

1 | INTRODUCTION

2 Growing human populations in tropical coastal areas and the associated coastal development and
3 changes in land use have caused declines in water quality (increased nutrients and suspended
4 sediments) in nearshore environments, with these declines in water quality being linked to changes
5 in benthic communities on nearshore coral reefs (Brodie et al., 2012; Brown et al., 2019; Fabricius et
6 al., 2012; Oliver et al., 2011). Increased dissolved nutrients, such as inorganic and organic forms of
7 nitrogen and phosphorus from agriculture, can result in increases in turf and macroalgae abundance
8 and/or benthic heterotrophic filter feeding taxa (Fabricius, 2005; Littler et al., 2006; Szmant, 2002).
9 Additionally, increased inputs of suspended particulate matter (SPM) can reduce light penetration and
10 hence photosynthetic efficiency of corals and algae, while increases in fine terrigenous sediments (<
11 20 μm) can smother corals and inhibit coral settlement, leading to changes in the assemblage
12 structure, composition, and cover of coral assemblages (Bainbridge et al., 2018; Weber et al., 2006).
13 Declining water quality, together with increasing seawater temperatures, is one of the most significant
14 stressors affecting the health and functions of coastal reef ecosystems, and provisioning of ecosystem
15 services (Burke et al., 2011; De'ath and Fabricius, 2010; Wooldridge, 2009). There is a clear need to
16 identify the key pollutants that impact water quality at localized coral reef sites so they can be better
17 managed.

18 Across tropical Pacific island countries, growing coastal populations, agriculture, coastal
19 development, and varying land uses have led to substantial and sustained declines in water quality
20 flowing into coastal marine environments (Falkland, 1999). Indeed, coastal development has been
21 estimated to threaten about 20% of reefs in the region, with another 25% of reefs being impacted by
22 watershed-based pollution (Burke et al., 2011). Given the tight coupling of watershed condition,
23 nearshore water quality, and coral reef health, a growing number of studies have focused on
24 integrated ridge-to-reef approaches to simultaneously address and manage upstream activities and
25 downstream ecological conditions (Adam et al., 2020; Comeros-Raynal et al., 2019; Delevaux et al.,
26 2018; Houk et al., 2020; Rodgers et al., 2012; Rude et al., 2016; Wenger et al., 2020). Despite
27 improved understanding of the effects of declining water quality on reef condition in the Pacific region,
28 our understanding of the relative effects of enriched nutrients and suspended particulate matter in
29 influencing the composition of benthic assemblages on nearshore reefs, particularly in high islands,
30 remains limited. Despite catchments on high islands being generally smaller than those found on
31 continents (Jupiter et al., 2017; Ruddle et al., 1992), high volcanic islands are especially vulnerable
32 to land use and land cover modifications due to their lithology (erosive andesite soils) and steep
33 topography (Verbist et al., 2010). The strong land-sea connection and intrinsic vulnerability of high
34 volcanic islands warrants expansion of research efforts in island communities (Carlson et al., 2019).

1 Tropical high islands present an opportunity to examine the effects of increasing nutrients and
2 sediment loads on the composition of benthic assemblages on nearshore reefs due to a combination
3 of complex geomorphology, strong seasonal precipitation patterns, coastline configuration comprising
4 replicate bays and fringing reef systems, and the varied distribution of human populations and land
5 use activities. Moreover, many high islands, such as Tutuila, American Samoa, are large enough to
6 provide a series of spatially distinct sampling sites, and are nested within similar oceanographic and
7 meteorological regimes. Concentrated human populations in the coastal plains, and associated soil
8 erosion from disturbance to vegetation highlight the predisposed vulnerabilities of nearshore coral
9 reefs to the negative impacts of human land uses in watersheds (Holst Rice et al., 2016; Houk et al.,
10 2010; Messina and Biggs, 2016; Shuler et al., 2019; Vargas-Angel and Huntington, 2020; Whitall et
11 al., 2019). Tutuila is located in the South Pacific Convergence Zone where there is abundant rainfall
12 (Kennedy, 1987; Shuler and El-Kady, 2017) potentially leading to higher transport rates of nutrients
13 and sediments to nearby coral reefs from watersheds with highly modified landscapes.

14 Here, we quantified water quality across six watersheds spanning a gradient in anthropogenic impact
15 on Tutuila, and relate this to variation in benthic assemblages on adjacent shallow coral reefs.
16 Specifically, we quantified total and dissolved nutrient concentrations from streams and adjacent reef
17 flats, and the accumulation rate, composition, and particle size of sediments captured in sediment
18 traps at 3-monthly intervals for one year. We examined the spatial groupings of nutrients and
19 sediments, and the composition of the benthos to determine if clustering followed local watershed
20 classifications. Lastly, we used exploratory path analyses to simultaneously examine the relationships
21 between natural (e.g., surface runoff, wave exposure), and anthropogenic drivers (intensive land use,
22 DIN load, stream and reef flat nutrients) and benthic cover. Monitoring terrestrial run-off and relating
23 its influence on benthic condition at sampling units relevant to management can help support
24 integrated scientific and management approaches in American Samoa and potentially can be adapted
25 to other oceanic high islands where ridge-to-reef approaches are implemented.

26

27 **2 | METHODS**

28 **2.1 | Study Location**

29 American Samoa has five main volcanic high islands (Tutuila, Aunu'u, Ofu, Olosega, and Ta'u) and
30 two atolls (Rose and Swains), and is the southernmost U.S. Territory in the South Pacific at 14.27°S,
31 170.13°W. Our study was conducted on Tutuila, the largest of the main high islands in American
32 Samoa with an area of 138 km² and the most populous with a population of 55,000 (US Census
33 Bureau, 2014). Tutuila is an extensively eroded volcanic island comprised of a central ridge of steep
34 mountains which lead sharply to a narrow coastline (Atkinson and Medeiros, 2005; Craig et al., 2010).
35 Tutuila has a tropical climate with uniform temperatures between 26 and 28 °C and high humidity

1 throughout the year. The mean annual rainfall on Tutuila is 3,810 mm/year with peak rainfall during
2 the wet season from October to April and lower rainfall occurring in the dry season from May to
3 September (Izuka et al., 2005; Wong, 1996). Precipitation on the islands generally increases with
4 elevation and ranges from 2,388 mm/year at the shorelines to 6,350 mm/year at ~480 m above sea
5 level elevation (Meyer et al., 2017).

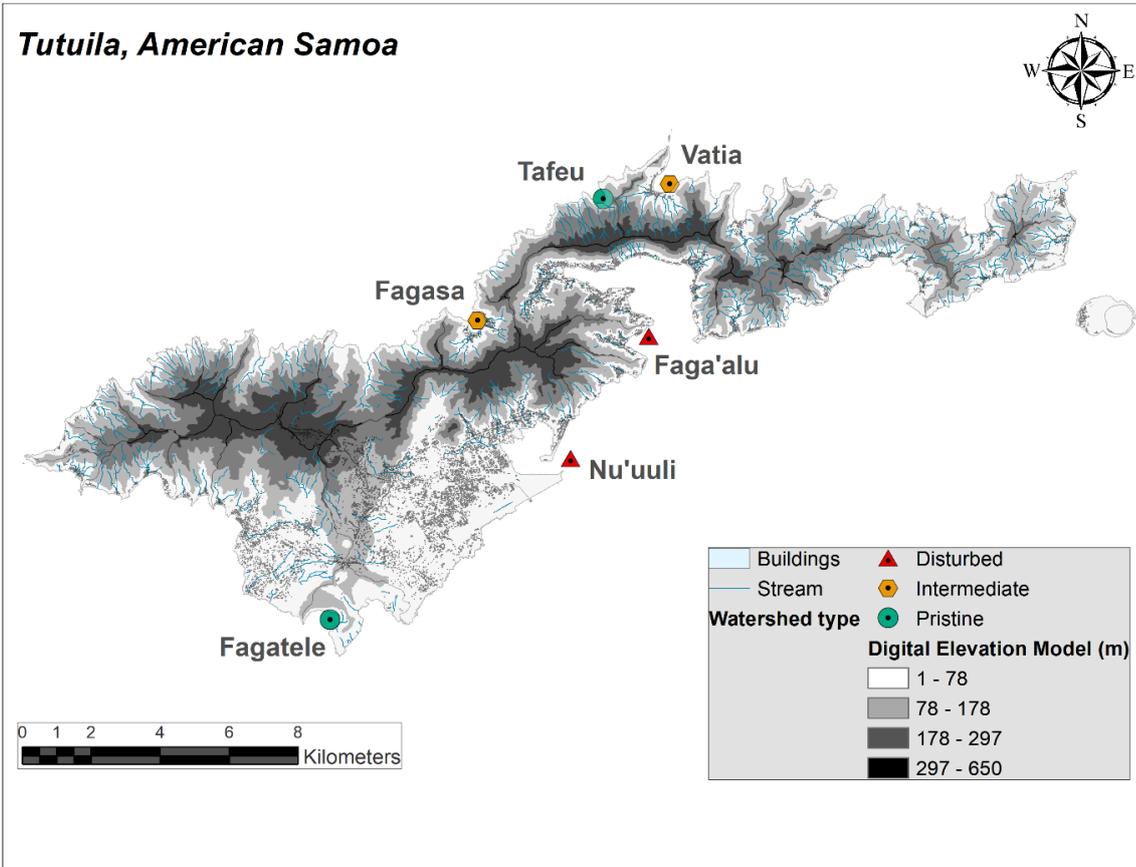
6

7 Land Use and Land Cover

8 Over 65% of Tutuila is natural forest while agriculture and development combined covers 24% of the
9 island and is concentrated on the south-western coast (Meyer et al., 2017). From 2004 to 2010, there
10 has been a 4.8% increase in developed land and a 6.8% rise of impervious surfaces, while agriculture
11 has increased by 17% (NOAA, 2020).

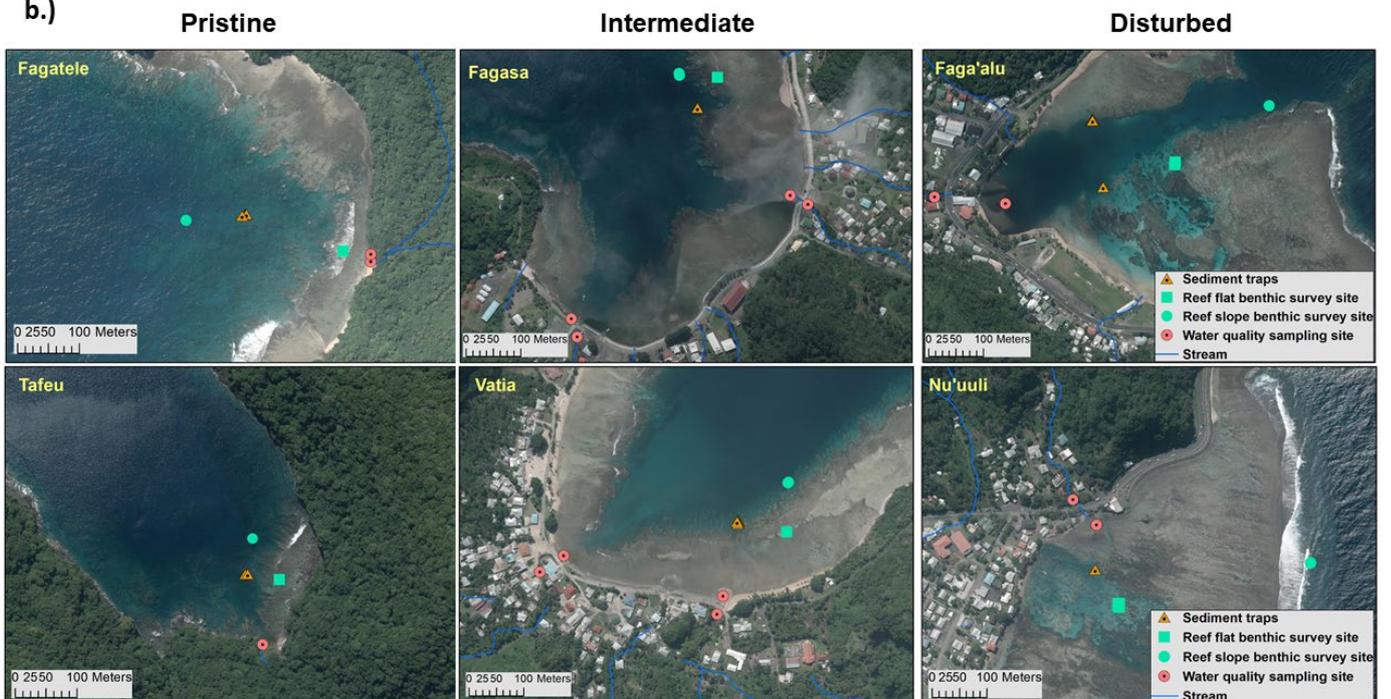
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37

1 a.)



2

b.)



3

4 Figure 1. Map of Tutuila, American Samoa showing the location of the six study sites. (a) Map of
5 Tutuila showing the location of two study sites within each of three watershed types as classified by
6 the American Samoa Environmental Protection Agency (AS-EPA). Red triangles: disturbed; orange

1 hexagon: intermediate; green circle: pristine. (b) Satellite images of the six sites showing the
2 approximate location of data collection sites for water quality, sediments, and benthic surveys.

3

4 **2.2 | Study design**

5 Water quality and benthic assemblage data were collected from six sites that spanned a gradient of
6 anthropogenic impact in the adjacent watersheds (Fig. 1). Specifically, we selected two sites within
7 each of three watershed classifications (pristine, intermediate, and extensive) characterized by the
8 American Samoa Environmental Protection Agency (DiDonato, 2004; Tuitele et al., 2016). For clarity,
9 the term 'disturbed' is used hereafter when referring to extensive watersheds, and is classified as
10 having a human population density of >290 individuals km^{-2} . Disturbed watersheds (Nu'uuli and
11 Faga'alu) had large watershed areas (i.e., >2.33 km^{-2}) and were characterized by greater proportions
12 of disturbed land (28% and 13% of the total watershed area, respectively). Similarly, intermediate
13 watersheds (Fagasa and Vatia) had large watershed areas and low to moderate proportions of
14 disturbed land (7% and 5%, respectively). Pristine watersheds (Fagatele and Tafeu) had the smallest
15 watershed sizes (i.e., <1.55 km^{-2}) and low human population density of <0.38 individuals km^{-2} .
16 However, the Fagatele watershed includes the major landfill on Tutuila, and had the highest agroforest
17 and cultivated land percent cover (Table 1).

18

19 **2.4 | Biological surveys**

20 Benthic composition was quantified along four replicate 50 m point-intercept transects within each of
21 two habitats, the reef flat (1 – 4 m depth) and reef slope (5 – 9 m depth), at each of the six sites in
22 May 2019. The substratum directly under the transect tape was recorded at 50cm intervals along
23 each transect (n=101 points per transect). Benthic categories were recorded as crustose coralline
24 algae (CCA), turf algae (primarily filamentous algae <10 mm in height), macroalgae (>10 mm in
25 height), and hard coral. Hard coral and macroalgae were identified to genus. Transects were laid
26 along the reef profile with a minimum of 10 m between adjacent transects.

27 **2.3 | Environmental data**

28 To evaluate the relationships of terrestrial run-off on nearshore shallow reef habitats, water quality
29 data were collected at 3-monthly intervals for 12 months across the six watersheds to account for
30 baseflow conditions and storm events over the two rainfall seasons in American Samoa: from June
31 through September representing the drier winter season, and from October through May, representing
32 the wetter summer season (Izuka et al., 2005). The major streams in each watershed were sampled,
33 with a single stream sampled in Faga'alu, Fagatele, Nu'uuli and Tafeu, and two streams in Vatia
34 (Gaoa and Faatafe) and Fagasa (Leele and Agasii). Water samples were collected every 3 months

1 from August 2018 – May 2019, and included two sampling periods in the wetter summer months:
2 November 2018 and February 2019; and in the drier winter months: August 2018 and May 2019.
3 Water samples were collected from the mouth of each stream at approximately 1 m above sea level
4 during low tide to minimize mixing of coastal water during each sampling period. Upstream land use
5 distance to stream mouth varied, with some point-source/cleared areas within 1 km of the sampling
6 site. Stream mouth distance to the reef flat and reef slope sampling sites varied across sites (50-550
7 m) due to inherent differences in reef geomorphology and stream locations across sites. Sediment
8 samples were collected using SediSampler® patented traps deployed on the reef slope at depths of
9 5-7 m.

10 Water samples were taken from the surface waters of the stream and reef flat by rinsing 500 ml and
11 60 ml polyethylene bottles three times with sample water prior to filling. Samples were placed on ice
12 after collection and returned to the laboratory. Samples in 60 ml bottles were immediately frozen
13 (unfiltered samples for total nitrogen and total phosphorus analysis), and the samples in 500 ml bottles
14 were filtered using 0.7 µm GF/F Whatman filters, and the filtrate stored frozen until analysis. Filtration
15 was conducted to remove most bacteria and other microorganisms that could affect the stability of
16 filtered nutrient constituents.

17 The frozen water samples were then sent to the University of Hawaii's SOEST Laboratory for
18 Analytical Biogeochemistry (S-LAB) for analysis of dissolved nutrients: sum of nitrate and nitrite (N+N;
19 and ammonium (NH⁴⁺), hereafter referred to as dissolved inorganic nitrogen (DIN)), ammonium
20 (NH⁴⁺), phosphate (PO₄³⁻), silicate (SiO₄⁴⁻); and Total Nitrogen (TN), and Total Phosphorus (TP)
21 (Armstrong et al., 1967; Grasshoff et al., 1983; Kérouel and Aminot, 1997; Murphy and Riley, 1962).
22 Subsequent analysis of stable isotope of dissolved nitrate was conducted by the Biogeochemical
23 Stable Isotope Facility at the University of Hawaii using the denitrifier method on a Thermo Finnigan
24 MAT 252 Mass Spectrometer using a continuous flow GC-interface with a Triplus autosampler
25 (McIlvin and Casciotti, 2010; Sigman et al., 2001).

26 *Suspended particulate matter*

27 To quantify suspended particulate matter (SPM) matter (i.e. sediments and associated particulate
28 matter), we deployed three SediSampler® patented traps (Integral Aqua Pty Ltd) on the reef slope at
29 each site in 5-7 m depth (Figure 1b). Each trap was attached to a steel bar driven into the substratum
30 so that the mouth of the trap was positioned approximately 1 m above the substratum (refer to Lewis
31 et al. 2020). The sediment traps were deployed at the time of the water sampling (i.e., November
32 2018, February 2019, May 2019, and September 2019) and collected after ~3 months. An additional
33 three SediSampler® traps were placed at the mouth of the stream in Faga'alu to account for the
34 previously reported differences in the spatial distribution of sediments at this site (Holst Rice et al.,
35 2016; Messina, 2016; Messina and Biggs, 2016). After three months deployment on the reef, the 1 L

1 sample bottles were carefully removed from the SediSampler[®] traps and the bottles capped
2 underwater to avoid loss of sediments. The sample bottles were placed on ice within 10 minutes of
3 collection and transferred to a refrigerator until sample processing.

4 In the laboratory, each 1 L sediment trap sample bottle from each site was transferred to individual
5 containers that had been pre-rinsed with distilled water, and the samples were well mixed for two
6 minutes to ensure even distribution of particles prior to subsampling. 21 aliquots (each 30 ml) were
7 collected from each sample for Total Suspended Solids (TSS) analysis (APHA et al., 2012). The 21
8 aliquots for TSS analysis were then placed in a refrigerator and the remaining sample (~370 ml)
9 prepared for salt removal. Similar to SPM, TSS includes both terrestrially-derived and marine-derived
10 organic matter and mineral sediment. Thus, we refer to TSS as SPM from here on. The remaining
11 wet sediment samples from each sediment trap were transferred to individual 1 L plastic beakers, and
12 left for 24-48 hours to allow sediment particles to settle. The supernatant was then decanted, taking
13 care to ensure sediment particles were not lost. Distilled water (900 ml) was then added to each
14 sample, agitated for two minutes to ensure mixing, allowed to settle for 24 hours and the supernatant
15 carefully decanted as described above. This process of rinsing in distilled water, settling, and
16 decanting was repeated until salinity was < 200 $\mu\text{s/cm}$ (i.e., 3-4 rinses). The three sediment trap
17 samples from each site were then combined in a pre-rinsed container and agitated for 2 minutes.
18 Seven 30ml aliquots were collected from the combined sample for particle size analysis.

19 The remaining bulk sample from the combined trap sediment samples were left to settle for 24 hours,
20 decanted, and transferred to a 1 L beaker. Samples were then dried in an oven at 60°C for 24-48
21 hours with approximately 7 individual bags per deployment for sediment composition analysis. A total
22 of 28 wet samples (30 ml tubes) and 7 dried samples from each deployment were then transported
23 to Australia and analyzed at the TropWATER Laboratory, James Cook University (JCU) for
24 Suspended Particulate Matter and Loss on Ignition (LOI), and the School of Earth and Environmental
25 Science laboratory, JCU for particle size analysis.

26 The Loss on Ignition method was used to determine proportions of mineral, organic, and carbonate
27 content by consecutively weighing the dried sediment samples after heating at suitable temperatures
28 (Heiri et al., 2001). Sample organic matter content was determined using Standard Method 2540E
29 (APHA et al., 2012). Briefly, a crucible containing a pre-weighed quantity of each sediment trap
30 sample (dried at 105°C to remove moisture) was ignited in a Carbolite (AAF1100) ashing furnace at
31 550°C for 4 to 5 hours, and reweighed. The weight lost during ignition represents the total volatile
32 solids, an approximation of the organic matter content of each sample (weight % LOI, 550°C)
33 (Bainbridge et al., 2012). To determine marine sediment trap carbonate content, the sample crucibles
34 were returned to the ashing furnace and heated at 950°C for at 2 hours, with the weight lost on ignition
35 representing carbon dioxide released from calcium carbonate (Bainbridge et al., In review).

1 The TSS method was used to measure the sediment accumulation within each sediment trap (n=21
2 per deployment). Briefly, we pipetted 1ml of wet sample from each sediment trap with-salt aliquot,
3 into a filtering manifold which was prefilled with 250 ml of RO water, and filtered the measured volume
4 through a pre-weighed Whatman GF/C (1.2 μm pore size) filter paper (Lewis et al., 2020). The filter
5 paper was then dried for ~24 hours at 105°C and weighed again to measure the weight of total
6 suspended solids (mg/L). This concentration was then converted to accumulation rate per day per
7 cm^2 using the length of each deployment and the internal cross-sectional area (21.52 cm^2) of the
8 sediment trap head (i.e. $\text{mg}/\text{cm}^2/\text{day}$).

9
10 Particle size distributions from salt removed wet samples were determined using the Malvern
11 Mastersizer 3000, a laser diffraction particle-size analyzer following the parameterization method of
12 Sperazza et al. (2004) and Bainbridge et al. (2012). This analysis was conducted on a sub-sample
13 of collected trap material, and includes mineral, organic and carbonate components. Particle sizes
14 were reported as percentage distributions D10, D50, and D90. For example, D50 refers to median
15 size particles, where the diameter of a sphere at which 50% of the particles in the sample is smaller.

16 *Environmental and land use variables*

17 We quantified land use, environmental (i.e., rainfall and wave energy), and anthropogenic (surface
18 runoff, and Dissolved Inorganic Nitrogen (DIN) load) factors for each of the six sites. We used ArcMap
19 10.4 to calculate percent cover of land that was forest, agroforest, cultivated, developed and other
20 land-use types using high resolution and LIDAR remote sensing habitat maps produced by the
21 American Samoa Department of Marine and Wildlife Resources (Meyer et al., 2017). Monthly
22 modeled average discharge rates (rainfall and surface runoff) for each of the six watersheds were
23 estimated using an open-source water budget model for Tutuila (Shuler et al., 2021). Wave energy
24 for each site was calculated using 10-year average wind speeds for Tutuila using the Wave Energy
25 tool in ArcGIS (Jenness and Houk, 2014). DIN loads exported from each watershed were taken from
26 Shuler and Comeros-Raynal (2020).

27 **2.5 | Data analysis**

28 We used principal component analyses (PCA) to visualize variation in (i) benthic assemblages, (ii)
29 nutrient concentrations, and (iii) suspended sediment characteristics among sampling locations. PCA
30 were conducted using the “vegan” package (Oksanen et al., 2019) in R version 4.0.2. Analysis of
31 benthic assemblages were based on the correlation matrix of the transect level data of each of the
32 benthic categories, for water quality data based on quarterly collections, and for suspended sediments
33 based on 3-month trap deployments. Separate PCA were performed for each of the two habitats
34 (reef flat and reef slope) for benthic assemblages, and each of two sampling locations (stream and
35 reef flat) for water quality.

1 We used path analysis to explore the potential influences of both natural and anthropogenic variables
2 relating to the catchment and wave exposure and dissolved nutrient concentrations on benthic cover
3 in shallow coastal reef flats. Natural variables comprised of wave energy, watershed size and
4 discharge rates, while anthropogenic variables included percent cover disturbed land area (% cover
5 of developed, cultivated and agroforest land uses), and a proxy of human population density from DIN
6 loads of onsite disposal systems and piggeries ($\text{kg}\cdot\text{day}^{-1}$). Of the water quality parameters quantified,
7 DIN and phosphate concentrations were used as explanatory variables as they are highly bioavailable
8 and are known indicators of anthropogenic nutrient loading on shallow reef systems. Benthic cover
9 was partitioned into two groups: turf and macroalgae, and hard coral and CCA as we expected that
10 responses to nutrient enrichment will vary between these different benthic components. Importantly,
11 hard corals together with CCA play key roles in CaCO_3 accretion into the reef matrix, and form
12 desirable reef health indicators from a management perspective (Houk et al., 2015; Littler and Littler,
13 2007).

14 Path analysis is a multivariate technique which uses a series of structured linear regression equations
15 to test the specified relationships between measured variables (Pedhazur, 1997). The relationships
16 are displayed in a path diagram where variables are linked by straight arrows indicating the direction
17 of the relationship between the variables (Streiner, 2005). In a path diagram, variables are
18 represented as rectangles and are either exogenous or endogenous (Lleras, 2005). The direct effects
19 of an independent variable on a dependent variable are expressed as path coefficients. Coefficients
20 are positive where an increase in the independent variable causes an increase in the dependent
21 variable when other causal variables are held constant, or negative where an increase in the causal
22 variables decreases the dependent variable. Model fit was assessed using four tests: X^2 , Comparative
23 Fit Index (CFI), Root Mean Square Error of Approximation (RMSEA) and Standardized Root Mean
24 Square Residual (S)RMR (Kline, 1998; Lleras, 2005). Assumptions of the linear models were
25 validated using the *gvlma* package (Peña and Slate, 2006). All linear regressions performed met
26 model assumptions.

27 **3 | RESULTS**

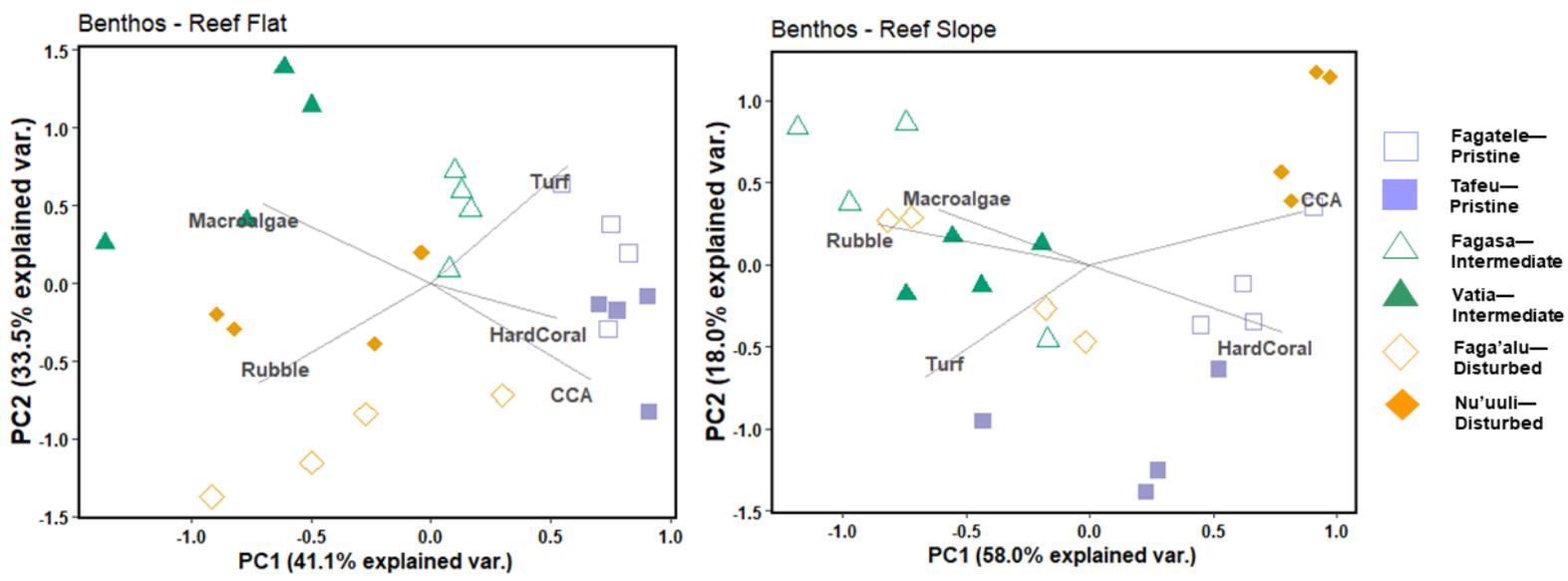
28 The PCA of both reef flat and reef slope benthic assemblages revealed a clear partitioning among
29 watershed types. For the reef flat, the first two principal components explained 41.1% and 33.5% of
30 the total variation, respectively, with sites within pristine watersheds being separated from
31 intermediate and disturbed watersheds along PC1, and intermediate and disturbed watersheds being
32 differentiated along PC2 (Figure 2a). Reef flat benthic assemblages within pristine watersheds were
33 represented by a relatively high percent cover of live coral, CCA, and turf algae, whereas those in
34 intermediate and disturbed watersheds were represented by a higher percent cover of macroalgae
35 and rubble, respectively (Figures 2a, S1). For the reef slope, the first two principal components

1 explained 58% and 18% of the total variation, respectively, with sites within pristine watersheds being
 2 represented by relatively high cover of hard coral cover and CCA and separated from intermediate
 3 watersheds that were represented by a high cover of macroalgae and rubble along PC1 (Figures 2b,
 4 S1). There were considerable variations in reef slope benthic assemblages between the two
 5 disturbed sites with Fagaalu being characterized by a relatively high cover of macroalgae and rubble,
 6 while Nu'uuli was defined by a high cover of CCA (Fig 2b).

7

8 a.)

b.)



9

10 Fig 2. Principal Component Analysis (PCA) showing variation in benthic assemblages on (a) the reef
 11 flat, and (b) the reef slope among watershed types (pristine, intermediate, disturbed), and six reef
 12 sites around Tutuila, American Samoa. Analyses are based on the cover of benthic categories along
 13 four 50 m point-intercept transects within each habitat at each site. Vector lengths are proportional to
 14 correlation strengths with the primary PCA axes.

15

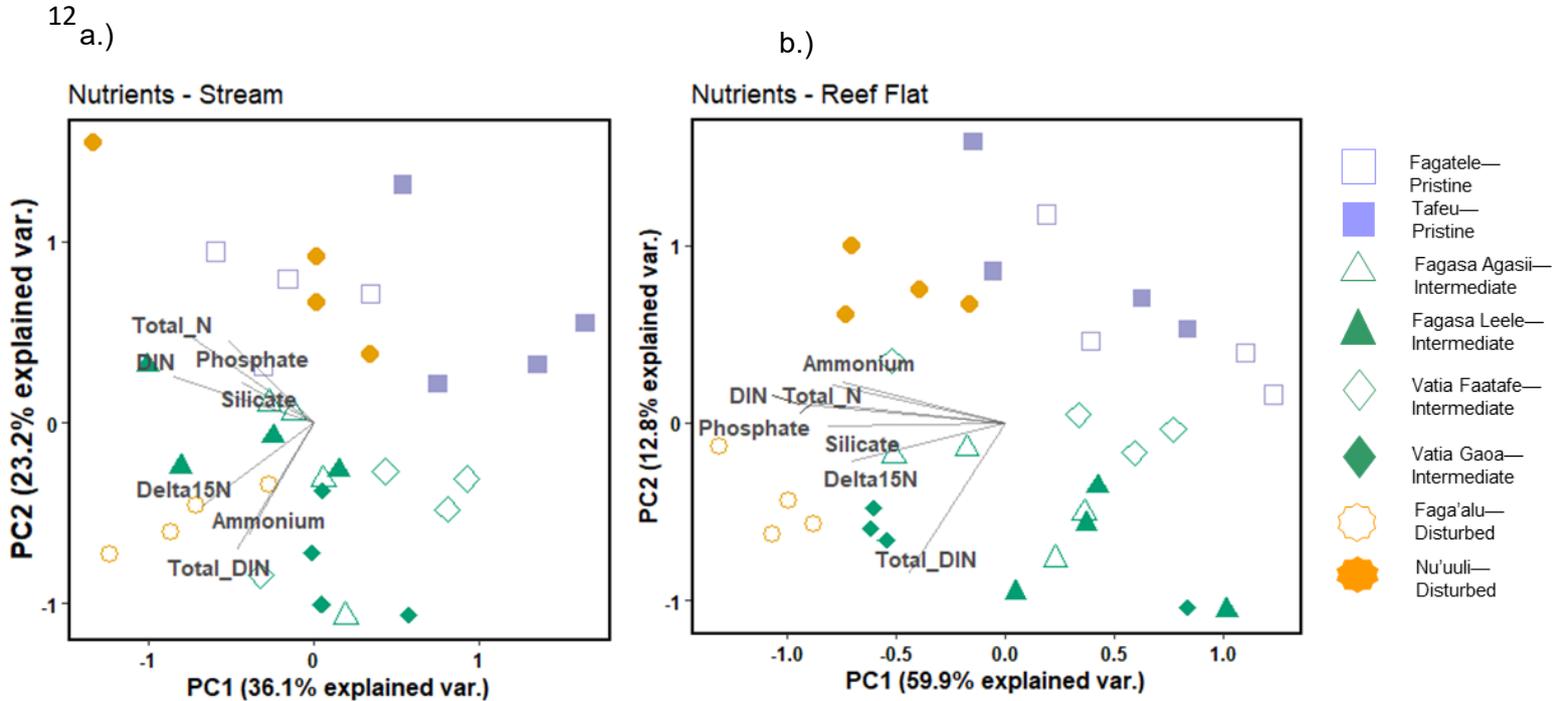
16 **3.1 | Water quality**

17 *Nutrients*

18 The PCA's of nutrient concentrations from stream and reef flat samples revealed a clear partitioning
 19 between pristine and intermediate watersheds (Figure 3). Nutrient concentrations exhibited spatial
 20 structure across watershed types with higher concentrations found in the intermediate and disturbed
 21 watersheds (Fig. S2) and a general trend of higher nutrient concentrations during the drier winter
 22 (August 2018) season (Fig. S3). For example, DIN stream concentrations were more variable across
 23 sites compared to reef flat DIN but in general had the highest average concentrations in disturbed
 24 watersheds, Nu'uuli and Faga'alu, in streams from 7.8 to 8.5 $\mu\text{mol/L}$ and reef flats from 4.0 to 6.7

1 $\mu\text{mol/L}$, respectively. From the stream samples, the first two principal components explained 36.1%
 2 and 23.2% of the total variation, respectively, with samples from pristine watersheds being
 3 differentiated from intermediate watershed samples along PC2 due to lower concentrations of total
 4 DIN and, ammonium, and lower values of $\delta^{15}\text{N}$ (Figure 3a).

5 For nutrients on reef flats, the first two principal components explained 59.9% and 12.8% of the total
 6 variation, respectively. Samples from pristine watersheds were largely differentiated from disturbed
 7 watersheds along PC1, with samples from disturbed sites characterized by higher concentrations of
 8 all nutrients than those from pristine sites (Figure 3b). Samples from intermediate watersheds were
 9 separated from pristine watersheds along PC2, and were characterized by a higher DIN load than
 10 pristine sites.



14 Figure 3. Principal Component Analysis (PCA) showing variation in nutrient concentration on (a) the
 15 stream, and (b) the reef flat among watershed types (pristine, intermediate, disturbed), and eight
 16 stream sites around Tutuila, American Samoa. The two major streams in Vatia (Gaoa and Faatafe)
 17 and Fagasa (Leele and Agasii) were sampled. Analyses are based on nutrient concentrations
 18 collected from quarterly sampling within each habitat at each site. Vector lengths are proportional to
 19 correlation strengths with the primary PCA axes.

20

21

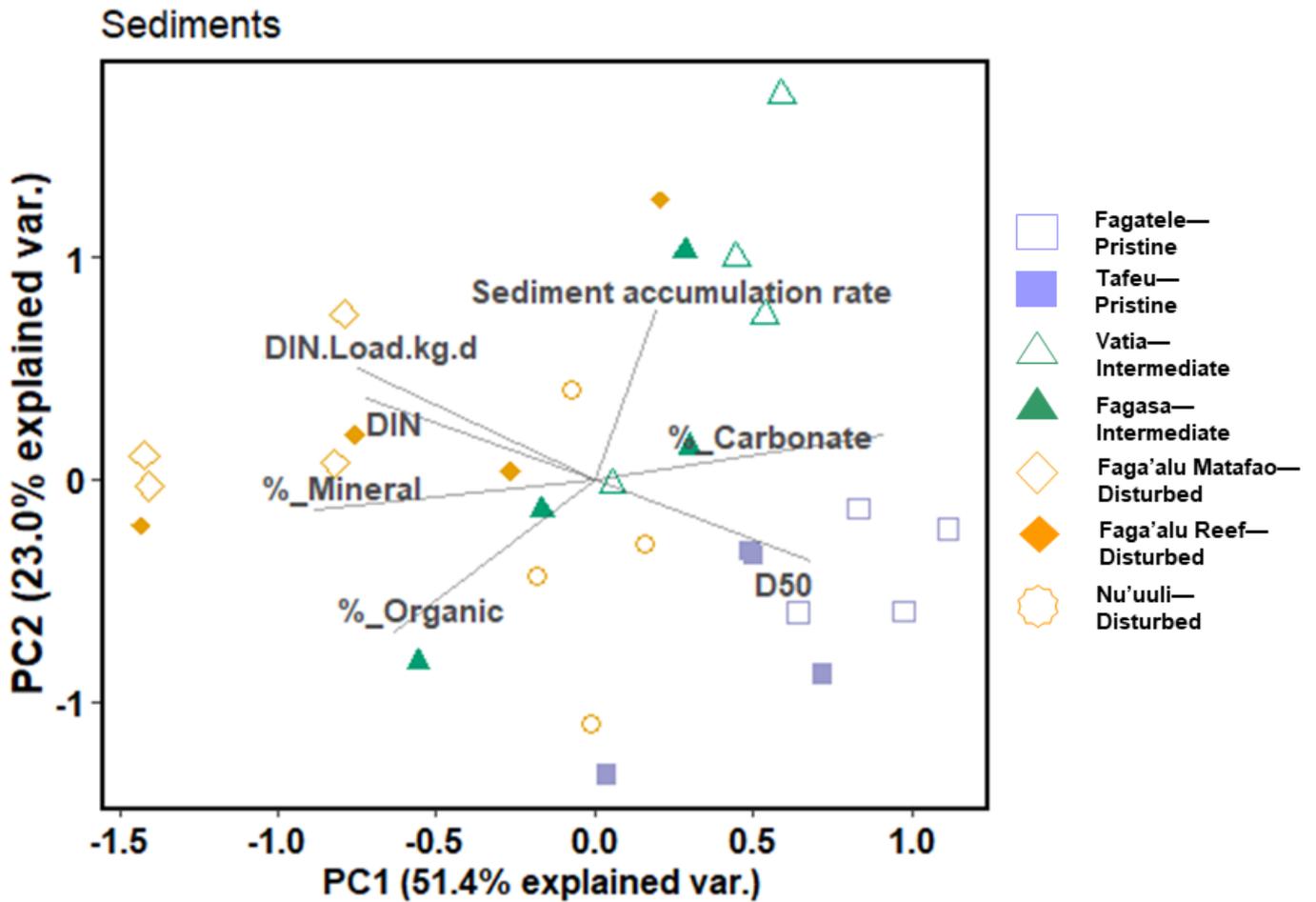
22

1 *Sediments*

2 For suspended particulate matter, the first two principal components explained 51.4% and 23% of the
3 total variation, respectively (Figure 4). The pristine watershed samples were closely clustered and
4 characterized by a larger median particle size (means: 48 – 70 μm), and greater carbonate content
5 (79%) than samples from intermediate or disturbed watersheds (Figure 4; Table 2). There was greater
6 variability in sediment characteristics among samples from the intermediate and disturbed watershed
7 sites, with the intermediate sites having higher sediment trap accumulation rates (i.e. >18
8 $\text{mg}/\text{cm}^2/\text{day}$), and with DIN load, DIN and mineral composition being generally higher in disturbed
9 samples (Figure 4).

10

11



12

13 Figure 4. Principal Component Analysis (PCA) showing variation in sediment accumulation rate,
14 carbonate, mineral, and organic content, DIN concentrations and DIN load ($\text{kg}\cdot\text{d}^{-1}$) among
15 watershed types (pristine, intermediate, disturbed), and seven sediment traps sites around Tutuila,

1 American Samoa. Analyses are based on samples collected from quarterly sampling at each site.
2 Vector lengths are proportional to correlation strengths with the primary PCA axes.

3

4 Sediment trap accumulation rates varied seasonally with higher rates during the wet deployments
5 (Nov-18 – Feb-19; Feb-19 – May-19) and lower accumulation during the dry season (May-19 – Sep-
6 19; Sep-19 - Nov-19) (Fig. 4a). Both intermediate watersheds and Tafeu had the highest sediment
7 accumulation rates during the first wet season deployment (Nov-18 – Feb-19). Disturbed watersheds
8 had more uniform sediment accumulation rates across sites and across deployments. In pristine
9 watersheds, sediment accumulation was higher in Tafeu during the two wet season deployments
10 (Nov-18 – Feb-19; Feb-19 – May-19) compared to Fagatele. Trap sediments were comprised
11 predominantly of carbonate material (Fig. S4), comprising 34 to 79% of the total sample across all
12 sites and deployments. The organic and mineral composition varied across deployments, with the
13 highest (terrigenous) mineral component measured in the two wet season deployments (Fig. 4b).
14 Average particle size range was largest for the pristine sites (8.1 to 298 μm and 10.5 to 349 μm ,
15 respectively for Fagatele and Tafeu) and disturbed site Nu'uuli (7.2 to 552 μm) (Table 2). Relatively
16 uniform percentile rank grain size distribution was measured across the remaining intermediate and
17 disturbed watersheds. Median particle size (i.e. D50) varied across sites but was smaller in the
18 disturbed watersheds (Table 2).

19

20

21

22

23

24

25

26

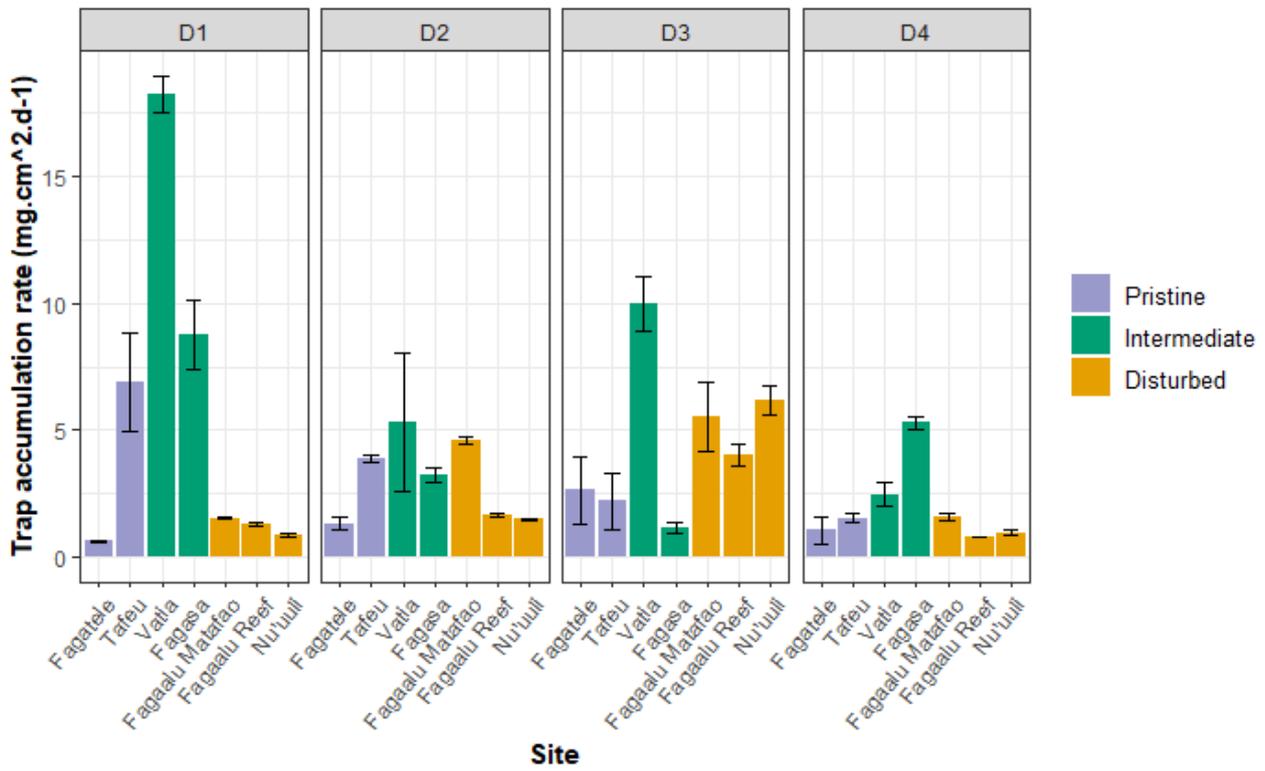
27

28

29

30

1 a)



2

3

4

5

6

7

8

9

10

11

12

13

14

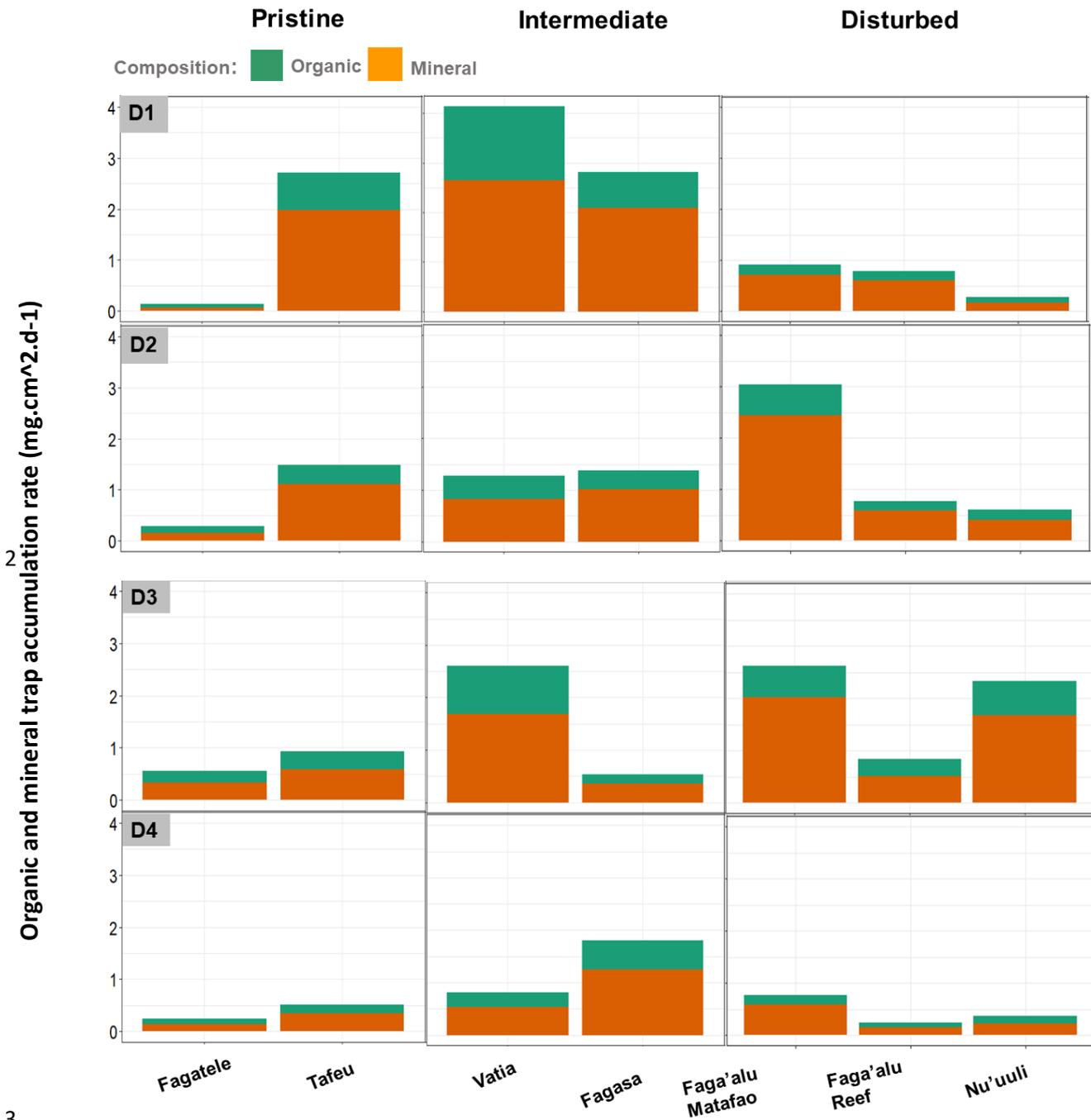
15

16

17

18

1 b)



3

4 Figure 4. Trap accumulation rate (mg.cm².d⁻¹) by watershed type. (a) Total trap accumulation rates
 5 (unit) by seasonal deployments. Error bars represent total trap accumulation standard error. (b) The
 6 terrigenous composition of each sample (organic and mineral sediment) is represented within each
 7 bar. D1 and D2 represent wet season deployments (Nov-18 – Feb-19; Feb-19 – May-19); D3 and D4
 8 represent dry season deployments (May-19 – Sep-19; Sep-19—Nov-19).

9

10

11

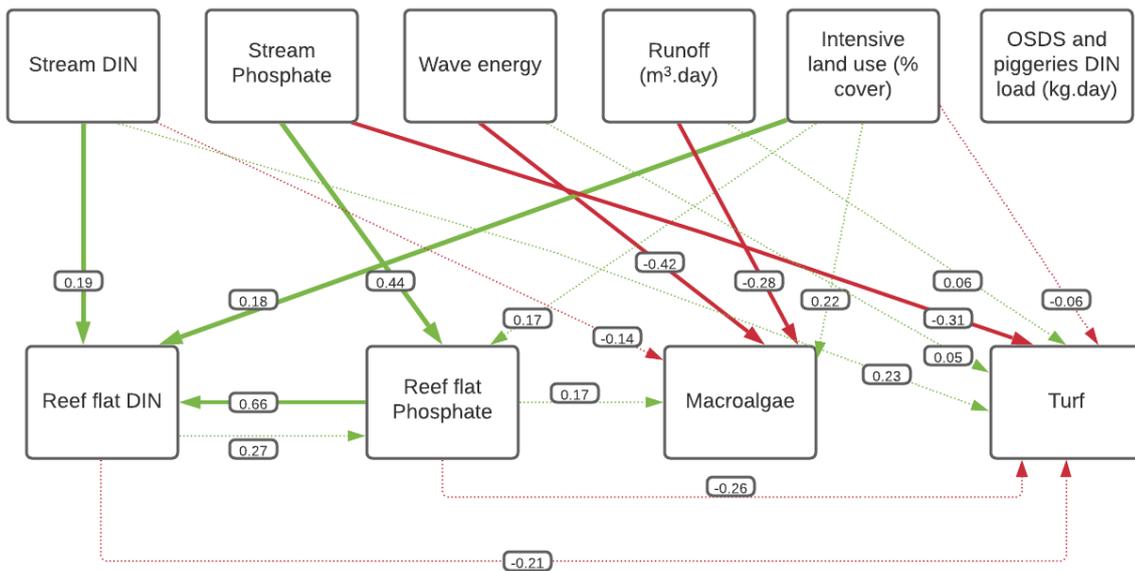
1 **3.3 | Path Analysis**

2 Path analysis revealed a direct and positive link between stream DIN and intensive land use with reef
3 flat DIN, and positive links between stream phosphate, reef phosphate and reef DIN (Fig. 5; Table 3).

4 Macroalgal cover on the reef flat was best predicted by wave energy and surface runoff, both having
5 a significant negative effect on macroalgal cover (Figure 5a). Interestingly, there was little evidence
6 to support any effect of nutrients, intensive land uses, or DIN loads from piggeries and On-Site
7 Disposal Systems on macroalgal cover, while the cover of turf algae was negatively associated to
8 stream phosphate concentrations (Fig 5a; Table 3).

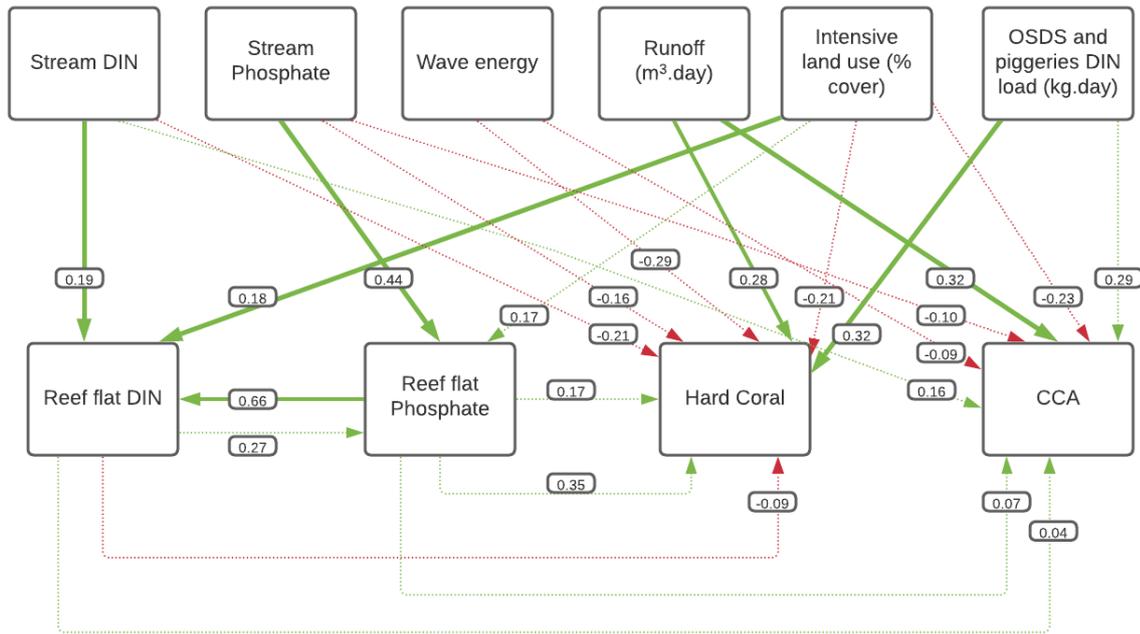
9 Hard coral cover was positively related to surface runoff and DIN load from On-Site Disposal Systems
10 and piggeries (path coefficient 0.28 and 0.32, respectively) (Fig 5b; Table 3). Nutrient concentrations
11 in the stream and reef flat, wave energy, and intensive land uses did not have direct significant effects
12 on hard coral cover. CCA cover was positively influenced mainly by surface runoff.

13 a)



14
15
16
17
18
19
20

1 b)



2

3 Fig. 5. Path analysis showing pathways through nutrients, wave, runoff, intensive land uses and DIN
4 loads affect (a) macroalgal and turf, and (b) hard coral and CCA cover on the reef flat. The two models
5 represent the two groupings of benthic assemblages (see Methods for more details). Arrows in red
6 and green with grey text boxes represent negative and positive path coefficients respectively. The
7 bold straight line arrows represent significant (p-value < 0.05) path coefficients, broken lines represent
8 non-significant path coefficients.

9

10 4 | DISCUSSION

11 Water quality parameters and the composition of shallow water benthic reef communities exhibited
12 distinct differences among watershed types on Tutuila, American Samoa, with sites adjacent to
13 pristine watersheds generally characterized by lower nutrient concentrations and sediment
14 accumulation rate, and higher cover of hard coral and CCA cover. In contrast, sites adjacent to
15 intermediate and disturbed watersheds had higher nutrient concentrations and sedimentation, and
16 higher cover of macroalgae, turf algae, and rubble. These findings are largely consistent with previous
17 studies linking patterns of land use and water quality to benthic composition on coral reefs (Brodie
18 and Pearson, 2016; Brown et al., 2017; Ennis et al., 2016; Oliver et al., 2011; Rodgers et al., 2012;
19 Wenger et al., 2020). Further, the exploratory path analyses highlighted the linkage between land use
20 and water quality which showed positive associations between intensive land uses and DIN
21 concentrations in both stream and reef flat waters, the analyses also revealed some unexpected
22 findings. The positive relationships between surface runoff and DIN load with hard coral cover was
23 unexpected because these drivers are generally thought to be detrimental to coral condition.

1 Conversely, factors generally thought to contribute to macroalgal growth and/or cover (i.e., nutrients)
2 were negatively correlated or absent in the path analysis, rather, incident wave energy was identified
3 as the most important driver of macroalgal cover. The results of the path analysis, although
4 exploratory, highlight the potential complexities of the processes shaping benthic communities on
5 coral reefs, and the need to consider multiple factors (including nutrient concentrations,
6 sedimentation, light, space availability and physical forces) simultaneously.

7 4.1 | Drivers of algal cover

8 The positive effects of nutrient availability on macroalgal growth and cover are widely accepted, with
9 numerous studies reporting correlations between nutrients and macroalgal cover (De'ath and
10 Fabricius, 2010; Fabricius et al., 2012) and elevated growth of macroalgae following nutrient-
11 enrichment (Lapointe, 1997; Larned, 1998; Littler et al., 1991). While the highest macroalgal cover in
12 the present study was at sites with the highest DIN and $\delta^{15}\text{N}$, the path analysis suggested that
13 macroalgal cover was primarily influenced by wave exposure and surface runoff. Macroalgal
14 responses to nutrient enrichment can vary among taxa. For example, the species cover and richness
15 of red and green macroalgae (i.e., Rhodophyta and Chlorophyta, respectively) on inshore reefs of the
16 Great Barrier Reef, Australia was shown to have a negative relationship with good water quality, yet
17 the cover and richness of brown macroalgae (i.e., Phaeophyceae) showed no relationship to water
18 quality (Fabricius, 2005; Fabricius and De'ath, 2004). While increased discharge rates can lead to
19 higher nutrient concentrations and increased macroalgae growth and cover, reduced salinity from
20 freshwater discharge and increased sedimentation can also potentially negatively affect macroalgae
21 photosynthesis and respiration, recruitment, growth, and survival (Kirst, 1990; Umar et al., 1998). The
22 negative relationship between wave exposure and percent cover of macroalgae in our study contrasts
23 with findings from inshore reefs within the Great Barrier Reef Marine Park, which showed a positive
24 effect of wave exposure on macroalgal cover (Ceccarelli et al., 2020), and in Palau where wave
25 exposure was attributed to macroalgal growth and dominance following disturbance from a super
26 typhoon (Roff et al., 2015). However, multiple factors influence the abundance and growth of
27 macroalgae including nutrient availability, light, surface availability and physical forces, and these
28 drivers can interact with each other leading to site-specific composition and abundance patterns
29 (Besterman et al., 2020). Further, wave exposure can enhance macroalgal productivity in shallow
30 environments (Leigh et al., 1987) but high exposures can also cause disturbance and inhibit
31 macroalgal stabilization and accumulation (Williams et al., 2013). While wave energy has been shown
32 to be a major predictor of benthic assemblage structure across regional (Jouffray et al., 2019) and
33 local scales (Ceccarelli et al., 2020), the strong influence of wave energy and its potential interaction
34 with terrestrial run-off warrants further consideration. For instance, higher wave energy can flush
35 terrestrial run-off away from sites exposed to strong waves and currents which can reduce the
36 vulnerability of exposed sites to terrestrial run-off (Rodgers et al., 2012), while low wave energy can

1 lead to retention of nutrients and sediments, increasing residence time and exposure to watershed-
2 derived materials (Storlazzi et al., 2018). The potential decoupling of watershed drivers (e.g.,
3 nutrients) and macroalgal cover in the present study could relate to the influence of localized water
4 movement in the transport of, and exposure to, terrestrial run-off.

5 The highest cover of turf algae around Tutuila was at sites adjacent to intermediate and pristine
6 watersheds on the northern aspect of the island (Tafeu, Vatia and Fagasa). Although these sites had
7 relatively high DIN and $\delta^{15}\text{N}$ concentrations, the path analysis suggested the cover of turf algae was
8 negatively related to phosphate concentration in stream waters. The lack of a positive relationship
9 between nutrients and turf algae cover is perhaps not surprising as numerous studies have linked the
10 cover of algal turfs with the spatial availability on the reef benthos, and in particular, recent coral
11 mortality (e.g., (Diaz-Pulido and McCook, 2002; Hughes et al., 2018), whereas the productivity and
12 biomass of turf algae may be more closely linked to nutrients (den Haan et al., 2016; Karcher et al.,
13 2020; Lapointe et al., 2019).

14 4.2 | Drivers of hard coral and CCA cover

15 The cover of CCA and hard coral was generally highest at the two sites adjacent to the pristine
16 watersheds (i.e., Fagatele and Tafua), and is consistent with previous studies on Tutuila (Birkeland
17 et al., 2003; Green, 1996; Green et al., 1999; Sudek and Lawrence, 2016). While such patterns of
18 higher coral and CCA cover adjacent to pristine watersheds may be interpreted as a positive effect of
19 higher water quality, the path analysis suggested hard coral and CCA cover were positively related
20 to surface runoff. Somewhat similar results were recently reported for the U.S. Virgin Islands where
21 coral diversity and reef rugosity were positively related to sedimentation (Oliver et al., 2018).
22 Terrestrially-derived organic matter in the SPM can comprise both light fraction particulate organic
23 matter, comprised of animal material and plants, and heavy-fraction organic matter, attached to
24 mineral sediment particles. The different fraction compositions likely interact with surface water
25 nutrient concentrations and influence bioavailable nutrient loads (Bainbridge et al., 2018). While
26 sediment accumulation rate was not included in the analysis due to potential interactions with nutrient
27 concentrations, the analysis was able to examine the direct effects of natural and anthropogenic
28 drivers on benthic cover. Runoff is one of a suite of factors affecting the transport of sediment and
29 nutrients into nearshore reef areas and is used as a proxy for potential sources of pollutants and
30 contaminants from the watersheds. However, terrestrial run-off is influenced by the changes in the
31 frequency, duration, and intensity of discharges via stream, river or groundwater into coastal
32 environments (Alvarez-Romero et al., 2013; Bainbridge et al., 2018; Messina and Biggs, 2016; Oliver
33 et al., 2011; Rodgers et al., 2012). Water flow can modulate the delivery of nutrients and sediment to
34 the reef, and as such further investigations on how discharge rates interact with tidal and wave forcing
35 are needed to better understand the effects of surface runoff on hard coral and CCA cover. Another

1 unexpected finding was the significant and positive effect of DIN load (kg/day) from piggeries and On-
2 Site Disposal Systems on hard coral cover, a surprising finding also shared between coral diversity
3 and proximity to impaired water quality sites in the US Virgin Islands (Oliver et al., 2018). Although
4 increased nutrients negatively affect corals for the most part, positive correlations does not always
5 necessarily infer causality. Elevated dissolved inorganic nutrients have also been attributed to
6 increased zooxanthellae density and photosynthetic rates (Fabricius, 2005).

7

8 4.3 | Links between land use, stream nutrients and reef nutrients, and suspended particulate matter

9 The spatial pattern of nutrient concentrations followed local watershed classifications, where higher
10 nutrient concentrations characterized intermediate and disturbed sites while lower concentrations
11 characterized pristine watersheds (Fig. S2). Path analysis further supports this relationship between
12 intensified land uses and stream DIN concentrations and the strong positive influence of stream
13 nutrient concentrations on reef flat nutrients. Disturbed sites had higher average concentrations of
14 reef flat nutrients, while pristine sites had lower average nutrient concentrations. Though there was
15 more spatial variability in the streams, disturbed sites also had higher average nutrient concentrations
16 (Fig. S2). For instance, phosphate concentrations (which dominated total phosphorus) were much
17 higher for disturbed sites. The higher nutrient concentrations in sites adjacent to disturbed watersheds
18 were similar to spatial patterns of DIN quantified at monthly intervals across 26 watersheds in Tutuila
19 (Comeros-Raynal et al. 2019), and broadly comparable to a baseline study of nutrient concentrations
20 around Vanuatu, another South Pacific high island (Devlin et al., 2020). Interestingly, the DIN
21 concentrations in stream waters for the pristine site, Fagatele, in the present study were higher than
22 those reported by Comeros-Raynal et al. (2019). This discrepancy could be attributed to the difference
23 in precipitation at the time of sampling, or potentially higher groundwater discharge in streams as
24 baseflow during sample collection.

25 Together with the spatial variation in nutrient concentrations, nutrient concentrations were generally
26 greater during the drier winter season (Fig. S3). Concordant with our findings, temporal patterns from
27 Comeros-Raynal et al. (2019) showed that DIN concentrations were higher during the drier winter
28 months (July – September), and were lowest during months with relatively low rainfall. We note that
29 four sampling points taken over a period of one year may not fully capture the temporal pattern in
30 nutrient concentrations, given other potential sources of variation. For instance, nutrient
31 concentrations may vary based on streamflow quantity and the hydrograph stage of sample collection.
32 Integrated, event-based sampling approaches that can account for the particular point in time when
33 the sample is taken from the hydrograph, is desirable because the timing of sampling, at the beginning
34 of a storm after a dry period as opposed to sampling as the storm progresses, will affect nutrient

1 concentrations. Further, nutrient concentrations can be impacted by varying stream flow rates or
2 submarine groundwater discharge (SGD), and thus, our sampling regime may not characterize the
3 true impact of terrestrial discharge into nearshore reef environments (Shuler and Comeros-Raynal,
4 2020). Nutrient loading, estimated by multiplying nutrient concentration and water discharge rate, ,
5 can potentially be a more effective indicator of nutrient impacts on coastal waters. However, local
6 water quality standards in surface waters are typically reported as nutrient concentrations, thus our
7 results can be directly compared to American Samoa's water quality standards for nitrogen and
8 phosphorus concentrations in freshwaters (e.g., streams) and embayments (e.g., reef flats), and can
9 be used to help establish nutrient thresholds (Houk et al., 2020).

10 *Suspended Particulate Matter*

11 Multiple factors affect the transport of sediments into nearshore reef environments including
12 precipitation patterns, land uses, topography, soil type, watershed size, shape, river network pattern,
13 and discharge rates (Devlin and Brodie, 2005; Lewis et al., 2020; Messina and Biggs, 2016). Higher
14 sediment accumulation rates recorded during the wet season (October to April) followed a temporal
15 trend for intermediate watersheds, Vatia and Fagasa. The pristine watershed, Tafeu, also had a
16 higher sediment accumulation rate at $6.9 \text{ mg.cm}^2.\text{d}^{-1}$ during the wet season (Fig S4). In addition to
17 high rainfall events, northern winds typical of the wet season from October to April could have
18 influenced the resuspension of large volumes of sediments at these three sites. The pattern of higher
19 sediment accumulation rates in the traps from flood events was similarly observed in inshore areas
20 of the Great Barrier Reef (Lewis et al., 2020) suggesting the important role of discharge in driving
21 increased suspended particulate matter on nearshore reefs. The reverse temporal trend, however,
22 was observed in the disturbed watershed, Faga'alu with the highest sediment trap accumulation rate
23 recorded during the dry season (May 2019 deployment). The high mountains in Faga'alu obstruct the
24 northerly winds from October to April during the wet season, and is instead exposed to dry season
25 south-easterly trade winds from May to September (Craig, 2009). The calm conditions during the wet
26 season coinciding with high rainfall events transporting sediments, nutrients and contaminants can
27 increase exposure of corals to contaminants and elevate their vulnerability to terrestrial run-off
28 (Storlazzi et al., 2018).

29 Sediment type in the traps was dominated by carbonate across all deployments and across all sites.
30 The islands of American Samoa are comprised mostly of basalt without carbonate rocks (Birkeland
31 et al., 2008), therefore, the predominance of carbonate sediment point to a marine provenance. Grain
32 size analysis from Messina (2016) of larger-sized particles comprised mostly of carbonate material in
33 more exposed reef sites support our findings. Terrigenous sediments (terrestrially-derived particulate
34 organic matter and mineral) comprised a smaller component of total sediment composition. However,
35 percent mineral content was higher than organic content across all sites and deployments. The higher

1 mineral and organic composition in Faga'alu Matafao compared to Faga'alu Reef support previous
2 work in Faga'alu that showed higher terrigenous sediment on the inner reef attributed to calmer
3 hydrodynamic conditions compared to the more exposed southern reef site (Messina, 2016). Although
4 the sediment composition at our six sites contrasts with sediment types at seven inshore sites in the
5 Great Barrier Reef, with higher proportions of mineral content (Lewis et al., 2020), the difference in
6 composition is not surprising because oceanic island lithology differs from continental shelves.
7 Further, land uses, catchment sizes, topography, and seasonal and hydrodynamic patterns vary
8 among the two locations.

9 Sediment particle size affects rates of sedimentation and water clarity, and can also determine
10 responses of the benthic biota to sedimentation. Finer-grained sediments (<63 μm) are more an issue
11 for corals because these sediments are easily resuspended and can remain in the water column
12 longer, reducing the light essential for photosynthesis in zooxanthellate corals (Storlazzi et al., 2015).
13 Land uses can significantly influence the fractions of organic matter. For instance, forested watershed
14 often have higher fractions of light fraction particulate matter, while intensive land uses such as
15 agriculture can affect the proportions of terrestrially-derived particulate organic matter transported
16 from streams (Bainbridge et al., 2018). Across sites, Tafu had the highest percent cover of forested
17 area (90%) while Nu'uuli had the lowest at 52% (Table 1). Fagatele, a pristine watershed, had 58%
18 forest cover and the highest percent cover of cultivated land and agroforest, 11% and 10%,
19 respectively. The larger sediment grain size measured in sediment traps at both pristine and one
20 disturbed (Nu'uuli) watershed sites potentially reflect the variability in space and time in the amount
21 of organic material delivered to streams (Table 1). Future work that can characterize and determine
22 the origin of particulate organic matter will improve our understanding of SPM properties and its
23 consequent impacts on nearshore reef environments.

24 4.4 | Limitations of the study

25 In fringing reefs adjacent to steep watersheds, tidal, wind, and wave forcing, and geomorphic controls
26 act in concert to influence the speed and, direction of currents, and residence times over the reef flats
27 influencing exposure of benthic assemblages to terrestrial run-off (Messina, 2016). Thus, our
28 exploratory path analysis provides at best, a simplified approximation of the ridge-to-reef continuum,
29 and should be interpreted with caution. The unexpected associations between surface runoff and hard
30 coral and CCA cover, in particular, warrant further examination because of the important role of
31 discharge in driving increased suspended particulate matter on nearshore reefs. Corals' response to
32 increased sedimentation is dependent on sediment transport and the level of exposure to this
33 stressor. Critical threshold values for deposited and suspended sediment on coral reefs range
34 between 10 and 300 $\text{mg}/\text{cm}^2/\text{d}$ or mg/l (Erftemeijer et al., 2012). However, adverse effects, including
35 mortality, can occur at lower sediment concentrations; 1 $\text{mg}/\text{cm}^2/\text{d}$ for deposited sediment and as low

1 as 3.2 mg/L for suspended sediment concentrations (Tuttle and Donahue, 2020). Sediment trap
2 accumulation rates of below 10 mg/cm²/d across all sites, could potentially negatively affect coral
3 growth and abundance. Future research efforts should be expanded to include sedimentation rates
4 in path analysis or similar analysis examining causal relationships. Further, additional sites along the
5 wave exposure gradients should be considered to examine variation due to inherent environmental
6 exposure and from anthropogenic influence, and the interactions between these drivers. Laboratory
7 and field experiments which quantify sedimentation rates, sediment type, grain size, and coral
8 responses should be expanded and locality-specific environmental regimes and taxa predisposed to
9 sedimentation stress should be considered as well as additive/synergistic effects from nutrient
10 enrichment. Because coral percent cover is the most widely used indicator of coral reef health and is
11 commonly available, our results can be used to compare to other sites that use integrated ridge-to-
12 reef management approaches. However, coral cover may be more sensitive to disturbances
13 compared to coral metrics such as evenness, heterogeneity, skewness, and size-class distribution.
14 Integrating terrestrial run-off information with coral demography data from local and national-level
15 coral reef monitoring programs in American Samoa could potentially help further understand the
16 impacts of terrestrial runoff in nearshore reef environments. While the exploratory path analysis
17 focused on direct effects and did not include indirect effects, it highlighted connection strengths
18 between land condition and drivers of benthic cover; and while exploratory, is expandable to emerging
19 datasets (e.g., precipitation projections, future % change of land use, fish recruitment, etc.), thus, can
20 be used as baseline for future ridge-to-reef investigations.

21 5 | Conclusions

22 The tight coupling between land and sea is an important factor in the health and functioning of
23 nearshore reef habitats. The quantification of nutrient concentrations and suspended particulate
24 matter across an environmental gradient, is the first study on Tutuila, American Samoa to
25 simultaneously examine the relative effects of two major contributors of terrestrial run-off on adjacent
26 reefs, and is also among the first in the South Pacific to use specifically-designed traps
27 (SediSampler[®]) to quantify SPM. Our findings build on a growing body of ridge-to-reef literature that
28 have linked patterns in land use to water quality and biological communities on adjacent shallow water
29 reefs (Delevaux et al., 2018; Oliver et al., 2011; Rodgers et al., 2012), and contributes to the baseline
30 for volcanic high island communities in the South Pacific. The increased coverage in islands can
31 potentially enable comparisons of water quality thresholds, compare benthic responses to increased
32 nutrients and sediments, and provide opportunities to scale up locality-driven findings to similar island
33 configurations in the Pacific region.

34

1 **Acknowledgements:**

2 We dedicate this article to the late Jon Brodie, and his dedication to student mentorship, water quality
3 science, and coral reefs. This research is funded by the National Oceanic and Atmospheric
4 Administration (NOAA) grant number NA18NOS4820114 and the Australian Research Council Centre
5 of Excellence for Coral Reef Studies (MCR). We thank Director Fa'amao Asalele, Jr., Deputy Director
6 William Sili at American Samoa Environmental Protection Agency (AS-EPA) for their support and
7 guidance. We are grateful for programmatic and laboratory support from Christianera Tuitele and
8 Jewel Tuiasosopo at AS-EPA. We thank the Department of Marine and Wildlife Resource for their
9 support. We are grateful to Atuatasi-Lelei Peau from the National Marine Sanctuaries of American
10 Samoa, Scott Burch, Eric Brown, and Ian Moffitt from the National Park Service for overall project
11 support. We would like to thank James Cook University's Michele Tink (TropWATER laboratory),
12 Professor Scott Smithers, Anna Purcell (School of Earth and Environmental Science), and Dr. Andrew
13 Cole (Centre for Macroalgal Resources and Biotechnology) for laboratory use and support. We thank
14 Danielle Hull at the University of Hawaii's SOEST Laboratory for Analytical Biogeochemistry (S-LAB)
15 and Natalie Wallsgrove at the Biogeochemical Stable Isotope Facility. Marine sediment sample
16 processing and analytical development under the Australian Government's National Environmental
17 Science Program - Tropical Water Quality Hub Project 5.8 (led by S. Lewis) also supported this
18 research. We are very grateful for technical, field and laboratory assistance from: Josephine Regis,
19 Hideyo Hattori, Michael Marsik, Hanae Spathias, Val Brown, Timothy Holland, Jonathan E. Brown,
20 Gemma Galbraith, Ben Creswell, David Whitall, Sabrina Woofter, Georgia Coward and Alice
21 Lawrence. We are grateful for laboratory support provided by the Division of Agriculture, Community,
22 and Natural Resources of the American Samoa Community College especially to Mark Schmaedick
23 and Ian Gurr. We are thankful to American Samoa Community College students: Warren Sevaaetasi,
24 Aveipepa Fua, Ingrid Papali'i, XuYi Pan, Perosi Vaofanua for sediment processing and laboratory
25 analysis.

26

27

28

29

30

31

32

33

1 **References:**

- 2 Adam, T.C., Burkepile, D.E., Holbrook, S.J., Carpenter, R.C., Claudet, J., Loiseau, C., Thiault, L., Brooks, A.J.,
3 Washburn, L., Schmitt, R.J., 2020. Landscape-scale patterns of nutrient enrichment in a coral reef ecosystem:
4 implications for coral to algae phase shifts. *Ecological Applications* n/a, e2227.
- 5 Alvarez-Romero, J.G., Devlin, M., Teixeira da Silva, E., Petus, C., Ban, N.C., Pressey, R.L., Kool, J., Roberts, J.J.,
6 Cerdeira-Estrada, S., Wenger, A.S., Brodie, J., 2013. A novel approach to model exposure of coastal-marine
7 ecosystems to riverine flood plumes based on remote sensing techniques. *J Environ Manage* 119, 194-207.
- 8 APHA, Association, A.W.W., Federation, W.E., 2012. *Standard Methods for the Examination of Water and*
9 *Waste Water* 22nd ed. American Water Works Association.
- 10 Armstrong, F.A.J., Stearns, C.R., Strickland, J.D.H., 1967. The measurement of upwelling and subsequent
11 biological process by means of the Technicon Autoanalyzer® and associated equipment. *Deep Sea Research*
12 *and Oceanographic Abstracts* 14, 381-389.
- 13 Atkinson, C.T., Medeiros, A.C., 2005. *Trip Report: Pilot Study of Factors Linking Watershed Function and*
14 *Coastal Ecosystem Health in American Samoa*. USGS Pacific Island Ecosystems Research Center, p. 34.
- 15 Bainbridge, Z., Lewis, S., Bartley, R., Fabricius, K., Collier, C., Waterhouse, J., Garzon-Garcia, A., Robson, B.,
16 Burton, J., Wenger, A., Brodie, J., 2018. Fine sediment and particulate organic matter: A review and case study
17 on ridge-to-reef transport, transformations, fates, and impacts on marine ecosystems. *Marine Pollution*
18 *Bulletin* 135, 1205-1220.
- 19 Bainbridge, Z., Lewis, S., Stevens, T., Petus, C., Lazarus, E., Gorman, J., Smithers, S.G., In review. Measuring
20 sediment grain size across the catchment to reef continuum: Improved methods and environmental insights
21 *Marine Pollution Bulletin*.
- 22 Bainbridge, Z.T., Wolanski, E., Álvarez-Romero, J.G., Lewis, S.E., Brodie, J.E., 2012. Fine sediment and nutrient
23 dynamics related to particle size and floc formation in a Burdekin River flood plume, Australia. *Marine*
24 *Pollution Bulletin* 65, 236-248.
- 25 Besterman, A.F., McGlathery, K.J., Reidenbach, M.A., Wiberg, P.L., Pace, M.L., 2020. Predicting benthic
26 macroalgal abundance in shallow coastal lagoons from geomorphology and hydrologic flow patterns.
27 *Limnology and Oceanography* n/a.
- 28 Birkeland, C., Craig, P., Fenner, D., Smith, L., Kiene, W.E., Riegl, B.M., 2008. *Geologic Setting and Ecological*
29 *Functioning of Coral Reefs in American Samoa*, in: Riegl, B.M., Dodge, R.E. (Eds.), *Coral Reefs of the USA*.
30 Springer Netherlands, Dordrecht, pp. 741-765.
- 31 Birkeland, C.E., Randall, R.H., Green, A.L., Smith, B., Wilkins, S., 2003. *Changes in the coral reef communities*
32 *of Fagatele Bay NMS and Tutuila Island (American Samoa), 1982-1995*, Technical Report Submitted to the
33 Fagatele Bay National Marine Sanctuary, Pago Pago, American Samoa
- 34 Brodie, J., Pearson, R.G., 2016. Ecosystem health of the Great Barrier Reef: Time for effective management
35 action based on evidence. *Estuarine, Coastal and Shelf Science* 183, 438-451.
- 36 Brodie, J.E., Kroon, F.J., Schaffelke, B., Wolanski, E.C., Lewis, S.E., Devlin, M.J., Bohnet, I.C., Bainbridge, Z.T.,
37 Waterhouse, J., Davis, A.M., 2012. Terrestrial pollutant runoff to the Great Barrier Reef: An update of issues,
38 priorities and management responses. *Marine Pollution Bulletin* 65, 81-100.
- 39 Brown, C.J., Jupiter, S.D., Albert, S., Anthony, K.R.N., Hamilton, R.J., Fredston-Hermann, A., Halpern, B.S., Lin,
40 H.-Y., Maina, J., Mangubhai, S., Mumby, P.J., Possingham, H.P., Saunders, M.I., Tulloch, V.J.D., Wenger, A.,
41 Klein, C.J., 2019. A guide to modelling priorities for managing land-based impacts on coastal ecosystems.
42 *Journal of Applied Ecology* 56, 1106-1116.
- 43 Brown, C.J., Jupiter, S.D., Albert, S., Klein, C.J., Mangubhai, S., Maina, J.M., Mumby, P., Olley, J., Stewart-Koster,
44 B., Tulloch, V., Wenger, A., 2017. Tracing the influence of land-use change on water quality and coral reefs
45 using a Bayesian model. *Sci Rep* 7, 4740.
- 46 Burke, L., Reytar, K., Spalding, M., Perry, A., 2011. *Reefs at Risk Revisited*. World Resources Institute,
47 Washington, D.C., USA, p. 114.
- 48 Carlson, R.R., Foo, S.A., Asner, G.P., 2019. Land Use Impacts on Coral Reef Health: A Ridge-to-Reef Perspective.
49 *Frontiers in Marine Science* 6, 562.

1 Ceccarelli, D.M., Evans, R.D., Logan, M., Mantel, P., Puotinen, M., Petus, C., Russ, G.R., Williamson, D.H., 2020.
2 Long-term dynamics and drivers of coral and macroalgal cover on inshore reefs of the Great Barrier Reef
3 Marine Park. *Ecological Applications* 30, e02008.

4 Comeros-Raynal, M.T., Lawrence, A., Sudek, M., Vaeoso, M., McGuire, K., Regis, J., Houk, P., 2019. Applying a
5 ridge-to-reef framework to support watershed, water quality, and community-based fisheries management in
6 American Samoa. *Coral Reefs* 38, 505–520.

7 Craig, P., 2009. *Natural History Guide to American Samoa*, Third ed. National Park Service, p. 134.

8 Craig, P., Didonato, G., Fenner, D., Hawkins, C., 2010. The State of Coral Reef Ecosystems of American Samoa.
9 De'ath, G., Fabricius, K., 2010. Water quality as a regional driver of coral biodiversity and macroalgae on the
10 Great Barrier Reef. *Ecological Applications* 20, 840-850.

11 Delevaux, J.M.S., Whittier, R., Stamoulis, K.A., Bremer, L.L., Jupiter, S., Friedlander, A.M., Poti, M., Guannel, G.,
12 Kurashima, N., Winter, K.B., Toonen, R., Conklin, E., Wiggins, C., Knudby, A., Goodell, W., Burnett, K., Yee, S.,
13 Htun, H., Oleson, K.L.L., Wiegner, T., Ticktin, T., 2018. A linked land-sea modeling framework to inform ridge-
14 to-reef management in high oceanic islands. *PLOS ONE* 13, e0193230.

15 den Haan, J., Huisman, J., Brocke, H.J., Goehlich, H., Latijnhouwers, K.R.W., van Heeringen, S., Honcoop, S.A.S.,
16 Bleyenbergh, T.E., Schouten, S., Cerli, C., Hoitinga, L., Vermeij, M.J.A., Visser, P.M., 2016. Nitrogen and
17 phosphorus uptake rates of different species from a coral reef community after a nutrient pulse. *Scientific*
18 *Reports* 6, 28821.

19 Devlin, M., Smith, A., Graves, C.A., Petus, C., Tracey, D., Maniel, M., Hooper, E., Kotra, K., Samie, E., Loubser,
20 D., Lyons, B.P., 2020. Baseline assessment of coastal water quality, in Vanuatu, South Pacific: Insights gained
21 from in-situ sampling. *Marine Pollution Bulletin* 160, 111651.

22 Devlin, M.J., Brodie, J., 2005. Terrestrial discharge into the Great Barrier Reef Lagoon: nutrient behavior in
23 coastal waters. *Mar Pollut Bull* 51, 9-22.

24 Diaz-Pulido, G., McCook, L., 2002. The fate of bleached corals: patterns and dynamics of algal recruitment.
25 *Marine Ecology Progress Series* 232, 115-128.

26 DiDonato, G.T., 2004. Developing an Initial Watershed Classification for American Samoa, Report to the
27 American Samoa Environmental Protection Agency, Pago Pago, American Samoa. American Samoa
28 Environmental Protection Agency, p. 14.

29 Ennis, R.S., Brandt, M.E., Wilson Grimes, K.R., Smith, T.B., 2016. Coral reef health response to chronic and
30 acute changes in water quality in St. Thomas, United States Virgin Islands. *Mar Pollut Bull* 111, 418-427.

31 Erftemeijer, P.L.A., Riegl, B., Hoeksema, B.W., Todd, P.A., 2012. Environmental impacts of dredging and other
32 sediment disturbances on corals: A review. *Marine Pollution Bulletin* 64, 1737-1765.

33 Fabricius, K.E., 2005. Effects of terrestrial runoff on the ecology of corals and coral reefs: review and synthesis.
34 *Mar Pollut Bull* 50, 125-146.

35 Fabricius, K.E., Cooper, T.F., Humphrey, C., Uthicke, S., De'ath, G., Davidson, J., LeGrand, H., Thompson, A.,
36 Schaffelke, B., 2012. A bioindicator system for water quality on inshore coral reefs of the Great Barrier Reef.
37 *Marine Pollution Bulletin* 65, 320-332.

38 Fabricius, K.E., De'ath, G., 2004. IDENTIFYING ECOLOGICAL CHANGE AND ITS CAUSES: A CASE STUDY ON CORAL
39 REEFS. *Ecological Applications* 14, 1448-1465.

40 Falkland, T., 1999. Water resources issues of small island developing states. *Natural Resources Forum* 23, 245-
41 260.

42 Grasshoff, K., Ehrhardt, M., Kremling, K., 1983. *Methods of Seawater Analysis* Verlag Chemie.

43 Green, A.L., 1996. Status of the Coral Reefs of the Samoan Archipelago. Department of Marine and Wildlife
44 Resources, American Samoa p. 126.

45 Green, A.L., Birkeland, C.E., Randall, R.H., 1999. Twenty Years of Disturbance and Change in Fagatele Bay
46 National Marine Sanctuary, American Samoa. *Pacific Science* 53, 376-400.

47 Heiri, O., Lotter, A.F., Lemcke, G., 2001. Loss on ignition as a method for estimating organic and carbonate
48 content in sediments: reproducibility and comparability of results. *Journal of Paleolimnology* 25, 101-110.

49 Holst Rice, S., Messina, A., Biggs, T., Vargas-Angel, B., Whittall, D., 2016. Baseline Assessment of Faga'alu
50 Watershed: A Ridge to Reef Assessment in Support of Sediment Reduction Activities and Future Evaluation of

1 their Success, NOAA Technical Memorandum CRCP. NOAA Coral Reef Conservation Program, Silver Spring,
2 Maryland, p. 44.

3 Houk, P., Camacho, R., Johnson, S., McLean, M., Maxin, S., Anson, J., Joseph, E., Nedlic, O., Luckymis, M.,
4 Adams, K., Hess, D., Kabua, E., Yalon, A., Buthung, E., Graham, C., Leberer, T., Taylor, B., van Woesik, R., 2015.
5 The Micronesia Challenge: Assessing the Relative Contribution of Stressors on Coral Reefs to Facilitate Science-
6 to-Management Feedback. PLOS ONE 10, e0130823.

7 Houk, P., Comerros-Raynal, M., Lawrence, A., Sudek, M., Vaeoso, M., McGuire, K., Regis, J., 2020. Nutrient
8 thresholds to protect water quality and coral reefs. Marine Pollution Bulletin 159, 111451.

9 Houk, P., Musburger, C., Wiles, P., 2010. Water quality and herbivory interactively drive coral-reef recovery
10 patterns in American Samoa. PLoS One 5, e13913.

11 Hughes, T.P., Kerry, J.T., Baird, A.H., Connolly, S.R., Dietzel, A., Eakin, C.M., Heron, S.F., Hoey, A.S.,
12 Hoogenboom, M.O., Liu, G., McWilliam, M.J., Pears, R.J., Pratchett, M.S., Skirving, W.J., Stella, J.S., Torda, G.,
13 2018. Global warming transforms coral reef assemblages. Nature 556, 492-496.

14 Izuka, S.K., Giambelluca, T.W., Nullet, M.A., 2005. Potential Evapotranspiration on Tutuila, American Samoa,
15 USGS Scientific Investigations Report 2005-5200. U.S. Geological Survey Department of the Interior, p. 40.

16 Jenness, J., Houk, P., 2014. UOGML Wave Energy ArcGIS Extension. University of Guam Marine Laboratory.

17 Jouffray, J.-B., Wedding, L.M., Norström, A.V., Donovan, M.K., Williams, G.J., Crowder, L.B., Erickson, A.L.,
18 Friedlander, A.M., Graham, N.A.J., Gove, J.M., Kappel, C.V., Kittinger, J.N., Lecky, J., Oleson, K.L.L., Selkoe, K.A.,
19 White, C., Williams, I.D., Nyström, M., 2019. Parsing human and biophysical drivers of coral reef regimes.
20 Proceedings of the Royal Society B: Biological Sciences 286, 20182544.

21 Jupiter, S.D., Wenger, A., Klein, C.J., Albert, S., Mangubhai, S., Nelson, J., Teneva, L., Tulloch, V.J., White, A.T.,
22 Watson, J.E.M., 2017. Opportunities and constraints for implementing integrated land–sea management on
23 islands. Environmental Conservation 44, 254-266.

24 Karcher, D.B., Roth, F., Carvalho, S., El-Khaled, Y.C., Tilstra, A., Kürten, B., Struck, U., Jones, B.H., Wild, C., 2020.
25 Nitrogen eutrophication particularly promotes turf algae in coral reefs of the central Red Sea. PeerJ 8, e8737-
26 e8737.

27 Kennedy, J., and Chilton Consulting Engineers, 1987. Groundwater contamination study Tafuna-Leone Plain
28 Tutuila Island, Final report for the Environmental Quality Commission, Office of the Governor, Tutuila,
29 American Samoa, p. 168.

30 Kérouel, R., Aminot, A., 1997. Fluorometric determination of ammonia in sea and estuarine waters by direct
31 segmented flow analysis. Marine Chemistry 57, 265-275.

32 Kirst, 1990. Salinity tolerance of eukaryotic marine algae. Annual review of plant physiology and plant
33 molecular biology 41.

34 Kline, R., 1998. Principles and Practices of Structural Equation Modeling. Guilford Press, New York

35 Lapointe, B.E., 1997. Nutrient thresholds for bottom-up control of macroalgal blooms on coral reefs in Jamaica
36 and southeast Florida. Limnology and Oceanography 42, 1119-1131.

37 Lapointe, B.E., Brewton, R.A., Herren, L.W., Porter, J.W., Hu, C., 2019. Nitrogen enrichment, altered
38 stoichiometry, and coral reef decline at Looe Key, Florida Keys, USA: a 3-decade study. Marine Biology 166,
39 108.

40 Larned, S.T., 1998. Nitrogen- versus phosphorus-limited growth and sources of nutrients for coral reef
41 macroalgae. Marine Biology 132, 409-421.

42 Leigh, E.G., Paine, R.T., Quinn, J.F., Suchanek, T.H., 1987. Wave energy and intertidal productivity. Proceedings
43 of the National Academy of Sciences 84, 1314.

44 Lewis, S., Bainbridge, Z., Stevens, T., Garzon-Garcia, A., Chen, C., Bahadori, M., Burton, J., Rezaei Rashti, M.,
45 James, C., Smithers, S.G., Olley, J., 2020. What’s really damaging the Reef?: Tracing the origin and fate of the
46 environmentally detrimental sediment and associated bioavailable nutrients, Report to the National
47 Environmental Science Program. Reef and Rainforest Research Centre Limited, Cairns, p. 248.

48 Littler, M.M., Littler, D.S., 2007. Assessment of coral reefs using herbivory/nutrient assays and indicator groups
49 of benthic primary producers: a critical synthesis, proposed protocols, and critique of management strategies.
50 Aquatic Conservation: Marine and Freshwater Ecosystems 17, 195-215.

1 Littler, M.M., Littler, D.S., Brooks, B.L., 2006. Harmful algae on tropical coral reefs: Bottom-up eutrophication
2 and top-down herbivory. *Harmful Algae* 5, 565-585.

3 Littler, M.M., Littler, D.S., Titlyanov, E.A., 1991. Comparisons of N- and P-limited productivity between high
4 granitic islands versus low carbonate atolls in the Seychelles Archipelago: a test of the relative-dominance
5 paradigm. *Coral Reefs* 10, 199-209.

6 Lleras, C., 2005. Path analysis. *Encyclopedia of social measurement* 3, 25-30.

7 McIlvin, M.R., Casciotti, K.L., 2010. Fully automated system for stable isotopic analyses of dissolved nitrous
8 oxide at natural abundance levels. *Limnology and Oceanography: Methods* 8, 54-66.

9 Messina, A., 2016. Terrigenous sediment dynamics in a small, tropical, fringing reef embayment, Department
10 of Geography. Dissertation for San Diego State University/UC Santa Barbara Joint-Doctoral Program.

11 Messina, A.M., Biggs, T.W., 2016. Contributions of human activities to suspended sediment yield during storm
12 events from a small, steep, tropical watershed. *Journal of Hydrology* 538, 726-742.

13 Meyer, R., Seamon, J., Fa'aumu, S., Lalogafuafua, I., 2017. Classification and Mapping of Wildlife Habitat in
14 American Samoa. An object-based approach using high resolution orthoimagery and LIDAR remote sensing
15 data, Report to the American Samoa Department of Marine and Wildlife Resources Department of Marine and
16 Wildlife Resources p. 140.

17 Murphy, J., Riley, J.P., 1962. A modified single solution method for the determination of phosphate in natural
18 waters. *Analytica Chimica Acta* 27, 31-36.

19 NOAA, 2020. CCAP Land Cover Atlas. NOAA.

20 Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlenn, D., Minchin, P.R., O'Hara, R.B.,
21 Simpson, G.L., Solymos, P., Stevens, M.H.H., Szoecs, E., Wagner, H., 2019. *vegan: Community Ecology Package*.

22 Oliver, L.M., Fisher, W.S., Fore, L., Smith, A., Bradley, P., 2018. Assessing land use, sedimentation, and water
23 quality stressors as predictors of coral reef condition in St. Thomas, U.S. Virgin Islands. *Environmental*
24 *Monitoring and Assessment* 190, 213.

25 Oliver, L.M., Lehrter, J.C., Fisher, W.S., 2011. Relating landscape development intensity to coral reef condition
26 in the watersheds of St. Croix, US Virgin Islands. *Marine Ecology Progress Series* 427, 293-302.

27 Pedhazur, E., 1997. *Structural Equation Models with Observed Variables: Path Analysis, Multiplpe Regression*
28 *in Behavioral Research Explanation and Prediction*, Third Edition ed. Christopher P. Klein, USA, pp. 765-841.

29 Peña, E.A., Slate, E.H., 2006. Global Validation of Linear Model Assumptions. *Journal of the American Statistical*
30 *Association* 101, 341-354.

31 Rodgers, K.S., Kido, M.H., Jokiel, P.L., Edmonds, T., Brown, E.K., 2012. Use of integrated landscape indicators
32 to evaluate the health of linked watersheds and coral reef environments in the Hawaiian islands. *Environ*
33 *Manage* 50, 21-30.

34 Roff, G., Chollett, I., Doropoulos, C., Golbuu, Y., Steneck, R.S., Isechal, A.L., van Woesik, R., Mumby, P.J., 2015.
35 Exposure-driven macroalgal phase shift following catastrophic disturbance on coral reefs. *Coral Reefs* 34, 715-
36 725.

37 Ruddle, K., Hviding, E., Johannes, R.E., 1992. Marine Resources Management in the Context of Customary
38 Tenure. *Marine Resource Economics* 7, 249-273.

39 Rude, J., Minks, A., Doheny, B., Tyner, M., Maher, K., Huffard, C., Hidayat, N.I., Grantham, H., 2016. Ridge to
40 reef modelling for use within land-sea planning under data-limited conditions. *Aquatic Conservation: Marine*
41 *and Freshwater Ecosystems* 26, 251-264.

42 Shuler, C., Brewington, L., El-Kadi, A.I., 2021. A participatory approach to assessing groundwater recharge
43 under future climate and land-cover scenarios, Tutuila, American Samoa. *Journal of Hydrology: Regional*
44 *Studies* 34, 100785.

45 Shuler, C.K., Comerros-Raynal, M., 2020. Ridge to Reef Management Implications for the Development of an
46 Open-Source Dissolved Inorganic Nitrogen-Loading Model in American Samoa. *Environmental Management*.

47 Shuler, C.K., D.W., A., V., G., Baker, L., Olguin, A.N., Dulai, H., C.M., S., Alegado, R.A., 2019. Assessment of
48 Terrigenous Nutrient Loading to Coastal Ecosystems along a Human Land-Use Gradient, Tutuila, American
49 Samoa. *Hydrology* 6, 18.

50 Shuler, C.K., El-Kady, A.I., 2017. Provisional Hydrogeologic Data and Recommendations for Sustainable
51 Groundwater Management Tutuila, American Samoa, December 2017 Draft. Prepared in Cooperation with

1 American Samoa Environmental Protection Agency through a US EPA Region IX Making a Visible Difference
2 Project Project No. C00543. University of Hawaii Water Resources Research Center, p. 77.

3 Sigman, D.M., Casciotti, K.L., Andreani, M., Barford, C., Galanter, M., Böhlke, J.K., 2001. A Bacterial Method for
4 the Nitrogen Isotopic Analysis of Nitrate in Seawater and Freshwater. *Analytical Chemistry* 73, 4145-4153.

5 Storlazzi, C.D., Cheriton, O.M., Messina, A.M., Biggs, T.W., 2018. Meteorologic, oceanographic, and
6 geomorphic controls on circulation and residence time in a coral reef-lined embayment: Faga'alu Bay,
7 American Samoa. *Coral Reefs* 37, 457-469.

8 Storlazzi, C.D., Norris, B.K., Rosenberger, K.J., 2015. The influence of grain size, grain color, and suspended-
9 sediment concentration on light attenuation: Why fine-grained terrestrial sediment is bad for coral reef
10 ecosystems. *Coral Reefs* 34, 967-975.

11 Streiner, D.L., 2005. Finding Our Way: An Introduction to Path Analysis. *Canadian Journal of Psychiatry* 50, 115-
12 122.

13 Sudek, M., Lawrence, A., 2016. American Samoa Coral Reef Monitoring Program Progress Report FY13/14,
14 Report to the Department of Marine and Wildlife Resources, Coral Reef Advisory Group and NOAA.
15 Department of Marine and Wildlife Resources p. 51.

16 Szmant, A.M., 2002. Nutrient enrichment on coral reefs: Is it a major cause of coral reef decline? *Estuaries* 25,
17 743-766.

18 Tuitele, C., Tuiasosopo, J., Faaiuasoo, S., 2016. Watershed Classification Update for American Samoa American
19 Samoa Environmental Protection Agency, Pago Pago, American Samoa p. 12.

20 Tuttle, L.J., Donahue, M.J., 2020. Thresholds for Sediment Stress on Corals: A systematic review and meta-
21 analysis. NOAA Fisheries Pacific Islands , Pacific Islands Regional Office, Habitat Conservation Division,
22 Honolulu, Hawaii, p. 76.

23 Umar, M.J., McCook, L.J., Price, I.R., 1998. Effects of sediment deposition on the seaweed *Sargassum* on a
24 fringing coral reef. *Coral Reefs* 17, 169-177.

25 US Census Bureau, 2014. 2010 Census of Population and Housing, American Samoa, Demographic Profile
26 Summary File: Technical Documentation.

27 Vargas-Angel, B., Huntington, B., 2020. Status and Trends Assessments for Land-based Sources of Pollution
28 Impacts on Benthic Reef Communities in Faga'alu Bay, American Samoa, NOAA Technical Memorandum
29 NMFS-PIFSC-109, p. 38.

30 Verbist, B., Poesen, J., van Noordwijk, M., Widiyanto, Suprayogo, D., Agus, F., Deckers, J., 2010. Factors affecting
31 soil loss at plot scale and sediment yield at catchment scale in a tropical volcanic agroforestry landscape.
32 *CATENA* 80, 34-46.

33 Weber, M., Lott, C., Fabricius, K.E., 2006. Sedimentation stress in a scleractinian coral exposed to terrestrial
34 and marine sediments with contrasting physical, organic and geochemical properties. *Journal of Experimental
35 Marine Biology and Ecology* 336, 18-32.

36 Wenger, A.S., Harris, D., Weber, S., Vaghi, F., Nand, Y., Naisilisili, W., Hughes, A., Delevaux, J.M.S., Klein, C.J.,
37 Watson, J., Mumby, P.J., Jupiter, S.D., 2020. Best-practice forestry management delivers diminishing returns
38 for coral reefs with increased land-clearing. *Journal of Applied Ecology* n/a.

39 Whittall, D., Curtis, M., Mason, A., Vargas-Angel, B., 2019. Excess Nutrients in Vatia Bay, American Samoa:
40 Spatiotemporal Variability, Source Identification and Impact on Coral Reef Ecosystems, NOAA Technical
41 Memorandum NOS NCCOS 266. NOAA Silver Spring, MD, p. 69.

42 Williams, G.J., Smith, J.E., Conklin, E.J., Gove, J.M., Sala, E., Sandin, S.A., 2013. Benthic communities at two
43 remote Pacific coral reefs: effects of reef habitat, depth, and wave energy gradients on spatial patterns. *PeerJ*
44 1, e81.

45 Wong, M.F., 1996. Analysis of Streamflow Characteristics for Streams on the Island of Tutuila, American
46 Samoa. U.S. Geological Survey, Honolulu, Hawaii, p. 173.

47 Wooldridge, S.A., 2009. Water quality and coral bleaching thresholds: Formalising the linkage for the inshore
48 reefs of the Great Barrier Reef, Australia. *Marine Pollution Bulletin* 58, 745-751.

49

50

1 Table 1. Site attributes including watershed area, percent land use, number of On-Site Disposal
 2 System units and pigs.

Site	Total Watershed Area (sqm)	Natural Land Use		Intensive Land Use		% All other land use (barren land, shrub/scrub land, grass/herbaceous land)	Total number of Onsite Disposal System units (cesspools, septic tanks)	No. of pigs
		% Forest	% Agro-forest	% Cultivated	% Develop Land			
Fagatele	1,917,759	58%	10%	11%	5%	15%	0	0
Tafeu	1,669,015	90%	0%	0%	0%	10%	0	0
Fagasa	3,483,148	78%	5%	5%	7%	6%	117	118
Vatia	3,574,377	79%	4%	3%	5%	9%	100	152
Nu'uuli	17,170,233	52%	5%	8%	28%	7%	65	221
Fagaalu	2,476,211	71%	4%	2%	13%	11%	124	132

3
 4
 5
 6
 7
 8
 9
 10
 11
 12
 13
 14
 15
 16

1 Table 2. Mean percentile rank of grain size distribution (in μm) of suspended particulate matter (SPM)
 2 for each site across all seasonal deployments.

Site	Watershed Type	D10 (mean \pm 1 SD)	D50 (mean \pm 1 SD)	D90 (mean \pm 1 SD)
Fagatele	Pristine	8.15 \pm 1.5	48.4 \pm 10.4	298 \pm 94
Tafeu	Pristine	10.5 \pm 1.7	69.6 \pm 15.9	349 \pm 109
Vatia	Intermediate	6.00 \pm 0.75	32.7 \pm 4.9	138 \pm 19
Fagasa	Intermediate	6.94 \pm 0.50	40.1 \pm 6.6	188 \pm 19
Fagaalu Matafao	Disturbed	6.32 \pm 0.88	28.0 \pm 5.2	114 \pm 5
Fagaalu Reef	Disturbed	6.64 \pm 0.99	34.3 \pm 9.8	195 \pm 80
Nu'uuli	Disturbed	7.17 \pm 0.92	36.4 \pm 5.5	552 \pm 773

3
 4
 5
 6
 7
 8
 9
 10
 11
 12
 13
 14
 15
 16
 17
 18
 19
 20
 21

1 Table 3. Path analysis model outputs. We used the default estimator in the lavaan package, maximum
 2 likelihood. Standard error (SE) is based on the expected information matrix, and the z-value
 3 represents the ratio of the parameter to the standard error. P-value <0.05 represents significant
 4 parameters. Path coefficients are standardized versions of linear regression weights. R-squared
 5 values reflect the proportion of variance in the dependent variables accounted for by the equation.

	Estimate	Std.Err	z-value	P(> z)	Std.all	R ²
Macroalgae Reef Flat ~						40%
Wave	-0.01	0.00	-3.49	0.00	-0.42	
Reef flat DIN ~						73%
Stream DIN	0.15	0.06	2.58	0.01	0.19	
Intensive land use	0.73	0.32	2.30	0.02	0.18	
Reef flat PO ₄	1.01	0.18	5.53	0.00	0.66	
Reef flat PO ₄ ~						61%
Stream PO ₄	0.55	0.14	3.90	0.00	0.44	
Intensive land use	0.44	0.26	1.71	0.09	0.17	
Reef Flat DIN	0.17	0.12	1.40	0.16	0.27	
Macroalgae Reef Flat ~						40%
Stream DIN	-0.04	0.03	-1.25	0.21	-0.14	
Reef flat PO ₄	0.08	0.05	1.56	0.12	0.17	
Runoff	-0.10	0.04	-2.54	0.01	-0.28	
Intensive land use	0.29	0.15	1.90	0.06	0.22	
Turf reef Flat ~						43%
Stream DIN	0.05	0.02	1.99	0.05	0.23	
Stream PO ₄	-0.16	0.06	-2.60	0.01	-0.31	
Reef Flat DIN	-0.06	0.05	-1.16	0.25	-0.21	
Reef flat PO ₄	-0.11	0.08	-1.36	0.17	-0.26	
Intensive land use	-0.07	0.12	-0.55	0.58	-0.06	
Wave energy	0.00	0.00	0.46	0.64	0.05	
Runoff	0.02	0.03	0.58	0.56	0.06	
Reef Flat DIN ~						73%
Stream DIN	0.15	0.06	2.58	0.01	0.19	
Intensive land use	0.73	0.32	2.30	0.02	0.18	
Reef flat PO ₄	1.01	0.18	5.53	0.00	0.66	
Reef flat PO ₄ ~						61%
Stream PO ₄	0.55	0.14	3.90	0.00	0.44	
Intensive land use	0.44	0.26	1.71	0.09	0.17	
Reef flat DIN	0.17	0.12	1.40	0.16	0.27	
Hard coral reef flat ~						20%
Stream DIN	-0.04	0.03	-1.49	0.14	-0.21	
Stream PO ₄	-0.07	0.06	-1.07	0.28	-0.16	

Reef flat DIN	-0.02	0.05	-0.43	0.67	-0.09	
Reef flat PO ₄	0.12	0.08	1.53	0.13	0.35	
Wave energy	0.00	0.00	-1.59	0.11	-0.29	
Runoff	0.07	0.03	2.12	0.03	0.28	
Intensive land use	-0.19	0.13	-1.52	0.13	-0.21	
DIN Load (OSDS + pigs) kg/day	0.06	0.03	2.10	0.04	0.32	
CCA Reef Flat ~						23%
Stream DIN	0.03	0.02	1.15	0.25	0.16	
Stream PO ₄	-0.04	0.06	-0.69	0.49	-0.10	
Reef flat DIN	0.01	0.05	0.17	0.87	0.04	
Reef flat PO ₄	0.02	0.08	0.32	0.75	0.07	
Wave energy	0.00	0.00	-0.49	0.63	-0.09	
Runoff	0.08	0.03	2.48	0.01	0.32	
Intensive land use	-0.20	0.12	-1.68	0.09	-0.23	
DIN Load (OSDS + pigs) kg/day	0.05	0.03	1.96	0.05	0.29	

- 1
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10
- 11
- 12
- 13
- 14
- 15
- 16
- 17

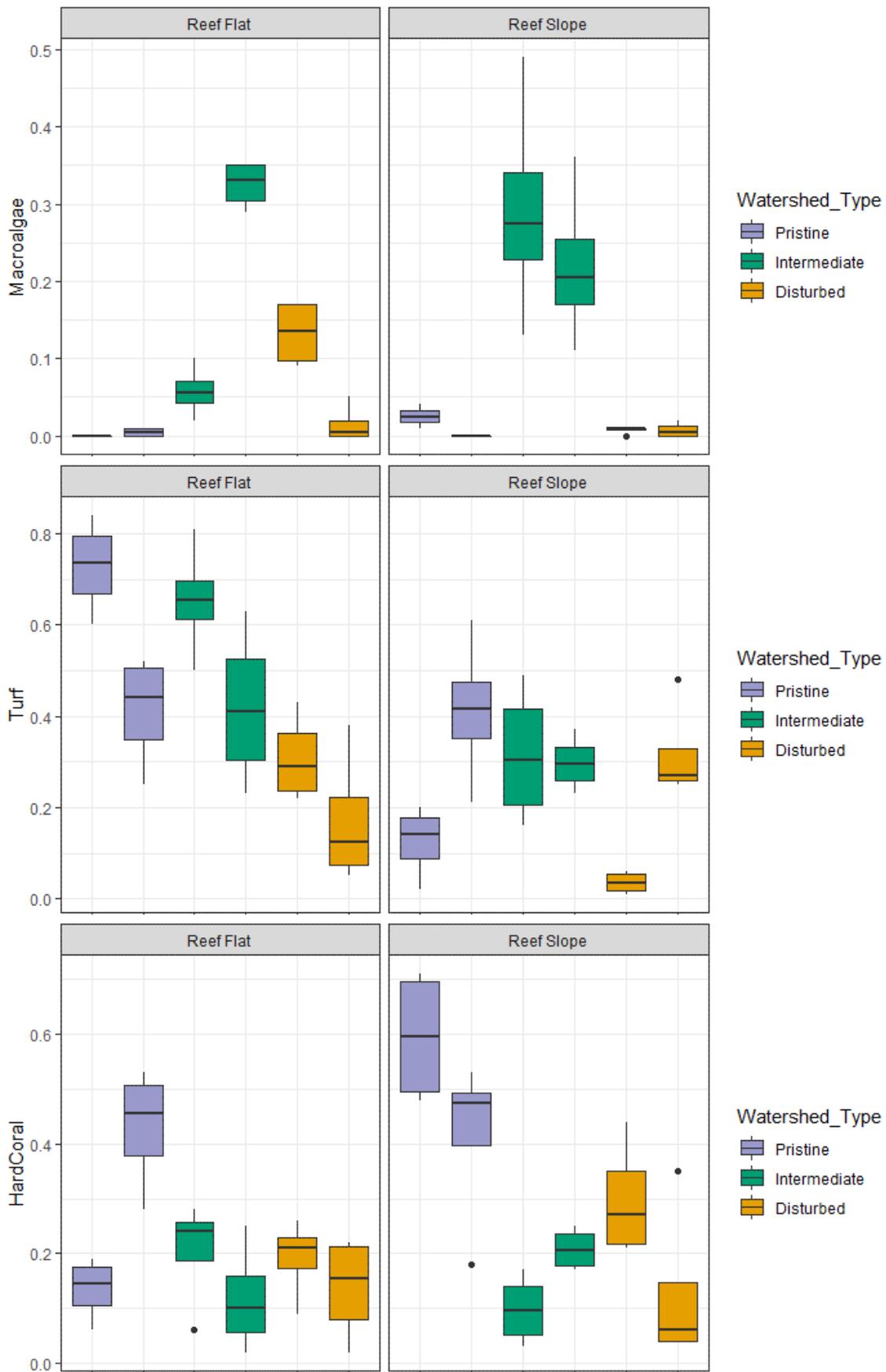
1 **Supplementary Information:**

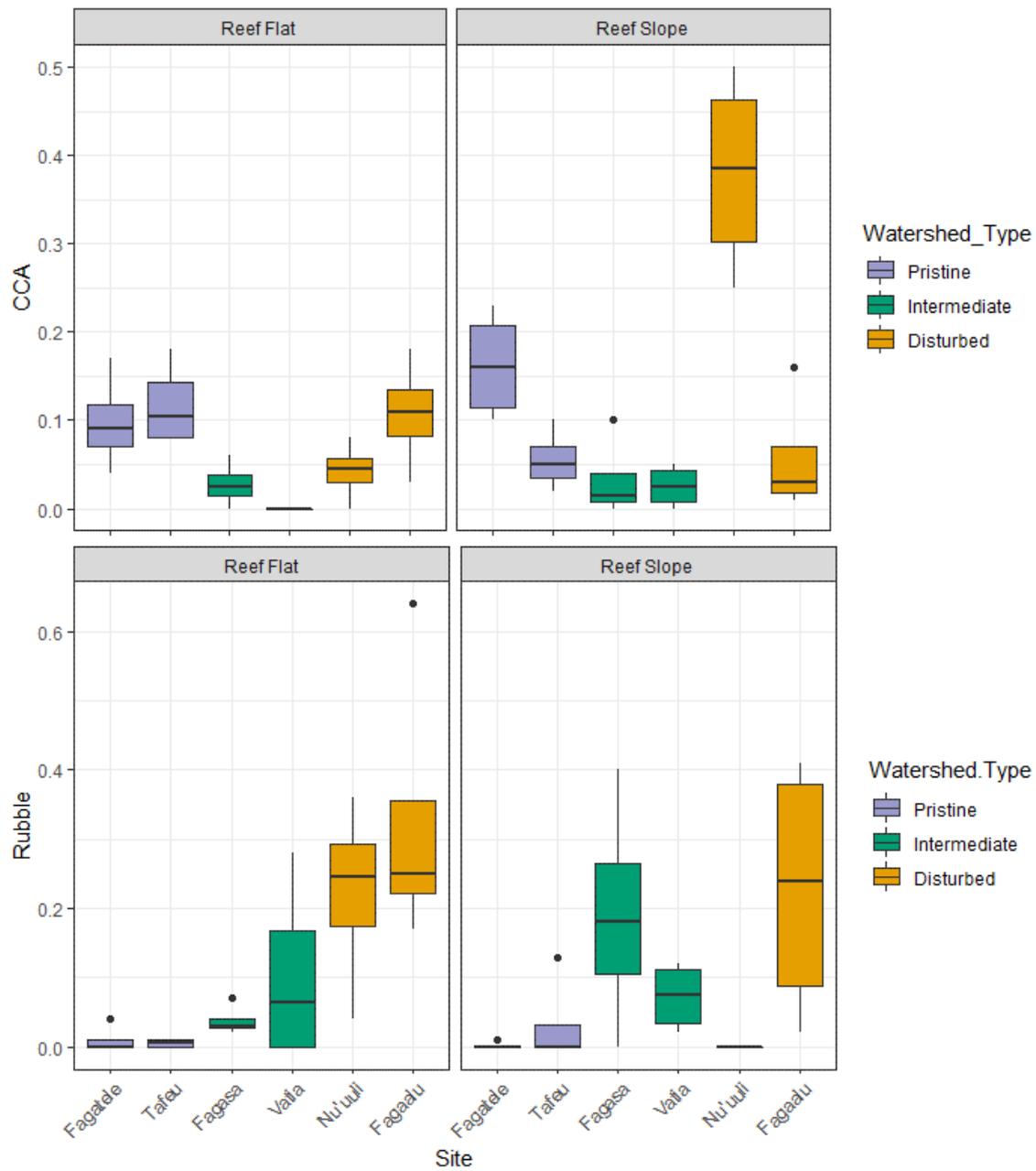
2 Table S1. Ridge and Reef site attributes

Site	Ridge					Reef	
	Watershed type	Human population density (pop/km²)	Average monthly rainfall (m3/day)	Average monthly runoff (m3/day)	DIN load (kg/day)	Mean wave exposure (joules/m²)	Sediment trap depth (m)
Fagatele	Pristine	0	2264.57	344	0.34	19.3	5- 7
Tafeu	Pristine	0	4840.26	1322.57	0.33	100.34	5- 7
Fagasa	Intermediate	831	3724.96	1079.06	3.3	230.63	5- 7
Vatia	Intermediate	640	4670.99	1159.68	2.65	220.35	5- 7
Nu'uuli	Extensive	3955	17375.57	3856.21	0.68	985.63	5- 7
Faga'alu	Extensive	910	2616.27	761	6.71	1663.78	5- 7

3

4





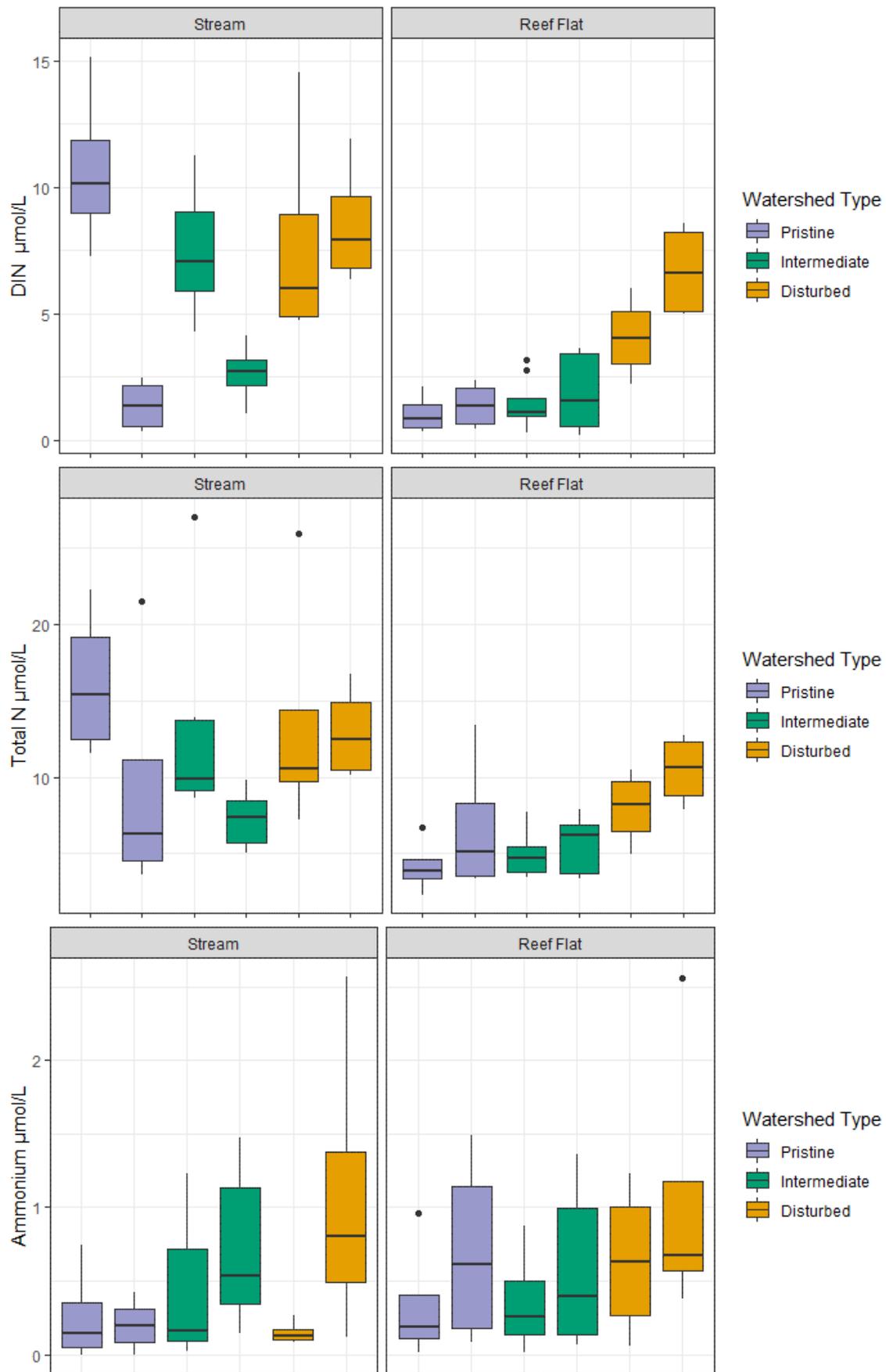
1

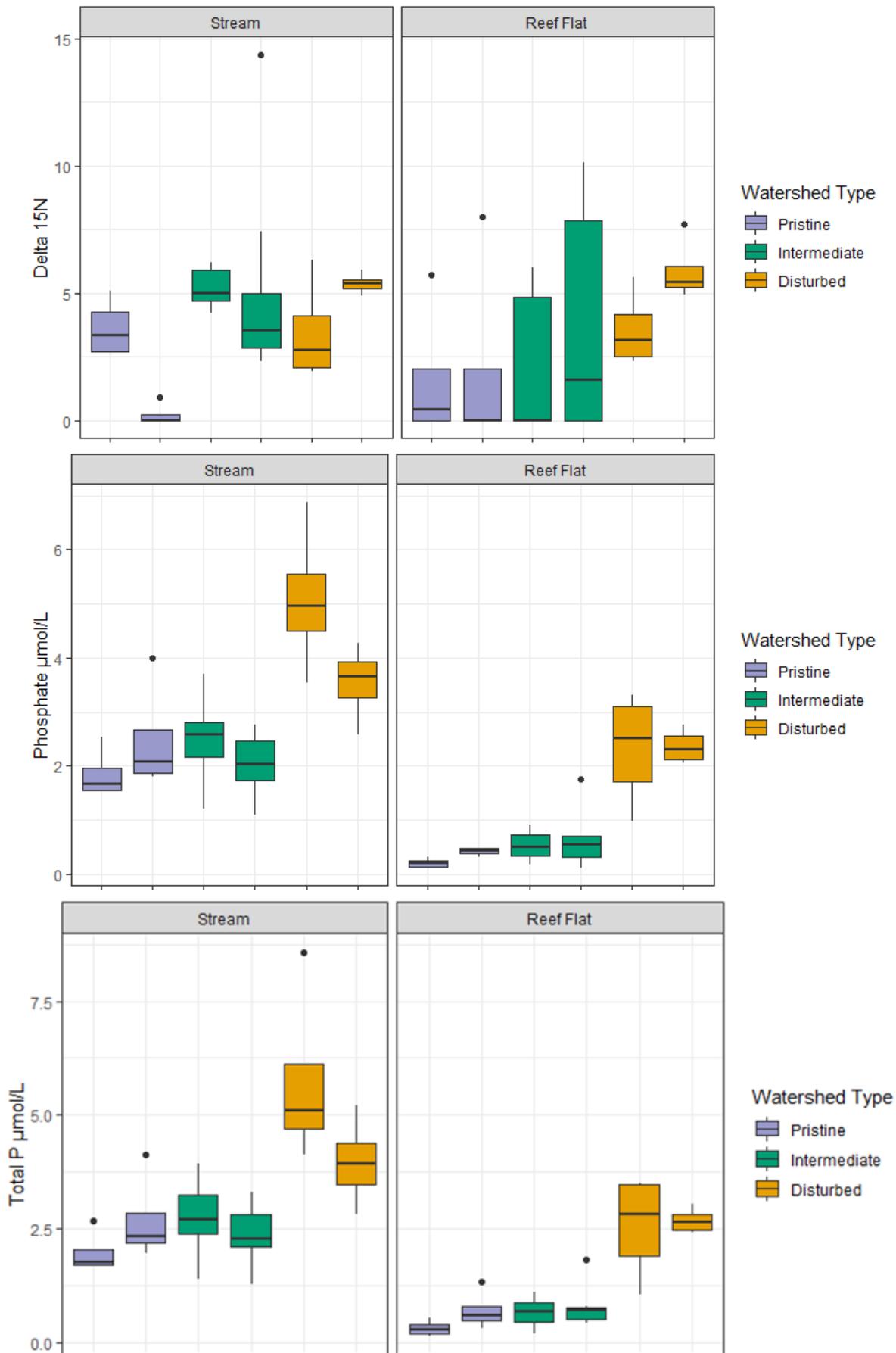
2 Figure S1. Percent cover of benthos across reef locations with black lines showing median values,
 3 boxes showing 25th and 75th percentile, and line showing 5th and 95th percentile of the data.

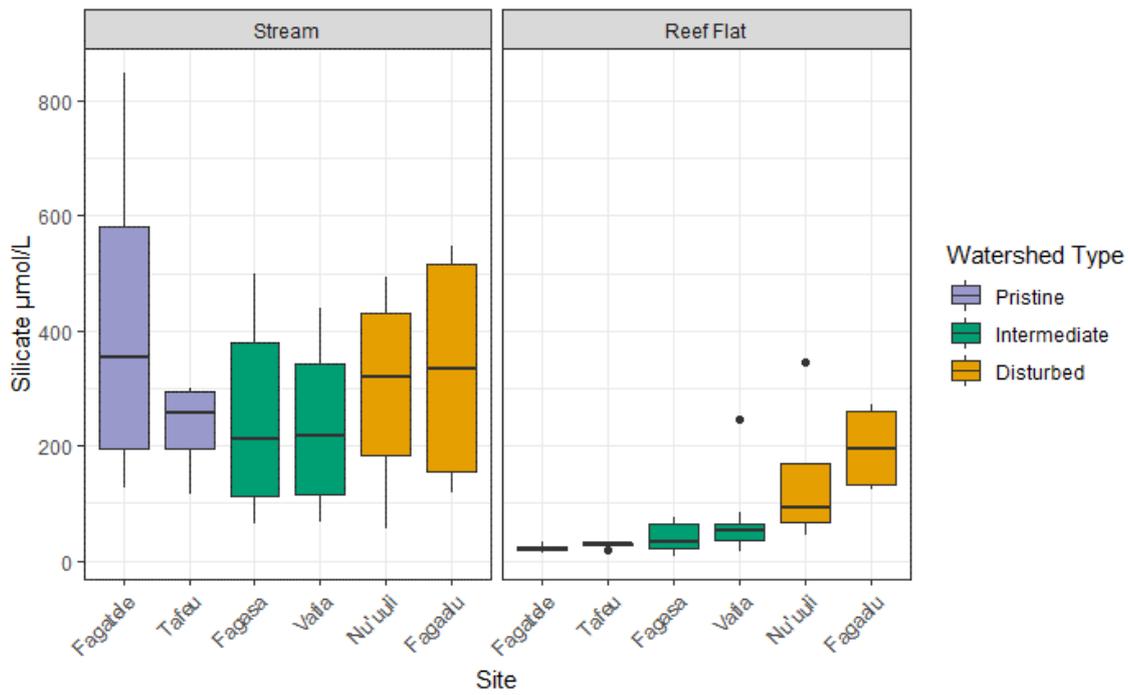
4

5

6







1

2 Figure S2. Nutrient concentrations ($\mu\text{mol/l}$) across all deployments by sampling location, with black
 3 lines showing median values, boxes showing 25th and 75th percentile, and line showing 5th and 95th
 4 percentile of the data.

5

6

7

8

9

10

11

12

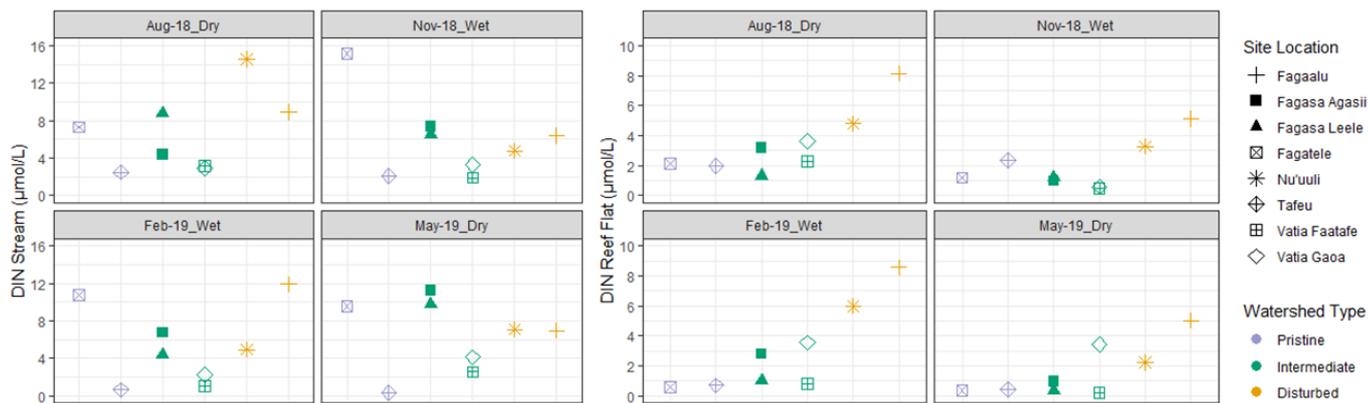
13

14

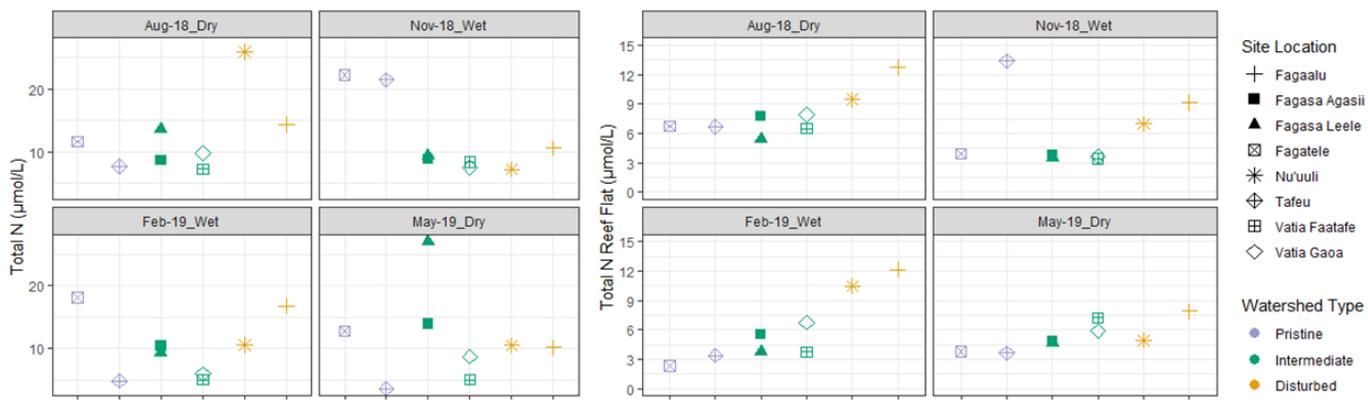
15

16

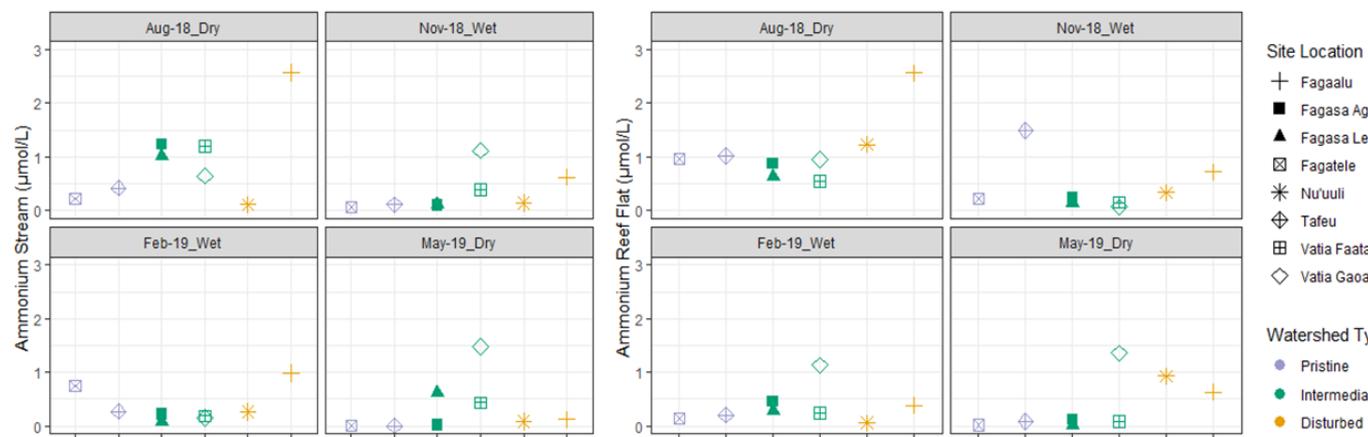
1



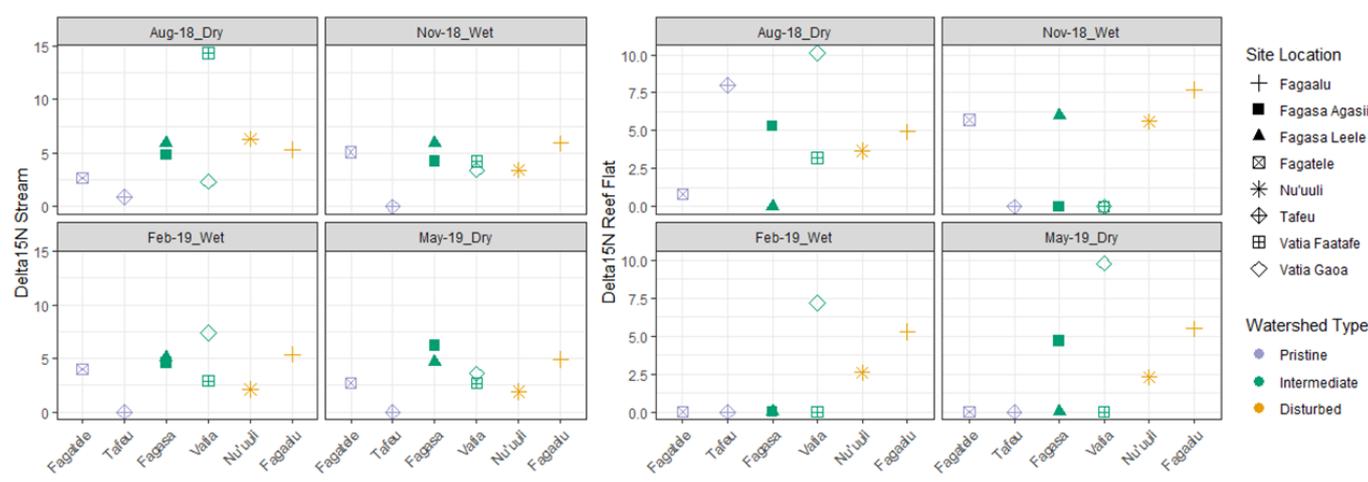
2



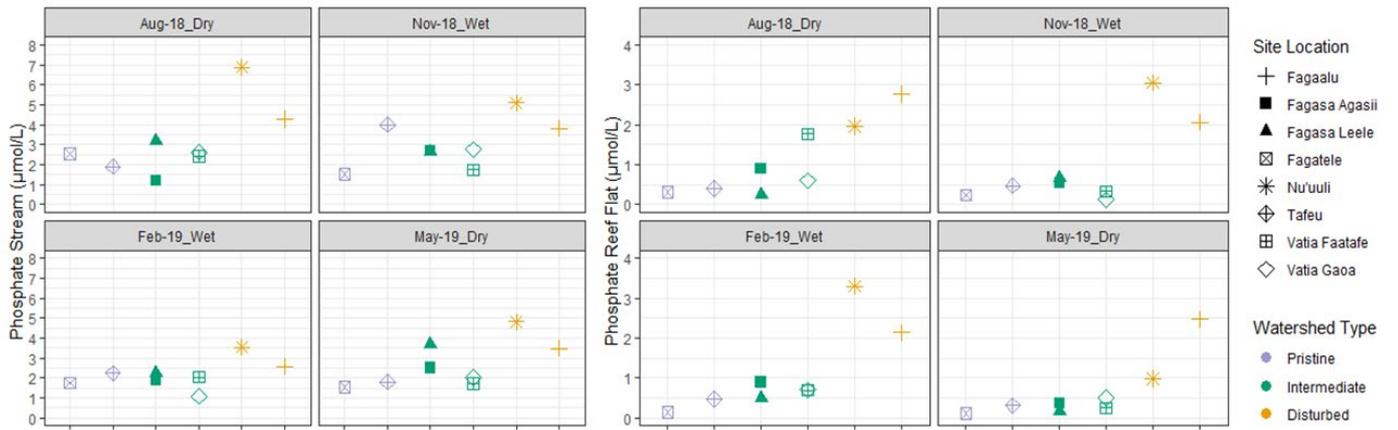
3



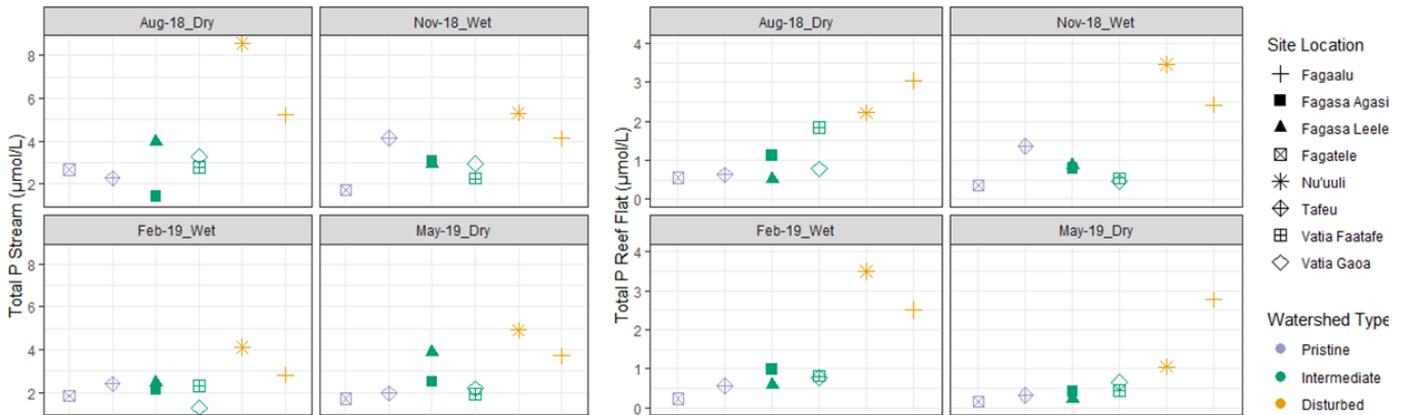
4



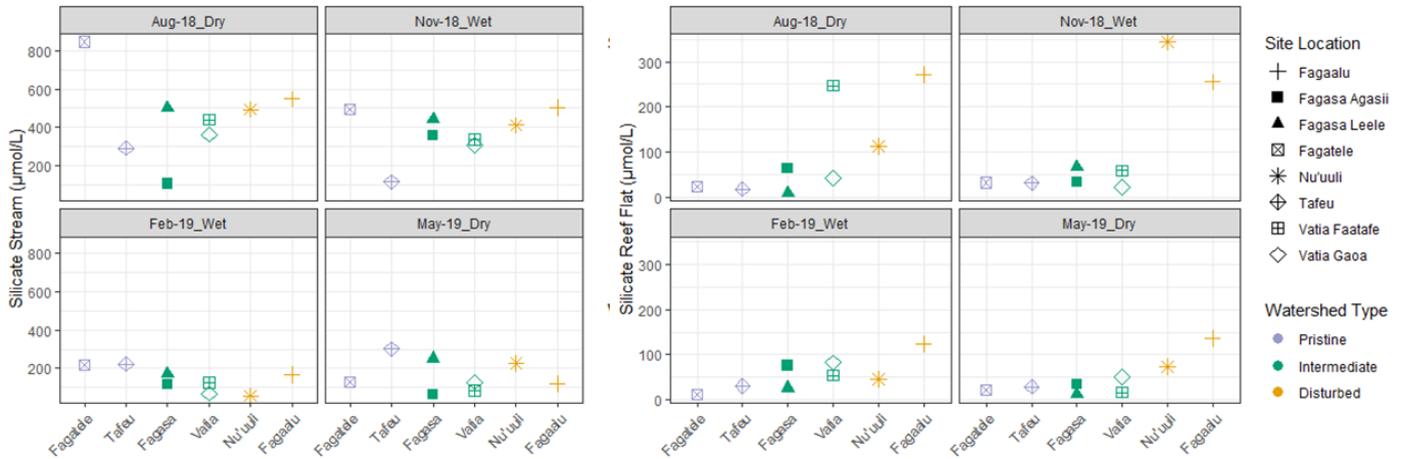
1



2



3

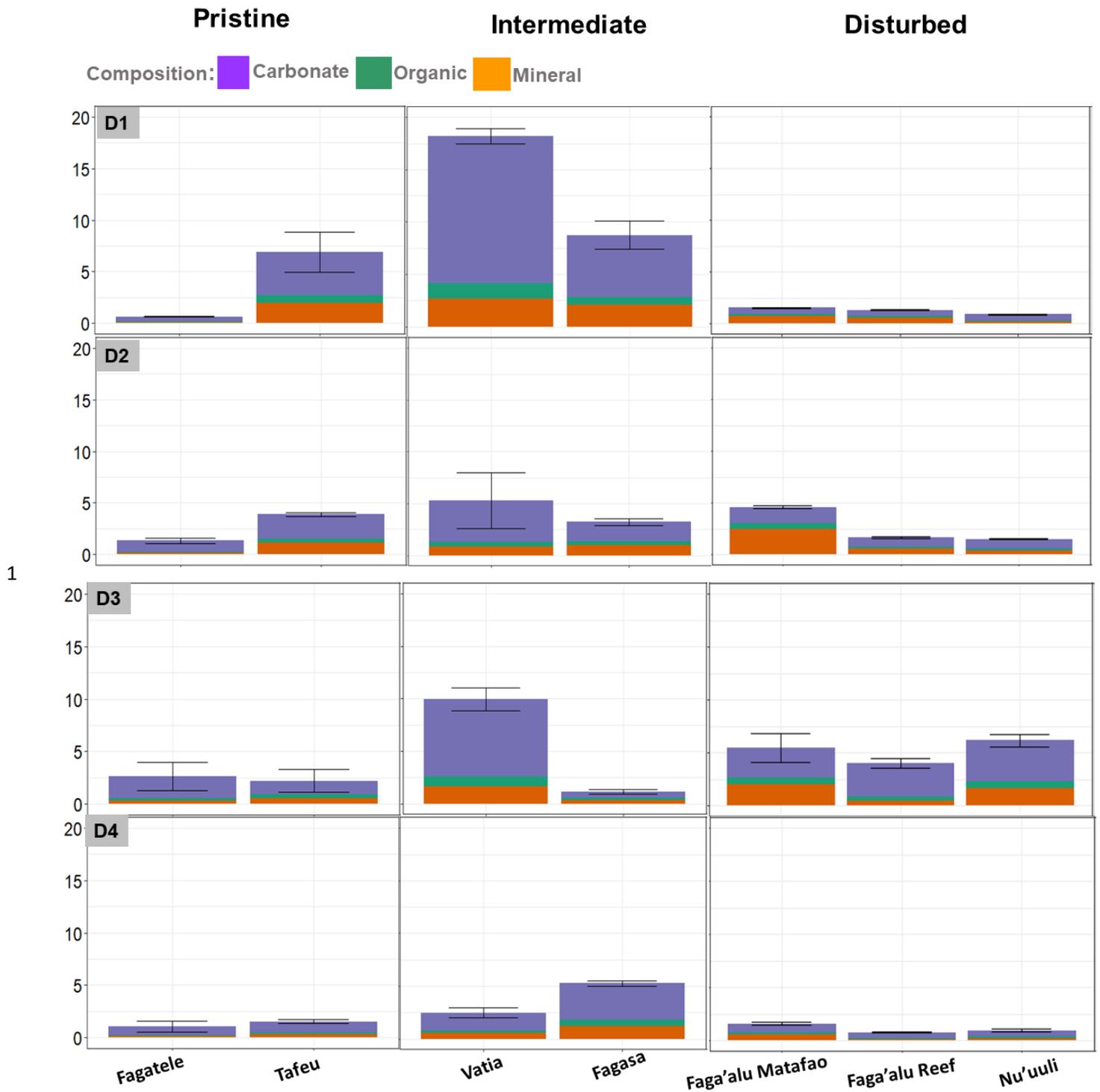


4 Figure S3. Nutrient concentrations for each sampling period collected from stream and reef flat
5 locations.

6

7

8



1

2

3 Figure S4. Pooled samples of carbonate, organic and mineral sediment composition is represented
 4 within each bar. D1 and D2 represent wet season deployments (Nov-18 – Feb-19; Feb-19 – May-
 5 19); D3 and D4 represent dry season deployments (May-19 – Sep-19; Sep-19—Nov-19).

6

7

8