

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23

The productivity and viability of Snake River Sockeye Salmon hatchery adults released into
Redfish Lake, Idaho

Suggested Running Head: Re-introduction of Sockeye Salmon into Redfish Lake, ID.

Christine C Kozfkay¹, Mike Peterson², Benjamin P. Sandford³, Eric Johnson⁴, and Paul Kline¹

¹Headquarters, Idaho Department of Fish and Game
600 South Walnut Street, Boise ID 83707

²Southwest Region, Idaho Department of Fish and Game
3101 S. Powerline Road, Nampa, ID 83686, USA

³Northwest Fisheries Science Center,
National Marine Fisheries Service, National Oceanic and Atmospheric Administration
2725 Montlake Boulevard East Seattle, WA 98112, USA

⁴Eagle Fish Hatchery, Idaho Department of Fish and Game
1800 Trout Road, Eagle, ID 83616, USA

24 <A>Abstract

25 In 1991, the Snake River Sockeye Salmon Captive Broodstock Program was initiated to
26 prevent extinction and preserve genetic diversity of this evolutionarily significant unit protected
27 by the Endangered Species Act. At the time of listing, the Redfish Lake population was
28 considered functionally extinct. One of the recovery strategies entails the release of adults for
29 volitional spawning in Redfish Lake for re-building of the natural population. In this paper, we
30 describe the productivity metrics from this strategy. We evaluated eight spawn years to address
31 three primary questions: 1. “What egg-to-smolt, smolts per female, and smolt-to-adult (SAR)
32 metrics result from recent adult releases? 2. How do these metrics compare to estimates for
33 Redfish Lake historically and estimates for other sockeye populations throughout the range?”
34 and 3. “Does the current combination of smolts per female and SRS result in population
35 replacement?” Replacement was determined as two adult recruits per female assuming an even
36 sex ratio. We found that the reintroduced adults, despite being derived from a multi-generational
37 captive broodstock, were able to successfully spawn and produce offspring that migrated to the
38 ocean and returned as adults. Smolt abundance, size, and age data suggest that the population is
39 functioning below density dependence. However, increased smolt production did not translate
40 into greater adult returns and this is likely due to out-of-basin factors. Productivity metrics were
41 similar to those of the wild population in Redfish Lake during the 1950-60s. However, both
42 current and historic productivity estimates were near the low end of the range for other sockeye
43 populations and have not resulted in population replacement. Until freshwater and out-of-basin
44 survival can be improved, our data suggest that adult releases will continue to be an important
45 recovery strategy to prevent cohort collapse and to re-build naturally spawning populations.

46 <A> Introduction

47 Captive broodstock programs have been established in both Europe and North America
48 as a safety net to conserve genetic diversity and reduce the extinction probability of highly
49 imperiled fish species and/or populations (Carr et al. 2004; Cooper et al. 2009; Withler et al.
50 2011; Saltzgiver et al. 2012; Osborne et al. 2013; Withler et al. 2014). Captive broodstock
51 programs are distinct from other hatchery programs in that fish remain in a hatchery environment
52 throughout their entire life-cycle (Flagg and Mahnken 1995; Miller and Kapuscinski 2003;
53 Berejikian et al. 2004; Hebdon et al. 2004) and are genetically managed to avoid inbreeding
54 depression and unintended selection (Kozfkay et al. 2008; Sturm et al. 2009; Kalinowski et al.
55 2012; Conrad et al. 2013; Fisch et al. 2012; O'Reilly and Kozfkay 2014; Fisch et al. 2015). The
56 primary goal of a captive broodstock is to retain the extant population (and its genetic diversity)
57 in protective culture until the causes that threaten persistence can be alleviated; but adults or
58 juveniles are released in the wild if numbers are available beyond what is needed for the
59 replacement broodstock. While captive broodstocks are less common relative to other types of
60 hatchery programs, they may become more widespread with increasing environmental and
61 climatic threats to population persistence and can be an important means of rebuilding declining
62 or extirpated populations (Flagg and Mahnaken 1995; Waters et al. 2015).

63 For Snake River Sockeye Salmon *Oncorhynchus nerka*, a captive broodstock program
64 was initiated prior to its listing as endangered under the U.S. Endangered Species Act (NMFS
65 1991). At the time of listing, only one remnant population remained in Redfish Lake located at
66 the headwaters of the Salmon River drainage in the Sawtooth Valley basin, Idaho (Figure 1).
67 This population exists at the extreme of the worldwide distribution as the most southerly, farthest
68 inland and highest elevation spawning population and was on the brink of extinction, with one
69 adult returning in 1989 and zero adults returning in 1990 (Waples et al. 1991). The captive

70 broodstock program was created over an eight-year period and captured multiple age-classes,
71 life-stages, and life-histories in the collection of the founding broodstock including all of the
72 wild, anadromous adults that returned from 1991-1998, smolts that emigrated from Redfish Lake
73 from 1991-1993, and residual adults collected in Redfish Lake from 1992-1995 (Kalinowski et
74 al. 2012; Kline and Flagg 2014). The creation of the captive broodstock prevented the imminent
75 extinction of the population.

76 Sockeye Salmon display life-history diversity in age structure and residency that allowed
77 it to persist at critically low levels prior to hatchery intervention. Many different age
78 combinations of freshwater and saltwater residency are represented within a cohort.
79 Anadromous sockeye salmon typically spend one or two years in the lake before they undergo
80 smoltification and migrate to the ocean and then spend an additional one to four years in the
81 ocean before returning to freshwater to spawn (Burgner 1991). Redfish Lake is also unique in
82 that it is one of only two lakes in the Pacific Northwest where three life histories of native *O.*
83 *nerka* reside: anadromous, residual, and kokanee (Nichols et al. 2016). Residual Sockeye
84 Salmon are a resident *O. nerka* ecotype, considered part of the listed population (Waples et al.
85 1997), that are capable of reproducing with the anadromous ecotype and producing both residual
86 and anadromous offspring (Bjornn et al. 1968; Burgner 1991; Rieman et al. 1994; Godbout et al.
87 2011). While Bjornn suspected the presence of residuals in the 1950's and 1960's, residuals
88 were not physically documented in Redfish Lake until 1992. Managers began to actively search
89 for the smaller, resident adults during spawning after otolith microchemistry results indicated
90 that many of the smolts leaving Redfish Lake in 1991 had a resident, female parent (Rieman et
91 al. 1994, Waples et al. 1997). It has been hypothesized that residual Sockeye in Redfish Lake
92 prevented extirpation of the population while Sunbeam Dam was in operation from 1910-1934

93 (Waples et al. 1997). Both the anadromous and residual ecotypes spawn on beach shoals and
94 spawn in late October and November, whereas kokanee spawn in a tributary of the lake from
95 August through September. The resident population of kokanee is genetically divergent from the
96 residual and anadromous ecotypes due to these differences in spawn-timing and location and not
97 considered part of the listed population (Cummings et al. 1997, Waples et al. 2011).

98 At the outset of the captive broodstock program, there was uncertainty regarding Sockeye
99 Salmon survival in captivity and their productivity and contribution to recovery once they were
100 released in the natural environment (Flagg et al. 2004). Given the range of possible outcomes,
101 the release of captive-reared adults has been widely debated among fishery professionals (Fraser
102 2008; Araki and Schmid 2010). In some cases, hatchery adults have been unsuccessful at
103 spawning after release (Carr et al. 2004; Griffiths et al. 2011) or have been able to successfully
104 spawn but have had negative impacts on reproductive fitness of natural populations (Araki et al.
105 2007; Araki et al. 2008; Christie et al. 2014). In other cases, salmon reintroductions have
106 produced demographic increases (Berejikian et al. 2009; Hess et al. 2012; Withler et al. 2014).
107 The variability in outcomes can be complex and multi-faceted depending on species, available
108 habitat, geographic location, phenotypic and behavioral traits, and approach.

109 The focus of this study was to assess the productivity of adult Snake River Sockeye
110 Salmon released into Redfish Lake, Idaho. An earlier evaluation in the program measured the
111 effectiveness of adult releases and response in freshwater productivity, however, the contribution
112 of released adults could not be independently quantified due to the inability to differentiate
113 natural production from eyed-eggs that were placed into lake incubation boxes (Hebdon et al.
114 2004). We selected eight spawn years (2004–2011) in which captive-reared and anadromous
115 adults were released to spawn volitionally. These years were chosen for analysis because eyed-

116 egg releases were not implemented in Redfish Lake during this time-frame. Captive-reared
117 adults have been reared exclusively in the hatchery from egg to sexual maturity. Anadromous
118 fish are the offspring of predominantly captive-reared parents that had been released to the wild
119 as adults to volitionally spawn or hatchery-reared juveniles that were released into the wild as
120 smolts and have successfully undergone seaward migration, and returned to their natal spawning
121 grounds as adults. Anadromous adults were trapped and those not spawned for the captive
122 broodstock were released into Redfish Lake for natural spawning. Anadromous adults were
123 released with captive-reared adults as part of the recovery strategy to increase naturally-
124 spawning Sockeye Salmon abundance and re-establish a self-sustaining population in Redfish
125 Lake (NMFS 2015).

126 Our objective was to evaluate the contribution from adult releases by answering three
127 primary questions: 1. “What freshwater productivity (smolts per female and egg-to-smolt
128 survival) and post-juvenile productivity (smolt-to-adult [SARs]) rates result from adult releases?
129 2. How do these productivity metrics compare to historic data from the 1950’s and 1960’s and to
130 other sockeye populations throughout the range?” and 3. “Does the current combination of
131 smolts per female and SARs result in population replacement?” Replacement was determined as
132 two adult recruits per female assuming an even sex ratio. Information presented here will
133 provide baseline data to monitor population status changes through time as recolonization efforts
134 continue using this recovery strategy in Redfish Lake and other natal lakes in the Sawtooth
135 Valley basin. Evaluation of this release strategy is critical to our understanding of how hatchery
136 fish can contribute to rebuilding natural spawning populations to meet recovery objectives.

137 <A> STUDY SITE

138 Adult Sockeye Salmon were released in Redfish Lake, located in the Sawtooth Valley

139 basin of central Idaho (Figure 1). Redfish Lake is located 1,996 m above sea level and is 1,448
140 km from the Pacific Ocean. Redfish Lake is the largest historic Sockeye Salmon rearing lake
141 within the Sawtooth Valley basin with a surface area of 615 ha. Lakes in the Sawtooth Valley
142 basin are glacial-carved and considered ultra-oligotrophic, but high in oxygen (Budy et al. 1998).
143 Redfish Lake has a relatively pristine watershed, with virtually no development because it lies
144 within a National Recreation Area (NMFS 2015).

145 <A>METHODS

146 We report the number of adults released into Redfish Lake for volitional spawning and
147 the resulting productivity metrics (egg-to-smolt, smolts per female, SAR return rates) from this
148 release strategy. Estimates of age and abundance for the different juvenile life-stages (deposited
149 eggs in the gravel, smolts, returning adults) were required to calculate these productivity
150 estimates for each spawn year. Spawn year (SY) is defined as the calendar year in which adults
151 were released to volitionally spawn and the year in which their offspring were born. Below, we
152 describe the specific methods and calculations used to estimate potential egg deposition (PED)
153 from the released females, the number and age composition of smolts, and the number and age
154 composition of returning anadromous adults that resulted from natural spawning in Redfish
155 Lake.

156 *Adult Releases.*----

157 Captive-reared (2004 – 2007) and a mixture of captive-reared and anadromous adults
158 (2008 – 2011) were released during September to spawn volitionally in Redfish Lake. Captive-
159 reared fish were cultured in freshwater at the National Marine Fisheries Service Burley Creek
160 hatchery near Port Orchard, Washington (NMFS-FW) as well as at the Idaho Department of Fish
161 and Game (IDFG) Eagle Fish Hatchery (IDFG-FW). Captive-reared fish were also cultured

162 from smolt to adult in seawater at the National Marine Fisheries Service Manchester Marine
163 Culture facility (NMFS-FW/SW). Captive broodstocks were maintained at separate facilities to
164 avoid catastrophic loss. Rearing methodologies are reported in Baker et al. (2009) for freshwater
165 and Frost et al. (2008a,b) for freshwater and saltwater rearing. Prior to release, the maturation
166 status and sex of captive-reared adults was determined using the ultrasound techniques described
167 in Frost et al. (2014) and fork-length was recorded. Beginning with SY 2005, tissue samples
168 were also taken from adults prior to release for genetic parentage analysis.

169 * Estimating potential egg deposition.----*

170 We developed regression equations using length-fecundity relationships for each rearing
171 group spawned in the hatchery to develop PED estimates for fish spawning in Redfish Lake in
172 the same year. We chose to evaluate these relationships by rearing group (freshwater [IDFG-
173 FW, NMFS-FW], saltwater [NMFS-FW/SW], anadromous), given the different rearing
174 conditions and hatchery practices that resulted in different proportions of females released from
175 each group annually and different sizes of the females, as well as different sets of years for each
176 group. Data used to estimate potential egg deposition were from SY 2010-2015 for the NMFS-
177 FW/SW females, 2004-2012 for the NMFS-FW females, 2004-2015 for the IDFG-FW females,
178 and 2008-2015 for the anadromous females. Models were used to estimate the effects of fish
179 size (fork length) and SY relative to fecundity to determine whether years could be pooled within
180 each rearing group. Only length was used in the model with NMFS-FW/SW females, given the
181 fact that the years when these fish were released were different from the years when these fish
182 were spawned in the hatchery. Linear regression analysis with fork length as a continuous
183 covariate, year as a factor, and their interaction was included in the modeling framework.
184 Akaike's Information Criterion (AICc) adjusted for sample size was used to compare relative

185 model support for the data (Burnham and Anderson 2002). The AICc values were compared,
 186 and the model with Delta AICc equal to 0.0 was determined to be best supported by the data.

187 After the best supported models were chosen, we assumed that all released females were
 188 successful at spawning and developed PED estimates based upon the fork length of the released
 189 females. Total annual egg deposition for Redfish Lake for SYs 2004 to 2011, with associated
 190 standard error (SE), was estimated using the following equations:

$$191 \quad Total\ Egg\ Deposition = \sum_{i=1}^n C_I + C_Y + C_L \times L_i + C_{Y \times L} \times L_i$$

192 where n was the number of released adults within which fecundity was predicted, C_I was the
 193 regression coefficient for the intercept, C_Y was for year, C_L was for length, $C_{Y \times L}$ was for their
 194 interaction, and L_i was the fork length for fish i and $Y \times L$ was their interaction.

$$195 \quad \frac{Var(Total\ Egg\ Deposition)}{n^2} = Var_I + Var_Y + L_{avg}^2 \times Var_L + L_{avg}^2 \times Var_{Y \times L} +$$

$$196 \quad 2[Cov_{I,Y} \times L_{avg} \times Cov_{I,L} + L_{avg} \times Cov_{I,Y \times L}] + 2 \times L_{avg} \times [Cov_{Y,L} + Cov_{Y,Y \times L}] + 2 \times L_{avg}^2$$

$$197 \quad \times Cov_{L,Y \times L} + Var_{Model\ Error}$$

198 where Var was the model-estimated variance for the subscripts defined above and for the
 199 unexplained or residual variance, Model Error. Cov was the model-estimated covariance
 200 between each value in the subscript pair, and L_{avg} was the average length of n adults. We used
 201 the square root of Var (Total Egg Deposition) as the Standard Error (SE) of total egg deposition.
 202 Statistical analysis was completed using the program R (R Core Team 2017).

203 *Smolt trapping and estimating smolt production, smolts per female, and egg-to-smolt*
 204 *survival.*-----

205 A fish trap located near the outlet of Redfish Lake was operated in each study year during
 206 the entire juvenile migration season from the first week of April through mid-June (Figure 1).

207 All captured Sockeye smolts were enumerated, anesthetized in buffered tricaine
208 methanesulfonate (MS-222), measured to fork length (nearest 1.0 mm), and weighed (nearest 0.1
209 g). The first 30-50 natural smolts captured per day were PIT tagged (Prentice et al. 1990) and
210 released approximately 250 m upstream of the trap one-half hour after sunset. Trap efficiency
211 was estimated daily by the proportion of PIT tagged fish recaptured in the trap. Annually, the
212 trapping operations were grouped into one to four intervals based on stream discharge and
213 consistent trapping probabilities to account for heterogeneous trapping efficiency across the
214 season (Steinhorst et al. 2004). The total number of natural-origin juvenile smolts was derived
215 using a modified Bailey adjusted Lincoln-Peterson estimator with 95% bootstrap confidence
216 intervals (software GSRUN 7.0; Steinhorst et al. 2004).

217 During trapping, scales were removed from a subsample of 5 natural-origin fish from
218 each 5-mm length group. Scales were separated and laid between microscope slides and aged
219 using the methods of Jearld (1983). Length-at-age values derived from length frequencies were
220 determined using the Rmix computer program. Rmix was developed by Du (2002) as an add-on
221 program to the R computing environment (R Core Team 2017) that utilized the original MIX
222 program developed by MacDonald and Green (1988). Rmix uses a maximum likelihood
223 estimation method to estimate the parameters of a mixture distribution with overlapping
224 components, such as the overlapping length distributions associated with smolt estimates of
225 different ages. Rmix proportions were multiplied by the total estimate of natural migrants to
226 determine the number of age-1 and age-2 smolts represented during each juvenile migration year.
227 Standard errors for the abundance of each age class and length of each age class were also
228 produced by Rmix.

229 Total smolt production (\hat{N}) for each SY was calculated as:

230
$$\hat{N} = \hat{S}_{y1}\hat{p}_{y1} + \hat{S}_{y2}\hat{p}_{y2}$$

231 where S_{y1} and S_{y2} are smolt numbers in years $y1$ and $y2$, and p_{y1} and p_{y2} are the proportions of
232 the SY in those years. Mean smolt production was also calculated for the evaluation period.

233 Given that smolt numbers and SY proportions were estimated independently, then the estimated
234 variance of the smolt total was:

235
$$\hat{V}(\hat{N}) \approx \hat{S}_{y1}^2 \hat{V}(\hat{p}_{y1}) + \hat{p}_{y1}^2 \hat{V}(\hat{S}_{y1}) + \hat{V}(\hat{S}_{y1})\hat{V}(\hat{p}_{y1})$$

236
$$+ \hat{S}_{y2}^2 \hat{V}(\hat{p}_{y2}) + \hat{p}_{y2}^2 \hat{V}(\hat{S}_{y2}) + \hat{V}(\hat{S}_{y2})\hat{V}(\hat{p}_{y2})$$

237 The number of smolts per female was estimated by dividing the number of females
238 released by the number of smolts produced from the corresponding SY. Egg-to-smolt survival
239 was calculated by dividing total smolt production by the potential egg deposition estimate for
240 each SY. Regression analyses were conducted to determine the relationship between the number
241 of released females and reproductive output as measured by the log-transformed number of
242 smolts and total number of deposited eggs. These results were compared to historic freshwater
243 productivity in Redfish Lake (Bjornn et al. 1968).

244 * Adult Trapping, Estimating smolt-to-adult return rates (SARs), Population Replacement----*

245 Anadromous adults were trapped annually across the entire adult migration period from
246 mid-July through mid-October at either the Redfish Lake Creek weir or at a weir located on the
247 upper Salmon River at the IDFG Sawtooth Fish Hatchery (Figure 1). Returning, natural-origin
248 anadromous adults (e.g. offspring of adult releases into Redfish Lake) were identified as having
249 an intact adipose fin. Data collected for natural-origin anadromous fish included fork length
250 (nearest 0.5 cm), sex, scales, and fin clips for genetic analysis.

251 In 2008 and 2009, adipose-intact adults returning to the Redfish Lake Creek trap were
252 assumed to be the progeny of natural production from Redfish Lake and scales were used to

253 assign ages because genetic samples were not taken from adults released in 2004. Four to five
254 scales from each fish were collected from the left side above the lateral line and slightly posterior
255 to the dorsal fin (as identified in Devries and Frie 1996). Program personnel viewed and aged
256 scales using methods identified in Schrader et al. (2011).

257 Parentage analyses (PBT) was used to assign adult offspring back to their respective SY
258 starting in 2010 since genetic samples were collected from adults released in 2005-forward.
259 Whole DNA was extracted using a Nexttec DNA isolation kit according to the manufacturer
260 instructions. Samples were genotyped with a panel of 13 to 16 microsatellite loci, and a
261 minimum of 9 loci per individual were needed for inclusion in the analyses (see the authors for
262 genotyping protocols). The software *Cervus* v. 3.0 (Kalinowski et al. 2007) was used to perform
263 the parentage analyses using parents with known sex. Up to one mismatch was allowed, and
264 only two parentage assignments were accepted. Once the parents were identified, the age and
265 origin of each returning fish could be determined.

266 Age could not be assessed for every returning adult with the above methods. In some
267 cases, scales were not collected or the scale was unreadable. Missing tissue samples, mutations,
268 genotyping errors and/or incomplete genotypes can lead to the inability to assign parentage to
269 every fish. Age/length keys (Isermann and Knight 2005) using known ages of fish as determined
270 by scales/genetics and corresponding fish lengths were used to annually assign ages to adults that
271 could not be aged by either of the above methods. The software FishR Vignette (Program R)
272 was used to assign ages using the semi-random method (Ogle 2013, 2016).

273 SARs (from Redfish Lake to Redfish Lake) were estimated by adding the age-3, age-4,
274 and age-5 anadromous returns from each SY and dividing by the estimated total smolt
275 production for that SY (\hat{N}). We estimated the variance of the SARs as:

276
$$\hat{V}(SAR) \approx \frac{SAR(1 - SAR)}{\hat{N}} + \frac{\sum_i \hat{V}(\hat{h}_i)}{\hat{N}^2} + \frac{SAR^2}{\hat{N}^2} \hat{V}(\hat{N})$$

277 and the 95% confidence interval as:

278
$$\left(SAR - 1.96\sqrt{\hat{V}(SAR)}, SAR + 1.96\sqrt{\hat{V}(SAR)} \right)$$

279 where \hat{h}_i was the estimated adult count from ages $I = 3-5$ and $\hat{V}(\hat{h}_i)$ was the estimated variance,
 280 and $\hat{V}(\hat{N})$ from above was the estimated variance around total smolt production \hat{N} . SARs were
 281 compared to historic estimates produced by Bjornn et al. (1968). The number of returning adults
 282 was also regressed against the number of released females for each SY.

283 Population replacement was defined as a minimum of two natural-origin adult recruits
 284 per released female and assumed an evenly split sex ratio. For this estimation, no density
 285 dependent effects or harvest was assumed. Replacement rates were calculated using the
 286 following equation:

287
$$Smolts\ per\ Female * SARs \geq 2$$

288 <A> RESULTS

289 <Adult releases>.-----

290 The number of released adults by rearing type is presented in Table 1. From 2004-2006,
 291 only adults from NMFS were available to release into Redfish Lake. Starting in 2007, releases
 292 also included adults from IDFG and in 2008, anadromous adults returned from this release
 293 strategy and other hatchery release strategies (Hebdon et al. 2004; Kline and Flagg 2014) and
 294 were released into Redfish Lake. The total number of adults released ranged from 176 in 2005 to
 295 1,621 in 2010 (Table 1). Within these releases, the number of total females ranged from 50 in
 296 2005 to 688 in 2010 (Table 1). While attempts were made to equalize sex-ratios between males
 297 and females upon release, this was not always possible.

298 *Potential egg deposition.*-----

299 Based upon AIC criteria, the model that included the year and length interaction was the
300 best fit for IDFG-FW and NMFS-FW females (Table 2). For anadromous fish, the year and
301 length additive model was determined to be the best fit (Table 2). For the NMFS FW/SW fish,
302 the length-only model was used. This resulted in separate linear regression equations for each
303 rearing group and SY.

304 Estimated potential egg deposition within Redfish Lake ranged from 91,748 eggs in 2005
305 (SE 10,800) when 50 females were released to a maximum of 1,697,192 eggs in 2010 when 688
306 females were released (SE 196,445; Table 3). Across study years, annual mean egg deposition
307 was 788,879. Anadromous females were longer and more fecund than captive-reared females,
308 averaging 2,679 eggs compared to 1,641 eggs per female in 2008-2011.

309 *Smolt production, egg-to-smolt survival, smolts per female.*

310 The total number of smolts resulting from natural production ranged from 4,822 (SE 654)
311 in SY 2007 to 27,765 (SE 1,638) in SY 2010 (Table 3, Figure 2). There was a significant,
312 positive relationship between the number of females released in Redfish Lake and smolt
313 production ($r^2 = 0.73$, $P = 0.004$, Figure 3). Mean annual smolt production in Redfish Lake was
314 estimated to be 11,593. Across all years, the majority (63% - 98%) of smolts migrated from
315 Redfish Lake at age-1 (Table 4). Average length of age-1 fish ranged 96-117 mm, while that of
316 age-2 fish ranged 125-146 mm (Table 4). There was a significant positive relationship between
317 the number of deposited eggs and the log-transformed number of smolts ($r^2 = 0.86$, $P < 0.001$,
318 Figure 2).

319 Egg-to-smolt survival ranged from 1.0% (SY 2009) to 6.6% (SY 2005; Table 3). Mean
320 egg-to-smolt survival for the study period was estimated to be 2.1%. The number of smolts per

321 female averaged 41.3 and ranged from 19.0-121.8 (Table 3). SY 2005 had the fewest number of
322 females released, but produced the highest number of smolts per female (Figure 3).

323 *Smolt-to-adult return rates (SAR) and replacement rates.*-----

324 From 2008 to 2016, 1,183 natural-origin adults returned to Redfish Lake and 916 were
325 offspring from SYs 2004–2011. In 2007, only three natural-origin adults returned and none of
326 these were age-3 adults from SY 2004. Of the 1,183 returning adults, age-length keys were used
327 to age 8% of the fish. The majority of natural-origin adults returned at age-4 (75%) and 22%
328 returned at age-5. Natural-origin adults recruiting from each SY ranged from 28 (SY 2011) to
329 374 (SY 2010; Table 3).

330 SARs across study years averaged 1.12% and ranged from 0.2% in 2011 to 3.2% in 2006
331 (Figure 4). The SAR for natural-origin adults from SY 2006 had the highest SAR values for any
332 other SY, with 2005 having the second highest estimate (Figure 4). We observed no significant
333 relationship between the number of females released and number of adult recruits returning from
334 a given SY ($r^2 = 0.14$, $P = 0.35$).

335 The relationship between the number of smolts per female and SAR is logarithmic
336 (Figure 5). At the current mean estimate of 41 smolts per female, a corresponding SARs > 4.9%
337 would be needed to reach population replacement. Conversely, with an estimated SAR of 3.0%
338 (SY 2006), 66 smolts per female would be required to reach replacement. At the average
339 observed SAR of 1.12%, 179 smolts per female are needed to reach replacement. While we have
340 observed SARs (3.19%) and smolts per female (120) that would have exceeded population
341 replacement if in accordance during these study years, high freshwater productivity and smolt to
342 adult survival rates have not occurred during the same SY.

343 <A> Discussion

344 This study reported productivity metrics for adult releases into Redfish Lake for the first
345 time since the late 1960's (Bjornn et al. 1968). Our results indicate that hatchery adults released
346 for re-introduction efforts has successfully begun to build a natural spawning population of
347 Sockeye Salmon in Redfish Lake. We present the productivity metrics from this release strategy
348 in order to evaluate the status of the Redfish Lake population relative to what existed in the latter
349 half of the nineteenth century and to provide a reference point for the next several decades. This
350 information is not only useful for tracking progress towards the establishment of a self-sustaining
351 natural spawning population but identifies key life-history events where survival may be limiting
352 replacement.

353 Our contemporary estimates of freshwater productivity indicate that the current
354 conditions in Redfish Lake do not appear to be limiting juvenile production. observed a
355 strong, positive response in smolt production with increasing numbers of released females and
356 deposited eggs. Estimates of smolt size and age at migration also suggest that juvenile Sockeye
357 Salmon are acquiring adequate dietary resources in Redfish Lake. Bjornn et al. (1968) observed
358 a positive relationship between the age that Sockeye juveniles migrated from Redfish Lake and
359 their growth during the first summer in the lake. When the mean length of a year class
360 approached 100 mm, over 90% of smolts migrated as yearlings (Bjornn et al. 1968). During this
361 evaluation, smolts were of similar size-at-age as those reported in the 1960's (Bjornn et al.
362 1968). We did not observe decreasing average smolt size or an increase in the proportion of age-
363 2 smolts as the total number of females or smolts increased, which might be expected if density
364 dependence was occurring (Kyle et al. 1988). We believe that smolt abundance has the
365 capability of increasing even further with the release of more females into Redfish Lake;
366 particularly anadromous females that are larger in size and capable of depositing more eggs.

367 Increased smolt abundances, however, did not translate into more returning natural-origin
368 adults as a result of highly variable and low SARs.

369

370

371 extreme temperatures in the
372 migratory corridor led to significant losses of adult Snake River Sockeye in migration year 2015
373 (NMFS 2016). These losses would have impacted the SARs for SY 2010 and SY 2011. Other
374 studies have shown that ocean productivity, as measured by the Pacific Decadal Oscillation
375 (PDO), plays a major role in salmon and steelhead survival and can drive adult return rates for
376 many populations (Mantua et al. 1997; Peterman and Dorner 2011; Petrosky and Schaller 2010;
377 Anderson et al. 2014; Williams et al. 2014). Snake River SARs were found to be highly
378 correlated with SARs from the nearest extant populations of Sockeye Salmon in the Columbia
379 River and there was a significant relationship between PDO and adult returns, indicating that a
380 common variable within the marine portion of their life-cycle was affecting post-juvenile
381 productivity (NOAA 2009). (2015) suggested that much of the life-cycle mortality
382 experienced by Snake River Sockeye Salmon occurred in the marine environment and was due to
383 low ocean productivity and shifts in preferred zooplankton food species. Nevertheless, it is
384 critically important to maximize the numbers of juvenile migrants as a safeguard against variable
385 marine and migratory conditions to ensure that some natural-origin adults return.

386 The adults that have been used to re-establish natural production have been exposed to
387 multiple generations within captivity but do not appear to exhibit reduced productivity when
388 compared to historic estimates from wild adults (Bjornn et al 1968). We found that
389 contemporary egg-to-smolt survival estimates fell within the range of those historically reported

390 for Redfish Lake (Range 0.58% -143%; Bjornn et al. 1968). Contemporary estimates of post-
391 juvenile productivity, as measured through SARs, even exceeded the historic range for SY2006
392 (Range 0.14% - 1.83%; Bjornn et al. 1968). It difficult to compare current estimates with those
393 observed during the 1960s due to downriver harvest rates potentially as high as 60% (Bjornn et
394 al. 1968) and fewer Snake River dams at that time.

395 Both the historic and current productivity estimates for Sockeye Salmon in Redfish Lake
396 were near the lower end of the range of other Sockeye Salmon populations. Bradford (1995)
397 reported that average egg-to-smolt survival ranged from 3.2% to 6.2% for seven populations of
398 Sockeye Salmon. Chapman et al. (1995) reported that egg-to-smolt survival for Lake Wenatchee
399 Sockeye ranged from 1.7 to 12.3% and egg-to-smolt survival for Okanogan sockeye ranged from
400 2.4 to 38%. Hyatt et al. (2005) reported wild sockeye salmon egg-to-smolt survival for
401 Tatsamenie and Tahltan lakes in British Columbia as 5.8 and 3.6 percent, respectively. SAR's
402 were also lower when compared to northern populations in British Columbia and Alaska (Range
403 1.34% to 3.4%, Bradford 1995; Chilko Lake BC, 2-5%; DFO 2017) and upper Columbia River
404 populations (Range 0.67% - 9.43%, NOAA 2009; Range 0.2% to 23.5% Williams et al. 2014).
405 Productivity levels were more similar to levels observed in other critically low populations such
406 as the Cultus Lake (avg 76 smolts per spawner; smolt-to-adult survival for 2003-2005 = 1%;
407 Bradford et al. 2010; Ackerman et al 2014) and Sakinaw Lake populations (egg-to-smolt survival
408 ranged 0.1% - 6%, SAR ranged 0% to 0.8%;Withler et al. 2014; COSEWIC 2016).

409 These comparisons suggest that the Redfish Lake population, when examined in the
410 1950s and 1960s, may have already experienced declines in productivity. There are intrinsic
411 differences in food availability, predation, and limnological characteristics in each rearing lake
412 (Finkle and Harding 2015) and differences related to migratory conditions and distance to the

413 ocean, impoundments, ocean rearing location, age structure, and harvest. The Redfish Lake
414 population had already experienced a population bottleneck prior to Bjornn's evaluation during
415 the 24 years when Sunbeam Dam was in place. It was hypothesized that the prior loss of the
416 anadromous return to Redfish Lake reduced nutrient loading and contributed to low production
417 (Wurtsbaugh et al. 1997). It is also possible that the Redfish Lake population always had lower
418 productivity relative to other Sockeye populations due to its location at the periphery of the range
419 in North America in a high-elevation, oligotrophic lake.

420 Smolt production from residual females in Redfish Lake likely introduced some degree of
421 bias in both the current and historic rates of freshwater productivity. Bjornn et al. (1968)
422 originally hypothesized that residual production could be an influence when egg-to-smolt
423 survival rates were as high as 21% and 143%, which is biologically impossible for the latter
424 estimate. The estimate for SY 2005 (6.6%) appears to be an outlier among current estimates and
425 indicates that residual production may have been a factor. Although the residual population is
426 difficult to enumerate, night-time snorkel surveys continue to document the presence of these
427 fish during spawning. Residuals are much smaller in size (i.e., similar to resident kokanee) and
428 their egg size and fecundity is low compared to captive-reared or anadromous females (Burgner
429 1991). The overall smolt production from residual spawning events is uncertain. However, we
430 suspect that the contribution may be greater when there is less competition or uneven sex ratios;
431 as the years with presumably greater residual contribution were the years when less than 50
432 anadromous or captive-reared females were released to spawn.

433 The SARs can also be biased if there are errors in aging or if there was adult straying
434 between trapping locations. Ageing errors generally decrease strong cohorts and inflate weak
435 cohorts that either precede or follow the strong cohort (Campana et al. 2001). These errors can

436 have more of an impact on the SAR values for the weaker cohorts and inflate mean SAR values
437 (Copeland et al. 2007). Parentage analysis provides an accurate estimate of age structure and
438 origin and removes this bias (Seamons et al. 2009). However, scale aging was used for two of
439 the return years, which may have led to some small degree of bias in the productivity metrics for
440 SY 2004 and SY 2005. For these same years, we also assumed that any natural-origin fish
441 trapped at Redfish Lake Creek was the offspring of adult releases into Redfish Lake and not the
442 product of another lake or release strategy (e.g. egg boxes in other lakes). Genetic parentage
443 assignments from the eight most recent years of anadromous returns indicated an average stray
444 rate of 1.0% between trapping locations. If this rate was consistent during the years we
445 evaluated, it likely had little effect on SARs and would not significantly change the interpretation
446 of our results.

447 In order for the Redfish Lake population to grow and become self-sustaining, survival
448 will need to increase at multiple life-stages. This is the case in spite of productivity metrics that
449 fell within historical ranges. The current survival rates are low compared to other Sockeye
450 Salmon populations and without an increase in freshwater survival, SARs would need to exceed
451 4.9% for population replacement. Redfish Lake resides in a national recreational area and there
452 has been little human development although boating activities and natural events could increase
453 siltation and reduce groundwater upwelling or substrate permeability during incubation (B.
454 Griswold, Biolines Consulting, personal communication). Natural production potential in
455 Redfish Lake may have also been subsumed by the resident, kokanee population and reductions
456 of the kokanee population might facilitate the re-establishment of Sockeye Salmon (Gross et al.
457 1998). Freshwater and post-juvenile productivity are inter-related as increased freshwater
458 growth rates can lead to increased SARs (Koenings and Burkett 1987; Henderson and Cass 1991;

459 Koenings et al. 1993). Conversely, increased numbers of anadromous adult spawners can
460 increase freshwater productivity (Gross et al. 1998; Cederholm et al. 1999). For Cultus Lake, the
461 number of smolts per spawner decreased by 50% when the number of adult spawners was below
462 7,000 adults (Cultus Sockeye Recovery Team 2005). Gross et al. (1998) indicated that
463 increasing SARs and the numbers of anadromous, spawning adults in Redfish Lake would
464 provide greater benefit than lake fertilization. Until survival can be improved, releasing captive-
465 reared adults into the lake to volitionally spawn will prevent cohort collapse during years of
466 unfavorable ocean productivity, when fewer anadromous adults return (Kline and Flagg 2014).
467 Additionally, both captive-reared and anadromous adult releases will continue to help build a
468 natural spawning population within Redfish Lake during periods of favorable marine growth and
469 survival (Kline and Flagg 2014).

470 Population growth rates and survival may increase as more anadromous adults return to
471 the program and are released to spawn in Redfish Lake. Not only can anadromous spawners
472 provide marine-derived nutrient loading that can boost survival and growth of juvenile sockeye
473 salmon, but they may also be more successful spawners. We assumed that all females spawned
474 successfully, deposited eggs into spawning gravel of equivalent quality, and had equivalent rates
475 of egg viability and survival. Sockeye salmon display high variance in reproductive success
476 (Mehranvar et al.2004) and there could be differences in reproductive success by rearing type,
477 especially between captive-reared and anadromous fish. The anadromous adults are larger and
478 have more body coloration and these traits may be advantageous during spawning (Fleming and
479 Gross 1994; Steen and Quinn 1999; Foote et al. 2004; Garcia de Leaniz et al. 2007). Berejekian
480 and Ford (2004) suggested that the duration of rearing in captivity can have an impact on
481 domestication selection and reproductive success. Other studies have indicated differential

482 reproductive success between hatchery and natural conspecifics (Fleming and Gross 1993;
483 Fleming and Petersson 2001; Williamson et al. 2010; Anderson et al. 2012; Ford et al. 2012;
484 Kozfkay et al. 2017) and have related this difference to age at maturation, fish size and
485 competition, spawn-timing, redd construction and location, or egg viability (Williamson et al.
486 2010; Anderson et al. 2012; Ford et al. 2012, Stark et al. 2018). Juvenile fitness-related traits
487 such as size and emergence timing have also been linked to maternal phenotype (Braun et al.
488 2013).

489 Adaptive evolution might be necessary before population increases are observed
490 (Anderson et al. 2014). Much of the available literature suggests that domestication selection
491 can occur during hatchery rearing (Araki et al. 2007; Araki et al. 2008; Christie et al. 2012), but
492 Fraser (2008) hypothesized that captive-reared fish could re-adapt to the wild within a timeframe
493 similar to that during which domestication selection occurred in the hatchery. Evans et al.
494 (2014) provided empirical support that increased survival of offspring can occur after one
495 generation of exposing parents to the natural environment and suggested that traits were being
496 selected that were adapted to natural conditions. For Snake River Sockeye Salmon, there is an
497 opportunity for adaptive evolution to occur as the population becomes more wild-exposed and
498 adult releases shift from predominantly captive-reared adults to hatchery, anadromous adults that
499 are reared in the hatchery until the smolt-stage, to natural-origin anadromous adults that are born
500 in Redfish Lake (IDFG 2010; NMFS 2015). Therefore, it is possible that domestication selection
501 can be reversed by limiting the time in captivity and with increased wild-exposure.

502 As natural Sockeye Salmon populations continue to be rebuilt, continued monitoring of
503 these productivity metrics will be important for understanding the production potential of
504 Redfish Lake and other natal lakes. This information not only provides a baseline to track the

505 status of the population through time but acts as a baseline to assess the outcomes for future
506 restoration and recovery actions. Additional research is needed regarding the factors that affect
507 survival in freshwater and marine environments. Evaluations of the reproductive performance of
508 fish released into Redfish Lake and the other natal lakes will also be important for assessing the
509 relative contributions by rearing type and life-time fitness. Our results suggest that the captive
510 broodstock program can be used to re-establish Snake River Sockeye Salmon throughout their
511 natal range and supports the supposition that survival and fitness may increase as the population
512 becomes more wild-exposed through anadromy. Ultimately the performance of the population
513 will depend on a combination of environmental, genetic, and ecological factors.

514 <A>Acknowledgements

515 This project was funded by the Bonneville Power Administration. Thanks to the field
516 crews who collected data during spring and summer 2004-2016. Special thanks to Kurtis Plaster,
517 who oversaw much of this data collection and to Debbie Frost and Dan Baker, who tracked fish
518 and provided data for adult releases. We also thank Josh McCormick, Beth Sanderson, Dan
519 Schill, John Powell, Lance Hebdon, Mark Scheuerell, and JoAnne Butzerin for earlier reviews of
520 this manuscript.

521

522

523

524

525

526

527

528 <A> References

- 529 Ackerman, P.A., Barnetson, S., Lofthouse, D., McClean, C., Stobbart, A., and Withler, R.E.
530 Back from the Brink: 2014. The Cultus Lake Sockeye Salmon Enhancement Program
531 from 2000 - 2014. Canadian Manuscript Report of Fisheries and Aquatic Sciences 3032:
532 vii + 63p.
- 533 Araki, H., B. Cooper, and M. S. Blouin. 2007. Genetic effects of captive breeding cause a rapid,
534 cumulative fitness decline in the wild. *Science* 318:100-103.
- 535 Araki, H., B. A. Berejikian, M. J. Ford, and M. S. Blouin. 2008. Fitness of hatchery-reared
536 salmonids in the wild. *Evolutionary Applications* 1:342–355.
- 537 Araki, H. B., and Co. Schmid. 2010. Is hatchery stocking a help or harm? Evidence, limitations
538 and future directions in ecological and genetic surveys. *Aquaculture* 308:S2-S11.
- 539 Anderson, J. H., P. L. Faulds, W. I. Atlas, and T. P. Quinn. 2012. Reproductive success of
540 captive bred and naturally spawned Chinook salmon colonizing newly accessible
541 habitat. *Evolutionary Applications* 6:165-179.
- 542 Anderson, J. H., G. R. Pess, R. W. Carmichael, M. J. Ford, T. D Cooney, C. M. Baldwin, and M.
543 M. McClure. 2014. Planning Pacific Salmon and Steelhead reintroductions aimed at
544 long-term viability and recovery. *North American Journal of Fisheries Management*
545 34:72-93.
- 546 Baker, D., T. Brown, D. Green, and J. Heindel. 2009. Snake River Sockeye Salmon Captive
547 Broodstock Program Hatchery Element, 2008. IDFG Report no. 10-09. Project no.
548 200740200. Bonneville Power Administration, Annual Report. Portland, Oregon.
- 549 Berejikian, B., T. Flagg, and P. Kline. 2004. Release of captive reared adult anadromous
550 salmonids for population maintenance and recovery: Biological trade-offs and
551 management considerations. Pages 233-245 in M. J. Nickum, P. M. Mazik, J. G. Nickum,

552 and D. D. MacKinlay, editors. Propagated fish in resource management. American
553 Fisheries Society, Symposium 44, American Fisheries Society, Bethesda, Maryland.

554 Berejikian, B. and M. Ford. 2004. A review of relative fitness of hatchery and natural salmon. U.
555 S. Department of Commerce. NOAA technical memorandum NMFS-NWFSC-61, 28 p.

556 Berejikian, B. A., D. M. Van Doornik, J. A. Scheurer, and R. Bush. 2009. Reproductive behavior
557 and relative reproductive success of natural- and hatchery-origin Hood Canal summer
558 Chum Salmon (*Oncorhynchus keta*). Canadian Journal of Fisheries and Aquatic Sciences
559 66:781-789.

560 Bjornn, T. C., D. R. Craddock, and D. R. Corley. 1968. Migration and survival of Redfish Lake,
561 Idaho, Sockeye Salmon, *Oncorhynchus nerka*. Transactions of the American Fisheries
562 Society 97:360-373.

563 Bradford, M. J. 1995. Comparative review of Pacific salmon survival rates. Canadian Journal of
564 Fisheries and Aquatic Sciences 52:1327-1338.

565 Bradford, M. J., J. M. B. Hume, R. E. Withler, D. Lofthouse, S. Barnettson, S. Grant, M. Folkes,
566 N. Schubert, and A.-M. Huang, 2010 Status of Cultus Lake Sockeye Salmon. Canadian
567 Science Advisory Secretariat Research Document 2010/123.

568 Braun, D.C., Patterson, D.A. and Reynolds, J.D., 2013. Maternal and environmental influences
569 on egg size and juvenile life-history traits in Pacific salmon. Ecology and evolution 3:
570 1727-1740.

571 Budy, P., C. Luecke, and W. A. Wurtsbaugh. 1998. Adding nutrients to enhance the growth of
572 endangered Sockeye Salmon: trophic transfer in an oligotrophic lake. Transactions of the
573 American Fisheries Society 127:19-34.

574 Burgner, R. L. 1991. Life History of Sockeye Salmon (*Oncorhynchus nerka*). Pages 3-117 in C.

575 Groot and L. Margolis, editors. Pacific Salmon Life Histories. University of British
576 Columbia Press, Vancouver, British Columbia.

577 Burnham, K. P., and D. R. Anderson. 2002. Model selection and multi-model inference, 2nd
578 Ed. Springer. New York.

579 Carr, J. W., F. Whoriskey, and P. O'Reilly. 2004. Efficacy of releasing captive reared broodstock
580 into an imperiled wild Atlantic salmon population as a recovery strategy. Journal of Fish
581 Biology 65:38-54 (Supplement A).

582 Cederholm, C. J., M. D. Kunze, T. Murota, and A. Sibatani. 1999. Pacific salmon carcasses:
583 essential contributions of nutrients and energy for aquatic and terrestrial ecosystems.
584 Fisheries 24:6-15.

585 Christie, M. R., M. L. Marine, R. A. French, and M. S. Blouin. 2012. Genetic adaptation to
586 captivity can occur in a single generation. Proceedings of the National Academy of
587 Sciences 109:238-242.

588 Christie, M. R. M. J. Ford, and M. S. Blouin. 2014. On the reproductive success of
589 early-generation fish in the wild. Evolutionary Applications 7:883-896.

590 Campana, S. E. 2001. Accuracy, precision and quality control in age determination, including a
591 review of the use and abuse of age validation methods. Journal of Fish Biology 59:197-
592 242.

593 Conrad, J. L., E. A. Gilbert-Horvath, and J. C. Garza. 2013. Genetic and phenotypic effects on
594 reproductive outcomes for captively-reared coho salmon, *Oncorhynchus kisutch*.
595 Aquaculture 404-405: 95-104

596 Cooper, A. M., L. M. Miller, and A. R. Kapuscinski. 2009. Conservation of population structure
597 and genetic diversity under captive breeding of remnant coaster Brook Trout (*Salvelinus*

598 *fontinalis*) populations. Conservation Genetics 11:1087-1093.

599 Copeland, T., M. W. Hyatt, and J. Johnson. 2007. Comparison of methods used to age spring-
600 summer chinook salmon in Idaho: validation and simulated effects on estimated age
601 composition. North American Journal of Fisheries Management 27: 1393-1401.

602 COSEWIC 2016. COSEWIC assessment and status report on the Sockeye Salmon
603 *Oncorhynchus nerka* Sakinaw population in Canada. Committee on the Status of
604 Endangered Wildlife in Canada. Ottawa.

605 Cultus Sockeye Recovery Team. 2005. National conservation strategy for sockeye salmon
606 (*Oncorhynchus nerka*), Cultus Lake population, in British Columbia. Recovery of
607 Nationally Endangered Wildlife (RENEW). Ottawa, Ontario, 49 pp.

608 Cummings, S. A., E. L. Brannon, K. J. Adams, and G. H. Thorgaard. 1997. Genetic analyses to
609 establish captive breeding priorities for endangered Snake River sockeye salmon.
610 Conservation Biology 11:662–669.

611 Devries, D. R., and R. V. Frie. 1996. Determination of age and growth. Pages 483-512 in B. R.
612 Murphy and D. W. Willis, editors. Fisheries Techniques, 2nd edition. American Fisheries
613 Society, Bethesda, Maryland.

614 DFO. 2017. Pre-season run size forecasts for Fraser River Sockeye (*Oncorhynchus nerka*) and
615 Pink (*O. gorbuscha*) salmon in 2017. DFO Can. Sci. Advis. Sec. Sci. Resp. 2017/016,
616 Canada Dept. of Fisheries and Oceans. Pacific Region; Canadian Science Advisory
617 Secretariat, Ottawa.

618 Du, Juan B.Sc. 2002. Combined algorithms for constrained estimation of finite mixture
619 distributions with grouped data and conditional data. Masters thesis. McMaster
620 University, Hamilton, Ontario, Canada.

621 Evans, M. L., N. F. Wilke, P. T. O'Reilly, and I. A. Fleming. 2014. Transgenerational effects of
622 parental rearing environment influence the survivorship of captive-born offspring in the
623 wild. *Conservation Letters* 7:371-379.

624 Finkle, H., and J. Harding. Karluk Sockeye Salmon Smolt Enumeration, 2014 Season Summary.
625 Alaska Department of Fish and Game, Division of Sport Fish, Research and Technical
626 Services, 2015.

627 Fisch, K. M., J. A. Ivy, R. S. Burton, and B. 2012. Evaluating the performance of captive
628 breeding techniques for conservation hatcheries: a case study of the delta smelt captive
629 breeding program. *Journal of Heredity* 104: 92-104.

630 Fisch, K. M., C. C. Kozfkay, J. A Ivy. O. A. Ryder and R. S. Waples. 2015. Fish hatchery
631 genetic management techniques: integrating theory with implementation. *North*
632 *American Journal of Aquaculture* 77: 343-357.

633 Flagg, T. A., and C. V. W. Mahnken. 1995. An assessment of the status of captive broodstock
634 technology for Pacific Salmon, Final Report to Bonneville Power Administration. Project
635 No. 93-56, p.298. Portland, Oregon.

636 Flagg TA, McAuley WC, Kline PA et al (2004) Application of captive broodstocks to
637 preservation of ESA-listed stocks of Pacific Salmon: Redfish Lake sockeye salmon case
638 example. In: Nickum MJ, Mazik PM, Nickum JG, MacKinlay DD (eds) Propagated fish
639 in resource management. *American Fisheries Society Symposium* 44, American Fisheries
640 Society, Bethesda, pp 387–400.

641 Fleming, I. A., and M. R. Gross. 1993. Breeding success of hatchery and wild Coho Salmon
642 (*Oncorhynchus Kisutch*) in competition. *Ecological Applications* 3: 230-245.

643 Fleming, I. A. and M. R. Gross. 1994. Breeding competition in a Pacific Salmon (Coho:

644 *Oncorhynchus kisutch*): Measures of natural and sexual selection. *Evolution* 48: 637-657.

645 Fleming, I. A., and E. Petersson. 2001. The ability of released, hatchery salmonids to breed and
646 contribute to the natural productivity of wild populations. *Nordic Journal of Freshwater*
647 *Research* 75:71-98.

648 Foote, C. J., G. S. Brown and C. W. Hawryshyn. 2004. Female colour and mate choice in
649 sockeye salmon: implications for the phenotypic convergence of anadromous and non-
650 andromous morphs. *Animal Behavior* 67: 69-83.

651 Ford, M. A. Murdoch and S. Howard. 2012. Early male maturity explains a negative correlation
652 in reproductive success between hatchery-spawned salmon and their naturally spawning
653 progeny. *Conservation Letters*: 450-458.

654 Fraser, D. J. 2008. How well can captive breeding programs conserve biodiversity? A review of
655 salmonids. *Evolutionary Applications* 1:535-586.

656 Frost, D. A., D. J. Maynard, W. C. McAuley, M. R. Wastel, B. Kluver, and T. A. Flagg. 2008a.
657 Redfish Lake Sockeye Salmon captive broodstock rearing and research, 2006 Annual
658 Report. Report to Bonneville Power Administration, Contract No. 00004464, p 38,
659 Portland, Oregon.

660 Frost, D. A., W. C. McAuley, D. J. Maynard, M. R. Wastel, B. Kluver, and T. A. Flagg. 2008b.
661 Redfish Lake Sockeye Salmon captive broodstock rearing and research, 2007 Annual
662 Report. Report to Bonneville Power Administration, Contract No. DE-AI79-92BP41841,
663 p 44. Portland, Oregon.

664 Frost, D. A. W. C. McAuley, B. Kluver, M. Wastel, D. Maynard, and T. A. Flagg. 2014.
665 Methods and Accuracy of Sexing Sockeye Salmon Using Ultrasound for Captive
666 Broodstock Management, *North American Journal of Aquaculture*, 76:2, 153-158.

667 Garcia de Leaniz, C., Fleming, I. A., Einum, S., Verspoor, E., Jordan, W. C., Consuegra, S., ... &
668 Webb, J. H. 2007. A critical review of adaptive genetic variation in Atlantic salmon:
669 implications for conservation. *Biological reviews* 82: 173-211.

670 Godbout, L., C. C. Wood, R. E. Withler, S. Latham, R. J. Nelson, L. Wetzel, R. Barnett-Johnson,
671 M. J. Grove, A. K. Schmitt, and K. D. McKeegan. 2011. Sockeye Salmon (*Oncorhynchus*
672 *nerka*) return after an absence of nearly 90 years: a case of reversion to anadromy.
673 *Canadian Journal of Fisheries and Aquatic Sciences* 68:1590-1602.

674 Griffiths, A. M., J. S. Ellis, D. Cliftn-Dey, G. Machado-Schiaffino, D. Bright, E.
675 Garcia-Vazquez, and J. R. Stevens. (2011). Restoration versus recolonisation: The
676 origin of Atlantic Salmon (*Salmon salar L.*) currently in the River Thames. *Biological*
677 *Conservation* 144:2733-2738.

678 Gross H. P., W. A. Wurtsbaugh, and C. Luecke. 1998. The role of anadromous Sockeye Salmon
679 in the nutrient loading and productivity of Redfish Lake, Idaho. *Transactions of the*
680 *American Fisheries Society* 127:1-18

681 Hebdon, J. L., P. Kline, D. Taki, and T. A. Flagg. 2004. Evaluating reintroduction strategies for
682 Redfish Lake Sockeye Salmon captive broodstock progeny. Pages 401-413 in M. J.
683 Nickum, P. M. Mazik, J. G. Nickum, and D. D. MacKinlay, editors. *Propagated fish in*
684 *resource management. American Fisheries Society, Symposium 44, American Fisheries*
685 *Society, Bethesda, Maryland.*

686 Hess, M. A., C. D. Rabe, J. L. Vogel, J. J. Stephenson, D. D. Nelson, and S. R. Narum. 2012.
687 Supportive breeding boosts natural population abundance with minimal negative impacts
688 on fitness of a wild population of Chinook Salmon. *Molecular Ecology* 21:5236-5250.

689 IDFG (Idaho Department of Fish and Game). 2010. Master Plan for the Snake River Sockeye

690 program. Idaho Department of Fish and Game, Boise, Idaho.

691 Isermann, D. A., and C. T. Knight. 2005. A computer program for age-length keys incorporating
692 age assignment to individual fish. *North American Journal of Fisheries Management*
693 25:1153–1160.

694 Jearld, A., Jr. 1983. Age determination. Pages 301-324 *in* L. A. Nielsen and D. L. Johnson,
695 editors. *Fisheries techniques*. American Fisheries Society, Bethesda, Maryland.

696 Kalinowski, S. T., M. L. Taper, and T. C. Marshall. 2007. Revising how the computer program
697 CERVUS accommodates genotyping error increases success in paternity assignment.
698 *Molecular Ecology* 16, 1099-1106.

699 Kalinowski, S. T., D. M. Van Doornik, C. C. Kozfkay, and R. S. Waples. 2012. Genetic diversity
700 in the Snake River Sockeye Salmon captive broodstock program. *Conservation Genetics*
701 13:1183-1193.

702 Kline, P. A., and T. A. Flagg. 2014. Putting the Red Back in Redfish Lake, 20 Years of Progress
703 Toward Saving the Pacific Northwest's Most Endangered Salmon Population. *Fisheries*
704 39:488-500.

705 Kozfkay, C.C., M R. Campbell, J. A. Heindel, D. J. Baker, P. M. S. Powell and T. Flagg. 2008.
706 A genetic evaluation of relatedness for broodstock management of captive, endangered
707 Snake River sockeye salmon, *Oncorhynchus nerka*. *Conservation Genetics* 9:1421-1430.

708 Kozfkay, C. C., E. J. Stark, D. J. Baker, B. S. Ayers, D. A Venditti, J. McCormick, and P. Kline.
709 2017. Captive Rearing Program for Salmon River Chinook Salmon. Completion Report
710 for Jan 1, 1995 – Dec 31, 2014. IDFG Report Number 16-11, Boise, Idaho.

711 Kyle, G. B., J. P. Koenings, and B. M. Barrett. 1988. Density-dependent, trophic level
712 responses to an introduced run of sockeye salmon (*Oncorhynchus nerka*) at Frazer Lake,

713 Kodiak Island, Alaska. Canadian Journal of Fisheries and Aquatic Sciences 45: 856-867.

714 MacDonald, P. D. M., and P. E. J. Green. 1988. User's Guide to Program MIX: an interactive
715 program for fitting mixtures of distributions. Release 2.3, January 1988. Ichthus Data
716 Systems, Hamilton, Ontario. iv+60 pp. ISBN 0-9692305-1-6.

717 Mantua, N. J., S. R Hare, Y. Zhang, J. M. Wallace and R. C. Francis. 1997. A Pacific
718 interdecadal climate oscillation with impacts on salmon production. Bulletin of the
719 american Meteorological Society, 78(6), pp.1069-1080.

720 Mehranvar, L. M. Healey, A Farrell, and S. Hinch. 2004. Social versus genetic measures of
721 reproductive success in sockeye salmon, *Oncorhynchus nerka*. Evolutionary Ecology
722 Research 6: 1167-1181.

723 Miller, L. M., and A. R. Kapuscinski. 2003. Genetic Guidelines for hatchery supplementation
724 programs. Pages 329-356 in E. M. Hallerman, editor. Population genetics: principles
725 and applications for fisheries scientists. American Fisheries Society, Bethesda, Maryland.

726 Nichols, K. M., C. C. Kozfkay, and S. R. Narum. 2016. Genomic signatures among
727 *Oncorhynchus nerka* ecotypes to inform conservation and management of endangered
728 Sockeye Salmon. Evolutionary Applications 9:1285-1300.

729 NMFS (National Marine Fisheries Service). 1991. Endangered and threatened species;
730 endangered status for Snake River Sockeye Salmon. Federal Register
731 56(224):58619-58624.

732 NMFS (National Marine Fisheries Service). 2015. ESA Recovery Plan for Snake River
733 Sockeye Salmon (*Oncorhynchus nerka*) – June 8, 2015. 431 p. Available at
734 [www.westcoast.fisheries.noaa.gov/publications/recovery_planning/salmon_steelhead/do](http://www.westcoast.fisheries.noaa.gov/publications/recovery_planning/salmon_steelhead/domains/interior_columbia/snake/snake_river_sockeye_recovery_plan_june_2015.pdf)
735 [mains/interior_columbia/snake/snake_river_sockeye_recovery_plan_june_2015.pdf](http://www.westcoast.fisheries.noaa.gov/publications/recovery_planning/salmon_steelhead/domains/interior_columbia/snake/snake_river_sockeye_recovery_plan_june_2015.pdf).

736 NMFS (National Marine Fisheries Service). 2016. 2015 Adult Sockeye Salmon Passage Report.
737 Available at www.westcoast.fisheries.noaa.gov/publications/hydropower
738 [/fcrps/2015_adult_sockeye_salmon_passage_report.pdf](http://www.westcoast.fisheries.noaa.gov/publications/hydropower/fcrps/2015_adult_sockeye_salmon_passage_report.pdf).

739 Ogle, D. H. 2013. FSA: Fisheries Stock Analysis. R package version 0.4.11.

740 Ogle, D. H. 2016. Introductory fisheries analyses with R. Chapman & Hall/CRC, Boca Raton,
741 FL.

742 O'Reilly, P. T., and C. C. Kozfkay. 2014. Use of microsatellite data and pedigree information in
743 the genetic management of two long-term salmon conservation programs. *Reviews in*
744 *Fish Biology and Fisheries* 24:819-848.

745 Osborne, M. J., T. L. Perez, C. S. Altenbach, and T. F. Turner. 2013. Genetic analysis of captive
746 spawning strategies for the endangered Rio Grande Silvery Minnow. *Journal of Heredity*
747 104:437-446.

748 Peterman, R. M., and B. Dorner. 2011. Fraser River sockeye production dynamics. Vancouver,
749 B.C. Cohen Commission Technical Report 10:133 pages. www.cohencommission.ca

750 Petrosky, C. E. and H. A. Schaller. 2010. Influence of river conditions during seaward migration
751 and ocean conditions on survival rates of Snake River Chinook salmon and steelhead.
752 *Ecology of Freshwater Fish* 19:520-536.

753 Prentice, E. F., T. A. Flagg, C. S. McCutcheon, D. F. Brastow, and D. C. Cross. 1990.
754 Equipment, methods, and an automated data-entry station for PIT tagging. Pages 335-340
755 in N. C. Parker, A. E. Giorgi, R. C. Hedinger, D. B. Jester, Jr., E. D. Prince, and G. A.
756 Winans, editors. *Fish-marking techniques*. American Fisheries Society, Symposium 7,
757 Bethesda, Maryland.

758 R Core Team (2017). R: A language and environment for statistical computing. R Foundation for

759 Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>.

760 Rieman, B. E., D. L Meyers, and R. L Nielsen. 1994. Use of otolith microchemistry to
761 discriminate *Oncorhynchus nerka* of resident and anadromous origin. Canadian Journal
762 of Fisheries and Aquatic Sciences 51:68-77.

763 Saltzgeber, M. J., E. J. Heist, and P. W. Hedrick. 2012. Genetic evaluation of the initiation of a
764 captive population: the general approach and a case study in the endangered Pallid
765 Sturgeon (*Scaphirhynchus albus*). Conservation Genetics 13:1381-1391.

766 Schrader, W. C., T. Copeland, M. W. Ackerman, K. Ellsworth, and M. R. Campbell. 2011. Wild
767 adult Steelhead and Chinook Salmon abundance and composition at Lower Granite Dam,
768 Spawn Year 2009. IDFG Report no. 11-24. Project no's. 1990-055-00, 1991-073-00,
769 2010-026-00. Bonneville Power Administration, Annual Report. Portland, Oregon.

770 Seamons, T. R., M. B. Dauer, J. Sneva, and T. P. Quinn. 2009. Use of parentage assignment and
771 DNA genotyping to validate scale analysis for estimating Steelhead age and spawning
772 history. North American Journal of Fisheries Management, 29:2, 396-403.

773 Stark, E. J., D. T. Vidergar, C. C. Kozfkay and P. A. Kline. 2018. Egg viability and egg-to-fry
774 survival of captive-reared Chinook Salmon released to spawn naturally. Transactions of
775 the American Fisheries Society 147:1-11.

776 Steinhorst, K., Y. Wu, B. Dennis, and P. Kline. 2004. Confidence intervals for fish out-migration
777 estimates using stratified trap efficiency methods. Journal of Agricultural, Biological, and
778 Environmental Statistics 9:284-299.

779 Steen, R. P. and T. P. Quinn. 1999. Egg burial depth by sockeye salmon (*Oncorhynchus nerka*):
780 implications for survival of embryos and natural selection for female body size. Canadian
781 Journal of Zoology 77:836-841.

782 Tucker, S., M. E. Thiess, J. F. T. Morris, D. Mackas, W. T. Peterson, J. R. Candy, T. D.
783 Beacham, E. M. Iwamoto, D. J. Teel, M. Peterson, and M. Trudel. 2015. Coastal
784 distribution and consequent factors influencing production of endangered Snake River
785 Sockeye Salmon. *Transactions of the American Fisheries Society*, 144:1, 107-123.

786 Waples, R. S. 1991. Pacific salmon and the definition of “species” under the Endangered Species
787 Act. *Marine Fisheries Review* 53:11-22.

788 Waples RS, Aebersold PB, Winans GA (1997) Population genetic structure and life history
789 variability in *Oncorhynchus nerka* from the Snake River Basin. Final Report of Research,
790 Bonneville Power Administration, Portland, 104 p

791 Waters, C. D., J. J. Hard, M. S. O. Briec, D. E. Fast, K. I. Warheit, R. S. Waples, C. M.
792 Knudsen, W. J. Bosch, and K. A. Naish. 2015. Effectiveness of managed gene flow in
793 reducing genetic divergence associated with captive breeding. *Evolutionary Applications*
794 8:956-971.

795 Williamson, K. S., A. R. Murdoch, T. N. Pearsons, E. J. Ward, and M. J. Ford. 2010. Factors
796 influencing the relative fitness of hatchery and wild spring Chinook salmon
797 (*Oncorhynchus tshawytscha*) in the Wenatchee River, Washington, USA. *Canadian*
798 *Journal of Fisheries and Aquatic Sciences* 67:1840-1851.

799 Williams, J.G., S. G. Smith, J. K. Fryer, M. D. Scheuerell, W. D. Muir, T. A. Flagg, R. W. Zabel,
800 J. W. Ferguson, and E. Casillas. 2014. Influence of ocean and freshwater conditions on
801 Columbia River Sockeye Salmon *Oncorhynchus nerka* adult return rates. *Fisheries*
802 *Oceanography* 23:2 210-224.

803 Withler, R. E., D. S. O’Brien, N. M. Watson, and K. J. Supernault. 2014. Maintenance of genetic
804 diversity in natural spawning of captive-reared endangered Sockeye Salmon,

805 *Oncorhynchus nerka*. Diversity 6:354-379.

806 Wurtsbaugh, W. A., H. P. Gross, C. Luecke and P. Budy. 1997. Nutrient limitation of
807 oligotrophic Sockeye Salmon lakes of Idaho (USA). Internationale Vereinigung für
808 theoretische und angewandte Limnologie Verhandlungen. 26:413-419.

809

810

811

812

813

814

815

816

817

818

819

820

821

822

823

824

825

826

827

828 Figure 1. Map of the upper Salmon River watershed and location of Redfish Lake in the
829 Sawtooth Valley basin in central Idaho. The trapping locations on Redfish Lake Creek and at the
830 Sawtooth Hatchery are presented along with the former location of Sunbeam Dam.

831 Figure 2. Estimated Sockeye Salmon potential egg deposition (PED) historically (Bjornn et al.
832 1968) and for current spawn years 2004-2011 and the number of smolts estimated as leaving
833 Redfish Lake.

834 Figure 3. Number of emigrating Sockeye Salmon smolts (right axis) resulting from captive and
835 anadromous adult releases (left axis) into Redfish Lake for volitional spawning.

836 Figure 4. Historic Sockeye Salmon smolt-to-adult survival as estimated by Bjornn et al. 1968
837 and current smolt-to-adult survival (SY 2004-2011) with 95% CI. *Year refers to the emigration
838 year for Bjornn et al. (1968) and the spawn year for which the fish were born for current data.

839 Figure 5. Diagram depicting the combinations of Sockeye Salmon freshwater productivity and
840 SARs which can result in population replacement. The current range of estimates observed
841 during SY 2004-2011 are presented for each year. The dark curved line represents population
842 replacement.

843

844

845

846

847

848

849

850
851
852
853
854
855
856
857
858
859
860
861
862
863
864
865
866
867
868

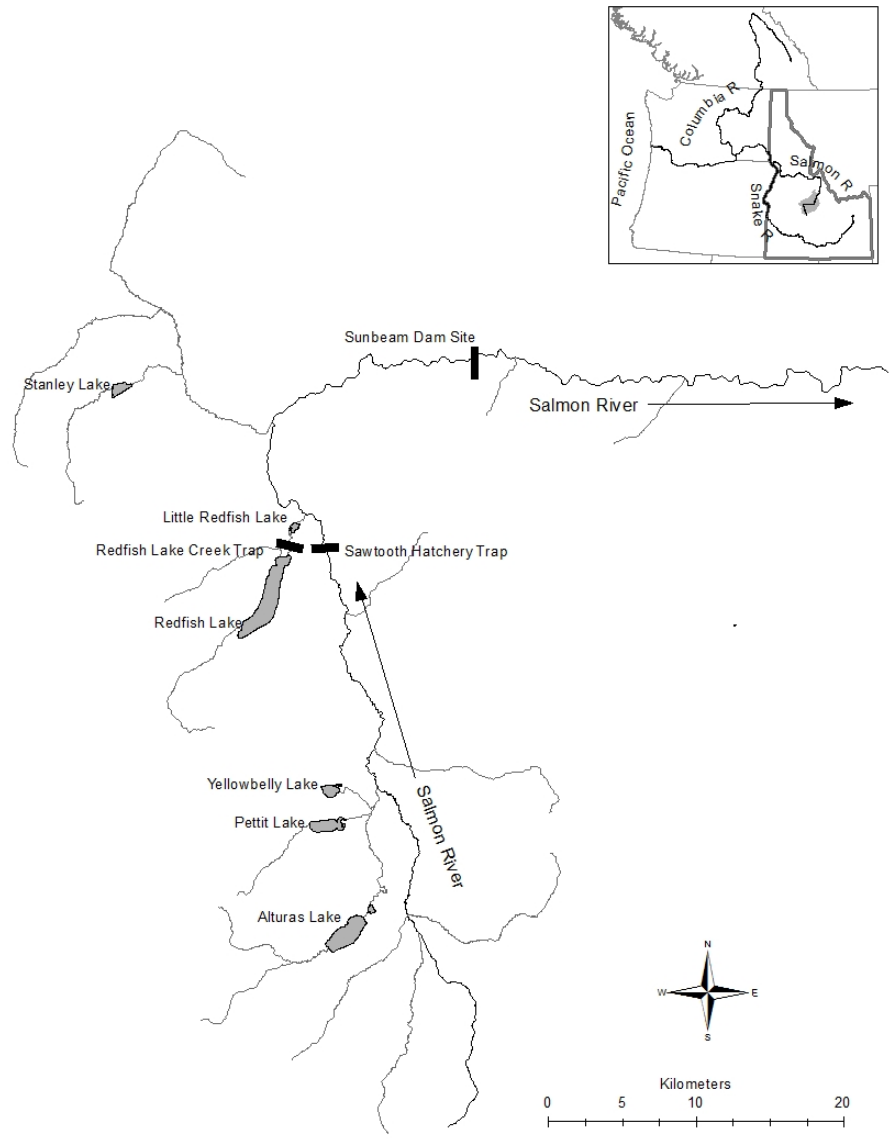
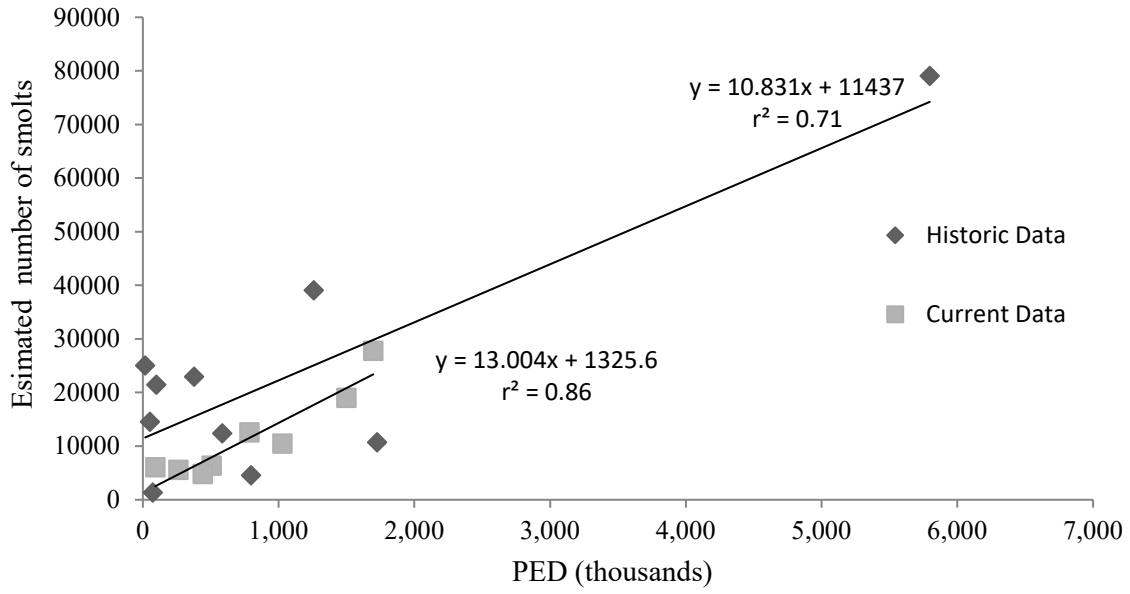
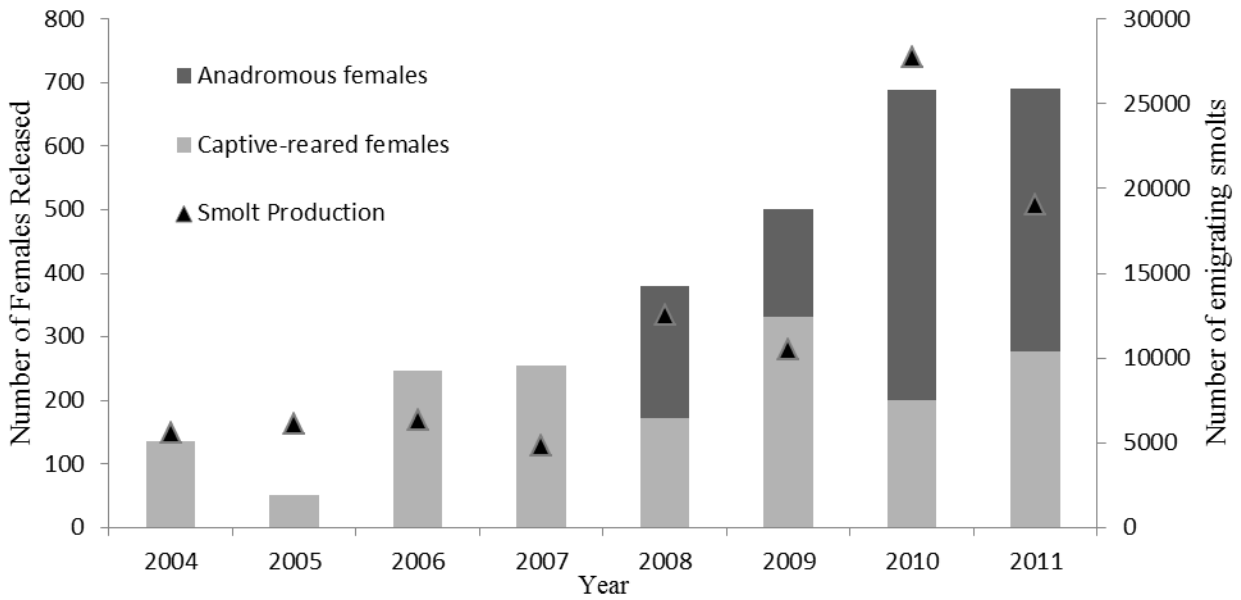


Figure 1

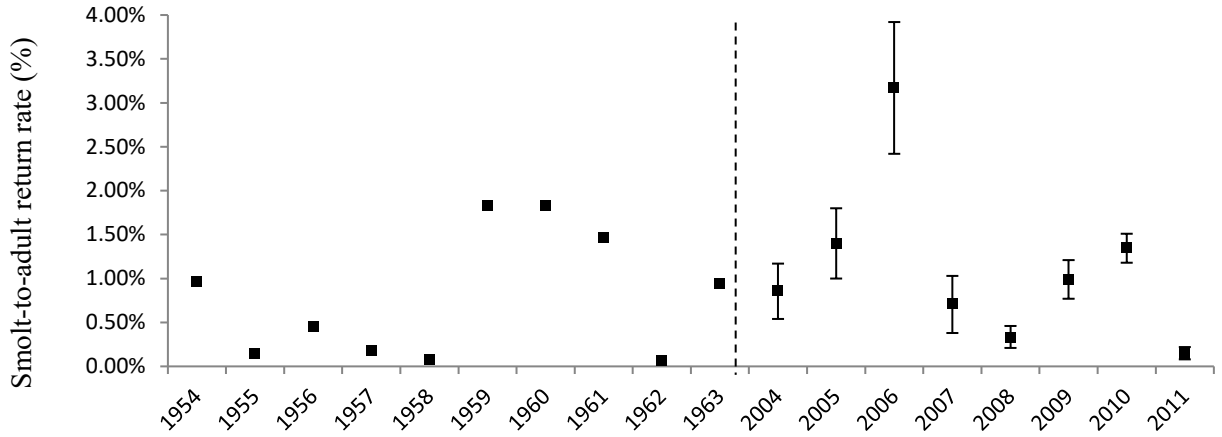


869
870 Figure 2

871



872
873 Figure 3



874
875 Figure 4

876

877

878

879

880

881

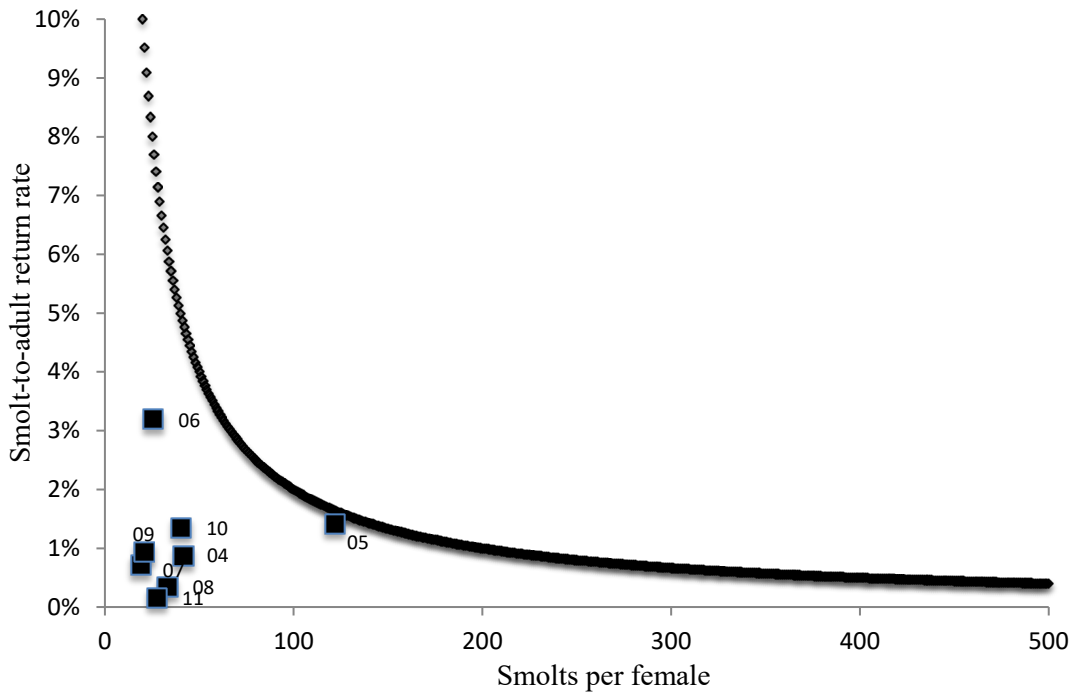
882

883

884

885

886



887 Figure 5

888

889

890

891

892

893 Table 1. Total number of Sockeye Salmon captive and anadromous adults by sex (females,
 894 males, and unknown) and rearing group (NMFS-FW/SW, NMFS-FW, IDFG-FW, Anadromous)
 895 released into Redfish Lake for volitional spawning from 2004 to 2011.

Spawn Year	Type	Females	Males	Unknown	Total
2004	NMFS-FW/SW	116	108	1	225
	NMFS-FW	19	0		19
2005	NMFS-FW/SW	20	60	3	83
	NMFS-FW	30	63		93
2006	NMFS-FW/SW	121	109		230
	NMFS-FW	126	109		235
2007	NMFS-FW/SW	96	144	1	241
	NMFS-FW	61	65		126
	IDFG-FW	97	34		131
2008	NMFS-FW/SW	61	74		135
	NMFS-FW	49	67		116
	IDFG-FW	62	82		144
	Anadromous	207	310	51*	568
2009	NMFS-FW/SW	9	89		98
	NMFS-FW	147	44		191
	IDFG-FW	175	216		391
	Anadromous	169	481		650
2010	NMFS-FW/SW	115	70	2	187
	NMFS-FW	10	0	1	11
	IDFG-FW	75	97		172
	Anadromous	488	719	1	1208
2011	NMFS-FW/SW	121	109		230
	NMFS-FW	0	0		0
	IDFG-FW	156	172		328
	Anadromous	414	574		988

896

897 *Estimated number of adults that passed through the Redfish Lake Creek adult trap and spawned in
 898 Redfish Lake.

899

900 Table 2. Results of linear regression modeling of fecundity on year and length and their
 901 interaction. AICc is Akaike's Information Criterion corrected for sample size. Delta AICc is the
 902 difference from the minimum AICc.

Rearing				
Strategy	Model	AICc	Delta AICc	r ²
NMFS-FW/SW	Length	N/A	N/A	0.68
NMFS-FW	Year + Length + (Year x Length)	12570.2	0.00	0.64
	Year + Length	12575.6	5.37	
	Year	13066.1	495.83	
	Length	12769.6	199.35	
IDFG-FW	Year + Length + (Year x Length)	23623.7	0.00	0.58
	Year + Length	23643	19.22	
	Year	24492.4	868.70	
	Length	23721	97.23	
Anadromous	Year + Length + (Year x Length)	7786.11	9.78	0.46
	Year + Length	7776.32	0.00	
	Year	8038.23	261.91	
	Length	7830.40	54.08	

903
 904
 905
 906
 907
 908
 909

910 Table 3. Natural productivity metrics resulting from releasing Sockeye Salmon adults to
 911 volitionally spawn within Redfish Lake, ID. Number of released females, potential egg
 912 deposition (PED) and smolt migration estimates are shown for each spawn year with standard
 913 error. Egg-to-smolt survival, smolts per female, and adult recruits are also presented as well as
 914 the arithmetic mean for each metric measured during the evaluation

Spawn Year	Female spawners	Estimated PED (SE)	Smolt migration (SE)	Egg-to-smolt survival (%)	Smolts per female	Adult Recruits
2004	135	262,101 (39,237)	5,609 (621)	2.14	41.54	48
2005	50	91,748 (10,800)	6,088 (489)	6.64	121.76	85
2006	247	506,640 (53,300)	6,338 (597)	1.25	25.69	201
2007	254	441,645 (45,852)	4,822 (654)	1.09	18.98	34
2008	379	785,577 (108,497)	12,588 (884)	1.60	33.13	42
2009	500	1,027,407 (93,732)	10,502 (475)	1.02	21.04	104
2010	688	1,697,192 (196,445)	27,765 (1,638)	1.64	40.35	374
2011	691	1,498,722 (171,411)	19,033 (795)	1.27	27.54	28
915 Mean		789,253	11,593	2.09	41.25	114

916 Table 4. Sockeye Salmon natural-origin smolt production from Redfish Lake. The total
 917 estimated abundance, proportion of age-1 and age-2 smolts, and smolt length (mm) with standard
 918 error is presented for each spawn year.

Spawn Year	Estimated smolts	Age-1 Percentage	Average length Age-1 (mm) (SE)	Age-2 Percentage	Average length Age-2 (mm) (SE)
2004	5,609	91%	96 (1.84)	9%	146 (2.27)
2005	6,088	78%	110 (2.10)	22%	125 (1.53)
2006	6,338	77%	98 (1.65)	23%	131 (2.04)
2007	4,822	65%	110 (2.19)	35%	131 (2.20)
2008	12,558	98%	106 (1.43)	2%	141 (2.77)
2009	10,502	63%	109 (1.83)	37%	140 (1.95)
2010	27,765	99%	102 (1.71)	1%	145 (2.15)
919 2011	19,033	96%	117 (1.71)	4%	140 (1.31)