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DOMOIC ACID

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Final Report of the Workshop

Oregon Institute of Marine Biology
February 21-23, 1992

Edited by A. Michelle Wood
and Lynda M. Shapiro



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Oregon State University
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THE WORKSHOP REPORT

Introduction

Amnesic shellfish poisoning (ASP) is a recently identified threat to the seafood consuming public. It is caused by the neurotoxin domoic acid, which can be concentrated by filter-feeding shellfish as they consume smaller organisms that produce the toxin. The first incidence of a human disease caused by domoic acid in shellfish was documented in 1987 on Prince Edward Island in eastern Canada. Scientists there identified the causative organism and helped develop a successful management strategy. This has allowed the shellfish industry there to continue to grow, and its harvest of cultured mussels there now exceeds 1987 levels. Despite the continued annual appearance of the diatom that produces domoic acid, tainted products have been kept off the market and public confidence in the Prince Edward Island mussel industry is high. This regional success story suggests that the domoic acid problems that emerged on the west coast of the United States in the fall of 1991 can be managed equally well.

The west coast incident started with the deaths of pelicans and cormorants in Monterey Bay in September 1991. Extensive testing ruled out pollutants, pesticides, bacteria, viruses, and heavy metals, but the behavior of mice being tested for possible paralytic shellfish poisoning (PSP) indicated that domoic acid was present in the birds. In late October, domoic acid was discovered in razor clams from the surf zone of Oregon and Washington coastal beaches, causing the closure of both the commercial and recreational fisheries. By early December, Dungeness crabs in California, Oregon, and Washington were found to contain the toxin, and that fishery was also closed. However, other clam species, mussels, and oysters never became toxic.

In response to these events, Sea Grant sponsored a workshop on domoic acid at the

University of Oregon's Institute of Marine Biology, February 20–23, 1992. The principal purpose was to identify scientific issues that must be addressed in order to achieve a successful management program. We recognize that a new public health threat of this type involves both an immediate, emergency response and a longer-term response. Closures and enhanced monitoring of the fisheries were some of the immediate responses. Our goal at the February meeting was to consider longer-term responses.

Participants at the workshop included some scientists who have studied problems related to domoic acid since the first outbreaks in Canada and many west coast scientists who have been part of the immediate response efforts in California, Washington, and Oregon. Among them were experts in toxic phytoplankton, domoic acid-producing phytoplankton, phytoplankton ecology, shellfish biology, molecular biology, and physical oceanography, as well as representatives from government agencies and industry. (See Appendix 1 for a list of participants and their affiliations.)

The participants identified three scientific priorities for a research plan that will provide the information needed to develop an optimal strategy for managing domoic acid. First, we need to determine the source of domoic acid and the mechanisms by which it accumulates in shellfish and other organisms. While workshop participants recommended continued focus on diatoms in the *Nitzschia/Pseudonitzschia* complex as the most likely source of domoic acid, sufficient differences exist between the domoic acid outbreaks in California, Oregon-Washington, and Canada to require considering the possibility that the source of domoic acid differs in all three areas.

Second, there is an amazingly small body of basic ecological information about phytoplankton dynamics off the Pacific Northwest coast. Process-oriented studies, which condensed

information on phytoplankton into taxa-insensitive measures such as chlorophyll concentration and rate of total primary production, have dominated oceanographic research off the northwest Pacific coast for some time. As a result, we have little information on phytoplankton species composition and on processes determining the abundance and distribution of species in coastal waters. Responses to the domoic acid problem and to other nuisance phytoplankton taxa that may arise in the future require an immediate and sustained effort to obtain this information. Further, the work must proceed on ecologically meaningful spatial and temporal scales despite the fact that individual states have different fisheries priorities and different mechanisms for supporting the research. Because the California Current links ecologically similar coastal environments from southern Alaska to Monterey Bay, investigation of phytoplankton species distribution and population dynamics must consider this regional ecosystem as well as local subsystems.

Third, all working groups recognized the need for a straightforward assay for domoic acid that can be applied to tissue samples as well as to water and sediment samples. An assay that could reliably, inexpensively, and rapidly indicate the presence of domoic acid at low levels and provide rough estimates of concentration at higher levels would be of tremendous value for both monitoring and research applications, even if it lacked the precision of HPLC-based methods.

Additionally, the participants recognized the need for training in methods of detection and standardization of detection methods throughout the affected area.

During the second day of the workshop, we met in working groups to draft research recommendations. The three working groups were (1) Phytoplankton, (2) Public Health and Detection, and (3) Shellfish. The separate reports of the working groups follow. Where the separate groups arrived at identical recommendations, we have allowed the redundancy to remain because the emphasis it conveys reflects the consensus of the participants.

Report of the Phytoplankton Working Group

Peter Franks (Oregon State University), Greta Fryxell (Texas A&M University), Lynda Shapiro (University of Oregon), and Michelle Wood (University of Oregon)

Working groups discussed phytoplankton ecology in the morning session and phytoplankton culture studies during the afternoon session. The results of these deliberations have been integrated and are presented below.

It is generally agreed that the question with highest priority is, What organisms are the source of domoic acid? This question must be answered before other high priority questions concerning the ecophysiology of domoic acid production and the ecology of the blooms can be addressed.

In eastern Canada, where clear cases of amnesic shellfish poisoning were documented in people who had eaten domoic acid-containing mussels, researchers have established that the causative organism is a form of the chain-forming diatom *Nitzschia pungens* (forma *multiseries*). In this taxon, studies with unialgal cultures indicate that domoic acid production increases with increasing senescence; nitrate and light appear to be necessary for domoic acid production, with cell growth limited by some other factor. Toxicity of isolates appears to decrease as the cells are maintained in culture for longer than a year, and some preliminary data show that levels of domoic acid production decrease in axenic culture relative to that of unialgal cultures that are not bacteria-free.

In Monterey Bay, no human health effects were observed, but pelican mortality was traced to the ingestion of anchovies that had been feeding on the chain-forming diatom *Pseudonitzschia australis*, the dominant member of the phytoplankton community at the time. Preliminary data from studies with unialgal (but not bacteria-free) cultures of *P. australis* indicate that this organism produces domoic acid. Research on *P. australis* is obviously more limited than research on *N. pungens* forma *multiseries* since its possible role as a toxic agent was not suspected until fall 1991.

The Monterey Bay pelican kill is, to date, the only example of transmission of toxic levels of domoic acid to vertebrates through a planktivorous fish vector.

Finally, in Oregon and Washington, where mild cases of neurologic disorder and gastric distress in humans have been traced to consumption of razor clams containing domoic acid, data on the phytoplankton species composition at the time of harvest are unavailable. However, *P. australis*, and *N. pungens* cells, with morphologies that conform to species descriptions for both the toxic and nontoxic form, have been obtained in phytoplankton samples collected from the region after the detection of domoic acid in shellfish. These taxa did not dominate the phytoplankton community, even at sites where the domoic acid concentration in razor clam tissue continued to increase. (This is a common feature of many nuisance blooms.)

Field and laboratory studies, as well as common sense, argue strongly that phytoplankton, probably *N. pungens* f. *multiseries*, *P. australis*, or close relatives, are the sources of domoic acid in fish and shellfish on the west coast of the United States. Because of the potential economic and human health problems associated with shellfish that has been contaminated with domoic acid, research on the environmental factors that control domoic acid production in *Nitzschia* and *Pseudonitzschia* species should be given high priority.

The differences between health and food chain effects observed in California, Washington-Oregon, and eastern Canada make it necessary to consider simultaneously the possibility that other organisms, either singly or in addition to *N. pungens* and *P. australis*, are responsible for the presence of domoic acid in shellfish and anchovies. In particular, other *Nitzschia* species, that is *N. delicatula* and *N. pseudodelicatissima*, may be capable of domoic acid production. It is important to examine dominant members of the surf zone phytoplankton community, epiphytes on kelp and celgrass, and decaying beach wrack as potential sources of domoic acid. The diatom *Amphora*

coffeaeformis is a suspected producer of domoic acid, and it may be part of the epiphyte community. Both its presence and its toxicity should be confirmed. Also the red alga *Chondria armata*, thought to be a relatively rare taxon on the Washington-Oregon coast, produces domoic acid and therefore deserves some additional study of its distribution and abundance. It should be noted that the affected shellfish in Washington and Oregon coastal waters live in more exposed environments than Monterey Bay or the embayments of Prince Edward Island.

As organisms that produce domoic acid are identified, the following questions take on the highest priority :

1. What factors govern domoic acid production?
2. What is the life history of each organism that produces domoic acid, and are there particular life history stages involved in domoic acid production or particular life history stages that can be exploited in a management program?
3. What is the physical ecology of population increases (for example, blooms) of the domoic acid-producing species?

Studies of the physical ecology of blooms of domoic acid-producing species must consider a number of topics: the environmental factors that govern species abundance and toxin production; the regularity, both temporal and spatial, of blooms of domoic acid-producing species; and the heterogeneity of the bloom community. Also, it is possible that the domoic acid-producing taxa have been introduced by human activities such as shipping; this possibility and its implications should be considered. A critical aspect of much of this work will be the identification of known domoic acid producers in field samples containing a large number of species. This is likely to be a difficult and time-consuming task. Therefore, we suggest that an effort be made to develop genetic probes that will distinguish between species in the field. In addition, an inexpensive and rapid domoic acid assay would allow easy identification of toxic waters.

The most fundamental aspect of bloom studies will be the development of a good sampling program. It is imperative that we understand the geographic scope of domoic acid outbreaks and the temporal range in which they occur. We must gain knowledge of the organism's life history to understand its distribution alongshore and offshore, the factors leading to domoic acid production, and the mechanisms of delivery to the amnesic shellfish poisoning vectors (for example, anchovy, razor clams, crabs, mussels). The plankton sampling must be integrated with hydrographic sampling (temperature, salinity and nutrient profiles at a minimum, also *in situ* fluorescence, beam transmittance, and currents). The studies should also consider historical data to examine environmental forcing of domoic acid outbreaks. Field studies should span a wide geographic area (southern California to northern Washington at minimum, but preferably including British Columbia and Alaska) to compare and seek links between adjacent outbreaks. Temporal coverage should be fine enough to identify the initiation of toxic outbreaks so that a sequence of events can be identified.

Laboratory work should examine the biochemical and genetic aspects of domoic acid production, the phylogenetic relationships of domoic acid-producing species, and particularly, the responses of these species to a variety of environmental stresses—for example, nutrient and light limitation and temperature effects. These experiments should also consider the role of other organisms (for example, bacteria) in domoic acid production and work toward understanding the genetic basis for the biosynthesis of domoic acid.

Finally, we must gain more knowledge of the vectors of domoic acid transmission. In particular, what can the feeding ecology of these organisms tell us about the toxic blooms? How is domoic acid distributed throughout the food web? And can these organisms take up dissolved domoic acid as opposed to actually ingesting toxic cells?

To better understand the dynamics of domoic acid outbreaks, the group identified special needs:

1. Sampling schemes that provide temporal or spatial surveys of plankton and hydrography
2. Establishment of additional new cultures of both toxic and nontoxic species.
3. More extensive culture studies
4. A "quick-and-dirty" assay for domoic acid
5. Elucidation of the biochemical pathway and genetic components leading to domoic acid production
6. Field studies of domoic acid "hot spots"
7. Preservation of field samples and subsequent gut analysis where appropriate

The working groups gave high priority to a recommendation for the establishment of an integrated sampling program for toxic algal blooms, including those producing paralytic shellfish poisoning, amnesic shellfish poisoning, and diarrhetic shellfish poisoning. This sampling program ideally would span from southern California to Alaska, including the coast of British Columbia. It is likely that novel toxic outbreaks will continue to occur. Such a sampling program will create a sample archive that would provide necessary information to researchers trying to understand the courses of events, and could aid in a fast, coordinated effort at protecting the public and the shellfish industry from harm.

Report of the Public Health and Detection Working Group

Bill Keene (Oregon Health Division) and John Wekell (National Marine Fisheries Service)

Overview of Monitoring Needs

- 1) There is an immediate need to protect consumers from the danger of acute illness and simultaneously to allow seafood to move commercially without (undue) hindrance, that is, by certifying it as "safe" (or at least as being below some recognized action level of contamination).
- 2) There is also a need for more research-oriented programs, designed to more efficiently or more sensitively predict and detect domoic acid

occurrences. A better understanding of the underlying phenomena will also improve planning capabilities of public health officials and of industry and consumers.

Routine Monitoring

Although these components need not be completely independent, it should be understood that a preliminary monitoring program is already in place, albeit in an ad hoc manner, and it is rapidly evolving under the aegis of the FDA and the respective state shellfish programs to become a routine program that will be managed by the states.

There are few significant variations among the several draft monitoring programs on the table in Washington, Oregon, and California. They are wholly analogous to pre-existing PSP programs, consisting of

- a. routine collection of commercially and (to a lesser extent) recreationally important species, especially oysters, mussels, clams, and crab, consumed by humans
- b. HPLC sampling of specimens for domoic acid by FDA, state, or other laboratories
- c. dissemination of results to interested parties, with regulatory action taken as levels reach (or approach) 20 ppm
- d. intensified monitoring activities as levels begin to rise significantly ("significant" to be defined later, but presumably at inflection points on any domoic acid level vs. time plot, for example, a shift from zero to 2–5 ppm)

These routine monitoring programs will be a reasonable source of baseline data (on those species in sampled areas, at least), but efforts to develop alternative monitoring programs should be designed as stand-alone scientific projects.

Research Needs for Domoic Acid Monitoring

Research efforts should concentrate on identifying and evaluating potential invertebrate and vertebrate indicator species or environmental conditions that might be cheaper or better than

the "brute force" method of testing food before people eat it. Seasonality or other cyclic phenomena may be important aspects of domoic acid occurrence and should be a focus of attention in all spheres of research.

To identify indicator species, we must begin with a broad effort to characterize the uptake and turnover of domoic acid by organisms in the food web. In addition to the commercially important species mentioned above, this effort should include finfish, other mollusks (including carnivores and snails), and krill. Uptake and turnover of domoic acid should be investigated both by species and by organ within each type of animal. It is anticipated that these efforts will be largely laboratory studies, but that they will be supplemented with field data collected during blooms of domoic acid-producing phytoplankton. The main purpose of these studies is to identify indicator or "sentinel" species in which the time frame of both uptake and retention are in appropriate balance.

Other desirable characteristics for indicator species are

- cosmopolitan distribution
- relative abundance
- ease of collection
- primary filter feeder (presumably)
- ease of domoic acid analysis

Mussels and barnacles were suggested as possibilities, assuming that uptake and retention studies gave promising results. (Of course, available evidence suggests that mussels of the Pacific Northwest may not take up or may turn over domoic acid too rapidly). Probably no single species will prove entirely satisfactory.

Direct Monitoring of Domoic Acid in Water

Research to develop simple, rapid, cheap methods for domoic acid detection would be very useful. This is primarily a biochemical problem. Having such a capability could prove of great value to biologists as well as to those monitoring for public health reasons.

Plankton Monitoring

Researchers in Canada have been highly successful in monitoring plankton. They have

provided an early warning of impending toxicity in mussels by monitoring the development of blooms of domoic acid-producing phytoplankton species. In only a few years, Canadians have collected an impressive data base that is already proving of great practical value. Work should begin along comparable lines here.

This approach obviously depends on confirmation of the domoic acid-producing species and a vastly improved ability to survey, identify, and quantify phytoplankton off our shores. Relevant research topics are indicated above, in the Phytoplankton Working Group, and are not repeated here. The applicability of the Canadian model of course rests on an improved understanding of the ecological dynamics of the species and of domoic acid production in both locales.

Specific research goals are to standardize plankton collection methods and to improve identification methods, particularly of toxic or potentially toxic species (for example, immunological techniques and molecular probes).

Biomedical Research

There is a major need for extensive research on the toxicology of domoic acid and the medical risk factors associated with illness, including both chronic and acute effects. Comprehensive evaluation of research needs in this area lies outside the sphere of marine biology, but two topics did arise during the workshop that should be conveyed to biomedical researchers investigating the domoic acid problem.

First, long-term or chronic health risks associated with multiple exposure to low levels of domoic acid are unknown. Consideration of this aspect is especially important for risk evaluation. Present permissible levels of domoic acid in shellfish are based on our knowledge of the acute response to relatively high levels of domoic acid. If domoic acid toxicity can also result from the cumulative effects of multiple exposures, then current standards may be inadequate to protect public health.

Second, the neurotoxic effects of domoic acid poisoning occur in the brain. This indication that the toxin can cross the blood-brain barrier leads to the reasonable conclusion that it may cross

other blood-tissue barriers. If so, then fetuses and pregnant women may be particularly vulnerable to domoic acid poisoning. An effort should be made to determine whether or not domoic acid can pass the placental barrier and, if so, what effect it has.

Epidemiology

Epidemiological studies may not be appropriate for Sea Grant, and, in any event, there are not many prospects for useful studies at this point. The relative risk of illness associated with exposure to available products is almost certainly so low as to demand impossibly large sample sizes for detection. Surveillance is difficult because of the impossibility of confirming the diagnosis under virtually any realistic conditions.

Report of the Shellfish Working Group

Chris Langdon (Oregon State University) and Neil Richmond (Oregon Department of Fish and Wildlife)

Judging from the experience of the Canadians, it is likely that domoic acid outbreaks will recur in subsequent years. It is clearly necessary to begin studies that will ultimately reduce health risks to shellfish consumers and reduce the adverse economic effects on recreational and commercial shellfisheries. The shellfish workshop consisted of representatives from the crab and oyster industries, state agencies (the Oregon Department of Fisheries and Wildlife and the Oregon Health Division), federal agencies (the Food and Drug Administration and the National Marine Fisheries Service), and academia. Four research areas that should be addressed to achieve these objectives were identified and prioritized.

1) The first priority is to establish a comprehensive shellfish-monitoring program on the west coast of the United States and Canada. The aim of this program would be to determine which species of shellfish are susceptible to domoic acid accumulation, with emphasis placed on edible species such as razor clams (deep and shallow water species), oysters, mussels, clams, Dungeness crab, sea urchins, and some fish species such as anchovy. From this information, indicator

species would be identified that could act as early-warning sentinels of domoic acid in the coastal environment. This information would be useful to state and federal health agencies who determine human health risks from the consumption of shellfish.

In addition, it is anticipated that the shellfish- and phytoplankton-monitoring programs be closely integrated, so that correlations can be made between oceanographic conditions, phytoplankton, and shellfish in regard to the spatial and temporal distribution of domoic acid in the coastal environment. Such an integrated program may lead to a comprehensive monitoring system that would provide shellfish harvesters, growers, and regulators with early warnings of potential domoic acid events. This type of program has been very effective in eastern Canadian waters in providing shellfish growers with almost two weeks of warning of domoic acid events, allowing growers to take precautionary measures, such as switching their harvesting to domoic acid-free estuaries.

2) Development of seafood processing methods that reduce concentrations of domoic acid in seafood would allow processors to minimize the health risks associated with the consumption of seafood contaminated by domoic acid. Research and testing should identify simple, economical procedures, such as depuration and frequent change of water used to boil crab meat.

3) We need to understand the mechanisms that determine the accumulation and fate of domoic acid in shellfish and fish. Such basic research would include studies of the feeding behaviors, uptake mechanisms, and rates of domoic acid accumulation from both particulate (phytoplankton, prey species, etc.) and dissolved phases. (In fact, the possibility of direct uptake of domoic acid from seawater has not, to our knowledge, been tested.) Investigations should include subsequent biochemical transformations in the tissues of shellfish and excretion of derivatives of domoic acid. Mechanisms and rates of loss of domoic acid from different species of shellfish would be of interest to biologists and would also form a basis for depuration procedures of domoic

acid-contaminated shellfish that could be adopted by industry.

It is also noteworthy that razor clams incorporate the domoic acid into body tissues (for example, foot and mantle), whereas mussels accumulate the toxin in digestive glands and rapidly depurate when placed in toxin-free water. Similarly, living Dungeness crabs probably accumulate the toxin only in the viscera, although toxin can enter crab meat during cooking if the crabs are not eviscerated before cooking. Depuration rates for razor clams and crabs are not known.

4) Domoic acid is highly toxic to mammals and is used as an insecticide and vermifuge in Japan. However, there have been no reports of adverse effects of domoic acid on the biology of shellfish, other marine invertebrates, or fish. Basic neurophysiological studies are required to understand why marine invertebrates, and possibly fish, appear immune to the effects of domoic acid. It is known that domoic acid mimics some neurotransmitter, most likely a glutamate type. Because it is water soluble, depuration is usually fast. However, the foot muscle of razor clams does not appear to depurate, and there does not appear to be a toxic effect. Some crustaceans are known to have glutamate receptors, yet crabs and other crustaceans do not appear to be affected. The answers to puzzles such as these are necessary if we are to identify indicator species. They are also likely to provide some basic comparative information on the mechanisms of neurotransmission.

5) Long-term, chronic health risks associated with consumption of low levels of domoic acid are also unknown. This aspect is especially important for setting the permitted level of domoic acid in tissue. If the toxicity is accumulative, then one could not help but worry about the levels currently being approved on the basis of acute responses in comparison to chronic dosages.

Such studies may lead to an increased understanding of the functioning of the nervous systems of animals and may result in the development of medical uses of domoic acid that would benefit human health or well-being.

Summary and Conclusions

The activities of utmost priority identified during the ASP Workshop are

1. to identify the source of domoic acid
2. to elucidate the factors governing domoic acid production
3. to establish a phytoplankton field study examining the processes that determine abundance and distributions of species
4. to establish routine toxicity monitoring in appropriate indicator species

Knowledge of the source of domoic acid, combined with information concerning its production, both spatially and temporally, may lead to a predictive capability for ASP outbreaks. While the predictions of outbreaks may be only one to two weeks in advance, this is sufficient time to modify harvesting procedures and protect the public, as shown by the Canadian example. The history of nuisance blooms along this coast and in most other regions of the world suggests that ASP will be a recurrent problem. By understanding the problem now, we may be able to save time, money, and lives in the future.

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Participants with an asterisk (*) before their name either helped write this document or commented on it.

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APPENDIX 2: DOMOIC ACID AND PSEUDONITZSCHIA REFERENCES

This bibliography was compiled by Steve Bates, of the Department of Fisheries and Oceans, New Brunswick, Canada. If you are aware of any new literature or additional older material that is relevant, please contact Dr. Bates at the following address so that he can maintain this list.

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Domoic Acid Analysis and Chemistry

- Bates, S.S., et al. 1989. The role of mass spectrometry combined with reversed phase HPLC in the identification of an unusual shellfish toxin, p. 1394-1395. P. Longevialle (ed.) *Advances in mass spectrometry*. Heyden & Son, Ltd., London.
- Bird, C.J., et al. 1988. Identification of domoic acid as the toxic agent responsible for the P.E.I. contaminated mussel incident. *Atlantic Research Lab. Tech. Rep.* 56.
- Boyd, R., et al. 1988. The role of mass spectrometry combined with reversed phase HPLC in the identification of an unusual shellfish toxin. *Proc. Am. Soc. Mass. Spec.* 36: 165.
- Dhoot, J.S., et al. 1992. An improved HPLC procedure for domoic acid analysis in seafood. *Intern. J. Environ. Anal. Chem.* (in press).
- Dickey, R. W., et al. 1992. Detection of the marine toxins okadaic acid and domoic acid in shellfish and phytoplankton in the Gulf of Mexico. *Toxicon* 30: 355-359.
- Falk, M. 1988. The infrared spectrum of domoic acid. *Can. J. Spectroscopy.* 33: 117-121.
- Falk, M., J.A. Walter, and P.W. Wiseman. 1989. Ultraviolet spectrum of domoic acid. *Can. J. Chem.* 67: 1421-1425.
- Falk, M., P.F. Seto, and J.A. Walter. 1991. Solubility of domoic acid in water and non-aqueous solvents. *Can. J. Chem.* 69: 1740-1744.
- Falk, M., et al. 1991. NMR study of domoic acid. Fourth European conference on spectroscopy of biological molecules; York, UK, 1991. *Spectrosc. Biol. Mol.* 94: 323-324.
- Hardstaff, W.R., et al. 1990. Reference materials for domoic acid, a marine neurotoxin. *J. Anal. Chem.* 338: 520-525.
- Hungerford, J.M. 1993. Seafood toxins and seafood products. *J. Assoc. Offic. Anal. Chem.* 76: 120-130.
- Lawrence, J.F., et al. 1989. Liquid chromatographic determination of domoic acid in shellfish products using the paralytic shellfish poison extraction procedure of the Association of Official Analytical Chemists. *J. Chromatogr.* 462: 349-356.
- Lawrence, J.F., et al. 1989. Confirmation of domoic acid in molluscan shellfish by chemical derivatization and reversed-phase liquid chromatography. *J. Chromatogr.* 462: 419-425.
- Lawrence, J.F., and C. Ménard. 1991. Confirmation of domoic acid in shellfish using butyl isothiocyanate and reversed-phase liquid chromatography. *J. Chromatogr.* 550: 595-601.
- Lawrence, J.F., C.F. Charbonneau, and C. Ménard. 1991. Liquid chromatographic determination of domoic acid in mussels, using AOAC analytic shellfish poison extraction procedure: collaborative study. *J. Assoc. Offic. Anal. Chem.* 74: 68-72.
- Newsome, H., et al. 1991. Determination of domoic acid in serum and urine by immuno-

- chemical analysis. *Bull. Environ. Contam. Toxicol.* 47: 329-334.
- Ohfuné, Y., and M. Tomita. 1982. Total synthesis of (-)-domoic acid. A revision of the original structure. *J. Am. Chem. Soc.* 104: 3511-3513.
- Pleasance, S., et al. 1990. Analysis of domoic acid and related compounds by mass spectrometry and gas chromatography/mass spectrometry as N-trifluoroacetyl-O-silyl derivatives. *Biomed. Environ. Mass Spectrom.* 19: 420-427.
- Pleasance, S., P. Thibault, and J. Kelly. 1992. Comparison of liquid-junction and coaxial interfaces for capillary electrophoresis-mass spectrometry with application to compounds of concern to the aquaculture industry. *J. Chromatogr.* 591: 325-339.
- Pocklington, R., et al. 1990. Trace determination of domoic acid in seawater and phytoplankton by high-performance liquid chromatography of the fluorenylmethoxycarbonyl (FMOC) derivative. *Intern. J. Environ. Anal. Chem.* 38: 351-368.
- Quilliam, M.A., et al. 1988. Determination of domoic acid in shellfish tissue by high-performance liquid chromatography. Atlantic Research Laboratory Technical Report 55.
- Quilliam, M.A., et al. 1989. High-performance liquid chromatography of domoic acid, a marine neurotoxin, with application to shellfish and plankton. *Internat. J. Environ. Anal. Chem.* 36: 139-154.
- Quilliam, M.A., et al. 1989. Ion-spray mass spectrometry of marine neurotoxins. *Rapid Comm. Mass Spectrom.* 3: 145-150.
- Quilliam, M.A., et al. 1992. Recent developments in instrumental analytical methods for marine toxins, p. 376-386. In: E.G. Bligh (ed.) *Seafood science and technology*. Fishing News Books, Oxford, England.
- Quilliam, M.A., and S. Pleasance. 1991. Liquid chromatography/mass spectrometry for the analysis of marine toxins, p. 131-135. In: J.M. Frey (ed.) *Proceedings of the international symposium on marine biotoxins*. CNEVA Pub. Series, Maisons-Alfort, Paris, France.
- Quilliam, M.A., M. Xie, and W.R. Hardstaff. 1991. A rapid extraction and clean-up procedure for the determination of domoic acid in tissue samples. Institute for Marine Bio-
- sciences Technical Report 64, National Research Council of Canada NRC 33001, 52 p.
- Shimizu, Y., S. Gupta, K. Masuda, L. Maranda, C.K. Walker, and R. Wang. 1989. Dinoflagellate and other microalgal toxins: chemistry and biochemistry. *Pure & Appl. Chem.* 61: 513-516.
- Thibault, P., et al. 1989. Mass spectrometry of domoic acid, a marine neurotoxin. *Biomed. Environ. Mass Spectrom.* 18: 373-386.
- Wright, J.L.C., et al. 1989. Identification of domoic acid, a neuroexcitatory amino acid, in toxic mussels from eastern Prince Edward Island. *Can. J. Chem.* 67: 481-490.
- Wright, J.L.C., et al. 1990. Identification of isodomoic acid D and two new geometrical isomers of domoic acid in toxic mussels. *J. Chem.* 68: 22-25.

Physiology, Ecology, Morphology, and Taxonomy of *Pseudonitzschia*

- Bates, S.S., et al. 1988. Investigations on the source of domoic acid responsible for the outbreak of amnesic shellfish poisoning (ASP) in eastern Prince Edward Island. Atlantic Research Laboratory Technical Report 57.
- Bates, S.S., et al. 1989. Pennate diatom *Nitzschia pungens* as the primary source of domoic acid, a toxin in shellfish from eastern Prince Edward Island, Canada. *Can. J. Fish. Aquat. Sci.* 46: 1203-1215.
- Bates, S.S., et al. 1991. Controls on domoic acid production by the diatom *Nitzschia pungens* f. *multiseries* in culture: nutrients and irradiance. *Can. J. Fish. Aquat. Sci.* 48: 1136-1144.
- Bates, S.S., et al. 1993. Discrimination between domoic-acid-producing and non-toxic forms of the diatom *Nitzschia pungens* using immunofluorescence. *Mar. Ecol. Progr. Ser.* (submitted).
- Bates, S.S., J. Worms, and J.C. Smith. 1993. Effects of ammonium and nitrate on domoic acid production by *Nitzschia pungens* in batch culture. *Can. J. Fish. Aquat. Sci.* (in press).
- Buck, K.R., et al. 1992. Autecology of the diatom *Pseudonitzschia australis* Frenguelli, a domoic acid producer, from Monterey Bay, California. *Mar. Ecol. Progr. Ser.* 84: 293-302.

- Douglas, D.J. 1991. Axenic cultures of the marine diatom *Nitzschia pungens* f. *multiseries* produce the neurotoxin domoic acid. *Appl. Phycol. Forum* 8: 10.
- Douglas, D.J., and S.S. Bates. 1992. Production of domoic acid, a neurotoxic amino acid, by an axenic culture of the marine diatom *Nitzschia pungens* f. *multiseries* Hasle. *Can. J. Fish. Aquat. Sci.* 49: 85-90.
- Douglas, D.J., et al. 1992. Biosynthesis of the neurotoxin domoic acid by the marine diatom *Nitzschia pungens* forma *multiseries*, determined with [¹³C]-labelled precursors and nuclear magnetic resonance. *J. Chem. Soc. Chem. Commun.* 1992: 714-716.
- Douglas, D.J., et al. 1993. Domoic acid production by axenic and non-axenic cultures of the pennate diatom *Nitzschia pungens* f. *multiseries*, p. 595-600. In: T.J. Smayda and Y. Shimizu (eds.) *Toxic phytoplankton blooms in the sea*. Elsevier Sci. Publ. B.V., Amsterdam.
- Forbes, J.R., and K.L. Denman. 1991. Distribution of *Nitzschia pungens* in coastal waters of British Columbia. *Can. J. Fish. Aquat. Sci.* 48: 960-967.
- Fritz, L., et al. 1992. An outbreak of domoic acid poisoning attributed to the pennate diatom *Pseudonitzschia australis*. *J. Phycol.* 28: 439-442.
- Fryxell, G.A., M.E. Reap, and D.L. Valencic. 1990. *Nitzschia pungens* Grunow f. *multiseries* Hasle: observations of a known neurotoxic diatom. *Beih. Nova Hedwigia* 100: 171-188.
- Fryxell, G.A., S.A. Garza, and D.L. Roelke. 1991. Auxospore formation in an Antarctic clone of *Nitzschia subcurvata* Hasle. *Diatom Res.* 6: 235-245.
- Garrison, D.L., et al. 1992. Confirmation of domoic acid production by *Pseudonitzschia australis* (Bacillariophyceae) cultures. *J. Phycol.* 28: 604-607.
- Hargraves, P.E., et al. 1993. Growth characteristics and toxins in *Pseudonitzschia pungens* and related taxa in ultraviolet light, (in press). In: H. van Dam (ed.) *Proceedings of the 12th international diatom symposium*, Hydrobiol.
- Hasle, G.R. 1964. *Nitzschia* and *Fragilariopsis* species studied in the light and electron microscopes. I. Some marine species of the groups *Nitzschia* and *Lanceolatae*. *Skr. Norske Vidensk-Akad. I. Mat.-Nat. Kl. Ny Series* 16: 1-48.
- Hasle, G.R. 1965. *Nitzschia* and *Fragilariopsis* species studied in the light and electron microscopes. II. The group *Pseudonitzschia*. *Skr. Norske Vidensk-Akad. I. Mat. -Nat. Kl. Ny Series* 18: 1-45.
- Hasle, G.R. 1993. Nomenclatural notes on marine planktonic diatoms. The family Bacillariacea. *Beih. Nova Hedwigia* 106: (in press). [Indicates change in nomenclature of some species of the genus *Nitzschia* to the genus *Pseudonitzschia*]
- Homer, R.A., and J.R. Postel. 1993. Toxic diatoms in western Washington waters, (in press). In: H. van Dam (ed.) *Proceedings of the 12th international diatom symposium*, Hydrobiol.
- Jackson, A.E. 1992. The effect of salinity on growth and amino acid composition in the marine diatom *Nitzschia pungens*. *Can. J. Bot.* 70: 2198-2201.
- Lewis, N.I., et al. 1993. Temperature effects on growth, domoic acid production, and morphology of the diatom *Nitzschia pungens* f. *multiseries*, p. 601-606. In: T.J. Smayda and Y. Shimizu (eds.) *Toxic phytoplankton blooms in the sea*. Elsevier Sci. Publ. B.V., Amsterdam.
- MacPhee, D.J., et al. 1992. Morphology of the toxin-producing diatom *Nitzschia pungens* Grunow forma *multiseries* Hasle. *Can. J. Fish. Aquat. Sci.* 49: 303-311.
- Martin, J.L., et al. 1990. *Nitzschia pseudodelicatissima*—a source of domoic acid in the Bay of Fundy, eastern Canada. *Mar. Ecol. Progr. Ser.* 67: 177-182.
- Martin, J.L., K. Kaya, and D.J. Wildish. 1993. Distribution and domoic acid content of *Nitzschia pseudodelicatissima* in the Bay of Fundy, p. 613-618. In: T.J. Smayda and Y. Shimizu (eds.) *Toxic phytoplankton blooms in the sea*. Elsevier Sci. Publ. B.V., Amsterdam.
- Pan, Y., D.V. Subba Rao, and R.E. Warnock. 1991. Photosynthesis and growth of *Nitzschia pungens* f. *multiseries* Hasle, a neurotoxin producing diatom. *J. Exp. Mar. Biol. Ecol.* 154: 77-96.

- Pan, Y., et al. 1993. Temperature dependence of growth and carbon assimilation in *Nitzschia pungens* f. *multiseries*, the causative diatom of domoic acid poisoning, p. 619-624. In: T.J. Smayda and Y. Shimizu (eds.) Toxic phytoplankton blooms in the sea. Elsevier Sci. Publ. B.V., Amsterdam.
- Parrish, C.C., et al. 1990. Unusual fatty acid composition of the toxic marine diatom *Nitzschia pungens*. Bull. Aquacult. Assoc. Can. 30: 15-18.
- Parrish, C.C., et al. 1991. Lipid composition of the toxic marine diatom, *Nitzschia pungens*. Phytochem. 30: 113-116.
- Pauley, K.E., et al. 1993. Occurrences of phycotoxins and related phytoplankton at winter temperatures in the southeastern Gulf of St. Lawrence, Canada, p. 311-316. In: T.J. Smayda and Y. Shimizu (eds.) Toxic phytoplankton blooms in the sea. Elsevier Sci. Publ. B.V., Amsterdam.
- Reap, M.E. 1991. *Nitzschia pungens* Grunow f. *multiseries* Hasle: growth phases and toxicity of clonal cultures isolated from Galveston Bay, Texas. M. Sc. Thesis, Texas A&M University. 78 p.
- Rivard, L. 1992. The effect of temperature on enzyme activity in *Nitzschia pungens* forma *pungens* and forma *multiseries*. Honours Thesis, Mount Allison University, Sackville, New Brunswick, Canada. 31 p.
- Rosowski, J.R., L.M. Johnson, and D.G. Mann. 1992. On the report of gametogenesis, oogamy, and uniflagellated sperm in the pennate diatom *Nitzschia pungens* (1991). J. Phycol. 27: 21-26). J. Phycol. 28: 570-574.
- Seguel, M.R. 1991. Interactive effects of temperature-light and temperature-salinity on growth of five phytoplanktonic species isolated from a shallow-water embayment of Nova Scotia. M.Sc. Thesis, Acadia University, Wolfville, Nova Scotia, Canada. 218 p.
- Smith, J.C., et al. 1990. Toxic blooms of the domoic acid containing diatom *Nitzschia pungens* in the Cardigan River, Prince Edward Island, p. 227-232. In: E. Granéli et al. (eds.) Toxic marine phytoplankton. Elsevier Sci. Publ. Co., Inc., New York.
- Smith, J.C., et al. 1990. Variation in domoic acid levels in *Nitzschia* species: implications for monitoring programs. Bull. Aquacult. Assoc. Can. 90-4: 27-31.
- Smith, J.C., et al. 1993. Growth and domoic acid production and retention by *Nitzschia pungens* forma *multiseries* at low temperatures, p. 631-636. In: T.J. Smayda and Y. Shimizu (eds.) Toxic phytoplankton blooms in the sea. Elsevier Science Publishers B.V., Amsterdam.
- Subba Rao, D.V., M.A. Quilliam, and R. Pocklington. 1988. Domoic acid—a neurotoxic amino acid produced by the marine diatom *Nitzschia pungens* in culture. Can. J. Fish. Aquat. Sci. 45: 2076-2079.
- Subba Rao, D.V., et al. 1990. Rates of production of domoic acid, a neurotoxic amino acid in the pennate marine diatom *Nitzschia pungens*, p. 413-417. In: E. Granéli et al. (eds.) Toxic marine phytoplankton. Elsevier Sci. Publ. Co., Inc., New York.
- Subba Rao, D.V., and G. Wohlgeschaffen. 1990. Morphological variants of *Nitzschia pungens* Grunow f. *multiseries* Hasle. Bot. Mar. 33: 545-550.
- Subba Rao, D.V., et al. 1991. Flow cytometry and microscopy of gametogenesis in *Nitzschia pungens*, a toxic, bloom-forming, marine diatom. J. Phycol. 27: 21-26.
- Subba Rao, D.V., et al. 1992. Gametogenesis in *Nitzschia pungens* f. *multiseries*. J. Phycol. 28: 574-576.
- Takano, H., and K. Kuroki. 1977. Some diatoms in the Section *Pseudonitzschia* found in coastal waters of Japan. Bull. Tokai Reg. Fish. Res. Lab. 91: 41-51.
- Takano, H., and K. Kikuchi. 1985. Anomalous cells of *Nitzschia pungens* Grunow found in eutrophic marine waters. Diatom 1: 18-20.
- Villac, M.C., et al. 1993. Comparison of two domoic acid producing diatoms: a review, (in press). In: H. van Dam (ed.) Proceedings of the 12th international diatom symposium, Hydrobiol.
- Villareal, T.A., D.L. Roelke, and G.A. Fryxell. 1993. Occurrence of the toxic diatom *Nitzschia pungens* f. *multiseries* in Massachusetts Bay, Massachusetts, U. S. A. Mar. Environ. Res. (in press).
- Wang, R., et al. 1993. Chemical variation of *Nitzschia pungens* as demonstrated by the

co-occurrence of domoic acid and bacillariolides, p. 637-642 In: T.J. Smayda and Y. Shimizu (eds.) Toxic phytoplankton blooms in the sea. Elsevier Sci. Publ. B.V., Amsterdam.

Worms, J., et al. 1990. Domoic acid, a neurotoxin produced by the pennate diatom *Nitzschia pungens* forma *multiseries*, p. 35-42. In: J.M. Frey (ed.) Proceedings of the international symposium on marine biotoxins. CNEVA Pub. Series, Maisons-Alfort, Paris, France.

Zou, J.Z., M.J. Zhou, and C. Zhang. 1993. Ecological features of toxic *Nitzschia pungens* Grunow in Chinese coastal waters, p. 651-657. T.J. Smayda and Y. Shimizu (eds.) Toxic phytoplankton blooms in the sea. Elsevier Sci. Publ. B.V., Amsterdam.

Domoic Acid in Shellfish

Gilgan, M.W., B.G. Burns, and G.J. Landry. 1990. Distribution and magnitude of domoic acid contamination of shellfish in Atlantic Canada during 1988, p. 469-474. In: E. Granéli et al. (eds.) Toxic marine phytoplankton. Elsevier Sci. Publ. Co., Inc., New York.

Haya, K., et al. 1991. Domoic acid in shellfish and plankton from the Bay of Fundy, New Brunswick, Canada. *J. Shellfish Res.* 10: 113-118.

MacKenzie, A., et al. 1993. Domoic acid and the New Zealand Greenshell mussel (*Perna canaliculus*), p. 607-612. In: T.J. Smayda and Y. Shimizu (eds.) Toxic phytoplankton blooms in the sea. Elsevier Sci. Publ. B.V., Amsterdam.

Madhyastha, M.S., et al. 1991. In vitro study of domoic acid uptake by digestive gland tissue of blue mussel (*Mytilus edulis* L.). *Aquat. Toxicol.* 20: 73-82.

Madhyastha, M.S., et al. 1991. A comparative study of uptake and release of glutamic and kainic acid by blue mussel (*Mytilus edulis* L.). *Aquat. Toxicol.* 21: 15-28.

Nassif, J., and R.J. Timperi. 1991. A quantitative assessment of the distribution of selected marine biotoxins in molluscan shellfish harvested from coastal Massachusetts, Nantucket Shoals and Georges Bank. Massachusetts Marine Biotoxin Monitoring Project FDA/PHS/HHS #223-89-4064—Interim Report, November 1989-March 1991.

Novaczek, I., et al. 1992. Depuration of domoic acid from live blue mussels (*Mytilus edulis*). *Can. J. Fish. Aquat. Sci.* 49: 312-318.

Novaczek, I., et al. 1991. Uptake, disposition and depuration of domoic acid by blue mussels (*Mytilus edulis*). *Aquat. Toxicol.* 21: 103-118.

Outerbridge, R. 1992. Effects of a toxic diatom, *Nitzschia pungens* forma *multiseries*, on selected aspects of bivalve physiology: implications for aquaculture management. M. Sc. Thesis, Dalhousie University, Halifax, Nova Scotia, Canada. 99 p.

Scarratt, D.J., et al. 1991. Detoxification of bivalve molluscs naturally contaminated with domoic acid, p. 239-245. In: W.S. Otwell, G.E. Rodrick, and R.E. Martin (eds.) Molluscan shellfish depuration. CRC Press, Boca Raton.

Scarratt, D.J. 1992. Aquatic toxicology—the wider implications of eastern Canadian shellfish toxins, p. 84-85. In: A.J. Niimi, and M.C. Taylor (eds.) Proceedings of the eighteenth annual aquatic toxicity workshop: September 30-October 3, 1991, Ottawa, Ontario. Can. Tech. Rep. Fish. Aquat. Sci. 1863: 381 p.

Shumway, S.E. 1989. Toxic algae: a serious threat to shellfish aquaculture. *World Aquacult.* 20: 65-74.

Shumway, S.E. 1990. A review of the effects of algal blooms on shellfish aquaculture. *J. World Aquacult. Soc.* 21: 65-104.

Shumway, S.E. 1992. Mussels and public health, p. 511-542. In: E. Gosling (ed.) The mussel *Mytilus*: ecology, physiology, genetics and culture. Elsevier, New York.

Silvert, W., and D.V. Subba Rao. 1991. Dynamic model of the flux of domoic acid, a neurotoxin, through a *Mytilus edulis* population. *Can. J. Fish. Aquat. Sci.* 48: 400-405.

Wohlgeschaffen, G.D. 1991. Uptake and loss of the neurotoxin domoic acid by mussels (*Mytilus edulis*) and scallops (*Placopecten magellanicus* Gmelin). M.Sc. Thesis, Dalhousie University, Halifax, Nova Scotia, Canada. 84 p.

Wohlgeschaffen, G.D., et al. 1992. Dynamics of the phycotoxin domoic acid: accumulation and excretion in two commercially important bivalves. *J. Appl. Phycol.* 4: 297-310.

Pharmacology and Epidemiology of Domoic Acid

- Biscoe, T.J., et al. 1975. Domoic and quisqualic acids as potent amino acid excitants of frog and rat spinal neurones. *Nature* 255: 166-167.
- Biscoe, T.J., et al. 1976. Structure-activity relations of excitatory amino acids on frog and rat spinal neurones. *Br. J. Pharmac.* 58: 373-382.
- Bose, R., G. Glavin, and C. Pinsky. 1989. Neurotoxicity and lethality of toxic extracts from Atlantic coast shellfish. *Progr. Neuropsychopharmacol. Biol. Psychiatry* 13: 559-562.
- Bose, R., et al. 1992. Effects of excitotoxins on free radical indices in mouse brain. *Toxicol. Lett.* 60: 211-219.
- Bruni, J.E., et al. 1991. Circumventricular organ origin of domoic acid-induced neuropathology and toxicology. *Brain Res. Bull.* 26: 419-424.
- Constanti, A., and A. Nistri. 1978. A study of the interactions between glutamate and aspartate at the lobster neuromuscular junction. *Br. J. Pharmac.* 62: 495-505.
- Debonnel, G., L. Beauchesne, and C. de Montigny. 1989. Domoic acid, the alleged "mussel toxin", might produce its neurotoxic effect through kainate receptor activation: an electrophysiological study in the dorsal hippocampus. *Can. J. Physiol. Pharmacol.* 67: 29-33.
- Fattorusso, E., and M. Piattelli. 1980. Amino acids from marine algae, p. 95-140. In: P.J. Scheuer (ed.) *Marine natural products: chemical and biological perspectives, Volume III.* Academic press, Toronto, Ontario.
- Fukami, J.-I. 1986. Effects of domoic acid and kainic acids on the neuromuscular junction of mealworm, *Tenebrio molitor* (Coleoptera: Tenebrionidae). *Appl. Entomol. Zool.* 21: 179-181.
- Glavin, G.B., R. Bose, and C. Pinsky. 1989. Kynurenic acid protects against gastroduodenal ulceration in mice injected with extracts from poisonous Atlantic shellfish. *Progr. Neuro-psychopharmacol. Biol. Psychiatry* 13: 569-572.
- Glavin, G.B., C. Pinsky, and R. Bose. 1989. Toxicology of mussels contaminated by neuroexcitant domoic acid. *Lancet* 336: 506-507.
- Glavin, G.B., C. Pinsky, and R. Bose. 1989. Mussel poisoning and excitatory amino acid receptors. *Trends Pharmacol. Sci.* 10: 15-16.
- Glavin, G.B., C. Pinsky, and R. Bose. 1990. Domoic acid-induced neurovisceral toxic syndrome: characterization of an animal model and putative antidotes. *Brain Res. Bull.* 24: 701-703.
- Grimmelt, B., et al. 1990. Relationship between domoic acid levels in the blue mussel (*Mytilus edulis*) and toxicity in mice. *Toxicon* 28: 501-508.
- Habermehl, G. 1990. Marine toxins: epidemiology and transfer, p. 43-55. In: E.M. Bernoth (ed.) *Public health aspects of seafood-borne zoonotic diseases: proceedings of the WHO-Symposium, Hannover, Fed. Republic of Germany (14.11.1989-16.11.1989).* Inst. für Veterinärmedizin des Bundesgesundheitsamtes, Berlin.
- Hampson, D.R., and R.J. Wenthold. 1988. A kainic acid receptor from frog brain purified using domoic acid affinity chromatography. *J. Biol. Chem.* 263: 2500-2505.
- Hampson, D.R., et al. 1992. Interaction of domoic acid and several derivatives with kainic acid and AMPA binding sites in rat brain. *European J. Pharmacol.* 218: 1-8.
- Hurst, J.W. 1990. WHO consultation on public health aspects of seafood-borne zoonotic diseases, p. 81-88. In: E.M. Bernoth (ed.) *Public health aspects of seafood-borne zoonotic diseases: proceedings of the WHO-Symposium, Hannover, Fed. Republic of Germany (14.11.1989-16.11.1989).* Inst. für Veterinärmedizin des Bundesgesundheitsamtes, Berlin.
- Iverson, F., et al. 1989. Domoic acid poisoning and mussel-associated intoxication: preliminary investigations into the response of mice and rats to toxic mussel extract. *Food Chem. Toxicol.* 27: 377-384.
- Maeda, M., et al. 1984. Insecticidal and neuromuscular activities of domoic acid and its related compounds. *J. Pesticide Sci.* 9: 27-32.

- Maeda, M., et al. 1986. Structures of isodomoic acids A, B and C, novel insecticidal amino acids from the redalga *Chondria armata*. *Chem. Pharm. Bull.* 34: 4892-4895.
- Maeda, M., et al. 1987. Neuromuscular action of insecticidal domoic acid on the American cockroach. *Pesticide Biochem. Physiol.* 28: 85-92.
- McGeer, E.G., J.W. Olney, and P.L. McGeer (eds.). 1978. *Kainic acid as a tool in neurobiology*, Raven Press, New York. 271 pp.
- Novelli, A., et al. 1992. The amnesic shellfish poison domoic acid enhances neurotoxicity by excitatory amino acids in cultured neurons. *Amino acids* 2: 233-244.
- Novelli, A., et al. 1992. Domoic acid-containing toxic mussels produce neurotoxicity in neural cultures through a synergism between excitatory amino acids. *Brain Res.* 577: 41-48.
- Perl, T.M., et al. 1990. An outbreak of toxic encephalopathy caused by eating mussels contaminated with domoic acid. *New England J. Med.* 322: 1775-1780.
- Petrie, B.F., et al. 1992. Parenteral domoic acid impairs spatial learning in mice. *Pharmacol. Biochem. Behav.* 41: 211-214.
- Pinsky, C., G. Glavin, and R. Bose. 1989. Kynurenic acid protects against neurotoxicity and lethality of toxic extracts from contaminated Atlantic coast mussels. *Neuropsychopharmacol. Biol. Psychiatry* 13: 595-598.
- Pinsky, C., et al. 1990. Kynurenate antagonism of domoic acid-provoked EEG seizure activity in the mouse. *European J. Pharmacol.* 183: 514-515.
- Shinozaki, H., and M. Ishida. 1976. Inhibition of quisqualate responses by domoic or kainic acid in crayfish opener muscle. *Brain Res.* 109: 435-439.
- Shinozaki, H., M. Ishida, and T. Okamoto. 1986. Acromelic acid, a novel excitatory amino acid from a poisonous mushroom: effects on the crayfish neuromuscular junction. *Brain Res.* 399: 395-398.
- Preston, E., and I. Hynie. 1991. Transfer constants for blood-brain barrier permeation of the neuroexcitatory shellfish toxin, domoic acid. *Can. J. Neurol. Sci.* 18: 39-44.
- Rogers, C.G., and B.G. Boyes. 1989. Evaluation of the genotoxicity of domoic acid in a hepatocyte-mediated assay with V79 Chinese hamster lung cells. *Mutation Res.* 226: 191-195.
- Stewart, G.R., et al. 1990. Domoic acid: a dementia-inducing excitotoxic food poison with kainic acid receptor specificity. *Exp. Neurology* 110: 127-138.
- Strain, S.M., and R.A.R. Tasker. 1991. Hippocampal damage produced by systemic injections of domoic acid in mice. *Neuroscience* 44: 343-352.
- Sutherland, R.J., J.M. Hoising, and I.Q. Whishaw. 1990. Domoic acid, an environmental toxin, produces hippocampal damage and severe memory impairment. *Neuroscience Lett.* 120: 221-223.
- Takeuchi, H., et al. 1984. Effects of alpha-kainic acid, domoic acid and their derivatives on a molluscan giant neuron sensitive to beta-hydroxyl-L-glutamic acid. *European J. Pharmacol.* 102: 325-332.
- Tasker, R.A.R., B.J. Connell, and S.M. Strain. 1990. Pharmacological, behavioural and morphological studies of domoic acid toxicity in vivo. *European J. Pharmacol.* 183: 959.
- Tasker, R.A.R., B.J. Connell, and S.M. Strain. 1991. Pharmacology of systemically administered domoic acid in mice. *Can. J. Physiol. Pharmacol.* 69: 378-382.
- Teitelbaum, J.S., et al. 1990. Neurologic sequelae of domoic acid intoxication due to the ingestion of contaminated mussels. *New England J. Med.* 322: 1781-1787.
- Tryphonas, L., and F. Iverson. 1990. Neuropathology of excitatory neurotoxins: the domoic acid model. *Toxicologic Pathology* 18: 165-169.
- Tryphonas, L., et al. 1990. Acute neurotoxicity of domoic acid in the rat. *Toxicologic Pathology* 18: 1-9.
- Tryphonas, L., J. Truelove, and F. Iverson. 1990. Acute parenteral neurotoxicity of domoic acid in cynomolgus monkeys (*M. fascicularis*). *Toxicologic Pathology* 18: 297-303.
- Tryphonas, L., et al. 1990. Experimental oral toxicity of domoic acid in cynomolgus monkeys (*Macaca fascicularis*) and rats.

- Preliminary investigations. *Food Chem. Toxic.* 28: 707-715.
- Work, T.M., et al. 1992. Epidemiology of domoic acid poisoning in brown pelicans and Brandt's cormorants in California. *J. Zool. Wildlife Medicine* (in press).
- Work, T.M., et al. 1993. Domoic acid intoxication of brown pelicans and Brandt's cormorants in Santa Cruz, California, p. 643-650. In: T.J. Smayda and Y. Shimizu (eds.) *Toxic phytoplankton blooms in the sea*. Elsevier Sci. Publ. B.V., Amsterdam.
- Zaczek, R., and J.T. Coyle. 1982. Excitatory amino acid analogues: neurotoxicity and seizures. *Neuropharmacol.* 21: 15-26.
-
- General and Review References**
- Addison, R.F., and J.E. Stewart. 1989. Domoic acid and the eastern Canadian molluscan shellfish industry. *Aquaculture* 77: 263-269.
- ARL Shellfish Toxin Team. 1988. Solving the toxic mussel problem. *Can. Chem. News* 10: 15-17.
- Bates, S.S., and J. Worms [eds]. 1989. Proceedings of the first Canadian workshop on harmful marine algae, Gulf Fisheries Centre, Moncton, New Brunswick, September 27-28, 1989. *Can. Tech. Rep. Fish. Aquat. Sci.* 1712: 60 p.
- Bird, C.J., and J.L.C. Wright. 1989. The shellfish toxin domoic acid. *World Aquacult.* 20: 40-41.
- Black, E.A. 1990. Forewarned and forearmed with algal blooms. *Fish Farmer*. Sept/Oct: 36-37.
- Cembella, A.D., and E.C.D. Todd. 1993. Seafood toxins of algal origins and their control in Canada, (in press). In: I.R. Falconer (ed.) *Algal toxins in seafood and drinking water*. Academic Press, London.
- Couturier, C. 1988. Shellfish toxins aplenty. *Bull. Aquaculture Assn. Can.* 88-2: 11-27.
- Daigo, K. 1959. Studies on the constituents of *Chondria armata*. II. Isolation of an antihelminthical constituent. *J. Pharm. Soc. Japan* 79: 353-356.
- Daigo, K. 1959. Studies on the constituents of *Chondria armata*. III. Constitution of domoic acid. *J. Pharm. Soc. Japan.* 79: 356-360.
- Drinkwater, K., and B. Petrie. 1988. Physical oceanographic observations in the Cardigan Bay region of Prince Edward Island. *Can. Tech. Rept. Hydrogr. Ocean Sci.* No. 110: 37 p.
- Forbes, J.R. (ed.). 1991. Pacific coast research on toxic marine algae. *Can. Tech. Rep. Hydrogr. Ocean Sci.* 135: 76 p.
- Gordon, D.C., Jr. (ed.). 1991. Proceedings of the second Canadian workshop on harmful marine algae, Bedford Institute of Oceanography, Dartmouth, Nova Scotia, October 2-4, 1990. *Can. Tech. Rep. Fish. Aquat. Sci.* 1799: 66 p.
- Gunner, S.W. 1990. Mussel poisoning in Canada. *J. Toxicol., Toxin Rev.* 9: 89.
- Hall, S. 1988. PSP and other shellfish toxins: recent outbreaks and research. *J. Shellfish Res.* 7: 565.
- Hynie, I., and E.C.D. Todd (eds.). 1990. Proceedings of a symposium on domoic acid toxicity. *Canada Diseases Weekly Report* 16 S1E: 123 p.
- Impellizzeri, G., et al. 1975. Amino acids and low-molecular-weight carbohydrates of some marine red algae. *Phytochem.* 14: 1549-1557.
- Johnson, K.J. 1988. The phytoplankton community structure of two estuaries in eastern Prince Edward Island. Honours Thesis, Mount Allison University, Sackville, New Brunswick, Canada. 34 p.
- Laycock, M.V., A.S.W. de Freitas, and J.L.C. Wright. 1989. Glutamate agonists from marine algae. *J. Appl. Phycol.* 1: 113-122.
- MacKenzie, D. 1988. Mystery of mussel poisoning deepens in Canada—as the chain of death spreads to whales. *New Scientist* 117: 30.
- Maranda, L., et al. 1990. Investigation of the source of domoic acid in mussels, p. 300-304. In: E. Granéli et al. (eds.) *Toxic marine phytoplankton*. Elsevier Sci. Publ. Co., Inc., New York.
- Quilliam, M.A., and J.L.C. Wright. 1989. The amnesic shellfish poisoning mystery. *Anal. Chem.* 61 (18): 1053A-1060A.
- Smayda, T.J. 1990. Novel and nuisance phytoplankton blooms in the sea: evidence for a global epidemic, p. 29-40. In: E. Granéli et al. (eds.) *Toxic marine phytoplankton*. Elsevier Sci. Publ. Co., Inc., New York.

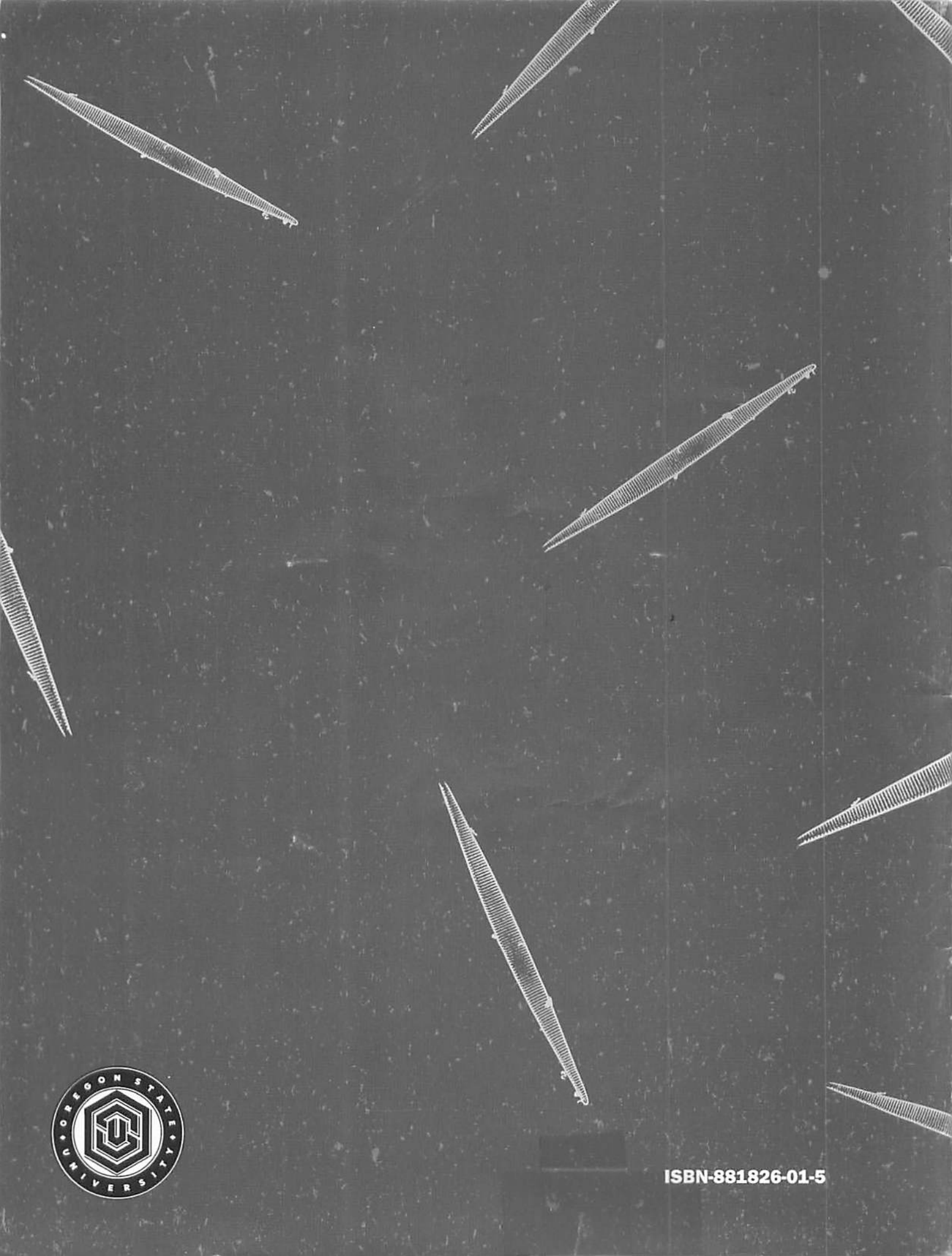
- Smith, J.C., and K.E. Pauley. 1993. A field and laboratory manual for the collection, identification and enumeration of toxic marine phytoplankton from southern and eastern regions of the Gulf of St. Lawrence. Can. Tech. Rep. Fish. Aquat. Sci. (in press).
- Stephen, K. 1990. Domoic acid follow up. Bull. Aquacult. Assoc. Can. 90-1: 35-36.
- Takemoto, T., and K. Daigo. 1958. Constituents of *Chondria armata*. Chem. Pharmaceutical Bull. 6: 578-580.
- Takemoto, T., and K. Daigo. 1960. Über die Inhaltsstoffe von *Chondria armata* und ihre pharmakologische Wirkung. Arch. Pharm. 293: 627-633.
- Therriault, J.-C., and M. Levasseur (eds.). 1992. Proceedings of the third Canadian workshop on harmful marine algae, Maurice Lamontagne Institute, Mont-Joli, Quebec, May 12-14, 1992. Can. Tech. Rep. Fish. Aquat. Sci. 1893: 154 p.
- Todd, E. 1990. Amnesic shellfish poisoning—a new seafood toxin syndrome, p. 504-508. In: E. Granéli et al. (eds.) Toxic marine phytoplankton. Elsevier Sci. Publ. Co., Inc., New York.
- Todd, E. 1993. Amnesic shellfish poisoning—a review. J. Food Protection 56: 69-83.
- Todd, E.C.D., et al. 1993. Recent illnesses from seafood toxins in Canada, p. 335-340. In: T.J. Smayda and Y. Shimizu (eds.) Toxic phytoplankton blooms in the sea. Elsevier Sci. Publ. B.V., Amsterdam.
- van Egmond, H.P., G.J.A. Speyers, and H.J. van den Top. 1992. Current situation on worldwide regulations for marine phycotoxins. J. Natural Toxins 1: 67-85.
- Waldichuk, M. 1989. Amnesic shellfish poison. Mar. Pollut. Bull. 20: 359-360.
- Windust, A. 1992. The response of bacteria, microalgae, and zooplankton to the diatom *Nitzschia pungens* forma *multiseries*, and its toxic metabolite domoic acid. M. Sc. Thesis, Dalhousie University, Halifax, Nova Scotia, Canada. 107 p.
- Wohlgeschaffen, G.D., D.V. Subba Rao, and K.H. Mann. 1992. Vat incubator with immersion core illumination—a new, inexpensive setup for mass phytoplankton culture. J. Appl. Phycol. 4: 25-29.
- Wood, A.M., N. Apelian, and L. Shapiro. 1992. Novel toxic phytoplankton: a component of global change? (in press). In: Proceedings of the sixth international symposium on microbial ecology.
- Wright, J.L.C. 1989. Domoic acid, a new shellfish toxin: the Canadian experience. J. Shellfish Res. 8: 444.
- Wright, J.L.C. 1990. Toxin research at the NRC Atlantic Research Laboratory. Can. Chem. News 42: 18-22.



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