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Climate sensitivities and uncertainties in food-web pathways supporting larval bluefin tuna in subtropical oligotrophic oceans

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**Abstract:**

Compared to high-latitude seas, the ecological implications of climate change for top consumers in subtropical regions are poorly understood. One critical area of knowledge deficiency is the nature of food-web connections to larvae during their vulnerable time in the plankton. Bluefin tuna (BFT) are highly migratory temperate species whose early life stages are spent in ultra-oligotrophic subtropical waters. Dietary studies of BFT larvae provide evidence of prey-limited growth coupled with strong selection for specific prey types – cladocerans and poecilostomatoid copepods – whose paradoxical or poorly resolved trophic characteristics do not fit the conventional understanding of open-ocean food web structure and flows. Current knowledge consequently leaves many uncertainties in climate change effects, including the possibility that increased nitrogen fixation by *Trichodesmium* spp. might enhance resiliency of BFT larvae, despite a projected overall decline in system productivity. To advance understanding and future predictions, the complementary perspectives of oceanographers and fisheries researchers need to come together in studies that focus on the trophic pathways most relevant to fish larvae, the factors that drive variability in spawning regions, and their effects on larval feeding, growth and survival.

## 37 Introduction

38 In high latitude systems, rising temperature and density stratification associated with climate  
39 change are dramatically altering sea-ice habitats and production cycle phenologies supporting  
40 high level consumers (Loeng and Drinkwater, 2007; Wassmann *et al.*, 2011; Constable *et al.*,  
41 2014). While the ultimate impacts are still to be determined, the directions of change are  
42 both measurable and being actively investigated to elucidate cause-and-effect mechanisms  
43 and predict future consequences (e.g., Laidre *et al.*, 2008; Ducklow *et al.*, 2013). In contrast,  
44 climate change effects on already warm and nutritionally poor regions of the oceans are less  
45 clearly understood and will be more difficult to quantify relative to background variability.  
46 For such regions, predicting the consequences of future changes will also strongly challenge  
47 our understanding of how these systems and the populations dependent on them actually  
48 function in the contemporary ocean. One specific concern is the nature of food webs that  
49 underlie successful recruitment of the ocean's top predators. Here, we will use the specific  
50 example of bluefin tuna (BFT) to illustrate knowledge and conceptual gaps, arguing that  
51 current understanding of the food web structure and dynamics of oligotrophic ocean systems  
52 largely ignores the components most relevant and important for understanding the successful  
53 development of BFT larvae and their sensitivities to climate change impacts.

54 The three species of BFT (Atlantic: *Thunnus thynnus*, Pacific: *T. orientalis* and  
55 Southern: *T. maccoyii*) are the largest and broadest-migrating tunas. All populations are  
56 highly valued economically, subject to heavy fishing pressure, and currently depleted  
57 (Fromentin *et al.*, 2014; Pons *et al.*, 2017). Primarily due to their ability to regulate their  
58 body temperature through endothermy (Graham & Dickson 2001), juvenile and adult BFT  
59 have exceptional environmental tolerances. This allows them to exploit food resources over  
60 vast expanses of temperate and subpolar waters. All three species, however, spawn in  
61 distinct restricted subtropical areas, which are currently among the most nutritionally  
62 deficient habitats in the oceans and expected to get even poorer in the future (Bopp *et al.*,  
63 2001; Behrenfeld *et al.*, 2006; Chust *et al.*, 2014; Doney *et al.*, 2014; Fu *et al.*, 2016). During  
64 their first 2-3 weeks of life, BFT larvae must grow rapidly or die in such waters. They are  
65 restricted by daylight feeding and light penetration to the upper 10-20 m of surface waters  
66 (Davis *et al.*, 1990a; Habtes *et al.*, 2014), and will thereby directly experience the major  
67 projected consequences of climate forcing – warmer surface temperature, stratification-  
68 diminished productivity and increased acidity – which will collectively increase metabolic  
69 requirements as food resources presumably decline. This is the climate-vulnerable  
70 recruitment bottleneck for BFT that needs to be understood.

71 Conventional understanding of the food-web structure of warm oligotrophic seas  
72 emphasizes the dominance of <1- $\mu\text{m}$  unicellular photosynthetic bacteria, the most efficient  
73 competitors when nutrient concentrations are chronically low. In the long-established  
74 microbial loop hypothesis (Azam *et al.*, 1983), bacteria are consumed by small flagellates,  
75 and they, in turn, by ciliated protozoa, leading to inefficient transfer of productivity to small  
76 animals of the size (~1-mm) of tuna larvae prey. Increasing oligotrophy would presumably  
77 work to lengthen this protistan food chain, decreasing transfer efficiency to higher levels.  
78 But is this relevant to BFT larvae? Here, we will highlight that the preferred prey of BFT  
79 larvae, namely poecilostomatoid copepods and cladocerans, are organisms associated with  
80 alternate and poorly explored trophic pathways in oligotrophic systems. We will outline how  
81 the characteristics and variability of different types of zooplankton prey may function in food  
82 webs to enhance vulnerability or resiliency of BFT larvae to climate change, the major areas  
83 of uncertainty that need to be resolved, and the types of research efforts needed to bridge the  
84 knowledge and conceptual gaps between oceanographic and fisheries approaches to  
85 elucidating the ecology of open-ocean ecosystems.

## 86 **Bluefin tuna spawning areas**

87 Bluefin tunas are among the top predators of pelagic ecosystems in all major oceans. They  
88 range widely, make large excursions through the water column and can effectively evade or  
89 mitigate unfavorable environmental conditions by virtue of their strong swimming abilities  
90 and physiological traits such as thermal regulation (Holland *et al.*, 1992; Block *et al.*, 2001).  
91 Unlike tropical tunas, which spawn over wide geographical areas throughout much of the  
92 year, the large temperate BFTs make long-distance migrations to small geographically  
93 restricted areas in oligotrophic subtropical seas (Figure 1) where they spawn over relatively  
94 short periods (Schaefer, 2001; Shimose and Farley, 2016). Successful development of their  
95 larvae thus depends on the food resources available in narrow portions of the subtropical  
96 oceans during limited periods of time.

97 Southern bluefin tuna (SBFT) forage widely in the Southern Ocean and migrate  
98 northward to the waters between Indonesia and northern Australia to spawn during the austral  
99 summer (Figure 1; Farley *et al.*, 2015; Hobday *et al.*, 2016). Pacific bluefin tuna (PBFT)  
100 range extensively across the northern Pacific Ocean but migrate to the areas between Japan  
101 and the Philippine Islands (Nansei area) and in the Sea of Japan to spawn during the boreal  
102 spring and summer months (Itoh, 2006; Tanaka and Suzuki, 2016; Fujioka *et al.*, 2016).  
103 Atlantic bluefin tuna (ABFT) feed throughout the North Atlantic but migrate to distinct  
104 western and eastern spawning areas in the Gulf of Mexico and the Mediterranean Sea,  
105 respectively, during boreal spring and early summer (Block *et al.*, 2005; Rooker *et al.*, 2007).  
106 ABFT larvae have also recently been recorded in the Slope Sea off the Mid-Atlantic Bight  
107 during summer (Richardson *et al.*, 2016), which has similar water temperatures to the Gulf of  
108 Mexico during spring and is oligotrophic during warmer months (Xu *et al.*, 2011).

109 The waters of all known BFT spawning grounds are warm ( $>24^{\circ}\text{C}$ ), highly stratified and  
110 especially nutritionally deficient during the spawning seasons (Matsuura *et al.*, 1997;  
111 Alemany *et al.*, 2010; Muhling *et al.*, 2013; Llopiz and Hobday, 2015; Druon *et al.*, 2016).  
112 The evolutionary advantages, and ecological consequences, of larvae developing in such  
113 waters remain largely unclear. Nonetheless, some hypotheses have been proposed, such as  
114 the advantage of reduced predation pressure on eggs and larvae by other pelagic fish species  
115 in oligotrophic environments (Bakun and Broad, 2003; Bakun, 2013). Further, though the  
116 spawning regions are characterized by mesoscale eddies and fronts, the waters selected for  
117 spawning generally have retention characteristics, often with less kinetic energy than at other  
118 times of the year (Reglero *et al.*, 2014). With the exception of the Mediterranean, however,  
119 these retention areas are also adjacent to strong poleward flowing boundary currents (Gulf  
120 Stream, Kuroshio and Leeuwin Currents), which facilitate the transport of surviving post  
121 larvae away from the spawning grounds and toward juvenile nursery habitats in more  
122 productive temperate ecosystems (Brothers *et al.*, 1983; Kitagawa *et al.*, 2010; Hobday *et al.*,  
123 2016).

## 124 **Feeding preferences of larval BFT**

125 Bluefin tuna spawn huge numbers of tiny pelagic eggs. Once hatched at  $\sim 2$  mm body length,  
126 the larvae quickly develop eyes, large mouths and a functioning gut (Yúfera *et al.*, 2014),  
127 which give them the capability to grow quickly, at rates of 0.3-0.4 mm body length per day,  
128 in warm waters with sufficient food (Malca *et al.*, 2017; Muhling *et al.*, 2017).  
129 Metamorphosis from the post-flexion larva to juvenile generally occurs at about 22 mm,  
130 about four weeks after hatching, after which swimming capabilities increase rapidly,  
131 including schooling behavior during daylight hours (Fukuda *et al.*, 2014). Feeding studies of  
132 the larvae have focused mainly on individuals  $<10$  mm, after which many switch from

133 zooplankton to a piscivorous, often cannibalistic, diet and eventually become too difficult or  
134 too rare to capture in plankton nets towed at moderate speeds.

135 BFT larvae live in near-surface waters (generally, the upper 20 m) and feed by visual cues  
136 during daylight hours (Davis *et al.* 1990; Satoh, 2010). Typical prey are epipelagic  
137 zooplankton of 1 mm size or less. While the smallest larvae are known to prey mostly on  
138 copepod nauplii, the dietary contributions of different copepod taxa to such prey are  
139 unresolved in stomach content analyses. For copepodid and adult stages, however, clear  
140 differences emerge among different taxonomic groups (Fig. 2). Among the more numerous  
141 and recognizable small copepods in most ocean ecosystems, *Oithona* spp. (Cyclopoida) are  
142 rarely reported in significant numbers in larval BFT stomachs. Small calanoid taxa (e.g.,  
143 *Paracalanus* and *Clausocalanus* spp.) are consumed, but in dietary studies of all BFT species  
144 they appear in significantly lower proportions than their contributions to the ambient prey  
145 community (Uotani *et al.*, 1981; Young and Davis, 1990; Llopiz *et al.*, 2015; Kodama *et al.*,  
146 2017). In contrast, poecilostomatoid copepods (typically *Corycaeus* or *Farranula* spp.),  
147 reported as cyclopoid copepods in earlier studies, appear to be strongly selected. This is clear  
148 in studies of SBFT that have expressly compared larval diets to the natural abundances of  
149 prey (Uotani *et al.*, 1981; Young and Davis 1990), as well as suggested by dietary results for  
150 ABFT and PBFT where the natural abundances of prey were not directly assessed (Uotani *et*  
151 *al.*, 1990; Tilley *et al.*, 2016). Kodama *et al.*'s (2017) atypical dietary results for PBFT in the  
152 Sea of Japan were during a time of insignificant natural abundances of *Corycaeus* and  
153 *Farranula* spp. as assessed by both microscopical and metagenomics analyses. Tilley *et al.*'s  
154 (2016) results for the northern Gulf of Mexico are confounded by unusual feeding on  
155 abundant barnacle (*Lepas* sp.) nauplii, which may have arisen from proximity to deep-water  
156 oil drilling platforms. We do not present those data in Figure 2 but note that  
157 poecilostomatoid copepods (29%) and cladocerans (24%) comprised the majority of the  
158 holozooplankton prey consumed by ABFT in that study (Tilley *et al.*, 2016).

159 Figure 2 shows that small marine cladocerans also contribute disproportionately to the  
160 diets of BFT larvae with evidence from each of the three species (Uotani *et al.*, 1981; Llopiz  
161 *et al.*, 2015; Kodama *et al.*, 2017). In addition, Catalán *et al.* (2011) documented high  
162 feeding incidence on cladoceran prey (33.7% of stomach items) and high selectivity relative  
163 to either copepod nauplii or copepodids by ABFT larvae in the Mediterranean Sea, which  
164 further extends the observations of this feeding preference to both eastern and western ABFT  
165 spawning regions. We did not include data from Catalán *et al.* (2011) in Figure 2 because  
166 they did not distinguish copepod orders – calanoida, cyclopoida, poecilostomatoid – in their  
167 analyses of prey items and natural abundances.

168 While there is some evidence that appendicularians can be important prey items for BFT  
169 larvae (e.g., Llopiz *et al.*, 2015; Gulf of Mexico), that is not a common finding from dietary  
170 analyses (Fig. 2). Catalán *et al.* (2011) commented about the low incidence of  
171 appendicularians in ABFT larvae from the Mediterranean Sea, and Tilley *et al.* (2016) found  
172 none in the diets of ABFT in the northern Gulf of Mexico. Kodama *et al.* (2017), however,  
173 did report significant feeding on *Oikopleura* spp. from metagenomic analyses of PBFT larval  
174 guts compared to no evidence from microscopical analysis, which raises the possibility that  
175 soft-bodied appendicularians are systematically underrepresented in gut content enumeration  
176 due to their more rapid digestion relative to the recognizable chitinous exoskeletons of  
177 crustacean prey. Nonetheless, it is notable that the larvae of other scombrid species, like  
178 *Auxis* spp. (bullet tuna), *Euthymmus alletteratus* (little tunny) and *Katsuwonus*  
179 *pelamis* (skipjack), are well documented to specialize on appendicularian prey, with  
180 numerical abundances exceeding 50%, and often 90%, of gut contents, often from the same  
181 studies, collection regions and analysts that found little or no feeding on appendicularians by



182 BFT larvae (Uotani *et al.*, 1981; Young and Davis, 1990; Llopiz *et al.*, 2010; Llopiz and  
183 Hobday, 2015). Thus, analytical bias does not easily explain the relatively low incidence of  
184 larval BFT feeding on appendicularians. In this regard, BFT larvae share stronger dietary  
185 characteristics with other *Thunnus* species (*T. alalunga* – albacore; *T. albacares* – yellowfin;  
186 *T. obesus* – bigeye), as well as the larvae of billfishes (*Makaira nigricans* – blue marlin;  
187 *Xiphias gladius* – swordfish; *Tetrapturus angustirostris* – shortbill spearfish; *Istiophorus*  
188 *platypterus* – sailfish), which all show very high feeding incidence, ranging from 33 to >90%,  
189 on the combination of *Corycaeus*, *Farranula* and cladoceran species (Uotani *et al.*, 1981;  
190 Young and Davis, 1990; Uotani and Ueyanagi, 1997; Govoni *et al.*, 2003; Llopiz and Cowen,  
191 2008; Llopiz and Hobday, 2015).

192 The specific characteristics of poecilostomatoid copepods and cladocerans that cause  
193 them to be preferred prey of *Thunnus* and billfish larvae are not known. Kodama *et al.*  
194 (2017) speculated that apparent prey selection could be driven by the superior escape abilities  
195 of copepods, leaving slower-swimming and less-evasive prey like cladocerans and  
196 appendicularians to be overrepresented in larval diets. Since poecilostomatoid copepods  
197 were largely absent from the water during the Kodama *et al.* (2017) study, however, a  
198 broader explanation of the selectivity results would need to consider differential  
199 vulnerabilities of the major copepod orders, the fact that appendicularians, with poor escape  
200 capabilities, are not typically dominant prey of *Thunnus* and billfish larvae, and inherent  
201 differences among species in the prey characteristics that trigger recognition and attack.  
202 Considering the characteristics of the preferred BFT prey, both cladocerans and  
203 poecilostomatoid copepods lack development of the appendages that generate the smooth  
204 swimming currents of calanoid copepods and swim with jerky strokes. Another common  
205 denominator is the presence of large and conspicuous eyes, with the paired compound eyes  
206 and cuticular lenses of poecilostomatoid copepods (Vaissière, 1961) potentially also  
207 enhancing their visibility by reflecting or refracting light. These could be key factors that set  
208 cladocerans and poecilostomatoid copepods apart from other zooplankton as prey of *Thunnus*  
209 and billfish larvae, whereas other scombrid larvae with similar size, vision and swimming  
210 capabilities clearly respond more strongly to the characteristics of appendicularians.

## 211 **Resource limitation of larval BFT**

212 Regardless of the mechanism(s) underlying prey capture success and dietary selectivity of  
213 BFT larvae, certain prey types clearly play disproportionate roles as food resources relative to  
214 their abundances in the plankton. Consequently, the quality of the prey environment for  
215 larvae cannot be assessed solely in terms of total zooplankton abundance or biomass, but  
216 depends on size structure and composition (Robert *et al.*, 2013). This would be of little  
217 consequence if larvae were always able to achieve optimal or equivalent rates of feeding,  
218 growth or survival success despite natural variations in prey. There is ample evidence,  
219 however, that food limitation can be a significant factor in the early life history of BFT.

220 For BFT larvae of a comparable size range (Table 1), the highest estimates of daytime  
221 larval feeding indices (>90% with prey and >7 prey per gut; Table 1) are for ABFT  
222 populations in the Gulf of Mexico (Llopiz *et al.*, 2015) and northwest Mediterranean (Catalán  
223 *et al.*, 2011). Llopiz *et al.* (2015) further reported an even higher 100% feeding incidence for  
224 >4 mm larvae and a 71% incidence of piscivory for larger 8-10 mm larvae, which laboratory  
225 and model studies have associated with improved growth and survival (Reglero *et al.*, 2011,  
226 2014). Catalán *et al.* (2011) also found that larger (>5 mm) larvae consumed more prey, 11.1  
227 prey/larvae on average (100% feeding incidence), but piscivory was not observed. Compared  
228 to these high estimates of larval feeding, the more typical results for SBFT (Uotani *et al.*,  
229 1981; Young and Davis, 1990), PBFT (Kodama *et al.*, 2017) and even occasionally ABFT

230 (Tilley *et al.*, 2016) show reduced feeding incidence of 53-78% and much lower numbers of  
231 prey (1.1-1.3 individuals) in stomach contents (Table 1). The wide range in these values is  
232 indicative of suboptimal feeding conditions at least some of the time in BFT spawning  
233 grounds, which is consistent with bioenergetic-foraging models that suggest food limitation  
234 on a planktivorous diet even when all zooplankton are assumed to be equally available as  
235 prey (Reglero *et al.*, 2011).

236 Young and Davis (1990) were the first to address food limitation as an ecological issue  
237 for BFT larvae, finding significant positive relationships between feeding indices and  
238 zooplankton biomass for SBFT larvae in the eastern Indian Ocean spawning region. For that  
239 study, stomach fullness doubled and % larval stomachs with prey increased 3-4 fold, from  
240 20-30% to ~90%, over a 3-fold natural variation in zooplankton biomass. A mean daily  
241 ration of 30% of body mass  $d^{-1}$  was determined from the prey ingested by 4.5 mm larvae and  
242 gut evacuation rates of 3-4 h, suggesting a relatively modest average rate of larval growth,  
243 though this is likely to be an underestimate given recent evidence of significantly faster  
244 digestion times (2 hours, P. Reglero, pers. com.). For larvae from the same study area,  
245 Jenkins *et al.* (1990) found that otolith estimates of growth rate declined by 25% on average  
246 in higher density patches of larvae, indicating the possibility for density-dependent depletion  
247 of prey resources. Growth rate estimates were positively correlated with feeding indices  
248 (feeding incidence, mean stomach fullness and prey number), but not with temperature,  
249 which varied narrowly during that study.

250 For PBFT larvae, Tanaka *et al.* (2008) found 40-90% mortality of hatchery-raised larvae  
251 after one day of non-feeding and virtually zero tolerance of 2 days without feeding. Up to  
252 26% of field-collected larvae had low RNA:DNA ratios comparable to larvae starved for 24  
253 h, with the proportion of starving larvae varying by a factor of 6 over a 4-fold range in  
254 copepod prey density. The effects of starvation or food-limited growth of larval survival  
255 were further tested by analyzing the otoliths of specimens from PBFT larval patches followed  
256 by satellite-tracked drifters in the Nansei spawning region (Satoh *et al.*, 2013). As  
257 hypothesized, the larvae that survived over several days of development were subsets of the  
258 original populations with wider otolith radii, higher daily growth rates and larger size at a  
259 given age. Such results help to explain the somewhat contradictory conclusions that can be  
260 made between demonstrable food-limitation impacts in BFT spawning regions and relatively  
261 rapid growth rates of the surviving larvae (Jenkins and Davis, 1990; Shimose and Farley,  
262 2016; Malca *et al.*, 2015, 2017). Successful development through the early life stages of BFT  
263 thus implies early and continued success in finding and capturing appropriate prey.

## 264 **Plankton food webs supporting larval BFT**

265 The ecological factors that determine the quality of the food web for successful larval feeding  
266 and growth in oligotrophic waters are major unknowns in the early life history of BFT.  
267 Based on general understanding of plankton trophic relationships, however, one can envision  
268 various pathways that connect the production base of ocean surface waters to potential and  
269 selected prey of larval tuna. These are represented in Figure 3 as a starting point for  
270 discussing possibilities and specific areas of uncertainty.

271 The main diagonal arrows in Figure 3 depict a conventional view of the pelagic food web  
272 of subtropical oligotrophic waters as might be represented in a simple ocean ecosystem  
273 model. Photosynthetic bacteria and picoeukaryotes are the dominant primary producers, and  
274 protists are major primary consumers and the main trophic link to mesozooplankton (Azam *et al.*  
275 *et al.*, 1983; Sherr and Sherr, 1994; Calbet and Landry, 2004). A more detailed food web would  
276 recognize alternate pathways of trophic coupling, including multiple steps of protistan

277 consumers up to mesozooplankton (e.g., Calbet and Landry, 1999) and direct consumption of  
278 some primary production by mesozooplankton feeding on larger phytoplankton with  
279 strategies (e.g., mixotrophy, nitrogen fixation) adapted to low-nutrient systems. Based on  
280 stable isotope analyses of amino acids, however, the three-step food chain from  
281 phytoplankton to phagotrophic protists to copepods appears to capture effectively the mean  
282 nitrogen enrichment and general trophic structure for such systems (Landry and Décima,  
283 2017), so we use that simple chain here to represent those mean flows.

284 Figure 3 separately represents large cells associated with nitrogen fixation, an important  
285 source of new nitrogen (Mulholland *et al.*, 2006; Raes *et al.*, 2014) and possibly micro-  
286 habitat structure in oligotrophic surface waters of BFT spawning grounds. These include the  
287 cyanobacterium *Trichodesmium* spp. and large clumping diatoms (*Rhizosolenia*, *Hemiaulus*  
288 and *Mastogloia* spp.) with endo-symbiotic diazotrophs (Kemp and Villareal, 2013) that  
289 contribute to nitrogen fixation directly, as well as diatoms that benefit indirectly from the  
290 fixed nitrogen released by diazotrophs (Devassy *et al.*, 1979; Bergman *et al.*, 2012; Bonnet *et al.*,  
291 2016).

292 Enhanced thermal stratification associated with climate warming is predicted to reduce  
293 new nutrient delivery to surface waters, reducing general productivity and trophic transfer  
294 efficiency, though likely providing relative growth advantages to picophytoplankton and  
295 diazotrophs (Ware and Thompson, 2005; Hutchins *et al.*, 2007; Brown *et al.*, 2010; Moore *et al.*,  
296 2013; Litchman *et al.*, 2015; Fu *et al.*, 2016; Follett *et al.*, 2018). If such impacts are  
297 passed uniformly by the conventional food web to all components of the zooplankton  
298 community, they would exacerbate the already prey-limited resource environment for BFT  
299 larvae. If, on the other hand, climate warming disproportionately impacts, positively or  
300 negatively, the preferred prey of larvae, the effect on BFT populations could be either  
301 positive or negative. We thus consider below some aspects of the trophic ecologies of  
302 specific groups of zooplankton prey that set them apart from the community as a whole.

303 First, we should note that, as is the case for many or most ichthyoplankton (Montagnes *et al.*,  
304 2010), the direct consumption of protistan zooplankton is likely underestimated in dietary  
305 analyses of BFT larvae because preservation and measurement techniques are not optimized  
306 for soft bodied prey. Since the size range of consumed prey increases rapidly with larval  
307 development, this bias should be most relevant for the smallest larvae, and thus potentially  
308 important for establishing their success in early feeding. While there is little evidence to  
309 evaluate dietary flows through this pathway for natural BFT populations, Nakagawa *et al.*  
310 (2007) documented active feeding by PBFT larvae in aquaculture rearing tanks on the large  
311 dinoflagellates and ciliates associated with a microbial loop food chain based on bacterial  
312 production. Despite comparable or higher standing biomass estimates of heterotrophic  
313 microplankton compared to the primary prey (rotifers), however, the microplankton provided  
314 a small and declining proportion of the energy flow to larvae (from 13 to 0.6% for 4 to 12 day  
315 old larvae, respectively). In addition, the diversion of energy from the primary food chain  
316 (algae-rotifers) to heterotrophic protists raised questions as to whether they actually provided  
317 a net positive benefit to larval growth and survival (Nakagawa *et al.*, 2007). Such results  
318 point to the many unresolved complexities and nuances of energy flows through the  
319 microbial food webs of open-ocean systems, which we do not further consider here.

320 Appendicularians are a specialized component of the zooplankton with the capability of  
321 feeding directly on bacterial-sized particles (Scheinberg *et al.*, 2005; Sutherland *et al.* 2010).  
322 They can consequently short-circuit the protistan trophic steps in the food web and provide a  
323 more efficient transfer to appropriately sized prey for fish larvae than the indirect flow  
324 through the conventional food chain. As noted previously, however, appendicularians are  
325 generally not preferred prey of BFT larvae. Based on known dietary selectivity, therefore, a

326 shift to greater energy flux through appendicularians would more likely enhance the larval  
327 feeding conditions for co-occurring skipjack, little tunny and *Auxis* species over *Thunnus* and  
328 billfish species, unless there was some compensatory indirect feedback to the preferred prey  
329 of BFT larvae. As will be considered further below, for example, such feedback could occur  
330 if large marine snow particles from discarded appendicularian houses were utilized as a food  
331 resource or refuge that selectively benefitted a subset of prey.

332 Compared to the relatively straightforward relationships considered to this point, the  
333 trophic strategies of the prey types that BFT larvae select for most strongly --  
334 poecilostomatoid copepods and cladocerans -- are more challenging to understand in an  
335 oligotrophic food-web context. For poecilostomatoid copepods, a key issue is trophic  
336 position. One important group of poecilostomatoids, the Oncaeidae, is strongly associated  
337 with marine snow particles, especially discarded appendicularian houses, to which they  
338 adhere and scavenge with modified grasping antennae and maxillipeds (Alldredge 1972,  
339 1974; Steinberg *et al.*, 1994; Ohtsuka *et al.*, 1996; Green and Dagg, 1997). Recent research  
340 in the Sea of Japan has highlighted the possibility that appendicularians enhance transfer to  
341 higher trophic levels via the production of *Oncaea* and *Microsetella* spp., whose population  
342 abundances are significantly correlated with appendicularians (Kodama *et al.*, 2018). Since  
343 appendicularian houses typically contain high remnant concentrations of phytoplankton  
344 filtered from the water, *Oncaea* spp. could have trophic positions close to the food web base  
345 and function relatively efficiently in trophic transfer. However, the rarity of *Oncaea* spp. in  
346 BFT larval stomachs suggests that they are largely inaccessible as prey in surface waters,  
347 possibly because of the large-particle association, although their population distributions also  
348 tend to be deeper in the water column (e.g., McKinnon *et al.*, 2013). A similar observation  
349 can also be made about the low feeding incidence of BFT larvae on the harpacticoid  
350 copepods, *Macrosetella*, *Miracia* and *Microsetella* spp., associated with macroscopic bundles  
351 and tufts of *Trichodesmium* (Roman *et al.*, 1978; O'Neil and Roman, 1992). Thus, based on  
352 the relative absence of such prey in BFT larval diets, one can reasonably conclude that the  
353 larvae do not effectively exploit the concentrated prey assemblages associated with large  
354 particle aggregates.

355 While the other abundant poecilostomatoid group, the Corycaeidae, are occasionally  
356 found on marine snow particles (e.g., Green and Dagg, 1997), their grasping appendages  
357 appear to function mainly for carnivorous feeding. Based on trait similarities, Benedetti *et al.*  
358 (2016) classified all 14 species of *Corycaeus* and *Farranula* in the Mediterranean Sea  
359 according to their function as small active-ambush carnivores. In laboratory experiments,  
360 *Corycaeus* spp. have been shown to feed effectively on nauplii or copepodid prey of  
361 appropriate size (Turner *et al.*, 1984; Landry *et al.*, 1985), using visual cues with their image-  
362 forming binocular eyes, as is evident in much higher feeding rates in the light than dark  
363 (Gophen and Harris, 1981). In field studies, however, *Corycaeus* spp. have been linked more  
364 strongly to predation on fish larvae and chaetognaths (Brewer *et al.* 1984; Palomares García  
365 and Alejandre, 1995). Assuming similar relationships in the BFT spawning regions, their  
366 trophic positions would be TP = 4 if feeding directly on omnivorous copepods, or TP = 5 if  
367 feeding on other carnivores. Therefore, relative to comparably sized appendicularians,  
368 calanoid and cyclopoid copepods and particle-associated poecilostomatoids or harpacticoids,  
369 the trophic pathways to these favored prey of larval tuna and billfish are poorly resolved in  
370 plankton trophic studies, but could be unusually long and inefficient.

371 Of the eight described species of marine cladocerans, most are associated with coastal  
372 waters, but at least one -- *Pseudevadne tergestina* (also called *Evadne tergestina* in many  
373 studies) -- is broadly distributed in oligotrophic open-ocean BFT spawning regions like the  
374 eastern Indian Ocean and Gulf of Mexico (Della Croce and Venugopal, 1972; Della Croce



375 and Angelino, 1987), and thus might be considered a model type for such systems. In a  
376 scanning electron microscopy study of the stomach contents of 5 cladoceran species from the  
377 Sea of Japan, Kim *et al.* (1989) reported no substantial differences between prey ingested by  
378 *P. tergestina* relative to other species. All demonstrated strong preference for centric diatoms  
379 over pennates and dinoflagellates, which were also abundant in the ambient plankton, and  
380 none showed any indication of feeding on animal prey. Similarly, in a comparative  
381 experimental study of 3 cladoceran species by Katechakis and Stibor (2004), each species  
382 exhibited a selection maximum on natural food particles between 10 and 100  $\mu\text{m}$ , with little  
383 indication of an ability to capture bacterial-sized ( $<2\text{-}\mu\text{m}$ ) prey, as also noted by Paffenhöfer  
384 and Orcutt (1986). While *P. tergestina* was not included in the Katechakis and Stibor (2004)  
385 study, the morphologically similar *Evadne nordmanni* was suggested to be inefficient in  
386 capturing motile prey, notably ciliates and dinoflagellates, in the intermediate size range of  
387 15-70  $\mu\text{m}$ , which points further to the presumptive importance of relatively large and  
388 immobile prey like diatoms in the diets of marine cladocerans. Despite seeming to be  
389 paradoxically maladaptive for success under the oligotrophic conditions of BFT spawning  
390 grounds where small and motile cells are expected to predominate, this feeding relationship is  
391 borne out in the observation of *P. tergestina* replacing calanoid copepods (*Paracalanus* spp.)  
392 as the zooplankton community dominant at an artificial upwelling installation off Korea that  
393 increased phytoplankton abundance (mostly diatoms) by 50 fold (Jeong *et al.*, 2013). In  
394 contrast, Atienza *et al.* (2006) have suggested that another cladoceran species, *Penilia*  
395 *avirostris*, does better feeding on nanophytoplankton during stratified water periods in the  
396 coastal NW Mediterranean because its filter feeding appendages get clogged when diatoms  
397 are abundant. Consequently, there is little consensus as to how the cladoceran prey of BFT  
398 larvae in oligotrophic open-ocean ecosystems derive nutrition from the alternate pathways  
399 depicted in Figure 3.

400 Given contradictory interpretations of the trophic ecology of cladocerans and their  
401 contrasts to the presumptive long food chains to carnivorous poecilostomatoid copepods,  
402 current knowledge does not provide coherent answers for how the preferred prey of BFT  
403 larvae relate to one another or might respond, separately or together, to climate impacts on  
404 open-ocean ecosystems. Sahu *et al.* (2015), however, recently observed that cladocerans (*P.*  
405 *tergestina* in particular, but also *P. avirostris*) became community dominants and carnivorous  
406 copepods (poecilostomatoids and cyclopoids) notably more important while calanoids  
407 declined during *Trichodesmium* blooms in the coastal Bay of Bengal. Underlying  
408 relationships may therefore exist that link these seemingly disparate prey types to each other  
409 and to a specific group of phytoplankton (diazotrophic microplankton) with significant  
410 ecological advantages in future ocean scenarios (Breitbarth *et al.*, 2007; Hutchins *et al.*, 2007;  
411 Follett *et al.*, 2018).

412 How this might work mechanistically is a matter of speculation that may or may not  
413 include an intermediate step involving diatoms. While most zooplankton cannot feed on  
414 *Trichodesmium* directly, diatoms can be stimulated indirectly from the nitrogen released by  
415 *Trichodesmium* (Bergman *et al.*, 2012) or contribute separately to open-ocean new  
416 production as diatom-diazotroph symbiotic associations (Carpenter *et al.*, 1999; Dore *et al.*,  
417 2008). When either occurs, a rapid cladoceran response might be expected due to their high  
418 feeding efficiency on diatoms, prodigious reproductive potential and fast development times  
419 (Atienza *et al.*, 2008). In an early study of plankton succession in coastal waters of the  
420 Arabian Sea, Devassy *et al.* (1979) showed exactly this scenario, with a declining  
421 *Trichodesmium* bloom fueling a *Chaetoceros*-dominated mixed-diatom bloom and a massive  
422 population outbreak of *P. tergestina*. This does not match, however, the events described by  
423 Sahu *et al.* (2015), who sampled phytoplankton but did not indicate a shadow diatom bloom  
424 preceding the cladoceran response. Given the ephemeral boom-bust nature of cladoceran

425 dynamics (Devassy *et al.*, 1979; Sahu *et al.*, 2015), how their growth response advantages  
426 translate to the more steady-state conditions of oligotrophic waters is an important open  
427 question, as are details of linkages to diazotrophy or other ecological conditions, that might  
428 selectively favor poecilostomatoid copepods. Lastly, recent studies have nicely illustrated  
429 how modern analytical methods, like nanoSIMS (nanometer-scale, secondary ion mass  
430 spectrometry), can be used to unravel the transfers of diazotroph-derived nitrogen in open-  
431 ocean food webs, but they also demonstrate that interpretations can be time or space  
432 dependent. Bonnet *et al.* (2016), for example, provided strong support for significant transfer  
433 of *Trichodesmium*-fixed nitrogen to diatoms in the ultra-oligotrophic southwest Pacific. With  
434 similar methodology and in the same region, however, Caffin *et al.* (2018) reported that  
435 picoplankton (*Prochlorococcus*, *Synechococcus* and bacteria) were the main beneficiaries  
436 (>90%) of *Trichodesmium* nitrogen, which was then transferred inefficiently to zooplankton.  
437 Therefore, to understand the specific circumstances underlying favorable food webs for BFT  
438 larvae in spawning regions, there is no substitute for studies conducted in those areas at the  
439 appropriate times.

#### 440 **Conclusions and recommendations**

441 Whether BFT populations will be strongly impacted by or resilient to climate change is a  
442 complex matter that depends on many factors, including physical and ecological effects on  
443 larval, juvenile and adult habitats, the ocean currents that connect them, physiological,  
444 behavioral and spawning plasticity, and management strategies (Karnauskas *et al.*, 2015;  
445 Kimura *et al.*, 2010; Llopiz *et al.*, 2014; Muhling *et al.*, 2011, 2015, 2017). While larval  
446 studies alone cannot answer the climate change question, they are nonetheless essential for  
447 connecting projections and models of future ocean conditions to a particularly vulnerable  
448 phase of BFT life history. In this regard, three major points or conclusions can be drawn  
449 from the present review. The first is that BFT have characteristics that can make them  
450 excellent model organisms for investigating the linkages between plankton dynamics and  
451 economically valuable fisheries in open-ocean ecosystems. Unlike wide-ranging tropical  
452 tunas, their use of surface mixed-layer waters in restricted spawning regions during specific  
453 times of the year lends itself to defined and tractable research agendas for evaluating the  
454 ecological conditions associated with larval success. At the same time, the broad similarities  
455 in diet between larval BFT and other *Thunnus* and billfish species suggest that lessons  
456 learned from studies in BFT spawning regions may also be broadly applicable to the early life  
457 histories of other valuable top consumers throughout warm oligotrophic regions of the  
458 oceans.

459 The second point is that conventional understanding and approaches to studying the food  
460 webs of open-ocean ecosystems have not yet led, and likely cannot lead, to the kind of  
461 understanding that is needed about habitat qualities that relate most directly to successful  
462 larval feeding, growth and survival. Biological oceanographers with the interest and  
463 expertise to study the structure and flows of open-ocean food webs have typically had a  
464 lower-level, biomass-based (as opposed to taxon-based) focus with little attention to, or  
465 apparent awareness of, the unusual trophic pathways to the favored prey of fish larvae.  
466 Fisheries researchers have similarly refined techniques for recognizing the physical  
467 characteristics of waters where larvae are found and for measuring larval feeding, age and  
468 growth, but have not connected to the underlying plankton dynamics of those systems that  
469 make them more or less favorable for larvae. Both of these perspectives and areas of  
470 expertise must come together in collaborative studies that focus on the trophic pathways most  
471 relevant to fish larvae, the factors that drive their variability in spawning regions, and their  
472 effects on larval feeding, growth and survival.

473 Lastly, there are abundant and interesting scientific issues, both basic and applied, to  
474 drive research of this kind, as well as modern tools and approaches for doing so. Research  
475 that addresses the hypothesized linkage between *Trichodesmium*/diazotrophy and the specific  
476 selected prey of tuna larvae would explore end-to-end food-web relationships, from  
477 biogeochemistry to fish, in ways that are broadly advocated (Moloney *et al.*, 2011; Mitra *et*  
478 *al.*, 2014), yet seldom realized. Related questions, such as the roles and implications of  
479 habitat variability on food webs and larval growth-survival outcomes, also need to be  
480 investigated, as mesoscale ocean features are disproportionately important as spawning sites  
481 of adult BFT (Alemany *et al.*, 2010; Lindo-Atichati *et al.*, 2012; Laiz-Carrion *et al.*, 2015;  
482 Domingues *et al.*, 2016). These are questions that challenge the boundaries of current  
483 knowledge about open-ocean food-web structure, highlight the importance of zooplankton  
484 differences and diversity (size is not everything) in ocean ecosystem function (Steinberg and  
485 Landry, 2017), and have management implications for the possible resiliency of BFT early  
486 life history to projected climate impacts. For both larvae and prey, open issues of dietary  
487 selectivity and trophic position are amenable for measurement by modern molecular and  
488 compound-specific isotopic techniques (Troedsson *et al.*, 2009; O'Rorke *et al.*, 2012; Décima  
489 *et al.*, 2017), as are detailed assessments of trophic cycling and transfer by advanced isotope-  
490 based technologies (Bonnet *et al.*, 2016; Caffin *et al.*, 2018). The egg/embryo bearing traits  
491 of cladocerans, Corycaeidae and various other potential prey can also be usefully exploited as  
492 indices of their relative growth and production (Saunders *et al.*, 1999; Nielsen *et al.*, 2002),  
493 allowing spatial and temporal mapping of the production environments for relevant prey at  
494 the same scales as hydrographic, biogeochemical, primary production and larval BFT feeding  
495 and growth measurements. These are not research issues that can be resolved by synthesizing  
496 existing data into bigger and better models, but require field-focused efforts designed to  
497 provide new knowledge that bridges the gap between oceanographic and fisheries  
498 perspectives in ocean ecology. We believe that such integrative studies will yield many new  
499 insights into the functioning of open-ocean systems that will advance modeling, ecosystem-  
500 based fisheries management, and future ocean predictions.

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## 508 **References**

- 509 Alemany, F., Quintanilla, L., Vélez-Belchí, P., García, A., Cortés, D., Rodríguez, J. M., de  
510 Puellas, M. F., González-Pola, C., López-Jurado, J. L. 2010. Characterization of the  
511 spawning habitat of Atlantic bluefin tuna and related species in the Balearic Sea (western  
512 Mediterranean). *Progress in Oceanography*, 86: 21–38.
- 513 Alldredge, A. L. 1972. Abandoned larvacean houses: A unique food source in the pelagic  
514 environment. *Science*, 117: 885–887.
- 515 Alldredge, A. L. 1974. Discarded appendicularian houses as sources of food, surface habitats,  
516 and particulate organic matter in planktonic environments. *Limnology and*  
517 *Oceanography*, 21: 14–23.

- 518 Atienza, D., Saiz, E., and Calbet, A. 2006. Feeding ecology of the marine cladoceran *Penilia*  
519 *avirostris*: natural diet, prey selectivity and daily ration. Marine Ecology Progress Series,  
520 315: 211–220.
- 521 Atienza, D., Saiz, E., Skovgaard, A., Trepát, I., and Calbet, A. 2008. Life history and  
522 population dynamics of the marine cladoceran *Penilia avirostris* (Branchiopoda:  
523 Cladocera) in the Catalan Sea (NW Mediterranean). Journal of Plankton Research, 30:  
524 345–357.
- 525 Azam, F., Fenchel, T., Gray, J. G., Meyer-Reil, L. A., and Thingstad, F. 1983. The ecological  
526 role of water-column microbes in the sea. Marine Ecology Progress Series, 10: 257–263.
- 527 Bakun, A. 2013. Ocean eddies, predator pits and bluefin tuna: implications of an inferred  
528 ‘low risk–limited payoff’ reproductive scheme of a (former) archetypical top predator.  
529 Fish and Fisheries, 14: 424–438.
- 530 Bakun, A., and Broad, K. 2003. Environmental 'loopholes' and fish population dynamics:  
531 comparative pattern recognition with focus on El Niño effects in the Pacific. Fisheries  
532 Oceanography, 12: 458–473.
- 533 Behrenfeld, M. J., O'Malley, R. T., Siegel, D. A., McClain, C. R., Sarmiento, J. L., Feldman,  
534 G. C., Milligan, A. J., Falkowski, P. G., Letelier, R. M., and Boss, E. S. 2006. Climate-  
535 driven trends in contemporary ocean productivity. Nature 444, 752–755.
- 536 Benedetti, F., Gasparini, S., and Ayata, S.-D. 2016. Identifying copepod functional groups  
537 from species functional traits. Journal of Plankton Research, 38: 159–166.
- 538 Bergman, B., Sandh, G., Lin, S., John Larsson, J., and Carpenter, E. J. 2012. *Trichodesmium*  
539 – a widespread marine cyanobacterium with unusual nitrogen fixation properties. FEMS  
540 Microbiology Reviews, 37: 286–302.
- 541 Block, B. A., Dewar, H., Blackwell, S. B., Williams, T. D., Prince, E. D., Farwell, C. J.,  
542 Boustany, A., *et al.* 2001. Migratory movements, depth preferences, and thermal biology  
543 of Atlantic bluefin tuna. Science, 293: 1310–1314.
- 544 Block B. A., Teo, S. L. H., Walli, A., Boustany, A., Stokesbury, M. J. W., Farwell, C. J.,  
545 Weng, K. C., *et al.* 2005. Electronic tagging and population structure of Atlantic bluefin  
546 tuna. Nature, 434: 1121–1127.
- 547 Bonnet, S., Hugo Berthelot, H., Turk-Kubo, K., Cornet-Barthaux, V., Fawcett, S., Berman-  
548 Frank, I., Barani, A., Grégori, G., Dekaezemacker, J., Benavides, M., and Capone, D. G.  
549 2016. Diazotroph derived nitrogen supports diatom growth in the South West Pacific: A  
550 quantitative study using nanoSIMS. Limnology and Oceanography, 61: 1549–1562.
- 551 Bopp, L., Monfray, P., Aumont, O., Dufresne, J. -L., Le Treut, H., Madec, G., Terray, L., *et*  
552 *al.* 2001. Potential impact of climate change on marine export production. Global  
553 Biogeochemical Cycles, 15: 81–99.
- 554 Breitbarth, E., Oschlies, A., and LaRoche, J. 2007. Physiological constraints on the global  
555 distribution of *Trichodesmium* – effect of temperature on diazotrophy. Biogeosciences, 4:  
556 53–61.
- 557 Brewer, D. G., Kleppel, G. S., and Dempsey, M. 1984. Apparent predation on  
558 ichthyoplankton by zooplankton and fishes in nearshore waters of southern California.  
559 Marine Biology, 80: 17–28.
- 560 Brothers, E. B., Prince, E. D., Lee, D. W. 1983. Age and growth of young-of-the-year bluefin  
561 tuna, *Thunnus thynnus*, from otolith microstructure. In: Proceedings of the international  
562 workshop on age determination of oceanic pelagic fishes: tunas, billfishes and sharks.  
563 Prince, E. D., and Pulos, L. M. (eds). NOAA Technical Report NMFS No. 8: pp. 49–59.



- 564 Brown, C. J., Fulton, E. A., Hobday, A. J., Matear, R. J., Possingham, H. P., Bulman C.,  
565 Christensen, V., Forrest, R. E., Gehrke, P. C., Gribble, N. A., Griffiths, S. P., Lozano-  
566 Montes, H., Martin, J. M., Metcalf, S., Okey, T. A., Watson, R., and Richardson, A. J.  
567 2010. Effects of climate-driven primary production change on marine food webs:  
568 implications for fisheries and conservation. *Global Change Biology*, 16: 1194–1212
- 569 Caffin, M., Berthelot, H., Cornet-Barthaux, V., Barani, A., and Bonnet, S. 2018. Transfer of  
570 diazotroph-derived nitrogen to the planktonic food web across gradients of N<sub>2</sub> fixation  
571 activity and diversity in the Western Tropical South Pacific. *Biogeosciences*, 15: 3795-  
572 3810.
- 573 Calbet, A., and Landry, M. R. 1999. Mesozooplankton influences on the microbial food web:  
574 Direct and indirect trophic interactions in the oligotrophic open-ocean. *Limnology and*  
575 *Oceanography*, 44: 1370–1380.
- 576 Calbet, A., and Landry, M.R. 2004. Phytoplankton growth, microzooplankton grazing and  
577 carbon cycling in marine systems. *Limnology and Oceanography*, 49: 51–57.
- 578 Carpenter, E. J., Montoya, J. P., Burns, J., Mulholland, M. R., Subramaniam, A., and Capone,  
579 D. G. 1999. Extensive bloom of a N<sub>2</sub>-fixing diatom/cyanobacterial association in the  
580 tropical Atlantic Ocean. *Marine Ecology Progress Series*, 185: 273–283.
- 581 Catalán, I. A., Tejedor, A., Alemany, F., and Reglero, P. 2011. Trophic ecology of Atlantic  
582 bluefin tuna *Thunnus thynnus* larvae. *Journal of Fish Biology*, 78: 1545–1560.
- 583 Chust, G., Allen, J., Bopp, L., Schrum, C., Holt, J., Tsiaras, K., Zavatarelli, M., *et al.* 2014.  
584 Biomass changes and trophic amplification of plankton in a warmer ocean. *Global*  
585 *Change Biology*, 20: 2124–2139.
- 586 Constable, A. J., Melbourne-Thomas, J., Corney, S. T., Arrigo, K. R., Barbraud, C., Barnes,  
587 D. K. A., Bindoff, N. L., Boyd, P. W., *et al.* 2014. Climate change and Southern Ocean  
588 ecosystems I: how changes in physical habitats directly affect marine biota. *Global*  
589 *Change Biology*, 20: 3004-3025.
- 590 Davis, T. L. O., Jenkins, G. P., and Young, J. W. 1990. Diel patterns of vertical distribution  
591 in larvae of southern bluefin *Thunnus maccoyii*, and other tuna in the East Indian Ocean.  
592 *Marine Ecology Progress Series*, 59: 63–74.
- 593 Décima, M., Landry, M. R., Bradley, C. J., and Fogel, M. L. 2017. Alanine  $\delta^{15}\text{N}$  trophic  
594 fractionation in heterotrophic protists. *Limnology and Oceanography*, 62: 2308–2322.
- 595 Della Croce, N., and Angelino, M. 1987. Marine cladocera in the Gulf of Mexico and the  
596 Caribbean Sea. *Cahiers de Biologie Marine*, 28: 263–268.
- 597 Della Croce, N., and Venugopal, P. 1972. Distribution of marine cladocerans in the Indian  
598 Ocean. *Marine Biology*, 15: 132–138.
- 599 Devassy, V. P., Bhattathiri, P. M. A., and Qasim, S. Z. *et al.*, 1979. Succession of organisms  
600 following *Trichodesmium* phenomenon. *Indian Journal of Marine Science*, 8: 89–93.
- 601 Domingues, R., Goni, G., Bringas, F., Muhling, B., Lindo-Atichati, D., and Walter, J. 2016.  
602 Variability of preferred environmental conditions for Atlantic bluefin tuna (*Thunnus*  
603 *thynnus*) larvae in the Gulf of Mexico during 1993–2011. *Fisheries Oceanography*, 25:  
604 320–336.
- 605 Doney, S. C., Ruckelshaus, M., Duffy, J. E., Barry, J. P., Chan, F., English, C. A., Galindo,  
606 H. M., *et al.* 2012. Climate change impacts on marine ecosystems. *Annual Review of*  
607 *Marine Science*, 4: 11–37.

- 608 Dore, J. E., Letelier, R. M., Church, M. J., Lukas, R., and Karl, D. M. 2008. Summer  
609 phytoplankton blooms in the oligotrophic North Pacific Subtropical Gyre: Historical  
610 perspective and recent observations. *Progress in Oceanography*, 76: 2–38.
- 611 Druon, J. N., Fromentin, J. M., Hanke, A. R., Arrizabalaga, H., Damalas, D., Tičina, V.,  
612 Quílez-Badia, G., *et al.* 2016. Habitat suitability of the Atlantic bluefin tuna by size class:  
613 An ecological niche approach. *Progress in Oceanography*, 142: 30–46.
- 614 Ducklow, H. W., Fraser, W. R., Meredith, M. P., Stammerjohn, S. E., Doney, S. C.,  
615 Martinson, D. G., Saille, S. F., Schofield, O. M., Steinberg, D. K., Venables, H. J., and  
616 Amsler, C. D. 2013. West Antarctic Peninsula: An ice-dependent coastal marine  
617 ecosystem in transition. *Oceanography*, 26: 190–203.
- 618 Farley, J. H., Davis, T. L. O., Bravington, M. V., Andamari, R., and Davies, C. R. 2015.  
619 Spawning dynamics and size related trends in reproductive parameters of Southern  
620 Bluefin Tuna, *Thunnus maccoyii*. *PLoS ONE*, 10: e0125744.
- 621 Fromentin, J. M., Bonhommeau, S., Arrizabalaga, H., and Kell, L. T. 2014. The spectre of  
622 uncertainty in management of exploited fish stocks: The illustrative case of Atlantic  
623 bluefin tuna. *Marine Policy*, 47: 8–14.
- 624 Fu, W., Randerson, J. T., and J. K. Moore. 2016. Climate change impacts on net primary  
625 production (NPP) and export production (EP) regulated by increasing stratification and  
626 phytoplankton community structure in the CMIP5 models. *Biogeosciences*, 13: 5151–  
627 5170.
- 628 Fujioka, K., Masujima, M., Boustany, A. M., and Kitagawa, T. 2016. Horizontal movements  
629 of Pacific bluefin tuna. Pages 101-122 in Kitagawa, T., and Kimura, S. (Eds): *Biology  
630 and Ecology of Bluefin Tuna*. CRC Press, Boca Raton, FL.
- 631 Fukuda, H., Torisawa, S., and Takagi, T. 2014. Ontogenetic changes in schooling behavior  
632 and visual sensitivity during larval and juvenile stages in Pacific bluefin tuna, *Thunnus  
633 orientalis*. *Bulletin of the Fisheries Research Agency*, 38: 135–139.
- 634 Follett, C. L., Dutkiewicz, S., Karl, D. M., Inomura, K., and Follows, M. J. 2018. Seasonal  
635 resource conditions favor a summertime increase in North Pacific diatom–diazotroph  
636 associations. *The ISME Journal*, 12: 1543–1557.
- 637 Gophen, M., and Harris, R. P. 1981. Visual predation by a marine cyclopoid copepod,  
638 *Corycaeus anglicus*. *Journal of the Marine Biological Association, United Kingdom*, 61:  
639 391–399.
- 640 Govoni, J. J., Laban, J. J., and Hare, J. A. 2003. The early life history of swordfish (*Xiphias  
641 gladius*) in the western Atlantic. *Fishery Bulletin*, 101: 778–789.
- 642 Graham, J. B., and Dickson K. A., 2001. Anatomical and physiological specializations for  
643 endothermy. In: Block, B. A., and Stevens, E. D. (Eds.): *Tuna - Physiology, Ecology, and  
644 Evolution*. *Fish Physiology*, 20: 121–165. Academic Press, San Diego.
- 645 Green, E. P., and Dagg, M. J. 1997. Mesozooplankton associations with medium to large  
646 marine snow aggregates in the northern Gulf of Mexico. *Journal of Plankton Research*,  
647 19: 435–447.
- 648 Habtes, S., Muller-Karger, F. E., Roffer, M. A., Lamkin, J. T., and Muhling, B. A. 2014. A  
649 comparison of sampling methods for larvae of medium and large epipelagic fish species  
650 during spring SEAMAP ichthyoplankton surveys in the Gulf of Mexico. *Limnology and  
651 Oceanography Methods*, 12: 86–101.
- 652 Hobday, A. J., Evans, K., Eveson, J. P., Farley, J. H., Hartog, J. R., Basson, M., Patterson, T.  
653 A. 2016. Distribution and migration – southern bluefin tuna (*Thunnus maccoyii*). Pages

- 654 189-210 in Kitagawa, T., and Kimura, S. (Eds): Biology and Ecology of Bluefin Tuna.  
655 CRC Press, Boca Raton, FL.
- 656 Holland, K. N., Brill, R. W., Chang, R. K. C., Sibert, J. R., and Fournier, D. A. 1992.  
657 Physiological and behavioural thermoregulation in bigeye tuna (*Thunnus obesus*). *Nature*,  
658 358: 410–412.
- 659 Hutchins, D. A., Fu, F. -X., Zhang, Y., Warner, M. E., Feng, Y., Portune, K., Bernhardt, P.  
660 W., Mulholland, M. R. 2007. CO<sub>2</sub> control of *Trichodesmium* N<sub>2</sub> fixation, photosynthesis,  
661 growth rates, and elemental ratios: Implications for past, present, and future ocean  
662 biogeochemistry. *Limnology and Oceanography*, 52: 1293–1304.
- 663 Itoh, T. 2006. Sizes of adult bluefin tuna *Thunnus orientalis* in different areas of the western  
664 Pacific Ocean. *Fisheries Science*, 72: 53–62.
- 665 Jenkins, G. P., and Davis, T. L. O. 1990. Age, growth rate, and growth trajectory determined  
666 from otolith microstructure of southern bluefin tuna *Thunnus maccoyii* larvae. *Marine*  
667 *Ecology Progress Series*, 63: 93–104.
- 668 Jenkins, G. P., Young, J. W., and Davis, T. L. O. 1991. Density dependence of larval growth  
669 of a marine fish, the southern bluefin tuna, *Thunnus maccoyii*. *Canadian Journal of*  
670 *Fisheries and Aquatic Science*, 48: 1358–1363.
- 671 Jeong, Y. K., Lee, H. N., Park, C.-I., Kim, D. S., and Kim M. C. 2013. Variation of  
672 phytoplankton and zooplankton communities in a sea area, with the building of an  
673 artificial upwelling structure. *Animal Cells and Systems*, 17: 63–72.
- 674 Karnauskas, M., Schirripa, M. J., Craig, J. K., Cook, G. S., Kelbe, C. R., Agar, J. J., Black, B.  
675 A., Enfield, D. B., Lindo-Atichati, D., Muhling, B. A., Purcell, K. M., Richards, P. M.,  
676 and Wang, C. 2015. Evidence of climate driven ecosystem reorganization in the Gulf of  
677 Mexico. *Global Change Biology*, 21: 2554–2568.
- 678 Katechakis, A., and Stibor, H. 2004. Feeding selectivities of the marine cladocerans *Penilia*  
679 *avirostris*, *Podon intermedius* and *Evadne nordmanni*. *Marine Biology*, 145: 529–539.
- 680 Kemp, A.E.S., Villareal, T.A. 2013. High diatom production and export in stratified waters -  
681 a potential negative feedback to global warming. *Progress in Oceanography*, 119: 4–23.
- 682 Kim, S. W., Onbé, T., and Yoon, Y. H. 1989. Feeding habits of marine cladocerans in the  
683 Inland Sea of Japan. *Marine Biology*, 100: 313–318.
- 684 Kimura, S., Kato, Y., Kitagawa, T., and Yamaoka, N. 2010. Impacts of environmental  
685 variability and global warming scenario on Pacific bluefin tuna (*Thunnus orientalis*)  
686 spawning grounds and recruitment habitat. *Progress in Oceanography*, 86: 39–44.
- 687 Kitagawa, T., Kato, Y., Miller, M. J., Sasai, Y., Sasaki, H., and Kimura, S. 2010. The  
688 restricted spawning area and season of Pacific bluefin tuna facilitate use of nursery areas:  
689 A modeling approach to larval and juvenile dispersal processes. *Journal of Experimental*  
690 *Marine Biology and Ecology*, 393: 23–31.
- 691 Kodama, T., Hirai, J., Tamura, S., Takahashi, T., Tanaka, Y., Ishihara, T., Tawa, A., *et al.*  
692 2017. Diet composition and feeding habits of larval Pacific bluefin tuna, *Thunnus*  
693 *orientalis*, in the Sea of Japan: Integrated morphological and metagenetic analysis.  
694 *Marine Ecology Progress Series*, 583: 211–226.
- 695 Kodama, T., Iguchi, N., Tomita, M., Morimoto, H., Ota, T., and Ohshimo, S. 2018.  
696 Appendicularians in the southwestern Sea of Japan during the summer: abundance and  
697 role as secondary producers. *Journal of Plankton Research*, 40:269–283.

- 698 Laidre, K. L., Stirling, I., Lowry, L. F., Wiig, Ø., Heide-Jørgensen, M. P., and Ferguson, S.  
699 H. 2008. Quantifying the sensitivity of arctic marine mammals to climate-induced habitat  
700 change. *Ecological Applications*, 18: S97-S125.
- 701 Laiz-Carrión, R., Gerard, T., Uriarte, A., Malca, E., Quintanilla, J. M. Muhling, B. A.,  
702 Alemany, F., Privoznik, S., Shiroza, A., Lamkin, J. T., and García, A. 2015. Trophic  
703 ecology of Atlantic bluefin tuna (*Thunnus thynnus*) larvae from the Gulf of Mexico and  
704 NW Mediterranean spawning grounds: a comparative stable isotope study. *PLoS ONE*,  
705 10: 1-22. DOI:10.1371/journal.pone.0133406.
- 706 Landry, M. R., and Décima, M. R. 2017. Protistan microzooplankton and the trophic position  
707 of tuna: quantifying the trophic link between micro- and mesozooplankton in marine  
708 foodwebs. *ICES Journal of Marine Science*, 74: 1885–1892.
- 709 Landry, M. R., Lehner-Fournier, J. M., and Fagerness, V. L. 1985. Predatory feeding  
710 behavior of the marine cyclopoid copepod *Corycaeus anglicus*. *Marine Biology*, 85:  
711 163–169.
- 712 Lindo-Atichati, D., Bringas, F., Goni, G., Muhling, B., Muller-Karger, F.E., and Habtes, S.  
713 2012. Varying mesoscale structures influence larval fish distribution in the northern Gulf  
714 of Mexico. *Marine Ecology Progress Series*, 463: 245-257.
- 715 Litchman, E., de Tezanos Pinto, P., Edwards, K. F., Klausmeier, C. A., Kremer, C. T., and  
716 Thomas, M. K. 2015. Global biogeochemical impacts of phytoplankton: a trait-based  
717 perspective. *Journal of Ecology*, 103: 1384–1396.
- 718 Llopiz, J. K., and Cowen, R. K. 2008. Precocious, selective and successful feeding of larval  
719 billfishes in the oceanic Straits of Florida. *Marine Ecology Progress Series*, 358: 231–  
720 244.
- 721 Llopiz, J. K., Cowen, R. K., Hauff, M. J., Ji, R., Munday, P. L., Muhling, B. A., Peck, M. A.,  
722 *et al.* 2014. Early life history and fisheries oceanography: New questions in a changing  
723 world. *Oceanography*, 27: 26–41.
- 724 Llopiz, J. K., and Hobday, A. J. 2015. A global comparative analysis of the feeding dynamics  
725 and environmental conditions of larval tunas, mackerels, and billfishes. *Deep-Sea*  
726 *Research II*, 113: 113–124.
- 727 Llopiz, J. K., Muhling, B. A., and Lamkin, J. T. 2015. Feeding dynamics of Atlantic bluefin  
728 tuna (*Thunnus thynnus*) larvae in the Gulf of Mexico. *Collective Volume of Scientific*  
729 *Papers, International Commission for the Conservation of Atlantic Tunas*, 71: 1710–1715.
- 730 Llopiz, J. K., Richardson, D. E., Shiroza, A., Smith, S. L., and Cowen, R. K. 2010.  
731 Distinctions in the diets and distributions of larval tunas and the important role of  
732 appendicularians. *Limnology and Oceanography*, 55: 983–996.
- 733 Loeng, H., and Drinkwater, K. 2007. An overview of the ecosystems of the Barents and  
734 Norwegian Seas and their response to climate variability. *Deep-Sea Research II*, 54: 23-  
735 26.
- 736 Malca, E., Muhling, B., Franks, J., García, A., Tilley, J., Gerard, T., Ingram Jr., W., and  
737 Lamkin, J. T. 2017. The first larval age and growth curve for bluefin tuna (*Thunnus*  
738 *thynnus*) from the Gulf of Mexico: Comparisons to the Straits of Florida, and the Balearic  
739 Sea (Mediterranean). *Fisheries Research*, 190: 24–33.
- 740 Malca, E., Muhling, B. A., Lamkin, J. T., Ingram Jr., G. W., Gerard, T., Tilley, J., and  
741 Franks, J. 2015. Age and growth of larval Atlantic bluefin tuna, *Thunnus thynnus*, from  
742 the Gulf of Mexico. *Collective Volume of Scientific Papers, International Commission*  
743 *for the Conservation of Atlantic Tunas*, 71:1728–1735.



- 744 Matsuura, H., Sugimoto, T., Nakai, M., and Tsuji, S. 1997. Oceanographic conditions near  
745 the spawning ground of Southern Bluefin Tuna, north-eastern Indian Ocean. *Journal of*  
746 *Oceanography*, 53: 421–433
- 747 McKinnon, A. D., Duggan, S., Bötter-Schnack, R., Gusmão, L. F. M., and O’Leary, R. A.  
748 2013. Depth structuring of pelagic copepod biodiversity in waters adjacent to an Eastern  
749 Indian Ocean coral reef. *Journal of Natural History*, 47: 639–665.
- 750 Mitra, A., Castellani, C., Gentleman, W. C., Jónasdóttir, S. H., Flynn, K. J., Bode, A.,  
751 Halsband, C., *et al.* 2014. Bridging the gap between marine biogeochemical and fisheries  
752 sciences; configuring the zooplankton link. *Progress in Oceanography*, 129: 176–199.
- 753 Moloney, C. L., St. John, M. A., Denman, K. L., Karl, D. M., Köster, F. W., Sundby, S., and  
754 Wilson, R. P. 2011. Weaving marine food webs from end to end under global change.  
755 *Journal of Marine Systems*, 84: 106–116.
- 756 Montagnes, D. J. S., Dower, J. F., and Figueiredo, G. M. 2010. The protozooplankton–  
757 ichthyoplankton trophic link: An overlooked aspect of aquatic food webs. *Journal of*  
758 *Eukaryotic Microbiology*, 57: 223–228.
- 759 Moore, J. K., Lindsay, K., Doney, S. C., Long, M. C., and Misumi, K. 2013. Marine  
760 ecosystem dynamics and biogeochemical cycling in the Community Earth System Model  
761 [CESM1(BGC)]: Comparison of the 1990s with the 2090s under the RCP4.5 and RCP8.5  
762 scenarios. *Journal of Climate*, 26: 9291–9312.
- 763 Muhling, B. A., Lamkin, J. T., Alemany, F., García, A., Farley, J., Ingram, G.W., Berastegui,  
764 D.A., *et al.* 2017. Reproduction and larval biology in tunas, and the importance of  
765 restricted area spawning grounds. *Reviews in Fish Biology and Fisheries*, 27: 697–732.
- 766 Muhling, B. A., Lee, S. –K., Lamkin, J. T., and Liu, Y. 2011. Predicting the effects of climate  
767 change on bluefin tuna (*Thunnus thynnus*) spawning habitat in the Gulf of Mexico. *ICES*  
768 *Journal of Marine Science*, 68: 1051–1062.
- 769 Muhling, B. A., Liu, Y., Lee, S. –K., Lamkin, J. T., Roffer, M. A., and Muller-Karger, F. E.  
770 2015. Potential impact of climate change on the Intra-Americas Seas: Part 2: Implications  
771 for Atlantic bluefin tuna and skipjack tuna adult and larval habitats. *Journal of Marine*  
772 *Systems*, 148: 1–13.
- 773 Muhling, B. A., Reglero, P., Ciannelli, L., Alvarez-Berastegui, D., Alemany, F., Lamkin, J.  
774 T., and Roffer, M. A. 2013. Comparison between environmental characteristics of larval  
775 bluefin tuna *Thunnus thynnus* habitat in the Gulf of Mexico and western Mediterranean  
776 Sea. *Marine Ecology Progress Series*, 486: 257–276.
- 777 Mulholland, M. R., Bernhardt, P. W., Heil, C. A., Bronk, D. A., and O’Neil, J. M. 2006.  
778 Nitrogen fixation and release of fixed nitrogen by *Trichodesmium* spp. in the Gulf of  
779 Mexico. *Limnology and Oceanography*, 51: 1762–1776.
- 780 Nakagawa Y., Eguchi, M., and Miyashita, S. 2007. Pacific bluefin tuna, *Thunnus orientalis*,  
781 larvae utilize energy and nutrients of microbial loop. *Aquaculture*. 267: 83–93.
- 782 Nielsen, T. G., Møller, E. F., Satapoomin, S., Ringuette, M., and Hopcroft, R. R. 2002. Egg  
783 hatching rate of the cyclopoid copepod *Oithona similis* in arctic and temperate waters.  
784 *Marine Ecology Progress Series*, 236: 301–306.
- 785 Ohtsuka, S., Böttger-Schnack, R., Okada, M., and Onbé, T. 1996. In situ feeding habits of  
786 *Oncaea* (Copepoda: Poecilostomatoida) from the upper 250 m of the central Red Sea,  
787 with special reference to consumption of appendicularian houses. *Bulletin of Plankton*  
788 *Society of Japan*, 43: 89–105.
- 789 O’Neil, J. M, and Roman, M. R. 1992. Grazers and associated organisms with  
790 *Trichodesmium*. *In*: Carpenter, E. J., Capone, D. G., and Rueter, J. G. (Eds): *Marine*

- 791 Pelagic Cyanobacteria: *Trichodesmium* and other Diazotrophs. NATO Advanced Science  
792 Institutes Series, Series C, Mathematical and Physical Sciences, 362: 61–73.
- 793 O’Rorke, R., Lavery, S., Chow, S., Takeyama, H., Tsai, P., Beckley, L. E., Thompson, P. A.,  
794 Waite, A. M., and Jeffs, A. G. 2012. Determining the diet of larvae of western rock  
795 lobster (*Panulirus cygnus*) using high-throughput DNA sequencing techniques. PLoS  
796 ONE 7: e42757. doi:10.1371/journal.pone.0042757.
- 797 Paffenhöfer, G. A., and Orcutt, J. D., 1986. Feeding, growth and food conversion of the  
798 marine cladoceran *Penilia avirostris*. Journal of Plankton Research, 8: 741–754.
- 799 Palomares García, R., and Alejandre, R. V. 1995. Predation upon larvae of the Pacific sardine  
800 *Sardinops sagax* by cyclopid copepods. Journal of Crustacean Biology, 15: 1996–2001.
- 801 Pons, M., Branch, T. A., Melnychuk, M. C., Jensen, O. P., Brodziak, J., Fromentin, J. M.,  
802 Harley, S. J., Haynie, A. C., Kell, L. T., Maunder, M. N., Parma, A. M., Restrepo, V. R.,  
803 Sharma, R., Ahrens, R., and Hilborn, R. 2017. Effects of biological, economic and  
804 management factors on tuna and billfish stock status. Fish and Fisheries, 18: 1–21.
- 805 Raes, E. J., Waite, A. M., McInnes, A. S., Olsen, H., Nguyen, H. M., Hardman-Mountford,  
806 N., and Thompson, P. A. 2014. Changes in latitude and dominant diazotrophic  
807 community alter N<sub>2</sub> fixation. Marine Ecology Progress Series, 516: 85–102.
- 808 Reglero, P., Tittensor, D. P., Alvarez-Berastegui, D., Aparicio-Gonzalez, A., and Worm, B.  
809 2014. Worldwide distributions of tuna larvae: revisiting hypotheses on environmental  
810 requirements for spawning habitats. Marine Ecology Progress Series, 501: 207–224
- 811 Reglero, P., Urtizberea, A., Torres, A. P., Alemany, F., and Fiksen, Ø. 2011. Cannibalism  
812 among size classes of larvae may be a substantial mortality component in tuna. Marine  
813 Ecology Progress Series, 433: 205–219.
- 814 Richardson, D. E., Marancik, K. E., Guyon, J. R., Lutcavage, M. E., Galuardi, B., Lam, C.  
815 H., Walsh, H. J., Wildes, S., Yates, D. A., and Hare, J. A. 2016. Discovery of a spawning  
816 ground reveals diverse migration strategies in Atlantic bluefin tuna (*Thunnus thynnus*).  
817 Proceedings of the National Academy of Sciences of the United States, 113: 3299–3304.
- 818 Robert, D., Murphy, H. M., Jenkins, G. P., Fortier, L. 2013. Poor taxonomical knowledge of  
819 larval fish prey preference is impeding our ability to assess the existence of a “critical  
820 period” driving year-class strength. ICES Journal of Marine Science, 71: 2042 – 2052
- 821 Roman, M. R. 1978. Ingestion of the blue-green alga *Trichodesmium* by the harpacticoid  
822 copepod, *Macrosetella gracilis*. Limnology and Oceanography, 23: 1245–1248.
- 823 Rooker, J. R., Alvarado Bremer, J. R., Block, B. A., de Metrio, G., Corriero, A., Krause, R.  
824 T., Prince, E. D., *et al.* 2007. Life history and stock structure of Atlantic bluefin tuna  
825 (*Thunnus thynnus*). Reviews in Fisheries Science, 15: 265–310.
- 826 Sahu, G., Mohanty, A. K., Smita Actary, M., Sarkar, S. K., and Satpathy, K. K. 2015.  
827 Changes in mesozooplankton community structure during *Trichodesmium erythraeum*  
828 bloom in the coastal waters of southwestern Bay of Bengal. Indian Journal of Geo-Marine  
829 Sciences, 44: 1292–1293.
- 830 Satoh, K. 2010. Horizontal and vertical distribution of larvae of Pacific bluefin tuna *Thunnus*  
831 *orientalis* in patches entrained in mesoscale eddies. Marine Ecology Progress Series, 404:  
832 227–240.
- 833 Satoh, K., Tanaka, Y., Masujima, M., Okazaki, M., Kato, Y., Shono, H., and Suzuki, K.  
834 2013. Relationship between the growth and survival of larval Pacific bluefin tuna,  
835 *Thunnus orientalis*. Marine Biology, 160: 691–702.

- 836 Saunders, P. A., Porter, K. G., and Taylor, B. E. 1999. Population dynamics of *Daphnia* spp.  
837 and implications for trophic interactions in a small, monomictic lake. *Journal of Plankton*  
838 *Research*, 21: 1823–1845.
- 839 Schaefer, K.M. 2001. Reproductive biology of tunas. *Fish Physiology*, 19: 225–270.
- 840 Scheinberg, R. D., Landry, M. R., and Calbet, A. 2005. Grazing impacts of two common  
841 appendicularians on the natural prey assemblage of a subtropical coastal ecosystem.  
842 *Marine Ecology Progress Series*, 294: 201–212.
- 843 Sherr, E. B., and Sherr, B. F. 1994. Bacterivory and herbivory: key roles of phagotrophic  
844 protists in pelagic food webs. *Microbial Ecology*, 28: 223–235.
- 845 Shimose, T., and Farley, J. H. 2016. Age, growth and reproductive biology of bluefin tunas.  
846 Pages 47-77 in Kitagawa, T., and Kimura, S. (Eds): *Biology and Ecology of Bluefin*  
847 *Tuna*. CRC Press, Boca Raton, FL.
- 848 Steinberg, D. K., Silver, M. W., Pilskaln, C. H., Coale, S. L., and Paduan, J. B. 1994.  
849 Midwater zooplankton communities on pelagic detritus (giant larvacean houses) in  
850 Monterey Bay, California. *Limnology and Oceanography*, 39: 1606–1620.
- 851 Steinberg, D. K., and Landry, M. R. 2017. Zooplankton and the ocean carbon cycle. *Annual*  
852 *Review of Marine Science*, 9:14.1–14.32, doi: 10.1146/annurev-marine-010814-015924.
- 853 Sutherland, K. R., Madin, L. P., and Stocker, R. 2010. Filtration of submicrometer particles  
854 by pelagic tunicates. *Proceedings of National Academy of Science*, 107: 15129–15134.
- 855 Tanaka, Y., Satoh, K., Yamada, H., Takebe, T., Nikaido, H., and Shiozawa, S. 2008.  
856 Assessment of the nutritional status of field-caught larval Pacific bluefin tuna by  
857 RNA/DNA ratio based on a starvation experiment of hatchery-reared fish. *Journal of*  
858 *Experimental Marine Biology and Ecology*, 354: 56–64.
- 859 Tanaka, Y., and Suzuki, N. 2016. Early life history. Pages 19-46 in Kitagawa, T., and  
860 Kimura, S. (eds). *Biology and ecology of bluefin tuna*. CRC Press Boca Raton FL.
- 861 Tilley, J. D., Butler, C. M., Suárez-Morales, E., Franks, J. S., Hoffmayer, E. R., Gibson, D.  
862 P., Comyns, B. H., *et al.* 2016. Feeding ecology of larval Atlantic bluefin tuna, *Thunnus*  
863 *thynnus*, from the central Gulf of Mexico. *Bulletin of Marine Science*, 92: 321–334.
- 864 Troedsson, C., Simonelli, P., Nägele, V., Nejstgaard, J. C., and Frischer, M. E. 2009.  
865 Quantification of copepod gut content by differential length amplification quantitative  
866 PCR (dla-qPCR). *Marine Biology*, 156: 253–259.
- 867 Turner, J. T., Tester, P. A., and Conley, W. J. 1984. Zooplankton feeding ecology: Predation  
868 by the marine cyclopoid copepod *Corycaeus amazonicus* F. Dahl upon natural prey.  
869 *Journal of Experimental Marine Biology and Ecology*, 84: 191–202.
- 870 Uotani, I., Matsuzaki, K., Makino, Y., Noda, K., Inamura, O., and Horikawa, M. 1981. Food  
871 habits of larval tunas and their related species in the area northwest of Australia. *Bulletin*  
872 *of the Japanese Society of Fisheries Science*, 47: 1165–1172.
- 873 Uotani, I., Saito, T., Hiranuma, K., and Nishikawa, Y. 1990. Feeding habit of Bluefin tuna  
874 *Thunnus thynnus* larvae in the western North Pacific Ocean. *Nippon Suisan Gakkaishi*,  
875 56: 713–717.
- 876 Uotani, I., and Ueyanagi, S. 1997. Feeding habits of Indo-Pacific blue marlin and shortbill  
877 spearfish larvae. *Journal of the School of Marine Science and Technology, Tokai*  
878 *University*, 43: 107–116.
- 879 Vaissière, R. 1961. Morphologie et histologie compares des yeux des crustacés copépodes.  
880 *Archives de Zoologie Expérimentale et Générale*, 100:1–125.

- 881 Ware, D. M., and Thomson, R. E. 2005. Bottom-up ecosystem trophic dynamics determine  
882 fish production in the Northeast Pacific. *Science*, 308: 1280–1284.
- 883 Wassmann, P., Duarte, C. M., Agusti, S., and Sejr, M. K. 2011. Footprints of climate change  
884 in the Arctic marine ecosystem. *Global Change Biology*, 17: 1235-1249.
- 885 Xu, Y., Chant, R., Gong, D., Castelao, R., Glenn, S., and Schofield, O. 2011. Seasonal  
886 variability of chlorophyll a in the Mid-Atlantic Bight. *Continental Shelf Research*, 31:  
887 1640–1650.
- 888 Young, J. W., and Davis, T. L. O. 1990. Feeding ecology of larvae of southern bluefin,  
889 albacore and skipjack tunas (Pisces: Scombridae) in the eastern Indian Ocean. *Marine*  
890 *Ecology Progress Series*, 61: 17–29.
- 891 Yúfera, M., Ortiz-Delgado, J. B., Hoffman, T., Sigüero, I., Urup, B., and Sarasquete, C. 2014.  
892 Organogenesis of digestive system, visual system and other structures in Atlantic bluefin  
893 tuna (*Thunnus thynnus*) larvae reared with copepods in mesocosm system. *Aquaculture*,  
894 426–427: 126–137.

## 895 **Figure Legends**

896 **Fig. 1.** Global map showing the known spawning areas for Atlantic, Pacific and Southern  
897 species of bluefin tuna superimposed on surface chlorophyll climatology from the MODIS  
898 Aqua satellite (2003-2016). Southern Hemisphere is the mean chlorophyll for the months  
899 of January-March, the peak spawning season of southern bluefin tuna. Northern  
900 Hemisphere is the mean surface chlorophyll for May-July, the spawning season of Atlantic  
901 and Pacific BFT.

902 **Fig. 2.** Summary of dietary studies of bluefin tuna larvae showing percentage composition of  
903 diet (% prey abundances) and significant trends in selection (positive = upward arrow;  
904 negative = downward arrow; relative to prey abundances in the ambient plankton  
905 community). SBT = southern; ABT = Atlantic; PBT = Pacific. Size (mm) is size range of  
906 larvae analyzed; n = number of larvae analyzed.

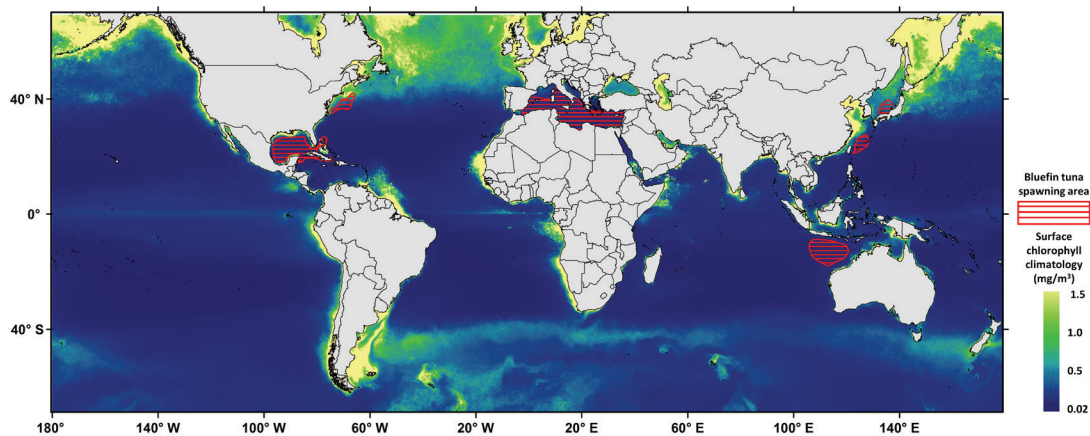
907 **Fig. 3.** Schematic diagram of food-web relationships supporting bluefin tuna larvae in open-  
908 ocean oligotrophic spawning regions plotted relative to body size and assumed trophic  
909 position. Arrows from picophytoplankton depict the conventional microbial food chain  
910 via phagotrophic protists to copepod-dominated mesozooplankton and the alternate route  
911 of direct picoplankton consumption by appendicularians. Arrows from comparably-sized  
912 zooplankton prey highlight the disproportionate importance of less-well-studied  
913 cladocerans and carnivorous poecilostomatoid copepods to the diets of BFT larvae  
914 compared to appendicularians, cyclopoid, calanoid and particle-associated  
915 poecilostomatoid copepods. Double-arrow dotted line represents the consortium of  
916 microphytoplankton taxa, *Trichodesmium* and diazotroph-diatom associations, that  
917 contribute directly to nitrogen fixation, as well as diatoms that utilize nitrogen released  
918 from *Trichodesmium*. Dashed lines with question marks indicate uncertainties in trophic  
919 relationships of the major selected prey of BFT larvae.

920



921 Table 1. Field estimates of % larvae feeding and number of prey per gut for larvae of  
 922 southern (SBFT), Atlantic (ABFT) and Pacific (PBFT) bluefin tuna. Med =  
 923 Mediterranean Sea; GoM = Gulf of Mexico. Size (mm) and *n* are the size range and  
 924 number of larvae analysed. All data, except for Uotani *et al.* (1981), are for larvae  
 925 collected during daylight hours only.

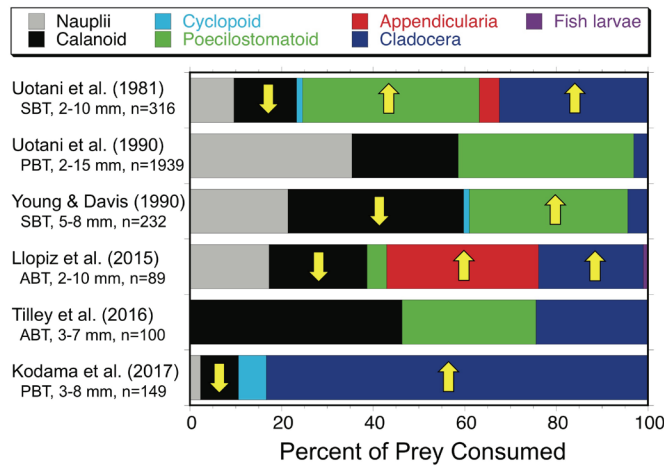
926	Stock	Size (mm)	<i>n</i>	% Feeding	Prey/gut	Reference
927	SBFT	2.2-9.6	316	78%	1.3	Uotani et al (1981)
928	SBFT	2.7-9.8	409	53%	1.2 ± 0.5	Young and Davis (1990)
929	ABFT-Med	2.6-8.7	107	94%	7.1 ± 5.7	Catalán et al. (2011)
930	ABFT-GoM	2.0-10	87	94%	7.9 ± 5.8	Llopiz et al. (2015)
931	ABFT-GoM	3.0-6.7	100	57%	1.1 ± 1.3	Tilley et al. (2016)
932	PBFT	2.7-7.6	96	64%	1.3 ± 1.7	Kodama et al. (2017)



933

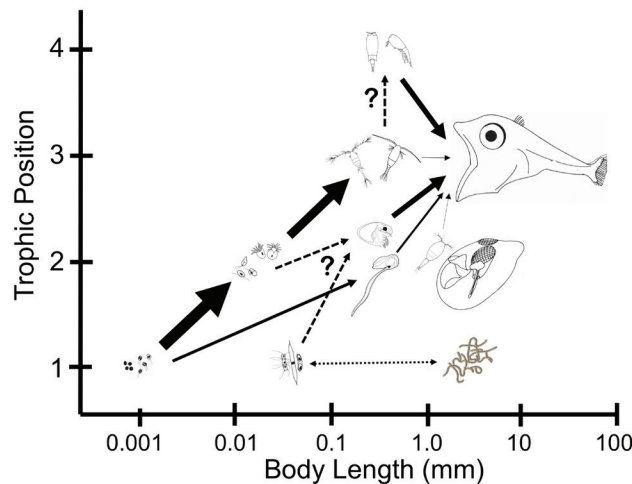
934 **Fig. 1.** Global map showing the known spawning areas for Atlantic, Pacific and Southern  
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941

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947

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