habitat for 686 species on the North American continental shelf. PLoS One. 2018; 13(5):e0196127. https://doi.org/10.1371/journal.pone.0196127 PMID: 29768423
- 15. Allyn AJ, Alexander MA, Franklin BS, Massiot-Granier F, Pershing AJ, Scott JD, et al. Comparing and synthesizing quantitative distribution models and qualitative vulnerability assessments to project

marine species distributions under climate change. PLoS One. 2020; 15(4):e0231595. https://doi.org/ 10.1371/journal.pone.0231595 PMID: 32298349

- Hoegh-Guldberg O, Mumby PJ, Hooten AJ, Steneck RS, Greenfield P, Gomez E, et al. Coral reefs under rapid climate change and ocean acidification. Science. 2007; 318(5857):1737. https://doi.org/ 10.1126/science.1152509 PMID: 18079392
- Morato T, Gonzalez-Irusta J-M, Dominguez-Carrio C, Wei C-L, Davies A, Sweetman AK, et al. Climate-induced changes in the suitable habitat of cold-water corals and commercially important deepsea fishes in the North Atlantic. Global Change Biology. 2020; 26:2181–202. https://doi.org/10.1111/ gcb.14996 PMID: 32077217
- Gazeau F, Quiblier C, Jansen JM, Gattuso J-P, Middelburg JJ, Heip CHR. Impact of elevated CO<sub>2</sub> on shellfish calcification. Geophysical Research Letters. 2007; 34(7). <u>https://doi.org/10.1029/</u> 2006gl028554
- Talmage SC, Gobler CJ. Effects of past, present, and future ocean carbon dioxide concentrations on the growth and survival of larval shellfish. Proceedings of the National Academy of Sciences of the United States of America. 2010; 107(40):17246–51. https://doi.org/10.1073/pnas.0913804107 PMID: 20855590
- Gazeau F, Parker LM, Comeau S, Gattuso J-P, O'Connor WA, Martin S, et al. Impacts of ocean acidification on marine shelled molluscs. Marine Biology. 2013; 160(8):2207–45. <u>https://doi.org/10.1007/s00227-013-2219-3</u>
- Waldbusser GG, Hales B, Langdon CJ, Haley BA, Schrader P, Brunner EL, et al. Ocean acidification has multiple modes of action on bivalve larvae. PLoS One. 2015; 10(6):e0128376. https://doi.org/10. 1371/journal.pone.0128376 PMID: 26061095
- Short FT, Neckles HA. The effects of global climate change on seagrasses. Aquatic Botany. 1999; 63 (3):169–96. https://doi.org/10.1016/S0304-3770(98)00117-X
- Björk M, Short F, Mcleod E, Beer S. Managing seagrasses for resilience to climate change. Gland, Switzerland: The International Union for the Conservation of Nature and Natural Resources (IUCN)/ The Nature Conservancy, 2008. 978-2-8317-1089-1.
- 24. Short FT, Kosten S, Morgan PA, Malone S, Moore GE. Impacts of climate change on submerged and emergent wetland plants. Aquatic Botany. 2016; 135:3–17. <u>https://doi.org/10.1016/j.aquabot.2016</u>. 06.006
- Nicholls RJ, Hoozemans FMJ, Marchand M. Increasing flood risk and wetland losses due to global sea-level rise: regional and global analyses. Global Environmental Change. 1999; 9:S69–S87. <a href="https://doi.org/10.1016/S0959-3780(99)00019-9">https://doi.org/10.1016/S0959-3780(99)00019-9</a>
- Gedan KB, Altieri AH, Bertness MD. Uncertain future of New England salt marshes. Marine Ecology Progress Series. 2011; 434:229–37. https://doi.org/10.3354/meps09084
- Kirwan ML, Mudd SM. Response of salt-marsh carbon accumulation to climate change. Nature. 2012; 489(7417):550–3. https://doi.org/10.1038/nature11440 PMID: 23018965
- Watson EB, Wigand C, Davey EW, Andrews HM, Bishop J, Raposa KB. Wetland loss patterns and inundation-productivity relationships prognosticate widespread salt marsh loss for southern New England. Estuaries and Coasts. 2017; 40(3):662–81. https://doi.org/10.1007/s12237-016-0069-1 PMID: 30008627
- Pörtner HO, Farrell AP. Physiology and climate change. Science. 2008; 322(5902):690–2. https://doi. org/10.1126/science.1163156 PMID: 18974339
- Kirwan ML, Blum LK. Enhanced decomposition offsets enhanced productivity and soil carbon accumulation in coastal wetlands responding to climate change. Biogeosciences. 2011; 8(4):987–93. <a href="https://doi.org/10.5194/bg-8-987-2011">https://doi.org/10.5194/bg-8-987-2011</a>
- Waldbusser GG, Voigt EP, Bergschneider H, Green MA, Newell RIE. Biocalcification in the eastern oyster (*Crassostrea virginica*) in relation to long-term trends in Chesapeake Bay pH. Estuaries and Coasts. 2011; 34(2):221–31. https://doi.org/10.1007/s12237-010-9307-0
- 32. Staudinger MD, Grimm NB, Staudt A, Carter SL, Chapin FS III, Kareiva P, et al. Impacts of climate change on biodiversity, ecosystems, and ecosystem services: technical input to the 2013 National Climate Assessment. Cooperative Report to the 2013 National Climate Assessment. Washington, D.C.: 2012.
- 33. Gobler CJ, DePasquale EL, Griffith AW, Baumann H. Hypoxia and acidification have additive and synergistic negative effects on the growth, survival, and metamorphosis of early life stage bivalves. PLoS One. 2014; 9(1):e83648. https://doi.org/10.1371/journal.pone.0083648 PMID: 24416169
- Auster PJ, Langton RW. The effects of fishing on fish habitat. American Fisheries Society Symposium 22. 1999:150–87.

- Nightingale B, Simenstad C. White paper on dredging activities: Marine issues. Washington Department of Fish and Wildlife, Washington Department of Ecology, Washington Department of Transportation, 2001.
- **36.** National Research Council. Effects of trawling and dredging on seafloor habitat. Washington, DC: The National Academies Press; 2002. 126 p.
- Lotze HK, Lenihan HS, Bourque BJ, Bradbury RH, Cooke RG, Kay MC, et al. Depletion, degradation, and recovery potential of estuaries and coastal seas. Science. 2006; 312(5781):1806–9. https://doi. org/10.1126/science.1128035 PMID: 16794081
- Johnson MR, Boelke C, Chiarella LA, Colosi PD, Greene K, Lellis-Dibble K, et al. Impacts to marine fisheries habitat from nonfishing activities in the northeastern United States. NOAA Technical Memorandum. Woods Hole, MA: National Oceanic and Atmospheric Administration, National Marine Fisheries Service, 2008. NMFS-NE-209.
- Gaichas SK, Seagraves RJ, Coakley JM, DePiper GS, Guida VG, Hare JA, et al. A framework for incorporating species, fleet, habitat, and climate interactions into fishery management. Frontiers in Marine Science. 2016; 3:17. https://doi.org/10.3389/fmars.2016.00105
- 40. U.S. Fish and Wildlife Service and National Oceanic and Atmospheric Administration. National Fish, Wildlife and Plants Climate Adaptation Strategy. Washington, DC: Association of Fish and Wildlife Agencies, Council on Environmental Quality, Great Lakes Indian Fish and Wildlife Commission, National Oceanic and Atmospheric Administration, and U.S. Fish and Wildlife Service, 2012.
- Staudt A, Leidner AK, Howard J, Brauman KA, Dukes JS, Hansen LJ, et al. The added complications of climate change: understanding and managing biodiversity and ecosystems. Frontiers in Ecology and the Environment. 2013; 11(9):494–501. https://doi.org/10.1890/120275
- Smith SL, Cunniff SE, Peyronnin NS, Kritzer JP. Prioritizing coastal ecosystem stressors in the Northeast United States under increasing climate change. Environmental Science & Policy. 2017; 78:49– 57. https://doi.org/10.1016/j.envsci.2017.09.009
- Nye JA, Link JS, Hare JA, Overholtz WJ. Changing spatial distribution of fish stocks in relation to climate and population size on the Northeast United States continental shelf. Marine Ecology Progress Series. 2009; 393:111–29. https://doi.org/10.3354/meps08220
- Pinsky ML, Worm B, Fogarty MJ, Sarmiento JL, Levin SA. Marine taxa track local climate velocities. Science. 2013; 341(6151):1239–42. https://doi.org/10.1126/science.1239352 PMID: 24031017
- Goode AG, Brady DC, Steneck RS, Wahle RA. The brighter side of climate change: How local oceanography amplified a lobster boom in the Gulf of Maine. Global Change Biology. 2019; 25(11):3906– 17. https://doi.org/10.1111/gcb.14778 PMID: 31344307
- Sunday JM, Bates AE, Dulvy NK. Thermal tolerance and the global redistribution of animals. Nature Climate Change. 2012; 2(9):686–90. https://doi.org/10.1038/nclimate1539
- Pörtner HO, Gutt J. Impacts of climate variability and change on (marine) animals: Physiological underpinnings and evolutionary consequences. Integrative and Comparative Biology. 2016; 56 (1):31–44. https://doi.org/10.1093/icb/icw019 PMID: 27371560
- Grimm NB, Chapin FS, Bierwagen B, Gonzalez P, Groffman PM, Luo Y, et al. The impacts of climate change on ecosystem structure and function. Frontiers in Ecology and the Environment. 2013; 11 (9):474–82. https://doi.org/10.1890/120282
- **49.** Donnelly JP, Bertness MD. Rapid shoreward encroachment of salt marsh cordgrass in response to accelerated sea-level rise. Proceedings of the National Academy of Sciences. 2001; 98(25):14218. https://doi.org/10.1073/pnas.251209298
- Kennedy VS, Twilley RR, Kleypas JA, Cowan JHJ, Hare SR. Coastal and marine ecosystems and global climate change: Potential effects on U.S. resources. Arlington, VA: Pew Center on Global Climate Change, 2002.
- Morris JT, Sundareshwar PV, Nietch CT, Kjerfve B, Cahoon DR. Responses of coastal wetlands to rising sea level. Ecology. 2002; 83(10):2869–77. <u>https://doi.org/10.1890/0012-9658(2002)083[2869:</u> ROCWTR]2.0.CO;2
- Scavia D, Field JC, Boesch DF, Buddemeier RW, Burkett V, Cayan DR, et al. Climate change impacts on US coastal and marine ecosystems. Estuaries. 2002; 25(2):149–64. <u>https://doi.org/10.1007/ Bf02691304</u>
- Andersen TJ, Svinth S, Pejrup M. Temporal variation of accumulation rates on a natural salt marsh in the 20th century—The impact of sea level rise and increased inundation frequency. Marine Geology. 2011; 279(1–4):178–87. https://doi.org/10.1016/j.margeo.2010.10.025
- Crosby SC, Sax DF, Palmer ME, Booth HS, Deegan LA, Bertness MD, et al. Salt marsh persistence is threatened by predicted sea-level rise. Estuarine, Coastal and Shelf Science. 2016; 181:93–9. <u>https:// doi.org/10.1016/j.ecss.2016.08.018</u>

- Minello TJ, Able KW, Weinstein MP, Hays CG. Salt marshes as nurseries for nekton: Testing hypotheses on density, growth and survival through meta-analysis. Marine Ecology Progress Series. 2003; 246:39. https://doi.org/10.3354/meps246039
- 56. Boesch DF, Turner RE. Dependence of fishery species on salt marshes: The role of food and refuge. Estuaries. 1984; 7(4):460–8. https://doi.org/10.2307/1351627
- Wiest WA, Correll MD, Olsen BJ, Elphick CS, Hodgman TP, Curson DR, et al. Population estimates for tidal marsh birds of high conservation concern in the northeastern USA from a design-based survey. The Condor. 2016; 118(2):274–88. https://doi.org/10.1650/condor-15-30.1
- Steneck RS, Graham MH, Bourque BJ, Corbett D, Erlandson JM, Estes JA, et al. Kelp forest ecosystems: biodiversity, stability, resilience and future. Environmental Conservation. 2002; 29(4):436–59.
- Witman JD, Lamb RW. Persistent differences between coastal and offshore kelp forest communities in a warming Gulf of Maine. PLoS One. 2018; 13(1):e0189388. https://doi.org/10.1371/journal.pone. 0189388 PMID: 29298307
- Orr JC, Fabry VJ, Aumont O, Bopp L, Doney SC, Feely RA, et al. Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. Nature. 2005; 437(7059):681–6. https://doi.org/10.1038/nature04095 PMID: 16193043
- Doney SC, Fabry VJ, Feely RA, Kleypas JA. Ocean acidification: the other CO<sub>2</sub> problem. Annual Review of Marine Science. 2009; 1:169–92. https://doi.org/10.1146/annurev.marine.010908.163834 PMID: 21141034
- Kroeker KJ, Kordas RL, Crim R, Hendriks IE, Ramajo L, Singh GS, et al. Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. Global Change Biology. 2013; 19(6):1884–96. https://doi.org/10.1111/gcb.12179 PMID: 23505245
- Cooley SR, Kite-Powell HL, Doney SC. Ocean acidification's potential to alter global marine ecosystem services. Oceanography. 2009; 22(4):172–81.
- Hoegh-Guldberg O, Poloczanska ES, Skirving W, Dove S. Coral reef ecosystems under climate change and ocean acidification. Frontiers in Marine Science. 2017; 4(158):1–20. <u>https://doi.org/10.3389/fmars.2017.00158</u>
- Auster PJ. Are deep-water corals important habitats for fishes? In: Freiwald A, Roberts JM, editors. Cold-Water Corals and Ecosystems. Erlangen Earth Conference. Berlin, Heidelberg: Springer-Verlag; 2005. p. 643–56.
- 66. Auster P, Packer D, Waller R, Auscavitch S, Kilgour M, Watling L, et al. Imaging surveys of select areas in the northern Gulf of Maine for deep-sea corals and sponges during 2013–2014. Report to the New England Fishery Management Council—1 December 2014. New England Fishery Management Council, 2015.
- Henderson MJ, Huff DD, Yoklavich MM. Deep-sea coral and sponge taxa increase demersal fish diversity and the probability of fish presence. Frontiers in Marine Science. 2020; 7:1–19. <u>https://doi.org/10.3389/fmars.2020.00548</u> PMID: 32802822
- Lyons MP, von Holle B, Caffrey MA, Weishampel JF. Quantifying the impacts of future sea level rise on nesting sea turtles in the southeastern United States. Ecological Applications. 2020; 30(5):e02100. https://doi.org/10.1002/eap.2100 PMID: 32086969
- Woodson CB, Litvin SY. Ocean fronts drive marine fishery production and biogeochemical cycling. Proceedings of the National Academy of Sciences of the United States of America. 2015; 112 (6):1710–5. https://doi.org/10.1073/pnas.1417143112
- Santora JA, Hazen EL, Schroeder ID, Bograd SJ, Sakuma KM, Field JC. Impacts of ocean climate variability on biodiversity of pelagic forage species in an upwelling ecosystem. Marine Ecology Progress Series. 2017; 580:205–20. https://doi.org/10.3354/meps12278
- 71. Galbraith H, Jones R, Park R, Clough J, Herrod-Julius S, Harrington B, et al. Global climate change and sea level rise: potential losses of intertidal habitat for shorebirds. Waterbirds. 2002; 25(2):173–83. https://doi.org/10.1675/1524-4695(2002)025[0173:Gccasl]2.0.Co;2
- 72. Von Holle B, Irish JL, Spivy A, Weishampel JF, Meylan A, Godfrey MH, et al. Effects of future sea level rise on coastal habitat. The Journal of Wildlife Management. 2019; 83(3):694–704. <u>https://doi.org/10.1002/jwmg.21633</u>
- 73. Wu L, Cai W, Zhang L, Nakamura H, Timmermann A, Joyce T, et al. Enhanced warming over the global subtropical western boundary currents. Nature Climate Change. 2012; 2(3):161–6. https://doi. org/10.1038/nclimate1353
- 74. Friedland KD, Morse RE, Manning JP, Melrose DC, Miles T, Goode AG, et al. Trends and change points in surface and bottom thermal environments of the US Northeast Continental Shelf Ecosystem. Fisheries Oceanography. 2020; 29(5):396–414. https://doi.org/10.1111/fog.12485

- Karmalkar AV, Horton RM. Drivers of exceptional coastal warming in the northeastern United States. Nature Climate Change. 2021; 11(10):854–60. https://doi.org/10.1038/s41558-021-01159-7
- 76. Saba VS, Griffies SM, Anderson WG, Winton M, Alexander MA, Delworth TL, et al. Enhanced warming of the Northwest Atlantic Ocean under climate change. Journal of Geophysical Research: Oceans. 2016; 121(1):118–32. https://doi.org/10.1002/2015jc011346
- 77. Sweet WV, Kopp RE, Weaver CP, Obeysekera J, Horton RM, Thieler ER, et al. Global and regional sea level rise scenarios for the United States. Silver Spring, Maryland: National Oceanic and Atmospheric Administration, National Ocean Service, 2017. NOAA Technical Report NOS CO-OPS 083.
- 78. Williams SE, Shoo LP, Isaac JL, Hoffmann AA, Langham G. Towards an integrated framework for assessing the vulnerability of species to climate change. PLoS Biology. 2008; 6(12):e325. <u>https://doi.org/10.1371/journal.pbio.0060325</u> PMID: 19108608
- Glick P, Stein BA, Edelson NA. Scanning the conservation horizon: a guide to climate change vulnerability assessment. Glick P, Stein BA, Edelson NA, editors. Washington, D.C.: National Wildlife Federation, 2011.
- Foden WB, Butchart SHM, Stuart SN, Vié J-C, Akcakaya HR, Angulo A, et al. Identifying the world's most climate change vulnerable species: A systematic trait-based assessment of all birds, amphibians and corals. PLoS One. 2013; 8(6):e65427. <u>https://doi.org/10.1371/journal.pone.0065427</u> PMID: 23950785
- Hare JA, Morrison WE, Nelson MW, Stachura MM, Teeters EJ, Griffis RB, et al. A vulnerability assessment of fish and invertebrates to climate change on the Northeast U.S. Continental Shelf. PLoS One. 2016; 11(2):e0146756. https://doi.org/10.1371/journal.pone.0146756 PMID: 26839967
- Crozier LG, McClure MM, Beechie T, Bograd SJ, Boughton DA, Carr M, et al. Climate vulnerability assessment for Pacific salmon and steelhead in the California Current Large Marine Ecosystem. PLoS One. 2019; 14(7):e0217711. https://doi.org/10.1371/journal.pone.0217711 PMID: 31339895
- Spencer PD, Hollowed AB, Sigler MF, Hermann AJ, Nelson MW. Trait-based climate vulnerability assessments in data-rich systems: An application to eastern Bering Sea fish and invertebrate stocks. Global Change Biology. 2019; 25(11):3954–71. https://doi.org/10.1111/gcb.14763 PMID: 31531923
- Sousa A, Alves F, Arranz P, Dinis A, Fernandez M, Gonzalez Garcia L, et al. Climate change vulnerability of cetaceans in Macaronesia: Insights from a trait-based assessment. Science of the Total Environment. 2021; 795:148652. https://doi.org/10.1016/j.scitotenv.2021.148652 PMID: 34247086
- 85. Manomet Center for Conservation Sciences and National Wildlife Federation. The vulnerabilities of fish and wildlife habitats in the northeast to climate change. A report to the Northeastern Association of Fish and Wildlife Agencies and the North Atlantic Landscape Conservation Cooperative. Galbraith H, editor. Manomet, MA: 2013.
- Comer P. Assessing climate change for landscapes and major vegetation types across the intermountain west [Internet]. Conservation Biology Institute; 2017. [updated 2019; cited 3 November 2021]. Available from: https://databasin.org/articles/1f0601a2715c44349fa1289807c468d6/.
- Teck S, Halpern B, Kappel C, Micheli F, Selkoe K, Crain C, et al. Using expert judgment to estimate marine ecosystem vulnerability in the California Current. Ecological Applications. 2010; 20:1402–16. https://doi.org/10.1890/09-1173.1 PMID: 20666257
- Morrison WE, Nelson MW, Howard JF, Teeters EJ, Hare JA, Griffis RB, et al. Methodology for assessing the vulnerability of marine fish and shellfish species to a changing climate. Technical Memorandum. U.S. Department of Commerce, National Oceanic and Atmospheric Administration, National Marine Fisheries Service, 2015. NMFS-OSF-3.
- Fortini L, Schubert O. Beyond exposure, sensitivity and adaptive capacity: a response based ecological framework to assess species climate change vulnerability. Climate Change Responses. 2017; 4 (1):1–7. https://doi.org/10.1186/s40665-017-0030-y
- Rosenberg A, Bigford TE, Leathery S, Hill RL, Bickers K. Ecosystem approaches to fishery management through essential fish habitat. Bulletin of Marine Science. 2000; 66(3):535–42.
- National Oceanic and Atmospheric Administration. NOAA National Habitat Policy. Department of Commerce, National Oceanic and Atmospheric Administration, 2015.
- 92. Federal Geographic Data Committee. Classification of wetlands and deepwater habitats of the United States. Adapted from Cowardin, Carter, Golet and LaRoe (1979). Washington, DC: Wetlands Subcommittee, Federal Geographic Data Committee and U.S. Fish and Wildlife Service, 2013. August 2013. FGDC-STD-004-2013.
- Cowardin LM, Carter V, Golet FC, LaRoe ET. Classification of wetlands and deepwater habitats of the United States. Washington, D.C.: U.S. Department of the Interior, Fish and Wildlife Service, Office of Biological Services, 1979. FWS/OBS-79/31.

- 94. Packer DB, Boelke D, Guida V, McGee L-A. State of deep coral ecosystems in the Northeastern US region: Maine to Cape Hatteras. The state of deep coral ecosystems of the United States. Lumsden SE, Hourigan TF, Bruckner AW, Dorr G, editors. Silver Spring, MD: National Oceanic and Atmospheric Administration, 2007. CRCP-3.
- Auster PJ, Kilgour M, Packer D, Waller R, Auscavitch S, Watling L. Octocoral gardens in the Gulf of Maine (NW Atlantic). Biodiversity. 2013; 14(4):193–4. <u>https://doi.org/10.1080/14888386.2013</u>. 850446
- Auster PJ, Packer D, Kilgour M, Watling L. Supplementary comment: conservation of deep-sea corals off the northeast United States. Biodiversity. 2013; 14(4):195–. <u>https://doi.org/10.1080/14888386</u>. 2013.850885
- **97.** Packer DB, Nizinski MS, Bachman MS, Drohan AF, Poti M, Kinlan BP. State of deep-sea coral and sponge ecosystems of the Northeast U.S. The state of deep-sea coral and sponge ecosystems of the United States. Hourigan T, Etnoyer PJ, Cairns SD, editors. Silver Spring, MD: National Oceanic and Atmospheric Administration, 2017. NMFS-OHC-4.
- Packer DB, Nizinski MS, Cairns SD, Hourigan TF. Deep-sea coral taxa in the U.S. Northeast Region: depth and geographical distribution (v. 2020) [Internet]. 2017. [updated 2020; cited 8 Dec. 2020]. Available from: https://deepseacoraldata.noaa.gov/NOAA\_SpeciesList\_Northeast\_Packer-et-al\_1-15-2021.pdf.
- 99. Riahi K, Rao S, Krey V, Cho C, Chirkov V, Fischer G, et al. RCP 8.5—A scenario of comparatively high greenhouse gas emissions. Climatic Change. 2011; 109(1–2):33–57. <u>https://doi.org/10.1007/s10584-011-0149-y</u>
- 100. van Vuuren DP, Edmonds J, Kainuma M, Riahi K, Thomson A, Hibbard K, et al. The representative concentration pathways: an overview. Climatic Change. 2011; 109(1–2):5–31. https://doi.org/10. 1007/s10584-011-0148-z
- 101. Lettrich MD, Asaro MJ, Borggaard DL, Dick DM, Griffis RB, Litz JA, et al. A method for assessing the vulnerability of marine mammals to a changing climate. NOAA Technical Memorandum. Silver Spring, MD: U.S. Department of Commerce National Oceanic and Atmospheric Administration National Marine Fisheries Service, 2019. NMFS-F/SPO-196.
- 102. Stock CA, Alexander MA, Bond NA, Brander KM, Cheung WWL, Curchitser EN, et al. On the use of IPCC-class models to assess the impact of climate on living marine resources. Progress in Oceanography. 2011; 88(1–4):1–27. https://doi.org/10.1016/j.pocean.2010.09.001
- 103. Deser C, Knutti R, Solomon S, Phillips AS. Communication of the role of natural variability in future North American climate. Nature Climate Change. 2012; 2(11):775–9. <u>https://doi.org/10.1038/nclimate1562</u>
- 104. Kirtman B, Power SB, Adedoyin AJ, Boer GJ, Bojariu R, Camilloni I, et al. Near-term climate change: projections and predictability. Climate change 2013: the physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Stocker TF, Qin D, Plattner G-K, Tignor M, Allen SK, Boschung J, et al., editors. Cambridge, United Kingdom and New York, NY, USA: Intergovernmental Panel on Climate Change, 2013.
- 105. U.S. Global Change Research Program. Climate Science Special Report: Fourth National Climate Assessment. Volume I. Wuebbles DJ, Fahey DW, Hibbard KA, Dokken DJ, Stewart BC, Maycock TK, editors. Washington, DC. USA: U.S. Global Change Research Program, 2017.
- 106. Demaria EMC, Palmer RN, Roundy JK. Regional climate change projections of streamflow characteristics in the Northeast and Midwest U.S. Journal of Hydrology: Regional Studies. 2016; 5:309–23. https://doi.org/10.1016/j.ejrh.2015.11.007
- Letcher BH, Hocking DJ, O'Neil K, Whiteley AR, Nislow KH, O'Donnell MJ. A hierarchical model of daily stream temperature using air-water temperature synchronization, autocorrelation, and time lags. PeerJ. 2016; 4:e1727. https://doi.org/10.7717/peerj.1727 PMID: 26966662
- 108. Linstone HA, Turoff M. The Delphi Method: Techniques and Applications. Reading, MA: Addison-Wesley Publishing Company; 2002.
- Hsu C-C, Sandford BA. The Delphi technique: making sense of consensus. Practical Assessment, Research, and Evaluation. 2007; 12. https://doi.org/10.7275/pdz9-th90
- Khan AH, Levac E, Chmura GL. Future sea surface temperatures in Large Marine Ecosystems of the Northwest Atlantic. ICES Journal of Marine Science. 2013; 70(5):915–21. <u>https://doi.org/10.1093/ icesjms/fst002</u>
- 111. Henson SA, Beaulieu C, Ilyina T, John JG, Long M, Séférian R, et al. Rapid emergence of climate change in environmental drivers of marine ecosystems. Nature Communications. 2017; 8:1–9. <u>https:// doi.org/10.1038/s41467-016-0009-6</u> PMID: 28232747
- Feely RA, Doney SC, Cooley SR. Ocean acidification: present conditions and future changes in a high-CO2 world. Oceanography. 2009; 22(4):36–47. https://doi.org/10.5670/oceanog.2009.95

- Gledhill D, White M, Salisbury J, Thomas H, Misna I, Liebman M, et al. Ocean and coastal acidification off New England and Nova Scotia. Oceanography. 2015; 25(2):182–97. https://doi.org/10.5670/ oceanog.2015.41
- 114. Colburn LL, Jepson M, Weng C, Seara T, Weiss J, Hare JA. Indicators of climate change and social vulnerability in fishing dependent communities along the Eastern and Gulf Coasts of the United States. Marine Policy. 2016; 74:323–33. https://doi.org/10.1016/j.marpol.2016.04.030
- **115.** National Oceanic and Atmospheric Administration. 2021 State of the Ecosystem: New England. Department of Commerce. NOAA, 2021.
- 116. Gawarkiewicz GG, Todd RE, Plueddemann AJ, Andres M, Manning JP. Direct interaction between the Gulf Stream and the shelfbreak south of New England. Scientific Reports. 2012; 2:553. <u>https://doi.org/ 10.1038/srep00553</u> PMID: 22870382
- 117. Pershing AJ, Alexander MA, Hernandez CM, Kerr LA, Le Bris A, Mills KE, et al. Slow adaptation in the face of rapid warming leads to collapse of the Gulf of Maine cod fishery. Science. 2015; 350(6262):1–8. https://doi.org/10.1126/science.aac9819 PMID: 26516197
- Neto AG, Langan JA, Palter JB. Changes in the Gulf Stream preceded rapid warming of the Northwest Atlantic Shelf. Communications Earth & Environment. 2021; 2(1). <u>https://doi.org/10.1038/s43247-021-00143-5</u>
- Cherry JA, McKee KL, Grace JB. Elevated CO<sub>2</sub> enhances biological contributions to elevation change in coastal wetlands by offsetting stressors associated with sea-level rise. Journal of Ecology. 2009; 97 (1):67–77. https://doi.org/10.1111/j.1365-2745.2008.01449.x
- Koch M, Bowes G, Ross C, Zhang X-H. Climate change and ocean acidification effects on seagrasses and marine macroalgae. Global Change Biology. 2013; 19(1):103–32. <u>https://doi.org/10.1111/j.1365-2486.2012.02791.x PMID: 23504724</u>
- 121. Langley JA, McKee KL, Cahoon DR, Cherry JA, Megonigal JP. Elevated CO2 stimulates marsh elevation gain, counterbalancing sea-level rise. Proceedings of the National Academy of Sciences of the United States of America. 2009; 106(15):6182–6. https://doi.org/10.1073/pnas.0807695106
- 122. Lellis-Dibble KA, McGlynn KE, Bigford TE. Estuarine fish and shellfish species in U.S. commercial and recreational fisheries: economic value as an incentive to protect and restore estuarine habitat. NOAA Technical Memorandum. Silver Spring, MD: U.S. Department of Commerce, 2008. November 2008. NMFSF/SPO-90.
- 123. National Research Council. Striking a balance: Improving stewardship of marine areas. Washington, DC: National Research Council, 1997. 0-309-06369-8.
- 124. Ayvazian SG, Deegan LA, Finn JT. Comparison of habitat use by estuarine fish assemblages in the Acadian and Virginian Zoogeographic Provinces. Estuaries. 1992; 15(3):368–83. https://doi.org/10. 2307/1352784
- 125. Able KW. A re-examination of fish estuarine dependence: Evidence for connectivity between estuarine and ocean habitats. Estuarine, Coastal and Shelf Science. 2005; 64(1):5–17. <u>https://doi.org/10.1016/j.ecss.2005.02.002</u>
- 126. National Research Council. Striking a balance: improving stewardship of narine areas. Committee on Marine Area Governance and Management MB, Commission on Engineering and Technical Systems, editor. Washington, D.C.: National Academy Press; 1997. 177 p.
- 127. Nicholls RJ. Coastal flooding and wetland loss in the 21st century: changes under the SRES climate and socio-economic scenarios. Global Environmental Change. 2004; 14(1):69–86. <u>https://doi.org/10.1016/j.gloenvcha.2003.10.007</u>
- 128. Valiela I, Rutecki D, Fox S. Salt marshes: biological controls of food webs in a diminishing environment. Journal of Experimental Marine Biology and Ecology. 2004; 300(1–2):131–59. <u>https://doi.org/10.1016/j.jembe.2003.12.023</u>
- 129. Shellenbarger Jones A, Bosch C, Strange E. Vulnerable species: the effects of sea-level rise on coastal habitats. Coastal sensitivity to sea-level rise: a focus on the Mid-Atlantic Region. Titus JG, Anderson KE, Cahoon DR, Gesch DB, Gill SK, Gutierrez BT, et al., editors. U.S. Climate Change Science Program, 2009.
- DePasquale EL, Baumann H, Gobler CJ. Vulnerability of early life stage Northwest Atlantic forage fish to ocean acidification and low oxygen. Marine Ecology Progress Series. 2015; 523:145–56. <u>https:// doi.org/10.3354/meps11142</u>
- **131.** Gedan KB, Kirwan ML, Wolanski E, Barbier EB, Silliman BR. The present and future role of coastal wetland vegetation in protecting shorelines: answering recent challenges to the paradigm. Climatic Change. 2011; 106(1):7–29. https://doi.org/10.1007/s10584-010-0003-7

- 132. Möller I, Kudella M, Rupprecht F, Spencer T, Paul M, van Wesenbeeck BK, et al. Wave attenuation over coastal salt marshes under storm surge conditions. Nature Geoscience. 2014; 7(10):727–31. https://doi.org/10.1038/ngeo2251
- Duarte CM, Middelburg JJ, Caraco N. Major role of marine vegetation on the oceanic carbon cycle. Biogeosciences. 2005; 2(1):1–8. https://doi.org/10.5194/bg-2-1-2005
- 134. Howard J, Sutton-Grier A, Herr D, Kleypas J, Landis E, McLeod E, et al. Clarifying the role of coastal and marine systems in climate mitigation. Frontiers in Ecology and the Environment. 2017; 15(1):42– 50. https://doi.org/10.1002/fee.1451
- Chmura GL, Anisfeld SC, Cahoon DR, Lynch JC. Global carbon sequestration in tidal, saline wetland soils. Global Biogeochemical Cycles. 2003; 17(4):1–12. https://doi.org/10.1029/2002gb001917
- **136.** Duarte CM, Cebrián J. The fate of marine autotrophic production. Limnology and Oceanography. 1996; 41(8):1758–66. https://doi.org/10.4319/lo.1996.41.8.1758
- 137. Pendleton L, Donato DC, Murray BC, Crooks S, Jenkins WA, Sifleet S, et al. Estimating global "blue carbon" emissions from conversion and degradation of vegetated coastal ecosystems. PLoS One. 2012; 7(9):e43542. https://doi.org/10.1371/journal.pone.0043542 PMID: 22962585
- Hoffmann AA, Sgro CM. Climate change and evolutionary adaptation. Nature. 2011; 470(7335):479– 85. <u>https://doi.org/10.1038/nature09670</u> PMID: <u>21350480</u>
- 139. Pörtner H-O, Karl DM, Boyd PW, Cheung WWL, Lluch-Cota SE, Nojiri Y, et al. Ocean systems. Climate change 2014. Impacts, adaptation, and vulnerability. Part A: global and sectoral aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Field CB, Barros VR, Dokken DJ, Mach KJ, Mastrandrea MD, Bilir TE, et al., editors. Cambridge, United Kingdom and New York, NY, USA: Intergovernmental Panel on Climate Change, 2014.
- 140. Brown AC, McLachlan A. Sandy shore ecosystems and the threats facing them: some predictions for the year 2025. Environmental Conservation. 2002; 29(1):62–77. <u>https://doi.org/10.1017/</u> S037689290200005X
- 141. Nicholls RJ, Wong PP, Burkett V, Codignotto J, Hay J, McLean R, et al. Coastal systems and low-lying areas. Climate change 2007: impacts, adaptation and vulnerability. Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Parry ML, Canziani OF, Palutikof JP, van der Linden PJ, Hanson CE, editors. Cambridge, UK: Intergovernmental Panel on Climate Change, 2007.
- 142. National Oceanic and Atmospheric Administration. 2020 State of the Ecosystem: Mid-Atlantic. Department of Commerce. NOAA, 2020.
- 143. Kearney MS, Rogers AS, Townshend JRG, Rizzo E, Stutzer D, Stevenson JC, et al. Landsat imagery shows decline of coastal marshes in Chesapeake and Delaware Bays. Eos, Transactions American Geophysical Union. 2002; 83(16):173–8. https://doi.org/10.1029/2002EO000112
- 144. Kolker AS, Kirwan ML, Goodbred SL, Cochran JK. Global climate changes recorded in coastal wetland sediments: Empirical observations linked to theoretical predictions. Geophysical Research Letters. 2010; 37(14):5. https://doi.org/10.1029/2010gl043874
- 145. Carey JC, Moran SB, Kelly RP, Kolker AS, Fulweiler RW. The declining role of organic matter in New England salt marshes. Estuaries and Coasts. 2017; 40(3):626–39. https://doi.org/10.1007/s12237-015-9971-1
- 146. Watson EB, Raposa KB, Carey JC, Wigand C, Warren RS. Anthropocene survival of southern New England's salt marshes. Estuaries and Coasts. 2017; 40(3):617–25. https://doi.org/10.1007/s12237-016-0166-1 PMID: 30271312
- Nicholls RJ, Cazenave A. Sea-level rise and its impact on coastal zones. Science. 2010; 328 (5985):1517–20. https://doi.org/10.1126/science.1185782 PMID: 20558707
- 148. National Oceanic and Atmospheric Administration. NOAA Tides and Currents: Sea Level Trends [Internet]. National Oceanic and Atmospheric Administration; 2021. [updated 2021; cited 1 Apr. 2021]. Available from: https://tidesandcurrents.noaa.gov/sltrends/sltrends.html.
- 149. Cahoon DR, Reed DJ, Kolker AS, Brinson MM, Stevenson JC, Riggs S, et al. Coastal wetland sustainability. Coastal Sensitivity to Sea-Level Rise: A Focus on the Mid-Atlantic Region. Titus JG, Anderson KE, Cahoon DR, Gesch DB, Gill SK, Gutierrez BT, et al., editors. Washington, D.C.: Report by the U. S. Climate Change Science Program and the Subcommittee on Global Change Research, 2009. EPA 430-R-09-023: Synthesis and Assessment Product 4.1.
- **150.** Kirwan ML, Guntenspergen GR, D'Alpaos A, Morris JT, Mudd SM, Temmerman S. Limits on the adaptability of coastal marshes to rising sea level. Geophysical Research Letters. 2010; 37(23):1–5. https://doi.org/10.1029/2010gl045489

- 151. Titus JG, Hudgens DE, Trescott DL, Craghan M, Nuckols WH, Hershner CH, et al. State and local governments plan for development of most land vulnerable to rising sea level along the US Atlantic coast. Environmental Research Letters. 2009; 4(4):1–7. https://doi.org/10.1088/1748-9326/4/4/044008
- 152. Deegan LA, Buchsbaum R. Chapter 5. The effect of habitat loss and degradation on fisheries. In: Buchsbaum R, Pederson J, Robinson WE, editors. The decline on fisheries resources in New England: evaluating the impact of overfishing, contamination, and habitat degradation. Cambridge, Massachusetts: Massachusetts Institute of Technology Sea Grant College Program; 2005. p. 67–96.
- 153. Kennish MJ, Brush MJ, Moore KA. Drivers of change in shallow coastal photic systems: an introduction to a special issue. Estuaries and Coasts. 2014; 37(S1):3–19. <u>https://doi.org/10.1007/s12237-</u> 014-9779-4
- 154. Guinotte JM, Orr J, Cairns S, Freiwald A, Morgan L, George R. Will human-induced changes in seawater chemistry alter the distribution of deep-sea scleractinian corals? Frontiers in Ecology and the Environment. 2006; 4(3):141–6. https://doi.org/10.1890/1540-9295(2006)004[0141:WHCISC]2.0.CO;2
- 155. Fabry VJ, Seibel BA, Feely RA, Orr JC. Impacts of ocean acidification on marine fauna and ecosystem processes. ICES Journal of Marine Science. 2008; 65(3):414–32. <u>https://doi.org/10.1093/icesjms/</u> fsn048
- 156. Watling L, Auster PJ. Distribution of deep-water Alcyonacea off the Northeast Coast of the United States. In: Freiwald A, Roberts JM, editors. Cold-Water Corals and Ecosystems. Berlin, Heidelberg: Springer Berlin Heidelberg; 2005. p. 279–96.
- 157. Hodgkins GA, Dudley RW. Changes in the magnitude of annual and monthly streamflows in New England, 1902–2002. Reston, Virginia: U.S. Department of the Interior, U.S. Geological Survey, 2005. Scientific Investigations Report 2005–5135.
- **158.** Lins HF, Slack JR. Streamflow trends in the United States. Geophysical Research Letters. 1999; 26 (2):227–30. https://doi.org/10.1029/1998gl900291
- **159.** Collins MJ. Evidence for changing flood risk in New England since the late 20th century. Journal of the American Water Resources Association. 2009; 45(2):279–90. <u>https://doi.org/10.1111/j.1752-1688</u>. 2008.00277.x
- Armstrong WH, Collins MJ, Snyder NP. Hydroclimatic flood trends in the northeastern United States and linkages with large-scale atmospheric circulation patterns. Hydrological Sciences Journal. 2014; 59(9):1636–55. https://doi.org/10.1080/02626667.2013.862339
- 161. Armstrong WH, Collins MJ, Snyder NP. Increased frequency of low-magnitude floods in New England. JAWRA Journal of the American Water Resources Association. 2012; 48(2):306–20. <u>https://doi.org/10.1111/j.1752-1688.2011.00613.x</u>
- 162. Hayhoe K, Wake CP, Huntington TG, Luo L, Schwartz MD, Sheffield J, et al. Past and future changes in climate and hydrological indicators in the US Northeast. Climate Dynamics. 2007; 28(4):381–407. https://doi.org/10.1007/s00382-006-0187-8
- Blum AG, Kanno Y, Letcher BH. Seasonal streamflow extremes are key drivers of brook trout youngof-the-year abundance. Ecosphere. 2018; 9(8):e02356. https://doi.org/10.1002/ecs2.2356
- Bradford MJ, Heinonen JS. Low flows, instream flow needs and fish ecology in small streams. Canadian Water Resources Journal. 2008; 33(2):165–80. https://doi.org/10.4296/cwrj3302165
- 165. Eaton JG, Scheller RM. Effects of climate warming on fish thermal habitat in streams of the United States. Limnology and Oceanography. 1996; 41(5):1109–15. https://doi.org/10.4319/lo.1996.41.5. 1109
- 166. Heino J, Virkkala R, Toivonen H. Climate change and freshwater biodiversity: detected patterns, future trends and adaptations in northern regions. Biological reviews of the Cambridge Philosophical Society. 2009; 84(1):39–54. https://doi.org/10.1111/j.1469-185X.2008.00060.x PMID: 19032595
- 167. Boyero L, Pearson RG, Dudgeon D, Ferreira V, Graça MAS, Gessner MO, et al. Global patterns of stream detritivore distribution: implications for biodiversity loss in changing climates. Global Ecology and Biogeography. 2012; 21(2):134–41. https://doi.org/10.1111/j.1466-8238.2011.00673.x
- 168. Pyne MI, Poff NL. Vulnerability of stream community composition and function to projected thermal warming and hydrologic change across ecoregions in the western United States. Global Change Biology. 2016; 23. https://doi.org/10.1111/gcb.13437 PMID: 27429092
- 169. DeWeber JT, Wagner T. Probabilistic measures of climate change vulnerability, adaptation action benefits, and related uncertainty from maximum temperature metric selection. Global Change Biology. 2018; 24(6):2735–48. https://doi.org/10.1111/gcb.14101 PMID: 29468779
- Daley ML, Potter JD, McDowell WH. Salinization of urbanizing New Hampshire streams and groundwater: effects of road salt and hydrologic variability. Journal of the North American Benthological Society. 2009; 28(4):929–40. https://doi.org/10.1899/09-052.1

- 171. Hall CJ, Jordaan A, Frisk MG. Centuries of anadromous forage fish loss: consequences for ecosystem connectivity and productivity. BioScience. 2012; 62(8):723–31. https://doi.org/10.1525/bio.2012.62.8.
- 172. U.S. Environmental Protection Agency. National Rivers and Streams Assessment 2008–2009: A collaborative survey. Washington, DC: U.S. Environmental Protection Agency, Office of Water and Office of Research and Development; 2016. March 2016. EPA/841/R-16/007.
- 173. Mattocks S, Hall CJ, Jordaan A. Damming, lost connectivity, and the historical role of anadromous fish in freshwater ecosystem dynamics. BioScience. 2017; 67(8):713–28. <u>https://doi.org/10.1093/biosci/ bix069</u>
- 174. Peterson MS, Comyns BH, Hendon JR, Bond PJ, Duff GA. Habitat use by early life-history stages of fishes and crustaceans along a changing estuarine landscape: differences between natural and altered shoreline sites. Wetlands Ecology and Management. 2000; 8(2):209–19. https://doi.org/10. 1023/A:1008452805584
- 175. Balouskus RG, Targett TE. Egg deposition by Atlantic silverside, *Menidia menidia*: substrate utilization and comparison of natural and altered shoreline type. Estuaries and Coasts. 2012; 35(4):1100–9. https://doi.org/10.1007/s12237-012-9495-x
- 176. Gittman RK, Peterson CH, Currin CA, Joel Fodrie F, Piehler MF, Bruno John F. Living shorelines can enhance the nursery role of threatened estuarine habitats. Ecological Applications. 2016; 26(1):249– 63. https://doi.org/10.1890/14-0716 PMID: 27039523
- 177. Dugan JE, Emery KA, Alber M, Alexander CR, Byers JE, Gehman AM, et al. Generalizing ecological effects of shoreline armoring across soft sediment environments. Estuaries and Coasts. 2017:1–17. https://doi.org/10.1007/s12237-017-0254-x
- 178. Geraldi NR, Smyth AR, Piehler MF, Peterson CH. Artificial substrates enhance non-native macroalga and N<sub>2</sub> production. Biological Invasions. 2013; 16(9):1819–31. <u>https://doi.org/10.1007/s10530-013-0629-2</u>
- 179. Pappal A. Marine invasive species. State of the Gulf of Maine Report. Currier P, Gould D, Hertz L, Huston J, Tremblay ML, editors. The Gulf of Maine Council on the Marine Environment, 2010.
- Tyrrell MC, Byers JE. Do artificial substrates favor nonindigenous fouling species over native species? Journal of Experimental Marine Biology and Ecology. 2007; 342(1):54–60. <u>https://doi.org/10.1016/j.jembe.2006.10.014</u>
- 181. Paxton BJ. Potential impact of common reed expansion on threatened high-marsh bird communities on the seaside: assessment of Phragmites invasion of high marsh habitats. Center for Conservation Biology Technical Report Series. Williamsburg, VA: College of William and Mary, 2006. CCBTR-06-17.
- 182. Smith JAM. The role of *Phragmites australis* in mediating inland salt marsh migration in a Mid-Atlantic estuary. PLoS One. 2013; 8(5):e65091. <u>https://doi.org/10.1371/journal.pone.0065091</u> PMID: 23705031
- Edwards M, Richardson AJ. Impact of climate change on marine pelagic phenology and trophic mismatch. Nature. 2004; 430:881–4. https://doi.org/10.1038/nature02808 PMID: 15318219
- Staudinger MD, Mills KE, Stamieszkin K, Record NR, Hudak CA, Allyn A, et al. It's about time: A synthesis of changing phenology in the Gulf of Maine ecosystem. Fisheries Oceanography. 2019; 28 (5):532–66. https://doi.org/10.1111/fog.12429 PMID: 31598058
- 185. Heck KL Jr., Able KW, Fahay MP, Roman CT. Fishes and decapod crustaceans of Cape Cod eelgrass meadows: species composition, seasonal abundance patterns and comparison with unvegetated substrates. Estuaries. 1989; 12(2):59–65.
- 186. Dionne M, Short FT, Burdick DM. Fish utilization of restored, created and reference salt-marsh habitat in the Gulf of Maine. American Fisheries Society Symposium 22: The American Fisheries Society; 1999. p. 384–404.
- 187. Lazzari MA. Habitat variability in young-of-the-year winter flounder, *Pseudopleuronectes americanus*, in Maine estuaries. Fisheries Research. 2008; 90(1–3):296–304. https://doi.org/10.1016/j.fishres. 2007.11.020
- Lazzari MA, Stone BZ. Use of submerged aquatic vegetation as habitat by young-of-the-year epibenthic fishes in shallow Maine nearshore waters. Estuarine, Coastal and Shelf Science. 2006; 69(3– 4):591–606. https://doi.org/10.1016/j.ecss.2006.04.025
- 189. Buchsbaum R, Pederson J, Robinson WE. The decline on fisheries resources in New England: evaluating the impact of overfishing, contamination, and habitat degradation. Cambridge, Massachusetts: Massachusetts Institute of Technology Sea Grant College Program; 2005. 1–175 p.
- **190.** Bell RJ, Richardson DE, Hare JA, Lynch PD, Fratantoni PS. Disentangling the effects of climate, abundance, and size on the distribution of marine fish: an example based on four stocks from the Northeast

US shelf. ICES Journal of Marine Science. 2015; 72(5):1311–22. https://doi.org/10.1093/icesjms/ fsu217

- 191. Brown CJ, Broadley A, Adame MF, Branch TA, Turschwell MP, Connolly RM. The assessment of fishery status depends on fish habitats. Fish and Fisheries. 2019; 20(1):1–14. <u>https://doi.org/10.1111/faf.</u> 12318
- **192.** Arkema KK, Guannel G, Verutes G, Wood SA, Guerry A, Ruckelshaus M, et al. Coastal habitats shield people and property from sea-level rise and storms. Nature Climate Change. 2013; 3(10):913–8. https://doi.org/10.1038/nclimate1944
- 193. Powell EJ, Tyrrell MC, Milliken A, Tirpak JM, Staudinger MD. A review of coastal management approaches to support the integration of ecological and human community planning for climate change. Journal of Coastal Conservation. 2018. https://doi.org/10.1007/s11852-018-0632-y
- 194. Reguero BG, Beck MW, Bresch DN, Calil J, Meliane I. Comparing the cost effectiveness of naturebased and coastal adaptation: a case study from the Gulf Coast of the United States. PLoS One. 2018; 13(4):e0192132. https://doi.org/10.1371/journal.pone.0192132 PMID: 29641611
- 195. Behrenfeld MJ, O'Malley RT, Siegel DA, McClain CR, Sarmiento JL, Feldman GC, et al. Climatedriven trends in contemporary ocean productivity. Nature. 2006; 444(7120):752–5. https://doi.org/10. 1038/nature05317 PMID: 17151666
- 196. Rabalais NN, Diaz RJ, Levin LA, Turner RE, Gilbert D, Zhang J. Dynamics and distribution of natural and human-caused hypoxia. Biogeosciences. 2010; 7(2):585–619. <u>https://doi.org/10.5194/bg-7-585-2010</u>