National Status and Trends Program

for Marine Environmental Quality

# Magnitude and Extent of Sediment Toxicity in Four Bays of the Florida Panhandle: Pensacola, Choctawhatchee, St. Andrew and Apalachicola. 



Silver Spring, Maryland
October 1997

## US Department of Commerce <br> noa@ National Oceanic and Atmospheric Administration

Coastal Monitoring and Bioeffects Assessment Division Office of Ocean Resources Conservation and Assessment
National Ocean Service
National Oceanic and Atmospheric Administration
U.S. Department of Commerce

N/ORCA2, SSMC4
1305 East-West Highway
Silver Spring, MD 20910

## Notice

This report has been reviewed by the National Ocean Service of the National Oceanic and Atmospheric Administration (NOAA) and approved for publication. Such approval does not signify that the contents of this report necessarily represents the official position of NOAA or of the Government of the United States, nor does mention of trade names or commerical products constitute endorsement or recommendation for their use.

# Magnitude and Extent of Sediment Toxicity in Four Bays of the Florida Panhandle: Pensacola, Choctawhatchee, St. Andrew and Apalachicola. 

Edward R. Long<br>National Oceanic and Atmospheric Administration<br>Gail M. Sloane<br>Florida Department of Environmental Protection<br>R. Scott Carr, Tom Johnson, and James Biedenbach<br>National Biological Service<br>K. John Scott and Glen B. Thursby<br>Science Applications International Corporation<br>Eric Crecelius and Carole Peven<br>Battelle Ocean Sciences<br>Herbert L. Windom, Ralph D. Smith and B. Loganathon<br>Skidaway Institute of Technology



Silver Spring, Maryland
October 1997

United States
Department of Commerce
William M. Daley
Secretary

National Oceanic and Atmospheric Administration
D. James Baker Under Secretary

National Ocean Service
W. Stanley Wilson

Assistant Administrator

## Table of Contents

List of Figures ..... - iii
List of Tables ..... -ix
Abstract ..... $-1$
Executive Summary ..... $-1$
Introduction ..... 4
Methods ..... $-7$
Results ..... 15
Discussion ..... 30
Conclusions ..... 35
References ..... 37
Figures ..... 41
Tables ..... 103
Appendices ..... 103

## List of Figures

1. Study area encompassing Pensacola, Choctawhatchee, St. Andrew, and Apalachicola Bays ..... 42
2. Total PAH concentrations in sediments from 10 NOAA NS\&T Program Sampling Sites ..... 42
3. Total DDT concentrations in sediments from 10 NOAA NS\&T Program Sites ..... 43
4. Lead concentrations in sediments from 10 NOAA NS\&T Program Sites ..... 43
5. Concentrations of organics and trace metals in sediments from NS\&T Program Sites sampled in Pensacola Bay in 1991 ..... 44
6. Concentrations of organics and trace metals in sediments from NS\&T Program Sites sampled in Choctawhatchee Bay in 1991 ..... 45
7. Concentrations of organics and trace metals in sediments from NS\&T Program Sites sampled in St. Andrew Bay in 1991 ..... 46
8. Concentrations of organics and trace metals in sediments from NS\&T Program Sites sampled in Apalachicola Bay in 1991 ..... 47
9. Pensacola Bay Study Area ..... 48
10. Choctawhatchee Bay Study Area ..... 48
11. St. Andrew Bay Study Area ..... 49
12. Apalachicola Bay Study Area ..... 49
13. Locations of sampling stations in Pensacola Bay ..... 50
14. Locations of sampling stations in Bayou Chico ..... 50
15. Locations of sampling stations in Choctawhatchee Bay ..... 51
16. Locations of sampling stations in St. Andrew Bay ..... 51
17. Locations of sampling stations in Apalachicola Bay ..... 52
18. Maximum concentrations of major classes of organic compounds (total PAHs, total PCBs, total pesiticides and total DDTs) in sediments from four western Florida bays ..... 52
19. Maximum, mean and minimum concentrations of total PAHs in western Florida bays ..... 53
20. Maximum, mean and minimum concentrations of total PCBs, total pesticides and total DDTs in western Florida bays ..... 54
21. Maximum, mean and minimum lead concentrations in western Florida bays ..... 54
22. Maximum, mean and minimum mercury concentrations in western Florida bays ..... 55
23. Distribution of total PAH concentrations among selected sampling stations in Pensacola Bay ..... 55
24. Distribution of total PCB concentrations among selected sampling stations in Pensacola Bay ..... 56
25. Distribution of mercury concentrations among selected sampling stations in Pensacola Bay ..... 56
26. Distribution of lead concentrations among selected sampling stations in Pensacola Bay ..... 57
27. Distribution of total PAH concentrations among selected sampling stations in Bayou Chico ..... 57
28. Distribution of total PCB concentrations among selected sampling stations in Bayou Chico ..... 58
29. Distribution of mercury concentrations among selected sampling stations in Bayou Chico ..... 58
30. Distribution of lead concentrations among selected sampling stations in Bayou Chico ..... 59
31. Distribution of total PAH concentrations among selected sampling stations in Choctawhatchee Bay ..... 59
32. Distribution of total PCB concentrations among selected sampling stations in Choctawhatchee Bay ..... 60
33. Distribution of mercury concentrations among selected sampling stations in Choctawhatchee Bay ..... 60
34. Distribution of lead concentrations among selected sampling stations in Choctawhatchee Bay ..... 61
35. Distribution of total PAH concentrations among selected sampling stations in St. Andrew Bay ..... 61
36. Distribution of total PCB concentrations among selected sampling stations in St. Andrew Bay ..... 62
37. Distribution of mercury concentrations among selected sampling stations in St. Andrew Bay ..... 62
38. Distribution of lead concentrations among selected sampling stations in St. Andrew Bay ..... 63
39. Distribution of total PAH concentrations among selected sampling stations in Apalachicola Bay ..... 63
40. Distribution of total PCB concentrations among selected sampling stations in Apalachicola Bay ..... 64
41. Distribution of mercury concentrations among selected sampling stations in Apalachicola Bay ..... 64
42. Distribution of lead concentrations among selected sampling stations in Apalachicola Bay ..... 65
43a. Relationship between arsenic and aluminum in 1993 Pensacola Bay samples ..... 65
43b. Relationship between cadmium and aluminum in 1993 Pensacola Bay samples ..... 66
43c. Relationship between chromium and aluminum in 1993 Pensacola Bay samples ..... 66
43d. Relationship between copper and aluminum in 1993 Pensacola Bay samples ..... 67
43e. Relationship between lead and aluminum in 1993 Pensacola Bay samples ..... 67
43f. Relationship between zinc and aluminum in 1993 Pensacola Bay samples ..... 68
44a. Relationship between arsenic and aluminum in 1994 Bayou Chico samples ..... 68
44b. Relationship between cadmium and aluminum in 1994 Bayou Chico samples ..... 69
44c. Relationship between chromium and aluminum in 1994 Bayou Chico samples ..... 69
44d. Relationship between copper and aluminum in 1994 Bayou Chico samples ..... 70
44e. Relationship between lead and aluminum in 1994 Bayou Chico samples ..... 70
44f. Relationship between nickel and aluminum in 1994 Bayou Chico samples ..... 71
44 g . Relationship between zinc and aluminum in 1994 Bayou Chico samples ..... 71
45a. Relationship between arsenic and aluminum in Choctawhatchee Bay samples ..... 72
45b. Relationship between cadmium and aluminum in Choctawhatchee Bay samples ..... 72
45c. Relationship between chromium and aluminum in Choctawhatchee Bay samples ..... 73
45d. Relationship between copper and aluminum in Choctawhatchee Bay samples ..... 73
45e. Relationship between lead and aluminum in Choctawhatchee Bay samples ..... 74
45f. Relationship between zinc and aluminum in Choctawhatchee Bay samples ..... 74
46a. Relationship between arsenic and aluminum in St. Andrew Bay samples ..... 75
46b. Relationship between cadmium and aluminum in St. Andrew Bay samples ..... 75
46c. Relationship between chromium and aluminum in St. Andrew Bay samples ..... 76
46d. Relationship between copper and aluminum in St. Andrew Bay samples ..... 76
46e. Relationship between lead and aluminum in St. Andrew Bay samples ..... 77
46f. Relationship between zinc and aluminum in St. Andrew Bay samples ..... 77
47a. Relationship between arsenic and aluminum in Apalachicola Bay samples ..... 78
47b. Relationship between cadmium and aluminum in Apalachicola Bay samples ..... 78
47c. Relationship between chromium and aluminum in Apalachicola Bay samples ..... 79
47d. Relationship between copper and aluminum in Apalachicola Bay samples ..... 79
47e. Relationship between lead and aluminum in Apalachicola Bay samples ..... 80
47f. Relationship between zinc and aluminum in Apalachicola Bay samples ..... 80
43. Sampling stations in Pensacola Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 81
44. Sampling stations in Bayou Chico in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 81
45. Sampling stations in Choctawhatchee Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 82
46. Sampling stations in St. Andrew Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 82
47. Sampling stations in Apalachicola Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 83
48. Sampling stations in Pensacola Bay in which sediments were either not toxic in Microtox ${ }^{\text {TM }}$ Tests, significantly different from controls, or highly toxic ..... 83
49. Sampling stations in Pensacola Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 84
50. Sampling stations in Choctawhatchee Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 84
51. Sampling stations in St. Andrew Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 85
52. Sampling stations in Apalachicola Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic ..... 85
53. Sampling stations in Choctawhatchee Bay in which sediments were either not toxic, suspected genotoxic, or genotoxic in Mutatox tests ..... 86
54. Sampling stations in Apalachicola Bay in which sediments were either not toxic, suspected genotoxic, or genotoxic in Mutatox tests ..... 86
55. Sampling stations in Pensacola Bay in which sediments were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters ..... 87
56. Sampling stations in Bayou Chico that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters ..... 87
57. Sampling stations in Choctawhatchee Bay that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters ..... 88
58. Sampling stations in St. Andrew Bay that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters ..... 88
59. Sampling stations in Apalachicola Bay that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters ..... 89
60. Sampling stations in Pensacola Bay that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters ..... 89
61. Sampling stations in Bayou Chico that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters ..... 90
62. Sampling stations in Choctawhatchee Bay that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters ..... 90
63. Sampling stations in St. Andrew Bay that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters ..... 91
64. Sampling stations in Apalachicola Bay that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters ..... 91
65. Relationship between normal urchin embryo development and the concentrations of zinc in Pensacola Bay ..... 92
66. Relationship between normal urchin embryo development and the concentrations of high molecular weight PAHs in Pensacola Bay ..... 92
67. Relationship between Microtox EC50s and the concentrations of total DDT in Pensacola Bay ..... 93
68. Relationship between Microtox EC50s and the sum of 18 chemical concentration/ERM quotients in Pensacola Bay ..... 93
69. Relationship between urchin percent normal development and the concentration of un-ionized ammonia in porewater from Pensacola Bay ..... 94
70. Relationship between sea urchin fertilization and the concentrations of zinc in samples from Bayou Chico and Apalachicola Bay ..... 94
71. Relationship between Microtox EC50s and the concentrations of fluoranthene (ug/goc) in samples from Bayou Chico and Apalachicola Bay ..... 95
72. Relationship between sea urchin fertilization and concentrations of total PAHs in Bayou Chico and Apalachicola Bay ..... 95
73. Relationship between urchin embryo development and the sum of 25 chemical concentration/ERM quotients in Bayou Chico and Apalachicola Bay ..... 96
74. Relationship between urchin embryo development and concentrations of un-ionized ammonia in porewater in Bayou Chico and Apalachicola Bay ..... 96
75. Relationship between sea urchin fertilization and concentrations of total DDTs in Choctawhatchee Bay ..... 97
76. Relationship between urchin embryo development and concentrations of un-ionized ammonia in porewater in Choctawhatchee Bay ..... 97
77. Relationship between amphipod survival and concentrations of total DDT in St. Andrew Bay sediments ..... 98
78. Relationship between sea urchin normal development and concentrations of un-ionized ammonia in the porewater of St. Andrew Bay sediments ..... 98
79. Relationship between percent sea urchin fertilization and the concentrations of zinc in St. Andrew Bay sediments ..... 99
80. Relationship between sea urchin fertilization and the concentrations of un-ionized ammonia in porewater of sediments from St. Andrew Bay ..... 99
81. Relationship between microbial bioluminescence (Microtox EC50s) and the concentrations of trans-nonachlor in St. Andrew Bay sediments ..... 100
82. Relationship between the sum of 25 chemical concentrations to ERM quotients and microbial bioluminescence (Microtox EC50s) sediments ..... 100
83. Relationship between Microtox EC50s and the concentrations of total DDTs in western Florida samples ..... 101
84. Relationship between urchin fertilization and the concentrations of all PAHs in western Florida samples ..... 101
85. Relationship between urchin embryo normal development and the concentrations of un-ionized ammonia in western Florida samples ..... 102

## List of Tables

1. Coordinates of sampling stations in each bay ..... 103
2. Numbers of stations sampled and tested for toxicity and numbers of stations tested for chemistry in four western Florida bays ..... 105
3. Summary of data from amphipod toxicity tests performed in both years on all samples; listed as mean percent survival, statistical significance and percent of control for each station ..... 106
4. Results from Microtox ${ }^{\top M}$ tests for both years and from all stations in the four western Florida bays; expressed as mean EC50s (mg equivalents, wet wt.) and standard deviations, statistical significance, and percent of controls ..... 109
5. Results of Mutatox ${ }^{T M}$ tests, expressed as categorical responses. This test was performed on samples collected during 1994 ..... 112
6. Mean percent fertilization success of sea urchins in 100 percent, 50 percent, and 25 percent concentrations of sediment porewater ..... 114
7. Mean normal embryo development in 100 percent, 50 percent, and 25 percent of sediment porewater ..... 119
8. Spearmen-Rank correlations (Rho) among the four toxicity tests performed on the 1993 samples from Pensacola Bay ..... 124
9. Spearmen-Rank correlations (Rho) among the four toxicity tests performed on the 1994 samples from Bayou Chico and Apalachicola Bays ..... 124
10. Spearmen-Rank correlations (Rho) among the four toxicity tests from Choctawhatchee Bay ..... 124
11. Spearmen-Rank correlations (Rho) among the four different toxicity tests performed in St. Andrew Bay ..... 125
12. Percent of samples from each bay that were either toxic (i.e., significantly different from controls) or highly toxic (i.e., significantly different from control and $<80 \%$ of controls) ..... 126
13. Estimates of the spatial extent of toxicity in western Florida bays with results of four different tests ..... 127
14. Test results for Pensacola Bay ..... 129
15. Stations within Pensacola Bay in which chemical concentrations equalled or exceeded their respective sediment quality guidelines or toxicity thresholds ( $\mathrm{n}=20$ for organics, $\mathrm{n}=40$ for metals). Stations with twofold or greater exceedances are listed in bold ..... 132
16. A comparison of the chemical concentrations (ave. $\pm$ std. dev.) in toxic and nontoxic samples in Pensacola Bay in Microtox ${ }^{T M}$ tests, ratios between the two averages, and ratios between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb) ..... 133
17. A comparison of the chemical concentrations (ave. $\pm$ std. dev.) in toxic and nontoxic samples in Pensacola Bay in urchin fertilization tests, ratios between the two averages, and ratios between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb) ..... 134
18. A comparison of the chemical concentrations (ave. $\pm$ std. dev.) in toxic and nontoxic samples in Pensacola Bay in urchin development tests, ratios between the two averages, and ratios between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb) ..... 135
19. Summary of toxicity/chemistry relationships for samples from Pensacola Bay ..... 136
20. Test results for Bayou Chico and Apalachicola Bay ..... 137
21. Stations within Bayou Chico in which chemical concentrations equalled or exceeded their respective sediment quality guidelines or toxicity thresholds ( $n=6$ for organics, $n=6$ for metals) stations with twofold or greater exceedances are listed in bold ..... 142
22. A comparison of the chemical concentrations (ave. $\pm$ std. dev.) in toxic and nontoxic samplesfrom Bayou Chico and Apalachicola Bay in urchin fertilization tests, the ratio between the twoaverages, and the ratio between the toxic average and applicable numerical guidelines (metalsin ppm, organics in ppb)143
23. Test results for Choctawhatchee Bay ..... 144
24. Stations within Choctawhatchee Bay in which chemical concentrations equalled or exceeded their respective sediment quality guidelines or toxicity thresholds ( $\mathrm{n}=20$ for organics, $\mathrm{n}=20$ for metals). Stations with twofold or greater exceedance are listed in bold ..... 148
25. A comparison of the chemical concentrations (ave $\pm$ std dev) in toxic and nontoxic samples from Choctawhatchee Bay in urchin fertilization tests, the ratio between the two averages, and the ratio between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb) ..... 149
26. Test results for St. Andrew Bay ..... 150
27. Stations within St. Andrew Bay in which chemical concentrations equalled or exceeded theirrespective sediment quality guidelines or toxicity thresholds. Stations with a twofold or greaterexceedance listed in bold152

# Magnitude and Extent of Sediment Toxicity in Four Bays of the Florida Panhandle: Pensacola, Choctawhatchee, St. Andrew and Apalachicola. 

Edward R. Long (NOAA), Gail M. Sloane (FDEP), R. Scott Carr, Tom Johnson, and James Biedenbach (NBS), K. John Scott and Glen B. Thursby (SAIC), Eric Crecelius and Carole Peven (Battelle), Herbert L. Windom, Ralph D. Smith and B. Loganathon (Skidaway)


#### Abstract

The toxicity of sediments in Pensacola, Choctawhatchee, St. Andrew and Apalachicola Bays was determined as part of bioeffects assessments performed by NOAA's National Status and Trends Program. The objectives of the survey were to determine: (1) the spatial patterns in toxicity throughout each bay, (2) the spatial extent of toxicity throughout and among the bays, (3) the severity or degree of toxicity, and (4) the relationships between chemical contamination and toxicity. The survey was conducted over two years: Pensacola Bay and St. Andrew Bay were sampled in 1993; and Choctawhatchee Bay, Apalachicola Bay and Bayou Chico (a sub-basin of Pensacola Bay) were sampled during 1994.

Surficial sediment samples were collected from 123 randomly-chosen locations throughout the five areas. Multiple toxicity tests were conducted on all samples, and chemical analyses were performed on 102 of the 123 samples.

Toxicological tests were conducted to determine survival, reproductive success, morphological development, metabolic activity, and genotoxicity; all bays showed toxicity in at least some of the samples. Toxicity was most severe in Bayou Chico, an industrialized basin adjoining Pensacola Bay. Other developed bayous adjoining Pensacola Bay and the other bays also showed relatively severe toxicity. The main basins of the bays generally showed lower toxicity than the adjoining bayous. The different toxicity tests, however, indicated differences in severity, incidence, spatial patterns, and spatial extent in toxicity. The most sensitive test, a bioassay of metabolic activity of a bioluminescent bacteria, indicated toxicity was pervasive throughout the entire study area. The least sensitive test, an acute bioassay performed with a benthic amphipod, indicated toxicity was restricted to a very small portion of the area.

Causes of toxicity were not determined in the survey. However, mixtures of potentially toxic substances, including pesticides, petroleum constituents, trace metals, and ammonia, were associated statistically with the measures of toxicity. The concentrations of many substances were highest in Bayou Chico, where the most severe toxicity was observed. At these toxic sites, some of the substances had considerably elevated concentrations, often exceeding numerical guidelines or known toxicity thresholds. The relationships between toxicity and chemical concentrations differed among the bays and toxicity tests.


## EXECUTIVE SUMMARY

The National Status and Trends (NS\&T) Program, administered by the National Oceanic and Atmospheric Administration (NOAA), conducts a nationwide program of monitoring and bioeffects assessments. As a part of this program, regional surveys are conducted to determine the toxicity of sediments in estuarine and marine environments.

Toxicity in this survey of four western Florida Bays was determined using a suite of four laboratory tests done on all samples: (1) percent survival of marine amphipods (Ampelisca abdita) in 10-day tests of solid-phase (bulk) sediments; (2) changes in bioluminescent activity of a marine bacterium, Photobacterium phosphoreum, in 5minute Microtox ${ }^{\text {TM }}$ bioassays of organic extracts; (3) fertilization success of the sea urchin Arbacia punctulata in one hour tests of the sediment porewater; and (4) normal embryological development of A. punctulata in 48-hour tests of the porewater. In addition, the Mutatox ${ }^{\text {TM }}$ variant of the microbial bioluminescence tests were performed
on samples collected during the second year from Bayou Chico Bay, Choctawhatchee Bay and Apalachicola Bay. The concentrations of trace metals, pesticides, other chlorinated compounds, polynuclear aromatic hydrocarbons, and sedimentological features of the sediments were determined on 102 of the 123 total samples.

Based upon the chemistry data acquired as a part of this survey, concentrations of many trace metals, pesticides, and polynuclear aromatic hydrocarbons (PAHs) were considerably higher in the urbanized bayous of Pensacola, Choctawhatchee, and St. Andrew Bays than in the main basins of those systems or in Apalachicola Bay. Trace metals concentrations exceeded background levels as predicted by geochemical normalization using metal-to-aluminum ratios, suggesting problematic metals loadings from upland sources. Concentrations of zinc, numerous high and low molecular weight PAHs, dieldrin, and DDT isomers were particularly high in Bayou Chico. To a lesser degree, bulk chemistry results showed elevated values of several contaminants in Watsons Bayou, Bayou Texar, Garnier Bayou, Boggy Bayou, and Massalino Bayou. Therefore, these data suggested that toxicity would be most probable and most severe in Bayou Chico, somewhat less likely in the other urban bayous, and least likely in the main basins of all four bays.

Some of the tests showed agreement and concordance among test results, while others failed to show significant concordance. Therefore, different toxicity tests identified overlapping but generally different patterns in toxicity. The amphipod test data showed a general lack of toxicity. Only three stations throughout all four bays were significantly toxic in the amphipod test; one in Apalachicola Bay, one in Choctawhatchee Bay, and one in Bayou Chico Bay in the Pensacola Bay estuary. Amphipod survival was less than $80 \%$ of controls in only one sample, which was collected from Bayou Chico.

The data from the Microtox ${ }^{\text {TM }}$ tests indicated that the majority of the samples from the four bays were toxic; 114 of 123 samples were significantly different from controls. Toxicity in this test was pervasive, extending throughout most or all of each bay. Mean test results often were less than $10 \%$ of reference response levels. All of the samples from Choctawhatchee Bay, St. Andrew Bay, and Bayou Chico were significantly different from controls. All except one sample from Apalachicola Bay were toxic and all except eight samples from Pensacola Bay were toxic. Nontoxic samples came from an upstream station in the Apalachicola River, several stations near the mouth of and scattered throughout Pensacola Bay.

Results of the Mutatox ${ }^{\text {TM }}$ tests conducted in Year 2 revealed that 22 of 52 samples produced a strong genotoxic response (G category). An additional 11 stations produced suspect results. All stations tested in Bayou Chico provided a genotoxic response. In contrast, only one of the nine samples from Apalachicola Bay showed a genotoxic response and five of the samples showed no genotoxicity.

The sea urchin fertilization tests showed relatively high toxicity in several bayous of Choctawhatchee Bay, Watsons Bayou in St. Andrew Bay, and Bayou Chico of Pensacola Bay, as compared to the remainder of the study area. There was relatively low toxicity in most of the main basins of Pensacola and St. Andrew Bays. Most of the 1994 samples from Bayou Chico were highly toxic in all porewater concentrations, whereas none collected in 1993 was toxic in any porewater concentrations. Two samples each in the lower Apalachicola River and lower Apalachicola Bay were toxic in $100 \%$ porewater. Among the 123 samples tested, $38(31 \%)$ were significantly toxic in tests of $100 \%$ porewater.

In the urchin embryo development tests, there was a relatively high incidence of toxicity in Choctawhatchee Bay, Apalachicola Bay and Bayou Chico as compared to other areas in the study area, and a low incidence of toxicity in St. Andrew Bay. Stations exhibiting toxicity in the embryo development tests were largely associated with urbanized tributaries to the bays in all systems, except in Apalachicola Bay. Toxicity was especially apparent in Bayou Chico, Watson Bayou, and Pensacola Harbor in Pensacola Bay, Destin Harbor, portions of Garnier Bayou, and part of Choctawhatchee Bay. Among the 123 samples tested, a total of 46 ( $37 \%$ ) were toxic in at least $100 \%$ porewater, a slightly higher proportion than observed in the fertilization tests. Results of the embryo development tests agreed relatively well with those from the urchin fertilization tests.

Overall, the highest incidence of toxicity occurred among the Bayou Chico samples, followed by Choctawhatchee Bay, Apalachicola Bay, St. Andrew Bay, and Pensacola Bay. All of the Bayou Chico samples were toxic in the sea urchin development, Microtox ${ }^{\text {TM }}$, and Mutatox ${ }^{\text {TM }}$ tests performed in 1994; all were highly toxic in the urchin development and Mutatox ${ }^{\text {TM }}$ tests; and all except one sample were highly toxic in the urchin fertilization tests. In addition, the only sample that was highly toxic in the amphipod survival tests was collected in Bayou Chico. In

Choctawhatchee Bay, all samples were toxic in Microtox ${ }^{\text {TM }}$ tests, most were toxic in Mutatox ${ }^{\text {TM }}$ tests, $57 \%$ were toxic in urchin fertilization tests, $49 \%$ were toxic in urchin development tests, and one sample was toxic in the amphipod tests. The incidence of toxicity in Pensacola Bay and St. Andrew Bay was relatively similar: zero and one sample toxic in amphipod tests (respectively), $10 \%$ and $13 \%$ toxic in urchin fertilization tests, $27 \%$ and $23 \%$ toxic in urchin development tests, and $80 \%$ and $100 \%$ toxic in Microtox ${ }^{\text {TM }}$ tests.

These data suggest that amphipod survival was affected in a tiny portion ( $0.005 \%$ ) of the entire study area and microbial bioluminescence was affected in nearly all ( $98.9 \%$ ) of the area. Both sea urchin fertilization and embryo development were affected in nearly one-half of Choctawhatchee Bay, and small portions of St. Andrew and Pensacola bays. In Apalachicola Bay, the majority of the area was affected in sea urchin development tests, whereas about one-third was affected in the fertilization tests. Usually, small portions of each bay were affected in both tests of $50 \%$ and $25 \%$ porewater. Throughout the entire study area, $23 \%$ of the area was toxic in urchin fertilization tests and $34 \%$ was toxic in the urchin development tests. Usually, small portions of each bay were affected in both tests of $50 \%$ and $25 \%$ porewater.

The relationships between toxicity measured in the four bioassays and solid-phase (bulk) sediment chemistry were explored in a multistep approach. Microtox ${ }^{\text {TM }}$ test results were highly correlated with the concentrations of chlorinated compounds, including DDTs, PCBs, and total pesticides. To a lesser degree, Microtox ${ }^{\text {TM }}$ test results were significantly correlated with the concentrations of some trace metals and PAHs. Urchin fertilization was highly correlated with the concentrations of PAHs, numerous trace metals, and DDT. Urchin embryo development was primarily correlated with the concentrations of unionized ammonia, and to a lesser degree, PAHs, three trace metals, and the pesticide dieldrin. The results of the Microtox ${ }^{\top M}$ and urchin fertilization tests were significantly correlated with the sum of the 25 chemical concentrations normalized to (i.e., divided by) effectsbased numerical guidelines, since these quotients account for the contribution of 25 different substances to toxicity. The correlations strongly suggest that mixtures of toxicants, co-varying with each other, contributed significantly to the observed toxicity. This association, on the other hand, was not observed in the embryo development tests, in which ammonia was a major contributor to toxicity. The relationships between measures of toxicity and chemical concentrations differed considerably among the bioassays and among the four bays.

Based upon these analyses, it appeared that the concentrations of zinc, high molecular weight PAHs, two DDD/ DDT isomers, total DDT, and dieldrin were most closely associated with toxicity in Pensacola Bay. To a lesser extent, cadmium, copper, lead, low molecular weight PAHs, and in the case of urchin embryo development, unionized ammonia, were moderately associated with toxicity in Pensacola Bay. Spearman-rank correlations failed to show significant correlations between toxicity and chemical concentrations in samples from Bayou Chico and Apalachicola Bay. However, there were numerous obvious associations between elevated chemical levels and toxicity. Of note, samples from Bayou Chico had considerably higher chemical concentrations than those from Apalachicola Bay.

The associations between toxicity and concentrations of potentially toxic substances in Choctawhatchee Bay were strongest for the urchin fertilization tests in which a large gradient in response was observed. Most notable among these were the concentrations of DDT isomers, total DDT, silver, the sum of PAHs, and dieldrin.

The four toxicity endpoints measured in St. Andrew Bay appeared to co-vary with different substances in the samples. Despite significant correlations between amphipod survival and the concentrations of copper and DDT, none of the bioassay results were significantly different from controls. Sea urchin development was significantly correlated only with unionized ammonia in the porewater tests, and zero percent normal development occurred in some samples with relatively high ammonia concentrations. Urchin fertilization was correlated with a number of trace metals, DDT, and ammonia. Concentrations of some metals and DDT exceeded effectsbased numerical guideline values. Microtox ${ }^{\text {TM }}$ test results were highly correlated with complex mixtures of substances, including many trace metals and organic compounds. Additionally, Microtox ${ }^{\text {TM }}$ test results showed a strong association with the cumulative ERM quotients, again suggesting that microbial bioluminescence responded to complex mixtures of substances in the organic solvent extracts.

The data from this survey indicated that sediments in some regions of the area were contaminated relative to background conditions and effects-based numerical guidelines, that toxicity occurred throughout the entire region as measured in the most sensitive tests, that the most severe toxic responses and the highest incidences
of toxicity occurred in Bayou Chico, that the toxicity test results generally paralleled the concentrations of potentially toxic substances in the samples, and that different mixtures of toxicants were associated with toxicity.

## INTRODUCTION

Toxic chemicals can enter the marine environment through numerous routes: stormwater runoff, industrial point source discharges, municipal wastewater discharges, atmospheric deposition, accidental spills, illegal dumping, pesticide applications and agricultural practices. Once they enter a receiving system, toxicants can become bound to suspended particles and increase in density sufficiently to sink to the bottom. Sediments are one of the major repositories of contaminants in aquatic environments. Furthermore, if they become sufficiently contaminated, sediments can act as sources of toxicants to important biota. Sediment quality data are direct indicators of the health of coastal aquatic habitats.

Sediment quality investigations conducted by the National Oceanic and Atmospheric Administration (NOAA) and the Florida Department of Environmental Protection (FDEP) have indicated that toxic chemicals are found in the sediments and biota of Florida estuaries, including those of the western Florida panhandle (NOAA, 1992; Seal et al., 1994). This report documents the toxicity of sediments collected within four large bays of the western Florida panhandle: Pensacola, St. Andrew, Choctawhatchee, and Apalachicola (Figure 1).

As a component of its National Status and Trends (NS\&T) Program, NOAA monitors toxicant concentrations in selected locations throughout the nation and surveys the biological significance of toxicant accumulations in selected regions. In the monitoring component of the program, mollusks and demersal fishes are captured annually for chemical analyses of their tissues. Sediments are collected and analyzed for a suite of metals and organic parameters. Spatial patterns and temporal trends in chemical concentrations are determined from the data (O'Connor and Ehler, 1991; O'Connor, 1991). Chemical analyses of sediments collected at each sampling site were performed at many of the sites the first year that each site was sampled. Nationwide, NS\&T monitoring activities were initiated in 1983 and have continued each year to the present.

Thus far, sediment toxicity surveys have been performed by NOAA in San Francisco Bay (Long and Markel, 1992), Tampa Bay (Long et al., 1994), Long Island Sound (Wolfe et al., 1994), Hudson-Raritan estuary (Long et al., 1995), Boston Harbor (Long et al., 1996), Los Angeles/Long Beach Harbor (Sapudar et al., 1994), San Diego Bay (Fairey et al., 1996), and in several other areas in which the surveys are still underway.

Each year, numerous sites ( $50-70$ ) have been sampled along the Gulf of Mexico, including the bays of the western Florida panhandle (Sericano et al., 1990). During the first years of the program, sediments and oysters collected at sites in Choctawhatchee Bay and St. Andrew Bay contained extremely high concentrations of DDTs, PCBs, and non-DDT pesticides (Sericano et al., 1990), exceeding the concentrations of those substances in all other Gulf coast sites.

NS\&T Program data from estuaries nationwide for the period 1984 through 1989 were summarized by NOAA (1991). These data revealed that the concentrations of a number of chemicals were significantly elevated in sediments, warranting further in-depth investigation. In ranking estuaries nationwide with respect to sediment contamination, the NS\&T Program ranked Choctawhatchee Bay $1^{\text {st }}$ (highest) in the nation for total chlorinated pesticides, $3^{\text {rd }}$ highest for total DDT, $6^{\text {th }}$ highest in lead, $6^{\text {th }}$ highest for total PAHs, and $4^{\text {th }}$ highest for arsenic. St. Andrew Bay ranked $19^{\text {th }}$ highest in the nation with respect to total chlorinated pesticides, $9^{\text {th }}$ highest for total DDTs, $2^{\text {nd }}$ highest for total PCB's, $3^{\text {rd }}$ highest for total PAHs, and $6^{\text {th }}$ for TOC.

The concentrations of total polynuclear aromatic compounds (PAHs), total DDTs, and lead in fine-grained sediments collected at each of the ten historic NS\&T Program monitoring sites in western Florida are compared in Figures 2-4. These samples accompanied collections of either fish or oysters and were intended to represent "average" conditions within major basins of each bay. They were not selected to represent highly contaminated areas within each bay. They may or may not represent conditions throughout each bay.

Stations PCMP and SAWB in St. Andrew Bay had the highest PAH concentrations. PAH concentrations were relatively low at three stations sampled in Apalachicola Bay, two locations in Pensacola Bay, station CBSR in Choctawhatchee Bay, and station PCLO in St. Andrew Bay. The concentrations of total DDTs showed a pattern very different from that of total PAHs (Figure 3). Elevated concentrations were found only at one location; station

CBPP in Choctawhatchee Bay. Another pattern in concentrations was evident with lead (Figure 4). Lead concentrations were elevated at station CBPP, intermediate at two locations in St. Andrew Bay (stations PCMP and SAWB), and moderate to relatively low in several other locations. The concentrations of total PAHs, total DDTs, and lead were uniformly relatively low in the Apalachicola Bay stations.

Concentrations of major classes of organic compounds and each trace element are compared among the NS\&T Program stations collected during 1991 in Figures 5-8. In Pensacola Bay, concentrations of PAHs, chromium, copper, lead, nickel, and zinc were relatively high at station PEN as compared to station PBIB (Figure 5). In Choctawhatchee Bay, PAH, DDT, and lead (Pb) concentrations were higher at station CBPP than at station CBSR (Figure 6). Among the three locations sampled in St. Andrew Bay, stations PCMP and SAWB had relatively high concentrations of several classes of organic compounds, chromium, copper, lead, and zinc (Figure 7). In Apalachicola Bay, the concentrations of both high molecular weight PAHs and total PAHs were relatively high at station APDB compared to the others (Figure 8). All other organic compounds occurred at relatively low concentrations. Among the trace elements, chromium and zinc occurred in relatively high concentrations at all three stations (Figure 8). The concentrations of each trace metal were similar among the three stations.

The Florida Department of Environmental Protection investigated coastal sediment quality from 1982 through 1991 throughout the state (Seal et al., 1994). The objectives of their investigations were several: to provide a statewide (spatial) perspective on the presence of contaminants in coastal systems, to investigate areas of special interest (ports projects) and to determine ways to interpret sediment quality data.

The FDEP produced an atlas that summarized sediment chemistry data from both the FDEP and NOAA NS\&T Program investigations in bay systems throughout the state (Seal et al., 1994). Elevated levels of both metals and organic contaminants were detected in samples collected statewide, including those from the four estuaries in northwest Florida. Metals contamination within the studied estuaries was usually within a factor of 5 times the expected background level, based on normalization to aluminum (Seal et. al., 1994, Schropp and Windom, 1988). PAHs were the most frequently detected organic contaminant, which followed a statewide trend (Seal et al, 1994). Other organic contaminants of concern included pesticides and PCBs. Brief synopses of conditions within each of the four bays are provided in the following discussion.

Pensacola Bay. Four major water bodies make up the Pensacola Bay estuarine system: Escambia Bay, Pensacola Bay, Blackwater Bay and East Bay (Figure 9). Together these waterbodies form one of the largest estuarine systems of Florida, encompassing over 152 square miles of estuarine surface water area (Seal et al., 1994). The estimated drainage basin covers 3,480 square miles of Florida and Alabama. Average water depth of the estuary is 19 feet (NOAA, 1985). Principle tributaries of the estuary include the Escambia, Blackwater, Yellow and East Bay Rivers.

The western portion of the Pensacola Bay estuary is predominantly urban, whereas the eastern portion is relatively undeveloped. The urban center of the city of Pensacola contains extensive industrial, commercial and residential development. Industrial facilities are located along Bayou Chico, the Escambia River, and near Escambia Bay. Other land uses within the watershed include urban development, military holdings, silviculture, agriculture, conservation and recreation (Seal et al., 1994, Paulic et al. 1994).

The Pensacola Bay estuary is included in the Surface Water Improvement and Management (SWIM) program of the State of Florida, overseen by the Florida Department of Environmental Protection and administered by the Northwest Florida Water Management. The SWIM plan prepared for the Pensacola Bay estuary reported that there has been an extensive loss of aquatic grass beds and widespread fluctuations in fish and shellfish harvests in both Escambia Bay and East Bay (Northwest Florida WMD, 1990).

FDEP sediment studies detected PAHs and PCBs in most stations located throughout the bay, and in NOAA NS\&T Program stations in Pensacola Bay and Indian Bayou showed elevated PAH concentrations. Sediments in the western portion of the estuary exceeded natural background levels of metals, especially in Bayou Chico and Bayou Grande (Seal et. al., 1994).

Choctawhatchee Bay. The Choctawhatchee Bay estuary has a surface area of 123 sq. miles with an estimated drainage area of 2259 square miles of Florida and Alabama (Seal et al., 1994) (Figure 10). Average depth of the estuary is about 22 feet (NOAA, 1985), however the bay is deepest to the west, and shallows to the east. The

Choctawhatchee River entering the bay on the far eastern shore is the most significant freshwater tributary. Several small tributary streams enter the Bay along the north shore (Seal et al., 1994).

Development in the basin along the north and west has been sparse. Eglin Air Force Base occupies most of the immediate northern drainage basin. The city of Fort Walton Beach is on the eastern shore, and the town of Destin is situated at East Pass, the mouth of the bay. Much of the peninsula that forms the southern boundary of the bay has been developed for single-family dwellings, hotels, and condominiums (Paulic et al., 1994). This coastal peninsula was severely impacted by Hurricane Opal in October of 1995.

Investigations of sediment quality throughout Choctawhatchee Bay have shown minimal enrichment by metals bay wide (Seal et al, 1994), however concentrations of several metals were elevated in Destin Harbor, and concentrations of lead were elevated near Boggy Point at the confluence of Boggy Bayou and the main basin of the Bay. Organic contaminants were elevated at several locations, including: PAH's in Destin Harbor (Seal et al, 1994); and PAH's, PCB's, and total DDT at the some of highest levels in the nation at both the Santa Rosa and Boggy Point NS\&T Program locations (NOAA, 1991).

St. Andrew Bay. The St. Andrew Bay estuarine system has a surface area of 98 square miles. The drainage basin of 1130 square miles is entirely within Florida. The average depth of the bay is 27 feet. Several embayments distinguish the St. Andrew Bay estuarine system (Figure 11 ): West Bay, North Bay, East Bay and the main basin of St. Andrew Bay. The drainage area includes the urban centers of Panama City, Lynn Haven, and Panama City Beach, however, the majority of the watershed is forested and in silviculture (Paulic et. al., 1994).

Elevated concentrations of metals and organic contaminants have been documented in the bay (FDEP, 1994; U.S. FWS, 1995; NOAA, 1991). The main source of elevated trace metals appears to be stormwater runoff from urbanized areas in and around Panama City (Seal, 1994). PAH's, PCB's and pesticides were detected at all NS\&T Program sites (NOAA, 1994).

An evaluation of dredged material from five stations in St. Andrew Bay, prepared by Battelle Ocean Sciences (1993) for the Army Corps of Engineers, showed that, in general, sediments from stations located in the bay had higher concentrations of the majority of contaminants of concern relative to reference sediments. Metals data were plotted using the FDEP metals normalization procedure, and were rarely found to be elevated above natural background levels. Sediments from most sites were nontoxic in multiple biological assays; sediments from only one station were toxic to the amphipod Rhepoxynius abronius. However, significant bioaccumulation of PAHs, pesticides and PCBs was measured in clams (Macoma nasuta) relative to reference sediment concentrations (Mayhew et.al, 1993), indicating that toxicants were bioavailable.

Apalachicola Bay. The Apalachicola Basin encompasses nearly 200 square miles of estuary, including St. Vincent Sound, East Bay, Apalachicola Bay and St. George Sound (Figure 12). The major freshwater inflow to the bay is the Apalachicola River, which originates at Lake Seminole, the impounded confluence of the Chattahoochee and Flint Rivers, formed by the Jim Woodruff dam at Chattahoochee, Florida. Headwaters for this alluvial river system originate in the Blue Ridge physiographic province. Annual flow into the bay is 25,000 cfs, varying seasonally from less than 15,000 to greater than 100,00 cfs. The estuary is highly productive, providing over $90 \%$ of the Florida oyster harvest, as well as supporting commercial fin fishing and other shellfish industries (Seal et al, 1994).

The Apalachicola estuary is unusual in that this system is composed of a river delta developing behind a sequence of barrier islands. This geomorphic situation will eventually eliminate the estuary by sediment infilling (Donoghue 1988). Finer grained materials are associated with higher burdens of contaminants; it follows that there exists good potential for contaminants to become trapped in the system in the finer grained depositional areas.

No metals enrichment had been detected by the DEP in the estuary, and only one sample out of three from NOAA's NS\&T Program surveys indicated slight enrichment with mercury. PAH's, PCB's and pesticides were detected in Apalachicola Bay, but they occurred in relatively low concentrations.

Current Survey Rationale. Several factors lead to the decision to conduct a survey of sediment bioeffects in this area. First, very high concentrations of organic compounds (DDT, PCB) had been found by the NOAA NS\&T Program in sediments and biota of Choctawhatchee and St. Andrew bays, bringing attention to the area from a
nationwide perspective. Second, the state of Florida wanted to further explore the potential for biological effects in the marine environment and field-validate recently drafted statewide sediment quality guidelines. Previously collected data were compared to effects-based numerical guidelines (Long and Morgan,1990; MacDonald, 1993, 1994). Sufficient numbers of sediment samples equaled or exceeded these values to warrant concern that sediments may pose a toxicological risk to resident biota. Third, the bays of the western Florida panhandle support abundant populations of living marine resources that could be at risk from toxicant contamination. Collectively, these factors lead to the decision to determine if contamination of the sediments in the western Florida panhandle was sufficiently high to warrant concern for resident biota.

This document reports the results of a survey of sediment toxicity in four selected bays of the western Florida panhandle. Sediments were collected throughout each of the bays over a two-year period to determine if there was an effect on biota based on the use of a battery of laboratory toxicity tests.
The objectives of the survey were to:
(1) determine the presence and severity of toxic responses,
(2) estimate the spatial extent of toxicity,
(3) identify spatial patterns of toxicity in each system, and
(4) characterize the relationships between toxicity and potential toxicants.

Sampling and testing methods used in previous surveys performed elsewhere in the USA were employed in this survey. A wide variety of candidate measures of toxicant effects were evaluated and compared to determine which would be most useful in NOAA's surveys (Wolfe, 1992; Long and Buchman, 1989). Batteries of assays performed with sediments, bivalve mollusks, and demersal fishes in selected regions have been used to form a weight of evidence with regard to the presence and incidence of toxicant-associated bioeffects. Analyses of sediment toxicity have been included in these regional assessments to provide an estimate of potential effects of sediment contaminants on resident benthic populations. Batteries of toxicity tests appropriate for analyzing sediment toxicity were selected following evaluations of a number of candidates (Long and Buchman, 1989).

Based upon sediment chemistry data from previous studies, toxicity was most probable in portions of Pensacola Bay (especially in the adjoining urban/industrial bayous), St. Andrew Bay, and Choctawhatchee Bay. Toxicity was least probable in Apalachicola Bay.

## METHODS

Sampling Design. The four estuaries were investigated during 1993 and 1994; Pensacola and St. Andrew bays were sampled during 1993, and Choctawhatchee and Apalachicola bays were sampled in 1994. Samples were collected in Bayou Chico, an embayment contiguous to Pensacola Bay, during both years.

The study area included saltwater portions of these four coastal bays. Stratified, random sampling designs patterned after those of the EMAP-Estuaries surveys (Schimmel et al., 1994) were used in each bay during the selection of sampling stations. Each bay was subdivided into irregular-shaped strata. Large strata were established in the open waters of the bays where toxicant concentrations were expected to be uniformly low. This approach provided the least intense sampling effort in areas known or suspected to be relatively homogenous in sediment type and water depth, and relatively distant from contaminant sources. In contrast, relatively small strata were established in urban harbors and bayous nearer suspected sources in which conditions were expected to be heterogeneous or transitional. Sampling effort was more intense in the small strata than in the large strata. The large strata were roughly equivalent in size as were the small strata.

This approach combines the strengths of a stratified design with the random-probabilistic selection of sampling locations. Data generated within each stratum can be attributed to the dimensions of the stratum. Therefore, these data can be used to estimate the spatial extent of toxicity with a quantifiable degree of confidence (Heimbuch, et al., 1995). Strata boundaries were established to coincide with the dimensions of major basins, bayous, waterways, etc. in which hydrographic, bathymetric and sedimentological conditions were expected to be relatively homogeneous.

The locations of individual sampling stations within each strata were chosen randomly using the EMAP computer software and hardware (Dr. Kevin Summers, U.S. EPA, Environmental Research Laboratory, Gulf Breeze, FL). One to three samples were collected within each stratum. Usually, four alternate locations were provided for
each station in a numbered sequence. The coordinates for each alternate were provided in tables and were plotted on the appropriate navigation chart. In a few cases the coordinates provided were inaccessible. They were rejected and the vessel was moved to the next alternate.

Sample Collection. At each station the sampling vessel was piloted to the first alternate location for the sample collection. If the station was inaccessible or if the material at the location was only coarse sand with no mud component, that alternate location was abandoned and the second (third, or fourth, if needed) alternate was sampled. In almost all cases the first or second alternates were acceptable and were sampled.

Vessel positioning and navigation were aided with a Trimble NavGraphic XL Global Positioning System (GPS) unit and a compensated LORAN C unit. Both systems generally agreed very well with each other when both were operational. Both were calibrated and their accuracy verified each morning at a known location within the study area.

Samples were collected with a Kynar-lined $0.1 \mathrm{~m}^{2}$ modified van Veen grab sampler (also known as a Young grab) deployed with an electric windless aboard the state of Florida R/V Raja. The grab sampler and sampling utensils were acid washed with $10 \% \mathrm{HCl}$ at the beginning of each survey, and thoroughly cleaned with site water and acetone before each sample collection. Usually 3 or 4 deployments of the sampler were required to provide a sufficient volume of material for the toxicity tests and chemical analyses. The upper $2-3 \mathrm{~cm}$. of the sediment were sampled to ensure the collection of recently arrived materials. Sediments were removed with a plastic scoop and accumulated in a stainless steel pot. The pot was covered with a Teflon plate between deployments of the sampler to minimize sample oxidation and exposure to shipboard contamination. The material was carefully homogenized in the field with a stainless steel spoon before it was distributed to prepared containers for each analysis.

Samples were shipped in ice chests packed with water ice or blue ice to the testing laboratories by overnight courier. Samples were accompanied by chain of custody forms which included the date and time of the sample collection, and station designation.

Locations of the individual sampling stations for each bay are illustrated in Figures 13-17, and coordinates for each are listed in Table 1. Forty samples were collected in 1993 throughout all the major basins of Pensacola Bay and in Bayou Grande, Bayou Texar, Bayou Chico and the mouth of the Escambia River (Figure 13). Twelve samples were collected in 1993/94 in Bayou Chico where toxicity was most probable (Figure 14). Samples ( $\mathrm{n}=36$ ) were collected throughout Choctawhatchee Bay, a very large system, and many adjoining bayous (Figure 15). Thirty-one samples were collected throughout St. Andrew Bay, near Panama City (Figure 16). In Apalachicola Bay, nine samples were collected throughout the bay and up the Apalachicola River (Figure 17). Field log notes containing information on depth and sediment characteristics at each station are listed in Appendix A.

Multiple toxicity tests were performed on all sediment samples. Chemical analyses were performed on a subset of samples from each bay system for trace metals, butyl tins, polynuclear aromatic hydrocarbons, chlorinated pesticides and PCBs following a review and evaluation of the toxicity test results. Because the study encompassed two years, different contract laboratories performed chemistry analyses and amphipod toxicity tests.

Amphipod Survival Test. The amphipod tests are the most widely and frequently used assays in sediment evaluations performed in North America. They are performed with adult crustaceans exposed to relatively unaltered, bulk sediments. Ampelisca abdita has shown relatively little sensitivity to nuisance factors such as grain size, ammonia, and organic carbon in previous surveys. In previous surveys, the NS\&T Program has observed wide ranges in responses among samples, strong statistical associations with toxicants, and small within-sample variability (Long et al., 1994; Wolfe et al., 1994; Long et al., 1995).

Ampelisca abdita is a euryhaline benthic amphipod that ranges from Newfoundland to south-central Florida, and along the eastern Gulf of Mexico. The amphipod test with $A$. abdita has been routinely used for sediment toxicity tests in support of numerous EPA programs, including EMAP in the Virginian, Louisianian, and Carolinian provinces (Schimmel et al., 1994). Amphipod toxicity tests followed ASTM protocols (ASTM, 1990, 1992). In the first year, amphipod tests of samples from Pensacola Bay and St. Andrew Bay were conducted by the National Biological Service (NBS, now USGS) laboratory in Port Aransas, Texas. In the second year, amphipod assays were conducted by Science Applications International Corporation, (SAIC) in Narragansett, R.I.

In Year 1, test animals were purchased from Brezina and Associates of Dillon Beach, California. Amphipods were packed in native sediment with $8-10$ liters of seawater in doubled plastic bags. Oxygen was injected into the bags and shipped via overnight courier to the testing lab at Port Aransas. Upon arrival, amphipods were acclimated and maintained at $20 \infty \mathrm{C}$ for one day prior to the initiation of the test.

Control sediments for Year 1 testing included sediment collected from the natural habitat of the amphipods in California, and a reference sediment from Redfish Bay, Texas. The Redfish Bay sediments had been used in previous sediment quality assessment studies by SAIC and by the U.S. Fish \& Wildlife Service (NBS, then USGS). Both reference sediments were handled in the same manner as the Pensacola Bay and St. Andrew Bay sediments.

For Year 2 testing, amphipods were collected by SAIC from tidal flats in the Pettaquamscutt (Narrow) River, a small estuary flowing into Narragansett Bay, Rhode Island. Animals were held in the laboratory in pre-sieved uncontaminated ("home") sediments under static conditions. Fifty percent of the water in the holding containers was replaced every second day when the amphipods were fed. During holding, A. abdita were fed laboratory cultured diatoms (Phaeodactylum tricornutum).

Control sediments were collected by SAIC from the Central Long Island Sound (CLIS) reference station of the U.S Army Corps of Engineers, New England Division. These sediments have been tested repeatedly with the amphipod survival test and other assays and found to be nontoxic (amphipod survival has exceeded $90 \%$ in $85 \%$ of the tests) and uncontaminated (Wolfe et al., 1994; Long et al., 1995). Sub-samples of the CLIS sediments were tested along with each series of samples from Pensacola, Choctawhatchee and Apalachicola bays.

Amphipod testing performed by both laboratories followed the procedures detailed in the Standard Guide for conducting 10 day Static Sediment Toxicity Tests with Marine and Estuarine Amphipods (ASTM, 1990, 1992). Briefly, amphipods were exposed to test and negative control sediments for 10 days with 5 replicates of 20 animals each under static conditions using filtered seawater. For the Year 1 (NBS) test, 250 milliliters (mls) of test or control sediments were delivered into 1 -liter glass jars, and 700 mls of seawater were added to each jar. The Year 2 SAIC procedure differed somewhat: 200 mls of test or control sediments were placed in the bottom of the test chamber and covered with approximately 600 mls . of filtered seawater ( $28-30 \mathrm{ppt}$ ). For both sets of tests, air was provided by air pumps and delivered into the water column through a pipette to ensure acceptable oxygen concentrations, but suspended in a manner to ensure that the sediments would not be disturbed. Temperature was maintained at $\sim 20 \infty$ C by either incubator (NBS) or water bath (SAIC). Lighting was continuous during the 10 day exposure period to inhibit the swimming behavior of the amphipods. Constant light inhibits emergence of the organisms from the sediment, thereby maximizing the amphipod's exposure to the test sediments.

Twenty healthy, active animals were placed into each test chamber, and monitored to ensure they burrowed into sediments. Non-burrowing animals were replaced, and the test initiated. The jars were checked daily, and records kept for dead animals and animals on the water surface, emerged on the sediment surface, or in the water column. Those on the water surface were gently freed from the surface film to enable them to burrow, and dead amphipods were removed.

Tests were terminated after ten days. Contents of each of the test chambers were sieved through a 0.5 mm mesh screen. The animals and any other material retained on the screen were examined under a stereomicroscope for the presence of amphipods. Total amphipod mortality was recorded for each test replicate.

During Year 1 the NBS laboratory had to terminate the first test trial due to poor amphipod survival in the controls. Major storm events in the San Francisco Bay area may have led to poor viability of test animals. The tests were repeated within 20 days of collection of test sediments. Animal quality and survival improved for the repeated test, although they were not optimal. The second test batch was initiated within 10 days of receipt of field collected sediments.

A positive control, or reference toxicant test, was used to document the sensitivity of each batch of test organisms. The positive control consisted of 96 hr. water-only exposures to sodium dodecyl sulfate (SDS). LC50 values were calculated for each test run.

Sea Urchin Fertilization and Embryological Development Tests. Tests of sea urchin fertilization and embryo development have been used in assessments of ambient water and effluents and in previous NS\&T Program surveys of sediment toxicity (Long et al., 1994). Test results have shown wide ranges in responses among test samples, excellent within-sample homogeneity, and strong associations with the concentrations of toxicants in the sediments. The tests, performed with the early life stages of sea urchins, have demonstrated high sensitivity.

In previous surveys, the tests of embryological development have shown higher sensitivity than tests of fertilization success and the two endpoints have shown relatively poor correlations with each other (Long, et al., 1990; Carr, 1993; NBS, 1994; Carr et al., in press). It appears that these two endpoints respond to different toxic substances in complex mixtures (Long, et al., 1990; Carr, 1993; NBS, 1994; Carr et al., in press).

Toxicity of sediment pore waters were conducted with the sea urchin Arbacia punctulata. These tests were performed during both years by the National Biological Service (NBS), National Fisheries Contaminant Research Center in Corpus Christi, Texas at their laboratory located in Port Aransas. Sea urchins used in this study were obtained either from jetties at Port Aransas, Texas, or from Gulf Specimen Company, Inc. (Panacea, Florida), and were acclimated to Port Aransas seawater before gametes were collected for testing.

Pore water was extracted from sediments with a pressurized squeeze extraction device (Carr and Chapman, 1992). Sediment samples were held refrigerated at $4^{\circ} \mathrm{C}$ until pore water was extracted. Pore water was extracted as soon as possible after receipt of the samples, but in no event were sediments held longer than 7 days from the time of collection before they were processed. After extraction, porewater samples were centrifuged in polycarbonate bottles (at 4200 g for 15 minutes in Year 1, and in Year 2, using a new centrifuge where 1200 g for 15 minutes was adequate) to remove any particulate matter, and then frozen. Two days before the start of a toxicity test, samples were moved from a freezer to a refrigerator at $4^{\circ} \mathrm{C}$, and one day prior to testing, thawed in a tepid water bath. Experiments performed by NBS have demonstrated no effects upon toxicity attributable to freezing of the pore water samples.

Sample temperatures were maintained at $20 \pm 1^{\circ} \mathrm{C}$. Sample salinity was measured and adjusted to $30 \pm 1 \mathrm{ppt}$, if necessary, using ultrapure sterile water or concentrated brine. Other water quality measurements were made for dissolved oxygen, pH , sulfide and total ammonia. Temperature and dissolved oxygen were measured with YSI meters; salinity was measured with Reichert or American Optical refractometers; and pH , sulfide and total ammonia (expressed as total ammonia nitrogen, TAN) were measured with Orion meters and their respective probes. The concentrations of unionized ammonia (UAN) were calculated using respective TAN, salinity, temperature, and pH values.

Each of the porewater samples was tested in a dilution series of $100 \%, 50 \%$, and $25 \%$ of the water quality adjusted sample with 5 replicates per treatment. Dilutions were made with clean, filtered ( 0.45 um ), Port Aransas laboratory seawater.

Tests followed the methods of Carr and Chapman (1992). Pore water from a reference area in Redfish Bay, Texas, an area located near the testing facility and in which sediment porewaters have been determined to be nontoxic in this test (e. g., Long et al., 1994), was included with each toxicity test as a negative (nontoxic) control. Adult male and female urchins were stimulated to spawn with a mild electric shock, and gametes collected separately.

For the sea urchin fertilization test, $50 u \mathrm{~L}$ of appropriately diluted sperm were added to each vial, and incubated at $20 \pm 2^{\circ} \mathrm{C}$ for 30 minutes. One ml of a well mixed dilute egg suspension was added to each vial, and incubated an additional 30 minutes at $20 \pm 2^{\circ} \mathrm{C}$. Two mls of a $10 \%$ solution of buffered formalin solution was added to stop the test. Fertilization membranes were counted, and fertilization percentages calculated for each replicate test.

For the sea urchin embryological development test, a well mixed dilute egg solution was added to each vial. Then, $50 u \mathrm{~L}$ of appropriately diluted sperm were added to each vial, and vials were incubated at $20 \pm 1^{\circ} \mathrm{C}$ for 48 hours. At the end of 48 hours, 2 mls of $10 \%$ buffered formalin were added to each vial to stop the test. One hundred embryos were counted, and recorded as normal, or as unfertilized, embryological development arrested or otherwise abnormal. The percent of the embryos that were normal was reported for each replicate test.

Microbial Bioluminescence Microtox ${ }^{\text {TM }}$ Test. This is a test of the relative toxicity of extracts of the sediments prepared with an organic solvent, and, therefore, it is immune to the effects of nuisance environmental factors, such as grain size, ammonia and organic carbon. Organic toxicants, and to a lesser degree trace metals, that may or may not be readily bioavailable were extracted with the organic solvent. Therefore, this test can be considered as a test of potential toxicity. In previous NS\&T Program surveys, the results of Microtox ${ }^{\text {TM }}$ tests have shown extremely high correlations with the concentrations of mixtures of organic compounds (Long et al., 1994; Long et al., 1995; Wolfe et al., 1994).

The Microtox ${ }^{\text {TM }}$ assay was performed with dichloromethane (DCM) extracts of sediments following the basic procedures used in testing Puget Sound sediments (U.S. EPA, 1986, 1990, 1994) and San Francisco Bay sediments (Long and Markel, 1992). Organic extracts were prepared by ABC laboratories, Inc. of Columbia, Missouri. The extractions and transfers were conducted under a laminar flow hood to limit exposure of the sample to light. All sediment samples and extracts were stored in the dark at $4 \infty \mathrm{C}$. Prior to initial homogenization of the sediment samples, any excess water was decanted and large debris (shells, pebbles, etc.) was discarded. Each sediment sample was centrifuged for five minutes at $1000 \times g$. Water was removed by decanting with a Pasteur pipette. Moisture content of each sample was determined and recorded. Five g. of sediment were weighed, recorded, and placed into a DCM rinsed 50 mL centrifuge tube. Residue or spectral grade DCM ( 30 ml ) was added and mixed. The mixture was shaken for 10 seconds, vented and tumbled overnight.

Each sample was centrifuged for 5 minutes at $1000 \times g$, and the extract poured into a Kuderna-Danish (KD) flask. Again, 30 ml DCM was added to each centrifuge tube. Each tube was shaken for 10 sec , vented, centrifuged for 5 min at 1000 xg , and the extract transferred to the K-D flask. This extraction procedure was repeated once more, and the extract combined in the flask.

A Snyder column was attached to the flask, and the DCM was concentrated with steam to a final volume of less than 2 mls . Acetone (approximately 5 mls ) was added to the flask, and the volume concentrated to approximately 2 mls . This acetone procedure was repeated. The extract was quantitatively transferred to a 10 ml DCM rinsed flask, using acetone to rinse the Kuderna-Danish flask. The extract was concentrated with a gentle stream of nitrogen gas to a volume of approximately 1 ml . Dimethylsulfoxide (DMSO) was added to make a final volume of 10 ml .

A suspension of luminescent bacteria, Photobacterium phosphoreum, (Microbics Corporation, Inc.) was thawed and hydrated with toxicant-free distilled water, covered and stored in a $4^{\circ} \mathrm{C}$ well on the Microtox ${ }^{\mathrm{TM}}$ analyzer. To determine toxicity, each sample was diluted into four test concentrations. Percent decrease in luminescence of each cuvette relative to the reagent blank was calculated. Based upon these data, the sediment concentrations that caused a 50\% decrease in light production (EC50's) were reported.

A negative control (extraction blank) was prepared using DMSO, the test carrier solvent. A positive control was prepared by adding phenol ( $50 \mathrm{mg} / \mathrm{ml}$ ) to an extraction blank. In addition to test sediment extracts, a sediment sample from Florissant, Missouri was used as a procedural control.

Microbial Bioluminescence Mutatox ${ }^{\text {TM }}$ Test. Samples collected during Year 2 (1994) from Bayou Chico, Pensacola Bay, Choctawhatchee Bay and Appalachicola Bay were additionally tested for toxicity with the Mutatox ${ }^{\top 1}$ assay. The Mutatox ${ }^{\text {TM }}$ assay used a dark mutant strain of the luminescent bacterium P. phosphoreum for detection of environmental genotoxins. DNA-damaging substances were detected by measuring the ability of a test extract to restore the luminescent capability in the bacterial cells (Johnson, 1992, 1994). The degree of light increase indicated the relative genotoxicity of the sample.

A 100 ml aliquot of Mutatox ${ }^{\text {TM }}$ Assay Medium (MAM) was prepared in a 250 ml beaker using a magnetic stirrer and adjusted to $25 \pm 1 \infty \mathrm{C}$. Rat hepatic S9 was thawed at $25 \infty \mathrm{C}$. An ampoule of freeze-dried bacteria was hydrated with 1.0 ml of MAM at $25 \infty \mathrm{C}$. MAM was immediately inoculated with the hydrated bacteria at a ratio of 100:1, stirred, and pre-incubated for 15 minutes at $25 \infty$ C. Rat hepatic $S 9$ was introduced into the MAM mixture at $1 \%$, stirred for 5 seconds and dispensed into sample cuvettes. Sediment extracts were tested in a dilution series. The treated cuvettes were covered with foil, preincubated in a water bath at $37 \infty \mathrm{C}$ for 15 minutes, transferred to a closed storage container, and placed in a dark incubator with a vented hood at $25 \pm 1 \infty$ C for 16 to 24 hours.

Three controls were used in the Mutatox ${ }^{\text {TM }}$ assay: positive, negative and procedural. The positive controls consisted of four known progenotoxins. An environmental control from a previous study was used in some batches to monitor the sensitivity of the assay. The negative control was the carrier solvent; this control identified the spontaneous light emission of the dark mutant strain. The procedural control was an organic extract of a fine silt and clay particle sediment from Florissant, MO.

The response of the luminescent bacteria was determined by measuring the light intensity of each cuvette with a model 500 analyzer (Microbics Corp.). A light value of 100 or more and at least three times the intensity of the negative control was defined as a "genotoxic response"; sensitivity limits were the maximum peak concentration and the lowest detected concentration in each dilution series. The "dose response number" was defined as the number of genotoxic responses recorded at different concentrations per dilution series. A dilution series with a dose response number of 3 or more was considered genotoxic, with a dose response number less than 3, the series was considered suspect, and without a genotoxic response, the series was designated negative. Therefore, the sample was considered genotoxic when replicate series indicated an average dose response number of three or more, suspect when at least three replicate series indicated an average dose response number less than three, or negative when at least three replicate series contained no genotoxic response. Replicate dilution series were conducted on different days before each test sample was finally evaluated.

Chemical Analyses. Not all samples collected were analyzed for bulk chemical content. Sediment samples were chosen for chemical analyses based upon an examination of the toxicity test results. Samples were chosen that represented gradients in the toxicity results and furthermore represented contiguous geographic strings of stations. In Year 1, chemical analyses of Pensacola Bay sediments were performed by Skidaway Institute of Oceanography, Savannah, Georgia. Battelle Ocean Sciences analyzed sediments collected from St. Andrew Bay in Year 1, and Choctawhatchee and Apalachicola Bays in Year 2. Both laboratories conformed with perfor-mance-based analytical protocols and employed quality-assurance steps of the NS\&T Program (Lauenstein and Cantillo, 1993).

During the 1993 studies of Pensacola Bay, Skidaway Institute of Oceanography received samples directly from the field. Upon receipt of the samples at the laboratory, sediments were frozen until selection for analysis. Total digestion was performed for the trace metal analyses with nitric, perchloric and hydrofluoric acids. Following digestion, samples were analyzed for lithium, aluminum, iron, manganese, cadmium, copper, chromium, nickel, lead, zinc, silver, arsenic, vanadium, barium, titanium, and total phosphorus by inductively coupled plasma mass spectrometry (ICP-MS). Mercury was quantified by ICP-MS (isotope dilution) methods (Smith, 1993). Total organic carbon and nitrogen were analyzed on a carbonate free basis, using a Perkin Elmer model 240C elemental analyzer. Total carbonate was determined from the loss in weight in acidified samples.

Procedures used in the analyses of organic compounds followed the basic methods of MacLeod et al (1985). For the polynuclear aromatic hydrocarbon (PAH) analyses, 50 g . of wet sediment was sequentially extracted with methanol, $1: 1$ methanol $-\mathrm{CH}_{2} \mathrm{Cl}_{2}$ and $\mathrm{CH}_{2} \mathrm{Cl}_{2}$. The organic phase was concentrated to several ml. and stored refrigerated until fractionation with column chromatography. The extracts were fractionated on columns of silica gel over alumina packed over activated copper to remove elemental sulfur. Aliphatic hydrocarbons were eluted with hexane (fraction SA1), while aromatic hydrocarbons/ PCBs/ pesticides were eluted with $1: 1$ pentane: $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ (fraction SA2). Further separation of the SA2 fraction was accomplished by Sephadex LH-20 chromatography. PAHs were quantified by capillary gas chromatography-mass spectrometry utilizing full scan and selected ion monitoring modes. PCB congeners and chlorinated pesticides were separated using silica gel column chromatography.

Pesticides and PCBs were quantified by high-resolution fused silica capillary gas chromatography (GC) with electron capture detection on a Varian 3400cx and Varian 8200 Auto sampler instruments. DB-5 capillary column ( 60 m length, $0-25 \mathrm{~mm}$ i.d. and 0.25 micron film thickness) was used to resolve the PCB congeners and pesticides. Pesticides were identified and quantified by comparison to authentic pesticide standards. PCBs could not be quantified after a high level of cleanup, due to the presence of interfering compounds, identified later as nitro aromatic compounds. Butlyltins were analyzed with a Hewett-Packard 5890 GC interfaced with a Finnigan Incos mass spectrometer operated in the electron impact mode.

Chemical analyses were performed according to the quality control/quality assurance procedures of the NS\&T Program, including instrument calibration, use of internal standards, replication of some analyses, percent recoveries of spiked blanks, and analyses of standard reference materials.

During the remaining portions of the study, samples stored either at Skidaway Institute of Oceanography or at the NBS laboratory were shipped to Battelle Ocean Sciences for analyses. Sediments were extracted by Battelle Ocean Sciences in two batches containing approximately 19 field samples each. One procedural blank, one standard reference material, a matrix spike sample and a matrix spike duplicate sample were extracted with each batch. Each field sample contained 30 g to 50 g of sediment. Sediment dry weight was determined using approximately 5 g of sample material. Analyses were performed for total trace metals, simultaneously-extracted metals (SEM), acid-volatile sulfides (AVS), PCB congeners, pesticides, and polynuclear aromatic hydrocarbons (PAHs). Analyses were performed for total organic carbon and sediment grain size.

Extraction and analytical methods followed those of Peven and Uhler (1993). Sediment was weighed into preweighed Teflon jars; surrogate internal standards (to monitor extraction efficiency), sodium sulfate, and 1:1 methylene chloride (DCM):acetone were added to each jar. Samples were extracted with the solvent mixture three times using shaker table techniques. After each extraction, the jar was centrifuged and the overlying solvent decanted into a labeled Erlenmeyer flask. Solvent from each of the three extractions was combined in the flask. The combined extract was chromatographed through a $5 \mathrm{~g}, 2 \%$ deactivated alumina column eluted with dichloromethane (DCM). After column cleanup, the sample extract was concentrated to approximately 900 $u L$ and further processed using a size-exclusion high performance liquid chromatography (HPLC) procedure. Six-hundred microliters of the extract were fractionated in this procedure, and the remaining 300 uL archived. After HPLC cleanup, the sample extract was concentrated to approximately 1000 uL and recovery internal standards were added to quantify surrogate recovery. The final sample was split in half by volume; one half was dedicated to GC/MS analysis of PAHs and the other half was solvent-exchanged with iso-octane and analyzed by GC/ECD for PCBs and pesticides.

The analytical methods for the trace metals followed those of Crecelius et al. (1993). Samples were completely digested with $4: 1 \mathrm{HNO}_{3} / \mathrm{HClO}_{4}$ and heated. The digestates were analyzed either by graphite furnace atomic absorption (Ag, Cd, Se) or cold vapor atomic absorption (Hg) or x-ray fluorescence (Al, As, Cr, Cu, Fe, Mn, Ni, Zn ) or inductively-coupled plasma mass spectrometry ( $\mathrm{Sb}, \mathrm{Sn}$ ). Two reagent blanks and three standard reference materials were analyzed in each analytical string of 50 samples.

The concentrations of acid volatile sulfides (AVS) and simultaneously-extracted metals (SEM) were determined in the samples. The analytical methods employed selective generation of hydrogen sulfide by acidifying the sample with 1 N HCl , cryogenic trapping of the evolved $\mathrm{H}_{2} \mathrm{~S}$, and gas chromatographic separation with photoionization detection. This method gives high sensitivity, low detection limits and very limited chemical interference with minimal sample handling. The AVS analytical system is made of glass and Teflon because of the reactivity of sulfide with metals. The filtered acid solution resulting from the AVS analysis was subsequently analyzed for SEM using graphite furnace atomic absorption, cold-vapor atomic absorption, and inductively-coupled/mass spectrometry.

Sediment samples were analyzed for total organic carbon (TOC) and total carbonate (TIC) by Global Geochemistry Corporation, Canoga Park, California. Before the samples were analyzed, LECO filtration crucibles were precombusted for at least 2 hours at $450^{\circ} \mathrm{C}$ and allowed to cool. Between approximately 175 mg and 250 mg of dried, finely ground and homogenized sample was placed in a pretreated crucible and 6 N HCl added to remove inorganic carbon. After approximately 1 hour deionized water was flushed through the crucible removing the acid, and the sample was dried overnight. Immediately prior to sample analysis, iron and copper chips were added to accelerate the combustion. A LECO model 761-100 carbon analyzer was used to determine both the TOC and TIC content. The analyzer converts all carbon in the sample to $\mathrm{CO}_{2}$ at high temperature in the presence of oxygen. The $\mathrm{CO}_{2}$ was then quantified by thermal conductivity detection. Before sample analysis for TIC, the filtration crucibles were precombusted for at least 2 hours at $450^{\circ} \mathrm{C}$ and allowed to cool. Between approximately 175 mg and 250 mg of dried, finely ground, homogenized sample was placed in a pretreated crucible, and the sample was placed in a $450^{\circ} \mathrm{C}$ oven for 2 hours to remove organic carbon.

The methods used to determine sediment grain size are those according to Folk (1974). Briefly, coarse and fine fractions were separated by wet-sieving. The fine fractions (silt and clay) were further separated by suspending
the sediment in a deflocculant solution and taking aliquots of the settling sediment at timed intervals after the solution was thoroughly mixed. The coarse fraction (sand and gravel) was dried and then separated by sieving through a 2 mm screen.

## Statistical Methods.

Amphipod test. Percent amphipod survival data from each station that had a mean survival less than that of the control was compared to the control using a one-way, unpaired $t$-test (alpha $=0.05$ ) assuming unequal variance. Data were not transformed since examination of data from previous tests have shown that $A$. abdita percentage survival data met the requirement for normality. A one-sample $t$-test was used to compare data from each sampling block with the mean performance control (CLIS) for each stratum.

Significant toxicity for $A$. abdita is defined here as survival statistically less than that in the performance control (alpha $=0.05$ ). In addition, samples in which survival was significantly less than controls and less than $80 \%$ of control values were regarded as "highly toxic" or "numerically significant". The $80 \%$ criterion is used by EPA as a critical statistical value for A. abdita test data in EMAP-Estuaries methods (Holland, 1990). Similarly, the EPA/ COE dredged material guidance manual (the "green book") also consider sediments toxic if survival relative to a reference sediment is less than $80 \%$ (U.S. EPA/U.S. ACOE, 1990). Furthermore, statistical power curves created from SAIC's extensive testing database with $A$. abdita show that the power to detect a $20 \%$ difference from the control is $90 \%$.

Microtox ${ }^{\text {TM }}$. Microtox ${ }^{\text {TM }}$ data were analyzed using the computer software package developed by Microbics Corporation to determine concentrations of the extract that inhibit luminescence by $50 \%$ (EC50). This value was then converted to mg dry wt. using the calculated dry weight of sediment present in the original extract. To determine significant differences of samples from each station, pair-wise comparisons were made between contaminated samples and results from control sediments using analysis of covariance (ANCOVA). Concentrations tested were expressed as mg dry wt. based on the percentage extract in the 1 ml exposure volume and the calculated dry wt. of the extracted sediment. Both the concentration and response data were log-transformed before the analysis. ANCOVA was used first to determine if two lines had equal slopes (alpha $=0.05$ ). If the slopes were equal, ANCOVA then determined the quality of the $Y$-intercepts (alpha $=0.05$ ). A one-sample $t$-test was used to compare data from each sampling block within each of the bays with the mean of the duplicate performance control data.

Significant toxicity for Microtox ${ }^{\text {TM }}$ is defined here as an EC50 statistically less than that in the performance control. Samples were considered highly toxic or numerically significant when the EC50s were significantly different from controls and less than $80 \%$ of the controls. The statistical significance of the $80 \%$ criterion has not been determined for this test, however, the $80 \%$ criterion was used to be consistent with the other toxicity tests. Also, in surveys performed in the Hudson-Raritan estuary and Long Island Sound, $81.4 \%$ and $90.0 \%$, respectively, of samples were significantly different from controls (alpha <0.05) when EC50 values were less than $80 \%$ of control responses (Long et al., 1995; Wolfe et al., 1994).

Sea urchin fertilization and morphological development. For both the sea urchin fertilization and morphological development tests, statistical comparisons among treatments were made using ANOVA and Dunnett's onetailed $t$-test (which controls the experiment-wise error rate) on the arcsine square root transformed data with the aid of SAS (SAS, 1989). The trimmed Spearman-Karber method (Hamilton et al., 1977) with Abbott's correction (Morgan, 1992) was used to calculate EC50 ( $50 \%$ effective concentration) values for dilution series tests. Prior to statistical analyses, the transformed data sets were screened for outliers (SAS, 1992). Outliers were detected by comparing the studentized residuals to a critical value from a t-distribution chosen using a Bonferroni-type adjustment. The adjustment is based on the number of observations $(n)$ so that the overall probability of a type 1 error is at most $5 \%$. The critical value (CV) is given by the following equation: $\mathrm{cv}=\mathrm{t}$ (dfError, $.05 /(2 \times n)$ ). After omitting outliers but prior to further analyses, the transformed data sets were tested for normality and for homogeneity of variance using SAS/LAB Software (SAS, 1992).

Spatial patterns in toxicity Spatial patterns in toxicity were estimated by plotting data on base maps of each bay. Estimates of the spatial extent of toxicity were determined with cumulative distribution functions in which the toxicity results from each station were weighted to the dimensions $\left(\mathrm{km}^{2}\right)$ of the sampling stratum in which the samples were collected (Schimmel et al., 1994). The size of each stratum ( $\mathrm{km}^{2}$ ) was determined by use of a planimeter applied to navigation charts, upon which the boundaries of each stratum were outlined. A critical
value of less than $80 \%$ of control response was used in the calculations of the spatial extent of toxicity for all tests.

Chemistry data. Similarly, chemical data from the sample analyses were plotted on base maps to identify spatial patterns, if any, in concentrations. Trace metal concentrations were plotted against aluminum concentrations and compared to expected ratios for uncontaminated sediments developed by Schropp et al., 1988.

Chemistry/toxicity relationships. Chemistry/toxicity relationships were determined in a five-step sequence (Long et al., 1995). First, simple Spearman-rank correlations were determined for each toxicity test and each physical/ chemical variable. The correlation coefficients and their statistical significance were recorded and compared among chemicals. Second, for those chemicals in which a significant correlation was observed, the data were examined in scatterplots to determine if there was a reasonable pattern of increasing toxicity with increasing chemical concentration and if any chemical in the toxic samples equaled or exceeded published numerical guidelines.

Chemical concentrations expressed in dry wt. were compared with the ERM values of Long et al. (1995) developed for NOAA and the PEL values of MacDonald (1994) developed for the state of Florida. Also, the concentrations of three PAHs (acenaphthene, fluoranthene, and phenanthrene) and two pesticides (dieldrin and endrin) expressed in units of organic carbon were compared to proposed National sediment quality criteria (SQCs) developed by U.S. EPA (1994). Finally, the concentrations of unionized ammonia were compared to LOEC concentrations determined for the sea urchin tests by the U.S. National Biological Service (Long et al., in press - Boston Harbor) and NOEC concentrations determined for amphipod survival tests published by Kohn et al. (1994).

Third, the numbers of samples out of those that were analyzed that exceeded the respective guidelines were determined. Fourth, the average concentrations of chemicals in nontoxic samples were compared with the average concentrations in significantly toxic samples, and ratios between the two averages were calculated and compared. In this step, the ratios of chemical concentrations in toxic samples to respective ERL/ERM values from Long et al. (1995) and TEL/PEL values from MacDonald (1994) were also determined. Finally, the average concentrations of chemicals in the toxic samples were compared with the respective numerical guidelines. The combined results of these steps were examined to determine which chemical(s), if any, may have contributed to the observed toxicity and which probably had a minor or no role in toxicity.

The data were treated separately for each bay. However, in the case of Bayou Chico, the 1994 data were merged with those from Apalachicola Bay since the data sets for both bays were too small to analyze alone and these two areas showed relatively high and relatively low toxicity, respectively. In addition, the toxicity/chemistry correlations were determined for the entire combined data set formed by merging the data from all the bays.

Correlations were determined for all the substances that were quantified in each bay, usually including total (bulk) trace metals, metalloids, trace metals simultaneously extracted (SEM) with acid volatile sulfides (AVS), unionized ammonia (UAN), percent fines, total organic carbon (TOC), chlorinated organic hydrocarbons (COHs), and polynuclear aromatic hydrocarbons (PAHs). In addition, a chemical index calculated as the sums of quotients formed by dividing the chemical concentrations in the samples by their respective ERM values (from Long et al., 1995) are shown. Those substances that showed significant correlations were indicated with asterisks. In correlation analyses involving a large number of variables, some correlations could appear to be significant by random chance alone. Adjustments are often needed to account for this possibility. Note that in the results tables only those correlations shown with two or three asterisks would remain significant if the number of variables were taken into account in these analyses.

## RESULTS

Information recorded in field sample logs for station coordinates, sampling dates, water depths, and water column salinity, temperature, and dissolved oxygen concentrations are listed in Appendix A for each bay.

## Physical Parameters.

Pensacola Bay. In Pensacola Bay, water depths at the sampling stations ranged from 1.2 through 10.0 meters, and the temperature, salinity, and dissolved oxygen concentrations indicated the water column at most stations
were relatively well mixed (Appendix A1). However, dissolved oxygen concentrations in some Blackwater Bay stations were very low and surface salinities were zero. Bottom water dissolved oxygen concentrations were low near the head of Bayou Chico in 1994, but not in 1993. Most sediments in Pensacola Bay were fine silts and clays, often sulfurous and odiforous, occasionally with petroleum sheens.

Choctawhatchee Bay. In Choctawhatchee Bay most sampling stations were 2 to 6 meters deep (Appendix A2). Bottom and surface temperatures were similar at most stations, however, in the Boggy Bayou/Rocky Bayou/ Tom's Bayou area surficial salinities were very low, indicative of freshwater inputs. Except for a few stations in small bayous, bottom water dissolved oxygen concentrations were relatively high at most stations. Sediments typically were olivine-colored silty clays, often accompanied with numerous benthic organisms, occasionally with sulfurous odors. Sediments in Tom's Bayou were noticeably enriched with organic matter.

St. Andrews Bay. Most stations in St. Andrews Bay were 2-6 meters deep, except for a few stations that were up to 12 meters deep (Appendix A3). All but station 55 at the head of Watsons Bayous showed good water column mixing. Bottom water dissolved oxygen at station 55 was noticeably depressed. Sediments usually were silty clays, often either grey or olivine in color, and usually with numerous benthic organisms. Sediments at some stations in Watsons Bayou were anoxic and had a sulfurous odor.

Apalachicola Bay. Apalachicola Bay stations ranged in depth from 1.6 to 5.4 meters (Appendix A4). Salinities were relatively low, especially in the stations located in Apalachicola River; however, dissolved oxygen concentrations were high at all stations. Most stations in the bay had brown to olivine sandy mud whereas several stations in the river had silty sand or silty clay with some sand.

Distribution and Concentrations of Chemical Contaminants. Most of the samples collected and testing for toxicity were also analyzed for chemical concentrations. During both years of the study, 123 samples were tested for toxicity; 101 of which were tested for chemistry. This division of analytical tests for each bay is summarized in Table 2.

All of the 40 samples from Pensacola Bay were analyzed for metals, and one-half (20) were analyzed for organics. All six of the 1994 Bayou Chico samples were analyzed for all substances. In 1994, 21 of the 37 samples from Choctawhatchee Bay were analyzed for chemistry. In St. Andrew Bay, organics were analyzed for all samples, and trace metals were analyzed on 22 samples. Three of the nine samples from Apalachicola Bay were analyzed for both organic and metals parameters. When sufficient funding was not available to perform analyses on all samples, a subset was chosen following a review of the data from all toxicity tests. These samples were not chosen randomly; rather, they were selected to represent toxicity gradients within selected regions of each bay or among contiguous stations. Raw chemical data from all bays and both years are listed in Appendix B. Distributions of representative substances among and within the four bays are illustrated in Figures 18-42. Note that the concentration scales differ among these figures.

Inter-bay comparisons. The maximum concentrations of total PAHs, total PCBs, total pesticides, and total DDTs are compared among the four bays in Figure 18. The concentrations of the PAHs were considerably higher than those of the other substances, obscuring between-bay patterns in concentrations. The maximum concentration of total PAHs was highest at a station in Pensacola Bay, intermediate in St. Andrew and Choctawhatchee bays, and lowest in Apalachicola Bay. To further define the distributions of PAHs, the maximum, mean, and minimum concentrations were compared (Figure 19). These summarized data indicated the same pattern: relatively high concentrations in Pensacola Bay, intermediate and nearly equivalent to each other in St. Andrew and Choctawhatchee bays, and lowest in Apalachicola Bay. The maximum and minimum concentrations of total PAHs among all samples differed by over four orders of magnitude.

Shown separately from the PAH data, between-bay distribution patterns are evident in the concentrations of total PCBs, total pesticides, and total DDTs (Figure 20). Samples from Pensacola, St. Andrew, and Choctawhatchee bays had considerably higher concentrations of these compounds than the samples from Apalachicola Bay. The maximum concentration of PCBs occurred in a St. Andrew Bay sample, but the mean concentrations were highest in Pensacola Bay samples. Total pesticide concentrations were highest in St. Andrew and Choctawhatchee bays and lowest in Pensacola and Apalachicola bays. Mean concentrations of total DDTs were roughly equivalent among Pensacola, St. Andrew, and Choctawhatchee bays.

Lead and total mercury were selected to illustrate patterns found with metals in the different bay systems. The maximum, mean, and minimum concentrations of lead were approximately equivalent among Pensacola, St. Andrew, and Choctawhatchee bays, and relatively low in Apalachicola Bay (Figure 21). In contrast, total mercury concentrations were relatively high in Pensacola and St. Andrew bays, intermediate in Choctawhatchee Bay, and lowest in Apalachicola Bay (Figure 22).

Overall, these data suggest that the chemical concentrations and mixtures differed among the four bays. All data suggest that, on average, the Apalachicola Bay samples were the least contaminated of the four bays. The following data summarize chemical gradients within each of the four bays and Bayou Chico, an industrialized tributary to Pensacola Bay.

Pensacola Bay. Chemical analyses were performed for metals on samples from all basins of Pensacola Bay and the three adjoining, urbanized and industrialized bayous (Texar, Chico, and Grande). Organic compounds were analyzed for samples from the three bayous and adjacent portions of Pensacola Bay. Among the Pensacola Bay samples analyzed for organics, the concentrations of total PAHs were considerably higher in Bayou Chico (stations 4-9) than in all other stations (Figure 23). PAH concentrations were intermediate in three samples from Bayou Texar, and lowest in the main basin of the system. PAH concentrations diminished rapidly outside the mouths of both bayous. Two samples from inner Bayou Chico had high PCB concentrations relative to all other samples from Pensacola Bay (Figure 24). Total mercury concentrations were elevated in Bayou Texar and near the Pensacola harbor (Figure 25). Lead concentrations were relatively high in all three urban bayous and near the Pensacola harbor (Figure 26). Overall, these data suggest that most chemical substances were elevated in concentration in the three urban bayous, especially Bayou Chico, but each substance had a unique distributional pattern within the bay.

Bayou Chico. Within Bayou Chico, total PAH concentrations were highest in samples collected in the middle and lower reaches of the system (Figure 27). In contrast the concentrations of total PCBs were highest in stations from the upper and middle reaches of the bayou (Figure 28). There was no clear pattern in the concentrations of mercury and lead; samples with relatively high concentrations were scattered along the length of the bayou (Figures 29,30). Overall, these data suggest highly variable distributional patterns of chemical substances within the bayou.

Choctawhatchee Bay. Bulk chemical analyses were done on 21 samples from Choctawhatchee Bay, including many from adjoining bayous. PAH concentrations were highest at station A1-1 from Garnier Bayou and relatively low in samples from the other stations (Figure 31). Concentrations of total PCBs were high in station A1-1 as well as in stations F1-1 and F2-1 in Boggy Bayou (Figure 32). Relatively high concentrations of total mercury occurred in samples from Garnier Bayou (A1-1, C2-1, C1-2), Boggy Bayou (F1-1, F2-1, E3-1), and Rocky Bayou (G2-1) and low concentrations were found elsewhere in the bay (Figure 33). The concentrations of lead showed a distributional pattern similar to that of mercury in this bay system (Figure 34).

St. Andrew Bay. Among the 31 samples from St. Andrew Bay analyzed for organics, samples from Watsons Bayou (stations 55-63) and Massalino Bayou (stations 53-54) had the highest PAH concentrations (Figure 35). PAH concentrations decreased relatively quickly beyond the mouths of these two urban bayous. The concentrations of total PCBs showed a similar pattern (Figure 36). The sample from station 53 had the highest PCB concentration. Among the 22 samples analyzed for metals, those collected in Watsons Bayou and Massalino Bay had the highest concentrations of both mercury and lead (Figures 37, 38).

Apalachicola Bay. Chemical concentrations in samples from Apalachicola Bay were considerably lower than those in the other three bays; note the differences in concentration scales between Figures 39-42 and the previous figures. Among the four samples analyzed for chemistry, station A2-1 had the highest PAH concentrations (Figure 39), but had the lowest concentrations of mercury and lead (Figures 41-42). PCB concentrations were relatively uniform among the four stations (Figure 40).

Comparisons of Trace Metals Concentrations to Background. The Florida Department of Environmental Regulation (Schropp et al., 1990, Schropp and Windom, 1988) determined that in Florida, and generally throughout the S.E. Coastal Plain, trace metal concentrations can be expressed as a function of the amount of aluminum and metals in sediments, based on naturally occurring geochemical relationships. The relationship between metals concentrations and aluminum content were determined in a series of field surveys involving collection of
sediments from sites throughout the state far removed from any known or suspect sources of contamination. The correlation of natural metals relationships with aluminum were determined and bracketed by the $95 \%$ confidence interval. Stations with corresponding values above the upper 95\% confidence limit were considered to be "enriched". The level of enrichment was expressed as a unitless ratio. Enrichment in most cases can be considered contributable to anthropogenic sources, whether introduced by point or nonpoint sources or by atmospheric deposition. The probability that the metal was contributed by anthropogenic sources increases with increasing enrichment ratios.

Concentrations of arsenic, cadmium, chromium, copper, lead, nickel and zinc in the western Florida sediments were compared to expected, background concentrations based upon normalization to aluminum. These relationships are summarized in Figures 43-47 for each bay and each trace metal, with the exception of nickel, which was enriched in only a few samples throughout the entire four bay study area. Mercury was considered, however, because of the lack of correlation of mercury with aluminum in the background dataset (Schropp et al., 1990, Schropp and Windom, 1988), the highest value found in reference stations was set as a background value ( $0.21 \mathrm{mg} / \mathrm{kg}$ dry wt.).

In the first year samples from Pensacola Bay, arsenic was not elevated above expected background (Figure 43a); however, cadmium, chromium, copper, lead, and zinc were elevated considerably in many samples (Figures 43b-43f). Pensacola Bay samples collected during the first year included six samples from Bayou Chico. In the second year, concentrations of copper, lead, and zinc were elevated above expected background in all six samples from Bayou Chico (Figures 44 d , and g ) and cadmium and chromium were elevated in three or four samples (Figures 44b, c).

Lead and zinc were enriched in most of the Choctawhatchee Bay sediments (Figures 45 e,f); correspondingly, cadmium, chromium and copper were elevated in many samples (Figures 45 b, c, d). In St. Andrew Bay a slightly different mixture; copper, lead, and zinc, were enriched (Figures 46d, e, f) in many samples along with cadmium in some samples (Figure 46b). None of the three samples from Apalachicola Bay that were analyzed had enriched metals concentrations (Figures 47a, b, c).

In Pensacola Bay, the enriched samples were collected mainly from sites in Bayou Chico, Bayou Grande, and Bayou Texar. In Choctawhatchee Bay, enriched samples were collected mainly from Garnier Bayou, Boggy Bayou, and Destin Harbor. In St. Andrew Bay, enriched samples came from Massamino Bayou and Watsons Bayou.

Stations that showed metals enrichment within the different bays usually were enriched with several of the metals analyzed. Lead, mercury, copper and zinc were most frequently enriched, with no stations showing enrichment with respect to arsenic and nickel.

Based upon the chemistry data acquired as a part of this survey, toxicant concentrations were considerably higher in the urbanized bayous of Pensacola, Choctawhatchee, and St. Andrew bays than in the main basins of those systems or in Apalachicola Bay. Chemical concentrations were particularly high in Bayou Chico, and to a lesser degree, Watsons Bayou, Bayou Texar, Garnier Bayou, Boggy Bayou, and Massamino Bayou. Therefore, the hypothesis is that higher levels of chemical contaminants will predict probable toxic effects, and that toxicity would be most probable in Bayou Chico, somewhat less likely in the other urban bayous, and least likely in the main basins of all four bays.

Toxicity Tests. Results of all toxicity tests are listed in Tables 3-7. Sample means and statistical significance relative to controls are listed for each sampling station. Each station is listed as either: not significantly different from controls (ns), or statistically significantly different from the controls ( $@ p<0.05$ *), or both significantly different from controls ( $\mathrm{p}<0.05$ ) and less than $80 \%$ of the control value (**).

Solid phase amphipod tests. Results of the amphipod tests performed with Ampelisca abdita are summarized for both years in Table 3. In the first year (1993) samples from Pensacola Bay and St. Andrew Bay were tested by the NBS (now, USGS) laboratory. Mean 96-h LC50 concentrations for the two test series were >20 mg SDS/l and 7.27 mg SDS/I. Mean survival in home (San Francisco Bay) sediments was $68 \pm 10.4 \%$ and $93 \pm 8.4 \%$, respectively, in the two test series. Mean survival in the Redfish Bay reference sediments was $55 \pm 3.5 \%$ and $97 \pm 2.7 \%$, respectively. The results of the two test series were considerably different, suggesting differences in
the health of the animals used in the two test series. The results of the tests of the positive controls (SDS) suggest that the animals used in the first tests were unusually insensitive, however, their low survival in the reference and control sediments suggests they were very sensitive. Collectively, the data from the first test series did not meet required quality assurance standards and, therefore, are of marginal value. The data from the second test series met quality assurance standards and were of higher quality.

In the second year (1994) tests were performed by the SAIC laboratory. Mean 96-h LC50 concentrations for the 4 test series ranged from 7.41 to 7.47 mg SDS/I, indicating a very small range in variability and animal sensitivity. Mean survival in Central Long Island Sound (CLIS) controls ranged from 85\% to 98\% and were acceptable.

In the first series of tests performed in 1993, mean percent survival for all samples from Pensacola Bay was $88 \%$ or more of controls (Table 3). In the second test series (St. Andrew Bay), amphipod survival ranged from 89\% to $106 \%$ of controls. No statistically significant differences in survival between treatments and controls were detected with ANOVA in either test series $1(p=0.9705)$ or test series $2(p=0.1144)$.

In Year 2, four test series were run with samples from the western Florida estuaries. Mean amphipod survival ranged from $74 \%$ to $105 \%$ of controls in the six samples from Bayou Chico (Table 3). One sample from station 5 3 was significantly different from controls and mean survival was less than $80 \%$ of the control response. Mean amphipod survival ranged from $87 \%$ to $113 \%$ of controls in the Choctawhatchee Bay samples and one sample from station K2-1 was significantly different from controls (Table 3). In the nine samples from Apalachicola Bay, mean survival ranged from $90 \%$ to $99 \%$ of controls and one sample from station A3-1 was different from controls.

Spatial patterns in toxicity determined in the amphipod survival tests are illustrated in Figures 48-52. As described above, none of the Pensacola Bay samples collected in 1993 were toxic, i.e. significantly different from controls in this test (Figure 48). However, one sample collected in 1994 near the mouth of Bayou Chico was highly toxic, i.e., different from controls and $<80 \%$ of controls (Figure 49). The one sample from Choctawhatchee Bay that was toxic in the amphipod tests was collected in the relatively undeveloped LaGrange Bayou adjoining the far eastern reaches of the estuary (Figure 50). None of the samples collected in St. Andrew Bay were toxic in the amphipod survival tests (Figure 51). One sample collected at the mouth of the Apalachicola River across the river from the town of Apalachicola was toxic in the amphipod tests (Figure 52).

Overall, the amphipod test data showed a general lack of toxicity. No spatial patterns in toxic effects could be measured from the results of this test. Only three stations throughout all four bays were significantly toxic in the amphipod test; one in Apalachicola Bay, one in Choctawhatchee Bay, and one in Bayou Chico Bay in the Pensacola Bay estuary. Amphipod survival was less than $80 \%$ of controls in only one sample, collected from Bayou Chico.

Microtox ${ }^{T M}$ and Mutatox ${ }^{T M}$ microbial bioluminescence tests of organic extracts. Microtox ${ }^{T M}$ tests were conducted on all samples collected in both years. Mutatox ${ }^{\top M}$ tests were performed only on the 1994 samples. In contrast to results from the amphipod toxicity test, there was a broad- scale toxic response in the Microtox ${ }^{\top M}$ and Mutatox ${ }^{T M}$ bioluminescence tests. Mean EC50 concentrations, standard deviations and results of one-tailed Dunnetts comparisons with reference materials in the Microtox ${ }^{T M}$ tests are summarized by bay and by station in Table 4. Additionally, results of the Mutatox ${ }^{\text {TM }}$ tests are reported in Table 5.

In Year 1, EC50 values for organic extracts of samples from Pensacola and St. Andrew Bays ranged from 0.07 to 12.34 mg equivalent sediment wet weight. Expressed as percent of controls, the Microtox ${ }^{\top \mathrm{TM}}$ test results ranged from $0.6 \%$ to $119 \%$. Mean EC50 values were statistically compared to the NFCRC standard reference sediment. In this initial analysis, all test samples had significantly reduced EC50s ( alpha $\leq 0.01$ ). Subsequently, test results for the station with the highest EC50 (station 22 in Pensacola Bay with an EC50 of 12.34 mg equivalent sediment wet weight) was used as the least toxic reference treatment. The majority of the stations in Year 1 sampling had significantly reduced EC50 values (alpha $\leq 0.05$ ) as compared to the 'reference' station 22 (Table 4). In Year 1 results, only 7 stations of 71 from both Pensacola and St. Andrew bays were not significantly different from the 'reference' EC50 values.

Fifty-two samples from the remainder of the study area were tested during Year 2, including those from all stations in Bayou Chico, Choctawhatchee and Apalachicola bays. Mean EC50 values were statistically compared to the reference sediments collected from Redfish Bay, Texas ( $E C 50$ value $=48.9 \pm 2.8$ ). With the excep-
tion of one sample in Apalachicola Bay, all stations were significantly different from the reference material (alpha $=0.01$ ). In addition, most EC50 concentrations were less than $80 \%$ of control means and many were less than $10 \%$ of controls. Expressed as percent of the control response, the range in response varied from 1.21 to $175 \%$ of the reference value.

Spatial patterns in Microtox ${ }^{\text {TM }}$ test results are illustrated in Figures 53-57. In Pensacola Bay all samples tested from Bayou Grande, Bayou Chico and inner Pensacola Harbor were highly toxic (different from controls and $<80 \%$ of controls) in this test (Figure 53). Many of bioluminescence responses were less than 10\% of controls, especially in samples from Bayou Grande, Bayou Chico, and the Pensacola inner harbor. Samples from the outer Pensacola Harbor were not toxic and one sample each in Escambia Bay, East Bay, and Bayou Texar were not toxic. Samples from three stations (16, 24, and 25) collected nearest the mouth of the estuary were not toxic and the sample from station 14 was toxic but did not show results less than $80 \%$ of controls. The combined 1993/1994 data (Figure 54) showed widespread toxicity in the Microtox ${ }^{T M}$ tests throughout Bayou Chico. All of the samples that showed the highest toxicity were collected near the mouth of Bayou Chico (Figure 54).

Spatial patterns in toxicity were difficult to discern in Choctawhatchee and St. Andrew bays, since all samples were highly toxic (Figures 55,56 ). Most of the samples in which responses were less than $10 \%$ of controls were collected in the peripheral bayous adjoining Choctawhatchee and St. Andrew bays. All of the samples from Watson's Bayou and Massamino Bayou, for example, were very highly toxic. In Apalachicola Bay, the upstream station in the Apalachicola River, which had a considerable amount of coarse sand, was not toxic in Microtox ${ }^{\text {TM }}$ tests; the other eight samples were highly toxic (Figure 57). Samples in which responses were less than $10 \%$ of controls were collected from stations scattered throughout the bay.

Collectively, the data from the Microtox ${ }^{\top M}$ tests indicated that the majority of the samples from the four bays were toxic; 114 of 123 samples were significantly different from controls. Toxicity in this test was pervasive, extending throughout most or all of each bay. Mean test results often were less than $10 \%$ of reference response levels. All of the samples from Choctawhatchee Bay, St. Andrew Bay, and Bayou Chico were significantly different from controls. All except one sample from Apalachicola Bay were toxic and all except eight samples from Pensacola Bay were toxic. Non-toxic samples came from an up-stream station in the Apalachicola River, several stations near the mouth of Pensacola Bay, and several stations scattered throughout Pensacola Bay.

An analysis of Mutatox ${ }^{\text {TM }}$ results from the Year 2 stations revealed that 22 of 52 samples produced a strong genotoxic response (G category, Table 5). An additional 11 stations produced suspect (S) results. All stations tested in Bayou Chico Bay provided a genotoxic response. In contrast only one of the nine samples from Apalachicola Bay showed a genotoxic response and five of the samples showed no genotoxicity.

No spatial patterns in Mutatox ${ }^{\text {TM }}$ test results were evident in Bayou Chico, since all samples collected in 1994 were genotoxic (therefore, no map was prepared). In Choctawhatchee Bay genotoxic samples were most evident in the adjoining bayous, including Destin Harbor ( D stations), Joes Bayou (H stations), the two arms of Garnier Bayou, Boggy Bayou, Rocky Bayou, and LaGrange Bayou (Figure 58). Most of the M and N stations in the main basin of the bay were not genotoxic and the three stations sampled near the north shore were suspected as genotoxic. One sample from Apalachicola Bay determined to be genotoxic was collected near the mouth of the Apalachicola River (Figure 59). Three samples, one from each sampling stratum in Apalachicola Bay, were suspected as genotoxic, whereas, two samples collected near the mouth of the bay were not toxic.

Sea urchin fertilization and embryological development. Sea urchin fertilization and embryological development tests were performed in two test series in 1993, and in one test series in 1994. Sulfide concentrations in all but two samples (BC1-2 and BC2-2, in Bayou Chico, Pensacola Bay 1994) were below the detection limit of 0.01 $\mathrm{mg} / \mathrm{l}$. Test porewater dissolved oxygen concentrations ranged from 86 to $128 \%$. Values for pH ranged from 6.97 to 8.67. Total ammonia nitrogen (TAN) concentrations ranged from $0.04-18.2 \mathrm{mg} / \mathrm{l}$ for both years, and unionized ammonia (UAN) ranged from 2.4-208.1 mg/l (test series one) and $1.2-239.4 \mathrm{mg} / \mathrm{l}$ (test series two) during 1993, and from $4.1-299.5 \mathrm{mg} / \mathrm{l}$ in the 1994 tests. The lowest observable effect concentrations (LOEC- the concentration above which toxicity begins) determined in the NBS laboratory for sea urchin development and fertilization tests are 90 mg UAN/L and 800 mg UAN/L, respectively (Carr et al., 1996). There were 11 samples in which the $90 \mathrm{mg} / \mathrm{LOEC}$ for urchin development tests was exceeded in 1993, and 5 samples in 1994. No samples in either year exceeded the $800 \mathrm{mg} / \mathrm{LOEC}$ for urchin fertilization.

Results of the urchin fertilization tests of $100 \%, 50 \%$, and $25 \%$ porewater are listed for each station in Table 6. In Pensacola Bay four samples were significantly different from controls in tests of $100 \%$ porewater. In tests of $100 \%$ and $50 \%$ porewater only one sample from Pensacola Bay was toxic. In contrast five of the six samples from Bayou Chico collected in 1994 were toxic in tests of $100 \%$ porewater and all remained toxic in tests of both $50 \%$ and $25 \%$ porewater. The 1994 test results for Bayou Chico contrast with those from 1993, in which none of the samples was toxic in this test. Of all the bays, percent fertilization was most significantly reduced in Choctawhatchee Bay at the $100 \%$ porewater concentration. The mean values were lower overall than other bays, and toxicity was apparent in a higher proportion of stations (21 of 37 stations). However, toxicity was reduced considerably in subsequent dilutions; only 7 of 37 stations showed a significant level of toxic response at the $25 \%$ porewater concentration. In Apalachicola Bay four of the nine samples showed a highly significant reduction in fertilization success in $100 \%$ porewater; but only one of these samples remained toxic in $50 \%$ porewater and none was toxic in $25 \%$ porewater.

A number of different spatial patterns in the urchin fertilization tests were evident. One sample from upper Bayou Grande, one from upper East Bay, and two from the lower main basin of Pensacola Bay were toxic in 100\% porewater (Figure 60). Only the sample from Bayou Grande was toxic in both $100 \%$ and $50 \%$ porewater. All stations sampled in Bayou Chico in 1993 were non-toxic and all sampled in 1994 were highly toxic (Figure 61), thus obscuring any possible spatial patterns in toxicity. In Choctawhatchee Bay the urchin fertilization tests showed spatial patterns in toxicity similar to those suggested with the Mutatox ${ }^{\top M}$ tests. That is, most of the highly toxic samples were collected within the adjoining bayous and many samples from the main basin were either non-toxic or toxic only in 100\% porewater (Figure 62). Samples toxic to urchin fertilization at all porewater concentrations were collected in Garnier Bayou, Toms Bayou (stratum F, an arm of Boggy Bayou), and one sample each from strata L and M along the north shore of the main basin (Figure 62).

Three stations (stations $56,58,59$ ) sampled in the Watsons Bayou tributary to St. Andrew Bay had significant responses in the fertilization tests at $100 \%$ porewater, but only one (station 56 at the upper end of the bayou) was toxic at $50 \%$ porewater, and none were toxic at $25 \%$ porewater (Figure 63). In addition, one sample collected off the mouth of Watsons Bayou was toxic in tests of $100 \%$ porewater. In Apalachicola Bay, four samples were toxic, two each in the lower Apalachicola River and the lower bay (Figure 64). The sample from station C31 was toxic in both $100 \%$ and $50 \%$ porewater.

Overall, the results of the sea urchin fertilization tests showed relatively high toxicity in several bayous of Choctawhatchee Bay, Watsons Bayou in St. Andrew Bay, and Bayou Chico of Pensacola Bay compared to the other areas and relatively low toxicity in most of main basins of Pensacola and St. Andrew bays. Most of the 1994 samples from Bayou Chico were highly toxic in all porewater concentrations, whereas none collected in 1993 was toxic in any porewater concentrations. Two samples each in the lower Apalachicola River and lower Apalachicola Bay were toxic in $100 \%$ porewater. Among the 123 samples tested, 38 ( $31 \%$ ) were significantly toxic in tests of $100 \%$ porewater.

Results of the sea urchin embryological development tests are summarized in Table 7. Normal development was reduced to some degree in samples from all of the bays. Test results ranged from $0.0 \%$ to $99.6 \%$ normal development among all 123 samples. Eleven of 40 samples from Pensacola Bay collected in 1993 were significantly toxic in this test. All except one of the samples collected in 1993 and 1994 in Bayou Chico were highly toxic in both $100 \%$ and $50 \%$ porewater. Ten of the 12 Bayou Chico samples showed $0.0 \%$ normal development in $100 \%$ porewater.

In Choctawhatchee Bay, 18 of 37 samples were significantly toxic in the embryological development test with $100 \%$ porewater (Table 7). Five of these samples showed $0.0 \%$ normal development. However, only four samples were significantly toxic in tests of $50 \%$ porewater and none was toxic in $25 \%$ porewater. In St. Andrew Bay 7 of 31 stations were significantly toxic in tests of $100 \%$ porewater. Watsons Bayou stations 55-59 in St. Andrew Bay had a highly significant response in tests of $100 \%$ porewater. Two of these five were toxic at the $50 \%$ porewater concentration, but none were toxic at the $25 \%$ concentration.

Urchin development was significantly reduced (both significantly different from control and less that $80 \%$ of control) in tests of $100 \%$ porewater at four of the nine stations in Apalachicola Bay (Table 7). Two of these samples showed $0.0 \%$ normal development. However, none of the samples was toxic at either the $50 \%$ or $25 \%$ concentrations.

As in the urchin fertilization tests, a number of different spatial patterns in toxicity were apparent with the embryo development tests. In Pensacola Bay, toxicity was restricted mainly to Pensacola Harbor and Bayou Chico (Figure 65, Figure 66). Several samples from Bayou Chico were toxic in all porewater concentrations (Figure 66) and several from the Pensacola Harbor area were toxic in either $100 \%$ or $50 \%$ porewater. Toxicity diminished rapidly toward the lower bay and the mouth of the estuary. One sample each from Bayou Grande, the mouth of the Escambia River and Bayou Texar were toxic in $100 \%$ porewater. Only the upper-most station (number 4) in Bayou Chico was non-toxic (Figure 66).

In Choctawhatchee Bay, samples most toxic to embryo development were those from Destin Harbor (D stratum), Joes Bayou (H stratum) and Toms Bayou (F stratum) in which results were significant in both 100\% and $50 \%$ porewater (Figure 67). In addition, samples from several other bayous, especially both arms of Garnier Bayou, were toxic only in the $100 \%$ porewater test. Only two samples from the main basin (strata L, M, N) were toxic. Toxicity in the embryo development test was restricted to five stations sampled in Watsons Bayou and two stations in the main basin of St. Andrew Bay (Figure 68). In Apalachicola Bay three samples from the main basin of the bay and one from the lower Apalachicola River were toxic, but only in tests of $100 \%$ porewater (Figure 69).

Overall, stations exhibiting toxicity in the embryo development tests largely were associated with urbanized tributaries to the bays in all cases except Apalachicola Bay. Toxicity was especially apparent in Bayou Chico, Watson Bayou, the Pensacola Harbor, Destin Harbor, and portions of Garnier Bayou. Among the 123 samples tested, a total of $46(37 \%)$ was toxic in at least $100 \%$ porewater, a slightly higher proportion than observed in the fertilization tests. There was a relatively high incidence of toxicity in Choctawhatchee Bay, Apalachicola Bay and Bayou Chico as compared to other areas and a very low incidence of toxicity in St. Andrew Bay. Results agreed relatively well with those from the urchin fertilization tests.

Concordance among toxicity tests. In the preceding section it was apparent that there were different levels of agreement or concordance among the different toxicity tests. Spearman-rank correlations were determined for pairs of tests to quantify these relationships. In Pensacola Bay the only statistically significant ( $\mathrm{p}<0.05$ ) correlation was between sea urchin fertilization and microbial bioluminescence (Table 8). Although test results for the sea urchin development and fertilization bioassays showed a positive correlation coefficient, the relationship was not significant. There were no significant correlations among tests for the 1994 combined Bayou Chico/ Apalachicola samples (Table 9). There were no significant correlations among tests in Choctawhatchee Bay (Table 10). In St. Andrew Bay, sea urchin fertilization was significantly correlated with both microbial bioluminescence and sea urchin development (Table 11).

The Mutatox ${ }^{\text {TM }}$ test results could not be correlated with other bioassay results because they were categorical scores, not numerical values. Nevertheless, all of the 21 samples that were scored as genotoxic in the Mutatox ${ }^{\mathrm{TM}}$ tests were highly toxic in the Microtox ${ }^{\text {TM }}$ tests, resulting in $100 \%$ agreement. There were 39 samples that were scored as either non-toxic or genotoxic in the Mutatox ${ }^{\text {TM }}$ tests: 22 ( $56 \%$ ) of these samples agreed as to toxicity or non-toxicity in the sea urchin fertilization tests and 23 (59\%) agreed in the urchin development tests.

Overall, these data suggest that the toxicity tests identified overlapping, but, generally different patterns in toxicity. The most consistent correlations were those between sea urchin fertilization and microbial bioluminescence and between the Mutatox ${ }^{\text {TM }}$ test results and Microtox ${ }^{\text {TM }}$ test results and both of the sea urchin tests. In the combined data set, urchin fertilization and development were significantly correlated ( $\mathrm{Rho}=+0.682, \mathrm{p}<0.0001$ ) and Microtox ${ }^{\text {TM }}$ EC50 values and amphipod survival were significantly correlated ( $\mathrm{Rho}=+0.363, \mathrm{p}=0.005$ ). Microtox ${ }^{\text {TM }}$ EC50 values were negatively correlated with both urchin fertilization and urchin development (Rho = -0.305 and -0.299 , respectively, $\mathrm{p}<0.05$ ). None of the other combinations of toxicity tests were significantly correlated

Incidence of toxicity. Table 12 summarizes the incidence of samples tested from each bay in which bioassay results were either significantly different from controls or both significantly different from controls and less than $80 \%$ of the control response. For the Mutatox ${ }^{\text {TM }}$ test, the toxic samples included those determined to be either Genotoxic or Suspect (highly toxic samples were those determined to be Genotoxic).

In the amphipod survival tests, only three of the 123 samples were toxic; one each from Bayou Chico, Choctawhatchee Bay, and Apalachicola Bay. One sample from Bayou Chico was highly toxic (Table 12). In the
urchin fertilization tests the incidence of toxicity was highest in Bayou Chico, followed by Choctawhatchee Bay, and Apalachicola Bay. However, the incidence of toxicity in this test decreased rapidly with porewater dilutions in all areas except Bayou Chico, where 5 of 6 samples were highly toxic in all porewater concentrations. Throughout the entire study area, approximately one-third (30.9\%) of the samples were toxic in tests of $100 \%$ porewater.

In the urchin development tests, the incidence of toxicity was, again, highest in Bayou Chico, followed by Choctawhatchee and Apalachicola bays (Table 12). As in the fertilization tests, toxicity also dropped rapidly with porewater dilutions, except in Bayou Chico. In Apalachicola Bay none of the samples was toxic in tests of $50 \%$ or $25 \%$ porewater. In Pensacola, Choctawhatchee, and St. Andrew bays none of the samples was toxic in tests of $25 \%$ porewater.

Of the 123 samples tested in Microtox ${ }^{T M}$ tests, 114 were toxic and of those, 113 were highly toxic, equivalent to $92.7 \%$ and $91.9 \%$ incidences of toxicity (Table 12). In the Mutatox ${ }^{\text {TM }}$ tests all 6 Bayou Chico samples, 24 of 37 Choctawhatchee Bay samples, and 4 of 9 Apalachicola Bay samples were either genotoxic or suspected as genotoxic.

Overall, the highest incidence of toxicity occurred among the Bayou Chico samples, followed by Choctawhatchee Bay, and the other areas. All of the Bayou Chico samples were toxic in the sea urchin development, Microtox ${ }^{\top \mathrm{TM}}$, and Mutatox ${ }^{T M}$ tests; all were highly toxic in the urchin development and Mutatox ${ }^{T M}$ tests; and all except one sample was highly toxic in the urchin fertilization tests. In addition, the only sample that was highly toxic in the amphipod survival tests was collected in Bayou Chico. In Choctawhatchee Bay all samples were toxic in Microtox ${ }^{\text {TM }}$ tests, most were toxic in Mutatox ${ }^{\text {TM }}$ tests, $57 \%$ were toxic in urchin fertilization tests, $49 \%$ were toxic in urchin development tests, and one sample was toxic in the amphipod tests. The incidence of toxicity in Pensacola Bay and St. Andrew Bay was relatively similar: zero and one sample toxic in amphipod tests (respectively); 10\% and $13 \%$ toxic in urchin fertilization tests; $27 \%$ and $23 \%$ toxic in urchin development tests; and $80 \%$ and $100 \%$ toxic in Microtox ${ }^{\text {TM }}$ tests. Among all areas Apalachicola Bay had the lowest incidence of toxicity in all tests.

Spatial extent of toxicity. Toxicity data were weighted to the sizes of each sampling stratum and the total area in each bay that was "highly" toxic (i.e., results were less than $80 \%$ of controls) was determined as the sum of the weighted areas. Data from both 1993 and 1994 sampling episodes in Bayou Chico were combined for these calculations. Calculations were not prepared for the Mutatox ${ }^{\top M}$ bioassay, since the results were categorical, not numerical.

In Apalachicola Bay, estimates of the spatial extent of toxicity were: $0.0 \mathrm{~km}^{2}$ for amphipod survival; $157.5 \mathrm{~km}^{2}$ ( $84 \%$ of total) for sea urchin deyelopment in $100 \%$ porewater; 63.6 km (33.9\%) for sea urchin fertilization in $100 \%$ porewater; and $186.8 \mathrm{~km}^{2}$ ( $99.6 \%$ ) for Microtox ${ }^{\text {TM }}$ (Table 13). In St. Andrew Bay the estimates were: 0.0 $\mathrm{km}^{2}$ for amphipod survival; $5.6 \%$ for sea urchin development; $1.8 \%$ for sea urchin fertilization; and $100 \%$ for Microtox ${ }^{T M}$. The estimates for Pensacola Bay were very similar to those for St. Andrew Bay: $0.015 \%$ for amphipod survival; $2 \%$ for sea urchin development; $5.3 \%$ for sea urchin fertilization; and $96.4 \%$ for Microtox ${ }^{\text {TM }}$. In Choctawhatchee Bay, estimates were: 0.0\% for amphipod survival; 45.4\% for sea urchin development; 44.5\% for sea urchin fertilization; and 100\% for Microtox ${ }^{\text {TM }}$.

These data suggest that amphipod survival was affected in a tiny portion of the entire study area and microbial bioluminescence was affected in nearly all of the area. Both sea urchin fertilization and embryo development were affected in nearly one-half of Choctawhatchee Bay, and small portions of St. Andrew and Pensacola bays. In Apalachicola Bay the majority of the area was affected in sea urchin development tests, whereas about onethird was affected in the fertilization tests. Usually, small portions of each bay were affected in both tests of $50 \%$ and 25\% porewater.

Relationships between Toxicity and Chemical Concentrations. The relationships between toxicity measured in the different toxicity tests and the concentrations of numerous chemical substances in the samples were examined for each bay and for the entire combined study area. The cause(s) of toxicity cannot be determined in field surveys such as those performed in these studies and determinations of causality were not among the objectives of the surveys. However, several statistical analyses were performed to identify those substances most clearly associated with measures of toxicity in the samples.

Pensacola Bay. In the 1993 Pensacola Bay samples, none of the substances were correlated with amphipod survival, not surprising because amphipod test results were similar among all samples (Table 14). Results of the

Microtox ${ }^{\text {TM }}$ tests were highly correlated with the concentrations of numerous chemicals, notably, cadmium, copper, lead, zinc, and the sum of the eight metals/ERM ratios. Microtox ${ }^{\text {TM }}$ results also were correlated with many individual PAHs, the sum of low molecular weight PAHs, total PAHs, the sum of the eight PAH/ERM quotients, several chlorinated organic hydrocarbons, several PCB congeners, and the sum of all 18 chemical/ERM quotients. The results of the urchin fertilization tests were significantly correlated with a few trace metals, many PAHs, a few chlorinated hydrocarbons, and the sum of all 18 chemical/ERM quotients. Urchin development was most correlated with the concentrations of un-ionized ammonia, followed by barium and the sum of the eight PAH/ERM quotients. In addition, urchin development was significantly correlated with the concentrations of a number of trace metals, PAHs, pesticides, and the sum of all 18 chemical/ERM quotients.

Collectively, these data suggest that toxicity in Pensacola Bay was associated with mixtures of many substances acting together, notably including cadmium, copper, lead, zinc, PAHs, several pesticides, and in the case of urchin development, ammonia. The significant relationship between the sum of the 18 chemical/ERM quotients and the results of both urchin tests and Microtox ${ }^{\top M}$ tests is noteworthy because it further suggests that mixtures of substances co-varying with each other contributed to toxicity.

Concentrations of several trace metals, PAHs, and chlorinated organic hydrocarbons equaled or exceeded respective guideline values in Pensacola Bay (Table 15). Notably, concentrations of zinc, dibenzo(a,h)anthracene, dieldrin, and isomers of DDT were elevated relative to guideline concentrations. These substances were correlated with toxicity, and they exceeded either the respective PEL or ERM concentration or both in some samples. None of the concentrations of un-ionized ammonia equalled or exceeded toxicity thresholds for amphipod survival (236 ug/l, Kohn et al., 1994) or urchin fertilization (800 ug/l, Long et al., 1996). However, seven un-ionized ammonia concentrations exceeded the low observable effects concentration (90 ug/l, Long et al., 1996) for urchin development and all seven samples were toxic.

Selected toxicity/chemistry relationships are illustrated in Figures 70-74 in which data are shown as bivariate scatterplots. These scatterplots include the Spearman-rank correlation coefficients, and either sediment quality guidelines or toxicity thresholds as reference points. Percent normal development of urchins dropped to zero in many samples with relatively high zinc concentrations, i.e., above the PEL concentration of 271 ppm (Figure 70). Conversely, percent normal development was relatively high in most samples with zinc concentrations below the TEL concentration of 124 ppm . Also, zero percent normal development occurred in some (but, not all) samples with total high molecular weight PAH concentrations above the PEL value of 6676 ppb (Figure 71). A sample from Bayou Chico was unusual in having over 25000 ppb total HPAH and over $90 \%$ normal urchin development. This sample also had an unusually high TOC concentration ( $6 \%$ ) which may have inhibited the bioavailability of hydrocarbons.

In the Microtox ${ }^{\text {TM }}$ tests, there was very strong association with concentrations of DDT, including the sum of the DDT isomers (Figure 72). Microtox ${ }^{\text {TM }}$ EC50 concentrations decreased rapidly with increasing concentrations of total DDT and all samples that exceeded the PEL concentration were highly toxic. To determine if mixtures of substances acting together may have contributed to toxicity, chemical concentrations were normalized to (i. e., divided by) their respective ERM values and these quotients were added to form cumulative ERM quotients. Microtox ${ }^{\text {TM }}$ test results were correlated with the cumulative ERM quotients; there was a strong pattern of decreasing EC50 concentrations with increasing ERM quotient values (Figure 73).

Ammonia is known to be a highly toxic substance in marine sediments (Kohn et al., 1994) and can be a major contributor to toxicity, especially in the un-ionized form. Results of the urchin embryo development tests were highly correlated with the concentrations of un-ionized ammonia. The scattergram of the data confirmed this strong association (Figure 74). All except one sample with a concentration of ammonia above the LOEC of 90 ug/I were highly toxic in tests of embryo development. The majority of the samples with low ammonia concentrations were not toxic.

In the Microtox ${ }^{\text {TM }}$ tests, the average concentrations of six trace metals and many organic compounds or classes of compounds in toxic samples exceeded the average concentrations in non-toxic samples by factors of up to 5.8 (Table 16). As expected in these analyses, there was considerable variability in these concentrations as indicated with the relatively high standard deviations relative to the means. Nevertheless, there was a pattern of higher concentrations, on average, of many substances in the 32 toxic samples than in the 8 non-toxic samples.

The average concentrations of all other substances that were measured, but not listed in Table 16, were lower in the toxic samples than in the non-toxic samples.

The average concentrations of many substances in the samples that were toxic in Microtox ${ }^{\text {TM }}$ tests exceeded either the ERL or TEL concentrations of Long et al. (1995) or MacDonald (1994), respectively. Only 4,4'-DDT, however, equaled or exceeded the ERM or PEL concentrations (Table 16). Furthermore, the average concentrations of total DDTs in the toxic samples exceeded the ERL value by a factor of 17.1 and the TEL value by a factor of 6.9. The concentrations of total DDTs and two isomers were correlated with toxicity (Table 14). Concentrations of high molecular weight PAHs, total PAHs, and total PCBs in toxic samples were high relative to the non-toxic samples and guideline values (Table 16).

In contrast, although cadmium was highly correlated with Microtox ${ }^{\text {TM }}$ test results (Table 14), the concentrations of this metal were not remarkably elevated compared to guideline values. Silver was neither correlated with toxicity test results nor particularly elevated in the toxic samples relative to either non-toxic samples or numerical guidelines. Both lead and zinc were correlated with Microtox ${ }^{\top \mathrm{TM}}$ results but only slightly elevated in toxic samples relative to both non-toxic samples and guideline concentrations.

These data suggest that the high molecular weight PAHs and DDTs, and to a lesser extent, total PCBs, total PAHs, and several trace metals (e.g., copper, lead, and zinc) may have contributed to toxicity in the Microtox ${ }^{\top \mathrm{TM}}$ tests. Most other trace metals and organic compounds probably had either a minor role or no role in contributing to toxicity.

In the urchin fertilization tests, only three trace metals and two pesticides were elevated in concentrations in the three toxic samples relative to the 36 non-toxic samples (Table 17). The ratios between the toxic and non-toxic averages, however, were very small ( $<2.0$ ) and the average concentrations in toxic samples exceeded respective guideline values by very small amounts (ratios of 1.1 to 3.6 ). None of these five substances showed significant correlations with fertilization success (Table 14).

Numerous substances were correlated with urchin embryological development (Table 14). The concentrations of many of these chemicals also were elevated in the toxic samples relative to non-toxic samples and numerical guidelines (Table 18). Notable among these chemicals were zinc, the sum of low molecular weight PAHs, 4, 4'DDD, 4,4'-DDT, and dieldrin, all of which had average concentrations in toxic samples 3.5 to 6.8 times higher than in the non-toxic samples and 3.3 to 26.4 times higher than respective numerical guidelines. The average concentrations of these five substances exceeded PEL values as well as the lower TEL concentrations. All of these substances were significantly correlated with urchin development. They may have contributed to toxicity in this test. The other substances listed in Table 18 may have made minor contributions to toxicity.

Table 19 provides a summary of the analyses of the relationships between toxicity and chemistry for Pensacola Bay. Table 19 includes the number of toxicity tests in which each substance was significantly correlated; the number of samples in which each substance exceeded either the ERL or TEL value; the ratio between the average chemical concentrations in toxic and non-toxic samples for each test; and the ratios between the average concentrations in toxic samples versus the respective TEL value. Based upon these summarized data, it appears that the concentrations of zinc, high molecular weight PAHs, two DDD/DDT isomers, total DDT, and dieldrin were most closely associated with toxicity. To a lesser extent, cadmium, copper, lead, low molecular weight PAHs, and in the case of urchin embryo development, un-ionized ammonia, were associated with toxicity in Pensacola Bay. The significant correlations between three measures of toxicity (Microtox ${ }^{\text {TM }}$, urchin fertilization, urchin development) and the sums of the chemical concentrations normalized to (i.e., divided by) the ERM values suggests that these substances acted together in complex mixtures to contribute to toxicity. Substances not included in Table 19 probably had no role or a minor role in contributing to toxicity. Other major contributors to toxicity may have included substances not measured in the chemical analyses, such as the extensive findings of nitro aromatic compounds later identified in cleanup of the PAH analyses.

Bayou Chico/Apalachicola Bay. Because the Bayou Chico and Apalachicola Bay datasets were relatively small, they were combined to examine chemistry/toxicity relationships. In the combined Bayou Chico/Apalachicola Bay data set from 1994, very few substances were correlated with toxicity (Table 20). Only six substances (five PAHs and one PCB congener) were correlated with toxicity and they were correlated only with urchin fertiliza-
tion. Many correlation coefficients were numerically relatively high (i.e., $>0.5$ ), but, because of considerable variability and the relatively small sample size ( $n=10$ ), they were not significant.

Scatterplots of the data showed that, despite the non-significant correlations, toxicity increased with increasing concentrations of many substances (Figures 75-79). Except for one sample, urchin fertilization dropped to zero as zinc concentrations increased to approximately 100 ppm and four samples exceeded both the PEL and ERM values for zinc (Figure 75). Similarly, Microtox ${ }^{\text {TM }}$ EC50 concentrations decreased rapidly with increasing concentrations of fluoranthene (Figure 76) and one sample exceeded the proposed national sediment quality criterion ( $300 \mathrm{ug} / \mathrm{goc}, \mathrm{U}$. S. EPA, 1994). Except for one sample with high urchin fertilization, test results dropped rapidly as total PAHs increased; two samples exceeded the PEL value of 16770 (Figure 77). Reflecting the association between the complex mixtures of substances in the samples and toxicity, urchin normal development decreased markedly with increasing cumulative ERM quotients (Figure 78).

Urchin development showed a strong association with un-ionized ammonia concentrations in the porewater and several samples exceeded the LOEC concentration of 90 ug/l (Figure 79). Dissolved oxygen concentrations were relatively high in the 1993 samples and very low in most 1994 samples; however, there was no apparent relationship between in situ bottom water dissolved oxygen concentrations and toxicity in either the fertilization or embryo development tests.

One sample from Bayou Chico (station 3-2) stood out as unusual in some scatterplots (i.e., Figures 75, 77) since the concentrations of many substances were relatively high, but the sample was not toxic in either of the amphipod or urchin fertilization tests. This sample had an unusually high concentration of total organic carbon (7.1\%), thereby possibly reducing the bioavailability of the toxicants. The Microtox ${ }^{\text {TM }}$ test indicated toxicity in this sample which is not unusual since this test was performed with an extract of the sediments prepared with an organic solvent. The solvent would be expected to elute mixtures of organics and, to a lesser extent, trace metals thus increasing their bioavailability in the extract.

As noted earlier, the concentrations of several trace metals (especially, copper, lead, and zinc) were anthropogenically-elevated relative to background concentrations in the Bayou Chico samples and not in the Apalachicola Bay samples. In addition, the concentrations of many substances in the Bayou Chico samples equalled or exceeded applicable, effects-based, sediment quality guidelines.

A summary of the exceedances of numerical guidelines is listed in Table 21. Numerous aromatic hydrocarbons were particularly elevated in concentration, including the sums of both low and high molecular weight compounds. One sample from Bayou Chico exceeded the national criterion for fluoranthene. The concentrations of zinc were particularly high in several samples relative to the guideline values. Two isomers of DDT and total PCBs were found in relatively high concentrations. All of the samples that had high contaminant concentrations were collected in Bayou Chico; none came from Apalachicola Bay, as expected. All stations except number 1-2 in the upper reach of Bayou Chico had particularly high chemical concentrations. The average of the cumulative ERM quotients among the three samples from Apalachicola Bay was 0.6 (range $=0.3$ to 1.0), considerably lower than the average cumulative ERM quotient for the six Bayou Chico samples ( 10.6 , range $=1.9$ to 14.2).

Comparisons between chemical concentrations in toxic and non-toxic samples could not be performed with amphipod test results because only one sample was classified as toxic. Conversely, all samples were identified as toxic in the Microtox ${ }^{\text {TM }}$ tests, providing no non-toxic samples for comparisons. In the urchin development tests only two samples were classified as non-toxic. Therefore, comparisons between toxic and non-toxic samples were restricted to the data from the urchin fertilization tests, in which three samples analyzed for chemistry were non-toxic and six were toxic in $100 \%$ porewater.

The average concentrations of many substances were elevated in the toxic samples as compared to the nontoxic samples from Bayou Chico and Apalachicola Bay (Table 22). Numerous individual PAHs occurred in high concentrations in the toxic samples; the sums of the low and high molecular weight compounds and all compounds listed in Table 22 reflect this pattern. Expressed in units of organic carbon, the concentrations of three individual PAHs for which national criteria have been developed, were particularly elevated in the toxic samples. The average concentrations of the low and high molecular weight PAHs in the toxic samples exceeded the ERL, TEL, and PEL concentrations, indicating a relatively high probability that they contributed to toxicity.

The concentrations of zinc were remarkably high in many samples, including those that were toxic in the urchin fertilization tests (Table 22). They exceeded both the ERM and PEL concentrations, the only substance that did so in Bayou Chico. Also, noteworthy were the concentrations of lindane, heptachlor epoxide, several isomers of DDT, total PCBs, and lead which occurred in moderately elevated concentrations in the toxic samples.

In summary, although Spearman-rank correlations failed to show significant correlations between toxicity in samples from Bayou Chico and Apalachicola Bay, there were numerous obvious associations between elevated chemical levels and toxicity. The concentrations of zinc and many organic compounds, notably the PAHs and to a lesser extent some chlorinated organic hydrocarbons, were relatively high in these samples, especially those that were toxic in the urchin fertilization tests. The toxicity of one sample with high chemical concentrations may have been inhibited by high concentrations of organic carbon which may have reduced the bioavailability of the organic compounds.

Choctawhatchee Bay. Spearman-rank correlations for Choctawhatchee Bay samples are listed in Table 23. Amphipod test results were significantly correlated with only the concentrations of total indeno-type pesticides. However, this correlation was positive, not negative as expected, and therefore meaningless. The Microtox ${ }^{\top M}$ tests were not significantly correlated with any chemical concentrations. In the amphipod tests, only one sample was significantly different from controls and none were highly toxic. In Microtox ${ }^{\text {TM }}$ tests, all samples were highly toxic, thus providing no meaningful toxicity gradients with which to compare chemical concentrations. Sea urchin embryo development was significantly correlated with the concentrations of only un-ionized ammonia and two individual PAHs (fluoranthene, phenanthrene) expressed in units of organic carbon (Table 23). None of the other substances were significantly correlated with the results of this toxicity test.

In the urchin fertilization tests, results were significantly correlated with the concentrations of numerous individual trace metals, the sum of the trace metals/ERMs quotients, the sum of 5 simultaneously-extracted metals (SEM), the sum of 5 SEM minus total AVS, many individual PAHs, the sums of classes of PAHs, the sum of 13 PAHs/ERMs quotients, numerous chlorinated hydrocarbons, total PCBs, and the sum of all 25 chemical concentrations/ERM quotients (Table 23). Expressed in units of dry wt., phenanathrene and fluoranthene were significantly correlated with urchin fertilization; however, when expressed in units of organic carbon, neither were significantly correlated with these test results.

The incidence of toxicity was similar in the urchin fertilization and embryological tests ( $46 \%$ and $43 \%$, respectively). However, the toxicity/chemistry correlations differed considerably between them.

Five samples from Choctawhatchee Bay (those from station A1-1 in Cinco Bayou, C3-1 in Garnier Bayou, D3-1 in Destin Harbor, and F1-1 and F2-1 in Tom's Bayou) had chemical concentrations that exceeded guideline values (Table 24). Concentrations of silver and several DDT isomers were elevated in the samples from Tom's Bayou and concentrations of four individual PAHs and total PAHs were elevated in the sample from Cinco Bayou. In addition, the concentrations of un-ionized ammonia exceeded the LOEC concentration (90 ug/l) for urchin embryo development in two samples (B3-1 and K2-1).

Average concentrations of chemicals in non-toxic and toxic samples were compared for the urchin fertilization tests (Table 25). Equivalent analyses of the data for the other tests could not be performed because the data either showed toxicity in all samples (Microtox ${ }^{\text {TM }}$ ) or too few samples (urchin development, amphipod survival) to provide comparison groups. Average concentrations of silver, dieldrin, DDT isomers, and total DDT were considerably higher in the toxic samples than in the non-toxic samples (ratios of about 20.0 of higher). Concentrations of many trace metals and PAHs were elevated in the toxic samples. However, concentrations of all substances were not very high compared to the sediment quality guidelines. None of the average concentrations in toxic samples equalled or exceeded respective PEL, ERM, or SQC values. These concentrations often approximated or were only slightly higher than the respective TEL or ERL values.

Although concentrations of silver were considerably higher in the toxic samples than in the non-toxic samples, average concentrations in the toxic samples were lower than the ERL and TEL values (Table 25). The same situation occurred with cadmium, zinc, the sum of low molecular weight PAHs, dieldrin, and total PCBs. Among the substances analyzed, the sum of DDT isomers was most elevated in concentration in toxic samples: average concentrations in toxic samples exceeded both the ERL and the TEL by factors of 11.4 and 4.7, respectively.

The relationship between results of the sea urchin fertilization tests and the concentrations of total DDT in sediments is illustrated in Figure 80. Fertilization success decreased remarkably with increasing concentrations of DDT. All samples with DDT concentrations that exceeded the TEL value were highly toxic in this test. Fertilization success was zero in the sample with the highest concentration of total DDT. The Spearman-rank correlation showed a significant association between fertilization success and total DDT.

The association between urchin embryo development and un-ionized ammonia was significant; the resulting scatterplot showed a pattern of decreasing normal development with increasing ammonia concentrations (Figure 81). Two samples either equalled or exceeded the lowest observed effects concentration (LOEC) of $90 \mathrm{ug} /$ I and both were highly toxic in this assay.

In summary, the associations between toxicity and concentrations of potentially toxic substances in Choctawhatchee Bay were strongest for the urchin fertilization tests in which a large gradient in response was observed. Microtox ${ }^{\text {TM }}$ and amphipod test results were not negatively correlated with any substances. Urchin embryo development test results were correlated with only three substances: un-ionized ammonia and two individual PAHs. In sharp contrast, numerous substances were correlated with urchin fertilization. Most notable among these were the concentrations of DDT isomers, total DDT, silver, the sum of PAHs, and dieldrin. However, the concentrations of these substances exceeded TEL and ERL values by small amounts and rarely equaled or exceeded respective PEL and ERM values. Therefore, there is insufficient evidence to suggest which measured substances may have contributed substantially to toxicity. As in the other bays, mixtures of contaminants and unmeasured compounds may be culprit in their contribution to toxicity.

St. Andrew Bay. Of the 30 samples collected within St. Andrew Bay, organic compounds were quantified in all samples, total organic carbon and grain size were determined in 25 samples, and both total trace metals in bulk sediments and simultaneously extracted metals in acid volatile sulfides were measured in 22 samples.

Percent amphipod survival was significantly correlated with concentrations of copper, two isomers of DDT, total DDT, and total pesticides (Table 26). The relationship between amphipod survival and copper is consistent with the copper/aluminum plot which indicates a high incidence of anthropogenic enrichment for copper in this area. Given that none of the samples was significantly toxic in this test and test results were relatively similar among all samples, these correlations were unexpected. The relationship between amphipod survival and total DDT concentrations is illustrated in Figure 82. Although most of the samples showed a pattern of decreasing amphipod survival with increasing total DDT concentrations, three samples in which amphipod survival was relatively high had elevated concentrations of DDT. The sample with the highest DDT concentration was collected in upper Massamino Bayou and probably had a high concentration of TOC (no analyses were performed). The concentration of total AVS was highest recorded among all St. Andrew Bay samples (>29 umoles/g). Some samples exceeded respective ERM values (Long et al., 1995) and PEL values (MacDonald, 1994), including several that were not toxic.

Concentrations of un-ionized ammonia were significantly correlated with toxicity to sea urchin development in $100 \%$ porewater (Table 26). This correlation was highly significant and showed a strong pattern of decreasing normal development with increasing ammonia concentrations (Figure 83). Samples were invariably toxic (less than $80 \%$ normal development) when un-ionized ammonia concentrations exceeded 40 ug/l. Furthermore, percent normal development was zero when un-ionized ammonia concentrations exceeded the LOEC of $90 \mathrm{ug} / \mathrm{l}$.

Results of the sea urchin fertilization tests were significantly correlated with many trace metals, the sum of the nine metals concentrations-to-ERM quotients, un-ionized ammonia, several DDT isomers, and the sum of the three chlorinated organic hydrocarbon-to-ERM ratios (Table 26). The relationship between sea urchin fertilization and zinc (Figure 84) is typical of that for the other trace metals. In four of the samples, fertilization success decreased with increasing zinc concentrations; however, in the other samples fertilization success remained relatively high despite exposure to elevated zinc concentrations.

The correlation between sea urchin fertilization and un-ionized ammonia was significant (Table 26). There was a strong pattern of decreasing fertilization success with increasing ammonia concentrations. However, none of the samples equalled or exceeded the LOEC concentration (800 ug/l) for this test (Figure 85), suggesting that this substance occurred at concentrations well below that which would have been expected to contribute to toxicity.

Microbial bioluminescence tests were significantly correlated with many substances (Table 26), including many trace metals, many chlorinated organic compounds, and all classes of PAHs. In addition, test results were correlated with the sums of the three classes of ERM quotients ( 9 metals, 3 chlorinated compounds, 13 PAHs ). These data indicated that the Microtox ${ }^{\top \mathrm{M}}$ test was sensitive to complex mixtures of substances in the sediments, many of which co-varied with each other. The correlation with trans-nonachlor was the strongest observed (Rho $=-0.722, \mathrm{p}<0.0001$ ) and the data showed a strong pattern of association (Figure 86). The Microtox ${ }^{\mathrm{TM}}$ data were scattered at trans-nonachlor concentrations below $0.5 \mathrm{ng} / \mathrm{g}$. As the concentrations of this substance increased, however, microbial bioluminescence EC50 values decreased markedly. Sample 53 had the highest trans-nonachlor concentration ( $9 \mathrm{ng} / \mathrm{g}$ ) and was the most toxic to microbial bioluminescence.

The relationship between the sum of the 25 chemical concentration-to-ERM quotients and microbial bioluminescence is illustrated in Figure 87. This chemical index accounts for variability among 25 substances in the samples, including 9 trace metals, 3 chlorinated organic compounds, and 13 PAHs , and, therefore, integrates the potential contributions of all of these substances in mixtures to the measure of toxicity. The index calculated with the ERM values was correlated with the microbial bioluminescence test results and the data showed a strong pattern of association (Figure 87). The two least toxic samples had among the lowest ERM index values and the most toxic sample (station 53 ) had the highest index value.

Sampling stations in which chemical concentrations equalled or exceeded either interpretive guideline values or toxicity thresholds are listed in Table 27. PEL values generally were lower than the corresponding ERM values, therefore, more samples exceeded the PELs than the ERMs (Table 27). Concentrations of several trace metals, PAHs, and chlorinated compounds exceeded the respective ERM values in the sample collected at station 53. Chemical concentrations in sample 53 as well as several others exceeded the respective PEL values. Chemical concentrations often were elevated in samples $53,54,56,57$, and 58 . Among the chemicals that were quantified, the concentrations of DDT isomers (particularly $p, p^{\prime}$-DDD) were most frequently elevated relative to the guidelines. The concentrations of un-ionized ammonia were elevated in five samples relative to the LOEC for the sea urchin development test ( $90 \mathrm{ug} / \mathrm{I}$ ), in one sample relative to the NOEC for the amphipod test ( $236 \mathrm{ug} / \mathrm{l}$ ), however, none of the samples exceeded the LOEC ( $800 \mathrm{ug} / \mathrm{l}$ ) for the sea urchin fertilization test.

The co-occurrence analyses of these data in which average chemical concentrations in toxic and non-toxic samples are compared was not performed with the St. Andrew Bay data. There were no samples that were toxic to amphipods, all samples were toxic to Microtox ${ }^{\text {TM }}$, only ammonia was correlated with sea urchin development, and only four samples were toxic to sea urchin fertilization. Therefore, we chose to forego these analyses with these data.

In summary, the four toxicity end-points measured in St. Andrew Bay appeared to co-vary with different substances in the samples. Despite significant correlations between amphipod survival and the concentrations of copper and DDT, none of the bioassay results were significantly different from controls. Sea urchin development was significantly correlated only with un-ionized ammonia in the porewater and zero percent normal development occurred in some samples with relatively high ammonia concentrations. Urchin fertilization was correlated with a number of trace metals, DDT, and ammonia and the concentrations of some metals and DDT exceeded numerical guideline values. Microtox ${ }^{\text {TM }}$ test results were highly correlated with complex mixtures of substances, including many trace metals and organic compounds. Microtox ${ }^{\text {™ }}$ test results showed a strong association with the cumulative ERM quotients, again, suggesting that microbial bioluminescence responded to complex mixtures of substances in the organic solvent extracts.

All Areas Combined. To determine if any of the toxicity/chemistry relationships observed in the individual bays also were significant throughout the entire study area, the data from all four bays were combined and correlations were calculated. Correlations between bioassay results and chemical concentrations for all 102 western Florida samples are summarized in Table 28. No data are included for the amphipod tests because none of the correlations were significant. Correlations are shown for major elements, classes of organic compounds, and organic compounds for which national sediment quality criteria have been proposed.

In the Microtox ${ }^{\text {TM }}$ tests, the strongest correlations were noted with concentrations of total DDT, total pesticides, and the sum of the chlorinated hydrocarbon/ERM quotients (Table 28). Concentrations of five trace metals, the sums of classes of individual PAHs, and total PCBs were correlated with Microtox ${ }^{\text {TM }}$ results. The sums of all 25 chemicals/ERM quotients were correlated with toxicity in this test, although the correlation with the sums of the

9 metals/ERM quotients were not significant. Likewise, correlations with the sums of the 5 simultaneouslyextracted metals were not significant. Among the strongest correlations was that of Microtox ${ }^{\top M}$ test results and the concentrations of total DDTs (Figure 88). Microtox ${ }^{\text {TM }}$ results showed considerable scatter and variability at DDT concentrations below the PEL value of 51.7 ppb ; however, as DDT concentrations increased above 51.7 ppb, all samples showed very high toxicity in this test. Two samples from Tom's Bayou adjacent to Choctawhatchee Bay had the highest DDT concentrations and were highly toxic in the microbial bioluminescence tests.

Urchin fertilization was highly correlated with the sum of concentrations of all PAHs ("sum all PAHs", Table 28). This sum included the parent aromatic hydrocarbons (sum PAHs) and many classes of substituted compounds, including alkylated- and methylated-compounds, and sulfurous compounds. The data illustrated in Figure 89 display a pattern of decreasing fertilization success in samples with total PAH concentrations of approximately 25000 or greater. There is no ERM or PEL value for the sum of all PAHs. Fertilization success also was significantly correlated with the concentrations of un-ionized ammonia, and to a lesser degree, many trace metals, total DDT, and the sum of the 25 chemical/ERM quotients.

Urchin development was highly correlated with un-ionized ammonia in the porewater test chambers (Table 28). This was the highest and most significant correlation coefficient observed for the combined data set. The data illustrated in Figure 90 show a sharp decrease in normal embryo development as un-ionized ammonia concentrations approach and exceed the LOEC concentration of 90 ug/l. Ten of the 102 samples exceeded the unionized ammonia LOEC concentration and all were highly toxic in tests of $100 \%$ porewater. Urchin embryo development also was correlated with the concentrations of copper, selenium, zinc, the sum of all PAHs, two individual PAHs normalized to organic carbon, and dieldrin normalized to organic carbon.

## DISCUSSION

This survey was conducted over a two-year period in the late spring of both 1993 and 1994 and extended throughout four large bays and adjoining bayous in the panhandle of western Florida. The objectives of the survey were to determine: (1) spatial patterns in toxicity throughout each bay, (2) the spatial extent of toxicity throughout and among bays, (3) the severity or degree of toxicity, (4) and relationships between chemical contamination and toxicity. Surficial sediments were collected to represent the quality of recently-deposited sediments. Four toxicity tests were conducted on each of the 123 samples collected in the four bays. A fifth toxicity test for genotoxic response was performed on samples collected in 1994. Chemical analyses were done on most (102) of the samples.

Chemical Concentrations in Sediments. Before the survey was conducted, chemical data available from previous studies in this region were reviewed to identify areas (including "hotspots") that contained levels of contaminants (and mixtures of contaminants) that had high potential to cause harm in marine ecosystems. These historical chemistry data indicated that toxicant concentrations were relatively high in the urbanized bayous of Pensacola, Choctawhatchee, and St. Andrew bays. Chemical concentrations were particularly high in Bayou Chico, and to a lesser degree, Watsons Bayou, Bayou Texar, Garnier Bayou, Boggy Bayou, and Massamino Bayou.

In these previous studies, concentrations of total DDT and other chlorinated organic hydrocarbons were extremely high at some locations in Choctawhatchee Bay. Other substances reported as occurring in high concentrations at several locations in the study area included lead, arsenic, other trace metals, total PCBs, and total PAHs. The hypothesis was, then, that in the presence of elevated toxic chemical concentrations, we would expect concomitant elevated incidence and severity of toxicity in these urbanized bayous.

Stratified-random sampling designs similar to the probabalistic designs of EPA's Environmental Monitoring and Assessment Program (EMAP), were used in the selection of sampling stations. Samples were collected both from strata with historically high to moderate chemical concentrations, where toxicity was expected, and from strata in which historic chemical concentrations were low, therefore, where toxicity was not expected.

Sediment chemistry data from this survey indicated concentrations of total PCBs, total pesticides, and total DDTs differed considerably among the four bays. Samples from Pensacola, St. Andrew, and Choctawhatchee bays had considerably higher concentrations of these compounds than samples from Apalachicola Bay. A simi-
lar pattern was evident with lead. Total mercury showed a different spatial distribution; concentrations of mercury were relatively high in samples from Pensacola and St. Andrew bays, intermediate in Choctawhatchee Bay, and lowest in Apalachicola Bay. Concentrations of PAHs were relatively high in samples from Pensacola Bay (especially in Bayou Chico), intermediate in samples from St. Andrew and Choctawhatchee bays, and lowest in samples from Apalachicola Bay. The maximum and minimum concentrations of total PAHs differed among all samples by over four orders of magnitude.

Some regions were clearly more contaminated with complex mixtures of substances than others. In Pensacola Bay, concentrations of PAHs, PCBs, mercury, and lead were elevated in three urban bayous (Bayou Chico, Bayou Texar, and Bayou Grande) when compared to the main basin of the system. Samples from Bayou Chico had especially high concentrations of many substances. In Choctawhatchee Bay, a similar pattern was evident; chemical concentrations were highest in the adjoining bayous (Garnier Bayou, Rocky Bayou, and Boggy Bayou) and lowest in the main basin of the bay. Chemical concentrations in St. Andrew Bay were highest in Watson's Bayou and Massamino Bayou when compared to other regions of the system. No clear pattern in contamination was evident in Apalachicola Bay, as all samples had relatively low chemical concentrations.

Based upon normalization to aluminum content (Schropp et al., 1990 ; Schropp and Windom, 1988), trace metals concentrations in all bays sampled during this survey except Apalachicola Bay were anthropogenically enriched. Pensacola Bay and Bayou Chico had the highest number of exceedances of background concentrations overall, followed by Choctawhatchee and St. Andrew Bays. The chemicals that most frequently exceeded background levels were cadmium, copper, lead, mercury and zinc. In addition, many substances equalled or exceeded effects-based, numerical guidelines (PEL and/or ERM values), indicating a high probability that they contributed to toxicity. Samples with chemical concentrations higher than effects-based numerical guidelines would be more likely to be toxic than those with all chemical concentrations below these levels. Notable among these substances were zinc, many individual PAHs, several isomers of DDT, total DDT, total PCBs, and unionized ammonia. Overall, the chemical data suggested that toxicity would be most probable and severe in Bayou Chico, somewhat less likely and severe in the other urban bayous, and least likely in the main basins of all four bays.

Incidence and Severity of Toxicity. Surficial sediment samples were collected from 123 randomly-chosen locations throughout the four bays. Toxicity was determined using a battery of four laboratory tests performed on all samples: (1) percent survival of marine amphipods (Ampelisca abdita) in 10-day tests of solid-phase (bulk) sediments; (2) changes in bioluminescent activity of a marine bacterium, Photobacterium phosphoreum, in 5minute assays of organic extracts; (3) fertilization success of the sea urchin Arbacia punctulata in one hour tests of the sediment porewater; and (4) normal embryological development of $A$. punctulata in 48-hour tests of the porewater. In addition, the Mutatox ${ }^{\top M}$ variant of the microbial bioluminescence test was performed on samples collected in Year 2 from Bayou Chico, Choctawhatchee and Apalachicola bays. The incidence and severity of toxicity differed considerably among the four tests and among the four bays.

Amphipod solid phase assay. The amphipod survival test showed a general lack of toxicity in all bays. From the total 123 samples, only $2.4 \%$ were toxic (i.e., significantly different from controls) and $0.8 \%$ were highly toxic (significantly different from controls and less than $80 \%$ of controls). Therefore, no spatial patterns in toxic effects could be measured from the results of this test. Only three stations throughout all four bays were significantly toxic in the amphipod test; one in Apalachicola Bay, one in Choctawhatchee Bay, and one in Bayou Chico in the Pensacola Bay estuary.

Microtox ${ }^{\text {TM }}$. In sharp contrast, the data from the Microtox ${ }^{T M}$ tests indicated that the majority of the samples from the four bays were toxic; 114 ( $92.7 \%$ ) of 123 samples were significantly different from controls. In all but one of these samples the test response was less than $80 \%$ of the controls. Test results ranged from $<1.0 \%$ to $>100 \%$ of control responses. Microbial bioluminescence EC50's were less than 10\% of controls in 79 (64\%) of the 123 samples. All except one sample from Apalachicola Bay were toxic and all except eight samples from Pensacola Bay were toxic. Nontoxic samples came from an upstream station in the Apalachicola River, several stations near the mouth of Pensacola Bay, and several stations scattered throughout Pensacola Bay.

Mutatox ${ }^{\text {TM }}$.An analysis of Mutatox ${ }^{\text {TM }}$ results revealed that $65.4 \%$ of the samples tested produced either a suspect (S category) or genotoxic (G category) test result and $40.4 \%$ produced a strong genotoxic response. All stations tested in Bayou Chico showed a genotoxic response. In contrast, only one of the nine samples from Apalachicola Bay showed a genotoxic response and five of the samples showed no genotoxicity.

Sea urchin fertilization. The incidence of toxicity in the urchin fertilization tests ( $100 \%$ porewater) was $10 \%$ in Pensacola Bay, $83.3 \%$ in Bayou Chico, $56.8 \%$ in Choctawhatchee Bay, $12.9 \%$ in St. Andrew Bay, and $44.4 \%$ in Apalachicola Bay. In $100 \%$ porewater, urchin fertilization was less than $50 \%$ in eight samples from Choctawhatchee Bay. Overall, among all 123 samples, $30.9 \%$ were toxic in this test. In $100 \%$ porewater, urchin fertilization ranged from $0.2 \%$ to $99.8 \%$ of controls. The overall incidence of toxicity diminished gradually in $50 \%$ and $25 \%$ porewater sample dilutions to $16.3 \%$ and $10.6 \%$, respectively. However, in Bayou Chico 5 of $6(83.3 \%)$ of the samples were both significantly toxic and highly toxic in all porewater concentrations; in addition, urchin fertilization was less than $1.0 \%$ of controls in 5 of the 6 samples of $100 \%$ porewater.

Sea urchin development.. Urchin development tests showed a wide range in test results. In tests of $100 \%$ porewater, normal urchin development was $0.0 \%$ in nine of twelve samples from Bayou Chico and in one sample from Bayou Texar. Zero normal development in $100 \%$ porewater also was observed in some samples from peripheral bayous of St. Andrew Bay and Choctawhatchee Bay. The two most toxic samples tested, both collected in upper Bayou Chico, showed zero normal development in all porewater concentrations. Overall, there was a relatively high incidence of toxicity in Choctawhatchee Bay (48.6\%), Apalachicola Bay (44.4\%) and Bayou Chico ( $100 \%$ ) in the embryo development tests as compared to Pensacola Bay ( $27.5 \%$ ) and St. Andrew Bay $(22.6 \%)$. Among the 123 samples tested, a total of $46(37 \%)$ were toxic in at least $100 \%$ porewater, a slightly higher proportion than observed in the fertilization tests.

Overall, the highest incidence of toxicity among all tests combined occurred among the Bayou Chico samples, followed by Choctawhatchee Bay, and the other areas. All of the Bayou Chico samples were toxic in the sea urchin development, Microtox ${ }^{T M}$, and Mutatox ${ }^{\top M}$ tests; all were highly toxic in the urchin development and Mutatox ${ }^{\text {TM }}$ tests; and all except one sample were highly toxic in the urchin fertilization tests. In addition, the only sample that was highly toxic in the amphipod survival tests was collected in Bayou Chico. In Choctawhatchee Bay all samples were toxic in Microtox ${ }^{\text {TM }}$ tests, most were toxic in Mutatox ${ }^{\text {TM }}$ tests, $57 \%$ were toxic in urchin fertilization tests, $49 \%$ were toxic in urchin development tests, and one sample was toxic in the amphipod tests. The incidence of toxicity in Pensacola Bay and St. Andrew Bay was relatively similar: zero and one sample toxic in amphipod tests (respectively); $10 \%$ and $13 \%$ toxic in urchin fertilization tests; $27 \%$ and $23 \%$ toxic in urchin development tests; and $80 \%$ and $100 \%$ toxic in Microtox ${ }^{\text {TM }}$ tests, respectively. Among all areas included in this survey, Apalachicola Bay had the lowest incidence of toxicity in all tests.

Spatial Extent of Toxicity. The data from the toxicity tests were weighted to the sizes of the strata within which they were collected. Therefore, the spatial significance of the toxicity could be estimated. Estimates of the spatial extent of toxicity were developed for all four bioassays for each of the four bays and for the entire survey area combined. Bioassay results that were less than $80 \%$ of control responses were treated as "toxic" in these calculations. The entire survey area encompassed approximately $840 \mathrm{~km}^{2}$.

These data suggest that amphipod survival was affected in a tiny portion of the entire study area ( $0.005 \%$ ), and in sharp contrast, microbial bioluminescence was affected in nearly all of the area ( $98.9 \%$ ). Both sea urchin fertilization and embryo development were affected in nearly one-half of Choctawhatchee Bay, and small portions of St. Andrew and Pensacola bays. In Apalachicola Bay, the majority of the area was affected in the development tests, whereas about one-third was affected in the fertilization tests. Throughout the entire study area, $23 \%$ was toxic in fertilization tests and $34 \%$ was toxic in the development tests. Usually, small portions of each bay were affected in both tests of $50 \%$ and $25 \%$ porewater.

As part of its Environmental Monitoring and Assessment Program (EMAP), U.S. EPA estimated that 7\% of the Louisianian estuarine province, which includes the western Florida panhandle, was toxic in laboratory tests with Ampelisca abdita (Summers et al., 1993). The Louisianian province extends from the Rio Grande, Texas to Anclote Key, Florida. The considerable difference in the estimated areas of toxicity (7\% by U.S. EPA and $0.005 \%$ by this study) is probably attributable to the differences in the two survey areas. In the EMAP-E studies, U.S. EPA included large riverine systems, especially the Mississippi River, and other urbanized regions that may be contaminated in their survey area, whereas in this study the survey area was restricted to four large bays of which only relatively small portions were highly urbanized. Nevertheless, both estimates suggest that toxicity as determined with the amphipod acute survival test was not extensive.

Spatial Patterns in Toxicity. Concordance among bioassay results was relatively poor and the four tests showed overlapping, but different patterns in toxicity. Spearman-rank correlations among tests were significant in only
three of 24 combinations of results. The most consistent correlations were those: (a) between sea urchin fertilization and microbial bioluminescence, (b) between the Mutatox ${ }^{\mathrm{TM}}$ and Microtox ${ }^{\mathrm{TM}}$ tests and (c) between both of the sea urchin tests.

Toxicity in the Microtox ${ }^{T M}$ tests was pervasive, extending throughout most or all of each bay. All of the samples from Choctawhatchee Bay, St. Andrew Bay, and Bayou Chico were significantly different from controls. Mean EC50 values were lowest, indicating highest toxicity, in samples from Bayou Chico and inner Pensacola Harbor in Pensacola Bay; Garnier Bayou, Destin Harbor, Boggy Bayou, Tom's Bayou, and Rocky Bayou in Choctawhatchee Bay; and Watsons Bayou and other portions of St. Andrew Bay. Therefore, toxicity in these tests followed the patterns often predicted by the available chemistry data from previous studies. Toxicity was relatively low in Apalachicola Bay compared to the other bays.

No spatial patterns in Mutatox ${ }^{\text {TM }}$ test results were evident in Bayou Chico, since all samples collected in 1994 were genotoxic. In Choctawhatchee Bay, genotoxic samples were most evident in the adjoining bayous, including Destin Harbor (D stations), Joe's Bayou (H stations), the two arms of Garnier Bayou, Boggy Bayou, Rocky Bayou, and La Grange Bayou. Most of the "M" and "N" series stations in the main basin of the bay were not genotoxic, and the three stations sampled near the north shore were suspected as genotoxic. One sample from Apalachicola Bay determined to be genotoxic was collected near the mouth of the Apalachicola River. Three samples, one from each sampling stratum in Apalachicola Bay, were suspected as genotoxic. Two samples collected near the mouth of the bay were not toxic.

The results of the sea urchin fertilization tests showed relatively high toxicity in several bayous of Choctawhatchee Bay, Watsons Bayou in St. Andrew Bay, and Bayou Chico of Pensacola Bay compared to the other areas, and relatively low toxicity in most of the main basin stations of Pensacola and St. Andrew bays. Most of the 1994 samples from Bayou Chico were highly toxic in all porewater concentrations, whereas none collected in 1993 was toxic in any porewater concentrations. Two samples each in the lower Apalachicola River and lower Apalachicola Bay were toxic in 100\% porewater.

Stations exhibiting toxicity in the embryo development tests were largely associated with urbanized tributaries to the bays in all cases except Apalachicola Bay. Toxicity was especially apparent in Bayou Chico, Watson Bayou, the Pensacola Harbor, Destin Harbor, and portions of Garnier Bayou.

Toxicity/Chemistry Relationships. The relationships between toxicity measured in the four bioassays and solid-phase (bulk) sediment chemistry were explored in a multistep approach. The causes of toxicity could not be determined in these studies. Additional, experimental work would be needed to tease out the substances that caused or significantly contributed to toxicity. Instead, the relative probability or likelihood of different substances contributing to toxicity was determined with analyses of the matching toxicity and chemistry data. Data from each of the four bays were treated separately to identify bay-specific toxicity/chemistry relationships and the data from all areas were combined to identify broad-scale relationships, if any.

In all cases, there was no single chemical or class of chemicals that stood out from the others as the primary cause of toxicity. The chemicals that were most associated with toxicity differed among the four bays. Furthermore, chemicals most associated with toxicity differed among the four different toxicity tests. In all cases it was apparent that complex mixtures of substances co-varied with toxicity and probably contributed to the toxic responses.

In Pensacola Bay, zinc, high molecular weight PAHs, two DDD/DDT isomers, total DDT, and dieldrin were most closely associated with toxicity. To a lesser extent, cadmium, copper, lead, low molecular weight PAHs, and in the case of urchin embryo development, unionized ammonia, were moderately associated with toxicity. The significant correlations between three measures of toxicity (Microtox ${ }^{\mathrm{TM}}$, urchin fertilization, urchin development) and cumulative ERM quotients suggests that these substances acted together in complex mixtures to contribute to toxicity. In addition, high levels of nitro aromatic compounds were found in samples that masked much of the PAH signal, and were not accounted for in the analyses of chemistry/toxicity correlations.

Spearman-rank correlations failed to show significant correlations between toxicity in samples from Bayou Chico and Apalachicola Bay, probably largely because of the small sample sizes. However, there were numerous obvious (but nonsignificant) associations between elevated chemical levels and toxicity. The concentrations of
zinc and many organic compounds, notably the PAHs and to a lesser extent some chlorinated organic hydrocarbons, were relatively high in these samples, especially those that were toxic in the urchin fertilization tests. The toxicity of one sample with high chemical concentrations may have been inhibited by high concentrations of organic carbon which may have reduced the bioavailability of the organic compounds. Samples from Bayou Chico had considerably higher chemical concentrations than those from Apalachicola Bay, as expected, and often higher concentrations than applicable numerical guidelines.

In Choctawhatchee Bay, the associations between toxicity and concentrations of potentially toxic substances were strongest for the urchin fertilization tests in which a large gradient in response was observed. Amphipod and Microtox ${ }^{\top M}$ test results were not correlated with any substances. Urchin embryo development test results were correlated with only three substances: unionized ammonia and two individual PAHs. In sharp contrast, numerous substances were correlated with urchin fertilization. Most notable among these were the concentrations of DDT isomers, total DDT, silver, the sum of PAHs, and dieldrin. However, the concentrations of these substances exceeded TEL and ERL values by relatively small amounts and rarely equaled or exceeded respective PEL and ERM values. Therefore, there is insufficient evidence to suggest which substances individually may have contributed substantially to toxicity.

The four toxicity endpoints measured in St. Andrew Bay appeared to co-vary with different substances in the samples. Despite significant correlations between amphipod survival and the concentrations of copper and DDT, none of the bioassay results were significantly different from controls. Sea urchin development was significantly correlated only with unionized ammonia in the porewater and zero percent normal development occurred in some samples with relatively high ammonia concentrations. Urchin fertilization was correlated with a number of trace metals, DDT, and ammonia and the concentrations of some metals and DDT exceeded numerical guideline values. Microtox ${ }^{\text {TM }}$ test results were highly correlated with complex mixtures of substances, including many trace metals and organic compounds. Microtox ${ }^{\mathrm{TM}}$ test results showed a strong association with the cumulative ERM quotients, again suggesting that microbial bioluminescence responded to complex mixtures of substances in the organic solvent extracts.

Although toxicity/chemistry relationships differed considerably among the four tests and four bays, there were a few common threads in these associations. In all four bays, the concentrations of selected trace metals (notably zinc, silver, and copper), DDT and its isomeric derivatives, and selected PAHs were associated with at least one measure of toxicity. Also, dieldrin and unionized ammonia showed associations with toxicity. In Pensacola Bay and St. Andrew Bay the cumulative ERM index, which takes into account the concentrations of 24 substances, showed a strong association with toxicity.

Some of the common associations observed in each bay were borne out in the correlations performed with the combined toxicity/chemistry data set of 102 samples. Microtox ${ }^{T M}$ test results were highly correlated with the concentrations of chlorinated compounds, including DDTs, PCBs, and total pesticides. To a lesser degree Microtox ${ }^{T M}$ test results were also significantly correlated with the concentrations of some trace metals and PAHs. Urchin fertilization was highly correlated with the concentrations of PAHs, numerous trace metals, and DDT. Urchin embryo development was primarily correlated with the concentrations of unionized ammonia, and to a lesser degree, PAHs, three trace metals, and one pesticide, dieldrin. It is significant that the results of the Microtox ${ }^{T M}$ and urchin fertilization tests were significantly correlated with the sum of the cumulative ERM quotients, since these quotients account for the contribution of 25 different substances to toxicity. The correlations strongly suggest that mixtures of toxicants, co-varying with each other, contributed significantly to the observed toxicity. This association, on the other hand, was not observed in the embryo development tests, in which ammonia was obviously a likely contributor to toxicity.

The differences in toxicity/chemistry relationships among the bays and toxicity tests are expected and have been observed in previous studies in other major estuaries of the USA (Long et al., 1994; 1996). The species of organisms used in the toxicity tests often display different responses to the many substances that occur in sediments. Different species have different responses to the same chemicals. The amphipod, sea urchin, and microbial bioluminescence toxicity tests were performed with three different phases of the sediments; the solidphase, porewater phase, and an extract prepared with an organic solvent. Therefore, substances bound to the sediment particles are expected to differ in their relative bioavailability - and toxicological response - among the different phases. Finally, there is a strong possibility that the substances quantified in the chemical analyses covaried with mixtures of chemicals that were not quantified. It is likely that in any survey, including the present
study, that there will be other unmeasured constituents that can add significantly to the toxic response, with no way to account for the extent of the contribution.

Relevance of Different Toxicity Tests. The ecological significance of sediment toxicity tests performed with amphipods has been established in correlative sediment quality studies performed elsewhere, including Commencement Bay, the Palos Verdes shelf and San Francisco Bay, California (Swartz et. al., 1982, 1986, 1994, respectively). Those studies indicated that resident benthic communities often were altered at locations shown to be highly toxic to the amphipod Rhepoxinius abronius in laboratory tests. Alterations to resident communities usually consisted of diminished abundance or a lack of certain sensitive crustacean groups, especially burrowing amphipods, relative to stations with high amphipod survival in laboratory tests. Other studies in San Francisco Bay (Chapman et. al., 1987) and Puget Sound (Long and Chapman, 1985) have shown similar results; that is missing or depauperate crustacean abundances in stations shown to be toxic in laboratory tests to amphipods. In Tampa Bay, some stations in which toxicity to Ampelisca abdita was observed had severely diminished benthic populations, but some of these alterations may have been attributable to local hypoxic conditions (Coastal Environmental, Inc., 1966). In estuaries, benthic populations are under frequent stress from short-term changes in salinity, cyclical hypoxia and/or ammonia events, and other natural factors such as depth, slope, sediment texture, and predators. Therefore, attribution of benthic population changes to only anthropogenic toxicants, as suggested in laboratory toxicity tests, is a difficult analytical step. Consequently, some have argued that toxicity tests are sufficiently robust to stand alone without the need for accompanying benthic data (Chapman, 1995).

Field validation data for the urchin and Microtox ${ }^{\top M} / M_{\text {Mato }}{ }^{\top M}$ tests currently are not available. However, in Tampa Bay the areas in which toxicity was most severe in both these and the amphipod tests had the most severely altered benthic populations (Coastal Environmental, Inc., 1996). Fertilization and developmental tests of urchin gametes and embryos exposed to the porewaters provide highly sensitive assays of sediment samples. These tests combine evidence of the disruption in essential reproductive functions in the test animals with exposures to the porewater in which toxicants are mainly in a dissolved state and, therefore, readily bioavailable. These are sublethal test endpoints, not acute toxicity tests. They can be viewed as indicators of slightly to moderately degraded conditions that, if allowed to deteriorate, may lead to toxicity expressed in the acute amphipod tests. They should be considered as important instruments for environmental managers in the prevention of further harm to environmental quality. As expected, based upon experience in previous surveys, these two tests showed more sensitivity than the amphipod tests.

The highly sensitive Microtox ${ }^{T M}$ test can be best viewed as an indicator of the potential for biological effects. Because the tests are performed with solvent extracts of the sediments, all solvent-extractable substances in the samples are potentially drawn out or extracted from the sediment matrix, and, therefore, artificially made bioavailable in the tests. Therefore, the test organisms are probably exposed to a greater dose of toxicants than the amphipods or urchins. In addition, the test is conducted with an assay of metabolic activity, not acute mortality, as in the amphipod tests. This test, therefore, is expected to be triggered by much lower chemical concentrations than the other assays. These microbial tests can be regarded as a flag or precursor for further harm in the environment. The Mutatox ${ }^{\top M}$ variant of this test indicates the presence of potentially mutagenic substances in the samples, most of which are not highly toxic in acute mortality tests, but would be highly problematic when one considers long-term impacts to a healthy, well balanced biotic population .

## CONCLUSIONS

- The concentrations of many different substances in some samples collected during this survey exceeded applicable toxicity thresholds or guideline values. Noteworthy among these chemicals were copper, lead, zinc, many high molecular weight PAHs, total PAHs, total DDT, several isomers of DDT, and dieldrin. Trace metals concentrations in many samples exceeded background levels.
- Chemical concentrations, on average, were most elevated in Pensacola, Choctawhatchee, and St. Andrew bays and lowest in Apalachicola Bay. Within these bays, contamination was highest in Bayou Chico, followed by several other bayous, all of which exceeded the main basins of all bays in contaminant levels. Samples from Bayou Chico equalled or exceeded guideline concentrations for the greatest number of substances and all samples there exceeded background concentrations of trace metals.
- All five laboratory tests indicated the presence of toxicity in western Florida samples. Amphipod survival was the least sensitive test, indicating highly significant toxicity in only one of the 123 samples ( $0.8 \%$ of the total). Microtox ${ }^{\top \mathrm{M}}$ tests, in contrast, indicated that $91.9 \%$ of the samples were highly toxic. In the sea urchin tests of $100 \%$ porewater, $26.0 \%$ and $35.0 \%$ of the samples were highly toxic in assays of fertilization success and normal embryological development.
- Some of the tests, notably the two urchin tests, the Microtox ${ }^{\text {TM }}$ and Mutatox ${ }^{\text {TM }}$ tests, and the Microtox ${ }^{\text {TM }}$ and amphipod survival tests, showed relatively strong concordance with each other, whereas the others showed no or weak concordance. The differences in toxicity among the tests were attributable to differences in the sediment phases tested and the differential sensitivities of the test organisms.
- The entire four bay survey area encompassed approximately $850 \mathrm{~km}^{2}$ of the western Florida panhandle. The spatial extent of toxicity throughout this area was estimated for each test except Mutatox ${ }^{\text {TM }}$. Approximately $90 \%$ of the area represented in the survey was toxic in the Microtox ${ }^{\text {TM }}$ test, $34 \%$ was toxic in the urchin development test, $23 \%$ was toxic in the urchin fertilization test, and $0.005 \%$ was toxic in the amphipod survival test.
- In 30 independent trials ( 6 samples, 5 toxicity tests), 23 ( $76.7 \%$ ) of the tests showed highly significant results in Bayou Chico. This area was clearly the most toxic region of the study area. The overall incidences of highly significant toxicity in the other areas were: $45.4 \%$ ( 84 of 185 trials) in Choctawhatchee Bay, $37.8 \%$ (17 of 45 trials) in Apalachicola Bay, 33.1\% (41 of 120 trials) in St. Andrew Bay, and 28.1\% (45 of 160 trials) in Pensacola Bay.
- Overall, the incidence and severity of toxicity were higher in Bayou Chico, an industrialized basin adjoining Pensacola Bay, than in all other bays. All of 1994 samples from Bayou Chico were highly toxic in the urchin embryo and Mutatox ${ }^{\text {TM }}$ tests. All except one sample was highly toxic in the Microtox ${ }^{\text {TM }}$ tests. The only sample in the entire survey showing a highly significant result in the amphipod tests was collected in Bayou Chico.
- Other bayous in which toxicity was apparent included Watson's Bayou and Massamino Bayou adjoining St. Andrew Bay; Garnier Bayou and Tom's Bayou adjoining Choctawhatchee Bay; and Bayou Grande and Bayou Texar adjoining Pensacola Bay. Toxicity also was apparent in the Pensacola Harbor and harbor entrance.
- The mixtures of chemical substances associated with toxicity differed among tests and among the four bays. Overall, however, the concentrations of DDT, ammonia, several trace metals (notably copper), and PAHs were most frequently associated with toxicity throughout the entire survey area.
- Overall, within the combined study area the Microtox ${ }^{T M}$ test results were most correlated with the concentrations of DDT and other pesticides, and to a lesser degree, with the concentrations of several trace metals and PAHs. Notably, this test showed a strong association with the sums of 25 toxicants normalized to their respective guideline values. In Pensacola Bay test results were highly correlated with cadmium, copper, lead, and zinc. None of the substances measured was correlated with Microtox ${ }^{\text {TM }}$ results in Choctawhatchee Bay. However, in St. Andrew Bay numerous trace metals, individual PAHs, classes of PAHs, chlordane, other pesticides and DDT were correlated with test results.
- Sea urchin fertilization showed a strong association with porewater ammonia, numerous trace metals and the sum of all PAHs quantified. In Pensacola Bay fertilization success was correlated with a mixture of trace metals, PAHs and a few pesticides. The concentrations of these substances were particularly elevated in the highly toxic samples collected in Bayou Chico. Similarly, numerous trace metals, many PAHs, PCB congeners, total PCBs, DDT and other pesticides were correlated with urchin fertilization success in Choctawhatchee Bay. In St. Andrew Bay, fewer substances were correlated with urchin fertilization, notably including unionized ammonia, several trace metals, total DDT and two DDT isomers.
- In the combined study area, sea urchin development was highly correlated with ammonia and, to a considerably lesser degree, three trace metals, PAHs, and dieldrin. In Pensacola Bay, embryo development was highly correlated with unionized ammonia, and to a lesser degree, many trace metals, PAHs, and total DDT. Zinc, PAH, DDT, and dieldrin concentrations in toxic samples exceeded applicable guideline concentrations in some Pensacola Bay samples. Although most of the samples from Bayou Chico exceeded guideline values for many substances, none of the correlations with urchin embryo development were significant. In both Choctawhatchee and St.

Andrew bays, urchin development was correlated strongly with unionized ammonia concentrations, which exceeded toxicity threshold values in numerous samples.

- The ecological relevance of the toxicity tests differs among the different bioassays. Severe toxicity in amphipod tests has been associated with altered benthic populations in several field studies. Therefore, this test may be an indicator of relatively degraded conditions in the environment. Results of the toxicity survey performed in the four western Florida bays suggest that resident benthic communities may be impacted in some of the regions, especially in the urbanized bayous in which chemical concentrations were highest and toxicity was most severe. The sea urchin test indicates reduced reproductive success of a sensitive marine invertebrate in exposures to the porewaters, a component of sediments in which toxicants are readily bioavailable. In the Microtox ${ }^{\text {TM }}$ tests, toxicants are drawn out of the sediment matrix with an organic solvent, and, therefore, are indicative of the potential for toxicity in the samples. The Mutatox ${ }^{T M}$ tests provide information on the mutagenic potential of sediment-associated toxicants.

In summary, the incidence of toxicity, whether acute or otherwise indicative of chronic problems, should be used to draw attention to these systems whose capability to sustain a healthy, well balanced benthic population is compromised. Changes in upland land management practices and better source controls are needed to prevent long term harm.

## REFERENCES

ASTM. 1992. Standard Guide for Conducting 10-day Static Toxicity Test with Marine and Estuarine Amphipods. Designation E 1367-92. Annual Book of Standards. 11.04. American Society for Testing and Materials. Philadelphia, PA.

Carr, R.S. 1993. Sediment Quality Assessment Survey of the Galveston Bay System. Galveston Bay National Estuary Program Report, GBNEP-30, 101 pp.

Carr, R.S. and D.C. Chapman. 1992. Comparison of solid-phase and porewater approaches for assessing the quality of marine and estuarine sediments. Chem. Ecol. 7:19-30.

Carr, R. S., E. R. Long, H. L. Windom, D. C. Chapman, G. Thursby, G. M. Sloane, and D. A. Wolfe. 1996. Sediment quality assessment studies of Tampa Bay, Florida. Environ. Toxicol. Chem 15(7): 1218-1231.

Chapman, P. M. 1995. Do sediment toxicity tests require field validation? Envir. Toxicol. \& Chem. 14 (9): 14511453.

Chapman, P. M., R. N. Dexter, and E. R. Long. 1987. Synoptic measures of sediment contamination, toxicity and infaunal community composition (the sediment quality triad) in San Francisco Bay. Marine Ecology-Progress Series 37: 75-96.

Coastal Environmental, Inc. 1996. An Assessment of Sediment Contamination in Tampa Bay, Florida. Draft report submitted to Tampa Bay National Estuary Program. St. Petersburg, FL.

Hamilton, M. A., R. C. Russo, and R. V. Thurston. 1977. Trimmed Spearman-Karber method for estimating median lethal concentrations in toxicity bioassays. Environ. Sci. Technol. 11: 714-719; correction, 1978: 12: 417.

Holland, A. F., editor. 1990. Near Coastal Program Plan for Estuaries. U.S. EPA600.14-90-033. U.S. EPA Office of Research and Development, ERL-Narragansett, R.I.

Johnson, B.T. 1992. An evaluation of genotoxicity assay with liver S 9 for activation and luminescent bacteria for detection. Environ. Toxicol. Chem. 11:473-480.

Johnson, B.T. 1993. Activated Mutatox ${ }^{\text {TM }}$ assay for detection of genotoxic substances. Environ. Toxicol. Water Qual. 8:108-113.

Kohn, N. P., J.Q. Word, D. K. Niyogi. 1994. Acute Toxicity of Ammonia to four species of Marine Amphipod. Mar. Env. Res. 38(1994):1-15.

Lauenstein, G. G. and A. Y. Cantillo, editors. 1993. Sampling and Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects. 1984-1992. NOAA Tech. Memo. NOS ORCA 71. National Oceanic and Atmospheric Administration. Silver Spring, MD.

Long, E. R. and P. M. Chapman. 1985. A sediment quality triad: Measures of sediment contamination, toxicity, and infaunal community composition in Puget Sound. Mar. Poll. Bull. 16(10): 4-5-415.

Long, E. R. and L.G. Morgan. 1990. The Potential for Biological Effects of Sediment-sorbed Contaminants Tested in the National Status and Trends Program. NOAA Technical Memorandum NOS OMA 52. National Oceanic and Atmospheric Administration. Seattle, Washington. 175 pp. + appendices.

Long, E. R. and M. F. Buchman. 1989. An Evaluation of Candidate Measures of Biological Effects for the National Status and Trends Program. NOAA Tech. Memo. NOS OMA 45. National Oceanic and Atmospheric Administration, Seattle, WA. 106 pp.

Long, E. R. and R. Markel. 1992. An Evaluation of the Extent and Magnitude of Biological Effects Associated with Chemical Contaminants in San Francisco Bay, California. NOAA Tech. Memo. NOS ORCA 64. National Oceanic and Atmospheric Administration, Seattle, WA. 86 pp.

Long, E. R., D. A. Wolfe, R. S. Carr, K. J. Scott, G. B. Thursby, H. L. Windom, R. Lee, F. D. Calder, G. M. Sloane, and T. Seal. 1994. Magnitude and Extent of Sediment Toxicity in Tampa Bay, Florida. NOAA Tech. Memo. NOS ORCA 78. National Oceanic and Atmospheric Administration, Silver Spring, MD. 138 pp.

Long, E. R., D. A. Wolfe, K. J. Scott, G. B. Thursby, E. A. Stern, C. Peven, and T. Schwartz. 1995. Magnitude and Extent of Sediment Toxicity in the Hudson-Raritan Estuary. NOAA Tech. Memo. NOS ORCA . National Oceanic and Atmospheric Administration, Silver Spring, MD. 230 pp.

Long, E. R., G. M. Sloane, R. S. Carr, K. J. Scott, G. B. Thursby, and T. Wade. Sediment Toxicity in Boston Harbor: Magnitude, Extent and Relationships with Chemical Toxicants. NOAA Tech. Memo. NOS ORCA 96. 133 pp.

MacDonald, D. D. 1993. Development of an Approach to Assessing Sediment Quality in Florida Coastal Waters. Report prepared for Florida Department of Environmental Regulation.

MacDonald, D. D. 1994. Approach to the assessment of sediment quality in Florida coastal waters. 4 Volumes and appendices. Report prepared for the Florida Department of Environmental Protection.

MacLeod, W.D., Jr., D.W. Brown, A.J. Friedman, O. Maynes, and R. W. Pierce. 1985. Standard Analytical Procedures of the NOAA National Analytical Facility, 1984-85. Extractable Toxic Organic Compounds. NOAA Tech. memo. NMFS F/NWC-64. National Marine Fisheries Service, Seattle, WA. 86 pp.

Mayhew, H.L., J.Q. Word, N.P. Kohn, M.R. Pinza, L.M. Karle and J.A. Ward. 1993. Ecological Evaluation of Proposed Dredged Material from St. Andrew Bay, Florida. Report PNL-8894/UC-000, 94 pp. and appendices. Prepared for the U.S. Army Corps of Engineers.

Morgan, B. J. T. 1992. Analysis of Quantal Response Data. Chapman and Hall, London, UK.
Moser, B. K. and G. R. Stevens. 1992. Homogeneity of variance in the two-sample means tests. American Statistician 46: 19-21.

National Oceanic and Atmospheric Administration. 1985. National Estuarine Inventory Data Atlas. Volume 1. Strategic Assessment Branch, Ocean Assessment Division.

National Oceanic and Atmospheric Administration. 1989. National Status and Trends Program for Marine Environmental Quality. Progress Report. A Summary of Data on Tissue Contamination from the First Three Years (1986-1988) of the Mussel Watch Project. NOAA technical Memorandum NOS OMA 49. Rockville, MD. 22 pp. + appendices.

National Oceanic and Atmospheric Administration. 1991. National Status and Trends Program for Marine Environmental Quality. Progress Report. Second Summary of Data on Chemical Contaminants in Sediments from the National Status and Trends Program. NOAA Technical Memorandum NOS OMA 59. Rockville, MD. 29 pp. + appendices.

National Oceanic and Atmospheric Administration. 1992. The National Status and Trends Program. Marine Environmental Quality. NOAA Special Publication. U.S. Department of Commerce, NOAA/NOS , Rockville, MD. 18 pp .

Northwest Florida Water Management District. 1990. S.W.I.M. Plan for Pensacola Bay, Florida.
O'Connor, T. P. 1991. Concentrations of organic contaminants in mollusks and sediments at NOAA National Status and Trend sites in coastal and estuarine United States. Envir. Health Perspectives 90:69-73.

O'Connor, T. P. and C. N. Ehler. 1991. Results from the NOAA National Status and Trends Program on distribution and effects of chemical contamination in the coastal and estuarine United States. Envir. Monit. \& Assessment 17:33-49.

Paulic, M. and J. Hand. 1994. Florida Water Quality Assessment 1994305 (b) Report. Miami: Florida Department of Environmental Protection. 261 pp. + technical appendices.

Sapudar, R. A., C. J. Wilson, M. L. Reid, E. R. Long, M. Stephenson, M. Puckett, R. Fairey, J. Hunt, B. Anderson, D. Holstad, J. Newman, and S. Birosik. 1994. Sediment chemistry and toxicity in the vicinity of the Los Angeles and Long Beach Harbors. California State Water Resources Control Board, Sacramento, CA. 77 pp.

SAS Institute, Inc. 1992. SAS/STAT User's Guide, Version 6, Fourth Edition, vol. 2. Cary, NC: SAS Institute, Inc. 846 pp.

Schimmel, S. C., B. D. Melzian, D. E. Campbell, C. J. Strobel, S. J. Benyi, J. S. Rosen, and H. W. Buffum. 1994. Statistical summary EMAP-Estuaries Virginian Province - 1991. EPA/620/R/94/005. U.S. Environmental Protection Agency, Narragansett, RI. 77 pp.

Schropp, S.J. and H.L. Windom. 1988. A guide to the Interpretation of Metal Concentrations in Estuarine Sediments. Florida Department of Environmental Regulation, Tallahassee, FI. 44 pp. + appendices.

Schropp, S. J., F. G. Lewis, H. L. Windom, J. D. Ryan, F. D. Calder, and L. C. Burney. 1990. Interpretation of metal concentrations in estuarine sediments of Florida using aluminum as a reference element. Estuaries 13(3):227-235.

Seal, Thomas L., Calder, F.D., Sloane, G. M., Schropp, S. J., Windom, H.L. 1994. Florida Coastal Sediment Contaminants Atlas. 112 pp. + technical appendices.

Sericano, J. L., E. L. Atlas, T. L. Wade, and J. M. Brooks. 1990. NOAA's Status and Trends Mussel Watch Program: Chlorinated pesticides and PCBs in oysters (Crassostrea virginica) and sediments from the Gulf of Mexico, 1986-1987. Mar. Envir. Res. 29:161-203.

Smith, R.G. 1993. Determination of mercury in environmental samples by isotope dilution/ICPMS. Analytical Chemistry 65:2485-2489.

Summers, J. K., J. M. Macauley, P. T. Heitmuller, V. D. Engle, A. Matt Adams, G. T. Brooks. 1993. Statistical Summary: EMAP-Estuaries. Louisianian Province - 1991. U. S. EPA/600/R-93-001. U. S. Environmental Protection Agency,Gulf Breeze, FL. 101 pp.

Swartz, R. C., W. A. DeBen, K. A. Sercu, and J. O. Lamberson. 1982. Sediment toxicity and the distribution of amphipods in Commencement Bay, Washington, USA. Mar. Pollu. Bull. 13: 359-364.

Swartz, R. C., F, A, Cole, D. W. Schults, and W. A. DeBen. 1986. Ecological changes on the Palos Verdes shelf near a large sewage outfall: 1980-1983. Marine Ecology-Progress Series 31: 1-13.

Swartz, R. C., F. A. Cole, J. O. Lamberson, S. P. Ferraro, D. W. Schults, W. A. DeBen, H. Lee II, and R. J. Ozretich. 1994. Sediment toxicity, contamination and amphipod abundance at a DDT-and dieldrin-contaminated site in San Francisco Bay. Envir. Toxicol. \& Chem. 13(6): 949-962.
U.S. EPA. 1986. Recommended protocols for conducting laboratory bioassays on Puget Sound sediments. 52 pp. Prepared for U.S. Environmental Protection Agency, Region 10, Puget Sound Estuary Program, Seattle, WA.
U.S. EPA. 1990. Recommended protocols for conducting laboratory bioassays on Puget Sound sediments. Final Report. 79 pp. Prepared for U.S. Environmental Protection Agency, Region 10, Puget Sound Estuary Program, Seattle, WA.
U.S. EPA. 1994. Recommended guidelines for conducting laboratory bioassays on Puget Sound sediments. 81 pp. Prepared for U.S. Environmental Protection Agency, Region 10, Puget Sound Estuary Program, Seattle, WA.

Wolfe, D. A. 1992. Selection of bioindicators of pollution for marine monitoring programmes. Chemistry and Ecology 6:149-167.

Wolfe, D. A., E. R. Long, and A. Robertson. 1993. The NS\&T bioeffects surveys: Design strategies and preliminary results. In: Proceedings, Coastal Zone ' 93 , the 8th Symposium on Coastal and Ocean Management Vol. 1: 298-312. O. T. Magoon, W. S. Wilson, H. Converse and L. T. Tobin (editors). New York. American Society of Civil Engineers.

Wolfe, D. A., S. B. Bricker, E. R. Long, K. J. Scott, and G. B. Thursby. 1994. Biological effects of toxic contaminants in sediments from Long Island Sound and environs. NOAA Tech. Memo. NOS ORCA 80. National Oceanic and Atmospheric Administration. Silver Spring, MD.

FIGURES


Figure 1. Study area encompassing Pensacola, Choctawhatchee, St. Andrew, and Apalachicola bays.


Figure 2. Total PAH concentrations in sediments from 10 NOAA NS \&T Program sampling sites.


Figure 3. Total DDT concentrations in sediments from 10 NOAA NS\&T Program sites.


Figure 4. Lead concentrations in sediments from 10 NOAA NS\&T Program sites.

## Pensacola Bay Organics



## Pensacola Bay Metals



Figure 5. Concentrations of organics and trace metals in sediments from NS\&T Program sites sampled in Pensacola Bay in 1991.

## Choctawhatchee Bay Organics



Choctawhatchee Bay Metals


Figure 6. Concentrations of organics and trace metals in sediments from NS\&T Program sites sampled in Choctawhatchee Bay in 1991.



Figure 7. Concentrations of organics and trace metals in sediments from NS\&T Program sites sampled in St. Andrew Bay in 1991.



Figure 8. Concentrations of organics and trace metals in sediments from NS\&T Program sites sampled in Apalachicola Bay in 1991.


Figure 9. Pensacola Bay study area.


Figure 10. Choctawhatchee Bay study area.


Figure 11. St. Andrew Bay study area.


Figure 12. Apalachicola Bay study area.


Figure 13. Locations of sampling stations in Pensacola Bay.


Figure 14. Locations of sampling stations in Bayou Chico.


Figure 15. Locations of sampling stations in Choctawhatchee Bay.


Figure 16. Locations of sampling stations in St. Andrew Bay.


Figure 17. Locations of sampling stations in Apalachicola Bay.


Figure 18. Maximum concentrations of major classes of organic compounds (total PAHs, total PCBs, total pesticides and total DDTs) in sediments from four western Florida bays.


Figure 19. Maximum, mean and minimum concentrations of total PAHs in western Florida bays.

20. Maximum, mean and minimum concentrations of total PCBs, total pesticides and total DDTs in western Florida bays.

21. Maximum, mean and minimum lead concentrations in western Florida bays.

22. Maximum, mean and minimum mercury concentrations in western Florida bays.

23. Distribution of total PAH concentrations among selected sampling stations in Pensacola Bay.

24. Distribution of total PCB concentrations among selected sampling stations in Pensacola Bay.

25. Distribution of mercury concentrations among selected sampling stations in Pensacola Bay.

26. Distribution of lead concentrations among selected sampling stations in Pensacola Bay.

27. Distribution of total PAH concentrations among selected sampling stations in Bayou Chico.

28. Distribution of total PCB concentrations among selected sampling stations in Bayou Chico.

29. Distribution of mercury concentrations among selected sampling stations in Bayou Chico.

30. Distribution of lead concentrations among selected sampling stations in Bayou Chico.

31. Distribution of total PAH concentrations among selected sampling stations in Choctawhatchee Bay.

32. Distribution of total PCB concentrations among selected sampling stations in Choctawhatchee Bay.

33. Distribution of mercury concentrations among selected sampling stations in Choctawhatchee Bay.

34. Distribution of lead concentrations among selected sampling stations in Choctawhatchee Bay.

35. Distribution of total PAH concentrations among selected sampling stations in St. Andrew Bay.

36. Distribution of total PCB concentrations among selected sampling stations in St. Andrew Bay.

37. Distribution of mercury concentrations among selected sampling stations in St. Andrew Bay.

38. Distribution of lead concentrations among selected sampling stations in St. Andrew Bay.

39. Distribution of total PAH concentrations among selected sampling stations in Apalachicola Bay.

40. Distribution of total PCB concentrations among selected sampling stations in Apalachicola Bay.

41. Distribution of mercury concentrations among selected sampling stations in Apalachicola Bay.

42. Distribution of lead concentrations among selected sampling stations in Apalachicola Bay.


Figure 43a. Relationship between arsenic and aluminum in 1993 Pensacola Bay samples.

## Pensacola Bay, Cadmium/Aluminum



Figure 43b. Relationship between cadmium and aluminum in 1993 Pensacola Bay samples.


Figure 43c. Relationship between chromium and aluminum in 1993 Pensacola Bay samples.


Figure 43d. Relationship between copper and aluminum in 1993 Pensacola Bay samples.


Figure 43e. Relationship between lead and aluminum in 1993 Pensacola Bay samples.


Figure 43f. Relationship between zinc and aluminum in 1993 Pensacola Bay samples.


Figure 44a. Relationship between arsenic and aluminum in 1994 Bayou Chico samples.

Bayou Chico, Cadmium/Aluminum


Figure 44b. Relationship between cadmium and aluminum in 1994 Bayou Chico samples.


Figure 44c. Relationship between chromium and aluminum in 1994 Bayou Chico samples.

## Bayou Chico, Copper/Aluminum



Figure 44d. Relationship between copper and aluminum in 1994 Bayou Chico samples.


Figure 44e. Relationship between lead and aluminum in 1994 Bayou Chico samples.


Figure 44f. Relationship between nickel and aluminum in 1994 Bayou Chico samples.


Figure 44g. Relationship between zinc and aluminum in 1994 Bayou Chico samples.


Figure 45a. Relationship between arsenic and aluminum in Choctawhatchee Bay samples.


Figure 45b. Relationship between cadmium and aluminum in Choctawhatchee Bay samples.

## Choctawhatchee Chromium/Aluminum



Figure 45c. Relationship between chromium and aluminum in Choctawhatchee Bay samples.


Figure 45d. Relationship between copper and aluminum in Choctawhatchee Bay samples.


Figure 45e. Relationship between lead and aluminum in Choctawhatchee Bay samples.


Figure 45f. Relationship between zinc and aluminum in Choctawhatchee Bay samples.


Figure 46a. Relationship between arsenic and aluminum in St. Andrew Bay samples.


Figure 46b. Relationship between cadmium and aluminum in St. Andrew Bay samples.


Figure 46c. Relationship between chromium and aluminum in St. Andrew Bay samples.


Figure 46d. Relationship between copper and aluminum in St. Andrew Bay samples.

St. Andrew, Lead/Aluminum


Figure 46e. Relationship between lead and aluminum in St. Andrew Bay samples.


Figure 46f. Relationship between zinc and aluminum in St. Andrew Bay samples.


Figure 47a. Relationship between arsenic and aluminum in Apalachicola Bay samples.


Figure 47b. Relationship between cadmium and aluminum in Apalachicola Bay samples.


Figure 47c. Relationship between chromium and aluminum in Apalachicola Bay samples.


Figure 47d. Relationship between copper and aluminum in Apalachicola Bay samples.


Figure 47e. Relationship between lead and aluminum in Apalachicola Bay samples.


Figure 47f. Relationship between zinc and aluminum in Apalachicola Bay samples.

48. Sampling stations in Pensacola Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic.


Figure 49. Sampling stations in Bayou Chico in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic.

## Amphipod Survival

- Not toxic
- Significantly different from controls
- Significantly different from controls and $<80 \%$ of controls


Figure 50. Sampling stations in Choctawhatchee Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic.


Figure 51. Sampling stations in St. Andrew Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic.


Figure 52. Sampling stations in Apalachicola Bay in which sediments were either not toxic to amphipod survival, significantly different from controls, or highly toxic.


Figure 53. Sampling stations in Pensacola Bay in which sediments were either not toxic in Microtox ${ }^{\mathrm{TM}}$ tests, significantly different from controls, or highly toxic.


Figure 54. Sampling stations in Bayou Chico in which sediments were either not toxic to Microtox ${ }^{\text {M }}$ tests, significantly different from controls, or highly toxic.

Microtox

- Not toxic
- Significantly different from control
- Significantly different from control and $<80 \%$ of control
A Significantly different from control and $<10 \%$ of control


Figure 55. Sampling stations in Choctawhatchee Bay in which sediments were either not toxic to Microtox ${ }^{\text {TM }}$ tests, significantly different from controls, or highly toxic.


Figure 56. Sampling stations in St. Andrew Bay in which sediments were either not toxic to Microtox ${ }^{\text {TM }}$ tests, significantly different from controls, or highly toxic.


Figure 57. Sampling stations in Apalachicola Bay in which sediments were either not toxic to Microtox ${ }^{\mathrm{TM}}$ tests, significantly different from controls, or highly toxic.

## Mutatox

- Not toxic
- Suspected genotoxic
- Genotoxic response


Figure 58. Sampling stations in Choctawhatchee Bay in which sediments were either not toxic, suspected genotoxic, or genotoxic in Mutatox ${ }^{\text {TM }}$ tests.


Figure 59. Sampling stations in Apalachicola Bay in which sediments were either not toxic, suspected genotoxic, or genotoxic in Mutatox ${ }^{\text {TM }}$ tests.


Figure 60. Sampling stations in Pensacola Bay in which sediments were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters.


Figure 61. Sampling stations in Bayou Chico that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters.

## Sea Urchin Fertilization

O Not toxic in $100 \%$ porewater

- Significantly different from control: $100 \%$ porewater
- Significantly different from control: $100 \%+50 \%$ porewater
- Significantly different from control: $100 \%+50 \%+25 \%$ porewater


Figure 62. Sampling stations in Choctawhatchee Bay that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters.


Figure 63. Sampling stations in St. Andrew Bay that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters.


Figure 64. Sampling stations in Apalachicola Bay that were either not toxic or significantly different from controls in sea urchin fertilization tests of sediment porewaters.


Figure 65. Sampling stations in Pensacola Bay that were either not toxic or significantly different from controls in sea urchin developmente tests of sediment porewaters.


Figure 66. Sampling stations in Bayou Chico that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters.

## Sea Urchin Development

- Not toxic in $100 \%$ porewater
- Significantly different from control: $100 \%$ porewater
- Significantly different from control: $100 \%+50 \%$ porewater
- Significantly different from control: $100 \%+50 \%+25 \%$ porewater


Figure 67. Sampling stations in Choctawhatchee that were either not toxic or significantly different from controls in sea urchin embryo development tests of sediment porewaters.


Figure 68. Sampling stations in St. Andrew Bay that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters.


Figure 69. Sampling stations in Apalachicola Bay that were either not toxic or significantly different from controls in sea urchin development tests of sediment porewaters.


Figure 70. Relationship between normal urchin embryo development and the concentrations of zinc in Pensacola Bay.


Figure 71. Relationship between urchin normal development and concentrations of high molecular weight PAHs in Pensacola Bay.


Figure 72. Relationship between Microtox ${ }^{\text {TM }}$ EC50's and the contrations of total DDT in Pensacola Bay.


Figure 73. Relationship between Microtox ${ }^{\text {TM }}$ EC50's and the sum of 18 chemical concentration/ERM quotients for Pensacola Bay.


Figure 74. Relationship between urchin percent normal development and the concentrations of un-ionized ammonia in porewater from Pensacola Bay.


Figure 75. Relationship between sea urchin fertilization and the concentrations of zinc in samples from Bayou Chico and Apalachicola Bay.


Figure 76. The relationship between Microtox ${ }^{\text {TM }}$ EC50's and the concentrations of fluoranthene (ug/goc) in samples from Bayou Chico and Apalachicola Bay.


Figure 77. Relationship between sea urchin fertilization and concentrations of total PAHs in Bayou Chico and Apalachicola Bay.


Figure 78. Relationship between urchin embryo development and the sum of 25 chemical concentration/ERM quotients in Bayou Chico and Apalachicola Bay.


Figure 79. Relationship between urchin embryo development and concentrations of un-ionized ammonia in porewater in Bayou Chico and Apalachicola Bay.


Figure 80. Relationship between sea urchin fertilization and the concentrations of total DDTs in Choctawhatchee Bay.


Figure 81. Relationship between urchin embryo development and concentrations of un-ionized ammonia in porewater in Choctawhatchee Bay.


Figure 82. Relationship between amphipod survival and the concentrations of total DDT in St. Andrew Bay sediments.


Figure 83. Relationship between sea urchin normal development and the concentrations of un-ionized ammonia in the porewater of St. Andrew Bay sediments.


Figure 84. Relationship between percent sea urchin fertilization and the concentrations of zinc in St. Andrew Bay sediments.


Figure 85. Relationship between sea urchin fertilization and the concentrations of un-ionized ammonia in porewater of sediments from St. Andrew Bay.


Figure 86. Relationship between microbial bioluminescence (Microtox EC50's) and the concentrations of trans-nonachlor in St. Andrew Bay sediments.


Figure 87. Relationship between the sum of 25 chemical concentration-to-ERM quotients and microbial bioluminescence (Microtox EC50's) in St. Andrew Bay sediments.


Figure 88. Relationship between Microtox ${ }^{\text {TM }}$ EC50's and the concentrations of total DDTs in western Florida samples.


Figure 89. The relationship between urchin fertilization and the concentrations of all PAHs in western Florida samples.


Figure 90. Relationship between urchin embryo normal development and the concentrations of un-ionized ammonia in porewater in western Florida samples.

Table 1. Coordinates of sampling stations in each bay.

| Apalachicola Bay, 1994 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Stratum | Station | Location | Latitude | Longitude |
| (A) B | 1-1 | Apalachicola Bay | $30^{\circ} 28.43$ ' N | $86^{\circ} 35.47{ }^{\prime} \mathrm{W}$ |
| (A)B | 3-1 | Apalachicola Bay | $29^{\circ} 42.65$ ' N | $84^{\circ} 53.98{ }^{\text {'W }}$ |
| (A) B | 2-1 | Apalachicola Bay | $29^{\circ} 39.65{ }^{\text {N }}$ | 84.56 .53 'W |
| (A)C | 3-1 | Apalachicola Bay | $29^{\circ} 36.81{ }^{\text {' }} \mathrm{N}$ | $84.59 .66{ }^{\text {'W }}$ W |
| (A)C | 2-1 | Apalachicola Bay | $29^{\circ} 39.28$ ' N | $85^{\circ} 03.21{ }^{\text {'W }}$ |
| (A)C | 1-1 | Apalachicola Bay | $29^{\circ} 42.96$ ' N | $84^{\circ} 59.23$ 'W |
| (A)A | 1-1 | Apalachicola Bay | $29^{\circ} 45.29$ ' N | $85^{\circ} 00.69$ 'W |
| (A)A | 2-1 | Apalachicola Bay | $29^{\circ} 44.47{ }^{\prime} \mathrm{N}$ | 84.59 .89 'W |
| (A)A | 3-1 | Apalachicola Bay | $29^{\circ} 43.63$ N | 84.58 .46 'W |
| Choctawhatchee Bay, 1994 |  |  |  |  |
| Stratum | Station | Location | Latitude | Longitude |
| (C) B | 1-1 | Garnier Bayou | $30^{\circ} 28.43 \mathrm{~N}$ | $86^{\circ} 35.46$ 'W |
| (C) B | 2-3 | Garnier Bayou | $30^{\circ} 27.80$ ' N | $86^{\circ} 35.75{ }^{\text {'W }}$ |
| (C) B | 3-1 | Dons Bayou | $30^{\circ} 27.05$ 'N | $86^{\circ} 36.18$ 'W |
| (C) C | 1-2 | Hand Cove, Garnier Bayou | $30^{\circ} 26.95{ }^{\text {' }} \mathrm{N}$ | $86^{\circ} 35.37{ }^{\text {'W }}$ |
| (C)C | 2-1 | Garnier Bayou | $30^{\circ} 26.65{ }^{\text {N }}$ | $86^{\circ} 35.58{ }^{\text {'W }}$ |
| (C) C | 3-1 | Garnier Bayou | $30^{\circ} 25.88{ }^{\text {' }}$ | $86^{\circ} 35.45{ }^{\text {'W }}$ |
| (C) L | 1-2 | Choctawhatchee Bay | $30^{\circ} 25.22$ N | $86^{\circ} 32.64{ }^{\text {'W }}$ |
| (C)A | 1-1 | Cinco Bayou | $30^{\circ} 25.74$ ' | $86^{\circ} 38.06$ 'W |
| (C)A | 2-1 | Cinco Bayou | $30^{\circ} 25.71 \mathrm{~N}$ | $86^{\circ} 37.12{ }^{\prime} \mathrm{W}$ |
| (C)A | 3-1 | Cinco Bayou | $30^{\circ} 25.54{ }^{\prime} \mathrm{N}$ | $86^{\circ} 36.51$ 'W |
| (C) ${ }^{\text {L }}$ | 3-1 | Choctawhatchee Bay | $30^{\circ} 28.51 \mathrm{~N}$ | $86^{\circ} 28.15{ }^{\text {'W }}$ |
| (C) L | 2-1 | Choctawhatchee Bay | $30^{\circ} 26.72{ }^{\text {N }}$ | $86^{\circ} 30.77$ 'W |
| (C) H | 1-1 | Joes Bayou | $30^{\circ} 24.55{ }^{\text {' }}$ | $86^{\circ} 29.30$ 'W |
| (C) H | 2-1 | Joes Bayou | $30^{\circ} 24.92$ ' N | $86^{\circ} 29.48{ }^{\text {'W }}$ |
| (C) D | 3-1 | Destin Harbor | $30^{\circ} 23.37 \mathrm{~N}$ | $86^{\circ} 29.57{ }^{\text {'W }}$ |
| (C) D | 2-2 | Destin Harbor | $30^{\circ} 23.37{ }^{\text {' }}$ | $86^{\circ} 29.94{ }^{\text {'W }}$ |
| (C) D | 1-1 | Destin Harbor | $30^{\circ} 23.44{ }^{\text {N }}$ | $86^{\circ} 30.20$ 'W |
| (C)E | 1-1 | Boggy Bayou | $30^{\circ} 30.95{ }^{\text {' }} \mathrm{N}$ | $86^{\circ} 29.49$ 'W |
| (C)E | 2-1 | Boggy Bayou | $30^{\circ} 29.83{ }^{\prime} \mathrm{N}$ | $86^{\circ} 29.13$ 'W |
| (C)E | 3-1 | Boggy Bayou | $30^{\circ} 29.72$ N | $86^{\circ} 28.93$ 'W |
| (C)F | 1-1 | Tom's Bayou | $30^{\circ} 30.18$ ' N | $86^{\circ} 30.02{ }^{\text {'W }}$ |
| (C)F | 2-1 | Tom's Bayou | $30^{\circ} 30.16$ ' N | $86^{\circ} 29.58{ }^{\prime} \mathrm{W}$ |
| (C)G | 1-1 | Boggy Bayou | $30^{\circ} 30.23$ ' | $86^{\circ} 25.15{ }^{\text {'W }}$ |
| (C)G | 2-1 | Rocky Bayou | $30^{\circ} 30.27{ }^{\prime} \mathrm{N}$ | $86^{\circ} 26.24$ 'W |
| (C)G | 3-1 | Rocky Bayou | $30^{\circ} 30.34{ }^{\prime} \mathrm{N}$ | $86^{\circ} 27.10$ 'W |
| (C)G | 4-1 | Rocky Bayou | $30^{\circ} 29.94{ }^{\prime} \mathrm{N}$ | $86^{\circ} 27.17{ }^{\text {'W }}$ |
| (C)M | 3-1 | Choctawhatchee Bay | $30^{\circ} 27.21$ ' N | $86^{\circ} 24.53$ 'W |
| (C)M | 2-1 | Choctawhatchee Bay | $30^{\circ} 27.77{ }^{\text {' }} \mathrm{N}$ | $86^{\circ} 19.08{ }^{\text {'W }}$ |
| (C)M | 1-1 | Choctawhatchee Bay | $30^{\circ} 28.16$ ' N | $86^{\circ} 16.87{ }^{\text {'W }}$ |
| (C) N | 1-1 | Choctawhatchee Bay | $30^{\circ} 26.37 \mathrm{~N}$ | $86^{\circ} 14.09$ 'W |
| (C) N | 2-1 | Choctawhatchee Bay | $30^{\circ} 24.61$ ' N | $86^{\circ} 10.55{ }^{\text {'W }}$ |
| (C) N | 3-1 | Choctawhatchee Bay | $30^{\circ} 23.59$ ' N | $86^{\circ} 10.02$ 'W |
| (C) J | 1-1 | Alaqua Bayou | $30^{\circ} 29.12$ ' N | $86^{\circ} 12.62$ 'W |
| (C) J | 2-1 | Alaqua Bayou | $30^{\circ} 29.02 \mathrm{~N}$ | $86^{\circ} 12.34{ }^{\prime} \mathrm{W}$ |
| (C)K | 1-1 | La Grange Bayou | $30^{\circ} 28.23$ ' N | $86^{\circ} 08.23{ }^{\text {'W }}$ |
| (C)K | 2-1 | La Grange Bayou | $30^{\circ} 28.09{ }^{\prime} \mathrm{N}$ | $86^{\circ} 08.65{ }^{\text {'W }}$ |
| (C)K | 3-1 | La Grange Bayou | $30^{\circ} 27.50$ ' N | $86^{\circ} 09.31{ }^{\text {'W }}$ |

## Table 1 Continued.

| Pensacola Bay, $\mathbf{1 9 9 3}$ |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Stratum | Station | Location |  | Latitude | Longitude

Table 1 continued.

| St. Andrew Bay, 1993 |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| Stratum | Station | Location | Latitude | Longitude |
| A | 41 | West Bay | $30^{\circ} 15.492^{\prime}$ | 85 ${ }^{\circ} 47.389^{\prime}$ |
| A | 42 | West Bay | $30^{\circ} 15.166^{\prime}$ | $85^{\circ} 45.219^{\prime}$ |
| B | 43 | North Bay | 30 ${ }^{\circ} 12.011{ }^{\prime}$ | 85 ${ }^{\circ} 44.116^{\prime}$ |
| B | 44 | North Bay | $30^{\circ} 13.752^{\prime}$ | 85 ${ }^{\circ} 41.849^{\prime}$ |
| B | 45 | North Bay | $30^{\circ} 14.781{ }^{\prime}$ | 85 ${ }^{\circ} 41.072^{\prime}$ |
| B | 46 | North Bay | $30^{\circ} 15.194^{\prime}$ | 85 ${ }^{\circ} 40.090$ |
| B | 47 | Lynn Haven | $30^{\circ} 14.741{ }^{\prime}$ | 85 ${ }^{\circ} 39.662^{\prime}$ |
| C | 48 | St. Andrew Bay | 3009.929' | $85^{\circ} 43.562^{\prime}$ |
| C | 49 | St. Andrew Bay | 3009.562' | 85 ${ }^{\circ} 42.415^{\prime}$ |
| C | 50 | St. Andrew Bay | $30^{\circ} 08.956^{\prime}$ | 85 ${ }^{\circ} 40.785^{\prime}$ |
| C | 51 | St. Andrew Bay | $30^{\circ} 10.016^{\prime}$ | 85 ${ }^{\circ} 42.345^{\prime}$ |
| C | 52 | St. Andrew Bay | $30^{\circ} 10.533^{\prime}$ | 85 ${ }^{\circ} 43.315^{\prime}$ |
| D | 53 | Massalina Bayou | 3009.239' | 85 ${ }^{\circ} 39.354{ }^{\prime}$ |
| D | 54 | Massalina Bayou | $30^{\circ} 09.140^{\prime}$ | $85^{\circ} 39.384^{\prime}$ |
| E | 55 | Watson Bayou | $30^{\circ} 09.346{ }^{\prime}$ | 85 ${ }^{\circ} 88.418^{\prime}$ |
| E | 56 | Upper Watson | $30^{\circ} 09.179^{\prime}$ | $85^{\circ} 38.380^{\prime}$ |
| E | 57 | Upper Watson | $30^{\circ} 09.077{ }^{\prime}$ | $85^{\circ} 38.336{ }^{\prime}$ |
| E | 58 | Mid Watson | 3008.834' | $85^{\circ} 38.006{ }^{\prime}$ |
| E | 59 | Mid Watson | $30^{\circ} 08.656$ | $85^{\circ} 37.960^{\prime}$ |
| E | 60 | Mid Watson | $30^{\circ} 08.862^{\prime}$ | 85 ${ }^{\circ} 37.707^{\prime}$ |
| E | 61 | Lower Watson | $30^{\circ} 08.480^{\prime}$ | 85 ${ }^{\circ} 38.469^{\prime}$ |
| E | 62 | Lower Watson | $30^{\circ} 08.454{ }^{\prime}$ | $85^{\circ} 38.146^{\prime}$ |
| E | 63 | Watson Bayou | $30^{\circ} 08.444^{\prime}$ | 85 ${ }^{\circ} 37.993{ }^{\prime}$ |
| F | 64 | St. Andrew Bay | $30^{\circ} 08.443^{\prime}$ | $85^{\circ} 39.369^{\prime}$ |
| F | 65 | St. Andrew Bay | $30^{\circ} 08.039^{\prime}$ | $85^{\circ} 38.948^{\prime}$ |
| F | 66 | Mouth of Watson Bayou | $30^{\circ} 08.058^{\prime}$ | 85 ${ }^{\circ} 88.092^{\prime}$ |
| F | 67 | St. Andrew Bay | $30^{\circ} 07.573^{\prime}$ | $85^{\circ} 36.759^{\prime}$ |
| F | 68 | Mouth of Pearl Bayou | $30^{\circ} 06.214^{\prime}$ | 85 ${ }^{\circ} 36.669^{\prime}$ |
| F | 69 | Smak Bayou | $30^{\circ} 07.729^{\prime}$ | $85^{\circ} 40.003{ }^{\prime}$ |
| G | 70 | West-East Bay | $30^{\circ} 05.284{ }^{\prime}$ | $85^{\circ} 33.068^{\prime}$ |
| G | 71 | East-East Bay | $30^{\circ} 02.498^{\prime}$ | 85 ${ }^{\circ} 3.093{ }^{\prime}$ |

Table 2. Numbers of stations sampled and tested for toxicity and numbers of stations tested for chemistry in four western Florida bays.

| Bay <br> name | Year <br> sampled | Total number <br> of stations | Number of stations <br> tested for chemistry |
| :--- | :--- | :---: | :---: |
| Pensacola Bay | 1993 | 40 | 20 (organics) |
| Bayou Chico | 1994 | 6 | 40 (metals) |
| Choctawhatchee Bay | 1994 | 37 | 6 |
| St. Andrew Bay | 1993 | 31 | 21 |
| Apalachicola Bay | 1994 | 9 | 31 (organics) |

Table 3. Summary of data from amphipod toxicity tests performed in both years on all samples; listed as mean percent survival, statistical significance and percent of control for each station.

| Pensacola Bay, 1993 | Station number | Location | Mean \% survival $\pm$ standard deviation | Statistical Significance | $\begin{array}{r} \% \text { of } \\ \text { Control } \end{array}$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Control ("home" sediments) |  | San Francisco Bay | $68 \pm 10.4$ |  |  |
| Reference |  | Redfish Bay, Texas | $55 \pm 3.5$ |  |  |
| Stratum |  |  |  |  |  |
| A | 1 | Bayou Grande | $72 \pm 8.4$ | ns | 106 |
| A | 2 | Bayou Grande | $68 \pm 15.6$ | ns | 100 |
| A | 3 | Bayou Grande | $72 \pm 12.0$ | ns | 106 |
| B | 4 | Bayou Chico | $71 \pm 11.4$ | ns | 104 |
| B | 5 | Bayou Chico | $73 \pm 14.4$ | ns | 107 |
| B | 6 | Bayou Chico | $67 \pm 5.7$ | ns | 98 |
| B | 7 | Bayou Chico | $68 \pm 16.0$ | ns | 100 |
| B | 8 | Bayou Chico | $69 \pm 13.9$ | ns | 101 |
| B | 9 | Bayou Chico | $75 \pm 7.9$ | ns | 110 |
| C | 10 | Bayou Texar | $66 \pm 16.7$ | ns | 97 |
| C | 11 | Bayou Texar | $76 \pm 10.2$ | ns | 112 |
| C | 12 | Bayou Texar | $74 \pm 16.7$ | ns | 109 |
| C | 13 | Bayou Texar | $66 \pm 12.9$ | ns | 97 |
| D | 14 | Warrington | $71 \pm 9.6$ | ns | 104 |
| E | 15 | Bayou Chico | $76 \pm 8.9$ | ns | 112 |
| E | 16 | Bayou Channel | $70 \pm 11.2$ | ns | 103 |
| F | 17 | Inner Harbor Channel | $75 \pm 16.6$ | ns | 110 |
| F | 18 | Inner Harbor | $75 \pm 10.0$ | ns | 110 |
| F | 19 | Inner Harbor | $74 \pm 16.4$ | ns | 109 |
| F | 20 | Inner Harbor | $70 \pm 12.7$ | ns | 103 |
| G | 21 | Pensacola Bay | $70 \pm 20.9$ | ns | 103 |
| G | 22 | Pensacola Bay | $74 \pm 14.3$ | ns | 109 |
| G | 23 | Inner Harbor | $69 \pm 16.7$ | ns | 101 |
| H | 24 | Lower Bay | $68 \pm 17.9$ | ns | 100 |
| H | 25 | Lower Bay | $63 \pm 9.7$ | ns | 93 |
| $J$ | 26 | East Bay | $77 \pm 13.5$ | ns | 113 |
| K | 27 | East Bay | $74 \pm 12.4$ | ns | 109 |
| K | 28 | East Bay | $77 \pm 7.6$ | ns | 113 |
| L | 29 | Blackwater Bay | $61 \pm 16.4$ | ns | 88 |
| L | 30 | Blackwater Bay | $72 \pm 14.8$ | ns | 106 |
| L | 31 | Blackwater Bay | $74 \pm 6.5$ | ns | 109 |
| M | 32 | Escambia Bay | $71 \pm 10.8$ | ns | 104 |
| N | 33 | Escambia Bay | $67 \pm 11.5$ | ns | 98 |
| N | 34 | Escambia Bay | $63 \pm 19.6$ | ns | 93 |
| 0 | 35 | Escambia Bay | $60 \pm 15.4$ | ns | 88 |
| 0 | 36 | Escambia Bay | $65 \pm 9.4$ | ns | 96 |
| 0 | 37 | Escambia River Mouth | $62 \pm 23.1$ | ns | 91 |
| P | 38 | Escambia | $73 \pm 7.6$ | ns | 107 |
| P | 39 | Floridatown | $73 \pm 7.6$ | ns | 107 |
| I | 40 | Central Bay | $66 \pm 11.9$ | ns | 97 |

Table 3 continued.

| Pensacola Bay, 1994 | Station <br> Number | Location | Mean \% survival <br> $\pm$ standard deviation | Statistical <br> Significance of <br> Control |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Control |  | Central Long Island Sd. | $92 \pm 4.5$ |  |  |
| Stratum |  |  |  |  |  |
| (BC) | $1-2$ | Bayou Chico | $93 \pm 2.7$ | ns | 101 |
| (BC) | $2-2$ | Bayou Chico | $97 \pm 4.5$ | ns | 105 |
| (BC) | $3-2$ | Bayou Chico | $94 \pm 2.2$ | ns | 102 |
| (BC) | $4-2$ | Bayou Chico | $86 \pm 9.6$ | ns | 93 |
| (BC) | $5-3$ | Bayou Chico | $68 \pm 15.7$ | $* *$ | 74 |
| (BC) | $6-2$ | Bayou Chico | $94 \pm 6.5$ | ns | 102 |

Choctawhatchee Bay, 1994

## Station number

Control
Control
Control
Control
Control

## Stratum

(C)A
(C)A
(C)A
(C) $B$
(C) B
(C)B
(C)C
(C)C
(C)C
(C)D
(C)D
(C)D
(C)E
(C) E
(C)E
(C)F
(C)F
(C)G
(C)G
(C)G
(C)G
(C) H
(C) H
(C) J
(C) J
(C)K
(C)K
(C)K
(C)

1-1
2-1
3-1
1-1
2-3
3-1
1-2
2-1
3-1
1-1
2-2
3-1
1-1
2-1
3-1
1-1
2-1
1-1
2-1
3-1
4-1
1-1
2-1
1-1
2-1
1-1
2-1
3-1
1-2

Central Long Island Sd.
Central Long Island Sd.
Central Long Island Sd.
Central Long Island Sd.
Central Long Island Sd.

| Mean \% survival | Statistical <br> SignificanceControl of |
| :---: | :---: |
| $\underline{\text { Cstandard deviation }}$ |  |

Table 3 continued.

| Choctawhatchee Bay, 1994 | Station <br> number |
| :---: | :---: |
| (C)L | $2-1$ |
| (C)L | $3-1$ |
| (C)M | $1-1$ |
| (C)M | $2-1$ |
| (C)M | $3-1$ |
| (C)N | $1-1$ |
| (C)N | $2-1$ |
| (C)N | $3-1$ |

St. Andrew Bay, 1993 | Station |
| ---: |
| number |

Control ("home") sediments Reference

Stratum

| A | 41 |
| :--- | :--- |
| A | 42 |
| B | 43 |
| B | 44 |
| B | 45 |
| B | 46 |
| B | 47 |
| C | 48 |
| C | 49 |
| C | 50 |
| C | 51 |
| C | 52 |
| D | 53 |
| D | 54 |
| E | 55 |
| E | 56 |
| E | 57 |
| E | 58 |
| E | 59 |
| E | 60 |
| E | 61 |
| E | 62 |
| E | 63 |
| F | 64 |
| F | 65 |
| F | 66 |
| F | 67 |
| F | 68 |
| F | 69 |
| G | 70 |
| G | 71 |


| Mean \% survival <br> $\pm$ standard deviation | Statistical <br> Significance | Control of |
| :---: | :---: | :---: |


| $88 \pm 7.6$ | ns | 104 |
| :---: | :---: | :---: |
| $97 \pm 4.5$ | ns | 99 |
| $98 \pm 2.7$ | ns | 100 |
| $92 \pm 7.6$ | ns | 105 |
| $93 \pm 2.7$ | ns | 106 |
| $96 \pm 4.2$ | ns | 109 |
| $91 \pm 10.8$ | ns | 94 |
| $98 \pm 2.7$ | ns | 101 |


| Mean \% survival | Statistical <br> $\pm$ standard deviation of <br> Significance <br> Control |
| :---: | :---: |

San Francisco Bay

$$
93 \pm 8.4
$$

Redfish Bay, Texas
$97 \pm 2.7$

| $92 \pm 5.7$ | ns | 99 |
| :---: | :---: | :---: |
| $94 \pm 5.5$ | ns | 101 |
| $84 \pm 8.9$ | ns | 90 |
| $91 \pm 4.2$ | ns | 98 |
| $90 \pm 10.0$ | ns | 97 |
| $88 \pm 11.5$ | ns | 95 |
| $86 \pm 4.2$ | ns | 92 |
| $95 \pm 5.0$ | ns | 102 |
| $96 \pm 6.5$ | ns | 103 |
| $89 \pm 6.5$ | ns | 96 |
| $85 \pm 3.5$ | ns | 103 |
| $89 \pm 6.5$ | ns | 96 |
| $96 \pm 5.5$ | ns | 103 |
| $84 \pm 7.4$ | ns | 90 |
| $93 \pm 5.7$ | ns | 100 |
| $84 \pm 14.7$ | ns | 90 |
| $93 \pm 2.7$ | ns | 100 |
| $94 \pm 6.5$ | ns | 101 |
| $87 \pm 10.4$ | ns | 94 |
| $83 \pm 29.9$ | ns | 89 |
| $88 \pm 5.7$ | ns | 95 |
| $85 \pm 17.3$ | ns | 103 |
| $94 \pm 6.5$ | ns | 101 |
| $96 \pm 4.2$ | ns | 103 |
| $89 \pm 8.9$ | ns | 96 |
| $89 \pm 8.9$ | ns | 96 |
| $94 \pm 6.5$ | ns | 101 |
| $92 \pm 4.5$ | ns | 99 |
| $88 \pm 7.6$ | ns | 95 |
| $99 \pm 2.2$ | ns | 106 |
| $92 \pm 5.7$ | ns | 99 |

Table 3 continued.

| Apalachicola Bay, 1994 | Station number | Station location | Mean \% survival $\pm$ standard deviation | Statistical \% of Significance Control |
| :---: | :---: | :---: | :---: | :---: |
| Control |  | Central Long Island Sd. | $97 \pm 4.5$ |  |
|  |  | Central Long Island Sd. | $98 \pm 2.7$ |  |
| Stratum |  |  |  |  |
| (A)A | 1-1 | Apalachicola Bay | $93 \pm 5.7$ | ns 96 |
| (A)A | 2-1 | Apalachicola Bay | $86 \pm 14.7$ | ns 89 |
| (A)A | 3-1 | Apalachicola Bay | $87 \pm 6.7$ | 90 |
| (A)B | 1-1 | Apalachicola Bay | $93 \pm 4.5$ | ns 96 |
| (A)B | 2-1 | Apalachicola Bay | $96 \pm 5.5$ | ns 99 |
| (A)B | 3-1 | Apalachicola Bay | $96 \pm 4.2$ | ns 99 |
| (A)C | 1-1 | Apalachicola Bay | $95 \pm 5.0$ | ns 97 |
| (A)C | 2-1 | Apalachicola Bay | $96 \pm 4.2$ | ns 99 |
| (A)C | 3-1 | Apalachicola Bay | $\underline{94 \pm 4.2}$ | ns 97 |

$\mathrm{ns}=$ not significant $(\mathrm{p}>0.05)$

* $=$ significantly different from controls ( $p<0.05$ )
** $=$ survival $<80 \%$ of controls
Table 4. Results from Microtox (тм) tests for both years and from all stations in the four western Florida Bays; expressed as mean EC 50s (mg equivalents, wet wt.) and standard deviations, statistical significance, and percent of controls.

| Microtox |  |  | EC 50 | significance |
| :---: | :---: | :---: | :---: | :---: |
|  |  |  | Mean $\pm$ SD |  |
| Apalachicola Bay, 1994 |  |  |  |  |
| Block | Station | Station Location | Mean $\pm \underline{\text { SD }}$ | significance |
| (A)A | 1-1 | Apalachicola Bay | $58.5 \pm 10.2$ | ns |
| (A)A | 2-1 | Apalachicola Bay | $3.1 \pm 0.8$ | ** |
| (A)A | 3-1 | Apalachicola Bay | $5.6 \pm 1.1$ | ** |
| (A)B | 1-1 | Apalachicola Bay | $2.8 \pm 0.4$ | ** |
| (A)B | 2-1 | Apalachicola Bay | $22.7 \pm 3.7$ | ** |
| (A) ${ }^{\text {B }}$ | 3-1 | Apalachicola Bay | $7.8 \pm 1.0$ | ** |
| (A)C | 1-1 | Apalachicola Bay | $2.2 \pm 0.5$ | ** |
| (A)C | 2-1 | Apalachicola Bay | $9.9 \pm 1.3$ | ** |
| (A)C | 3-1 | Apalachicola Bay | $1.2 \pm 0.1$ | ** |
| Choctawhatchee Bay, 1994 |  |  |  |  |
| Block | Station | Station Location | Mean $\pm \underline{\text { SD }}$ | significance |
| (C)A | 1-1 | Cinco Bayou | $2.1 \pm 0.2$ | ** |
| (C)A | 2-1 | Cinco Bayou | $4.7 \pm 1.3$ | ** |
| (C)A | 3-1 | Cinco Bayou | $1.2 \pm 0.1$ | ** |
| (C)B | 1-1 | Garnier Bayou | $1.6 \pm 0.2$ | ** |
| (C)B | 2-3 | Garnier Bayou | $6.3 \pm 1.3$ | ** |
| (C)B | 3-1 | Dons Bayou | $2.3 \pm 0.2$ | ** |
| (C)C | 1-2 | Hand Cove, Garnier Bayou | $12.8 \pm 2.1$ | ** |

Table 4 continued.
Choctawhatchee Bay, 1994
Block Sta
(C)C

Station Location
Mean $\pm$ SD
significance

| $0.4 \pm 0.1$ | $* *$ |
| ---: | :--- |
| $4.3 \pm 0.5$ | $* *$ |
| $3.2 \pm 0.5$ | $* *$ |
| $0.8 \pm 0.2$ | $* *$ |
| $0.3 \pm 0.0$ | $* *$ |
| $2.2 \pm 0.6$ | $* *$ |
| $1.7 \pm 0.3$ | $* *$ |
| $3.1 \pm 0.2$ | $* *$ |
| $1.6 \pm 0.4$ | $* *$ |
| $3.8 \pm 0.7$ | $* *$ |
| $1.0 \pm 0.3$ | $* *$ |
| $10.8 \pm 1.8$ | $* *$ |
| $2.5 \pm 0.4$ | $* *$ |
| $0.8 \pm 0.2$ | $* *$ |
| $0.3 \pm 0.1$ | $* *$ |
| $0.5 \pm 0.0$ | $* *$ |
| $17.3 \pm 5.1$ | $* *$ |
| $7.3 \pm 2.1$ | $* *$ |
| $1.0 \pm 0.3$ | $* *$ |
| $0.9 \pm 0.2$ | $* *$ |
| $2.7 \pm 0.4$ | $* *$ |
| $1.1 \pm 0.2$ | $* *$ |
| $16.8 \pm 4.9$ | $* *$ |
| $1.3 \pm 0.1$ | $* *$ |
| $8.6 \pm 0.2$ | $* *$ |
| $15.8 \pm 2.0$ | $* *$ |
| $2.4 \pm 0.7$ | $* *$ |
| $25.9 \pm 5.3$ | $* *$ |
| $18.3 \pm 3.7$ | $* *$ |
| $17.6 \pm 3.4$ | $* *$ |

Pensacola Bay, 1993

Station

1
2
3
4
5
6
7
8

Location
Bayou Grande
Bayou Grande
Bayou Grande
Bayou Chico
Bayou Chico
Bayou Chico
Bayou Chico
Bayou Chico
Bayou Chico
Bayou Texar
Bayou Texar

Mean $\pm$ SD
$0.07 \pm 0.02$ **
$0.76 \pm 0.03$ **
$0.2: \pm 0.07$ **
$0.77 \pm 0.0: \quad * *$
$0.25 \pm 0.06$ **
$0.24 \pm 0.07$ **
$0.63 \pm 0.17$ **
$0.57 \pm 0.06$ **
$0.45 \pm 0.13$ **
$0.32 \pm 0.03$ **
$0.53 \pm 0.10$ **

Table 4 continued.
Choctawhatchee Bay, 1994
Block Station Station Location

Mean $\pm$ SD

Bayou Texar
Bayou Texar
Warrington
$1.21 \pm 0.16$
$9.12 \pm 1.76$
$7.38 \pm 1.92$

12

13
14
Bayou Texar
Bayou Texar
Warrington
ns

Pensacola Bay, 1993

| Station | Location |
| :---: | :---: |
|  |  |
| 15 | Bayou Chico |
| 16 | Bayou Channel |
| 17 | Inner Harbor Channel |
| 18 | Inner Harbor |
| 19 | Inner Harbor |
| 20 | Inner Harbor |
| 21 | Pensacola Bay |
| 22 | Pensacola Bay |
| 23 | Inner Harbor |
| 24 | Lower Bay |
| 25 | Lower Bay |
| 26 | East Bay |
| 27 | East Bay |
| 28 | East Bay |
| 29 | Blackwater Bay |
| 30 | Blackwater Bay |
| 31 | Blackwater Bay |
| 32 | Escambia Bay |
| 33 | Escambia Bay |
| 34 | Escambia Bay |
| 35 | Escambia Bay |
| 36 | Escambia Bay |
| 37 | Escambia River Mouth |
| 38 | Escambia |
| 39 | Floridatown |
| 40 | Central Bay |

Mean $\pm$ SD

| $5.89 \pm 0.23$ | $* *$ |
| ---: | :--- |
| $3.56 \pm 0.74$ | $* *$ |
| $4.33 \pm 0.32$ | $* *$ |
| $0.09 \pm 0.01$ | $* *$ |
| $2.68 \pm 0.55$ | $* *$ |
| $0.75 \pm 0.09$ | $* *$ |
| $10.03 \pm 2.6:$ | $n s$ |
| $12.34 \pm 1.22$ | ns |
| $9.08 \pm 1.37$ | ns |
| $9.23 \pm 2.16$ | $n s$ |
| $11.49 \pm 1.19$ | $n s$ |
| $1.11 \pm 0.24$ | $* *$ |
| $3.36 \pm 0.61$ | $* *$ |
| $1.62 \pm 0.35$ | $* *$ |
| $0.66 \pm 0.16$ | $* *$ |
| $8.01 \pm 1.24$ | ns |
| $1.05 \pm 0.11$ | $* *$ |
| $4.72 \pm 1.02$ | $* *$ |
| $2.33 \pm 0.21$ | $* *$ |
| $0.66 \pm 0.14$ | $* *$ |
| $9.84 \pm 1.53$ | ns |
| $6.65 \pm 0.07$ | $* *$ |
| $3.76 \pm 0.58$ | $* *$ |
| $2.29 \pm 0.14$ | $* *$ |
| $4.49 \pm 0.88$ | $* *$ |
| $1.84 \pm 0.25$ | $* *$ |

Pensacola Bay, 1994

| Block | Station | Location | Mean $\pm \mathbf{S D}$ | significance |
| :---: | :---: | :---: | ---: | :---: |
|  |  |  |  |  |
| $(B C)$ | $1-2$ | Bayou Chico | $1.93 \pm 0.35$ | $* *$ |
| $(B C)$ | $2-2$ | Bayou Chico | $40.53 \pm 2.35$ | $*$ |
| $(B C)$ | $3-2$ | Bayou Chico | $13.47 \pm 3.66$ | $* *$ |
| $(B C)$ | $4-2$ | Bayou Chico | $0.62 \pm 0.09$ | $* *$ |
| $(B C)$ | $5-3$ | Bayou Chico | $1.73 \pm 0.21$ | $* *$ |
| $(B C)$ | $6-2$ | Bayou Chico | $0.81 \pm 0.22$ | $* *$ |

Table 4 continued.
St. Andrew Bay, 1993

| Block | Station | Location | Mean $\pm \mathbf{S D}$ |
| :---: | :---: | :---: | :---: |
|  | significance |  |  |
| 41 | West Bay | $3.35 \pm 0.83$ | $* *$ |
| 42 | West Bay | $0.89 \pm 0.08$ | $* *$ |
| 43 | North Bay | $1.66 \pm 0.27$ | $* *$ |
| 44 | North Bay | $0.31 \pm 0.08$ | $* *$ |
| 45 | North Bay | $0.68 \pm 0.04$ | $* *$ |
| 46 | North Bay | $0.1: \pm 0.02$ | $* *$ |
| 47 | Lynn Haven | $0.74 \pm 0.07$ | $* *$ |
| 48 | St. Andrew Bay | $1.32 \pm 0.3:$ | $* *$ |
| 49 | St. Andrew Bay | $0.42 \pm 0.06$ | $* *$ |
| 50 | St. Andrew Bay | $0.18 \pm 0.03$ | $* *$ |
| 51 | St. Andrew Bay | $0.48 \pm 0.10$ | $* *$ |
| 52 | St. Andrew Bay | $2.1: \pm 0.28$ | $* *$ |
| 53 | Massalina Bayou | $0.15 \pm 0.04$ | $* *$ |
| 54 | Massalina Bayou | $0.20 \pm 0.02$ | $* *$ |
| 55 | Watson Bayou | $0.68 \pm 0.08$ | $* *$ |
| 56 | Upper Watson | $0.14 \pm 0.02$ | $* *$ |
| 57 | Upper Watson | $0.23 \pm 0.06$ | $* *$ |
| 58 | Mid Watson | $0.14 \pm 0.02$ | $* *$ |
| 59 | Mid Watson | $0.50 \pm 0.03$ | $* *$ |
| 60 | Mid Watson | $0.23 \pm 0.05$ | $* *$ |
| 61 | Lower Watson | $0.22 \pm 0.04$ | $* *$ |
| 62 | Lower Watson | $0.99 \pm 0.25$ | $* *$ |
| 63 | Watson Bayou | $0.7: \pm 0.16$ | $* *$ |
| 64 | St. Andrew Bay | $0.51 \pm 0.09$ | $* *$ |
| 65 | St. Andrew Bay | $0.66 \pm 0.05$ | $* *$ |
| 66 | Mouth of Watson Bayou | $0.22 \pm 0.01$ | $* *$ |
| 67 | St. Andrew Bay | $7.29 \pm 2.17$ | $* *$ |
| 68 | Mouth of Pearl Bayou | $0.73 \pm 0.19$ | $* *$ |
| 69 | Smak Bayou | $0.23 \pm 0.02$ | $* *$ |
| 70 | West-East Bay | $0.35 \pm 0.05$ | $* *$ |
| 71 | East-East Bay | $0.62 \pm 0.16$ | $* *$ |
|  |  |  |  |

Table 5. Results of Mutatox tests, expressed as categorical responses. This test was performed on samples collected during 1994.

## Pensacola Bay, 1994

## Stratum Station

| (BC) | $1-2$ |
| :--- | :--- |
| (BC) | $2-2$ |
| (BC) | $3-2$ |
| (BC) | $4-2$ |
| (BC) | $5-3$ |
| (BC) | $6-2$ |

Genotoxic Response

## Category

G
G
G
G
G
G

Table 5 continued.

Choctawhatchee Bay, 1994
Stratum Station

| (C)A | $1-1$ |
| :--- | :--- |
| (C)A | $2-1$ |

$\begin{array}{ll}\text { (C)A } & 3-1\end{array}$
(C) $\mathrm{B} \quad 1-1$
(C) $\mathrm{B} \quad$ 2-3
(C)B $\quad 3-1$
(C)C 1-2
(C)C 2-1
(C)C $\quad 3-1$
(C)D 1-1
(C)D 2-2
(C)D 3-1
(C)E 1-1
(C)E 2-1
(C)E 3-1
(C)F $\quad$ 1-1
(C)F 2-1
(C)G 1-1
(C)G 2-1
(C)G 3-1
(C)G 4-1
(C) $\mathrm{H} \quad$ 1-1
(C) $\mathrm{H} \quad$ 2-1
(C) J 1-1
(C) J 2-1
(C)K $\begin{aligned} & \text { 1-1 }\end{aligned}$
(C)K 2-1
(C)K 3-1
(C)L 1-2
(C)L 2-1
(C)L 3-1
(C) $\mathrm{M} \quad 1-1$
(C) $\mathrm{M} \quad$ 2-1
(C) $\mathrm{M} \quad 3-1$
(C)N $\quad$ 1-1
(C) $\mathrm{N} \quad$ 2-1
(C)N $\quad$ 3-1

Apalachicola Bay, 1994
Stratum Station
(A)A $\quad 1-1$
(A)A 2-1
(A)A $\quad$ 3-1
(A)B $\quad 1-1$
(A)B $\quad$ 2-1
(A)B $\quad 3-1$
(A)C $\quad 1-1$
(A)C $\quad$ 2-1
(A)C $\quad$ 3-1

Genotoxic Response
Category

Location

| Cinco Bayou | G |
| :---: | :---: |
| Cinco Bayou | N |
| Cinco Bayou | G |
| Garnier Bayou | S |
| Garnier Bayou | N |
| Dons Bayou | N |
| Hand Cove, Garnier Bayou | N |
| Garnier Bayou | G |
| Garnier Bayou | S |
| Destin Harbor | G |
| Destin Harbor | G |
| Destin Harbor | G |
| Boggy Bayou | G |
| Boggy Bayou | S |
| Boggy Bayou | N |
| Tom's Bayou | S |
| Tom's Bayou | G |
| Boggy Bayou | G |
| Rocky Bayou | N |
| Rocky Bayou | S |
| Rocky Bayou | G |
| Joes Bayou | G |
| Joes Bayou | G |
| Alaqua Bayou | N |
| Alaqua Bayou | S |
| La Grange Bayou | G |
| La Grange Bayou | G |
| La Grange Bayou | S |
| Choctawhatchee Bay | N |
| Choctawhatchee Bay | S |
| Choctawhatchee Bay | S |
| Choctawhatchee Bay | N |
| Choctawhatchee Bay | N |
| Choctawhatchee Bay | N |
| Choctawhatchee Bay | N |
| Choctawhatchee Bay |  |
| Choctawhatchee Bay |  |

N
G
S
N
N
N
G

G
G

N

N
s
G
G
G
Alaqua Bayou N
Alaqua Bayou S
La Grange Bayou G
La Grange Bayou -
Choctawhatchee Bay N
Choctawhatchee Bay S
Choctawhatchee Bay N
Choctawhatchee Bay N
Choctawhatchee Bay S
Choctawhatchee Bay N
Choctawhatchee Bay N
$\left.\begin{array}{cc}\text { Location } & \begin{array}{c}\text { Genotoxic Response } \\ \text { Category }\end{array} \\ \text { Apalachicola Bay } & \mathrm{N}\end{array}\right)$
(N=Negative, or not toxic, $\mathrm{G}=$ Genotoxic and $\mathrm{S}=$ Suspect).
Table 6. Mean percent fertilization success of sea urchins in $100 \%, 50 \%$ and $25 \%$ concentrations of
Signifi-
cance







 $\begin{array}{r}100 \% \\ \text { WQAP } \\ \hline\end{array}$





Table 6 continued.













 $83.2 \pm 7.8$ $99.4 \pm 0.5$ $99.6 \pm 0.5$ $99.8 \pm 0.4$ $99.8 \pm 0.4$
$99.0 \pm 1.7$ $99.4 \pm 0.5$ $99.4 \pm 0.9$ $\stackrel{\circ}{\dot{+}}$
+
+1
ले -
-
+1
$\infty$
$\infty$
$\infty$ $98.6 \pm 1.7$

 $98.0 \pm 1.2$





[^0] $\begin{array}{cc}\text { Apalachicola Bay, } 1994 \\ \text { Block } & \text { Station } \\ & \text { Control } \\ \text { (A)A } & 1-1 \\ \text { (A)A } & 2-1 \\ \text { (A)A } & 3-1 \\ \text { (A)B } & 1-1 \\ \text { (A)B } & 2-1 \\ \text { (A)B } & 3-1 \\ \text { (A)C } & 1-1 \\ \text { (A)C } & 2-1 \\ \text { (A)C } & 3-1\end{array}$

Table 7. Mean normal embryo development in $100 \%, 50 \%$ and $25 \%$ concentrations of sediment porewater.

| 100\% WQAP |  | Significance | $\begin{array}{r} 50 \% \\ \text { WQAP } \end{array}$ | Significance | $\begin{array}{r} 25 \% \\ \text { WQAP } \end{array}$ |  | Significance |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Mean $\pm$ | SD |  | Mean $\pm$ SD |  | Mean $\pm$ | SD |  |
| $88.0 \pm$ | 3.2 |  | $94.2 \pm 3.1$ |  | $92.6 \pm$ | 2.1 |  |
| $1.2 \pm$ | 1.8 | ** | $82.8 \pm 11.4$ | ns | $90.4 \pm$ | 5.2 | ns |
| $91.5 \pm$ | 1.3 | ns | $92.4 \pm 3.4$ | ns | $89.6 \pm$ | 5.0 | ns |
| $86.0 \pm$ | 4.7 | ns | $93.2 \pm 5.7$ | ns | $88.2 \pm$ | 2.6 | ns |
| $78.8 \pm$ | 4.3 | ns | $90.4 \pm 4.3$ | ns | $91.0 \pm$ | 2.1 | ns |
| $0.0 \pm$ | 0.0 | ** | $0.0 \pm 0.0$ | ** | $85.2 \pm$ | 5.0 | ns |
| $0.0 \pm$ | 0.0 | ** | $47.4 \pm 7.3$ | ** | $91.2 \pm$ | 2.8 | ns |
| 0.0 $\pm$ | 0.0 | ** | $0.0 \pm 0.0$ | ** | $88.2 \pm$ | 2.3 | ns |
| $0.0 \pm$ | 0.0 | ** | $1.2 \pm 0.8$ | ** | $88.8 \pm$ | 4.1 | ns |
| $0.0 \pm$ | 0.0 | ** | $0.0 \pm 0.0$ | ** | $84.8 \pm$ | 5.1 | ns |
| $0.0 \pm$ | 0.0 | ** | $88.8 \pm 4.6$ | ns | $88.0 \pm$ | 3.6 | ns |
| $88.6 \pm$ | 3.5 | ns | $90.0 \pm 3.0$ | ns | $90.8 \pm$ | 3.1 | ns |
| $91.4 \pm$ | 5.5 | ns | $91.2 \pm 1.6$ | ns | $92.6 \pm$ | 3.0 | ns |
| $88.8 \pm$ | 2.9 | ns | $91.4 \pm 1.5$ | ns | $90.8 \pm$ | 3.6 | ns |
| $90.2 \pm$ | 0.8 | ns | $91.0 \pm 3.5$ | ns | $90.6 \pm$ | 3.4 | ns |
| $84.8 \pm$ | 3.9 | ns | $91.2 \pm 3.7$ | ns | $88.4 \pm$ | 1.5 | ns |
| $72.3 \pm$ | 3.3 | * | $84.0 \pm 4.2$ | ns | $91.4 \pm$ | 4.2 | ns |
| $88.4 \pm$ | 4.2 | ns | $88.4 \pm 4.7$ | ns | $92.0 \pm$ | 2.2 | ns |
| $92.6 \pm$ | 2.3 | ns | $91.0 \pm 1.9$ | ns | $92.4 \pm$ | 1.7 | ns |
| $98.8 \pm$ | 0.8 | ns | $99.8 \pm 0.4$ | ns | $98.4 \pm$ | 1.7 | ns |
| $98.4 \pm$ | 0.9 | ns | $99.0 \pm 0.7$ | ns | $98.8 \pm$ | 1.6 | ns |
| $99.4 \pm$ | 0.9 | ns | $98.6 \pm 1.5$ | ns | $98.2 \pm$ | 1.6 | ns |
| $73.3 \pm$ | 10.1 | ** | $97.6 \pm 0.5$ | ns | $99.0 \pm$ | 1.2 | ns |
| $4.0 \pm$ | 7.4 | ** | $97.0 \pm 1.6$ | ns | $98.8 \pm$ | 1.1 | ns |
| $98.6 \pm$ | 0.9 | ns | $98.6 \pm 0.9$ | ns | $98.6 \pm$ | 1.1 | ns |
| $85.8 \pm$ | 4.9 | ns | $95.4 \pm 2.3$ | ns | $95.2 \pm$ |  | ns |
| $95.4 \pm$ | 2.4 | ns | $93.0 \pm 1.0$ | ns | $94.8 \pm$ | 2.4 | ns |
| $95.0 \pm$ | 2.1 | ns | $93.4 \pm 4.5$ | ns | $95.8 \pm$ |  | ns |
| 89.8土 | 3.8 | ns | $95.6 \pm 1.1$ | ns | $95.0 \pm$ | 1.9 | ns |



[^1]






亥比



$100 \%$
WQAP




＊＊＊＊＊＊＊＊＊ロ の ロ の
＊＊＊＊＊＊＊＊の の の の ロ＊


＊＊＊＊＊＊＊$\underset{*}{*} \underset{*}{*}$





| Table 7 continued. |  |  | $100 \%$ <br> WQAP |  | Significance | WQAP <br> $50 \%$ WQAP | Signifi canc | \% <br> QAP |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Apalachicola Bay, 1994 |  |  |  |  |  |  |  |  |  |  |
| Stratum | Station control | Station Location Redfish Bay, Texas | Mean $\pm$$90.4 \pm$ | SD |  | Mean $\pm$ SD | Mean $\pm$ |  | SD |  |
|  |  |  |  | 5.1 |  | $80.2 \pm 2.6$ |  | $79.2 \pm$ | 5.2 |  |
| (A)A | 1-1 | Apalachicola Bay | $66.4 \pm$ | 6.1 | ns | $72.6 \pm 7.7$ | ns | $77.2 \pm$ | 7.7 | ns |
| (A)A | 2-1 | Apalachicola Bay | $0.0 \pm$ | 0.0 | ** | $77.0 \pm 7.7$ | ns | $72.0 \pm$ | 5.5 | ns |
| (A)A | 3-1 | Apalachicola Bay | $73.0 \pm$ | 4.6 | ns | $75.4 \pm 5.5$ | ns | $77.2 \pm$ | 3.6 | ns |
| (A)B | 1-1 | Apalachicola Bay | $67.4 \pm$ | 5.9 | ns | $75.4 \pm 3.3$ | ns | $80.2 \pm$ | 5.0 | ns |
| (A)B | 2-1 | Apalachicola Bay | $78.0 \pm$ | 8.3 | ns | $79.4 \pm 2.5$ | ns | $81.2 \pm$ | 2.9 | ns |
| (A)B | 3-1 | Apalachicola Bay | $36.6 \pm$ | 43.1 | ** | $82.2 \pm 5.3$ | ns | $77.4 \pm$ | 6.0 | ns |
| (A)C | 1-1 | Apalachicola Bay | $46.8 \pm$ | 9.7 | ** | $81.8 \pm 6.7$ | ns | $82.8 \pm$ | 4.0 | ns |
| (A)C | 2-1 | Apalachicola Bay | 0.0 $\pm$ | 0.0 | ** | $75.6 \pm 3.4$ | ns | $80.4 \pm$ | 5.4 | ns |
| (A)C | 3-1 | Apalachicola Bay | $71.8 \pm$ | 5.4 | ns | $78.8 \pm 5.3$ | ns | $80.4 \pm$ | 7.4 | ns |

[^2]Table 8. Spearman-rank correlations (Rho) among the four toxicity tests performed on 1993 samples from Pensacola Bay.

|  | Microbial <br> bioluminescence <br> EC50's | Sea urchin <br> fertilization <br> $(100 \%$ WQAP $)$ | Sea urchin <br> development <br> $(100 \%$ WQAP) |  |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| Amphipod survival | -0.164 | ns | $-0.1429999:$ | ns | $0.1189999:$ | ns |

Table 9. Spearman-rank correlations (Rho) among the four toxicity tests performed on 1994 samples from Bayou Chico and Apalachicola Bays.

|  | Microbial bioluminescence EC50's | $\begin{gathered} \text { Sea urchin } \\ \text { fertilization } \\ (100 \% \text { WQAP) } \end{gathered}$ | Sea urchin development (100\% WQAP) |
| :---: | :---: | :---: | :---: |
| Amphipod survival | 0.18199999: ns | -0.049 ns | 0.1779999: ns |
| Microbial bioluminescence |  | 0.350 ns | -0.0609999: ns |
| Sea urchin fertilization |  |  | 0.0619999: ns |

WQAP = water quality adjusted porewater

Table 10. Spearman-rank correlations (Rho) among the four toxicity tests from Choctawhatchee Bay.

|  | Microbial <br> bioluminescence <br> EC50's | Sea urchin <br> Fertilization <br> 100\% WQAP | Sea urchin <br> Development <br> 100\% WQAP |  |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| Amphipod Survival | 0.029 | ns | 0.127 | ns | $0.1859999:$ | ns |

WQAP = water quality adjusted porewater

Table 11. Spearman-rank correlations (Rho) among the four different toxicity tests performed in St. Andrew Bay.

|  | Microbial <br> bioluminescence <br> EC50's | Sea urchin <br> fertilization <br> @ 100\% WQAP | Sea urchin <br> development <br> @ 100\% WQAP |
| :--- | :---: | :---: | :---: |
| Amphipod <br> survival | +0.212 ns | +0.211 ns | +0.075 ns |
| Microbial <br> bioluminescence | $+0.408 *$ |  |  |
| Sea urchin <br> fertilization |  | +0.179 ns |  |

WQAP = water quality adjusted porewater

* $p<0.05$
Table 12. Percent of samples from each bay that were either toxic (i.e., significantly different from controls) or highly toxic (i.e., significantly different from controls and <80\% of controls).


[^3]Table 13. Estimates of the spatial extent of toxicity in Western Florida bays with results of four different tests.

| Apalachicola Bay |  |  |
| :---: | :---: | :---: |
| Toxicity test | Kilometer ${ }^{2}$ | Percent of Total |
| Amphipod Survival | 0.0 | 0.0 |
| Sea Urchin Development |  |  |
| 100\% porewater | 157.5 | 83.96 |
| 50\% porewater | 0.0 | 0.0 |
| 25\% porewater | 0.0 | 0.0 |
| Sea Urchin Fertilization |  |  |
| 100\% porewater | 63.56 | 33.89 |
| 50\% porewater | 0.0 | 0.0 |
| 25\% porewater | 0.0 | 0.0 |
| Microtox | 186.84 | 99.6 |
| Survey area | 187.58 |  |
| St. Andrew Bay |  |  |
| Toxicity test | Kilometer ${ }^{2}$ | Percent of Total |
| Amphipod Survival | 0.0 | 0.0 |
| Sea Urchin Development |  |  |
| 100\% porewater | 7.17 | 5.6 |
| 50\% porewater | 0.123 | 0.1 |
| 25\% porewater | 0.0 | 0.0 |
| Sea Urchin Fertilization |  |  |
| 100\% porewater | 2.28 | 1.8 |
| 50\% porewater | 0.0 | 0.0 |
| 25\% porewater | 0.0 | 0.0 |
| Microtox | 127.22 | 100 |
| Survey area | 127.22 |  |
| Choctawhatchee Bay |  |  |
| Toxicity test | Kilometer ${ }^{2}$ | Percent of Total |
| Amphipod Survival | 0.0 | 0.0 |
| Sea Urchin Development |  |  |
| 100\% porewater | 116.06 | 45.4 |
| 50\% porewater | 0.75 | 0.30 |
| 25\% porewater | 0.0 | 0.0 |
| Sea Urchin Fertilization |  |  |
| 100\% porewater | 113.14 | 44.46 |
| 50\% porewater | 35.73 | 13.87 |
| 25\% porewater | 0.09 | 0.04 |
| Microtox | 254.47 | 100 |
| Survey area | 254.47 |  |

Table 13 continued.
Pensacola Bay

| Toxicity test | Kilometer $^{2}$ | Percent of Total |
| :--- | :--- | :--- |
| Amphipod Survival | 0.04 | 0.015 |
| Sea Urchin Development | 5.41 | 1.98 |
| 100\% porewater | 0.61 | 0.22 |
| 50\% porewater | 0.19 | 0.07 |
| 25\% porewater | 14.4 | 5.28 |
| Sea Urchin Fertilization | 0.28 | .102 |
| 100\% porewater | 0.28 | .102 |
| 50\% porewater | 262.75 | 96.37 |
| 25\% porewater | 272.63 |  |

Combined Western Florida survey area

| Toxicity test | Kilometer $^{2}$ | Percent of Total |
| :--- | :--- | :--- |
| Amphipod Survival | 0.04 | 0.005 |
| Sea Urchin Development | 286.1 |  |
| 100\% porewater | 1.5 | 34.0 |
| 50\% porewater | 0.2 | 0.2 |
| 25\% porewater |  | 0.02 |
| Sea Urchin Fertilization | 193.4 | 23.0 |
| 100\% porewater | 36.0 | 4.3 |
| 50\% porewater | 0.4 | 0.04 |
| 25\% porewater | 831.2 | 98.7 |
| Microtox | 841.9 |  |
| Total Survey area |  |  |

Table 14. Test results for Pensacola Bay.
Amphipod survival
Microtox
Rho


$\underbrace{\text { Urchin development }}_{\text {Rho }}$









|  |  |
| :---: | :---: |
|  |  |
|  |  |





## 

Amphipod survival
Rho

| ORGANIC COMPOUNDS |  |
| :--- | ---: |
| Polycyclic aromatic hydrocarbons (PAH) |  |
| fluorene | -0.216999999 |
| phenanthrene | -0.17699999 |
| anthracene | -0.290999999 |
| Sum of 3 LMW PAH | -0.10799999 : |
| fluoranthene | -0.15599999 : |
| fluoranthene (ug/goc) | -0.041 |
| benzo(a)anthracene | -0.158 |
| chrysene | -0.033 |
| benzo(b,k)fluoranthene | -0.10399999 : |
| benzo(e)pyrene | -0.114 |
| benzo(a)pyrene | -0.21 |
| perylene | -0.254 |
| indeno(1,2,3)pyrene | -0.12199999 : |
| dibenz(a,h)anthracene | -0.245999999 |
| benzo(g,h,i)perylene | -0.162 |
| Sum of 11 HMW PAH | -0.10399999 |
| Total PAH | -0.111 |
| Sum of 8 PAH ERMs | -0.15199999 : |

[^4] benzo(a)anthracene
chrysene benzo(b,k)fluora benzo(e)pyrene benz(a) Total PAH

Urchin development



| Table 14 continued. | Amphipod survival |  |
| :---: | :---: | :---: |
|  | Rho | signif. |
| Pesticides |  |  |
| Dieldrin | -0.01499999: | ns |
| Dieldrin (ug/goc) | -0.408999999 | ns |
| Mirex | -0.14699999: | ns |
| PCB congeners |  |  |
| PCBCON8 | nd | nd |
| PCBCON18 | nd | nd |
| PCBCON29 | nd | nd |
| PCBCON52 | nd | nd |
| PCBCON44 | nd | nd |
| PCBCON66 | -0.286999999 | ns |
| PCBCON101 | -0.17399999: | ns |
| PCBCON77 | -0.17399999: | ns |
| PCBCON118 | -0.068 | ns |
| PCBCON153 | -0.07099999: | ns |
| PCBCON105 | -0.03699999: | ns |
| PCBCON138 | -0.06099999: | ns |
| PCBCON126 | nd | nd |
| PCBCON187 | 0.03099999 : | ns |
| PCBCON128 | 0.10599999 : | ns |
| PCBCON180 | nd | nd |
| PCBCON170 | nd | nd |
| PCBCON195 | nd | nd |
| PCBCON206 | nd | nd |
| PCBCON209 | nd | nd |
| Sum of 20 PCBs | -0.083 | ns |
| Total PCBs | -0.083 | ns |
| Sum of 2 COH ERMs | 0.138 | ns |
| Sum of 18 ERMs | 0.198 | ns |

[^5]Table 15. Stations within Pensacola Bay in which chemical concentrations equalled or exceeded their respective greater exceedances are listed in bold.

Greater than or equal to ERM value

na

na
๔ ฮ
na
na
na
none

* ERL and ERM values from Long et al. (1995); TEL and PEL values from MacDonald (1994),SQCs from U. S. EPA, 1994;
urchin development LOEC from Long et al, In press; and amphipod NOEC from Kohn et al., 1994.
na $=$ no applicable values.
Table 16. A comparison of the chemical concentrations (ave $\pm$ std dev) in toxic and non-toxic samples in Pensacola Bay in Microtox ${ }^{\mathrm{TM}}$ tests, ratios between the two averages, and ratios between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb)*.

| Chemical substance | Not toxic ( $\mathrm{n}=8$ ) | $\begin{gathered} \text { Toxic } \\ (n=32) \end{gathered}$ | Ratio of averages | Guidelines Exceeded (ratio to toxic average) guideline value |  | guideline value | $\underline{\text { ratio }}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Cadmium | $0.2 \pm 0.2$ | $0.6 \pm 0.9$ | 4.1 | <ERL (1.2 ppm) | <1.0 | < TEL (0.67 ppm) | <1.0 |
| Copper | $21.2 \pm 4.6$ | $41.9 \pm 43.9$ | 2.0 | >ERL ( 34.0 ppm ) | 1.2 | >TEL (18.7 ppm) | 2.2 |
| Lead | $39.9 \pm 17.6$ | $63.4 \pm 54.7$ | 1.6 | >ERL (46.7 ppm) | 1.3 | >TEL (30.2 ppm) | 2.1 |
| Mercury | $0.15 \pm 0.06$ | $0.25 \pm 0.25$ | 1.7 | <ERL (0.15 ppm) | 1.0 | >TEL (0.13 ppm) | 1.1 |
| Silver | $0.3 \pm 0.2$ | $0.3 \pm 0.2$ | 1.2 | <ERL (1.0 ppm) | <1.0 | <TEL (0.73 ppm) | <1.0 |
| Zinc | $147.5 \pm 56.2$ | $252.2 \pm 261.0$ | 1.7 | >ERL (150 ppm) | 1.7 | >TEL (124 ppm) | 2.0 |
| Sum LMW PAH | 206.0さ298.0 | $540.5 \pm 740.0$ | 2.6 | <ERL (552 ppb) | <1.0 | >TEL (312 ppb) | 1.7 |
| Sum HMW PAH | $1826 \pm 2896$ | $5360 \pm 7247$ | 2.9 | >ERL (1700 ppb) | 3.1 | >TEL (655 ppb) | 8.2 |
| Sum total PAH | $2032 \pm 3193$ | $5900 \pm 7896$ | 2.9 | >ERL (4022 ppb) | 1.5 | >TEL (1684 ppb) | 3.5 |
| Heptachlor epoxide | $0.1 \pm 0.0$ | $0.2 \pm 0.2$ | 1.6 | na |  | na |  |
| alpha chlordane | $4.8 \pm 7.2$ | $6.8 \pm 8.3$ | 1.4 | na |  | na |  |
| trans-nonachlor | $1.3 \pm 0.0$ | $1.7 \pm 1.8$ | 1.3 | na |  | na |  |
| 2,4'-DDD | $1.0 \pm 0.0$ | $5.0 \pm 7.7$ | 5.0 | na |  | na |  |
| 4,4'-DDD | $5.5 \pm 10.1$ | $13.7 \pm 17.4$ | 2.5 | na |  | $>$ TEL (7.81 ppb) | 1.7 |
| 4,4'-DDT | $4.5 \pm 7.9$ | $6.8 \pm 10.9$ | 1.5 | na |  | >TEL (1.19 ppb) | 5.7 |
|  |  |  |  |  |  | >PEL ( 4.77 ppb ) | 1.4 |
| Sum DDTs | $12.5 \pm 18.8$ | $27.0 \pm 32.0$ | 2.2 | >ERL (1.58 ppb) | 17.1 | >TEL (3.89 ppb) | 6.9 |
| Endrin | $1.6 \pm 1.2$ | $4.5 \pm 9.1$ | 2.8 | na |  | na |  |
| Mirex | $0.5 \pm 0.0$ | $2.9 \pm 6.0$ | 5.8 | na |  | na |  |
| Total PCBs | $33.5 \pm 0.0$ | $56.0 \pm 53.0$ | 1.7 | >ERL (22.7 ppb) | 2.5 | >TEL (21.6 ppb) | 2.6 |

[^6]Table 17. A comparison of the chemical concentrations (ave $\pm$ std dev) in toxic and non-toxic samples in Pensacola Bay in urchin fertilization tests, ratios between the two averages, and ratios between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb)*.

| Chemical substance | $\begin{aligned} & \text { Not toxic } \\ & (\mathrm{n}=37) \end{aligned}$ | Highly toxic $(\mathrm{n}=3)$ | Ratio of averages | Guidelines Exceeded (ratio to toxic averag guideline value | e) ratio | guideline value | $\underline{\text { ratio }}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Arsenic | $16.6 \pm 8.9$ | $26.2 \pm 3.8$ | 1.6 | >ERL (8.2 ppm) | 3.2 | >TEL (7.2 ppm) | 3.6 |
| Chromium | $71.4 \pm 42.4$ | $114.7 \pm 34.9$ | 1.6 | >ERL (81.0 ppm) | 1.3: | >TEL (52.3 ppm) | 2.2 |
| Nickel | $19.0 \pm 8.9$ | $23.5 \pm 1.6$ | 1.2 | >ERL (20.9 ppm) | 1.1 | >TEL (15.9 ppm) | 1.5 |
| 2,4'-DDE | $1.0 \pm 0.0$ | $1.4 \pm 0.5$ | 1.39999: | na |  | na |  |
| Dieldrin | $0.1 \pm 0.1$ | $0.1 \pm 0.0$ | 1.1 | na |  | <TEL (0.71 ppb) |  |

* ERL and ERM values from Long et al. (1995) and TEL and PEL values from MacDonald (1994). na $=$ no applicable values.
Table 18. A comparison of the chemical concentrations (ave $\pm$ std dev) in toxic and non-toxic samples in Pensacola Bay in urchin development tests, ratios between the two averages, and ratios between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb)*.

| Ratio of averages | Guidelines Exceede (ratio to toxic avera guideline value |  | guideline value | ratio |
| :---: | :---: | :---: | :---: | :---: |
| 4.2 | =ERL (1.2 ppm) | 1.0 | >TEL (0.67 ppm) | 1.8 |
| 1.1 | <ERL (81.0 ppm) | <1.0 | >TEL (52.3 ppm) | 1.5 |
| 3.5 | >ERL (34.0 ppm) | 2.4 | >TEL (18.7 ppm) | 4.4 |
| 2.1 | >ERL (46.7 ppm) | 2.0 | >TEL (30.2 ppm) | 3.2 |
| 2.6 | >ERL (0.15 ppm) | 2.6 | >TEL (0.13 ppm) | 3.3 |
| 1.0 | <ERL (20.9 ppm) | <1.0 | >TEL (15.9 ppm) | 1.3 |
| 3.5 | >ERL (150 ppm) | 3.3 | >TEL (124 ppm) | 4.0 |
|  |  |  | >PEL (271 ppm) | 1.8 |
| 3.1 | >ERL (552 ppb) | 1.5 | >TEL (312 ppb) | 2.6 |
| 2.4 | >ERL (1700 ppb) | 4.4 | >TEL (655 ppb) | 11.3 |
|  |  |  | >PEL (6676 ppb) | 1.1 |
| 2.4 | >ERL (4022 ppb) | 2.0 | >TEL (1684 ppb) | 5.0 |
| 2.2 | na |  | na |  |
| 2.3 | na |  | na |  |
| 2.9 | na |  | na |  |
| 2.6 | na |  | >TEL (1.22 ppb) | 16.4 |
|  |  |  | >PEL (7.81 ppb) | 2.6 |
| 2.1 | na |  | na |  |
| 5.2 | na |  | >TEL (1.2 ppb) | 10.8 |
|  |  |  | >PEL (4.8 ppb) | 2.7 |
| 3.0 | >ERL (1.58 ppb) | 26.4 | > TEL (3.89 ppb) | 10.7 |
| 6.8 | na |  | na |  |
| 3.0 | na |  | > TEL (0.71 ppb) | 8.7 |
|  |  |  | >PEL (4.3 ppb) | 1.4 |
| 2.3 | >ERL (22.7 ppb) | 3.5 | > TEL (21.6 ppb) | 3.7 |

[^7]Table 19. Summary of toxicity/chemistry relationships for samples from Pensacola Bay.

| Chemical | No. of significant correlations | No. of samples> SQGs |  | Toxic/non-toxic ratios |  |  | Toxic/SQGs ratios (TELs) |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | Micro- | Urchin | Urchin | Micro- | Urchin | Urchin |
|  |  | ERLs | TELs | tox | fert'n. | dev’t. | tox | fert'n. | dev’t. |
| Cadmium | 3 | 0.0 | 0.0 | 4.1 | $<1.0$ | 4.2 | $<1.0$ | <1.0 | 1.8 |
| Copper | 3 | 0.0 | 3 | 2.0 | $<1.0$ | 3.5 | 2.2 | <1.0 | 4.4 |
| Lead | 2 | 1 | 4 | 1.6 | <1.0 | 2.1 | 2.1 | <1.0 | 3.2 |
| Zinc | 2 | 5 | 10 | 1.7 | <1.0 | 3.5 | 2.0 | <1.0 | 4.0 |
| Sum LMW PAH | 3 | 0.0 | 4 | 2.6 | <1.0 | 3.1 | 1.7 | <1.0 | 2.6 |
| Sum HMW PAH | 3 | 3 | 7 | 2.9 | <1.0 | 2.4 | 8.2 | <1.0 | 11.3 |
| 4,4'-DDD | 2 | na | 9 | 2.5 | <1.0 | 2.6 | 1.7 | <1.0 | 16.4 |
| 4,4'-DDT | 2 | na | 7 | 1.5 | <1.0 | 5.2 | 5.7 | <1.0 | 10.7 |
| Total DDTs | 3 | 6 | 6 | 2.2 | <1.0 | 3.0 | 6.9 | <1.0 | 10.7 |
| Dieldrin | 2 | na | 6 | <1.0 | 1.1 | 3.0 | <1.0 | <1.0 | 8.7 |

na $=$ no applicable Sediment Quality Guidelines available
Table 20. Test Results for Bayou Chico and Apalachicola Bay.

|  | Amphipod survival |  | Microtox |  | Urchin fertilization |  | Urchin development |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Rho | signif. | Rho | signif. | Rho | signif | Rho | signif. |
| Unionized ammonia in toxicity test chambers |  |  |  |  |  |  |  |  |
| UAN - amphipod | 0.413 | ns |  |  |  |  |  |  |
| UAN - porewater |  |  |  |  | -0.497 | ns | 999-0.464 | ns |
| Major and trace elements, and grain size |  |  |  |  |  |  |  |  |
| Silver | 0.245 | ns | 0.218 | ns | -0.443 | ns | -0.296 | ns |
| Aluminum | 0.494 | ns | 0.383 | ns | -0.153 | ns | -0.365 | ns |
| Arsenic | 0.285 | ns | 0.383 | ns | -0.186 | ns | -0.602 | ns |
| Cadmium | 0.142 | ns | 0.267 | ns | -0.458 | ns | -0.493 | ns |
| Chromium | 0.243 | ns | 0.367 | ns | -0.322 | ns | -0.639 | ns |
| Copper | 0.193 | ns | 0.243 | ns | -0.477 | ns | -0.486 | ns |
| Iron | 0.234 | ns | 0.550 | ns | -0.186 | ns | -0.639 | ns |
| Mercury | 0.067 | ns | 0.250 | ns | -0.458 | ns | -0.493 | ns |
| Manganese | 0.286 | ns | 0.469 | ns | 0.417 | ns | 0.303 | ns |
| Nickel | 0.201 | ns | 0.517 | ns | -0.153 | ns | -0.365 | ns |
| Lead | 0.092 | ns | 0.083 | ns | -0.593 | ns | -0.475 | ns |
| Antimony | 0.142 | ns | 0.267 | ns | -0.458 | ns | -0.493 | ns |
| Selenium | 0.261 | ns | 0.259 | ns | -0.433 | ns | -0.559 | ns |
| Silicon | -0.360 | ns | -0.383 | ns | 0.119 | ns | 0.456 | ns |
| Tin | 0.142 | ns | 0.267 | ns | -0.458 | ns | -0.493 | ns |
| Zinc | 0.067 | ns | 0.250 | ns | -0.458 | ns | -0.493 | ns |
| Sum of 8 metals ERMs | 0.167 | ns | 0.200 | ns | -0.49199 | ns | -0.475 | ns |
| \% gravel | 0.291 | ns | 0.111 | ns | -0.388 | ns | -0.319 | ns |
| \% sand | -0.644 | ns | -0.350 | ns | 0.322 | ns | 0.337 | ns |
| \%silt | 0.628 | ns | 0.300 | ns | -0.373 | ns | -0.248 | ns |
| \% clay | 0.611 | ns | 0.367 | ns | -0.271 | ns | -0.228 | ns |
| \% fines (silt and clay) | 0.644 | ns | 0.356 | ns | -0.322 | ns | -0.337 | ns |
| \% TOC | 0.519 | ns | 0.433 | ns | -0.186 | ns | -0.139 | ns |
| AVS | 0.209 | ns | -0.200 | ns | -0.661 | ns | -0.329 | ns |
| SEM-Cd | 0.126 | ns | 0.150 | ns | -0.542 | ns | -0.402 | ns |
| SEM-Cu | 0.142 | ns | 0.267 | ns | 0.458 | ns | -0.493 | ns |







Table 20 continued.
Major and trace elements, and grain size

|  | Amphipod survival |  |
| :--- | :---: | :---: |
| Rho | signif. |  |
| SEM-Hg | -0.143 | ns |
| SEM-Ni | 0.611 | ns |
| SEM-Pb | 0.109 | ns |
| SEM-Zn | 0.209 | ns |
| Sum 5SEM | 0.209 | ns |
| Sum 5SEM/AVS | -0.017 | ns |
| Sum 5SEM-AVS | 0.142 | ns |

ORGANIC COMPOUNDS Polycyclic aromatic hydrocarbons (PAH) $\begin{array}{lr}\text { \% TOC } & 0.519 \\ \text { NAPHTHALENE } & -0.128 \\ \text { 2-METHYLNAPHTHALENE } & -0.182\end{array}$ 1-METHYLNAPHTHALENE -0.182
C1-NAPHTHALENES 2,6-DIMETHYLNAPHTHALENE $\quad-0.182$ C2-NAPHTHALENES C3-NAPHTHALENES
BIPHENYL
ACENAPHTHYLENE $\begin{array}{ll}\text { ACENAPHTHENE } & -0.207 \\ \text { DIBENZOFURAN } & -0.100\end{array}$ FLUORENE $\begin{array}{ll}\text { C1-FLUORENES } & -0.208 \\ \text { C2-FLUORENES } & -0.195\end{array}$ C3-FLUORENES PHENANTHRENE $\begin{array}{lc}\text { 1-METHYLPHENANTHRENE } & -0.092 \\ \text { ANTHRACENE } & -0.123 \\ \text { C1-PHENANTHRENE/ANTHRACENES-0.178 } \\ \text { C2-PHENANTHRENE/ANTHRACENES-0.082 }\end{array}$ $\begin{array}{lc}\text { 1-METHYLPHENANTHRENE } & -0.092 \\ \text { ANTHRACENE } & -0.123 \\ \text { C1-PHENANTHRENE/ANTHRACENES-0.178 } \\ \text { C2-PHENANTHRENE/ANTHRACENES-0.082 }\end{array}$ $\begin{array}{ll}\text { 1-METHYLPHENANTHRENE } & -0.092 \\ \text { ANTHRACENE } & -0.123 \\ \text { C1-PHENANTHRENE/ANTHRACENES-0.178 } \\ \text { C2-PHENANTHRENE/ANTHRACENES-0.082 }\end{array}$
Urchin development



| Microtox |  |
| :---: | :---: |
| Rho | signif. |
| -0.406 | ns |
| -0.281 | ns |
| -0.369 | ns |
| -0.319 | ns |
| -0.294 | ns |
| -0.094 | ns |
| -0.309 | ns |
| -0.333 | ns |
| -0.200 | ns |
| -0.188 | ns |
| -0.236 | ns |
| -0.042 | ns |
| -0.069 | ns |
| -0.081 | ns |
| -0.081 | ns |
| -0.200 | ns |
| -0.006 | ns |
| -0.02: | ns |
| -0.02: | ns |
| 0.127 | ns |
| -0.006 | ns |
| -0.006 | ns |
| -0.006 | ns |
| -0.345 | ns |
| -0.212 | ns |
| -0.224 | ns |
| -0.224 | ns |
| -0.567 | ns |
| -0.667 | ns |
| -0.683 | ns |
| -0.333 | ns |

Table 20 continued.
Polycyclic aromatic hydrocarbons (PAH) $\begin{array}{lr}\text { SC3-PHENANTHRENE/ANTHRACENES-0.025 } \\ \text { C4-PHENANTHRENE/ANTHRACENES } 0.138 \\ \text { DIBENZOTHIOPHENE } & -0.207 \\ \text { C1-DIBENZOTHIOPHENES } & -0.182 \\ \text { C2-DIBENZOTHIOPHENES } & 0.082 \\ \text { C3-DIBENZOTHIOPHENES } & 0.119 \\ \text { FLUORANTHENE } & -0.316 \\ \text { PYRENE } & -0.237 \\ \text { C1-FLUORANTHENES/PYRENES } & -0.061 \\ \text { BENZ(A)ANTHRACENE } & -0.043 \\ \text { CHRYSENE } & -0.067 \\ \text { C1-CHRYSENES } & -0.055 \\ \text { C2-CHRYSENES } & 0.038 \\ \text { C3-CHRYSENES } & 0.019 \\ \text { C4-CHRYSENES } & 0.019 \\ \text { BENZO(B)FLUORANTHENE } & 0.000 \\ \text { BENZO(K)FLUORANTHENE } & 0.128 \\ \text { BENZO(E)PYRENE } & -0.036 \\ \text { BENZO(A)PYRENE } & -0.036 \\ \text { PERYLENE } & 0.097 \\ \text { INDENO(1,2,3-C,D)PYRENE } & 0.036 \\ \text { DIBENZ(A,H)ANTHRACENE } & 0.128 \\ \text { BENZO(G,H,I)PERYLENE } & -0.024 \\ \text { Sum 10 LPAH } & -0.243 \\ \text { Sum 12 HPAH } & -0.055 \\ \text { sum 22 PAHs } & -0.073 \\ \text { sum all PAHs } \\ \text { acenaphthene (ug/goc) } & -0.073 \\ \text { fluoranthene (ug/goc) } & 0.176 \\ \text { phenanthrene (ug/goc) } & -0.092 \\ \text { Sum of 13 PAH ERMs } & -0.134 \\ & -0.237\end{array}$

Table 20 continued.

|  | Amphipod survival |  |
| :--- | ---: | ---: |
|  | Rho | signif. |
| Chlorinated organic hydrocarbons |  |  |
| CL8(195) | 0.259 | ns |
| CL9(206) | 0.082 | ns |
| CL10(209) | 0.037 | ns |
| sum of 20 PCB congeners | 0.061 | ns |
|  |  |  |
| Pesticide Total : | -0.091 | ns |
| DDT Total : | -0.170 | ns |
| Indeno Total : | 0.251 | ns |
|  |  |  |
| PCB Total (No MDL): | 0.061 | ns |
| Pesticide Total (No MDL): | -0.091 | ns |
| DDT Total (No MDL): | -0.170 | ns |
| Indeno Total (No MDL): | 0.251 | ns |
|  |  |  |
| dieldrin (ug/goc) | 0.293 | ns |
| endrin (ug/goc) | -0.369 | ns |
| Sum of 3 COH ERMs |  |  |
| Sum of 25 ERMs | -0.073 | ns |

Table 21. Stations within Bayou Chico in which chemical concentrations equalled or exceeded their respective sediexceedances are listed in bold.

Greater than or equal to
ERM value ERM value none
none
2-2, 3-3, 4-2, 6-2
none
none
none
none
none
6-2
none
none әuou әuou

2-2, 3-2, 4-2, 5-3, 6-2
none
$2-2,3-2,4-2,5-3,6-2$
$4-2,5-3,6-2$
$2-2,3-2,6-2$
$2-2,3-2,4-2,5-3,6-2$
$2-2,3-2,4-2,6-2$
$2-2,3-2,4-2,5-3,6-2$
$4-2,6-2$
$2-2,3-2,4-2,5-3,6-2$
$3-2,4-2,5-3,6-2$
$4-2$
$4-2,6-2$
2-2, 3-2, 4-2, 6-2
$4-2,5-3,6-2$
$2-2,3-2,6-2$
$2-2,3-2,5-3,6-2$

## $\stackrel{\otimes}{\circ}$

na
na
na
na
ERL and ERM values from Long et al. (1995); TEL and PEL values from MacDonald (1994),SQCs from U. S. EPA, 1994; urchin development LOEC rom Long et al, In press; and amphipod NOEC from Kohn et al., 1994.
Table 22. A comparison of the chemical concentrations (ave $\pm$ std dev) in toxic and non-toxic samples from Bayou toxic average and applicable numerical guidelines (metals in ppm, organics in ppb)*.

| Chemical substance | Not toxic ( $\mathrm{n}=3$ ) | Toxic $(\mathrm{n}=6)$ | Ratio of averages | Guidelines Exceeded (ratio to toxic average) |  | guideline value | ratio |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Silver | $0.1 \pm 0.1$ | $0.2 \pm 0.2$ | 1.3 | <ERL (8.2 ppm) | <1.0 | <TEL (7.2 ppm) | <1.0 |
| Cadmium | $0.5 \pm 0.6$ | $0.6 \pm 0.5$ | 1.3 | <ERL (1.2 ppm) | <1.0 | <TEL (0.67 ppm) | <1.0 |
| Chromium | $41.3 \pm 24.4$ | $42.5 \pm 28.0$ | 1.0 | <ERL (81.0 ppm) | <1.0 | <TEL (52.3 ppm) | <1.0 |
| Copper | $63.7 \pm 77.2$ | $89.9 \pm 61.8$ | 1.4 | >ERL (34.0 ppm) | 2.6 | >TEL (18.7 ppm) | 4.8 |
| Lead | $39.0 \pm 39.4$ | $68.3 \pm 43.8$ | 1.8 | >ERL (46.7 ppm) | 1.5 | >TEL (30.2 ppm) | 2.3 |
| Mercury | $0.2 \pm 0.3$ | $0.3 \pm 0.2$ | 1.3 | >ERL (0.15 ppm) | 1.3 | >TEL (0.13 ppm) | 1.5 |
| Nickel | $19.3 \pm 8.7$ | $20.0 \pm 7.1$ | 1.0 | <ERL (20.9 ppm) | <1.0 | >TEL (15.9 ppm) | 1.3 |
| Zinc | $329.2 \pm 422.0$ | $438.0 \pm 330.3$ | 1.3 | >ERL ( 150 ppm ) | 2.9 | >TEL (124 ppm) | 3.5 |
|  |  |  |  | >ERM (410 ppm) | 1.1 | >PEL (271 ppm) | 1.6 |
| Sum LMW PAH | $581 \pm 815$ | $1723 \pm 1708$ | 3.0 | >ERL (552 ppb) | 3.1 | >TEL (312 ppb) | 5.5 |
|  |  |  |  |  |  | >PEL (1442 ppb) | 1.2 |
| Sum HMW PAH | $3850 \pm 5387$ | $7716 \pm 6533$ | 2.0 | >ERL (1700 ppb) | 4.5 | >TEL (655 ppb) | 11.8 |
|  |  |  |  |  |  | >PEL (6676 ppb) | 1.2 |
| Sum total PAH | $4430 \pm 6202$ | $9440 \pm 8106$ | 2.1 | >ERL (4022 ppb) | 2.3 | >TEL (1684 ppb) | 5.6 |
| Acenaphthene (ug/goc) | $0.9 \pm 1.2$ | $6.8 \pm 6.8$ | 7.6 | <SQC (230 ug/goc) | <1.0 |  |  |
| Fluoranthene (ug/goc) | $9.0 \pm 11.6$ | $107.3 \pm 135.7$ | 11.9 | <SQC (300 ug/goc) | <1.0 |  |  |
| Phenanthrene (ug/goc) | $1.7 \pm 2.3$ | $26.8 \pm 37.6$ | 15.8 | <SQC (240 ug/goc) | <1.0 |  |  |
| Lindane | 0.06 $\pm 0.01$ | $0.5 \pm 1.1$ | 8.2 | na |  | >TEL (0.32 ppb) | 1.6 |
| Heptachlor epoxide | $0.03 \pm 0.0$ | $0.24 \pm 0.5$ | 8.0 | na |  | na |  |
| 2,4' DDE | $0.04 \pm 0.0$ | $0.3 \pm 0.5$ | 8.1 | na |  | na |  |
| 4,4'-DDE | $4.2 \pm 5.7$ | $5.6 \pm 4.7$ | 1.3 | >ERL (2.2 ppb) | 1.9 | >TEL (2.07 ppb) | 2.7 |
| 2,4'-DDD | $2.1 \pm 2.9$ | $2.8 \pm 2.6$ | 1.3 | na |  | na |  |
| 4,4'-DDD | $6.1 \pm 8.5$ | $7.2 \pm 8.1$ | 1.2 | na |  | >TEL (1.22 ppb) | 5.9 |
| 2,4'-DDT | $0.1 \pm 0.0$ | $0.2 \pm 0.4$ | 3.2 | na |  | na |  |
| 4,4'-DDT | $0.3 \pm 0.4$ | $3.8 \pm 3.9$ | 12.1 | na |  | >TEL (1.2 ppb) | 3.2 |
| Sum total DDTs | $12.9 \pm 17.4$ | $20.0 \pm 17.4$ | 1.6 | >ERL (1.58 ppb) | 12.6 | >TEL (3.89 ppb) | 5.1 |
| Dieldrin (ug/goc) | $0.1 \pm 0.1$ | $0.2 \pm 0.1$ | 2.4 | <SQC (20 ug/goc) | <1.0 |  |  |
| Sum total PCBs | $79.7 \pm 108.4$ | 103.9 ${ }^{\text {d }} 87.9$ | 1.3 | >ERL (22.7 ppb) | 4.6 | > TEL (21.6 ppb) | 4.8 |

[^8]Table 23. Test Results for Choctawhatchee Bay.

|  | Amphipod survival |  |
| :--- | :---: | :---: |
|  | Rho | signif. |
|  |  |  |
| Un-ionized ammonia in toxicity test chambers |  |  |
| UAN - amphipod | -0.006 | ns |
| UAN - porewater | nd | nd |
|  |  |  |
| Major and trace elements, and grain size |  |  |
| Silver | 0.247 | ns |
| Aluminum | -0.163 | ns |
| Arsenic | -0.062 | ns |
| Cadmium | 0.185 | ns |
| Chromium | -0.126 | ns |
| Copper | 0.139 | ns |
| Iron | -0.207 | ns |
| Mercury | 0.109 | ns |
| Manganese | -0.056 | ns |
| Nickel | 0.042 | ns |
| Lead | 0.019 | ns |
| Antimony | 0.026 | ns |
| Selenium | 0.117 | ns |
| Silicon | 0.061 | ns |
| Tin | 0.091 | ns |
| Zinc | 0.012 | ns |
| Sum of 9 metals ERMs | 0.048 | ns |
| \% gravel | -0.109 | ns |
| \% sand | 0.026 | ns |
| \%silt | 0.050 | ns |
| \% clay | -0.198 | ns |
| \% fines (silt \& clay) | -0.026 | ns |
| \% TOC | -0.013 | ns |
| AVS | 0.201 | ns |
| SEM-Cd | 0.277 | ns |
| SEM-Cu | 0.276 | ns |
| SEM-Hg | -0.069 | ns |



の の の の の



の๓* 』』


Microtox

の ๑の の の
䓂




』の ๓の』』
Table 23 continued.

| Polycyclic aromatic hydrocarbons (PAH) |  |
| :--- | :--- |
| C3-DIBENZOTHIOPHENES | -0.154 |

    \(\begin{array}{lr}\text { C3-DIBENZOTHIOPHENES } & -0.154 \\ \text { FLUORANTHENE } & 0.062\end{array}\)
    FLUORANTHENE
    CYRELUORANTHENES/PYRENES
                BENZ(A)ANTHRACENE
                    CHRYSENE
    C1-CHRYSENES
                                0.035
    0.057
0.147
0.147
0.112
0.11
0.119
0.129
0.129
-0.058
0.148
0.152
0.147
0.098
$-0.05:$

 | Chlorinated organic hydrocarbons |
| :--- |
| LINDANE |
| CL3（18） |
| CL3（28） |
| HEPTACHLOR |
| CL4（52） |
| ORGANIC COMPOUNDS |

 | Chlorinated organic hydrocarbons |
| :--- |
| LINDANE |
| CL3（18） |
| CL3（28） |
| HEPTACHLOR |
| CL4（52） |
| ORGANIC COMPOUNDS | acenaphthene（ug／goc）

fluoranthene $(\mathrm{ug} / \mathrm{goc})$
phenanthrene $(\mathrm{ug} / \mathrm{goc})$
Sum of 13 PAH ERMs
 BENZO（E）PYRENE BENZO（A）PYRENE PERYLENE
INDENO（1，2，3－C，D）PYRENE DIBENZ（A，H）ANTHRACENE BENZO（G，H，I）PERYLENE Sum 10 LPAH Sum 12 HPAH sum 24 PAHs
sum all PAHs
Chlorinated organic hydrocarbons
Chlorinated organic hydrocarbons

CL4(66)
CL5(101)

TRANS-NONACHLOR DIELDRIN
山
CL5(118)
4,4-DDD
CL5(105)
CL6(138) 0
$\stackrel{N}{N}$
0
0
 CL7(170)
 CL9(206)
CL10(209) sum of 20 PCBs
total PCBs
Pesticide Total :
Indeno Total :


Table 24. Stations within Choctawhatchee Bay in which chemical concentrations equalled or exceeded their respective sediment quality guidelines or toxicity thresholds ( $\mathrm{n}=20$ for organics, $\mathrm{n}=20$ for metals). Stations with two-fold or greater exceedances are listed in bold.

| Chemical $\quad$ Greater substance | Greater than or equal to ERM value | Greater than or equal to PEL value | Greater than or equal to SQC or LOEC/NOEC |
| :---: | :---: | :---: | :---: |
| Silver | F1-1, F2-1 | F1-1, F2-1 | na |
| Dibenzo (a, h) anthracene | none | A1-1 | na |
| Chrysene | none | A1-1 | na |
| Benzo(a) pyrene | none | A1-1 | na |
| Benz(a)anthracene | none | A1-1 | na |
| Dieldrin | na | A1-1 | none |
| Endrin | na | na | C3-1, D1-1 |
| p, p'-DDE | F1-1 | none | na |
| p,p'-DDD | na | F1-1 | na |
| p,p'-DDT | na | F1-1 | na |
| Total DDTs | F1-1 | F1-1 | na |
| UAN LOEC for urchin development | na | na | B3-1, K2-1 |
| UAN LOEC for amphipod survival | na | na | none |

* ERL and ERM values from Long et al. (1995); TEL and PEL values from MacDonald (1994), SQCs from U. S. EPA, 1994; urchin development LOEC from Long et al, In press; and amphipod NOEC from Kohn et al., 1994. na $=$ no applicable values.
Table 25. A comparison of the chemical concentrations (ave $\pm$ std dev) in toxic and non-toxic samples from Choctawhatchee Bay in urchin fertilization tests, the ratio between the two averages, and the ratio between the toxic average and applicable numerical guidelines (metals in ppm, organics in ppb).

| $\begin{aligned} & \text { Toxic } \\ & (\mathrm{n}=15) \end{aligned}$ | Ratio of averages | Guidelines Exceeded (ratio to toxic average) |  | guideline value | ratio |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | guideline value | ratio |  |  |
| $0.65 \pm 1.2$ | 19.8 | <ERL (8.2 ppm) | <1.0 | <TEL (7.2 ppm) | <1.0 |
| $14.6 \pm 8.1$ | 2.7 | >ERL (8.2 ppm) |  | 7:>TEL (7.2 ppm) | 2.0 |
| $0.5 \pm 0.6$ | 8.1 | <ERL (1.2 ppm) | <1.0 | <TEL (0.67 ppm) | <1.0 |
| $66.1 \pm 35.7$ | 3.0 | <ERL (81.0 ppm) | <1.0 | >TEL (52.3 ppm) | 1.3 |
| $28.4 \pm 25.6$ | 3.3 | <ERL (34.0 ppm) | <1.0 | >TEL (18.7 ppm) | 1.5 |
| $53.1 \pm 49.2$ | 7.1 | >ERL (46.7 ppm) | 1.1 | >TEL (30.2 ppm) | 1.8 |
| $0.13 \pm 0.1$ | 5.2 | <ERL (0.15 ppm) | <1.0 | >TEL (0.13 ppm) | 1 |
| $18.4 \pm 10.0$ | 3.1 | <ERL (20.9 ppm) | <1.0 | >TEL (15.9 ppm) | 1.2 |
| 80.5 $\pm 50.1$ | 3.1 | <ERL (150 ppm) | <1.0 | <TEL (124 ppm) | <1.0 |
| $118.8 \pm 183.2$ | 9.4 | <ERL (552 ppb) | <1.0 | <TEL (312 ppb) | <1.0 |
| $2164 \pm 3618$ | 3.3 | >ERL (1700 ppb) | 1.3 | >TEL (655 ppb) | 3.3 |
| $2283 \pm 3801$ | 11.9 | <ERL (4022 ppb) | <1.0 | >TEL (1684 ppb) | 1.4 |
| $0.29 \pm 0.34$ | 4.4 | na |  | $<$ TEL (0.32 ppb) | <1.0 |
| $0.69 \pm 1.4$ | 17.3 | na |  | $<$ TEL (0.71 ppb) | <1.0 |
| $10.0 \pm 18.3$ | 33.5 | >ERL (2.2 ppb) | 4.5 | >TEL (2.07 ppb) | 4.8 |
| $5.2 \pm 12.9$ | 33.2 | na |  | >TEL (1.22 ppb) | 4.3 |
| $1.2 \pm 2.4$ | 6.8 | na |  | >TEL (1.2 ppb) | 1.0 |
| $18.3 \pm 36.5$ | 21.6 | >ERL (1.58 ppb) | 11.4 | >TEL (3.89 ppb) | 4.7 |
| $19.1 \pm 21.0$ | 6.8 | <ERL (22.7 ppb) | <1.0 | <TEL (21.6 ppb) | <1.0 |

* ERL/ERM values from Long et al. (1995), TEL/PEL values from MacDonald (1994), SQC values from U. S. EPA (1994). na $=$ no applicable values.
Table 26．Test Results for St．Andrew Bay． ๑
 の ๑ ๑






- 0
の * か の


##  <br> 


 ＊＊＊＊』 ＊＊＊＊の＊の＊の＊＊＊＊＊＊＊＊の ๓ ＊＊＊＊」

|  | 으ㅈㅡㅣ |  <br>  <br> óo ó ó ó óo óo óo óo ó o |  |  |
| :---: | :---: | :---: | :---: | :---: |

 Amphipod survival
Rho signif．

| Unionized ammonia in toxicity test chambers |  |
| :--- | ---: |
| UAN－amphipod | 0.056 |
| UAN－porewater |  |

$\begin{array}{ll}\text { Major and trace elements，and grain size } \\ \text { Silver } & -0.324\end{array}$

-0.336
-0.283
0.056


|  | Amphipod survival |  | Microtox |  | Urchin fertilization |  | Urchin development |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Rho | signif. | Rho | signif. | Rho | signif. | Rho | signif. |
| Polycyclic aromatic hydrocarbons (PAH) |  |  |  |  |  |  |  |  |
| fluoranthene (ug/goc) | -0.076 | ns | -0.414 | * | -0.007 | ns | -0.050 | ns |
| phenanthrene (ug/goc) | -0.201 | ns | -0.395 | ns | -0.035 | ns | -0.098 | ns |
| Sum of 13 PAH ERMs | -0.286 | ns | -0.535 | * | -0.211 | ns | -0.180 | ns |
| Chlorinated organic hydrocarbons |  |  |  |  |  |  |  |  |
| 2,4-DDE | -0.292 | ns | -0.609 | ** | -0.434 | * | -0.134 | ns |
| CIS-CHLORDANE | -0.302 | ns | -0.470 | * | -0.245 | ns | -0.253 | ns |
| TRANS-NONACHLOR | -0.110 | ns | -0.722 | *** | -0.276 | ns | -0.149 | ns |
| DIELDRIN | -0.179 | ns | -0.627 | ** | -0.213 | ns | -0.099 | ns |
| 4,4-DDE | -0.317 | ns | -0.540 | * | -0.394 | * | -0.141 | ns |
| 2,4-DDD | -0.540 | * | -0.377 | * | -0.337 | ns | -0.151 | ns |
| ENDRIN | <mdl |  | <mdl |  | <mdl |  | <mdl |  |
| 4,4-DDD | -0.311 | ns | -0.520 | * | -0.315 | ns | -0.039 | ns |
| 2,4-DDT | -0.370 | * | -0.328 | ns | -0.201 | ns | 0.074 | ns |
| 4,4-DDT | -0.340 | ns | -0.487 | * | -0.23: | ns | 0.119 | ns |
| sum of 23 PCBs | -0.124 | ns | -0.516 | * | -0.315 | ns | -0.222 | ns |
| DDT Total : | -0.374 | * | -0.552 | * | -0.360 | * | -0.055 | ns |
| Pesticide Total (No MDL): | -0.375 | * | -0.556 | * | -0.356 | ns | -0.062 | ns |
| dieldrin (ug/goc) | -0.066 | ns | -0.450 | * | -0.121 | ns | -0.068 | ns |
| endrin (ug/goc) | 0.308 | ns | 0.472 | * | 0.229 | ns | 0.127 | ns |
| Sum of 3 COH ERMs | -0.348 | ns | -0.538 | * | -0.386 | * | -0.129 | ns |
| Sum of 25 ERMs | -0.351 | ns | -0.621 | * | -0.380 | ns | -0.219 | ns |

[^9]

[^10]Table 28. Correlations between bioassay results and chemical concentrations for all 102 western Florida samples.

|  | Microtox |  | Urchin fertilization |  | Urchin development |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Rho | signif. | Rho | signif. | Rho | signif. |
| Unionized ammonia in toxicity test chambers |  |  |  |  |  |  |
| UAN - porewater | nd | -0.453 | ** | -0.6919: | *** |  |
| Major and trace elements |  |  |  |  |  |  |
| Silver | -0.341 | * | -0.188 | ns | -0.177 | ns |
| Arsenic | -0.061 | ns | -0.308 | * | -0.222 | ns |
| Cadmium | -0.307 | * | -0.270 | ns | -0.238 | ns |
| Chromium | -0.043 | ns | -0.243 | ns | -0.183 | ns |
| Copper | -0.287 | * | -0.309 | * | -0.291 | * |
| Mercury | -0.351 | * | -0.211 | ns | -0.142 | ns |
| Nickel | -0.042 | ns | -0.295 | * | -0.201 | ns |
| Lead | -0.262 | ns | -0.329 | * | -0.255 | ns |
| Antimony | -0.239 | ns | -0.282 | * | -0.166 | ns |
| Selenium | -0.020 | ns | -0.426 | * | -0.279 | * |
| Tin | -0.324 | * | -0.259 | * | -0.186 | ns |
| Zinc | -0.271 | ns | -0.370 | * | -0.365 | * |
| Sum of 9 metals ERMs | -0.245 | ns | -0.330 | * | -0.273 | ns |
| Sum 5 SEM | -0.342 | ns | -0.298 | * | -0.270 | ns |
| Organic compounds |  |  |  |  |  |  |
| Sum LPAH | -0.421 | * | -0.079 | ns | -0.090 | ns |
| Sum HPAH | -0.341 | * | -0.238 | ns | -0.172 | ns |
| Sum PAHs | -0.355 | * | -0.206 | ns | -0.158 | ns |
| Sum all PAHs | +0.224 | ns | -0.601 | *** | -0.454 | ** |
| Acenaphthylene (ug/goc) | +0.084 | ns | -0.341 | * | -0.379 | * |
| Fluoranthene (ug/goc) | -0.065 | ns | -0.250 | ns | -0.335 | * |
| Phenanthrene (ug/goc) | -0.224 | ns | -0.121 | ns | -0.248 | ns |
| Sum of 13 PAH ERMs | -0.375 | * | -0.175 | ns | -0.143 | ns |
| Total PCBs | -0.397 | * | -0.159 | ns | -0.107 | ns |
| Total DDTs | -0.486 | ** | -0.060 | * | +0.057 | ns |
| Total Pesticides | -0.443 | ** | -0.125 | ns | +0.001 | ns |
| dieldrin (ug/goc) | +0.094 | ns | -0.268 | ns | -0.360 | * |
| endrin (ug/goc) | +0.616 | *** | -0.212 | ns | -0.225 | ns |
| Sum of 3 COH ERMs | -0.442 | ** | -0.124 | ns | -0.015 | ns |
| Sum of 25 ERMs | -0.293 | * | -0.312 | * | -0.252 | ns |

SEM = simultaneously-extracted metals
UAN = un-ionized ammonia
ERM = effects range-median
$\mathrm{COHs}=$ chlorinated organic hydrocarbons
PAHs = polynuclear aromatic hydrocarbons
LPAHs = low moluecular weight PAHs
HPAHs = high molecular weight PAHs
nd $=$ no data

## APPENDICES

Appendix A1: Field notes from Pensacola Bay; 1993 and 1994.
APPENDIX A2. Field notes from Choctawhatchee Bay.
Appendix A3. Field notes from St. Andrews Bay.
APPENDIX A4. Field notes from Apalachicola Bay.
Appendix B1. Chemistry and toxicity data from Pensacola Bay, 1993.
Appendix B2. Chemistry and toxicity data from Choctawhatchee Bay, 1994.

Appendix B3. Chemistry and toxicity data from St. Andrew Bay.
Appendix B4. Chemistry and toxicity data from Bayou Chico and Appalachicola Bay.

| APPENDIX <br> Pensacola $\begin{array}{\|r} 1993 \\ \hline \end{array}$ | A1. Field <br> Bay <br> Station | Notes No. | from Pensacola B <br> Location | 1993 a Date | 1994. <br> Time | $\begin{gathered} \text { Latitude } \\ \circ N \mathrm{~N} \\ \hline \end{gathered}$ | $\begin{gathered} \text { Longitude } \\ \circ W \\ \hline \end{gathered}$ | $\begin{gathered} \text { Water } \\ \text { Depth }(M) \\ \hline \end{gathered}$ | Air $\begin{aligned} & \text { Temp. } \\ & { }^{\circ} \mathbf{C}\end{aligned}$ | $\begin{array}{cc} \text { Top } & \text { Temp. } \\ & { }^{\circ} \mathbf{C} \\ \hline \end{array}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 1 |  | Bayou Grande | 3/31/93 | 15:45 | $30^{\circ} 22.548^{\prime}$ | $87^{\circ} 16.743^{\prime}$ | 3.4 | Not Recorded | 22 |
|  | 2 |  | Bayou Grande | 3/31/93 | 15:05 | $30^{\circ} 22.095{ }^{\prime}$ | $87^{\circ} 17.091^{\prime}$ | 2.9 | " | 22 |
|  | 3 |  | Bayou Grande | 3/31/93 | 14:44 | $30^{\circ} 22.258^{\prime}$ | $87^{\circ} 18.771^{\prime}$ | 3 | " | 22 |
|  | 4 |  | Bayou Chico | 3/31/93 |  | $30^{\circ} 24.425^{\prime}$ | $87^{\circ} 15.387^{\prime}$ | 3.2 | " | 22.8 |
|  | 5 |  | Bayou Chico | 3/31/93 | 10:32 | $30^{\circ} 24.317^{\prime}$ | $87^{\circ} 15.377{ }^{\prime}$ | 2.6 | " | 21.5 |
|  | 6 |  | Bayou Chico | 3/31/93 | 10:07 | $30^{\circ} 24.196^{\prime}$ | $87^{\circ} 15.245^{\prime}$ | 2 | " | 22 |
|  | 7 |  | Bayou Chico | 3/31/93 | 9:40 | $30^{\circ} 24.041^{\prime}$ | $87^{\circ} 14.786^{\prime}$ | 3 | " | 20.8 |
|  | 8 |  | Bayou Chico | 3/31/93 | 9:10 | $30^{\circ} 23.966{ }^{\prime}$ | $87^{\circ} 14.667^{\prime}$ | 3.7 | " | 20.8 |
|  | 9 |  | Bayou Chico | 3/31/93 | 8:50 | $30^{\circ} 23.935^{\prime}$ | $87^{\circ} 14.385^{\prime}$ | 5.1 | " | 19.9 |
|  | 10 |  | Bayou Texar | 3/30/92 | 8:55 | $30^{\circ} 27.113^{\prime}$ | $80^{\circ} 12.078^{\prime}$ | 1.1 | " | 18 |
|  | 11 |  | Bayou Texar | 3/30/92 | 9:26 | $30^{\circ} 26.646^{\prime}$ | $87^{\circ} 11.241^{\prime}$ | 2 | " | 18 |
|  | 12 |  | Bayou Texar | 3/30/92 | 9:49 | $30^{\circ} 25.910^{\prime}$ | $87^{\circ} 11.168^{\prime}$ | 2.1 | $\cdots$ | 19.1 |
|  | 13 |  | Bayou Texar | 3/30/92 | 11:17 | $30^{\circ} 25.661^{\prime}$ | $87^{\circ} 11.362^{\prime}$ | 2.5 | " | 19.5 |
|  | 14 |  |  | 3/31/93 | 16:43 |  | $87^{\circ} 15.130^{\prime}$ | 7 | " | 20 |
|  | 15 |  | Bayou Chico | 4/2/93 | 7:22 | $30^{\circ} 23.389^{\prime}$ | $87^{\circ} 13.587^{\prime}$ | 7.5 | " | 18 |
|  | 16 |  | Bayou Channel | 4/1/93 | 11:34 | $30^{\circ} 23.558^{\prime}$ | $8713.989^{\prime}$ | 3.5 | " | 20 |
|  | 17 |  |  | 3/31/93 | 13:12 | $30^{\circ} 24.141^{\prime}$ | $87^{\circ} 13.622^{\prime}$ | 2.2 | " | 21 |
|  | 18 |  | Inner Harbor | 3/31/93 | 13:39 | $30^{\circ} 24.175^{\prime}$ | $87^{\circ} 13.296^{\prime}$ | 3.5 | " | 21 |
|  | 19 |  | Inner Harbor | 4/1/93 | 9:38 | $30^{\circ} 24.134$ | $87^{\circ} 12.895$ | 4.5 | " | 19.5 |
|  | 20 |  | Inner Harbor | 4/1/93 | 10:10 | $30^{\circ} 24.138^{\prime}$ | $87^{\circ} 12.855^{\prime}$ | 4.5 | " | 19.1 |
|  | 21 |  |  | 3/30/93 | 12:15 | $30^{\circ} 23.010^{\prime}$ | $87^{\circ} 13.012^{\prime}$ |  | " | 20 |
|  | 22 |  |  |  | 12:46 | $30^{\circ} 23.463^{\prime}$ | $87^{\circ} 12.949^{\prime}$ | 9.5 | " | 20 |
|  | 23 |  | Inner Harbor | 4/1/93 | 9:00 | $30^{\circ} 23.853^{\prime}$ | $87^{\circ} 13.010^{\prime}$ | 10 | " | 19.5 |
|  | 24 |  | Lower Bay | 4/1/93 | 8:23 | $30^{\circ} 22.081^{\prime}$ | $87^{\circ} 12.650^{\prime}$ | 5.5 | " | 19.5 |
|  | 25 |  | Lower Bay | 3/31/93 | 16:15 | $30^{\circ} 22.521^{\prime}$ | $87^{\circ} 14.464^{\prime}$ | 7.8 | " | 21 |
|  | 26 |  | East Bay | 4/2/93 | 12:36 | $30^{\circ} 28.289^{\prime}$ | $87^{\circ} 03.152^{\prime}$ | 2 | " | 19 |
|  | 27 |  | East Bay | 4/2/93 | 13:01 | $30^{\circ} 26.634^{\prime}$ | $86^{\circ} 59.533^{\prime}$ | 3 | " | 19.5 |
|  | 28 |  | East Bay | 4/2/93 | 13:38 | $30^{\circ} 27.744^{\prime}$ | $86^{\circ} 58.686^{\prime}$ | 2.7 | " | 19.5 |
|  | 29 |  | Blackwater Bay | 4/2/93 | 14:15 | $30^{\circ} 31.726^{\prime}$ | $87^{\circ} 00.973{ }^{\prime}$ | 1.7 | " | 19.9 |
|  | 30 |  | Blackwater Bay | 4/2/93 | 14:49 | $30^{\circ} 32.602^{\prime}$ | $87^{\circ} 00.788^{\prime}$ | 3.2 | " | 20.5 |
|  | 31 |  | Blackwater Bay | 4/2/93 | 15:18 | $30^{\circ} 34.817^{\prime}$ | $87^{\circ} 00.661^{\prime}$ | 4.5 | " | 19.9 |
|  | 32 |  |  | 3/29/93 | 15:39 | $30^{\circ} 28.886^{\prime}$ | $87^{\circ} 07.199^{\prime}$ | 3.1 | " | 21 |
|  | 33 |  |  | 3/29/93 | 14:52 | $30^{\circ} 31.169^{\prime}$ | $87^{\circ} 07.219^{\prime}$ | 2.4 | " | 18.5 |
|  | 34 |  |  | 3/29/93 | 14:21 | $30^{\circ} 31.621^{\prime}$ | $87^{\circ} 08.723^{\prime}$ | 2.5 | " | 20 |
|  | 35 |  |  | 3/29/93 | 13:49 | $30^{\circ} 32.137{ }^{\prime}$ | $87^{\circ} 10.797^{\prime}$ | 1.7 | " | 20.5 |
|  | 36 |  |  | 3/29/93 | 13:16 | $30^{\circ} 32.923{ }^{\prime}$ | $87^{\circ} 11.299^{\prime}$ | 1.3 | " | 21.2 |
|  | 37 |  |  | 3/29/93 | 12:02 | $30^{\circ} 33.754^{\prime}$ | $87^{\circ} 10.479^{\prime}$ | 1 | " | 20 |
|  | 38 |  | Escambia | 3/29/93 | 11:24 | $30^{\circ} 33.337{ }^{\prime}$ | $87^{\circ} 09.150^{\prime}$ | 2.4 | " | 18.5 |
|  | 39 |  | Floridatown | 3/29/93 | 10:21 | $30^{\circ} 34.470^{\prime}$ | $87^{\circ} 09.960^{\prime}$ | 2.5 | " | 22 |
|  | 40 |  | Central Bay | 3/30/93 | 7:46 | $30^{\circ} 24.335^{\prime}$ | $87^{\circ} 07.090^{\prime}$ | 5.5 | " | 20 |



Appendix A1. Field notes from Pensacola Bay; 1993 and 1994 (continued).

| 1 | Bayou Grande | Chocolate brown pudding, oxic brown fine silt, sulfurous below 1 cm . |
| :---: | :---: | :---: |
| 2 | Bayou Grande | 1 cm . oxic silty sediment surface over light grey clayey sand. No odors. No benthos |
| 3 | Bayou Grande | Puddin extraordinaire, fine silty clay,oxic lt. brn oozy over drk green silty clay. No benthos. |
| 4 | Bayou Chico | Clayey, light brown oxic layer over dark sticky clay (consolidated), sand component. |
| 5 | Bayou Chico | Very sulfurous, stinko yukko. Silty brown mud overlying sulfurous dark anoxic silt. |
| 6 | Bayou Chico | Soupy silt. 1st cm. thick oxic layer, lt. brn over blk silt, sm. sand component, organic silty mud. |
| 7 | Bayou Chico | Slight H2S, slight petro. sheen, brn. silty mud, thin oxic layer over drk silt, nonconsolidated goo. |
| 8 | Bayou Chico | Petro sheen mainly gooey Drk. brn silty mud, very organic w/thin brn layer on top. |
| 9 | Bayou Chico | Oily sheen, drk brn, 1 cm blk anoxic under runny silty clay w/distinct petro sheen, Petro blobs. |
| 10 | Bayou Texar | Puddin w/ debris. Very silty, light brown on surface, runny... CAN'T READ Very light H2S |
| 11 | Bayou Texar | Lt. brn, $2-3 \mathrm{~cm}$ deep, dark gray brown, runny silt. Very slight H2S odor,sm. organic component. |
| 12 | Bayou Texar | Same as number 11. |
| 13 | Bayou Texar | Silty, soupy brown puddin 2 cm . Rich in organics (leaf debris) over gray clay puddin layer. |
| 14 |  | Brown 1 cm oxic layer over black silty clay. |
| 15 | Bayou Chico | Homogeneous,one layer of drk olive green, grey clay-no odor,brn oxic on olive consoldated clay. |
| 16 | Bayou Channel | Fine to med grade sand with silt component over brn-grey mud, a few shell fragments. |
| 17 |  | Lt. brn on drk sandy retro sheen from sediments,silt fractions, mostly sand, blk organic detritus. |
| 18 | Inner Harbor | Dark brown 1 cm muddy silt,dark grey beneath small sand component-some leaves. |
| 19 | Inner Harbor | Thin flocculent surface. Lt. brn, very small shell frags. on surface over green/grey silty clay. |
| 20 | Inner Harbor | 1 cm brown oxic flocculent over green/black, muddy, silty clay--shell frags. \& some rocks. |
| 21 |  | Lt. brn clayey silt over a dark grey clay: small shrimp. |
| 22 |  | Same as 21 |
| 23 | Inner Harbor | Oxic, light, brown, silty clay. Over green/grey, silty clay. |
| 24 | Lower Bay | 5 cm thin, solid, brown, oxic, silty, clay layer over dark greenish/grey, silty clay. |
| 25 | Lower Bay | Silty brn goo, $1 / 2 \mathrm{in}$ oxic floc surface, tan lt. brown over grey silty over semi-consolodated clay. |
| 26 | East Bay | Dark olive, brown clay 2 cm . No distinct odor. 1st layer over dark brown clay. No vis benthos. |
| 27 | East Bay | 1 cm oxic, brown,silty, clay over semi CAN'T READ |
| 28 | East Bay | 1 cm brown/gray, oxic clay surface layer over grey clay w/ slit. |
| 29 | Blackwater Bay | Brn oxic layer 1 cm on drk grey, silty clay. Worm tube, organic matter,wood decay, lot of worms. |
| 30 | Blackwater Bay | Brown oxic $1-2 \mathrm{~cm}$ clayey over dark grayish green clay silt |
| 31 | Blackwater Bay | Brown oxic thin silty clay surface layer over brown sandy clay. Sandy dark gray clay. |
| 32 |  | 2.5 cm silty,oxic, tan brown layer over grayish clay. |
| 33 |  | Brown, runny, oozy clay. 2.5 cm surface, darker clay below. |
| 34 |  | Runny, gooey, brown/black clay. |
| 35 |  | 2 cm brown, oxic clay layer over dark green/black clay. NUSL in first grabs. Wormtubes. |
| 36 |  | Silty tan mud, 2 cm oxic, sm. sand component, amphipods, leaf debris on blk anoxic layer w/sand |
| 37 |  | 1.3 cm thick sandy oxic layer on top of black organic silty sand. |
| 38 | Escambia | Soupy,runny silty clay,med. tan clay w/drk blue-grey lens under 1 cm brn layer, organic debris. |
| 39 | Floridatown | Green filamentous simple cell scum layer on top lt. tan few shell frags. Clay, sand composition. |
| 40 | Central Bay | Olive gray clay "like pudding". 1 cm layer over darker grayish black clay. |



Appendix A1. Field notes from Pensacola Bay; 1993 and 1994 (continued).
Air
Temperature
${ }^{\circ} \mathrm{C}$



APPENDIX A2. Field notes from Choctawhatchee Bay.
Choctawhatchee Bay

$\sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i} \sum_{i}$

 $\begin{array}{lcrl}\text { Choctawhatchee Bay } & \\ \text { Block Station } & \text { Station Location }\end{array}$

Appendix A1. Field notes from Pensacola Bay; 1993 and 1994 (continued).


Appendix A2. Field notes from Choctawhatchee Bay; 1993 and 1994.


Appendix A2. Field notes from Choctawhatchee Bay; 1993 and 1994 (continued).

| Appendix A3. Field notes from St. Andrews Bay. |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| St. Andrews Bay |  |  |  |  |  |  |  |  |  |
|  |  |  |  |  |  | Water | Surface | Top Sal. | Surface |
| Station | \# Location | Date | Time | Latitude | Longitude | Depth(m) | Temp. ${ }^{\circ} \mathrm{C}$ | ppt | D. O.mg/l |
| 41 |  | 4/20/93 | 8:32 | $30^{\circ} 15.492^{\prime}$ | 85047.389' | 3.25 | 21 | 19 | 7.2 |
| 42 | West Bay | 4/20/93 | 9:22 | $30^{\circ} 15.166^{\prime}$ | $85^{\circ} 45.219^{\prime}$ | 5 | 20 | 19.8 | 7.4 |
| 43 | North Bay | 4/20/93 | 12:25 | $30^{\circ} 12.011^{\prime}$ | $85^{\circ} 44.116^{\prime}$ | 11 | 21 | 22.8 | 7.6 |
| 44 | North Bay | 4/20/93 | 11:40 | $30^{\circ} 13.752^{\prime}$ | $85^{\circ} 41.849^{\prime}$ | 6.9 | 18.9 | 21 | 7.5 |
| 45 | North Bay | 4/20/93 | 11:00 | $30^{\circ} 14.781^{\prime}$ | $85^{\circ} 41.072^{\prime}$ | 4.6 | 19 | 13 | 7.8 |
| 46 | North Bay | 4/20/93 | 10:12 | $30^{\circ} 15.194^{\prime}$ | $85^{\circ} 40.090^{\prime}$ | 3.9 | 20.2 |  | 7.7 |
| 47 |  | 4/20/93 | 10:32 | $30^{\circ} 14.741^{\prime}$ | $85^{\circ} 39.662^{\prime}$ |  | 20.5 |  | 7.5 |
| 48 |  | 4/20/93 | 13:40 | $30^{\circ} 09.929{ }^{\prime}$ | $85^{\circ} 43.562^{\prime}$ | 7 | 20.9 | 23 | 7.4 |
| 49 | St. Andrews | 4/20/93 | 14:10 | $30^{\circ} 09.562^{\prime}$ | $85^{\circ} 42.415^{\prime}$ | 40 ft . | 20.8 | 22.5 | 7.6 |
| 50 | St. Andrews | 4/20/93 | 15:00 | $30^{\circ} 08.956^{\prime}$ | $85^{\circ} 40.785^{\prime}$ | 12 | 20 | 25.3 | 7.4 |
| 51 |  | 4/21/93 | 9:20 | $30^{\circ} 10.016^{\prime}$ | $85^{\circ} 42.345^{\prime}$ | 6.1 | 20.1 | 26.2 | 7.7 |
| 52 | St. Andrews | 4/20/93 | 13:00 | $30^{\circ} 10.533^{\prime}$ | $85^{\circ} 43.315^{\prime}$ | 6.5 | 21.2 | 22.5 | 7.8 |
| 53 | Massalina Bayou |  | 10:40 | $30^{\circ} 09.239^{\prime}$ | $85^{\circ} 39.354^{\prime}$ | 2.2 | 19 | 24 | 6.7 |
| 54 | Massalina Bayou | 4/22/93 | 11:10 | $30^{\circ} 09.140^{\prime}$ | $85^{\circ} 39.384^{\prime}$ | 2.3 | 19 | 23.2 | 6.9 |
| 55 | Watson Bayou | 4/21/93 | 10:15 | $30^{\circ} 09.346{ }^{\prime}$ | $85^{\circ} 38.418^{\prime}$ | 4 | 22 | 21.3 | 7.1 |
| 56 | Upper Watson | 4/21/93 | 10:40 | $30^{\circ} 09.179^{\prime}$ | $85^{\circ} 38.380^{\prime}$ | 2.9 | 22 | 25 | 7.3 |
| 57 | Upper Watson | 4/21/93 | 11:03 | $30^{\circ} 09.077^{\prime}$ | $85^{\circ} 38.336{ }^{\prime}$ | 3.6 | 22.1 | 23.6 | 7.4 |
| 58 | Mid Watson | 4/21/93 | 11:21 | $30^{\circ} 08.834^{\prime}$ | $85^{\circ} 38.006^{\prime}$ | 4.4 | 22.9 | 20.9 | 7.6 |
| 59 | Mid Watson | 4/21/93 | 11:52 | $30^{\circ} 08.656^{\prime}$ | $85^{\circ} 37.960^{\prime}$ | 4.7 | 23.2 | 19.5 | 7.4 |
| 60 | Mid Watson | 4/21/93 | 12:17 | $30^{\circ} 08.862^{\prime}$ | $85^{\circ} 37.707^{\prime}$ | 2.4 | 23 | 19.5 | 7.6 |
| 61 | Lower Watson | 4/21/93 | 12:51 | $30^{\circ} 08.480^{\prime}$ | $85^{\circ} 38.469^{\prime}$ | 2.7 | 23 | 20.5 | 7.3 |
| 62 | Lower Watson | 4/21/93 | 13:52 | $30^{\circ} 08.454^{\prime}$ | $85^{\circ} 38.146^{\prime}$ | 2 | 23 | 22.2 | 7.4 |
| 63 | Watson Bayou | 4/21/93 | 14:24 | $30^{\circ} 08.444^{\prime}$ | $85^{\circ} 37.993^{\prime}$ | 4.4 | 23 | 22.1 | 7.7 |
| 64 | St. Andrew Bay | 4/22/93 | 9:07 | $30^{\circ} 08.443^{\prime}$ | $85^{\circ} 39.369^{\prime}$ | 10.4 | 19 | 25.2 | 7.3 |
| 65 | St. Andrew Bay | 4/22/93 | 8:40 | $30^{\circ} 08.039^{\prime}$ | $85^{\circ} 38.948^{\prime}$ | 7.2 | 19 | 24.5 | 7.2 |
| 66 | Mouth of Watson Bayou | 4/21/93 | 14:50 | $30^{\circ} 08.058^{\prime}$ | $85^{\circ} 38.092^{\prime}$ | 5.4 | 23 | 22 | 7.6 |
| 67 |  | 4/19/93 | 15:46 | $30^{\circ} 07.573^{\prime}$ | $85^{\circ} 36.759^{\prime}$ | 7.5 | 21 | 19 | 8.2 |
| 68 | Mouth of Pearl Bayou | 4/19/93 | 17:08 | $30^{\circ} 06.214^{\prime}$ | $85^{\circ} 36.669^{\prime}$ | 6.7 | 21 | 17.2 | 8.2 |
| 69 | Smak Bayou | 4/22/93 | 9:50 | $30^{\circ} 07.729^{\prime}$ | $85^{\circ} 40.003^{\prime}$ | 3.4 | 19 | 22.6 | 7.1 |
| 70 | West-East Bay | 4/19/93 | 16:24 | $30^{\circ} 05.284^{\prime}$ | $85^{\circ} 33.068^{\prime}$ | 3.4 | 20.5 | 19.1 | 8.3 |
| 71 | East-East Bay | 4/19/93 | 15:36 | $30^{\circ} 02.498^{\prime}$ | $85^{\circ} 30.093^{\prime}$ | 2 | 24 | 12 | 8.3 |

Appendix A3. Field notes from St. Andrew Bay.

| Appendix A3. Field notes from St. Andrews Bay. |  |  |  |  | Bottom | Bottom |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| St. Andrews Bay |  |  |  |  |  |  |  |
|  |  | Top | Bottom | Bottom |  |  |  |
| tation | Location | Cond. | Temp. | Sal. | D. 0 . | Cond. | Sediment Description |
| 41 |  | 28300 | 21 | 19.7 | 6.6 | 28800 | Lt. brn silty oxic 1cm on top of 2cm sand (lt. gray) over drk gray clay, silty sand |
| 42 | West Bay | 28900 | 20 | 26.8 | 7.4 | 38100 | Olive, gray, silty clay homogenous. Brown flocculent layer on top. |
| 43 | North Bay | 32600 | 21 | 24.3 | 3.3 |  |  |
| 44 | North Bay | 31100 | 20.2 | 24.2 | 5.3 | 36100 | Olive 1 cm silt surface over black, anoxic, silty, clay. Plant debris. |
| 45 | North Bay | 19800 | 19.1 | 25.9 | 6.8 | 37900 |  |
| 46 | North Bay | 22800 | 20.5 | 26 | 5.8 |  |  |
| 47 |  |  | 23.9 |  | 7 |  |  |
| 48 |  | 33800 | 19 | 28.5 | 5.9 | 41000 | Olive gray brown sandy clay. Some shell frags. Polychaetes, amphipods--signs. |
| 49 | St. Andrews | 33000 | 18.9 | 29.9 | 6.6 | 42800 | Sandy mud oxic--surface greenish brown 1 cm over clay sand layer CAN'T READ |
| 50 | St. Andrews | 36900 | 21 | 30.7 |  | 39100 | Grey-brn silt clay on drk grey anoxic silty clay, poorly consolidated, no sand,sulfur odor. |
| 51 |  | 32300 | 19.5 | 34.9 | 6.4 | 40000 | 5 cm brown oxic surface over dark grey-silty, sandy clay. Few shell frags. |
| 52 | St. Andrews | 32900 | 20.5 | 28.5 | 6.3 | 40900 | Brown oxic layer over dark gray silty sand. Shell fragments. |
| 53 | Massalina Bayou | 34000 | 20.2 | 28.7 | 6.3 | 39000 | Brown green flocculent surface over grey black silty clay-leaf debris. |
| 54 | Massalina Bayou | 32900 | 20.5 | 29 | 5.7 | 40000 | .5 cm brown flocculent over grey black sandy silty clay. Plant debris abundant. |
| 55 | Watson Bayou | 31800 | 20.8 | 25.5 | 1.1 | 36200 | No surface layer=homogenous sandy, silty clay drk, gray-blk color, sulfur odor, anoxic. |
| 56 | Upper Watson | 31800 | 21 | 29.6 | 3.1 | 36500 | Gray-blk anoxic silty clay (H2S), flocculent layer, blk flecks on surface, 1 mm brn silty top. |
| 57 | Upper Watson | 31400 | 20.9 | 26 | 6.4 | 36600 | Black grey anoxic silty clay (H2S) poorly consolidated. 1 mm light brown silty surface. |
| 58 | Mid Watson | 31000 | 20.5 | 28.1 | 6.3 | 38100 | Black grey anoxic silty clay (H2S) poorly consolidated. 1 mm brown silty surface. |
| 59 | Mid Watson | 30500 | 20.8 | 27.2 | 7 | 39900 | .5 cm grey surface over sandy silty dark grey clay. Shell fragments. |
| 60 | Mid Watson | 30900 | 22.2 | 21.1 | 6.3 | 33300 | Med gray top 1 cm silt on sandy clay-med gray. Slight H2S, organic debris, flocculent on top. |
| 61 | Lower Watson | 30100 | 21.9 | 27.1 | 7.6 | 37200 | Brn flocculent layer on top,brn oxic 1cm-over drk gray blk silty clay,tiny amt. plant debris. |
| 62 | Lower Watson | 32000 | 22.5 | 22.8 | 6.9 | 32500 | Light brown oxic no H2S. Sandy mud, some pine needles CAN'T READ |
| 63 | Watson Bayou | 32000 | 20.9 | 29 | 7.7 | 31.9 | Olive surface layer 1 mm on top. Olive gray silty clay. |
| 64 | St. Andrew Bay | 35900 | 18.8 | 31.2 | 6.3 | 42200 | Fine shell frags. on surface. Brownish green 1 cm over gray green sandy clay. |
| 65 | St. Andrew Bay | 36500 | 18.9 | 30.2 | 6.2 | 41500 | Brown, olive, green, over grayish green sandy clay w/ shell frags. "luminous surface" |
| 66 | Mouth of Watson Bayou | 32700 | 20.1 | 28.1 | 7.3 | 40900 | Algal scum on top over grey green clayey mud. Diatom scum. |
| 67 |  | 27900 | 18.5 | 30 | 6.3 |  |  |
| 68 | Mouth of Pearl Bayou | 25300 | 18.1 | 29.8 | 6 | 39800 | Surface brown grey 1 cm . Silty clay over dark green grey clay. |
| 69 | Smak Bayou | 32200 | 19.5 | 26.9 | 6.6 | 37500 | Brn .5 cm silty surface over grey silty sand worm tubes. Algae green-brn scum flocculent. |
| 70 | West-East Bay | 27900 | 20 | 23 | 7.6 | 33000 | Floc. layer,olive brn semi-consolidated 2 cm on grey grn silty clay w/fine sand,H2S light. |
| 71 | East-East Bay | 19100 | 20 | 19 | 8.4 | 27000 | Lt. brn 1 cm over brn-blk med-fine grained silty sand. Worm tubes, silty clayey fraction. |

Appendix A3. Field notes from St. Andrew Bay (continued).

| APPEN | DIX A | Field not | es fr | $m$ Apal | hicola |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Apalach | cola |  |  |  |  |  |  |  |  |  |  |
|  |  |  |  |  |  |  |  |  | Air | Surface | Bottom |
| Block | Station | Station Lo | cation | Date | Time | Latitude | Longitude | Depth <br> m | Temp. | Temp | Temp. <br> ${ }^{\circ} \mathrm{C}$ |
| (A) B | 1-1 | Apalachicola | Bay | 6/11/94 | 8:58 AM | $30^{\circ} 28.43{ }^{\prime} \mathrm{N}$ | $86^{\circ} 35.47$ 'W | 1.7 | 26.0 | 28.0 | 27.4 |
| (A)B | 3-1 | Apalachicola | Bay | 6/11/94 | 9:35 AM | $29^{\circ} 42.65{ }^{\text {'N }}$ | $84^{\circ} 53.98$ 'W | n d | n d | 2.7 | 28.5 |
| (A)B | 2-1 | Apalachicola | Bay | 6/11/94 | 10:20 AM | $29^{\circ} 39.65{ }^{\text {'N }}$ | $84^{\circ} 56.53$ 'W | 3.4 | nd | 27.9 | 27.8 |
| (A)C | 3-1 | Apalachicola | Bay | 6/11/94 | 11:08 AM | $29^{\circ} 36.81$ 'N | $84^{\circ} 59.66{ }^{\text {'W }}$ | 3.6 | nd | 28.5 | 29.0 |
| (A) C | 2-1 | Apalachicola |  | 6/11/94 | 12:11 PM | $29^{\circ} 39.28{ }^{\text {'N }}$ | $85^{\circ} 03.21{ }^{\text {'W }}$ | 2.7 | 30.0 | 29.9 | 29.0 |
| (A) C | 1-1 | Apalachicola |  | 6/11/94 | 13:36 PM | $29^{\circ} 42.96{ }^{\prime} \mathrm{N}$ | $84^{\circ} 59.23$ 'W | 1.6 | nd | 25.1 | 30.1 |
| (A) A | 1-1 | Apalachicola | River | 6/11/94 | 15:04 PM | $29^{\circ} 45.29{ }^{\prime} \mathrm{N}$ | $85^{\circ} 00.69$ 'W | 4.4 | 30.5 | 28.5 | 28.1 |
| (A) A | 2-1 | Apalachicola | River | 6/11/94 | 15:50 PM | $29^{\circ} 44.47{ }^{\prime} \mathrm{N}$ | $84^{\circ} 59.89$ 'W | 5.4 | n d | 29.1 | 28.9 |
| (A) A | 3-1 | Apalachicola | River | 6/11/94 | 16:49 PM | $29^{\circ} 43.63{ }^{\prime} \mathrm{N}$ | $84^{\circ} 58.46$ 'W | 1.7 | n d | 29.8 | 29.5 |

Appendix A4. Field notes from Apalachicola Bay.

| APPEN | X A4. | eld $n$ | S from | palachic | a Bay. |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Apalachi | ola Bay |  |  |  |  |  |  |  |
|  |  | Air | Surface | Bottom | Surface | Bottom | Surface | Bottom |
| Block | Station | Temp. | Temp | Temp. | Salinity | Salinity | D.O. | D.O. |
| \# | \# | ${ }^{\circ} \mathrm{C}$ | ${ }^{\circ} \mathrm{C}$ | ${ }^{\circ} \mathrm{C}$ | ppt | ppt | mg/L | mg/L |
| (A) B | 1-1 | 26.0 | 28.0 | 27.4 | 12.4 | 13.0 | 7.0 | 6.9 |
| (A)B | 3-1 | nd | 2.7 | 28.5 | 10.4 | 10.6 | 6.5 | 5.7 |
| (A)B | 2-1 | nd | 27.9 | 27.8 | 15.9 | 28.7 | 6.9 | 5.7 |
| (A) C | 3-1 | nd | 28.5 | 29.0 | 29.8 | 28.9 | 6.2 | 4.7 |
| (A) C | 2-1 | 30.0 | 29.9 | 29.0 | 28.5 | 28.6 | 6.7 | 6.4 |
| (A) C | 1-1 | n d | 25.1 | 30.1 | 2.6 | 17.2 | 7.7 | 7.4 |
| (A) A | 1-1 | 30.5 | 28.5 | 28.1 | 0.0 | 0.0 | 6.2 | 6.1 |
| (A) A | 2-1 | n d | 29.1 | 28.9 | 0.0 | 22.8 | 6.5 | 3.6 |
| (A) A | 3-1 | n d | 29.8 | 29.5 | 0.3 | 0.2 | 6.6 | 6.6 |

Appendix A4. Field notes from Apalachicola Bay (continued).



Appendix A4. Field notes from Apalachicola Bay (continued).





$\frac{1}{5}$





Appendix B1. Chemistry and toxicity data from Pensacola Bay, 1993.


$$
1
$$

$$
\frac{0}{2}
$$

SAMPLE



앙ㅇㅇㅇㅁN








으은


[^11] ppb


28 East Bay
29 Blackwater Bay
30 Blackwater Bay
31 Blackwater Bay
32 Escambia Bay
33 Escambia Bay
34 Escambia Bay
35 Escambia Bay
36 Escambia Bay
28 East Bay
29 Blackwater Bay
30 Blackwater Bay
31 Blackwater Bay
32 Escambia Bay
33 Escambia Bay
34 Escambia Bay
35 Escambia Bay
36 Escambia Bay 36 Escambia Bay
37 Escambia River Mouth 37 Escambia River Mouth
39 Fscambia
39 Floridatown 39 Floridatown $-$


| Station No. | Location | Mirex ppb | SAMPLE | PCBCON8 ppb | PCBCON18 ppb | PCBCON29 ppb | PCBCON52 ppb | PCBCON44 ppb | PCBCON66 ppb | PCBCON101 ppb | PCBCON77 ppb |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | Bayou Grande | 0.5 | 1 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 11.31 | 7.17 |
| 2 | Bayou Grande | 0.5 | 2 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 3 | Bayou Grande | 0.5 | 3 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 4 | Bayou Chico | 0.5 | 4 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 5 | Bayou Chico | 24.28 | 5 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 6 | Bayou Chico | 0.5 | 6 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 6.48 | 18 | 49.92 |
| 7 | Bayou Chico | 11.18 | 7 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 8 | Bayou Chico | 11.22 | 8 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 9 | Bayou Chico | 0.5 | 9 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 10 | Bayou Texar | 0.5 | 10 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 11 | Bayou Texar | 0.5 | 11 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 12 | Bayou Texar | 0.5 | 12 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 13 | Bayou Texar | 0.5 | 13 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 14 | Warrington | 0.5 | 14 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 15 | Bayou Chico | 0.5 | 15 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 16 | Bayou Channel | 0.5 | 16 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 17 | Inner Harbor Channel | 0.5 | 17 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 18 | Inner Harbor | 0.5 | 18 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 19 | Inner Harbor | 0.5 | 19 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 20 | Inner Harbor | 0.5 | 20 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 21 | Pensacola Bay | 0.5 | 21 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 22 | Pensacola Bay | 0.5 | 22 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 23 | Inner Harbor | 0.5 | 23 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 24 | Lower Bay | 0.5 | 24 | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |
| 25 | Lower Bay |  | 25 |  |  |  |  |  |  |  |  |
| 26 | East Bay |  | 26 |  |  |  |  |  |  |  |  |
| 27 | East Bay |  | 27 |  |  |  |  |  |  |  |  |
| 28 | East Bay |  | 28 |  |  |  |  |  |  |  |  |
| 29 | Blackwater Bay |  | 29 |  |  |  |  |  |  |  |  |
| 30 | Blackwater Bay |  | 30 |  |  |  |  |  |  |  |  |
| 31 | Blackwater Bay |  | 31 |  |  |  |  |  |  |  |  |
| 32 | Escambia Bay |  | 32 |  |  |  |  |  |  |  |  |
| 33 | Escambia Bay |  | 33 |  |  |  |  |  |  |  |  |
| 34 | Escambia Bay |  | 34 |  |  |  |  |  |  |  |  |
| 35 | Escambia Bay |  | 35 |  |  |  |  |  |  |  |  |
| 36 | Escambia Bay |  | 36 |  |  |  |  |  |  |  |  |
| 37 | Escambia River Mouth |  | 37 |  |  |  |  |  |  |  |  |
| 38 | Escambia |  | 38 |  |  |  |  |  |  |  |  |
| 39 | Floridatown |  | 39 |  |  |  |  |  |  |  |  |
| 40 | Central Bay |  | 40 |  |  |  |  |  |  |  |  |
| MDL |  | 0.5 |  | 0.75 | 0.51 | 0.94 | 0.51 | 0.58 | 0.96 | 0.83 | 1.03 |







| Appendix <br> Battelle ID $1994$ | B2. Chemistry and |  | toxicity data from Choctawhatchee Bay, 1994. |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Site Location | C3-NAPHTHALENES | C4-NAPHTHALENES | BIPHENYL | ACENAPHTHYLENE | ACENAPHTHENE | DIBENZOFURAN | FLUORENE | C1-FLUORENES |
| PE02 | A 1-1 | Cinco Bay | 0.21 | 0.21 | 7.71 | 82.15 | 16.79 | 20.66 | 24.77 | 11.02 |
| PE19 | A 2-1 | Cinco Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE22 | B 3-1 | U. Garnier Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE05 | C1-2 | L. Garnier Bayou | 0.21 | 0.21 | 0.2 | 35.83 | 0.28 | 0.23 | 4.54 | 0.23 |
| PE03 | C2-1 | L. Garnier Bayou | 0.21 | 0.21 | 0.2 | 46.37 | 0.28 | 4.82 | 7.41 | 0.23 |
| PE16 | C3-1 | L. Garnier Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE12 | D 1-1 | Destin Harbor | 0.21 | 0.21 | 0.2 | 1.58 | 0.28 | 0.23 | 0.23 | 0.23 |
| PD98 | D 2-2 | Destin Harbor | 0.21 | 0.21 | 0.2 | 12.34 | 0.28 | 0.23 | 0.23 | 0.23 |
|  | D 3-1 | Destin Harbor |  |  |  |  |  |  |  |  |
| PE13 | E3-1 | Boggy Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE23 | F1-1 | Toms Bayou | 0.21 | 0.21 | 0.2 | 50.53 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE11 | F2-1 | Tom's Bayou | 0.21 | 0.21 | 0.2 | 39.25 | 0.28 | 0.23 | 0.23 | 0.23 |
| PD91 | G1-1 | Rocky Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PD96 | G2-1 | Rocky Bayou | 0.21 | 0.21 | 0.2 | 34.56 | 0.28 | 0.23 | 0.23 | 0.23 |
| PD99 | G3-1 | Rocky Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE06 | K2-1 | La Grange Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE04 | K3-1 | La Grange Bayou | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE01 | L1-2 | western basin | 0.21 | 0.21 | 0.2 | 7.46 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE21 | L3-1 | western basin | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 1.13 | 0.23 |
| PE09 | M 1-1 | central basin | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PE07 | N3-1 | eastern basin | 0.21 | 0.21 | 0.2 | 3.37 | 0.28 | 0.23 | 0.23 | 0.23 |
| PO15PB | NA | Procedural Blank | K 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |
| PO19SSRM | $\begin{aligned} & \text { BOS ID } \\ & \# 940504-4 \end{aligned}$ | SRM 1941a | 286.32 | 165.88 | 101.39 | 89.71 | 38.09 | 105.71 | 76.75 | 75.79 |
| MDLs |  |  | 0.21 | 0.21 | 0.2 | 0.12 | 0.28 | 0.23 | 0.23 | 0.23 |


| $\begin{array}{\|} \hline \text { Appendix } \\ \text { Battelle ID } \\ 1994 \end{array}$ | B2. Chemistry and toxicity data from Choctawhatchee Bay, 1994. |  |  |  |  |  | ANTHRACENE C1-PHENANTHRENE/ANTHRACENES C2-PHENANTHRENE/ANTHRACENES |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Sta. No. | Site Location | C2-FLUORENES | C3-FLUORENES | PHENANTHRENE | 1-METHYLPHENANTHRENE |  |  |  |
| PE02 | A 1-1 | Cinco Bay | 26.37 | 362.11 | 360.54 | 72.45 | 156.94 | 152.74 | 154.64 |
| PE19 | A 2-1 | Cinco Bayou | 0.23 | 0.23 | 0.19 | 0.19 | 0.17 | 0.19 | 0.19 |
| PE22 | B 3-1 | U. Garnier Bayou | 0.23 | 0.23 | 1.84 | 0.19 | 1.38 | 0.19 | 0.19 |
| PE05 | C 1-2 | L. Garnier Bayou | 0.23 | 0.23 | 67.23 | 8.44 | 42.31 | 44.31 | 47.46 |
| PE03 | C 2-1 | L. Garnier Bayou | 0.23 | 0.23 | 104.47 | 14.14 | 56.13 | 64.3 | 69.55 |
| PE16 | C3-1 | L. Garnier Bayou | 0.23 | 0.23 | 0.19 | 0.19 | 0.17 | 0.19 | 0.19 |
| PE12 | D 1-1 | Destin Harbor | 0.23 | 0.23 | 0.77 | 0.19 | 1.9 | 0.19 | 0.19 |
| PD98 | D 2-2 | Destin Harbor | 0.23 | 0.23 | 5.67 | 3.11 | 15.83 | 7.68 | 0.19 |
|  | D 3-1 | Destin Harbor |  |  |  |  |  |  |  |
| PE13 | E3-1 | Boggy Bayou | 0.23 | 0.23 | 33.66 | 0.19 | 7.25 | 31.27 | 66.93 |
| PE23 | F1-1 | Toms Bayou | 0.23 | 0.23 | 57.33 | 0.19 | 53.84 | 0.19 | 0.19 |
| PE11 | F 2-1 | Tom's Bayou | 0.23 | 0.23 | 87.99 | 14.81 | 49.4 | 76.45 | 96.11 |
| PD91 | G1-1 | Rocky Bayou | 0.23 | 0.23 | 0.19 | 0.19 | 0.17 | 0.19 | 0.19 |
| PD96 | G2-1 | Rocky Bayou | 0.23 | 0.23 | 20.51 | 12.86 | 31.85 | 23.28 | 31.94 |
| PD99 | G3-1 | Rocky Bayou | 0.23 | 0.23 | 0.19 | 0.19 | 0.17 | 0.19 | 0.19 |
| PE06 | K2-1 | La Grange Bayou | 0.23 | 0.23 | 1.59 | 1.53 | 1.26 | 8.37 | 26.21 |
| PE04 | K3-1 | La Grange Bayou | 0.23 | 0.23 | 3.48 | 2.17 | 0.83 | 8.8 | 16.21 |
| PE01 | L1-2 | western basin | 0.23 | 0.23 | 12 | 1.98 | 8.61 | 12.06 | 18.21 |
| PE21 | L3-1 | western basin | 0.23 | 0.23 | 3.58 | 1.1 | 1.3 | 4.99 | 0.19 |
| PE09 | M 1-1 | central basin | 0.23 | 0.23 | 0.66 | 0.19 | 0.38 | 0.19 | 0.19 |
| PE07 | N3-1 | eastern basin | 0.23 | 0.23 | 1.08 | 0.19 | 2.54 | 0.19 | 0.19 |
| PO15PB | NA | Procedural Blank | 0.23 | 0.23 | 0.19 | 0.19 | 0.17 | 0.19 | 0.19 |
| PO19SSRM | $\begin{aligned} & \text { BOS ID } \\ & \# 940504-4 \end{aligned}$ | SRM 1941a | 171.11 | 310.9 | 514.99 | 100.1 | 203.91 | 437.06 | 408.36 |
| MDLs |  |  | 0.23 | 0.23 | 0.19 | 0.19 | 0.17 | 0.19 | 0.19 |


Appendix B2. Chemistry and toxicity data from Choctawhatchee Bay, 1994.









| 0.11 | 0.6 |
| ---: | ---: |
| 1147.36 | 914.28 |

$0.11 \quad 0.6$

 $\stackrel{\sim}{\sim}$ PHEN Site Location
Cinco Bay
Cinco Bayou
U. Garnier Bayou
L. Garnier Bayou
L. Garnier Bayou
L. Garnier Bayou
Destin Harbor
Destin Harbor
Destin Harbor
Boggy Bayou




87.22
$\begin{array}{r}87 \\ 0 \\ 0 \\ 15 \\ 15 \\ \hline\end{array}$
000
No 둥
$\stackrel{\sim}{\oplus}$
202

##  <br> 

$\frac{0}{4} \frac{0}{4}$
$\stackrel{N}{\stackrel{\infty}{4}}$
$\stackrel{\stackrel{\infty}{山}}{\sim}$ $\stackrel{\cong}{\dddot{\sim}}$ Toms Bayou
Tom's Bayou
Rocky Bayou Rocky Bayou La Grange Bayou La Grange Bayou western basin central basin eastern basin
צue|a fe.nnpooold
SRM 1941a


| PO15PB | NA |
| :--- | :--- |
| PO19SSRM | BOS ID |
|  | $\# 940504-4$ |
| MDLs |  |



| $\begin{array}{\|c} \hline \text { Appendix } \\ \text { B2. } \\ \text { Battelle ID } \\ 1994 \end{array}$ | Chemistry and to |  | data from Choctawhatchee Bay, 1994. |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Sta. No. | Site Location | DIBENZ(A,H)ANTHRACENE BENZO(G,H,I)PERYLENE Sum 10LPAH |  |  | Sum 12HPAH | sum 24 PAHs | sum all PAHs | acenaphthene/oc oranthene/phenanthrene/oc ug/goc ug/goc ug/goc |  |  |
| PE02 | A 1-1 | Cinco Bay | 201 | 863 | 734.32 | 14466 | 15200.32 | 18497.7 | 1.08 | 30.66 | 4.74 |
| PE19 | A 2-1 | Cinco Bayou | 0.92 | 4.93 | 2.61 | 62.33 | 64.94 | 75.28 | 0.59 | 41.31 | 0.93 |
| PE22 | B 3-1 | U. Garnier Bayou | 1.49 | 7.14 | 5.64 | 90.42 | 96.06 | 110.52 | 0.17 | 14.78 | 2.63 |
| PE05 | C1-2 | L. Garnier Bayou | 43.11 | 187.18 | 178.4 | 2994.85 | 3173.25 | 3954.98 | 0.68 | 8.24 | 1.28 |
| PE03 | C2-1 | L. Garnier Bayou | 66.87 | 271.74 | 236.18 | 4522.13 | 4758.31 | 5871.08 | 0.87 | 13.04 | 1.96 |
| PE16 | C3-1 | L. Garnier Bayou | 0.7 | 3.34 | 8.31 | 40.28 | 48.6 | 56.89 | 2.94 | 119.06 | 4.63 |
| PE12 | D 1-1 | Destin Harbor | 1.68 | 9.03 | 6.44 | 103.76 | 110.21 | 125.04 | 39.62 | 266.53 | 19.18 |
| PD98 | D 2-2 | Destin Harbor | 9.69 | 49.19 | 37.51 | 533.74 | 571.25 | 655.75 | 4.57 | 16.24 | 2.1 |
|  | D 3-1 | Destin Harbor |  |  |  |  |  |  |  |  |  |
| PE13 | E3-1 | Boggy Bayou | 14.47 | 67.13 | 56.85 | 993.78 | 1050.63 | 1489.94 | 0 | 2.59 | 0.6 |
| PE23 | F 1-1 | Toms Bayou | 47.64 | 211.91 | 190.97 | 3452.14 | 3643.12 | 4080.94 | 0.4 | 4.75 | 0.45 |
| PE11 | F2-1 | Tom's Bayou | 54.68 | 225.06 | 191.99 | 3685.52 | 3877.51 | 5131.35 | 0.51 | 6.88 | 1.14 |
| PD91 | G1-1 | Rocky Bayou | 1.38 | 5.9 | 5.38 | 124.9 | 130.28 | 151.22 | 0 | 0.31 | 0 |
| PD96 | G2-1 | Rocky Bayou | 16.71 | 59.74 | 95.88 | 1024.76 | 1120.64 | 1524.59 | 0.46 | 1.45 | 0.27 |
| PD99 | G3-1 | Rocky Bayou | 0.28 | 0.26 | 2.68 | 2.35 | 5.02 | 9.57 | 0.59 | 1.25 | 0.93 |
| PE06 | K2-1 | La Grange Bayou | 1.41 | 5.6 | 13.9 | 274.29 | 288.19 | 424.55 | 0 | 0.41 | 0.02 |
| PE04 | K3-1 | La Grange Bayou | 0.92 | 3.84 | 13.26 | 109.56 | 122.82 | 252.58 | 0 | 0.73 | 0.14 |
| PE01 | L1-2 | western basin | 7.91 | 36.69 | 40.38 | 560.43 | 600.81 | 740.15 | 0.17 | 1.88 | 0.28 |
| PE21 | L3-1 | western basin | 1.66 | 7.57 | 9.01 | 122.13 | 131.14 | 175.63 | 0.01 | 1.35 | 0.2 |
| PE09 | M 1-1 | central basin | 0.42 | 2.18 | 3.97 | 32.51 | 36.47 | 43.9 | 0.01 | 0.29 | 0.04 |
| PE07 | N3-1 | eastern basin | 2.04 | 7.56 | 10.75 | 168.52 | 179.28 | 213.58 | 0.18 | 1.32 | 0.06 |
| PO15PB | NA | Procedural Blank | 0.28 | 1.63 | 2.68 | 8.74 | 11.42 |  |  |  |  |
| PO19SSRM | BOS ID | SRM 1941a | 110.14 | 457.97 | 2628.13 | 7292.92 | 9921.05 |  |  |  |  |
|  | \#9405 |  |  |  |  |  |  |  |  |  |  |
| MDLs |  |  | 0.28 | 1.63 |  |  |  |  |  |  |  |


| $\begin{aligned} & \stackrel{\mathcal{F}}{\underset{J}{J}} \end{aligned}$ |  |  | －8．80 | $\stackrel{\circ}{\circ}$ |
| :---: | :---: | :---: | :---: | :---: |
| z |  |  | 208 | $\stackrel{\circ}{\circ}$ |
| $\begin{aligned} & \overline{\overline{\mathrm{x}}} \\ & \stackrel{\rightharpoonup}{\text { 人}} \end{aligned}$ | 0000000 | 0000000000 | $\bigcirc 0^{\circ}$ | 。 |
| $$ |  |  | ㅇㅇㅇ | $\stackrel{\circ}{\circ}$ |
|  | $\bigcirc 000^{\circ} 0$ | $00000000^{\circ} 000$ | $0 \cdot 1$ | $\bigcirc$ |
|  |  |  | $\stackrel{\sim}{\circ}$ | $\stackrel{\sim}{\circ}$ |
|  | $\bigcirc 0000000$ | $\bigcirc 00-1$ | $\bigcirc{ }^{\circ}$ | － |
|  |  |  |  |  |
| $\cdots$ |  | $\sim \infty \sim \infty \sim \infty$ |  |  |
| $\stackrel{\sim}{\sim}$ | ○○○ | $\cdots$ | مٌo | $\bigcirc$ |
|  |  |  |  |  |
| ¢ |  |  | $\stackrel{\sim}{\circ} \stackrel{0}{\sim}$ | $\stackrel{\circ}{\circ}$ |
| $\stackrel{-}{\square}$ | 0000000 | 0000000000 | $\bigcirc$ | $\bigcirc$ |
| 0 |  |  |  |  |
|  |  |  |  | $\stackrel{\circ}{\circ}$ |
|  | －0000000 | $00^{\circ} 0000000$ | $\bigcirc$ |  |
|  |  |  | ${ }_{0}^{\circ}$ | $\stackrel{\circ}{\circ}$ |
|  | $0000000$ | 0000000000 | $\bigcirc$ | $\bigcirc$ |
|  |  |  |  |  |
|  |  |  |  |  |
|  |  |  |  |  |
|  |  |  |  |  |
| $\infty$ |  |  | 8） | $\stackrel{\circ}{\circ}$ |
| $\stackrel{\bigcirc}{\sim}$ | 00000000 | 00000000000 | $\bigcirc$ | $\bigcirc$ |
| J |  |  |  |  |
|  |  |  | N |  |
|  | かののハトーじ ゥゅのஸーの人 |  <br>  |  |  |
|  |  |  |  |  |
|  |  |  |  |  |
| © © ® |  |  | $\Sigma$ |  |
|  |  |  | 릏룽 |  |
|  |  |  |  |  |
|  | 可可 |  | ¢ |  |
| $\begin{aligned} & \frac{3}{0} \\ & \stackrel{0}{0} \\ & 0 \\ & 0 \\ & \hline \end{aligned}$ |  | $\overline{-}$ | $\stackrel{+}{\infty}$ |  |
|  |  |  |  |  |
| － | －mN－${ }^{\text {－}}$ |  | $\stackrel{\infty}{\sim}$ |  |
| $E_{0}^{0}{ }_{3}^{3}$ |  |  |  |  |
| 흔 름 |  |  |  |  |
|  |  |  |  |  |
| \ouc |  |  |  |  |
|  |  |  | no |  |
|  |  |  | 5た |  |
| －${ }_{\text {¢ }}$ |  |  |  |  |
|  |  |  |  |  |
|  |  | 僉 | － |  |
|  | ¢ ¢ | mimmmmm |  |  |
|  |  |  |  |  |
|  |  |  | $\bigcirc \stackrel{3}{\circ}$ |  |
|  |  |  |  |  |
|  |  |  | ミ ¢ |  |
| － |  |  | m $\sum_{0}^{5}$ |  |
| － |  <br>  |  <br>  |  | $\stackrel{\text { ® }}{\text { ® }}$ |


| Appendix B2. Battelle ID Sta. No. 1994 |  | Chemistry | Choctawhatchee Bay, 1994. |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Site Location | HEPTACHLOREPOXIDE | CL4(66) | 2,4-DDE | CL5(101) | CIS-CHLORDANE | TRANS-NONACHLOR | DIELDRIN | 4,4-DDE | CL4(77) | 2,4-DDD | ENDRIN | CL5(118) |
| PE02 | A1-1 | Cinco Bay | 0.91 | 0.87 | 0.04 | 1.38 | 4.77 | 5.25 | 5.42 | 14.29 | 0.06 | 4.52 | 0.06 | 1.33 |
| PE19 | A2-1 | Cinco Bayou | 0.03 | 0.06 | 0.04 | 0.07 | 0.06 | 0.07 | 0.22 | 0.15 | 0.06 | 0.04 | 0.06 | 0.04 |
| PE22 | B3-1 | U. Garnier Bayou | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.17 | 0.04 | 0.31 | 0.06 | 0.11 | 0.06 | 0.04 |
| PE05 | C1-2 | L. Garnier Bayou | 0.03 | 0.26 | 0.44 | 0.63 | 0.29 | 0.48 | 1.11 | 11.31 | 0.06 | 1.24 | 0.06 | 0.43 |
| PE03 | C2-1 | L. Garnier Bayou | 0.03 | 0.2 | 0.32 | 0.6 | 0.25 | 0.4 | 0.85 | 11.49 | 0.06 | 1.41 | 0.06 | 0.29 |
| PE16 | C3-1 | L. Garnier Bayou | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.05 | 0.04 | 0.1 | 0.06 | 0.11 | 0.06 | 0.04 |
| PE12 | D1-1 | Destin Harbor | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.05 | 0.04 | 0.04 | 0.06 | 0.11 | 0.06 | 0.04 |
| PD98 | D2-2 | Destin Harbor | 0.03 | 0.09 | 0.04 | 0.07 | 0.09 | 0.05 | 0.04 | 0.3 | 0.06 | 0.11 | 0.06 | 0.11 |
|  | D3-1 | Destin Harbor |  |  |  |  |  |  |  |  |  |  |  |  |
| PE13 | E3-1 | Boggy Bayou | 0.03 | 0.06 | 0.04 | 0.54 | 0.21 | 0.44 | 0.04 | 7.9 | 0.06 | 1.09 | 0.06 | 0.69 |
| PE23 | F1-1 | Toms Bayou | 0.03 | 1.29 | 0.04 | 2.82 | 1.86 | 30.38 | 0.04 | 75.32 | 0.06 | 13.14 | 0.06 | 4.01 |
| PE11 | F2-1 | Tom's Bayou | 0.03 | 0.84 | 0.04 | 1.24 | 0.35 | 0.87 | 1.92 | 17.06 | 0.06 | 2.63 | 0.06 | 1.64 |
| PD91 | G1-1 | Rocky Bayou | 0.03 | 0.06 | 0.04 | 0.09 | 0.25 | 0.05 | 0.04 | 3.05 | 0.06 | 0.11 | 0.06 | 0.04 |
| PD96 | G2-1 | Rocky Bayou | 0.03 | 0.15 | 1 | 0.22 | 0.05 | 0.86 | 0.52 | 4.37 | 0.06 | 0.39 | 0.06 | 0.04 |
| PD99 | G3-1 | Rocky Bayou | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.36 | 0.04 | 0.03 | 0.06 | 0.18 | 0.06 | 0.04 |
| PE06 | K2-1 | La Grange Bayou | 0.03 | 0.06 | 0.04 | 0.06 | 0.05 | 0.05 | 0.04 | 1.21 | 0.06 | 0.21 | 0.06 | 0.29 |
| PE04 | K3-1 | La Grange Bayou | 0.03 | 0.06 | 0.04 | 0.04 | 0.05 | 0.05 | 0.04 | 0.96 | 0.06 | 0.19 | 0.06 | 0.18 |
| PE01 | L1-2 | western basin | 0.03 | 0.07 | 0.19 | 0.12 | 0.1 | 0.05 | 0.04 | 2.21 | 0.06 | 0.32 | 0.06 | 0.11 |
| PE21 | L3-1 | western basin | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.05 | 0.04 | 0.69 | 0.06 | 0.12 | 0.06 | 0.04 |
| PE09 | M 1-1 | central basin | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.05 | 0.04 | 0.45 | 0.06 | 0.11 | 0.06 | 0.04 |
| PE07 | N3-1 | eastern basin | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.05 | 0.04 | 0.82 | 0.06 | 0.12 | 0.06 | 0.1 |
| P015PB | NA | Procedural Blank | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.05 | 0.04 | 0.04 | 0.06 | 0.11 | 0.06 | 0.04 |
| PO19SSRM | $\begin{aligned} & \text { BOS ID } \\ & \# 9405 \end{aligned}$ | $\begin{aligned} & \text { SRM 1941a } \\ & 504-4 \end{aligned}$ | 1.99 | 7.91 | 0.04 | 12.23 | 2.17 | 0.05 | 3.99 | 6.55 | 0.06 | 5.93 | 0.06 | 5.81 |
| MDLs |  |  | 0.03 | 0.06 | 0.04 | 0.07 | 0.05 | 0.05 | 0.04 | 0.07 | 0.06 | 0.11 | 0.06 | 0.04 |





| $\begin{array}{\|c\|} \hline \text { Appendix } \\ \text { Battelle ID } \\ 1994 \end{array}$ | B2. Chemistry and |  | toxicity data fros |  | from Choctawhatchee Bay, 1994. |  |  |  |  | \% clay | \% fines | \%TOC | MSL Code | $\begin{array}{r} \text { AVS } \\ \mu \mathrm{mole} / \mathrm{g} \end{array}$ | $\begin{aligned} & \text { SEM-Cd } \\ & \mu \mathrm{g} / \mathrm{g} \end{aligned}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Sta. No. | Site Location | Se | Si | Sn | Zn | \% gravel | \% sand | \%silt |  |  |  |  |  |  |
|  |  |  | GFAA | XRF | ICP/MS | XRF |  |  |  |  |  |  |  |  |  |
| PE02 | A 1-1 | Cinco Bay | 1.35 | 266000 | 3.1274 | 163.8 | 0 | 47.4 | 40.9 | 11.8 | 52.7 | 7.6 | 779EDL-9 | 37.5135822 | 0.49701219 |
| PE19 | A2-1 | Cinco Bayou | 0.183 | 450000 | 0.1374 | 5.56 | 0 | 98.9 | 0.3 | 0.8 | 1.1 | 0.02 | 779EDL-10 | 0.10403844 | 0.012 |
| PE22 | B 3-1 | U. Garnier Bayou | 0.183 | 425000 | 0.2994 | 8.8 | 0.8 | 96.6 | 1.1 | 1.5 | 2.6 | 0.07 | 779EDL-11 | 0 | 0 |
| PE05 | C1-2 | L. Garnier Bayou | 2.4 | 205000 | 3.5474 | 127.5 | 0 | 6 | 66.1 | 27.9 | 94 | 5.25 | 779EDL-12 | 28.399242 | 0.68847999 |
| PE03 | C2-1 | L. Garnier Bayou | 2.19 | 187000 | 3.6574 | 128.7 | 0.3 | 6.2 | 62.3 | 31.3 | 93.6 | 5.32 | 779EDL-13 | 43.7039586 | 0.46713037 |
| PE16 | C3-1 | L. Garnier Bayou | 0.183 | 451000 | 0.184 | 5.4 | 0 | 98.7 | 0.3 | 0.9 | 1.2 | 0.001 | 779EDL-14 | 0.006 | 0.012 |
| PE12 | D 1-1 | Destin Harbor | 0.183 | 467000 | 0.0934 | 8.6 | 0 | 99 | 0.1 | 0.9 | 1 | 0.004 | $779 \mathrm{EDL-15}$ | 0.15962 | 0.012 |
| PD98 | D2-2 | Destin Harbor | 0.31 | 453000 | 0.9584 | 39 | 0 | 94.6 | 3.4 | 2 | 5.4 | 0.27 | 779EDL-16 | 6.14563721 | 0.14420135 |
|  | D 3-1 | Destin Harbor | 0.42 | 359000 | 1.6074 | 154.5 |  |  |  |  |  |  | 779EDL-17 | 32.8609213 | 0.34146903 |
| PE13 | E3-1 | Boggy Bayou | 2.08 | 208000 | 3.9474 | 107.1 | 1.1 | 6.5 | 66.4 | 26.1 | 92.5 | 5.6 | 779EDL-18 | 56.4498026 | 0.48734703 |
| PE23 | F1-1 | Toms Bayou | 4.89 | 169000 | 5.1674 | 132.3 | 0 | 5.8 | 68.1 | 26.1 | 94.2 | 12.72 | 779EDL-19 | 70.4323521 | 0.79850721 |
| PE11 | F2-1 | Tom's Bayou | 2.71 | 193000 | 4.7874 | 130.8 | 0 | 13.2 | 63.8 | 22.9 | 86.7 | 7.7 | 779EDL-20 | 110 | 0.53734938 |
| PD91 | G1-1 | Rocky Bayou | 1.26 | 337000 | 1.2374 | 26.9 | 0 | 76.5 | 19.4 | 4.1 | 23.5 | 5.2 | 779EDL-21 | 90.2161761 | 0.6679283 |
| PD96 | G2-1 | Rocky Bayou | 2.62 | 173000 | 3.2874 | 84.4 | 0 | 2 | 63.5 | 34.6 | 98.1 | 7.48 | 779EDL-22 | 74.7870387 | 0.47397426 |
| PD99 | G3-1 | Rocky Bayou | 0.183 | 441000 | 0.9424 | 21.8 | 0 | 99.2 | 0.4 | 0.4 | 0.8 | 0.02 | 779EDL-23 | 2.21861879 | 0.012 |
| PE06 | K2-1 | La Grange Bayou | 1.46 | 225000 | 2.4774 | 80.6 | 0 | 7.7 | 55.4 | 36.8 | 92.2 | 6.51 | 779EDL-24 | 10.0052446 | 0.29146387 |
| PE04 | K3-1 | La Grange Bayou | 0.73 | 350000 | 1.3574 | 35.8 | 0.3 | 66.3 | 22.8 | 10.1 | 32.9 | 2.42 | 779EDL-25 | 9.64944881 | 0.08238125 |
| PE01 | L1-2 | western basin | 1.68 | 177000 | 3.1974 | 98.9 | 0 | 19.5 | 52.4 | 28.1 | 80.5 | 4.36 | 779EDL-26 | 6.25617086 | 0.1126173 |
| PE21 | L3-1 | western basin | 0.183 | 413000 | 0.7694 | 25.5 | 0 | 85.9 | 8.8 | 5.2 | 14 | 1.77 | 779EDL-27 | 7.14201249 | 0.04292124 |
| PE09 | M 1-1 | central basin | 0.52 | 297000 | 1.6974 | 54.4 | 0 | 78.2 | 13.2 | 8.5 | 21.7 | 1.62 | 779EDL-29 | 8.68040944 | 0.043529 |
| PE07 | N3-1 | eastern basin | 0.52 | 298000 | 1.7574 | 52.7 | 0 | 68.3 | 18.1 | 13.6 | 31.7 | 1.85 | $779 E D L-28$ | 2.70196833 | 0.041236 |


| Appendix | B2. | Chemistry | and toxicit | ty | Choctaw | whatchee Bay, 1994. |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Battelle ID | Sta. No. | Site Location | SEM-Cd | SEM-Cu | SEM-Cu | SEM-Hg | SEM-Hg |
| 994 |  |  | $\mu \mathrm{mole} / \mathrm{g}$ | $\mu \mathrm{g} / \mathrm{g}$ | $\mu \mathrm{mole} / \mathrm{g}$ | $\mu \mathrm{g} / \mathrm{g}$ | m |
| PE02 | A 1-1 | Cinco Bay | 0.00442182 | 6.7602147 | 0.10639305 | 0.00014408 | 7.18E-07 |
| PE19 | A 2-1 | Cinco Bayou | 0.0001 | 0.27342741 | 0.00430323 | 0.00014408 | 7.18E-07 |
| PE22 | B 3-1 | U. Garnier Bay | 0 | 0 | 0 | 0 | 0 |
| PE05 | C1-2 | L. Garnier Bayı | 0.00612527 | 22.1808466 | 0.34908477 | 0.00321225 | 1.60E-05 |
| PE03 | C2-1 | L. Garnier Bayı | 0.00415596 | 15.3623242 | 0.24177407 | 0.02291044 | 0.00011422 |
| PE16 | C3-1 | L. Garnier Bayı | 0.0001 | 0.28566499 | 0.00449583 | 0.00060825 | 3.03E-06 |
| PE12 | D 1-1 | Destin Harbor | 0.0001 | 1.33342339 | 0.02098557 | 0.00038747 | 1.93E-06 |
| PD98 | D 2-2 | Destin Harbor | 0.00128293 | 12.9608499 | 0.20397938 | 0.00675231 | $3.37 \mathrm{E}-05$ |
|  | D 3-1 | Destin Harbor | 0.00303798 | 23.568283 | 0.37092041 | 0.00129637 | 6.46E-06 |
| PE13 | E3-1 | Boggy Bayou | 0.00433583 | 10.0626899 | 0.1583678 | 0.0029069 | $1.45 \mathrm{E}-05$ |
| PE23 | F1-1 | Toms Bayou | 0.00710416 | 0.65952477 | 0.01037968 | 0.00014408 | 7.18E-07 |
| PE11 | F2-1 | Tom's Bayou | 0.00478069 | 15.9883751 | 0.25162693 | 0.00097312 | $4.85 \mathrm{E}-06$ |
| PD91 | G 1-1 | Rocky Bayou | 0.00594242 | 8.32394994 | 0.1310033 | 0.0005586 | $2.78 \mathrm{E}-06$ |
| PD96 | G2-1 | Rocky Bayou | 0.00421685 | 4.36093175 | 0.06863286 | 0.00289692 | $1.44 \mathrm{E}-05$ |
| PD99 | G3-1 | Rocky Bayou | 0.0001 | 0.01713002 | 0.00026959 | 0.00014408 | 7.18E-07 |
| PE06 | K2-1 | La Grange Bayc | 0.00259309 | 0.72855304 | 0.01146605 | 0.00577489 | $2.88 \mathrm{E}-05$ |
| PE04 | K 3-1 | La Grange Bayc | 0.00073293 | 1.40852437 | 0.02216752 | 0.00014408 | 7.18E-07 |
| PE01 | L1-2 | western basin | 0.00100193 | 7.93231074 | 0.12483964 | 0.01110954 | $5.54 \mathrm{E}-05$ |
| PE21 | L3-1 | western basin | 0.00038186 | 2.5454102 | 0.04005997 | 0.00179503 | 8.95E-06 |
| PE09 | M 1-1 | central basin | 0.00038727 | 3.46287923 | 0.0544992 | 0.00926947 | 4.62E-05 |
| PE07 | N3-1 | eastern basin | 0.00036687 | 1.39625289 | 0.02197439 | 0.00437983 | 2.18E-05 |


| Appendix Battelle ID 1994 | B2. C <br> Sta. No. | hemistry and Site Location | toxicity data from Choctawhatchee Bay, 1994. |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | SEM-Ni | SEM-Ni ole/g | SEM-Pb | SEM-Pb umole/g | SEM-Zn | SEM-Zn | Sum 5SEM umole/g | Sum 5SEM/AVS |  |
| PE02 | A 1-1 | Cinco Bay | 0.55705497 | 0.00948825 | 91.8252399 | 0.4431934 | 97.594929 | 1.49296204 | 2.0565 | 0.05481904 | 2.0016 |
| PE19 | A2-1 | Cinco Bayou | 0.03865132 | 0.00065834 | 1.25701263 | 0.00606696 | 1.0630723 | 0.01626239 | 0.0274 | 0.26327692 | -0.2359 |
| PE22 | B3-1 | U. Garnier Bayou | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| PE05 | C 1-2 | L. Garnier Bayou | 2.89069972 | 0.04923692 | 51.8849622 | 0.25042214 | 69.747804 | 1.06696962 | 1.7218 | 0.06062974 | 1.6612 |
| PE03 | C2-1 | L. Garnier Bayou | 2.65592688 | 0.04523807 | 49.381244 | 0.23833797 | 31.860996 | 0.48739477 | 1.0169 | 0.02326793 | 0.9936 |
| PE16 | C3-1 | L. Garnier Bayou | 0.05748627 | 0.00097916 | 0.52840752 | 0.00255035 | 0.7568896 | 0.01157855 | 0.0197 | 3.28398074 | -3.2643 |
| PE12 | D 1-1 | Destin Harbor | 0.08299636 | 0.00141367 | 0.67603813 | 0.00326289 | 2.6383431 | 0.04036015 | 0.0661 | 0.41424811 | -0.3481 |
| PD98 | D2-2 | Destin Harbor | 0.29053794 | 0.0049487 | 7.9644046 | 0.0384401 | 37.186701 | 0.56886494 | 0.8175 | 0.13302381 | 0.6845 |
|  | D3-1 | Destin Harbor | 0.90552751 | 0.01542374 | 28.4784263 | 0.13745078 | 140 | 2.0704889 | 2.5973 | 0.07903984 | 2.5183 |
| PE13 | E3-1 | Boggy Bayou | 1.71444962 | 0.029202 | 34.1830743 | 0.16498419 | 45.127547 | 0.69034032 | 1.0472 | 0.01855153 | 1.0287 |
| PE23 | F1-1 | Toms Bayou | 0.34096841 | 0.00580767 | 24.1689982 | 0.11665137 | 57.04505 | 0.87264876 | 1.0126 | 0.0143768 | 0.9982 |
| PE11 | F2-1 | Tom's Bayou | 1.05286301 | 0.01793328 | 63.5924036 | 0.30692796 | 53.989758 | 0.82591032 | 1.4072 | 0.01279254 | 1.3944 |
| PD91 | G1-1 | Rocky Bayou | 0.69691571 | 0.01187048 | 43.8807009 | 0.21178967 | 55.517404 | 0.84927954 | 1.20988541 | 0.01358467 | 1.19630074 |
| PD96 | G2-1 | Rocky Bayou | 1.19930967 | 0.02042769 | 19.3523764 | 0.09340401 | 27.298456 | 0.41759915 | 0.6043 | 0.00808002 | 0.5962 |
| PD99 | G3-1 | Rocky Bayou | 0.01543302 | 0.00026287 | 0.7012136 | 0.0033844 | 0.4451153 | 0.00680917 | 0.0108 | 0.00487963 | 0.0059 |
| PE06 | K2-1 | La Grange Bayou | 0.42007911 | 0.00715515 | 4.74463301 | 0.02289991 | 29.473104 | 0.4508659 | 0.495 | 0.04947207 | 0.4455 |
| PE04 | K3-1 | La Grange Bayou | 0.8251356 | 0.01405443 | 4.6820474 | 0.02259784 | 12.896908 | 0.19729092 | 0.2568 | 0.02661744 | 0.2302 |
| PE01 | L1-2 | western basin | 3.95634341 | 0.0673879 | 22.7338762 | 0.10972478 | 35.323863 | 0.54036811 | 0.8433 | 0.13479849 | 0.7085 |
| PE21 | L3-1 | western basin | 36.3825829 | 0.61969993 | 7.21579394 | 0.03482694 | 8.8970771 | 0.13610337 | 0.8311 | 0.11636385 | 0.7147 |
| PE09 | M 1-1 | central basin | 1.61126322 | 0.02744444 | 10.269278 | 0.04956454 | 17.89222 | 0.2737069 | 0.4056 | 0.04672618 | 0.3589 |
| PE07 | N3-1 | eastern basin | 1.60178633 | 0.02728302 | 8.07653284 | 0.03898129 | 18.981566 | 0.29037121 | 0.379 | 0.14025952 | 0.2387 |















$\stackrel{\circ}{0}$




| Appendix | B3. | emistry | ci | data f | rom St. Andrew | Bay. |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| MSL Code | Station Number | Signif. | \% urchdevp@100\% | Signif. | \% urchdevp@50\% | signif. | \%urchdevp@25\% | signif. | Sample I.D. | $\begin{gathered} \mathrm{Ag}, \mathrm{ug} / \mathrm{g} \\ \text { GFAA } \end{gathered}$ | $\begin{gathered} \mathrm{Al}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ | $\begin{gathered} \text { As,ug/g } \\ \text { XRF } \\ \hline \end{gathered}$ | $\begin{gathered} \hline \mathrm{Cd}, \mathrm{ug} / \mathrm{g} \\ \text { GFAA } \end{gathered}$ | $\begin{gathered} \mathrm{Cr}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ | $\begin{gathered} \mathrm{Cu}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ | $\begin{gathered} \hline \mathrm{Fe}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ |
|  | controls |  | 96.6 | ns | 86.4 | ns | 86.0 | ns |  |  |  |  |  |  |  |  |
| 782-TOX 1 | 41 | ns | 97.8 | ns | 87.4 | ns | 90.0 | ns | 00-22 | 0.008 | 12000 | 1.3 | 0.017 | 12 | 4.7 | 2400 |
| 782-TOX 2 | 42 | ns | 99.2 | ns | 90.0 | ns | 85.0 | ns | 00-23 |  |  |  |  |  |  |  |
| 782-TOX 3 | 43 | ns | 96.6 | ns | 88.6 | ns | 91.6 | ns | 00-28 |  |  |  |  |  |  |  |
| 782-TOX 4 | 44 | ns | 97.6 | ns | 98.2 | ns | 96.0 | ns | 00-27 |  |  |  |  |  |  |  |
| 782-TOX 5 | 45 | ns | 99.2 | ns | 98.4 | ns | 98.6 | ns | 00-26 | 0.11 | 59000 | 13 | 0.22 | 97 | 19 | 32000 |
|  | 46 | ns | 98.8 | ns | 99.2 | ns | 98.6 | ns |  |  |  |  |  |  |  |  |
|  | 47 | ns | 99.2 | ns | 98.6 | ns | 99.0 | ns |  |  |  |  |  |  |  |  |
| 782-TOX 6 | 48 | ns | 97.6 | ns | 98.8 | ns | 98.8 | ns | 00-30 | 0.011 | 12000 | 2.1 | 0.029 | 10 | 4.7 | 4400 |
| 782-TOX 7 | 49 | ns | 98.2 | ns | 99.2 | ns | 98.2 | ns | 00-31 | 0.030 | 19000 | 4.4 | 0.041 | 21 | 6.5 | 6500 |
| 782-TOX 8 | 50 | ns | 6.4 | ** | 98.4 | ns | 99.0 | ns | 00-32 | 0.11 | 44000 | 20 | 0.21 | 61 | 20 | 23000 |
| 782-TOX 9 | 51 | ns | 99.2 | ns | 99.0 | ns | 98.2 | ns | 00-02 | 0.019 | 12000 | 1.4 | 0.029 | 10 | 2.5 | 2500 |
|  | 52 | ns | 97.4 | ns | 98.6 | ns | 97.2 | ns |  |  |  |  |  |  |  |  |
| 782-TOX 10 | 53 | ns | 97.8 | ns | 98.4 | ns | 98.0 | ns | 00-16 | 1.6 | 48000 | 30 | 1.7 | 84 | 350 | 30000 |
| 782-TOX 11 | 54 | ns | 99.6 | ns | 98.2 | ns | 98.8 | ns | 00-17 | 0.59 | 21000 | 13 | 0.67 | 33 | 150 | 12000 |
| 782-TOX 12 | 55 | ns | 0.0 | ** | 98.0 | ns | 98.6 | ns | 00-03 | 2.1 | 59000 | 16 | 1.0 | 74 | 63 | 27000 |
| 782-TOX 13 | 56 | ns | 0.2 | ** | 0.0 | ** | 98.4 | ns | 00-04 | 1.9 | 51000 | 20 | 1.1 | 69 | 110 | 27000 |
| 782-TOX 14 | 57 | ns | 0.0 | ** | 99.2 | ns | 99.0 | ns | 00-05 | 2.5 | 60000 | 20 | 1.12 | 77 | 96 | 30000 |
| 782-TOX 15 | 58 | ns | 0.2 | ** | 0.0 | ** | 98.2 | ns | 00-06 | 1.9 | 67000 | 24 | 1.24 | 100 | 140 | 32000 |
| 782-TOX 16 | 59 | ns | 0.0 | ** | 98.8 | ns | 98.2 | ns | 00-07 | 0.33 | 23000 | 5.7 | 0.26 | 46 | 53 | 8900 |
| 782-TOX 17 | 60 | ns | 74.2 | ns | 97.2 | ns | 99.2 | ns | 00-08 | 1.0 | 38000 | 12 | 0.87 | 51 | 160 | 15000 |
| 782-TOX 18 | 61 | ns | 98.8 | ns | 98.8 | ns | 98.2 | ns | 00-09 ave | 0.645 | 25000 | 8.2 | 0.74 | 35.15 | 31.95 | 9700 |
| 782-TOX 19 | 62 | ns | 98.2 | ns | 99.4 | ns | 98.8 | ns | 00-10 | 0.060 | 10000 | 1.3 | 0.081 | 9.0 | 5.7 | 2100 |
| 782-TOX 20 | 63 | ns | 99.4 | ns | 98.8 | ns | 98.6 | ns | 00-11 | 0.96 | 39000 | 15 | 0.90 | 50 | 53 | 16000 |
|  | 64 | ns | 97.0 | ns | 98.6 | ns | 98.4 | ns | 00-14 | 0.045 | 19000 | 5.6 | 0.080 | 21 | 5.0 | 6600 |
| 782-TOX 21 | 65 | ns | 99.4 | ns | 98.2 | ns | 98.2 | ns | 00-13 | 0.16 | 42000 | 10 | 0.20 | 62 | 15 | 17000 |
| 782-TOX 22 | 66 | ns | 36.0 | ** | 99.0 | ns | 97.6 | ns | 00-12 | 0.55 | 63000 | 20 | 0.52 | 5.0 | 35 | 27000 |
|  | 67 | ns | 96.0 | ns | 96.6 | ns | 94.8 | ns |  |  |  |  |  |  |  |  |
|  | 68 | ns | 88.0 | ns | 99.2 | ns | 96.8 | ns |  |  |  |  |  |  |  |  |
|  | 69 | ns | 98.4 | ns | 98.4 | ns | 97.6 | ns |  |  |  |  |  |  |  |  |
| MDL | 70 | ns | 97.8 | ns | 95.0 | ns | 95.4 | ns | 00-18 | 0.11 | 50000 | 11 | 0.22 | 56 | 12 | 25000 |
|  | 71 | ns | 86.0 | ns | 86.4 | ns | 97.6 | ns | 00-19 | 0.007 | 9900 | 1.3 | 0.017 | 8.7 | 2.1 | 3100 |
| 782-TOX 15 |  |  |  |  |  |  |  |  |  | 0.01 | 5400.00 | 1.30 | 0.01 | 11.0 | 2.30 | 20.00 |
| 782-TOX 15 |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
|  | 61 |  |  |  |  |  |  |  | 00-09 R1 | 0.86 | 26000 | 9.2 | 0.89 | 36 | 32 | 10000 |
|  | 61R |  |  |  |  |  |  |  | 00-09 R2 | 0.43 | 24000 | 7.2 | 0.59 | 34 | 32 | 9400 |
|  | AVE. |  |  |  |  |  |  |  | ave | 0.65 | 25000.00 | 8.20 | 0.74 | 35.15 | 31.95 | 9700.00 |


| Appendix | B3. Chemistry |  | and toxicity data from St. Andrew Bay. |  |  |  |  |  | $\begin{aligned} & \hline \text { Sn,ug/g } \\ & \text { ICP/MS } \end{aligned}$ | $\begin{gathered} \hline \mathrm{Zn}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ | MSL Code | Sponsor ID | AVS umole/g |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Station <br> Number | $\begin{gathered} \mathrm{Hg}, \mathrm{ug} / \mathrm{g} \\ \text { CVAA } \\ \hline \end{gathered}$ | $\begin{gathered} \mathrm{Mn}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ | $\begin{gathered} \hline \mathrm{Ni}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ | $\begin{gathered} \mathrm{Pb}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ | Sb,ug/g <br> ICP/MS | Se,ug/g GFAA |  | $\begin{gathered} \mathrm{Si}, \mathrm{ug} / \mathrm{g} \\ \text { XRF } \\ \hline \end{gathered}$ |  |  |  |  |  |
| controls |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 41 | 0.012 | 31 | 3.5 | 2.6 | 0.071 | 0.15 | U | 440000 | 0.35 | 6.3 | 782TOX*1 | 00-22 | 0.146339474 |
| 42 l |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 43 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 44 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 45 | 0.12 | 180 | 27 | 36 | 0.51 | 1.3 |  | 230000 | 2.7 | 82 | 782TOX*2 | 00-26 | 3.143570281 |
| 46 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 47 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 48 | 0.020 | 66 | 4.9 | 7.3 | 0.15 | 0.15 |  | 400000 | 0.51 | 15 | 782TOX*3 | 00-30 | 0.13576619 |
| 49 | 0.033 | 79 | 6.9 | 7.4 | 0.21 | 0.17 |  | 360000 | 0.81 | 18 | 782TOX*4 | 00-31 | 3.005709829 |
| 50 | 0.10 | 280 | 21 | 25 | 0.40 | 0.93 |  | 220000 | 2.1 | 63 | 782TOX*5 | 00-32 | 1.506374985 |
| 51 | 0.020 | 50 | 4.5 | 6.8 | 0.13 | 0.15 | U | 420000 | 0.53 | 12 | 782TOX*6 ave | 00-02R ave | 0.948582904 |
| 52 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 53 | 1.3 | 130 | 25 | 320 | 1.1 | 1.3 |  | 210000 | 19 | 480 | 782TOX*7 | 00-16 | 29.74485158 |
| 54 | 0.51 | 55 | 9.2 | 96 | 0.54 | 0.33 |  | 350000 | 7.6 | 180 | 782TOX*8 | 00-17 | 3.912578603 |
| 55 | 0.33 | 120 | 21 | 84 | 0.81 | 1.5 |  | 280000 | 6.9 | 240 | 782TOX*9 | 00-03 | 3.235691091 |
| 56 | 0.41 | 120 | 19 | 94 | 0.80 | 1.4 |  | 260000 | 7.6 | 310 | 782TOX*10 | 00-4 | 11.03398336 |
| 57 | 0.45 | 130 | 21 | 110 | 1.0 | 1.5 |  | 260000 | 8.8 | 310 | 782TOX*11 | 00-5 | 7.407787974 |
| 58 | 0.51 | 150 | 27 | 110 | 1.0 | 1.9 |  | 220000 | 8.7 | 370 | 782TOX*12 | 00-6 | 21.82112956 |
| 59 | 0.088 | 61 | 11 | 33 | 0.22 | 0.15 | U | 410000 | 1.4 | 190 | 782TOX*13 | 00-7 | 2.809585862 |
| 60 | 0.49 | 68 | 12 | 120 | 0.72 | 0.75 |  | 340000 | 6.1 | 230 | 782TOX*14 | 00-8 | 8.045293711 |
| 61 | 0.216 | 60.8 | 7.45 | 37.25 | 0.41 | 0.555 | \#DIV/0! | 350000 | 3.04465 | 115 | 782TOX*15 | 00-9 | 6.229896057 |
| 62 | 0.044 | 20.8 | 2.2 | 7.0 | 0.10 | 0.15 |  | 390000 | 0.39 | 21 | 782TOX*16 | 00-10 | 0.552155522 |
| 63 | 0.54 | 110 | 14 | 53 | 0.77 | 1.0 |  | 290000 | 4.7 | 170 | 782TOX*17 | 00-11 | 2.554487182 |
| 64 | 0.047 | 83 | 5.7 | 6.6 | 0.22 | 0.17 |  | 360000 | 0.79 | 28 | 782TOX*18 | 00-14 | 1.677563213 |
| 65 | 0.11 | 260 | 15 | 22 | 0.41 | 0.68 |  | 290000 | 2.1 | 53 | 782TOX*19 | 00-13 | 2.580121115 |
| 66 | 0.24 | 400 | 24 | 48 | 0.74 | 1.3 |  | 200000 | 3.9 | 130 | 782TOX*20 | 00-12 | 6.786758358 |
| 67 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 68 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 69 |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 70 | 0.17 | 210 | 15 | 25 | 0.38 | 0.76 |  | 280000 | 2.1 | 23 | 782TOX*21 | 00-18 | 4.8105311 |
| 71 | 0.0070 | 37 | 2.2 | 2.6 | 0.40 | 0.15 |  | 410000 | 0.15 | 6.9 | 782TOX*22 | 00-19 | 0.578764363 |
|  | 0.003 | 10 | 2.7 | 2.9 | 0.021 | 0.15 |  | 2000 | 0.024 | 3 | MDL |  | 0.11 |
| 61 | 0.28 | 59 | 9.3 | 36 | 0.47 | 0.67 |  | 350000 | 3.7 | 120 | 782TOX*6 R1 | 00-02R | 0.999079825 |
| 61R | 0.15 | 62 | 5.6 | 38 | 0.35 | 0.44 |  | 350000 | 2.4 | 110 | 782TOX*6 R2 | 00-02R | 0.898085983 |
| AVE. | 0.22 | 60.80 | 7.45 | 37.25 | 0.41 | 0.56 | \#DIV/0! | 350000 | 3.04 | 115 | ave. | ave | 0.948582904 |


| Appendix | 3. Chemistry | and toxicity | ta from St. | Andrew Bay. |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Station Number | SEM-Cd $\mu \mathrm{g} / \mathrm{g}$ | SEM-Cd umole/g | SEM-Cu $\mu \mathrm{g} / \mathrm{g}$ | SEM-Cu umole/g | $\begin{gathered} \text { SEM-Hg } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ | SEM-Hg umole/g | SEM-Ni <br> $\mu \mathrm{g} / \mathrm{g}$ | SEM-Ni umole/g | $\begin{gathered} \text { SEM-Pb } \\ \mu \mathrm{g} / \mathrm{g} \end{gathered}$ | SEM-Pb umole/g | $\begin{gathered} \text { SEM-Zn } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ |
| controls |  |  |  |  |  |  |  |  |  |  |  |
| 41 | 0.005544442 | $4.93278 \mathrm{E}-05$ | 0.51756852 | 0.008145554 | 0.001467995 | 7.32E-06 | 0.180769248 | 0.00307902 | 1.523804867 | 0.007354626 | 2.12501271 |
| 42 |  |  |  |  |  |  |  |  |  |  |  |
| 43 |  |  |  |  |  |  |  |  |  |  |  |
| 44 |  |  |  |  |  |  |  |  |  |  |  |
| 45 | 0.100704406 | 0.000895947 | 4.478080864 | 0.070476564 | 0.00014 | 0.00000067 | 1.677854729 | 0.028578687 | 20.6050889 | 0.099450209 | 41.65141471 |
| 46 |  |  |  |  |  |  |  |  |  |  |  |
| 47 |  |  |  |  |  |  |  |  |  |  |  |
| 48 | 0.017323255 | 0.000154121 | 1.334318243 | 0.020999658 | 0.0006571 | $3.28 \mathrm{E}-06$ | 0.663778468 | 0.011306055 | 4.272221475 | 0.020619825 | 5.957743706 |
| 49 | 0.059129596 | 0.000526064 | 3.615334141 | 0.056898554 | 0.002225821 | $1.10964 \mathrm{E}-05$ | 2.424249326 | 0.041291932 | 5.995708294 | 0.028938213 | 9.619752974 |
| 50 | 0.180048321 | 0.001601853 | 9.727843731 | 0.15309795 | 0.00536523 | 2.67472E-05 | 6.780830876 | 0.115497034 | 17.4711912 | 0.084324491 | 25.58714455 |
| 51 | 0.021095014 | 0.000187678 | 1.782877891 | 0.028059142 | 0.001076738 | 5.36786E-06 | 0.621313721 | 0.010582758 | 4.213531683 | 0.020336559 | 6.402610261 |
| 52 |  |  |  |  |  |  |  |  |  |  |  |
| 53 | 1.139211713 | 0.010135336 | 200 | 3.11167847 | 0.00607007 | $3.02611 \mathrm{E}-05$ | 19.59255 | 0.333717425 | 260 | 1.249224814 | 310 |
| 54 | 0.380736736 | 0.003387338 | 86.72478164 | 1.364884823 | 0.007029311 | $3.50432 \mathrm{E}-05$ | 1.520376106 | 0.025896374 | 84.34747054 | 0.407102035 | 140 |
| 55 | 0.693404616 | 0.00616908 | 25.92012133 | 0.407933921 | 0.001402791 | $6.99 \mathrm{E}-06$ | 1.775115148 | 0.030235312 | 37.82360481 | 0.182555166 | 170 |
| 56 | 0.915217353 | 0.008142503 | 61.79456705 | 0.972530171 | 0.002416913 | 1.2049E-05 | 1.825027001 | 0.031085454 | 58.33260555 | 0.281541607 | 220 |
| 57 | 0.830684685 | 0.007390433 | 54.53988612 | 0.858355148 | 0.0038 | 0.000019 | 1.859900532 | 0.03167945 | 57.07263944 | 0.275460396 | 230 |
| 58 | 1.008452948 | 0.008972001 | 79.95739145 | 1.258378839 | 0.003350289 | 1.67022E-05 | 3.180955838 | 0.054180818 | 66.70447769 | 0.321948345 | 270 |
| 59 | 0.216660052 | 0.001927581 | 12.12984135 | 0.190900871 | 0.000549948 | $2.74 \mathrm{E}-06$ | 0.814348695 | 0.013870698 | 12.09323497 | 0.058367851 | 62.76139394 |
| 60 | 0.461590567 | 0.004106678 | 56.62023105 | 0.891095862 | 0.002974091 | $1.48267 \mathrm{E}-05$ | 1.040815575 | 0.01772808 | 46.62206788 | 0.22502084 | 150 |
| 61 | 0.370262717 | 0.003294152 | 22.849469287 | 0.359607638 | 0.002946830 | 0.000014691 | 1.352495750 | 0.023036889 | 26.347155312 | 0.127164223 | 89.439119842 |
| 62 | 0.069322607 | 0.000616749 | 3.640985957 | 0.057302266 | 0.000771669 | $3.84699 \mathrm{E}-06$ | 0.215979823 | 0.003678757 | 4.312861828 | 0.020815975 | 13.32715389 |
| 63 | 0.636198441 | 0.005660128 | 28.75974433 | 0.452624242 | 0.001979565 | $9.86871 \mathrm{E}-06$ | 1.760455816 | 0.029985621 | 30.06848407 | 0.14512517 | 110 |
| 64 | 0.039077686 | 0.000347666 | 2.720289264 | 0.042812233 | 0.001422786 | 7.09E-06 | 1.157688318 | 0.019718759 | 5.393158092 | 0.026030012 | 8.729211192 |
| 65 | 0.134866316 | 0.001199878 | 7.882882577 | 0.124061734 | 0.006666654 | 3.32352E-05 | 2.442759164 | 0.041607208 | 14.90593867 | 0.071943331 | 26.98685887 |
| 66 | 0.389751444 | 0.00346754 | 21.37332521 | 0.336375908 | 0.004304257 | $2.1458 \mathrm{E}-05$ | 4.128769221 | 0.070324804 | 31.27479928 | 0.150947436 | 68.08732757 |
| 67 |  |  |  |  |  |  |  |  |  |  |  |
| 68 |  |  |  |  |  |  |  |  |  |  |  |
| 69 |  |  |  |  |  |  |  |  |  |  |  |
| 70 | 0.182484394 | 0.001623527 | 6.51507916 | 0.102535083 | 0.004417741 | 2.20237E-05 | 1.926846806 | 0.032819738 | 17.66578579 | 0.085263699 | 24.59042977 |
| 71 | 0.010561669 | $9.3965 \mathrm{E}-05$ | 0.441739295 | 0.006952145 | 0.000664147 | $3.31 \mathrm{E}-06$ | 0.229411779 | 0.003907542 | 1.533419407 | 0.00740103 | 1.957825396 |
|  | 0.011 | 0.000097 | 0.016 | 0.00025 | 0.00014 | 0.00000067 | 0.011 | 0.000097 | 0.0033 | 0.000016 | 0.006480962 |
| 61 | 0.022877142 | 0.000203533 | 1.677659409 | 0.026403201 | 0.000629417 | $3.14 \mathrm{E}-06$ | 0.682460702 | 0.011624267 | 4.224833735 | 0.020391108 | 6.536385475 |
| 61R | 0.019312886 | 0.000171823 | 1.888096374 | 0.029715083 | 0.00152406 | 7.60E-06 | 0.56016674 | 0.009541249 | 4.20222963 | 0.02028201 | 6.268835046 |
| AVE. | 0.021095014 | 0.000187678 | 1.782877891 | 0.028059142 | 0.001076738 | $5.36786 \mathrm{E}-06$ | 0.621313721 | 0.010582758 | 4.213531683 | 0.020336559 | 6.402610261 |


| Appendix | 3. Chemistry | and toxicity | ta from St. | Andrew Bay. |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Station <br> Number | $\begin{gathered} \hline \text { SEM-Cd } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ | SEM-Cd umole/g | $\begin{gathered} \hline \text { SEM-Cu } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ | SEM-Cu $\mu \mathrm{mole} / \mathrm{g}$ | $\begin{gathered} \hline \text { SEM-Hg } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ | SEM-Hg umole/g | $\begin{gathered} \hline \text { SEM-Ni } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ | $\begin{array}{r} \text { SEM-Ni } \\ \mu \mathrm{mole} / \mathrm{g} \\ \hline \end{array}$ | $\begin{gathered} \text { SEM-Pb } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ | SEM-Pb umole/g | $\begin{gathered} \text { SEM-Zn } \\ \mu \mathrm{g} / \mathrm{g} \\ \hline \end{gathered}$ |
| controls |  |  |  |  |  |  |  |  |  |  |  |
| 41 | 0.005544442 | $4.93278 \mathrm{E}-05$ | 0.51756852 | 0.008145554 | 0.001467995 | 7.32E-06 | 0.180769248 | 0.00307902 | 1.523804867 | 0.007354626 | 2.12501271 |
| 42 |  |  |  |  |  |  |  |  |  |  |  |
| 43 |  |  |  |  |  |  |  |  |  |  |  |
| 44 |  |  |  |  |  |  |  |  |  |  |  |
| 45 | 0.100704406 | 0.000895947 | 4.478080864 | 0.070476564 | 0.00014 | 0.00000067 | 1.677854729 | 0.028578687 | 20.6050889 | 0.099450209 | 41.65141471 |
| 46 |  |  |  |  |  |  |  |  |  |  |  |
| 47 |  |  |  |  |  |  |  |  |  |  |  |
| 48 | 0.017323255 | 0.000154121 | 1.334318243 | 0.020999658 | 0.0006571 | 3.28E-06 | 0.663778468 | 0.011306055 | 4.272221475 | 0.020619825 | 5.957743706 |
| 49 | 0.059129596 | 0.000526064 | 3.615334141 | 0.056898554 | 0.002225821 | $1.10964 \mathrm{E}-05$ | 2.424249326 | 0.041291932 | 5.995708294 | 0.028938213 | 9.619752974 |
| 50 | 0.180048321 | 0.001601853 | 9.727843731 | 0.15309795 | 0.00536523 | 2.67472E-05 | 6.780830876 | 0.115497034 | 17.4711912 | 0.084324491 | 25.58714455 |
| 51 | 0.021095014 | 0.000187678 | 1.782877891 | 0.028059142 | 0.001076738 | 5.36786E-06 | 0.621313721 | 0.010582758 | 4.213531683 | 0.020336559 | 6.402610261 |
| 52 |  |  |  |  |  |  |  |  |  |  |  |
| 53 | 1.139211713 | 0.010135336 | 200 | 3.11167847 | 0.00607007 | $3.02611 \mathrm{E}-05$ | 19.59255 | 0.333717425 | 260 | 1.249224814 | 310 |
| 54 | 0.380736736 | 0.003387338 | 86.72478164 | 1.364884823 | 0.007029311 | $3.50432 \mathrm{E}-05$ | 1.520376106 | 0.025896374 | 84.34747054 | 0.407102035 | 140 |
| 55 | 0.693404616 | 0.00616908 | 25.92012133 | 0.407933921 | 0.001402791 | 6.99E-06 | 1.775115148 | 0.030235312 | 37.82360481 | 0.182555166 | 170 |
| 56 | 0.915217353 | 0.008142503 | 61.79456705 | 0.972530171 | 0.002416913 | 1.2049E-05 | 1.825027001 | 0.031085454 | 58.33260555 | 0.281541607 | 220 |
| 57 | 0.830684685 | 0.007390433 | 54.53988612 | 0.858355148 | 0.0038 | 0.000019 | 1.859900532 | 0.03167945 | 57.07263944 | 0.275460396 | 230 |
| 58 | 1.008452948 | 0.008972001 | 79.95739145 | 1.258378839 | 0.003350289 | 1.67022E-05 | 3.180955838 | 0.054180818 | 66.70447769 | 0.321948345 | 270 |
| 59 | 0.216660052 | 0.001927581 | 12.12984135 | 0.190900871 | 0.000549948 | $2.74 \mathrm{E}-06$ | 0.814348695 | 0.013870698 | 12.09323497 | 0.058367851 | 62.76139394 |
| 60 | 0.461590567 | 0.004106678 | 56.62023105 | 0.891095862 | 0.002974091 | $1.48267 \mathrm{E}-05$ | 1.040815575 | 0.01772808 | 46.62206788 | 0.22502084 | 150 |
| 61 | 0.370262717 | 0.003294152 | 22.849469287 | 0.359607638 | 0.002946830 | 0.000014691 | 1.352495750 | 0.023036889 | 26.347155312 | 0.127164223 | 89.439119842 |
| 62 | 0.069322607 | 0.000616749 | 3.640985957 | 0.057302266 | 0.000771669 | $3.84699 \mathrm{E}-06$ | 0.215979823 | 0.003678757 | 4.312861828 | 0.020815975 | 13.32715389 |
| 63 | 0.636198441 | 0.005660128 | 28.75974433 | 0.452624242 | 0.001979565 | $9.86871 \mathrm{E}-06$ | 1.760455816 | 0.029985621 | 30.06848407 | 0.14512517 | 110 |
| 64 | 0.039077686 | 0.000347666 | 2.720289264 | 0.042812233 | 0.001422786 | 7.09E-06 | 1.157688318 | 0.019718759 | 5.393158092 | 0.026030012 | 8.729211192 |
| 65 | 0.134866316 | 0.001199878 | 7.882882577 | 0.124061734 | 0.006666654 | $3.32352 \mathrm{E}-05$ | 2.442759164 | 0.041607208 | 14.90593867 | 0.071943331 | 26.98685887 |
| 66 | 0.389751444 | 0.00346754 | 21.37332521 | 0.336375908 | 0.004304257 | $2.1458 \mathrm{E}-05$ | 4.128769221 | 0.070324804 | 31.27479928 | 0.150947436 | 68.08732757 |
| 67 |  |  |  |  |  |  |  |  |  |  |  |
| 68 |  |  |  |  |  |  |  |  |  |  |  |
| 69 |  |  |  |  |  |  |  |  |  |  |  |
| 70 | 0.182484394 | 0.001623527 | 6.51507916 | 0.102535083 | 0.004417741 | 2.20237E-05 | 1.926846806 | 0.032819738 | 17.66578579 | 0.085263699 | 24.59042977 |
| 71 | 0.010561669 | $9.3965 \mathrm{E}-05$ | 0.441739295 | 0.006952145 | 0.000664147 | $3.31 \mathrm{E}-06$ | 0.229411779 | 0.003907542 | 1.533419407 | 0.00740103 | 1.957825396 |
|  | 0.011 | 0.000097 | 0.016 | 0.00025 | 0.00014 | 0.00000067 | 0.011 | 0.000097 | 0.0033 | 0.000016 | 0.006480962 |
| 61 | 0.022877142 | 0.000203533 | 1.677659409 | 0.026403201 | 0.000629417 | 3.14E-06 | 0.682460702 | 0.011624267 | 4.224833735 | 0.020391108 | 6.536385475 |
| 61R | 0.019312886 | 0.000171823 | 1.888096374 | 0.029715083 | 0.00152406 | 7.60E-06 | 0.56016674 | 0.009541249 | 4.20222963 | 0.02028201 | 6.268835046 |
| AVE. | 0.021095014 | 0.000187678 | 1.782877891 | 0.028059142 | 0.001076738 | $5.36786 \mathrm{E}-06$ | 0.621313721 | 0.010582758 | 4.213531683 | 0.020336559 | 6.402610261 |





Appendix B3. Chemistry and toxicity data from St. Andrew Bay.


| Appendix B3. Chemistry and toxicity data from St. Andrew Bay. |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Station Number | C4-CHRYSENES $\mathrm{ng} / \mathrm{g}$ dry wt. | BENZO(B)FLUORANTHENE $\mathrm{ng} / \mathrm{g}$ dry wt. | BENZO(K)FLUORANTHENE | BENZO(E)PYRENE $\mathrm{ng} / \mathrm{d}$ dry wt . | BENZO(A)PYRENE $\mathrm{ng} / \mathrm{g}$ dry wt. | PERYLENE ng/g dry wt. | J(1,2,3-C,D)P ng/g dry wt. | DIBENZ(A,H)ANTHRACENE $\mathrm{ng} / \mathrm{g}$ dry wt. | BENZO(G,H,I)PERYLENE <br> $\mathrm{ng} / \mathrm{g}$ dry wt. |
| controls |  |  |  |  |  |  |  |  |  |
| 41 | 0.137 | 2.888 | 0.849 | 1.352 | 1.245 | 5.198 | 1.572 | 0.325 | 1.622 |
| 42 | 17.362 | 45.379 | 14.830 | 23.402 | 19.677 | 20.463 | 20.971 | 5.780 | 22.328 |
| 43 | 35.633 | 154.370 | 58.013 | 75.562 | 79.010 | 32.386 | 72.754 | 20.269 | 62.381 |
| 44 | 17.863 | 58.331 | 18.959 | 29.384 | 25.998 | 17.633 | 25.064 | 6.905 | 25.548 |
| 45 | 0.137 | 48.376 | 16.113 | 24.846 | 21.122 | 24.485 | 21.853 | 5.535 | 23.254 |
| 46 | 32.977 | 86.590 | 27.281 | 41.769 | 23.349 | 39.676 | 35.184 | 8.953 | 31.444 |
| 47 | 14.685 | 49.615 | 16.710 | 23.685 | 20.731 | 16.233 | 19.997 | 5.089 | 19.364 |
| 48 | 7.998 | 28.195 | 9.843 | 13.181 | 13.002 | 7.081 | 12.219 | 3.066 | 11.765 |
| 49 | 14.737 | 41.498 | 14.604 | 19.675 | 19.214 | 10.371 | 17.828 | 4.438 | 17.056 |
| 50 | 20.845 | 66.589 | 24.369 | 35.271 | 35.356 | 16.544 | 32.870 | 7.525 | 30.312 |
| 51 | 12.719 | 68.425 | 23.278 | 29.511 | 23.339 | 9.326 | 23.659 | 5.488 | 20.215 |
| 52 | 5.047 | 24.401 | 8.784 | 12.000 | 12.983 | 5.413 | 11.911 | 2.805 | 11.398 |
| 53 | 592.840 | 2517.991 | 805.818 | 1081.070 | 1176.201 | 278.227 | 1103.179 | 361.182 | 1097.058 |
| 54 | 155.567 | 1192.460 | 360.272 | 528.626 | 576.926 | 138.766 | 486.027 | 185.445 | 391.427 |
| 55 | 61.658 | 194.609 | 62.415 | 102.119 | 95.833 | 114.869 | 95.927 | 22.953 | 90.970 |
| 56 | 107.406 | 332.353 | 104.112 | 170.212 | 167.513 | 122.071 | 165.628 | 39.294 | 160.536 |
| 57 | 129.055 | 453.517 | 149.477 | 228.184 | 241.046 | 104.547 | 224.676 | 53.789 | 200.278 |
| 58 | 161.419 | 747.469 | 245.021 | 359.890 | 381.408 | 139.343 | 335.427 | 83.747 | 275.112 |
| 59 | 47.987 | 227.330 | 75.912 | 109.343 | 115.627 | 51.540 | 92.562 | 22.376 | 85.612 |
| 60 | 109.794 | 948.890 | 352.084 | 479.342 | 509.153 | 200.936 | 404.984 | 105.777 | 325.900 |
| 61 | 97.259 | 593.706 | 207.197 | 273.089 | 292.203 | 85.000 | 209.108 | 58.971 | 165.696 |
| 62 | 14.713 | 71.788 | 22.534 | 35.026 | 33.575 | 13.534 | 26.376 | 6.607 | 24.446 |
| 63 | 87.751 | 326.471 | 109.069 | 169.564 | 163.047 | 71.972 | 154.950 | 39.195 | 143.217 |
| 64 | 15.532 | 43.716 | 14.269 | 19.188 | 20.359 | 10.593 | 18.858 | 4.679 | 17.495 |
| 65 | 46.881 | 161.424 | 53.980 | 80.485 | 85.190 | 31.164 | 70.268 | 19.030 | 62.904 |
| 66 | 98.995 | 328.286 | 113.339 | 169.576 | 178.070 | 63.709 | 143.270 | 38.085 | 128.749 |
| 67 | 22.778 | 48.116 | 15.967 | 23.181 | 23.137 | 11.583 | 20.937 | 5.550 | 19.093 |
| 68 | 132.695 | 383.307 | 136.848 | 185.381 | 221.947 | 73.096 | 197.628 | 57.744 | 167.687 |
| 69 | 14.783 | 40.655 | 12.813 | 19.714 | 18.432 | 6.878 | 16.528 | 4.114 | 16.635 |
| 70 | 0.137 | 44.975 | 14.473 | 24.888 | 22.363 | 22.985 | 23.388 | 5.669 | 29.394 |
| 71 | 0.137 | 2.776 | 0.917 | 1.640 | 1.157 | 3.581 | 1.759 | 0.424 | 1.804 |
|  | 0.137 | 0.145 | 0.204 | 0.151 | 0.151 | 25.748 | 0.254 | 0.280 | 7.957 |
| 61 | 230.049 | 1313.582 | 430.247 | 700.937 | 537.707 | 414.304 | 664.911 | 164.335 | 594.667 |
| $\begin{aligned} & 61 R \\ & \text { AVE. } \end{aligned}$ | ND | ND | ND |  |  | B | ND | ND | B |
|  | 0.137 | 0.145 | 0.204 | 0.151 | 0.151 | 0.151 | 0.254 | 0.280 | 0.201 |
|  | 453.870 | 413.456 | 640.786 | 153.617 | 571.725 |  |  |  |  |
|  | BENZO(A)PYRENE | PERYLENE | INDENO(1,2,3-C,D)PYRENE DIBENZ(A,H)ANTHRACENE BENZO(G,H,I)PERYLENE |  |  |  |  |  |  |

Appendix B3. Chemistry and toxicity data from St. Andrew Bay.


| Appendix | B3. Chemistry | and toxicity data | from St. Andrew | Bay. |
| :---: | :---: | :---: | :---: | :---: |
| Station | CL3(18) | CL3(28) | HEPTACHLOR | CL |
| Number | ng $/ \mathrm{g}$ dry wt. | ng $/ \mathrm{g}$ dry wt. | ng/g dry wt. | ng $/ \mathrm{g}$ |


| Station Number | $\begin{gathered} \text { CL3(18) } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \\ \hline \end{gathered}$ | $\begin{gathered} \text { CL3(28) } \\ \text { ng/g dry wt. } \\ \hline \end{gathered}$ | HEPTACHLOR $\mathrm{ng} / \mathrm{g}$ dry wt. | $\begin{gathered} \text { CL4(52) } \\ \text { ng/g dry wt. } \end{gathered}$ | ALDRIN $\mathrm{ng} / \mathrm{g}$ dry wt. | $\begin{gathered} \text { CL4(44) } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | $\begin{gathered} \hline \text { CL5(103) } \\ \text { ng/g dry wt. } \end{gathered}$ | HEPTACHLOREPOXIDE $\mathrm{ng} / \mathrm{g}$ dry wt. | $\begin{gathered} \hline \text { CL4(66) } \\ \text { ng/g dry wt. } \\ \hline \end{gathered}$ | $\begin{gathered} \text { 2,4-DDE } \\ \mathrm{ng} / \mathrm{g} \text { dry } \mathrm{wt.} \end{gathered}$ | $\begin{gathered} \hline \text { CL5(101) } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | CIS-CHLORDANE ng/g dry wt. |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| controls |  |  |  |  |  |  |  |  |  |  |  |  |
| 41 | 0.045781151 | 0.054233864 | 0.046883679 | 0.051871305 | 0.045676149 | 0.049515551 | 2.260398266 | 0.030477016 | 0.015122 | 0.01399818 | 0.01565762 | 0.037176811 |
| 42 | 0.167417016 | 0.054233864 | 0.046883679 | 0.127127051 | 0.045676149 | 0.258908813 | 7.453491034 | 0.030477016 | 0.186646 | 0.309958031 | 0.217932087 | 0.046883679 |
| 43 | 0.257017834 | 0.054233864 | 0.046883679 | 0.296647952 | 0.045676149 | 0.041817302 | 5.904474901 | 0.030477016 | 0.173217 | 0.144484808 | 0.462846764 | 0.076122853 |
| 44 | 0.29696053 | 0.054233864 | 0.046883679 | 0.144987035 | 0.045676149 | 0.255617978 | 6.558628637 | 0.030477016 | 0.121363 | 0.223638721 | 0.220685681 | 0.077931432 |
| 45 | 0.229907937 | 0.054233864 | 0.046883679 | 0.139918719 | 0.045676149 | 0.041817302 | 7.696276022 | 0.030477016 | 0.190761 | 0.720909016 | 0.298747615 | 0.044123746 |
| 46 | 0.165974354 | 0.054233864 | 0.046883679 | 0.073866715 | 0.045676149 | 0.364276684 | 8.595358497 | 0.030477016 | 0.496478 | 1.544789597 | 0.256817771 | 0.046883679 |
| 47 | 0.101551814 | 0.054233864 | 0.046883679 | 0.084099807 | 0.045676149 | 0.203858309 | 4.9607094 | 0.030477016 | 0.102731 | 0.203103627 | 0.204518655 | 0.057167115 |
| 48 | 0.163135854 | 0.054233864 | 0.046883679 | 0.023740608 | 0.045676149 | 0.041817302 | 2.697561276 | 0.030477016 | 0.065402 | 0.090867102 | 0.09850351 | 0.025742086 |
| 49 | 0.12180952 | 0.054233864 | 0.046883679 | 0.051871305 | 0.045676149 | 0.041817302 | 3.25554824 | 0.030477016 | 0.12363 | 0.122313025 | 0.198226113 | 0.00828847 |
| 50 | 0.248818256 | 0.054233864 | 0.046883679 | 0.051871305 | 0.045676149 | 0.041817302 | 7.103667482 | 0.030477016 | 0.150041 | 0.215077425 | 0.397799511 | 0.087775061 |
| 51 | 0.420258489 | 0.054233864 | 0.046883679 | 0.096506737 | 0.045676149 | 0.407029202 | 2.810095714 | 0.030477016 | 0.075261 | 0.132780589 | 0.223373773 | 0.214655856 |
| 52 | 0.076492537 | 0.054233864 | 0.046883679 | 0.051871305 | 0.045676149 | 0.041817302 | 2.401463835 | 0.030477016 | 0.022158 | 0.142192882 | 0.061394948 | 0.1163031 |
| 53 | 3.624953966 | 0.054233864 | 0.046883679 | 6.312292848 | 0.045676149 | 3.974810341 | 6.253590631 | 0.030477016 | 4.580762 | 3.864034765 | 10.96007955 | 14.55063711 |
| 54 | 1.202265304 | 0.054233864 | 0.046883679 | 2.009035761 | 0.045676149 | 1.416111653 | 3.609850252 | 0.030477016 | 1.430789 | 0.734569211 | 3.642345056 | 3.407881899 |
| 55 | 1.448478901 | 0.054233864 | 0.046883679 | 1.443976217 | 0.045676149 | 1.526179068 | 4.456560642 | 0.030477016 | 1.327888 | 0.82179761 | 2.680655776 | 0.988570109 |
| 56 | 1.220627435 | 0.054233864 | 0.046883679 | 2.986740483 | 0.045676149 | 1.494566332 | 5.353701046 | 0.030477016 | 1.010731 | 2.447747932 | 3.464766591 | 1.344063974 |
| 57 | 2.569012666 | 0.054233864 | 0.046883679 | 2.275259067 | 0.045676149 | 1.581102476 | 6.644142199 | 0.030477016 | 1.724813 | 2.76460852 | 3.538212435 | 1.427101324 |
| 58 | 3.051955585 | 0.054233864 | 0.046883679 | 2.723152138 | 0.045676149 | 2.329714949 | 7.645343056 | 0.030477016 | 2.438598 | 10.1182466 | 4.964285714 | 1.140536957 |
| 59 | 0.995374653 | 0.054233864 | 0.046883679 | 0.934050262 | 0.045676149 | 0.896584952 | 3.100794018 | 0.030477016 | 0.709451 | 0.237897009 | 1.60021585 | 0.342275671 |
| 60 | 2.484995764 | 0.054233864 | 0.046883679 | 2.800998796 | 0.045676149 | 2.449235297 | 3.783118562 | 0.030477016 | 2.668391 | 0.488384536 | 3.733847594 | 2.161813885 |
| 61 | 1.799331913 | 0.054233864 | 0.046883679 | 1.139378389 | 0.045676149 | 0.961367157 | 4.580412471 | 0.030477016 | 1.4253 | 0.33341402 | 2.948489543 | 1.367931836 |
| 62 | 0.347262706 | 0.054233864 | 0.046883679 | 0.329800201 | 0.045676149 | 0.295034183 | 2.029201336 | 0.030477016 | 0.315835 | 0.037468864 | 0.547750278 | 0.084980656 |
| 63 | 2.357749469 | 0.054233864 | 0.046883679 | 2.619367775 | 0.045676149 | 1.734666195 | 5.291696155 | 0.030477016 | 1.924628 | 0.784383109 | 5.905874027 | 0.660061335 |
| 64 | 0.492470305 | 0.054233864 | 0.046883679 | 0.370616516 | 0.045676149 | 0.277891686 | 3.027891686 | 0.030477016 | 0.238546 | 0.046486143 | 0.482678167 | 0.054227941 |
| 65 | 0.970965038 | 0.054233864 | 0.046883679 | 0.526400862 | 0.045676149 | 0.393019636 | 4.762631705 | 0.030477016 | 0.308908 | 0.161518199 | 1.483716475 | 0.211147031 |
| 66 | 2.567722076 | 0.054233864 | 0.046883679 | 2.209058927 | 0.045676149 | 0.601934916 | 7.907124011 | 0.030477016 | 1.644855 | 0.71292876 | 4.262445031 | 0.427616535 |
| 67 | 0.562042671 | 0.054233864 | 0.046883679 | 0.498883155 | 0.045676149 | 0.345451745 | 2.898559655 | 0.030477016 | 0.369637 | 0.049949935 | 1.149618732 | 0.179696526 |
| 68 | 0.975232631 | 0.054233864 | 0.046883679 | 0.895193451 | 0.045676149 | 0.760092353 | 5.542713216 | 0.030477016 | 0.748058 | 0.293780172 | 2.16217729 | 0.3227454 |
| 69 | 0.507691804 | 0.054233864 | 0.046883679 | 0.116391726 | 0.045676149 | 0.119272061 | 3.567982456 | 0.030477016 | 0.072303 | 0.151544907 | 0.27117701 | 0.105721393 |
| 70 | 0.361031709 | 0.054233864 | 0.046883679 | 0.247025194 | 0.045676149 | 0.319816526 | 6.222429037 | 0.030477016 | 0.269162 | 0.11613375 | 0.532473576 | 0.007977132 |
| 71 | 0.045781151 | 0.054233864 | 0.046883679 | 0.028813938 | 0.045676149 | 0.05036855 | 2.492628993 | 0.030477016 | 0.016724 | 0.035018381 | 0.022392227 | 0.046883679 |
|  | 0.045781151 | 0.054233864 | 0.046883679 | 0.051871305 | 0.045676149 | 0.157585175 | 6.093261922 | 0.030477016 | 0.063973 | 0.035018381 | 0.068015461 | 0.046883679 |
|  | 3.264386 | 5.630215543 | 1.813130314 | 4.68736405 | 0.045676149 | 6.36919122 | 16.574649 | 2.729879375 | 10.41685 | 0.035018381 | 10.69290093 | 2.042119834 |
|  | 0.045781151 | 0.054233864 | 0.046883679 | 0.051871305 | 0.045676149 | 0.110756402 | 5.11280482 | 0.030477016 | 0.063973 | 0.035018381 | 0.068015461 | 0.046883679 |
| 61 | 2.901851483 | 5.905833167 | 1.116663349 | 3.783197292 | 0.045676149 | 5.956599642 | 14.44017519 | 2.433605415 | 5.301015 | 0.035018381 | 9.704957197 | 1.709735218 |
| $\begin{gathered} 61 \mathrm{R} \\ \text { AVE. } \end{gathered}$ |  |  |  |  |  |  |  |  |  |  |  |  |


| Appendix | Chemistry and | xicity d | rom St. | ew Bay. |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Station Number | TRANS-NONACHLOR $\mathrm{ng} / \mathrm{g}$ dry wt. | $\begin{gathered} \text { CL5(112) } \\ \text { ng/g dry wt. } \end{gathered}$ | $\begin{gathered} \text { DIELDRIN } \\ \mathrm{ng} / \mathrm{g} \text { dry } \mathrm{wt} . \end{gathered}$ | $\begin{gathered} \text { 4,4-DDE } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | $\begin{gathered} \text { CL4(77) } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | $\begin{gathered} \hline \text { 2,4-DDD } \\ \text { ng/g dry wt. } \end{gathered}$ | $\begin{gathered} \text { ENDRIN } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | $\begin{gathered} \hline \text { CL5(118) } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | $\begin{gathered} \hline \text { 4,4-DDD } \\ \text { ng/g dry wt. } \end{gathered}$ | $\begin{gathered} \text { 2,4-DDT } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | $\begin{gathered} \hline \text { CL6(153) } \\ \text { ng/g dry wt. } \end{gathered}$ | $\begin{gathered} \hline \text { CL5(105) } \\ \mathrm{ng} / \mathrm{g} \text { dry wt. } \end{gathered}$ | $\begin{gathered} \hline \text { 4,4-DDT } \\ \text { ng/g dry wt. } \end{gathered}$ |
| controls |  |  |  |  |  |  |  |  |  |  |  |  |  |
| 41 | 0.045902254 | 0 | 0.020823296 | 0.27514587 | 0.063972859 | 0.142123013 | 0.057987708 | 0.030244634 | 0.13666292 | 0.06838297 | 0.031823778 | 0.015898507 | 0.158262406 |
| 42 | 0.080045784 | 0 | 0.150247997 | 3.27333079 | 0.063972859 | 0.843113316 | 0.057987708 | 0.467149943 | 1.510034338 | 0.06838297 | 0.564059519 | 0.215413964 | 0.183899275 |
| 43 | 0.034180978 | 0 | 0.176931968 | 2.882678336 | 0.063972859 | 3.044996697 | 0.057987708 | 0.726799868 | 2.377229194 | 0.06838297 | 1.055812417 | 0.316875826 | 3.421895641 |
| 44 | 0.038389513 | 0 | 0.127845001 | 3.589527514 | 0.063972859 | 1.240276577 | 0.057987708 | 0.407519447 | 2.091112071 | 0.228392394 | 0.586862576 | 0.155862864 | 1.99891962 |
| 45 | 0.046883679 | 0 | 0.240772995 | 4.370075475 | 0.063972859 | 0.823422078 | 0.057987708 | 0.397279589 | 1.842746952 | 0.06838297 | 0.565729452 | 0.245749357 | 0.496723895 |
| 46 | 0.202185299 | 0 | 0.201191981 | 6.134368792 | 0.063972859 | 1.806664259 | 0.057987708 | 0.442206971 | 3.010926494 | 0.06838297 | 4.443200289 | 1.602853531 | 6.131208236 |
| 47 | 0.046883679 | 0 | 0.083486628 | 6.321069761 | 0.063972859 | 1.602518749 | 0.057987708 | 0.180557521 | 7.673175794 | 0.313522947 | 0.202160275 | 0.497523702 | 17.50813641 |
| 48 | 0.012686291 | 0 | 0.064509176 | 0.732725705 | 0.063972859 | 0.383113684 | 0.057987708 | 0.228630373 | 0.54470994 | 0.06838297 | 0.328704274 | 0.094500554 | 0.046157162 |
| 49 | 0.030326504 | 0 | 0.106936752 | 0.84201557 | 0.063972859 | 0.821371858 | 0.057987708 | 0.330880359 | 1.007087804 | 0.06838297 | 0.498818699 | 0.08431775 | 1.01328479 |
| 50 | 0.097310513 | 0 | 0.212469438 | 1.317603912 | 0.063972859 | 1.299592502 | 0.057987708 | 0.650203749 | 1.46609617 | 0.06838297 | 0.913854931 | 0.25590872 | 0.352648737 |
| 51 | 0.083978541 | 0 | 0.246204963 | 0.809059318 | 0.063972859 | 1.146558556 | 0.057987708 | 0.239041639 | 1.079954886 | 0.06838297 | 1.923306712 | 0.707919283 | 0.168048528 |
| 52 | 0.008438576 | 0 | 0.078616533 | 0.358926521 | 0.063972859 | 0.408754305 | 0.057987708 | 0.127583238 | 0.376722158 | 0.06838297 | 0.210074627 | 0.106917336 | 0.061165327 |
| 53 | 8.649186124 | 0 | 5.045518156 | 66.39561314 | 0.063972859 | 36.02393754 | 0.057987708 | 19.12079252 | 43.68188533 | 0.06838297 | 24.01966561 | 6.581792738 | 2.913235619 |
| 54 | 2.181606075 | 0 | 1.446188436 | 15.46628738 | 0.063972859 | 11.61303186 | 0.057987708 | 5.962075256 | 14.15616387 | 0.67577313 | 5.652483774 | 3.068171213 | 0.915793493 |
| 55 | 0.548634763 | 0 | 0.479939964 | 17.43433585 | 0.063972859 | 2.876811176 | 0.057987708 | 5.412284246 | 6.124747446 | 0.06838297 | 3.993188247 | 2.257692086 | 0.276741904 |
| 56 | 0.619438179 | 0 | 0.981409336 | 23.20552252 | 0.063972859 | 6.079830497 | 0.057987708 | 6.779851001 | 9.409473037 | 0.06838297 | 5.78231153 | 3.787095892 | 0.564691409 |
| 57 | 0.626871042 | 0 | 0.791090961 | 19.55857801 | 0.063972859 | 4.779504893 | 0.057987708 | 6.376079447 | 9.339090386 | 0.06838297 | 5.728195164 | 2.871761658 | 0.578871618 |
| 58 | 0.652055022 | 0 | 1.689923765 | 22.71486576 | 0.063972859 | 7.0391117 | 0.057987708 | 10.58352668 | 19.18727212 | 1.143602917 | 8.638133908 | 4.066291018 | 2.45782234 |
| 59 | 0.155257478 | 0 | 0.305003084 | 6.802073697 | 0.063972859 | 2.701048412 | 0.057987708 | 3.077243293 | 5.897818378 | 0.06838297 | 2.571500154 | 1.275747764 | 0.156953438 |
| 60 | 1.077228341 | 0 | 0.640344228 | 28.04561466 | 0.063972859 | 16.73803897 | 0.057987708 | 6.847282294 | 17.80153601 | 0.750167209 | 5.354126722 | 3.163196148 | 5.438979801 |
| 61 | 0.747821456 | 0 | 1.581041828 | 13.24089853 | 0.063972859 | 4.204541053 | 0.057987708 | 4.791150271 | 13.11952943 | 0.06838297 | 4.288584431 | 2.338594113 | 0.555383424 |
| 62 | 0.054427898 | 0 | 0.125708835 | 1.753643542 | 0.063972859 | 0.956065504 | 0.057987708 | 0.776962213 | 2.732365255 | 0.06838297 | 0.755074461 | 0.343632413 | 0.323493561 |
| 63 | 0.199339467 | 0 | 0.653043171 | 25.4445624 | 0.063972859 | 8.398030196 | 0.057987708 | 8.700283086 | 18.85365014 | 0.06838297 | 9.317763623 | 3.47287096 | 0.475406936 |
| 64 | 0.110506222 | 0 | 0.058010464 | 1.161587952 | 0.063972859 | 0.7109375 | 0.057987708 | 0.758802319 | 1.713235294 | 0.06838297 | 0.768417704 | 0.343820701 | 0.109375 |
| 65 | 0.067528736 | 0 | 0.194085249 | 3.208213602 | 0.063972859 | 2.620150862 | 0.057987708 | 2.471264368 | 4.691810345 | 0.06838297 | 2.826688218 | 1.212164751 | 0.250778257 |
| 66 | 0.124890062 | 0 | 0.434828496 | 8.523658751 | 0.063972859 | 5.365171504 | 0.057987708 | 5.522427441 | 10.69199648 | 0.06838297 | 7.872031662 | 3.027616535 | 0.419876869 |
| 67 | 0.046883679 | 0 | 0.093660941 | 1.652468613 | 0.063972859 | 1.232380806 | 0.057987708 | 1.764230147 | 2.694215513 | 0.06838297 | 1.737849495 | 0.840791805 | 0.141954864 |
| 68 | 0.046883679 | 0 | 0.103827048 | 7.142517316 | 0.063972859 | 6.730707339 | 0.057987708 | 3.488280977 | 8.468690968 | 0.06838297 | 3.386482894 | 1.515357168 | 0.926257609 |
| 69 | 0.072401152 | 0 | 0.036396963 | 6.550602252 | 0.063972859 | 2.459217073 | 0.057987708 | 0.305282797 | 22.74718513 | 0.153377848 | 0.402035873 | 0.191018591 | 2.536789736 |
| 70 | 0.251346141 | 0 | 0.14478495 | 2.728578076 | 0.063972859 | 1.063551153 | 0.057987708 | 0.728112743 | 2.014957123 | 0.06838297 | 1.048859935 | 0.338030978 | 0.893571761 |
| 71 | 0.068321421 | 0 | 0.014937458 | 0.194159035 | 0.063972859 | 0.108498995 | 0.057987708 | 0.019488497 | 0.114753183 | 0.06838297 | 0.038334822 | 0.008599509 | 0.06838297 |
|  | 0.046883679 | 0 | 0.035018381 | 0.035018381 | 0.063972859 | 0.106498928 | 0.057987708 | 0.035569645 | 0.136268164 | 0.06838297 | 0.034283362 | 0.048563721 | 0.06838297 |
|  | 0.046883679 | 0 | 2.381253708 | 6.368004746 | 0.063972859 | 8.483290488 | 0.057987708 | 6.902313625 | 4.600158197 | 0.06838297 | 8.386592842 | 3.275459759 | 1.790587305 |
|  | 0.046883679 | 0 | 0.035018381 | 0.035018381 | 0.063972859 | 0.106498928 | 0.057987708 | 0.035569645 | 0.039991428 | 0.06838297 | 0.034283362 | 0.048563721 | 0.06838297 |
| 61 | 0.046883679 | 0 | 1.964961179 | 5.932709536 | 0.063972859 | 10.3014135 | 0.057987708 | 6.434999005 | 3.776826598 | 0.06838297 | 7.002588095 | 4.132391001 | 0.86860442 |
| $\begin{aligned} & \text { 61R } \\ & \text { AVE. } \end{aligned}$ |  |  |  |  |  |  |  |  |  |  |  |  |  |



| 41 | 0.041030994 | 0.048563721 | 3.550639687 | 0.026880676 | 0.0434 | 0.01228521 | 0.0684 | 0.011027247 | 0.005942 | 0.040426 | 0.004335956 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 42 | 0.591835177 | 0.048563721 | 12.95719191 | 0.335062953 | 0.1641 | 0.199618466 | 0.0684 | 0.117054559 | 0.057077 | 0.1209462 | 0.109576498 |
| 43 | 1.010898283 | 0.048563721 | 12.05242734 | 0.467140026 | 0.275 | 0.555399604 | 0.0684 | 0.304243725 | 0.132596 | 0.1386229 | 0.40596103 |
| 44 | 0.545736099 | 0.048563721 | 10.54271103 | 0.208225295 | 0.125 | 0.286949006 | 0.0684 | 0.099539038 | 0.066695 | 0.0675598 | 0.195692884 |
| 45 | 0.660860911 | 0.048563721 | 12.95206104 | 0.316745459 | 0.1961 | 0.208509579 | 0.0684 | 0.15036908 | 0.056316 | 0.064278 | 0.444472091 |
| 46 | 0.628318584 | 0.048563721 | 13.31406899 | 0.420444284 | 0.1525 | 0.210854253 | 0.0684 | 0.14999097 | 0.063392 | 0.0680874 | 0.348925411 |
| 47 | 0.322437621 | 0.048563721 | 7.9128343 | 0.240790529 | 0.0877 | 0.198575539 | 0.0684 | 0.144332814 | 0.062874 | 0.033206 | 0.098580256 |
| 48 | 0.329412489 | 0.048563721 | 5.193773864 | 0.136069713 | 0.0945 | 0.146169479 | 0.0684 | 0.043293509 | 0.031346 | 0.0264503 | 0.058443158 |
| 49 | 0.504938224 | 0.048563721 | 6.228126573 | 0.264688795 | 0.1314 | 0.316743483 | 0.0684 | 0.133816182 | 0.136334 | 0.3056664 | 0.2067857 |
| 50 | 0.782314588 | 0.048563721 | 12.08891606 | 0.47791361 | 0.2804 | 0.448410758 | 0.0684 | 0.245476773 | 0.108476 | 0.0572127 | 0.187123064 |
| 51 | 0.30210937 | 0.048563721 | 5.755867829 | 0.249771383 | 0.1466 | 0.164512589 | 0.0684 | 0.394440041 | 0.078217 | 0.0481924 | 0.25013717 |
| 52 | 0.196727899 | 0.048563721 | 4.253329506 | 0.079104478 | 0.051 | 0.090585534 | 0.0684 | 0.051349024 | 0.024713 | 0.0279277 | 0.17445465 |
| 53 | 36.2288429 | 0.048563721 | 13.66288576 | 11.21654268 | 6.3376 | 14.15901893 | 0.0684 | 8.747072255 | 4.445017 | 5.0877955 | 4.400235693 |
| 54 | 9.289483731 | 0.048563721 | 6.618122428 | 3.025664956 | 1.8974 | 4.568022738 | 0.0684 | 3.139057396 | 1.457048 | 1.6405209 | 1.368769355 |
| 55 | 6.415632396 | 0.048563721 | 8.075044738 | 1.437106737 | 1.3725 | 2.504185187 | 0.0684 | 1.525313167 | 0.422213 | 0.3525948 | 0.581019454 |
| 56 | 8.598933771 | 0.048563721 | 11.54316178 | 2.088989133 | 1.9649 | 3.666393275 | 0.0684 | 2.495249812 | 0.767617 | 0.4910122 | 1.054951815 |
| 57 | 8.287852619 | 0.048563721 | 13.78662925 | 2.405080599 | 1.9365 | 3.854058722 | 0.0684 | 2.078727691 | 0.825921 | 0.6373777 | 0.897380541 |
| 58 | 12.16050713 | 0.048563721 | 14.11725224 | 3.82424594 | 3.0834 | 5.752817368 | 0.0684 | 3.015081206 | 1.180892 | 1.8169539 | 1.299386808 |
| 59 | 3.545251311 | 0.048563721 | 6.938328708 | 1.146238051 | 0.9215 | 1.497032069 | 0.0684 | 0.952397471 | 0.310052 | 0.2379741 | 0.133749615 |
| 60 | 7.634726 | 0.048563721 | 6.664779061 | 2.780309448 | 1.9613 | 4.796762831 | 0.0684 | 3.081508896 | 1.168413 | 1.3145762 | 1.881749677 |
| 61 | 6.158549574 | 0.048563721 | 8.28718048 | 2.951684741 | 1.6511 | 3.98813904 | 0.0684 | 2.070826878 | 0.817777 | 0.4092273 | 0.782339272 |
| 62 | 0.850548519 | 0.048563721 | 4.303169219 | 0.240235307 | 0.2352 | 0.429090042 | 0.0684 | 0.194366421 | 0.094573 | 0.0570512 | 0.120091155 |
| 63 | 11.72464025 | 0.048563721 | 10.28373437 | 2.852441614 | 2.0538 | 5.621078084 | 0.0684 | 2.880219391 | 0.94468 | 0.6233192 | 0.396614768 |
| 64 | 0.937747455 | 0.048563721 | 5.828089649 | 0.451887726 | 0.2084 | 0.682656957 | 0.0684 | 0.25 | 0.120829 | 0.1202277 | 0.112980769 |
| 65 | 2.944264847 | 0.048563721 | 10.1272749 | 1.078603927 | 0.5626 | 1.687559866 | 0.0684 | 1.207136015 | 0.431573 | 0.4178041 | 0.356920498 |
| 66 | 8.928056288 | 0.048563721 | 17.22102023 | 3.344151275 | 1.4648 | 5.622515391 | 0.0684 | 2.095602463 | 0.94978 | 0.7913808 | 0.888566403 |
| 67 | 2.05106678 | 0.048563721 | 6.186898252 | 0.784641454 | 0.4521 | 0.99453131 | 0.0684 | 0.310444427 | 0.160441 | 0.1831241 | 0.055495648 |
| 68 | 4.247393829 | 0.048563721 | 12.44147485 | 2.41558805 | 1.0611 | 2.433289023 | 0.0684 | 0.817323165 | 0.717064 | 0.6056811 | 0.553347793 |
| 69 | 0.399286462 | 0.048563721 | 5.941051322 | 0.248985336 | 0.1405 | 0.285840534 | 0.0684 | 0.114231474 | 0.072172 | 0.0816313 | 0.193244305 |
| 70 | 0.968024995 | 0.048563721 | 10.89443595 | 0.406102506 | 0.2837 | 0.440204746 | 0.0684 | 0.160007977 | 0.082962 | 0.0396862 | 0.01721731 |
| 71 | 0.055813044 | 0.048563721 | 4.303048917 | 0.035179808 | 0.0189 | 0.018790485 | 0.0684 | 0.014099844 | 0.015356 | 0.040426 | 0.017980791 |
|  | 0.142601822 | 0.048563721 | 16.19535079 | 0.026880676 | 0.0434 | 0.036882178 | 0.3524 | 0.064518226 | 0.023284 | 0.1029456 | 0.155647265 |
|  | 9.856041131 | 0.048563721 | 35.21198339 | 6.515523037 | 3.1244 | 8.1908246 | 0.0684 | 9.986948784 | 3.279415 | 4.7609254 | 12.76666008 |
|  | 0.038693473 | 0.048563721 | 7.877460466 | 0.026880676 | 0.0434 | 0.036882178 | 0.0684 | 0.032157059 | 0.023284 | 0.040426 | 0.038147852 |
| 61 | 8.357555246 | 0.048563721 | 34.27095361 | 5.449333068 | 2.6327 | 7.217997213 | 0.0684 | 6.502886721 | 2.950627 | 5.1919172 | 10.93509855 |
| $61 R$ <br> AVE. |  |  |  |  |  |  |  |  |  |  |  |

Appendix











[^0]:    WQAP = water quality adjusted porewater

    * significantly different from controls (t-test, $a<0.05$ );
    ** significantly different from controls and $80 \%$ or less than the control response.

[^1]:    Pensacola Bay, 1993
    Stratum Station

[^2]:    WQAP = water quality adjusted porewater

    * significantly different from controls (t-test, $a<0.05$ );
    ** significantly different from controls and $80 \%$ or less than the control response.

[^3]:    a Suspect or genotoxic = toxic category; genotoxic = highly toxic category; nd = no data.

[^4]:    Pesticides
    Lindane
    Heptachlor epoxide Alpha CHL Trans Nono
     4,4-DDD 2,4'-DDE 2,4-DDT 4,4-DDT
    endrin (ug/goc)

[^5]:    $n s=$ not significant $(p>0.05)$ $*$ significant ( $p<0.05$ )
    nd $=$ no data

[^6]:    * ERL and ERM values from Long et al. (1995) and TEL and PEL values from MacDonald (1994). na = no applicable values.

[^7]:    * ERL and ERM values from Long et al. (1995) and TEL and PEL values from MacDonald (1994). na = no applicable values.

[^8]:    * ERL/ERM values from Long et al. (1995), TEL/PEL values from MacDonald (1994), SQC values from U. S. EPA (1994). na $=$ no applicable values.

[^9]:    AVS = acid volatile sulfides
    SEM = simultaneously-extracted metals UAN = un-ionized ammonia ERM = effects range-median TOC = total organic carbon Fines = silt + clay

    COH = chlorinated organic hydrocarbons PAHs = polynuclear aromatic hydrocarbons LPAHs = low moluecular weight PAHs HPAHs = high molecular weight PAHs < mdl $=$ less than method detection limit nd = no data

[^10]:    ERL/ERM values from Long et al. (1995); TEL/PEL values from MacDonald (1994); SQC's from U. S. EPA (1994). $\mathrm{na}=$ no applicable guidelines available

    Urchin UAN LOEC from Long et al. (in press); amphipod UAN NOEC from Kohn et al. (1994).

[^11]:    ppb

