

1 **Ontogenetic spatial constraints of sub-arctic marine fish species**

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27

28 **Abstract**

29 Marine species may respond and adapt to climate change through shifting spatial distributions,  
30 but options may be limited by the occupancy of essential habitats which are anchored in space.  
31 Limited knowledge of when spatial constraints are most likely to occur in marine fish life cycles  
32 has impeded the development of realistic distribution forecasts. In this study, we develop and  
33 implement analytical techniques to identify spatial constraints, defined by both the *consistency*  
34 through which a particular geographic area is used year after year, and by the *extent* of such area  
35 with respect to the entire population range. This approach is applied to simulated data and to ten  
36 case studies including six groundfish species from three subarctic marine systems. Our analyses  
37 illustrate that the early phase of the species' life cycle is more spatially constrained than older life  
38 stages. We detected significant species-specific variability in both the degree to which species are  
39 anchored in space throughout their life cycle, and the ontogenetic changes of the geographic  
40 association. There is an indication that this variability can be explained by the species life history  
41 strategy, highlighting the need to extend similar analyses to other species and regions. The  
42 presence of ontogenetic spatial constraints, particularly during early life stages, indicates  
43 restrictions exist to changes of spatial distribution and questions the assertion that global  
44 warming will uniformly result in an increase in abundance and harvest at higher latitudes and  
45 decreases at lower latitudes. Our study develops ecological and analytical insights that are critical  
46 for accurate projections of species distributions under different climate change scenarios.

47 Key words: Adaptation, climate change, marine subarctic, spatial distribution, spatial ecology,  
48 spawning

49

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## 61 **Introduction**

62 A focal point in the study of fisheries oceanography is to understand and predict how fish  
63 populations respond to climate change, and to regional and local scale oceanographic variability  
64 (Bograd et al., 2014; Cury et al., 2008; Kendall & Duker, 1998) Within this framework, one of  
65 the remaining great challenges is to quantitatively characterize the adaptive potential of marine  
66 organisms (Merilä & Hendry, 2014). Climate change is affecting ecological processes and  
67 biogeochemical cycles of many marine environments. Species may respond and adapt to these  
68 changes through both phenotypic plasticity and microevolutionary adaptations (Crozier &  
69 Hutchings, 2014; Donelson et al., 2019). For example, one way in which adult marine fish  
70 quickly adapt to climate change is by shifting their spatial distribution (Poloczanska et al., 2013).  
71 Commercial exploitation has also shown to result in spatial shifts, in particular a deepening  
72 among older age groups (Frank et al., 2018). Retrospective analyses of long-term monitoring  
73 programs have documented several shifts of marine fish spatial distribution across both  
74 latitudinal and bathymetric gradients (Nye et al., 2009; Perry, 2005; Pinsky et al., 2013, 2020).  
75 These trends are particularly evident in sub-arctic and temperate environments (Kleisner et al.,  
76 2017), where the effects of global warming appear to be the primary driver (Saba et al., 2016).  
77 Range shifts of marine fish are now impacting coastal communities that are dependent on  
78 commercial fisheries (Rogers et al., 2019; Selden et al., 2020).

79       There are limits to how far individuals in a population can adapt to a changing climate.  
80 Marine fish species have complex life cycles composed of multiple life history stages. Each life  
81 stage frequently has unique spatial distributions and habitat requirements (Barbeaux & Hollowed,  
82 2018; Bartolino et al., 2011). During some life stages individuals are spatially constrained due to  
83 the presence of critical habitats and oceanographic conditions, such as bathymetry, sediment  
84 characteristics, or topographically induced currents and tides, which are also fixed in space and

85 time (Ciannelli et al., 2010; Lough et al., 2006; Stuart-Smith et al., 2021). Imprinted behavior  
86 (Cury, 2019) and social facilitations (Corten, 2002) also contribute to the presence of persistent  
87 spatial aggregations of individuals in geographically fixed habitats. Examples of spatially  
88 constrained habitats include spawning sites (Ciannelli et al., 2014), nursery areas (Beck et al.,  
89 2001), and dispersal pathways between them (Petitgas et al., 2013). During other life stages the  
90 same individuals are able to change their distribution and track climate velocities (e.g., Alvarez-  
91 Berastegui et al., 2014).

92         The effects of climate variability on species distributions are often assessed by examining  
93 a single or few life stages utilizing readily available survey data (e.g., Kleisner et al., 2017;  
94 Thorson et al., 2016; Wisz et al., 2015). However, parameterizing species distribution models in  
95 this manner may produce results that over-estimate the degree of spatial re-distribution in  
96 response to climate change, and result in the misinterpretation of the processes involved in  
97 driving such spatial responses (Asch & Erisman, 2018). Recognizing where and when in the  
98 species' life cycle spatial constraints occur is important to produce realistic distributional  
99 forecasts. Spatially constrained stages may act as a bottleneck to adaptation which can have  
100 fitness repercussions (Caddy, 2014). Therefore, identifying these constraints has important  
101 implications for monitoring and managing marine fish species in a changing climate. To date,  
102 there is no systematic knowledge of when in marine fish life cycles such spatial constraints are  
103 most likely to occur.

104         Our study has two objectives: i) develop analytical techniques to identify spatial  
105 constraints during a species' life cycle, ii) implement these new analytical techniques to compare  
106 patterns of life history spatial constraints across species with contrasting life histories. We chose  
107 to evaluate data rich species/populations from sub-arctic systems in both the North Atlantic  
108 (Scotian Shelf and Barents Sea) and the North Pacific (Bering Sea) (Fig. 1). The selection of

109 species in this study balances the need to include contrasting life history strategies and highly  
110 resolved life cycle data. Here we define the degree of spatial constraint by both the *consistency* at  
111 which a particular geographic area is used year after year, and by the *extent* of such an area  
112 relative to the entire population range. Consequently, life history stages that occupy the same  
113 areas over time, and are concentrated in an area less than the population range, are deemed highly  
114 constrained. In discussing our results, relevance is given to global climate change and the ways in  
115 which species can adapt in the presence of strong spatial constraints. With these study goals and  
116 methodologies in mind, we anticipate defining the possible existence of bottlenecks to species  
117 adaptability through changes of spatial distribution, to generate hypotheses linking a species' life  
118 history and the processes affecting the selection of habitat through the life cycle, and to develop  
119 spatial distribution models that consider multiple life history stages and that can be used to  
120 project species distributions in relation to future climate change scenarios.

121

## 122 **Methods**

123 The methods are organized in two sections. We first describe the quantitative metrics for  
124 assessing habitat constraints, and then describe the application of these metrics to a simulated set  
125 of data and ten sub-arctic fish populations that are representative of contrasting life history  
126 strategies and habitat requirements.

127

### 128 *Metrics for assessing habitat constraint*

129 We defined the habitat constraint (*HC*) for a species (*s*) at a target age or size (*a*), as the product  
130 of the geographic *consistency* *C*, dimensionless, ranging from 0 (not consistently occupied) to 1  
131 (consistently occupied) at which a specific geographic location is occupied, and the geographic

132 *extent E*, dimensionless, ranging from 0 (small extent) to 1 (large extent) of that geographic  
133 location:

134

$$135 \quad (HC)_a = C_a(1 - E_a) \quad (1)$$

136

137 As such, the habitat constraint metric ranges from 0 to 1. It approaches 1 when, over the studied  
138 period, fish at a target age or size consistently occupy a very small fraction of the area where the  
139 population is known to occur. Conversely, the constraint metric will be close to zero when target  
140 age or size distribution is highly variable from year to year and widely spread over the population  
141 range. Note that the habitat constraint changes for each species, however in equation (1) the sub  
142 index for species has been omitted.

143 We used Generalized Additive Models (GAMs) to determine  $C_a$ . GAMs allow for  
144 nonlinearities between covariates and response, and therefore are well suited to capture unique  
145 patterns of spatial dependency (Wood, 2017). Let  $X_{a,lat,lon,y}$  be the natural log of the local  
146 abundance of the studied species at the target age or size  $a$ , collected over multiple years  $y$ , at  
147 locations defined by geographic coordinates ( $lat$ ,  $lon$ ). We quantify the consistency of habitat use  
148 ( $C_a$ ) as the difference of the residual error between the *base* model (2), which only includes  
149 yearly variable intercepts ( $\beta_{a,y}$ ) and the day of the year effect (i.e., phenology  $d$ ),

150

$$151 \quad X_{a,lat,lon,y} = \beta_{a,y} + s_a(d) + \varepsilon_{b_{a,lat,lon,y}} \quad (2)$$

152

153 and the *habitat* model (3), which includes also the effect of geographic coordinates ( $lat$ ,  $lon$ )

154

155  $X_{a,lat,lon,y} = \beta_{a,y} + s_a(d) + g_a(lat, lon) + \varepsilon_{h_{a,lat,lon,y}}$  (3)

156

157 The parameter  $\beta_{a,y}$  is a stage and year specific intercept, included as a fixed effect to account for  
 158 interannual changes in abundance. Consideration of including year as a fixed effect is most  
 159 important when population abundance varies from year to year but distribution does not. In such  
 160 circumstances a model that only includes year can explain a large portion of the data variance ( $R^2$   
 161 is high) even though there is no spatial variability in the data. The function  $g$  is a two-  
 162 dimensional smoother that varies by stage,  $s$  is a one-dimensional smoother that also varies by  
 163 stage, and  $\varepsilon$  is a random error assumed to be normally distributed (on the log scale) with mean  
 164 equal to zero and variance equal to the residual variance of the model. We run models with a  
 165 Tweedie distribution and soap film smoother which yielded very similar results to the models  
 166 with lognormal distribution and thin plate regression splines. Models 2 and 3 are fitted separately  
 167 for each age or size-group of a population.

168 Given the base and habitat models in 2 and 3, the consistency metric is derived as

169 follows:

170

171 
$$C_a = \frac{\sum(\varepsilon_{b_{a,lat,lon,y}})^2 - \sum(\varepsilon_{h_{a,lat,lon,y}})^2}{\sum(\varepsilon_{b_{a,lat,lon,y}})^2}$$
 (4)

172

173 If a target age or size group consistently uses the same habitat each year, then the error of the  
 174 habitat model ( $\varepsilon_{h_{a,lat,lon,y}}$ ) will be small, the ensuing  $C_a$  metric will be high, and approach 1. For  
 175 a habitat used very inconsistently from one year to the next,  $C_a$  will approach 0. Note that when  
 176 using field data, it is improbable for the consistency and constraint metrics to reach the  
 177 theoretical maximum of 1, because of uncertainty in the data.



178 The determination of  $C_a$  from models (2) and (3) is executed on presence only data (i.e.,  
179 positive, non-zero catches). However, corresponding logistic regression models are also  
180 implemented using the presence-absence data to estimate the probability of occurrence of the  
181 target stage. The presence-absence and the presence only models are used together to predict  
182 density over a regularly spaced array of stations for the calculation of habitat extent, as discussed  
183 in the next paragraph.

184 Given an age or size group  $a$ , its geographical *extent* ( $E_a$ ), is determined by the smallest  
185 fractions of the total occupied habitat that contains a pre-specified fraction (e.g.,  $f=0.75$ ) of the  
186 predicted total abundance at the raw scale, of the species at the targeted age or size. Namely,  
187

$$188 \quad E_a = \frac{A_{f,a}}{A_t} / f \quad (5)$$

189  
190 where  $A_{f,a}$  is the smallest area containing a fraction  $f$  of the species total abundance at the targeted  
191 age/size, and  $A_t$  is the total area occupied by the population. Because the surveys we evaluate  
192 cover areas where the targeted species has never occurred and because observations are collected  
193 on irregular or inconsistently sampled grids,  $A_t$  is here defined by a regular array of equally  
194 spaced grid points that are less than 30 km from a location with a positive occurrence of the  
195 targeted species.  $A_{f,a}$  is equal to the areal sum of all the grid cells (H), each with area ( $h$ ), and  
196 whose cumulative fraction of predicted abundance is less than or equal to  $f$ :

$$197 \quad A_{f,a} = \sum_{i=1}^H h_i$$

199

200 We choose the fraction  $f$  equal to 75% of the predicted abundance within the area  $A_t$ , and for  
201 three of the ten case studies examined, we assessed the sensitivity of  $f$  to the habitat extent and  
202 habitat constraint metrics (Appendix S1, Figs. S1-S3). To predict the density of the targeted  
203 species and stage at each location in the array ( $\hat{d}_{h_i,a}$ ), we use the delta method (Porter &  
204 Ciannelli, 2018). This was achieved by taking the product of the probability of occurrence ( $u$ ) and  
205 the abundance ( $x$ ) of the target stage and species predicted from a presence-absence and presence  
206 only habitat model (3), respectively:

207

$$208 \quad \hat{d}_{h_i,a} = \hat{u}_{h_i,a} \hat{x}_{h_i,a}.$$

209

210 To predict  $A_{f,a}$ , the array units ( $h$ ) are first sorted in decreasing order of the respective  
211 predicted density, such that  $\hat{d}_{h_i} \geq \hat{d}_{h_{i+1}}$ , and their respective areas are summed until the  
212 fractional abundance threshold  $f$  is reached. When so derived, the extent metrics ( $E_a$ ) ranges from  
213 0 to 1. It approaches 0 when the target stage is spatially clustered within the species distribution  
214 range, and 1, when the target stage is uniformly distributed across the species distribution range.

215

216 *Applying constraint metrics to simulated data and species with contrasting life histories*

217 We implemented the habitat constraint models to a simulated data set and to ten case  
218 studies, including six groundfish species from the Eastern Bering Sea, three from the Scotian  
219 Shelf, and one from the Barents Sea (Table 1). For the simulated data set we formulated four  
220 spatial distribution scenarios (Table S1), each varying in the degree of spatial autocorrelation and  
221 conditional effects on position: 1. *High constraint*, with yearly stable conditional distribution on  
222 the coordinate system, indicative of highly spatially correlated distribution with high spatial

223 gradients that do not change over time (Figure S4); 2. *Moderate constraint*, with yearly stable  
224 conditional distribution on the coordinate system, indicative of moderately spatially correlated  
225 distribution with moderate spatial gradients that do not change over time, but have greater spatio-  
226 temporal variability compared to the previous scenario (Figure S5); 3. *Environmental constraint*,  
227 with high and yearly variable conditional distribution on position, indicative of a highly spatially  
228 correlated distribution with a latent environmental variable (e.g., water temperature), which also  
229 affects distribution (Figure S6); 4. *Unconstrained*, characterized by unconditional distribution,  
230 indicative of interannually variable distribution and limited spatial autocorrelation (Figure S7).  
231 All scenarios were formulated using the function `gstat` in R (version 2.0-7), which produces  
232 Gaussian random fields given variogram models and the linear coefficients specified in the  
233 Appendix S2, Table (S1). For all scenarios we reproduced 20 random fields, each representative  
234 of a yearly distribution, and in the Appendix S2 we show the first 10 of these fields. Results did  
235 not change by increasing the number of random fields beyond 20.

236 We expect the geographic consistency metric ( $C$ ) to be highest for the high constraint  
237 scenario, and progressively smaller for the moderate, environmental constraint, and  
238 unconstrained scenarios. We expect the habitat extent metric ( $E$ ) to be low for the high constraint  
239 scenario, and progressively higher for the moderate and environmental constraint scenarios, and  
240 highest for the unconstrained scenario. Consequently, we anticipate the habitat constraint metric  
241 ( $HC$ ) to be highest for the high constraint scenario due to high geographic consistency and low  
242 habitat extent, and progressively smaller for the moderate, environmental and unconstrained  
243 scenarios, due to low consistency and high habitat extent.

244 Fish distribution data used in the ten case studies were collected as part of long-term  
245 survey programs conducted by the agencies that oversee fisheries management and assessment in  
246 their regions, including the National Oceanic and Atmospheric Administration (NOAA) for the

247 USA (Lauth et al., 2019), the Canadian Department of Fisheries and Oceans for Canada (DFO;  
248 Frank et al., 2018), the Institute of Marine Research (IMR) for Norway (Fall et al., 2018), and the  
249 Polar Branch of Russian Federal Research Institute of Fisheries and Oceanography (VNIRO) for  
250 Russia (Stige et al., 2019). We only included stages of species for which the available survey  
251 coverage clearly reflected the core distribution of the targeted age or size, and for years with  
252 more than 40 stations sampled. In general, the timing of the juvenile and adult survey is summer,  
253 while that of eggs and larval stages changes depending on the spawning phenology of the species.  
254 Additional information about each survey, including years and months covered, is provided in the  
255 Appendix S3.

256         To compare patterns of habitat constraint across the ten case studies we first derived an  
257 index of ontogenetic development ( $O_{s,a}$ ) as the ratio between the size or age of the examined life  
258 history stage and the maximum size or age of the species (Mindel et al., 2016). If the ontogenetic  
259 categories were age based, we assigned an age of 0.05 year of age to eggs and 0.15 year to larvae.  
260 If the categories were size based, we assigned 1 mm to eggs and 1.5 mm to larvae. For older or  
261 larger categories, we assigned the age or size measured from the survey. We are aware that  
262 ontogeny does not linearly scale with either age or size. Therefore, we also considered alternative  
263 ways of calculating the ontogenetic index, e.g., as the log of the age or size over the log of  
264 maximum age or size. Because the results were not sensitive to the way in which the ontogenetic  
265 index was calculated, we opted for the simpler linear formulation. We then fit a GAM to the  
266 habitat constraint versus ontogenetic index metrics. In these analyses, the inter-species variability  
267 was first included as a random term, which allowed us to estimate an average relationship across  
268 the ten combinations of species and populations, and the variance components associated with the  
269 fixed and random terms:

270

271  $HC_{s,a} = \beta + g(O_a) + \delta_s + \varepsilon_a$  (6)

272  
273 where  $\beta$  is a model intercept equal to the average habitat constraint across all species and stages,  
274  $g$  is a smooth non-parametric function,  $\delta_s$  is a random effect varying by combinations of species  
275 and population (10 estimates) assumed to be normally distributed, and  $\varepsilon_a$  is the residual variation,  
276 also normally distributed.

277 In a second GAM formulation, the inter-population variability was modeled as a fixed  
278 effect, which allowed us to estimate a specific smooth term and average constraint for each  
279 examined population:

280

281  $HC_{s,a} = \beta_s + g_s(O_{s,a}) + \varepsilon_a$  (7)

282

283 In GAM formulations (6) and (7), the maximum degrees of freedom for the one-dimensional  
284 smooth terms were limited to three, to avoid overfitting. Significance level for the fixed effects of  
285  $g_s$  were set at 0.1.

286 Where possible, the above analyses were applied to multiple life stages, including eggs,  
287 larvae, and eight or nine progressively larger or older juvenile and adult stages. Some case studies  
288 did not include the egg and larval stages due to lack of taxonomic resolution (arrowtooth  
289 flounder, *Atheresthes stomias*, Pleuronectidae) or because of unique life history characteristics  
290 (Pacific cod, *Gadus macrocephalus*, Gadidae, possess demersal eggs); all the Scotian Shelf case  
291 studies did not include the egg stage. For the Bering Sea case studies, juvenile and adult stages  
292 were partitioned into eight size groups, with size thresholds chosen to create uniform sample  
293 sizes. For the Scotian Shelf and Barents Sea case studies, juvenile and adult stages were

294 partitioned into nine age groups, (age-0 to age-8 and older). For each species, we only included  
295 data from sampled stations that are less than 30 km away from a sampled location that had at  
296 least one positive occurrence of the targeted species at any age or size. Thus, we eliminated  
297 surveyed stations where the targeted species had never occurred.

298         For a given species, with the exception of the Barents Sea case study, we limited the  
299 analyses to overlapping years of data availability across the stages examined. So, for the Scotian  
300 Shelf case studies, we restricted the analyses from 1978 to 1982, corresponding to the years when  
301 ichthyoplankton surveys were conducted. Given the limited number of ichthyoplankton years  
302 available for the Scotian Shelf cases, we repeated the analyses including the full extent of the  
303 adult and juvenile stages, based on annual surveys conducted during 1970-2015, and for these  
304 stages we found no differences with the results obtained by only including the 1978-1982 time  
305 interval. For the Bering Sea, data were available from 1982 to 2018 for adults and juveniles, and  
306 from 1988 to 2018 for eggs and larvae, however, the exact years changed among the species  
307 examined due to the necessity of including no less than 40 sampled stations within each year. For  
308 the Barents Sea, temporal coverage differed between stages. Egg and larval data were only  
309 available from 1981 to 1993, while the autumn juvenile and adult surveys were only available  
310 from 2003 onward. So, in this case we could not restrict the analysis to overlapping years.  
311 Further information about the years and months included in the analyses is provided in the  
312 Appendix S3.

313         To estimate the relationship between habitat constraint and the ontogenetic index (Eqs. 6  
314 and 7), we included species and stages that had greater than 400 stations with positive catches  
315 through all years – a number of stations that correspond to less than the 10% quantile of available  
316 sample sizes across all examined species and stages. A minimum of 400 stations was deemed  
317 necessary to simultaneously fit one- and two-dimensional smooth functions of phenology and

318 distribution in the model (3). This caused the removal of yellowfin sole (*Limanda aspera*,  
319 Pleuronectidae) eggs (n = 33), Alaska plaice (*Pleuronectes quadrituberculatus*, Pleuronectidae)  
320 larvae (n = 253), Scotian Shelf haddock (*Melanogrammus aeglefinus*, Gadidae, n = 160) and cod  
321 (*Gadus morhua*, Gadidae, n = 262) larvae, Scotian Shelf cod age-0 (n = 262) and age-1 (n = 333),  
322 Scotian Shelf hake (*Merluccius bilinearis*, Gadidae) age-0 (n = 372), and Scotian Shelf haddock  
323 age-0 (n = 283) and age-1 (n = 342). To reduce model uncertainties, the size or age specific data  
324 should ideally capture the distribution range and have similar sample size across size or age  
325 groups. In our applications, we removed yellowfin sole larvae because their distribution was  
326 clearly truncated at the northern end of the sampled area, in spite of this stage including 603  
327 stations. The majority of arrowtooth flounder abundance in the Bering Sea is within the surveyed  
328 area (Ciannelli et al., 2012), thus they were still retained in the analyses. For the remaining cases,  
329 the area monitored include clusters of high numerical density, and covers bathymetric and  
330 oceanographic features that are representative of the habitat variability experienced throughout  
331 the population range. Thus, we are confident that the data analyzed effectively captures the  
332 degree to which different size or age groups distribute over space. To determine whether our data  
333 inclusion criteria affected the results, we also implemented the analyses in Eqs. 6 and 7 on the  
334 unrestricted set of available data. All analyses were conducted in R (version 3.6.2) and GAM  
335 models were fitted with the mgcv library (1.8-33). With the exception of data from the Barents  
336 Sea case study, all other data and R code are deposited in:  
337 <https://github.com/lciannel/lifehistoryconstraint>.

338

## 339 **Results**

340 *Implementing habitat constraints analyses to simulated and field data*

341 *Simulated data*

342 The results of the constraint analyses on simulated data corroborate our expectations. The habitat  
343 constraint metric was lowest in the unconditional (HC = 0.001) and environmental (HC = 0.01)  
344 constraint scenarios and progressively higher for the moderate (HC = 0.360) and high constraint  
345 scenarios (HC = 0.759, Appendix 2, Fig. S8). The high constraint scenario had highest  
346 consistency (C = 0.841) and lowest habitat extent (E = 0.10) metrics, and the unconditional  
347 scenario had the opposite pattern (C = 0.002, E = 0.550). The environmental constraint had high  
348 explained variance ( $R^2$ ) but low geographic consistency metrics, due to the interannually variable  
349 abundance and spatial gradients of this scenario (Appendix 2, Fig. S6).

### 350 *Field data*

351 We show results for two species, walleye pollock (*Gadus chalgogrammus*, Gadidae) in the  
352 Bering Sea and haddock on the Scotian Shelf. These two species are chosen because they are  
353 considered representative of the other species examined. Results for the other eight case studies  
354 are provided in the Appendix S4.

### 355 *Bering Sea walleye pollock*

356 The distribution of walleye pollock in the Eastern Bering Sea changes for different  
357 ontogenetic stages and size groups (Fig. 2). At the egg and larval stages, the majority of the  
358 estimated abundance is confined to the Alaska Peninsula and to the Pribilof Islands regions,  
359 which are known spawning locations. The slight difference in distribution between egg and larval  
360 stages is attributable to passive transport by currents from spawning to juvenile habitats. An  
361 isolated cluster of larval abundance was identified in the Bering Sea basin region, as a result of  
362 isolated high larval catches in the early year of the available time series. As individuals grow and  
363 develop, they occupy larger areas of the outer (depth 100-200 m) and middle (depth 50-100 m)  
364 shelf regions, illustrated by a progressive increase of the habitat extent for these stages (Fig. 2).  
365 Estimates of habitat constraint decline with ontogeny, ranging from 0.22 at the egg stage to 0.06



366 at an intermediate adult stage. This is due to both a higher interannual spatio-temporal variance  
367 associated with larger sized individuals, and a progressive increase of habitat extent (Fig. 3). The  
368 consistency at which a stage occupies a specific geographic area decreases notably for individuals  
369 in the middle of size ranges, between 383 mm and 444 mm in length, denoting large interannual  
370 variability of spatial distribution at corresponding size ranges (Fig. 3).

### 371 *Scotian Shelf Haddock*

372 The spatial distribution of haddock on the Scotian Shelf does not change through different  
373 life stages, which is unlike the pollock case study where distribution changes occurred across life  
374 stages. There are, however, differences in the consistency and extent of habitat use across their  
375 life history. High concentrations of haddock larvae are located at the southern and western  
376 portion of the Scotian Shelf, corresponding to Browns and Emerald Banks, which are known  
377 spawning grounds. Consequently, the majority of the estimated larval abundance is confined to  
378 these two locations (Fig. 4). Habitat expansion and increased dispersion is observed during the  
379 age-0 stage, (due to greater mobility of these juveniles), while distribution becomes concentrated  
380 and patchier at older and reproductively mature life stages (age-1+) (Fig. 4).

381 Haddock's metrics of habitat constraints are higher than those of pollock, and are highest  
382 between age-2 (0.26) and age-4 (0.27) and then decline for the older life stages, at age-8 (0.17)  
383 and age-9+ (0.18). During the larval and age-0 stages, haddock metrics of constraint are also low,  
384 but the sample size for these stages is small compared to that of other stages, and may have  
385 contributed to the high interannual variability and low consistency of spatial distribution (Fig. 5).

386

### 387 *Inter-species comparisons*

388 By including population as a random term (Eq. 6), we found a significant and linear effect  
389 of the ontogenetic index on habitat constraint across the ten case studies examined (Fig. 6). This

390 outcome implies that younger stages and size groups, on average, tend to be more highly  
391 constrained in space than older stages and larger size groups. However, the variance component  
392 of the random term (population) is three orders of magnitude greater than the variance explained  
393 by the ontogeny effect, underscoring high inter-population (or species) variability in this  
394 relationship (Table 2). Results did not change when we included species and stages that did not  
395 meet our criteria for retention in the inter-specific comparisons.

396         The variability in the shape and elevation of the smoothed ontogenetic index is evident  
397 from the results of the fixed effect model (Eq. 7). Three of the populations exhibit habitat  
398 constraints that persist into the juvenile or even adult stages, including Alaska plaice, yellowfin  
399 sole, and flathead sole (*Hippoglossoides elassodon*, Pleuronectidae) and for two of these species  
400 the relationship between the ontogenetic index and habitat constraint is nonlinear (e.g., flathead  
401 sole and yellowfin sole) (Fig. 7). Five populations have a gradual linear decline of habitat  
402 constraint with ontogeny (Barents Sea cod, Bering Sea pollock and cod, and Scotian Shelf  
403 haddock and cod), while the remaining two populations have no significant relationship between  
404 habitat constraint and ontogeny (arrowtooth flounder, silver hake). In addition to variability in the  
405 shape of the ontogenetic effect we also found significant variability in the elevation associated  
406 with each population (Table 2). Yellowfin sole, Alaska plaice and flathead sole have the highest  
407 intercept estimates, indicating a stronger overall affinity of these populations to location or  
408 substrate, compared to other cases. The relationships between the metrics of habitat constraint  
409 and extent with the ontogenetic index did not change with changes of the parameter  $f$  (fraction of  
410 abundance for the habitat extent metric) (Appendix S1).

411

## 412 **Discussion**

413 *Drivers of life history spatial constraints*

414 Our analyses clearly illustrate a tendency for the early phase of the life cycle to be more  
415 constrained in space than other life stages for the various marine fish species evaluated. The  
416 existence of life history spatial constraints is common and well documented in terrestrial  
417 vertebrates and invertebrate species, and have practical implications for conservation. For  
418 example, stopover sites of long-distance migratory birds, such as the bar-tailed godwits (*Limosa*  
419 *lapponica*) in the eastern Atlantic flyway (Scheiffarth et al., 2002), are critical to the completion  
420 of the species' annual migration. Likewise, migrating and overwintering generations of the  
421 eastern monarch butterflies *Danaus plexippus* are known to concentrate from a 4.5 million km<sup>2</sup>  
422 breeding ground during spring and summer, down to an approximately 1 km<sup>2</sup> area in winter  
423 (Reppert & de Roode, 2018). In contrast to these examples of strong spatial affinity and site  
424 fidelity for terrestrial species, the current paradigm for distribution models of marine fish species  
425 (especially pelagic species; Cheung et al., 2015), is that of a loose association with discrete  
426 geographic locations. This is in part because of their presumed high dispersing life history  
427 strategy, and in part because of the spatially dynamic nature of pelagic seascapes. This is the  
428 basic premise behind the postulate that fish species can adapt to global warming by shifting their  
429 spatial distribution, leading to a redistribution of global catches, with loss of abundance at the  
430 tropics and gains at higher latitudes (Cheung et al., 2010; Bindoff et al., 2019). While there is  
431 mounting evidence that many marine fish species are currently redistributing, here we show that  
432 some species from sub-arctic seas consistently occupy restricted geographic locations during the  
433 early phases of their life cycle. The existence of these spatial constraints will limit the degree to  
434 which these species can adapt to climate change through shifting distributions to higher latitudes.

435 We did not address the drivers of these spatial constraints, but it is reasonable to expect  
436 that natal homing (Cury, 2019) and a life history strategy tuned to the presence and persistence of  
437 topographic, sediment, and bathymetric characteristics are the primary reasons. In the Bering Sea

438 for example, organic content and sediment characteristics are correlated with higher trophic level  
439 community composition and abundance in benthic habitats (Grebmeier et al., 1989). Flatfish  
440 abundance has strong associations with sediment characteristics, and it is hypothesized that these  
441 associations are the result of diet preferences and benthic prey availability (McConnaughey &  
442 Smith, 2000). Similarly, on the Scotian Shelf there are strong habitat associations of the three  
443 gadids studied here (cod, silver hake and haddock) and sediment types (Scott, 1982b; Waiwood  
444 & Buzeta, 1989), and in the nearby Georges Bank newly settled juvenile cod and haddock are  
445 associated with pebble-gravel deposits on the northeast side of the bank (Lough et al., 1989).  
446 Bathymetric-driven circulation features, important for the dispersal and retention of fish eggs and  
447 larvae, are also potential drivers of spatial constraint during early life stages (Iles & Sinclair,  
448 1982). In the Bering Sea, canyons along the continental slope drive topographically steered  
449 circulation (Ladd, 2014) and affect the transport pathways of slope spawned flatfish eggs and  
450 larvae (Sohn et al., 2016). On the shelf, circulation features (e.g., gyres) can also aggregate and  
451 retain eggs and larvae in spatially restricted areas around banks (Lough et al., 2006; O'Boyle et  
452 al., 1984; Shackell et al., 1999; Smith & Morse, 1985). In our analyses of the Scotian Shelf,  
453 haddock and cod larvae generally occupied Browns and Emerald banks, and silver hake was  
454 found in the vicinity of Western Bank. These are known gadid spawning locations in the Scotian  
455 Shelf region (Brickman, 2003; Hurley & Campana, 1989), and previous studies have underscored  
456 the importance of larval retention around these banks (O'Boyle et al., 1984). While in principle  
457 during spatially constrained phases of a fish life cycle, water temperature may not be an  
458 important driver of spatial distribution, temperature may still act as an important covariate during  
459 other life history stages. For example, the spawning habitats of walleye pollock, in both the  
460 Eastern Bering Sea and western Gulf of Alaska, do not geographically change in relation to

461 variability in water temperature (Bacheler et al., 2010, 2012), while adult stages of walleye  
462 pollock outside of the spawning season shift northward during warm years (Thorson et al., 2020).

463       Exploitation and predator-prey interactions may also contribute to changes of habitat  
464 constraints through ontogeny. Sub-arctic systems are characterized by relatively few species,  
465 some of which are commercially harvested, with strong trophic interactions (Frank et al., 2006;  
466 Livingston, 2000). Strong exploitation erodes spatial structure and exploitation status is another  
467 important driver of spatial constraint (Ciannelli et al., 2013). On the Scotian Shelf, Frank et al.,  
468 (2018) found that exploitation drives ontogenetic-like deepening of Atlantic cod. Thus, one  
469 recommendation for future analyses would be compare patterns of range shifts in relation to  
470 exploitation status of the stock. Trophic interactions may shape the degree of patchiness,  
471 especially in pelagic fish species. For example, (Frank et al., 1993) found that capelin larvae on  
472 the Southeast Shoal of the Grand Banks were patchy during early stages, followed by increased  
473 dispersion, and then, with the onset of schooling behavior, distributions became patchier and  
474 were closely associated with their primary food sources. Some species, such as adult Atlantic cod  
475 and walleye pollock, are prone to eating smaller and younger conspecific individuals. So, their  
476 movement and distribution are linked to that of their prey (Ciannelli et al., 2007; Fall et al.,  
477 2018).

478  
479 *Expanding these analyses to other case studies*

480 The assessment of ontogenetic spatial constraints requires spatial data collected over multiple age  
481 or size groups of a species' life cycle, ideally during oceanographically contrasting years, and  
482 throughout the distributional range of the population. When data are available over different  
483 seasons, it is best to separate the analyses accordingly, as species can undergo seasonal  
484 contraction and expansions in their distribution. This is evident in the Barents Sea cod, with

485 winter distribution range limited by low temperature under sea ice, and therefore more  
486 constrained compared to summer (Appendix S4, Figs. S23 and S25). When using field data, it is  
487 impossible for the consistency and constraint metrics to reach the theoretical maximum of 1,  
488 because of uncertainty in the data. The results from the simulated data are instrumental to  
489 appreciate the influence of spatial and interannual variability, because they show that even for  
490 scenarios in which the population distribution is highly conditional on position, the habitat  
491 constraint metrics is less than 0.8. In the moderate constraint scenario, the metric is about 0.4,  
492 and in the environmental and random scenarios the metrics are close to zero.

493 Not surprisingly, the availability of data for ontogenetic habitat constraint analyses is  
494 often limited. Even for well-monitored systems examined here there were data gaps for specific  
495 life stages, especially during the egg and larval phases. In the Scotian Shelf, for example, we only  
496 had five overlapping years among larval, juvenile and adult collections, and for the Bering Sea  
497 there were no overlapping years among ichthyoplankton, juvenile and adult surveys. For all three  
498 species examined in the Scotian Shelf, results obtained on the full set of years in which juvenile  
499 and adult data were available (1970-2015), were similar to those obtained when only the five  
500 overlapping years (1978-1982) were analyzed, suggesting that a minimum of five years is  
501 necessary to obtain a spatial representation of geographic consistency and extent.

502 Ontogenetic spatial constraints inferred from data rich cases could provide insights for  
503 data poor species. In the cases we examined, three patterns of spatial constraint across life  
504 histories were evident (Fig. 7). The first pattern is characterized by a gradual and monotonic  
505 decline of spatial constraint, resulting in spawning individuals and eggs being most spatially  
506 constrained, and the non-spawning adult individuals being more spatially dispersed. Gadids,  
507 including walleye pollock, Scotian Shelf cod and haddock, Pacific cod and Barents Sea cod  
508 follow this pattern in so far as their populations have the simultaneous presence of spatially

509 constrained and highly mobile individuals. These species have strong site fidelity at spawning  
510 (Hurley & Campana, 1989; Neidetcher et al., 2014; Rose, 1993) and highly mobile adults during  
511 the nonbreeding seasons. Tagging studies of adult stages of Pacific cod (> 50 cm length) indicate  
512 widespread movement and occupancy during summer, and directional movement to slope  
513 spawning areas during winter (Shimada & Kimura, 1984). Similarly, mature Barents Sea cod  
514 undertake extensive southwards countercurrent migrations during winter toward spawning  
515 grounds along the Norwegian coast and migrate northwards to vast feeding grounds in the  
516 summer (Ottersen et al., 2014).

517         The second pattern is characterized by a delayed (flathead sole) or protracted (Alaska  
518 plaice, yellowfin sole) period of high spatial constraint, persisting into juvenile and sub-adult  
519 stages, before declining at later stages. Previous work has pointed to strong associations of  
520 yellowfin sole abundance in sandy substrate, flathead sole in mixed sand and mud, and Alaska  
521 plaice in mixed gravel and sand (McConnaughey & Smith, 2000). In the Bering Sea shelf,  
522 bathymetry is inversely correlated with sediment size and, not surprisingly, all three species have  
523 a strong affinity with specific depth ranges (Swartzman et al., 1992). Adult yellowfin sole  
524 performs annual spring and fall migrations to and from spawning areas (Wilderbuer et al., 1992).  
525 The timing of these migrations is influenced by water temperature and sea ice extent on the  
526 Bering Sea shelf, and in turn influences availability to the survey (Nichol et al., 2019). Therefore,  
527 it is possible that the rapid decline of habitat constraint of adult yellowfin sole may be driven by  
528 the changes in timing of their migration and availability to the summer survey, rather than  
529 interannual distribution changes.

530         The third pattern includes cases with a relatively constant level of spatial constraint  
531 through the species life cycle, indicating either a loose (e.g., arrowtooth flounder) or high (e.g.,  
532 silver hake) lifetime spatial associations. Species that conform to this pattern are at the

533 northernmost extreme of their range, and their spatial distribution and annual abundance may be  
534 particularly sensitive to water temperature changes. For example, arrowtooth flounder in the  
535 Bering Sea avoids water less than 2°C, which is typically observed in the middle shelf during  
536 summer on the Bering Sea (Ciannelli et al., 2012). Similarly, silver hake on the Scotian Shelf  
537 tends to occupy warmer water of the central and southwest portion of the shelf while avoiding the  
538 colder northeast shelf (Scott, 1982a), and bottom water temperature on the shelf is an important  
539 driver of their abundance (Reed et al., 2018).

540         We note that our cross-section of species only includes those residing in sub-arctic  
541 systems, and with either benthic or demersal strategies during the adult phase. Thus, augmenting  
542 these analyses with more species, including those from temperate and tropical reef species, as  
543 well as species with pelagic life history strategies during the adult phase (Muhling et al., 2013),  
544 would be instrumental to validate the trends of habitat constraint across the life cycle, and to  
545 establish the link with life history strategies. Ultimately, this broader level of investigation would  
546 further enable inference about habitat use and adaptability to climate change for data poor  
547 species, for which life history trait information is known but spatial data for multiple life stages is  
548 limiting.

549

#### 550 *Implications for species distribution models*

551 Assessment of habitat constraints through the species life cycle have important implications for  
552 accurate predictions of species distribution in relation to climate change. The presence of strong  
553 spatial dependency for some stages indicates that within the range of examined values, water  
554 temperature may not be a primary driver of spatial distribution (Stuart-Smith et al., 2021), and  
555 calls into question projections of species or catch distributions based solely on water temperature.  
556 Additionally, species distribution models are often parameterized using data from stages that are



557 highly variable, such as the adult stages during the summer feeding months, which is when most  
558 groundfish surveys occur in sub-arctic areas. Our analyses indicate that adult stages are more  
559 likely to follow temperature or climate velocities because they are less constrained in space.  
560 Consistently, stage-specific models have different prediction outcomes compared to those  
561 parameterized on only one life stage (Asch & Erisman, 2018). Life history stages with high  
562 habitat constraint will limit future distribution shifts and can be used as a “mask” increasing  
563 accuracy of current species distribution model predictions.

564

565 *To what degree can range shift offset climate change?*

566 Populations that have both a spatially constrained and a spatially unconstrained life stages can  
567 experience a further separation among their stage-specific habitats and therefore be challenged to  
568 close their life cycle. This appears to be the case for both Pacific cod and walleye pollock in the  
569 Bering Sea, where in recent years the juvenile and adult summer distributions have been shifting  
570 progressively northward (Stevenson & Lauth, 2019), while spawning habitats are fixed in space  
571 at the southern edge of their distribution range (Bacheler et al., 2012; Neidetcher et al., 2014).  
572 Spawning habitats can indeed change, as shown for some of the species included in this study  
573 (e.g., Barents Sea cod), however, it is difficult to determine the underlying causes (Langangen et  
574 al., 2019; Opdal, 2010; Sundby & Nakken, 2008).

575         When species are constrained in space during a certain life stage they will either adapt to  
576 new habitat conditions in order to change their distribution or eventually fail to survive. Adaptive  
577 changes are required when the distribution of the most limiting stage is 1) geographically  
578 controlled, and/or 2) the physiological tolerance limits (e.g., temperature-dependent  
579 development) of the most limiting stage have been reached. In 1), for species to shift distribution,  
580 adaptation must occur by linking the new geographic cues (e.g., deeper waters) to the

581 environmental conditions associated with suitable habitat (e.g., Bonamour et al., 2019). In 2), if  
582 physiological tolerance limits have been reached, adaptation in the physiological response to the  
583 environment would be needed before further shifts could occur. In the Gulf of Alaska for  
584 example, water temperature is significantly warmer than the Bering Sea, and laboratory and  
585 modeling analyses indicate that Pacific cod eggs may be experiencing thermal stress at their  
586 traditional spawning sites, which may further limit recruitment (Laurel & Rogers, 2020). To  
587 adapt to these changes Pacific cod may evolve new cues and spawn in cooler habitats within the  
588 Gulf of Alaska, and/or shift the thermal tolerance of embryos, or perish. Models that account for  
589 life history constraints as outlined here, that are extended to also include physiologically based  
590 relationships (Kearney & Porter, 2009) of environmental covariates in addition to a fixed spatial  
591 term, can be used to simulate plastic and adaptive responses of fish to climate change.  
592 Furthermore, modelling the dispersion of free-drifting life stages and energy budget modelling of  
593 migratory adults might be needed to assess whether different spatial constraints across life stages  
594 will necessitate evolutionary adaptations in order for the species to close its life cycle in a future  
595 climate.

596         There is unfortunately a chasm in the marine sciences literature with fisheries  
597 oceanographers being more keenly aware of plastic responses of fish to climate change and life-  
598 history biologists focused more on evolutionary adaptations (Hutchings, 2011). Bridging this gap  
599 between plasticity and adaptations (Anderson et al., 2012; Schoener, 2011) to understand and  
600 model how marine species respond to climate change and variability (Donelson et al., 2019), as  
601 well as other anthropogenic stressors, will be an important advancement in marine ecology.

602

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612

### 613 **Data availability**

614 With the exception of data from the Barents Sea case study, all other data and R code are  
615 deposited in: <https://github.com/lciannel/lifehistoryconstraint>

616

617 **Literature cited**

- 618 Alvarez-Berastegui, D., Ciannelli, L., Aparicio-Gonzalez, A., Reglero, P., Hidalgo, M., López-  
619 Jurado, J. L., Tintoré, J., & Alemany, F. (2014). Spatial Scale, Means and Gradients of  
620 Hydrographic Variables Define Pelagic Seascapes of Bluefin and Bullet Tuna Spawning  
621 Distribution. *PLoS ONE*, 9(10), e109338. <https://doi.org/10.1371/journal.pone.0109338>
- 622 Anderson, J. T., Panetta, A. M., & Mitchell-Olds, T. (2012). Evolutionary and Ecological  
623 Responses to Anthropogenic Climate Change: Update on Anthropogenic Climate Change.  
624 *PLANT PHYSIOLOGY*, 160(4), 1728–1740. <https://doi.org/10.1104/pp.112.206219>
- 625 Asch, R. G., & Erisman, B. (2018). Spawning aggregations act as a bottleneck influencing  
626 climate change impacts on a critically endangered reef fish. *Diversity and Distributions*,  
627 24(12), 1712–1728. <https://doi.org/10.1111/ddi.12809>
- 628 Bacheler, N. M., Ciannelli, L., Bailey, K. M., & Bartolino, V. (2012). Do walleye pollock exhibit  
629 flexibility in where or when they spawn based on variability in water temperature? *Deep*  
630 *Sea Research Part II: Topical Studies in Oceanography*, 65–70, 208–216.  
631 <https://doi.org/10.1016/j.dsr2.2012.02.001>
- 632 Bacheler, N. M., Ciannelli, L., Bailey, K. M., & Duffy-Anderson, J. T. (2010). Spatial and  
633 temporal patterns of walleye pollock ( *Theragra chalcogramma* ) spawning in the eastern  
634 Bering Sea inferred from egg and larval distributions. *Fisheries Oceanography*, 19(2),  
635 107–120. <https://doi.org/10.1111/j.1365-2419.2009.00531.x>
- 636 Barbeaux, S. J., & Hollowed, A. B. (2018). Ontogeny matters: Climate variability and effects on  
637 fish distribution in the eastern Bering Sea. *Fisheries Oceanography*, 27(1), 1–15.  
638 <https://doi.org/10.1111/fog.12229>

639 Bartolino, V., Ciannelli, L., Bacheler, N. M., & Chan, K.-S. (2011). Ontogenetic and sex-specific  
640 differences in density-dependent habitat selection of a marine fish population. *Ecology*,  
641 92(1), 189–200. <https://doi.org/10.1890/09-1129.1>

642 Beck, M. W., Heck, K. L., Able, K. W., Childers, D. L., Eggleston, D. B., Gillanders, B. M.,  
643 Halpern, B., Hays, C. G., Hoshino, K., Minello, T. J., Orth, R. J., Sheridan, P. F., &  
644 Weinstein, M. P. (2001). The Identification, Conservation, and Management of Estuarine  
645 and Marine Nurseries for Fish and Invertebrates. *BioScience*, 51(8), 633.  
646 [https://doi.org/10.1641/0006-3568\(2001\)051\[0633:TICAMO\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2001)051[0633:TICAMO]2.0.CO;2)

647 Bindoff, N. L., Cheung, W. W. L., Kairo, J. G., Aristegui, J., Guinder, V. A., Hallberg, R., Hilmi,  
648 N., Jiao, N., Karim, M. S., Levin, L. A., O'Donoghue, S., Purca Cuicapusa, S. R.,  
649 Rinkevich, B., Suga, T., Tagliabue, A., and Williamson, P. (2019). Changing Ocean,  
650 Marine Ecosystems, and Dependent Communities. In: H.-O. Pörtner, D. C. Roberts, V.  
651 Masson-Delmotte, P. Zhai, M. Tignor, E. Poloczanska, K. Mintenbeck, A. Alegría, M.  
652 Nicolai, A. Okem, J. Petzold, B. Rama and N. M. Weyer, editors, IPCC Special Report on  
653 the Ocean and Cryosphere in a Changing Climate. IPCC.

654 Bograd, S., Hazen, E., Howell, E., & Hallowed, A. (2014). The Fate of Fisheries Oceanography:  
655 Introduction to the Special Issue. *Oceanography*, 27(4), 21–25.  
656 <https://doi.org/10.5670/oceanog.2014.83>

657 Bonamour, S., Chevin, L.-M., Charmantier, A., & Teplitsky, C. (2019). Phenotypic plasticity in  
658 response to climate change: The importance of cue variation. *Philosophical Transactions*  
659 *of the Royal Society B: Biological Sciences*, 374(1768), 20180178.  
660 <https://doi.org/10.1098/rstb.2018.0178>

661 Brickman, D. (2003). Controls on the distribution of Browns Bank juvenile haddock. *Marine*  
662 *Ecology Progress Series*, 263, 235–246. <https://doi.org/10.3354/meps263235>

663 Caddy, J. F. (2014). Why do assessments of demersal stocks largely ignore habitat? *ICES Journal*  
664 *of Marine Science*, 71(8), 2114–2126. <https://doi.org/10.1093/icesjms/fss199>

665 Cheung, W. W. L., Brodeur, R. D., Okey, T. A., & Pauly, D. (2015). Projecting future changes in  
666 distributions of pelagic fish species of Northeast Pacific shelf seas. *Progress in*  
667 *Oceanography*, 130, 19–31. <https://doi.org/10.1016/j.pocean.2014.09.003>

668 Cheung, W. W. L., Lam, V. W. Y., Sarmiento, J. L., Kearney, K., Watson, R., Zeller, D., &  
669 Pauly, D. (2010). Large-scale redistribution of maximum fisheries catch potential in the  
670 global ocean under climate change: Climate change impacts on catch potential. *Global*  
671 *Change Biology*, 16(1), 24–35. <https://doi.org/10.1111/j.1365-2486.2009.01995.x>

672 Ciannelli, L., Bailey, K., & Olsen, E. M. (2014). Evolutionary and ecological constraints of fish  
673 spawning habitats. *ICES Journal of Marine Science*, 72(2), 285–296.  
674 <https://doi.org/10.1093/icesjms/fsu145>

675 Ciannelli, L., Bartolino, V., & Chan, K.-S. (2012). Non-additive and non-stationary properties in  
676 the spatial distribution of a large marine fish population. *Proceedings of the Royal Society*  
677 *B: Biological Sciences*, 279(1743), 3635–3642. <https://doi.org/10.1098/rspb.2012.0849>

678 Ciannelli, L., Dingsør, G. E., Bogstad, B., Ottersen, G., Chan, K., Gjørseter, H., Stiansen, J. E., &  
679 Stenseth, N. Chr. (2007). Spatial anatomy of species survival: Effects of predation and  
680 climate-driven environmental variability. *Ecology*, 88(3), 635–646.  
681 <https://doi.org/10.1890/05-2035>

682 Ciannelli, L., Fisher, J., Skern-Mauritzen, M., Hunsicker, M., Hidalgo, M., Frank, K., & Bailey,  
683 K. (2013). Theory, consequences and evidence of eroding population spatial structure in  
684 harvested marine fishes: A review. *Marine Ecology Progress Series*, 480, 227–243.  
685 <https://doi.org/10.3354/meps10067>

686 Ciannelli, L., Knutsen, H., Olsen, E. M., Espeland, S. H., Asplin, L., Jelmert, A., Knutsen, J. A.,  
687 & Stenseth, N. C. (2010). Small-scale genetic structure in a marine population in relation  
688 to water circulation and egg characteristics. *Ecology*, *91*(10), 2918–2930.  
689 <https://doi.org/10.1890/09-1548.1>

690 Corten, A. (2002). The role of “conservatism” in herring migrations. *Reviews in Fish Biology*  
691 *and Fisheries*, *11*(4), 339–361. <https://doi.org/10.1023/A:1021347630813>

692 Crozier, L. G., & Hutchings, J. A. (2014). Plastic and evolutionary responses to climate change in  
693 fish. *Evolutionary Applications*, *7*(1), 68–87. <https://doi.org/10.1111/eva.12135>

694 Cury, P. (2019). Obstinate nature. *ICES Journal of Marine Science*, *76*(2), 384–391.  
695 <https://doi.org/10.1093/icesjms/fsy188>

696 Cury, P., Shin, Y., Planque, B., Durant, J., Fromentin, J., Kramerschadt, S., Stenseth, N., Travers,  
697 M., & Grimm, V. (2008). Ecosystem oceanography for global change in fisheries. *Trends*  
698 *in Ecology & Evolution*, *23*(6), 338–346. <https://doi.org/10.1016/j.tree.2008.02.005>

699 Donelson, J. M., Sunday, J. M., Figueira, W. F., Gaitán-Espitia, J. D., Hobday, A. J., Johnson, C.  
700 R., Leis, J. M., Ling, S. D., Marshall, D., Pandolfi, J. M., Pecl, G., Rodgers, G. G., Booth,  
701 D. J., & Munday, P. L. (2019). Understanding interactions between plasticity, adaptation  
702 and range shifts in response to marine environmental change. *Philosophical Transactions*  
703 *of the Royal Society B: Biological Sciences*, *374*(1768), 20180186.  
704 <https://doi.org/10.1098/rstb.2018.0186>

705 Fall, J., Ciannelli, L., Skaret, G., & Johannesen, E. (2018). Seasonal dynamics of spatial  
706 distributions and overlap between Northeast Arctic cod (*Gadus morhua*) and capelin  
707 (*Mallotus villosus*) in the Barents Sea. *PLOS ONE*, *13*(10), e0205921.  
708 <https://doi.org/10.1371/journal.pone.0205921>

709 Frank, K. T., Carscadden, J. E., & Leggett, W. C. (1993). Causes of spatio-temporal variation in  
710 the patchiness of larval fish distributions: Differential mortality or behaviour? *Fisheries*  
711 *Oceanography*, 2(3–4), 114–123. <https://doi.org/10.1111/j.1365-2419.1993.tb00129.x>

712 Frank, K. T., Petrie, B., Leggett, W. C., & Boyce, D. G. (2018). Exploitation drives an  
713 ontogenetic-like deepening in marine fish. *Proceedings of the National Academy of*  
714 *Sciences*, 115(25), 6422–6427. <https://doi.org/10.1073/pnas.1802096115>

715 Frank, K. T., Petrie, B., Shackell, N. L., & Choi, J. S. (2006). Reconciling differences in trophic  
716 control in mid-latitude marine ecosystems: Reconciling differences in trophic control.  
717 *Ecology Letters*, 9(10), 1096–1105. <https://doi.org/10.1111/j.1461-0248.2006.00961.x>

718 Grebmeier, J. M., Feder, H. M., & McRoy, C. P. (1989). Pelagic-benthic coupling on the shelf of  
719 the northern Bering and Chukchi Seas. II. Benthic community structure. *Marine Ecology.*  
720 *Progress Series (Halstenbek)*, 51(3), 253–268.

721 Hurley, P. C. F., & Campana, S. E. (1989). Distribution and Abundance of Haddock (  
722 *Melanogrammus aeglefinus*) and Atlantic Cod (*Gadus morhua*) Eggs and Larvae in the  
723 Waters off Southwest Nova Scotia. *Canadian Journal of Fisheries and Aquatic Sciences*,  
724 46(S1), s103–s112. <https://doi.org/10.1139/f89-282>

725 Hutchings, J. A. (2011). Old wine in new bottles: Reaction norms in salmonid fishes. *Heredity*,  
726 106(3), 421–437. <https://doi.org/10.1038/hdy.2010.166>

727 Iles, T. D., & Sinclair, M. (1982). Atlantic Herring: Stock Discreteness and Abundance. *Science*,  
728 215(4533), 627–633.

729 Kearney, M., & Porter, W. (2009). Mechanistic niche modelling: Combining physiological and  
730 spatial data to predict species' ranges. *Ecology Letters*, 12(4), 334–350.  
731 <https://doi.org/10.1111/j.1461-0248.2008.01277.x>



- 732 Kendall, A. W., & Duker, G. J. (1998). The development of recruitment fisheries oceanography  
733 in the United States. *Fisheries Oceanography*, 7(2), 69–88.  
734 <https://doi.org/10.1046/j.1365-2419.1998.00056.x>
- 735 Kleisner, K. M., Fogarty, M. J., McGee, S., Hare, J. A., Moret, S., Perretti, C. T., & Saba, V. S.  
736 (2017). Marine species distribution shifts on the U.S. Northeast Continental Shelf under  
737 continued ocean warming. *Progress in Oceanography*, 153, 24–36.  
738 <https://doi.org/10.1016/j.pocean.2017.04.001>
- 739 Ladd, C. (2014). Seasonal and interannual variability of the Bering Slope Current. *Deep Sea*  
740 *Research Part II: Topical Studies in Oceanography*, 109, 5–13.  
741 <https://doi.org/10.1016/j.dsr2.2013.12.005>
- 742 Langangen, Ø., Färber, L., Stige, L. C., Diekert, F. K., Barth, J. M. I., Matschiner, M., Berg, P.  
743 R., Star, B., Stenseth, N. C., Jentoft, S., & Durant, J. M. (2019). Ticket to spawn:  
744 Combining economic and genetic data to evaluate the effect of climate and demographic  
745 structure on spawning distribution in Atlantic cod. *Global Change Biology*, 25(1), 134–  
746 143. <https://doi.org/10.1111/gcb.14474>
- 747 Laurel, B. J., & Rogers, L. A. (2020). Loss of spawning habitat and prerecruits of Pacific cod  
748 during a Gulf of Alaska heatwave. *Canadian Journal of Fisheries and Aquatic Sciences*,  
749 77(4), 644–650. <https://doi.org/10.1139/cjfas-2019-0238>
- 750 Lauth, R., Dawson, E. J., & Conner, J. (2019). *Results of the 2017 eastern and northern Bering*  
751 *Sea continental shelf bottom trawl survey of groundfish and invertebrate fauna* (NOAA  
752 Tech. Memo NMFS-AFSC-396; p. 260).  
753 [file:///Users/lciannel/Downloads/noaa\\_20734\\_DS1-1.pdf](file:///Users/lciannel/Downloads/noaa_20734_DS1-1.pdf)
- 754 Livingston, P. (2000). Fisheries in boreal ecosystems. *ICES Journal of Marine Science*, 57(3),  
755 619–627. <https://doi.org/10.1006/jmsc.2000.0728>

- 756 Lough, R., Hannah, C., Berrien, P., Brickman, D., Loder, J., & Quinlan, J. (2006). Spawning  
757 pattern variability and its effect on retention, larval growth and recruitment in Georges  
758 Bank cod and haddock. *Marine Ecology Progress Series*, 310, 193–212.  
759 <https://doi.org/10.3354/meps310193>
- 760 Lough, R., Valentine, P., Potter, D., Auditore, P., Bolz, G., Neilson, J., & Perry, R. (1989).  
761 Ecology and distribution of juvenile cod and haddock in relation to sediment type and  
762 bottom currents on eastern Georges Bank. *Marine Ecology Progress Series*, 56, 1–12.  
763 <https://doi.org/10.3354/meps056001>
- 764 McConnaughey, R. A., & Smith, K. R. (2000). Associations between flatfish abundance and  
765 surficial sediments in the eastern Bering Sea. *Canadian Journal of Fisheries and Aquatic  
766 Sciences*, 57(12), 2410–2419.
- 767 Merilä, J., & Hendry, A. P. (2014). Climate change, adaptation, and phenotypic plasticity: The  
768 problem and the evidence. *Evolutionary Applications*, 7(1), 1–14.  
769 <https://doi.org/10.1111/eva.12137>
- 770 Mindel, B. L., Webb, T. J., Neat, F. C., & Blanchard, J. L. (2016). A trait-based metric sheds new  
771 light on the nature of the body size–depth relationship in the deep sea. *Journal of Animal  
772 Ecology*, 85(2), 427–436. <https://doi.org/10.1111/1365-2656.12471>
- 773 Muhling, B., Reglero, P., Ciannelli, L., Alvarez-Berastegui, D., Alemany, F., Lamkin, J., &  
774 Roffer, M. (2013). Comparison between environmental characteristics of larval bluefin  
775 tuna *Thunnus thynnus* habitat in the Gulf of Mexico and western Mediterranean Sea.  
776 *Marine Ecology Progress Series*, 486, 257–276. <https://doi.org/10.3354/meps10397>
- 777 Neidetcher, S. K., Hurst, T. P., Ciannelli, L., & Logerwell, E. A. (2014). Spawning phenology  
778 and geography of Aleutian Islands and eastern Bering Sea Pacific cod (*Gadus*

779 macrocephalus). *Deep Sea Research Part II: Topical Studies in Oceanography*, 109, 204–  
780 214. <https://doi.org/10.1016/j.dsr2.2013.12.006>

781 Nichol, D. G., Kotwicki, S., Wilderbuer, T. K., Lauth, R. R., & Ianelli, J. N. (2019). Availability  
782 of yellowfin sole *Limanda aspera* to the eastern Bering Sea trawl survey and its effect on  
783 estimates of survey biomass. *Fisheries Research*, 211, 319–330.  
784 <https://doi.org/10.1016/j.fishres.2018.11.017>

785 Nye, J., Link, J., Hare, J., & Overholtz, W. (2009). Changing spatial distribution of fish stocks in  
786 relation to climate and population size on the Northeast United States continental shelf.  
787 *Marine Ecology Progress Series*, 393, 111–129. <https://doi.org/10.3354/meps08220>

788 O’Boyle, R. N., Sinclair, M., Conover, R. J., Mann, K. H., & Kohler, A. C. (1984). Temporal and  
789 spatial distribution of ichthyoplankton communities of the Scotian Shelf in relation to  
790 biological, hydrological, and physiographic features. *Rapp. P.-v. Réun. Cons. Int. Explor.*  
791 *Mer*, 183, 27–40.

792 Opdal, A. F. (2010). Fisheries change spawning ground distribution of northeast Arctic cod.  
793 *Biology Letters*, 6(2), 261–264. <https://doi.org/10.1098/rsbl.2009.0789>

794 Ottersen, G., Bogstad, B., Yaragina, N. A., Stige, L. C., Vikebø, F. B., & Dalpadado, P. (2014).  
795 A review of early life history dynamics of Barents Sea cod (*Gadus morhua*). *ICES*  
796 *Journal of Marine Science*, 71(8), 2064–2087. <https://doi.org/10.1093/icesjms/fsu037>

797 Perry, A. L. (2005). Climate Change and Distribution Shifts in Marine Fishes. *Science*,  
798 308(5730), 1912–1915. <https://doi.org/10.1126/science.1111322>

799 Petitgas, P., Rijnsdorp, A. D., Dickey-Collas, M., Engelhard, G. H., Peck, M. A., Pinnegar, J. K.,  
800 Drinkwater, K., Huret, M., & Nash, R. D. M. (2013). Impacts of climate change on the  
801 complex life cycles of fish. *Fisheries Oceanography*, 22(2), 121–139.  
802 <https://doi.org/10.1111/fog.12010>

803 Pinsky, M. L., Selden, R. L., & Kitchel, Z. J. (2020). Climate-Driven Shifts in Marine Species  
804 Ranges: Scaling from Organisms to Communities. *Annual Review of Marine Science*,  
805 12(1), 153–179. <https://doi.org/10.1146/annurev-marine-010419-010916>

806 Pinsky, M. L., Worm, B., Fogarty, M. J., Sarmiento, J. L., & Levin, S. A. (2013). Marine Taxa  
807 Track Local Climate Velocities. *Science*, 341(6151), 1239–1242.  
808 <https://doi.org/10.1126/science.1239352>

809 Poloczanska, E. S., Brown, C. J., Sydeman, W. J., Kiessling, W., Schoeman, D. S., Moore, P. J.,  
810 Brander, K., Bruno, J. F., Buckley, L. B., Burrows, M. T., Duarte, C. M., Halpern, B. S.,  
811 Holding, J., Kappel, C. V., O’Connor, M. I., Pandolfi, J. M., Parmesan, C., Schwing, F.,  
812 Thompson, S. A., & Richardson, A. J. (2013). Global imprint of climate change on  
813 marine life. *Nature Climate Change*, 3(10), 919–925.  
814 <https://doi.org/10.1038/nclimate1958>

815 Porter, S. M., & Ciannelli, L. (2018). Effect of temperature on Flathead Sole (*Hippoglossoides*  
816 *elassodon*) spawning in the southeastern Bering Sea during warm and cold years. *Journal*  
817 *of Sea Research*, 141, 26–36. <https://doi.org/10.1016/j.seares.2018.08.003>

818 Reed, D., Plourde, S., Cook, A., Pepin, P., Casault, B., Lehoux, C., & Johnson, C. (2018).  
819 Response of Scotian Shelf silver hake (*Merluccius bilinearis*) to environmental  
820 variability. *Fisheries Oceanography*. <https://doi.org/10.1111/fog.12406>

821 Reppert, S. M., & de Roode, J. C. (2018). Demystifying Monarch Butterfly Migration. *Current*  
822 *Biology*, 28(17), R1009–R1022. <https://doi.org/10.1016/j.cub.2018.02.067>

823 Rogers, L. A., Griffin, R., Young, T., Fuller, E., St. Martin, K., & Pinsky, M. L. (2019). Shifting  
824 habitats expose fishing communities to risk under climate change. *Nature Climate*  
825 *Change*, 9(7), 512–516. <https://doi.org/10.1038/s41558-019-0503-z>

826 Rose, G. A. (1993). Cod spawning on a migration highway in the north-west Atlantic. *Nature*,  
827 366(6454), 458–461. <https://doi.org/10.1038/366458a0>

828 Saba, V. S., Griffies, S. M., Anderson, W. G., Winton, M., Alexander, M. A., Delworth, T. L.,  
829 Hare, J. A., Harrison, M. J., Rosati, A., Vecchi, G. A., & Zhang, R. (2016). Enhanced  
830 warming of the North West Atlantic Ocean under climate change. *Journal of*  
831 *Geophysical Research: Oceans*, 121(1), 118–132. <https://doi.org/10.1002/2015JC011346>

832 Scheiffarth, G., Wahls, S., Ketzenberg, C., & Exo, K.-M. (2002). Spring migration strategies of  
833 two populations of bar-tailed godwits, *Limosa lapponica*, in the Wadden Sea: Time  
834 minimizers or energy minimizers? *Oikos*, 96(2), 346–354. <https://doi.org/10.1034/j.1600->  
835 0706.2002.960216.x

836 Schoener, T. W. (2011). The Newest Synthesis: Understanding the Interplay of Evolutionary and  
837 Ecological Dynamics. *Science*, 331(6016), 426–429.  
838 <https://doi.org/10.1126/science.1193954>

839 Scott, J. S. (1982a). Depth, Temperature and Salinity Preferences of Common Fishes of the  
840 Scotian Shelf. *Journal of Northwest Atlantic Fishery Science*, 3, 29–39.  
841 <https://doi.org/10.2960/J.v3.a2>

842 Scott, J. S. (1982b). Selection of Bottom Type by Groundfishes of the Scotian Shelf. *Canadian*  
843 *Journal of Fisheries and Aquatic Sciences*, 39(7), 943–947. <https://doi.org/10.1139/f82->  
844 128

845 Selden, R. L., Thorson, J. T., Samhour, J. F., Bograd, S. J., Brodie, S., Carroll, G., Haltuch, M.  
846 A., Hazen, E. L., Holsman, K. K., Pinsky, M. L., Tolimieri, N., & Willis-Norton, E.  
847 (2020). Coupled changes in biomass and distribution drive trends in availability of fish  
848 stocks to US West Coast ports. *ICES Journal of Marine Science*, 77(1), 188–199.  
849 <https://doi.org/10.1093/icesjms/fsz211>

850 Shackell, N. L., Frank, K. T., Petrie, B., Brickman, D., & Shore, J. (1999). Dispersal of early life  
851 stage haddock (*Melanogrammus aeglefinus*) as inferred from the spatial distribution and  
852 variability in length-at-age of juveniles. *Canadian Journal of Fisheries and Aquatic  
853 Sciences*, 56(12), 2350–2361.

854 Shimada, A. M., & Kimura, D. K. (1984). *Seasonal movements of Pacific cod, Gadus  
855 macrocephalus, in the eastern Bering Sea and adjacent waters based on tag-recapture  
856 data*. 17.

857 Smith, W., & Morse, W. (1985). Retention of larval haddock *Melanogrammus aeglefinus* in the  
858 Georges Bank region, a gyre-influenced spawning area. *Marine Ecology Progress Series*,  
859 24, 1–13. <https://doi.org/10.3354/meps024001>

860 Sohn, D., Ciannelli, L., & Duffy-Anderson, J. T. (2016). Distribution of early life Pacific halibut  
861 and comparison with Greenland halibut in the eastern Bering Sea. *Journal of Sea  
862 Research*, 107, 31–42. <https://doi.org/10.1016/j.seares.2015.09.001>

863 Stevenson, D. E., & Lauth, R. R. (2019). Bottom trawl surveys in the northern Bering Sea  
864 indicate recent shifts in the distribution of marine species. *Polar Biology*, 42(2), 407–421.  
865 <https://doi.org/10.1007/s00300-018-2431-1>

866 Stige, L. C., Rogers, L. A., Neuheimer, A. B., Hunsicker, M. E., Yaragina, N. A., Ottersen, G.,  
867 Ciannelli, L., Langangen, Ø., & Durant, J. M. (2019). Density- and size-dependent  
868 mortality in fish early life stages. *Fish and Fisheries*, 20(5), 962–976.  
869 <https://doi.org/10.1111/faf.12391>

870 Stuart-Smith, R. D., Mellin, C., Bates, A. E., & Edgar, G. J. (2021). Habitat loss and range shifts  
871 contribute to ecological generalization among reef fishes. *Nature Ecology & Evolution*.  
872 <https://doi.org/10.1038/s41559-020-01342-7>

873 Sundby, S., & Nakken, O. (2008). Spatial shifts in spawning habitats of Arcto-Norwegian cod  
874 related to multidecadal climate oscillations and climate change. *ICES Journal of Marine*  
875 *Science*, 65(6), 953–962. <https://doi.org/10.1093/icesjms/fsn085>

876 Swartzman, G., Huang, C., & Kaluzny, S. (1992). Spatial Analysis of Bering Sea Groundfish  
877 Survey Data Using Generalized Additive Models. *Canadian Journal of Fisheries and*  
878 *Aquatic Sciences*, 49(7), 1366–1378. <https://doi.org/10.1139/f92-152>

879 Thorson, J. T., Ciannelli, L., & Litzow, M. A. (2020). Defining indices of ecosystem variability  
880 using biological samples of fish communities: A generalization of empirical orthogonal  
881 functions. *Progress in Oceanography*, 181, 102244.  
882 <https://doi.org/10.1016/j.pocean.2019.102244>

883 Thorson, J. T., Pinsky, M. L., & Ward, E. J. (2016). Model-based inference for estimating shifts  
884 in species distribution, area occupied and centre of gravity. *Methods in Ecology and*  
885 *Evolution*, 7(8), 990–1002. <https://doi.org/10.1111/2041-210X.12567>

886 Waiwood, K. G., & Buzeta, M.-I. (1989). Reproductive Biology of Southwest Scotian Shelf  
887 Haddock ( *Melanogrammus aeglefinus* ). *Canadian Journal of Fisheries and Aquatic*  
888 *Sciences*, 46(S1), s153–s170. <https://doi.org/10.1139/f89-286>

889 Wilderbuer, T. K., Walters, G. E., & Bakkala, R. G. (1992). Yellowfin Sole, *Pleuronectes asper*,  
890 of the Eastern Bering Sea: Biological Characteristics, History of Exploitation, and  
891 Management. *Marine Fisheries Review*, 54(4), 18.

892 Wisz, M. S., Broennimann, O., Grønkjær, P., Møller, P. R., Olsen, S. M., Swingedouw, D.,  
893 Hedeholm, R. B., Nielsen, E. E., Guisan, A., & Pellissier, L. (2015). Arctic warming will  
894 promote Atlantic–Pacific fish interchange. *Nature Climate Change*, 5(3), 261–265.  
895 <https://doi.org/10.1038/nclimate2500>

896 Wood, S. N. (2017). *Generalized Additive Models: An Introduction with R* (Second edition).  
897 Taylor & Francis Group. <https://doi.org/10.1201/9781315370279>  
898



899  
900 Table 1. List of case studies examined. NOAA: National Oceanic and Atmospheric  
901 Administration; DFO: Department of Fisheries and Ocean; IMR: Institute of Marine Research,  
902 VNIRO: Russian Federal Institute of Fisheries and Oceanography. For additional information  
903 about sample sizes, years, and months included in each case study, see Appendix S3 (Table S1).  
904 Ich= Ichthyoplankton survey, BT = bottom trawl survey, MT = midwater trawl.

Common and scientific names	Area	Max size or age	Survey type	Stages available	Source
Walleye pollock ( <i>Gadus chalcogrammus</i> , Gadidae)	Eastern Bering Sea	910 mm	Ich, BT	Eggs, larvae, juveniles, adults	NOAA
Flathead sole (FH, <i>Hippoglossoides elassodon</i> , Pleuronectidae)	Eastern Bering Sea	650 mm	Ich, BT	Eggs, larvae, juveniles, adults	NOAA
Arrowtooth flounder (ATF, <i>Atheresthes stomias</i> , Pleuronectidae)	Eastern Bering Sea	980 mm	BT	Juveniles, adults	NOAA
Pacific cod ( <i>Gadus macrocephalus</i> , Gadidae)	Eastern Bering Sea	1170 mm	BT	Juveniles, adults	NOAA

Alaska plaice (AP, <i>Pleuronectes</i> <i>quadrituberculatus</i> , Pleuronectidae)	Eastern Bering Sea	720 mm	BT	Eggs, larvae†, juveniles, adults	NOAA
Yellowfin sole (YFS, <i>Limanda aspera</i> , Pleuronectidae)	Eastern Bering Sea	700 mm	BT	Eggs†, larvae, juveniles, adults	NOAA
Atlantic cod ( <i>Gadus morhua</i> , Gadidae)	Barents Sea	15 years	Ich, MT, BT	Eggs, larvae, juveniles, adults	IMR, VNIRO
Atlantic cod ( <i>Gadus morhua</i> , Gadidae)	Scotian Shelf	15 years	Ich, BT	Larvae†, juveniles†, adults	DFO
Silver hake ( <i>Merluccius bilinearis</i> , Gadidae)	Scotian Shelf	15 years	Ich, BT	Larvae, juveniles†, adults	DFO
Haddock ( <i>Melanogrammus</i> <i>aeglefinus</i> , Gadidae)	Scotian Shelf	15 years	Ich, BT	Larvae†, juveniles†, adults	DFO

905 (+) Indicates stages that were not included in the analyses of habitat constraints vs ontogenetic  
906 index (Equations 6 and 7) due to a small sample size.  
907

908 Table 2. Results of the GAM models for the analyses of habitat constraints versus ontogenetic  
 909 index. Model results are shown for two different formulations, including the first in which  
 910 population is a random term ( $\delta$ , Eq. 6), and the second with population as a fixed effect (Eq. 7).  
 911 The values reported for the intercept terms ( $\beta$ ) indicate their estimated mean, and the values  
 912 reported for the smooth terms ( $g(O)$ ) indicate the estimated degrees of freedom.  $I$  is in indicator  
 913 variable (0,1) specifying the species/population considered ( $s$ ). For the fixed effect model, species  
 914 are sorted in decreasing order of their p-value for the smooth term. Signif. codes: ‘\*\*\*\*’ <0.001,  
 915 ‘\*\*\*’ < 0.01, ‘\*\*’ < 0.05, ‘.’ < 0.1. YFS: Yellowfin sole. ATF: Arrowtooth flounder

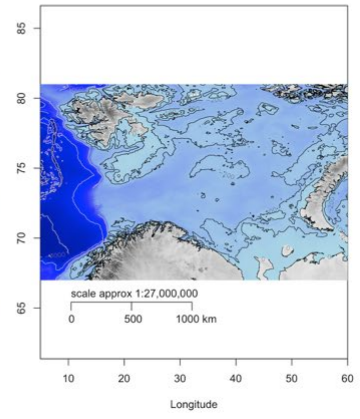
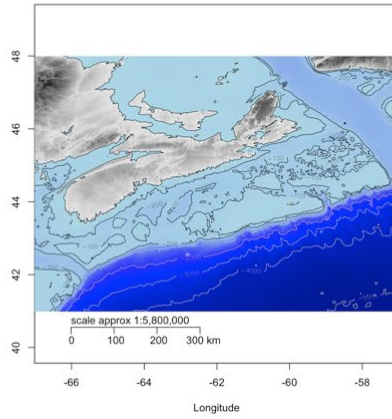
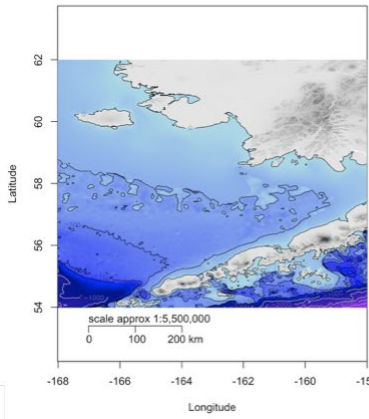
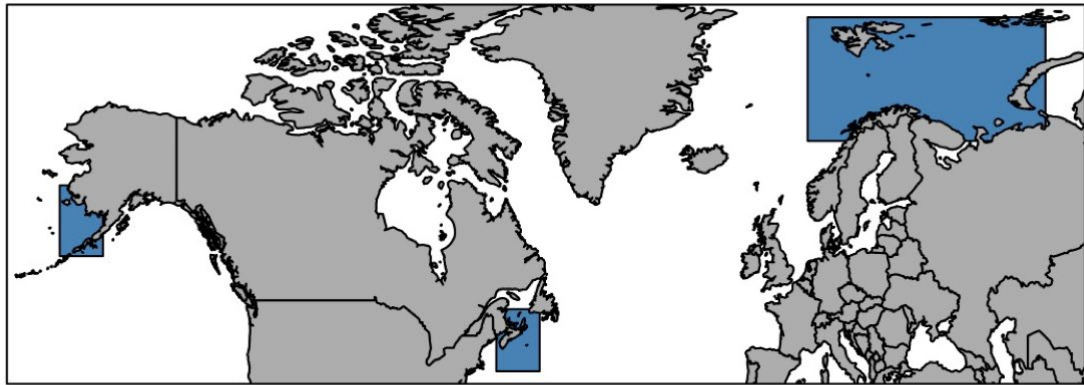
<b>Mixed effect model</b>	$HC_a = \beta + g(O_a) + \delta_s + \varepsilon_a$		
	$\beta$	$g(O_a)$	$\delta_s$
	Adjusted $R^2 = 0.673$ , Dev = 71.0%, n = 86		
Estimate	0.201 ***	1.000 ***	
Variance component		$4.05 \times 10^{-5}$	$8.12 \times 10^{-2}$
<b>Fixed effect model</b>	$HC_{s,a} = \beta * I_s + g(O_{s,a}) * I_s + \varepsilon_a$		
	$\beta * I_s$	$g(O_{s,a}) * I_s$	
	$R^2 = 0.831$ , Dev = 97.7%, n = 86		
ATF	0.217 ***	1.691	
Silver hake	0.249 ***	1.289	
Pacific cod	0.065 ***	1.299	
Scotian cod	0.153 ***	1.831 .	
Barents cod	0.129 ***	1.000 *	
Haddock	0.244 ***	1.000 *	

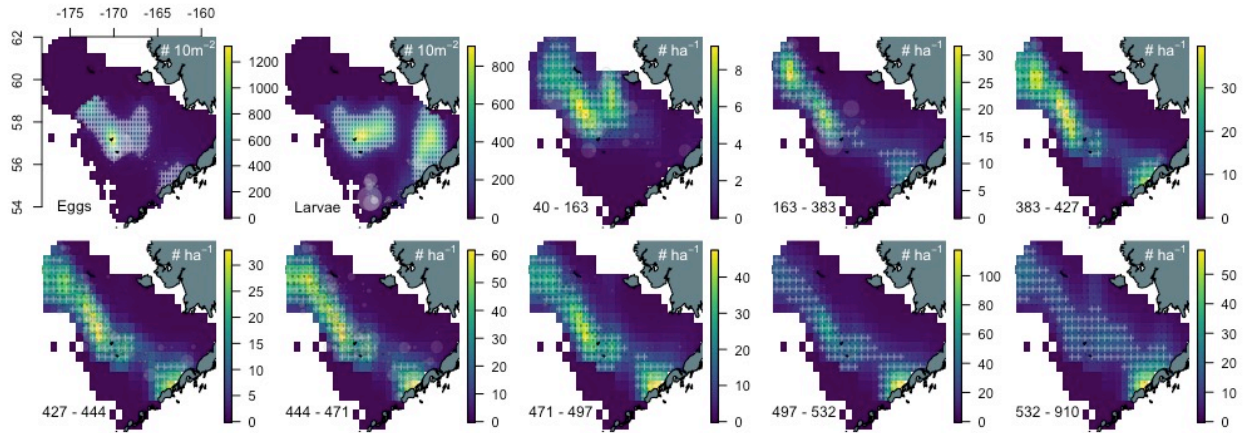
Pollock	0.111 ***	1.710 *
Alaska plaice	0.279 ***	1.591 ***
Flathead sole	0.281 ***	1.970 ***
YFS	0.370 ***	1.983 ***

916

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923

924 **Figure 2.** Spatial distribution of walleye pollock in the Bering Sea over different ontogenetic

925 stages and size groups. Predictions are shown as color image in the units of number of

926 individuals  $10\text{m}^{-2}$  (eggs and larvae), or numbers of individuals per hectare ( $\text{ha}^{-1}$ ) for all other size

927 groups. Predictions were obtained from presence/absence and presence only GAM models (delta

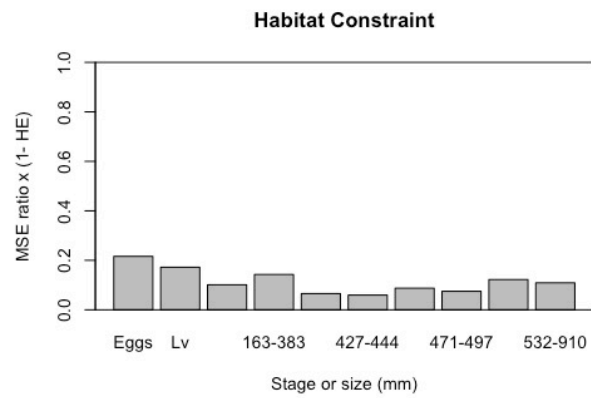
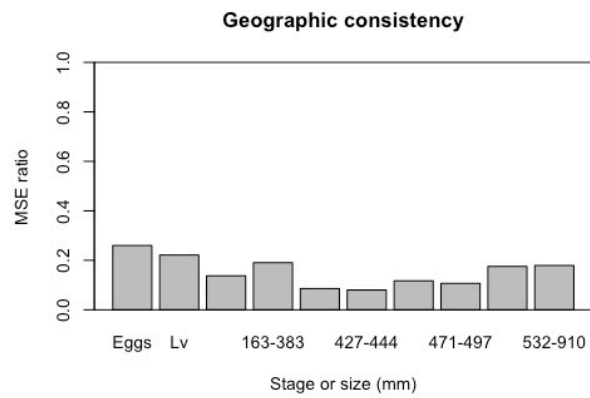
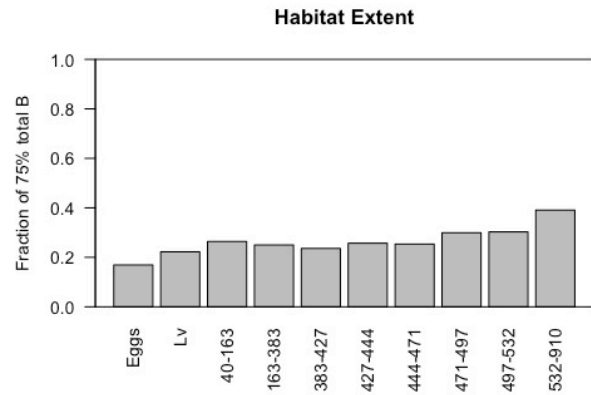
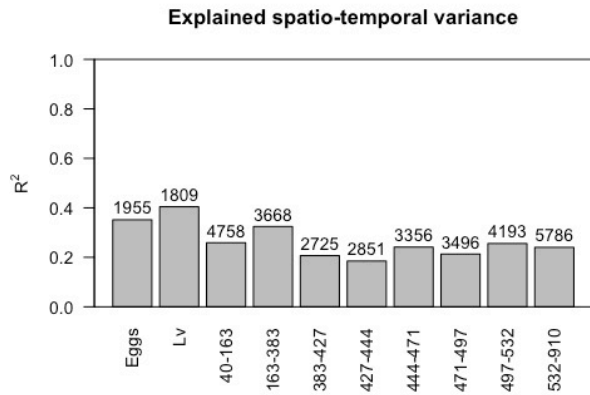
928 method, equation 3). Semitransparent grey bubbles are observed densities and the region

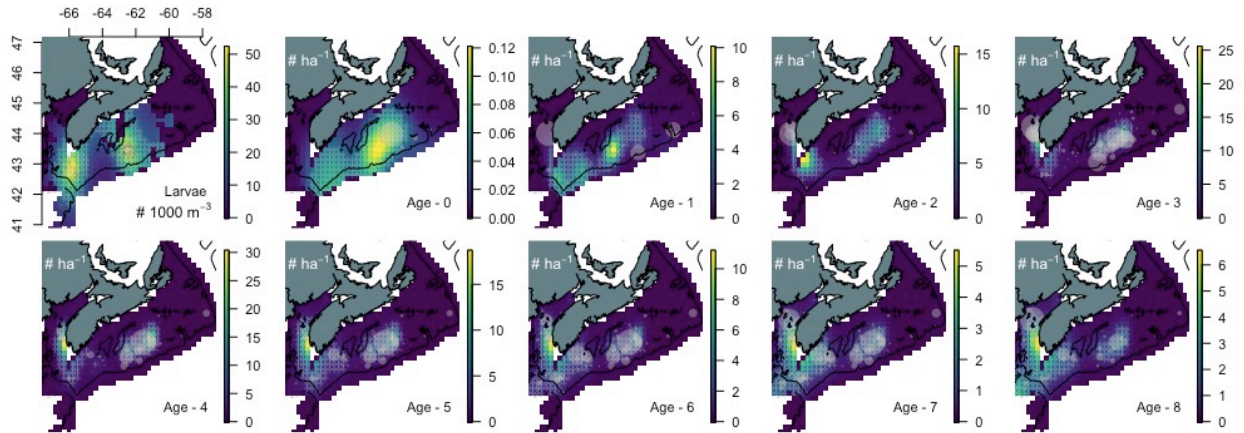
929 delineated by a white cross ('+') represents the estimated habitat extent, or the portion of the

930 surveyed area that contains 75% of the estimated total abundance. The stage or size range (in

931 mm) examined is indicated in the lower left corner of each plot. The 200m depth isobath is

932 shown.





943

944 **Figure 4.** Spatial distribution of haddock in the Scotian Shelf over different ontogenetic stages

945 and age groups. Predictions are shown as color image in the units of number of individuals  $m^{-3}$

946 (larvae), or numbers of individuals per hectare ( $ha^{-1}$ ) for all other age groups. Predictions were

947 obtained from presence/absence and presence only GAM models (delta method, equation 3).

948 Grey bubbles are observed densities and the region delineated by a white cross ('+') represents

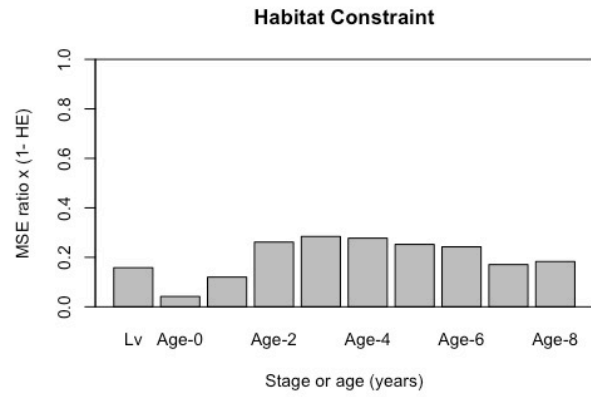
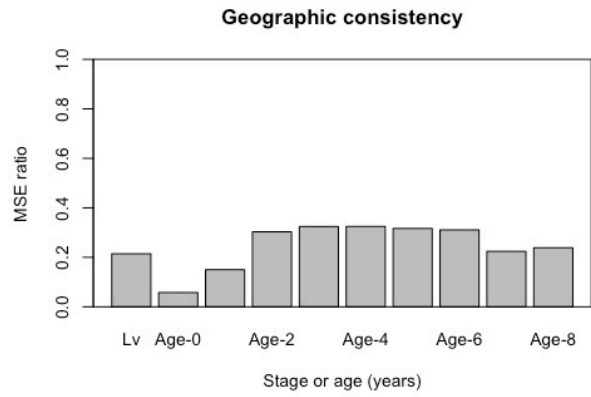
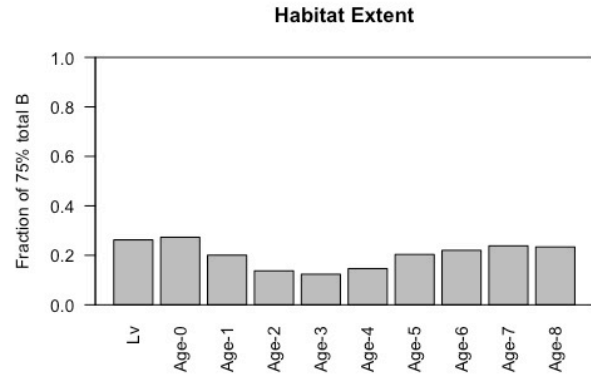
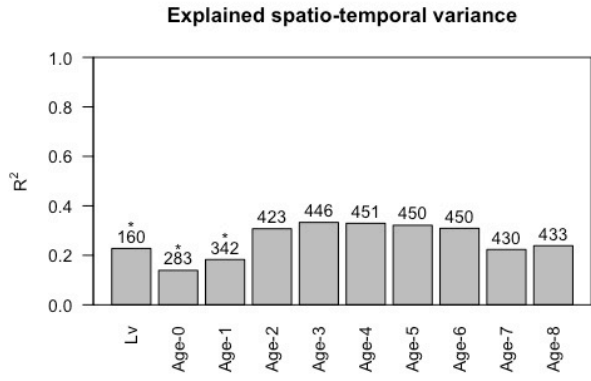
949 the estimated habitat extent, or the portion of the surveyed area that contains 75% of the

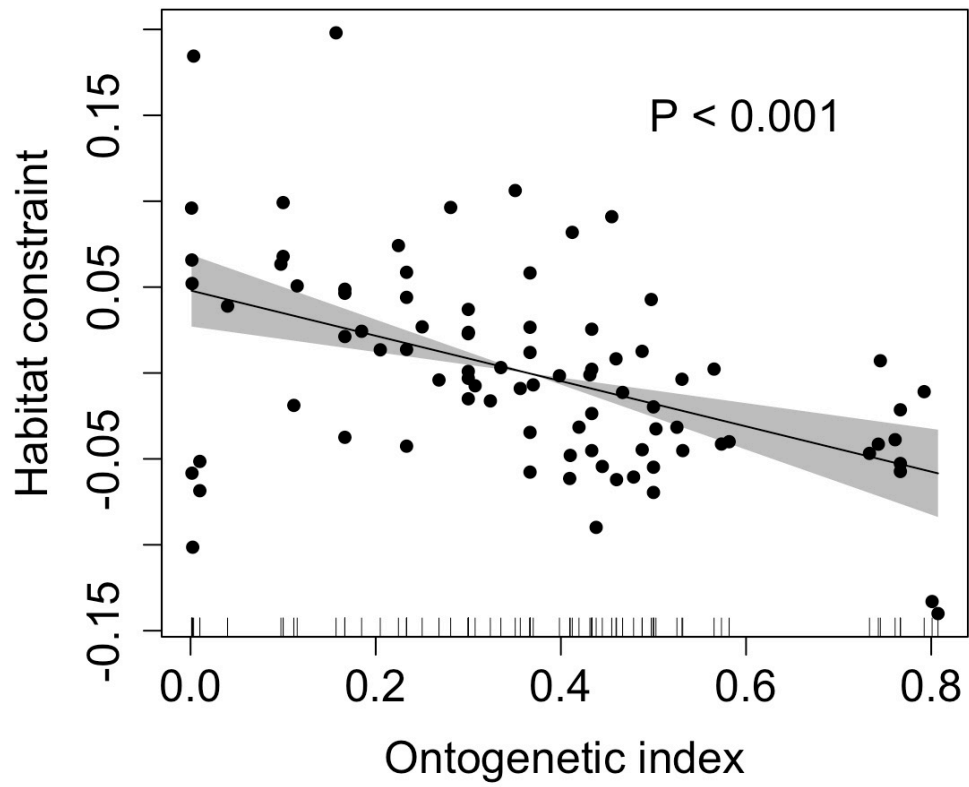
950 estimated total abundance. The stage or age (in years) examined is indicated in the lower left

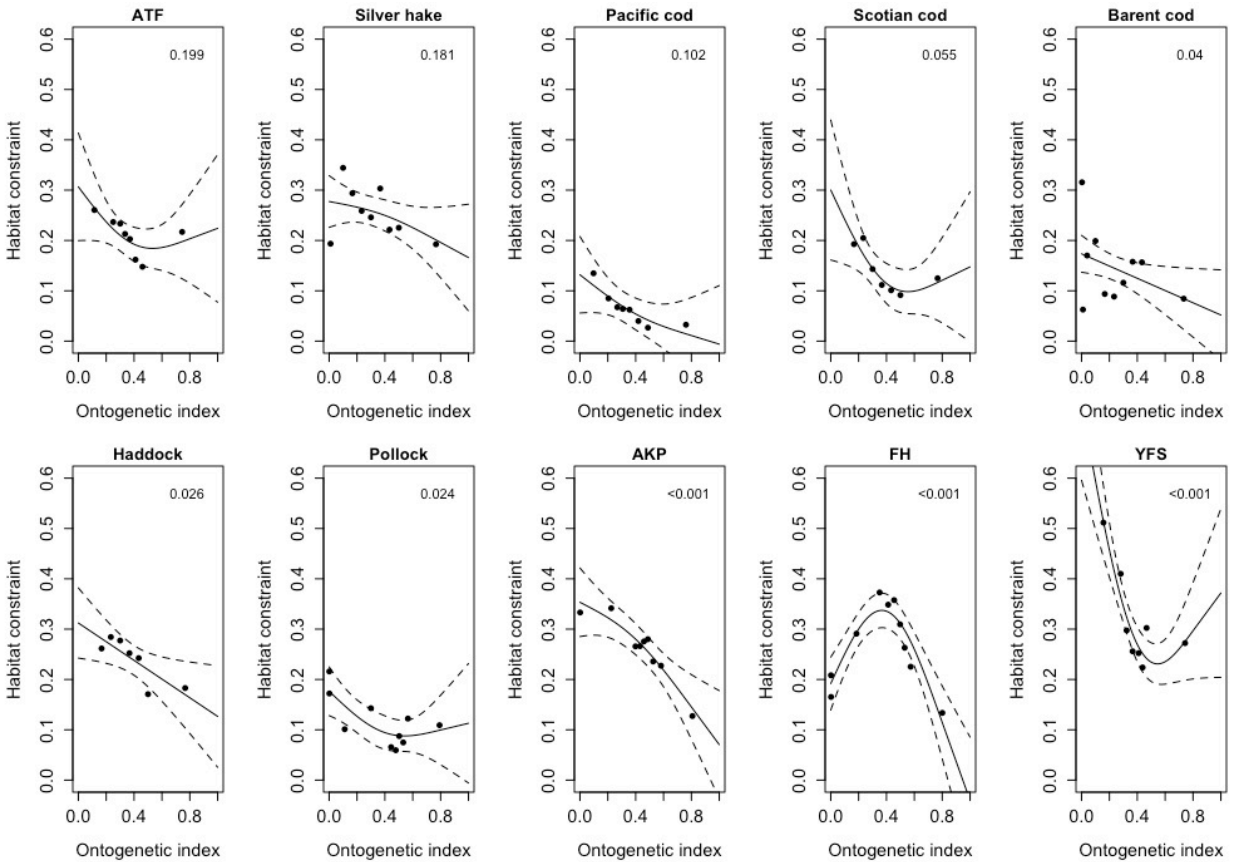
951 corner of each plot. The 200m depth isobaths is shown.

952









972 **Figure 7.** Relationship between the habitat constraint metrics and the ontogenetic index for the  
 973 ten populations examined in this study (Equation 7). The smooth solid lines are average  
 974 relationships determined by a fixed effect GAM between constraint and the linear ontogeny  
 975 index. Dashed lines are 95% confidence intervals. The scale of the y-axis represents the response  
 976 variable. Species are sorted in decreasing order of their p-value for the ontogenetic index (upper  
 977 right corner on each plot). For species abbreviations see Table 1.