



Refining Estimates of Greenhouse Gas Emissions From Salt Marsh “Blue Carbon” Erosion and Decomposition

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Coastal wetlands have sediments that contain organic matter preserved against decomposition for timespans that can range up to millennia. This “blue carbon” in wetland sediments has been proposed as a sink for atmospheric carbon dioxide and a potential source of greenhouse gases if coastal habitats are lost. A missing gap in the role of coastal habitats in the global carbon cycle is elucidating the fate of wetland sediment carbon following disturbance events, such as erosion, that can liberate organic matter to an oxygenated environment where decomposition can more readily occur. Here, we track the fate of previously stored salt marsh sediment by measuring the production of carbon dioxide (CO₂) and methane (CH₄) during an oxygenated incubation. Sediments from two depth horizons (5–10 cm and 20–25 cm) were incubated at two temperatures (20 and 30°C) for 161 days. Q₁₀ of the decomposition process over the entire course of the experiment was 2.0 ± 0.1 and 2.2 ± 0.2 for shallow and deep horizons, respectively. Activation energy for the decomposition reaction (49.7 kJ · mol⁻¹ and 58.8 kJ · mol⁻¹ for shallow and deep sediment horizons, respectively) was used to calculate temperature-specific decomposition rates that could be applied to environmental data. Using high-frequency water temperature data, this strategy was applied to coastal states in the conterminous United States (CONUS) where we estimated annual *in situ* decomposition of eroded salt marsh organic matter as 7–24% loss per year. We estimate 62.90 ± 2.81 Gg C · yr⁻¹ is emitted from eroded salt marsh sediment decomposition in the CONUS.

Keywords: salt marsh, decomposition, blue carbon, greenhouse gas emissions, erosion, sediment organic matter, carbon fluxes and pools, coastal wetlands

INTRODUCTION

The 3,000 petagrams (Pg) of stored organic carbon (OC) in living biomass and preserved biogenic organic matter (OM) in Earth’s vegetated ecosystems is enormous compared to the growth rate of atmospheric carbon dioxide (CO₂) of 4.7 Pg · C yr⁻¹ (Campbell et al., 2008; Tarnocai et al., 2009; Le Quéré et al., 2018). Among vegetated ecosystems, coastal wetlands, including salt marshes, have received attention for their “blue carbon” sediments that contain disproportionately large carbon stocks compared to their relative surface area. OM formed by salt marshes can exist for

millennia in sediments that are meters deep and are protected in place from photo-oxidation and aerobic microbial breakdown (Brevik and Homburg, 2004; Johnson et al., 2007; McTigue et al., 2019). Purposefully expanding blue carbon stocks has been suggested as part of a mitigation strategy to combat the rising CO₂ concentration in the atmosphere (Trumper et al., 2009), although only a small fraction of annual primary production is preserved in wetland sediments for centuries. Therefore, preserving this slowly accumulating C stock against anthropogenically facilitated decomposition is also critically important to limit and avoid greenhouse gas (GHG) emissions, namely CO₂ and methane (CH₄) hereafter. Few empirical data track the diagenetic fate of salt marsh sediment OM after it is released to the environment by erosion, drowning, or disturbance events (Macreadie et al., 2013; Theuerkauf et al., 2015; FitzGerald and Hughes, 2019; Schepers et al., 2020). This study specifically measures the conversion of eroded salt marsh sediment carbon to CO₂ and CH₄. This experiment mimicked the conditions of eroded sediment deposited to the marsh surface that would have been exposed to non-submerged conditions during low tide (**Supplementary Figure 1**).

Quantifying the range and temperature sensitivity of decomposition rates of this previously buried OM specifically addresses a major knowledge gap identified for advancing blue carbon science (Macreadie et al., 2019). Formerly stored wetland OC is expected to decompose when it is liberated to oxic environments after disturbance as remineralization to CO₂ occurs more readily via aerobic respiration than anaerobic processes (Trevathan-Tackett et al., 2017; Chapman et al., 2019; Spivak et al., 2019). However, biolability alone does not appear to be a straight-forward predictor of how wetland OC will remineralize (Lovelock et al., 2017). Given the varied composition (e.g., source and age) of marsh organic matter and the unpredictable, dynamic environment to which it might be exposed after disturbance, predicting decomposition is difficult without direct experimentation. For example, a global-scale estimate of greenhouse gas emissions that could occur after loss of salt marsh habitats conducted by Pendleton et al. (2012) applied a range of 25–100% decomposition to the belowground carbon stock after habitat loss. While the wide range of decomposition is appropriate for a broad-scale estimation, it illustrates the need for experimentally derived values that measure greenhouse gas production directly to refine regional and global models.

Temperature plays a critical role in regulating OM decomposition. Essentially, the fate of OM is governed by microbial enzyme kinetics, which follow Arrhenius kinetics (Davidson and Janssens, 2006). Microbial decomposition rates typically exhibit temperature sensitivity by increasing with temperature, which is often expressed in terms of the ratio of respiration rates given a 10°C difference, or Q₁₀. Additionally, the carbon-quality temperature (CQT) hypothesis states that more chemically complex OM has greater temperature sensitivity to decomposition compared to less chemically complex counterparts (Bosatta and Ågren, 1999). Thus, more stable OM that can resist biochemical and/or enzymatic degradation at one temperature can be remineralized to CO₂ at

higher temperatures (Conant et al., 2008; Craine et al., 2010). This suggests that pools of presumably stable OM stored in wetland sediments could potentially decompose under higher temperatures. Understanding the decomposition response to temperature between depth horizons allows us to extrapolate laboratory results to any temperature conditions that eroded OC would experience and test the CQT hypothesis.

Few experimental data exist that specifically track decomposing wetland OC to GHGs. In mangrove forests, sediment organic carbon was reduced by 12–34% · yr⁻¹ after the clearing of *Rhizophora mangle* (red mangrove) forest based upon sediment core carbon stocks across a 6-year space-for-time (i.e., a chronosequence) experiment (Sweetman et al., 2010). This approach provided potential decomposition rates but did not track the fate of the lost material from treatment plots and assumed the difference in carbon stock was remineralized to CO₂ as opposed to translocated elsewhere and still remaining intact. In salt marshes, *Spartina alterniflora* (salt marsh cordgrass) belowground biomass was reduced by 55% over 18 months during a litter bag incubation (Benner et al., 1991). Similar to the chronosequence experiment, the litterbag approach does not account for the physical loss of small particulate organic carbon and leached dissolved organic carbon that does not ultimately remineralize to CO₂; thus, this technique might overestimate actual decomposition and greenhouse gas emissions. Another study attributed the loss of sediment OC from wrack-induced disturbance of *S. alterniflora* to the loss of belowground biomass (Macreadie et al., 2013). However, the authors present a clear framework that while the fate of the OC lost after habitat disturbance could be microbial decomposition to CO₂, it was also possible the OC was physically exported to another location or consumed by detritivores. Therefore, to provide context to field observations, greenhouse gas production needs to be measured explicitly to quantify the rate of sediment OC loss.

Marsh surface sediments are generally protected from erosion by the reduction of wave energy and turbulence from aboveground marsh vegetation, which in turn promotes long-term sediment accretion and surface elevation gain (Möller et al., 1999; Leonard and Croft, 2006; Mudd et al., 2010). However, due to factors including bank erosion, sea level rise, eutrophication, and land-use change, the disturbance of buried wetland OM is common (Gedan et al., 2009; Deegan et al., 2012; Fagherazzi et al., 2013; Leonardi et al., 2016). Deep, anoxic marsh sediment containing decades to millennia-old marsh carbon can be exposed and enter tidal waters in a number of ways: wave erosion of the lower layer of a marsh cliff edge (Tonelli et al., 2010), barrier-island erosion and rollover (Theuerkauf and Rodriguez, 2017), and ponding following loss of marsh vegetation (Wilson et al., 2014; **Figure 1**). The erosion and redistribution of marsh shoreline sediment is a constant process and wave energy and geomorphology are primary drivers of erosion rates (Fagherazzi et al., 2013; Mariotti and Carr, 2014; Leonardi et al., 2016; Ganju et al., 2020). Development of coastal property and maintenance of navigation channels can also result in the degradation of salt marshes and release of buried sediment carbon. Once exposed and released into tidal waters, marsh sediment C may be transported by incoming tides back onto the

salt marsh platform or deposited in adjacent subtidal sediments (Canuel et al., 2009; Hopkinson et al., 2018; **Figure 1** and **Supplementary Figure 1**). In each of these scenarios eroded marsh C may spend days to months exposed to oxygen before reburial or transport into an anoxic sedimentary environment (**Figure 1**). We conducted experiments to reproduce conditions where sub-surface marsh sediments are released from the anoxic marsh platform and remain exposed to oxygen in a light-limited environment for months.

In this study, we address what proportion of salt marsh belowground organic carbon will remineralize to CO₂ or CH₄ after an erosion event. Salt marsh erosion to oxic horizons was simulated by incubating sediment organic matter from two depths in oxic chambers at two different temperatures and directly measuring CO₂ and CH₄ production. We utilized these results to estimate the amount of belowground carbon released annually via shoreline erosion in the conterminous United States (CONUS). Because few experimental data exist to substantiate the current range of belowground organic carbon conversion to GHGs, this study provides empirical data to refine the range of potential emissions from the post-disturbance decomposition of salt marsh sediment carbon. These data will provide context for estimates of the effect of global warming on marsh C cycles.

MATERIALS AND METHODS

Sample Collection

Replicate cores ($n = 10$) were collected from Freeman Creek marsh located on Marine Corps Base Camp Lejeune on the coast of North Carolina (34° 35.69' N, 77° 15.12' W). Cores were collected with a Russian peat corer (5 cm diameter) from a section of interior marsh dominated by *Spartina alterniflora*. Cores were collected at 0.062 m NAVD88 at the interface of short-form and tall-form *S. alterniflora*. Samples were transferred to PVC cradles and wrapped in cling wrap to minimize contact with air for transport to the NOAA lab in Beaufort, NC. 5 L of creek water (salinity = 32) was collected from the site.

Incubation Chamber Design

Decomposition incubations were designed to measure the production of CO₂ and CH₄ after simulated erosion of marsh sediment to oxygenated conditions (**Supplementary Figure 2**). 500 mL PETG (polyethylene terephthalate copolymer) bottles were used as incubation chambers due to their low CO₂ permeability (8.06 cm³-mil/m²-24 h-atm). Each bottle was sealed with a Nalgene 38–430 cap with two 1/4–28 ports fitted with a polypropylene spin collar (Western Fluids Engineering, CA, United States). Each port was then fitted with a quick-connect coupling insert with matching threads and a valved shut-off system so that when the insert is not connected to its female counterpart, it is closed to air exchange (Part # PMCD24042812; Colder Products, MN, United States). Teflon tape was wrapped around the insert's threads to ensure a tight fit. The mating part was a quick-connect coupling body (Part # PMC130412) with no valve, which was attached to the inlet and outlet of the greenhouse gas analyzer using 1/4-inch PTFE tubing. Without a valve, the

inflow and outflow of air to the greenhouse gas analyzer was unimpeded when disconnected from the coupling counterpart. During the experiment, the cap ports were submerged in water further decreasing their chance of contamination with the ambient atmosphere. Chambers were wrapped in aluminum foil to ensure darkness during incubation.

Experimental Design

A 2-way factorial experiment was conducted testing the effect of temperature and sediment depth, a proxy for age and OM quality, on the decomposition rate of salt marsh sediment OM. Two 5-cm segments were cut from each replicate core and designated as shallow (5–10 cm below surface) or deep (20–25 cm below surface) sediment. The segment depths were decided after visual inspection of the cores confirmed the shallower depth material was situated in the rhizosphere while the latter was beneath the rhizosphere. The shallow and deep sections of wet sediment from all 10 cores were homogenized by hand for 5 min in a clean Ziploc bag before dividing ~1 g wet weight of sediment into tared incubation chambers. Sediment wet weight was recorded. Two replicate aliquots were also prepared from each homogenized sample and placed in previously combusted (550°C) aluminum boats for subsequent analysis of water and organic carbon content.

Creek water was filtered with a 0.22 μm membrane filter. An inoculum of unfiltered creek water was reintroduced to the filtered water at a concentration of 6 mL L⁻¹ after passing over 55 μm Nitex mesh to remove large particulates. To ensure that any microautotrophs present in the inoculate would not fix inorganic carbon during the incubation, diuron (3-(3,4-dichlorophenyl)-1,1-dimethylurea) was added to inhibit photosynthesis (Morin et al., 2018). Diuron is a common algicide that inhibits photosynthesis by specifically interrupting photosystem II but does not interrupt other metabolisms to our knowledge (Rutherford and Krieger-Liszskay, 2001). For every 1L of creek water, 60 μL of diuron solution (1.0134 g dissolved in 25 mL of 100% ethanol) was added. 3 mL of inoculated creek water was added to each incubation chamber and was shaken to mix with the sediment. Each chamber was sealed with the previously described 2-ported cap and submerged in an incubation bath at either 20 or 30°C. Replicate ($n = 3$) incubations were conducted simultaneously for each depth interval (shallow vs. deep) and temperature (20 vs. 30°C) for a total of 12 incubation chambers.

Greenhouse Gas Analysis

Concentrations of CO₂ and CH₄ were simultaneously measured in each incubation chamber using an ultra-portable greenhouse gas analyzer (GGA; Los Gatos Research, Inc., CA, United States). The GGA utilizes off-axis integrated cavity output spectroscopy to detect instantaneous gas concentrations at part-per-billion sensitivity. CO₂ and CH₄ production were measured as change in concentration over time. When the GGA is coupled to the incubation chamber by connecting its inlet and outlet ports to the chamber's ported cap, a closed system is created. System equilibration occurred when the incubation chamber's gas concentration homogenizes with the gases in the instrument's

1. Pond erosion*
2. Scarp erosion*
3. Sediment becomes suspended via wave and current energy
4. Transportation in water column
5. Deposition to estuarine bottom or export to coastal ocean
6. Tidal transportation to marsh surface*

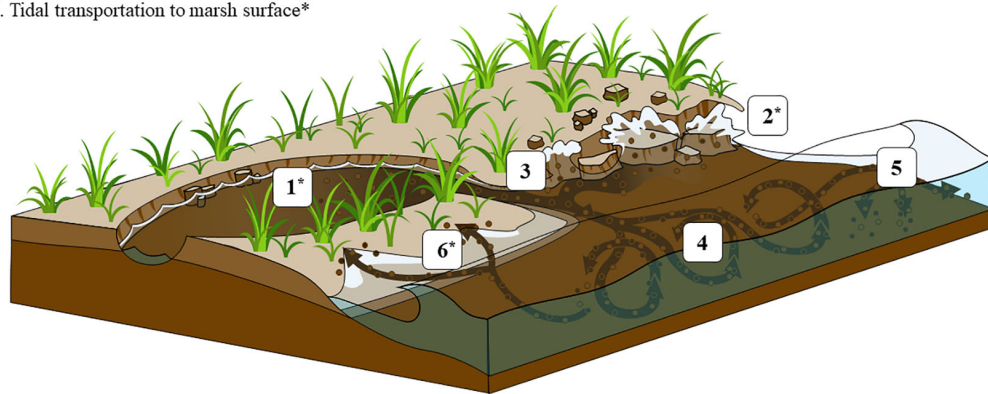


FIGURE 1 | Conceptual figure depicting salt marsh carbon liberation and redistribution vectors. 1–2 illustrate processes that release buried sediment carbon from intact marshes. 3–6 show physical processes that move sediment to locations where decomposition can occur. This experiment mimics the conditions of wet sediment exposed to air, which occurs after either pond erosion (1) or edge erosion (2) and subsequent tidal transport of eroded material to the sediment surface at either the erosion site or elsewhere (6). The decomposition of sediment suspended in the water column and transported away from the erosion site was not measured by this experiment.

headspace and a constant concentration reading is reported by the instrument. During the days between readings while the chamber was detached from the GGA, the port caps were sealed with closed valves and CO_2 (and CH_4) accumulated as respiration of OM occurred. To measure the concentration at the next time point, the GGA would again be connected to the chamber ports and the concentration of CO_2 (and CH_4) from the headspace inside the GGA would equilibrate with chamber gases.

The concentration of gases within the chamber (C_{chamber}) was derived from the equation

$$C_{\text{TOTAL}} \cdot V_{\text{TOTAL}} = V_{\text{GGA}} \cdot C_{\text{GGA}} + V_{\text{chamber}} \cdot C_{\text{chamber}}$$

where V = headspace volume and C = gas concentration, GGA = the greenhouse gas analyzer, chamber = decomposition chamber, and TOTAL = chamber connected to the GGA by the inlet and outlet ports. This approach accounts for the CO_2 and CH_4 concentrations in the GGA volume prior to mixing with the decomposition chamber's atmosphere. The Ideal Gas Law was assumed since temperature and pressure were known. This methodology of measuring gas accumulation also replenishes oxygen to the incubation chamber every time they were connected to the GGA ensuring oxic decomposition could continue. The incubation chamber was never less than 90% initial oxygen if we assumed the CO_2 produced during incubation required an equal amount of oxygen using a 1:1 molar ratio.

Gas concentrations were measured approximately every 3 days for the initial 6 weeks of the incubation, after which measurements were less frequent at about once per week. After each measurement, the amount of C respired was subtracted from the available substrate. The experiment was maintained until all incubation chamber replicates from every treatment reached

at least 4% loss of substrate, which occurred after 161 days. A reference gas mixture (402 ppmv CO_2 , 1,999 ppmv CH_4 ; Scott-Marrin Inc., Riverside, CA) was measured periodically to ensure there was no drift in the instrument.

Elemental Analysis of Sediments

Representative aliquots ($n = 4$) of incubated sediment were used to estimate the initial carbon content in the wet sediment dispersed into each incubation chamber. Replicate sediment aliquots were dried to a constant weight at 60°C . Water content was measured as wet weight minus dry weight. Once dry, sediments were homogenized by hand with mortar and pestle and acidified with 0.1 M HCl. No bubbling was observed. A small subsample of each was weighed to the nearest 10^{-6} g and wrapped in a silver tin for elemental analysis (Costech ECS 4010). Internal acetanilide standards consistently produced a coefficient of variance less than 1.5%.

Calculating Decomposition Rates, Q_{10} , and Activation Energy

Decomposition rate was calculated as the amount of CO_2 produced per available OC substrate in a measured time interval, standardized as $\mu\text{mol C} \cdot \text{mol C}^{-1} \cdot \text{day}^{-1}$. Q_{10} represents the temperature sensitivity for a reaction with a 10°C increase, defined as:

$$Q_{10} = \left(\frac{R_w}{R_c} \right)^{\frac{10}{(T_w - T_c)}}$$

where R_w is the decomposition rate at the warmer temperature (in this case 30°C), R_c is the decomposition rate at the cooler temperature (in this case 20°C), and $(T_w - T_c)$ is the difference

between the two temperatures. As decomposition of OM occurs in sediment, the more labile fraction of OM might be consumed initially while less labile fractions within the same parcel of sediment take longer to decompose, which would bias Q_{10} over the duration of the experiment. The metric Q_{10-q} developed by Conant et al. (2008) standardizes Q_{10} to substrate quality to ensure that Q_{10} is not influenced by changing lability of organic matter substrate. Following Conant et al. (2008), Q_{10-q} was used to check if the changing quality of OM was a driver of differing decomposition rates by comparing the time required to decompose a fraction of OM at the cooler temperature (t_c) to the time required to decompose that same fraction at the warmer temperature (t_w), calculated as

$$Q_{10-q} = \left(\frac{t_c}{t_w} \right)^{\frac{10}{(T_w - T_c)}}$$

If the decreasing quality of sediment substrate is affecting the decomposition rate, then t_c should increase for later fractions of OM and cause Q_{10-q} to increase.

To calculate activation energy (E_a), we used the form of the Arrhenius equation

$$\ln k_2 = -\frac{E_a}{R} \left(\frac{1}{T_2} - \frac{1}{T_1} \right) + \ln k_1$$

where R is the gas constant ($0.008314 \text{ kJ} \cdot \text{K}^{-1} \cdot \text{mol}^{-1}$), k_1 and k_2 are rates of reactions at temperatures T_1 and T_2 in Kelvin. Once E_a was derived for the decomposition reaction for shallow and deep sediments, then the temperature-dependent rate (k_2 , or k_T hereafter) at any temperature (T) could be determined using E_a and setting k_1 to the rate at 20°C .

In situ Water Temperature Records for Site-Specific Decomposition

Water temperature time series data were gathered to apply the previously determined k_T to *in situ* conditions in order to calculate site-specific, temperature-dependent decomposition rates. Long-term monitoring has produced continuous water temperature data from a tide gauge located in an estuarine embayment inside of the New River Inlet, approximately 8 km from where cores were collected. A 600LS data sonde (YSI Inc., OH, United States) was secured to a dock piling and recorded estuarine water temperature every 6 min. The temperature sensor utilizes a thermistor of sintered metallic oxide that changes predictably in resistance as temperature varies. The sonde was regularly checked every 1–2 months for fouling and battery life. Temperature was factory calibrated with an accuracy of $\pm 0.15^\circ\text{C}$. Due to the proximity of sites adjacent to the New River Estuary, we assume the temperature at the monitoring station approximates the temperature distribution of the coring station.

Water temperature data from coastal states in the CONUS were gathered as well. Six-minute water temperature data from 171 NOAA Center for Operational Oceanographic Products and Services (CO-OPS) water level stations were downloaded using the R package *rnoaa* (Chamberlain, 2021). Of those, 155 stations met the requirements of containing a continuous 12-month period at least 95% temporally complete. 149 of those

were >99% complete. Temperature was rounded to the nearest integer to sum temperature durations to derive k_2 from Eq. (4) in subsequent calculations.

For our study site and each CO-OPS station, duration at each temperature was integrated with k_T and summed to determine annual decomposition rates for each station ($\text{mol C} \cdot \text{mol C}^{-1}$, or %). Results from each station were standardized to annual rates by dividing the integrated sum by the proportion of record completeness. Applying the most-complete temperature record from each station regardless of start year or month effectively randomizes sampling date and ensures regional trends are not an ephemeral artifact of sampling period. A mean decomposition rate was determined for each state. Standard deviation of the mean state decomposition rate was calculated to represent variation of temperature records in each state.

C Emissions From Eroded Marsh Sediment in CONUS

In order to determine annual C emissions from decomposition of eroded salt marsh sediment, the previously determined average statewide decomposition rate was multiplied by the C in estimated eroded salt marsh sediment volume. Annual input (m^3) of eroded marsh sediment was estimated by multiplying state-by-state estimates of marsh shoreline length by an estimate of horizontal annual erosion and vertical bank height (Currin et al., 2015). For all coastal states except Louisiana, we used marsh shoreline length from Gittman et al. (2015). A wide range in marsh shoreline has been reported for Louisiana, which we narrowed by extracting the ‘salt and brackish water marsh’ objects from the Louisiana Environmental Sensitivity Index (Office of Response and Restoration, 2020). For all marsh shorelines, we assumed an erosion rate of $0.3 \text{ m} \cdot \text{yr}^{-1}$ and a vertical bank height of 0.3 m, which fall within the low end of published values (Currin et al., 2015; Leonardi et al., 2016; Sapkota and White, 2019). A marsh sediment C density value of $27.0 \text{ kg} \cdot \text{m}^{-3}$ (Holmquist et al., 2018) was multiplied by erosion volume to determine the amount of marsh C liberated annually via bank erosion from shorelines. The amount of eroded marsh carbon was then multiplied by the average decomposition rate for each state to determine carbon emissions from eroded salt marsh. The k_T experimentally determined for shallow sediment was applied to the top 0.1 m of the hypothetical 0.3 m tall erosional bank, while the k_T determined for the deeper horizon from our experiment was applied to the sediment from 0.1 to 0.3 m. Raw data from the experiment and R scripts used for analysis are available in the **Supplementary Material** and at <https://github.com/QAWalker/BlueCarbonErosionAndDecomp>.

RESULTS

Sediment Carbon Content

Sediment from the shallow (5–10 cm) horizon contained $11.9 \pm 0.2 \text{ \%OC}$, whereas sediment from the deep (20–25 cm) horizon contained $9.5 \pm 0.1 \text{ \%OC}$ (Table 1). The molar C:N ratio for shallow and deep sediment horizons was 22.9 ± 0.8 and 21.4 ± 0.4 , respectively. Decomposition rates were standardized

to available carbon substrate, which differed in each treatment. The shallow sediment treatments contained 93.1 ± 5.6 mg C in the 20°C chambers and 89.0 ± 4.2 mg C in the 30°C chambers. The deep sediment treatment contained 128.6 ± 30.3 mg C in the 20°C chambers and 123.7 ± 6.9 mg C in the 30°C chambers.

Decomposition Rates

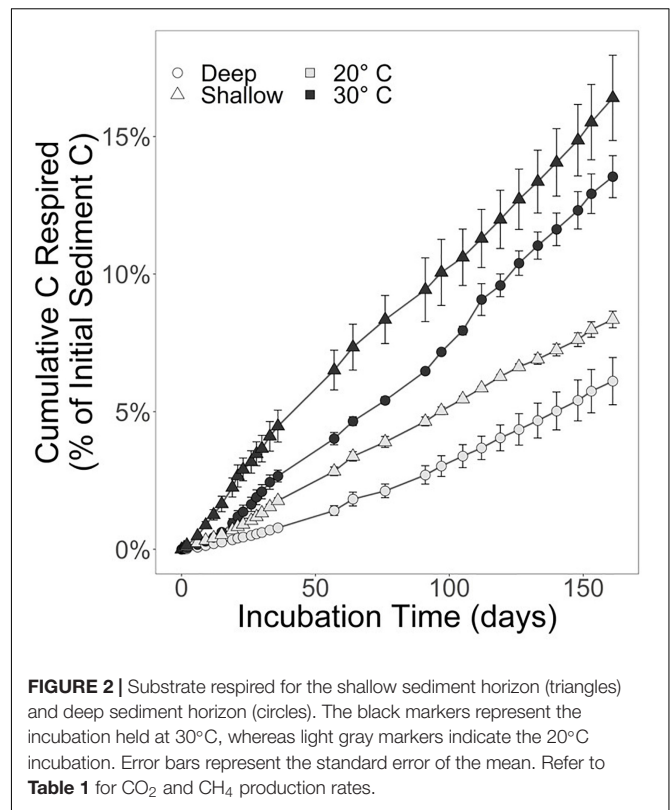
Over the course of the 161-day experiment, the CO₂ production rates were relatively linear. By day 161, $8.4 \pm 0.5\%$ of the shallow sediment treatment at 20°C was respired. In the shallow treatment at 30°C, $16.4 \pm 2.7\%$ of the starting content was respired. The deep sediment horizon had lost $6.1 \pm 1.5\%$ and $13.5 \pm 1.3\%$ of available substrate at 20 and 30°C, respectively (Figure 2).

The respiration rates at 30°C were faster than at 20°C for both sediment depth treatments (Figure 2). Moreover, the shallow sediment treatments at either temperature exhibited faster respiration rates than the deeper sediment treatments at either temperature. On average, the CO₂ production rates for the shallow sediment treatments were 518.0 ± 32.4 and $1,018 \pm 166$ $\mu\text{mol CO}_2 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$ for 20 and 30°C, respectively (Table 1). The CO₂ production rates for the deep sediment treatments at 20 and 30°C were 379.4 ± 92.3 and 840.5 ± 82.4 $\mu\text{mol CO}_2 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$, respectively.

CH₄ was produced at detectable levels during the experiment, albeit at fractional concentrations compared to CO₂ (Table 1). The shallow sediment treatments exhibited rates of CH₄ production at 0.20 ± 0.09 and 0.71 ± 0.43 $\mu\text{mol CH}_4 \text{ mol C}^{-1} \cdot \text{d}^{-1}$ at 20 and 30°C, respectively. The deep sediment treatments evolved CH₄ at rates of 0.20 ± 0.09 and 0.47 ± 0.33 $\mu\text{mol CH}_4 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$ at 20 and 30°C, respectively.

Q₁₀, Q_{10-q}, and E_a

Q₁₀ for OC loss from sediments as CO₂ and CH₄ production in shallow and deep sediment horizons was 2.0 ± 0.1 and 2.2 ± 0.2 , respectively, for the entire course of the experimental incubation. Q₁₀ between the depth horizons was not significantly



different ($p > 0.05$). In order to account for the potential change in lability of decomposable substrate, Q_{10-q} was determined for both sediment horizons (Figure 3). Q_{10-q} in the shallow sediment horizon was 2.4 ± 0.2 for the first-1% of OC substrate remineralized compared to 2.1 ± 0.3 for the fifth-1%. For the deep sediment horizon, Q_{10-q} decreased from 2.2 ± 0.2 to 1.3 ± 0.1 from the first-1% to the fifth-1% interval of substrate lost. The E_a for decomposition of sediment OM was 49.7 $\text{kJ} \cdot \text{mol}^{-1}$ for the shallow sediment horizon compared to the deeper sediment horizon at 58.8 $\text{kJ} \cdot \text{mol}^{-1}$ (Table 1).

Site Decomposition Predicted by *in situ* Temperature

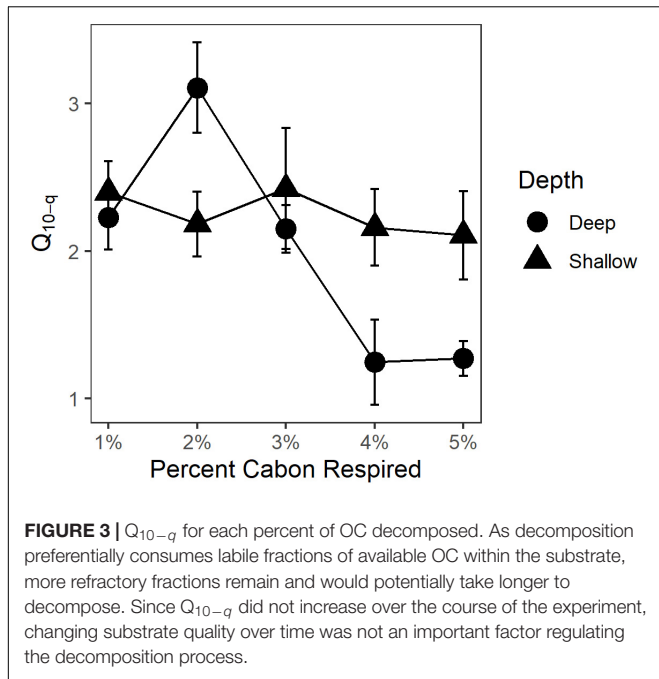
High-frequency (6-min) water temperature data from the study site collected over a year from April 2015 to March 2016 were employed to calculate annual decomposition rates of sediment OM using the respective E_a of shallow and deep sediment (Figure 4). The temperature-integrated annual decomposition rate for shallow sediment was 223 $\text{mmol C} \cdot \text{mol C}^{-1} \cdot \text{yr}^{-1}$, or 22.3% (Figure 4; Table 1). Using the same annual temperature profile, the decomposition rate for the deeper sediment with a higher E_a was 172 $\text{mmol C} \cdot \text{mol C}^{-1} \cdot \text{yr}^{-1}$, or 17.2%.

Estimated GHG Emissions From Eroded Marsh Sediment in CONUS

Salt marsh shorelines in the CONUS erode an estimated sediment volume of 12.81×10^6 $\text{m}^3 \cdot \text{yr}^{-1}$ (Table 2). The majority (>96%) of this occurs in the Atlantic and Gulf of Mexico regions where

TABLE 1 | Sediment characteristics and decomposition rates for the two experimental depth horizons.

	Shallow	Deep
Depth horizon (cm)	5–10	20–25
%OC	11.9 ± 0.2	9.5 ± 0.1
C:N (mol:mol)	22.9 ± 0.8	21.4 ± 0.4
CO ₂ production at 20°C ($\mu\text{mol CO}_2 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$)	518.0 ± 32.4	379.4 ± 92.3
CO ₂ production at 30°C ($\mu\text{mol CO}_2 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$)	1018 ± 166	840.5 ± 82.4
CH ₄ production at 20°C ($\mu\text{mol CH}_4 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$)	0.20 ± 0.09	0.20 ± 0.09
CH ₄ production at 30°C ($\mu\text{mol CH}_4 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$)	0.71 ± 0.43	0.47 ± 0.33
Q ₁₀	2.0 ± 0.1	2.2 ± 0.2
Activation Energy (kJ mol^{-1})	49.7	58.8
Annual C Loss (%)	22.3	17.2



of $62.90 \pm 2.81 \text{ Gg C} \cdot \text{yr}^{-1}$ from decomposition processes in dark, oxic conditions. A latitudinal gradient was produced where southern states exhibited higher decomposition rates (>20%) than those with more northern or Pacific Ocean coasts (**Figure 5**). Despite long coastlines, the states along the Pacific Ocean have relatively small amounts of salt marsh shoreline and cooler water year-round resulting in low amounts of eroded C (<4% of CONUS total) and C emissions (<2% of CONUS total) (**Table 2**).

DISCUSSION

The Role of Temperature in Wetland OM Decomposition

Temperature rather than organic content and C:N played the larger role in regulating salt marsh sediment organic matter decomposition because the warmer treatments of both sediment horizons decomposed faster than the lower temperature treatments (**Figure 2**). Unlike decomposition experiments that use fresh OM to simulate the fate of recently deposited materials, this experiment utilized portions of the sediment carbon pool considered “stored” and not available for decomposition under undisturbed circumstances. These materials did not exhibit the classic exponential decay of fresh OM (e.g., Benner et al., 1991; Conant et al., 2008), but rather established a roughly linear decomposition rate from the experiment’s onset (**Figure 2**).

most of the salt marsh in the CONUS exists. Annual erosion yields an estimated 345.9 Gg C liberated from intact salt marsh banks each year resulting in potential greenhouse gas emissions

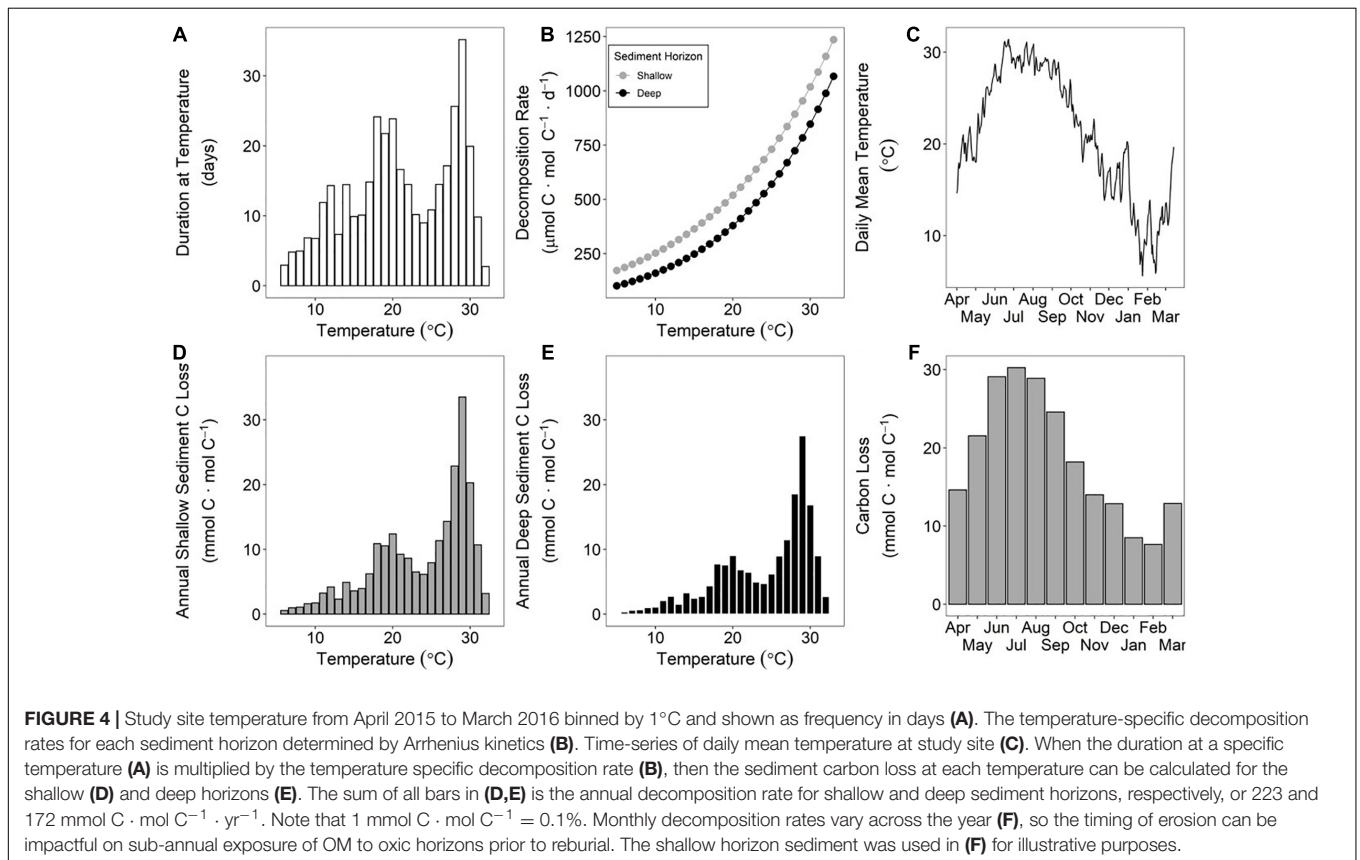


TABLE 2 | Summary of erosion and subsequent potential C emissions from CONUS.

Region	Marsh shoreline ^a (km)	Eroded sediment ($10^6 \text{ m}^3 \cdot \text{yr}^{-1}$)	Eroded carbon ($\text{Gg C} \cdot \text{yr}^{-1}$)	Potential carbon emissions ($\text{Gg C} \cdot \text{yr}^{-1}$)
Atlantic	55,719	5.01	135.4	20.82 ± 0.71
Gulf of Mexico	81,846	7.37	198.9	41.01 ± 2.72
Pacific	4,797	0.43	11.7	1.07 ± 0.12
CONUS	142,362	12.81	345.9	62.90 ± 2.81

An eroded bank height of 0.3 m and erosion rate of $0.3 \text{ m} \cdot \text{yr}^{-1}$ were applied to marsh shoreline length to determine eroded volume. Sediment volume was converted to C using $27.0 \text{ kg} \cdot \text{m}^{-3}$ (Holmquist et al., 2018). The potential carbon emissions were the summed product of state mean decomposition rate and eroded C. Error was propagated from the standard deviation of the mean decomposition value from each state. Individual state values reported in **Supplementary Table 1**.

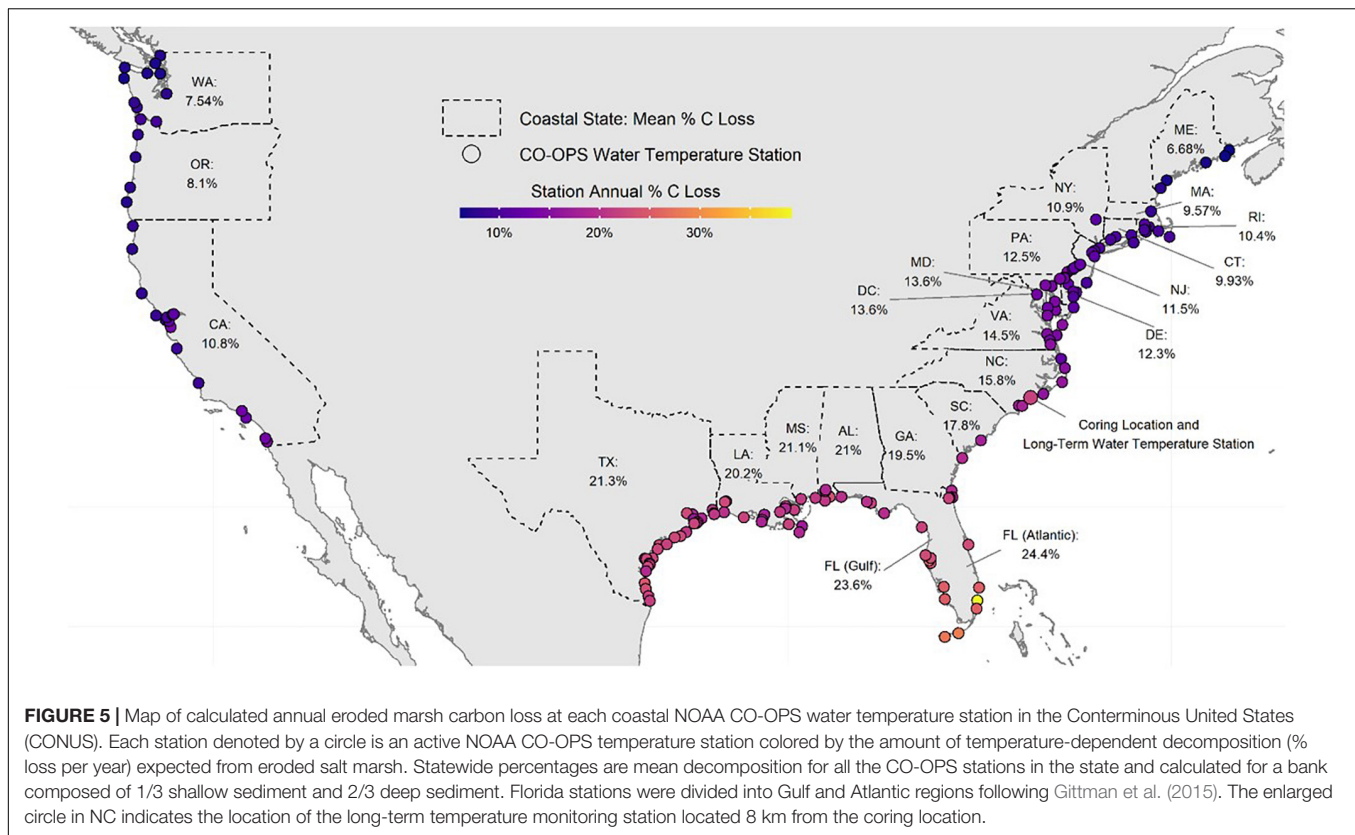
^aFrom Gittman et al. (2015) for all states except LA (Office of Response and Restoration, 2020).

The initial rapid decomposition associated with fresher OM is attributed to the preferential microbial remineralization of the more labile components before the portion of OM that is more resistant to microbial enzymatic breakdown is all that remains (Benner et al., 1991). After the labile fractions have been lost, there is a relatively slower, linear remineralization phase. Decomposition occurs in multiple phases because sediment OM consists of more than one-pool (Davidson et al., 2000). For the context of decomposition experiments, three-pools of sediment organic carbon exist with distinct lability and response to temperature: one fraction is labile, one is more resistant to degradation but will slowly remineralize, and the third is stable over the timescales that incubation experiments operate (Knorr et al., 2005). One explanation for the trend observed during this experiment is that the labile fraction of the sediment OM in both horizons had been remineralized prior to our collection and subsequent incubation. Therefore, decomposition rates shown here are for the more stable components of sediment OM, and notably, there was no reactive, quickly degrading fraction in the stored salt marsh sediment obtained from the cores, even at the 5–10 cm interval below the surface. This is further supported by the lack of increase in Q_{10-q} over the course of the experiment (**Figure 3**). An increase in Q_{10-q} over the incubation period would indicate that over time the quality of substrate is changing due to labile fractions preferentially being remineralized earlier in the incubation, and the less labile fractions taking longer to decompose later in the experiment (Conant et al., 2008, 2011). Contrary to this expectation, the slight decrease in Q_{10-q} reflects that decomposition rates were slowest at the beginning of the incubation. This could be caused by the microbial community in the lower temperature incubations for both sediment horizons taking longer to achieve maximum respiration rates.

Apart from temperature or substrate quality, other factors, such as mineral protection, water content, redox zonation, and plant-microbe interactions are important factors that determine the decomposition of wetland OM (Dungait et al., 2012; Reynolds et al., 2017; Spivak et al., 2019). These factors no doubt regulate the accumulation and loss of OM in wetland sediments but might operate more prominently within intact sediments. Our experimental design controlled some of these factors by homogenizing the microbial community from the water column and conducting oxic incubations as the intent was to simulate erosion from an intact marsh. The incubated sediments were also physically homogenized and spread thin to maximize

oxygenation and contact with microbes. Moreover, mineral protection of organic matter contained in the sediment matrix is likely a prominent factor that causes differences in decomposition rates between the incubation of homogenized sediment vs. plant biomass (Craine et al., 2010; Dungait et al., 2012).

We presume the deeper horizon contained older OM. Using a ^{210}Pb geochronology for salt marsh cores near our study site within a similar elevation range -0.159 to 0.128 m NAVD88, McKee et al. (2018) derived a mean (\pm standard deviation) sediment accretion rate of $2.4 \pm 0.9 \text{ mm} \cdot \text{yr}^{-1}$ using the Constant Flux model and $2.7 \pm 1.4 \text{ mm} \cdot \text{yr}^{-1}$ using the Constant Flux Constant Sedimentation model. These rates fall on the low end of shorter-term measures of marsh surface elevation change measured at nearby *S. alterniflora* marshes of $2\text{--}12 \text{ mm} \cdot \text{yr}^{-1}$ (Davis et al., 2017), and are consistent with reduced sediment accretion at higher elevation, interior marshes. Assuming these rates determined by the ^{210}Pb geochronology encompass the sediment accretion rate experienced at the coring location for this study, the shallow horizon age was between 19 to 42 years old before collection while the deeper horizon was between 74 to 104 years old before collection. The deeper, older horizon of sediment was respired slower compared to the shallower horizon at either temperature for the entire duration of the experiment, but there is not a dramatic difference between the chemical composition of the two horizons. Carbon content was slightly higher in the shallow horizon ($11.9 \pm 0.2\%$) vs. the deeper horizon ($9.5 \pm 0.1\%$), but the molar C:N ratios were similar (22.9 ± 0.8 vs. 21.4 ± 0.4 for shallow and deep, respectively), suggesting that the sediment OM pools were similar (**Table 1**). The sediment accretion rate could be conservative for estimating OM age since “pre-aged” material eroded from the marsh edge can be deposited on the marsh platform and reincorporated into marsh sediment (Canuel et al., 2009; Hopkinson et al., 2018; **Figure 1**). The “pre-aging” of the material incorporated into the sediment matrix could explain the similarity in chemical composition between the two horizons since they might contain OM of the age. Still, these results reveal a paradox of similar chemical character yielding different decomposition rates. The coarse elemental analysis conducted here does not provide the resolution required to elucidate the character of the more reactive shallower treatment other than demonstrating a slightly higher carbon content. Moreover, the conundrum of both sediment horizons exhibiting no detectable difference in quality precluded us from the testing the CQT hypothesis directly. The sediment



horizons exhibited similar temperature sensitivity (i.e., similar Q_{10}) albeit at different respiration rates due to the different E_a measured for each horizon.

The temperature sensitivity of salt marsh OM has implications in scaling up our results to landscape or regional climate models and would have a positive feedback on CO_2 -induced warming since the deeper (>0.1 m) sediment that acts as a carbon reservoir prior to erosion becomes an increasing source of GHGs as it experiences warmer temperatures post-erosion. To demonstrate this concept, we will use a hypothetical incubation chamber comprised of equal parts of the two depth horizons used in this experiment that continue to exhibit consistent decomposition rates presented above. At 5°C the decomposition rate would total $137 \mu\text{mol CO}_2 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$ of which 63% of the CO_2 is from the shallow pool and 37% is from the deep (Figure 6). However, at 30°C the total decomposition increases to $933 \mu\text{mol CO}_2 \cdot \text{mol C}^{-1} \cdot \text{d}^{-1}$ but the proportion of decomposition changes to 55% and 45% for shallow and deep fractions, respectively. Given the continued disturbance of wetlands via erosion (Leonardi et al., 2016; Sapkota and White, 2019; Donatelli et al., 2020), sea-level rise (FitzGerald and Hughes, 2019), and eutrophication (Deegan et al., 2012), a large portion of the presently stable deeper horizons of wetland sediment could be remineralized to CO_2 . Our experiment was conducted under conditions similar to those of marsh sediment exposed to the atmosphere, which occurs during low tides. When decomposition occurs while the marsh sediment is submerged or after tidal flushing of pore spaces, CO_2 from marsh decomposition enters the DIC pool and can be

advected into estuarine and coastal ocean waters where it could exchange with the atmosphere or become retained in deeper waters (Wang et al., 2016; Maher et al., 2018; Najjar et al., 2018).

While the initial transport of eroded material can be to oxygenated settings, including the salt marsh surface and water column (Hopkinson et al., 2018), eventually most of that material is reburied into anoxic sediment horizons where oxic breakdown will halt (Figure 1). Since oxic decomposition will not likely continue indefinitely, the timing and frequency of the erosion event is an important consideration for decomposition of material that remains in an oxic environment less than a year. At the North Carolina coring site, if salt marsh was hypothetically eroded in April but reburied 6 months later, it would experience the highest temperatures of the year and decompose with correspondingly higher rates (Figures 4C,F). Comparatively, material eroded in October and reburied 6 months later would experience the colder winter temperatures and decompose approximately half that of what would have occurred in the previous 6 months.

OC accumulation in wetlands occurs, in part, due to the anoxic state of their waterlogged sediments. Phenolic compounds (e.g., lignin) that are integral to the physical structure of vascular plants can resist decomposition within wetland sediments since the enzyme phenol oxidase required to breakdown phenolic compounds requires oxygen (Janusz et al., 2017). Therefore, the oxygenation of wetland sediment upon disturbance (e.g., wetland erosion or drainage) can unfasten the enzymatic “latch” that preserves stored belowground OM from remineralization to CO_2

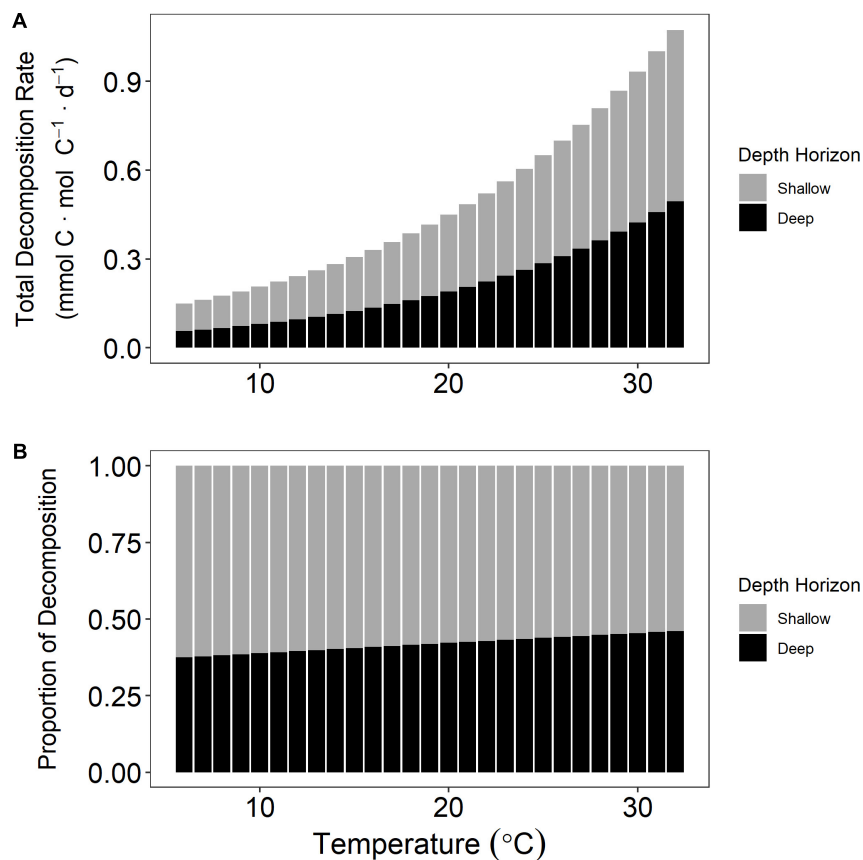


FIGURE 6 | Decomposition rate at multiple temperatures for a hypothetical incubation chamber that contained equal parts of both sediment horizons. Total decomposition rate increases with temperature (A). The proportional contribution of the deep horizon that possesses a higher activation energy (E_a) increases with temperature (B).

(Freeman et al., 2001). A portion of the CO_2 flux we measured could be from lignin and other phenolic compounds that were previously stored in the sediments unable to degrade prior to the simulated erosion. We note that our results are specific to salt marsh sediments that have been oxygenated.

We measured small fluxes of CH_4 production, an anaerobic process, despite the oxic and near-ocean salinity conditions of the experiment. This suggests that anoxic microsites formed within the sediment matrix of the experimental substrate (Ribas et al., 2019). CH_4 is a more potent greenhouse gas than CO_2 with a 100-year global warming potential (GWP) of 28 (Boucher et al., 2009). Considering this factor, the daily CH_4 production was still dwarfed by the CO_2 production. For example, in the shallow sediment incubation at 30°C , there was still 51 times more CO_2 production than GWP-standardized CH_4 production. Using oxygen to respire sediment carbon yields far more energy than methanogenesis, so these proportions are not surprising. However, it should be noted that even in an oxygenated, saline experimental setting, detectable amounts of CH_4 were produced, which further supports that environmental factors cannot always predict methane emission (Al-Haj and Fulweiler, 2020). Given that methanogenesis occurred during the incubation, other anaerobic processes like sulfate reduction or denitrification may

have occurred as well until the terminal electron acceptors for those processes were exhausted.

It should be noted that sediments with inorganic carbon constituents (e.g., oyster reefs, seagrass beds, coral reefs, etc.) would require additional consideration if used in this experimental design. Since a high CO_2 atmosphere forms in the decomposition chambers, porewater pH would decrease along with aragonite saturation. As carbonate minerals dissolve, some CO_2 would be consumed altering the measurable respiration. A high CO_2 headspace and, thus, a low pH porewater could impact microbial activity as well; however, this was unmeasured in our study. The relatively linear rates of CO_2 production do not suggest there were intermittent interruptions in microbial performance (Figure 2).

Refining Estimates of Carbon Emissions Post-disturbance Through Extrapolation

Extrapolated decomposition rates throughout CONUS were not an exact linear function of salt marsh shoreline due to regionally different temperature regimes. For example, North Carolina, South Carolina, and the Gulf Coast of Florida erode less C each year than Maryland, but each has higher total C

emissions from decomposition since their coastal waters are warmer (**Supplementary Table 1**). Louisiana, with the longest marsh shoreline tallied here, erodes almost seven times more C annually than Virginia with the next longest marsh shoreline, but the warmer temperatures in Louisiana account for almost 10 times the C emissions.

The calculations conducted for this study are conservative for several reasons. The decomposition rates assume no photo-oxidation takes place, which most likely would take place *in situ* and would increase the decomposition rate (Wang et al., 2020). The introduction of sediment into a nutrient-rich water column may prime the microbial community and enhance decomposition (Ward et al., 2016; Bulseco et al., 2019). The rates reported here are specifically microbial respiration, and decomposition rates could potentially be higher *in situ* in the presence of grazers and detritivores. Our estimate also does not include additional losses that take place after habitat disturbance, as loss of salt marsh plants via erosion results in the loss of additional plant carbon fixation via photosynthesis or facilitation of sedimentation and subsequent burial of carbon. The experimentally determined decomposition rates exhibited variation of about 10%, which was not propagated into the state-by-state C emission rates. Lastly, tidal creeks where salt marshes exist can experience warmer temperatures than NOAA CO-OPS tide gauges, which are often located closer to the coastal ocean; therefore, the state-by-state decomposition estimates may be conservative. Despite these limitations, the decomposition rates extrapolated to CONUS provide context to previous estimates of C emissions.

Previous studies that estimate C emissions from salt marsh loss have been forced to make assumptions about decomposition rates due to the lack of empirical data that track the fate of sediment organic carbon to GHGs after habitat loss. Pendleton et al. (2012) estimated between 0.2 and 0.24 Pg C · yr⁻¹ of global C emissions from salt marshes after land-use change or habitat loss. Their emission rates were broadly bounded with decomposition rates ranging between 25 and 100%. Similarly, Sapkota and White (2019) estimated C emissions from eroded salt marsh from Barataria Basin, LA, assuming a decomposition rate of 75% loss of sediment organic carbon. Conversely, our estimates for salt marsh sediment organic matter decomposition throughout CONUS range from 7 to 24% annual loss, which accounts for *in situ* temperature of the environments into which salt marsh OM is eroded. We assumed that E_a remains unchanged for 1 year to facilitate our calculation of an annual decomposition rate, but our laboratory measurements of GHG production lasted for only 161 days. It is possible that the rates would slow as decomposition progresses, thus, increasing E_a over the time during which the OM can decompose (Conant et al., 2008). For this reason, it is unrealistic to extrapolate these rates over a time frame required to reach 100% loss of OM. For example, while we report a mean state decomposition rate for Texas at $\sim 20\% \cdot \text{yr}^{-1}$, it is not implied that eroded salt marsh OM will completely decompose in 5 years. Moreover, a major question remains unanswered that is required to understand the long-term fate of eroded materials: will the eroded salt marsh OM remain in oxic conditions indefinitely or might they eventually

become reburied? Evidence that eroded marsh edge OM is deposited on the marsh platform and reburied suggests that there is a finite time during which aerobic decomposition can take place (Hopkinson et al., 2018; **Figure 1**). We used the utility of reporting rates as per year; however, this does not imply the duration of decomposition of disturbed sediment *in situ*. One approach to estimate the anoxic duration would be to divide the depth that the anoxic zone in the marsh sediment begins by the sediment accretion rate if both were known.

The estimated rates of C emissions from eroded salt marsh were calculated with several caveats. We assumed all salt marsh decomposition occurs with the same E_a as measured in the laboratory experiment. It is plausible that E_a differs geographically as emergent vegetation and soil types change (Craine et al., 2010). Two states (SC and GA) had only one CO-OPS station that met our requirements; in these cases, the entire state's salt marsh C emissions depended on the temperature record from one location. The decomposition calculation also hinges on quantifying salt marsh length. As this value becomes more accurately measured over time, especially in states with extensive wetlands like Louisiana, the estimate for decomposition from salt marshes will too refine. We also used a conservative, fixed bank height and erosion rate for the entire CONUS, but these vary spatially as they are dependent on a multitude of hydrographic, morphologic, and chemical variables. Opting for lower erosion rates also helps balance the occurrence of prograding and non-eroding marsh shorelines. This extrapolation exercise is one example of an application for measuring temperature-based decomposition rates of coastal sediment organic matter to predict future GHG emission scenarios. Blue carbon ecosystems are renowned for their abilities to accrete and store carbon but refining how losses to their extent feed a positive GHG feedback loop is essential to understanding their full value.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, and further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

NM and CC conceived and designed the experiment. NM, QW, and CC collected, analyzed, and interpreted the data. NM and QW generated the tables and figures. NM wrote the manuscript with input, guidance, and revisions from QW and CC. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmars.2021.661442/full#supplementary-material>

REFERENCES

- Al-Haj, A. N., and Fulweiler, R. W. (2020). A synthesis of methane emissions from shallow vegetated coastal ecosystems. *Glob. Change Biol.* 26, 2988–3005. doi: 10.1111/gcb.15046
- Benner, R., Fogel, M., and Sprague, E. (1991). Diagenesis of belowground biomass of spartina alterniflora in salt-marsh sediments. *Limnol. Oceanogr.* 36, 1358–1374. doi: 10.4319/lo.1991.36.7.1358
- Bosatta, E., and Ågren, G. I. (1999). Soil organic matter quality interpreted thermodynamically. *Soil Biol. Biochem.* 31, 1889–1891. doi: 10.1016/S0038-0717(99)00105-4
- Boucher, O., Friedlingstein, P., Collins, B., and Shine, K. P. (2009). The indirect global warming potential and global temperature change potential due to methane oxidation. *Environ. Res. Lett.* 4:044007. doi: 10.1088/1748-9326/4/4/044007
- Brevik, E. C., and Homburg, J. A. (2004). A 5000 year record of carbon sequestration from a coastal lagoon and wetland complex, Southern California, USA. *CATENA* 57, 221–232. doi: 10.1016/j.catena.2003.12.001
- Bulsecò, A. N., Giblin, A. E., Tucker, J., Murphy, A. E., Sanderman, J., Hiller-Bittroff, K., et al. (2019). Nitrate addition stimulates microbial decomposition of organic matter in salt marsh sediments. *Global Change Biol.* 25, 3224–3241. doi: 10.1111/gcb.14726
- Campbell, A., Lera, M., Igor, L., Holly, G., and Adam, H. (2008). *Carbon Storage in Protected Areas: Technical Report*.
- Canuel, E. A., Lerberg, E. J., Dickhut, R. M., Kuehl, S. A., Bianchi, T. S., and Wakeham, S. G. (2009). Changes in sediment and organic carbon accumulation in a highly-disturbed ecosystem: the Sacramento-San Joaquin River Delta (California, USA). *Mar. Pollut. Bull.* 59, 154–163. doi: 10.1016/j.marpolbul.2009.03.025
- Chamberlain, S. (2021). *rnoaa: 'NOAA' Weather Data from R. R package version 1.3.2*. Available online at: <https://CRAN.R-project.org/package=rnoaa>
- Chapman, S. K., Hayes, M., Kelly, B., and Langley, J. A. (2019). Exploring the oxygen sensitivity of wetland soil carbon mineralization. *Biol. Lett.* 15:407. doi: 10.1098/rsbl.2018.0407
- Conant, R. T., Drijber, R. A., Haddix, M. L., Parton, W. J., Paul, E. A., Plante, A. F., et al. (2008). Sensitivity of organic matter decomposition to warming varies with its quality: temperature sensitivity of organic matter decomposition. *Glob. Change Biol.* 14, 868–877. doi: 10.1111/j.1365-2486.2008.01541.x
- Conant, R. T., Ryan, M. G., Ågren, G. I., Birge, H. E., Davidson, E. A., Eliasson, P. E., et al. (2011). Temperature and soil organic matter decomposition rates – synthesis of current knowledge and a way forward. *Glob. Change Biol.* 17, 3392–3404. doi: 10.1111/j.1365-2486.2011.02496.x
- Craine, J. M., Fierer, N., and McLaughlan, K. K. (2010). Widespread coupling between the rate and temperature sensitivity of organic matter decay. *Nat. Geosci.* 3, 854–857. doi: 10.1038/ngeo1009
- Currin, C., Davis, J., Baron, L. C., Malhotra, A., and Fonseca, M. (2015). Shoreline change in the New River Estuary, North Carolina: rates and consequences. *J. Coast. Res.* 315, 1069–1077. doi: 10.2112/JCOASTRES-D-14-00127.1
- Davidson, E. A., and Janssens, I. A. (2006). Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature* 440, 165–173. doi: 10.1038/nature04514
- Davidson, E. A., Trumbore, S. E., and Amundson, R. (2000). Soil warming and organic carbon content. *Nature* 408, 789–790. doi: 10.1038/35048672
- Davis, J., Currin, C., and Morris, J. T. (2017). Impacts of fertilization and tidal inundation on elevation change in microtidal, low relief salt marshes. *Estuar. Coasts* 40, 1677–1687. doi: 10.1007/s12237-017-0251-0
- Deegan, L. A., Johnson, D. S., Warren, R. S., Peterson, B. J., Fleeger, J. W., Fagherazzi, S., et al. (2012). Coastal eutrophication as a driver of salt marsh loss. *Nature* 490, 388–392. doi: 10.1038/nature11533
- Donatelli, C., Zhang, X., Ganju, N. K., Aretxabaleta, A. L., Fagherazzi, S., and Leonardi, N. (2020). A nonlinear relationship between marsh size and sediment trapping capacity compromises salt marshes' stability. *Geology* 48, 966–970. doi: 10.1130/G47131.1
- Dungait, J. A. J., Hopkins, D. W., Gregory, A. S., and Whitmore, A. P. (2012). Soil organic matter turnover is governed by accessibility not recalcitrance. *Glob. Change Biol.* 18, 1781–1796. doi: 10.1111/j.1365-2486.2012.02665.x
- Fagherazzi, S., Mariotti, G., Wiberg, P., and McGlathery, K. (2013). Marsh collapse does not require sea level rise. *Oceanography* 26, 70–77. doi: 10.5670/oceanog.2013.47
- FitzGerald, D. M., and Hughes, Z. (2019). Marsh processes and their response to climate change and sea-level rise. *Annu. Rev. Earth Planet. Sci.* 47, 481–517. doi: 10.1146/annurev-earth-082517-010255
- Freeman, C., Ostle, N., and Kang, H. (2001). An enzymic “latch” on a global carbon store. *Nature* 409, 149–149. doi: 10.1038/35051650
- Ganju, N. K., Defne, Z., and Fagherazzi, S. (2020). Are elevation and open-water conversion of salt marshes connected? *Geophys. Res. Lett.* 47:e2019GL086703. doi: 10.1029/2019GL086703
- Gedan, K. B., Silliman, B. R., and Bertness, M. D. (2009). Centuries of human-driven change in salt marsh ecosystems. *Ann. Rev. Mar. Sci.* 1, 117–141. doi: 10.1146/annurev.marine.010908.163930
- Gittman, R. K., Fodrie, F. J., Popowich, A. M., Keller, D. A., Bruno, J. F., Currin, C. A., et al. (2015). Engineering away our natural defenses: an analysis of shoreline hardening in the US. *Front. Ecol. Environ.* 13:301–307. doi: 10.1890/150065
- Holmquist, J. R., Windham-Myers, L., Bliss, N., Crooks, S., Morris, J. T., Megonigal, J. P., et al. (2018). Accuracy and Precision of Tidal Wetland Soil Carbon Mapping in the Conterminous United States. *Sci. Rep.* 8:9478. doi: 10.1038/s41598-018-26948-7
- Hopkinson, C. S., Morris, J. T., Fagherazzi, S., Wollheim, W. M., and Raymond, P. A. (2018). Lateral marsh edge erosion as a source of sediments for vertical marsh accretion. *J. Geophys. Res. Biogeosci.* 123, 2444–2465. doi: 10.1029/2017JG004358
- Janusz, G., Pawlik, A., Sulej, J., Świdorska-Burek, U., Jarosz-Wilkolazka, A., and Paszczyński, A. (2017). Lignin degradation: microorganisms, enzymes involved, genomes analysis and evolution. *FEMS Microbiol. Rev.* 41, 941–962. doi: 10.1093/femsre/fux049
- Johnson, B. J., Moore, K. A., Lehmann, C., Bohlen, C., and Brown, T. A. (2007). Middle to late holocene fluctuations of C₃ and C₄ vegetation in a northern New England salt marsh, Sprague marsh, Phippsburg, Maine. *Organic Geochem.* 38, 394–403. doi: 10.1016/j.orggeochem.2006.06.006
- Knorr, W., Prentice, I. C., House, J. I., and Holland, E. A. (2005). Long-term sensitivity of soil carbon turnover to warming. *Nature* 433, 298–301. doi: 10.1038/nature03226
- Le Quéré, C., Andrew, R. M., Friedlingstein, P., Sitch, S., Hauck, J., Pongratz, J., et al. (2018). Global carbon budget 2018. *Earth Syst. Sci. Data* 10, 2141–2194. doi: 10.5194/essd-10-2141-2018
- Leonard, L. A., and Croft, A. L. (2006). The effect of standing biomass on flow velocity and turbulence in *Spartina alterniflora* canopies. *Estuar. Coast. Shelf Sci.* 69, 325–336. doi: 10.1016/j.ecss.2006.05.004
- Leonardi, N., Ganju, N. K., and Fagherazzi, S. (2016). A linear relationship between wave power and erosion determines salt-marsh resilience to violent storms and hurricanes. *Proc. Natl. Acad. Sci. U.S.A.* 113, 64–68. doi: 10.1073/pnas.1510095112

- Lovelock, C. E., Atwood, T., Baldock, J., Duarte, C. M., Hickey, S., Lavery, P. S., et al. (2017). Assessing the risk of carbon dioxide emissions from blue carbon ecosystems. *Front. Ecol. Environ.* 15:257–265. doi: 10.1002/fee.1491
- Macreadie, P. I., Anton, A., Raven, J. A., Beaumont, N., Connolly, R. M., Friess, D. A., et al. (2019). The future of blue carbon science. *Nat. Commun.* 10:3998. doi: 10.1038/s41467-019-11693-w
- Macreadie, P. I., Hughes, A. R., and Kimbro, D. L. (2013). Loss of 'Blue Carbon' from coastal salt marshes following habitat disturbance. *PLoS One* 8:e69244. doi: 10.1371/journal.pone.0069244
- Maher, D. T., Call, M., Santos, I. R., and Sanders, C. J. (2018). Beyond burial: lateral exchange is a significant atmospheric carbon sink in mangrove forests. *Biol. Lett.* 14:20180200. doi: 10.1098/rsbl.2018.0200
- Mariotti, G., and Carr, J. (2014). Dual role of salt marsh retreat: long-term loss and short-term resilience. *Water Resour. Res.* 50, 2963–2974. doi: 10.1002/2013WR014676
- McKee, B., Bost, M., Atencio, A., Rodriguez, A., Currin, C., McTigue, N., et al. (2018). "Carbon burial in estuarine, wetland, and coastal barrier environments," in *Defense Coastal/Estuarine Research Program 2 Final Report*, ed. P. Cunningham (Durham, NC: RTI International).
- McTigue, N., Davis, J., Rodriguez, A. B., McKee, B., Atencio, A., and Currin, C. (2019). Sea level rise explains changing carbon accumulation rates in a salt marsh over the past two millennia. *J. Geophys. Res. Biogeosci.* 124, 2945–2957. doi: 10.1029/2019JG005207
- Möller, I., Spencer, T., French, J. R., Leggett, D. J., and Dixon, M. (1999). Wave transformation over salt marshes: a field and numerical modelling study from North Norfolk, England. *Estuar. Coastal Shelf Sci.* 49, 411–426. doi: 10.1006/ecss.1999.0509
- Morin, S., Chaumet, B., and Mazzella, N. (2018). A time-dose response model to assess diuron-induced photosynthesis inhibition in freshwater biofilms. *Front. Environ. Sci.* 6:131. doi: 10.3389/fenvs.2018.00131
- Mudd, S. M., D'Alpaos, A., and Morris, J. T. (2010). How does vegetation affect sedimentation on tidal marshes? Investigating particle capture and hydrodynamic controls on biologically mediated sedimentation. *J. Geophys. Res.* 115:1566. doi: 10.1029/2009JF001566
- Najjar, R. G., Herrmann, M., Alexander, R., Boyer, E. W., Burdige, D. J., Butman, D., et al. (2018). Carbon budget of tidal wetlands, estuaries, and shelf waters of eastern north America. *Glob. Biogeochem. Cycle* 32, 389–416. doi: 10.1002/2017GB005790
- Office of Response and Restoration (2020). Louisiana and Lower Mississippi River 2014 ESIL (Environmental Sensitivity Index – Lines) from 2010-06-15 to 2010-08-15. Available online at: <https://www.fisheries.noaa.gov/inport/item/53935> (accessed October 7, 2020).
- Pendleton, L., Donato, D. C., Murray, B. C., Crooks, S., Jenkins, W. A., Sifleet, S., et al. (2012). Estimating global "Blue Carbon" emissions from conversion and degradation of vegetated coastal ecosystems. *PLoS One* 7:e43542. doi: 10.1371/journal.pone.0043542
- Reynolds, L. L., Lajtha, K., Bowden, R. D., Johnson, B. R., and Bridgman, S. D. (2017). The carbon quality-temperature hypothesis does not consistently predict temperature sensitivity of soil organic matter mineralization in soils from two manipulative ecosystem experiments. *Biogeochemistry* 136, 249–260. doi: 10.1007/s10533-017-0384-z
- Ribas, A., Mattana, S., Llubra, R., Debouk, H., Sebastià, M. T., and Domene, X. (2019). Biochar application and summer temperatures reduce N₂O and enhance CHN₄ emissions in a Mediterranean agroecosystem: role of biologically-induced anoxic microsites. *Sci. Total Environ.* 685, 1075–1086. doi: 10.1016/j.scitotenv.2019.06.277
- Rutherford, A. W., and Krieger-Liszczak, A. (2001). Herbicide-induced oxidative stress in photosystem II. *Trends Biochem. Sci.* 26, 648–653. doi: 10.1016/S0968-0004(01)01953-3
- Sapkota, Y., and White, J. R. (2019). Marsh edge erosion and associated carbon dynamics in coastal Louisiana: a proxy for future wetland-dominated coastlines world-wide. *Estuar. Coast. Shelf Sci.* 226:106289. doi: 10.1016/j.ecss.2019.106289
- Schepers, L., Brennand, P., Kirwan, M. L., Guntenspergen, G. R., and Temmerman, S. (2020). Coastal marsh degradation into ponds induces irreversible elevation loss relative to sea level in a microtidal system. *Geophys. Res. Lett.* 47:e2020GL089121. doi: 10.1029/2020GL089121
- Spivak, A. C., Sanderman, J., Bowen, J. L., Canuel, E. A., and Hopkinson, C. S. (2019). Global-change controls on soil-carbon accumulation and loss in coastal vegetated ecosystems. *Nat. Geosci.* 12, 685–692. doi: 10.1038/s41561-019-0435-2
- Sweetman, A. K., Middelburg, J. J., Berle, A. M., Bernardino, A. F., Schander, C., Demopoulos, A. W. J., et al. (2010). Impacts of exotic mangrove forests and mangrove deforestation on carbon remineralization and ecosystem functioning in marine sediments. *Biogeosciences* 7, 2129–2145. doi: 10.5194/bg-7-2129-2010
- Tarnocai, C., Canadell, J. G., Schuur, E. A. G., Kuhry, P., Mazhitova, G., and Zimov, S. (2009). Soil organic carbon pools in the northern circumpolar permafrost region: soil organic carbon pools. *Glob. Biogeochem. Cycles* 23:3327. doi: 10.1029/2008GB003327
- Theuerkauf, E. J., and Rodriguez, A. B. (2017). Placing barrier-island transgression in a blue-carbon context: barrier island carbon budget model. *Earth's Fut.* 5, 789–810. doi: 10.1002/2017EF000568
- Theuerkauf, E. J., Stephens, J. D., Ridge, J. T., Fodrie, F. J., and Rodriguez, A. B. (2015). Carbon export from fringing saltmarsh shoreline erosion overwhelms carbon storage across a critical width threshold. *Estuar. Coast. Shelf Sci.* 164, 367–378. doi: 10.1016/j.ecss.2015.08.001
- Tonelli, M., Fagherazzi, S., and Petti, M. (2010). Modeling wave impact on salt marsh boundaries. *J. Geophys. Res.* 115:6026. doi: 10.1029/2009JC006026
- Trevathan-Tackett, S. M., Seymour, J. R., Nielsen, D. A., Macreadie, P. I., Jeffries, T. C., Sanderman, J., et al. (2017). Sediment anoxia limits microbial-driven seagrass carbon remineralization under warming conditions. *FEMS Microbiol. Ecol.* 93:Fix033. doi: 10.1093/femsec/fix033
- Trumper, K., Bertzky, M., Dickson, B., van der Heijden, G., Jenkins, M., and Manning, P. (2009). *The Natural Fix? The Role of Ecosystems in Climate Mitigation: A UNEP Rapid Response Assessment*. Cambridge: United Nations Environment Programme.
- Wang, H., Hu, X., Wetz, M. S., Hayes, K. C., and Lu, K. (2020). Photomineralization of organic carbon in a eutrophic, semi-arid estuary. *Limnol. Oceanogr. Lett.* 5, 246–253. doi: 10.1002/lol2.10146
- Wang, Z. A., Kroeger, K. D., Ganju, N. K., Gonneea, M. E., and Chu, S. N. (2016). Intertidal salt marshes as an important source of inorganic carbon to the coastal ocean: marsh lateral export of inorganic carbon. *Limnol. Oceanogr.* 61, 1916–1931. doi: 10.1002/lno.10347
- Ward, N. D., Bianchi, T. S., Sawakuchi, H. O., Gagne-Maynard, W., Cunha, A. C., Brito, D. C., et al. (2016). The reactivity of plant-derived organic matter and the potential importance of priming effects along the lower amazon river: organic matter reactivity in the amazon. *J. Geophys. Res. Biogeosci.* 121, 1522–1539. doi: 10.1002/2016JG003342
- Wilson, C. A., Hughes, Z. J., FitzGerald, D. M., Hopkinson, C. S., Valentine, V., and Kolker, A. S. (2014). Saltmarsh pool and tidal creek morphodynamics: dynamic equilibrium of northern latitude saltmarshes? *Geomorphology* 213, 99–115. doi: 10.1016/j.geomorph.2014.01.002

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Conflict of Interest: QW was employed by the company CSS-Inc.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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