1 2	Wood Traits and Tidal Exposure Mediate Shipworm Infestation and Biofouling in Southeastern U.S. Estuaries
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25 Abstract

26 Annually, shipworms and other biofouling species cause millions of dollars in damage to 27 wooden marine infrastructure across the world. Given their abundant larval supply and high 28 dispersal potential, bioeroders and biofoulers are ubiquitous threats that shorten the lifespan of 29 wooden docks, piers, boats and shoreline stabilization structures in coastal environments. Despite 30 these impacts, there are no treatments that completely protect wood against shipworms and 31 biofouling. To explore potential approaches for extending the lifespan of wooden shoreline 32 stabilization structures, we conducted two field experiments to evaluate the resistance to 33 shipworms and biofouling of small and large diameter branches of four trees – laurel oak 34 (Quercus hemisphaerica), sweetgum (Liquidambar styraciflua), crepe myrtle (Lagerstroemia 35 spp.), and black mangrove (Avicennia germinans) – positioned at varying distances from the sediment surface in southeastern US estuaries. We discovered that the wood volume lost to 36 37 shipworm burrows was concentrated near the sediment surface, more prevalent in tree species 38 with lower wood densities, and varied markedly between years. Barnacle fouling was far higher 39 on branches > 30cm from the surface and on laurel oak and sweetgum branches. In a third field 40 experiment, we tested two chemical and two non-chemical wood treatments and found chemical 41 treatments to be more effective at deterring barnacle fouling and shipworm burrowing of wooden 42 posts, especially beneath the sediment surface. By identifying desirable characteristics of the 43 wood employed and elevations at which the impacts of shipworms and biofouling are especially 44 prevalent, this experimental study informs the design of more durable wooden stabilization 45 structures in coastal environments.

46

47 **1.** Introduction

48 An important challenge for the functioning and durability of marine infrastructure is 49 biofouling and bioerosion- the growth of barnacles, algae, sponges, and other sessile organisms 50 on or within submerged or partially submerged structures (Richmond & Seed 1991, Callow & 51 Callow 2002, Srivutha Murthy et al. 2009). For instance, biofouling can corrode and degrade 52 metal structures including offshore oil rigs (Edyvean et al. 1988, Stevenson et al. 2011, Yang et 53 al. 2014, Lingvay et al. 2018) and obstruct the optical window of submerged sensors (Sriyutha 54 Murthy et al. 2009). Because this pervasive and widespread problem affects a variety of marine 55 structures and instruments, significant investment continues to be made to combat biofouling 56 (Alberte et al. 1990, Schultz et al. 2011).

57 One material particularly vulnerable to biofouling is wood. Wooden ships can be 58 adversely affected by the settlement and growth of oysters, barnacles and other sessile 59 invertebrates, and by the infestation of boring bivalve molluscs of the genera Teredo, Bankia, 60 and Lyrodus, collectively known as shipworms. Barnacles, for instance, increase drag on wooden 61 ships, resulting in increased fuel consumption (Schultz et al. 2011, Lindholdt et al. 2015). In 62 contrast, shipworms bioerode wooden structures with the aid of symbiotic gut bacteria (Rice et 63 al. 1990, Lopez-Anido et al. 2004, Nelson 2015) and for centuries have posed a problem for 64 humans. In the 1730s, shipworms caused extensive damage to wooden 'wave breakers', which 65 left Dutch dikes and the cities that they protected vulnerable to storm surge and damage (Sundberg 2015). Likewise, the introduction of *Teredo navalis* into the West Coast of the United 66 67 States reached epidemic proportions between 1880 and 1920 and caused massive damage to 68 infrastructure due to this shipworm bioeroding wharves, piers, and docks to the point of collapse. 69 The costs of rebuilding and treating structures against future boring and biofouling in San

70 Francisco Bay after this epidemic, along with the economic losses due to lost business from 71 damaged infrastructure, totaled approximately half a billion US dollars (Nelson 2015). In the US, 72 somewhat outdated estimates (current estimates do not exist to the best of our knowledge) 73 indicate that costs associated with damages to infrastructure due to shipworms alone is \$205 74 million US dollars per year (Pimentel et al. 2000), further highlighting the substantial drain on 75 marine and coastal economies that these burrowing molluscs and other biofoulers impose. 76 Although shipworms and biofouling species are prevalent pests in marine systems, they 77 are especially problematic in coastal and estuarine environments that harbor many wooden 78 structures (e.g. wharves, piers, docks) and significant wooden debris (e.g. fallen trees). 79 Shipworms can tolerate salinities ranging from 5-35 ppt, although their boring activity stops at 80 salinities below 10 ppt, and peaks at higher salinities resembling those of the open ocean 81 (Barrows 1917, Paalvast & van der Velde 2011a). Optimal water temperature for shipworm 82 growth ranges between 15-25°C (Paalvast & van der Velde 2011a), however shipworms are able 83 to spawn as soon as temperatures exceed 11-12°C (Graves 1928). In addition, females spawn 3-4 84 times per reproductive season, releasing between 1-5 million larvae at a time. These veliger 85 larvae occur in a planktonic stage for up to three weeks before settling on wooden substrates 86 (Grave 1928, Grave 1942). During this stage, shipworms have the potential to disperse hundreds 87 of kilometers in currents and ballast water (Scheltema 1971). Barnacles and oysters, the 88 biofouling taxa observed in this study, exhibit similar life history traits and tolerances to salinity 89 and temperature fluctuations, although they are encrusting, sessile filter feeders rather than 90 wood-consumers (Strathmann et al. 1981, Trager et al. 1990, Qiu & Qian 1999). Collectively, 91 these life history traits make shipworms, barnacles and oysters well-equipped to survive, persist 92 and damage wooden structures in tropical and temperate estuaries worldwide.

93 To prolong the lifespan of wooden structures in estuarine environments, humans have 94 employed different methods. Historic accounts show that ancient Egyptians and Chinese 95 protected wooden structures with resin, pitch, and paint (Borges 2014). Other cultures placed copper or lead plates on wooden ships, as well as used paraffin, tar, and asphalt on piers, 96 97 wharves, and other structures, to protect against bioerosion and biofouling (Paalvast & van der 98 Velde 2011b). Creosote – a material derived from the carbonization of coal and one of the most 99 effective protective measures against marine wood borers - has also been applied commonly to 100 marine timber but has been banned or highly regulated in many countries due to its carcinogenic 101 properties (Hoppe 2002, Ohgami et al. 2015). More modern approaches include treating wood 102 with chromated copper arsenate (CCA), which is relatively effective at deterring biofouling 103 (Weis & Weis 1992). CCA is widely used to protect wood but remains a controversial approach 104 due to its negative environmental effects (Edwin & Sreeja 2011, Paalvast & van der Velde 105 2011b). Although shipworms, barnacles, and other biofouling organisms have posed a problem 106 for centuries, no method has been developed that is one hundred percent effective at preventing 107 their settlement and growth on wooden structures (Borges 2014). 108 The lack of treatment against shipworms and biofouling, and limited understanding of 109 their ecological impacts on various wood types is problematic because wood continues to be a 110 commonly used construction material in estuaries (Borges et al. 2003). In particular, wood is

often used in the construction of shoreline stabilization structures, including bulkheads and breakwalls. Breakwalls, also known as groynes in Europe, are composed of wooden piles or fence posts that are filled with brush, branches or small trees. Given their porous nature and construction just offshore, breakwalls are designed to decrease wave or boat wake energy acting on the shoreline edge and facilitate sediment deposition (Herbert et al. 2018). First built in the 116 North Sea in Germany in 1815, breakwalls continued to be constructed today and are preferred 117 over conventional hardened, non-permeable breakwalls because they are less expensive and 118 result in less scouring and erosion of sediment in adjacent unprotected shorelines (Bakker et al. 119 1984, Orford 1988, Weichbrodt 2008, Herbert et al. 2018). For these same reasons, shoreline 120 stabilization and restoration efforts in Germany, the Netherlands, US, and other countries are 121 now adopting more natural approaches including breakwalls (Poff et al. 2004, Borsje et al. 2011, 122 Lippert et al. 2017). In the coming years, the use of wooden shoreline stabilization structures 123 may increase in the face of sea level rise and increased shoreline erosion (Bulleri and Chapman 124 2010). Despite the forecasted increase in their use, it remains unclear how their design might be 125 optimized to enhance their longevity in the face of shipworms and other forms of biofouling.

126 To better understand the environmental and substrate characteristics that modulate 127 shipworm infestation and biofouling of intertidal breakwall branches and posts, we conducted a 128 6-month field experiment to test how distance from the sediment surface, tree species identity, 129 branch diameter, and site interact to mediate shipworm burrow density and percentage of wood 130 volume lost to burrowing in two northeast Florida estuaries (Experiment 1). We then tested how 131 tree species identity and distance from sediment mediate patterns in shipworm infestation and 132 barnacle and oyster settlement across southeastern US estuaries by replicating this experiment 133 for three months in the same two sites and at four additional sites (Experiment 2). Finally, we 134 compared barnacle and oyster colonization as well as shipworm infestation of two non-chemical 135 (tape and silicone wraps) and two chemical techniques (pressure-treated wood and copper-based 136 antifouling paint) meant to protect wooden posts against biofouling and enhance their longevity 137 to unprotected, control posts (Experiment 3).

138 For Experiments 1 and 2, we hypothesized that: (1) shipworm burrow density and wood 139 volume loss as well as barnacle and oyster density would be highest on branches located close to 140 the sediment surface that are inundated for longest and negligible for branches buried underneath 141 the sediment surface due to anoxic conditions, (2) small branches will lose a higher percentage of 142 wood volume to shipworms, but large branches will have higher shipworm burrow densities, and 143 (3) branches with high wood densities (laurel oak and mangrove) will experience less damage 144 than those with low wood densities (crepe myrtle and sweetgum). For Experiment 3, we 145 hypothesized that chemical treatments would result in wooden posts having fewer barnacles, 146 oysters, and shipworm burrows than non-chemical treatments, but that non-chemical treatments 147 would have less damage than unprotected controls. Together, these three experiments inform the 148 ecologically-engineered design of wooden breakwalls for shoreline protection in the southeastern 149 US, a region where lateral loss of shorelines is pervasive due to boat traffic and high-energy 150 wave environments (Morton 2003, Herbert et al. 2018). 151 Materials and methods 2. 152 2.1 **Study sites** 153 Experiments 1 and 3 were conducted in two tidal creeks within the Matanzas River Estuary in St. Augustine, Florida, USA (Site 1: 29° 45' 47.9592" N, 81° 15' 46.242" W and Site 154 155 2: 29° 51' 57.7584" N, 81° 18' 48.5316" W, Fig. 1). These sites are exposed to semidiurnal tides 156 ranging from -0.25 m to 1.25 m above Mean Lower Low Water (MLLW), experience 157 temperatures between 22-35°C in summer and from 4-25°C in winter, and receive an annual 158 average of 113 mm of precipitation per month (NOAA National Centers for Environmental 159 Information 2018). The tidal creeks were surrounded by salt marsh dominated by smooth 160 cordgrass (Spartina alterniflora). Black (Avicennia germinans) mangroves were also present at 161 both sites and occurred as isolated trees. Eastern oyster, Crassostrea virginica, reefs were also

162 common at the lower intertidal margins of the salt marsh habitat. The experiments were deployed
163 approximately 30cm below the lower elevation of naturally occurring oyster reefs in exposed
164 intertidal mudflats as this elevation is where breakwalls are typically deployed for shoreline
165 stabilization in the region (Herbert et al. 2018).

166 2.2 Experiment 1: Tree Species, Branch Diameter, Elevation and Site Effects

167 We tested four tree species' susceptibility to shipworm infestation and biofouling: laurel 168 oak (Quercus hemisphaerica), sweetgum (Liquidambar styraciflua), crepe myrtle 169 (Lagerstroemia spp.), and black mangrove (Avicennia germinans). The first three species were 170 selected because of their abundance in the region and thus availability for use in breakwall 171 construction. Laurel oak and sweetgum are native to Florida, while crepe myrtle is an introduced 172 ornamental species that is now well established in the region. Black mangroves are also common 173 in Florida and, due to their natural exposure to shipworms, barnacles and oysters as a result of 174 their intertidal estuarine distribution, we anticipated that this species would be more resistant to 175 shipworm infestation and biofouling. However, mangroves cannot be harvested without a permit 176 and were investigated in this study as a useful comparison from which to gauge the vulnerability 177 of the other tree species to bioerosion and biofouling.

For each tree species, we tested two branch diameter classes relevant for filling breakwalls given their availability and ease of handling, large and small. Because of the natural distribution of branch sizes, diameter classes differed slightly among species. Laurel oak had a small diameter class ranging from 1-2.5 cm and a large diameter class ranging from 2.5-5 cm. Crepe myrtle and sweetgum had a small diameter class ranging from 1-2 cm and a large diameter class ranging from 2-4 cm. Mangrove branches had a small diameter class ranging from 1-1.5 cm and a large diameter class ranging from 1.5-3.5 cm.

185 To compare shipworm infestation and biofouling prevalence between tree species, 186 diameter classes, and distances from the sediment surface, we built 'ladders' using PVC poles as 187 the ladder sides and tree branches as rungs (Fig. 2). Each ladder from the small diameter class 188 had a total of nine, 50 cm-long branch rungs secured to the PVC pipes with cable ties at 189 distances of -10, -5, 0, 5, 10, 20, 30, 40, and 50 cm from the sediment surface. Each ladder in the 190 large diameter classes had a total of seven, 50 cm-long branches secured at distances of -10, 0, 191 10, 20, 30, 40, and 50 cm from the sediment; note that the -5 and 5 cm rungs were omitted from 192 large branch ladders due to space constraints. Five replicate ladders were built for each study site 193 and diameter class for laurel oak, sweetgum, and crepe myrtle branches. However, due to a lack 194 of available tree branches and legislation in Florida restricting mangrove trimming, small and 195 large diameter mangrove ladders had only 5 branches each, positioned at -10, 0, 10, 20, and 30 196 cm from the sediment (N=5 small diameter and N=4 large diameter class ladders). In July 2016, 197 the ladders were driven by hand into the sediment at a spacing of 1m in the intertidal mudflat and 198 2-3m from the salt marsh shoreline edge at each study site. The elevations of the ladders were -199 0.57 to -0.73 m and -0.51 to -0.59 m above mean low water at sites 1 and 2, respectively. These 200 locations were selected to mimic the location where breakwalls are typically built to stabilize 201 eroding shorelines. The ladders were retrieved six months later in January 2017. 202 In the lab, number of barnacles and oysters were counted on each branch to evaluate

biofouling. Oysters were rarely observed on branches, averaging only 0.5 oysters per branch across the 562 branches deployed, so are not discussed further in the main text (see Appendix A1 for summary of results). We then measured the length and diameter of each branch in order to calculate the initial wood volume by multiplying the cross-sectional area of the branch times its length. We then cut each branch into 5cm-long segments and, for each segment, counted the 208 number of shipworm burrows. Using calipers, we then measured the diameter and depth of 10 209 burrows per segment. If less than 10 burrows were observed, all burrows were measured. We 210 used the burrow diameter and depth to calculate the volume of each of these 10 burrows by 211 multiplying the area of the burrow opening times the burrow depth. We then averaged these 212 volumes, multiplied this value by the number of burrows per segment, and summed these values 213 across each branch to estimate the total wood volume lost to shipworm burrowing. Finally, we 214 calculated the percent of branch wood volume lost to shipworms by dividing the total volume of 215 wood lost to shipworm burrowing by the initial wood volume.

216 2.3 Experiment 2: Regional Study of Tree Species, Elevation and Site Effects

217 To evaluate potential spatial variation across the region and assess interannual variability 218 in shipworm infestation rates and biofouling at the two sites evaluated in Experiment 1, we 219 repeated this experiment in June 2017 in the same two sites used in Experiment 1 and deployed 220 ladders at 4 additional sites along the southeastern US coast (Cedar Key, FL, Withlacoochee 221 Bay, FL, Suwannee River, FL, and Sapelo Island, GA). Based on results from Experiment 1, we 222 used only small diameter laurel oak and sweetgum ladders with branches positioned at distances 223 from 0-30 cm above the sediment in 10 cm intervals as these tree species varied in their 224 vulnerability to shipworms and biofouling, and these diameter class and positions were most 225 vulnerable to shipworms and thus best suited for assessing spatial and interannual variation in 226 bioerosion and biofouling rates. At each site, we deployed 5 replicates of each ladder in June 227 2017 and retrieved them September 2017. Branches were brought back to the lab and analyzed in 228 the same way as branches in Experiment 1 – barnacles and oysters were counted on each branch 229 and branches were cut into 5-cm segments and inspected for shipworm burrows. Similar to 230 Experiment 1, oysters were relatively rare and only reported on further in Appendix A2.

231 **2.4** Experiment 3: Anti-Fouling Techniques for Wooden Substrates

232 We tested five wood protection treatments against biofouling: CCA pressure-treated 233 fence posts, Rust-Oleum 207012 Marine Flat Boat Bottom commercial copper anti-fouling paint, 234 1.5-cm thick silicone wraps made with Smooth-On Mold Max silicone mold-making rubber, 235 Gorilla duct tape, and an untreated control (Appendix B). The pressure-treated fence posts were 236 8.9-cm diameter by 1.5m long, livestock fence posts that are often used in groyne, breakwall, 237 seawall and bulkhead construction, while the other three treatments were applied to 5.08cm 238 x5.08cm by 1.5m long untreated spruce pine fir wooden posts. The treatments were applied to 239 the middle 1m of the post, which was then driven into the ground making sure that 0.5m of 240 treated surface was underground and 0.5m was above the sediment. The copper paint, silicone 241 wrap and duct tape were utilized as treatments because all three are easy to acquire and/or 242 inexpensive and easy-to-apply materials. Five replicates of each treatment were deployed in July 243 2016 in sites 1 and 2 (same sites as in Experiment 1). They were retrieved on January 2018, after 244 18 months, and brought to the laboratory for processing. Biofouling was assessed for each post 245 by counting all barnacles and oysters per post and dividing this number by the surface area of 246 each post to standardize these values. Shipworm damage was quantified by cutting each post at 247 the -10, -5, 0, 5, 10, and 20 cm mark and estimating the percent of the cross-sectional area 248 burrowed by shipworms. These posts were left in the ground for significantly longer than the 249 branches from Experiments 1 and 2 to explore the potential long-term efficacy of these 250 protective treatments in reducing the rate of biofouling and bioerosion on wooden breakwalls. 251 (See Table 1 for summary of all three experiments.)

252 2.5 Statistical Analyses

To evaluate the significance and relative importance of site, distance from the sediment surface, tree species, and diameter in explaining variation in the number of barnacles per branch, 255 the number of oysters per branch (Appendix A), percent wood volume lost, and shipworm 256 burrow density in Experiment 1, we developed a regression tree using the analysis of variance 257 (ANOVA) method of recursive partitioning for each response variable. We then pruned over-258 fitted trees using k-fold cross-validation (see Gittman et al. 2015 for details). Regression trees 259 were made using R version 3.2.2 and the R package "rpart" (Therneau et al. 2018). We utilized 260 regression trees to both facilitate identification of the relative importance of the four fixed factors 261 (i.e. the factors that explain the most variation in each response metric are found at the base of 262 the regression tree) and overcome challenges associated with interpreting complex, multi-factor 263 interactions that can arise from four-factor ANOVA. However, because the regression tree 264 revealed branch diameter to be of little significance (p>0.5) in predicting the number of 265 barnacles per branch, we simplified our analytical approach and evaluated the effect size and significance of site, distance from the sediment, and tree species on this specific response 266 267 variable with a three-way ANOVA. Post hoc analyses were performed using Tukey HSD test. 268 For Experiment 2, we used a three-way site*distance from the sediment*tree species 269 ANOVA to evaluate the effect size and significance of these fixed factors and their interactions 270 on the number of barnacles per branch. Due to the lack of shipworm burrows obtained in 271 Experiment 2 (3 total burrows in 240 branches), no statistical analyses were run for the percent 272 wood volume lost and shipworm burrow density response variables. 273 To evaluate the significance and relative importance of site and wood protection 274 treatment on the number of barnacles and oysters (Appendix A) per unit surface area of post and

276 run with these two variables as fixed factors. Separate ANOVAs were run for the percentage of

percent area burrowed by shipworms on wooden posts in Experiment 3, two-way ANOVAs were

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area burrowed by shipworms and the number of barnacles per branch at each distance from the
sediment (i.e. -10, -5, 0, 5, 10, and 20 cm from the sediment surface).

279 3. Results

280 **3.1 Biofouling**

3.1.1 Experiment 1

282 Interactions between distance from the sediment and tree species and between distance 283 from the sediment and site, as well as the main effects of distance from sediment, tree species 284 and site (all: p<0.0001) significantly affected barnacle fouling of branches in Experiment 1 (Fig. 285 3A). At farther distances from the sediment (\geq 30 cm), the number of barnacles per branch 286 increased dramatically, reaching > 50 barnacles per branch, on laurel oak and sweetgum 287 branches, but remained below 30 barnacles on crepe myrtle and mangrove branches (distance 288 from sediment * tree species: $F_{3.546}=15.4$, p<0.0001). At distances close to the sediment (i.e. 289 between -10 and 10 cm), all tree species had few barnacles per branch, densities that steadily 290 increased with increasing distance from the sediment (distance from sediment * site: $F_{1.546}=16.7$, 291 p<0.0001), especially at site 2 where approximately 50% more barnacles were observed (site: 292 $F_{1.546}=23.7$, p<0.0001). In general, branches positioned ≥ 30 cm supported significantly more 293 barnacles than those positioned at lower distances (distance from the sediment: $F_{1,546}=348.2$, 294 p < 0.0001) and barnacle density was considerably higher on laurel oak (36 ± 3 barnacles per branch, mean \pm SEM here and below), sweetgum (27 \pm 3 barnacles per branch) and crepe myrtle 295 296 $(18 \pm 2 \text{ barnacles per branch})$ than mangrove $(7 \pm 1 \text{ barnacles per branch})$, tree species: 297 F_{3,546}=20.7, p<0.0001, Fig. 3, Table 2, Appendix C). 298 3.1.2 Experiment 2

For Experiment 2 in which ladders with laurel oak and sweetgum branches spanning
heights from 0-30 cm were deployed across six estuaries, the interaction between distance from

301 the sediment and site (F_{5.216}=14.4, p<0.0001), as well as the main effects of these fixed factors 302 (distance from sediment: $F_{1,216}=14.5$, p<0.0002; site: $F_{5,216}=55.2$, p<0.0001) influenced barnacle 303 fouling of branches. While we observed significantly higher barnacle densities on branches 304 positioned 10, 20 and 30 cm above the sediment at Cedar Key, the site where the most barnacles 305 were observed, this pattern was the opposite at Withlacoochee. At the other sites, barnacle 306 densities were relatively low and did not vary much with distance from the sediment (Table 3, 307 Appendix C). Across sites, laurel oak branches were fouled by more barnacles than sweetgum 308 ($F_{1,216}$ =4.6, p<0.05), a pattern consistent with results from Experiment 1.

309 3.1.3 Experiment 3

310 Although barnacle density did not differ between sites for posts treated with copper paint, 311 silicone, or duct tape in experiment 3, pressure-treated fence posts and control posts were fouled by more barnacles at Site 1 (0.75 and 0.17 barnacles cm⁻¹, respectively) than at Site 2 (0.25 and 312 313 0.03 barnacles cm⁻¹, respectively, treatment * site: $F_{4,40}=3.40$, p<0.02). Significantly higher 314 densities of barnacles colonized pressure-treated fence posts than all other treatments with an average of 0.5 barnacles cm⁻¹. In contrast, zero barnacles were observed on silicone-treated posts 315 and only 0.03-0.1 barnacles cm⁻¹ were observed on the copper paint, duct tape, and control post 316 317 treatments (treatment: F_{4,40}=12.97, p<0.0001). Barnacle density was also more than 2-times higher at Site 1 (0.2 barnacles cm⁻¹ on average) compared to Site 2 (0.07 barnacles cm⁻¹ on 318 319 average, site: F_{1,40}=7.36, p<0.01, Table 4).

- 320 **3.2** Shipworm Damage
- 321 3.2.1 Experiment 1

Despite sites 1 and 2 having a number of branches with evidence of shipworm boring (75 of 278 branches with at least one shipworm burrow at Site 1 and 79 of 284 branches at Site 2), damage, measured both in terms of burrow density and wood volume lost, differed between the 325 two sites. Regression tree analyses explained 99.9 and 94.4% of the variation in the percent of 326 wood volume lost (Fig. 4A, tree root node error = 0.015) and burrow density (Fig. 4B, tree root 327 node error = 5.56), respectively, and revealed that site was the strongest driver of both shipworm 328 damage metrics. While on average 2.6% of wood volume loss to burrows was observed at Site 2, 329 only 0.4% wood volume loss was observed at Site 1. Similarly, while 10 burrows were observed 330 per branch on average at Site 2, only 1 burrow per branch was observed at Site 1. Shipworm 331 burrowing also varied significantly with distance from the sediment at both sites and across the 332 four species. While we detected shipworm burrows at all distances, the percent of wood volume 333 lost peaked in branches located between 0-20 cm from the sediment layer. Specifically, 334 regression trees for both shipworm damage metrics (Fig. 4) identified -2.5 cm (i.e. 2.5 cm below 335 the sediment surface) to be the lower limit and 25 cm to be the upper limit of the zone at which 336 most shipworm damage occurs. Within this zone, between 3.7 and 5% of wood volume was lost 337 to shipworm burrows compared to only 0.4% wood volume loss outside this area. Similarly, 338 shipworm burrow densities ranged between 14-19 burrows per branch if the branch was 339 positioned within this zone versus 1-2 burrows per branch if the branch was positioned further 340 into the sediment or higher in the water column.

Tree species was the third most important factor mediating variation in shipworm burrow damage such that sweetgum and crepe myrtle branches experienced >2.5-times higher percent wood volume loss and 3-times higher shipworm burrow density than laurel oak and mangrove branches (Figs. 4 and 5). While branch diameter had no significant effect on the percentage of wood volume lost, it did explain variation in burrow density such that large diameter branches had almost three times more burrows than small diameter branches (Fig. 4B, Table 5).

347 **3.2.2 Experiment 2**

Of 240 branches deployed across six sites in Experiment 2, only one sweetgum branch
deployed at Site 2 was burrowed by shipworms. A total of three burrows were found in this
branch, accounting for approximately a 1% wood volume loss in the branch.

351 3.2.3 Experiment 3

352 Two-way ANOVAs performed to assess variation in the percent of wooden post volume 353 lost to shipworms at each distance from the sediment found treatment and site to have significant 354 effects on this metric of shipworm damage, but only at -10cm (Treatment: F_{4,36}=4.45, p<0.01, 355 Site: F_{1,36}=10.09, p<0.005) and -5cm distances (Treatment: F_{4,36}=4.58, p<0.005, Site: F_{1,36}=7.53, 356 p < 0.01). At -10 and -5cm, shipworm burrowing was significantly higher on control posts (28%) 357 and 32% of post area lost to burrows, respectively) and significantly lower on copper paint and 358 duct tape treated posts (0% wood volume lost at both distances) (Table 4, Appendix D). 359 Similar to the branch ladders (see Experiment 1 results above), wooden posts deployed at 360 Site 2 experienced significantly higher burrowing at -10 and -5 cm from the sediment (17% and 361 19% of area burrowed, respectively) than those deployed at Site 1 at those same distances (1.9%

and 3.7% of area burrowed, respectively). At the 10cm distance, control posts experienced

burrowed). At all other distances, shipworm damage ranged from 0-27% area burrowed but did

significantly higher burrowing (23% of area burrowed) than all other treatments (<15% of area

- 365 not differ between treatments or sites.
- 366 **4. Discussion**

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In this experimental field study, we discovered spatial complementarity in wood
vulnerability to biofouling and bioeroding organisms whereby branches and posts located at
greater distances from the sediment (≥30cm) were more susceptible to biofouling by barnacles,
while those at elevations close (0-20cm) to the sediment surface were more intensively damaged

by shipworms. In addition, we found that trees with lower wood and tannin densities – i.e.
sweetgum and crepe myrtle - were more vulnerable to shipworm burrowing than higher wood
density tree species and that copper-based paint and duct tape offered the greatest protection
against barnacles and shipworms for wood in intertidal environments. Together, these results
indicate that biofouling and bioerosion of wooden marine infrastructure can be reduced through
the strategic use of certain tree species and easy-to-implement treatments that interfere with the
settlement and growth of these biota.

378 Similar barnacle biofouling results were found in Experiments 1 and 2, with the number 379 of barnacles per branch increasing with increasing height (Fig. 3). This pattern of enhanced 380 barnacle colonization of higher elevation surfaces is consistent with that reported in the literature 381 for intertidal barnacles (e.g. Grosberg 1982, Wethey 1983, Raimondi 1988) and is thought to be 382 driven both by larval behavior (e.g. barnacle larvae are buoyant, responsive to light, and favor 383 low-pressure environments found near the water surface) and by enhanced vulnerability of 384 barnacles, once settled, to predation at lower intertidal elevations (Connell 1970). In Experiment 385 2, we also found barnacle abundance to be relatively high in Cedar Key but did not differ 386 between the other five sites, variability that we suspect was driven by higher barnacle larval 387 delivery to this more open-water rather than estuarine site (Minchinton & Scheibling 1991), 388 although variation in predation pressure cannot be ruled out as a contributing factor to the 389 observed variation.

When considering potential treatments against barnacle biofouling in Experiment 3, we found silicone to be the most effective (i.e. no barnacles settled on silicone-treated posts), and pressure-treated wood to be the least effective. Savoya & Schwindt (2010) quantified barnacle settlement on substrates of varying texture in supporting and found rough-textured substrates to

394 be most suitable for barnacle growth. These results might explain why no barnacles were 395 observed on silicone: its surface texture is very smooth. In contrast, the relative ineffectiveness 396 of pressure-treated wood in warding off barnacles could be due to both the rougher surface of 397 these posts promoting colonization as well the amount of time these posts were in the water in 398 our experiment (18 months). Weis & Weis (1996) and Edwin & Sreeja (2011) found that when 399 pressure-treated wood is submerged, it leaches copper, chromium, and arsenic (CCA) into the 400 water such that, after two months, these chemicals are no longer in high enough concentrations 401 to deter biofoulers. Thus, it is possible that, although the pressure-treated posts initially 402 supported little to no barnacle growth, their anti-fouling capacity was diminished by the end of 403 the experiment. Viewed in the context of wooden marine infrastructure, these results suggest that 404 barnacles are likely to induce the largest drag and ecological effects (e.g. forming a physical 405 barrier to shipworm larval settlement (Singh and Sasekumar 1996)) on the rough-surfaced, upper 406 sections (\geq 30 cm) of wooden break walls and posts that do not emit strong chemical deterrents in 407 coastal environments.

408 Shipworms and other marine borers have been attacking wood for centuries (Nicholas 409 1982) and, in response, many techniques (e.g. fish-, coconut-, cashew-oils as coatings or 410 application of sand, cement, black tar, and copper chromate arsenate) to deter shipworms have 411 been employed (Nagabhushanam 1997). The average and maximum percent wood volume lost to 412 shipworms recorded in our study was < 7% and 55%, respectively, over six months (Fig. 4). 413 Although generally low, this level of damage can be enough to compromise the structural 414 integrity of wooden structures, especially those exposed to high and/or frequent wave and wake 415 loading (Charles et al. 2016). Most importantly, the shipworm infestation patterns observed 416 across site, distances from the sediment, and tree species give an indication of when and how

often wooden structures, like breakwalls and bulkheads, will need maintenance. In particular, the
bottom of walls and walls built from less dense sweetgum and crepe myrtle branches – species
that lost the most wood volume to shipworms (Fig 5) - are likely to need more maintenance,
especially at sites experiencing high shipworm recruitment. Our results correspond to previous
studies that also found tree species to vary in resistance to borers depending on certain traits; for
example, resistance to marine borers has been shown to increase with wood silica and alkaloid
contents (Nicholas 1982, Roszaini & Salmiah 2015).

424 According to regression tree analyses, site was the most important factor mediating 425 shipworm burrow density and amount of wood volume lost (Fig. 4). One might expect that 426 ladders at lower elevations, given their longer inundation time, to be exposed to shipworm-427 infested waters for a longer and thus suffer higher shipworm damage. This rationale cannot explain our results in Experiment 1, however, since Site 1 ladders, positioned at an average 428 429 elevation of -0.65 m above sea level, experienced less damage than those at Site 2 which were 430 positioned higher in elevation at at -0.59 m above sea level. Additionally, one might also expect 431 proximity to a saltwater source to correspond to shipworm damage given shipworms' preference 432 for higher salinities (Barrows 1917). However, we found average salinities during the study 433 period to be nearly the same at the two sites according to nearby water quality monitoring 434 stations (34.8 vs. 34.1 ppt at Sites 1 and 2, respectively) despite variation in site proximity to 435 tidal inlets to the open ocean (Site 1 and 2 are 8.3 and 6.5 km, respectfully, away from the closest 436 tidal inlet to the Atlantic Ocean). Given that Site 1 consisted of a narrow, meandering creek 437 surrounded by denser vegetation, while Site 2 consisted of a wider creek that was closer and had 438 a wide, unvegetated connection to the main channel, it is possible that local geomorphology 439 drove the significant variation in shipworm damage between the sites. Specifically, it is likely

that a greater water volume was exchanged per tidal cycle at Site 2 relative to Site 1, resulting in
higher delivery of shipworm larvae, based on Leonard and Reed's (2002) finding that creek
vegetation reduced water flow speed and on Roegner's (2000) calculations that narrow creeks
transport a lower volume of water than wider creeks.

444 The second most important indicator of shipworm boring was distance from the sediment 445 surface. Results from Experiment 1 show shipworm damage to be concentrated in the top 20 cm 446 above the sediment layer, consistent with findings from Tuente et al. (2002) who also found that 447 shipworm burrow densities on wooden piles in German harbors increase with decreasing height 448 above the sea floor. Scheltema and Truitt (1956) found similar results in Maryland's coastal 449 waters, with higher shipworm densities on wooden panels positioned closer to the sediment 450 surface over a range of depths from 0-2.1 m. Finally, Paalvast and van der Velde (2011a) 451 similarly report a negative correlation between shipworm burrowing and distance from the sea 452 floor at depths of 0-1 m. This general pattern of high shipworm colonization of wood close to the 453 sediment surface likely arises because shipworms cannot access and survive within branches 454 found deep in the substrate (-5 and -10 cm) due to anoxic conditions and because branches at the 455 upper limits (≥30cm) are inundated, and thus exposed to shipworm larvae, for less time. Given 456 these dynamics, shipworm activity would be expected to be more prevalent closer to the 457 sediment and would explain patterns seen in this and previous experiments.

Finally, tree species identity also influenced the extent of shipworm bioerosion (Fig 5). These differences may be due to tree species' differing hardness, with shipworm burrows being more prevalent in softer branches that require less energy investment to burrow into than in harder branches (Paalvast and van der Velde's 2011a). One measure of tree hardness is wood density, which is typically calculated by the ratio of dry weight of wood divided by its green 463 volume (Zobel and Jett 1995). In our first experiment, we found significantly more shipworm 464 damage on sweetgum and crepe myrtle branches, which have wood densities of 0.42 and 0.55 g cm⁻³, respectively (Holbrook and Putz 1989, Reyes et al. 1992). In contrast, black mangrove and 465 466 oak species -those that experienced less shipworm damage - have higher wood densities of 0.87 467 and 0.70 g cm⁻³, respectively (Reyes et al. 1992, Saenger 2002). These shipworm burrowing 468 patterns are consistent with Paalvast and van der Velde's (2011a) who also found higher 469 shipworm damage in softer fir than in harder oak panels. In addition, oak and mangrove trees 470 produce tannins, a compound known to limit protein availability to organisms consuming their 471 bark and leaves (Hathway 1958, Robbins et al 1987, Kimura and Wada 1989), potentially 472 limiting digestibility of these wood types in shipworms. These results suggest that the tree 473 species used can be a significant driver in the long-term vulnerability of wooden structures in 474 coastal environments.

475 From the regional study carried out during the second year of this experiment we can see 476 that there can be interannual variability in shipworm activity. One possibility is that patterns seen 477 from one year to the next are a result of the time the experimental branches was deployed. The 478 ladders were in the field for six months (July-January) in experiment 1 and three months (June-479 September) due to logistical constraints in experiment 2. In their field experiment conducted in 480 Port of Rotterdam, Netherlands, Paalvast and van der Velde (2011a) report that although 481 shipworm larvae were present in the water from April-November, they did not observe 482 infestations in their wooden panels before September. This suggests that shipworm larvae might 483 have been present but the wood was not in the water long enough for larvae to grow, develop, 484 and cause significant, visible damage. These findings also importantly suggest that the timing of 485 when wood gets placed in the water matters. Structures, such as wooden breakwalls, might have

486 a longer life span if they are strategically built around the period of minimum reproductive487 activity to minimize their larval exposure and thus reduce shipworm boring.

488 However, as stated previously, shipworms prefer high-salinity environments. This may 489 have also influenced our regional study results given that 2016 experienced higher drought levels 490 and coincident salinity levels in our study sites than 2017. During the 2016 study period, the 491 Palmer Drought Severity Index ranged from -2.73 to -2.06, while during 2017 it ranged from -492 0.76 to 2.48 (Appendix E1) in this region. Salinities between the two years differed mainly in the 493 minimum values reached. Sites 1 and 2 experienced minimum salinities of 28.6 and 23.2 ppt in 494 2016, respectively, but these values dropped to 16.2 and 9.7 ppt in 2017 (Appendix E2). It is 495 important then to consider how climate change and other anthropogenic drivers interact to affect 496 the severity and duration of drought, thus creating conditions for persistent shipworm activity.

497

5.

Conclusions: Enhancing Wooden Structure Longevity in Coastal Environments

498 Barnacle biofouling and shipworm boring preferences seen here can be used to inform 499 the construction of wooden structures in coastal environments, such as wooden breakwalls used 500 as living shorelines techniques. Particularly, understanding how shipworm burrowing varies 501 across different tree species can help identify the optimal building materials that will prolong the 502 life span of these structures. For instance, when selecting filling for wooden breakwalls, 503 choosing tree species with higher wood densities and high tannin content may result in structures 504 that are more resistant to shipworm damage and that may not require as much maintenance 505 compared to lower wood density species. In addition, knowing where shipworm burrowing and 506 biofouling are concentrated along the water column can help predict which will be the most 507 vulnerable areas of wooden structures. With this knowledge, priority can be given to the 508 bottommost 20 cm of wooden structures when maintenance to combat bioerosion is required. 509 Furthermore, being aware of the spatial variability in shipworm boring and biofouling, and of

510 characteristics such as larval delivery to a site, can help determine how long wooden structures 511 will last in a particular area. Knowing if a site has high or low larval delivery can indicate the 512 extent of shipworm damage that can be expected and thus inform the timing of potential 513 maintenance. Also, being aware of the environmental conditions (e.g. drought, decreased river 514 discharge, warmer temperatures) that can create a hospitable environment for shipworms can 515 help coordinate the timing of the deployment of new structures in order to maximize their life 516 span or can dictate maintenance efforts for wooden structures already installed in coastal areas. 517 Finally, treating wooden marine infrastructure at strategic locations (e.g. parts of the structure 518 found near the sediment surface or underground) with easy-to-use materials with minimal 519 environmental risk (e.g. inexpensive adhesive wraps or concrete slurries that prevent biofouling 520 organisms from adhering or burrowing into the surfaces of posts or pilings) can protect areas key 521 to maintaining structural stability. Ultimately, shipworms will be a persistent threat to wood in 522 coastal environments, but a better understanding of the conditions under which shipworm boring 523 occurs can promote a smarter approach to prolonging the life span of wooden structures. 524 Knowing the areas that are vulnerable to shipworm damage and addressing these vulnerabilities 525 through innovative techniques such as combinations of natural and manmade materials can help

526 build more resistant and longer-lasting structures.

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533 References

- Alberte, R.L.S., Zahuranec, B.J., Whetstone, M., Snyder, S., 1992. Biofouling research needs for
 the United States Savy: Program history and goals. Biofouling 6, 91–95.
- 536 https://doi.org/10.1080/08927019209386214
- Bakker, W.T., Hulsbergen, C.H., Roelse, P., de Smit, C., Svasek, J.N., 1984. Permeable
 Groynes: Experiments and Practice in the Netherlands, in: Coastal Engineering. American
 Society of Civil Engineers, Houston, TX, pp. 2026–2041.
- 540 https://doi.org/https://doi.org/10.1061/9780872624382.137
- Barrows, A.L., 1917. An unusual extension of the distribution of the shipworm in San Francisco
 Bay, California. Univ. Calif. Publ. Zool. 18, 27–43.
- Borges, L.M.S., Cragg, S.M., Williams, J.R., 2003. Comparing the Resistance of a Number of
 Lesser Known Species of Tropical Hardwoods to the Marine Borer Limnoria Using a Short
- 545 Term Laboratory Assay. Stockholm.
- Borges, L.M.S., 2014. Biodegradation of wood exposed in the marine environment: Evaluation
 of the hazard posed by marine wood-borers in fifteen European sites. Int. Biodeterior.
 Biodegrad. 96, 97–104. https://doi.org/10.1016/j.ibiod.2014.10.003
- 549 Borsje, B.W., van Wesenbeeck, B.K., Dekker, F., Paalvast, P., Bouma, T.J., van Katwijk, M.M.,
 550 de Vries, M.B., 2011. How ecological engineering can serve in coastal protection. Ecol.
 551 Eng. 37, 113–122. https://doi.org/https://doi.org/10.1016/j.ecoleng.2010.11.027
- Bulleri, F., Chapman, M.G., 2010. The introduction of coastal infrastructure as a driver of
 change in marine environments. J. Appl. Ecol. 47, 26–35. https://doi.org/10.1111/j.13652664.2009.01751.x
- 555 Callow, M.E., Callow, J.A., 2002. Marine biofouling: A sticky problem. Biologist 49, 10–14.
- Charles, F., Coston-Guarini, J., Guarini, J.-M., Fanfard, S., 2016. Wood decay at sea. J. Sea Res.
 114, 22–25. https://doi.org/https://doi.org/10.1016/j.seares.2016.05.002
- Connell, J.H., 1970. A Predator-Prey System in the Marine Intertidal Region. I. Balanus glandula
 and Several Predatory Species of Thais. Ecol. Monogr. 40, 49–78.
 https://doi.org/10.2307/1942441
- 561 Edyvean, R.G.J., Thomas, C.J., Brook, R., 1988. The Effect of Marine Fouling on Fatigue and
- 562 Corrosion-Fatigue of Offshore Structures, in: Houghton, D.R., Smith, R.N., Eggins, H.O.W.
- 563 (Eds.), Biodeterioration 7. Springer Netherlands, Dordrecht, pp. 385–390.
- 564 https://doi.org/10.1007/978-94-009-1363-9_50

- Gittman, R.K., Fodrie, F.J., Popowich, A.M., Keller, D.A., Bruno, J.F., Carolyn, A., Peterson,
 C.H., Piehler, M.F., 2015. Engineering away our natural defenses: an analysis of shoreline
 hardening in the US. Front. Ecol. Environ. 13, 301–307. https://doi.org/10.1890/150065
- Grave, B.H., 1928. Natural history of shipworm, Teredo navalis, at Woods Hole,
 Massachussetts. Biol. Bull. 55, 260–282.
- 570 Grave, B.H., 1942. The Sexual Cycle of the Shipworm, Teredo navalis. Biol. Bull. 82, 438–445.
- Grosberg, R.K., 1982. Intertidal Zonation of Barnacles: The Influence of Planktonic Zonation of
 Larvae on Vertical Distribution of Adults. Ecology 63, 894–899.
 https://doi.org/https://doi.org/10.2307/1937228
- 574 Hathway, D.E., 1958. Oak-bark Tannins. Biochem. J. 70, 34–42.
- 575 Herbert, D., Astrom, E., Bersoza, A.C., Batzer, A., McGovern, P., Angelini, C., Wasman, S.,
- 576 Dix, N., Sheremet, A., 2018. Mitigating erosional effects induced by boat wakes with living 577 shorelines. Sustain. 10, 1–19. https://doi.org/10.3390/su10020436
- Holbrook, M., Putz, F.E., 1989. Influence of Neighbors on Tree Form : Effects of Lateral Shade
 and Prevention of Sway on the Allometry of Liquidambar styraciflua (Sweet Gum) Author
 (s): N. Michele Holbrook and Francis E. Putz Source : American Journal of Botany , Vol
 . 76, No. 1 76, 1740–1749.
- Hoppe, K.N., 2002. Teredo Navalis --- the Cryptogenic Shipworm, in: Leppäkoski, E., Gollasch,
 S., Olenin, S. (Eds.), Invasive Aquatic Species of Europe. Distribution, Impacts and
 Management. Springer Netherlands, Dordrecht, pp. 116–119. https://doi.org/10.1007/97894-015-9956-6_12
- 586 Kimura, M., Wada, H., 1989. Tannins in mangrove tree roots and their role in the root
 587 environment. Soil Sci. Plant Nutr. 35, 101–108.
 588 https://doi.org/10.1080/00380768.1989.10434741
- Leonard, L.A., Reed, D.J., 2002. Hydrodynamics and sediment transport through tidal marsh
 canopies. J. Coast. Res. 36, 459–469. https://doi.org/10.2112/1551-5036-36.sp1.459
- Lindholdt, A., Dam-Johansen, K., Olsen, S.M., Yebra, D.M., Kiil, S., 2015. Effects of biofouling
 development on drag forces of hull coatings for ocean-going ships: a review. J. Coatings
 Technol. Res. 12, 415–444. https://doi.org/10.1007/s11998-014-9651-2
- Lingvay, I., Fortuna, L., Varga, E., Bors, A.-M., Nicula Butoi, N.O., Lingvay, D., 2018.
 Durability and anticorrosive protection capability of paint layers Biological factors influence. Electroteh. Electron. Autom. 66, 52–58.

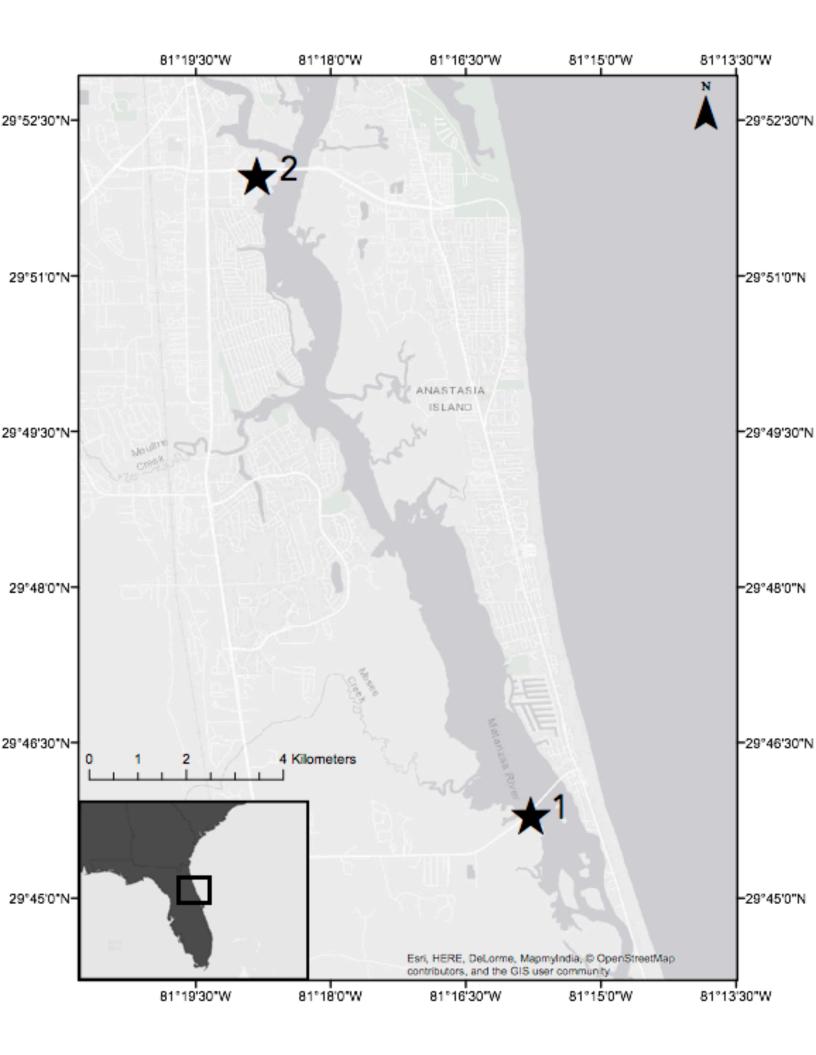
- Lippert, H., Weigelt, R., Glaser, K., Krauss, R., Bastrop, R., Karsten, U., 2017. Teredo navalis in
 the Baltic Sea: Larval Dynamics of an Invasive Wood-Boring Bivalve at the Edge of Its
 Distribution. Front. Mar. Sci. 4, 1–12. https://doi.org/10.3389/fmars.2017.00331
- Lopez-Anido, R., Michael, A.P., Goodell, B., Sandford, T.C., 2004. Assessment of Wood Pile
 Deterioration due to Marine Organisms. J. Waterw. Port, Coastal, Ocean Eng. 130, 70–76.
- Minchinton, T.E., Scheibling, R.E., 1991. The Influence of Larval Supply and Settlement on the
 Population Structure of Barnacles. Ecology 72, 1867–1879.
 https://doi.org/10.2307/1940984
- Morton, R.A., 2003. An Overview of Coastal Land Loss: With Emphasis on the Southeastern
 United States. St. Petersburg, FL.
- 607 Nagabhushanam, R., 1997. Fouling Organisms of the Indian Ocean. Taylor & Francis.
- Nelson, D.L., 2015. The Ravages of Teredo: The Rise and Fall of Shipworm in US History,
 1860-1940. Environ. Hist. Durh. N. C. 21, 100–124. https://doi.org/10.1093/envhis/emv118
- 610 Nicholas, D.D., 1982. Wood Deterioration and Its Prevention by Preservative Treatments:
 611 Degradation and protection of wood, Degradation & Protection of Wo. Syracuse University
 612 Press.
- Ohgami, N., Yamanoshita, O., Thang, N.D., Yajima, I., Nakano, C., Wenting, W., Ohnuma, S.,
 Kato, M., 2015. Carcinogenic risk of chromium, copper and arsenic in CCA-treated wood.
 Environ. Pollut. 206, 456–460. https://doi.org/https://doi.org/10.1016/j.envpol.2015.07.041
- Paalvast, P., van der Velde, G., 2011a. Distribution, settlement, and growth of first-year
 individuals of the shipworm Teredo navalis L. (Bivalvia: Teredinidae) in the Port of
 Rotterdam area, the Netherlands. Int. Biodeterior. Biodegrad. 65, 1822–1829.
 https://doi.org/10.1016/j.marpolbul.2011.05.009
- Paalvast, P., van der Velde, G., 2011b. New threats of an old enemy: The distribution of the
 shipworm Teredo navalis L. (Bivalvia: Teredinidae) related to climate change in the Port of
 Rotterdam area, the Netherlands. Mar. Pollut. Bull. 62, 1822–1829.
 https://doi.org/10.1016/j.marpolbul.2011.05.009
- Pimentel, D., Lach, L., Zuniga, R., Morrison, 2000. Environmental and Economic Costs of
 Nonindigenous Species in the United States. Bioscience 50, 53–65.
- Poff, M.T., Stephen, M.F., Dean, R.G., Mulcahy, S., Beach, P., Pofff, T., Stephen, F., Hall, W.,
 Dean, G., Mulcahyt, S., 2004. Permeable Wood Groins : Case Study on their Impact on the
 Coastal System Coastal Groins : The Interaction of Groins and the Beach Process and
 Planning.

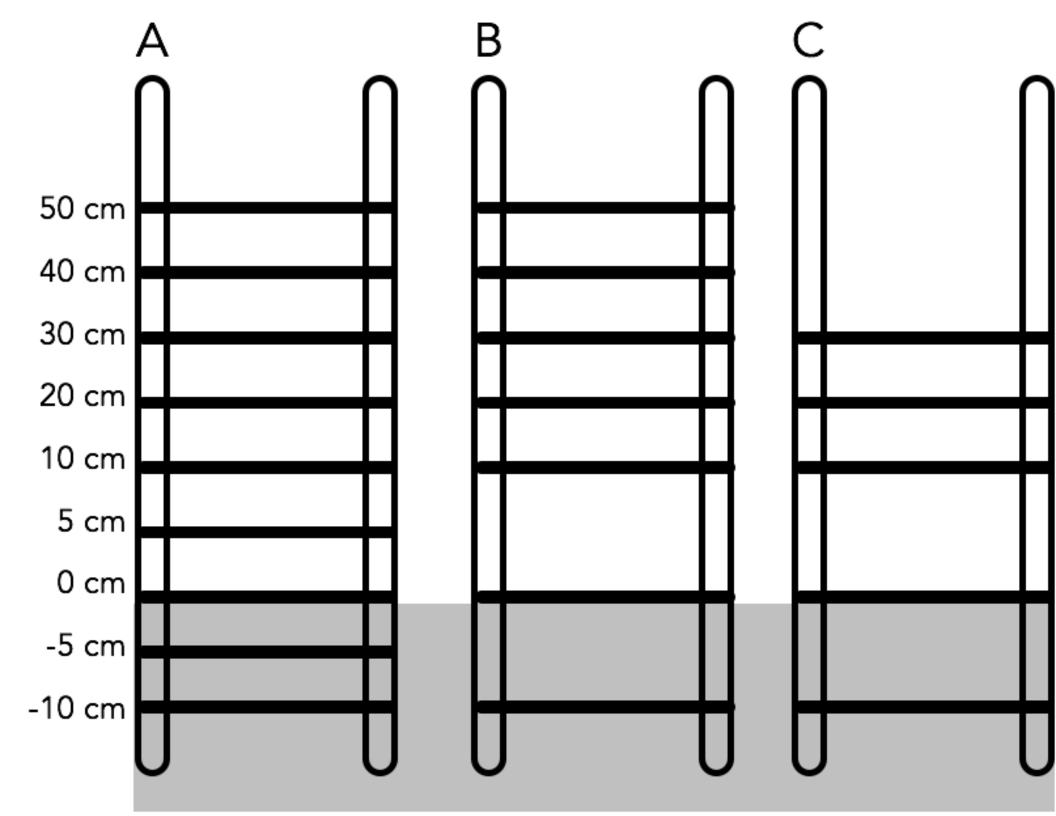
- Qiu, J.-W., Qian, P.-Y., 1999. Tolerance of the barnacle Balanus amphitrite amphitrite to salinity
 and temperature stress: effects of previous experience. Mar. Ecol. Prog. Ser. 188, 123–132.
- Raimondi, P.T., 1988. Settlement Cues and Determination of the Vertical Limit of an Intertidal
 Barnacle. Ecology 69, 400–407. https://doi.org/10.2307/1940438
- Reyes, G., Brown, S., Chapman, J., Lugo, A.E., 1992. Wood densities of Tropical tree species,
 Technical report.
- Rice, S.A., Johnson, B.R., Estevez, E.D., 1990. Wood-boring marine and estuarine animals in
 Florida.
- Richmond, M.D., Seed, R., 1991. A review of marine macrofouling communities with special
 reference to animal fouling. Biofouling 3, 151–168.
 https://doi.org/10.1080/08927019109378169
- Robbins, C.T., Hanley, T.A., Hagerman, A.E., Hjeljord, O., Baker, D.L., Schwartz, C.C., Mautz,
 W.W., 1987. Role of Tannins in Defending Plants Against Ruminants: Reduction in Protein
 Availability. Ecology 68, 98–107.
- Roegner, G.C., 2000. Transport of molluscan larvae through a shallow estuary. J. Plankton Res.
 22, 1779–1800. https://doi.org/10.1093/Plankt/22.9.1779
- Roszaini, K., Salmiah, U., 2015. Resistance of five timber species to marine borer attack. J.
 Trop. For. Sci. 27, 400–412.
- 648 Saenger, P., 2002. Mangrove Ecology, Silviculture and Conservation. Springer Science &
 649 Business Media.
- Savoya, V., Schwindt, E., 2010. Effect of the substratum in the recruitment and survival of the
 introduced barnacle Balanus glandula (Darwin 1854) in Patagonia, Argentina. J. Exp. Mar.
 Bio. Ecol. 382, 125–130. https://doi.org/10.1016/j.jembe.2009.10.012
- Scheltema, R.S., 1971. Dispersal of phytoplanktotrophic shipworm larvae (Bivalvia:
 Teredinidae) over long distances by ocean currents. Mar. Biol. 11, 5–11.
 https://doi.org/10.1007/BF00348015
- 656 Scheltema, R.S., Truitt, R.V., 1956. The Shipworm Teredo Navalis in Maryland Coastal Waters.
 657 Ecology 37, 841–843.
- Schultz, M.P., Bendick, J.A., Holm, E.R., Hertel, W.M., 2011. Economic impact of biofouling
 on a naval surface ship. Biofouling 27, 87–98.
 https://doi.org/10.1080/08927014.2010.542809
- 661 Singh, H.R., Sasekumar, A., 1996. Wooden Panel Deterioration by Tropical Marine Wood
 662 Borers 755–769.

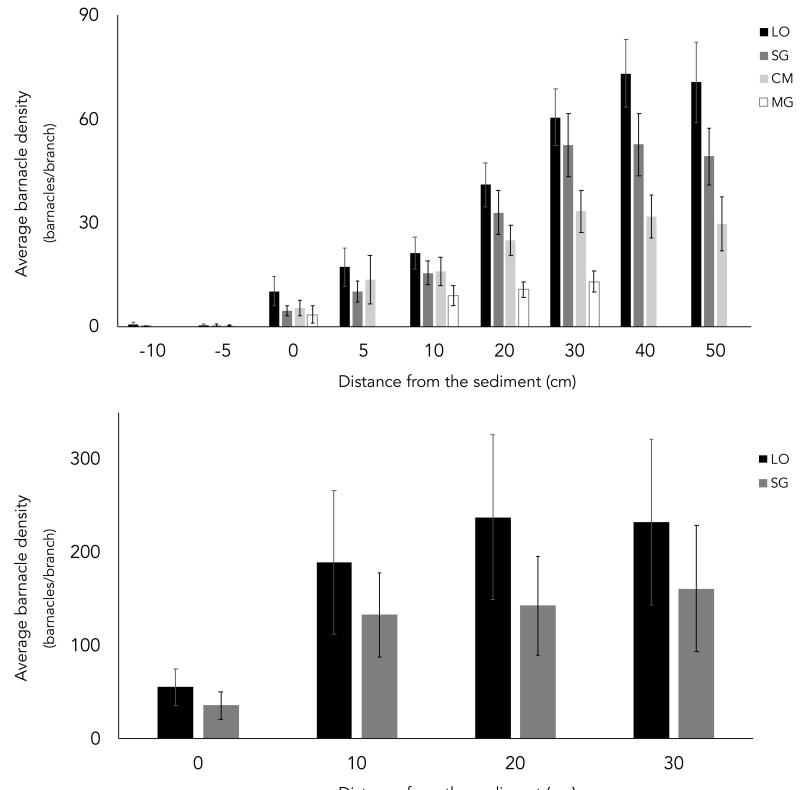
663	Sriyutha Murthy, P., Venugopalan, V.P., Nair, K.V.K., Subramoniam, T., 2009. Larval
664	Settlement and Surfaces: Implications in Development of Antifouling Strategies, in:
665	Costerton, J.W. (Ed.), Marine and Industrial Biofouling. Springer, pp. 233–264.
666	https://doi.org/10.1007/b136878
667	Stevenson, C.F., Demes, K.W., Salomon, A.K., 2016. Accounting for size-specific predation
668	improves our ability to predict the strength of a trophic cascade. Ecol. Evol. 6, 1041–1053.
669	https://doi.org/10.1002/ece3.1870
670 671 672	Strathmann, R.R., Branscomb, E.S., Vedder, K., 1981. Fatal errors in set as a cost of dispersal and the influence of intertidal flora on set of barnacles. Oecologia 48, 13–18. https://doi.org/10.1007/BF00346982
673	Sundberg, A., 2015. Molluscan Explosion: The Dutch Shipworm Epidemic of the 1730s.
674	Environ. Soc. Portal 14. https://doi.org/doi.org/10.5282/rcc/7307
675	Therneau, T., Atkinson, B., Ripley, B., 2018. An Introduction to Recursive Partitioning Using
676	the RPART Routines [WWW Document]. R Packag. version 4.1-10. URL https://cran.r-
677	project.org/web/packages/rpart/vignettes/longintro.pdf
678	Trager, G.C., Hwang, J.S., Strickler, J.R., 1990. Barnacle suspension-feeding in variable flow.
679	Mar. Biol. 105, 117–127. https://doi.org/10.1007/BF01344277
680	Tuente, U., Piepenburg, D., Spindler, M., 2002. Occurrence and settlement of the common
681	shipworm Teredo navalis (Bivalvia: Teredinidae) in Bremerhaven harbours, northern
682	Germany. Helgol. Mar. Res. 56, 87–94. https://doi.org/10.1007/s10152-002-0101-7
683	Weis, J.S., Weis, P., 1992. Construction materials in estuaries: reduction in the epibiotic
684	community on chromated copper arsenate (CCA) treated wood. Mar. Ecol. Prog. Ser. 83,
685	45–53.
686 687 688	Weis, J.S., Weis, P., 1996. The effects of using wood treated with chromated copper arsenate in shallow-water environments: A review. Estuaries 19, 306–310. https://doi.org/10.2307/1352236
689 690 691	Wethey, D.S., 1983. Geographic Limits and local zonation: the barnacles Semibalanus (Balanus) and Chthamalus in New England. Biol. Bull. 165, 330–341. https://doi.org/10.2307/1541373
692	Yang, W.J., Neoh, K.G., Kang, E.T., Teo, S.L.M., Rittschof, D., 2014. Polymer brush coatings
693	for combating marine biofouling. Prog. Polym. Sci. 39, 1017–1042.
694	https://doi.org/10.1016/j.progpolymsci.2014.02.002
695	Zobel, B.J., Jett, J.B., 1995. The Importance of Wood Density (Specific Gravity) and Its

Component Parts BT - Genetics of Wood Production, in: Zobel, B.J., Jett, J.B. (Eds.), . 696

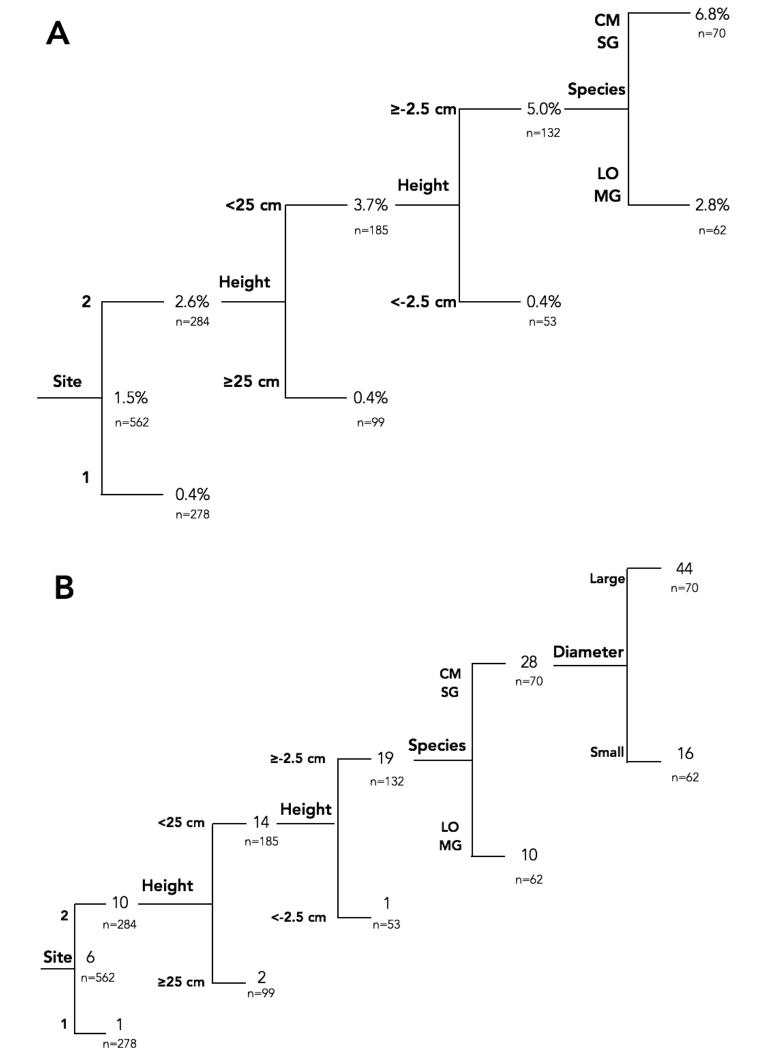
697Springer Berlin Heidelberg, Berlin, Heidelberg, pp. 78–97. https://doi.org/10.1007/978-3-698642-79514-5_4

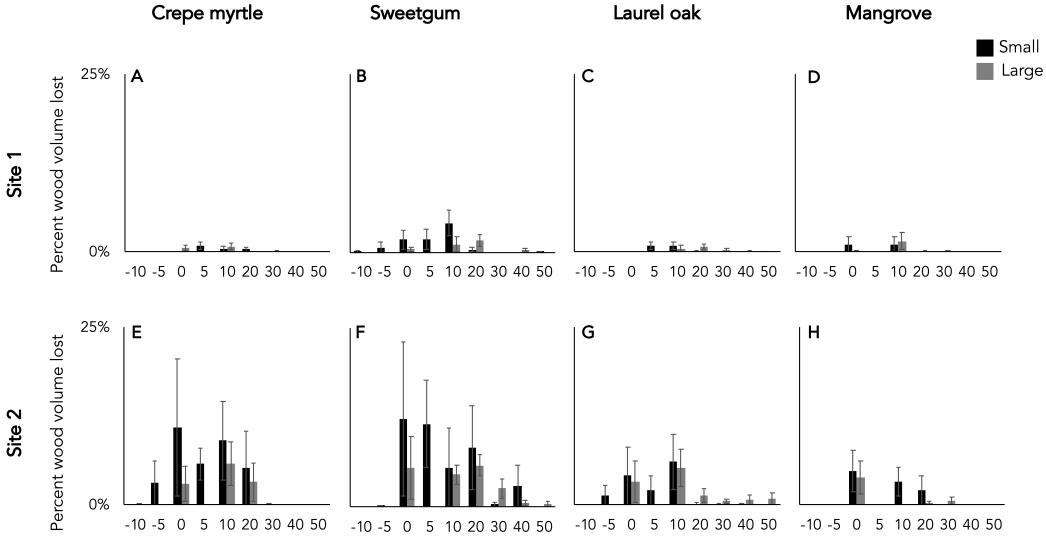






Distance from the sediment (cm)





Distance from the sediment (cm)

Site 2

Experiment	Experimental factors	Tree Species	Branch diameter	Distances from the sediment	Sites	Duration
1	Site, distance from sediment, tree species identity, and branch diameter	Crepe myrtle (<i>Lagerstroemia</i> <i>spp.</i>), Laurel oak (<i>Quercus</i> <i>hemisphaerica</i>), Sweetgum (<i>Liquidambar</i> <i>styraciflua</i>), Mangrove (<i>Avicennia</i> <i>germinans</i>)	Small (1-2.5 cm) and Large (2.5-5 cm)	-10, -5, 0, 5, 10, 20, 30, 40, 50 cm	Saint Augustine Sites 1 and 2	6 months (July 2016 - Jan. 2017)
2	Site, distance from sediment, and tree species	Laurel oak (Quercus hemisphaerica), Sweetgum (Liquidambar styraciflua)	Small (1-2.5 cm)	0, 10, 20, and 30 cm	Saint Augustine Sites 1 and 2, Cedar Key, Withlacoochee Bay, Suwannee River, and Sapelo Island	3 months (June 2017 - Sep. 2017)
3	Chemical and non-chemical wood treatment methods	Treated (pressure- treated, tape, silicon, copper paint) and untreated wooden posts	Pressure treated wood post dimensions: 8.9 cm diameter x 1.5 m long. All other post dimensions: 1.5 m x 5.08 cm x 5.08 cm	-10, -5, 0, 5, 10, and 20 cm	Saint Augustine Sites 1 and 2	6 months (July 2016 - Jan. 2018)

Table 1. Objective, study species, branch diameters, distance from sediment, treatments, site, and duration of experiments 1, 2, and 3.

		Barnacle Density (# per m ²)		
Variable	df	F value	p value	
Distance from sediment	1, 546	348.2	< 0.0001	
Tree species	3, 546	20.7	< 0.0001	
Site	1, 546	23.7	< 0.0001	
Distance from sediment*tree species	3, 546	15.4	< 0.0001	
Distance from sediment*site	1, 546	16.7	< 0.0001	

Table 2. Summary of ANOVA results for barnacle biofouling in Experiment 1.

		Barnacle Density (# per m ²)		
Variable	df	F value	p value	
Distance from sediment	1,216	14.5	< 0.0002	
Tree species	1,216	4.6	< 0.05	
Site	5,216	55.2	< 0.0001	
Distance from sediment*site	5,216	14.4	< 0.0001	

Table 3. Summary of ANOVA results for barnacle biofouling in Experiment 2.

		Barnacle Density (# per cm ²)		Shipworm burrow percent cover at -10 cm		Shipworm burrow percent cover at -5 cm	
Variable	df	F value	p value	F value	p value	F value	p value
Treatment	4,40	12.97	< 0.0001	4.45	< 0.01	4.58	< 0.005
Site	1,40	7.36	< 0.01	10.09	< 0.005	7.53	< 0.01
Treatment*Site	4,40	3.40	< 0.02	2.1	< 0.1	1.6	< 0.5

Table 4. Summary of ANOVA results for barnacle and shipworm biofouling in Experiment 3.

		% Wood Volume Lost	Burrows per branch
Site	Site 1	0.4% ± 0.1%	1 ± 0.2
Site	Site 2	$2.6\% \pm 0.4\%$	10 ± 1.6
	> 0 cm	$0.4\% \pm 0.3\%$	1 ± 0.9
Distance from	0-20 cm	5% ± 0.8%	19 ± 3.2
sediment	< 25 cm	$0.5\% \pm 0.2\%$	2 ± 1.0
	Crepe myrtle	6.1% ± 1.7%	28 ± 9.4
Tree	Sweetgum	7.6% ± 2.0%	27 ± 6.0
Species	Laurel oak	3.1% ± 1%	12 ± 4.0
	Mangrove	$2.4\% \pm 0.8\%$	6 ± 2.0
Branch	Small	NSD	16 ± 4.5
diameter	Large	NSD	44 ± 11.0

Table 5. Summary of statistical results for shipworm biofouling in Experiment 1.