2	Tributary loading and sediment desorption as
3	sources of PFAS to receiving waters
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## 24 Abstract

25 Transport of per- and polyfluoroalkyl substances (PFAS) to the Great Lakes is of great 26 importance as this large freshwater system provides drinking water to over 40 million people. 27 Tributary PFAS loading to the Great Lakes is poorly characterized and the role of sediments as a 28 source or sink of PFAS is largely unknown. We quantified 10 perfluoroalkyl acids (PFAAs) in 29 water (4 - 1.310 ng/L) and sediment (below detection to 3.255 ng/kg) of 41 tributaries to Green 30 Bay of Lake Michigan. We demonstrate that tributary discharge plays a major role in PFAS 31 contribution to receiving waters. In this system, three large rivers (i.e., Fox, Menominee, and 32 Peshtigo Rivers) contribute two-thirds of the total tributary PFAA loading to Green Bay despite 33 their relatively low concentrations and despite the current regulatory focus on sites with high 34 concentrations. Tributary PFAA composition is linked to likely sources, including a fire-fighting 35 foam manufacturer, other industrial activity, and airports. In addition to tributary discharge, we 36 that tributary sediments can contribute to PFAA transport via desorption. show 37 Perfluorooctanesulfonate rapidly desorbs from contaminated riverbed sediments when 38 equilibrated with Lake Michigan water, indicating that sediments may act as a PFAS source if 39 water concentrations are reduced by pollution mitigation methods.

40

#### 41 Keywords

42 Perfluorinated substances, Great Lakes, loadings, partitioning, Green Bay

43

### 44 Synopsis

45 PFAA transport in tributaries is largely linked to tributary discharge rate and is impacted by the46 ability of sediment to act as either a source or a sink of PFAAs.

### 47 Introduction

48 Per- and polyfluoroalkyl substances (PFAS) are a large group of synthetic chemicals that have been used in applications such as cookware and fire-fighting products since the 1950s.<sup>1-3</sup> 49 Many PFAS are persistent, bioaccumulative, and harmful to humans and wildlife.<sup>4-8</sup> 50 51 Perfluoroalkyl acids (PFAAs) are an important class of PFAS compounds that do not degrade naturally and therefore accumulate in the environment.<sup>1,9–11</sup> PFAS are the active ingredients in 52 53 many aqueous film-forming foams (AFFF), which are often used at military sites, airports, and flammable storage facilities to extinguish petroleum-based fires.<sup>12,13</sup> AFFF is a significant source 54 55 of environmental PFAS contamination as its use dispenses relatively high quantities of PFAS directly into the environment.<sup>14,15</sup> Other routes of environmental contamination include municipal 56 wastewater treatment facilities either from their effluents9,16,17 or from land-spreading of 57 biosolids.18-20 58

Preventing PFAS contamination in the Laurentian Great Lakes of North America is of utmost concern as it is the largest freshwater system in the world, supplies drinking water to 40 million people,<sup>21–23</sup> and supports a vibrant commercial and recreational fishery.<sup>21,24</sup> The Great Lakes Basin includes Lakes Superior, Michigan, Huron, Erie, and Ontario. PFAS have been detected in water, sediment, and biota of the Great Lakes, with higher concentrations measured in the eastern lakes.<sup>25–27</sup> Lake Michigan has moderate PFAS concentrations and yet is understudied compared to the other Great Lakes.

Green Bay, a major 4,210 km<sup>2</sup> embayment in the northwest side of Lake Michigan, is of
particular concern for contamination given potential PFAS sources in its watershed. A fire-fighting
products facility located in Marinette, WI has manufactured, tested, and trained with PFAScontaining AFFF since the 1970s (Figure 1). This company started under the name of Ansul,

70 which remains as a major brand of AFFF manufactured by the current company, Tyco Fire Products (Tyco) of Johnson Controls International.<sup>28</sup> Tyco first measured PFAS in groundwater at 71 the testing site in 2013, but did not publicly disclose that PFAS had migrated off site until 2017.<sup>29</sup> 72 73 PFAS contamination attributed to activities at the Tyco facility is prevalent in the surrounding area including private drinking water wells,30 soil,29 and surface water.31 Furthermore, the local 74 wastewater treatment facility has elevated PFAS in its effluent<sup>32</sup> and biosolids.<sup>33</sup> Treated 75 76 wastewater effluent is discharged into the Menominee River, which flows into Green Bay, and biosolids were land-applied on agricultural fields until 2018,<sup>34</sup> potentially distributing PFAS 77 78 contamination throughout the region. Other potential PFAS sources to the bay include the Fox and Menominee Rivers, which both have industrial legacies,<sup>35</sup> and small airports that use or used AFFF 79 80 onsite.

81 The unique properties of PFAS (e.g., both hydrophobic and lipophobic moieties) make 82 their cycling and bioavailability in the environment challenging to predict. PFAS mobility in 83 stream systems can be influenced by partitioning to sediment, but the extent of partitioning 84 depends on several factors, including PFAS chain length, head group chemistry, and differences 85 in water and sediment chemistry (e.g., aqueous cation concentrations and sediment organic carbon content, respectively).<sup>2,36–40</sup> Sediment-water partition coefficients derived from field 86 measurements are consistently higher than those calculated in laboratory studies,<sup>41</sup> indicating a 87 88 knowledge gap in partitioning behavior. Previous research shows that soils and sediments readily act as a sink for some PFAS compounds.<sup>36,39,42,43</sup> Several studies have suggested that irreversible 89 sorption of certain PFAS compounds can occur,<sup>44–49</sup> but the ability of contaminated sediments to 90 91 serve as a PFAS source in tributaries remains unclear.

92 This study quantifies water and sediment PFAA concentrations in 41 Green Bay tributaries 93 to investigate the role of tributaries and sediments as PFAA sources to receiving waters. Green 94 Bay is an ideal location to study PFAA inputs to surface waters due to the extensive groundwater 95 contamination in the region, the variability of tributary sizes and PFAA sources around the large 96 bay, and the critical importance of this freshwater resource. Importantly, we use a mass balance 97 approach to provide the first baseflow estimates of PFAA loadings from tributaries to receiving 98 waters. We also use desorption experiments to demonstrate that riverbed sediments may act as a 99 source of PFAAs following remediation efforts. This study provides insight into the fate of PFAAs 100 in surface waters and has implications for managing and mitigating PFAA contamination in 101 tributaries of the Great Lakes and other freshwater systems.

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## 103 Methods and Materials

104 Materials. PFAS standards were purchased from Wellington Laboratories. A list of 105 analytes, surrogates, and internal standards is shown in **Supporting Information Table S1**. Other 106 chemicals are described in **Section S1**. Ultrapure water was supplied by a Milli-Q water system 107 maintained at  $18.2 \text{ M}\Omega$ ·cm.

**Field Sample Collection.** Samples were collected July 7-10, 2020 at 41 sites under baseflow conditions with no recent storm events (**Figures 1** and **S1**; **Table S2**). Sites included tributaries associated with likely PFAS sources, all major Green Bay tributaries, and sites with anticipated low concentrations to serve as background measurements. We focused on a wide range of sites to assess spatial variability in loadings and sources rather than temporal variability, as aqueous PFAS concentrations in this region exhibit minimal temporal variation.<sup>50</sup> Water samples for PFAS analysis were collected by submerging a 250 mL polypropylene bottle approximately 115 15 cm below the surface to fill. Samples for other water analyses were filtered (0.45 µm nylon) on 116 site and stored in amber glass vials. A polypropylene dipper was used to retrieve riverbed sediment, 117 which was transferred to 50 mL polypropylene Falcon tubes. Only 34 of the 41 sample locations 118 had accessible sediment. Samples were stored at 4°C prior to analysis. Equipment was chosen to 119 avoid PFAS-containing materials, such as Teflon. Nitrile gloves were worn at all times and only 120 PFAS-free clothing and materials were used. See Section S2 for quality control measures.

121 Water Extraction, PFAA Analysis, and Characterization. Water was analyzed for 122 seven PFCAs and three PFSAs (Table S1) following EPA method 533, modified to use a shorter 123 analyte list.<sup>51</sup> Analytes were chosen based on previous detections in this region,<sup>50</sup> their importance 124 in surface water regulations, and their known persistence, bioaccumulation potential, and toxicity.<sup>4-8</sup> Briefly, 250 mL samples were buffered using 1 g/L ammonium acetate and mass-125 126 labeled surrogate standards were added. PFAA compounds were extracted by solid-phase 127 extraction (SPE) onto weak anion exchange cartridges (Phenomenex Strata X-AW) and eluted 128 with 2% ammonium hydroxide in methanol. Extracts were dried under a gentle stream of nitrogen, 129 amended with mass-labeled internal standards, and diluted to 1.0 mL with methanol:Milli-Q water 130 (80:20, v:v). Final extracts were analyzed by liquid chromatography-tandem mass spectrometry 131 (LC-MS/MS; Sections S1 and S2). PFAA concentrations were adjusted based on surrogate 132 standard recovery (Table S3). All PFAAs measured in aqueous samples can be assumed to be 133 dissolved due to the low levels of suspended solids ( $5.3 \pm 12.3 \text{ mg/L}$ ; 70% below detection). Using 134 field-derived partition coefficients, <1% of total aqueous concentrations is expected to be 135 associated with suspended solids in these tributaries. All reported PFAA concentrations are 136 summed linear and branched isomers unless otherwise specified. A separate water sample was used for geochemical characterization, including pH, dissolved organic carbon (DOC), and cation
measurements (Section S1).

139 Sediment Extraction, PFAA Analysis, and Characterization. Sediment samples were analyzed for the same 10 PFAAs following the protocol ASTM D7968-14.52 Briefly, 2g of dry 140 141 sediment was weighed into a 15 mL falcon tube and mass-labeled surrogate standards were added. 142 A 10 mL extraction solution of methanol:Milli-Q water (50:50, v:v) was added and samples were 143 placed on a shaker table for one hour. Samples were adjusted to pH 9-10 using ammonium 144 hydroxide, centrifuged, and filtered (0.2 µm nylon). The filtered supernatant was adjusted to pH 145 3-4 using acetic acid and analyzed by LC-MS/MS (Section S1 and S2). Samples were screened 146 for acceptable surrogate recovery (Table S4). Additional sediment was characterized for organic 147 carbon content, point of zero charge (PZC), surface area, and metal content (Section S1).

148 Laboratory PFAA Equilibrium and Desorption Experiments. Laboratory experiments 149 were performed in sealed 50 mL falcon tubes. For equilibrium experiments, 2 g of dry sediment 150 was added to 50 mL of water from the same site and placed on a shaker table for 10 days. The equilibrium time was based on information from a previous sorption study.<sup>39</sup> For desorption 151 152 experiments, 2 g of sediment was added to 50 mL of Lake Michigan water and placed on a shaker 153 table for up to 10 days. The tubes were centrifuged, supernatant decanted, and refilled with 50 mL 154 of Lake Michigan water every two days. Lake Michigan water served as a low-PFAA surface 155 water surrogate (Table S8) and was renewed to ensure it remained low in PFAAs. At the end of 156 each experiment, each tube was centrifuged, supernatant decanted, and sediment was dried at 157 60°C. The sediments were re-weighed to account for loss of colloids, extracted, and analyzed for 158 PFAAs via LC-MS/MS.

160 **Results & Discussion** 

161 PFAA Concentrations in Tributary Water. The sum of measured PFAA concentrations 162 in the water range from 4 ng/L in Renard Creek to 1,310 ng/L in Ditch B (Figures 2a and S2; 163 **Table S8**). When Ditch B is excluded, the mean and median PFAA concentrations are  $27 \pm 15$ 164 ng/L and 23 ng/L, respectively. The summed PFAA concentrations at these sites are similar to 165 other rivers in the area. For example, the average summed PFAA concentrations in the Mississippi 166 and Wisconsin Rivers are  $28 \pm 12$  ng/L (average of the same 10 PFAAs at four sites at three time 167 points) and  $38 \pm 16$  ng/L (average of the same 10 PFAAs at three sites at three time points), respectively.<sup>50</sup> Concentrations in Green Bay tributaries are generally higher than those reported in 168 Lake Michigan (11 ng/L).<sup>25</sup> The majority of the detected compounds are PFCAs, which make up 169 170  $86 \pm 11\%$  of PFAAs across all sites (Figure S4).

Aqueous tributary concentrations are below most current and proposed regulatory standards. Only three of the sites (Ditch B, Portage Creek, and Duck Creek) exceed the proposed surface water criteria for Wisconsin for PFOA (20 ng/L) and PFOS (8 ng/L).<sup>53</sup> Three sites (Ditch B, Portage Creek, and Thomas Slough) exceed the promulgated drinking water and groundwater standards in Michigan for PFOA (8 ng/L), PFNA (6 ng/L), and PFOS (16 ng/L).<sup>54</sup> Only Ditch B exceeds the drinking water health advisory level of 70 ng/L combined PFOS and PFOA issued by the U.S. Environmental Protection Agency.<sup>55</sup>

We hypothesized that elevated PFAA concentrations would be present in the Marinette area (**Figure S1**) due to the long history of AFFF use in the region. The Tyco Fire Products facility is located between two major rivers, the Menominee and Peshtigo Rivers, and is associated with a large PFAS groundwater plume. These rivers receive wastewater influent from the cities of Marinette and Peshtigo, respectively, which could also serve as PFAS sources. Additionally, several smaller tributaries, including Ditch B, lead directly from the Tyco facility to the bay.
Biosolids from the Marinette City Wastewater Plant, which are known to contain high levels of
PFAS,<sup>33</sup> were spread on agriculture fields northwest of Marinette until 2018.<sup>34</sup>

186 Despite these potential distribution pathways, high aqueous PFAA concentrations are only 187 observed in Ditch B. A PFAS treatment system was installed in Ditch B upstream of our sampling site in 2019 but was not operating properly at the time of sampling.<sup>56,57</sup> The distribution of PFAA 188 189 compounds (Figure S3) in this sample is dominated by PFOA and is consistent with the signature 190 of legacy fluorotelomer-based AFFF. Ansul uses fluorotelomerization in AFFF manufacturing,<sup>58</sup> 191 with 6:2 and 8:2 fluorotelomer thioether amido sulfonates (FtTAoS) as likely active ingredients.<sup>59,60</sup> FtTAoS compounds can degrade to produce PFCAs including PFPeA, PFHxA, 192 and PFOA as terminal products.<sup>58</sup> The current AFFF formulation produced by Ansul uses short 193 194 chain compounds (C  $\leq$  6), suggesting that the high amount of PFOA in Ditch B may be attributable 195 to historical manufacture and/or use of C<sub>8</sub> fluorotelomer AFFF.

196 PFAA concentrations measured in the Menominee River are low (16 ng/L) and are similar 197 to concentrations reported by the Wisconsin Department of Natural Resources (WDNR) at the mouth of the river (4 – 7 ng/L summed PFAS in June, July, and September 2019).<sup>50</sup> Smaller 198 199 tributaries in this region generally have higher aqueous PFAA concentrations than larger 200 tributaries (Figure 2a), which can be attributed to their lower flow rate and proximity to PFAS 201 sources. Two smaller tributaries in the Marinette area have elevated PFAAs: a tributary near the 202 Menominee Regional Airport (Kirby Creek; 71 ng/L) and a creek adjacent to fields that received 203 biosolids (Mudbrook Creek; 60 ng/L).

Overall, summed PFAAs levels in the greater Marinette area (Little River through Gravelly
 Brook in Figure 2a) are not elevated compared to other areas. However, the distribution of PFAAs

206 in these sites suggests they are impacted by similar sources. Principal component analysis (PCA) 207 using the PFAA distribution in aqueous samples shows that sites in the Marinette region are closely 208 related (Figure 3a), which we attribute to higher percentages of PFHxA and/or PFOA (i.e., terminal degradation products of 6:2 and 8:2 FtTAoS).<sup>59,60</sup> An increase in linear isomers could 209 also indicate fluorotelomer AFFF use;<sup>61,62</sup> however, almost all sites in this study contain high 210 211 percentage of linear isomers with no notable difference in Marinette (Figure S5; Table S9). The 212 PFAA compounds measured in this study are important from a regulatory and toxicity perspective. 213 Given that they represent terminal products associated with fluorotelomer AFFF use, more 214 research, including the quantification of PFCA precursors using total oxidizable precursor (TOP) 215 assay or other methods, is needed to fully assess the extent and potential for long-term release of 216 PFAS in this region.

217 Several sites outside of the Marinette area have notably higher PFAA concentrations 218 (Figure 2a). Duck Creek is located in the city of Green Bay and is near the Green Bay Austin 219 Straubel International Airport. Thomas Slough in Oconto, WI does not have any known PFAS 220 sources, but clusters with the Marinette region samples by PCA (Figure 3a). Portage Creek 221 receives runoff from the Delta County Airport in Escanaba, Michigan, where significant PFAS contamination has been found,<sup>63</sup> and clusters with Green Bay sites due to its high PFHxS content. 222 223 In all sites that show elevated aqueous PFAAs, the tributary discharge is relatively low ( $<1 \text{ m}^3/\text{s}$ ).<sup>64</sup> 224 Areas with low PFAA concentrations include the eastern and northern shores of the bay (i.e., the 225 Door County peninsula and the upper peninsula of Michigan, respectively), corresponding to a 226 regional background level of 22 ng/L for summed PFAAs (Table S8). Areas with elevated aqueous 227 PFAAs tend to be individual tributaries, rather than entire regions around the bay, suggesting that 228 PFAAs are closely associated with nearby sources. However, PCA shows that aqueous PFAA

229 distribution generally clusters by region (Figure 3a), indicating that PFAA sources may impact 230 tributaries throughout a region.

# 231

PFAA Loading to the Bay of Green Bay from Tributaries. While concentrations of 232 PFAAs in individual tributaries are important for evaluating the potential for human and ecosystem 233 exposure, stream concentrations do not directly translate into fluxes to receiving waters. Therefore, 234 we calculated baseflow loading of PFAAs to quantify transport to Green Bay by tributaries:

$$Loading = [PFAA] * tributary discharge$$
(1)

236 where loading is in units of ng/s, [PFAA] is in units of ng/L, and discharge is in units of L/s. Tributary discharge estimates are taken from Mooney et al.<sup>64</sup> where the discharge-watershed area 237 238 ratio method was used and verified using U.S. Geological Survey stream gauges in the Great Lakes 239 Basin (Table S2). All major rivers that drain to the bay, as well as many smaller tributaries, are 240 included in the study. This analysis considers PFAA concentrations measured under baseflow 241 conditions, thus providing a snapshot of loading at the time of sampling. Discharge can range 242 widely (e.g., during snowmelt) and relationships between PFAS concentrations and discharge in 243 these tributaries have not been established. Further work is needed to quantify other loading 244 sources to the bay including groundwater flow (e.g., from the PFAS plume that flows southeast 245 from Tyco) and precipitation.

246 The largest rivers, including the Fox, Peshtigo, and Menominee Rivers, have the largest 247 PFAA loadings (Figure 2b), despite having relatively low aqueous concentrations. These three 248 rivers contribute over two-thirds of the total tributary PFAA loading to Green Bay. In total, the 249 sampled tributaries contribute 320 g/day of summed PFAAs into the bay under baseflow 250 conditions. The Fox River contributes 32% of the PFAA loading to the bay from these tributaries,

the Peshtigo contributes 12%, and the Menominee contributes 23%. Ditch B, which has high
PFAA concentrations but very low discharge, contributes only 4% of the PFAA loading to the bay.

253 PFAA loading from tributaries is strongly correlated with discharge rate, rather than PFAA 254 concentration (Figure S6). These results demonstrate that larger rivers dominate PFAA inputs to 255 the bay despite their low concentration relative to many of the smaller tributaries. Current and proposed PFAS regulations focus on concentrations at individual sites,<sup>53–55</sup> which is critical for 256 257 protecting human health (e.g., in the context of drinking water or fish consumption). However, the 258 results of this study indicate that regulatory efforts should place greater emphasis on discharge and 259 loadings in order to protect receiving waters. This concept is commonly applied to nutrients and 260 other stressors by applying Total Maximum Daily Loads and should be considered for PFAS.

261 **PFAA Concentrations in Tributary Sediment.** Certain PFAS compounds, like PFOS, 262 exhibit a high potential for sorption and can accumulate in sediments.<sup>1,40,65</sup> In Green Bay 263 tributaries, as in most fluvial systems, summed PFAA concentrations are much higher in bed 264 sediments than in overlying water, reaching levels of over 1,000 ng/kg in many sites (Figure 2c; 265 Table S10). Interestingly, PFAA sediment concentrations are more variable than surface water 266 concentrations. Sediment PFAA concentrations range from below detection to 3,255 ng/kg (mean: 267  $732 \pm 851$  ng/kg; median: 304 ng/kg). Lake Michigan open water sediments have reported total PFAA concentrations of 4 ng/kg.<sup>25</sup> 268

Sediment PFAA concentrations are highest at sites near urban or industrially impacted areas and near airports (**Figures 1** and **2c**). For example, tributaries near the city of Green Bay have elevated sediment PFAA concentrations ranging 258 – 2,508 ng/kg. Portage Creek (2,095 ng/kg), which receives runoff from the Delta County Airport, also has very high sediment PFAAs. Near Marinette, there are elevated sediment PFAAs in Ditch B (1,582 ng/kg), which discharges directly off of the Tyco property, and two upstream sites (>2,000 ng/kg) that drain fields that
 previously received contaminated biosolids.<sup>33</sup>

276 Sites outside of the Marinette and Green Bay areas are considered rural or minimally 277 impacted by known point sources. Most of these sites are low in summed sediment PFAAs (below 278 detection to 400 ng/kg), corresponding to a regional background level of 211 ng/kg (**Table S10**). 279 However, several sites have elevated sediment concentrations with no identified PFAS source 280 (Figure 2c). Thomas Slough (3,255 ng/kg) is in a rural area between Green Bay and Marinette and 281 contains the highest sediment PFAA concentration in this study. Two large rivers in the upper 282 peninsula of Michigan also have elevated PFAAs: Rapid River (1,309 ng/kg) and Fishdam River 283 (1,101 ng/kg). The source(s) of elevated PFAAs in these three sites are not known.

284 The distribution of specific PFAA compounds in these sediments (Figure S7) is dominated 285 by PFOS (57  $\pm$  21%). Across all sites, PFCAs represented 39  $\pm$  20% of summed PFAAs (Figure 286 S8), which is a much lower fraction than in the aqueous samples ( $86 \pm 11\%$  PFCAs; Figure S4). This difference is likely driven by the high sorptivity of PFOS<sup>1,40,65</sup> and the increased sorption of 287 PFSAs compared to PFCAs of the same carbon chain length.<sup>37,39</sup> The percentage of linear isomers 288 289 is higher in sediments than in the aqueous phase (Figure S9; Table S11). For example,  $47 \pm 10\%$ 290 of PFOS is linear in aqueous samples compare to  $85 \pm 10\%$  in sediment samples. This trend is 291 consistent with higher sorption of linear compounds compared to branched compounds observed in the Great Lakes.<sup>25</sup> 292

Ditch B, Mudbrook Creek, and Gravelly Brook, which are all connected to the Tyco facility, cluster near each other by PCA based on the relative distribution of PFAAs in the sediment (**Figure 3b**) and exhibit elevated levels of PFPeA, PFHxA, and PFOA, which are terminal products of the active ingredients in fluorotelomer-based AFFF.<sup>58</sup> These three sites contain an average of 297 62% PFCAs, which is much higher than the overall average in sediment. The similarity in 298 compound distributions between Ditch B and the two streams draining biosolids-impacted fields 299 suggests that PFAAs from Tyco-impacted biosolids have impacted nearby streams (**Figure 3b**). 300 According to one analysis, biosolids from the Marinette City Wastewater Plant are elevated in 301 PFHxA and PFOS (290  $\mu$ g/kg and 210  $\mu$ g/kg, respectively) and include some PFOA (10  $\mu$ g/kg).<sup>33</sup> 302 The continued presence of these compounds indicate that agricultural sediments can retain legacy 303 PFAS for years.

Sediment-water partition coefficients (Section S4) calculated from measured sediment and water PFAA concentrations show that log  $K_d$  values from Green Bay field samples (Figure S10) increase with increasing number of carbons in the hydrophobic tail and that PFSAs have a higher  $K_d$  than PFCA counterparts, in agreement with previous laboratory measurements.<sup>37,39</sup> While these data indicate that PFAA structure is important for determining the extent of sorption, the high variability in  $K_d$  values suggests that other factors may contribute.

310 Variability in PFAS sorption among sediment samples has been largely attributed to 311 organic carbon content,<sup>1,40,65</sup> although variation in PFAS sorption across sediments cannot be predicted by organic carbon content alone.<sup>36</sup> To evaluate the role of organic carbon content on 312 313 PFAA sorption in Green Bay tributaries, we calculate  $\log K_{OC}$  values (Figure S10; Table S12) 314 after quantifying organic carbon content in each sediment sample (average =  $7.1 \pm 9.7\%$ ; Table 315 S13). There is a significant relationship between sorption of individual PFAA compounds and 316 organic carbon content (Figure S11; p-values <0.001), indicating that  $f_{OC}$  is a factor in PFAA 317 sorption in these tributaries. No significant correlations were found between PFAA sorption and 318 aqueous cations (Figure S12; Table S14) or sediment PZC (Figure S13; Table S15).

319 Field sediment-water partition coefficients calculated here and in other freshwater field studies<sup>41</sup> are consistently higher than those measured in laboratory studies, suggesting that 320 321 laboratory studies underestimate PFAS sorption. For example, reported laboratory log  $K_d$  values 322 for PFCAs range from -0.75 L/kg for PFPeA to 1.0 L/kg for PFDA,<sup>37</sup> which are considerably lower than the values observed here. The log  $K_d$  values for PFOA and PFOS in this study agree with 323 published global field averages of 1.9 and 2.3 L/kg, respectively,<sup>41</sup> which are higher than the 324 reported laboratory averages of 0.8 and 1.4 L/kg, respectively.<sup>66</sup> The difference between laboratory 325 326 and field partition coefficients may be attributable to the use of water column concentrations rather 327 than porewater or the fact that field samples are not at equilibrium at the time of sampling, as 328 discussed below.

329 Significance of PFAAs Sorbed to Tributary Sediment. The large amount of sediment-330 associated PFAAs identified in tributaries leading to the bay of Green Bay generates questions 331 regarding the mobility of sediment-associated PFAAs. While sorptive compounds like PFOS can accumulate in riverbed sediments if aqueous concentrations are elevated,<sup>1,40,65</sup> it is unclear if those 332 333 sediments can later act as a PFAS source once overlying water concentrations decline. Several studies conclude that certain compounds, including PFOS, PFOA, and PFHxS, undergo sorptive 334 hysteresis where sorption to environmental sediments is partially irreversible,<sup>44–49</sup> suggesting that 335 336 sediments primarily act as a sink for certain compounds and do not release them back into the 337 aqueous phase. Alternatively, others propose that the non-ideal, hysteretic desorption of PFOS has minimal effect on its mobility,<sup>67</sup> suggesting that sediments can act as either a source or a sink of 338 339 PFOS depending on aqueous phase concentrations. Many studies use high concentration 340 isotherms, which are useful to characterize PFAS behavior, but do not directly address questions 341 about transport under conditions representative of tributaries. Furthermore, the dependence of 342 PFAS sorption and desorption on a wide variety of variables, many of which are unknown, makes 343 transport modeling challenging.<sup>68</sup> Therefore, simulating environmental conditions as closely as 344 possible is critical for understanding and ultimately modeling PFAS mobility in natural 345 environments.

346 Sediment and water will not be at equilibrium in a riverine system unless either the 347 sorption/desorption processes happen very rapidly or PFAS concentrations in the flowing water is 348 constant over time. In order to test whether PFAAs in these sediments are at equilibrium with the 349 overlying water, we first equilibrate sediment with water from the same site for 10 days. We 350 selected six sites based on their importance (Menominee and Fox Rivers are current or delisted Areas of Concern;<sup>35</sup> Peshtigo River and Hay Creek are near a site of high groundwater 351 352 contamination) and/or high PFAA concentrations (Portage Creek and Thomas Slough have 353 sediment PFAA concentrations >2,000 ng/kg). Sediment PFAS concentrations change by up to 354 150% over the 10-day period, showing that the water and sediments were not at equilibrium when 355 the field samples were collected (Figure 4a and S14a; Table S16). Notably, PFOS concentrations 356 in the bed sediments decrease in four of the six samples, indicating that PFOS is susceptible to 357 desorption from these sediments over time. Initial and final PFOS partition coefficients (log  $K_d$  of 358 1.92 and 1.26 L/kg, respectively; Table S17) show that these sediments approach the reported laboratory partitioning coefficient (1.4 L/kg)<sup>66</sup> after equilibration. This result may support the 359 360 hypothesis that differing concentrations between water column and porewater drive differences in 361 field and laboratory K<sub>d</sub> values. Additionally, PFNA and PFDA decrease in four and one sediment 362 samples, respectively.

However, the opposite trend is observed for several other compounds with sediment PFAA
 concentrations increasing after equilibration. For example, sediment samples increase in

365	concentrations of PFOA ( $n = 6$ ), PFDA ( $n = 5$ ), PFNA ( $n = 2$ ), and PFOS ( $n = 2$ ). The increase in
366	concentrations of certain PFAAs indicates that the sediment is undersaturated with respect to
367	sorption of these compounds, driving sorption to the sediment. In the case of PFOA, the increase
368	in log $K_d$ from 1.08 to 1.56 L/kg after equilibration ( <b>Table S17</b> ) moves the $K_d$ even further from
369	the global laboratory average of 0.8 L/kg.41 The reason for this deviation from equilibrium is
370	unknown, but may be related to the low PFOA concentrations in the selected sediment samples
371	(14 - 132  ng/kg). None of the samples increase or decrease in concentrations of all compounds,
372	indicating that the sorptive and desorptive behavior is not inherent to sediment type but rather a
373	function of water and sediment concentrations. Changes in shorter chain compounds were not
374	recorded as either the initial, final, or both concentrations were below the minimum reporting limit.
375	To further quantify the ability of sediment to serve as a PFAA source, we equilibrated site
376	sediment with Lake Michigan water for 10 days. Lake Michigan water was used as an
377	environmentally relevant surrogate for surface water with low PFAA concentrations (Table S8),
378	representing the surface water in these tributaries if PFAS sources were substantially reduced. Four
379	sediments were selected based on their high PFAA concentrations (>1,500 ng/kg; Figure 2c),
380	while Menominee River sediment was selected based on its proximity to the Tyco facility.
381	Sediment PFAA concentrations decrease in all five samples after 10 days of equilibrium with Lake
382	Michigan water (Figure 4b and S14b; Table S18). Notably, PFOS concentrations decrease in all
383	samples, showing desorption of up to 2,000 ng/kg in one sample. PFOA concentrations decrease
384	in four of the five sediments, with desorption of up to 644 ng/kg in one sample. This further
385	demonstrates that these sediments can act as a source of PFAAs depending on water
386	concentrations. It should be noted that sediment desorption is dependent on further deposition of

sediment, as fresh, non-contaminated sediment can potentially cap existing contaminated

388 sediment. PFOS desorption happens rapidly (i.e., within two days; Figure S15). Sediment 389 concentrations do not reach zero after desorption experiments, which could be due to either 390 equilibrium with Lake Michigan water or sorption hysteresis.<sup>44–49</sup>

391 **Conclusions.** This study has implications for both the Green Bay watershed and 392 environmental PFAS contamination as a whole. Baseflow loading calculations for these tributaries 393 demonstrate that rivers with high discharge rates but low concentrations contribute the large 394 majority of PFAAs to the bay. This indicates that tributary discharge must be considered when 395 developing regulations to protect the bay of Green Bay, the Great Lakes, and other receiving 396 waters, an approach markedly different from the current focus on concentrations of individual 397 PFAS on a site-by-site basis.

398 Sediment-water partitioning analyses shows that sediments can act as a source or a sink of 399 PFAAs, but a fraction of PFOS may be irreversibly sorbed. Our findings suggest that tributary 400 sediments with high PFAA concentrations can act as a source of PFAAs if the aqueous 401 concentrations are decreased via pollution mitigation measures.

402 Certain PFAS compounds, including the PFAAs evaluated in this study, can lead to 403 negative health effects in humans and wildlife. Long-chain PFAAs (PFOA and longer, PFHxS and 404 longer) are persistent, bioaccumulative, and toxic.<sup>3,69,70</sup> The Great Lakes and other waterways are 405 important to protect as they are used as a source of drinking water and food. They are also 406 considered sacred to many indigenous peoples and support the lives of many plant and animal 407 species. Once PFAS compounds have made it into waterways, they are difficult to remove due to 408 their unique lipophobic and hydrophobic characteristics and strong carbon-fluorine bonds.

We focused on PFAAs because these terminal compounds are most closely associated with
 adverse health outcomes. While these PFAA compounds are driving regulations across the United

411	States, including in Wisconsin, future work should focus on fluorotelomers and other precursors
412	in order to more fully assess the potential for long-term release of PFAS in this area.
413	
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419	
420	Supporting Information
421	The Supporting Information is available free of charge at
422	Additional experimental details, quality control information, and supplementary figures
423	and tables (PDF).

424



Figure 1. Map of sites where samples were collected from Green Bay tributaries and in the bay itself created using ArcGIS software (10.6.1) by Esri. Data provided by the National Atlas of the United States, USGS. Black airplane symbols show where regional airports are located. Urban areas, including Green Bay and Marinette, are shaded dark gray. See Figure S1 in Supporting Information for a detailed map of the Marinette region, including the Tyco facility.

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Figure 2. PFAA concentrations in Green Bay tributaries including (a) concentrations in the water column, (b) loading rates into the bay, and (c) concentrations in sediment. Loading is calculated using aqueous PFAA concentrations and estimated discharge rates.<sup>64</sup> Loading is not calculated for sites denoted with an X because these sites do not drain directly into the bay. Sediment data was not collected for sites denoted by a \* due to inaccessibility or rocky riverbeds. See Figure S2 for

- 440 a detailed view of panel (a).
- 441



442Standardized PC1 (33.6% explained var.)Standardized PC1 (30.7% explained var.)443Figure 3. Principal components analysis of the normalized distribution of 10 PFAAs in (a) aqueous444samples and (b) sediment samples. Labels correspond to site numbers in Figure 1.



447 Site Site
448 Figure 4. Change in PFAA concentrations of sediments that were equilibrated with (a) surface
449 water from the same site for 10 days, and (b) Lake Michigan water for 10 days. See Figure S14
450 for plots of concentration percent change.

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# 453 References

- 454 (1) Xiao, F. Emerging poly- and perfluoroalkyl substances in the aquatic environment: A review of current literature. *Water Res.* 2017, *124*, 482–495.
- 456 (2) Ahrens, L. Polyfluoroalkyl compounds in the aquatic environment: A review of their occurrence and fate. *J Env. Monitor.* **2011**, *13* (1), 20–31.
- 458 (3) Lindstrom, A. B.; Strynar, M. J.; Libelo, E. L. Polyfluorinated compounds: Past, present, and future. *Environ. Sci. Technol.* 2011, 45 (19), 7954–7961.
- 460 (4) Ankley, G. T.; Cureton, P.; Hoke, R. A.; Houde, M.; Kumar, A.; Kurias, J.; Lanno, R.;
  461 McCarthy, C.; Newsted, J.; Salice, C. J.; Sample, B. E.; Sepúlveda, M. S.; Steevens, J.;
  462 Valsecchi, S. Assessing the ecological risks of per- and polyfluoroalkyl substances: Current
  463 state-of-the science and a proposed path forward. *Environ. Toxicol. Chem.* 2021, 40 (3),
  464 564–605.
- 465 (5) Brown, J. B.; Conder, J. M.; Arblaster, J. A.; Higgins, C. P. Assessing human health risks
  466 from per- and polyfluoroalkyl substance (PFAS)-impacted vegetable consumption: A tiered
  467 modeling approach. *Environ. Sci. Technol.* 2020, *54* (23), 15202–15214.
- 468 (6) Houde, M.; Czub, G.; Small, J. M.; Backus, S.; Wang, X.; Alaee, M.; Muir, D. C. G.
  469 Fractionation and bioaccumulation of perfluorooctane sulfonate (PFOS) isomers in a Lake
  470 Ontario food web. *Environ. Sci. Technol.* 2008, *42* (24), 9397–9403.
- 471 (7) Garg, S.; Kumar, P.; Mishra, V.; Guijt, R.; Singh, P.; Dumée, L. F.; Sharma, R. S. A review
  472 on the sources, occurrence and health risks of per-/poly-fluoroalkyl substances (PFAS)
  473 arising from the manufacture and disposal of electric and electronic products. *J. Water*474 *Process Eng.* 2020, *38* (101683), 1–15.
- 475 (8) Brase, R. A.; Mullin, E. J.; Spink, D. C. Legacy and emerging per- and polyfluoroalkyl substances: Analytical techniques, environmental fate, and health effects. *Int. J. Mol. Sci.*477 2021, 22 (3), 995.
- 478 (9) Rayne, S.; Forest, K. Perfluoroalkyl sulfonic and carboxylic acids: A review of physicochemical properties, levels and patterns in waters and wastewaters, and treatment methods. J. Environ. Sci. Health Part A Tox. Hazard. Subst. Environ. Eng. 2009, 44 (12), 1145–1199.
- (10) Rahman, M. F.; Peldszus, S.; Anderson, W. B. Behaviour and fate of perfluoroalkyl and polyfluoroalkyl substances (PFASs) in drinking water treatment: A review. *Water Res.*2014, 50, 318–340.
- 485 (11) Giesy, J. P.; Kannan, K. Global distribution of perfluorooctane sulfonate in wildlife.
  486 *Environ. Sci. Technol.* 2001, *35* (7), 1339–1342.
- 487 (12) DOD. Performance Specification: Fire Extinguishing Agent, Aqueous Film-Forming Foam
   488 (AFFF) Liquid Concentrate, for Fresh and Sea Water; MIL-PRF-24385; U.S. Department
   489 of Defense, 2018.
- 490 (13) Darwin, R. L. *Estimated Quantities of Aqueous Film Forming Foam (AFFF) in the United*491 *States*; Fire Fighting Foam Coalition: Baltimore, MD, 2004; p 45.
- 492 (14) Anderson, R. H.; Long, G. C.; Porter, R. C.; Anderson, J. K. Occurrence of select 493 perfluoroalkyl substances at U.S. Air Force aqueous film-forming foam release sites other 494 than fire-training areas: Field-validation of critical fate and transport properties. 495 *Chemosphere* 2016, *150*, 678–685.
- 496 (15) Moody, C. A.; Field, J. A. Determination of perfluorocarboxylates in groundwater impacted
  497 by fire-fighting activity. *Env. Sci. Technol.* 1999, *33* (16), 2800–2806.

- 498 (16) Hu, X. C.; Andrews, D. Q.; Lindstrom, A. B.; Bruton, T. A.; Schaider, L. A.; Grandjean, P.;
  499 Lohmann, R.; Carignan, C. C.; Blum, A.; Balan, S. A.; Higgins, C. P.; Sunderland, E. M.
  500 Detection of poly- and perfluoroalkyl substances (PFASs) in U.S. drinking water linked to
  501 industrial sites, military fire training areas, and wastewater treatment plants. *Environ. Sci.*502 *Technol. Lett.* 2016, *3* (10), 344–350.
- (17) Houtz, E.; Wang, M.; Park, J.-S. Identification and fate of aqueous film forming foam
   derived per- and polyfluoroalkyl substances in a wastewater treatment plant. *Environ. Sci. Technol.* 2018, *52* (22), 13212–13221.
- 506 (18) Gottschall, N.; Topp, E.; Edwards, M.; Payne, M.; Kleywegt, S.; Lapen, D. R. Brominated
  507 flame retardants and perfluoroalkyl acids in groundwater, tile drainage, soil, and crop grain
  508 following a high application of municipal biosolids to a field. *Sci. Total Environ.* 2017, *574*,
  509 1345–1359.
- (19) Ghisi, R.; Vamerali, T.; Manzetti, S. Accumulation of perfluorinated alkyl substances
  (PFAS) in agricultural plants: A review. *Environ. Res.* 2019, *169*, 326–341.
- (20) Chu, S.; Letcher, R. J. Side-chain fluorinated polymer surfactants in aquatic sediment and
   biosolid-augmented agricultural soil from the Great Lakes basin of North America. *Sci. Total Environ.* 2017, 607–608, 262–270.
- (21) Crimmins, B. S.; McCarty, H. B.; Fernando, S.; Milligan, M. S.; Pagano, J. J.; Holsen, T.
  M.; Hopke, P. K. Commentary: Integrating non-targeted and targeted chemical screening in Great Lakes fish monitoring programs. J. Gt. Lakes Res. 2018, 44 (5), 1127–1135.
- 518 (22) Quinn, F. H. Hydraulic residence times for the Laurentian Great Lakes. J. Gt. Lakes Res.
  519 1992, 18 (1), 22–28.
- 520 (23) Facts and Figures about the Great Lakes. US Environmental Protection Agency April 6,
   521 2021, https://www.epa.gov/greatlakes/facts-and-figures-about-great-lakes.
- 522 (24) Clement, R. E.; Reiner, E. J.; Bhavsar, S. P. Organohalogen contaminants of emerging
  523 concern in Great Lakes fish: A review. *Anal. Bioanal. Chem.* 2012, 404 (9), 2639–2658.
- (25) Remucal, C. K. Spatial and temporal variability of perfluoroalkyl substances in the
   Laurentian Great Lakes. *Environ. Sci. Process. Impacts* 2019, 21, 1816–1834.
- Muir, D.; Miaz, L. T. Spatial and temporal trends of perfluoroalkyl substances in global
   ocean and coastal waters. *Environ. Sci. Technol.* 2021, 55 (14), 9527–9537.
- (27) Gewurtz, S. B.; Bradley, L. E.; Backus, S.; Dove, A.; McGoldrick, D.; Hung, H.; DryfhoutClark, H. Perfluoroalkyl acids in Great Lakes precipitation and surface water (2006–2018)
  indicate response to phase-outs, regulatory action, and variability in fate and transport
  processes. *Environ. Sci. Technol.* 2019, *53* (15), 8543–8552.
- 532 (28) ANSUL: Who We Are; F-2006141; ANSUL Innovative Fire Solutions: Wayback Machine,
   533 2010,
- https://web.archive.org/web/20100917210930/http://ansul.com/en/whoweare/pdf\_docs/F 2006141.pdf
- (29) Records: Tyco Knew of Contamination 4 Years before Reporting. Associated Press.
   February 4, 2019, https://apnews.com/article/f23e955a2e8a4cb0b739a649fd0b36e8.
- Kaeding, D. Peshtigo Residents Reach \$17.5M Class Action Settlement with Tyco over 538 (30) 539 PFAS Contamination. Wisconsin Public Radio. January 7. 2021. 540 https://www.wpr.org/peshtigo-residents-reach-17-5m-class-action-settlement-tyco-over-541 pfas-contamination.
- 542 (31) Elevated PFAS Levels Found in Ditch Downstream of JCI/Tyco. *Wisconsin Department of* 543 *Natural Resources*. August 28, 2020, https://dnr.wi.gov/news/releases/article/?id=5234.

- 544 (32) Howard, W. *WWTP Sample Results*; 20180612 43; Wisconsin Department of Natural
   545 Resources: 02-38-580694 Tyco Fire Technology Center, 2018.
- 546 (33) Test America. *Biosolids Sample Results*; 20180622 43; Wisconsin Department of Natural
   547 Resources: 02-38-580694 Tyco Fire Technology Center, 2018.
- 548 (34) Biosolids Map; City of Marinette Official Website, March 8, 2021.
   549 https://www.marinette.wi.us/421/Biosolids-Map
- 550 (35) List of Great Lakes AOCs https://www.epa.gov/great-lakes-aocs/list-great-lakes-aocs.
- (36) Campos Pereira, H.; Ullberg, M.; Kleja, D. B.; Gustafsson, J. P.; Ahrens, L. Sorption of
   perfluoroalkyl substances (PFASs) to an organic soil horizon Effect of cation composition
   and pH. *Chemosphere* 2018, 207, 183–191.
- (37) Guelfo, J. L.; Higgins, C. P. Subsurface Transport potential of perfluoroalkyl acids at aqueous film-forming foam (AFFF)-impacted sites. *Environ. Sci. Technol.* 2013, 47 (9), 4164–4171.
- Johnson, R. L.; Anschutz, A. J.; Smolen, J. M.; Simcik, M. F.; Penn, R. L. The adsorption
  of perfluorooctane sulfonate onto sand, clay, and iron oxide surfaces. *J. Chem. Eng. Data*2007, 52 (4), 1165–1170.
- 560 (39) Higgins, C. P.; Luthy, R. G. Sorption of perfluorinated surfactants on sediments. *Environ.* 561 Sci. Technol. 2006, 40 (23), 7251–7256.
- 562 (40) Du, Z.; Deng, S.; Bei, Y.; Huang, Q.; Wang, B.; Huang, J.; Yu, G. Adsorption behavior and
  563 mechanism of perfluorinated compounds on various adsorbents—A review. *J. Hazard.*564 *Mater.* 2014, 274, 443–454.
- 565 (41) Zareitalabad, P.; Siemens, J.; Hamer, M.; Amelung, W. Perfluorooctanoic acid (PFOA) and
  566 perfluorooctanesulfonic acid (PFOS) in surface waters, sediments, soils and wastewater –
  567 A review on concentrations and distribution coefficients. *Chemosphere* 2013, *91* (6), 725–
  568 732.
- (42) Codling, G.; Hosseini, S.; Corcoran, M. B.; Bonina, S.; Lin, T.; Li, A.; Sturchio, N. C.;
  Rockne, K. J.; Ji, K.; Peng, H.; Giesy, J. P. Current and historical concentrations of poly
  and perfluorinated compounds in sediments of the northern Great Lakes Superior, Huron,
  and Michigan. *Environ. Pollut.* 2018, 236, 373–381.
- 573 (43) Awad, E.; Zhang, X.; Bhavsar, S. P.; Petro, S.; Crozier, P. W.; Reiner, E. J.; Fletcher, R.;
  574 Tittlemier, S. A.; Braekevelt, E. Long-term environmental fate of perfluorinated compounds
  575 after accidental release at Toronto Airport. *Environ. Sci. Technol.* 2011, 45 (19), 8081–8089.
- 576 (44) Milinovic, J.; Lacorte, S.; Vidal, M.; Rigol, A. Sorption behaviour of perfluoroalkyl substances in soils. *Sci. Total Environ.* **2015**, *511*, 63–71.
- 578 (45) Miao, Y.; Guo, X.; Dan Peng; Fan, T.; Yang, C. Rates and equilibria of perfluorooctanoate
  579 (PFOA) sorption on soils from different regions of China. *Ecotoxicol. Environ. Saf.* 2017,
  580 *139*, 102–108.
- 581 (46) Zhao, L.; Zhang, Y.; Fang, S.; Zhu, L.; Liu, Z. Comparative sorption and desorption
  582 behaviors of PFHxS and PFOS on sequentially extracted humic substances. *J. Environ. Sci.*583 2014, 26 (12), 2517–2525.
- (47) Pan, G.; Jia, C.; Zhao, D.; You, C.; Chen, H.; Jiang, G. Effect of cationic and anionic
  surfactants on the sorption and desorption of perfluorooctane sulfonate (PFOS) on natural
  sediments. *Environ. Pollut.* 2009, 157 (1), 325–330.
- 587 (48) Jia, C.; You, C.; Pan, G. Effect of temperature on the sorption and desorption of 588 perfluorooctane sulfonate on humic acid. *J. Environ. Sci. China* **2010**, *22* (3), 355–361.

- (49) You, C.; Jia, C.; Pan, G. Effect of salinity and sediment characteristics on the sorption and desorption of perfluorooctane sulfonate at sediment-water interface. *Environ. Pollut.* 2010, *158* (5), 1343–1347.
- 592 (50) 2019 PFAS Surface Water Sampling Results; Wisconsin Department of Natural Resources,
   593 2019, https://www.dhs.wisconsin.gov/publications/p02434v.pdf.
- (51) Rosenblum, L.; Wendelken, S. Method 533: Determination of per-and Polyfluoroalkyl
   Substances in Drinking Water by Isotope Dilution Anion Exchange Solid Phase Extraction
   and Liquid Chromatography/Tandem Mass Spectrometry. United States Environmental
   Protection Agency 2020.
- 598 (52) Standard Test Method for Determination of Perfluorinated Compounds in Soil by Liquid
   599 Chromatography Tandem Mass Spectrometry (LC/MS/MS). ASTM International 2014.
- 600 (53) Cole, P. D. Order of the State of Wisconsin Natural Resources Board Amending and
   601 Creating Rules. Wisconsin Department of Natural Resources. September 2021,
   602 <u>https://dnr.wisconsin.gov/topic/SurfaceWater/PFASCriteria.html</u>.
- 603(54)Michigan adopts strict PFAS in drinking water standards; State of Michigan Department of604Environment, Great Lakes, and Energy. July 2020,605https://www.michigan.gov/egle/0,9429,7-135--534660--,00.html
- 606 (55) Fact Sheet: PFOA & PFOS Drinking Water Health Advisories; EPA 800-F-16-003; US
   607 Environmental Protection Agency, 2016.
- (56) Cox, J. Tyco Submits New Sampling Results for Ditches A and B as Well as Green Bay,
   2020. https://tycomarinette.com/tyco-submits-new-sampling-results-for-ditches-a-and-b as-well-as-green-bay/.
- 611 (57) Cox, J. Ditch B Surface Water Ongoing Testing Results, 2021.
   612 https://tycomarinette.com/ditch-b-surface-water-ongoing-testing-results-5/
- 613 (58) Houtz, E. F.; Higgins, C. P.; Field, J. A.; Sedlak, D. L. Persistence of perfluoroalkyl acid
  614 precursors in AFFF-impacted groundwater and soil. *Environ. Sci. Technol.* 2013, 47 (15),
  615 8187–8195.
- 616 (59) Place, B. J.; Field, J. A. Identification of novel fluorochemicals in aqueous film-forming
  617 foams used by the US military. *Environ. Sci. Technol.* 2012, 46 (13), 7120–7127.
- (60) Backe, W. J.; Day, T. C.; Field, J. A. Zwitterionic, cationic, and anionic fluorinated
  chemicals in aqueous film forming foam formulations and groundwater from U.S. Military
  bases by nonaqueous large-volume injection HPLC-MS/MS. *Env. Sci. Technol.* 2013, 47,
  5226–5234.
- (61) Harding-Marjanovic, K. C.; Houtz, E. F.; Yi, S.; Field, J. A.; Sedlak, D. L.; Alvarez-Cohen,
  L. Aerobic biotransformation of fluorotelomer thioether amido sulfonate (Lodyne) in AFFFamended microcosms. *Env. Sci. Technol.* 2015, *49*, 7666–7674.
- (62) Kärrman, A.; Elgh-Dalgren, K.; Lafossas, C.; Møskeland, T. Environmental levels and distribution of structural isomers of perfluoroalkyl acids after aqueous fire-fighting foam (AFFF) contamination. *Environ. Chem.* 2011, *8* (4), 372–380.
- (63) TV6 News Team. Delta County Airport Works to Mitigate PFAS Contamination. Upper
   Michigans Source. Escanaba, MI January 21, 2021.
   https://www.uppermichiganssource.com/2021/01/21/delta-county-airport-works-to mitigate-pfas-contamination/.
- (64) Mooney, R. J.; Stanley, E. H.; Rosenthal, W. C.; Esselman, P. C.; Kendall, A. D.; McIntyre,
  P. B. Outsized nutrient contributions from small tributaries to a Great Lake. *Proc. Natl. Acad. Sci.* 2020, *117* (45), 28175–28182.

- (65) Li, Y.; Oliver, D. P.; Kookana, R. S. A critical analysis of published data to discern the role
  of soil and sediment properties in determining sorption of per and polyfluoroalkyl
  substances (PFASs). *Sci. Total Environ.* 2018, 628–629, 110–120.
- 638 (66) Ahrens, L.; Yeung, L. W. Y.; Taniyasu, S.; Lam, P. K. S.; Yamashita, N. Partitioning of
  639 perfluorooctanoate (PFOA), perfluorooctane sulfonate (PFOS) and perfluorooctane
  640 sulfonamide (PFOSA) between water and sediment. *Chemosphere* 2011, *85* (5), 731–737.
- (67) Brusseau, M. L.; Khan, N.; Wang, Y.; Yan, N.; Van Glubt, S.; Carroll, K. C. Nonideal
  transport and extended elution tailing of PFOS in soil. *Environ. Sci. Technol.* 2019, *53* (18),
  10654–10664.
- 644 (68) Brusseau, M. L.; Yan, N.; Van Glubt, S.; Wang, Y.; Chen, W.; Lyu, Y.; Dungan, B.; Carroll,
  645 K. C.; Holguin, F. O. Comprehensive retention model for PFAS transport in subsurface
  646 systems. *Water Res.* 2019, *148*, 41–50.
- (69) DeWitt, J. C. (Ed). *Toxicological Effects of Perfluoroalkyl and Polyfluoroalkyl Substances*;
   Molecular and Integrative Toxicology; Humana Press, 2015.
- 649 (70) Wang, Z.; DeWitt, J. C.; Higgins, C. P.; Cousins, I. T. A never-ending story of per- and polyfluoroalkyl substances (PFASs)? *Environ. Sci. Technol.* 2017, *51* (5), 2508–2518.

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