1	Rapid age estimation of longnose skate (Raja rhina) vertebrae using near infrared
2	spectroscopy
3	
4	Morgan B. Arrington ¹ *, Thomas E. Helser ² , Irina M. Benson ² , Timothy E. Essington ¹ , Mary
5	Elizabeth Matta ² , André E. Punt ¹
6	
7	¹ School of Aquatic and Fishery Sciences, University of Washington, 1122 Boat Street, Seattle,
8	WA 98195, USA
9	² Resource Ecology and Fisheries Management Division, Alaska Fisheries Science Center,
10	National Marine Fisheries Service, National Oceanic and Atmospheric Administration, 7600
11	Sand Point Way NE, Seattle, WA 98115, USA
12	

13 *corresponding author

14 Abstract

15

There is a paucity of age data for chondrichthyan fishes due, in large part, to limitations in 16 17 traditional age estimation methods. Fourier transform near infrared (FT-NIR) spectroscopy has shown promise as an alternative, more efficient method for acquiring age data from 18 chondrichthyans. However, studies are limited to sharks in the Southern Hemisphere. We explored 19 FT-NIR spectroscopy to predict age for a batoid species in the Northern Hemisphere. The longnose 20 skate (Raja rhina) is one of a small number of batoids for which annual band periodicity in 21 22 vertebral centra has been validated, allowing for traditional age estimation and making it an ideal candidate for this study. We fit a multivariate partial least squares predictive model between FT-23 NIR spectra collected from vertebral centra and traditional age estimates, and tested model 24 25 predictive skill using external validation. Using FT-NIR spectroscopy, we were able to predict age for longnose skates between the ages of 1 and 14 years with near equal precision and bias as 26 traditional methods in less than a quarter of the time. These results support potential for FT-NIR 27 spectroscopy to increase the amount of age data available for assessments used to inform the 28 29 conservation and management of this sensitive group of species.

30 1. Introduction

31

Information on the age of fishes is an essential component of fisheries research and 32 management. Accurate age data allow for a more robust understanding of population dynamics 33 and contribute to our ability to conserve and manage species effectively (Lai and Gunderson 1987; 34 35 Campana 2001). Fish age data are essential for age-structured methods of stock assessment as they are used to estimate the size of population cohorts as well as important life-history parameters 36 related to growth, mortality, maturity, and longevity. Because these metrics vary over time, 37 38 monitoring temporal changes in their values is increasingly recognized as an important component of population assessment. Fish growth can be altered by factors such as ecological and 39 environmental conditions (Shelton and Mangel 2012), fishing mortality (Heino and Dieckmann 40 2008), and density-dependent effects (Lorenzen and Enberg 2001). For example, changes in 41 temperature have been shown to affect fish growth (Matta et al. 2010; Pistevos et al. 2015; Matta 42 et al. 2018), increased mortality can reduce population density leading to increased food 43 availability and faster growth (Heino and Dieckmann 2008), and mortality that is size-selective 44 toward older and larger individuals can favor genotypes that grow faster (Stokes and Law 2000). 45

Despite its importance, monitoring population parameters that rely on age data is challenging because of the time and expense involved, as well as the difficulties with producing reliable age estimates for some vulnerable species such as chondrichthyans. Historically there has not been a large research focus on chondrichthyan species because they have not supported as many economically valuable fisheries as teleost fishes (Fowler *et al.* 2005; Dulvy *et al.* 2014). Additionally, acquiring information on the age of chondrichthyans is especially difficult because they do not possess otoliths, which are commonly used for age estimation of teleost fishes (Cailliet

et al. 2006; Matta et al. 2017). Growth band patterns visible in thin sections of vertebral centra are 53 often used for ageing chondrichthyans, but this method is time consuming and has not been 54 validated for many species (Matta et al. 2017). Furthermore, a growing number of studies have 55 demonstrated that these banding patterns may not form annually throughout life in all species, 56 raising doubts about the accuracy of age estimates generated from vertebral centra (Natanson et 57 al. 2018). Validation across species and age groups using independent methods of age 58 determination is important to ensure that the age-estimation protocol yields biologically accurate 59 age estimates. Consequently, age information is generally lacking for chondrichthyan populations 60 61 (Cailliet et al. 2006; Matta et al. 2017) even though many species face elevated risk of extinction due to the expansion of fishing, habitat loss, and climate change (Dulvy *et al.* 2014). 62

The longnose skate (*Raja rhina*) is one of a few species of chondrichthyan for which age 63 estimation methods have been validated (King et al. 2017). However, traditional growth-band age 64 estimation is expensive and time consuming for this species, which has precluded routine age 65 estimation. Life history traits such as late age-at-maturity relative to total lifespan and probable 66 low fecundity may make this species more sensitive to exploitation (King and McFarlane 2003), 67 yet a high percentage of bycatch is retained in some areas and there has been interest by industry 68 69 in developing a target fishery (Farrugia 2017). Thus, more age data would be beneficial to monitor population status with higher spatial and temporal resolution. 70

Fourier transform near infrared (FT-NIR) spectroscopy is a technology that could be used to improve the efficiency of longnose skate age estimation and enable the enhanced collection of age data for this species. Routinely used in pharmaceutical and agricultural industries to analyze chemical composition (McClure *et al.* 2002; Roggo *et al.* 2007), FT-NIR measures the interaction of near-infrared light with the chemistry of biological materials such that unique molecular

structures result in a measure of absorbance across a range of wavenumbers from 12,000 to 4,000 76 cm⁻¹ (Robins *et al.* 2015). It is being increasingly utilized in ecological research including for 77 species identification, physiological status, sex, detection of disease, and diet composition (Vance 78 et al. 2016). In the field of fish ageing, FT-NIR is an emerging technology that has been used to 79 more efficiently estimate ages of teleost fish using otoliths and of sharks using vertebrae, fin 80 81 spines, and fin clips (Rigby et al. 2014; Wedding et al. 2014; Robins et al. 2015; Rigby et al. 2016; Helser et al. 2018; Passerotti et al. 2020a; Passerotti et al. 2020b, Wright et al. 2021). The ability 82 to predict animal age using FT-NIR spectroscopy is based on the concept that there is a relationship 83 84 between an ageing structure's chemical composition and the specimen's age. While this technology has shown promise for estimating chondrichthyan ages, it has not yet been applied to 85 vertebrae in the northern hemisphere or from batoids. 86

Because ages estimated based on annual band periodicity have been validated for longnose skates (King *et al.* 2017), we were presented a unique opportunity to evaluate the use of FT-NIR spectroscopy to estimate ages from the vertebral centra of a batoid species in the northeastern Pacific Ocean. The objective of this study is to evaluate the utility of FT-NIR spectroscopy to estimate longnose skate age and compare results relative to traditional methods.

92

93 **2. Materials and methods**

94

The longnose skate vertebral centra used in this study were provided by the National Marine Fisheries Service (NMFS) Northwest Fisheries Science Center's (NWFSC) bottom trawl research surveys (Keller *et al.* 2017). A segment of thoracic vertebrae was collected from specimens and frozen at sea following approved NMFS procedures and guidance from the NMFS

Animal Care and Use Policy (Policy Directive 04-112). Collections occurred during 2011-2012 99 along the U.S. West Coast between May and October. These samples were processed for a separate 100 study on regional variability in the age and growth of longnose skates and more details and sample 101 collection locations are fully described in Arrington (2020). A group of three to four adjacent 102 thoracic vertebrae were dissected out of the frozen vertebral column in the laboratory, neural and 103 104 hemal arches were removed, and centra were stored in ethanol for preservation (Cailliet and Goldman 2004). One centrum per specimen was used for age estimation and another one was used 105 for spectroscopic evaluation in this study. Age readers estimated ages by counting annually 106 107 deposited growth bands on vertebral thin sections according to the validated protocol (Gburski et al. 2007; King et al. 2017). Hereinafter, we refer to reader-generated age estimates as "traditional 108 ages", while "predicted ages" represent the ages estimated using the calibration model in the FT-109 NIR spectroscopy approach. The primary reader (Reader 1) aged all specimens. Then, to evaluate 110 ageing error, two additional readers independently aged a random subset of vertebral thin sections. 111 We only evaluated a single vertebra per individual to maximize the number of individuals, but 112 future studies might explore variation in growth across vertebrae within individuals. Traditional 113 age estimates from the full longnose skate dataset ranged from 0 to 19 years (Arrington 2020). For 114 115 the purpose of the present study, we truncated the dataset at 14 years due to poor representation of the older age classes (fewer than 5 samples per age class), which could have a disproportionately 116 large influence on the predictive model. 117

To collect FT-NIR spectral absorbance data (hereinafter called spectral data), we placed one whole centrum per specimen under a fume hood and allowed it to air-dry for 48 hours so that ethanol could fully evaporate. We then placed each centrum on the sampling window of a Bruker TANGO-R Fourier transform near infrared spectrometer (Bruker Optics, Ettlingen, Germany) in

a standardized orientation with centrum cone side down and covered it with a reflector cap.
Spectral data were acquired at 16 cm⁻¹ resolution with 64 co-added scans. We collected spectral
data from 633 vertebrae (Fig. 1a).

Partial Least Squares (PLS) regression analysis was used to build a predictive model for 125 longnose skate age based on spectral data of whole centra. A PLS regression is a multivariate 126 regression method commonly used for chemometric analysis (Wold et al. 1984). In PLS, the 127 independent (here, spectral data) and dependent (here, Reader 1's traditional age estimate) data 128 matrices are decomposed into a set of scores and loadings. Loadings are determined by maximizing 129 130 the correlation between scores based on least squares. We conducted data analysis in R statistical computing software version 3.6.3 (R Core Team 2020), with chemometric packages *mdatools* 131 (Kucheryavskiy 2020) and simplerspec (Baumann 2020). 132

Raw spectral data often need to be transformed to enhance variation between specimens 133 and remove unwanted noise before modeling (Rinnan et al. 2009). First, we estimated the first 134 135 derivative of the spectral data to enhance separation between spectra while reducing any baseline drift in the data (Brown et al. 2000). We calculated the first derivative using a polynomial least 136 squares estimation known as the Savitzky-Golay first derivative (17-point smooth, polynomial 137 138 order = 1; Savitzky and Golay 1964). The Savitzky-Golay approach is commonly used in spectroscopy due to desirable properties such as reducing unwanted noise in spectral data while 139 140 preserving the chemical signal of interest. After calculating the first derivative, we mean centered 141 the data (Fig. 1b). We then split the data into a set used to calibrate the model and a set to externally validate it. These sets were selected so that the validation set contained all specimens that each had 142 143 three independent traditional age estimates. This allowed us to directly compare ageing error 144 between the FT-NIR spectroscopy approach and the traditional method. The calibration set

contained 413 samples and the validation set contained 220 samples (Fig. 2). We applied PLS to 145 the pre-processed spectral data from the calibration set and Reader 1's corresponding traditional 146 age estimates using a 10-fold venetian blind cross validation. This is a version of k-fold cross 147 validation in which each fold is constructed from samples in the dataset in the 1 through 10 position 148 until all data have been sorted. Each fold is systematically left out and a PLS regression is applied 149 150 to the remaining samples. The parameter estimates are then used to estimate the age of the left-out fold and the mean error of all predictions versus Reader 1's estimated ages can be calculated as 151 the root mean square error of cross validation (RMSECV). This is an estimate of the predictive 152 153 ability of the model on new data. We then applied this model to the validation data set and calculated a root mean square error of prediction (RMSEP) between predicted and Reader 1's 154 traditional age estimates to measure the true predictive ability of the model on an external data set. 155 We also report percentage root mean square error (% RMSE) = (RMSECV/maximum age \times 100) 156 to estimate the average predictive ability of the PLS model within the range of the age data 157 158 (Couture et al. 2016; Passerotti et al. 2020a). This metric also provides a standardized measure of error that can be compared across studies (Passerotti et al. 2020a). 159

Lastly, we assessed age estimation error in the FT-NIR spectroscopy approach compared 160 161 to the traditional method. There are two types of error associated with traditional age and growth studies: process error and observation error. Process error occurs when growth zones in the ageing 162 163 structure do not reflect true age. We did not address process error in this study. However, King et 164 al. (2017) suggest annual band periodicity in the vertebrae of longnose skates. Observation error is due to interpretation of the ageing structure and can be evaluated by comparing age estimates 165 166 among multiple age readers. In the FT-NIR approach, we considered observation error to be error 167 between FT-NIR predicted age and Reader 1's traditional age estimates.. This definition of

observation error is based on the simplifying assumption that there is no process error in FT-NIR 168 measurements that would result in different age estimates for the same specimens, though this has 169 vet to be evaluated or quantified for this species. To evaluate the precision of each method, we 170 compared the coefficient of variation (CV) between Reader 1's traditional age estimates and the 171 FT-NIR predicted ages to the CV among the three age reader estimates (Chang 1982). McBride 172 173 (2015) found that CV tracks precision better than other commonly used measures, such as average percent agreement (APE). We evaluated relative bias among traditional methods and the FT-NIR 174 approach visually using age bias plots and Bowker's test of symmetry (Hoenig et al. 1995; 175 176 McBride 2015). Since the PLS regression returns fractional age estimates, we first had to round predictions to the nearest whole number (i.e. 2.45 years = 2 years) so that precision and relative 177 bias could be compared between the two methods. We evaluated current levels of observation error 178 based on precision and relative bias among traditional age estimates to compare to the performance 179 of the FT-NIR approach and to evaluate the utility of this approach for generating age estimates 180 181 for longnose skates.

182

183 **3. Results**

184

The FT-NIR spectra of longnose skate vertebral centra correlated with Reader 1's traditionally estimated age with a coefficient of determination (R^2) of 0.86, a RMSECV of 1.38 years, and a %RMSE of 9.87% (Fig. 3). The RMSECV of 1.38 years indicates that 67% of the predicted ages from cross-validation fell within 1.38 years of the traditionally estimated age. When the model was applied to the external validation set and predictions compared to Reader 1's traditional age estimates, RMSEP was 1.32 years (Fig. 3).

The FT-NIR approach predicted longnose skate ages between 1 and 14 years in the external 191 validation set with as much precision as the traditional method but with a slight increase in bias 192 for individuals at either end of the age range. Specimens with Reader 1's traditional age estimate 193 of 0 years were imprecisely estimated by the PLS regression (Fig. 3). When age-0 specimens were 194 included, the traditional method had a CV of 22.6% among the three age readers and no systematic 195 disagreement between Reader 1 and 2 (Bowker's $\chi^2 = 32.10$, d.f. = 29, P = 0.315) or Reader 1 and 196 3 (Bowker's $\chi^2 = 46.91$, d.f. = 40, P = 0.210). Precision was lower between the FT-NIR predictions 197 and the Reader 1's traditional age estimates with a CV of 34.2% and the null hypothesis of no 198 systematic disagreement between methods was rejected (Bowker's $\chi^2 = 61.06$, d.f. = 37, P = 0.008). 199 When specimens with Reader 1's traditional age estimate of 0 were removed, the precision 200 between FT-NIR predictions and Reader 1's traditional age estimates (CV of 19.6%) was 201 comparable to that among readers (CV of 19.1%) and Bowker's test of symmetry indicated no 202 systematic disagreement between methods (Bowker's $\chi^2 = 42.86$, d.f. = 35, P = 0.170). We visually 203 observed a slight bias in FT-NIR predictions, with a tendency to over-estimate the age of younger 204 skates (< 2 years) and under-estimate the age of older skates (> 13 years) relative to the three age 205 reader estimates (Fig. 4). When the magnitude of discrepancies in age estimates was compared 206 207 between traditional and FT-NIR spectroscopy methods, we found that the percentage of samples with complete agreement (discrepancy of 0 years) was slightly higher among age readers using 208 traditional methods (41.5-45.1% agreement) than between Reader 1's estimates and FT-NIR 209 210 predictions (33.0% agreement), but otherwise had a similar distribution (Fig. 5).

211

212 **4. Discussion**

The results of this study suggest that FT-NIR spectra of longnose skate centra can be used 214 to estimate skate ages, similar to results published for other chondrichthyan species (Rigby et al. 215 2014; Rigby et al. 2016). When applied to an external validation set, the FT-NIR spectroscopy 216 method produced age predictions with comparable precision to traditional methods for longnose 217 skates between the ages of 1 and 14 years (the full age range in this study was 0-14). Since the 218 219 traditional method has been shown to produce biologically accurate age estimates (King et al. 2017), these findings support the validity of using FT-NIR predicted ages in age-based assessments 220 of longnose skates. Additionally, this method provides considerable efficiency gains -- a scan takes 221 222 just one minute and requires minimal preparation of the sample. Traditional methods for longnose skate age determination require extensive preparation and take between 15 and 30 minutes per 223 sample. Estimates can also be biased among age readers, especially among different agencies 224 (King et al. 2017). FT-NIR spectroscopy may allow age estimation to be standardized among age 225 readers and agencies due to its reproducibility. This technology has the potential to improve the 226 frequency, quantity, and reproducibility of longnose skate age data for use in stock assessments 227 and management. 228

This study marks the first known application of this technology to a batoid and results were comparable to those found by Rigby *et al.* (2014, 2016) for shark vertebrae. For *Sphyrna mokarran, Carcharhinus sorrah*, and *Squalus megalops*, Rigby *et al.* (2014, 2016) developed calibration models that yielded R^2 values between 0.78 and 0.89, RMSECV values of 1.23 to 2.48 years, and uncertainty between 7 and 9% (%RMSE of 7.40 to 8.97). The FT-NIR spectroscopy analysis of longnose skate vertebrae in this study produced comparable calibration models with an R^2 of 0.86, RMSECV of 1.38 years, and uncertainty of 9.87% (Fig. 3).

The biased age predictions we observed for age-0 longnose skates in the external validation relative to traditional methods could be due to the exceedingly small size of age-0 vertebrae (1.5 mm). Passerotti *et al.* (2020b) hypothesized that excess stray light from the FT-NIR spectrometer might confound their results for otoliths of juvenile red snapper that were between 1.5 and 7.0 mm. They found better calibration models when using a Teflon aperture to constrain the field of light. Future work for longnose skates could include using an aperture that may improve the predictive ability of the FT-NIR spectroscopy approach for younger skates.

Except for age 0 skates, the FT-NIR spectroscopy approach yielded age predictions for the 243 external validation set that had comparable precision to the traditional method currently used for 244 production ageing. However, neither method was as precise as is typical for teleost age 245 determination. Poor precision in age estimation is common for chondrichthyan species due to the 246 difficulty of interpreting their ageing structures. Published CVs for chondrichthyan vertebrae are 247 generally much higher (>10%) than those for teleost otoliths (~5%) (Campana 2001). The current 248 study reports slightly poorer precision in the traditional method (CV = 19.1%) than prior ageing 249 studies for longnose skates by Thompson (2006), Gburski et al. (2017), and King et al. (2017) that 250 found CVs between 11.9% and 15.2%. However, this is likely due to the large number of young 251 252 skates included in the current study. It is difficult to distinguish growth increments on longnose skate vertebrae under the age of 2 years and any disagreement within this age range is weighed 253 more heavily in the calculation of the overall CV. For instance, when one reader assigns an age of 254 255 0 and another assigns an age of 1 to a given individual, the disagreement dramatically inflates the CV. 256

The low precision of traditional age estimates in this study relative to teleosts is likely due
to observation error. However, the equivalently low precision of FT-NIR predictions could be due

to several factors. First, any observation error associated with the traditional age estimates used 259 to calibrate the model could reduce the precision of age predictions. Second, some process error is 260 likely present in vertebrae which could cause a difference in the visual versus spectral 261 interpretation of the structure. Third, any natural variation in chemical composition of vertebrae 262 may affect the spectral data collected using FT-NIR spectroscopy. This may be amplified because 263 264 we were unable to standardize the exact position of the thoracic vertebral samples used for spectroscopy. Size of vertebrae is variable along the vertebral column, and it is possible that larger 265 thoracic vertebrae could impart different FT-NIR spectra than smaller thoracic vertebrae within an 266 267 individual. Finally, the calibration model did not contain as many older skates as young skates, which could affect its predicative capability in that age range. Ideally, calibration samples would 268 have even representation from all age classes. Rigby et al. (2014) found that age predictions 269 markedly improved when older age classes were better represented in the calibration model. 270

An increasing number of studies show that vertebral banding patterns may not form 271 annually throughout life in all species (Natanson et al. 2018). This finding raises doubts about the 272 accuracy of age estimates from vertebral centra in general and suggests that process error may be 273 common. However, Rigby et al. (2014) found FT-NIR to be a promising alternative method for 274 275 estimating age from vertebrae even when no visible banding pattern is present. Rigby et al. (2016) 276 also found that using known-age samples to build predictive models based on FT-NIR spectra had 277 improved precision relative to using traditionally estimated ages with RMSECV of +/- less than a 278 year (0.87 and 0.88 years). This demonstrates the potential for FT-NIR spectroscopy to estimate age despite process error if accurate ages are used to calibrate the model. These findings indicate 279 280 that as the accuracy and representation of age estimates used to calibrate the model improve, we 281 may be able to improve the precision of age predictions. Future work for longnose skates could

include using a more balanced dataset to fit the calibration model between spectra and age
estimates, utilizing known-age specimens, and standardizing the position of centra along the
vertebral column. This may help to resolve bias in younger and older ages and improve precision
while also improving the accuracy of age estimates.

The results of this study as well as those of Rigby et al. (2014, 2016) indicate that there is 286 287 a correlation between age and the chemical composition of vertebrae. It is unknown what specific chemical compound(s) correlate with age in chondrichthyans, but it could be related to the quantity 288 of calcified phosphate mineral, hydroxyapatite, in their cartilaginous vertebral centra, as 289 290 mineralization occurs incrementally with age in many chondrichthyans (Cailliet 1990; Kerr and Campana 2014). Calcium hydroxyapatite is detectable in the NIR spectrum due to stretching and 291 292 bending of OH bonds in surface hydroxyl groups when exposed to near infrared light (Elkabouss et al. 2004). Relative quantities of the unmineralized components of vertebrae may also contribute 293 to the observed relationship between NIR spectra and age. The unmineralized components of 294 chondrichthyan vertebrae include water, proteoglycan, and collagen fibers, which are also 295 detectable in the NIR region between 5,400 and 3,800 cm⁻¹ (Baykal *et al.* 2010). Future work is 296 needed to determine the specific chemistry of chondrichthyan vertebrae related to age to better 297 298 understand the mechanism driving the observed correlation between age and FT-NIR spectra.

The results of this study show promise for the use of FT-NIR spectroscopy to more rapidly and efficiently estimate ages for longnose skates. The FT-NIR approach provides considerable efficiencies over the labor-intensive traditional process of preparing vertebrae for age determination. This is important because it may allow for a larger quantity and higher frequency of age estimates to be generated for use in stock assessments to monitor the status of this species. Additionally, modern stock assessment software allows the inclusion of ageing error in population

models and the results of this study provide the basis for quantifying this error. The most recent 305 assessment for longnose skate off the US West Coast included age-reading error matrices 306 estimated using the approach of Punt et al. (2008) based on various models of the relationship 307 between the CV of age-reading error and true age (Gertseva et al. 2019). This study also adds to a 308 growing body of literature demonstrating the successful application of this technology to estimate 309 310 age for chondrichthyan species. Fourier transform near infrared spectroscopy may provide a way to estimate age for other members of this sensitive group of fishes that previously had little to no 311 age data available. 312

313

314 Acknowledgments

We thank the vessel crews from the National Marine Fisheries Service's (NFMS) bottom trawl 315 surveys for providing vertebrae samples. Many thanks to Christopher Gburski for generously 316 assisting with this study by providing training, age estimates, and logistical support. We thank 317 project interns Emma Evans and Melinda Carr for wonderful assistance with sample preparation 318 and processing. Olav Ormseth, Brenna Groom, and two anonymous reviewers provided helpful 319 reviews that improved this manuscript. Fish research by the National Marine Fisheries Service is 320 321 not required to undergo ethical review as per the National Marine Fisheries Service Animal Care and Use Policy (Policy Directive 04-112). This policy (04–112) is currently limited to research 322 323 on free-living marine mammals, seabirds, and sea turtles and does not cover research on captive 324 or wild fish. However, fish samples used in this study were considered dead in the net upon arrival on board and collected and handled in strict accordance within the guidelines of the U.S. 325 326 Government Principles for the Utilization and Care of Vertebrate Animals Used in Testing, 327 Research and Training and the American Fisheries Society Guidelines for the Use of Fishes in

328	Research (https://fisheries.org/docs/policy_useoffishes.pdf; Chapter V). Permits acquired for							
329	sample collection including NOAA, NMFS, WCR (West Coast Region) Scientific Research							
330	Permit 2011: SRP-06-2011 and 2012: SRP-06-2012, Oregon Scientific Taking Permit for Fish							
331	and Marine and Freshwater Invertebrates: 2011: Permit 16328 and 2012: Permit 17200, CA							
332	Scientific Collecting Permit from the California Department of Fish and Game for 2011-2013:							
333	SC-11678, National Marine Sanctuary Permit for 2010 -2012: Multi-2010-004. The authors							
334	thank Aimee Keller for providing permit information. This research was supported by a grant							
335	from the NOAA Fisheries Office of Science and Technology's Improve a Stock Assessment							
336	program and a graduate fellowship from the University of Washington's School of Aquatic and							
337	Fishery Sciences. The scientific results and conclusions, as well as any views or opinions are							
338	those of the author(s) and do not necessarily reflect those of NOAA or the Department of							
339	Commerce. Reference to trade names does not imply endorsement by the National Marine							
340	Fisheries Service, NOAA.							
341								
342								
343	Conflicts of Interest							
344	The authors declare no conflicts of interest.							
345								
346	Data Availability Statement							
347	The data that support this study will be shared upon reasonable request to the corresponding							
348	author. More information available at https://www.fisheries.noaa.gov/resource/data/age-and-							
349	growth-otolith-collections-ageing-methods.							
250								

351 **References**

- Arrington, M. B. (2020). Growth and maturity of Longnose Skates (*Raja rhina*) along the North
- 353 American West Coast. M.Sc. Thesis, University of Washington, Seattle, WA.
- Baykal, D., Irrechukwu, O., Lin, P., Fritton, K., Spencer, R. G., and Pleshko, N. (2010).
- 355 Nondestructive assessment of engineered cartilage constructs using near-infrared spectroscopy.
- 356 *Applied Spectroscopy* **64**, 1160-1166.
- 357 Baumann, P. (2020). simplerspec: Soil and plant spectroscopic model building and prediction. R
- *package version 0.1.0.9001*. Available at https://github.com/philipp-baumann/simplerspec.
- Brown, C. D., Vega-Montoto, L., and Wentzell, P. D. (2000). Derivative preprocessing and
- optimal corrections for baseline drift in multivariate calibration. *Applied Spectroscopy* 54, 10551068.
- Cailliet, G. M. (1990). Elasmobranch age determination and verification: an updated review, pp.
- 363 157–165. In H. L. Pratt, S. H. Gruber, and T. Taniuchi (eds.), Elasmobranchs as Living
- Resources: Advances in the Biology, Ecology, Systematics, and the Status of the Fisheries.
- Proceedings of the Second United States–Japan Workshop, 9–14 December 1987, Honolulu, HI,
- 366 USA. U.S. Department of Commerce, NOAA Technical Report NMFS 90.
- 367 Cailliet, G. M., and Goldman, K. J. (2004). Age determination and validation in Chondrichthyan
- fishes. Pages 399-447 in J. C. Carrier, J. A. Musick, and M. R. Heithaus, editors. Biology of
- 369 Sharks and Their Relatives. CRC Press, Boca Raton, FL.

- 370 Cailliet, G. M., Smith, W. D., Mollet, H. F. and Goldman, K. J. (2006). Age and growth studies
- 371 of chondrichthyan fishes: the Need for consistency in terminology, verification, validation, and
- growth function fitting. *Environmental Biology of Fishes* **77**, 211-228.
- 373 Campana, S. E. (2001). Accuracy, precision and quality control in age determination, including a
- review of the use and abuse of age validation methods. *Journal of Fish Biology* **59**, 197-242.
- 375 Chang, W.Y.B. (1982). A statistical method for evaluating the reproducibility of age
- determination. *Canadian Journal of Fisheries and Aquatic Sciences* **39**, 1208-1210.
- Couture, J. J., Singh, A., Rubert-Nason, K. F., Serbin, S. P., Lindroth, R. L., and Townsend, P.
- A. (2016). Spectroscopic determination of ecologically relevant plant secondary metabolites.

379 *Methods in Ecology and Evolution* **7**, 1402-1412.

- 380 Dulvy, N. K., Fowler, S. L., Musick, J. A., Cavanagh, R. D., Kyne, P. M., Harrison, L. R.,
- 381 Carlson, J. K., Davidson, L., Fordham, S. V., Francis, M. P., Pollock, C. M., Simpfendorfer, C.
- A, Burgess, G. H., Carpenter, K. E., Compagno, L., Ebert, D. A., Gibson, C., Heupel, M. R.,
- Livingstone, S. R., Sanciangco, J. C., Stevens, J. D., Valenti, S., and White, W. T. (2014).
- Extinction risk and conservation of the world's sharks and rays. *eLife* **3**, 1-34.
- 385 Elkabouss, K., Kacimi, M., Ziyad, M., Ammar, S. and Bozon-Verduraz, F. (2004). Cobalt-
- exchanged hydroxyapatite catalysts: Magnetic studies, spectroscopic investigations, performance
- in 2-butanol and ethane oxidative dehydrogenations. *Journal of Catalysis* **226**, 16-24.
- Farrugia, T. J. (2017). Interdisciplinary assessment of the skate fishery in the Gulf of Alaska.
- 389 Ph.D. Diss., University of Alaska Fairbanks, Fairbanks, AK.

- Fowler, S. L., Cavanagh, R. D., Camhi, M., Burgess, G. H., Cailliet, G. M., Fordham, S.V.,
- 391 Simpfendorfer, C. A., and Musick, J.A. (2005). Sharks, Rays and Chimaeras: The Status of the
- 392 Chondrichthyan Fishes. IUCN/SSC Shark Specialist Group. IUCN, Gland, Switzerland and

393 Cambridge, UK.

- 394 Gertseva, V. Matson, S., Taylor, I. Bizzarro, J, Wallace, J. (2019). Stock assessment of the
- 395 Longnose Skate (*Beringraja rhina*) in state and Federal waters off California, Oregon and

396 Washington. Pacific Fishery Management Council, Portland, OR.

- 397 Gburski, C.M., Gaichas, S.K., and Kimura, D.K. (2007). Age and growth of Big Skate (*Raja*
- *binoculata*) and Longnose Skate (*R. rhina*) in the Gulf of Alaska. *Environmental Biology of Fishes* 80, 337-349.
- 400 Heino, M. and Dieckmann, U. (2008). Detecting fisheries-induced life-history evolution: an

401 overview of the reaction-norm approach. *Bulletin of Marine Science* **83**, 69-93.

- 402 Helser, T. E., Benson, I., Erickson, J., Healy, J., Kastelle, C., and Short, J.A. (2018). A
- transformative approach to ageing fish otoliths using Fourier transform near infrared
- 404 spectroscopy: a Case study of eastern Bering Sea walleye pollock (*Gadus chalcogrammus*).

405 *Canadian Journal of Fisheries and Aquatic Sciences* **76**, 1-10.

- 406 Hoenig, J. M., Morgan, M. J., and Brown, C. A. (1995). Analysing differences between two age
- 407 determination methods by tests of symmetry. *Canadian Journal of Fisheries and Aquatic*
- 408 *Sciences* **52**, 364-368.

409	Keller, A. K.	, Wallace, J. R.	, Methot, R. D.	(2017). The North	west Fisheries	Science Center's
-----	---------------	------------------	-----------------	-------	--------------	----------------	------------------

410 West Coast Groundfish Bottom Trawl Survey: History, Design, and Description. U.S.

411 Department of Commerce, NOAA Technical Memorandum NMFS-NWFSC-136.

- 412 Kerr, L.A., and Campana, S.E. (2014). Chemical composition of fish hard parts as a natural
- 413 marker of fish stocks. Pages 205-234 in S. X. Cadrin, L. A. Kerr, and S. Mariani, editors. Stock
- 414 Identification Methods 2nd Edition. Academic Press, San Diego.
- King, J. R., and McFarlane, G. A. (2003). Marine fish life history strategies: Applications to

fishery management. *Fisheries Management and Ecology* **10**, 249-264.

417 King, J. R., Helser, T., Gburski, C., Ebert, D. A., Cailliet, G., and Kastelle, C. R. (2017). Bomb

418 radiocarbon analyses validate and inform age determination of Longnose Skate (*Raja rhina*) and

- 419 Big Skate (*Beringraja binoculata*) in the North Pacific Ocean. *Fisheries Research* **193**, 195-206.
- 420 Kucheryavskiy, S. (2020). mdatools R package for chemometrics. Chemometrics and
- 421 Intelligent Laboratory Systems. Available at https://doi.org/10.1016/j.chemolab.2020.103937.
- 422 Lai, H. L., and Gunderson, D. R. (1987). Effects of ageing errors on estimates of growth,

423 mortality and yield per recruit for walleye pollock (*Theragra chalcogramma*). *Fisheries*

424 *Research* **5**, 287-302.

- Lorenzen, K., and Enberg, K. (2002). Density-dependent growth as a key mechanism in the
- 426 regulation of fish populations: evidence from among-population comparisons. *Proceedings of the*

427 *Royal Society B: Biological Sciences* **269**, 49-54.

- 428 Matta, M.E, Black, B.A, and Wilderbuer, T.K. (2010). Climate-driven synchrony in otolith
- growth-increment chronologies for three Bering Sea flatfish species. *Marine Ecology Progress Series* 413, 137-145.
- 431 Matta, M.E., Tribuzio, C. A., Ebert, D. A., Goldman, K. J., and Gburski, C. M. (2017). Age and
- 432 growth of elasmobranchs and applications to fisheries management and conservation in the

433 Northeast Pacific Ocean. *Advances in Marine Biology* 77, 179-220.

- 434 Matta, M. E., Helser, T. E., and Black, B. A. (2018). Intrinsic and environmental drivers of
- growth in an Alaskan rockfish: an Otolith biochronology approach. *Environmental Biology of Fishes* 101, 1571-1587.
- 437 McClure, W., Crowell, B., Stanfield, D., Mohapatra, S., Morimoto, S., and Batten, G. (2002).
- 438 Near infrared technology for precision environmental measurements: Part 1. Determination of
- 439 nitrogen in green-and dry-grass tissue. *Journal of Near Infrared Spectroscopy* **10**, 177-185.
- McBride, R. (2015). Diagnosis of paired age agreement: a simulation of accuracy and precision
 effects. *ICES Journal of Marine Science* 72, 2149-2167.
- 442 Natanson, L. J., Skomal, G. B., Hoffman, S. L., Porter, M. E., Goldman, K. J., and Serra, D.
 443 (2018). Age and growth of sharks: Do vertebral band pairs record age? *Marine Freshwater*
- 444 *Research* **69**, 1440-1452.
- 445 Passerotti, M. S., Helser, T. E., Benson, I. M., Barnett, B. K., Ballenger, J. C., Bubley, W. J.,
- 446 Reichert, M. J. M., and Quattro, J. M. (2020a). Age estimation of Red Snapper (Lutjanus
- 447 *campechanus*) using FT-NIR spectroscopy: Feasibility of application to production ageing for
- 448 management. *ICES Journal of Marine Science* **77**, 2144–2156.

- 449 Passerotti, M. S., Jones, C. M., Swanson, C. E., and Quattro, J. M. (2020b). Fourier-transform
- 450 near infrared spectroscopy (FT-NIRS) rapidly and non-destructively predicts daily age and
- 451 growth in otoliths of juvenile red snapper *Lutjanus campechanus* (Poey, 1860). *Fisheries*

452 *Research* **223**, 1-8.

- 453 Pistevos, J. C. A., Nagelkerken, I., Rossi, T., Olmos, M., and Connell, S. D. (2015). Ocean
 454 acidification and global warming impair shark hunting behaviour and growth. *Scientific Reports*455 5, 1-10.
- 456 Punt, A. E., Smith, D. C., KrusicGolub, K., Roberston, S. (2008). Quantifying age-reading error
- 457 for use in fisheries stock assessments, with application to species in Australia's southern and
- 458 eastern scalefish and shark fishery. *Canadian Journal of Fisheries and Aquatic Sciences* 65,
 459 1991-2005.
- 460 R Core Team (2020). R: A Language and Environment for Statistical Computing, Vienna,
- 461 Austria. Available at: https://www.R-project.org/.
- 462 Rigby, C.L., Wedding, B. B., Grauf, S., and Simpfendorfer, C. A. (2014). The utility of near
- 463 infrared spectroscopy for age estimation of deepwater sharks. *Deep-sea Research Part I:*
- 464 *Oceanographic Research Papers* **94**, 184-194.
- Rigby, C. L., Wedding, B. B., S. Grauf, and Simpfendorfer, C. A. (2016). Novel method for
 shark age estimation using near infrared spectroscopy. *Marine and Freshwater Research* 67,
- 467 537-545.
- 468 Rinnan, A., van den Berg, F., Engelsen, S. B. (2009). Review of the most common pre-
- 469 processing techniques for near-infrared spectra. *Trends in Analytical Chemistry* **28**, 1201-1222.

- 470 Robins, J. B., Wedding, B. B., Wright, C., Grauf, S., Sellin, M., Fowler, A., Saunders, T., and
- 471 Newman, S. J. (2015). Revolutionising Fish Ageing: Using Near Infrared Spectroscopy to Age
- 472 Fish. Department of Agriculture, Fisheries and Forestry. Brisbane, April, 2015. CC BY 3.0.
- 473 Roggo, Y., Chalus, P., Maurer, L., Lema-Martinez, C., Edmond, A., and Jent, N. (2007). A
- 474 review of near infrared spectroscopy and chemometrics in pharmaceutical technologies. *Journal*
- 475 *of Pharmaceutical and Biomedical Analysis* **44**, 683-700.
- 476 Savitzky, A., and Golay, M. J. E. (1964). Smoothing and Differentiation of Data by Simplified
- 477 Least Squares Procedures. *Analytical Chemistry* **36**, 1627-1639.
- 478 Shelton, A. O., and Mangel, M. (2012). Estimating von Bertalanffy parameters with individual
- and environmental variations in growth. *Journal of Biological Dynamics* **6**, 3-30.
- 480 Stokes, K., and Law, R. (2000). Fishing as an evolutionary force. *Marine Ecology Progress*481 *Series* 208, 307-309.
- 482 Thompson, J. E. (2005). Age, Growth and Maturity of the Longnse Skate (*Raja rhina*) for the US
- 483 West Coast and Sensitivity to Fishing Impacts. M.Sc. Thesis. State University, Oregon.
- 484 Vance, C.K., Tolleson, D.R., Kinoshita, K., Rodriguez, J., and Foley, W.J. (2016). Near infrared
- 485 spectroscopy in wildlife and biodiversity. *Journal of Near Infrared Spectroscopy* 24, 1-25.
- 486 Wold, S., Ruhe, A., Wold, H., and Dunn III, W. J. (1984). The collinearity problem in linear
- 487 regression. The Partial least squares (PLS) approach to generalized inverses. Society for
- 488 *Industrial and Applied Mathematics* **5**: 735-743.

- Wedding, B.B., Forrest, A.J., Wright, C., Grauf, S., Exley, P., and Poole, S. E. (2014). A novel
- 490 method for the age estimation of saddletail snapper (*Lutjanus malabaricus*) using Fourier
- 491 transform near infrared (FT-NIR) spectroscopy. *Marine and Freshwater Research* **65**, 894-900.
- 492 Wright, C., Wedding, B.B., Grauf, S., and Whybird, O.J. (2021). Age estimation of barramundi
- 493 (Lates calcarifer) over multiple seasons from the southern Gulf of Carpentaria using FT-NIR
- 494 spectroscopy. *Marine and Freshwater Research.*

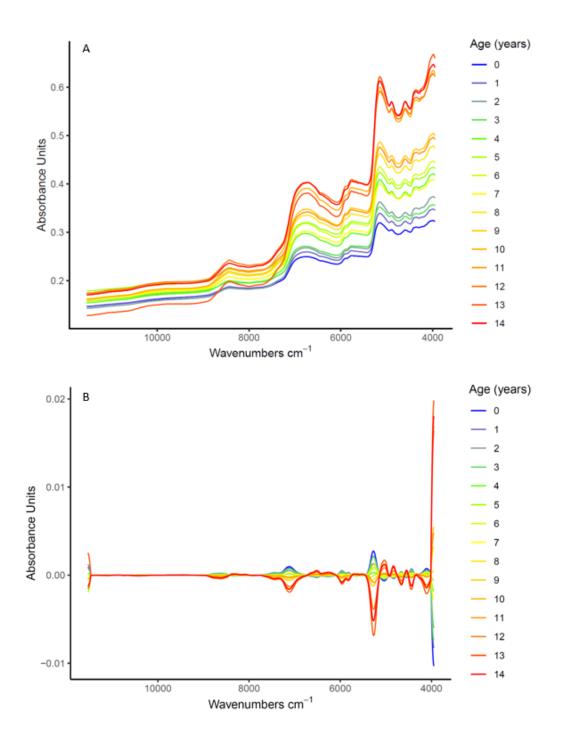


Figure 1. Fourier transform near infrared spectral absorbance data of all Longnose Skate
vertebrae. Spectra were averaged by traditionally estimated age, represented in colors: (A) raw
absorbance data and (B) pre-processed spectral data using a 1st derivative Savitzky-Golay
transform (17-point smooth, polynomial order = 1) and mean centering.

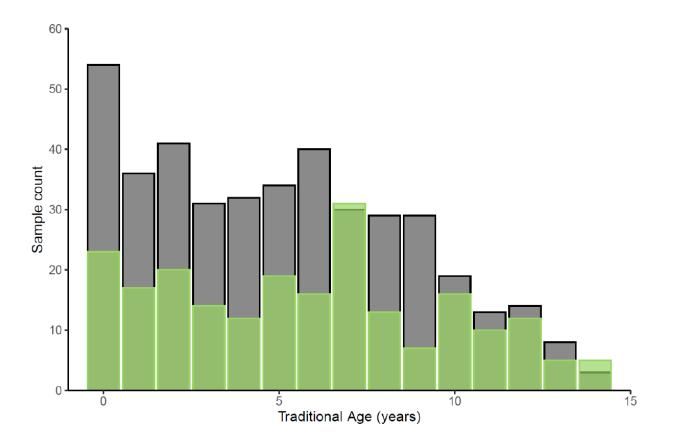


Figure 2. Age-specific sample sizes for the calibration set (grey) overlapped by external
validation set (green) used in the age estimation of longnose skate by Fourier transform near
infrared (FT-NIR) spectroscopy.

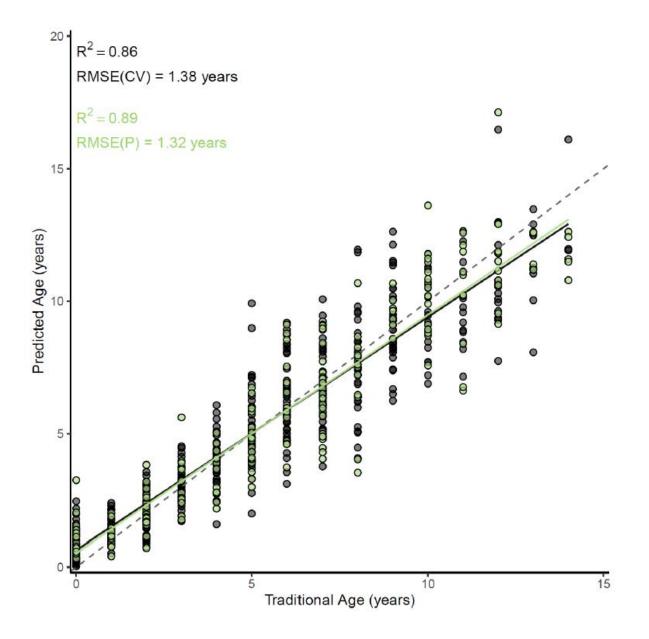


Figure 3. Results of a partial least squares model fit between reader-estimated age (traditional age) and Fourier transform near infrared spectra (predicted age) of Longnose Skate vertebral centra. Age predictions relative to traditionally estimated ages shown for the cross-validation (black) and for the external validation set (green) for each specimen. The solid lines are the regression lines for the cross-validation (black) and external validation (green). The dashed line represents one-to-one agreement between prediction and traditional estimate. Transparency in point shading shows overlapping data points.

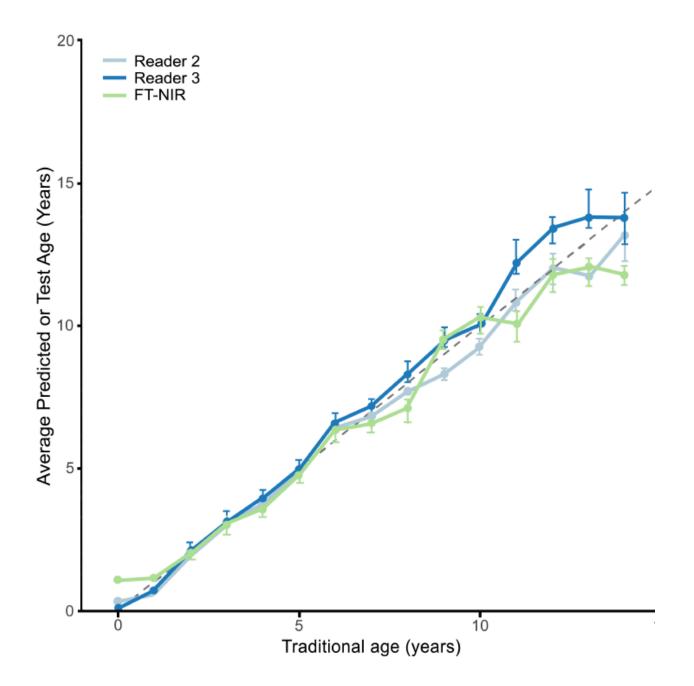


Figure 4. Bias plot comparing Reader 1's traditional age estimates to: average age estimates of
Reader 2 (light blue), Reader 3 (in dark blue) and Fourier transform near infrared (FT-NIR)
predictions (green) for longnose skates. Standard error bars shown for age groups with multiple
samples. Dashed line represents 1:1 agreement. Sample count for each age category are
represented by grey bars.

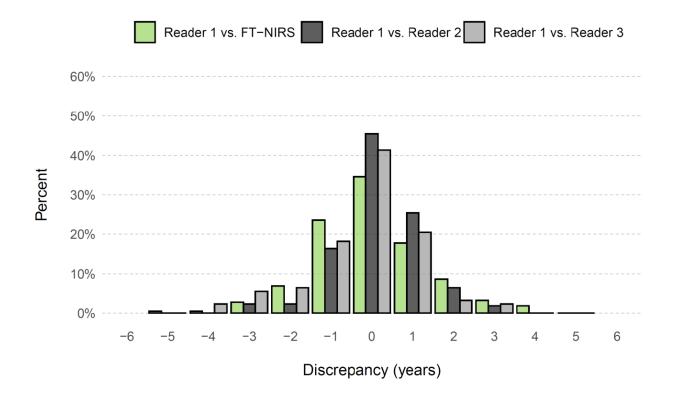


Figure 5. Comparing discrepancies in longnose skate (Raja rhina) age estimates for the validation
set from Fourier transform near infrared spectroscopy (FT-NIRS) and three age readers using
traditional methods. Discrepancies are shown as percentage of specimens with age estimate
differences of 0 to 6 years. In grey: traditional age estimated by Reader 1 – 2 and Reader 1 – 3,
and in green: Reader 1 – FT-NIR prediction.