SUMMARY REPORT OF ENHANCED MONITORING AND POLLUTION SOURCE TRACKING EFFORTS IN THE NEW SALT RD. TRIBUTARY, GOOSEFARE BROOK OLD ORCHARD BEACH, MAINE

2012-2017

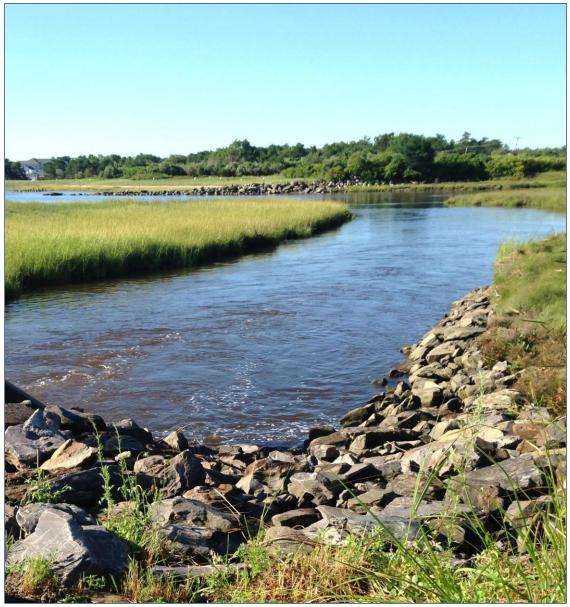


Photo: Meagan Sims

Prepared by: Meagan Sims, University of Maine Cooperative Extension-Maine Healthy Beaches program & Noah Sargent, Maine Conservation Corps Environmental Steward-Maine Department of Environmental Protection.

TABLE OF CONTENTS

Executi	ive Summary	
Acknow	wledgements	
1.	Background	
2.	Project Method	ls6
3.	Results & Disc	pussion7
	3.1	Enterococci and Optical Brighteners
	3.2	Microbial Source Tracking
	3.3	Risk Factor Analysis
4.	Local Actions	to Improve Water Quality
5.	Recommendati	ons
	5.1	Target Human Sources
	5.2	Implement Precautionary Rainfall Advisories
	5.3	Promote Best Practices
6.	Disclaimer	
7.	Appendices	
	7.1	Appendix A: Monitoring Data
	7.2	Appendix B: Additional Source Tracking Efforts
	7.3	Appendix C: Local Assessments

Executive Summary

The Goosefare Brook (GFB) forms the border between the City of Saco to the south and Town of Old Orchard Beach (OOB) to the north. In response to concerns over water quality issues in the mouth and adjacent beach water, the Maine Healthy Beaches (MHB) program has supported multi-year enhanced monitoring and pollution source tracking efforts, held Stakeholder Workshops, and more to address impaired water quality throughout the watershed. Over the past five years, the MHB program has focused primarily on paired enterococci and optical brightener samples in OOB's New Salt Rd. Tributary (NSRT). This work identified widespread bacterial contamination throughout the tributary as well as two priority regions likely impacted by human-sourced fecal contamination.

In 2017, 44 samples were collected during eight events at nine sites within the two priorty areas identified through previous source-tracking efforts. Parameters tested include enterococci (ENT), optical brighteners (OBs), and microbial source tracking (MST) DNA analyses. ENT values ranged from 5 to 6,870 MPN/100mls with a combined geometric mean of 772 MPN for all sites. OB values ranged from 33 to 157 µg/l with a combined mean of 94 µg/l for all sites. In 2016-2017, microbial source tracking (MST) techniques were utilized to identify specific host source(s) of fecal contamination within the NSRT. The majority of samples were analyzed for mammal and human DNA (presence/absence) and a subset of samples were tested for the presence of bird, ruminant, and canine DNA. All samples tested positive for mammal DNA whereas percent detection of human sources varied between the two priority regions (mouth vs. marsh). Human sources were detected consitently in the mouth region with a mid-season peak in signal strength (July/August), the portion of the year when OOB likely experiences the greatest population numbers. Human sources were detected intermittely at marsh locations, indicating potential contamination from the use of seasonal residence(s) and possible groundwater transport of pollution sources. Additionally, regrowth and persistance of FIBs in this area as well as additional untested mammalian source(s) are likely contributing to elevated ENT levels given the relatively consistent but high mammalian signal, irregular human and canine detections, and very elevated ENT concentrations. For both regions, Bird DNA was detected in the majority of samples at relatively consistent concentrations throughout each season. Canine DNA was detected during 3 monitoring events in the marsh region (2017) and no ruminant DNA was detected for either region.

The pollution source tracking tools used as part of this study were combined into a risk factor analysis highlighting sites potentially impacted by human sources of fecal contamination. It is recommended that OOB prioritize investigations in these regions to identify and remove sources of human sewage. As part of ongoing efforts to address water quality in the GFB, both municipalities have investigated and removed numerous sources of human wastewater, have expanded and upgraded sewer and stormwater infrastructure, and have worked together to acquire supplemental funding to complete a watershed management plan and begin implementation of outlined objectives in that plan. Continued human-sourced contamination issues underscore the need to continue investigations to ensure the integrity of wastewater disposal methods throughout the GFB watershed.

Acknowledgements

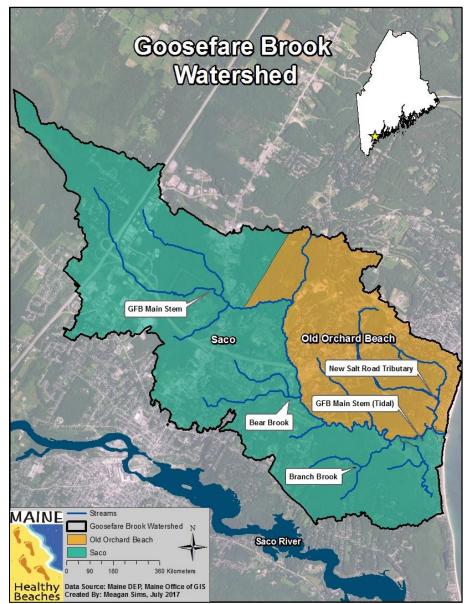
Special thanks to all of the dedicated staff/volunteers who have helped collect data since this study began, especially the late John Bird who was instrumental in transforming the data to information to action. Also thank you to Maine DEP, US EPA, OOB, and Saco for their support. Funding for the Maine Healthy Beaches program is provided by US EPA and Maine DEP.

Background

The Goosefare Brook (GFB) Watershed is approximately 9.83mi² and is shared by the City of Saco (approximately 4,000 acres) and Town of Old Orchard Beach (OOB) (approximately 1,000 acres). The mouth of the GFB demarcates the beach and boundary between Saco and OOB. Just inland from the mouth, the brook splits into two branches, one draining primarily from Saco (Main Stem) and the other from an OOB tributary named the New Salt Road Tributary (NSRT) for purposes of this study (Figure 1). Progressing upland in the watershed (the land area draining to the brook), the two major sections of the brook continue to branch into a network of smaller tributaries. Municipal and private sewer services the majority of the GFB watershed, yet some properties have subsurface wastewater disposal (septic, cesspool) systems. Additionally, both municipalities are designated as "MS4" communities requiring them to implement a multifaceted approach to improving the quality of stormwater. A 5.54-mile segment of the GFB and several upstream tributaries are listed on ME-DEP's 303(d) list of urban impaired waters for bacteria and other stressors.

Since 2003, Saco and OOB have participated in routine beach monitoring as part of the MHB program. Monitoring at Ocean Park beach sites near the mouth of the GFB frequently revealed elevated bacteria levels and prompted the need to expand monitoring further upland. Enhanced efforts began with routine monitoring of two sites (GFB-01 and Saco-00) (Figure 2) located just above the mouth where the brook splits into the Main Stem and the NSRT. Subsequent monitoring was initiated further upland to address impaired water quality throughout the entire watershed in 2010. Results of this larger assessment revealed extensive bacterial pollution with a high likelihood of human fecal contributions particularly in Saco's Bear Brook and OOB's NSRT (Figure 1).

Figure 1. Goosefare Brook Watershed boundary, GFB main stem, and several major tributaries including the NSRT.



In response, MHB planned/facilitated meetings with representatives from Saco, OOB, ME DEP, and US EPA to share data and develop remediation strategies in 2011. From 2012-2017, MHB program efforts have concentrated primarily on the OOB branch (NSRT) and ME DEP efforts have focused on several upland regions of the GFB impaired for a number of criteria including Bear Brook (Figure 1). MHB staff continue to use local knowledge of potential suspect areas and collected data hone in on problematic areas

In an effort to pinpoint human sources, the pollution source tracking toolbox approach has been utilized incorporating multiple parameters (Table 1). Typically, as the number of parameters that exceed a threshold (or detectable) limit increases, so does the confidence that human sources are impacting water quality. Toolbox parameters utilized are largely dependent on staff availability and funding, and have therefore varied for each monitoring season. For 2017, parameters monitored included enterococci (ENT), optical brighteners (OB), and microbial source tracking (MST). The incorporation of MST was made possible through a continued applied research partnership with the University of New Hampshire Jackson Estuarine Laboratory (UNH JEL). Parameter results were combined (for NSRT monitoring efforts) to create a risk factor analysis highlighting suspect areas warranting further investigation by OOB (Table 3). Although wildlife, pet, and waterfowl waste can contribute to impaired water quality, it is recommended to target human sources first.

Parameter	Method	Source Target	Cost/Expertise
		Warm blooded	
Enterococci (ENT)	Grab sample, Enterolert	animals	Low/Low
	Grab sample,		
Optical brighteners (OB)	Fluorometry	Human	Low/Low
Pharmaceuticals and personal	Grab sample, metabolite		
care products (PPCP)	analysis	Human	High/High
	Grab sample in tandem		
Canine detection	with canines	Human	Low/Med
Microbial source tracking	Grab sample, DNA	Variety of human and	
(MST)	extraction	non-human sources	High/High

Table 1. Source tracking toolbox parameters used in the GFB by MHB and associated partners.

ENT indicate the presence of fecal contamination from warm-blooded animals and the possible presence of disease-causing microorganisms. However, fecal indicator bacteria (FIBs) like ENT do not differentiate the source(s) of bacterial pollution and have been found to persist and regrow in sand and sediments.¹ OBs are commonly used in commercial/retail products such as clothing detergents, dishwashing agents, and personal care products to brighten the whiteness of materials. These products are typically flushed down the drain and when concentrations are coupled with elevated fecal bacteria levels, can be indicative of human-sourced fecal contamination.

MST methods are used to complement traditional FIB monitoring, specifically targeting DNA of individual source markers using PCR², allowing for the differentiation between human and non-human fecal sources potentially contributing to observed elevated FIB levels. In contrast to FIBs, DNA source markers quickly degrade outside of their host (approximately 1 week) and therefore, a positive PCR assay suggests a recent contamination event. This method is ideal for the NSRT because of the potential for FIBs to persist in several

¹ Badgley B.D., Thomas F.I., & Harwood V.J. 2011. Quantifying environmental reservoirs of fecal indicator bacteria associated with sediment and submerged aquatic vegetation. Environmental microbiology 13.4: 932-942.

 $^{^{2}}$ PCR= Polymerase chain reaction. It is a method used to amplify segments DNA resulting in a copy number for specific DNA targets.

low-lying marsh regions. These markers demonstrate host specificity, allowing for the quantification of numerous specific host sources from one sample. MST methods can also provide the relative strength of the fecal source marker by using quantitative PCR (qPCR). This test provides a DNA copy number that can be used to better track fecal contamination to the source(s) and give a sense of the relative pollution contribution from human waste. There are currently no established thresholds for qPCR copy numbers as there are for FIBs. For this reason, it is useful to compare data to similar watersheds to gain a greater context of the results. Data can also be compared to known human-associated contamination events to better understand the potential human-sourced contribution given observed concentrations.

Project Methods

Since 2012, the MHB program has supported over 600 paired ENT and OB samples at 23 routine sites stratified througout the NSRT portion of the GFB watershed. Monitoring locations targeted suspect areas identified through previous monitoring efforts, suspected human-sourced fecal contamination "hot-spots", and local information keeping in mind ease of accessibility and avoidance of private property. Due to this approach, site locations and monitoring frequency have varied each year.



Multi-year pollution tracking efforts highlighted two priority regions within the NSRT with the highest likelihood of human fecal contributions. These are the GFB-01 region located at the mouth of the brook near the tide gate where the NSRT combines with the GFB main stem before it reaches popular downstream swimming beaches and the marsh region located upstream at the outlet of the NSRT drainage from the Jordan Marsh (Marsh-1, Marsh-2). Monitoring sites were reduced in 2016 & 2017 to further hone in and bracket suspected hot spots and prioritize resources for DNA analyses to confirm suspected human-sourced fecal contributions to observed elevated bacteria levels. In order to assess NSRT water quality before mixing with seawater, MHB staff monitored during outgoing tides at six routine sites and seven FYI sites in 2016-17 (Figure 2).

Figure 2. 2016-17 NSRT monitoring sites.

Efforts for 2017 included 44 samples collected over 8 events at 9 monitoring stations from late June to early October to document potential baseline ENT, OB, and DNA readings before and after the majority of seasonal residents arrived in the region of Ocean Park. Collections included a combination of dry and wet weather events although the MHB program is most concerned with potential point sources of human fecal pollution (malfunctioning septics, faulty sewer infrastructure) indicated by FIB exceedances during dry weather conditions. During wet weather events, multiple sources (human and non-human) act together and often result in extremely elevated fecal bacteria levels that often do not provide insight as to what the problem(s) are and where they are located.

Five DNA markers were targeted for this study and include general mammal, human, canine, ruminant,³ and bird. For all samples, initial tests were conducted to determine the presence (PCR) of mammal and human DNA markers to confirm suspected contamination hot spots. Subsequent qPCR analyses were conducted for sites testing positive for the presence of the human DNA marker to determine the strength of the signal and its fluctuation over time relative to the general mammal marker. This is meant to give an indication of human contibution relative to other mammals. To assess potential mammalian fecal inputs in the absence of consistent human DNA detection, follow-up ruminant DNA tests were conducted for sites with suspected wildlife contributions. Canine and bird DNA analyses were also performed. qPCR general mammal DNA values are greater than human specific qPCR results because the general marker represents all mammal sources contributing with human sources as a component of that value. Birds represent a separate animal class and while they cannot be directly compared to the mammal source marker as a component, they can provide information regarding other potential fecal sources contributing to elevated FIBs.

Results & Discussion

Enterococci and Optical Brighteners

In general, all identified suspect sites demonstrated elevated ENT levels over the past five years, and for many sites particularly in the GFB-01 (mouth) and GFB-05 (marsh) series, those levels have increased over time (Figure A4). Additionally, OB concentrations at these locations have generally been greater compared to less problematic sites within the NSRT drainage area (Figure A5).

For the three routinely monitored sites in 2017, all exceeded the ENT geometric mean⁴ safety threshold⁵ for marine waters. Single sample ENT values ranged from 5 to 6,870 MPN/100ml. ENT geometric mean levels varied between monitoring stations and ranged from 315 to 1,836 MPN/100ml with a combined geometric mean value for all NSRT sites of 772 MPN/100ml. This is over 22 times the EPA geometric mean safety criteria for recreational water contact (Table A2, Figure A6). While the NSRT-wide ENT geometric mean value has fluctuated since 2012, primarily as result of changes in site locations, monitoring frequency, and changes in the number of wet weather monitoring events, results have remained well over the EPA safety threshold for all monitoring seasons. OB single sample concentrations ranged from 33 to 157 μ g/l with a combined NSRT mean of 94 μ g/l (Table A2, Figure A7).

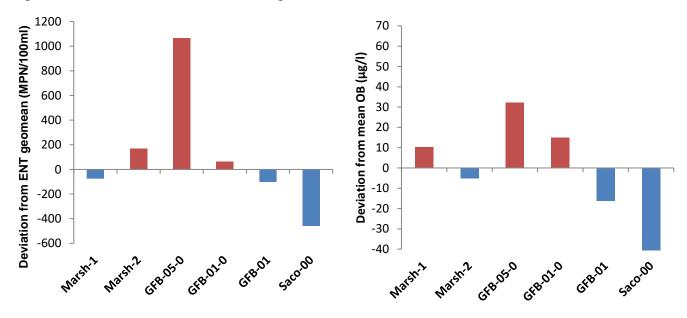
³ Ruminants include cattle, sheep, goats, deer, giraffes, antelopes, and camels. Canine and ruminant source markers were tested based on local feedback of potential sources contributing in this watershed. Ruminants tested in 2016 only.

⁴ A geometric mean represents the typical value of a set of numbers. It is calculated using the product of a set of values rather than using their sum as when calculating an arithmetic mean (average). Any ENT single sample results of <10 MPN/100ml were considered 5 MPN/100ml for report calculations.

⁵ US Environmental Protection Agency (EPA) recommend single sample maximum value for enterococci in marine waters is 104 (MPN/100 ml) and 61 (MPN/100 ml) for fresh water sites. EPA recommended geometric mean values are 35 (MPN/100 ml) and 33 (MPN/100 ml) respectively.

Goosefare Brook 2017 Monitoring Report

For the NSRT watershed, the OB 100µg/l threshold may not be a good metric for indicating human-sourced pollution due to interference from humic substances (tannins and other dissolved organic compounds) that can elevate OB readings and cause a "background level" contribution to measured OBs in systems like the NSRT that have tea colored water, an indicator of humic content. To help identify "hot-spots" of contamination, calculating individual site deviations from the overall mean can help pull a meaningful signal when most sites exhibit elevated ENT levels and are impacted by organic matter/interference (i.e. the most problematic sites within the system). Sites with positive deviations for both ENT and OB levels represent suspect locations potentially impacted by human sources. Sites with historical positive deviations for both parameters were targeted for further source identification using MST in 2016-17.



Figures 3-4. Deviations from 2017 ENT geometric mean and mean OB value for all NSRT sites. Bars above the x-axis indicate sites where ENT/OB values were greater than the geomean/mean and bars below represent those lower than the geomean/mean. See Table A2 for sample sizes. Sites GFB-01-OB, GFB-01-01, and CB-C/WG were single sample events and not included in geomean/mean comparisons.

Microbial Source Tracking

Confirm suspected contamination hot-spots

All samples submitted for DNA analyses were tested for the presence of mammal and human DNA markers⁶. PCR (presence/absence) analyses confirmed the presence of human DNA in both priority regions. Human DNA was consistently detected at the mouth of the NSRT (GFB-01 region) whereas human source(s) were observed sporadically for marsh sites. Specifically, human sources were detected 96.6% of the time in mouth sites vs 54.8% of the time in marsh associated sites. All sites tested positive for mammal sourced DNA, a trend not uncommon in regions like the GFB where numerous potential mammalian fecal sources exist throughout the watershed. Of the sites tested, bird DNA was detected in almost every sample (mouth-100%; marsh-97.1% detection) (Figure 5). Percent detection information was used to identify sites for additional human DNA analyses, prioritizing the mouth region as results indicated consistent human source(s) present and the presence of downstream beaches extensively used by the public Because human DNA was not consistently detected at marsh locations, follow up PCR analyses for canine and ruminant⁷ DNA were conducted.

⁶ MST results were combined for the 2016 and 2017 monitoring seasons.

⁷ 2016 monitoring only.

Goosefare Brook 2017 Monitoring Report

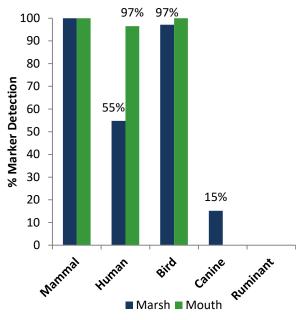


Figure 5. Total % detection of each source marker for marsh and mouth regions $(2016-17)^8$.

Canine sources were also tested for in the mouth region⁹, as local input suggested this residental area just upstream from popular beaches may be impacted by canine fecal waste. All samples tested were negative for canine and ruminant markers in 2016 for both regions. In 2017, 25% of samples tested positive for canine DNA at marsh sites. Consistent mammal DNA detections with intermittent human and canine detection suggests there may be other mammalian source(s) contributing to elevated FIBs in the marsh region, regrowth and persistence of FIBs in favorable conditions, or a combination of the two.

Subsequent qPCR analyses were used to better assess the strength of the source marker, resulting in a DNA copy number. Data were combined for sites within both regions to obtain an overall signal strength for each marker¹⁰. Results indicate a stronger human signal for mouth sites compared to marsh locations. Bird DNA copy numbers were

Table 2. 2016-17 PCR summary for GFB monitoring sites including
total sample number and number of samples tested for each marker.
M=mammal, H=human, B=bird, C=canine, R=ruminant.

Site	Sample #	Sources Tested (# of samples)
Marsh-1	15	M (15), H (15), B (12), C (12), R (5)
Marsh-2	15	M (15), H (15), B (12), C (12), R (5)
GFB-05-0	9	M(9),H(9),B(9),C(9),R(3)
GFB-05-1	3	M(3),H(3),B(2)
Marsh Region	42	M(42) ,H(42), B(35), C(33), R(13)
GFB-01	15	M (15), H (15), B (8), C (8)
GFB-01-0	10	M (10), H (10), B (2), C (2)
GFB-01-C	2	M(2),H(2), B(2),C(2)
GFB-01-B	1	M (1), H (1), B (1), C (1)
Porter Ave	1	M (1), H (1)
Mouth Region	29	M(29), H(29), B(13), C(13)

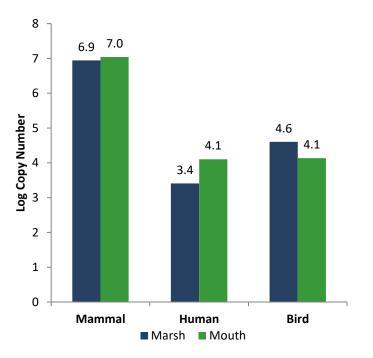


Figure 6. 2016-17 qPCR log copy numbers for marsh and mouth regions of the GFB.

⁸ Summaries include 9 samples for which positive PCR results corresponded with negative qPCR copy numbers indicating the target sequence was likely detected at low copy numbers. These samples are classified as "low level positive" and detection of target sequences at these concentrations cannot be reproduced 100% of the time. Corresponding qPCR copy numbers were not used in subsequent summaries.

⁹ 2017 monitoring only.

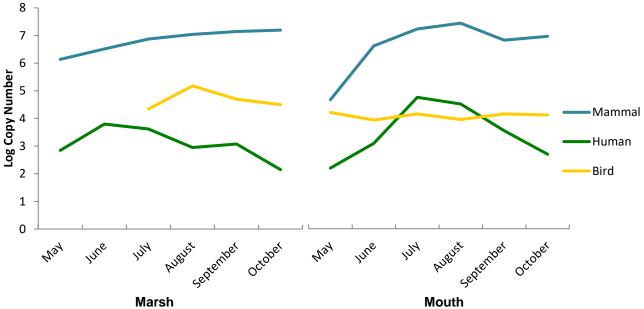
¹⁰ Mammal, human, and bird.

similar between the two regions, but slightly higher for the marsh. Mammal DNA signal strength was nearly identical between the two regions. There are currently no established thresholds for qPCR copy number values, therefore results were put into context through comparisons to studies conducted in similar watersheds. Results indicate greater human DNA levels overall in the OOB mouth region compared to results from similar watersheds.

Seasonal fluctuations

Samples were collected throughout the summer season for both years¹¹, allowing for the use of qPCR results to assess persistence of fecal sources and seasonal fluctuations in source signal strength over time. Combined data for 2016 and 2017 revealed fairly consistent bird copy numbers over the course of the season for both regions with a slight peak in August for marsh sites. More more notable fluctuations were observed for human and mammal qPCR concentrations with distinctly different signal strength patterns observed between the two priority regions (Figures 7-8).

For the marsh region, the human signal was detected 54.8% of the time with two concentration peaks, one in June/July and another smaller peak in September. No distinct relationship was observed between the human and mammal signal strength for this area, indicating the possibility of another mammalian fecal source driving the strength and persistence of the mammal DNA signal (Figure 7). Given the fairly consistent mammalian signal and irregular human DNA increases, regrowth and persistance of FIBs may be contributing in part to elevated ENT levels recorded in this low lying marsh with little consistent stream flow.



Figures 7-8. 2016-17 qPCR copy numbers/ENT MPN for all sources tested in GFB priority regions (marsh and mouth).

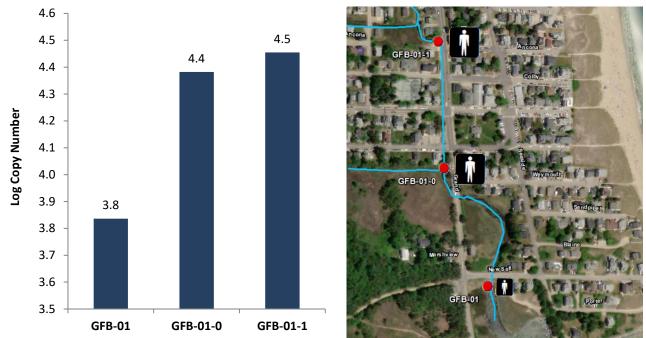
For the mouth region where the human signal was detected throughout both monitoring seasons, human copy numbers generally mirrored mammal values (excluding the October decrease in human copy number), suggesting that human source(s) in this region may be the primary driver of the strength of the mammal DNA signal detected. A seasonal peak in human DNA signal strength was observed mid-season (July/August) (Figure 8) and this peak coincides with historical ENT seasonal patterns, suggesting this portion of the season may when the greatest pressure is being placed on subsurface waste systems (Table A8).

¹¹ May-October for 2016; June-October 2017.

Refine hot spots

To better understand the signal strength of potential human source(s), samples were collected at multiple sites progressing upland in the watershed (when feasible) and analyzed using qPCR. Pollution source refinement was considered preliminary as funding constraints limited the number of samples analyzed for a given date.

For example, samples were collected at 3 sites on 8/15/2017 starting at the mouth of the NSRT (GFB-01) and progressively moving upstream (GFB-01-0 then GFB-01-1). Human qPCR concentrations increased slightly for upstream sites, indicating those samples may have been collected closer to potential human source(s) contributions for that date (Figures 9-10). Given the number of potential human sourced impairments within this region (i.e. aging/faulty sewer infrastructure, septic malfunctions, cross connections) and the seasonal variability in flow conditions of the tributary, source strengths (copy numbers) varied depending on the monitoring day. While more work is needed to further hone in on suspect properties/infrastructure, this information can be used to more effectively prioritize ongoing investigative efforts that are often costly and time consuming.



Figures 9-10. qPCR copy numbers (Fig. 9) and sites monitored (Fig. 10) on 8/15/2017 to refine priority areas needing further investigation in the mouth region of the NSRT.

Risk Factor Analysis

Given the pervasiveness of fecal contamination in the GFB watershed, it is important to use multiple source tracking tools to identify human-sourced contributions. This work is meant to help inform local pollution remediation efforts. The pollution source-tracking tools applied in the NSRT for 2012-2017 were combined into a risk factor analysis where ENT results were analyzed in conjunction with other co-indicators of human sewage. Risk factors included ENT geomean and OB mean threshold exceedances, deviations from geomean ENT and mean OB values, presenece of 4 or more detectable PPCP compounds, positive canine detections, and positive human DNA source detections (Table 3).

Monitoring stations with \geq 4 elevated/positive parameters are highlighted as priority sites with the potential for point sources of human associated fecal pollution. The risk factor analysis is meant as a guide and is not a

definitive or conclusive indicator that illicit source(s) are present. Further investigations are needed to ensure the integrity of nearby wastewater disposal.

< 5 times). Sites with 5	S times). Sites with 5 EN17OB samples ofe more included. See Appendix B for cannie and FFCF data.							
MONITORING STATION	ENT ≥ 35 MPN/100ml	OB≥100 μg/l	+ Dev. from ENT Mean	+ Dev. from OB Mean	≥4 PPCPs ng/l	+ Canine Det.	+ Human PCR	
GFB-01	Y	Ν	Y	Ν	Ν	Y	Y	
GFB-01-0	Y	Ν	Y	Y	Ν	Ν	Y	
GFB-01-1	Y	Ν	Y	Y	Ν	Ν	-	
GFB-04	Y	Ν	Ν	Ν	-	Ν	-	
GFB-04-0	Y	Ν	Ν	Ν	Ν	Ν	-	
GFB-04-0-1	Y	Ν	Ν	Ν	Y	Y	-	
GFB-04-1	Y	Ν	Y	Y	-	Ν	-	
GFB-04-2	Y	Ν	Ν	Y	Ν	Ν	-	
GFB-04-3	Y	Ν	Ν	Ν	Ν	Ν	-	
GFB-05	Y	Y	Y	Y	-	Ν	-	
GFB-05-0	Y	Y	Y	Y	Y	Ν	Y	
GFB-05-1	Y	Y	Y	Y	Y	Y	-	
GFB-05-2	Y	Ν	Ν	Y	Ν	Ν	-	
Marsh-1	Y	Y	Y	Y	-	-	Y	
Marsh-2	Y	Ν	Y	Ν	-	-	Y	

Table 3. 2012-2017 Pollution source tracking toolbox risk factor analysis. Y= Yes, N=No, (-) = not monitored (or monitored< 5 times). Sites with 5 ENT/OB samples ore more included. See Appendix B for canine and PPCP data.</td>

Impaired bacterial water quality in the NSRT is likely a combination of human, wild, and domestic animal waste. Human sources may include but are not limited to faulty sewer lines, cross-connections between sewer and stormwater infrastructure, and malfunctioning septic systems/cesspools. Segments of the sewer infrastructure in the NSRT are aging and comprised of sub-optimal materials (clay, asbestos) (Figures C3-C4). Additionally, stormwater drains directly to the NSRT sub-watershed at no fewer than 20 locations and polluted runoff transports waste from various diffuse sources throughout the watershed.

Local Actions to Improve Water Quality

Saco and OOB continue to work creatively to use limited resources to address water quality impairments in the GFB. In 2017, Saco and OOB continued their collaborative work to protect and restore water quality in the GFB by implementing a number of stormwater retrofits, erosion/buffer control projects, and education/outreach initiatives utilizing 319 Phase I Implementation grant funds. As part of this work, both communities have worked with diverse partners to collect data, identify sites for installation of best management practices (BMPs), and promote education/outreach throughout their communities. A Restoration Committee was formed including members of each municipality, Conservation Commissions, state agencies, local schools, and non-profits to guide the implementation of current and future restoration efforts. Additionally, an Outreach Committee was formed to ensure outreach initiatives are effective and targeting appropriate audiences. Outreach work for 2017 included stream cleanups, partnering with local schools to conduct buffer plantings and storm drain stenciling, and hosting the innagurel April Stools Day event.

Saco and OOB have continued routine maintenance (catch basin and sewer line cleaning, street sweeping, etc.), completed illicicit discharge detection and elimination studies, performed line and catch basin replacements,

and conducted smoke, dye, and CCTV surveys to ensure the integrity of storm and sanitary infrastructure. As a result, faulty sewer lines, cross connections between sewer/stormwater infrastructure, and malfunctioning subsurface wastewater disposal (septic/cesspool) systems have been identified and eliminated throughout the watershed. The MHB program continued working with OOB Public Works to document previous investigations and identify priority areas within the sanitary system for follow up CCTV investigations. The town continued implementing a new GIS program to better track and access future inspection information including catch basin cleanings, sewer investigations, street sweeping, etc. MHB partnered with the OOB Conservation Commission to continue an applied research partnership with UNH for MST testing in priority areas where human sources were identified in 2016. The City of Saco partnered with MHB, ME DEP, and UNH to initiate MST efforts in an impaired upstream tributary (Bear Brook). Both communities continued to post supplemental signage at the mouth of the GFB in 2017, alerting the public of the potential risk of water contact at this location.

In 2018, both communities will continue implementing 319 grant-associated objectives, continue enhanced monitoring and pollution source tracking work, and investigate the integrity of and make improvements to sewer/stormwater infrastructure. As part of this work, OOB will develop a fill ordinance conferring greater protections to GFB water quality, will continue to prioritize CCTV and other maintenance efforts by collaborating with MHB staff to document investigations, and will follow up on suspect properties identified through smoke testing efforts in 2015. MHB plans to conduct follow-up MST testing for priority hot spots of human fecal contamination identified in 2016-2017.

Recommendations and Next Steps

Target Human Sources

It is recommended that the towns continue investigations of suspect areas to rule out sources of human sewage, as research indicates human sources present the greatest health risk due to the host-specificity of associated pathogens.¹² Of particular concern are potential wastewater sources in the vicinity of documented contamination hotspots with elevated bacteria and co-indicators of human sewage including the presence of human DNA sourced from fecal matter. Recommendations include:

- Follow-up on identified parcels from smoke tests (2015) indicating home to sewer connection issues (OOB PW & LPI jurisdiction) (Figure C2).
- Continue to maintain and update septic inventory/pump out records (Figure C1).
- Provide education/outreach material to the public on septic best practices and promote the Town's pump out tax credit.
- Continue partnering with community and state organizations to implement outreach initiatives and BMPs in priority regions.

¹² Ferguson C.M., Coote B.G., Ashbolt N.J., & Stevenson I.M. 1996. Relationships between indicators, pathogens and water quality in an estuarine system. Water Res. 30:2045–2054.

Wade T.J., Calderon R.L., Brenner K.P., Sams E., Beach M., Haugland R., Dufour A.P. 2008. High sensitivity of children to swimming-associated gastrointestinal illness: results using a rapid assay of recreational water quality. Epidemiology. 19:375–383.

- Continue supporting bacteria/DNA monitoring of priority sites to hone in on potential sources and to ensure existing sources have been removed/new ones haven't emerged.
- Use DNA results (2016-17) to prioritize future investigations in the following areas:

- Mouth/Outlet of NSRT Region (GFB-01 series)

- Human sources were detected in this region throughout the 2016-17 monitoring season with a mid-season peak in source strength.
- The historical trend at the mouth of the GFB has been higher ENT results on an incoming tide (Figure A17, Table A8) suggesting potential source(s) in the vicinity of the mouth and/or conditions favoring persistence and possibly regrowth of ENT.
- Although the town has tested the tide gate and areas directly upland, it is recommended to continue investigations in this region to ensure a tight system at the tide gate.
- Human DNA strength appears to increase moving upland from the tide gate. Target investigations along West Grand from New Salt Rd. to Winona Ave.
- Survey potential septic systems in the region.
- Investigate infrastructure integrity near GFB-01-0 where the NSRT goes underground (in a closed box culvert parallel to Rt. 9) between sites GFB-01-0 (Randall Ave.) and GFB-01-1 (Ancona Ave) (Figure A1).

- Marsh Region (GFB-05 & Jordan Marsh series)

- Investigate where the brook runs along a residential area between sites GFB-05-1 and Marsh-1 (Oceana Ave.), branching to the right at GFB-05-0 (Rt.9 near Casco Ave.) and to the left at Marsh-2.
- Rule out human sources in this region. MST data indicates a recent fecal source. Irregular human pulses may indicate an issue with a residence used intermittently during the summer season.
- The area may be impacted by water table/groundwater overland flow. Sources may be further away if ideal transport conditions are present. Priority homes and associated infrastructure to test: 24 Oceana, 22 Oceana, 170 West Grand.
- The culvert in this region is decaying. Replacement is recommended to ensure no pollution sources can infiltrate the NSRT.
- Additional camera and dye testing is recommended to determine potential infiltration/exfiltration issues.
- Investigate potential septic systems in the area.

As time and resources allow, it is also recommended to continue expanding and improving sewer and stormwater infrastructure. More qPCR data is recommended to further hone in on the source(s) of human fecal DNA detected by stratifying monitoring sites in priority areas and tracking the strength of the DNA signal to isolate contamination sources. On a broad scale, it is recommended the towns incorporate water quality assessment and investigation of these sites into their MS4 Permit/Plan that requires the towns to develop and implement a stormwater management program. The MHB program will continue to meet with the town to discuss results and will provide recommendations on future remediation efforts based on continued source tracking work.

Implement Precautionary Advisories

Due to the history of impaired water quality in the brook and its impact on adjacent coastal beaches, it is recommended that Saco and OOB beach managers post precautionary rainfall advisories at Bay View, Kinney Shores, and Ocean Park beaches when local precipitation levels are greater than one inch within 24hrs. The advisory should be kept in place for at least 24hrs after the rainfall ceases to allow flushing of the system. Additionally, recreational water contact occurs in the mouth of GFB including swimming and jumping off of the Rt. 9 Bridge. It is recommended that Saco and OOB continue to post permanent signage at the bridge and on both banks of the river mouth alerting the public to the potential hazards of swimming at this location until ENT levels are consistently within acceptable limits.

Promote Best Practices

The towns are encouraged to follow low impact development practices throughout the watershed such as reducing impervious surfaces to allow rainwater to naturally percolate into the ground, preserving and recreating natural landscapes to treat polluted runoff, restoring vegetative buffers (sections of vegetation adjacent to bodies of water used to minimize runoff effects), etc. It is suggested that the towns continue to work with partners (e.g. MHB, OOB Conservation Commission) on outreach and education campaigns such as septic system maintenance, responsible pet waste management, and storm drain stenciling (e.g. no dumping, drains to ocean).

Disclaimer

This report has been compiled to the best of the Maine Healthy Beaches program's knowledge. Please submit and comments or additions to MHB staff.

Appendix A: Monitoring Data

2012-2017 Monitoring Data

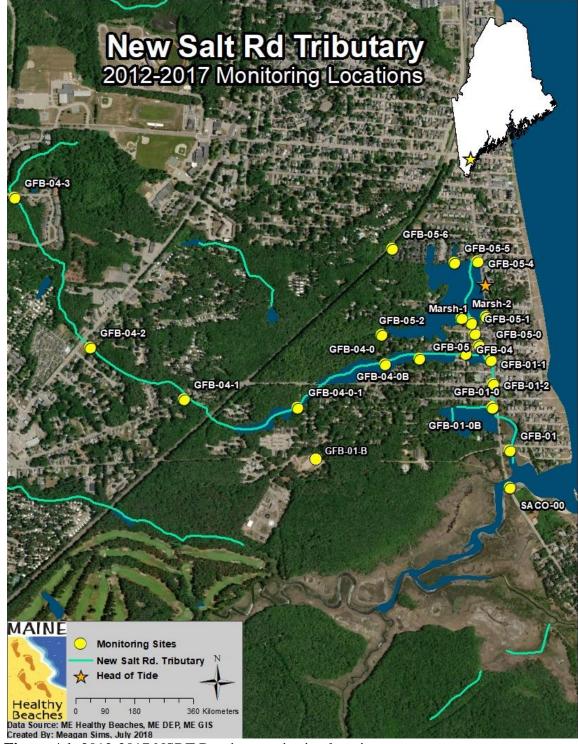


Figure A1. 2012-2017 NSRT Routine monitoring locations.

Site	Year	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
GFB-04-1	2012	339.6	88.4	4	5
GFB-04-2	2012	199.7	89.5	4	5
GFB-04-3	2012	131.9	46.3	4	5
GFB-01-0B	2012-13	274.6	79.7	4	5
GFB-01-2	2012-14	504.7	87.0	13	13
GFB-01	2012-17	349.8	78.1	63	54
GFB-01-0	2012-17	464.5	97.2	46	47
GFB-01-1	2012-15	288.8	95.3	36	37
GFB-04	2012-15	169.6	86.5	35	36
GFB-04-0	2012-15	118.4	78.6	34	35
GFB-04-0-1	2012-15	116.1	76.2	33	34
GFB-05	2012-15	564.7	101.4	36	37
GFB-05-0	2012-17	980.5	119.9	45	46
GFB-05-1	2012-16	739.2	100.9	39	40
GFB-05-2	2012-15 2012-15;	59.3	95.7	34	34
SACO-00	2017	46.7	45.9	33	24
GFB-05-6	2013	44.6	81.7	9	9
GFB-04-0B	2013-15	129.8	78.6	29	29
GFB-05-4	2013-15	46.1	64.6	30	30
GFB-05-5	2013-15	13.5	85.9	30	29
Marsh-1	2015-17	1382.3	102.3	24	24
Marsh-2	2015-17	1017.9	86.6	24	24
GFB-01-C*	2016	1806.0	95.1	2	2
Total		224	88	611	604

Table A1. 2012-2017 data summary for Goosefare Brook watershed monitoring including the year sampled, mean ENT concentration, geometric mean ENT concentration, mean optical brightener concentration, and the sample size at each site.

*GFB-01-C mean value given.

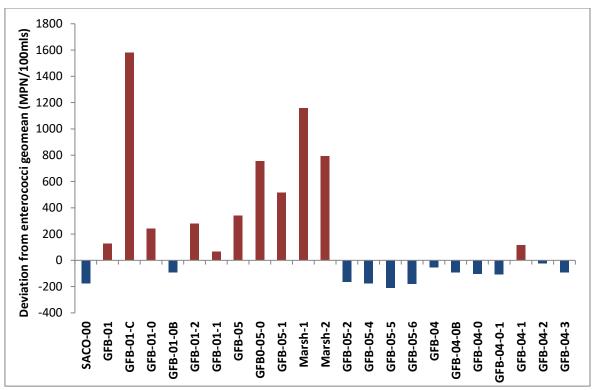


Figure A2. Deviations from the 2012-2017 combined ENT geometric mean for all NSRT sites. Bars above the X-axis indicate sites where ENT values were greater than the overall geomean and bars below represent those lower than the overall geomean (See table A1 for sample sizes).

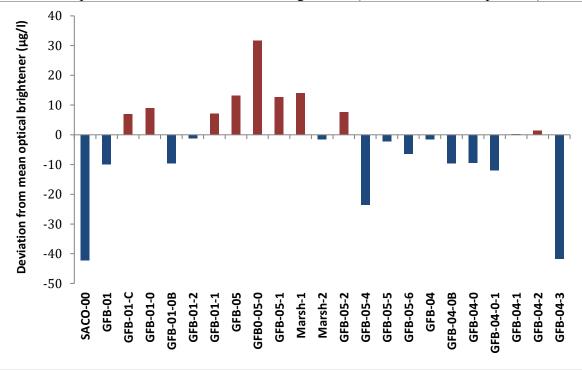


Figure A3. Deviations from the 2012-2017 combined mean OB value for all NSRT sites. Bars above the X-axis indicate sites where OB values were greater than the average value and bars below represent those that were lower than the average value (See table A1 for sample sizes).

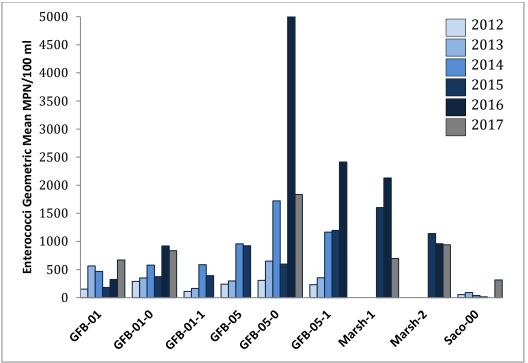


Figure A4. ENT geometric mean for priority sites within the GFB-01 and GFB-05/Marsh series from 2012-2017 (Note differences in sample size (Table A1)).

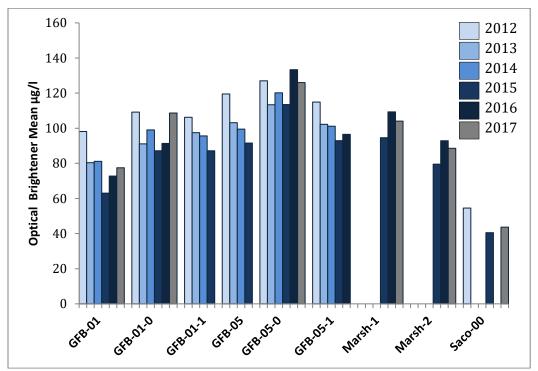


Figure A5. OB mean values for priority sites within the GFB-01 and GFB-05 series from 2012-2017 (Note differences in sample size (Table A1)).

Table A2. 2017 data summary for Goosefare Brook watershed monitoring including the mean enterococci concentration, geometric mean enterococci concentration, mean optical brightener concentration and the sample size at each site for enterococci and optical brightener samples.

Site	Mean ENT	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
Marsh-1	1757.43	696.88	104.04	7	7
Marsh-2	1969.14	940.64	88.57	7	7
GFB-05-0	2055.14	1835.55	126.00	7	7
GFB-01-0	1452.13	835.84	108.66	8	8
GFB-01	1661.00	670.35	77.49	7	7
Saco-00	574.80	314.70	43.68	5	5
GFB-01-0B*	959.00	959.00	112.00	1	1
GFB-01-1*	833.00	833.00	108.00	1	1
CB-C/WG*	109.00	109.00	44.60	1	1
Total	1556.61	772.08	93.75	44	44

*Single sample values given.

Site	Mean ENT	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
GFB-01	428.88	323.91	72.76	8	8
GFB-01-0	920.00	656.03	91.35	2	2
GFB-01-B*	1236	1236	94.4	1	1
GFB-01-C	1806.00	1536.57	95.10	2	2
Porter Ave*	31	31	50.6	1	1
Marsh-1	3430.38	2129.67	109.31	8	8
Marsh-2	1640.75	959.63	92.88	8	8
GFB-05-0	5150.33	5068.33	133.33	3	3
GFB-05-1	2413.67	2238.24	96.52	3	3
Total	2039	1021	95	36	36

Table A3. 2016 data summary for Goosefare Brook watershed monitoring including the mean enterococci concentration, geometric mean enterococci concentration, mean optical brightener concentration and the sample size at each site for enterococci and optical brightener samples.

*Single sample values given.

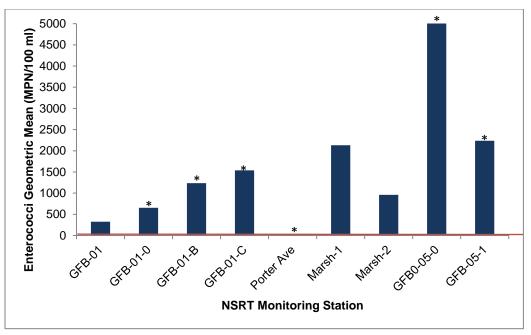


Figure A6. The 2016 enterococci geometric mean (MPN/100ml) values by monitoring station as indicated by blue bars. Red solid line indicates safety level of 35 MPN/100ml. Asterisks indicate values based on fewer than 5 samples.

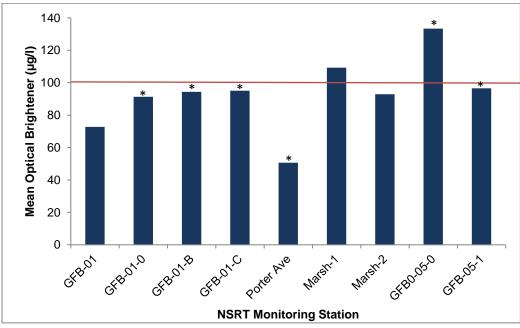


Figure A7. NSRT mean optical brightener ($\mu g/l$) concentrations by monitoring station for 2016. Red solid line indicates optical brightener lower threshold (100 $\mu g/l$) indicating the potential for human wastewater contamination. Asterisks indicate values based on fewer than 5 samples.

Site	Mean ENT	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
GFB-01	264.3	182.0	63.0	11	11
GFB-01-0	460.4	373.6	87.2	11	11
GFB-01-1	434.6	391.6	87.2	11	11
GFB-04	182.0	132.1	80.6	11	11
GFB-04-0	95.6	83.3	70.5	10	10
GFB-04-0-1	55.1	43.9	68.1	10	10
GFB-04-0B	97.8	77.4	70.1	10	10
GFB-05	1071.9	923.8	91.6	11	11
GFB-05-0	865.3	597.1	113.5	11	11
GFB-05-1	1330.2	1196.5	92.9	11	11
GFB-05-2	51.4	30.9	91.0	10	10
GFB-05-4	97.6	25.9	54.6	10	10
GFB-05-5	14.8	9.0	74.6	10	10
Marsh-1	2266.4	1603.5	94.6	9	9
Marsh-2	1581.8	1140.6	79.5	9	9
SACO-00	66.5	14.5	40.5	11	11
Total	538	160	80	166	166

Table A4. 2015 data summary for Goosefare Brook watershed monitoring including the mean enterococci concentration, geometric mean enterococci concentration, mean optical brightener concentration and the sample size at each site for enterococci and optical brightener samples.

Total53816080166166*Note sample size does not reflect duplicates (field and lab) or FYI sampling events. Sample size including FYI sites = 171 for both parameters.

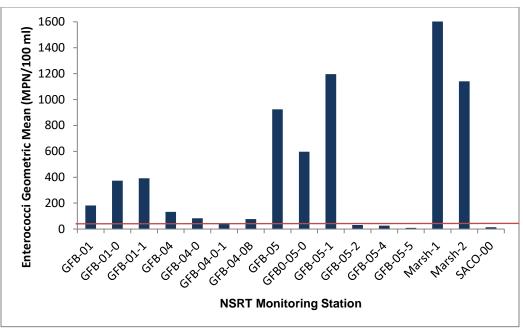


Figure A8. The 2015 enterococci geometric mean (MPN/100ml) values by monitoring station as indicated by blue bars. Red solid line indicates safety level of 35 MPN/100ml.

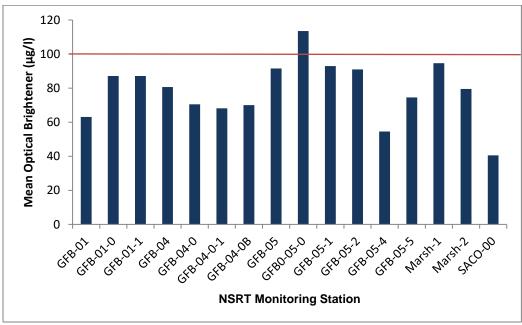


Figure A9. NSRT mean optical brightener ($\mu g/l$) concentrations by monitoring station for 2015. Red solid line indicates optical brightener lower threshold (100 $\mu g/l$) indicating the potential for human wastewater contamination.

Table A5. 2014 data summary for Goosefare Brook watershed monitoring including the mean enterococci concentration, geometric mean enterococci concentration, mean optical brightener concentration and the sample size at each site for enterococci and optical brightener samples.

Site	Mean ENT	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
GFB-01	627.9	467.1	81.2	16	11
GFB-01-0	650.4	578.4	99.1	11	11
GFB-01-1	647.4	586.4	95.6	10	10
GFB-01-2	579.3	476.5	89.8	10	10
GFB-04	233.6	191.8	88.4	10	10
GFB-04-0	266.4	193.4	81.4	11	11
GFB-04-0-1	276.3	186.4	78.7	11	11
GFB-04-0B	226.3	188.6	82.5	10	10
GFB-05	1143.4	958.2	99.5	11	11
GFB-05-0	2276.4	1721.9	120.2	10	10
GFB-05-1	1500.1	1165.7	101.2	11	11
GFB-05-2	121.1	82.5	115.5	11	10
GFB-05-4	209.6	139.6	63.4	10	10
GFB-05-5	28.1	14.1	91.1	10	10
SACO-00	509.4	37.8	NA	4	NA
Total	624	276	92	156	146

*Note sample size does not reflect duplicates (field and lab) and includes 8/14/15 sampling event. Those results are not included in analyses.

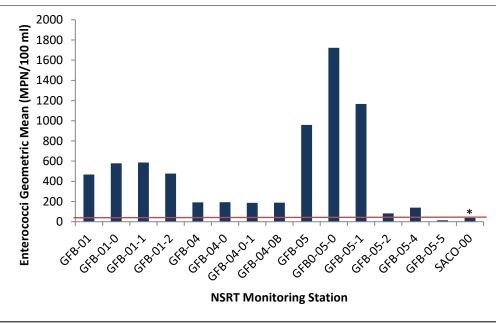


Figure A10. The 2014 enterococci geometric mean (MPN/100ml) values by monitoring station as indicated by blue bars. Red solid line indicates safety level of 35 MPN/100ml.

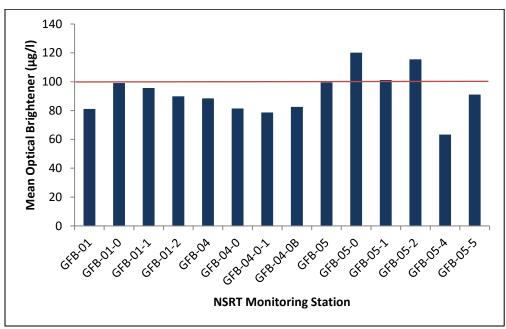


Figure A11. NSRT mean optical brightener $(\mu g/l)$ concentrations by monitoring station for 2014. Red solid line indicates optical brightener lower threshold (100 $\mu g/l$) indicating the potential for human wastewater contamination.

Site	Mean ENT	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
GFB-01	1347.3	564.2	80.4	14	10
GFB-01-0	449.0	350.7	91.1	10	10
GFB-01-0B	181.0	179.5	37.6	2	3
GFB-01-1	213.2	163.9	97.5	10	10
GFB-04	207.9	183.3	88.1	9	9
GFB-04-0	132.9	108.9	81.9	9	9
GFB-04-0-1	188.1	131.3	79.8	9	9
GFB-05	315.6	297.2	103.2	10	10
GFB-05-0	729.9	650.1	113.4	9	9
GFB-05-1	381.9	354.2	102.2	10	10
GFB-05-2	89.8	52.8	102.8	9	9
SACO-00	2039.2	91.0	-	5	-
GFB-01-2	658.3	611.4	77.6	3	3
GFB-04-0B	181.1	152.5	83.7	9	9
GFB-05-4	37.7	27.1	75.9	10	10
GFB-05-5	25.7	19.4	92.8	10	5
GFB-05-6	74.7	44.6	81.7	9	6
Total	409	148	89	147	131

Table A6. 2013 data summary for Goosefare Brook watershed monitoring including the mean enterococci concentration, geometric mean enterococci concentration, mean optical brightener concentration and the sample size at each site for enterococci and optical brightener samples.

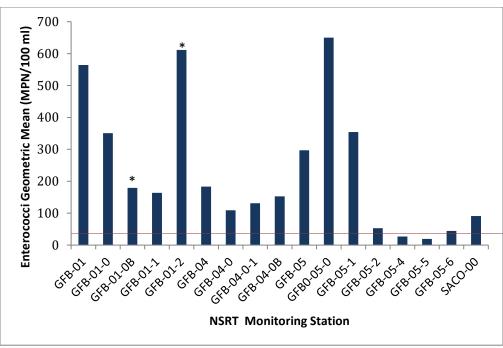


Figure A12. The 2013 geometric mean enterococci (MPN/100ml) values by monitoring station as indicated by blue bars. Red solid line indicates safety level of 35 MPN/100ml. Asterisks indicate values based on fewer than 5samples.

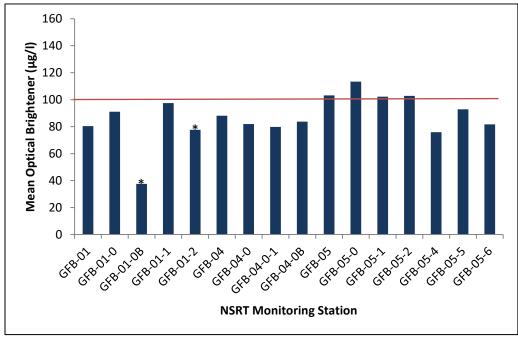


Figure A13. NSRT mean optical brightener ($\mu g/l$) concentrations by monitoring station for 2013. Red solid line indicates optical brightener lower threshold (100 $\mu g/l$) indicating the potential for human wastewater contamination. Asterisks indicate values based on fewer than 5 samples.

Site	Mean ENT	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
GFB-01	268.1	151.4	98.1	8	8
GFB-01-0	334.0	288.6	109.2	5	6
GFB-01-0B	509.5	419.9	143.0	2	2
GFB-01-1	239.6	111.1	106.2	5	6
GFB-04	292.8	200.2	91.7	5	6
GFB-04-0	226.0	103.8	82.6	5	6
GFB-04-0-1	535.0	305.6	80.7	4	5
GFB-04-1	494.5	339.6	88.4	4	5
GFB-04-2	282.0	199.7	89.5	4	5
GFB-04-3	158.5	131.9	46.3	4	5
GFB-05	271.0	239.9	119.5	5	6
GFB-05-0	337.2	307.6	127.0	5	6
GFB-05-1	253.2	230.8	114.9	5	6
GFB-05-2	182.6	140.1	63.5	5	6
SACO-00	117.9	54.2	54.5	7	8
Total	282	174	92	73	86

Table A7. 2012 data summary for Goosefare Brook watershed monitoring including the mean enterococci concentration, geometric mean enterococci concentration, mean optical brightener concentration and the sample size at each site for enterococci and optical brightener samples.

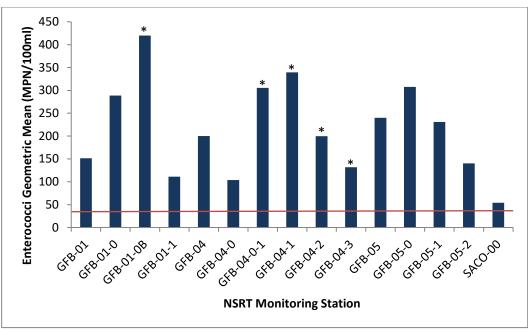


Figure A14. The 2012 geometric mean enterococci (MPN/100ml) values by monitoring station in the NSRT as indicated by blue bars. Red solid line indicates safety level of 35 MPN/100ml. Asterisks indicate values based on fewer than 5 samples.

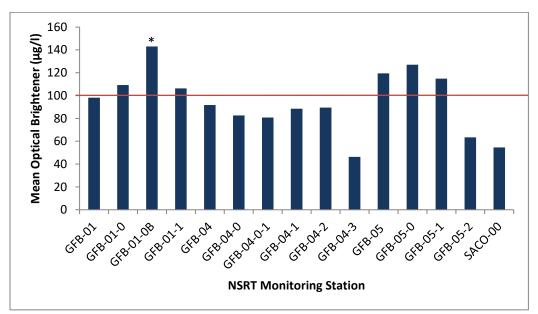


Figure A15. NSRT mean optical brightener ($\mu g/l$) concentrations by monitoring station for 2012. Red solid line indicates optical brightener lower threshold (100 $\mu g/l$) indicating the potential for human wastewater contamination. Asterisks indicate values based on fewer than 5 samples.

ADDITIONAL MONITORING DATA

Seasonal Shifts

Table A8. Total 2012-2017 ENT geometric mean concentration, OB mean concentration, ENT sample size, and OB sample size for each month monitored.

Month	GeoMean ENT	Mean OB	Sample Size ENT	Sample Size OB
May	71.8074	175.0645	31	31
June	92.50282	342.3107	93	103
July	93.04955	922.9624	128	133
August	81.99766	635.6765	146	136
September	80.3215	752.8444	132	135
October	71.50114	706.2603	73	73

Flood vs Ebb Tidal Conditions

Comparison of ENT geometric mean results (2012-2014) for weekly samples collected during all tidal conditions at two sites (GFB-01 and Saco-00) at the mouth of the brook revealed distinct differences between ebb and flood tidal stages (Figure A16). In all years, ENT geometric mean results were greater during flood (incoming) conditions vs. ebb (outgoing) and in many cases, the flood bacteria values were more than double those observed during ebb conditions. Also, for GFB-01 in particular, the bacteria results during both incoming and outgoing tidal conditions appear to be increasing over time (Figures A17, Table A8). Given the documented bacteria issues throughout the GFB watershed, it was expected that ebbing tide conditions would result in greater ENT results compared to flood conditions. Presumably, outgoing tides pull water from tributaries (including contaminates from upland areas) compared to incoming tides when ocean waters mix with the brook. Higher flood tide ENT levels suggest potential pollution source(s) in or near the mouth and/or conditions in this area favor persistence and possibly regrowth of ENT.



Figure A16. Monitoring stations GFB-01 and Saco-00 located at the mouth of the Goosefare Brook.

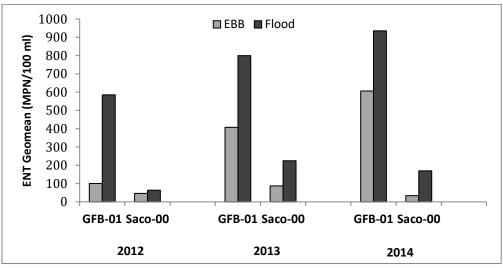


Figure A17. Season-wide ENT geomean results for GFB-01 and Saco-00 samples collected at ebb and flood tidal conditions.

Table A8. 2014 data summary for Goosefare Brook watershed ebb vs. flood
monitoring including the geometric mean ENT concentration and sample size
for both tidal conditions.

Site	Year	GeoMean ENT Ebb	GeoMean ENT Flood	Sample Size Ebb	Sample Size Flood
GFB-01	2012	100.4	584.8	6	8
	2013	407.2	799.7	7	8
	2014	606.0	935.3	7	8
Saco-00	2012	46.4	64.0	6	9
	2013	87.4	225.0	7	8
	2014	34.3	169.9	7	8

Appendix B: Additional Source Tracking Efforts

Pharmaceutical and Personal Care Products (PPCP)

With the help of US EPA, the source tracking toolbox was expanded to include the analysis of 7 PPCPs in 2012 (table B1). The presence of these compounds can be indicative of human sourced fecal contamination. In 2012, US-EPA analyzed PPCPs at 11 of the 15 monitoring locations within the NSRT sub-watershed for 4 of the 6 enhanced monitoring dates (Table B2). US EPA did not provide PPCP support in 2013-2016.

РРСР	Description		
Atenolol	Control high blood pressure		
Acetaminophen	Pain killer		
Cotinine	Metabolite of nicotine		
1,7-Dimethylxanthine	Metabolite of caffeine		
Caffeine	Stimulant		
Carbamazepine	Control seizures		
Metoprolol	Control high blood pressure		

Table B1. PPCPs monitored at selected stations within the NSRT in 2012.

PPCP results indicated that all 11 sites monitored had detectable limits of 1,7-dimethylxanthine, caffeine and cotinine (Table B2). These results are likely due to human sources in the NSRT watershed as 1,7-dimethylxanthine (caffeine metabolite) and cotinine (nicotine metabolite) pass through the human body.

Table B2. Mean concentration (n=4) of pharmaceutical compounds (ng/l) for 11 monitoring stations within the New Salt Road Tributary sub-watershed. Rows highlighted in red indicate \geq 4/7 pharmaceutical compounds present at or above the assay detection limit.

MONITORING STATION	1,7- DIMETHYLXANTHINE	ACETAMINOPHEN	ATENOLOL	CAFFEINE	CARBAMAZEPINE	COTININE	METOPROLOL
GFB-01	2.60	-	-	21.97	-	1.90	-
GFB-01-0	2.30	-	-	7.50	-	3.10	-
GFB-01-1	3.80	-	-	16.75	-	1.55	-
GFB-01-B	1.80	-	-	9.70	-	3.60	-
GFB-04-0	3.80	-	-	15.00	-	4.25	-
GFB-04-0-1	4.30	19.00	-	16.00	-	1.50	-
GFB-04-2	10.60	-	-	18.48	-	2.13	-
GFB-04-3	4.70	-	-	36.00	-	0.76	-
GFB-05-0	3.47	7.35	-	23.43	-	6.70	1.00
GFB-05-1	2.90	2.60	-	23.00	-	5.70	-
GFB-05-2	9.20	-	-	21.40	-	1.33	_

As all sites monitored for PPCPs had detectable limits for the same 3 compounds, monitoring sites (GFB-04-0-1, GFB-05-0, and GFB-05-1) with 4 or more detectable limits out of the 7 PPCP compounds tested were considered to "stand out" in this context. The presence of multiple compounds is likely a "red flag" prompting the need for further investigation into potential illicit discharges in the areas surrounding those monitoring locations. In general, as the number of PPCP compounds with detectable limits increases, so

does the likelihood of human sources of pollution are impacting water quality at or near the monitoring site. Further monitoring is recommended to increase the sample size.

Canine Detection Services

A separate study funded by the Ocean Park Conservation Society and conducted by FB Environmental Associates in partnership with Environmental Canine Detection Services was conducted to "sniff" our human sources contributing to elevated bacteria concentrations. This study involved the collection of Enterococci samples while employing 2 sewage-sniffing dogs at 14 of the 15 locations throughout the NSRT watershed in 2012. The canines are trained to alert their trainers to the presence of human sources at distinct locations or in water samples collected from suspect areas. All the sites monitored during this event, excluding Saco-00, exceeded the US EPA-recommended single sample threshold of 61 MPN/100mls for freshwater sites and 104 MPN/100mls for tidally influenced sites. Human sewage was detected by one dog at GFB-04-0-1, by the other dog at GFB-05-1, and both dogs alerted for human sewage at GFB-01. The canines were not part of the GFB source tracking work in 2013-2017.

Appendix C: Local Assessments

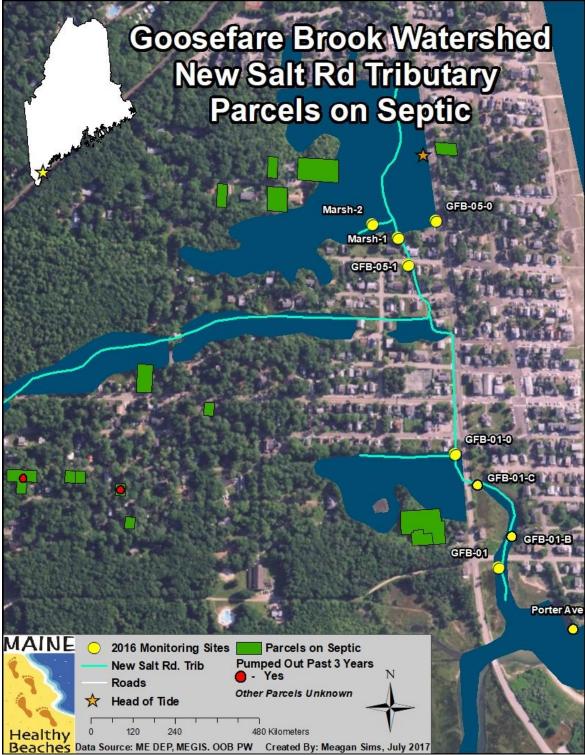


Figure C1. Known parcels on septic in close proximity to the NSRT and 2016 MHB monitoring locations. This figure may not contain all relevant information and it will be periodically updated as new information is received by MHB.

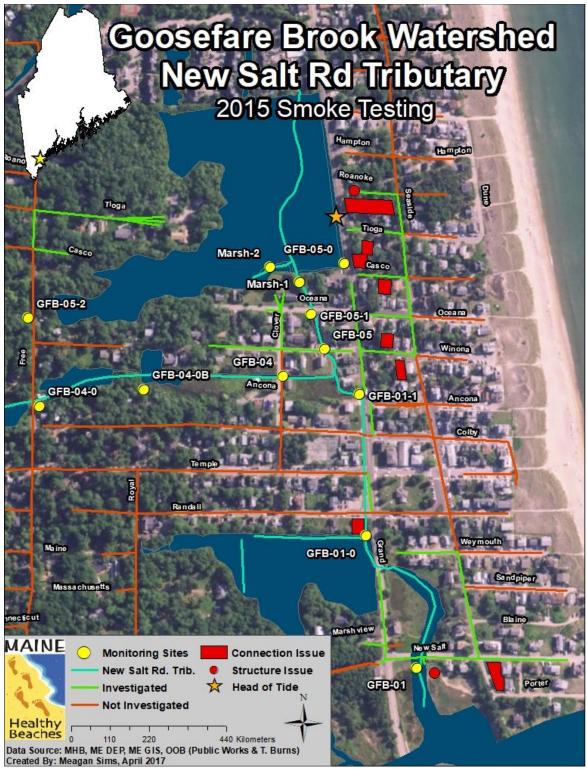


Figure C2. Results from 2015 sanitary system smoke testing and MHB monitoring locations along the New Salt Rd. Tributary. This figure may not contain all relevant information and it will be periodically updated as new information is received by MHB.



Figure C3. Old Orchard Beach wastewater infrastructure pipe installation year and 2016 MHB monitoring locations along the New Salt Rd. Tributary. This figure may not contain all relevant information and it will be periodically updated as new information is received by MHB.

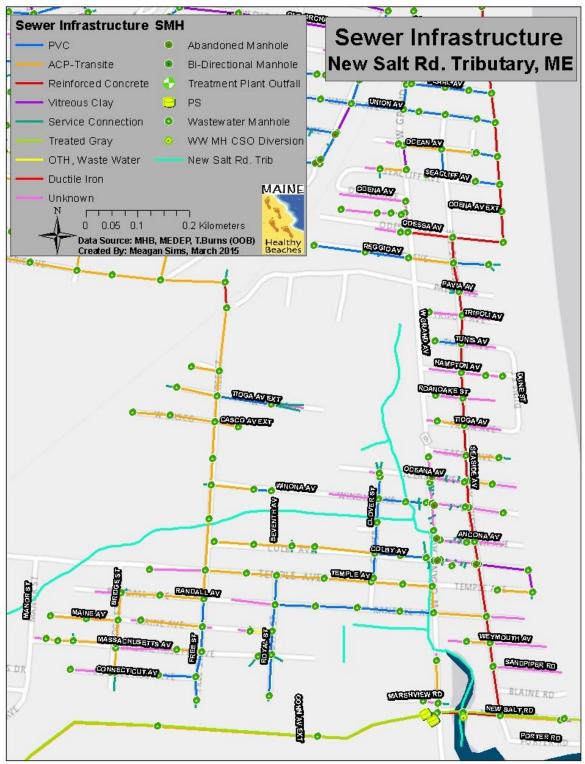


Figure C4. Old Orchard Beach wastewater infrastructure materials (pipe type) and MHB monitoring locations along the New Salt Rd. Tributary. This figure may not contain all relevant information and it will be periodically updated as new information is received by MHB (No updates received for the 2016 season).

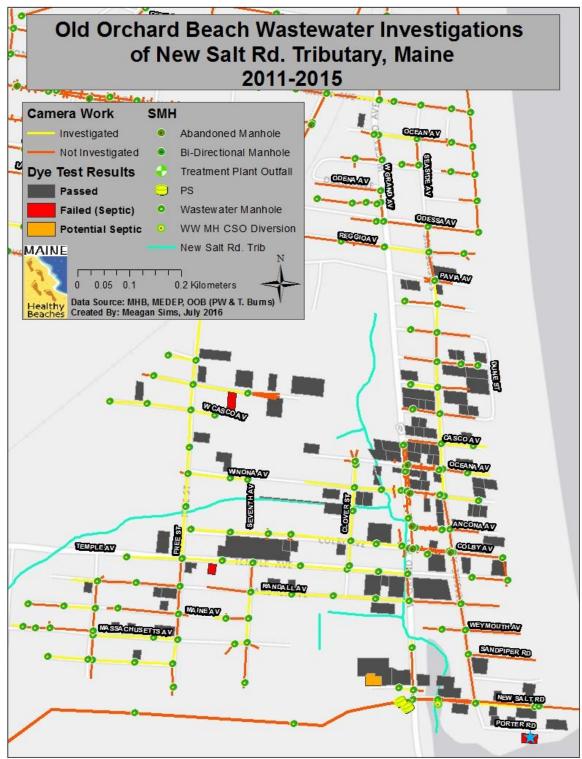


Figure C5. OOB wastewater camera and dye test investigations conducted by Public Works from 2011 to 2015 along the NSRT. This figure may not contain all work completed and it will be periodically updated as new information is received by MHB. Parcel on Porter road (\uparrow) identified as being served by a cesspool. Cesspool removed (2014) and property tied into sanitary system (No updates received for the 2016 season).