

Morphometric forms, biovolume and cellular carbon content of dinoflagellates from polluted waters on the Karachi coast, Pakistan

*Sonia Munir¹, Zaib-un-nisa Burhan¹, Tahira Naz¹, Steve L. Morton² & Pirzada Jamal Ahmed Siddiqui¹

¹ Centre of Excellence in Marine Biology, University of Karachi-75270-Karachi Pakistan.

² National Oceanic Atmospheric Administration, 219 Fort Johnson Road, Charleston SC-29412 USA.

*[E.Mail:soniaku2003@yahoo.com]

Received 21 November 2013; revised 25 February

Present study reports new information on the biovolume and carbon biomass estimates for dinoflagellates from Manora Channel, Karachi coast, Pakistan. Biovolume per cell was calculated using the geometric shape of dinoflagellates at species level. Both thecate and athecate species were examined under light and scanning electron microscope. A total of 45 species were measured and their cell size was ranged between 20-450 μm . Geometric forms of the species were seven classed into as ellipsoidal, spherical, double cone shape, prolate sphere, cone and half sphere, "cone+3 cylinder" shape, "ellipsoidal + 2 cone+cylinder" shape, "cylinder+ cone" shaped. Total biovolume ranged from 3.743×10^3 to $2.2 \times 10^5 \mu\text{m}^3 \text{cell}^{-1}$ and estimated cellular carbon content per cell ranged from 397×10^2 to $26.5 \times 10^4 \text{pg C cell}^{-1}$. Carbon and biovolume relationship was significant for thecate species which can thus be used for carbon flux studies.

[**Keywords:** *Biovolume*, Carbon biomass, Dinoflagellates, Pakistan]

*Corresponding author

Introduction

Dinoflagellates ranging from (5–2000 μm in diameter) are a significant proportion of total phytoplankton biomass in oceanic ecosystems¹. Some of these aquatic microorganism are toxic and harmful, and can then negatively impact water quality and cause public health risks as well as economic loss to fisheries resources^{2,3}. Dinoflagellates also contribute to carbon flux and thereby provide energy to deep-water and demersal fish^{3,4}. To determine the ecology as fully as possible, abundance, cell biovolume, cell biomass, species succession, growth and cell loss processes should be estimated^{4,5,6,7}. Sizes and shapes plays important in numerical censuses of abundance, grazing, cellular processes and other metabolic process. Coastal physical forcing (wind, circulation, horizontal transport and eddy turbulence) affects cell density and surface area to volume ratio⁴. Surface areas to volume ratio have been used to estimate the contribution of small cells, which generally contribute a significant fraction of the total biomass, rather than large cells alone⁸. Small species have a reduced boundary layer and larger surface area per unit

volume, which allows them to acquire relatively more nutrients⁹.

Carbon fluxes estimation requires data on cell biovolume^{5,6}. Several techniques have been employed to calculate phytoplankton biovolume, such as optical and image analysis software¹⁰, electronic particle counting¹¹, flow cytometry^{12,13}, and computer tomography such as holographic Scanning techniques¹⁴. Cell biovolume and cellular carbon content can be estimated from chlorophyll *a*, ATP, cell-nitrogen concentration and depth variation^{15, 16,17}, as well as from geometric shape measurement^{5,6, 18,19}.

Geometric shape models and biovolume estimates for dinoflagellates and other phytoplankters have been made for Chinese Sea^{6, 20}, the Baltic Sea²¹, the Mediterranean Sea⁷ and India²². Biovolume and carbon biomass for dinoflagellates reported in this paper provide original baseline data for the Pakistan coast, northern Arabian Sea.

Materials and Methods

Phytoplankton samples were collected from two sites in the Manora Channel, off the coast from Karachi, Pakistan (Fig. 1). Bimonthly samples (n=180) were collected using a 1.7 L Niskin bottle from May 2002 to July 2003, and fixed with Lugol's preservative. Samples were examined by light microscope using the Utermöhl method²³ and species identifications were confirmed by scanning and fluorescence microscopy.

Species' biovolumes (as $\mu\text{m}^3 \text{ cell}^{-1}$) were calculated from assigned geometric shape Sun and Liu 2003, and at least 30 to 300 cells of each species were measured⁴. The cellular carbon content (pg C cell^{-1}) was calculated from the formula of Eppley (1970):

$$\text{Log pg C cell}^{-1} = \text{Power } 10^{0.94 \times \text{Log V}(\mu\text{m}^3) - 0.60}$$

Results

Community structure and morphometric shapes

At both stations, a total of 45 species of 20-450 μm were divided into sizes classes. Seven geometric forms, determined from the measurements under light/scanning electron microscopy, were used to calculate biovolume.

Ellipsoidal-shaped species were *Akashiwo sanguinea*; *Alexandrium ostenfeldii*; *A. tamarensis*; *A. tamiyavanichi*; *Dinophysis caudata*; *D. acuminata*; *Gymnodinium* sp.; *Gyrodinium spirale*; *Gyrodinium* sp.; *Prorocentrum arcuatum*; *P. micans*; *P. gracile*; *P. donghaiense*; *P. minimum*; *Phyrophacus steinii* (Plate I, Fig 2). Double-cone-shaped species were *Ceratium fusus*; *C. inflatum*; *Gonyaulax spinifera*; *Protoperidinium steinii*; *P. divergens*; *P. depressum* (Plate II, Fig 3), a cone-and-half-sphere shaped species was *Scrippsiella trochoidea*, and spherical species were *Protoceratium reticulatum*; *P. balticum* (Plate III, Fig 4) and complex-shaped species were 2 cones + 3 cylinders *C. furca*, *C. lineatum*; ellipsoidal + 2 cones + 1 cylinder was *C. macroceros*, (Plate IV, Fig 5), while a prolate sphere shaped species (*Cochlodinium* cf. *fulvescens*).

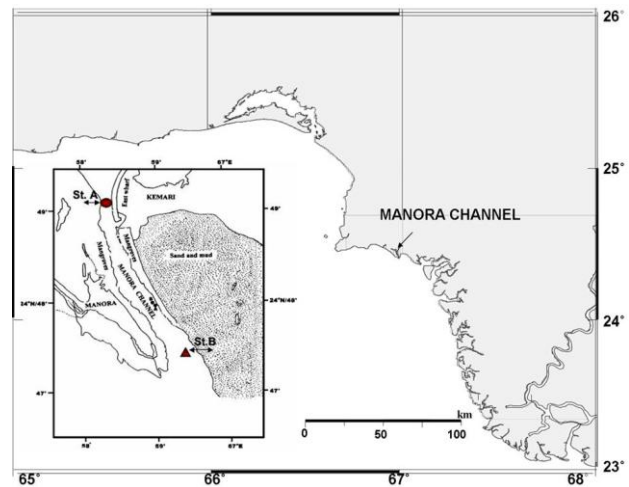


Fig. 1. Map of Karachi coast, Pakistan. Locations indicated by circle (red dot) for St. A and triangle (red triangle) for St. B.

Biovolume and carbon-biomass

Biovolumes and carbon content for all species are listed in Table 1. Biovolumes of species ranged between 2535 to 220893 $\mu\text{m}^3 \text{ cell}^{-1}$ and carbon content ranged from 397 to 33016 pg C cell^{-1} . A total of 31 autotrophic/mixotrophic and 14 heterotrophic species had cell densities ranging from 20 cells/L to 48,166 cells/L (Table 1). Biovolume and carbon content of the autotrophic/mixotrophic species ranged from 4088 to 105500 μm^3 and 623 to 16684 pg C cells^{-1} , respectively, at station A and 2535 to 147262 μm^3 and 397 to 18114 pg C cells^{-1} , respectively, at station B (Table 1). Biovolume and carbon content of heterotrophic species at Station A had values ranging from 3151 to 220893 μm^3 and 571 to 33916 pg C cells^{-1} , respectively, and 5473 to 149500 μm^3 and 810 to 23316 pg C cells^{-1} , respectively, at station B (Table 1). Lowest carbon per cell was observed for *P. minimum*/*P. balticum* at station B and highest carbon per cells was observed for *P. depressum* at station A.

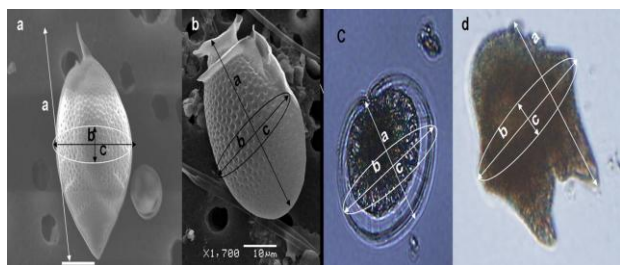


Fig. 2, Plate I. LM and SEM's of ellipsoidal shape *Prorocentrum micans* (A), *Dinophysis acuminata* (B), *Pyrophacus steinii* (C), *Akashiwo sanghainea* (D) represented by length (a), breadth (b), depth (c) and biovolume V: $3.14/6 \cdot a \cdot b \cdot c$ (Sun and Liu 2003).

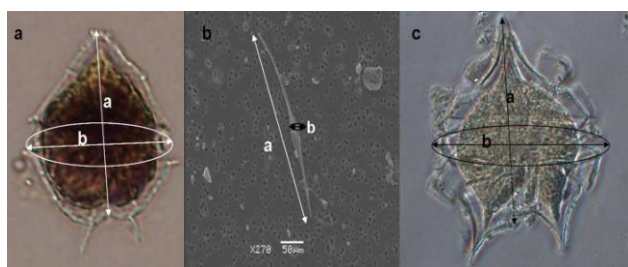


Fig. 3, Plate II. Light and scanning electron microscopy, double cone shape species, *Gonayaulax spinifera* (A), *Ceratium fusus* (B), *Protoperidinium divergens* (C) represented by length (a), width (b), depth (c). Formula computed by biovolume V: $\pi/12 \cdot a \cdot b^2$. Scale bar= 10 μm , 50 μm

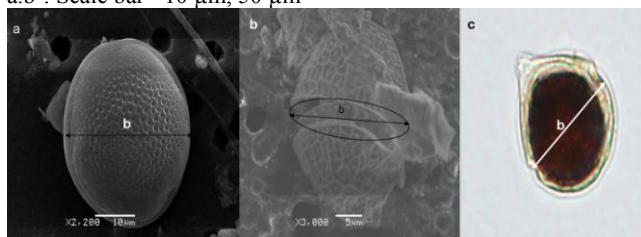


Fig. 4, Plate III. SEMs of sphere shaped *Prorocentrum compressum* (A), *Protoperidinium reticulatum* (B) represented by geometric calculation diameter (b) and biovolume computed by V: $\pi/6 \cdot a^3$ and , cone and half shape *Scripsiella trochoidea* (C) and biovolume computed by V: $3.14/4 \cdot a \cdot b^2$

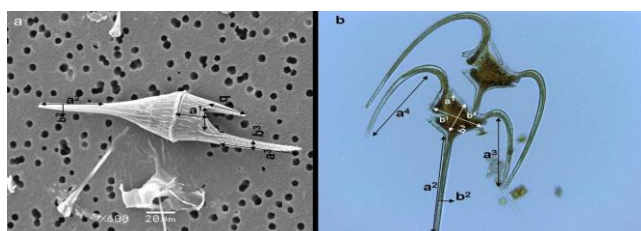


Fig. 5, Plate IV. Lm and SEMs of complex shape species *Ceratium furca* (Cone+3 cylinder) by geometric calculation by formula $V: 3.14/4 \cdot a_2 \cdot b_2^2 + 3.14/2 \cdot a_3 \cdot b_3^2 + 3.14/12 \cdot a_1 \cdot (b_1^2 + b_1 \cdot b_2 + b_2^2)$ and *C. macroceros* (Ellipsoid + 2 cones+ cylinder) represented by V: $3.14/4 \cdot a_2 \cdot b_2^2 + 3.14/12 \cdot (a_3 + a_4)$.

Biovolume and carbon biomass calculated on single cell basis of all species studied had significant

relationship (linear coefficient relationship). A total of 33 thecates and 4 athecates species showed significant correlation with carbon biomass (Figures 6 & 7), although thecates species appeared to have slightly higher calculated carbon content at Stn A ($R^2 = 0.993$, $P > 0.0001$) and Stn B ($R^2 = 0.978$, $P > 0.0001$) than athecate species ($R^2 = 0.82$, Stn A) and ($R^2 = 0.99$, $P > 0.01$, Stn B). It is interesting to note that smaller cells contribute more carbon biomass per liter of water compared to large cells.

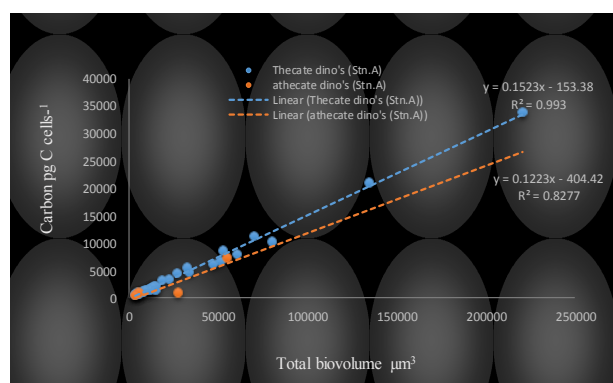


Fig. 6, Linear regression between biovolume and carbon per cells of thecate and athecate dino's at Station A.

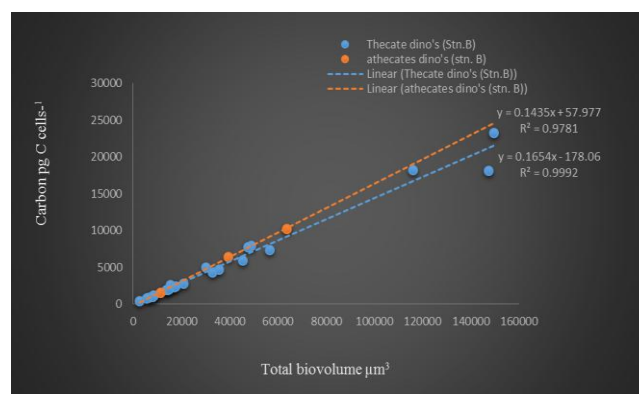


Fig. 7, Linear regression between biovolume and carbon per cells of thecate and athecate dino's at Station B.

Table 1. The biovolume and carbon content estimation for dinoflagellates from Karachi waters.

Autotrophic/ Mixotrophic	Station A			Station B		
	Max cells/L	Biovolume μm^3	Carbon Pg C cells^{-1}	Max cells/L	Biovolume μm^3	Carbon Pg C cells^{-1}
<i>Akashiwo sanguinea</i>	40	27750	1000	-	-	-
<i>Alexandrium ostenfeldii</i>	1440	8181	1197	2120	14137	2001
<i>A. tamarense</i>	520	5236	787	-	-	-
<i>A. tamiyavanichi</i>	225	14392	2035	-	-	-
<i>Ceratium furca</i>	75	12371	1765	640	20697	2864
<i>C. fusus</i>	80	51516	6749	906	32598	4389
<i>C. inflantum</i>	80	46952	6185	106	45363	5988
<i>C. kofoid</i>	-	-	-	40	15332	2619
<i>C. lineatum</i>	106	14729	2080	80	17338	2425
<i>C. macroceros var macroceros</i>	-	-	-	160	6313	938
<i>C. massiliense</i>	-	-	-	40	30050	4997
<i>C. pelluchellum</i>	-	-	-	40	48782	7956
<i>C. longipes</i>	-	-	-	40	47476	7752
<i>Cochlodinium</i> sp	160	5724	856	926	39208	6451
<i>Dinophysis caudata</i>	94	80799	10303	392	35540	4761
<i>D. acuminata</i>	80	34088	4634	-	-	-
<i>D. miles</i>	80	15228	1382	-	-	-
<i>D. fortii</i>	40	52675	8565	-	-	-
<i>D. infundibula</i>	20	12828	1826	-	-	-
<i>Gonyaulax spinifera</i>	80	4088	623	666	8356	1221
<i>G.polygramma</i>	20	4500	807	-	-	-
<i>G.verior</i>	60	27040	4516	-	-	-
<i>Prorocentrum micans</i>	545	10996	1580	1000	16965	2376
<i>P. minimum/ P. balticum</i>	1400	9948	1439	4500	2535	397
<i>P. gracile/P.sigmoidea</i>	160	15134	2134	250	13614	1932
<i>P. arcuatum</i>	233	13614	1932	986	14632	2067
<i>P. donghaiense</i>	3000	5320	799	6060	8181	1197
<i>P. compressum</i>	160	8181	1197	40	147262	18114
<i>Pyrophacus steinii</i>	140	61324	7950	840	56566	7369
<i>Scrippseilla trochoidea</i>	1153	11611	1419	1120	7942	1047
<i>Protoceratium reticulatum</i>	40	22437	3375	-	-	-
Heterotrophic						
<i>D. rotundatum</i>	80	9533	1382	-	-	-
<i>Gyrodinium</i> sp	14053	3743	574	48166	11240	1613
<i>G. spirale</i>	187	55127	7244	140	63689	10277
<i>Gymnodinium catenatum</i>	106	18750	3177	-	-	-
<i>Oblea rotunda</i>	100	70685	11359	-	-	-
<i>Protoperdinium steinii</i>	313	3983	608	780	5403	810
<i>P. divergens</i>	50	87967	14013	50	-	18283
<i>P. depressum</i>	40	220893	33916	-	-	-
<i>P. curvipes</i>	20	134628	21084	-	-	-
<i>P cf. minutum</i>	20	3515	636	-	-	-
<i>P. subinermis</i>	20	18840	3192	-	-	-
<i>P. granii</i>	20	5272	939	-	-	-
<i>P. leonis</i>	40	32500	5387	-	-	-
<i>P. oblongum</i>	-	-	-	40	149500	23316

Table 2 shows carbon biomass values in microgram per Liter in different cells size classes. Species with cell size between 20-45 μm contribute high carbon biomass ranging from 0.04 to 8.06 $\mu\text{g CL}^{-1}$ at Stn A and 0.63 to 77.4 $\mu\text{g CL}^{-1}$ at Stn B; cells with sizes 45-60 μm had low carbon biomass at Stn A (0.04 to 1.13 $\mu\text{g CL}^{-1}$) and high at Stn B (0.48 to 6.80 $\mu\text{g CL}^{-1}$); cells having 100-150 μm had carbon biomass ranged from 0.01 to 1.35 $\mu\text{g CL}^{-1}$ at station A and 0.10 to 1.86 $\mu\text{g CL}^{-1}$ at station B; Cell sizes between 200-450 μm had carbon biomass ranging from 0.49 to 0.53 $\mu\text{g CL}^{-1}$ at station A and 0.19 to 3.0 $\mu\text{g CL}^{-1}$ at station B.

Discussion

This study provides new data on biovolume and carbon biomass of 45 marine dinoflagellate species from Pakistani coastal waters. Linear measurement of simple shapes, such as, ellipsoidal, spherical, double cone form, prolate ellipses, cones and half spheres were easily measured, but some species are more complex in shape, such as, “cone+3 cylinders”, “ellipsoidal + 2 cones + cylinder”, or “cylinder + cone” used for *Ceratium* sp. required more measurements²⁰. A double cone shape has been recommended for the measurement of *C. fusus* and *C. inflatum* (Sun Jun, personal communication)..

In present study, biovolume and carbon content of dinoflagellates was recorded on single cell basis fall well within the range reported earlier^{5,24,25,26}. Contribution of carbon biomass in a litter of water appeared to be controlled by the abundance of dinoflagellates species which vary with geographical location and other nutritional and environmental conditions. For example, heterotrophic species have been reported to have high carbon biomass (5 to 438 $\mu\text{g C L}^{-1}$)²⁷, (0.5-272 $\mu\text{g CL}^{-1}$)²⁵. It is interesting to note that mixotrophs from the same area (North Sea) had lower range (0.2 to 455 $\mu\text{g CL}^{-1}$)²⁵. Carbon contribution in Pakistani coastal waters although lower but lie within the range reported in previous studies^{25, 27}. As small dinoflagellates species are abundant in coastal waters of Pakistan, they contribute high amount of carbon compare to larger species regardless of nutritional modes, for example, both *P. minimum* (mixotroph) and *Gyrodinium* sp. (heterotroph) were observed in abundance having small volume but contribute more carbon (77 $\mu\text{g CL}^{-1}$

¹) which is good agreement with the results of same size

Table 2. The total carbon biomass microgram per Liter values (μg

Size classes	Taxa	Station A	Station B
13-18 μm	<i>Prorocentrum minimum</i> / <i>P. balticum</i>	0.86	1.79
20-45 μm	<i>Alexandrium ostenfeldii</i>	1.72	4.24
	<i>A. tamarense</i>	0.40	-
	<i>Dinophysis rotundatum</i>	0.16	-
	<i>Gonyaulax spinifera</i>	0.04	0.81
	<i>G. polygramma</i>	0.01	-
	<i>Gyrodinium</i> sp	8.06	77.6
	<i>Gymnodinium catenatum</i>	0.33	-
	<i>P. donghaiense</i>	2.39	7.25
	<i>P. compressum</i>	0.19	1.45
	<i>Protopteridinium steinii</i>	0.19	0.63
45-60 μm	<i>P. curvipes</i>	0.42	-
	<i>P. cf. minutum</i>	0.01	-
	<i>P. subinerme</i>	0.06	-
	<i>P. granii</i>	0.01	-
	<i>Protoceratium reticulatum</i>	0.13	-
	<i>Scrippseilla trochoidea</i>	1.63	1.17
	<i>A. tamiyaunichivi</i>	0.45	-
	<i>Akashiwo sanguinea</i>	0.04	-
	<i>Cochlodinium</i> sp	0.13	5.97
	<i>D. fortii</i>	0.34	-
	<i>D. infundibula</i>	0.04	-
	<i>G. verior</i>	0.27	-
	<i>P. micans</i>	0.86	2.38
	<i>P. gracile</i> / <i>P. sigmoides</i>	0.34	0.48
	<i>P. arcuatum</i>	0.45	2.03
75-150 μm	<i>Pyrophacus steinii</i>	1.11	6.18
	<i>Oblea rotunda</i>	1.13	-
	<i>Ceratium furca</i>	0.13	1.83
	<i>C. kofoid</i>	-	0.1
	<i>C. lineatum</i>	0.22	0.19
	<i>C. longipes</i>	0.01	-
	<i>C. macroceros</i> var <i>macroceros</i>	-	0.15
	<i>Dinophysis caudate</i>	0.96	1.86
	<i>D. miles</i>	0.05	-
	<i>D. acuminata</i>	0.37	-
200-450 μm	<i>Gyrodinium spirale</i>	-	1.44
	<i>Protopteridinium divergens</i>	0.70	0.91
	<i>P. depressum</i>	1.35	-
	<i>P. oblongum</i>	1.35	0.93
	<i>Ceratium fusus</i>	0.53	3.9
	<i>C. inflatum</i>	0.49	0.63
	<i>C. massiliense</i>	-	0.19

(CL^{-1}) of different size classes dinoflagellates estimated from Station A and Station B.

species (*Gyrodinium* sp) reported from Mediterranean waters²⁵. Carbon contribution of large size dinoflagellates in Pakistani waters is much lower compared to other studies^{6, 24, 25, 26, 28} primarily due to low cells densities of large size dinoflagellates species (Table 1).

The only species, *Gymnodinium* sp., has been studied for assessment of carbon biomass from eastern part of the Arabian Sea bordering India²². Dinoflagellates from Pakistani waters appear to have more cellular carbon (269-39116 pg C Cell⁻¹) as oppose to carbon levels reported by Ravi Kumar et al.²². Therefore, results obtained in the present study (269-39116 pg C Cell⁻¹ or 0.33-77.42 µg CL⁻¹) fall well within the range of carbon biomass reported from various waters (0.1 to 3.5 µg C L⁻¹)²⁹ and (0.05 to 2.0 µg C L⁻¹)³⁰. Variation in carbon biomass is regulated by species composition, their cell sizes and nutritional conditions. For example, coastal species appear to have more carbon compared to open water species²⁷. Similarly, thecate species have more carbon as oppose to athecate species⁵. In spite of their high biovolume, athecate species were less significant than the thecate species in terms of carbon contribution. Most of thecate species were small, but produce more carbon per unit volume per unit time⁵.

In the present study Lugol's fixed samples were used for the estimation of biovolume and carbon biomass. However, Lugol's fixation is reported to causes shrinkage of cells, for example, in cyanobacteria³¹, diatoms and dinoflagellates^{32,33} and ciliates^{34,35,36}. Although live samples have been used³³, but it is difficult to culture all species present in the natural waters.

Acknowledgments

Special thanks to Dr. Jun Sun (Tianjin University of Science and Technology No. 29, 300457, P. R China) for helpful comments.

References

- 1 Encyclopedia Britannica. Dinoflagellates (2011) <http://www.britannica.com/EBchecked/topic/163945/dinoflagellate>. © 2000 Britannica.com Inc.- 53.
- 2 Smayda T J. Harmful algal blooms their ecophysiology and general relevance to phytoplankton blooms in the sea. *Limnol. Oceanogr.*, 42 (1997a) 1137-1153.
- 3 Shumway S E. A review of the effects of algal blooms on shellfish and aquaculture. *J. World. Aquac. Soc.*, 21 (1990) 65-104.
- 4 Smayda T J. From phytoplankton to biomass. In: (Sournia A ed). *Phytoplankton Manual. Monographs on Oceanographic Methodology* 6. UNESCO, Paris, (1978) , pp 273-279.
- 5 Menden-Deuer S, Lessard E J. Carbon to volume relationship for dinoflagellates, diatoms and other protest plankton. *Limnol. Oceanogr.*, 45 (2000) 596-579.
- 6 Sun J, Liu D, & Qian S. 2000. Estimating biomass of phytoplankton in Jiaozhou Bay 1. Phytoplankton biomass estimated from cell volume and plasma volume. *Act. Ocean. Sin.*, 19(2) (2000) 97-110.
- 7 Vadrucchi MR, Cabrini M, & Basset A. Biovolume determination of phytoplankton guilds in transitional water ecosystems of Mediterranean Ecoregion. *Transit. Waters Bull.*, 2 (2007) 83-102.
- 8 Chisholm SW. Phytoplankton size. In : Falkowski, P.G, A.D, Woodhead (Eds), *Primary Productivity and Biogeochemical Cycles in the Sea*. Plenum Press, New York, (1992), pp 213-237.
- 9 Raven JA. The twelfth Tansley lecture, small is beautiful, the picoplankton. *Funct. Ecol.*, 12 (1998) 503-513.
- 10 Culverhouse, P, Williams R, & Reguera B. Do experts make mistakes? A comparison of human and machine identification of dinoflagellates. *Mar. Ecol. Prog. Ser.*, 247 (2003) 17-25.
- 11 Boyd C M, Johnson GW. Precision of size determination of resistive electronic particle counters. *J. Plank.Res.*, 17 (1995) 223-234.
- 12 Cunningham A, & Buonnacorsi G A Narrow-angle forward light scattering from individual algal cells implications for size and shape discrimination in flow cytometry. *J. Plank.Res.*, 14 (1992) 223-234.
- 13 Collier J L, & Campbell L. Flow cytometry in molecular aquatic ecology. *Hydrobiol.*, 33 (2000) 401.
- 14 Brown L.M, Gargantini I, Brown D J, Atkinson H J, Govindarajan J, Vanlerberghe GC. Computer- based image analysis for the automated counting and morphological description of microalgae in culture. *J. Appl.Phycol.*, 1 (1989) 211-225.
- 15 Eppley R W, Reid F M H & Strickland J D H. Estimates of phytoplankton crop size, growth rate and primary production. *Bull. Scripps Inst. Oceanogr.*, 17 (1970) 33-42.
- 16 Yamaguchi A, Watanabe Y, Ishida H, Harimoto T, Furusawa K, Suzuki S, Ishizaka J, Ikeda T, & Takahashi M M. Structure and size distribution of plankton communities down to the greater depths in the Western North Pacific Ocean. *Deep-Sea Res II*. 49 (2002a) 5513-5529.
- 17 Gin K Y H, Chisholm SW, Olson R J. Seasonal and depth variation in microbial size spectra at the Bermuda Atlantic time series station. *Deep-Sea Res.*, 46 (1999) 122-1245.
- 18 Hillebrand H, & Sommer U. Response of epilithic microphytobenthos of the Western Baltic Sea to *in situ* experiments with nutrient enrichment. *Mar. Ecol. Prog. Ser.*, 160 (1997) 35-46.
- 19 Hillebrand H, Durselen C D, Kirschtel D, Pollinger, D & Zohary T. Bio volume calculation for pelagic and benthic microalgae. *J. Phycol.*, 35 (1999) 403-424.
- 20 Sun J, & Liu D. Geometric models for calculating cell biovolume and surface area of phytoplankton. *J. Plank.Res.*, 25 (2003) 1331-1346.
- 21 Helsinki Commission. Phytoplankton species composition, abundance and biomass. HELCOM (2000). <http://sea.helcom.fi/manual/ anxc6.html>.

- 22 Ravi-Kumar M S, Ramaiah N, & Tang D. Morphometry and Cell volumes of Diatoms from a tropical estuary India. *Ind. Jr. Mar. Sci.*, 38 (2) (2009) 160-165.
- 23 Utermöhl H. Zur Vervollkommnung der quantitative Phytoplankton-Methodik. *Ass. Intern. Limnol. Théor.*, 9 (1958) 1-38.
- 24 Fanuko N, Valcic M. Phytoplankton composition and biomass of the northern Adriatic lagoon of Stella Maris, Croatia. *Acta. Bot. Croat.*, 68(1) (2009) 29-44.
- 25 Loder M J G, Kraberg AC, Aberle N, Peters S, & Wiltshire KH. Dinoflagellates and ciliates at Helgoland Roads, North Sea. *Helgol. Mar. Res.*, (2011) DOI 10.1007/s10152-010-0242-z
- 26 Garzion LM, Steinberg DK. Microzooplankton community composition along the Western Antarctic Peninsula. *Deep. Sea. Res. I.*, (2013) 77 36-49.
- 27 Ismael A A. Succession of heterotrophic and mixotrophic dinoflagellates as well as autotrophic microplankton in the harbour of Alexandria, Egypt. *J. Plankton Res.*, 25 (2) (2003) 193- 202
- 28 Gallegos C L, Jordan T E, Hedrick. SS. Long-term Dynamics of Phytoplankton in the Rhode River, Maryland (USA). *Estuaries.Coasts.*, (2009). DOI 10.1007/s12237-009-9172-x
- 29 Verity P G, Stoecker D K, Sieracki M E, Burkill P H, Edwards E S, & Tronzo C R. Abundance, biomass and distribution of heterotrophic dinoflagellates during the North Atlantic \spring bloom. *Deep. Sea. Res. II.*, 40 (1993) 227-244.
- 30 Lessard E J. The trophic role of heterotrophic dinoflagellates in diverse marine environments. *Mar.Micro. Food webs.*, 5 (1991) 49-58.
- 31 Hawkins P R, Holliday J, Kathuria A, Bowling L. Change in *Cyanobacterial* biovolume due to preservation by Lugol's Iodine. *Harmful Algae.*, 4 (6) (2005) 1033-1043.
- 32 Choi J W, & Stoecker D K . Effects of fixation on cell volume of marine phytoplankton protozoa. *Appl. Environ. Microbiol.*, 55 (1989) 1761-1765.
- 33 Menden-Deuer S, Lessard E J, & Satterberg J. Effect of preservation on dinoflagellate and diatom cell volume and consequences for carbon biomass predictions. *Mar. Ecol. Prog. Ser.*, 222 (2001) 41-50.
- 34 Throndsen J. Preservation and storage. In: (Sournia, A. (Ed.) *Phytoplankton Manual*. UNESCO, Paris, (1978), pp 69-74.
- 35 Karayanni H, Christaki U, Van Wambeke F, & Dalby A P. Evaluation of double formalin-Lugol's fixation in assessing number and biomass of ciliates An example of estimations at mesoscale in NE Atlantic. *J. Microbiol. Meth.*, 56 (2004) 349-358.
- 36 Leakey R J G, Archer S D, and Grey J. Microbial dynamics in coastal waters of East Antarctica bacterial production and nano flagellate grazing. *Mar. Ecol. Progr.Ser.*, (1996) 1423-17.