1 Distribution and Abundance of Juvenile Demersal Fishes in Relation to

2 Summer Hypoxia and Other Environmental Variables in Coastal

3 Oregon, USA

- 4
- 5 Kathryn L. Sobocinski^{*+1}, Lorenzo Ciannelli¹, W. Waldo Wakefield², Matthew E. Yergey³,
- 6 Angela Johnson-Colegrove¹
- 7
- 8 ¹College of Earth, Ocean, and Atmospheric Sciences
- 9 Oregon State University
- 10 104 CEOAS Administration
- 11 Corvallis, Oregon, 97331, U.S.A.
- 12
- 13 ² Fishery Resource Analysis and Monitoring Division
- 14 Northwest Fisheries Science Center
- 15 National Marine Fisheries Service
- 16 National Oceanic and Atmospheric Administration
- 17 2032 SE OSU Drive
- 18 Newport, Oregon 97365, U.S.A.
- 19
- 20 ³ Pacific States Marine Fisheries Commission
- 21 2032 SE OSU Drive
- 22 Newport, Oregon 97365, U.S.A.
- 23
- 24 *Corresponding Author, kathryn.sobocinski@noaa.gov
- 25 +Current Address:
- 26 Kathryn L. Sobocinski
- 27 NOAA-Affiliate, Long Live the Kings
- 28 Fish Ecology Division
- 29 NOAA Fisheries Northwest Fisheries Science Center
- 30 2725 Montlake Blvd. E
- 31 Seattle, WA 98112
- 32 (206) 302-2466
- 33
- 34

35 Abstract

36 The juvenile demersal fish assemblage along the Pacific Northwest coast has received little 37 attention relative to adult life history stages since pioneering work in the 1970s. Increasing 38 severity of hypoxia along the Oregon coast in recent years has prompted investigations into 39 the response of biota in this region. We used summer data (2008 to 2013) from a beam 40 trawl survey targeting juvenile demersal fishes in soft-bottom habitats along the Oregon coast to describe patterns of distribution and abundance at fixed sampling stations (from 41 42 30m to 100m depth). We relate the assemblage and abundance of the common species to 43 environmental variables and analyze condition of recently settled fish (<50 mm SL). Most 44 of the captured fishes were young-of-the-year flatfishes, dominated by Butter Sole 45 (Isopsetta isolepis), English Sole (Parophrys vetulus), Speckled Sanddab (Citharichthys stigmaeus), and Pacific Sanddab (Citharichthys sordidus). Community analysis of the full 46 47 dataset showed some variation in species richness among years and high evenness across 48 all sampling sites and years. Depth was a structuring variable for the community, indicated 49 by multivariate nonmetric multidimensional scaling analysis. Generalized additive models 50 for common flatfish species abundances during the summer months indicated depth preferences, with Butter Sole, English Sole, and Speckled Sanddab at shallower locations 51 and Pacific Sanddab occurring at deeper locations farther offshore. Additionally, while 52 53 most common species were collected in high abundances in hypoxic conditions, dissolved 54 oxygen was a significant factor in determining flatfish abundance. Condition factor was 55 weakly negatively impacted by low dissolved oxygen for the species assessed, but the strength of the relationship varied by species. Increased sampling frequency and spatial 56 57 coverage would improve our understanding of this community, especially in light of 58 changing environmental drivers such as decreasing pH, warming water, and episodic 59 periods of low dissolved oxygen coinciding with settlement for many species.

60

61 Keywords:

62 Demersal fishes, hypoxia, flatfishes, nearshore, dissolved oxygen, condition factor, Oregon

- 63
- 64

65 **1. Introduction**

66 Shallow, coastal waters are nursery areas for many fish and invertebrate species. The use of nursery habitats—those that afford favorable physiological conditions, ample 67 food supply, and protection from predators—optimizes the trade-off between growth and 68 69 mortality for juvenile fishes (Sogard, 1997; Beck et al., 2001). Along the Pacific Northwest 70 coast, nursery habitats for many fish species tend to be associated with the coastal shelf, in 71 depths of less than 100m. While some species along this coast use estuaries as juvenile 72 habitats (Krygier and Pearcy, 1986; Gunderson et al., 1990; Rooper et al., 2003; Hughes et 73 al. 2015), many other species remain in coastal waters where they settle, some changing 74 depth with ontogeny (Gunderson et al., 1990). Understanding distribution and abundance 75 patterns, movement across habitats, and juvenile fish response to environmental 76 conditions is important for management of these species, especially within an ecosystem 77 context (Pikitch at al., 2004; NMFS 2013). Also, because year-class strength can be 78 determined by environmental factors (Houde, 2002; Thannesekos et al., 2016) 79 understanding early life histories of fishes can aid in population assessment and 80 management. In this study, we evaluate the depth distribution and response to 81 environmental variables, specifically dissolved oxygen, for a community of fishes in near-82 coastal soft-bottom nursery grounds.

The dominant geomorphology along the Oregon coastal shelf is unconsolidated softsediments (Romsos et al., 2007). While lesser studied than rocky reef habitats, which are also important features in this region (Pearcy et al., 1989; Tissot et al., 2007; 2008; Donnellan et al., 2009), this substratum provides habitat for a number of demersal fish and invertebrate species (Jay, 1996; Keller et al., 2010; Toole et al., 2011), many of which are of

88 commercial importance (Miller et al., 2006; Yoklavich and Wakefield, 2015; NOAA 89 Commercial Landings Database¹). A predominant component of the demersal fish 90 community is flatfishes of the families Pleuronectidae and Paralichthyidae (Toole et al., 91 2011). Despite an abundance of juvenile flatfishes in Oregon coastal waters and the 92 importance of these species to commercial fisheries, little attention has been paid to early 93 life history of these fishes on the open coast since pioneering work in the 1960s and 1970s 94 (but see Toole et al., 2011; in addition to Pearcy, 1964; Pearcy et al., 1977; Richardson and 95 Pearcy, 1977; Pearcy, 1978). In addition to seeking greater understanding of early life 96 history of fishes in this region, juvenile fishes are often vulnerable to variable 97 environmental conditions and predation, and may be good indicators of environmental 98 change in this region.

99 Demersal marine fishes off the Oregon coast spawn primarily during winter months 100 (Shanks and Eckert, 2005; Brodeur et al., 2008) with pelagic larvae captured from February 101 to July (Richardson and Pearcy, 1977; Gadomski and Boehlert, 1984), although there is 102 variation in timing by species, spawning mode, and depth of occurrence (Shanks and 103 Eckert, 2005). Settlement of juvenile flatfishes to benthic habitats occurs primarily during 104 the summer months (Pearcy et al., 1977), although variation exists among species (Krygier 105 and Pearcy, 1986). The current study focuses on newly settled individuals over soft-106 sediments during the summer months from 2008-2013.

107In recent years, periodic hypoxia events (dissolved oxygen <1.4 mL L-1, Grantham et</th>108al., 2004) have occurred during the summer months along the Oregon coast (Grantham et

¹ https://www.st.nmfs.noaa.gov/commercial-fisheries/commercial-landings/index

109 al., 2004; Chan et al., 2008; Adams et al., 2013; Peterson et al., 2013). Particularly severe 110 events occurred in 2002 and 2006, with as much as 60% of the shelf affected by hypoxia 111 (Peterson et al., 2013). These events occur when surface waters are driven offshore by the 112 prevailing northwesterly summer winds, resulting in upwelling of low-oxygen, nutrient-113 rich waters (Chan et al., 2008; Peterson et al., 2013). If the upwelling-producing conditions 114 are strong but interspersed with periods of upwelling relaxation, low-oxygen zones can 115 form in the water column of the coastal shelf, resulting in die-offs of fauna that are not 116 adapted to prolonged periods of hypoxia (Chan et al., 2008). These periodic events appear 117 to be occurring more frequently in recent years (Chan et al., 2008; Peterson et al., 2013) 118 and are of concern for the management of valuable marine fisheries in the region. How 119 early life stages—which are less motile than adults, and thus, more susceptible to *in situ* 120 conditions—are affected by periodic low-oxygen events remains unclear. 121 The ephemeral nature of hypoxic events, occurring with varying intensity and 122 extent during summer months (Peterson et al., 2013), makes describing causal 123 relationships for fauna challenging. However, Keller et al. (2010; 2015) linked decreased 124 fish and invertebrate biomass and species diversity to depressed oxygen levels using data 125 from a regularly-conducted NOAA-Fisheries bottom trawl survey targeting adults in this 126 region. While the NOAA survey has a time series of catch across the shelf from Washington 127 to California, our understanding of the juvenile fish community in this same region is 128 limited in both time and space. To better understand the impacts of hypoxic events on 129 young-of-the-year (YOY) fishes, a beam trawl sampling program was initiated in 2008. This 130 survey targets nearshore soft-sediment areas where hypoxic events have been identified 131 during the summer months (Stinton et al., 2014) and aimed to collect juvenile life stages. In

132 recent years, the survey has expanded temporally, with year-round monthly sampling to 133 better understand flatfish communities in nearshore waters in general (see Yergey et al., in 134 review). The objectives of the current study were to 1.) describe the soft-bottom benthic 135 species assemblage 2.) relate summer distribution and abundance of common species to 136 environmental variables (specifically depth, temperature, and dissolved oxygen), and 3.) 137 investigate linkages between flatfish condition and environmental variables, especially low 138 dissolved oxygen. Juvenile demersal fishes may be especially susceptible to changing ocean 139 conditions along the Oregon coast due to their limited motility. This study draws 140 connections between dissolved oxygen and individual fish condition, abundance of 141 common species, and the fish assemblage in an under-sampled habitat.

142 **2.** Methods

143 **2.1** Fish Sampling

144 Demersal fishes were collected during the summer months from 2008-2013 at fixed 145 stations ranging from 30-100 m along a transect extending from Yaquina Head to the west 146 (known as the Newport Hydrographic Line, or NH Line, ~ 44.66°N). An effort was made to 147 sample all depth strata on each cruise; however, weather and gear did not always allow for 148 complete transects. Fishes were collected using a 2.0 m-wide, 0.5 m-tall beam trawl 149 constructed of 38-mm mesh and lined throughout with 2.5 x 3-mm mesh; a tickler chain 150 was attached to the beam trawl skids forward of the trawl footrope to disturb the substrate 151 and flush fishes into the net. Tows were generally 10-min. in duration, from cable out to the 152 start of the retrieval (assumed to be bottom contact time) and actual time of the tow was 153 noted. The trawl was equipped with a paired odometer wheel system that measured

distance sampled. The distance measured on the wheel was converted to meters and used
as the distance towed. For tows lacking a distance measure (~30%), we generated a linear
model including depth and time towed from other samples collected and imputed the
missing values so that all catch could be standardized to area swept. Vessel speed over
ground was maintained at approximately 1.0 kt for all trawls.

159 Catch was sorted on deck, with fishes \geq 150 mm (standard length, SL) being 160 identified and measured onboard before being released and fishes <150 mm SL placed in 161 storage bags and flash frozen on dry ice for later processing. Samples were stored in a -80°C 162 freezer until processed. In the lab, thawed fish were identified, and lengths and damp 163 weights were taken. All data were entered into a database containing vessel, trip, and 164 sample data. For all analyses, a sample was defined as one net tow at any of the set stations, 165 with correlating fish abundances and environmental variables; the catch within this net 166 tow was standardized to density (# m⁻²) based upon area swept (distance towed x 2 m for 167 the width of the beam trawl).

A CTD cast (Conductivity-Temperature-Depth, Seabird Model 25 or SBE 19 with flow-through dissolved oxygen sensor) was made at the starting point of each tow to measure the salinity, temperature, and bottom dissolved oxygen at each station. Where casts were not made (<25% of all samples), available data on environmental parameters from nearby buoys or other sampling efforts were used to populate the data set.

173 2.2 Data Analysis

The dataset was analyzed to address multiple objectives, using both univariate and multivariate statistical methods. We conducted all analyses in R (R Core Team, 2016). To describe the benthic fish community (including multivariate analysis and description of species richness, diversity, and evenness) the full dataset was retained. Reduced datasets
were used for univariate analyses relating individual species abundances to environmental
variables and for condition factor analysis (see below). Specifically, "YOY" included all
young-of-the-year fishes (<100 mm SL) and "recently settled" included all newly settled
juvenile flatfish (typically <50 mm SL, e.g., Van Cleve and El-Sayed, 1969; Rosenberg, 1982,
for *Parophrys vetulu;* and Rackowski and Pikitch, 1989, for *Citharichthys sordidus* and *C.*stigmaeus).

184 2.2.1 Community

185 To characterize the fish community, we used two approaches. The first was to model 186 ecological response (species richness, diversity, and evenness) as a function of year, month, 187 station/depth, dissolved oxygen, and temperature at the bottom depth using general linear 188 models (function *lm* in R). Species richness (SR) was defined as the number of species in a 189 sample, species diversity (H') was defined as the Shannon–Wiener index, $H' = -sum p_i \ln p_i$, 190 where p_i is the proportional abundance of species *i*, and higher H' values indicate greater 191 diversity (Whittaker, 1972). Sample evenness () was defined as Pielou's evenness, where 192 *J*=*H*′/*SR*, where *H*′ is species diversity as defined above and *SR* is species richness. Evenness 193 values ranged from 0 to 1, with larger values indicating that all occurring taxa were present 194 in the same relative concentrations (Pielou, 1969). We calculated rarefied values of species 195 richness (Heck et al., 1975) to account for differences in sampling effort and used these in 196 addition to species richness values in our models. These responses were normally 197 distributed and as such, general linear models were suitable. Month, station, and year were 198 all treated as factors, while all other variables were continuous. Since depth and station are 199 highly correlated, we only included the factor with the best model fit. All model selection

was done using Akaike's Information Criterion (AIC, Burnham and Anderson, 2002; Zuur etal., 2009).

202 Secondly, we used nonmetric multidimensional scaling (NMDS) to analyze drivers of 203 community structure across the sampling locations and suite of environmental variables. 204 The dataset was comprised of species densities, standardized by area swept. NMDS is an 205 unconstrained ordination technique that maps multidimensional data in reduced 206 dimensional space (2d or 3d) using the rank order of dissimilarity values among samples. A 207 Bray-Curtis distance measure was used to generate the rank order of samples on square 208 root transformed data (Clarke and Warwick, 2001). NMDS solutions were determined in 209 two and three dimensions using multiple random starts to maximize the likelihood of 210 reaching a global minimum stress value. Stress, a goodness-of-fit criterion that measures 211 the discrepancy between the distance in ordination space and the distances of the original 212 data set, was used to determine model fit. Stress values were plotted against the number of 213 dimensions (scree plot), to determine how increased dimensionality and stress interacted; 214 the greatest reduction in stress occurred at 3 dimensions, but 2d solutions are presented 215 here for ease of interpretation. To determine differences in factors of interest (year, month, 216 depth, and dissolved oxygen and temperature at bottom depth), we conducted 217 permutational multivariate analyses of variance (PERMANOVA), a procedure that does not 218 require multivariate normality (Anderson, 2001). We used library "vegan" with function 219 *metaMDS* for NMDS and function *adonis* for PERMANOVA analyses (Oksanen et al., 2015). 220 Within significant groups (e.g., site/depth), we used the indicator species analysis package 221 "indicspecies" to identify important species within each group (De Caceres and Legendre, 222 2009).

223 *2.2.2 Abundance*

224 For total fish abundance and for each of the most abundant flatfish species, we 225 modeled YOY fish abundance (CPUE, catch per unit effort) over the summer season using 226 Generalized Additive Models (GAMs). GAMs allow for nonlinearities in the relationships 227 between response and explanatory variables. Responses (abundance, in this case) can be 228 modeled with both a parametric component (equivalent to generalized linear modeling) 229 and also with a non-parametric component, which relies on smoothing functions for 230 covariates (Wood, 2006; Zuur et al., 2009; Zuur, 2013). The gam function in "mgcv" 231 package in R (R Core Team, 2016) was used for all GAM modeling. The gam function 232 estimates the optimal smoothed relationship in model fitting. Effective degrees of freedom 233 (edf) is a calibration tool to determine the shape of the curve, where a value of 1 indicates a 234 straight line and higher values a progressively non-linear pattern (Zuur, 2013). The 235 smoothers used in this application were thin plate regression splines for all parameters. 236 The models were fitted using the "GCV.Cp" method (Generalized Cross Validation with an 237 unknown scale parameter) and best models were re-estimated by Restricted Maximum 238 Likelihood (REML) to check for stability.

For all abundance models, the response variable was count of a fish species (number per tow) and the predictor variables of interest were: year (2008-2013), bottom depth (m, from ~30m-120m), the hydrographic parameters temperature and dissolved oxygen (DO) at the bottom, and monthly upwelling index. For the upwelling index, we took the average of the monthly value in which sampling occurred from the three closest index stations (42°, 45°, and 48° N, Pacific Fisheries Environmental Laboratory http://www.pfeg.noaa.gov) as an indicator of larger oceanic processes that may influence recruitment and/or distribution

246 of fishes. We used a negative binomial distribution with natural log link in the models to 247 account for the overdispersed nature of the abundance data and we included the natural 248 log of area swept (m⁻²) as an offset to account for variation in tow distance. We also 249 considered the interaction between depth and dissolved oxygen, as exploratory plots 250 showed possible collinearity, although it should be noted that at deeper stations dissolved 251 oxygen was always low (<3 ml/L) with no higher measurements observed during our summer sampling periods. Collinearity among the predictors was assessed with coplots, by 252 253 plotting Pearson residuals (Zuur, 2013), and by analyzing variance inflation factors (VIF) 254 using the *aed* function (Zuur et al., 2009); all variables had VIF scores <4, suggesting 255 collinearity was not strong. The full model form was:

256 Abundance= offset(ln(distance towed)) + Year+ s(Depth) + s(DO) + s(Temp)

257

+ s(Upwell.Index) + ε

(Equation 1)

258 All variables except year, which was treated as a factor, were smoothed (*s*). We constrained 259 the wiggliness of the smoothed terms by setting the knots, k, equal to 4, which sets upper limits for degrees of freedom and aids in preventing model overparameterization. The 260 261 error term (ε) is assumed to be independent and identically distributed (Zuur et al., 2009). 262 The output of all models was assessed using the function *gam.check*, which plots residuals 263 and presents other diagnostic measures. We used AIC for model selection and an 264 assessment of residuals, deviance explained, and the Unbiased Risk Estimator (UBRE), a 265 procedure designed to measure error, provided an additional evaluation of model fit and 266 validity (Zuur 2009; 2013). We present model forms, the amount of deviance explained, 267 and significance levels for variables for the models.

268 2.2.3 Condition Factor

For three of the most common species, Butter Sole (*Isopsetta isolepis*), English Sole
(*Parophrys vetulus*), Speckled Sanddab (*Citharichthys stigmaeus*), we evaluated condition of
newly recruited individuals as a function of environmental variables, to determine
specifically whether low dissolved oxygen resulted in reduced fitness. Individual condition
was evaluated through the use of both Fulton's K and residuals from a length-weight
relationship. Fulton's K is a commonly used metric of fish condition (Gilliers et al., 2004;
Amara et al., 2007; De Raedemaecker et al., 2012):

276 $K=100^{*}(W/L^{3})$

(Equation 2)

where: *W*=fish blotted wet weight and *L*=length in cm. This metric was calculated for each
individual fish for which both length and weight data were available (N=2400, 1426, and
1041 for Butter Sole, English Sole, and Speckled Sanddab, respectively). Fulton's K was
then used as the response variable in our models.

281 Similarly, we used the residuals from a L-W relationship as a response (Schulte-282 Hostedde et al., 2005). To derive the residuals, we constructed GAMs for each of the three 283 species with *Weight~s(Length)*, where the length component was smoothed to allow for 284 non-linear response. Residuals from model fits were extracted and used in multivariable 285 model analysis. For both condition metrics, larger values indicated fish that had higher 286 weight for their length class, suggesting better individual condition. A similar study on 287 offshore adult demersal fishes, found that for several species, body condition was positively 288 correlated with the dissolved oxygen concentration; however, this was a not a universal 289 response (Keller et al., 2010). In our analysis, we selected newly settled (<50 mm SL) 290 individuals on the premise that these organisms would be most responsive to changes in 291 habitat condition, such as with hypoxia, because they are less motile than their larger

292 conspecifics, and would therefore have limited means to avoid unfavorable environmental293 conditions.

294 To relate the response variables (Fulton's K and L-W Residuals) to the explanatory 295 variables (year, temperature, dissolved oxygen, depth, and depth*dissolved oxygen 296 interaction), we used general linear mixed models using the function *lme* in R. We included 297 tow as a random factor in the models to account for similarity of individuals captured at the 298 same time and location. Models were fit using restricted maximum likelihood (REML). AIC 299 was used for model selection and model fit (R^2) for both the fixed and random components 300 were estimated with the function *r.squaredGLMM* using the methods of Nakagawa and 301 Schielzeth (2013).

302 3. Results

We made a total of 135 tows from 2008-2013, although effort was not divided equally among years and not all months/stations were sampled each year due to weather and vessel availability (Table 1). We collected over 80 species of fish from 23 families, in depths from 30 m to 120 m across the six summers of the survey (Table 2 shows species occurring in >1% of tows).

308 3.1 Environmental Parameters

Bottom water temperature ranged from 6.8 -11.8°C, with a mean of 7.8°C, and with deeper sampling sites being the coldest (Figure 1, Table 1). While some variability in temperature existed among years and by month (Figure 2), depth had the most influence on temperature. Similarly, bottom dissolved oxygen decreased with depth, with highest values (5.2 ml/L) in the shallowest sites and lower values (<2 ml/L) observed at all depths, but with deeper sites generally having lower values (Figure 1, Table 1). Both temperature
and dissolved oxygen were most variable at shallow depths. Unlike temperature, dissolved
oxygen levels decreased through the summer months (Figure 2), suggesting that while
temperature and dissolved oxygen may be correlated, these variables vary according to
different processes.

Mean dissolved oxygen across all sampling periods was 2.4 ml/L with 22 of 135 measurements below the 1.4 ml/L hypoxia threshold. Hypoxia occurred at depths from 30 m-125 m and in all summer months, except May. Hypoxic measurements were most numerous (and broadest in depth distribution) in 2012, but were also observed in 2008, 2010, and 2013. Salinity ranged from 32.8 to 33.9 (practical salinity units), with a mean of 33.8. Because of the narrow range of salinities observed, we did not include this variable as a factor in our analyses.

326 3.2 Juvenile Fishes

327 Pleuronectidae and Paralichthyidae (flatfishes) were the two most frequently 328 occurring and abundant families and made up over 88% of the overall catch, with over 329 11,000 individuals collected. The most abundant species were the flatfishes: Butter Sole, 330 English Sole, Speckled Sanddab, Pacific Sanddab (Citharichthys sordidus) and Slender Sole 331 (Lyopsetta exilis). Other abundant species included: Pacific Tomcod (Microgadus proximus), 332 Pacific Sandlance (Ammodytes hexapterus), Warty Poacher (Occella verrucosa), and Pacific 333 Staghorn Sculpin (*Leptocottus armatus*), although these were much less frequently 334 occurring and were collected in <20% of the tows (Table 2). Many species (>50%) were 335 rare, occurring in low abundances and in less than 10% of the samples collected.

336 Over half the fishes collected were <50 mm (53.8%), with 22.7% sized 50-100 mm, 337 and 23.5% >100 mm, demonstrating that the catch was comprised mostly of recently 338 settled and juvenile individuals, as targeted by the gear type (Figure 3). Among the 339 common flatfish species, Butter Sole \sim 35 mm SL were the most abundant (Figure 4). 340 Recently settled (<50 mm SL) English Sole, Speckled Sanddab, and Slender Sole were also captured in high abundance, although Slender Sole were only abundant in 2011 and 2012. 341 342 Pacific Sanddab collected by the beam trawl were slightly larger, around 60 mm SL. 343 Minimum size of fish captured varied by year; however, not all months were sampled in all 344 years, so comparisons of timing and size differences from year to year should be made with 345 caution.

346 **3.3 Community Analysis**

347 Mean species richness was 7 (min=1, max=21) and was variable among years, with 348 the greatest variance in 2013. Evenness and species diversity were less variable among 349 years (Figure 5), and in general, evenness was high, showing the dominance of a few 350 common species. General linear models showed species richness (SR) and evenness (J) to 351 be influenced by year and station; temperature was also a significant variable for SR (these 352 results were the same for raw and rarefied SR values and we present raw values here). For 353 species diversity (H'), year was not a significant predictor, but month and station were, 354 showing the importance of station for all three metrics (Table 3). Station proved to be a 355 better predictor for all metrics than depth (by AIC), although the two are highly collinear 356 and station can be seen as a categorical proxy for depth. Species richness, evenness, and diversity were highest at moderate depths (Stations NH3, MB40, and NH5, from 30-60 m). 357

358 To better understand assemblage associations with environmental parameters, we 359 used nonmetric multidimensional scaling (NMDS). The two-dimensional model solution 360 had a final stress=0.17 and the three-dimensional model had stress=0.13 (stress < 0.20 has 361 been indicated as an interpretable result, with lower values indicating better fit (Clark 362 1993)). Visual inspection of the Shepherd plot, an indicator of model fit, showed the 363 predicted distances to be representative of the observed distances. Using PERMANOVA, 364 with possible explanatory factors including: year, month, depth, temperature and dissolved 365 oxygen, only year was not a significant predictor. Depth was the strongest predictor of 366 community, with assemblages at greater depths being distinct from assemblages at 367 shallower depths (Stations MB30 and MB40) (Figure 6). We repeated this analysis for the 368 subset of data with juveniles only (<100 mm SL) and found similar results with improved 369 model fit (two-dimensional model stress=0.12): depth was the strongest predictor, but 370 month, temperature and dissolved oxygen were also significant predictors when tested 371 using PERMANOVA. Because depth was the strongest structuring variable, we used 372 indicator species analysis for each station (a categorical surrogate for depth). Indicator 373 species analysis showed that the species driving the depth associations were Slender Sole, 374 Rex Sole (Glyptocephalus zachirus), and several uncommon poacher (family Agonidae) and 375 eelpout species (family Zoarcidae) at the deepest sites and the common flatfishes (English 376 Sole, Butter Sole, and Speckled Sanddab) at shallow-water sites. Pacific Sanddab was 377 characteristic of intermediate sites (50m-90m in depth).

378 3.4 Juvenile Flatfishes and Environmental Variables

We analyzed abundance (CPUE) of YOY related to environmental variables of the
four most common flatfish species: Butter Sole, English Sole, Speckled Sanddab, Pacific

381 Sanddab, all of which occurred in high abundances and in a large proportion of the tows 382 (YOY of each species were in 75%, 54%, 79%, and 43% of tows, respectively). We explored 383 relationships for Slender Sole as well, but this species was not commonly occurring (YOY 384 found in <20% of the tows) and was found in high abundance only in 2012, indicating that 385 a zero-inflated modeling approach would be a better fit and it was dropped from further 386 analysis. Exploratory plots revealed few obvious patterns in abundance as a function of the 387 environmental variables. Depth, however, was the primary driver of abundance, with 388 English Sole, Butter Sole, and Speckled Sanddab found at shallower depths (<70 m) and 389 Pacific Sanddab being more abundant at mid- to deep depths (50-100m). 390 We explored two general model formulations in our GAM analysis: models with each 391 factor considered independently and models that included a depth*dissolved oxygen 392 interaction. In all cases, the best model fit was: 393 Species count ~ offset (ln(distance towed))+ s(Depth) + s(DO) + s(Temp)+ 394 s(Avg.Upwelling.Index) + factor(Year) (Equation 3) 395 where all terms were significantly different from zero in at least one level or interval within 396 their respective range. This was consistent with our analysis of variance inflation factors, 397 which showed weak collinearity; however, our understanding of local oceanographic 398 processes and interest in fish response to dissolved oxygen led us to investigate the 399 interaction. For all species (including total abundance of all species) model fits had 41-73% 400 of deviance explained, with normal residuals (Table 4). When plots of smoothed terms of 401 each factor were evaluated (Figure 7), the effect of upwelling index appeared neutral (not 402 shown in figure) and abundances appeared positively correlated with temperature and 403 negatively correlated with dissolved oxygen for all species and total fish abundance. The

404 effects of depth varied by species, but in most cases this variable was the strongest 405 predictor. When predicted abundances were plotted as a function of dissolved oxygen and 406 depth, we found high concordance between the predictions and our raw catch data (Figure 407 8). While all species were found across a range of dissolved oxygen measurements, depth 408 appeared to be the stronger structuring variable. Trends in annual summer abundance 409 (Figure 9, plotted as log density by year) varied by species, with Butter Sole and Pacific 410 Sanddab showing variation among all years, English Sole having a slight downward trend 411 across the time series, and Speckled Sanddab having highest densities in the middle of the 412 time series.

413 3.5 Juvenile Flatfish Condition

414 We evaluated both Fulton's K and residuals from length-weight regression models 415 (L-W Residuals) for indications of variable fitness within newly settled individuals of 416 Butter Sole, English Sole, and Speckled Sanddab. While Pacific Sanddab was also abundant, 417 we collected fewer new recruits of this species and did not include this species in the 418 analysis. There was a slight positive and non-linear correlation between Fulton's K and L-W 419 Residuals for all three species, as both are weight-based measures of condition. Fulton's K 420 for all three species ranged from 0.13 to 4.49, with a mean of 1.37 (50% of the values were 421 between 1.22-1.50); these values are similar to those found for other flatfishes in this size 422 range (De Raedemaecker et al., 2012). Fulton's K varied among the three species (ANOVA, *f*= 354.6, p<.001), with mean values for English Sole (1.52) significantly higher than Butter 423 424 Sole (1.32) and Speckled Sanddab (1.40). Summary plots showed few clear trends among 425 species and within the two condition metrics with regard to environmental variables. 426 However, for both Butter Sole and English Sole, condition as measured by Fulton's K and L-

W Residuals appeared lower in 2013 than in other years; this same observation was not
made for Speckled Sanddab (Fig. 10) and our mixed models only indicated year as an
important explanatory variable for Butter Sole (see below, Table 5).

When analyzed with general linear mixed models, three of the six condition models
had low explanatory power (*R*² ranging from 0.026 to 0.136), with slightly higher values for
the other three (*R*² ranging from 0.208 to 0.384). The random effect (tow) contributed
marginally to model fit (Table 5). For all three species and both metrics of condition,
dissolved oxygen was included in all best models selected through AIC (lowest scores) with
a slight positive relationship between condition and dissolved oxygen. However,
coefficients for dissolved oxygen were very close to zero and effect sizes were small.

437 **4. Discussion**

438 4.1 Depth is a Primary Structuring Variable

439 The beam trawl was effective at collecting juvenile demersal fishes along a depth 440 gradient on the Oregon coast. While the overall fish diversity was high, with over 80 441 species collected in 135 tows made over the summer months, the samples were dominated 442 by juvenile flatfishes, which became the focus of our study. Species richness within a tow 443 was low (mean=7), but with high overall diversity, the soft-bottom regions of the near-444 coastal shelf serve as habitat for many species, and appear to be critical rearing areas for 445 young-of-the-year flatfishes (Laroche and Holton, 1979; Kryger and Pearcy, 1986). Toole et 446 al. (2011) sampled deeper habitats (50m-400m) and found similar dominance by juvenile 447 flatfishes. Pacific Sanddab and Slender Sole were common in that study, as at deeper 448 sampling sites in this study; however, other species (Rex Sole and Dover sole, *Microstomus*

449 *pacificus*), which were rare in our study, were more common. This difference likely reflects 450 the deeper waters sampled in the Toole et al. (2011) study and preference for different 451 depths among species, but also could be a result of differing gear types and methods (larger 452 net opening and 6.4 mm cod end mesh in the Toole et al. gear). The species we sampled 453 also differ from those collected in the NOAA groundfish survey, which targets adults using a 454 larger mesh net (~14 cm) and cod-end (3.8 cm) (Keller 2010; 2015, and A. Keller, NOAA-455 NWFSC, Seattle, WA, personal communication). Without being duplicative, this study offers 456 insight into the variables structuring the juvenile demersal fish community and the 457 response of species, and individuals within species, to environmental variables such as low 458 dissolved oxygen and temperature.

459 Depth was the strongest structuring variable in our analysis of community and 460 species abundance. In analyses where we considered hydrography (bottom temperature 461 and dissolved oxygen) as well, depth was a stronger predictor. The NMDS and 462 PERMANOVA analyses showed modest segregation of community by depth (station). 463 Similar results were also found by Stinton et al. (2014) on a subset of data from this study. 464 We note that heterogeneity within assemblage groups (Anderson and Walsh, 2013) and 465 uneven sample size (fewer samples collected at deep sites) may leave the community 466 underrepresented at those depths.

In our community analysis and generalized additive models of abundance, English
Sole and Butter Sole juveniles were collected at depths <50 m, while Speckled Sanddab
were evenly distributed within the sampled depth range and Pacific Sanddab were found at
the deeper stations. As adults, English Sole are mostly found in water deeper than 50m, but
decrease in abundance with increasing depth across the shelf (Bradburn et al., 2011), while

472 Butter Sole remain in shallow habitats and are caught only in low numbers in the 473 shallowest stratum (<184 m) by the NOAA West Coast Groundfish Survey (Bradburn et al., 474 2011, A. Keller, NOAA-NWFSC, Seattle, WA, personal communication). The patterns we 475 observed, in addition to known estuarine residence (Krygier and Pearcy, 1986; Da Ben et 476 al. 1990), suggest that English Sole use a variety of habitats during their life cycle. Butter Sole and Speckled Sanddab, which were abundant in this nearshore juvenile survey, appear 477 478 confined to shallower areas throughout their life cycle, as they are not commonly collected 479 as adults further offshore (and Butter Sole are largely absent in coastal estuaries as well, 480 Gunderson et al., 1990). Pacific Sanddab, which we found occupying the deeper regions in 481 our survey, appear to shift habitats as they mature, being found at even deeper sites as 482 adults (Bradburn et al., 2011). Rooper et al. (2006) showed that among four species of 483 flatfish in a Pacific estuary, the species were spatially segregated based upon physical 484 habitat characteristics, even in a relatively shallow environment. Similarly, the observed 485 depth preferences among species in our study may serve as a means of niche partitioning, 486 especially during juvenile life stages, when densities are high and rapid growth may help 487 mitigate predation mortality.

While depth was a strong predictor of distribution, many other environmental variables, some of which were not measured explicitly in this study, correlate with depth: temperature, dissolved oxygen, sediment grain size, wave stress, biogenic and physical features of the benthic/sedimentary environment, etc. Additionally, food availability driven by upwelling-induced nutrient delivery, subsequent primary and secondary production, and the flux of organic carbon can be depth-related in this region (Hill and Wheeler, 2002). While these factors may all play a role in structuring the fish community, our results,

combined with those of Keller et al. (2010), Toole et al. (2011) and other work in this
region, suggest strong depth preferences among species. Similarly, in the Baltic Sea, flatfish
distribution was dictated by physical habitat variables rather than by prey availability
(Florin et al., 2009), and in estuaries, spatial segregation among competitors has been
observed in the presence of abundant prey (Rooper et al., 2006).

500 4.2 Response to Hypoxia

501 The initial phase of this survey was designed to collect fishes during episodic 502 summer hypoxic events to better understand biotic response to these disturbances. 503 Therefore, sampling effort was highest in July and August, when these events are typically 504 strongest and most widely distributed, covering a broad expanse of the shelf area 505 (Peterson et al., 2013). But because our sampling effort was not allocated evenly across 506 years and months, temporal comparisons of biotic response should be made with caution. 507 In most years we did not have before/after comparisons to know which species were 508 present before hypoxic events occurred, and there are likely differences in recruitment 509 timing irrespective of environmental disturbance (Shanks and Eckert, 2005). The 510 phenology of settlement for the common species is of interest, but without sampling at a 511 higher temporal resolution, this question remains unresolved. Continuation of the beam 512 trawl study with monthly sampling, as has been implemented in recent years, will improve 513 our understanding of recruitment timing (Yergey et al., in review), and potentially, 514 movement across the shelf in response to hypoxia for sensitive species.

515 While our study did not encompass the years of most severe hypoxia along the 516 Oregon coast (2002 and 2006), sampling in several years occurred during periods of 517 moderate hypoxia, which covered as much as 40% of the shelf by area (Peterson et al., 2013). The hypoxic events in this region result from upwelling and other advective
processes carrying low dissolved oxygen waters into the shallow regions sampled as part
of our study (Grantham et al., 2004; Pierce et al., 2012). Climate-related changes and
decreases in dissolved oxygen of source waters may further exacerbate these events
(Peterson et al., 2013).

523 Despite collecting a number of samples in hypoxic conditions, dissolved oxygen was 524 not observed to be the primary driver in structuring the fish assemblage nor in influencing 525 abundances of the common flatfish species in this region. However, it was a significant 526 covariate in all of the metrics that were statistically assessed in this study (community 527 composition, species abundance, and fish condition). Because hypoxic events are initiated 528 at depth from oxygen-depleted upwelled water along this coast, organisms associated with 529 the benthos in deeper habitats would be most likely to be negatively affected. However, we 530 caught high abundances of the flatfish species over a range of dissolved oxygen, including 531 at depth. Increased sampling at deeper stations could improve balance within the dataset 532 and potentially yield different results, especially since our dataset did not include any 533 normoxic (DO>2.0 ml/L) conditions at deeper sites.

In recently settled fishes, which we assumed to be most susceptible to environmental stresses, we expected to see some correlation between low dissolved oxygen and negative fish condition based upon the work of Keller et al. (2010, 2015) with adults. However, models were poorly fitting with high unexplained variance. The Fulton's K values we observed were similar to those reported for other YOY flatfishes (Gilliers et al., 2004, De Raedemaecker et al., 2012), but this metric has been shown to be less sensitive than others (e.g., RNA:DNA ratio, hepatosomatic index, lipid content) to environmental

541 variation (Gilliers et al., 2004; Steirhoff et al., 2009; Schloesser and Fabrizio, 2015). While 542 relationships between condition and dissolved oxygen were not strong, there are a number 543 of factors which were not sampled as part of this effort but which may be better predictors 544 for condition. For example, we have no record of the prey field to which these recently 545 settled fishes were exposed. Weight-based condition factors would be highly influenced by 546 prey availability (De Raedemaecker et al., 2012) and with an absence of gut data, we could 547 not directly evaluate stomach fullness or other diet-based attributes. Additionally, the 548 hypoxic events that we captured may have been too limited in persistence to understand 549 the sub-lethal impacts of hypoxia, such as compromised foraging ability, on recently settled 550 fishes.

551 One common immediate response to hypoxia in fishes is change in behavior. 552 Behavioral response to hypoxia can include changes in activity, increased use of air 553 breathing, increased use of aquatic surface respiration, and vertical or horizontal 554 movement (Kramer, 1987; Hughes et al., 2015). While increased air breathing, and 555 increased use of surface respiration are unlikely for demersally-associated fishes, changes 556 in behavior related to movement or change in activity are plausible, although were not 557 directly measured in this study. Consistency in depth preference among years and at 558 varying dissolved oxygen conditions in our study suggests that, at the dissolved oxygen 559 levels experienced, species generally prefer the same depth range during this juvenile stage 560 even when dissolved oxygen levels are lower.

It is also possible that the dominant species are tolerant of, or have adapted to, low oxygen. Recent work with English Sole in a lab setting on the Oregon coast showed English Sole growth to be negatively impacted at the lowest oxygen levels (1.3 ml/L) only when

564 temperature was high (10°C and 13°C treatments) but not impacted when temperature 565 was nearer the ambient conditions we observed in this study (7°C, Bancroft, 2015). In our 566 study, temperature and dissolved oxygen were correlated, with low oxygen values 567 occurring in areas of colder (upwelled) water; however, with recent warming along the 568 Pacific Coast (Bond et al., 2015) warmer temperatures may adversely affect fishes in low 569 oxygen areas. With the availability of modern respirometry equipment, lab studies 570 evaluating animal response to low oxygen are illuminating some capacity for tolerance of 571 these events (Brill et al., 2015), especially when fishes are at rest (Nelson and Lipkey, 572 2015). Laboratory studies would provide a mechanistic link between hypoxia and fish 573 response and may illustrate a range of responses fishes in the wild could exhibit under 574 unfavorable environmental conditions.

575 **5.** Conclusions

576 While we found weak evidence for juvenile flatfish response to low dissolved 577 oxygen, we observed a stronger relationship between fish distribution and depth. We also 578 observed that the inshore juvenile soft-sediment community is different than the 579 community found further offshore in deeper waters, both juveniles and adults, when we 580 compared our findings to those of other researchers in this region. Thus, the inshore 581 community may provide a trophic connection between coastal waters, including estuaries, 582 and offshore demersal fish habitats. Additionally, with ocean conditions in this region 583 changing rapidly, understanding this nearshore community will afford better 584 understanding of the impacts to biota across the coastal shelf. This study contributes to our

- 585 understanding of newly settled demersal fishes, which may be important for determining
- 586 year-class strength, in soft-bottom near-coastal habitats.

588 Acknowledgements

589 We thank vessel captains and crews of Oregon State University's *R/V Elakha* and other

vessels for assisting in data collection. S. Henkel and J. Phillips provided additional data on

591 fish lengths and weights for this analysis. C. Rooper and M. McClure (NOAA-Fisheries) and

anonymous reviewers provided comments improving the manuscript. The field portion of

593 this work was funded by Oregon Sea Grant (award number NA060AR4170010, project

number R/ECO-23). KS and LC were funded by the NSF SEES Research Coordination
Network, Grant 1140207, "Sustainability of Marine Renewable Resources in Subarctic

596 Systems Under Incumbent Environmental Variability and Human Exploitation." Funding

597 sources did not contribute to the study design, data collection, analysis, or development of

this manuscript.

Literature Cited

Adams, K. A., Barth, J. A. and Chan, F. 2013. Temporal variability of near-bottom dissolved oxygen during upwelling off central Oregon. *Journal of Geophysical Research: Oceans* 118: 4839–4854.

Amara, R., Meziane, T., Gilliers, C., Hermel, G., and Laffargue, P. 2007. Growth and condition indices in juvenile sole *Solea solea* measured to assess the quality of essential fish habitat. *Marine Ecology Progress Series* 351: 201-208.

Anderson, M. J. 2001. A new method for non-parametric multivariate analysis of variance. *Austral Ecology* 26: 32–46.

Anderson, M.J. and Walsh, D.C.U. 2013. PERMANOVA, ANOSIM, and the Mantel test in the face of heterogeneous dispersions: What null hypothesis are you testing? *Ecological Monographs* 83:557-574.

Bankcroft, M.P. 2015. *An Experimental Investigation of the Effects of Temperature and Dissolved Oxygen on the Growth of Juvenile English Sole and Juvenile Dungeness Crab.* MS Thesis, Oregon State University, Corvallis, OR, USA.

Beck, M.W., Heck, K.L., Able, K.W., Childers, D.L., Eggleston, D.B., Gillanders, B.M., Halpern, B., Hays, C.G., Hoshino, K., Minello, T.J., Orth, R.J., Sheridan, P.F. and Weinstein, M.P. 2001. The identification, conservation, and management of estuarine and marine nurseries for fish and invertebrates. *BioScience* 51(8): 633–641

Bond, N.A., Cronin, M.F., Freeland, H. and Mantua, N. 2015. Causes and impacts of the 2014 warm anomaly in the NE Pacific. *Geophysical Research Letters* 42: 3414–3420.

Bradburn, M.J., Keller, A.A., and Horness, B.H. 2011. The 2003 to 2008 U.S. West Coast bottom trawl surveys of groundfish resources off Washington, Oregon, and California: Estimates of distribution, abundance, length, and age composition. U.S. Dept. of Commerce, NOAA Tech. Memo. NMFS-NWFSC-114, 323 p.

Brill, R.W., Bushnell, P.G., Elton, T.A. and Small, H.J. 2015. The ability of blue crab (*Callinectes sapidus*, Rathbun 1886) to sustain aerobic metabolism during hypoxia. *Journal of Experimental Marine Biology and Ecology* 471: 126–136.

Brodeur, R.D., Peterson, W.T., Auth, T.D., Soulen, H.L., Parnel, M.M. and Emerson, A.A. 2008. Abundance and diversity of coastal fish larvae as indicators of recent changes in ocean and climate conditions in the Oregon upwelling zone. *Marine Ecology Progress Series* 366: 187-202.

Burnham, K. P. and Anderson, D.R. 2002. *Model Selection and Multimodel Inference: A Practical Information-Theoretical Approach.* 2d ed. New York: Springer-Verlag.

Chan, F., Barth, J.A., Lubchenco, J., Kirincich, A., Weeks, H., Peterson, W.T. and Menge, B.A. 2008. Emergence of anoxia in the California current large marine ecosystem. *Science* 319: 920.

Clarke, K.R., and R.M. Warwick. 2001. *Change in marine communities: an approach to statistical analysis and interpretation.* 2nd edition. Plymouth: PRIMER-E.

De Ben, W.A., Clothier, W.D., Ditsworth, G.R. and Baumgartner, D.J. 1990. Spatio-temporal fluctuations in the distribution and abundance of demersal fish and epibenthic crustaceans in Yaquina Bay, Oregon, *Estuaries*, 113: 469-478.

De Caceres, M. and Legendre, P. 2009. Associations between species and groups of sites: indices and statistical inference. *Ecology* 90(12): 3566-3574.

De Raedemaecker, F., Brophy, D., O'Connor, I. and Comerford, S. 2012. Habitat characteristics promoting high density and condition of juvenile flatfish at nursery grounds on the west coast of Ireland. *Journal of Sea Research* 73: 7-17.

Donnellan, M., Merems, A., Miller, B. and Dinsmore, M. 2009. Resolving spatial scales of fishhabitat relationships on nearshore rocky reefs. Oregon Department of Fish and Wildlife, Final Report for State Wildlife Grant Number T-17, Salem.

Florin, A.B., Sundbald, G. and Bergström, U. 2009. Characterization of juvenile flatfish habitats in the Baltic Sea. *Estuarine, Coastal and Shelf Science*, 82: 294-300.

Gadomski, D.M. and Boehlert, G.W. 1984. Feeding ecology of pelagic larvae of English sole *Parophrys vetulus* and butter sole *Isopsetta isolepis* off the Oregon coast. *Marine Ecology Progress Series* 20: 1-12.

Gilliers, C, Amara, R., Bergeron, J-P. and Le Pape, O. 2004. Comparison of growth and condition indices of juvenile flatfish in different coastal nursery grounds. *Environmental Biology of Fishes* 71: 189–198.

Grantham, B.A., Chan, F., Nielsen, K.J., Fox, D.S., Barth, J.A., Huyer, A., Lubchenco, J. and Menge, B.A. 2004. Upwelling-driven nearshore hypoxia signals ecosystem and oceanographic changes in the northeast Pacific. *Nature* 429:749–754.

Gunderson, D.R., Armstrong, D.A., Shi, Y.-B., and McConnaughey, R. A. 1990. Patterns of estuarine use by juvenile English sole (*Parophrys vetulus*) and Dungeness crab (*Cancer magister*). *Estuaries* 13:59–71.

Heck, K.L., van Belle, G. and Simberloff, D. 1975. Explicit calculation of the rarefaction diversity measurement and the determination of sufficient sample size. *Ecology*, 56, 1459–1461.

Hill, J.K. and Wheeler, P.A. 2002. Organic carbon and nitrogen in the northern California current system: comparison of offshore, river plume, and coastally upwelled waters. *Progress in Oceanography* 53: 369–387.

Houde, E.D. 2002. Mortality. In *Fishery Science: the Unique Contribution of Early Life Stages,* pp. 64–87. Ed. by L.A. Fuiman, and Werner, R.G. Blackwell Publishing, Oxford.

Hughes, B.B., Levey, M.D., Fountain, M.C., Carlisle, A.B., Chavez, F.P. and Gleason, M.G. 2015. Climate mediates hypoxic stress on fish diversity and nursery function at the land-sea interface. *Proceedings of the National Academy of Sciences* 112: 8025-8030.

Jay, C. 1996. Distribution of bottom-trawl fish assemblages over the continental shelf and upper slope of the U.S. west coast, 1977-92. *Canadian Journal of Fisheries and Aquatic Sciences*. 53:1203-1225.

Keller, A.A., Simon, V., Chan, F., Wakefield, W.W., Clarke, M.E., Barth, J.A., Kamikawa, D. and Fruh. E.L. 2010. Demersal fish and invertebrate biomass in relation to an offshore hypoxic zone along the US West Coast. *Fisheries Oceanography* 19: 76-87.

Keller, A.A., Ciannelli, L., Wakefield, W.W., Simon, V., Barth, J.A., and Pierce, S.D. 2015. Occurrence of demersal fishes in relation to near-bottom oxygen levels within the California Current large marine ecosystem. *Fisheries Oceanography* 24: 162-176.

Kramer, D. 1987. Dissolved oxygen and fish behavior. *Environmental Biology of Fishes* 2: 81-92.

Krygier, E.E. and Pearcy, W. G. 1986. The role of estuarine and offshore nursery areas for young English sole, *Parophrys vetulus* Girard, of Oregon. *Fishery Bulletin* 84: 119-132.

Laroche, W.A. and Holton, R.L. 1979. Occurrence of 0-age English sole, *Parophrys vetulus*, along the Oregon coast: an open coast nursery area? *Northwest Science* 53:94-96.

Miller, S.D., Clarke, M.E., Hastie, J.D. and Hamel, O.S. 2009 Pacific Coast Fisheries. In: Our living oceans. Report on the status of U.S. living marine resources, 6th edition. US Dep. Commer., NOAA Memo. NMFS-=F/SPO-80, 369 p.

Nakagawa, S. and Schielzeth, H. 2013. A general and simple method for obtaining R2 from Generalized Linear Mixed-effects Models. *Methods in Ecology and Evolution* 4: 133–142.

National Marine Fisheries Service (NMFS). 2013. Groundfish Essential Fish Habitat Synthesis: A Report to the Pacific Fishery Management Council. NOAA NMFS Northwest Fisheries Science Center, Seattle, WA, April 2013. 107 p.

Nelson, J.A. and Lipkey, G.K. 2015. Hypoxia tolerance variance between swimming and resting striped bass *Morone saxatilis. Journal of Fish Biology* 87: 510–518.

Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., O'Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H. and Wagner, H. 2015. 'Vegan:' Community Ecology Package, R Package version 2.3-0. Published: 2015-05-21. http://cran.rproject.org/web/packages/vegan/vegan.pdf

Pearcy, W.G. 1964. Some distributional features of mesopelagic fishes off Oregon. *Journal of Marine Research* 22: 83–102.

Pearcy W.G. 1978. Distribution and abundance of small flatfishes and other demersal fishes in a region of diverse sediments and bathymetry off Oregon. *Fishery Bulletin* 76: 629-639.

Pearcy, W.G., Hosie, M.J. and Richardson, S.L. 1977. Distribution and duration of pelagic life of larvae of Dover sole, *Microstomus pacificus*; rex sole, *Glyptocephalus zachirus*; and petrale sole, *Eopsetta jordani*, in waters off Oregon. *Fishery Bulletin* 75:173–183.

Pearcy, W.G., Stein, D.L., Hixon, M.A., Pikitch, E.K., Barss, W.H. and Starr, R.M. 1989. Submersible observations of deep-reef fishes of Heceta Bank, *Oregon. Fish. Bull.*, 87: 955–965

Peterson, J.O., Morgan, C.A., Peterson, W.T. and Di Lorenzo, E. 2013. Seasonal and interannual variation in the extent of hypoxia in the northern California current from 1998–2012. *Limnology and Oceanography* 58: 2279–2292.

Pielou, E.C. 1969. *An Introduction to Mathematical Ecology*. New York, NY: John Wiley and Sons.

Pierce, S.D., Barth, J.A., Shearman, K.R. and Erofeev, A.Y. 2012. Declining oxygen in the northeast Pacific. *American Meteorological Society*, 42: 495-501.

Pikitch, E.K., Santora, C., Babcock, E.A., Bakun, A., Bonfil, R., Conover, D.O., Dayton, P., Doukakis, P., Fluharty, D., Heneman, B., Houde, E.D., Link, J., Livingston, P., Mangel, M., McAllister, M.K., Pope, J. and Sainsbury, K. J. 2004. Ecosystem-based fishery management. *Science* 305:346-347.

R Core Team. 2016. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL http://www.R-project.org/.

Rackowski, J.P., Pikitch, E.K. 1989. Species profiles: life histories and environmental requirements of coastal fishes and invertebrates (Pacific Southwest)--Pacific and speckled

sanddabs. U.S. Fish and Wildlife Service Biol. Rep. 82(11.107). U.S. Army Corps of Engineers, TR EL-82-4. 18 pp.

Richardson, S.L. and Pearcy, W.G. 1977. Coastal and oceanic fish larvae in an area of upwelling off Yaquina Bay, Oregon. *Fishery Bulletin* 75:125-145.

Romsos, C.G., Goldfinger, C., Robison, R., Milstein, R.L., Chaytor, J.D., and Wakefield, W.W. 2007. Development of a regional seafloor surficial geologic habitat map for the continental margins of Oregon and Washington, USA. *in* Todd, B.J., and Greene, H.G., eds., *Mapping the Seafloor for Habitat Characterization*, Geological Association of Canada, Special Paper 47, p. 209-234.

Rooper, C.N., Gunderson, D.R. and Armstrong, D.A. 2003. Patterns in use of estuarine habitat by juvenile English sole (*Pleuronectes vetulus*) in four eastern north Pacific estuaries. *Estuaries* 26:1142–1154.

Rooper, C.N., Gunderson, D.R. and Armstrong, D.A. 2006. Evidence for resource partitioning and competition in nursery estuaries by juvenile flatfish in Oregon and Washington. *Fishery Bulletin* 104: 616-622.

Rosenberg, A.A. 1982. Growth of juvenile English sole, *Parophrys vetulus*, in estuarine and open coastal nursery grounds. *Fishery Bulletin*. 80: 245-252.

Schloesser, R.W. and Fabrizio, M.C. 2015. Relationship among proximate components and energy density of juvenile Atlantic estuarine fishes. *Transactions of the American Fisheries Society* 144: 942-955.

Schulte-Hostedde, A., Zinner, B., Millar, J.S. and Hickling, G.J. 2005. Restitution of mass-size residuals: validating body condition indices. *Ecology* 86: 155–163.

Shanks, A.L. and Eckert, G.L. 2005. Population persistence of California Current fishes and benthic crustaceans: A marine drift paradox. *Ecological Monographs* 75: 505–524.

Sogard, S.M. 1997. Size-selective mortality in the juvenile stage of teleost fishes: A review. *Bulletin of Marine Science* 60:1129–1157.

Stierhoff, K.L., Targett, T.E. and Power, J.H. 2009. Hypoxia-induced growth limitation of juvenile fishes in an estuarine nursery: assessment of small-scale temporal dynamics using RNA:DNA. Canadian Journal. Fish. Aquat. Sci. 66: 1033–1047.

Stinton, A., Ciannelli, L., Reese, D.C. and Wakefield, W.W. 2014. Using in situ video analysis to assess juvenile flatfish behavior along the Oregon central coast. *CalCOFI Reports* 55: 158-168.

Thanassekos, S., Latour, R.J. and Fabrizio, M.C. 2016. An individual-based approach to yearclass strength estimation. *ICES Journal of Marine Science* 73: 2252–2266. Tissot, B.N., Hixon, M.A. and Stein, D.L. 2007. Habitat-based submersible assessment of groundfish assemblages at Heceta Bank, Oregon from 1988 to 1990. *J. Exp. Mar. Biol. Ecol.* 352: 50-64.

Tissot, B.N., Wakefield, W.W., Hixon, M.A. and Clemons, J.E.R. 2008. Twenty years of fishhabitat studies on Heceta Bank, Oregon. In J.R. Reynolds and H.G. Greene (eds). *Marine Habitat Mapping Technology for Alaska*, Alaska Sea Grant College Program, Fairbanks, AK. CD-ROM. doi:10.4027/mhmta.2008.15

Toole, C.L., Brodeur, R.D., Donohoe, C.J. and Markle, D.F. 2011. Seasonal and interannual variability in the community structure of small demersal fishes off the Oregon coast. *Marine Ecological Progress Series* 428: 201-217.

Van Cleve, R., and El-Sayed, S.Z. 1969. Age, growth, and productivity of an English sole (*Parophrys vetulus*) population in Puget Sound, Washington. *Pac. Mar. Fish. Comm. Bull.* 7: 51-71.

Whittaker, R.H. 1972. Evolution and measurement of species diversity. Taxon 21:213–251.

Wood, S.N. 2006. *Generalized Additive Models: an introduction with R*, CRC Press, Boca Raton, FL.

Yergey, M., Wakefield, W.W., Ciannelli, L. and Sobocinski, K.L. In review. Distribution and abundance of young-of-the-year groundfishes off the central Oregon coast: One year of monthly observations and trends. *Fishery Bulletin*

Yoklavich, M. and Wakefield, W.W. 2015. Pacific Coast Region. In: NMFS. Our living oceans: habitat. Status of the habitat of U.S. living marine resources. U.S. Dep. Commerce, NOAA Tech. Memo. NMFS-F/SPO-75, 327 p.

Zuur, A.F., Ieno, E.N., Walker, N.J., Saveliev, A.A. and Smith, G.M. 2009. *Mixed Effects Models and Extensions in Ecology with R.* Springer, New York. 574 pp

Zuur, AF. 2013. *A Beginner's Guide to Generalized Additive Models with R.* Highland Statistics, Newburgh, U.K. 188 pp.

						Station					
		MB30	MB40	MB50	NH03	NH05	NH07	NH10	NH15	NH20	
Physical	Mean Depth										
Properties	(m)	31.2	41.0	50.7	43.7	58.5	73.2	79.8	110.9	120.0	
	Temperature										
	(°C)	8.2	7.9	7.5	7.7	7.5	7.5	7.3	7.3	7.3	
	Dissolved										-
	Oxygen										
	(mL/L)	2.8	2.3	1.8	1.9	1.9	1.2	1.6	1.4	1.2	
Samples											
Collected											Total
2008	July	1	1			1		1			4
	August	2	4			4	1	4			15
		3	5			5	1	5			19
2009	July	1	1			1					3
	August	1	1			1		1			4
		2	2			2		1			7
2010	June	3	3	3							9
	July	1	1	1		1		1			5
	August	4	4	3		1		1			13
		8	8	7		2		2			27
2011	May	3	3	3							9
	June	5	3	1							9
	July	8	8								16
	August	3	3	3							9
		19	17	7							43
2012	July	3	3		3	3		3			15
	August	1	1		1	1		1	1	1	7
	September	1	1			1		1	1		5
		5	5		4	5		5	2	1	27
2013	June	1	1		1	1		1	1		6
	August	1	1		1	1		1	1		6
		2	2		2	2		2	2		12
	Grand Total	39	39	14	6	16	1	15	4	1	135

Table 1. Physical properties and number of samples collected by year and month for each station with the beam trawl, summers 2008-2013. Temperature and dissolved oxygen are mean bottom water values.

Table 2. Fish species collected and total abundance by year for the six sampling years for species occurring in >1% of all tows. Overall abundance (Total) and frequency of occurrence (Freq.) are also shown. Frequency of occurrence refers to the proportion of tows out of the total number of tows (135) in which the species was present. This table omits 40+ species which occurred in <1% of the tows, as well as 33 unidentified specimens.

	Total Abundance								
Common Name	Scientific Name	2008	2009	2010	2011	2012	2013	Total	Freq.
Butter Sole	Isopsetta isolepis	305	80	347	861	1216	316	3125	0.84
Speckled Sanddab	Citharichthys stigmaeus	226	68	1072	811	413	104	2694	0.83
English Sole	Parophrys vetulus	632	200	444	333	634	103	2346	0.85
Pacific Sanddab	Citharichthys sordidus	387	120	365	68	417	208	1565	0.58
Slender Sole	Lyopsetta exilis	8	5	10	0	761	8	792	0.21
Sanddab spp.	Citharichthys spp.	0	0	76	270	68	10	424	0.27
Pacific Tomcod	Microgadus proximus	70	6	1	4	25	186	292	0.17
Smelt spp.	Osmeridae spp.	1	0	21	135	1	10	168	0.09
Sand Sole	Psettichthys melanostictus	2	2	27	91	23	10	155	0.40
Pacific Sandlance	Ammodytes hexapterus	111	0	16	2	2	17	148	0.16
Warty Poacher	Occella verrucosa	12	4	76	18	8	16	134	0.30
Righteye Flounder	Pleuronectidae spp.	0	0	0	7	54	41	102	0.11
Pacific Staghorn Sculpin	Leptocottus armatus	24	8	3	28	6	20	89	0.24
Slim Sculpin	Radulinus asprellus	10	0	0	0	60	1	71	0.05
Rex Sole	Glyptocephalus zachirus	1	0	0	21	43	1	66	0.10
Snailfish spp.	Liparidae spp.	1	0	0	24	22	4	51	0.17
Roughback Sculpin	Chitonotus pugetensis	9	9	5	7	8	8	46	0.13
Dover Sole	Microstomus pacificus	11	3	7	7	14	2	44	0.21
Dungeness Crab	Metacarcinus magister	0	0	0	0	13	23	36	0.09
Black Rockfish	Sebastes melanops	0	0	28	1	0	0	29	0.05
Tubenose Poacher	Pallasina barbata	6	2	9	1	2	1	21	0.09
Lingcod	Ophiodon elongatus	7	1	0	5	0	6	19	0.08
Night Smelt	Spirinchus starksi	0	0	0	0	16	1	17	0.03
Sculpin spp.	Artedius spp.	5	0	1	1	7	1	15	0.05
Pertrale Sole	Eopsetta jordani	2	0	1	1	3	5	12	0.07
Canary Rockfish	Sebastes pinniger	1	0	2	5	1	3	12	0.06
Big Skate	Beringraja binoculata	2	0	0	5	3	0	10	0.06
Brown Irish Lord	Hemilepidotus spinosus	1	0	1	0	1	7	10	0.03
Northern Ronquil	Ronquilus jordani	0	0	0	0	8	1	9	0.03
Poacher spp.	Agonidae spp.	7	0	1	1	0	0	9	0.03
Bluebarred Prickleback	Plectobranchus evides	0	0	0	0	8	0	8	0.02
Smooth Alligatorfish	Anoplagonus inermis	0	0	3	1	2	0	6	0.04
Northern Spearnose Poacher	Agonopsis vulsa	0	1	0	0	4	0	5	0.04
Pygmy Poacher	Odontopyxis trispinosa	0	0	2	3	0	0	5	0.04
Rockfish spp.	Sebastes spp.	2	0	1	1	0	1	5	0.03
Flathead Sole	Hippoglossoides elassodon	1	1	0	0	0	2	4	0.02
Thornback Sculpin	Paricelinus hopliticus	1	0	2	0	0	0	3	0.02

Table 3. Linear model output for diversity metrics, Species Richness (SR), Evenness (J), and Diversity (H') for the beam trawl community. The full model evaluated for each metric was: *Response ~ Year + Month + Station + Dissolved Oxygen + Temperature*. The best fitting models were: *SR ~ Year + Station + Temperature*, *J ~ Year + Station*, and *H' ~ Month + Station*. Coefficient values for significant factors are shown for each explanatory variable and for each level for categorical variables; for categorical variables, the coefficient is relative to the first level of the factor. NS=Not a significant factor and - = reference level.

Explanatory Variable	Levels	Species Richness	Evenness	Diversity	
		(SR)	(L)	(H')	
Intercept		3.69	0.64	1.39	
Dissolved Oxygen		NS	NS	NS	
Temperature		0.59	NS	NS	
Year	2008	-	-	NS	
	2009	-0.43	-0.02	NS	
	2010	-1.12	-0.02	NS	
	2011	-2.36	0.06	NS	
	2012	1.06	-0.06	NS	
	2013	1.70	0.07	NS	
Month	May	NS	NS	-	
	June	NS	NS	-0.13	
	July	NS	NS	-0.23	
	August	NS	NS	-0.03	
	September	NS	NS	-0.34	
Station	MB30	-	-	-	
	MB40	0.41	0.07	0.16	
	MB50	0.30	0.06	0.02	
	NH03	-1.13	-0.06	-0.08	
	NH05	-1.64	0.06	0.02	
	NH07	-4.05	-0.31	-0.90	
	NH10	-3.46	0.02	-0.29	
	NH15	-2.60	-0.05	-0.27	
	NH20	1.97	-0.04	-0.34	

Table 4. GAM output for the four most common flatfish species. The best model for all species (and total abundance) was $Y \sim s(Depth) + s(DO) + s(Temp) + s(Avg.Upwelling.Index) + factor(Year)$. For all models, all explanatory variables were significant (p<0.001 in all cases). The delta AIC indicates the difference in AIC between the model with and without the Depth*DO interaction term.

Species	Deviance Explained	Δ AIC (difference from model with interaction)
Total Abundance (all species)	41.3%	22.5
Butter Sole	52.1%	12.5
English Sole	73.5%	183.6
Speckled Sanddab	55.6%	118.6
Pacific Sanddab	50.1%	113.4

Table 5. Best fitting models for two metrics of condition factor for the three most abundant species of juvenile flatfish. The full model for each response was: Response ~ *Year* + *Depth* + *Dissolved Oxygen* + *Temperature* + *Dissolved Oxygen* : *Depth (interaction term)*. The coefficient for the dissolved oxygen parameter (the main factor of interest) and R² for both the fixed and random effects are provided for the best fitting model. Effect size refers to the change in condition for one unit change is dissolved oxygen. *For the models for English Sole and Speckled Sanddab, the model form only including dissolved oxygen had the lowest AIC score; however, dissolved oxygen was not a significant explanatory variable, as evidenced by coefficients near zero and low explanatory power.

Species	Metric	Best Fitting Model	Dissolved Oxygen Coefficient	Effect Size	R2 (random)	R2 (fixed)
Butter Sole	Residuals	Response ~ Year + DO	0.007	0.9%	0.052	0.384
(n=2400)	Fulton's K	Response ~ Year + DO	0.035	1.0%	0.067	0.271
English Sole	Residuals	Response ~ DO*	0.005	0.6%	0.002	0.208
(n=1426)	Fulton's K	Response ~ DO*	0.001	0.04%	0.001	0.026
Speckled Sanddab	Residuals	Response ~ DO*	0.007	0.4%	0.003	0.136
(n=1041)	Fulton's K	Response ~ DO*	0.007	0.2%	0.001	0.119



Figure 1. Dissolved oxygen (mL/L) and temperature (°C) as a function of depth as measured concurrently with beam trawl sampling.



Figure 2. Dissolved oxygen (A., ml/L) and Temperature (B., °C) across all sampling sites by month, from May (5) to September (9). Dark lines show median values with boxes representing first and third quartiles.



Figure 3. Length frequency of all fishes collected in the beam trawl survey. Vertical lines denote newly settled (<50 mm) and young-of-the-year (<100 mm) individuals.



Figure 4. Probability density function of length-frequency for the five most abundant flatfish species. Young-of-the-year fishes dominated the catch, with Butter Sole (*Isopsetta isolepis*) being the most abundant and captured in highest abundance when recently settled (~35 mm SL).



Figure 5. Species richness (SR, black squares), diversity (H', triangles) and evenness (J, gray diamonds, secondary y-axis), for all stations across the six sampling years. Error bars on species richness represent standard deviation of the mean.



Figure 6. NMDS plot showing assemblage structure with significant explanatory variables shown as vectors. Sampled depth is represented by contour lines, with increasing depth to the right side of the plot. Depth was the strongest predictor of community structure as shown by the vector arrow across the depth contours. 2-dimensional stress for the model was 0.17.



Figure 7. GAM output for species abundance anomalies for four species (rows) showing smoothed terms for each of three predictors: Depth (left column), Dissolved Oxygen (center column), and Temperature (right column). Upwelling index was included in the model, but is not shown because its effect on mean fish abundance was small and predominantly flat throughout the examined range for all species. Note that the scale of the effect size (y-axis) varies by species and predictor.



Figure 8. Model output for the most common flatfish species from GAM. Bubbles represent abundance data from the beam trawl, scaled by count (log scale, as per legend for each species). Tows with zero catches of the species are indicated by (+). Shading indicates the predicted abundance of each species based upon depth (x-axis) and dissolved oxygen (y-axis), with lighter shading indicating greater predicted abundance and darker areas indicating less predicted abundance.



Figure 9. Density by year for four species from the beam trawl survey. Density is shown as log(n+0.01) for best resolution. Black bars within boxes show median values.



Figure 10. Fish condition, Fulton's K, across the sampling years for three common species, Butter Sole, English Sole, and Speckled Sanddab. Dark lines in the center of boxes show medians with boxes showing first and third quartiles. Black dots indicate outliers.