

U.S. DEPARTMENT OF COMMERCE National Oceanic and Atmospheric Administration

Pacific Islands Fisheries Science Center 1845 Wasp Blvd. Bldg. 176 • Honolulu, Hawaii 96818

Final Project Instructions

Date	SII	hm	itted:
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June 21, 2016

Platform:

NOAA Ship Hi'ialakai

Project Number:

HA-16-06

Project Title:

HARAMP (Hawaiian Archipelago RAMP)

Project Dates:

July 12, 2016 to September 30, 2016

Prepared by:

Dated:

6/22/16

Joao Garriques

For: Bernardo Vargas-Ángel, Ph.D., Chief Scientist Legs I & II

Brett D. Schumacher, Ph.D., Chief Scientist Leg III

Coral Reef Ecosystem Program

Pacific Islands Fisheries Science Center

Approved by:

Datad

6/22/16

Michael Seki, Ph.D., Director

Pacific Islands Fisheries Science Center

Approved by:

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Dated: 24JUNE 2016

Commander Matthew J. Wingate, NOAA

Commanding Officer

Marine Operations Center - Pacific Islands



I. Overview

A. Brief Summary and Project Period

NOAA Ship *Hi'ialakai* will be engaged as support for the Hawaiian Archipelago Reef Assessment and Monitoring Program (HARAMP) from July 12 through September 30, 2016, for a total of 72 days at sea (DAS).

HARAMP is a component of an integrated coral reef ecosystem assessment led by the Coral Reef Ecosystem Program (CREP) of the Pacific Islands Fisheries Science Center in some 50 U.S.-affiliated Pacific Islands. This comprehensive, multi-agency research and education effort is sponsored by NOAA's Coral Reef Conservation Program (CRCP), a partnership between the National Marine Fisheries Service, National Ocean Service, and other NOAA agencies with the objective of improving understanding and management of coral reef ecosystems.

Small boats will be deployed from *Hi'ialakai* to reach dive survey areas around Hawai'i, Maui, Lāna'i, Moloka'i, Kaho'olawe, Kaua'i, Ni'ihau and O'ahu, in the Main Hawaiian Islands (MHI), including areas along windward coasts and near channels between the islands. In addition, small boat operations will also occur in the Northwestern Hawaiian Islands (NWHI); specifically the reef systems at French Frigate Shoals, Neva Shoals/Lisianski Island, Pearl and Hermes Atoll, and Kure Atoll. Teams of scuba divers will conduct fine-scale, rapid ecological assessment (REA) surveys of reef fishes, corals, other invertebrates, and algae. Taxonomic diversity of coral reefs will be evaluated by retrieving autonomous reef monitoring structures (ARMS) previously placed on the seafloor.

Scientists will collect data to monitor nearshore physical and ecological factors associated with ocean acidification and general water quality, including data on water temperature, salinity, and other physical and biological characteristics of the coral reef environment using an assortment of oceanographic sampling and monitoring instruments, including systems deployed from the ship, underwater moored instruments, and sensors on the ship.

Data collected during this mission are pivotal to long-term biological and oceanographic monitoring of coral reef ecosystems in the Hawaiian Archipelago. The 2016 expedition will add to information collected during monitoring and mapping surveys conducted in 2005, 2006, 2008, 2010 and 2013. Data on the abundance and spatial distribution of reef fishes, invertebrates, corals, and algae will allow scientists to evaluate potential changes in the condition and integrity of coral reef ecosystems in the Hawaiian Archipelago and enable federal and state resource managers to more effectively conserve coral reefs ecosystems of the MHI and manage ecosystem services. Data collected during the project also support monitoring components of the CRCP Coral Reef Ecosystem Integrated Observing System (CREIOS) in the Pacific.

B. Service Level Agreements

Of the <u>72</u> DAS scheduled for this project, <u>0</u> DAS are funded by the program and <u>72</u> DAS are funded by OMAO. This project is estimated to exhibit a High Operational Tempo.

C. Operating Area

The Operating Area for of HA-16-06 (*Appendix 1*) encompasses the waters around the Main Hawaiian Islands and the Northwestern Hawaiian Islands, specifically:

- Leg I: The operating area includes the nearshore waters Maui County (*Appendix 2*) and Kaua'i County (*Appendix 3*), with dive operations occurring at the islands of Moloka'i, Lāna'i, Maui, Kaho'olawe and Kaua'i, Ni'ihau respectively.
- Leg II: The operating are includes the nearshore waters of Hawai'i Island (Appendix 4) and O'ahu (Appendix 5).

Leg III: The operating area encompasses the water within and surrounding the Northwestern Hawaiian Archipelago. Dive operations will occur specifically at:

- French Frigate Shoals (Appendix 6)
- Lisianski (Appendix 7)
- Pearl and Hermes (Appendix 8)
- Kure Atoll (Appendix 9).

The Station/Waypoint List for instrument deployments and retrievals of the project is presented as an attached file (*Appendix 10*).

D. Summary of Objectives

The ship will support assessment and monitoring operations in the waters surrounding the Hawaiian Archipelago. The scientific objectives of this project are to:

- 1. Conduct ecosystem monitoring of the species composition, abundance, percent cover, size distribution, recruitment and general health of the fishes, corals, other invertebrates, and algae of the shallow water (< 35 m) coral reef ecosystems of the Hawaiian Archipelago
- 2. Deploy, retrieve and/or service an array of Subsurface Temperature Recorders (STRs), Sea Surface Temperature (SST) Buoys, Autonomous Reef Monitoring Structures (ARMS), Calcification Accretion Units (CAUs), Bioerosion Monitoring Units (BMUs), Ecological Acoustic Recorders (EARs), moored Acoustic Doppler Current Profilers (ADCPs) as well as anchored arrays consisting of a portable underwater collector (PUC), ADCP, a Conductivity Temperature Pressure (CTP) recorder, and a thermistor string to allow remote long-term monitoring of oceanographic and environmental conditions affecting the coral reef ecosystems of the Hawaiian Archipelago. This effort is in support of the Coral Reef Ecosystem Integrated Observing Systems (CREIOS).
- 3. Monitor nearshore physical and ecological factors associated with ocean acidification and general water quality, including analysis of seawater for

nutrients, chlorophyll concentration, salinity, temperature, dissolved oxygen, transmissivity, total alkalinity, and dissolved inorganic carbon. These parameters will be measured via the collection of water in Niskin bottles and conductivity-temperature-depth (CTD) casts. Shallow-water CTDs are conducted from small boats to a depth of ~30 m.

- 4. Collect shallow water coral cores to examine calcification/growth rates in recent decades and assess potential early impacts of ocean acidification. Coring operations will be conducted opportunistically (as a scientific dive).
- 5. Conduct plankton tows to expand upon the documentation of the marine biota on coral reefs.
- 6. Conduct shipboard Acoustic Doppler Current Profiler (ADCP) surveys around reef ecosystems to examine physical and biological linkages supporting and maintaining the island ecosystems.
- 7. Collect oceanographic data utilizing ship-based measurement systems (ADCP, ThermoSalinoGraph TSG, and the Scientific Computer System SCS) during all transits for the duration of the project.
- 8. Conduct investigations of marine microbial communities, including the collection of specimens via water sampling and benthic grab samples.
- 9. Determine the existence of threats to the health of these coral reef resources from anthropogenic sources, including marine debris.

E. Participating Institutions

- Joint Institute for Marine and Atmospheric Research (JIMAR)
- NOAA Pacific Islands Fisheries Science Center (PIFSC):
 - o Coral Reef Ecosystem Program (CREP)
 - o PIFSC Scientific Operations Division (SOD)
- NOAA Diving Program (NDP)
- Division of Aquatic Resources (DAR)
- Papahānaumokuākea Marine National Monument (PMNM)
- San Diego State University (SDSU)
- NOAA Commissioned Officer Corps (NOAA CORPS)
- The Nature Conservancy (TNC)
- Scrips Institute of Oceanography (SIO)
- Moss Landing Marine Labs (MLML)

F. Personnel/Science Party

Leg I

Name (Last, First)	Title	Gender	Affiliation	Nationality
Ayotte, Paula	Fish REA Diver	Female	CREP/JIMAR	USA
Raja, Kristin	Fish REA Diver	Female	NOAA CORPS	USA
Zamzow, Jill	Fish REA Diver	Female	CREP/JIMAR	USA
Purves, Andrew	Fish REA Diver	Male	CREP/JIMAR	USA
Tejchma, Kendall	Fish REA Diver	Female	DAR	USA
Vargas-Angel, Bernardo	Benthic REA Diver	Male	CREP/JIMAR	USA
Schumacher, Brett	Benthic REA Diver	Male	CREP/JIMAR	USA
White, Darla	Benthic REA Diver	Female	DAR	USA
Gray, Andrew	Fish REA/Tow Diver	Male	CREP/JIMAR	USA
McCoy, Kaylyn	Fish REA/Tow Diver	Female	CREP/JIMAR	USA
Bailey, Hatsue	Benthic REA/Tow Diver	Female	CREP/JIMAR	JPN (USA Perm. Resident
Garriques, Joao	Benthic REA/Tow Diver Male CREP/JIMA		CREP/JIMAR	USA
Slater, David	Oceanography Diver	Male	CREP/JIMAR	USA
Pomeroy, Noah	Oceanography Diver	Male	CREP/JIMAR	USA
Reardon, Russell	Oceanography Diver	Male	CREP/JIMAR	USA
Ferguson, Marie	Benthic REA Diver	Female	CREP/JIMAR	USA
Weible, Rebecca	Fish REA Diver	Female	UH	USA
Edwards, Clint	Mosaic Diver	Male	SIO	USA
Brandon, Reyes	Microbial Biologist	Male	SDSU	USA
Trick, Kevin	Data Manager	Male	CREP/JIMAR	USA
Gordon, Bill	Chamber Operator/ Dive Master	vive Male NDP		USA
TBD	TBD	Male	TBD	TBD

Leg II

Name (Last, First)	Title	Gender	Affiliation	Nationality
Ayotte, Paula	Fish REA Diver	Female	CREP/JIMAR	USA
Purves, Andrew	Fish REA Diver	Male	TNC	USA
Zamzow, Jill	Fish REA Diver	Female	CREP/JIMAR	USA
Carr, Ryan	Fish REA Diver	Male	TNC	USA
Raja, Kristin	Fish REA Diver	Female	NOAA CORPS	USA
Vargas-Angel, Bernardo	Benthic REA Diver	Male	CREP/JIMAR	USA
Ferguson, Marie	Benthic REA Diver	Female	CREP/JIMAR	USA
Lager, Daniel	Benthic REA Diver	Male	DAR	USA
Gray, Andrew	Fish REA/Tow Diver	Male	CREP/JIMAR	USA
Suka, Rhonda	Fish REA/Tow Diver	Female	CREP/JIMAR	USA
Lichowsky, Frances	Benthic REA/Tow Diver	Female	CREP/JIMAR	GER (USA Perm. Resident)
Garriques, Joao	Benthic REA/Tow Diver	Male	CREP/JIMAR	USA
Morioka, James	Oceanography Diver	Male	CREP/JIMAR	USA
Pomeroy, Noah	Oceanography Diver	Male	CREP/JIMAR	USA
Reardon, Russell	Oceanography Diver	Male	CREP/JIMAR	USA
Reardon, Kerry	ARMS Diver	Female	CREP/JIMAR	USA
Godwin, Scott	ARMS Diver	Male	PMNM	USA
Bonito, Lindsay	Mosaic Diver	Female	SIO	USA
Roach, Ty	Microbial Biologist	Male	SDSU	USA
Trick, Kevin	Data Manager	Male	CREP/JIMAR	USA
Hileman, Zack	Chamber Operator/ Dive Master	Male	NDP	USA
Heidt, Amanda	ARMS Lab Assistant	Female	MLML	USA

Leg III

Name (Last, First)	Title	Gender	Affiliation	Nationality
Hennan, Adel	Fish REA Diver	Female	CREP/JIMAR	UK
McCoy, Kaylyn	Fish REA Diver	Female	CREP/JIMAR	USA
TBD	Fish REA Diver	TBD	PMNM	USA
Gorospe, Kelvin	Fish REA Diver	Male	CREP/JIMAR	USA
Giuseffi, Louise	Fish REA Diver	Female	SOD/PIFSC	USA
Swanson, Dione	Benthic REA Diver	Female	CREP/JIMAR	USA
TBD	Benthic REA Diver	TBD	PMNM	TBD
Bailey, Hatsue	Benthic REA Diver	Female	CREP/JIMAR	JPN (USA Perm. Resident
Gray, Andrew	Fish REA/Tow Diver	Male	CREP/JIMAR	USA
Suka, Rhonda	Fish REA/Tow Diver	Female	CREP/JIMAR	USA
Schumacher, Brett	Benthic REA Diver	Female	CREP/JIMAR	USA
Garriques, Joao	Benthic REA/Tow Diver	Male	CREP/JIMAR	USA
Morioka, James	Oceanography Diver	Male	CREP/JIMAR	USA
Oliver, Thomas	Oceanography Diver	Male	CREP/JIMAR	USA
Clark, Jeannette	Oceanography Diver	Female	CREP/JIMAR	USA
Reardon, Kerry	ARMS Diver	Female	CREP/JIMAR	USA
Barba, Evan	ARMS Diver	Female	HIMB	USA
Sullivan, Chris	Mosaic Diver	Male	SIO	USA
Little, Mark	Microbial Biologist	Male	SDSU	USA
Akridge, Michael	Data Manager	Male	CREP/JIMAR	USA
Mahaffey, Katie	Chamber Operator/ Dive Master	Female	NDP	USA
TBD	TBD	TBD	TBD	TBD

G. Administrative

1. **Points of Contacts:**

Chief Scientists*:

Dr. Bernardo Vargas-Ángel, (Legs I, II) Bernardo. Vargas Angel@noaa.gov **NOAA IRC** Attn: NMFS / PIFSC / CREP 1845 Wasp Blvd, Building 176, Honolulu, HI 96818 808-725-5423

Dr. Brett Schumacher (Leg III) Brett.Schumacher@noaa.gov **NOAA IRC** Attn: NMFS / PIFSC / CREP 1845 Wasp Blvd, Building 176, Honolulu, HI 96818 808-725-5405

Project Operations Lead*:

Joao Garriques (Legs I, II, III) Joao.Garriques@noaa.gov **NOAA IRC** Attn: NMFS / PIFSC / CREP 1845 Wasp Blvd, Building 176, Honolulu, HI 96818 808-725-5407

Ship Operations Officer:

LT Faith O. Knighton OPS.Hiialakai@noaa.gov Faith.Knighton@noaa.gov NOAA Ship Hi'ialakai 1897 Ranger Loop, Building 184, Honolulu, HI 96818 In-Port: 808-725-5780

At Sea: 808-684-3235

* The Leg I Chief Scientist and Project Operations Lead are the designated program points of contact for all project planning and pre-departure correspondence with the ship.

2. **Diplomatic Clearances**

This project involves Marine Scientific Research in waters under the jurisdiction of the United States. Diplomatic clearance is not applicable.

3. Licenses and Permits

Activities to be conducted during HA-16-06 have been authorized under the following permits:

- State of Hawai'i Department of Land and Natural Resources, Division of Aquatic Resources, Special Activity Permit No. [Pending] issued for 2016 MHI operations. Permittee: Dr. Russell Brainard (Program Manager, NOAA PIFSC CREP).
- Scientific Research Permit No. [Pending] issued by the Papahānaumokuākea Marine National Monument.
 - NOAA ship *Hi'ialakai* Conservation and Management Permit PMNM-2016-006 issued by PMNM to Commanding Officer CDR Elizabeth I. Kretovic, NOAA ship *Hi'ialakai*.
- c. Department of the Army Nationwide Permit Verification File No: POH-2009-00083 (issued: 18 March 2014; expiration: 18 March 2017). The installation and maintenance of scientific measurement devices associated with the Pacific RAMP project is currently authorized under U.S. Army Corps of Engineers (USACE) Nationwide Permit No. 5, Scientific Measurement Devices (authorization date: March 2014) and is authorized until modified, revoked, or reissued by USACE.
- d. National Environmental Policy Act (NEPA), Programmatic Environmental Assessment for Research Activities Conducted by the Coral Reef Ecosystem Division, PIFSC, 2010-2015*. Finding of No Significant Impact (FONSI) signed 7 May 2010. (http://www.pifsc.noaa.gov/nepa/CRED_Programmatic%20Environmental%20Assessment_Final.pdf) *It is noted that while this NEPA document was originally intended to carry out program-level research activities for a period of five years, the analysis in the EA was intended to span an indefinite period of time, and is not limited to a five year period (see section 2.3 in EA).

A copy of these research permits will be provided to the Command prior to commencing scientific operations.

II. Operations

The Chief Scientist is responsible for ensuring the scientific staff are trained in planned operations and are knowledgeable of project objectives and priorities. The Commanding Officer is responsible for ensuring all operations conform to the ship's accepted practices and procedures.

A. Project Itinerary

Weather, equipment failures, and scheduling problems are unpredictable. As such, the following intended itinerary should be considered as only a guide for survey progression. In particular, the order in which survey areas are worked within a single island area or among islands within close proximity may be altered as appropriate based on weather, sea conditions, or the progress of the survey. Transit estimates have been calculated based on a ship's speed of 9.7 knots westbound, 9.2 knots north and southbound, and 8.7 knots eastbound.

Pre-Project

Jul 6-8

Loading of 10' ARMS Lab container, boat cradle and chest freezer with Navy crane. Small boat fuel (gasoline). Conduct small boat and davit familiarization for scientific personnel. Conduct station walk-throughs and dive neurological examinations for scientific personnel joining on Leg I and on later legs.

Leg I:

Jul 12

Depart Pearl Harbor: Embark full scientific complement at Ford Island. Depart the NOAA pier and being transit to Maui County (~50nm, 6h). Conduct remaining dive stations and small boat walkthroughs.

Jul 13-22

Maui County: Arrive Maui, commence full day of scientific operations. Operations include: fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 14 STRs and 85 CAUs, as well as a temporary deployment [up to 48 hours] of an anchored array consisting of a PUC, ADCP, CTP, and thermistor string; retrieve: 14 STRs, 85 CAUs, and the temporarily deployed anchored array), collection of up to six 20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 20 Plankton tows will be collected. Night operations include shipboard ADCP transects each night around Maui County. Anticipated order of survey polygons: MAI D, LAN B, MAI A, MAI B MOL A, MOL C, MOL B, LAN A, KAH A, KAH B. Upon securing operation on July 22, depart Maui County transit to Kaua'i (~175 nm, 18h).

Jul 23

Continue transit. Arrive Kaua'i County. Based on dive tempo the day may be utilized as the rest day.

Jul 24-29

Kaua'i County: Commence scientific operations, operations include: fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 12 STRs and 20 CAUs, 3 anchors, 1 EAR, as well as a temporary deployment [up to 48 hours] of an anchored array

consisting of a PUC, ADCP, CTP, and thermistor string; retrieve: 12 STRs, 20 CAUs, and the temporarily deployed anchored array), collection of up to six 20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 12 plankton tows will be collected. Night operations include shipboard ADCP transects each night around Kaua'i County. Anticipated order of survey polygons: KAU_B, KAU_D, KAU_01, NII_A, NII_B. Personnel drop off of Kristin Raja will occur when logistically feasible at Kaua'i Island via small boat. Upon securing operation on July 29, depart Kaua'i and transit to O'ahu (~120 nm, 14h).

Jul 30

Arrive O'ahu. To effectively utilize this day at sea, the Chief Scientist may request the ship to facilitate additional small boat training for program coxswains. End Leg I.

Jul 31- Aug 4

In-Port, Pearl Harbor: Resupply small boat fuel and loading of ethanol.

Leg II:

Aug 5

Pearl Harbor: Embark full scientific complement at Ford Island. Depart Pier and transit to fuel pier. Upon completion of fueling, transit to survey polygon HAW_A. North tip of Hawai'i Island (~150 nm, 17h). Complete remaining neuros and dive stations and small boat walk-throughs.

Aug 6-15

Hawai'i Island: Arrive Hawai'i County, anticipated half day of dive operations. Operations include fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 9 STRs, 50 CAUs, 20 BMUs, 12 ARMS; retrieve: 9 STRs, one anchor, 50 CAUs, 50 BMUs, and 12 ARMS), collection of up to six 20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 20 plankton tows will be collected. Night operations include shipboard ADCP transects each night around Hawai'i Island. Anticipated order of survey polygons: HAW_A, HAW_B, HAW_C, HAW_D, HAW_E, HAW_F, HAW_G. Upon securing operation on Aug 15, depart Hawai'i island and transit to O'ahu (~200 nm, 20h).

Aug 16

Continue Transit, arrive O'ahu. Based on dive tempo the day may be utilized as the rest day. Conduct personnel transfers via small boat.

Aug 17 - 23

O'ahu: Commence scientific operations, operations include fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 15 STRs, 65 CAUs, 25 BMUs, 15 ARMS; retrieve: 14 STRs, one SST buoy, one anchor, 60 CAUs, and 12 ARMS), collection of up to six

20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 16 tows will be collected. Night operations include shipboard ADCP transects each night around O'ahu. Anticipated order of survey polygons: OAH_B, OAH_A, OAH_E, OAH_D, OAH_C.

Aug 24

Conduct partial day of scientific operations in the general vicinity of Pearl Harbor prior to pulling into port (small boats could meet the ship alongside the pier). End leg II

Aug 25-28

In-Port, Pearl Harbor: Neurological exams and station walkthrough for new divers. Resupply small boat fuel. Conduct required Hull Inspection before operating in the PMNM.

Leg III:

Aug 29

Pearl Harbor: Embark full scientific complement at Ford Island. Depart Pier and transit to French Frigate Shoals (~490 nm, 2d 3h).

Aug 30

Transit

Aug 31-Sep 6

French Frigate Shoals (FFS): Arrive at FFS, commence partial day of scientific operations. Operations include fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 16 STRs, 50 CAUs, 20 BMUs, 12 ARMS; retrieve: 16 STRs, 2 anchors, 1 EARS, 50 CAUs, and 12 ARMS), collection of up to six 20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 12 plankton tows will be collected. Night operations include shipboard ADCP transects each night around FFS. Upon securing operation on Sep 6, depart FFS and transit to Lisianski Island (LIS) (~455 nm, 1d 16h).

Sep 7-8

Transit

Sep 9-13

Lisianski Island (LIS): Arrive at LIS, commencefull a day of scientific operations. Operations include: fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 15 STRs and 55 CAUs, as well as a temporary deployment [up to 48 hours] of an anchored array consisting of a PUC, ADCP, CTP, and thermistor string; retrieve: 15 STRs, 55 CAUs, 3 anchors, 1 EAR, and the temporarily deployed anchored array), collection of up to six 20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 10 plankton tow will be collected. Night operations include shipboard ADCP transects each night around Lisianski Island. Upon securing

operation on Sep 13, depart LIS and transit to Pearl and Hermes Atoll (~ 150 nm, 16h).

Sept 14

Transit to Pear and Hermes Atoll, this will be used as a rest day

Sep 15-20

Pearl and Hermes Atoll (PHR): Arrive at PHR, commence full day of scientific operations. Operations include fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 17 STRs, 60 CAUs, 20 BMUs, 12 ARMS; retrieve: 17 STRs, 5 anchors, 60 CAUs, and 12 ARMS), collection of up to six 20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 12 plankton tows will be collected. Night operations include shipboard ADCP transects each night around PHR. Upon securing operation on Sep 20, depart PHR and transit to Kure Atoll (~150 nm, 16h).

Sep 21-24

Kure Atoll (KUR): Arrive at KUR, commence full day of scientific operations. Operations include: fish and benthic Rapid Ecological Assessment (REA) surveys, towed-diver surveys, moored instrument deployments and retrievals (deploy: 15 STRs and 65 CAUs, as well as a temporary deployment [up to 48 hours] of an anchored array consisting of a PUC, ADCP, CTP, and thermistor string; retrieve: 15 STRs, 65 CAUs, 5 anchors, 1 EAR and the temporarily deployed anchored array), collection of up to six 20-40 cm coral cores as well as collection of carbonate chemistry water samples and small boat CTD casts at all CAU and coral coring sites, and microbial collections at one to two sites per day. A total of 8 Plankton tows will be collected. Night operations include shipboard ADCP transects each night around Kure Atoll. Upon securing operation on Sep 24, depart KUR and transit to Pearl Harbor (1,240 nm, 5d 22 h)

Sep 25-29

Transit

Sep 30

Pearl Harbor: Arrive Ford Island, Pearl Harbor. End of Project.

B. Staging and Destaging

Staging: Staging of large scientific gear and equipment will begin the week of July 4th, or as otherwise coordinated with the Command. Assistance from the ship's personnel for craning aboard large gear and for loading small boat gasoline (delivered by truck and pumped directly into the ship's hip tanks) will be necessary. Ethyl alcohol will be delivered in 5 gallon carboys to be transferred to the ship's quick release rack. Hand carried items will be loaded in lab areas throughout the week prior to departure. All scientists anticipate embarking the vessel at Ford Island, Pearl Harbor, as per ship's sailing board, on July 12, 2016, and will be aboard for the fueling. The ship itself will not return to the pier prior to beginning transit to Maui County.

<u>Mid-project Refueling</u>: Replenishment of unleaded gasoline will be required in Ford Island, Pearl Harbor during all scheduled in-ports. The fuel will be delivered by truck to be pumped directly into the ship's deck tanks. Re-fueling of *Hi'ialakai* will occur the start of Leg II.

<u>Destaging</u>: Full off-load of all program-provided gear and small boats will begin in coordination with the Command upon return to Pearl Harbor, September 30. The program boats and cradles will be priority items for offload as they will be needed for a subsequent project on another ship.

C. Operations to be Conducted

The Chief Scientist has the authority to revise or alter the technical portion of the instructions as work progresses, provided that, after consultation with the Commanding Officer, it is ascertained that the proposed changes will not (1) jeopardize the safety of personnel or the ship, (2) exceed the overall time allotted for the project, (3) result in undue additional expenses, and (4) alter the general intent of the project instructions. In addition, the Chief Scientist must notify the Office of the Science Director of the Pacific Islands Fisheries Science Center at the earliest opportunity prior to making (1) deviations from the general project track or area of operations noted in the project instructions, (2) changes or additions of research operations to those specified in the project instructions, or (3) port calls not specifically identified in the project instructions.

Scientific Operations

The ship will support assessment and monitoring operations within the project Operations Area. Specifically, the ship will support Rapid Ecological Assessments and the continuation of long-term monitoring of reef fish, corals, other invertebrates, and algae, and oceanographic monitoring of the coral reef ecosystems of the region.

Research and monitoring efforts will require extensive diving operations (both scuba and snorkeling) to be supported by *Hi'ialakai*. Up to five small boats will be operating simultaneously during daylight hours to maximize productivity. *Hi'ialakai*'s 29-ft Metal Shark (HI-1), 26-ft Ambar (HI-2), and 17-ft Northwind (HI-3), as well as two program-provided 19-ft SAFE Boats will be required to support the REA, towed diver, instrumentation, and microbial survey teams on a daily basis.

The REA and towed-diver surveys will include monitoring of species composition, abundance, size distribution, and spatial distribution of reef fishes, corals, other invertebrates, and algae of the region and will further ground-truth shallow-water benthic habitat maps. Benthic community structure and demography parameters, including percent cover, taxonomic richness, density, size-class distribution, and health condition, will be assessed along line and belt transects to provide spatial-temporal appraisals of coral reef dynamics in the region. Scientists may also obtain limited specimens of algae, corals, and other invertebrates. All deployments, retrievals, and specimen collections will be conducted as stipulated through pertinent Agency-approved Special Activity Permit guidelines.

A number of Autonomous Reef Monitoring Structures (ARMS) will be recovered and replaced or newly installed at select locations by the Ocean and Climate Change Team.

These small units are designed to mimic the complexity of the coral reef structure and to attract small invertebrates to recruit within them. ARMS samples will be processed onboard in a program-provided lab space, preserved in ethanol, and stored in the scientific freezer until the ship returns to Pearl Harbor.

Calcification Accretion Units (CAUs) will be recovered and replaced or newly installed at select locations. The CAUs serve as a mechanism to quantify accretion rates of calcifying coralline algae and scleractinian corals; this information will allow for comparisons to determine possible consequences of increased ocean acidity and lowered aragonite saturation states. Additionally, a Bioerosion Monitoring Unit (BMU) will be installed on CAUs at select locations. Each BMU is made of a 1cm x 2cm x 5cm block of calcium carbonate rock mounted on a similarly sized piece of polyvinylchloride plate. The BMUs rest flush on the seafloor and serve as a mechanism to quantify bioerosion rates across coral reefs experiencing different physical oceanography and local human impacts. BMUs will be retrieved on a subsequent mission and measured for changes in weight and scanned for changes in density using microcomputed tomography.

Approximately 123 STRs, 445 CAUs, 48 ARMS, and 85 BMUs will be deployed and 123 STRs, 445 CAUs, 48 ARMS, 28 anchors, 1 SST buoy, and 5 EARs will be retrieved. In addition, the following instruments will be deployed and retrieved during the same project: eight moored ADCPs deployed and retrieved within one month and nine anchored arrays consisting of a PUC, ADCP, CTP, and thermistor string deployed and retrieved within 48 hours. Retrieved anchors will be secured on stackable aluminum pallets for transport back to Pearl Harbor.

As part of CREP's effort to investigate ocean acidification and carbonate chemistry, the Ocean and Climate Change Team may also obtain coral cores at several locations, if time and operational conditions allow. Up to three cores may be collected with a diver-held pneumatic drill at each given site. Each core would measure 20-40 cm in length and 3.8 cm in diameter. Locations for coring will not be planned in advance. Coring operations will be conducted opportunistically as appropriate, and as a scientific dive. In addition, underway shipboard oceanographic measurements (ADCP, TSG, and SCS) will be recorded throughout the duration of the project and water samples may be collected from small boats for nutrient, chlorophyll, salinity, and carbonate chemistry analyses.

As part of the ongoing effort to understand the microbial community, two types of water samples will be collected at select REA sites using diver-deployable Niskin bottles (four bottles; two liters per bottle). Two of the Niskin bottles will be filled with water collected from approximately one meter above the benthos, and two will be filled with water from within the reef (pore-water). The pore-water samples will be collected using Niskin bottles with a flexible stainless-steel hose attachment. These water samples will be returned to the ship and processed for dissolved organic carbon (DOC), particulate organic matter (POM), nutrients, microbial (Bacteria and Archaea) and viral abundance (fluorescent microscopy), FACS (heterotrophs vs. autotrophs), and microbial and viral community composition (coarse analysis: 16s rRNA). At two REA sites per island, approximately 60-80 L of reef water will be collected from reef crevices and surfaces for metagenomic analysis of the microbial and viral community associated with the reef benthos.

In addition to understanding water-column microbial dynamics, investigating shifts in the microbial community associated with benthic composition is important as it can serve as

an indicator of reef-ecosystem health. If time permits, six fist-sized samples of rubble (three of these will also contain a handful of the first 5-10 cm of sediment from different sand pockets), and six pieces of the most dominant algae-type will be collected in zip-top bags. Both the algae and the rubble/sediment samples will be frozen at -20°C. These samples will remain on the ship until it returns to Honolulu. The 16s bacterial rRNA genes associated with these samples will be sequenced to characterize the microbial communities associated with the benthos (rubble and algae).

Coral reef biodiversity remains greatly under described, especially with regards to the smaller invertebrate animals that reside in and on the reef. Many of these organisms have planktonic larvae that can be collected and thus their occurrence documented via plankton samples. A plankton net 50 cm diameter with a 80 µm mesh size having a 1 L cod end jar attached to net with flow meter will be trailing a few yards behind the stern of a small boat. Each tow will be assigned a unique station number with corresponding GPS coordinates, date & time recorded for start and finish of each tow. The tows will be just below the surface, for 5 minutes and the small boat will be going just fast enough for net to be tight and flow meter to run (1–2 knots). The net will be brought back onto the small boat and processed in the field. The outside of the plankton net will be washed down with seawater applied to outside collecting flora and fauna in the cod end. Depending on the number of days spent at each island, we expect to conduct 5–20 plankton tows per island, for a maximum of 110 plankton tow samples for the entire cruise. This project is led by Gustav Pauli at the Florida Museum of Natural History.

Snorkeling Operations

All snorkeling shall be conducted in accordance with the NOAA Scientific Diving Manual (Section 4.13).

Small Boat Operations

Per OMAO Supplement to the NOAA Small Boat Standards and Procedures Manual, March 2010, Section 4.03a2, a program certified Operator in Charge (OIC) must "earn the full confidence of both the Commanding Officer (CO) and Designated Examiner (DE) and has successfully completed the shipboard training requirements." As part of any OIC evaluation, it is understood that a small boat OIC will be designated to accompany and evaluate an OIC-in-training. This may limit the number of small boats the ship can deploy during this evaluation period, but every effort will be made to limit any impact to operations. An OIC-in-training is not guaranteed to be qualified by the CO and DE during a project.

Small boat deployment and recovery operations from a ship at sea are inherently dangerous. Experience levels of all personnel involved and environmental conditions are limiting factors regarding the decision to proceed with said operations. Proficiency levels of deck officers, deck department, or small boat crews may impact operations. All small boat crewmembers must have the full confidence of the CO and DE. At any time, the CO may call for a halt to boat deployment and recovery operations. If indicated, a Safety Stand Down, extra training or practice may be required to begin operations again. This is especially true when the ship has been in port or when program personnel have not been aboard for an extended period of time, as well as when boat operations are called for within 48 hours of departure.

While minimizing impact to science operations, ship's diver/coxswain training and proficiency regulations may require the use of a ship's small boat during an extended project. The CO will work with the Chief Scientist to plan and minimize impacts to fulfill such requirements.

CTD Operations

No shipboard CTD operations are required under HARAMP 2016. However, should multibeam mapping be undertaken as an Ancillary Project by *Hi'ialakai*, CTD casts for the purposes of calculating sound velocity profiles may need to be conducted. Such surveying would be undertaken on a not-to-interfere basis with the ship's primary mission.

D. Dive Plan

All dives are to be conducted in accordance with the requirements and regulations of the NOAA Diving Program (http://www.ndc.noaa.gov/dr.html) and require the approval of the ship's Commanding Officer.

The Dive Plans encompassing Legs I – III of HA-16-06 are presented in *Appendix 11* (Attached file).

E. Applicable Restrictions

Conditions which preclude normal operations: Poor weather and sea conditions, equipment failure, safety concerns, and/or unforeseen circumstances, may alter or prohibit operations as planned. At these times, the Chief Scientist and Commanding Officer will determine the appropriate plan of action.

NMFS employees are not exempt from the requirements of the Marine Mammal Protection Act (MMPA) or the Endangered Species Act (ESA). PIFSC has developed mitigation measures for its fisheries and ecosystem research projects to avoid take and to comply with the Lecky, Murawski, and Merrick guidance. A copy of these documents is available at https://sites.google.com/a/noaa.gov/pifsc-science-operations/nepa-permits/protected-species-mitigation-measures and on the ship's bridge.

1. "Take" of Protected Species

- a. Under the MMPA and ESA it is unlawful to take a protected species. The MMPA defines take as "harass, hunt, capture, kill or collect, or attempt to harass, hunt, capture, kill or collect." The ESA defines take as "to harass, harm, pursue, hunt, shoot, wound, kill, trap, capture, or collect, or to attempt to engage in any such conduct." An incidental take is one that is incidental to, but not the purpose of, otherwise lawful activities.
- b. In the event of an incidental take of a marine mammal or federally listed threatened or endangered species during the project, the Chief Scientist will report the incident to the ship's Commanding Officer then the PIFSC Director and Deputy Director IMMEDIATELY via IRIDIUM,

INMARSAT, and email. Samples should not be collected from any incidentally taken marine mammals, sea turtles, or seabirds.

- 2. Activities in the Hawaiian Islands Humpback Whale National Marine Sanctuary
 - a. The humpback whale season in Hawai'i is November through May. Unless otherwise authorized under the MMPA and ESA, it is unlawful to approach, or cause a vessel or other object to approach, within 100 yards of any humpback whale within the Sanctuary. Please reference the complete list of prohibited activities and boundary maps at https://sites.google.com/a/noaa.gov/pifsc-science-operations/nepa-permits/protected-species-mitigation-measures. A copy of these materials will also be available on the ship's bridge.

III. Equipment

A. Equipment and Capabilities provided by the ship (itemized)

1. Equipment: To successfully meet the project objectives, the scientific compliment aboard will need the ship to provide the items listed below. Prior to sailing, the ship's crew will inspect these items to ensure they are in proper working order for the project:

J-frame

A-frame

Aft deck crane

30-ft Metal Shark launch, HI-1

26-ft Ambar launch, HI-2

17-ft Northwind launch, HI-3

SCUBA compressor (Nitrox and Air)

Recompression chamber

Dive lockers

Stainless Steel Lockers (4)

Scientific freezer with shelving

Wet Lab faucets and drains

Acoustic Doppler Current Profiler (ADCP)

Scientific Computer System (SCS)

ThermoSalinoGraph (TSG)

Sea Surface Sound Velocity (SSSV)

Operational multibeam echo sounder

CTD (Would only be required if multibeam echo sounder project is possible; See

"Additional Projects" section below.)

Adequate fresh water for gear and small boat wash-down

Iridium phone

VHF radios for ship's small boats

Global Positioning System (GPS) for ship's small boats

Depth sounders for ship's small boats

Gasoline storage tanks

Sufficient HAZMAT storage for 240 L gallons of ethanol

2. Capabilities: It is requested that the ship provide the following:

- a. Assistance from the Command and ship's Deck Department in conducting davit hook checkouts for program personnel prior to departure of HA-16-06.
- b. Permission for Scientists to ready scientific work spaces (e.g., set up computer server) during the week prior to departure.
- c. Assistance from the ship's Deck Department in craning and staging large gear during loading and off-loading.
- d. Support from the Engineering and Deck departments prior to sailing to transfer 3.78 kL (1000 gallons) of program-provided gasoline into the ship's stainless steel fuel tanks to be used as outboard engine fuel. The gasoline will be delivered by truck to be pumped directly into the deck tanks.
- e. Support from the Engineering and Deck departments prior to sailing to transfer 190L of program-provided ethyl alcohol into the port fantail releasable rack. Additional storage capacity for 4 standard 19L (5 gal) carboys of waste ethanol generated during the project is necessary.
- f. Hi'ialakai's HI-1 (Metal Shark), HI-2, and HI-3 will be required to support the program's dive teams on a daily basis. The ship should plan to provide coxswains for HI-1 (Metal Shark), HI-2, and HI-3 during all days of diving operations.
- g. Support from the medical officer and deck department to conduct neurological exams, boat familiarizations and station walk-throughs prior departure (as appropriate) and for new divers each leg.
- h. Mid-project support from the engineering department is requested to assist in the transfer of additional unleaded gasoline during all Pearl Harbor in-ports. *Hi'ialakai* will be responsible for providing diesel fuel for HI-1 and HI-2.
- i. Operable Wet Lab facilities are necessary to support water sampling and the cleaning of field equipment. Approximately 16 0.9 m x 0.6 m totes of the program's glass sample bottles and plastic sample bags filled with seawater will need to be stored onboard in an air conditioned space.
- To support the ARMS Lab (10-ft container box), working power, freshwater, saltwater and compressed air sources and connections on the fantail will be necessary.
- k. To support CAU processing, deck power connection for one chest freezer (~ 6 ft. x 3 ft x 3 ft) will be necessary. Note: Similar program freezers were carried previously on the bridge deck (starboard rail) and on the boat deck (port side).
- 1. To be consistent with the mission objectives, the ship and its compliment of small boats will employ all methods feasible to minimize damage to coral reef habitats during any anchoring operations that may be required.

m. Provide necessary dive support towards the required Hull Inspection that needs to be performed prior to departure to the Papahānaumokuākea Marine National Monument.

B. Equipment and Capabilities provided by the scientists (itemized)

- **1. Equipment:** The program's full equipment list is presented in *Appendix 12* (Attached file).
- **2. Capabilities:** In addition to scientific expertise, the program will provide coxswains and routine maintenance for program provided boats.

IV. Hazardous Materials

A. Policy and Compliance

The Chief Scientist is responsible for complying with FEC 07 Hazardous Materials and Hazardous Waste Management Requirements for Visiting Scientific Parties (or the OMAO procedure that supersedes it). By Federal regulations and NOAA Marine and Aviation Operations policy, the ship may not sail without a complete inventory of all hazardous materials by name and quantity, MSDS, appropriate spill cleanup materials (neutralizing agents, buffers, or absorbents) in amounts adequate to address spills of a size equal to the amount of chemical brought aboard, and chemical safety and spill response procedures. Documentation regarding those requirements will be provided by the Chief of Operations, Marine Operations Center, upon request.

Per OMAO procedure, the scientific party will include with their project instructions and provide to the CO of the respective ship 30 days before departure:

- List of chemicals by name with anticipated quantity
- List of spill response materials, including neutralizing agents, buffers, and absorbents
- Chemical safety and spill response procedures, such as excerpts of the program's Chemical Hygiene Plan or SOPs relevant for shipboard laboratories
- For bulk quantities of chemicals in excess of 50 gallons total or in containers larger than 10 gallons each, notify ship's Operations Officer regarding quantity, packaging and chemical to verify safe stowage is available as soon as chemical quantities are known.

Upon embarkation and prior to loading hazardous materials aboard the vessel, the scientific party will provide to the CO or their designee:

- An inventory list showing actual amount of hazardous material brought aboard
- An MSDS for each material
- Confirmation that neutralizing agents and spill equipment were brought aboard sufficient to contain and cleanup all of the hazardous material brought aboard by the program

Confirmation that chemical safety and spill response procedures were brought aboard

Upon departure from the ship, scientific parties will provide the CO or their designee an inventory showing that all chemicals were removed from the vessel. The CO's designee will maintain a log to track scientific party hazardous materials. MSDS will be made available to the ship's complement, in compliance with Hazard Communication Laws.

Scientific parties are expected to manage and respond to spills of scientific hazardous materials. Overboard discharge of hazardous materials is not permitted aboard NOAA ships.

B. Inventory of Hazardous Materials

See Appendix 13.

C. Chemical safety and spill response procedures

See Appendix 14.

D. Radioactive Materials

No Radioactive Isotopes are planned for this project.

V. Additional Projects

Supplementary (piggyback) and ancillary projects are secondary to the objectives of the project and should be treated as additional investigations. The difference between the two types of secondary projects is that an ancillary project does not have representation aboard and is accomplished by the ship's force.

A. Supplementary ("Piggyback") Projects

No Supplementary Projects are planned

B. NOAA Fleet Ancillary Projects

Collection of multibeam echosounder data: This ancillary project requires the assistance of the ship's Survey Technician, if aboard, or similarly trained individual, to run the multibeam surveying operation. Although extensive multibeam surveying has been conducted throughout the Pacific Islands Region, gaps in coverage remain that preclude characterizing the suitability of uncovered areas as habitat for deep sea corals, and may limit the ability to effectively survey and study these areas with other technologies. Using multibeam bathymetry data supplied by the Pacific Islands Fisheries Science Center/Coral Reef Ecosystem Program, services of the ship's Survey Technician or other trained shipboard personnel are requested to identify data gaps that are within range of shipboard multibeam echosounders and conduct survey operations to collect multibeam data to fill them. Multibeam data should be collected following standard practices for insuring high quality data, including collecting sound velocity profiles with

the ship's CTD and applying them before beginning a multibeam survey session, and every six hours thereafter, or more frequently if required. Surveying will be conducted on a not-to-interfere basis with the ship's primary mission. Data collected, including both multibeam and sound velocity data, should be provided to the Chief Scientist at the conclusion of the project using a program supplied hard drive and will include information on all system specifications, settings and offsets required to enable the development of metadata records meeting ISO 1911 standards.

Surveying should prioritize the shallowest gaps first and work progressively deeper. Depths as shallow as the ship is able to safely navigate may contain black corals, wire corals or other azoothanthellate coral species and are a high priority. It is anticipated that the NOAA Ship *Okeanos Explorer* will conduct multibeam surveying at depths below 500 m throughout the region between FY15 – FY17, so gaps at those depths should receive the lowest priority. However, if gaps shallower than 500 m have been filled and survey time is available, gaps between 500 m and 3000 m those deeper areas should also be surveyed.

VI. Disposition of Data and Reports

The project will follow the current PIFSC and CRED data management plans, both of which comply with NOAA requirements. Disposition of data gathered aboard NOAA ships will conform to NAO 216-101 *Ocean Data Acquisitions* and NAO 212-15 *Management of Environmental Data and Information*. To guide the implementation of these NAOs, NOAA's Environmental Data Management Committee (EDMC) provides the *NOAA Data Documentation Procedural Directive* (data documentation) and *NOAA Data Management Planning Procedural Directive* (preparation of Data Management Plans). OMAO is developing procedures and allocating resources to manage OMAO data and Programs are encouraged to do the same for their Project data.

A. Data Classifications: Under Development

- 1. OMAO Data
- 2. Program Data *Under Development*. Contact Nori Shoji (<u>noriko.shoji@noaa.gov</u>), Director, Science Operations Division, for PIFSC data policy updates.

B. Responsibilities: Under Development

Integrated ecosystem observations of coral reefs are collected to characterize the spatial and temporal variability of the distribution, abundance, and diversity of corals, algae, other macroinvertebrates, and fishes in the context of their benthic habitats and oceanographic environments. All data are quality assured, processed, and made available to region managers and stakeholders.

VII. Meetings, Vessel Familiarization, and Project Evaluations

A. <u>Pre-Project Meeting</u>: The Chief Scientist and Commanding Officer will conduct a meeting of pertinent members of the scientific party and ship's crew to discuss required equipment, planned operations, concerns, and establish mitigation strategies for all

concerns. This meeting shall be conducted before the beginning of the project with sufficient time to allow for preparation of the ship and project personnel. The ship's Operations Officer usually is delegated to assist the Chief Scientist in arranging this meeting.

- B. <u>Vessel Familiarization Meeting</u>: The Commanding Officer is responsible for ensuring scientific personnel are familiarized with applicable sections of the standing orders and vessel protocols, e.g., meals, watches, etiquette, drills, etc. A vessel familiarization meeting shall be conducted in the first 24 hours of the project's start and is normally presented by the ship's Operations Officer.
- C. Post-Project Meeting: The Commanding Officer is responsible for conducting a meeting no earlier than 24 hrs before or 7 days after the completion of a project to discuss the overall success and shortcomings of the project. Concerns regarding safety, efficiency, and suggestions for future improvements shall be discussed and mitigations for future projects will be documented for future use. This meeting shall be attended by the ship's officers, applicable crew, the Chief Scientist, and members of the scientific party and is normally arranged by the Operations Officer and Chief Scientist.
- Project Evaluation Report: Within seven days of the competition of project, a Customer Satisfaction Survey is to be completed by the Chief Scientist. The form is available at https://sites.google.com/a/noaa.gov/omao-intranet-dev/operations/marine/customer-satisfaction-survey and provides a "Submit" button at the end of the form. It is also located at https://docs.google.com/a/noaa.gov/forms/d/1a5hCCkgIwaSII4DmrHPudAehQ9HqhRqY3J_FXqbJp9g/viewform. Submitted form data is deposited into a spreadsheet used y OMAO management to analyze the information. Though the complete form is not shared with the ships, specific concerns and praises are followed up on while not divulging the identity of the evaluator.

VIII. Miscellaneous

A. Meals and Berthing

The ship will provide meals for the scientists listed above. Meals will be served 3 times daily beginning one hour before scheduled departure, extending throughout the project, and ending two hours after the termination of the project. In addition, packed lunches will be required on all full-day operations. Since the watch schedule is split between day and night, the night watch may often miss daytime meals and will require adequate food and beverages (for example a variety of sandwich items, cheeses, fruit, milk, juices) during what are not typically meal hours. Special dietary requirements for scientific participants will be made available to the ship's command at least seven days prior to the survey. Berthing requirements, including number and gender of the scientific party, will be provided to the ship by the Chief Scientist.

The Chief Scientist and Commanding Officer will work together on a detailed berthing plan to accommodate the gender mix of the scientific party taking into consideration the current make-up of the ship's complement. The Chief Scientist is responsible for ensuring the scientific berthing spaces are left in the condition in which they were received; for stripping bedding and linen return; and for the return of any room keys which were issued. The Chief Scientist is also responsible for the cleanliness of the laboratory spaces and the storage areas utilized by the scientific party, both during the project and at its conclusion prior to departing the ship.

All NOAA scientists will have proper travel orders when assigned to any NOAA ship. The Chief Scientist will ensure that all non NOAA or non Federal scientists aboard also have proper orders. It is the responsibility of the Chief Scientist to ensure that the entire scientific party has a mechanism in place to provide lodging and food and to be reimbursed for these costs in the event that the ship becomes uninhabitable and/or the galley is closed during any part of the scheduled project.

All persons boarding NOAA vessels give implied consent to comply with all safety and security policies and regulations which are administered by the Commanding Officer. All spaces and equipment on the vessel are subject to inspection or search at any time. All personnel must comply with OMAO's Drug and Alcohol Policy dated May 7, 1999 which forbids the possession and/or use of illegal drugs and alcohol aboard NOAA Vessels.

B. Medical Forms and Emergency Contacts

The NOAA Health Services Questionnaire (NHSQ, NF 57-10-01 (3-14)) must be completed in advance by each participating scientist. The NHSQ can be obtained from the Chief Scientist or the NOAA website http://www.corporateservices.noaa.gov/noaaforms/eforms/nf57-10-01.pdf.

All NHSQs submitted after March 1, 2014 must be accompanied by <u>NOAA Form (NF)</u> <u>57-10-02</u> - Tuberculosis Screening Document in compliance with <u>OMAO Policy 1008</u> (Tuberculosis Protection Program).

The completed forms should be sent to the Regional Director of Health Services at the applicable Marine Operations Center. The NHSQ and Tuberculosis Screening Document should reach the Health Services Office no later than 4 weeks prior to the start of the

project to allow time for the participant to obtain and submit additional information should health services require it, before clearance to sail can be granted. Please contact MOC Health Services with any questions regarding eligibility or completion of either form. Ensure to fully complete each form and indicate the ship or ships the participant will be sailing on. The participant will receive an email notice when medically cleared to sail if a legible email address is provided on the NHSQ.

The participant can mail, fax, or email the forms to the contact information below. Participants should take precautions to protect their Personally Identifiable Information (PII) and medical information and ensure all correspondence adheres to DOC guidance (http://ocio.os.doc.gov/ITPolicyandPrograms/IT_Privacy/PROD01_008240).

The only secure email process approved by NOAA is Accellion Secure File Transfer which requires the sender to setup an account. Accellion's Web Users Guide is a valuable aid in using this service, however to reduce cost the DOC contract doesn't provide for automatically issuing full functioning accounts. To receive access to a "Send Tab", after your Accellion account has been established send an email from the associated email account to accellionAlerts@doc.gov requesting access to the "Send Tab" function. They will notify you via email usually within 1 business day of your approval. The 'Send Tab" function will be accessible for 30 days.

Contact information:

Regional Director of Health Services Marine Operations Center – Pacific 2002 SE Marine Science Dr. Newport, OR 97365 Telephone <u>541-867-8822</u> Fax <u>541-867-8856</u> Email <u>MOP.Health-Services@noaa.gov</u>

Prior to departure, the Chief Scientist must provide an electronic listing of emergency contacts to the Executive Officer for all members of the scientific party, with the following information: contact name, address, relationship to member, and telephone number.

C. Shipboard Safety

Wearing open-toed footwear or shoes that do not completely enclose the foot (such as sandals or clogs) outside of private berthing areas is not permitted. Steel-toed shoes are required to participate in any work dealing with suspended loads, including CTD deployments and recovery. The ship does not provide steel-toed boots. Hard hats are also required when working with suspended loads. Work vests are required when working near open railings and during small boat launch and recovery operations. Hard hats and work vests will be provided by the ship when required.

All scientists will comply with standing safety regulations of PIFSC and that of the vessel's Standing Orders from the Commanding Officer:

E. Communications

A progress report on operations prepared by the Chief Scientist may be relayed to the program office. Sometimes it is necessary for the Chief Scientist to communicate with another vessel, aircraft, or shore facility. Through various means of communications, the ship can usually accommodate the Chief Scientist. Special radio voice communications requirements should be listed in the project instructions. The ship's primary means of communication with the Marine Operations Center is via e-mail and the Very Small Aperture Terminal (VSAT) link. Standard VSAT bandwidth at 128kbs is shared by all vessels staff and the science team at no charge. Increased bandwidth in 30 day increments is available on the VSAT systems at increased cost to the scientific party. If increased bandwidth is being considered, program accounting is required it must be arranged at least 30 days in advance.

IT Security

Any computer that will be hooked into the ship's network must comply with the *NMAO* Fleet IT Security Policy 1.1 (November 4, 2005) prior to establishing a direct connection to the NOAA WAN. Requirements include, but are not limited to:

- 1. Installation of the latest virus definition (.DAT) file on all systems and performance of a virus scan on each system.
- 2. Installation of the latest critical operating system security patches.
- 3. No external public Internet Service Provider (ISP) connections.
- 4. Provide the Electronics Technician with a spreadsheet of the following information:

Device	Name	Operating System	LAN MAC Address
Iphone	Scientist	MAC OS	21:34:6K:P8:W6:77
Laptop	Scientist	Windows XP	23:34:6K:P8:M6:77

Completion of these requirements prior to boarding the ship is required.

Non-NOAA personnel using the ship's computers or connecting their own computers to the ship's network must complete NOAA's IT Security Awareness Course prior to embarking. Arrangements to take the Course and/or achieve security clearance for any non-NOAA computers should be coordinated with PIFSC/CREP administration well in advance of the project.

F. Foreign National Guests Access to OMAO Facilities and Platforms

The foreign national participants for project HA-16-06 are Adel Heenan and no others. Katie Mahaffey will serve as the onboard foreign national sponsor for the participant.

All foreign national access to the vessel shall be in accordance with NAO 207-12 and RADM De Bow's March 16, 2006 memo (http://deemedexports.noaa.gov). National Marine Fisheries Service personnel will use the Foreign National Registration System (FRNS) to submit requests for access to NOAA facilities and ships. The Departmental Sponsor/NOAA (DSN) is responsible for obtaining clearances and export licenses and for providing escorts required by the NAO. DSNs should consult with their designated NMFS Deemed Exports point of contact to assist with the process.

The following are basic requirements. Full compliance with NAO 207-12 is required.

Responsibilities of the Chief Scientist:

- Provide the Commanding Officer with the e-mail generated by the FRNS granting approval for the foreign national guest's visit. This e-mail will identify the guest's DSN and will serve as evidence that the requirements of NAO 207-12 have been complied with.
- Escorts The Chief Scientist is responsible to provide escorts to comply with NAO 207-12 Section 5.10, or as required by the vessel's DOC/OSY Regional Security Officer.
- 3. Ensure all non-foreign national members of the scientific party receive the briefing on Espionage Indicators (NAO 207-12 Appendix A) at least annually or as required by the servicing Regional Security Officer.
- 4. Export Control Ensure that approved controls are in place for any technologies that are subject to Export Administration Regulations (EAR).

The Commanding Officer and the Chief Scientist will work together to implement any access controls necessary to ensure no unlicensed export occurs of any controlled technology onboard regardless of ownership.

Responsibilities of the Commanding Officer:

- 1. Ensure only those foreign nationals with DOC/OSY clearance are granted access.
- Deny access to OMAO platforms and facilities by foreign nationals from countries controlled for anti-terrorism (AT) reasons and individuals from Cuba or Iran without written NMAO approval and compliance with export and sanction regulations.
- 3. Ensure foreign national access is permitted only if unlicensed deemed export is not likely to occur.
- 4. Ensure receipt from the Chief Scientist or the DSN of the FRNS e-mail granting approval for the foreign national guest's visit.
- 5. Ensure Foreign Port Officials, e.g., Pilots, immigration officials, receive escorted access in accordance with maritime custom to facilitate the vessel's visit to foreign ports.
- 6. Export Control 8 weeks in advance of the project, provide the Chief Scientist with a current inventory of OMAO controlled technology onboard the vessel and a copy of the vessel Technology Access Control Plan (TACP). Also notify the Chief Scientist of any OMAO-sponsored foreign nationals that will be onboard while program equipment is aboard so that the Chief Scientist can take steps to prevent unlicensed export of Program controlled technology. The Commanding Officer and the Chief Scientist will work

- together to implement any access controls necessary to ensure no unlicensed export occurs of any controlled technology onboard regardless of ownership.
- 7. Ensure all OMAO personnel onboard receive the briefing on Espionage Indicators (NAO 207-12 Appendix A) at least annually or as required by the servicing Regional Security Officer.

Responsibilities of the Foreign National Sponsor:

- 1. Export Control The foreign national's sponsor is responsible for obtaining any required export licenses and complying with any conditions of those licenses prior to the foreign national being provided access to the controlled technology onboard regardless of the technology's ownership.
- The DSN of the foreign national shall assign an on-board Program
 individual, who will be responsible for the foreign national while on board.
 The identified individual must be a U.S. citizen, NOAA (or DOC) employee.
 According to DOC/OSY, this requirement cannot be altered.
- 3. Ensure completion and submission of Appendix C (Certification of Conditions and Responsibilities for a Foreign National Guest) as required by NAO 207-12 Section 5.03.h.

Appendices

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Appendix 1: Operating Area for HA-16-06 HARAMP
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Appendix 2: Maui County

Appendix 3: Kaua'i County

Appendix 4: Hawai'i Island

Appendix 5: O'ahu Island

Appendix 6: French Frigate Shoals

Appendix 7: Lisianski Island

Appendix 8: Pearl and Hermes Atoll

Appendix 9: Kure Atoll

Appendix 10: Station/Waypoint List (attached file; coordinates in Latitude, Longitude: decimal degrees)

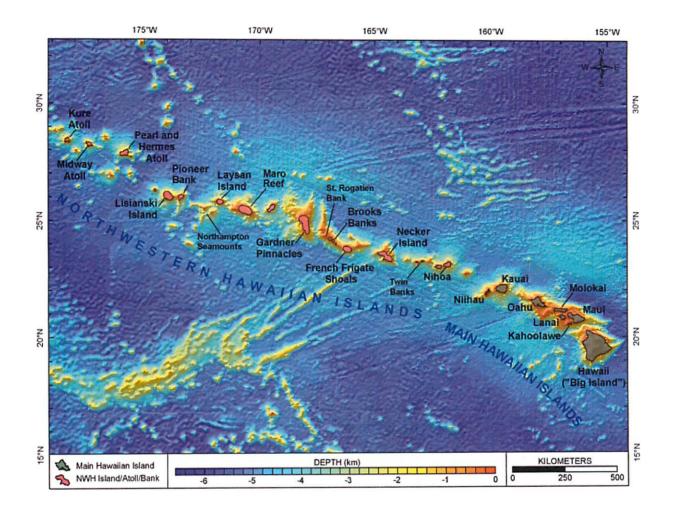
Appendix 11: Dive plans (attached file)

Appendix 12: Program Equipment List (attached file)

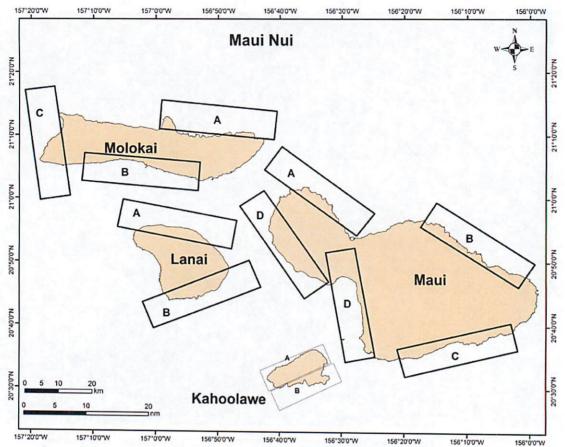
Appendix 13: Section IV.B. Hazardous Materials Inventory

Appendix 14: Section IV.C. Chemical Safety and Spill Response Procedures

Appendix 1: Operating Area for HA-16-06 HARAMP

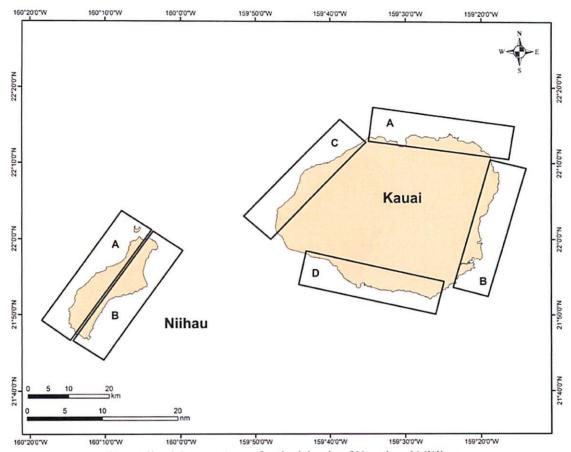


Appendix 2: Maui County



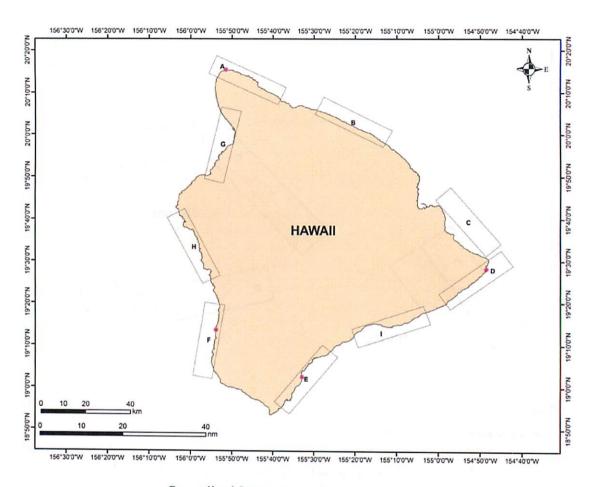
Generalized Survey Areas for the islands of Maui, Lāna'i, Moloka'i and Kaho'olawe

Appendix 3: Kaua'i County



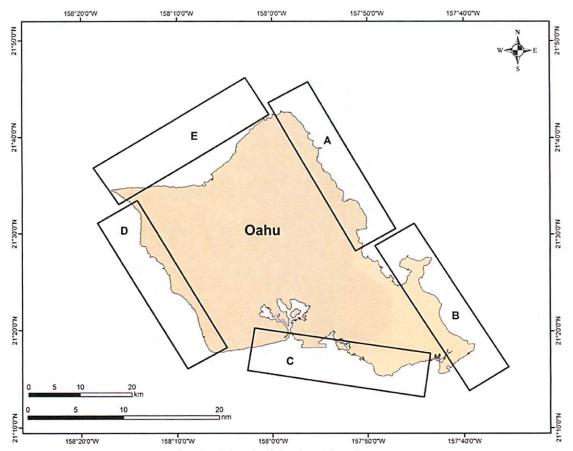
Generalized Survey Areas for the islands of Kauai and Ni'ihau

Appendix 4: Hawai'i Island



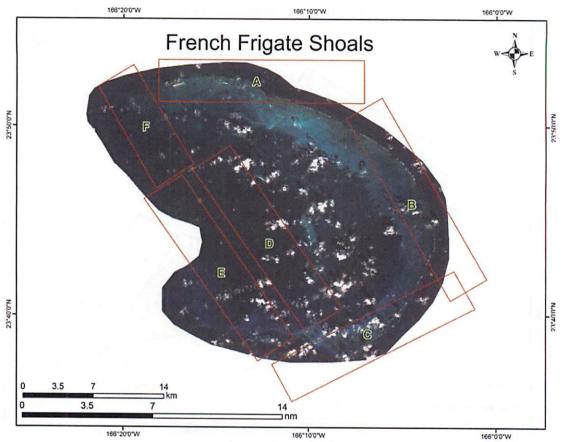
Generalized Survey Areas for Hawai'i Island

Appendix 5: O'ahu Island



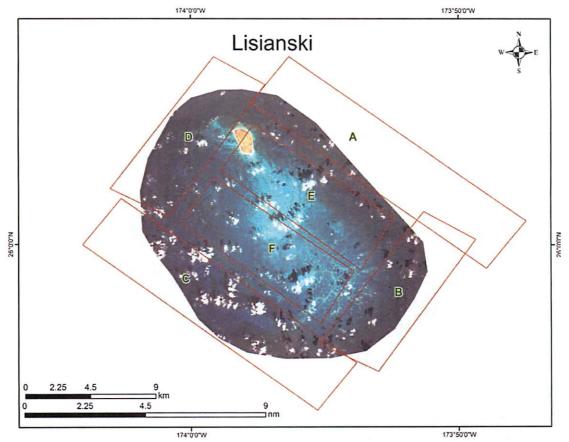
Generalized Survey Areas for the island of O'ahu Island Appendix 6: French Frigate Shoals

Appendix 6: French Frigate Shoals



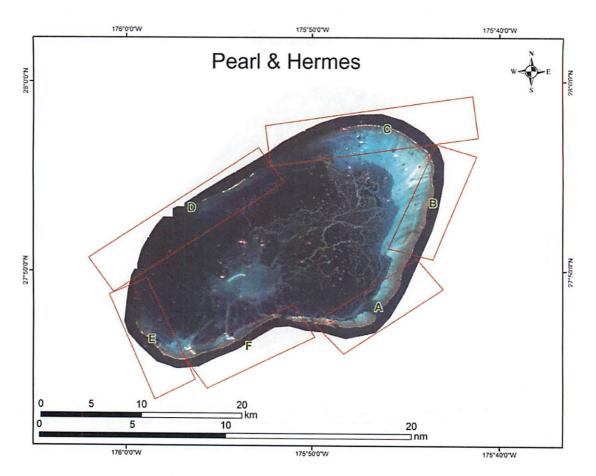
Generalized Map of French Frigate Shoals

Appendix 7: Lisianski Island



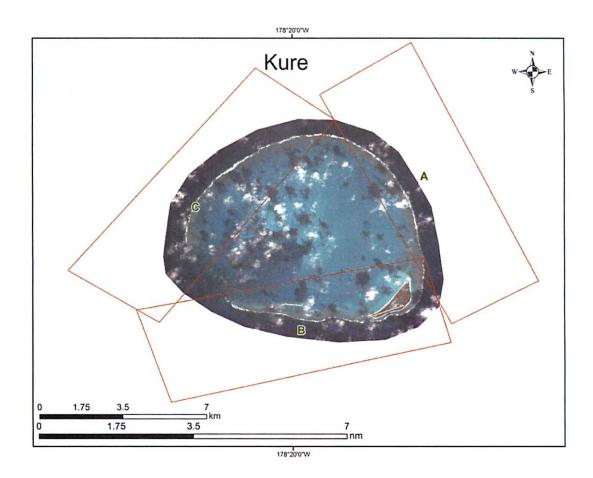
Generalized Map of Lisianski Island

Appendix 8: Pearl and Hermes Atoll



Generalized Map of Pearl and Hermes Atoll

Appendix 9: Kure Atoll



Generalized Map of Pearl and Hermes Atoll

Appendix 10: Station/Waypoint List (coordinates in Latitude, Longitude: decimal degrees)

See attached file

Appendix 11: Dive plans

See Attached File

Appendix 12: Program Equipment List

(See Attached File)

Appendix 13: Section IV.B. Hazardous Materials Inventory

Inventory of Hazardous Materials

Common Name	Quantity	Notes	Trained Individual	Spill Control
10,000X SYBR Gold (nucleic acid stain)	1 ml	Stored in sealed container in Scientific refrigerator	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	NT
25% Glutaraldehyde (disinfectant/fixative)	10 ml	Stored in sealed plastic container in Scientific refrigerator	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	F
32% Paraformaldehyde (disinfectant/fixative)	10 ml	Stored in Hazmat cabinet in wetlab	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	F
5% Hydrochloric Acid	5 L	Stored in sealed plastic bucket in wet-lab	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	A
95% Ethyl Alcohol (190 proof)	240 L	Highly Volatile, Flammable Bulk stored in carboys in port fantail release rack Daily use quantity (19 L carboy) stored in ARMS lab in secondary containment. Preserved samples stored in secondary containment in Scientific freezer.	Bulk: Ship's Chief Engineer Daily use and sample quantities: (I) Andrew Gray (II/III) Kerry Reardon	Ship SOP (bulk) AL (Daily use)
Aquamend underwater Epoxy	200sticks/ 63.5 kgs	Stored in Stainless Steel lockers on Fantail and Dry Lab	(I)Noah Pomeroy (II/III) Thomas Oliver	NT
Chloroform	70 ml	Stored in Hazmat cabinet in wet-lab within sealed metal secondary container and padding	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	F
Commercial Bleach	75 L	Stored in ARMS lab in secondary containment	(I) Joao Garriques (II-III) Kerry Reardon	F

Common Name	Quantity	Notes	Trained Individual	Spill Control
DAPI Nucleic Acid Stain	500 μg	Stored in sealed plastic container in Scientific fridge	(I) Brandon Reyes (II) Ty Roach (III)Mark Little	NT
DMSO Buffer	10 L	Stored in Hazmat cabinet in wetlab	(II-III) Kerry Reardon	F
Dynamic Descaler (Aqueous Hydrogen Chloride)	57 L	Biodegradable; neutralize with copious amounts of water Inventory stored in ½ pallet tote on fantail In-use quantity held in ½ pallet tote behind crane	(II-III) Kerry Reardon	Α
Gasoline, unleaded	3.7 kL	Volatile, Flammable Stored in ship's fantail tanks	Ship's Chief Engineer	Ship SOP
Liquid Nitrogen	12 L	Stored in wetlab Scientific dewar	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	LN
Mercuric Chloride (Saturated solution, 7g HgCl ₂ in 60 ml of deionized water)	60 ml	Scientific samples consist of 200 µl HgCl ₂ solution in 500 ml of seawater	(I/II)Noah Pomeroy (III) Thomas Oliver	М
Methanol 95%	1L	Stored in wet lab	(II) Amanda Heidt	AL
Methanol 70%	3L	Daily use quantity (19L carboy) stored in Microbial lab in secondary containment Preserved samples stores in secondary containment in Microbial Fridge	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	AL
Mytomycin (antibiotic)	lmg	Stored in program mini- fridge	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	P

Common Name	Quantity	Notes	Trained Individual	Spill Control *
Pool Time Shock XtraBlue 6 in 1 Pool Shock		Corrosive	(I) Joao Garriques	
(primarily Sodium Dichloro-s-Triazinetrione- Dihydrate)	4.6 kg	Contained in ten 1-lb bags within lidded 5-gal bucket on Grated Deck	(II/III)Kerry Reardon	Р
RNAlater	1000ml	Stored in undersink cabinet	(I) Brandon Reyes (II) Ty Roach and Amanda Hedit (III) Mark Little	NT
Scothcast Flame Retardant Compound 2130 (Insulating resin)	225ml	Stored undersink cabinet	(I/II/III) Andrew Gray	NT
Sodium Hydroxide (NaOH) pellets	500 g	Highly caustic Stored in Hazmat cabinet in wetlab	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	В
TCBS agar culture plate (Thiosulfate citrate bile salts sucrose agar)	1L	Stored in microbial fridge	(I) Brandon Reyes (II) Ty Roach (III) Mark Little	NT
Weld-On PVC glue, solvent	273ml	Flammable Stores in Hazmat cabinet in wetlab	(I/II/III) Andrew Gray	NT
Z Fix (buffered zinc formalin fixative)	3.8L	Toxic Stored in Hazmat cabinet in wetlab	(I/II) Bernardo Vargas-Ángel (III) Hatsue Bailey	F

Appendix 14: Section IV.C. Chemical Safety and Spill Response Procedures

*Spill Control Key

A: Acids

- Wear appropriate personal protective equipment (PPE) and clothing during clean-up.
- Keep upwind. Keep out of low areas.
- Ventilate closed spaces before entering them.
- Stop the flow of material, if this is without risk. Dike the spilled material, where this is possible.
- Large Spills: Dike ahead of spill for containment. Use a non-combustible material like vermiculite, sand or earth to soak up the product and place into a container for later disposal.
- **Small Spills**: Wipe up with absorbent material (e.g. cloth, fleece). Clean surface thoroughly to remove residual contamination.
- Never return spills to original containers for re-use.
- Neutralize spill area and washings with soda ash or lime. Collect in a non-combustible container for disposal.
- J. T. Baker NEUTRASORB® acid neutralizers are recommended for spills of this type.

AL: Alcohols (daily use quantities)

- Extinguish smoking lamp. Remove all sources of ignition.
- Wear appropriate PPE and clothing during clean-up.
- Ventilate closed spaces before entering them.
- Use absorbent socks to surround spills or to divert fluid flow.
- Use vermiculite or kitty litter to soak up and absorb fluid.
- Do not use combustible materials, such as saw dust.
- Use absorbent pads/diapers to wipe up the spill or a dust pan to sweep up vermiculite/kitty litter.
- Place used absorbents in plastic bag or pail.
- Clean surface thoroughly to remove residual contamination.
- Bags containing used absorbents will be properly disposed of once the ship returns to port.

B: Bases

- Wear appropriate PPE and clothing during clean-up.
- Keep upwind. Keep out of low areas.
- Ventilate closed spaces before entering them.
- Stop the flow of material, if this is without risk. Dike the spilled material, where this is possible.
- Large Spills: Dike ahead of spill for containment. Use a non-combustible material like vermiculite, sand or earth to soak up the product and place into a container for later disposal.
- Small Spills: Wipe up with absorbent material (e.g. cloth, fleece). Clean surface thoroughly to remove residual contamination.
- Never return spills to original containers for re-use.
- Neutralize spill area and washings with product such as Grainger Base Eater Spill Kit.
 Collect in a non-combustible container for prompt disposal.

F: Fixatives/Formalin/Formaldehyde

- Wear appropriate PPE (gloves, goggles, breathing mask).
- Ventilate area of leak or spill. Remove all sources of ignition.
- Isolate hazard area. Keep unnecessary and unprotected personnel from entering.
- Contain and recover liquid when possible.
- Use non-sparking tools and equipment. Collect liquid in an appropriate container or absorb with an inert material (e.g., vermiculite, kitty litter, absorbent pads), and place in a chemical waste container. A dust pan and plastic bags are available to aid in cleanup and disposal.
- Do not use combustible materials, such as saw dust.

LN: Liquid Nitrogen

- Wear appropriate PPE (close-toed shoes, cryogloves, goggles, long-sleeved and long-legged clothes are of particular importance).
- Ventilate area.
- Contain spill where safe to do so.
- Nitrogen is more harmful in its liquid state than in its gaseous state, in a well-ventilated area. Minimally handle or interfere with the spilled LN, and allow it to sublimate off after restricting personnel access to the contained spill area under well maintained ventilation.

M: Mercury

- Wear appropriate PPE and clothing during clean-up (a minimum of nitrile gloves and eyewear).
- Stop the flow of fluid by using absorbent material (e.g. cloth, fleece, paper) to dike and soak up the spilled solution.
- Use Mercury Spill Kit if need be.
- Sprinkle area with sulfur or calcium polysulfide to suppress mercury.
- Contaminated area should be wiped with water dampened absorbent material, until one feels the area is sufficiently clean.
- Pick up used absorbents and place in a suitable container for reclamation or disposal in a method that does not generate dust
- If all the HgCl₂ solution from a spill is not wiped up, then potential exists for the HgCl₂ to come out of solution, and HgCl₂ crystals are more problematic (from a health perspective) than HgCl₂ in solution.
- All PPE and absorbent material contaminated with HgCl₂ should be contained in a ziptop bag labeled "HgCl₂ Waste," kept within the ship's HAZMAT locker, and properly disposed of once the ship returns to port.
- The concentration of HgCl₂ in solution, once mixed with copious amounts of fresh/salt water, will rapidly dilute the concentration of HgCl₂ relieving concern for further contamination by effluent, as concentrations will be below environmental toxicity, see MSDS for toxicological information.
- Areas of skin contact should be thoroughly rinsed under fresh/salt water for a minimum of 15 minutes.
- HgCl₂ solution contact with eyes/ingestion should be immediately addressed by the ship's doctor, rinse eyes for a minimum of 15 minutes.

NT: Non-toxic

• Wear appropriate PPE and clothing during clean-up.

- Ventilate area.
- Contain spill where safe to do so.
- Absorb liquid with paper towel while wearing gloves; place waste in sealed plastic container until processed on land.

P: Powdered Chlorine Salts

- Wear appropriate PPE (gloves, eyewear, dust mask, etc.) and clothing during clean-up.
- Ventilate area.
- Keep upwind. Avoid inhalation of salts, granules or dust.
- Large Spills: Sweep or scoop all spilled material, contaminated soil or other materials and place into clean, dry containers for disposal. Do not close containers containing wet or damp material. If wet or damp, container should be left open in a well-ventilated area to disperse any hazardous gases that may form. Once cleaned, neutralize/flood the spill area with large amounts of water as appropriate.
- Small Spills: Sweep or scoop up spilled material and add it to dive gear "disinfectant" rinse tote if available and full of water. If dive gear "disinfectant" rinse tote is not available, dispose of collected material into a clean, dry container. Once cleaned, neutralize/flood spill area with large amounts of water as appropriate.
- Never return spills to original containers for re-use.

GNF: Non-Flammable gases (ex. Oxygen, Nitrox)

- Ventilate area.
- Isolate cylinder and move exposed personnel to fresh air.

Inventory of Spill Kit Supplies

Product Name	Amount	Chemicals useful against	Amount of clean up possible
Absorbent pads	20	A, AL, F, M	~4 L
Base Eater	Large Kit	В	~19 L
Dust pan	1 set	A, F, P	n/a
Goggles	5 pair	A, F, G, LN, M	n/a
Kitty litter	5.4 kg	A, AL, F	~4 L
Nitrile gloves	12 pairs	A, F, G, LN, M	n/a
NEUTRASORB®	3.2 kg	A	Varies with acid concentration
Plastic bags	5	A, AL, F, P	~4 L (each)
Vermiculite	2.5 kg	AL, F, NT, M	~6 L of chemical spilled
Vinyl gloves	20 pairs	A, F, G	n/a